



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification<sup>4</sup> :</b> <b>G01N 33/553, 33/543, 21/55</b> <b>// G01N 33/94, 33/74</b>	<b>A1</b>	<b>(11) International Publication Number: WO 89/ 08260</b> <b>(43) International Publication Date: 8 September 1989 (08.09.89)</b>
<b>(21) International Application Number:</b> PCT/GB89/00156 <b>(22) International Filing Date:</b> 23 February 1989 (23.02.89) <b>(31) Priority Application Number:</b> 8804669 <b>(32) Priority Date:</b> 27 February 1988 (27.02.88) <b>(33) Priority Country:</b> GB <b>(71) Applicant:</b> AMERSHAM INTERNATIONAL PLC [GB/GB]; Amersham Place, Little Chalfont, Buckinghamshire HP7 9NA (GB). <b>(72) Inventors:</b> CORRIE, John ; 50 Hornbeam Spring, Knebworth, Hertfordshire SG3 6AY (GB). FAIRCLOUGH, Lynne ; 97 Heath Street, Hampstead, London NW3 6SS (GB). CHARLES, Stephen, Alexander ; 5 Eliot Close, Aylesbury, Buckinghamshire HP19 3JB (GB). FINLAN, Martin, Francis ; 7 Ballard Close, Aylesbury, Buckinghamshire HP21 9UY (GB).	<b>(74) Agent:</b> PENNANT, Pyers; Stevens, Hewlett & Perkins, 5 Quality Court, Chancery Lane, London WC2A 1HZ (GB). <b>(81) Designated States:</b> AT (European patent), BE (European patent), CH (European patent), DE (European patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent), SU. <b>Published</b> <i>With international search report.          Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>	
<b>(54) Title: IMMOBILISATION OF HAPTENS AND MEASUREMENT</b>		
<b>(57) Abstract</b> <p>A metal surface carries a coating comprising spacer units, e.g. protein molecules, to which haptens are linked. These metal surfaces are useful for assays, e.g. in which dissolved haptens in a sample compete with immobilised haptens for binding to antibodies. The coated metal surfaces are adapted for use in surface plasmon resonance (SPR) techniques. Also included are immunoassays in which antibodies are immobilised on the metal surface with hapten conjugates reversibly bound to them, displacement of conjugate, as a result of addition of a sample containing the hapten, being monitored by SPR.</p>		

***FOR THE PURPOSES OF INFORMATION ONLY***

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	FR	France	ML	Mali
AU	Australia	GA	Gabon	MR	Mauritania
BB	Barbados	GB	United Kingdom	MW	Malawi
BE	Belgium	HU	Hungary	NL	Netherlands
BG	Bulgaria	IT	Italy	NO	Norway
BJ	Benin	JP	Japan	RO	Romania
BR	Brazil	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	LI	Liechtenstein	SN	Senegal
CH	Switzerland	LK	Sri Lanka	SU	Soviet Union
CM	Cameroon	LU	Luxembourg	TD	Chad
DE	Germany, Federal Republic of	MC	Monaco	TG	Togo
DK	Denmark	MG	Madagascar	US	United States of America
FI	Finland				

IMMOBILISATION OF HAPTENS AND MEASUREMENT

This invention concerns methods of assaying for analytes using the phenomenon of surface plasmon resonance (SPR). The method is applicable to analytes generally, but is likely to be of particular interest where the analyte is a hapten (a small molecule capable of being bound by antibody but not of itself immunogenic).

The phenomenon of SPR is well known and will not be described in detail. Briefly, the intensity of monochromatic plane-polarised light (conveniently obtained from a laser) reflected from the interface between an optically transparent material, e.g. glass, and metal depends on the refractive index of material in a thin layer, at most a few hundred nm thick, on the downstream side of the metal. Accordingly, by measuring changes in intensity of reflected light an indication can be obtained of changes in refractive index of material on the metal. The intensity of reflected light also varies with the angle of incidence, and reflectivity drops sharply to a minimum at a particular angle characteristic of the equipment. The metal surface is generally of silver, although this is not critical to the invention.

The immunoassay of haptens by Surface Plasmon Resonance Spectrometry (SPRS) poses a particular problem because the haptens are, by definition of low molecular weight and therefore cause only very small changes in refractive index when they bind to or dissociate from an antibody-coated SPRS silver-coated surface.

This invention is also concerned with immobilisation of haptens, for use in such immunoassays. Difficulties arise when attempting to immobilise haptens on metal surfaces e.g. for use in immunoassays: although haptens

- 2 -

can be immobilised directly on a metal surface, the immobilised haptens may be unusable in immunoassays.

The present invention aims to overcome these difficulties.

5 According to one aspect of the present invention there is provided an immobilised hapten comprising a metal surface which has immobilised thereon spacer units to which haptens are linked.

Preferably, the metal surface carries a  
10 macromolecular coating comprising macromolecular spacer units to which haptens are covalently linked. The coating may comprise spacer units and blocking agent which may be of organic polymeric material which may be of biological or synthetic origin. To minimise non-  
15 specific binding, the coating should be continuous, and may be covalently attached to the metal surface.

The spacer unit preferably comprises a protein which may be chemically linked to a hapten, e.g. by a suitable bridge structure. Known techniques, such  
20 as the mixed anhydride technique, may be used for this purpose. Examples of suitable hapten-bridge-protein conjugates include phenytoin-glucuronide-gamma globulin, and phenytoin-glucuronyl-lysozyme. Suitable bridges for other combinations are known to those  
25 skilled in the art.

It is thought desirable that the protein has an excess of basic groups (i.e. at least one more basic than acidic amino acid residue), and preferred proteins include gamma globulin, thyroglobulin and methylated  
30 ovalbumin, with gamma globulin currently being the most preferred protein. It is believed that the presence of an excess of basic groups provides a beneficial effect by promoting adsorption of the protein to a metal, e.g. silver, surface. It is, however, also thought that the  
35 presence of an excess of basic groups may have the undesirable effect of enhancing non-specific protein

binding, probably not at the metal surface but by association of non-specific proteins with the spacer protein. A degree of non-specific binding is tolerable and in any event can be reduced by use of a blocking material, e.g. by coating the immobilised hapten with material such as non-immune serum or gamma globulin.

5 The metal desirably comprises silver or gold, conveniently in the form of a layer e.g. deposited by evaporation on a carrier such as a glass slide.

10 This aspect of the invention is applicable to all haptens, including e.g. most drugs, steroid hormones, thyroxine.

It is found that by use of a suitable spacer unit, with the hapten indirectly immobilised near the metal surface, non-specific binding problems can be overcome. Although some non-specific binding may occur, as discussed above, a degree of non-specific binding is tolerable and dose dependent antibody binding can be obtained.

15 The present invention also provides a method of immobilising a hapten on a metal surface, comprising linking the hapten to a spacer unit to form a conjugate and immobilising the conjugate on the metal surface.

20 An immobilised hapten in accordance with this aspect of the invention finds application, inter alia, in immunoassays.

Hence in a further aspect the present invention provides an immunoassay method using hapten linked to a spacer unit and immobilised on a metal surface.

30 The method of the invention may be used for determining the presence in a sample of antibody to a particular hapten, by contacting immobilised hapten with the sample and determining whether hapten-antibody binding has occurred, although this possibility is unlikely to be of practical importance.

35 In a further aspect the invention provides an

immobilised hapten comprising a metal surface which has immobilised thereon spacer units to which haptens are linked, with corresponding antibody bound to the hapten.

5           These concepts lead to immunoassays of two kinds for analytes, preferably but not necessarily haptens.

          Thus in a further aspect, the invention provides a method of assaying for an analyte in a sample, by the use of the analyte or an analogue thereof immobilised  
10       via spacer units on a metal surface, which method comprises bringing an antibody to the analyte, and the sample containing the analyte, into contact with the metal surface, and monitoring antibody binding to the immobilised analyte or analogue by means of surface  
15       plasmon resonance as indicative of the presence or the concentration of the analyte in the sample.

          An analogue of the analyte is a substance which competes with the analyte for binding to a specific binder, such as an antibody, therefor. Often the  
20       analogue will be arranged to be as near as possible or even completely identical to the analyte. The use in assays of analyte analogues is well known.

          The order of addition of reagent is not critical. Preferably antibody is first added to the immobilised  
25       hapten or analogue, which is then washed to remove any unbound material. Then the sample is brought into contact with the metal surface, on which the antibody has previously been reversibly bound to the immobilised analyte or analogue, displacement of antibody being  
30       monitored as indicative of the presence or the concentration of the analyte in the sample. This arrangement has the advantage that bringing the sample into contact with the metal surface generates the SPR signal, no other reagent being required.

35           In a further aspect, the invention provides a method of assaying for an analyte in a sample, by the

- 5 -

use of a metal surface having immobilised thereon antibody to the analyte, which method comprises bringing a conjugate of the analyte or an analogue thereof, and the sample containing the analyte, into  
5 contact with the metal surface, and monitoring binding of the conjugate by means of surface plasmon resonance as indicative of the presence or concentration of the analyte in the sample.

The conjugate of the analyte or analogue should be  
10 with some substance of sufficient molecular weight to yield a significant SPR signal on the displacement of the conjugate by added free hapten, and may typically be a macromolecule such as a protein.

Again, the order of addition of reagents is not  
15 critical. Preferably conjugate is first reversibly bound to the immobilised antibody, and the sample thereafter brought into contact with the metal surface, displacement of the conjugate being monitored to assay the analyte.

20 These methods can be used qualitatively or quantitatively.

When producing antibodies to a hapten, the hapten is commonly linked to a larger unit using a bridge structure similar to the bridge structures mentioned  
25 above for linking a hapten to a protein spacer unit. For use in a particular immunoassay, different bridge structures should be used for the two purposes, to prevent recognition of the hapten-protein bridge structure by the antibodies. For example, where the  
30 hapten phenytoin is linked to lysozyme by use of glucuronyl bridge for immobilisation purposes, phenytoin-3-omega-valeryl-bovine serum albumin may be used as the immunogen against which antiserum is raised.

35 It is possible to use antibody fragments

incorporating the hapten binding site, such as Fab' or Fab fragments, in place of the entire antibody molecule, and references to antibodies should be construed as including such fragments.

5           References to antibodies should thus be construed as including modified antibodies.

Work carried out using SPR for refractive index detection indicates that the method is very sensitive and is capable of giving accurate results within a few  
10           seconds, with detection limits of below 1 ng/l.

The invention finds particular application in monitoring a wide range of analytes of clinical importance, typically having serum concentrations in the range  $10^{-6}$  to  $10^{-12}$  mol/l. In particular the  
15           invention is useful in monitoring serum levels of haptens including drugs such as theophylline, methotrexate, aminoglycoside antibiotics etc.

The invention will be further described, by way of example, with reference to the accompanying drawings, in which:  
20

Figure 1 illustrates schematically one embodiment of apparatus for carrying out the method of the invention, using surface plasmon resonance;

Figure 2 is a diagram showing two assay systems  
25           (a) and (b) according to the invention;

Figure 3 is a graph of reflectivity versus time showing results obtained for phenytoin using the apparatus of Figure 1;

Figure 4 is a graph of reflectivity versus time  
30           showing results obtained on theophylline using SPR equipment described in EPA 305109; and

Figure 5 is a graph of reaction rate against thyroxine concentration in an immunoassay of thyroxine using SPR.

35           Figure 6 is a graph of reflectivity versus time in



- 7 -

an assay for theophylline in which anti-theophylline antibodies were attached to the metal surface.

Detailed description of the drawings

5 The apparatus illustrated schematically uses the phenomenon of surface plasmon resonance (SPR) for measuring the refractive index of a layer absorbed on a thin metallic film.

10 The phenomenon of SPR is well known and will not be described in detail. Briefly the intensity of monochromatic plane-polarised light (conveniently obtained from a laser) reflected from the interface between an optically transparent material, e.g. glass, and metal depends on the refractive index of material on the downstream side of the metal. Accordingly, by  
15 measuring changes in intensity of reflected light an indication can be obtained of changes in refractive index of material on the metal at a particular point of the metal. The intensity of reflected light also varies with the angle of incidence, and reflectivity  
20 drops sharply to a minimum at a particular angle X for any given set up. The apparatus is most sensitive when the angle of incidence Y of the light is more acute than the angle X, roughly half way along the linear part of the dip in SPR curve.

25 The illustrated apparatus comprises a glass prism 10 to which is attached a glass microscope slide 12 covered on one side by a thin (about 50 to 60nm thick) film 14 (not shown to scale) of silver deposited on the slide by evaporation. A hapten is indirectly  
30 immobilised near to the surface of the silver film, as will be described below. A laser light source 16, e.g. a Uniphase Model 1108P He-Ne laser is located on the prism side of the slide, with a light detector 18 located to receive light reflected from the glass-  
35 silver interface. The apparatus is adjusted so that

- 8 -

the angle of incidence of the light has a value  $\gamma$  at which sensitivity is maximised. The intensity of reflected light received by detector 18 is monitored while the immobilised hapten is contacted with a sample  
5 to be analysed.

Figure 2 shows two alternative assay systems in diagrammatic form. In 2(a), the hapten 20 is immobilised to the silver surface 22 used for SPRS detection, and binds the corresponding antibody 24.  
10 Introduction of free hapten 26 (whose concentration it is wished to determine) displaces antibody by competing with surface bound hapten. This displacement of antibody from the surface is detected as an SPRS signal. In 2(b), the antibody 24 is bound to the  
15 surface 22 and binds a conjugate 28 of the hapten 20 and (typically) a protein 30 of sufficient molecular weight to yield a significant SPRS signal on displacement of the conjugate by added free hapten.

The assays in Examples 1 to 3 were carried out using the apparatus of Figure 1. The assays in  
20 Examples 4 to 6 were carried out using the improved equipment described in EPA 305109.

#### Example 1

Phenytoin linked to glucuronide bridge, as  
25 described by J.A. Hinds, C.F. Pincombe, S. Smith and P. Duffy in J. Immunol. Methods, 80 239-53 (1985), was coupled to rabbit gamma globulin using the mixed anhydride technique, as described by B.F. Erlanger, F. Borek, S.M. Beiser and S. Leibermann in J. Biol. Chem.  
30 234, 1090-4 (1959) to produce a phenytoin-glucuronide-rabbit gamma globulin conjugate.

A glass microscope slide covered on one side by a thin (50-60 nm) film of silver was immersed for 30-45 minutes in a 1  $\mu\text{mol/l}$  solution of the conjugate in  
35 buffer (10 mmol/l sodium phosphate, pH 7.5). The

gamma globulin adsorbed to the silver and indirectly immobilised the phenytoin near to the surface. The slide was then immersed in a solution of 5  $\mu\text{mol/l}$  rabbit gamma globulin in phosphate-buffered saline for 30 minutes. The gamma globulin forms a blocking coating and acts to minimise non-specific binding. The slide was then rinsed once in buffer, then immersed for 16 hours in a solution of phenytoin antiserum (raised in a rabbit against a phenytoin-3-(omega-valeryl)-bovine serum albumin conjugate, essentially as described by C.E. Cook, J. F. Kepler and H.D. Christensen in Res. Commun. Chem. Pathol. Pharmacol., 5, 767 (1973)) diluted 1/800 in buffer containing ovalbumin. This results in binding of antibody to the immobilised phenytoin. The slide was then rinsed 4 times in buffer containing 0.05% Tween 20 and a further time in buffer without Tween 20, resulting in a removal of unbound material, and the non-silvered surface was cleaned by wiping with a tissue moistened with isopropanol.

The surface plasmon resonance properties of the slide (i.e. reflectance versus angle of incidence, and reflectance at fixed angle versus time) were then examined essentially as described by B. Liedberg, C. Nylander and I. Lundstroem in Sensors and Actuators, 4, 299-304 (1983). Thus reflectance versus time was measured with the coated face of the slide initially exposed to buffer, and then following injection into a flow cell of a solution of phenytoin in buffer. The presence of phenytoin causes dose-dependent displacement of antibody from the slide, and hence a decrease in refractive index of the biolayer attached to the metal surface, as shown by the decrease in reflectivity. Typical results are shown in Figure 3, where the points indicated by arrows represent

- 10 -

injection of phosphate buffered saline containing 0, 1 or 5  $\mu\text{g/l}$  phenytoin. The initial slope after injection is measured and the values related to hapten concentration.

5           Because the injection of buffer or hapten sometimes causes small spikes in the reflectivity traces, measurement of the initial slope of the reflectivity vs time curves is taken over a 1 minute period commencing 30 seconds after the injection.  
10 Results expressed in this way are set out in the Table below, together with results from Example 2.

Example 2

Progesterone. The  $11\alpha$ -glucuronyl derivative of progesterone, prepared as described by J.E.T. Corrie, 15 W.M. Hunter and J.S. Macpherson, Clin. Chem., 27, anhydride technique of Erlanger et al (op. cit.) was used to produce a progesterone-glucuronyl-rabbit gamma globulin conjugate. Other details exactly as for theophylline, except that the progesterone conjugate 20 was coated onto the slide at a concentration of 1  $\mu\text{mol/l}$ , and the antiserum (raised in a rabbit against a progesterone- $11\alpha$ -hemisuccinyl-bovine serum albumin conjugate as described by K.K. Dighe and W.M. Hunter, Biochem., J. 143, 219 (1974)) was used at a dilution of 25 1 in 400. The Table shows the results for progesterone doses of 0, 12.5 and 50 ng/ml in PBS.

<u>Example</u>	<u>Hapten</u>	<u>Concentration</u>	<u>Slope</u>	
30	1	Phenytoin	5 $\mu\text{g/ml}$	-0.45
			1 $\mu\text{g/ml}$	-0.35
			0	-0.14
35	2	Progesterone	50ng/ml	-0.36
			12.5ng/ml	-0.28
			0	-0.13

Example 3

Other spacers. Theophylline-7-propionic acid, prepared as described above, was conjugated to ovine fibrinogen, equine haemoglobin and lysozyme, all by the technique of Erlanger et al (op. cit.). Each conjugate, together with the theophylline-7-propionyl-rabbit gamma globulin conjugate described above, was coated onto separate silvered glass slides using the conditions described above (i.e. 1  $\mu$ mol/l in phosphate buffer for 45 min). Blocking of residual binding sites on the metal surface was effected by immersion for 30 minutes in 1 in 200 diluted normal rabbit serum in 10 mmol/l sodium phosphate, pH 7.4. The slides were then incubated with the theophylline antiserum and washed as described above and the SPR properties determined in response to doses of theophylline at 0, 1 and 10  $\mu$ g/ml. Initial slopes (% reflectivity change/min) of the reflectivity vs. time curves were as follows:

Theophylline Dose ( $\mu$ g/ml)	Protein Spacer			
	Fibrinogen	Gamma Globulin	Haemoglobin	Lysozyme
0	-0.05	-0.05	-0.09	-0.07
1	-1.06	-0.97	-0.91	-0.46
10	-1.57	-2.04	-1.66	-1.64

Example 4

Theophylline. The derivative theophylline-7-propionic acid, prepared as described by T. Nishikawa, M. Saito and H. Kubo, Chem. Pharm. Bull., 27, 893-8 (1979), was coupled with rabbit gamma globulin using the mixed anhydride technique of Erlanger et al (1959) (as in Example 1) to produce a theophylline-7-propionyl-rabbit gamma globulin conjugate.

- 12 -

A glass microscope slide covered on one side by a thin (50-60 nm) film of silver was immersed for 30-45 min in an 8  $\mu\text{mol/l}$  solution of the conjugate in buffer (10 mmol/l sodium phosphate, pH 7.4). The coated slide was then immersed for 30 min in a solution of 5  $\mu\text{mol/l}$  rabbit gamma globulin solution in the same buffer to block residual binding sites on the metal surface. The slide was then incubated overnight in a solution of theophylline antiserum (raised in a rabbit against a theophylline-8-butryl-bovine serum albumin conjugate, essentially as described by T. Nishikawa, M. Saito and H. Kubo, Chem. Pharm. Bull, 32, 4951-7 (1984) diluted 1 in 500 in a buffer (50 mmol/l sodium phosphate/0.154 mol/l sodium chloride, pH 7.4, hereafter called PBS) which also contained 0.1% ovalbumin. The slide was then rinsed twice in PBS buffer containing 0.05% Tween 20, and twice in PBS, and stored until use in PBS. For use, the non-silvered surface was cleaned with isopropanol and the SPR properties of the slide determined before and after exposure to theophylline. Typical results are shown in Figure 4.

#### Example 5

Thyroxine. Tri-iodothyronine (T3) linked to disuccinimidyl suberate (DSS) was coupled to human gamma globulin to produce a T3-DSS-human gamma globulin conjugate.

A glass microscope slide covered on one side by a thin (approx 56 nm) film of silver was exposed for 5 minutes to a 1  $\mu\text{mol/l}$  solution of the conjugate in buffer (10 mmol/l sodium phosphate, pH7.4). The gamma globulin adsorbed to the silver surface and indirectly immobilised the T3 near to the surface. After washing with buffer, the slide was exposed for 5 minutes to a 5  $\mu\text{mol/l}$  solution of horse gamma globulin in buffer.

- 13 -

The gamma globulin forms a blocking coating and acts to minimise non-specific binding. After washing with buffer and then PBS, the slide was exposed for a further 5 minutes to a 1  $\mu\text{mol/l}$  solution of monoclonal anti-thyroxine (anti-T4) in PBS. This results in binding of the anti-T4 to the immobilised T3 (the anti-T4 exhibits approx 10% cross reactivity with T3). Unbound material was removed by rinsing in PBS.

Figure 5 shows the initial rate of change in reflectivity for different doses of added T4 in PBS + 0.5% bovine serum albumin + 0.2% anilino naphthalene sulfonic acid.

#### Example 6

Theophylline. A glass microscope slide covered on one side by a thin (approx 56 nm) film of silver was exposed for 5 minutes to a 1  $\mu\text{mol/l}$  solution of monoclonal anti-theophylline in buffer (10 mmol/l sodium phosphate, pH7.4). After washing with buffer, the slide was exposed for 5 minutes to a 5  $\mu\text{mol/l}$  solution of rabbit gamma globulin in buffer. The gamma globulin forms a blocking coating and acts to minimise non-specific binding. After washing with buffer and then PBS, the slide was exposed for a further 5 minutes to a 1  $\mu\text{mol/l}$  solution of a theophylline-rabbit gamma globulin conjugate (prepared as described in Example 4) in PBS + 0.1% ovalbumin. Unbound material was removed by rinsing in PBS.

Figure 6 shows the SPR response of the slide when exposed to 0 and 25  $\mu\text{g/ml}$  theophylline in PBS.

CLAIMS

1. An immobilised hapten comprising a metal surface which has immobilised thereon spacer units to which haptens are linked.
2. An immobilised hapten as claimed in claim 1, comprising a metal surface carrying a macromolecular coating comprising macromolecular spacer units to which haptens are covalently linked.
3. An immobilised hapten as claimed in claim 1 or claim 2, wherein the spacer units are protein molecules.
4. An immobilised hapten as claimed in any one of claims 1 to 3, wherein the metal surface is gold or silver.
5. An immobilised hapten as claimed in any one of claims 1 to 4, wherein corresponding antibodies are bound to the haptens.
6. A method of making the product of any one of claims 1 to 5, which method comprises linking haptens to spacer units to form conjugates and immobilising the conjugates on a metal surface, and thereafter applying a blocking agent to coat the metal surface.
7. A method as claimed in claim 6, wherein the blocking agent is a protein.
8. A method of assaying for an analyte in a sample, by the use of the analyte or an analogue thereof immobilised via spacer units on a metal surface, which method comprises bringing an antibody to the analyte, and the sample containing the analyte, into contact with the metal surface, and monitoring antibody binding to the immobilised analyte or analogue by means of surface plasmon resonance as indicative of the presence or the concentration of the analyte in the sample.
9. A method as claimed in claim 8, wherein the sample



is brought into contact with the metal surface, on which the antibody has previously been reversibly bound to the immobilised analyte or analogue, displacement of antibody being monitored as indicative of the presence or the concentration of the analyte in the sample.

5

10. A method of assaying for an analyte in a sample, by the use of a metal surface having immobilised thereon antibody to the analyte, which method comprises bringing a conjugate of the analyte or an analogue thereof, and the sample containing the analyte, into contact with the metal surface, and monitoring binding of the conjugate by means of surface plasmon resonance as indicative of the presence or concentration of the analyte in the sample.

10

11. A method as claimed in claim 10, wherein the conjugate of the analyte or analogue is with a protein.

15

12. A method as claimed in claim 10 or claim 11, wherein the sample is brought into contact with the metal surface, on which the conjugate has previously been reversibly bound to the immobilised antibody, displacement of the conjugate being monitored as indicative of the presence or the concentration of the analyte in the sample.

20

13. A method as claimed in any one of Claims 8 to 12, wherein the analyte is a hapten.

25

30

35

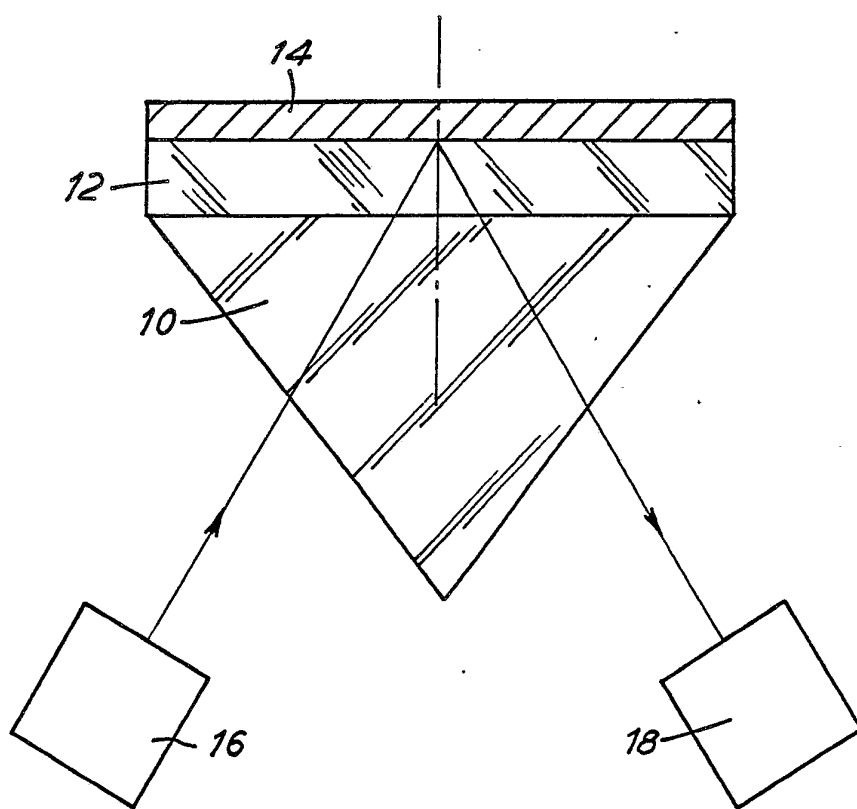
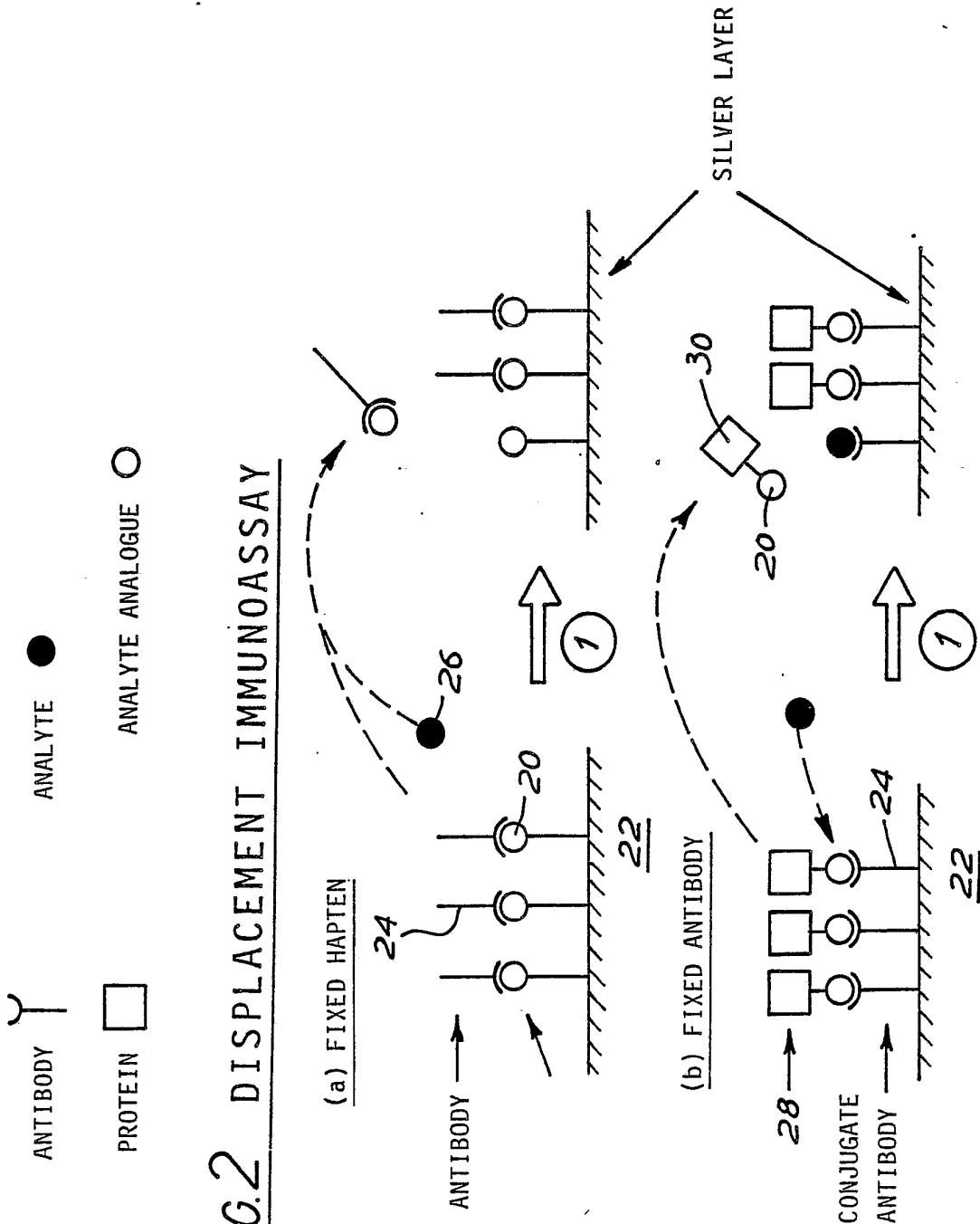


FIG.1



**FIG.2 DISPLACEMENT IMMUNOASSAY**

3/6

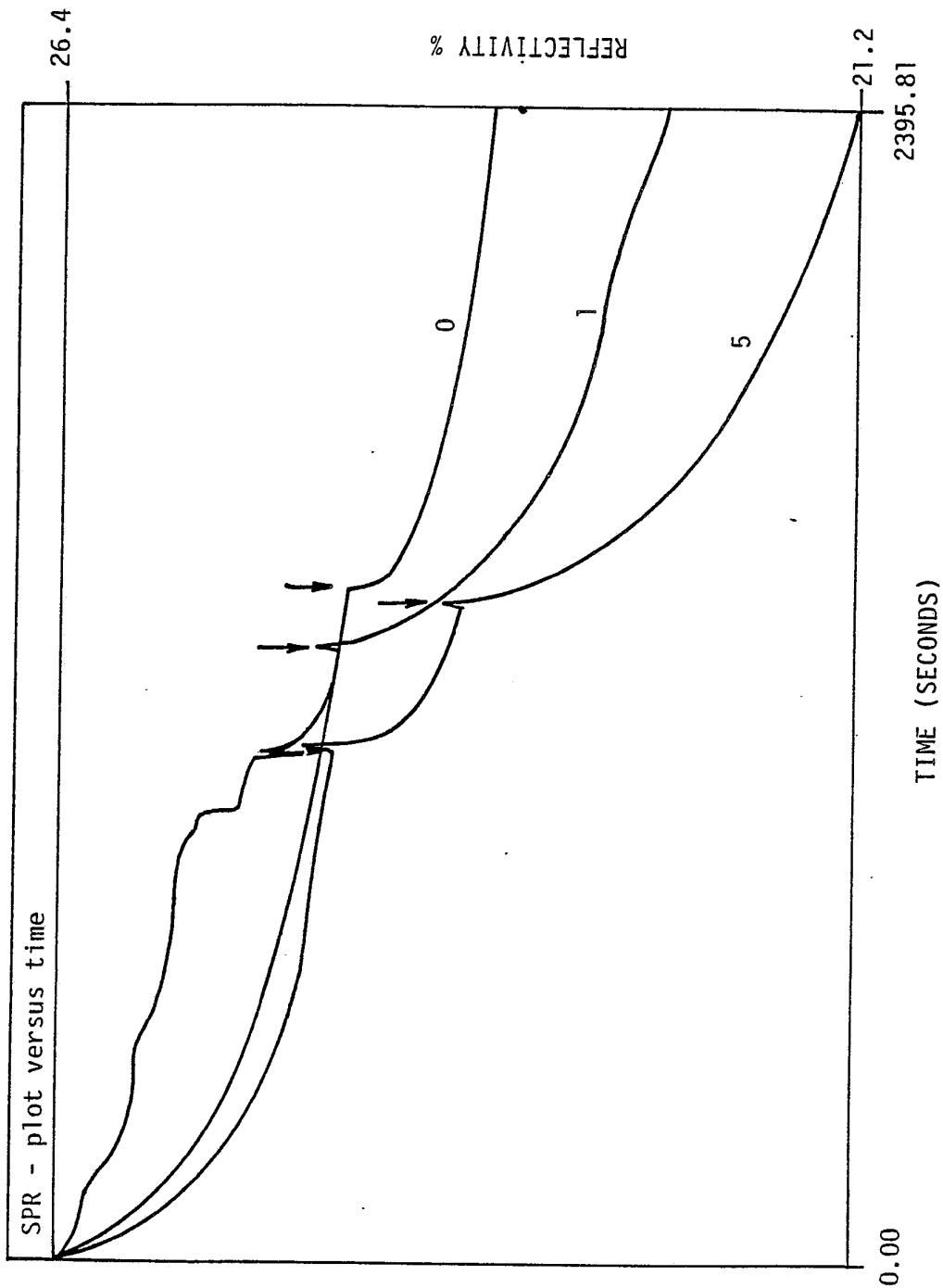


FIG. 3

4/6

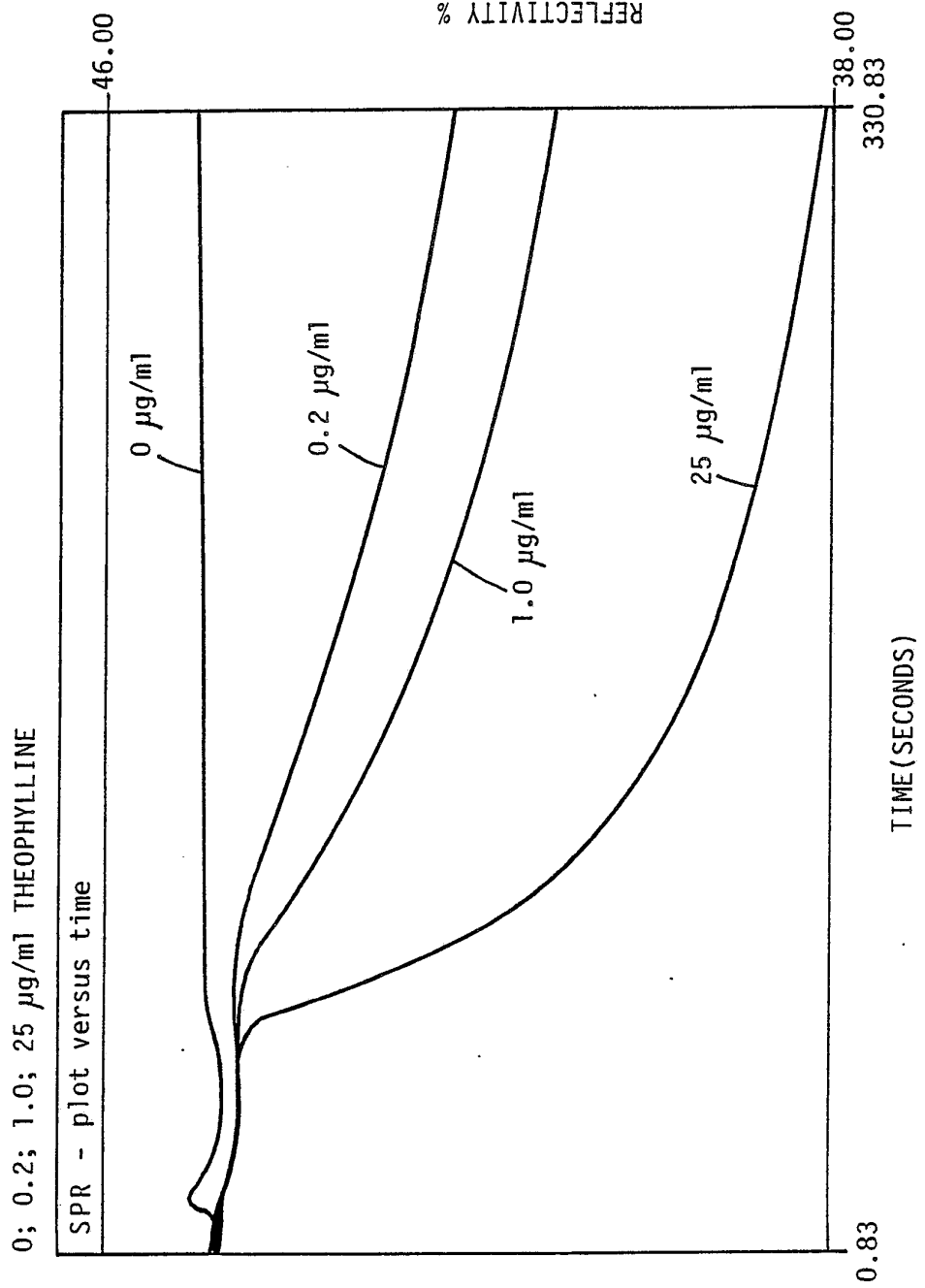
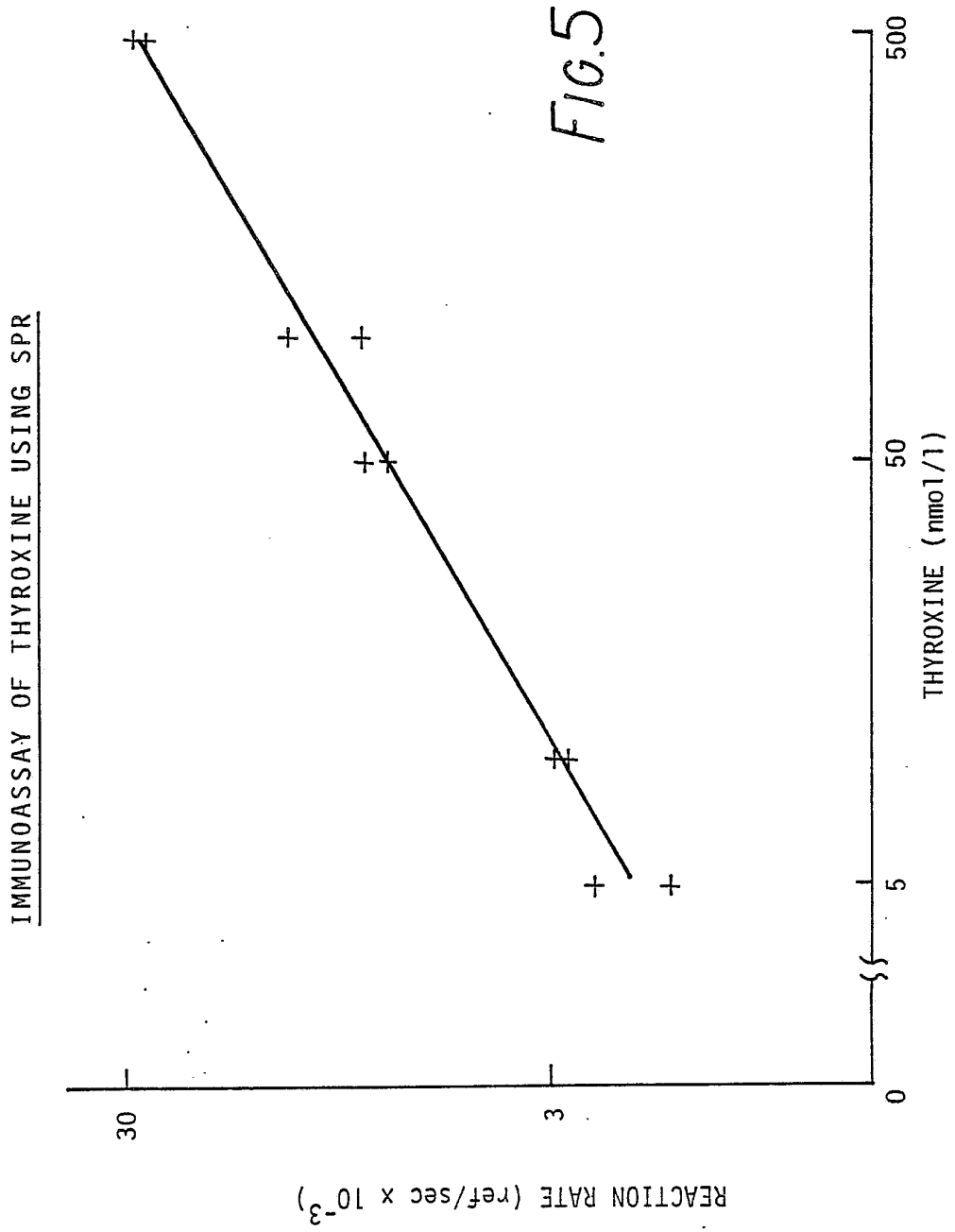


FIG.4

5/6



6/6

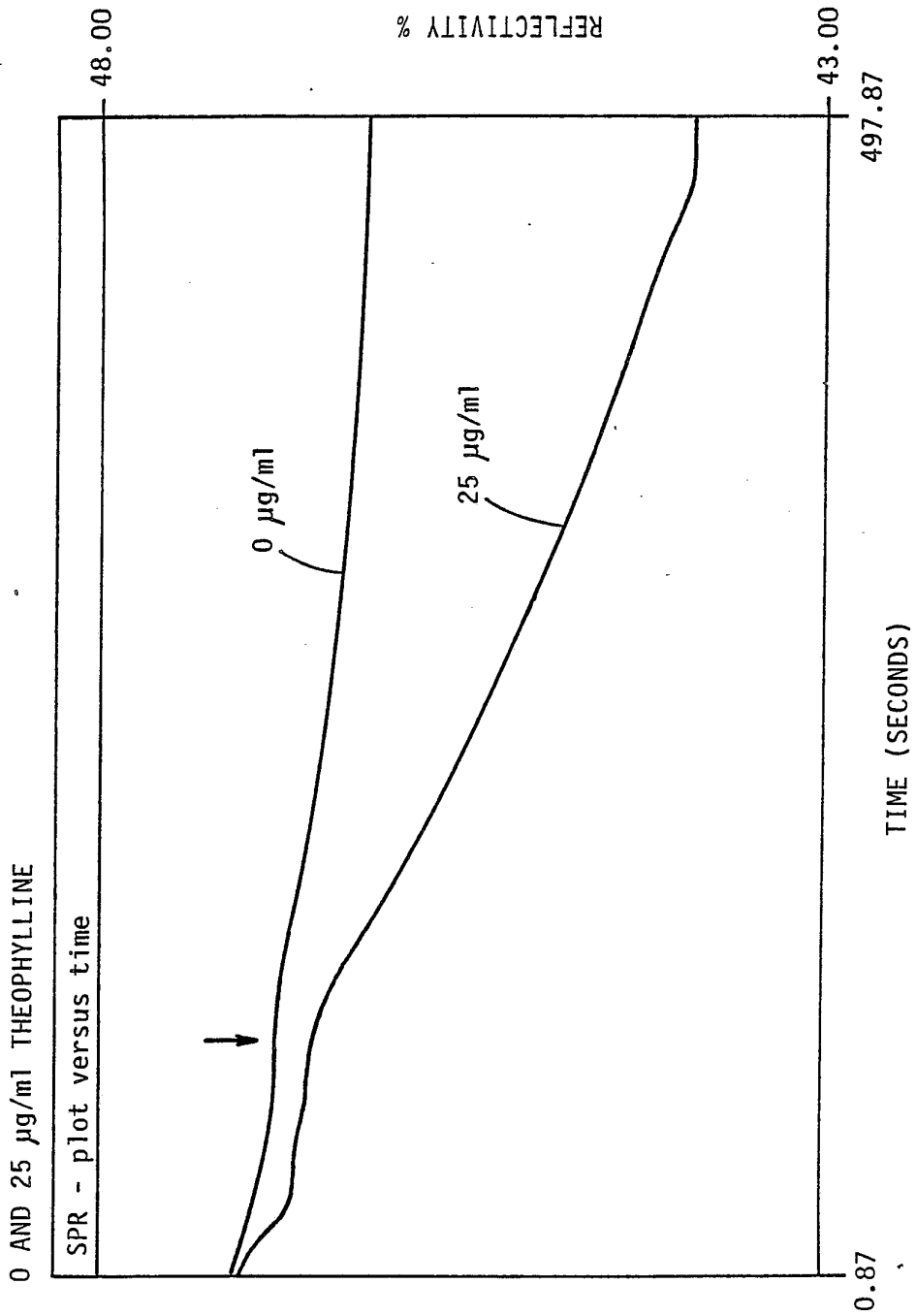



FIG.6

# INTERNATIONAL SEARCH REPORT

International Application No PCT/GB 89/00156

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>6</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC <sup>4</sup> G 01 N 33/553; G 01 N 33/543; G 01 N 21/55; //G 01 N 33/94; IPC : G 01 N 33/74		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>7</sup>		
Classification System	Classification Symbols	
IPC <sup>4</sup>	G 01 N	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched <sup>8</sup>		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT</b> <sup>9</sup>		
Category <sup>10</sup>	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
X	EP, A, 0254575 (ARES-SERONO RESEARCH & DEVELOPMENT) 27 January 1988 see page 1, line 1 - page 6, line 4	1, 2, 4, 5
Y	--	3, 6, 7
Y	EP, A, 0007654 (AKZO N.V.) 6 February 1980 see page 5, lines 8-17; page 7, line 17 - page 9, line 18	3, 6, 7
Y	--	1-6
Y	Sensors and Actuators, vol. 4, 1983, Elsevier Sequoia (NL), B. Liedberg et al.: "Surface plasmon resonance for gas detection and biosensing", pages 299-304 see page 304, lines 4-5	1-6
Y,P	--	1-6
Y,P	Journal of Immunological Methods, vol. 115, 1988, Elsevier Science Publishers B.V. (Biomedical Division),	1-6
	./.	
<p><sup>10</sup> Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&amp;" document member of the same patent family</p>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
12th June 1989	30 JUN 1989	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	 P.C.G. VAN DER PUTTEN	



III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category*	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
	M. Werthen et al.: "Effect of antibody affinity on the isotherm of antibody binding to surface-immobilized antigen", pages 71-78 see abstract --	
X	WO, A, 87/03095 (THE JOHN HOPKINS UNIVERSITY/APPLIED PHYSICS LAB.) 21 May 1987 see page 4, line 31 - page 5, line 10; page 10, lines 23-31; page 13, line 35 - page 15, line 12; page 24, line 23 - page 27, line 7	1,2,5
Y	--	4,8,9,13
Y,P	Sensors and Actuators, vol. 15, no. 1, September 1988, Elsevier Sequoia, (Lausanne, CH), P.B. Daniels et al.: "Surface plasmon resonance applied to immunosensing", pages 11-18 see page 11 - page 13, line 16 --	4,8,9,13
Y	US, A, 4070246 (J.F. KENNEDY et al.) 24 January 1978 see abstract; column 4, lines 9-30 --	1-3
Y	EP, A, 0028132 (DYNASCIENCES CORP.) 6 May 1981 see abstract; page 5, line 1 - page 6, line 8 -----	1-3

**ANNEX TO THE INTERNATIONAL SEARCH REPORT  
ON INTERNATIONAL PATENT APPLICATION NO.**

GB 8900156  
SA 27523

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 20/06/89. The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A- 0254575	27-01-88	AU-A- 7600787	28-01-88
		JP-A- 63100355	02-05-88
EP-A- 0007654	06-02-80	NL-A- 7807532	15-01-80
		AT-T- 786	15-04-82
		AU-B- 530217	07-07-83
		AU-A- 4883479	17-01-80
		CA-A- 1135183	09-11-82
		JP-A- 55015100	01-02-80
		US-A- 4313734	02-02-82
WO-A- 8703095	21-05-87	EP-A- 0245477	19-11-87
		JP-T- 63501446	02-06-88
US-A- 4070246	24-01-78	None	
EP-A- 0028132	06-05-81	AU-B- 533026	27-10-83
		AU-A- 6295280	30-04-81
		CA-A- 1146853	24-05-83
		JP-A- 56071023	13-06-81
		US-A- 4410634	18-10-83