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(54) **Title: VISIBLE** LIGHT CHROMOPHORE **EXCITATION** FOR MICROORGANISM CONTROL

(57) Abstract: Visible light disinfection is a healthcare technology wherein violet light is used to inactivate pathogens such as bacteria, fungi, and viruses. The present invention overcomes the limitations of continuous irradiance in whole-room environments **by** pulse width modulation of the light sources and increasing the instantaneous irradiance while maintaining average irradiance and hence light are synchronized and the phase of the spectral components are offset in order to maximize synergistic or antagonistic responses to intracellular chromophore excitation.

VISIBLE LIGHT CHROMOPHORE EXCITATION FOR MICROORGANISM CONTROL

TECHNICAL FIELD

[0001] The subject matter of the present invention relates to the use of optical radiation to influence microbial activity. In particular, the optical radiation is visible, with one or more monochromatic wavelengths.

BACKGROUND

[0002] Visible light disinfection is a healthcare technology wherein violet light is used to inactivate pathogens such as bacteria, fungi, and viruses. Unlike germicidal lamps emitting ultraviolet-C radiation, visible light does not pose the considerable risks of photokeratitis ("snow blindness") and erythema ("sunburn") for room occupants. With the availability of inexpensive and efficient violet LEDs (light-emitting diodes), the technology is being adopted **by** major luminaire manufacturers for everything from hospital operating theatres to residential kitchens. **[0003]** Despite appearances, visible light disinfection is **by** no means a new technology. The ancient Egyptians reported the health benefits of sun exposure some six millennia ago, while ancient Greek, Roman and Arabic cultures similarly recognized the therapeutic values of sunlight. The benefits of phototherapy (or more specifically heliotherapy) were given scientific support **by** Downes, L. R., and T. Blunt. **1877.** "Research on the Effect of Light Upon Bacteria and Other Organisms," Proc. Royal Society of London **26:488-500,** who reported that bacteria were inactivated **by** sunlight, and that violet-blue light was the most effective. [0004] The work of Downes and Blunt inspired Niels Finsen, a Danish medical researcher, to investigate further. After initial experiments in the early 1890s with natural light, he developed an apparatus using electric carbon arcs that would later become known as the "Finsen light" (Grzybowski, **A.,** and K. Pietrzak. 2012. "From Patient to Discoverer **-** Niels Ryberg Finsen **(1860 -** 1904) **-** the Founder of Phototherapy in Dermatology," Clinics in Dermatology 30:451 455).

[0005] Finsen began his experiments using common glass lenses to focus the electric arc emission, but knew from the work of Ward, H. M. 1894. "The Action of Light on Bacteria **- III,"** Philosophical Trans. Royal Society of London B: Biology **185:961-986,** and others that ultraviolet radiation was germicidal, and so he replaced these lenses with fused quartz. However, as reported **by** Moller, K. **I.,** et al. **2005.** "How Finsen's Light Cured Lupus vulgaris," Photodermatology, Photoimmunology **&** Photomedicine 21(3):118-124, Finsen used methylthioninium chloride ("methylene blue") in solution as a heat-absorbing filter. Likely unknown to Finsen, this dye also absorbs ultraviolet radiation with wavelengths shorter than 340

nm. The Finsen light therefore produced ultraviolet-A radiation and visible light, but no germicidal ultraviolet-B radiation.

[0006] In WO **2007/012875,** Anderson et al. disclosed a system and method for inactivating gram-positive bacteria such as methicillin-resistant *Staphylococcus aureus* (MRSA), coaugulese *negative Staphylococcus (CONS), Streptococcus, Enterococcus and Clostridium* species **by** means of exposure to visible light within the spectral range of 400 to **500** nm. It has since been determined (e.g., Tomb, R. M., et al. **2018.** "Review of the Comparative Susceptibility of Microbial Species to Photoinactivation Using **380-480** nm Violet-Blue Light, Photochemistry and Photobiology 94(3):445-458) that many different species of gram-positive and gram negative bacteria, fungi, yeasts, and some viruses can be inactivated **by** species-specific doses of visible light. The optimal wavelength has been determined to be 405 ± 5 nm for maximum disinfection efficacy (Maclean, M., et al. **2008.** "High-Intensity Narrow-Spectrum Light Inactivation and Wavelength Sensitivity of *Staphylococcus aureus,"* **FEMS** Microbiology Letters **285:227-232).** This system and method have recently been commercialized in the form of architectural luminaires that emit both blue light for disinfection and white light for general illumination, for example SpectraCleanTM luminaires as manufactured by Hubbell Lighting Components (Elgin, IL). These luminaires are typically equipped with arrays of 405 nm lighting emitting diodes to provide visible light disinfection radiation, and phosphor-coated "white light" **LED** arrays to provide general illumination.

[0007] A disadvantage of visible light disinfection for whole rooms such as hospital in-patient rooms and operating theaters, however, is that the irradiance levels on exposed surfaces are typically limited to **0.1** to *0.5* mW/cm2 (e.g., Halstead, F. **D.,** et al. **2019.** "Violet-Blue Light Arrays at 405 nm Exert Enhanced Antimicrobial Activity for Disinfection of Monomicrobial Nosocomial Biofilms," Applied and Environmental Microbiology **85(21): 1-16)** in order to comply with International Commission on Non-Ionizing Radiation Protection (ICNIRP) guidelines on exposure limits for incoherent visible radiation, and **by** building standards (e.g., California Energy Commission **2019** Building Energy Efficiency Standards for Residential and Nonresidential Buildings, Table 140.6-C) that mandate maximum allowable power per unit area for lighting (typically less than 10 W/m^2 , or 1.0 mW/cm^2).

[0008] A limit of, for example, **0.5** mW/cm2 , means that the surface will receive a dose of **25** J/cm2 of disinfecting blue light over a period of 14 hours, which is a typical daily operating time for hospital in-patient rooms. Such doses have been shown to inactivate on the order of **90** percent of some infectious bacterial pathogens commonly found in hospital settings (e.g., Bache, **S. E.,** et al. **2017,** "Universal Decontamination of Hospital Surfaces in an Occupied

Inpatient Room with a Continuous 405 nm Light Source," **J.** Hospital Infection **98(1):P67-73),** but there are many other infectious pathogens (including bacteria, yeasts, fungi, and viruses) that require considerably greater doses to achieve **90** percent inactivation, e.g., in TABLE 1 Common pathogen doses required for **90** percent reduction. (Source: Tomb et al. **2018.).**

TABLE 1

[0009] Gillespie, **J.** B., et al. **2017.** "Efficacy of Pulsed 405-nm Light-Emitting Diodes for Antimicrobial Photodynamic Inactivation: Effects of Intensity, Frequency, and Duty Cycle," Photomedicine and Laser Surgery **35(3):150-156,** reported that when exposing MRSA bacteria to **116** mW/cm2 of 405 nm radiation, varying the pulse width duty cycle from **25** to **100** percent had

little effect on the dose required to achieve the same degree of inactivation. The pulse **width** modulation (PWM) frequency was varied from **100** Hz to **10** kHz, with **1000** Hz showing the best performance, and with *35* percent energy savings for a *50* percent duty factor. The authors speculated that the cell porphyrins become saturated with continuous exposure, and that the off period of each cycle enables the absorbed light to generate ROS (reactive oxygen species) with fewer photons being absorbed unnecessarily.

[0010] Masson-Meyers, **D.,** et al. **2019.** "Pulsed 450 nm Blue Light Significantly Inactivates *Propionibacterium acnes* more than Continuous Wave Blue Light," **J.** Photochemistry and Photobiology B: Biology **202:111719,** reported that a duty cycle of **33** percent and a PWM frequency of **33** kHz was optimal for *450* nm radiation treating acne vulgaris caused **by** the *Propionibacterium acnes* bacterium. The experiments involved microLEDs mounted on flexible plastic sheets that were applied to the infected skin, but remarkably the required irradiance to achieve **100** percent eradication of the bacteria was 40 to **100** times less than previously reported results for *in vitro* experiments involving MRSA bacteria. However, this involved repeated treatments every four hours that were timed to coincide with the replication cycle of *P. acnes, as* opposed to single exposures of bacterial cultures in the laboratory.

[0011] Bumah et al. (2020) demonstrated that pulsed *450* nm blue light suppresses the formation of MRSA and *Propionibacterium acnes* bacteria in planktonic cultures and bacterial biofilms. Irradiances of 3mW/cm² and three doses of 7.6 J/cm² were sufficient to completely eradicate MRSA bacteria in solution, while 2 mW/cm² and 5 J/cm² were sufficient to eradicate *P. acnes* bacteria. Assuming that "eradication" means **99.9** percent inactivation, this implies that the doses required for 90 percent inactivation would be 1.9 J/cm² and 1.3 J/cm² respectively.

[0012] **A** common assumption is that visible light disinfection is the result of intracellular (endogenous) porphyrins absorbing and being excited **by** 405 nm photons, and then generating ROS such as singlet oxygen and hydrogen peroxide, which being cytotoxic inactivate the cells. However, Enwemeka, **C. S.** et al. 2021. "Pulsed Blue Light Inactivates Two Strains of Human Coronavirus," **J.** Photochemistry and Photobiology B: Biology **222:112282,** reported that pulsed blue light inactivates enveloped coronaviruses, so there are presumably other intracellular mechanisms involved, and which do not necessarily require 405 nm irradiation.

[0013] Most visible light disinfection studies conducted to date have focused on the spectral region of **385** nm to 420 nm. However, bactericidal effects of visible light have been demonstrated with wavelengths of up to 740 nm, as shown in **FIG.** 1 (from Hessling, M., et al. **2017.** "Photoinactivation of Bacteria **by** Endogenous Photosensitizers and Exposure to Visible Light of Different Wavelengths **- A** Review on Existing Data," **FEMS** Microbiology Letters

367). These are presumably due to the absorption characteristics of different endogenous photosensitizers that absorb photons and generate cytotoxic ROS that inactivate the bacterial or fungal cells. However, the dose required for inactivation generally increases exponentially with wavelength.

[0014] As noted **by** Gwynne, P. **J.,** and M. P. Gallagher. **2018.** "Light as a Broad-Spectrum Antimicrobial," Frontiers in Microbiology **9** Article **119,** however, much of the visible light spectrum has yet to be explored. There are known endogenous photosensitizers with absorptance peaks at 405 nm (porphyrins) and 450 nm (riboflavin), but there are other potential endogenous chromophores that may result in anomalous susceptibility for particular pathogen species or broad classes.

[0015] There is a similar situation with plant biology research, wherein many different photopigments, including chlorophylls, phytochromes, cryptochromes, carotenes, xanthophylls, anthocyanins, phototropins and more are being explored to elucidate their photobiological functions. More recently, it has been realized that there are complex interactions between these photopigments, and that the spectral power distribution of the photobiologically active radiation from **280** nm to **800** nm incident on the plants can be manipulated to achieve desirable results, such as producing compact plants, advancing or delaying the onset of flowering, and the production of secondary metabolites for food and medicinal purposes.

[0016] As reported **by** Gillespie et al. **(2017)** and Masson-Meyers et al. **(2019),** pulsed blue light appears to be more efficacious than continuous blue light, with the duty cycle appearing to have little effect on the required dose. From the perspective of excited chromophores generating ROS or initiating other photochemical and non-photochemical reactions, this implies chromophore saturation.

[0017] Pathak, **J.,** et al. **2018** "Mechanisms of Photoprotection in Cyanobacteria," in **A.** K. Mishra et al. (Eds.), Cyanobacteria: From Basic Science to Applications. Academic Press, reports that while cyanobacteria rely on photosynthetically active radiation (PAR) in order to perform photosynthesis, the photosynthetic process may become saturated at **high** light levels. (The PAR spectrum of 400 nm to **700** nm roughly corresponds to that of the visible light spectrum.)

[0018] It is chlorophyll **A** and B in photosynthetic organisms that absorb blue and red light respectively, assisted **by** various other accessory photopigments such as beta-carotene. Each such photopigment acts a chromophore, but what they have in common is that the organisms have evolved mechanisms to protect themselves from excess energy when the photosynthetic process becomes saturated, including nonphotochemical processes, fluorescence, and photochemistry.

These processes are triggered **by** the generation of ROS, which initially serve as signaling molecules, but whose overproduction can lead to oxidative stress and cell inactivation. **[0019]** The cited literature has established that visible light induces the overproduction of ROS in cells, leading to oxidative stress and, in sufficient doses, cellular damage and cell inactivation. As indicated **by** Hessling et al. **(2016)** in **FIG. 1,** the studies have examined the effects of quasimonochromatic LEDs from **360** nm to 740 nm. In addition, a few studies have examined the susceptibility of bacteria to broadband visible light. For example, Lipovsky, **A.,** et al. **2009.** "Sensitivity *of Staphylococcus aureus* Strains to Broadband Visible Light," Photochemistry and Photobiology *85:255-260* employed a halogen lamp with a filter having a spectral bandwidth of approximately 400 nm to **800** nm, while Rathnasinghe, R., et al. **2021.** "The Virucidal Effects of 405 nm Visible Light on SARS-CoV-2 and Influenza **A** Virus," Scientific Reports **11(11):19470** considered fluorescent and phosphor-coated light-emitting diode arrays emitting "white" light with varying correlated color temperatures (CCTs). Rutala, W. **A.,** et al. **2018.** "Antimicrobial Activity of a Continuous Visible Light Disinfection System" Infectious Control Hospital Epidemiology **39(10):1250-1258,** further investigated visible light disinfection luminaires with both blue and "white" light-emitting diode arrays.

[0020] Examples of interactions in plant biology include Meng, **Q.,** et al. **2019,** "Blue Radiation Interacts with Green Radiation to Influence Growth and Predominantly Controls Quality Attributes of Lettuce," **J.** American Society of Horticultural Science 145(2):1-13, and Zhen, **S.** and B. Bugbee. **2019.** "Far-red Photons Have Equivalent Efficiency to Traditional Photosynthetic Photons: Implications for Redefining Photosynthetically Active Radiation," Plant, Cell **&** Environment 2020:1-14, where far-red photons synergistically interact with shorter-wavelength photons to increase leaf photochemical efficiency. However, this relates to plant biology and not microbiology.

[0021] For whole-room disinfection, building standards mandate maximum irradiances, based on maximum allowable power per unit area. However, these limitations are based on average power with continuous irradiation. **If** the visible light for disinfection purposes is pulsed through pulsed modulation or similar techniques, the instantaneous irradiance during the on cycle is inversely proportional to the pulsed modulation duty factor. For example, with a duty factor of ten percent, the maximum allowable irradiance is ten times that of continuous irradiance.

[0022] This background is not intended, nor should be construed, to constitute prior art against the present invention.

SUMMARY

[0023] The inventor has recognized a need for a system and method of visible light disinfection that avoids the need for unachievably **high** irradiances and doses to achieve **90** percent inactivation or more of common pathogens. The activity of the pathogens, or more broadly microorganisms, may be inhibited or promoted with the application of visible light. The microorganisms are illuminated with two different wavelengths of pulsed, visible light. During the pulses of light, two or more chromophores in the microorganism are excited, which leads to the activity of the microorganism being either inhibited or promoted, depending on the microorganism and the chromophores. The pulses of the different wavelengths are at a frequency that is a whole number multiple of a fundamental frequency. There may be a phase difference between the two different pulse trains.

[0024] Disclosed is a method for exciting two different chromophores in a microorganism, the method comprising illuminating the microorganism with first light having a first pulse frequency and a first wavelength between **380** nm and *750* nm, while illuminating the microorganism with second light having a second pulse frequency and a second wavelength between **380** nm and *750* nm, wherein the second wavelength is different to the first wavelength, wherein the first and second pulse frequencies are integral multiples of a fundamental frequency, and wherein a cycle of the first light has a zero, positive or negative phase difference with respect to a corresponding cycle of the second light.

[0025] Also disclosed is a luminaire comprising: a first light emitting element **(LEE)** that emits first light at a first pulse frequency with a first wavelength between **380** nm and *750* nm; and a second **LEE** that emits second light at a second pulse frequency with a second wavelength between **380** nm and *750* nm; wherein the second wavelength is different to the first wavelength; wherein the first and second pulse frequencies are integral multiples of a fundamental frequency; and wherein a cycle of the first light has a controllable phase difference with respect to a corresponding cycle of the second light.

[0026] Further disclosed is a system for exciting two different chromophores in a microorganism, the system comprising: a luminaire having a first light emitting element **(LEE)** that emits first light at a first pulse frequency with a first wavelength between **380** nm and *750* nm, and a second **LEE** that emits second light at a second pulse frequency with a second wavelength between **380** nm and *750* nm, wherein the second wavelength is different to the first wavelength, wherein the first and second pulse frequencies are integral multiples of a fundamental frequency, and wherein a cycle of the first light has a controllable phase difference with respect to a corresponding cycle of the second light; a driver coupled to drive each **LEE**

with an independently variable duty factor and an independently variable peak radiant flux output and to control the phase difference; a sensor configured to detect a parameter of an environment of the microorganism; and a controller configured to adjust, based on the parameter, either or both of a peak radiant flux and a duty factor, in either or both of the first light and the second light, in order to maintain a predetermined rate of chromophore excitation in other microorganisms that are the same as said microorganism.

[0027] This summary provides a simplified, non-exhaustive introduction to some aspects of the invention, without delineating the scope of the invention.

BRIEF **DESCRIPTION** OF DRAWINGS

[0028] FIG. 1 shows prior art bacterial susceptibility versus wavelength.

[0029] FIG. 2 is a block diagram of a system, according to an embodiment of the present invention.

[0030] FIG. 3 shows two PWM intensity profiles according to an embodiment of the present invention.

[0031] FIG. 4 is a schematic diagram of a luminaire, according to an embodiment of the present invention.

[0032] FIG. 5 is a schematic diagram of a light-treatment chamber, according to an embodiment of the present invention.

[0033] FIG. 6 is a schematic diagram of a plant-illumination apparatus, according to an embodiment of the present invention.

[0034] **FIG. 7** is a schematic diagram of a water-disinfecting device, according to an embodiment of the present invention.

[0035] FIG. 8 is a flowchart of a method for inhibiting pathogens, according to an embodiment of the present invention.

DETAILED DESCRIPTION

A. Definitions

[0036] A "chromophore" is a chemical group capable of spectrally selective absorption of optical radiation resulting in coloration of certain organic compounds. The absorption is often incidental to any biological function of the chemical group.

[0037] A "driver" means an electrical device that regulates the pulsed modulation to LEEs. **[0038]** The term "general illumination" means lighting that is typically used in a home, office or factory, which is comfortable for the occupants in relation to the tasks they are performing. **[0039]** The term light-emitting element **("LEE")** is used to define any device that emits radiation

in any region or combination of regions of the electromagnetic spectrum for example, the visible region, infrared and/or ultraviolet region, when activated **by** applying a potential difference across **it** or passing a current through it, for example. Therefore, an **LEE** can have monochromatic, quasi-monochromatic, polychromatic or broadband spectral emission characteristics. Examples of LEEs include semiconductor, organic, or polymer/polymeric light emitting diodes, optically pumped phosphor coated light-emitting diodes, optically pumped nano-crystal light-emitting diodes or other similar devices as would be readily understood **by** a worker skilled in the art. Each **LEE** generates optical radiation with one or more monochromatic wavelengths in the range of 200 nm to **3000** nm. Specifically, each **LEE** may generate optical radiation in the visible spectrum with one or more wavelengths from about **380** nm to about **750** nm. An **LEE** may have multiple constituent LEEs of the same or different types. [0040] The term "microorganism" or "microbe" refers to bacteria, fungi, yeasts, viruses,

[0041] The term "monochromatic light" herein refers to light with a very narrow bandwidth,

bacteriophages and protozoans. Microorganisms may be beneficial or pathogenic.

such as typically emitted **by** a laser or laser diode.

[0042] The term "quasimonochromatic light" herein refers to light with a narrow bandwidth, such as light that is typically emitted **by** an **LED.**

[0043] The term "white light" is used herein in the layman's sense, to refer to light that appears to be white or close to white, rather than in the scientific sense in which it refers to the continuum of the visible spectrum.

[0044] Individual parts of the system may be referred to as modules. The term "module" can refer to any component in this invention and to any or all of the features of the invention without limitation. **A** module may be a software, firmware or hardware module, and may be or be located in a luminaire, driver, controller or other computing device.

[0045] The term "processor" is used to refer to any electronic circuit or group of circuits that perform calculations, and may include, for example, single or multicore processors, multiple processors, an **ASIC** (Application Specific Integrated Circuit), and dedicated circuits implemented, for example, on a reconfigurable device such as an **FPGA** (Field Programmable Gate Array). The processor performs the steps in the flowchart, whether they are explicitly described as being executed **by** the processor or whether the execution thereby is implicit due to the steps being described as performed **by** code or a module. The processor, **if** comprised of multiple processors, may be located together or geographically separate from each other. The term includes virtual processors and machine instances as in cloud computing or local virtualization, which are ultimately grounded in physical processors.

[0046] The terms "pulse modulation" and "pulse modulated" mean any technique utilizing a digitally modulated pulse to switch LEEs on and off at a desired frequency, and includes pulse width modulation (PWM), pulse code modulation (PCM), randomized PCM, pulse position modulation (PPM), pulse-amplitude modulation (PAM), pulse-duration modulation (PDM), and any other similar techniques or pulse modes as would be readily understood **by** a worker skilled in the art. One or more different pulse modulation techniques may be used concurrently or alternately on one or more serial or parallel channels. As would be readily understood **by** a worker skilled in the art the "on" periods relative to the "off" periods represent the pulse modulation duty factor.

[0047] Reactive oxygen species ("ROS") are **highly** reactive chemicals formed from dioxygen molecules (O_2) by means of one-electron excitation to produce either singlet oxygen $({}^1O_2)$ or one-electron reduction to produce a superoxide radical (O_2) , which may be a precursor to a peroxide radical (HO_2^-) , a peroxide ion $(O_2^2^-)$, hydrogen peroxide (H_2O_2) , or a hydroxyl radical (OH). In a biological context, ROS are byproducts of the normal metabolism of oxygen, and have roles in cell signaling and homeostasis. They are normally eliminated **by** antioxidative intracellular mechanisms, but they otherwise can cause irreversible damage **if** they oxidize cellular components such as proteins, lipids (cell membrane), and **DNA,** thereby preventing from performing their functions, in a process called oxidative stress.

B. Principle

[0048] None of the prior art studies considered the possibility of synergetic or antagonistic interactions between different wavelength bands in microorganisms. The existence of interactions between chromophores in bacterial and yeast cells, and possibly in the shells of enveloped viruses, which upon excitation **by** multiple wavelengths of visible light, as well as ultraviolet-A **(UV-A)** and near-infrared (NIR) radiation, may stress pathogens and so prevent them from multiplying.

[0049] When an endogenous chromophore such as a porphyrin or a flavin in a bacterial cell absorbs a photon, one of its electrons is promoted into a higher-energy orbital, elevating the chromophore from its ground state into a short-lived, electronically excited state. It may then decay from this excited state to the ground state, typically within a nanosecond to a microsecond. **[0050] All** pathogens exposed to light have evolved under the spectral distribution of natural daylight, and so it is possible that cellular processes involved in managing oxidative stress may rely on different chromophores. Whether or not these chromophores harvest energy from incident photons to perform their respective functions, exciting them with photons whose

wavelengths are within the chromophores' spectral absorption peaks may disrupt their activity and so increase oxidative stress within the cells.

[0051] There are synergetic or antagonistic interactions between chromophores involved in the processes responsible for removing ROS under oxidative stress conditions. It is not necessary that these chromophores, including proteins and lipids, be involved in photochemical reactions, but rather that they may be electronically excited **by** incident photons in a manner that disrupts their normal activity. This activity may not necessarily involve the removal of the ROS **if** the disruption results in environmental stress leading to cell inactivation. **If** these interactions are influenced **by** the application of multispectral but not necessarily broadband radiation, it **is** possible to employ a multispectral **LEE** with wavelengths tuned to the absorption peaks of two or more of the corresponding chromophores such that its antimicrobial efficacy is greater than that of a quasimonochromatic or broadband **LEE.** In other words, a radiant flux that is sufficient to have an effect, when it is incident upon a microorganism from a combination of two monochromatic LEEs, is less than the radiant flux that would be required to achieve the same effect using quasimonochromatic or broader band light that encompasses the two wavelengths of the monochromatic LEEs.

[0052] By synchronizing the frequencies and phases of two or more spectral components of a multispectral **LEE,** further benefits may be obtained compared to a pulsed, single spectral component. Such benefits may include a further increase in efficacy of pathogen reduction, for example. As one example, an **LEE** with a blue component with a spectral range of 400 nm to **410** nm and a green component with a spectral range of **520** nm to *530* nm may be modulated with a frequency of **1000** Hz and a duty factor of 20 percent, but where the phase of the green component is offset **by 90** degrees (i.e., *250* microseconds delay) from that of the blue component.

C. Exemplary System

[0053] Referring to **FIG.** 2, an exemplary system is shown. **A** driver **10** is connected to LEEs 12, 14, and may drive them identically or differently. **A** controller **15** connected to the driver includes a processor **16** connected to a computer readable memory **18,** which stores computer readable instructions in the form of an application 20. This application controls the driver **10,** e.g. **by** specifying the output parameters of the LEEs, such as the frequency of the pulse modulation, the peak intensity, the duty factor, the phase between two or more **LEE** driving signals, etc. Also stored in the memory are data 21 and an **Al** (artificial intelligence) application 22. **A** user interface **26** is also connected to the processor and one or more sensors **28** are connected to the

processor. Cultures **30** of pathogens grown on agar may be measured before and after being subjected to the illumination from the LEEs and the data fed back into the system via the user interface, for example. The processor **16** executes the instructions in the application 20 to provide one or more functions of the system.

[0054] Referring to **FIG. 3,** the intensity outputs of two LEEs are shown. The lower graph shows the output 40 of one LEE 12 with pulses 41 at a pulse frequency of f_1 and peak radiant flux I_1 . The average intensity is shown **by** the dashed line 42. The upper graph shows the output *45* of another LEE 14 with pulses 46 at a pulse frequency of f_2 and peak radiant flux I_2 . The average intensity is shown by dashed line 47. A time difference Δt is present between the start of pulses 41 and the start of corresponding pulses selected from pulses 46. The time difference Δt is a positive, zero or negative value and is measured between the start of a cycle 48 of the output from **LEE** 12 and the start of a corresponding cycle 49 of the output from **LEE** 14. Some or all of the parameters Δt , I_1 , I_2 , f_1 , f_2 , pulse modes and duty factors may be adjusted by the controller 15, subject to any constraints on the system, such as f_1 and f_2 being integral multiples of a common or fundamental frequency.

D. Exemplary Embodiments

[0055] In a first embodiment, a pulse modulated **LEE** has a radiant flux output with variable duty factor and variable radiant flux output, such that the radiant flux output can be increased with decreasing duty factor to provide constant average radiant flux. The **LEE** may have a wavelength of 405 nm, for example, or a wavelength in the range 400 **-** 410 nm, or in the range 200 **- 3000** nm for embodiments in which non-visible light is used. Referring to **FIG.** 2, such an **LEE** may be represented **by LEE** 12 or **LEE** 14. In variations of this embodiment, multiple identical LEEs 12, 14 may be combined and driven in synchronization with each other to provide the pulsed light output.

[0056] In a second embodiment, a multispectral pulse modulated **LEE** has a radiant flux output with variable duty factor and variable radiant flux output, wherein the radiant **flux** is comprised of two or more constituent LEEs with different spectral ranges. The constituent LEEs may have two different wavelengths in the range 400 **- 700** nm, or in the range 200 **- 3000** nm, for example, as there are embodiments in which both visible and non-visible wavelengths are used. Referring to **FIG.** 2, such a composite **LEE** may be represented **by** the combination of **LEE** 12 and **LEE** 14, for example where each of **LEE** 12 and **LEE** 14 have a different wavelength. **[0057]** In a third embodiment, a multispectral pulsed modulated **LEE** has a radiant flux output with variable duty factor and variable radiant flux output, wherein the frequencies of the spectral

component modulations are multiples of a fundamental frequency. The radiant flux is provided **by** two or more constituent LEEs with different spectral ranges, or with different monochromatic wavelengths. The constituent LEEs may have two different wavelengths in the range 400 **- 700** nm, or in the range 200 **- 3000** nm, for example. Referring to **FIG.** 2, such a composite **LEE** may be represented **by** the combination of **LEE** 12 and **LEE** 14, for example. Referring to **FIG. 3,** one of the constituent LEEs may have a first frequency **fi** of pulse modulation and a second one of the constituent LEEs may have a frequency f_2 of pulse modulation, where $f_2 = 2f_1$. In other variations of this embodiment, $f_2 = 3f_1$, or $f_2 = 4f_1$, for example, with other multiples also possible. In yet other variations of this embodiment, $f_1 = nf_0$ and $f_2 = mf_0$, where f_0 is the fundamental frequency and n and m are different integers, e.g. 2 and **3.** That is, the ratio between the pulse frequencies is non-integral.

[0058] As discussed above, different chromophores have peak susceptibilities in different wavelength ranges (e.g., 400 to 410 nm and *520* to **530** nm) and at different pulse frequencies. It is reasonable to assume that different chromophores have different relaxation constants. Ideally, the pulse rate for each wavelength would be determined **by** this constant. However, having different pulse frequencies could lead to undesirable beat frequencies that could be visually noticeable. Assuming that the dependence on pulse frequency is reasonably low, such that a range of pulse frequencies does not significantly affect the susceptibility, then different chromophores may matched to different pulse frequencies that are multiples of a fundamental frequency. **By** applying light to one or both chromophores at a frequency that is not at, but relatively close to, the frequency of peak susceptibility allows for a simplification of the **LEE** driver **by** avoiding beat frequencies, while maintaining a significant and effective amount of susceptibility. **A** luminaire may therefore have one or more LEEs emitting in the range 400 410 nm and one or more other LEEs emitting in the range **520-530** nm, where the different LEEs have different pulse frequencies.

[0059] In a fourth embodiment, a multispectral pulsed modulated **LEE** has a radiant flux output with variable duty factor and variable radiant flux output, wherein the frequencies of the spectral component modulations are multiples of a fundamental frequency, and where the phases of the spectral components are offset from each other **by** predetermined amounts. The radiant flux **is** provided **by** two or more constituent LEEs with different spectral wavelengths. The constituent LEEs may have two different wavelengths in the range 400 **- 700** nm, or in the range 200 **- 3000** nm, for example. Referring to **FIG.** 2, such a composite **LEE** may be represented **by** the combination of **LEE** 12 and **LEE** 14, for example. Referring to **FIG. 3,** one of the constituent LEEs may have a pulsed intensity output 40 and a second one of the constituent LEEs may have

a pulsed intensity output 45. The start of the pulses 41 of the output 40 are shown to lag the start of corresponding pulses 46 of the output 45 **by** a time At relative to the start of every second pulse 46. The time Δt represents a phase lag of output 40 compared to output 45. In other variations of this embodiment, the pulses 41 start in advance of the corresponding pulses 46, representing a phase lead of output 40 compared to output 45. In other variations of this embodiment, $f_1 = f_2$. In yet other variations of this embodiment, the pulses 41 and 46 do not overlap, or pulses 41 start at the same time as the respective ones of pulses 46 end. **[0060]** In a fifth embodiment, a multispectral pulsed modulated **LEE** has a radiant flux output with variable duty factor and variable radiant flux output, wherein the frequencies of the spectral component modulations are multiples of a fundamental frequency, where the phases of the spectral components are offset from each other **by** predetermined amounts, and wherein the amplitudes of the spectral component modulations differ with respect to each other **by** predetermined amounts. Referring to **FIG.** 2, such a composite **LEE** may be represented **by** the combination of **LEE** 12 and **LEE** 14, for example. Referring to **FIG. 3,** one of the constituent LEEs may have a pulsed intensity output 40 with a peak radiant flux output I_1 and a second one of the constituent LEEs may have a pulsed radiant flux output 45 with a peak **of 12,** where **I1** and **¹²**are different. In a variation of this embodiment, one or both of the peak radiant fluxes may be independently adjusted, or they may both be adjustable while maintaining a constant ratio between the two.

[0061] In a sixth embodiment, the duty factors and radiant flux outputs of a pulse modulated **LEE** or multispectral pulsed modulated **LEE** may vary in response to external conditions, including time, ambient illumination, relative humidity, ambient temperature, air movement, salinity, and other environmental factors which may affect the growth or inhibition of pathogens. For example, in terms of determining a consistent exposure time, it may be advantageous to modulate the radiant flux in response to different or changing external conditions. These external conditions may be determined as a result of electronic monitors or may be entered manually through a user interface. In addition, feedback systems may be employed to automate the duty factors and radiant flux outputs of a pulse modulated **LEE** or multispectral pulse modulated **LEE** in response to external conditions. For example, a method may involve detecting, with a sensor, a parameter of an environment of the microorganism; and

in response, adjusting either or both of a peak radiant flux and a duty factor, in either or both of the first light and the second light, in order to maintain a predetermined rate of chromophore excitation in other microorganisms that are the same as said microorganism.

[0062] Common means of determining pathogen inactivation are colony counts on agar plates.

With sufficient time and patience, the relationship between different external conditions and inactivation rates may be explored and enumerated.

[0063] Referring to **FIG.** 2, one or more sensors **28** form part of or are connected to the system. The sensors may monitor a duration of a particular environment, ambient illumination, relative humidity, ambient temperature, air movement, salinity or any other factor which may influence the multiplication rate of pathogens. For example, **if** the environmental factors are known to promote a slower reproduction of a pathogen, then the peak intensity, duty cycle or frequency of the **LEE** output may be reduced to save energy while still maintaining a satisfactory pathogen reduction rate. Conversely **if** the environmental factors are known to promote a more rapid reproduction of a pathogen, then the peak intensity, duty cycle or frequency of the **LEE** output may be increased to maintain a satisfactory pathogen reduction rate.

[0064] In an additional embodiment, machine learning may be used to help determine the effect of specific conditions which affect a particular pathogen, including external conditions. This machine learning also helps to assess pathogen mutations, and determine changing conditions that affect the pathogen over time. Referring to **FIG.** 2, an **Al** application 22 is included in the system. The **Al** application takes as input the environmental conditions as indicated **by** the sensors **28,** the intensity profiles of the LEEs, and the level or change in level of pathogens in the environment as determined from time to time **by** pathogen colony growth **30.** Results from the colony growth may be input manually into the system via a user interface **26,** or photographs (including at the microscope level) of the culture growth may be taken manually or automatically and input into the **Al** application.

E. Exemplary Applications

[0065] In a first application and referring to **FIG.** 2, pulsed LEEs 12, or 12 and 14, are incorporated in a dedicated architectural, industrial, or medical light source to provide visible light disinfection radiation.

[0066] In a second application and referring to **FIG.** 4, pulsed LEEs *50, 54* with different monochromatic or quasimonochromatic wavelengths are incorporated in an architectural or industrial luminaire **56** with a "white light" **LEE** *52.* For example, **LEE 52** has a broad spectral output, or multiple broad spectral ranges, or multiple monochromatic or quasimonochromatic wavelengths that appear to the human eye as white. The luminaire *56,* as well as providing pathogen-reducing light, also provides general "white light" illumination from **LEE 52** with a fixed or variable **CCT** and color rendering properties. Wavelengths of LEEs **50,** 54 may be tuned to the absorption peaks of different chromophores in a microorganism. For example, **LEE 50**

may be a 405 nm **LED** tuned to a porphyrin and **LEE** 54 may be a 450 nm **LED** tuned to nboflavin.

[0067] In a third application, pulsed multispectral LEEs are incorporated in an architectural or industrial luminaire that also provides general "white light" illumination with a fixed or variable correlated color temperature, wherein the spectral power distribution **(SPD)** of the white light illuminant **(LEE 52)** is such that the combination of its **SPD** and that of the pulsed multispectral LEEs **50,** 54 generates white light with a predetermined **CCT** and color rendering properties. For example, compared to a white light luminaire that would traditionally be used only for illumination of an interior space, one or more portions of the spectrum of the white light are removed and provided instead as pathogen-reducing light. As such, the majority of the white light for illumination may be provided at a higher frequency (e.g. 2 to **500** times higher than the fundamental frequency) than the pathogen-reducing light, while the luminaire **56** still appears to the human eye to output the same spectrum as the traditional luminaire. For example, the PWM frequency of commercial **LED** drives is between 20 kHz and **50** kHz, which minimizes the size of the smoothing capacitors. The PWM frequencies explored so far for disinfection purposes have been **100 Hz to 10** kHz.

[0068] In a fourth application, the pulsed multispectral LEEs are realized as pulse modulation LEEs in a horticultural luminaire, e.g. luminaire **56** in **FIG.** 4. In a variation of this embodiment, **LEE** light emanates from optical components that are mounted on a substrate, each optical component being coupled to an optical fiber that delivers optical radiation combined from LEEs 12, 14 **(FIG.** 2) or **50, 52,** 54, for example. **A** linear or elliptical holographic diffuser is located to diffuse the light emanating from the optical components. An example of this set-up of optical components may be seen in patent **US11,122,747.** The **LEE** wavelengths excite plant photopigments for predetermined physiological responses, and the light source intensities may be temporally modulated to maximize photosynthesis and control photomorphogenesis responses. The light may also **kill** the pathogens on the plants as well. However, the dependence of the photosynthetic process on visible light may complicate matters. This is where pulsed LEEs with small duty factors may be useful, in that the pulses of light do not significantly contribute to the Daily Light Integral for the plants, but they may be sufficient to inactive pathogens on the plant leaves, such as powdery mildew. Each **LEE** may be independently controlled. Optical fiber textiles may be used to provide the light output.

[0069] In a fifth application and referring to **FIG. 5,** the pulse modulated LEEs **60, 62** are incorporated in a medical light treatment system. The light treatment system comprises one or more LEEs for illuminating a part (e.g. skin) of a human or animal body 64 for light treatment.

The light treatment system further comprises a housing **66** for emitting light towards the part of the human or animal body, and for at least partially covering the illuminated part of the human or animal body from view. The housing further comprises a window **68** for emitting light from an inner part of the housing away from the housing. An effect of the light treatment system according to the invention is that the window in the housing may be used as an indicator whether the light treatment system is operating and/or functioning well. Furthermore, the presence of a luminescent material, for example, on the window, may be used as an indicator for, for example, ultraviolet light, as part of the ultraviolet light may be converted **by** the luminescent material into visible light. Optical fiber textiles may be used to provide the light output and placed near on the part of the human or animal body.

[0070] In a sixth application and referring to **FIG. 6,** the pulse modulated LEEs **70, 72** are incorporated in a biological lighting system 74 for controlling root growth of plants **76,** wherein the optical radiation either prevents or promotes bacterial activity in the soil or other growth medium **78.** Such a growth medium may be transparent or translucent. In some cases the growth medium may be soil, through which there is at least some penetration of the illumination. Bacterial activity that may be promoted **by** the applied light includes, for example, nutrient recycling, decomposition of organic matter, inhibition of diseases and reproduction. In another application, the **LEE 79** may be used to illuminate the shoot system of the plants, to inhibit pathogens that may be present on **it.**

[0071] In addition to the pathogen-influencing illumination, the biological lighting system also provides further, temporally and spatially modulated photon flux output and spectral power distributions to plants on a circadian and circannual basis, or circadian and life cycle basis, to maximize effective and efficient growth in a horticultural setting. This additional irradiation, e.g. **by LEE 79,** may be applied to the shoot sytem, the roots, or both. The photon flux or irradiance output and the spectral power distribution are modulated to match circadian and circannual rhythms, with individual or multiple luminaires controlled through one or more controllers. Different lighting spectra can be employed depending on the direction of illumination. The photon flux or irradiance output and the spectral power distribution may be set as best suited for any particular plant species, and the system is also useful for raising animals.

[0072] In a seventh application in **FIG. 7,** the pulsed LEEs **80, 82** are incorporated in a water purification system 84. As the water 86 flows through a pipe, for example, it is illuminated with the light from the LEEs **80, 82.** The light reduces the number of pathogens in the water. In a variation of this embodiment, the water may be static, e.g. in a tank, while it is being illuminated.

F. Exemplary Method

[0073] Referring to **FIG. 8,** a method is shown **by** which data is fed back to the system in order to better control the luminaire. In step **90,** an environment in which a pathogen is present or may be present is illuminated with the luminaire. The illumination may be, for example, illumination with two different monochromatic wavelengths of light, with pulse frequencies that are integral multiples of a fundamental frequency, and which may have a zero, positive or negative phase difference between the two. In step **92,** the amount of pathogen in the environment is measured, which may involve, for example, taking a sample and growing a culture on an agar plate. In step 94, the results of the culture growth are recorded in a database **96,** in relation to the parameters of light that was used for the illumination of the environment prior to taking the sample. In step **98,** the settings of the parameters of the luminaire are changed, e.g. **by** changing the duty factor, the peak intensity, the frequency, the phase between two different wavelengths etc. The environment with the pathogen or possible presence of the pathogen is again illuminated in step **90,** with the adjusted parameters, and the process cycles through multiple times. Over time, enough data **is** obtained for a person or an **Al** machine to determine optimum settings for the luminaire. In other cases, analytic or algorithmic relationships between the parameters of the luminaire and the inhibition of growth of the pathogen may be determined.

G. Variations

[0074] The present invention is herein described more fully with reference to the accompanying drawings, in which embodiments and applications of the invention are shown. This invention may, however, be embodied and applied in many different forms, and should not be construed as limited to the embodiments and applications described herein. Rather, these embodiments and applications are provided so that this disclosure will be thorough and complete, and will fully convey the scope of the invention to those skilled in the art.

[0075] Modules may be divided into constituent modules or combined into larger modules. **[0076]** The system may be configured so that its output may be changed depending on the type of pathogen that is present in the space that is to be disinfected.

[0077] The system may be used for research purposes for the further investigation of how multiple pulsed wavelengths of illumination inhibit or promote the activity of different microorganisms.

[0078] Throughout the description, specific details have been set forth in order to provide a more thorough understanding of embodiments of the invention. However, the invention may be practised without these specific details. In other instances, well known elements have not been

shown or described in detail and repetitions of steps and features have been omitted to avoid unnecessarily obscuring the invention. Embodiments, depending on their configuration, may exhibit all or fewer than all of the advantages described herein. Other advantages not mentioned may be present in one or more of the embodiments. Features from any of the embodiments may be combined with features from any of the other embodiments to form another embodiment within the scope of the invention. In general, unless otherwise indicated, singular elements may be in the plural and vice versa with no loss of generality. **All** quantities, parameters, wavelengths are examples only and others may be used in other embodiments. **All** ranges given include all subranges within the range. For example, **if** a range is given as m-q, then the ranges m-n, n-p and **p-q are included, where n and p are any values that satisfy m<n<p>>s-q.**

[0079] Accordingly, the scope of the invention is to be construed in accordance with the substance defined **by** the claims.

CLAIMS

I claim:

1. A method for exciting two different chromophores in a microorganism, the method comprising:

illuminating the microorganism with first light having a first pulse frequency and a first wavelength between **380** nm and *750* nm; while

illuminating the microorganism with second light having a second pulse frequency and a second wavelength between **380** nm and *750* nm;

wherein the second wavelength is different to the first wavelength;

wherein the first and second pulse frequencies are different;

wherein the first and second pulse frequencies are integral multiples of a fundamental frequency; and

wherein a pulse train of the first light has a zero, positive or negative phase difference with respect to a corresponding pulse train of the second light.

2. The method of claim **1,** wherein the microorganism is in air, in water, in a horticultural growth medium, on mammalian skin, on a plant root or on a plant shoot system.

3. The method of claim **1,** comprising providing further illumination, which, in combination with the first light and the second light, provides white light for general illumination with a predetermined correlated color temperature and predetermined color rendering properties.

4. The method of claim **3,** wherein the further illumination has a pulse frequency of **2-500** times greater than the fundamental frequency.

5. The method of claim **1,** comprising providing further illumination, which, in combination with the first light and the second light, provides light corresponding to a circadian cycle, circannual cycle or life cycle of a plant.

6. The method of claim **1,** wherein each of the first and second wavelengths is tuned to an absorption peak of the corresponding chromophore.

7. The method of claim **1,** wherein:

the first light and the second light are monochromatic; and

a radiant flux that is incident upon the microorganism from the first light and the second light combined is less than a radiant flux that would be required to achieve said excitation of the chromophores using quasimonochromatic or broadband light that encompasses the first and second wavelengths.

8. The method of claim **1,** wherein the phase difference is non-zero.

9. The method of claim **1,** further comprising: detecting, with a sensor, a parameter of an environment of the microorganism; and in response, adjusting either or both of a peak radiant flux and a duty factor, of either or both of the first light and the second light.

10. The method of claim **1,** wherein each chromophore is a flavin or a porphyrin.

11. A luminaire comprising:

a first light emitting element **(LEE)** that emits first light at a first pulse frequency with a first wavelength between **380** nm and *750* nm; and

a second **LEE** that emits second light at a second pulse frequency with a second wavelength between **380** nm and *750* nm;

wherein the second wavelength is different to the first wavelength;

wherein the first and second pulse frequencies are different;

wherein the first and second pulse frequencies are integral multiples of a fundamental frequency; and

wherein a pulse train of the first light has a controllable phase difference with respect to a corresponding pulse train of the second light.

12. The luminaire of claim **11,** wherein the first light and the second light are monochromatic.

13. The luminaire of claim 12, wherein the first wavelength is between 400-410 nm and the second wavelength is between **520-530** nm.

14. The luminaire of claim 11 further comprising one or more further LEEs that generate optical radiation with one or more wavelengths in a range of 200 nm to **3000** nm, wherein the optical radiation is monochromatic, polychromatic, multispectral or quasimonochromatic.

15. The luminaire of claim 14, wherein the optical radiation corresponds to a circadian cycle, a circannual cycle or a life cycle of a plant.

16. The luminaire of claim 11 further comprising one or more further LEEs that generate optical radiation with one or more wavelengths in a range of **380** nm to *750* nm.

17. The luminaire of claim **16,** wherein the optical radiation is emitted at a pulse frequency **2-500** times greater than the fundamental frequency.

18. The luminaire of claim **11,** wherein a ratio between the first and second pulse frequencies is non-integral.

19. A system for exciting two different chromophores in a microorganism, the system comprising:

a luminaire having:

a first light emitting element **(LEE)** that emits first light at a first pulse frequency with a first wavelength between **380** nm and **750** nm; and

a second **LEE** that emits second light at a second pulse frequency with a second wavelength between **380** nm and **750** nm;

wherein the second wavelength is different to the first wavelength;

wherein the first and second pulse frequencies are different;

wherein the first and second pulse frequencies are integral multiples of a fundamental frequency; and

wherein a pulse train of the first light has a controllable phase difference with respect to a corresponding pulse train of the second light;

a driver coupled to drive each **LEE** with an independently variable duty factor and an independently variable peak radiant flux output and to control the phase difference;

a sensor configured to detect a parameter of an environment of the microorganism; and a controller configured to adjust, based on the parameter, either or both of a peak radiant flux and a duty factor, in either or both of the first light and the second light.

20. The method of claim **1,** comprising controlling the phase difference.

FIG. 1 (PRIOR ART)

FIG. 2

2/3

INTENSITY 49 46 \mathbf{I}_2 $\begin{array}{ccc} \hline \end{array}$ $\begin{array}{ccc} \hline \end$ 0 $f₂$ 48 I_1 $\overline{I_1}$ $\overline{I_2}$ $\overline{I_2}$ $\overline{I_3}$ $\overline{I_4}$ 42 40 0 f₁ $+$ $+$ $+$ $+$ \rightarrow TIME Δt Δt

FIG. 3

FIG. 4

FIG. 5

3/3

FIG. 6

FIG. 7

FIG. 8