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Description

The present invention relates to methods, as defined in the claims, for constructing libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family. In a preferred embodiment, the displayed polypeptides are human Fabs.

More specifically, the methods of the invention comprise cleaving single-stranded nucleic acids at chosen locations, the cleaved nucleic acids encoding, at least in part, the peptides, polypeptides or proteins displayed on the genetic packages of the libraries of the invention. In a preferred embodiment, the genetic packages are filamentous phage or phagemids.

The present specification describes methods of screening the libraries of genetic packages that display useful peptides, polypeptides and proteins and to the peptides, polypeptides and proteins identified by such screening.

BACKGROUND OF THE INVENTION

It is now common practice in the art to prepare libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family. In many common libraries, the displayed peptides, polypeptides or proteins are related to antibodies. Often, they are Fabs or single chain antibodies.

In general, the DNAs that encode members of the families to be displayed must be amplified before they are cloned and used to display the desired member on the surface of a genetic package. Such amplification typically makes use of forward and backward primers.

Such primers can be complementary to sequences native to the DNA to be amplified or complementary to oligonucleotides attached at the 5' or 3' ends of that DNA. Primers that are complementary to sequences native to the DNA to be amplified are disadvantaged in that they bias the members of the families to be displayed. Only those members that contain a sequence in the native DNA that is substantially complementary to the primer will be amplified. Those that do not will be absent from the family. For those members that are amplified, any diversity within the primer region will be suppressed.

For example, in European patent 368, 684 B1, the primer that is used is at the 5' end of the V_H region of an antibody gene. It anneals to a sequence region in the native DNA that is said to be "sufficiently well conserved" within a single species. Such primer will bias the members amplified to those having this "conserved" region. Any diversity within this region is extinguished.

It is generally accepted that human antibody genes arise through a process that involves a combinatorial selection of V and J or V, D, and J followed by somatic mutations. Although most diversity occurs in the Complementary Determining Regions (CDRs), diversity also occurs in the more conserved Framework Regions (FRs) and at least some of this diversity confers or enhances specific binding to antigens (Ag). As a consequence, libraries should contain as much of the CDR and FR diversity as possible.

To clone the amplified DNAs for display on a genetic package of the peptides, polypeptides or proteins that they encode, the DNAs must be cleaved to produce appropriate ends for ligation to a vector. Such cleavage is generally effected using restriction endonuclease recognition sites carried on the primers. When the primers are at the 5' end of DNA produced from reverse transcription of RNA, such restriction leaves deleterious 5' untranslated regions in the amplified DNA. These regions interfere with expression of the cloned genes and thus the display of the peptides, polypeptides and proteins coded for by them.

Zhu D., Analytical Biochemistry, Vol. 177(1), 1989, pages 120-124, describes oligodeoxynucleotide-directed cleavage and repair of a single-stranded vector a method of site-specific mutagenesis.

Thielking V et al., Biochemistry, Vol. 29(19), 1990, pages 4682-4691, describes the accuracy of the ECO-RI restriction endonuclease binding and cleavage studies with oligodeoxynucleotide substrates containing degenerate recognition sequences.

Alves Juergen et al., Biochemistry, Vol. 34(35), 1995, pages 11191-11197, describes the accuracy of the EcoRV restriction endonuclease in binding and cleavage studies with oligodeoxynucleotide substrates containing degenerate recognition sequences.

Kim S.C. et al., Science, Vol. 240, No. 4851, 1988, pages 504-506, describes cleaving DNA at any predetermined site with adapter-primers and class-IIS restriction enzymes.

Podhajski A.J. & Szybalski W., Gene, Vol. 40(2-3), 1985, pages 175-182, describes conversion of the FOK-I endonuclease to a universal restriction enzyme cleavage of phage M-13-MP-7 DNA at predetermined sites.

WO 97/20923 describes preparation of a multi-combinatorial library of antibody gene expression vectors.

WO 97/49809 describes polypeptides capable of forming antigen binding structures with specificity for the Rhesus D antigens, the DNA encoding them and the process for their preparation and use.

SUMMARY OF THE INVENTION

It is an object of this invention to provide novel methods for constructing libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of this family. These methods are not biased toward DNAs that contain native sequences that are complementary to the primers used for amplification. They also enable any sequences that may be deleterious to expression to be removed from the amplified DNA before cloning and displaying.

The specification describes a method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:

(i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and

(ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

The specification describes an alternative method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:

(i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and

(ii) cleaving the nucleic acid solely at the cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

The specification describes a method of capturing DNA molecules that comprise a member of a diverse family of DNAs and collectively comprise at least a portion of the diversity of the family. These DNA molecules in single-stranded form have been cleaved by one of the methods of this invention. This method involves ligating the individual single-stranded DNA members of the family to a partially duplex DNA complex. The method comprises the steps of:

(i) contacting a single-stranded nucleic acid sequence that has been cleaved with a restriction endonuclease with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region that remains after cleavage, the double-stranded region of the oligonucleotide including any sequences necessary to return the sequences that remain after cleavage into proper reading frame for expression and containing a restriction endonuclease recognition site 5' of those sequences; and

(ii) cleaving the partially double-stranded oligonucleotide sequence solely at the restriction endonuclease recognition site contained within the double-stranded region of the partially double-stranded oligonucleotide.

It is another object of this invention to prepare libraries, that display a diverse family of peptides, polypeptides or proteins and collectively display at least part of the diversity of the family, using the methods and DNAs described above, as defined in the claims.

It is an object to screen those libraries to identify useful peptides, polypeptides and proteins and to use those substances in human therapy.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a schematic of various methods that may be employed to amplify VH genes without using primers specific for VH sequences.

FIG. 2 is a schematic of various methods that may be employed to amplify VL genes without using VL sequences.

FIG. 3 depicts gel analysis of cleaved kappa DNA from Example 2.

FIG. 4 depicts gel analysis of cleaved kappa DNA from Example 2.

FIG. 5 depicts gel analysis of amplified kappa DNA from Example 2.

FIG. 6 depicts gel purified amplified kappa DNA from Example 2.

TERMS

In this application, the following terms and abbreviations are used:

Sense strand	The upper strand of ds DNA as usually written. In the sense strand, 5'-ATG-3' codes for Met.
Antisense strand	The lower strand of ds DNA as usually written. In the antisense strand, 3'-TAC-5' would correspond to a Met codon in the sense strand.
Forward primer:	A "forward" primer is complementary to a part of the sense strand and primes for synthesis of a new antisense-strand molecule. "Forward primer" and "lower-strand primer" are equivalent.
Backward primer:	A "backward" primer is complementary to a part of the antisense strand and primes for synthesis of a new sense-strand molecule. "Backward primer" and "top-strand primer" are equivalent.
Bases:	Bases are specified either by their position in a vector or gene as their position within a gene by codon and base. For example, "89.1" is the first base of codon 89, 89.2 is the second base of codon 89.
Sv	Streptavidin
Ap	Ampicillin
ap ^R	A gene conferring ampicillin resistance.
RE	Restriction endonuclease
URE	Universal restriction endonuclease
Functionally complementary	Two sequences are sufficiently complementary so as to anneal under the chosen conditions.
RERS	Restriction endonuclease recognition site
AA	Amino acid

PCR	Polymerization chain reaction
GLGs	Germline genes
Ab	Antibody: an immunoglobulin. The term also covers any protein having a binding domain which is homologous to an immunoglobulin binding domain. A few examples of antibodies within this definition are, <i>inter alia</i> , immunoglobulin isotypes any the Fab, F(ab') ₂ , scFv, Fv, dAb and Fd fragments.
Fab	Two chain molecule comprising an Ab light chain and part of a heavy-chain.
scFv	A single-chain Ab comprising either VH: :linker: :VL or VL: :linker: :VH
w.t.	Wild type
HC	Heavy chain
LC	Light chain
VK	A variable domain of a Kappa light chain.
VH	A variable domain of a heavy chain.
VL	A variable domain of a lambda light chain.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The nucleic acid sequences that are useful in the methods of this invention, i.e., those that encode at least in part the individual peptides, polypeptides and proteins displayed on the genetic packages of this invention, may be naturally occurring, synthetic or a combination thereof. They may be mRNA, DNA or cDNA. In the preferred embodiment, the nucleic acids encode antibodies. Most preferably, they encode Fabs.

The nucleic acids useful in this invention may be naturally diverse, synthetic diversity may be introduced into those naturally diverse members, or the diversity may be entirely synthetic. For example, synthetic diversity can be introduced into one or more CDRs of antibody genes.

Synthetic diversity may be created, for example, through the use of TRIM technology (U.S. 5,869,644). TRIM technology allows control over exactly which amino-acid types are allowed at variegated positions and in what proportions. In TRIM technology, codons to be diversified are synthesized using mixtures of trinucleotides. This allows any set of amino acid types to be included in any proportion.

Another alternative that may be used to generate diversified DNA is mixed oligonucleotide synthesis. With TRIM technology, one could allow Ala and Trp. With mixed oligonucleotide synthesis, a mixture that included Ala and Trp would also necessarily include Ser and Gly. The amino-acid types allowed at the variegated positions are picked with reference to the structure of antibodies, or other peptides, polypeptides or proteins of the family, the observed diversity in germline genes, the observed somatic mutations frequently observed, and the desired areas and types of variegation.

In a preferred embodiment of this invention, the nucleic acid sequences for at least one CDR or other region of the peptides, polypeptides or proteins of the family are cDNAs produced by reverse transcription from mRNA. More preferably, the mRNAs are obtained from peripheral blood cells, bone marrow cells, spleen cells or lymph node cells (such as B-lymphocytes or plasma cells) that express members of naturally diverse sets of related genes. More preferably, the mRNAs encode a diverse family of antibodies. Most preferably, the mRNAs are obtained from patients suffering from at least one autoimmune disorder or cancer. Preferably, mRNAs containing a high diversity of autoimmune diseases, such as systemic lupus erythematosus, systemic sclerosis, rheumatoid arthritis, antiphospholipid syndrome and vasculitis are used.

In a preferred embodiment of this invention, the cDNAs are produced from the mRNAs using reverse transcription. In this preferred embodiment, the mRNAs are separated from the cell and degraded using standard methods, such that only the full length (i.e., capped) mRNAs remain. The cap is then removed and reverse transcription used to produce the cDNAs.

The reverse transcription of the first (antisense) strand can be done in any manner with any suitable primer. See, *e.g.*, HJ de Haard et al., *Journal of Biological Chemistry*, 274(26):18218-30 (1999). In the preferred embodiment of this invention where the mRNAs encode antibodies, primers that are complementary to the constant regions of antibody genes may be used. Those primers are useful because they do not generate bias toward subclasses of antibodies. In another embodiment, poly-dT primers may be used (and may be preferred for the heavy-chain genes). Alternatively, sequences complementary to the primer may be attached to the termini of the antisense strand.

In one preferred embodiment of this invention, the reverse transcriptase primer may be biotinylated, thus allowing the cDNA product to be immobilized on streptavidin (Sv) beads. Immobilization can also be effected using a primer labeled at the 5' end with one of a free amine group, b) thiol, c) carboxylic acid, or d) another group not found in DNA that can react to form a strong bond to a known partner on an insoluble medium. If, for example, a free amine (preferably primary amine) is provided at the 5' end of a DNA primer, this amine can be reached with carboxylic acid groups on a polymer bead using standard amide-forming chemistry. If such preferred immobilization is used during reverse transcription, the top strand RNA is degraded using well-known enzymes, such as a combination of RNaseH and RNaseA, either before or after immobilization.

The nucleic acid sequences useful in the methods of this invention are generally amplified before being used to display the peptides, polypeptides or proteins that they encode. Prior to amplification, the single-stranded DNAs may be cleaved using either of the methods described before. Alternatively, the single-stranded DNAs may be amplified and then cleaved using one of those methods.

Any of the well known methods for amplifying nucleic acid sequences may be used for such amplification. Methods that maximize, and do not bias, diversity are preferred. In a preferred embodiment of this invention where the nucleic acid sequences are derived from antibody genes, the present invention preferably utilizes primers in the constant regions of the heavy and light chain genes and primers to a synthetic sequence that are attached at the 5' end of the sense strand. Priming at such synthetic sequence avoids the use of sequences within the

variable regions of the antibody genes. Those variable region priming sites generate bias against V genes that are either of rare subclasses or that have been mutated at the priming sites. This bias is partly due to suppression of diversity within the primer region and partly due to lack of priming when many mutations are present in the region complementary to the primer. The methods disclosed in this invention have the advantage of not biasing the population of amplified antibody genes for particular V gene types.

The synthetic sequences may be attached to the 5' end of the DNA strand by various methods well known for ligating DNA sequences together. RT CapExtension is one preferred method.

In RT CapExtension (derived from Smart PCR[™]), a short overlap (5'...GGG-3' in the upper-strand primer (USP-GGG) complements 3'-CCC...5' in the lower strand) and reverse transcriptases are used so that the reverse complement of the upper-strand primer is attached to the lower strand.

In a preferred embodiment of this invention the upper strand or lower strand primer may be also biotinylated or labeled at the 5' end with one of a) free amino group, b) thiol, c) carboxylic acid and d) another group not found in DNA that can react to form a strong bond to a known partner as an insoluble medium. These can then be used to immobilize the labeled strand after amplification. The immobilized DNA can be either single or double-stranded.

FIG. 1 shows a schematic of the amplification of VH genes. FIG. 1, Panel A shows a primer specific to the poly-dT region of the 3' UTR priming synthesis of the first, lower strand. Primers that bind in the constant region are also suitable. Panel B shows the lower strand extended at its 3' end by three Cs that are not complementary to the mRNA. Panel C shows the result of annealing a synthetic top-strand primer ending in three GGGs that hybridize to the 3' terminal CCCs and extending the reverse transcription extending the lower strand by the reverse complement of the synthetic primer sequence. Panel D shows the result of PCR amplification using a 5' biotinylated synthetic top-strand primer that replicates the 5' end of the synthetic primer of panel C and a bottom-strand primer complementary to part of the constant domain. Panel E shows immobilized double-stranded (ds) DNA obtained by using a 5'-biotinylated top-strand primer.

FIG. 2 shows a similar schematic for amplification of VL genes. FIG. 2, Panel A shows a primer specific to the constant region at or near the 3' end priming synthesis of the first, lower strand. Primers that bind in the poly-dT region are also suitable. Panel B shows the lower strand extended at its 3' end by three Cs that are not complementary to the mRNA. Panel C shows the result of annealing a synthetic top-strand primer ending in three GGGs that hybridize to the 3' terminal CCCs and extending the reverse transcription extending the lower strand by the reverse complement of the synthetic primer sequence. Panel D shows the result of PCR amplification using a 5' biotinylated synthetic top-strand primer that replicates the 5' end of the synthetic primer of panel C and a bottom-strand primer complementary to part of the constant domain. The bottom-strand primer also contains a useful restriction endonuclease site, such as *Asc*I. Panel E shows immobilized ds cDNA obtained by using a 5'-biotinylated top-strand primer.

In FIGS. 1 and 2, each V gene consists of a 5' untranslated region (UTR) and a secretion signal, followed by the variable region, followed by a constant region, followed by a 3' untranslated region (which typically ends in poly-A). An initial primer for reverse transcription may be complementary to the constant region or to the poly A segment or the 3'-UTR. For human heavy-chain genes, a primer of 15 T is preferred. Reverse transcriptases attach several C residues to the 3' end of the newly synthesized DNA. RT CapExtension exploits this feature. The reverse transcription reaction is first run with only a lower-strand primer. After about 1 hour, a primer ending in GGG (USP-GGG) and more RTase are added. This causes the lower-strand cDNA to be extended by the reverse complement of the USP-GGG up to the final GGG. Using one primer identical to part of the attached synthetic sequence and a second primer complementary to a region of known sequence at the 3' end or the sense strand, all the V genes are amplified irrespective of their V gene subclass.

After amplification, the DNAs of this invention are rendered single-stranded. For example, the strands can be separated by using a biotinylated primer, capturing the biotinylated product on streptavidin beads, denaturing the DNA, and washing away the complementary strand. Depending on which end of the captured DNA is wanted, one will choose to immobilize either the upper (sense) strand or the lower (antisense) strand.

To prepare the single-stranded amplified DNAs for cloning into genetic packages so as to effect display of the peptides, polypeptides or proteins encoded, at least in part, by those DNAs, they must be manipulated to provide ends suitable for cloning and expression. In particular, any 5' untranslated regions and mammalian signal sequences must be removed and replaced, in frame, by a suitable signal sequence that functions in the display host. Additionally, parts of the variable domains (in antibody genes) may be removed and replaced by synthetic segments containing synthetic diversity. The diversity of other gene families may likewise be expanded with synthetic diversity.

According to the methods of this invention, there are two ways to manipulate the single-stranded amplified DNAs for cloning, as defined in the claims. The first method comprises the steps of:

- (i) contacting the single-stranded nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being complementary to the single-stranded nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

In this first method, short oligonucleotides are annealed to the single-stranded DNA so that restriction endonuclease recognition sites formed within the now locally double-stranded regions of the DNA can be cleaved. In particular, a recognition site that occurs at the same position in a substantial fraction of the single-stranded DNAs is identical.

For antibody genes, this can be done using a catalog of germline sequences. See, e.g., "<http://www.mrc-cpe.cam.ac.uk/imt-doc/restricted/ok.htm> 1." Updates can be obtained from this site under the heading "Amino acid and nucleotide sequence alignments." For other families, similar comparisons exist and may be used to select appropriate regions for cleavage and to maintain diversity.

For example, Table 195 depicts the DNA sequences of the FR3 regions of the 51 known human VH germline genes. In this region, the genes contain restriction endonuclease recognition sites shown in Table 200. Restriction endonucleases that cleave a large fraction of germline

genes at the same site are preferred over endonuclease that cut at a variety of sites. Furthermore, it is preferred that there be only one site for the restriction endonucleases within the region to which the short oligonucleotide binds on the single-stranded DNA, e.g., about 10 bases on either side of the restriction endonuclease recognition site.

An enzyme that cleaves downstream in FR3 is also more preferable because it captures fewer mutations in the framework. This may be advantageous in some cases. However, it is well known that framework mutations exist and confer and enhance antibody binding. The present invention, by choice of appropriate restriction site, allows all or part of FR3 diversity to be captured. Hence, the method also allows extensive diversity to be captured.

Finally, in the methods of this invention restriction endonucleases that are active between about 45° and about 75°C are used. Preferably enzymes that are active above 50°C, and more preferably active about 55°C, are used. Such temperatures maintain the nucleic acid sequence to be cleaved in substantially single-stranded form.

Enzymes shown in Table 200 that cut many of the heavy chain FR3 germline genes at a single position include: *MaeIII*(24@4), *Tsp45I*(21@4), *HphI*(44@5), *BsaI*(23@65), *AluI*(23@47), *BlpI*(21@48), *DdeI*(29@58), *BglII*(10@61), *MslI*(44@72), *BsiEI*(23@74), *EaeI*(23@74), *EagI*(23@74), *HaeIII*(25@75), *Bst4CI*(51@36), *HpyCH4III*(51@86), *HinI*(38@2), *MlyI*(18@2), *PleI*(18@2), *MnlI*(31@67), *HpyCH4V*(21@44), *BsmAI*(16@11), *BpmI*(19@12), *XmnI*(12@30), and *SacI*(11@51). (The notation used means, for example, that *BsmAI* cuts 16 of the FR3 germline genes with a restriction endonuclease recognition site beginning at base 11 of FR³.)

For cleavage of human heavy chains in FR3, the preferred restriction endonucleases are: *Bst4CI* (or *TaaI* or *HpyCH4III*), *BlpI*, *HpyCH4V*, and *MslI*. Because ACNGT (the restriction endonuclease recognition site for *Bst4CI*, *TaaI*, and *HpyCH4III*) is found at a consistent site in all the human FR3 germline genes, one of those enzymes is the most preferred for capture of heavy chain CDR3 diversity. *BlpI* and *HpyCH4V* are complementary. *BlpI* cuts most members of the VH1 and VH4 families while *HpyCH4V* cuts most members of the VH3, VH5, VH6, and VH7 families. Neither enzyme cuts VH2s, but this is a very small family, containing only three members. Thus, these enzymes may also be used in preferred embodiments of the methods of this invention.

The restriction endonucleases *HpyCH4III*, *Bst4CI*, and *TaaI* all recognize 5'-ACnGT-3' and cut upper strand DNA after n and lower strand DNA before the base complementary to n. This is the most preferred restriction endonuclease recognition site for this method on human heavy chains because it is found in all germline genes. Furthermore, the restriction endonuclease recognition region (ACnGT) matches the second and third bases of a tyrosine codon (tay) and the following cysteine codon (tgy) as shown in Table 206. These codons are highly conserved, especially the cysteine in mature antibody genes.

Table 250 E shows the distinct oligonucleotides of length 22 (except the last one which is of length 20) bases. Table 255 C shows the analysis of 1617 actual heavy chain antibody genes. Of these, 1511 have the site and match one of the candidate oligonucleotides to within 4 mismatches. Eight oligonucleotides account for most of the matches and are given in Table 250 F.1. The 8 oligonucleotides are very similar so that it is likely that satisfactory cleavage will be achieved with only one oligonucleotide (such as H43.77.97.1-02#1) by adjusting temperature, pH, salinity, and the like. One or two oligonucleotides may likewise suffice whenever the germline gene sequences differ very little and especially if they differ very little close to the restriction endonuclease recognition region to be cleaved. Table 255 D shows a repeat analysis of 1617 actual heavy chain antibody genes using only the 8 chosen oligonucleotides. This shows that 1463 of the sequences match at least one of the oligonucleotides to within 4 mismatches and have the site as expected. Only 7 sequences have a second *HpyCH4III* restriction endonuclease recognition region in this region.

Another illustration of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of human heavy chains. Cleavage in FR1 allows capture of the entire CDR diversity of the heavy chain.

The germline genes for human heavy chain FR1 are shown in Table 217. Table 220 shows the restriction endonuclease recognition sites found in human germline genes FR1s. The preferred sites are *BsgI*(GTGCAG;39@4), *BsoFI*(GCngc;43@6,11@9,2@3,1@12), *TseI*(Gcwg;43@6,11@9,2@3,1@12), *MspAII*(CMGckg;46@7,2@1), *PvuII*(CAGctg;46@7,2@1), *AluI*(AGct;48@82@2), *DdeI*(Ctnag;22@52,9@48), *HphI* (tcacc; 22@80), *BssKI*(Nccngg;35@39,2@40), *BsaJI* (Ccnngg; 32@40, 2@41), *BstNI*(CCwgg; 33@40), *ScrFI* (CCngg;35@40, 2@41), *EcoO109I*(RGgnccy;22@46, 11@43), *Sau96I*(Ggncc;23@47,11@44), *AvaII* (Ggwcc; 23@47, 4@44), *PpuMI*(RGgwccy;22@46,4@43), *BsmFI*(gtccc;20@48), *HinI*(Gantc;34@16,21@56, 21@77), *TfiI*(21@77), *MylI*(GAGTC;34@16), *MlyI*(gactc;21@56), and *AlwNI*(CAGnnctg;22@68). The more preferred sites are *MspAI* and *FvuII*. *MspAI* and *PvuII* have 46 sites at 7-12 and 2 at 1-6. To avoid cleavage at both sites, oligonucleotides are used that do not fully cover the site at 1-6. Thus, the DNA will not be cleaved at that site. We have shown that DNA that extends 3, 4, or 5 bases beyond a *PvuII*-site can be cleaved efficiently.

Another illustration of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of human kappa light chains. Table 300 shows the human kappa FR1 germline genes and Table 302 shows restriction endonuclease recognition sites that are found in a substantial number of human kappa FR1 germline genes at consistent locations. Of the restriction endonuclease recognition sites listed, *BsmAI* and *PfFI* are the most preferred enzymes. *BsmAI* sites are found at base 18 in 35 of 40 germline genes. *PfFI* sites are found in 35 of 40 germline genes at base 12.

Another example of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of the human lambda light chain. Table 400 shows the 31 known human lambda FR1 germline gene sequences. Table 405 shows restriction endonuclease recognition sites found in human lambda FR1 germline genes. *HinI* and *DdeI* are the most preferred restriction endonuclease for cutting human lambda chains in FR1.

After the appropriate site or sites for cleavage are chosen, one or more short oligonucleotides are prepared so as to functionally complement, alone or in combination, the chosen recognition site. The oligonucleotides also include sequences that flank the recognition site in the majority of the amplified genes. This flanking region allows the sequence to anneal to the single-stranded DNA sufficiently to allow cleavage by the restriction endonuclease specific for the site chosen.

The actual length and sequence of the oligonucleotide depends on the recognition site and the conditions to be used for contacting and cleavage. The length must be sufficient so that the oligonucleotide is functionally complementary to the single-stranded DNA over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and solely at the desired location.

Typically, the oligonucleotides of this preferred method of the invention are about 17 to about 30 nucleotides in length. Below about 17 bases, annealing is too weak and above 30 bases there can be a loss of specificity. A preferred length is 18 to 24 bases.

Oligonucleotides of this length need not be identical complements of the germline genes. Rather, a few mismatches taken may be tolerated. Preferably, however, no more than 1-3 mismatches are allowed. Such mismatches do not adversely affect annealing of the oligonucleotide to the single-stranded DNA. Hence, the two DNAs are said to be functionally complementary.

The second method to manipulate the amplified single-stranded DNAs of this invention for cloning comprises the steps (as defined in the claims) of:

- (i) contacting the single-stranded nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being complementary to the single-stranded nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and
- (ii) cleaving the nucleic acid solely at the cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and aft the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

This second method employs Universal Restriction Endonucleases ("URE"). UREs are partially double-stranded oligonucleotides, The single-stranded portion or overlap of the URE consists of a DNA adapter that is functionally complementary to the sequence to be cleaved in the single-stranded DNA. The double-stranded, portion consists of a type II-S restriction endonuclease recognition site.

The URE method of this invention is specific and precise and can tolerate some (e.g., 1-3) mismatches in the complementary regions, i.e., it is functionally complementary to that region. Further, conditions under which the URE is used can be adjusted so that most of the genes that are amplified can be cut, reducing bias in the library produced from those genes.

The sequence of the single-stranded DNA adapter or overlap portion of the URE typically consists of about 14-22 bases. However, longer or shorter adapters may be used. The size depends on the ability of the adapter to associate with its functional complement in the single-stranded DNA and the temperature used for contacting the URE and the single-stranded DNA at the temperature used for cleaving the DNA with the type II-S enzyme. The adapter must be functionally complementary to the single-stranded DNA over a large enough region to allow the two strands to associate such that the cleavage may occur at the chosen temperature and at the desired location. We prefer single-stranded or overlap portions of 14-17 bases in length, and more preferably 18-20 bases in length.

The site chosen for cleavage using the URE is preferably one that is substantially conserved in the family of amplified DNAs. As compared to the first cleavage method of this invention, these sites do not need to be endonuclease recognition sites. However, like the first method, the sites chosen can be synthetic rather than existing in the native DNA. Such sites may be chosen by references to the sequences of known antibodies or other families of genes. For example, the sequences of many germline genes are reported at <http://www.mrc-cpe.cam.ac.uk/mt-doc/restricted/ok.html>. For example, one preferred site occurs near the end of FR3 -- codon 89 through the second base of codon 93. CDR3 begins at codon 95.

The sequences of 79 human heavy-chain genes are also available at <http://www.ncbi.nlm.nih.gov/entre2/nucleotide.html>. This site can be used to identify appropriate sequences for URE cleavage according to the methods of this invention. See, e.g., Table 8B.

Most preferably, one or more sequences are identified using these sites or other available sequence information. These sequences together are present in a substantial fraction of the amplified DNAs. For example, multiple sequences could be used to allow for known diversity in germline genes or for frequent somatic mutations. Synthetic degenerate sequences could also be used. Preferably, a sequence (s) that occurs in at least 65% of genes examined with no more than 2-3 mismatches is chosen

URE single-stranded adapters or overlaps are then made to be complementary to the chosen regions.

Conditions for using the UREs are determined empirically. These conditions should allow cleavage of DNA that contains the functionally complementary sequences with no more than 2 or 3 mismatches but that do not allow cleavage of DNA lacking such sequences.

As described above, the double-stranded portion of the URE includes a Type II-S endonuclease recognition site. Any Type II-S enzyme that is active at a temperature necessary to maintain the single-stranded DNA substantially in that form and to allow the single-stranded DNA adapter portion of the URE to anneal long enough to the single-stranded DNA to permit cleavage at the desired site may be used.

The preferred Type II-S enzymes for use in the URE methods of this invention provide asymmetrical cleavage of the single-stranded DNA. Among these are the enzymes listed in Table 800. The most preferred Type II-S enzyme is FokI.

When the preferred Fok I containing URE is used, several conditions are preferably used to effect cleavage:

- 1) Excess of the URE over target DNA should be present to activate the enzyme. URE present only in equimolar amounts to the target DNA would yield poor cleavage of ssDNA because the amount of active enzyme available would be limiting.
- 2) An activator may be used to activate part of the FokI enzyme to dimerize without causing cleavage. Examples of appropriate activators are shown in Table 510.
- 3) The cleavage reaction is performed at a temperature between 45°-75°C, preferably above 50°C and most preferably above 55°C.

The UREs used in the prior art contained a 14-base single-stranded segment, a 10-base stem (containing a FokI site) , followed by the palindrome of the 10-base stem. While such UREs may be used in the methods of this invention, the preferred UREs of this invention also include a segment of three to eight bases (a loop) between the FokI restriction endonuclease recognition site containing segments. In the preferred embodiment, the stem (containing the FokI site) and its palindroms are also longer than 10 bases. Preferably, they are 10-14 bases in length. Examples of these "lollipop" URE adapters are shown in Table 5.

One example of using a URE to cleave a single-stranded DNA involves the FR3 region of human heavy chain. Table 508 shows an analysis of 840 full-length nature human heavy chains with the URE recognition sequences shown. The vast majority (718/840=0.85) will be recognized with 2 or fewer mismatches using five UREs (VHS881-1.1, VHS881-1.2, VHS881-2.1, VHS881-4.1, and VHS881-9.1). Each has a 20-base adaptor sequence to complement the germline gene, a ten-base stem segment containing a FokI site, a five base loop, and the reverse complement of the first stem segment. Annealing those adaptors, alone or in combination, to single-stranded antisense heavy chain DNA and treating with *FokI* in the presence of, *e.g.*, the activator FOKIact, will lead to cleavage of the antisense strand at the position indicated.

Another example of using a URE (s) to cleave a single-stranded DNA involves the FR1 region of the human Kappa light chains. Table 512 shows an analysis of 182 full-length human kappa chains for matching by the four 19-base probe sequences shown. Ninety-six percent of the sequences match one of the probes with 2 or fewer mismatches. The URE adapters shown in Table 512 are for cleavage of the sense strand of Kappa chains. Thus, the adaptor sequences are the reverse complement of the germline gene sequences. The URE consists of a ten-base stem, a five base loop, the reverse complement of the stem and the complementation sequence. The loop shown here is TTGTT, but other sequences could be used. Its function is to interrupt the palindrome of the stems so that formation of a lollipop monomer is favored over dimerization. Table 512 also shows where the sense strand is cleaved.

Another example of using a URE to cleave a single-stranded DNA involves the human lambda light chain. Table 515 shows analysis of 128 human lambda light chains for matching the four 19-base probes shown. With three or fewer mismatches, 88 of 128 (69%) of the chains match one of the probes. Table 515 also shows URE adapters corresponding to these probes. Annealing these adapters to upper-strand ssDNA of lambda chains and treatment with *FokI* in the presence of FOKIact at a temperature at or above 45°C will lead to specific and precise cleavage of the chains.

The conditions under which the short oligonucleotide sequences of the first method and the UREs of the second method are contacted with the single-stranded DNAs may be empirically determined. The conditions must be such that the single-stranded DNA remains in substantially single-stranded form. More particularly, the conditions must be such that the single-stranded DNA does not form loops that may interfere with its association with the oligonucleotide sequence or the URE or that may themselves provide sites for cleavage by the chosen restriction endonuclease.

The effectiveness and specificity of short oligonucleotides (first method) and UREs (second method) can be adjusted by controlling the concentrations of the URE adapters/oligonucleotides and substrate DNA, the temperature, the pH, the concentration of metal ions, the ionic strength, the concentration of chaotropes (such as urea and formamide), the concentration of the restriction endonuclease (*e.g.*, *FokI*), and the time of the digestion. These conditions can be optimized with synthetic oligonucleotides having: 1) target germline gene sequences, 2) mutated target gene sequences, or 3) somewhat related non-target sequences. The goal is to cleave most of the target sequences and minimal amounts of non-targets.

In this invention, the single-stranded DNA is maintained in substantially that form using a temperature between 45°C to 75°C. More preferably, a temperature between 50°C and 60°C, most preferably between 55°C and 60°C, is used. These temperatures are employed both when contacting the DNA with the oligonucleotide or URE and when cleaving the DNA using the methods of this invention.

The two cleavage methods of this invention have several advantages. The first method allows the individual members of the family of single-stranded DNAs to be cleaved solely at one substantially conserved endonuclease recognition site. The method also does not require an endonuclease recognition site to be built in to the reverse transcription or amplification primers. Any native or synthetic site in the family can be used.

The second method has both of these advantages. In addition, the URE method allows the single-stranded DNAs to be cleaved at positions where no endonuclease recognition site naturally occurs or has been synthetically constructed.

Most importantly, both cleavage methods permit the use of 5' and 3' primers so as to maximize diversity and then cleavage to remove unwanted or deleterious sequences before cloning and display.

After cleavage of the amplified DNAs using one of the methods of this invention, the DNA is prepared for cloning. This is done by using a partially duplexed synthetic DNA adapter, whose terminal sequence is based on the specific cleavage site at which the amplified DNA has been cleaved.

The synthetic DNA is designed such that when it is ligated to the cleaved single-stranded DNA, it allows that DNA to be expressed in the correct reading frame so as to display the desired peptide, polypeptide or protein on the surface of the genetic package. Preferably, the double-stranded portion of the adapter comprises the sequence of several codons that encode the amino acid sequence characteristic of the family of peptides, polypeptides or proteins up to the cleavage site. For human heavy chains, the amino acids of the 3-23 framework are preferably used to provide the sequences required for expression of the cleaved DNA.

Preferably, the double-stranded portion of the adapter is about 12 to 100 bases in length. More preferably, about 20 to 100 bases are used. The double-stranded region of the adapter also preferably contains at least one endonuclease recognition site useful for cloning the DNA into a suitable display vector (or a recipient vector used to archive the diversity). This endonuclease restriction site may be native to the germline gene sequences used to extend the DNA sequence. It may be also constructed using degenerate sequences to the native germline gene sequences. Or, it may be wholly synthetic.

The single-stranded portion of the adapter is complementary to the region of the cleavage in the single-stranded DNA. The overlap can be from about 2 bases up to about 15 bases. The longer the overlap, the more efficient the ligation is likely to be. A preferred length for the overlap is 7 to 10. This allows some mismatches in the region so that diversity in this region may be captured.

The single-stranded region or overlap of the partially duplexed adapter is advantageous because it allows DNA cleaned at the chosen site, but not other fragments to be captured. Such fragments would contaminate the library with genes encoding sequences that will not fold into proper antibodies and are likely to be non-specifically sticky.

One illustration of the use of a partially duplexed adaptor in the methods of this invention involves ligating such adaptor to a human FR3

region that has been cleaved, as described above, at 5'-ACnGT-3' using HpyCH4III, Bst4CI or TaaI.

Table 250 F.2 shows the bottom strand of the double-stranded portion of the adaptor for ligation to the cleaved bottom-strand DNA. Since the HpyCB4III-Site is so far to the right (as shown in Table 206), a sequence that includes the *Afl*III-site as well as the *Xba*I site can be added. This bottom strand portion of the partially-duplexed adaptor, H43.XAExt, incorporates both *Xba*I and *Afl*III-sites. The top strand of the double-stranded portion of the adaptor has neither site (due to planned mismatches in the segments opposite the *Xba*I and *Afl*III-Sites of H43.XAExt), but will anneal very tightly to H43.XAExt. H43.XAExt contains only the *Afl*III-site and is to be used with the top strands H43.ABr1 and H43.ABr2 (which have intentional alterations to destroy the *Afl*III-site).

After ligation, the desired, captured DNA can be PCR amplified again, if desired, using in the preferred embodiment a primer to the downstream constant region of the antibody gene and a primer to part of the double-standard region of the adapter. The primers may also carry restriction endonuclease sites for use in cloning the amplified DNA.

After ligation, and perhaps amplification, of the partially double-stranded adapter to the single-stranded amplified DNA, the composite DNA is cleaved at chosen 5' and 3' endonuclease recognition sites.

The cleavage sites useful for cloning depend on the phage or phagemid into which the cassette will be inserted and the available sites in the antibody genes. Table 1 provides restriction endonuclease data for 75 human light chains. Table 2 shows corresponding data for 79 human heavy chains. In each Table the endonucleases are ordered by increasing frequency of cutting. In these Tables, Nch is the number of chains cut by the enzyme and Ns is the number of sites (some chains have more than one site).

From this analysis, *Sfi*I, *Not*I, *Afl*III, *Apa*LI, and *Asc*I are very suitable. *Sfi*I and *Not*I are preferably used in pCES1 to insert the heavy-chain display segment. *Apa*LI and *Asc*I are preferably used in pCES1 to insert the light-chain display segment.

*Bst*EII-sites occur in 97% of germ-line JH genes. In rearranged V genes, only 54/79 (68%) of heavy-chain genes contain a *Bst*EII-site and 7/61 of these contain two sites. Thus, 47/79 (59%) contain a single *Bst*EII-Site. An alternative to using *Bst*EII is to cleave *via* UREs at the end of JH and ligate to a synthetic oligonucleotide that encodes part of CHI.

One example of preparing a family of DNA sequences using the methods of this invention involves capturing human CDR 3 diversity. As described above, mRNAs from various autoimmune patients is reverse transcribed into lower strand cDNA. After the top strand RNA is degraded, the lower strand is immobilized and a short oligonucleotide used to cleave the cDNA upstream of CDR3. A partially duplexed synthetic DNA adapter is then annealed to the DNA and the DNA is amplified using a primer to the adapter and a primer to the constant region (after FR4). The DNA is then cleaved using *Bst*EII (in FR4) and a restriction endonuclease appropriate to the partially double-stranded adapter (e.g., *Xba*I and *Afl*III (in FR3)). The DNA is then ligated into a synthetic VH skeleton such as 3-23,

One example of preparing a single-stranded DNA that was cleaved using the URE method involves the human Kappa chain. The cleavage site in the sense strand of this chain is depicted in Table 512. The oligonucleotide kapextURE is annealed to the oligonucleotides (kaBR01UR, kaBR02UR, kaBR03UR, and ka3R04UR) to form a partially duplex DNA. This DNA, is then ligated to the cleaved soluble Kappa chains. The ligation product is then amplified using primers kapextUREPCR and CKForeAsc (which inserts a *Asc*I site after the end of C Kappa). This product is then cleaved with *Apa*LI and *Asc*I and ligated to similarly cut recipient vector.

Another example involves the cleavage illustrated in Table 515. After cleavage, an extender (ON_LamEx133) and four bridge oligonucleotides (ON_LAMB1-133, ON_LamB2-133, ON_LamBS-133, and ON_LamB4-133) are annealed to form a partially duplex DNA. That DNA is ligated to the cleaved lambda-chain sense strands. After ligation, the DNA is amplified with ON_Lam133PCR and a forward primer specific to the lambda constant domain, such as CL2ForeAsc or CL7ForeAsc (Table 130).

In human heavy chains, one can cleave almost all genes in FR4 (downstream, *i.e.* toward the 3' end of the sense strand, of CDR3) at a *Bst*EII-Site that occurs at a constant, position in a very large fraction of human heavy-chain V genes. One then needs a site in FR3, if only CDR3 diversity is to be captured, in FR2, if CDR2 and CDR3 diversity is wanted, or in FR1, if all the CDR diversity is wanted. These sites are preferably inserted as part of the partially double-stranded adaptor.

The preferred process of this invention is to provide recipient vectors having sites that allow cloning of either light or heavy chains. Such vectors are well known and widely used in the art. A preferred phage display vector in accordance with this invention is phage MALIA3. This displays in gene III. The sequence of the phage MALIA3 is shown in Table 120A (annotated) and Table 120B (condensed).

The DNA encoding the selected regions of the light or heavy chains can be transferred to the vectors using endonucleases that cut either light or heavy chains only very rarely. For example, light chains may be captured with *Apa*LI and *Asc*I. Heavy-chain genes are preferably cloned into a recipient vector having *Sfi*I, *Nco*I, *Xba*I, *Afl*III, *Bst*EII, *Apa*I, and *Not*I sites. The light chains are preferably moved into the library as *Apa*LI-*Asc*I fragments. The heavy chains are preferably moved into the library as *Sfi*I-*Not*I fragments.

Most preferably, the display is had on the surface of a derivative of M13 phage. The most preferred vector contains all the genes of M13, an antibiotic resistance gene, and the display cassette. The preferred, vector is provided with restriction sites that allow introduction and excision of members of the diverse family of genes, as cassettes. The preferred vector is stable against rearrangement under the growth conditions used to amplify phage.

In another embodiment of this invention the diversity captured by the methods of the present invention may be displayed in a phagemid vector (*e.g.* pCES1) that displays the peptide, polypeptide or protein on the III protein. Such vectors may also be used to store the diversity for subsequent display using other vectors or phage.

In another embodiment, the mode of display may be through a short linker to three possible anchor domains. One anchor domain being the final portion of M13 III ("IIIstump"), a second anchor being the full length III mature protein, and the third being the M13 VIII mature protein.

The IIIstump fragment contains enough of M13 III to assemble into phage but not the domains involved in mediating infectivity. Because the w.t. III and VIII proteins are present, the phage is unlikely to delete the antibody genes and phage that do delete these segments receive

only a very small growth advantage. For each of the anchor domains, the DNA encodes the w.t. AA sequence, but differs from the w.t. DNA sequence to a very high extent. This will greatly reduce the potential for homologous recombination between the display anchor and the w.t. gene that is also present.

Most preferably, the present invention uses a complete phage carrying an antibiotic-resistance gene (such as an ampicillin-resistance gene) and the display cassette. Because the w.t. *iii* and *viii* genes are present, the w.t. proteins are also present. The display cassette is transcribed from a regulatable promoter (e.g., P_{lacZ}). Use of a regulatable promoter allows control of the ratio of the fusion display gene to the corresponding w.t. coat protein. This ratio determines the average number of copies of the display fusion per phage (or phagemid) particle.

The specification describes a method of displaying peptides, polypeptides or proteins (and particularly Fabs) on filamentous phage. In the most preferred embodiment this method displays FABS and comprises:

a) obtaining a cassette capturing a diversity of segments of DNA encoding the elements:

P_{reg} : :RBS1: :SS1: :VL: :CL: :stop: :RBS2: :SS2: :VH: :CH1: :linker: :anchor: :stop: :

where P_{reg} is a regulatable promoter, RBS1 is a first ribosome binding site, SS1 is a signal sequence operable in the host strain, VL is a member of a diverse set of light-chain variable regions, CL is a light-chain constant region, stop is one or more stop codons, RBS2 is a second ribosome binding site, SS2 is a second signal sequence operable in the host strain, VH is a member of a diverse or heavy-chain variable regions, CH1 is an antibody heavy-chain first constant domain, linker is a sequence of amino acids of one to about 50 residues, anchor is a protein that will assemble into the filamentous phage particle and stop is a second example of one or more stop codons; and

b) positioning that cassette within the phage genome to maximize the viability of the phage and to minimize the potential for deletion of the cassette or parts thereof.

The DNA encoding the anchor protein in the above preferred cassette should be designed to encode the same (or a closely related) amino acid sequence as is found in one of the coat proteins of the phage, but with a distinct DNA sequence. This is to prevent unwanted homologous recombination with the w.t. gene. In addition, the cassette should be placed in the regions, The positioning and orientation of the display cassette can influence the behavior of the phage.

A transcription terminator may be placed after the second stop of the display cassette above (e.g., Trp). This will reduce interaction between the display cassette and other genes in the phage antibody display vector (PADV).

In another embodiment, the phage or phagemid can display proteins other than Fab, by replacing the Fab portions indicated above, with other protein genes.

Various hosts can be used for growth of the display phage or phagemids. Such hosts are well known in the art. In the preferred embodiment, where Fabs are being displayed, the preferred host should grow at 30°C and be RecA⁻ (to reduce unwanted genetic recombination) and EndA⁻ (to make recovery of RF DNA easier). It is also preferred that the host strain be easily transformable by electroperation.

XL1-Blue MRF⁺ satisfies most of these preferences, but does not grow well at 30°C. XL1-Blue MRF⁺ does grow slowly at 38°C and thus is an acceptable host. TG-1 is also an acceptable host although it is RecA⁺ and EndA⁺. XL1-Blue MRF⁺ is more preferred for the intermediate host used to accumulate diversity prior to final construction of the library.

After display, the libraries of this invention, may be screened using well known and conventionally used techniques. The selected peptides, polypeptides or proteins may then be used to treat disease. Generally, the peptides, polypeptides or proteins for use in therapy or in pharmaceutical compositions are produced by isolating the DNA encoding the desired peptide, polypeptide or protein from the member of the library selected. That DNA is used in conventional methods to produce the peptide, polypeptides or protein it encodes in appropriate host cells, preferably mammalian host cells, e.g., CHO cells. After isolation, the peptide, polypeptide or protein is used alone or with pharmaceutically acceptable compositions in therapy to treat disease.

EXAMPLES

Example 1: Capturing kappa chains with BsmAI:

A repertoire of human-kappa, chain mRNAs was prepared by treating total or poly(A⁺) RNA isolated from a collection of patients having various autoimmune diseases with calf intestinal phosphatase to remove the 5'-phosphate from all molecules that have them, such as ribosomal RNA, fragmented mRNA, tRNA and genomic DNA. Full length mRNA (containing a protective 7-methyl cap structure) is unaffected. The RNA is then treated with tobacco acid pyrophosphatase to remove the cap structure from full length mRNAs leaving a 5'-monophosphate group.

Full length mRNA¹ were modified with an adaptor at the 5' end and then reversed transcribed and amplified using the GeneRACE™ method and kit (Invitrogen). A 5' biotinylated primer complementary to the adaptor and a 3' primer complementary to a portion of the construct region were used.

Approximately 2 micrograms (ug) of human kappa-chain (Igkappa) gene RACE material with biotin attached to 5'-end of upper strand was immobilized on 200 microliters (uL) of Seradyn magnetic beads. The lower strand was removed by washing the DNA with 2 aliquots 200 uL of 0.1 M NaOH (pH 13) for 3 minutes for the first aliquot followed by 30 seconds for the second aliquot. The beads were neutralized with 200 uL of 10 mM Tris (pH 7.5) 100 mM NaCl. The short oligonucleotides shown in Table 525 were added in 40 fold molar excess in 100 uL of NEB buffer 2 (50 mM NaCl, 10 mM Tris-HCl, 10 mM MgCl₂, mM dithiothreitol pH 7.9) to the dry beads. The mixture was incubated at 95°C for 5 minutes then cooled down to 55°C over 30 minutes. Excess oligonucleotide was washed away with 2 washes of NEB buffer 3 (100 mM NaCl, 50 mM Tris-HCl, 10 mM MgCl₂, 1 mM dithiothreitol pH 7.9). Ten units of BsmAI (IEB) were added in NEB buffer 3 and incubated for 1 h at 55°C. The cleaved downstream DNA was collected and purified over a Qiagen PCR purification column (FIGs. 3 and 4).

A partially double-stranded adaptor was prepared using the oligonucleotide shown in Table 525. The adaptor was added to the single-stranded DNA in 100 fold molar excess along with 1000 units of T4 DNA ligase (NEB) and incubated overnight at 16°C. The excess oligonucleotide was removed with a Qiagen PCR purification column. The ligated material was amplified by PCR using the primers kapPCRt1 and kapfor shown in Table 525 for 10 cycles with the program shown in Table 530.

The soluble PCR product was run on a gel and showed a band of approximately 700 n, as expected (FIGs. 5 and 6). The DNA was cleaved with enzymes *Apa*I and *Ase*I, gel purified, and ligated to similarly cleaved vector pCES1. The presence of the correct size insert was checked by PCR in several clones as shown in FIG. 15.

Table 500 shows the DNA sequence of a kappa light chain, captured by this procedure. Table 501 shows a second sequence captured by this procedure. The closest bridge sequence was complementary to the sequence 5'-agccacc-3', but the sequence captured reads 5'-Tgccacc-3', showing that some mismatch in the overlapped region is tolerated.

Example 2: Construction of Synthetic CDR1 and CDR2 Diversity in V-3-23 VH Framework

A synthetic Complementary Determinant Region (CDR) 1 and 2 diversity was constructed in the 3-23 VH framework in a two step process: first, a vector containing the 3-23 VH framework was constructed, and then, a synthetic CDR 1 and 2 was assembled and cloned into this vector.

For construction of the V3-23 framework, 8 oligos and two PCR primers (long oligonucleotides: TOPFR1A, BOTFR1B, BOTFR2, BOTFR3, F06, BOTFR4, ON-vgC1, and ON-vgC2 and primers: SFPRMET and BOTPCRPRIM, shown in Table 600) that overlap were designed based on the Genbank sequence of V323 VH. The design incorporated at least one useful restriction site in each framework region, has shown in Table 600. In Table 600, the segments that were synthesized are shown as bold, the overlapping regions are underscored, and the PCR priming regions at each end are underscored. A mixture of these 8 oligos was combined at a final concentration of 2.5uM in a 20ul Polymerase Chain Reaction (PCR) reaction. The PCR mixture contained 200uM dNTPs, 2.5mM MgCl₂, 0.02U *Pfu Turbo*TM DNA Polymerase, 1U Qiagen HotStart Taq DNA Polymerase, and 1X Qiagen PCR buffer. The PCR program consisted of 10 cycles of 94°C for 30s, 55°C for 30s, and 72°C for 30s. The assembled V3-23 DNA sequence was then amplified, using 2.5ul of a 10-fold dilution from the initial PCR in 100ul PCR reaction. The PCR reaction contained 200uM dNTPs, 2-5mM MgCl₂, 0.02U *Pfu Turbo*TM DNA Polymerase, 1U Qiagen HotStart Taq DNA Polymerase, 1X Qiagen PCR Buffer and 2 outside primers (SFPRMET and BOTPCRPRIM) at a concentration of 1uM. The PCR program consisted of 23 cycles at 94°C for 30s, 55°C for 30s, and 72°C for 60s. The V3-23 VH DNA sequence was digested and cloned into pCES1 (phagemid vector) using the *Sfi*I and *Bst*EII restriction endonuclease sites (All restriction enzymes mentioned herein were supplied by New England BioLabs, Beverly, MA and used as per manufacturer's instructions).

Stuffer sequences (shown in Table 610 and Table 620) were introduced into pCES1 to replace CDR1/CDR2 sequences (900 bases between *Bsp*EI and *Xba*I RE sites) and CDR3 sequences (358 bases between *Afl*II and *Bst*EII), prior to cloning the CDR1/CDR2 diversity. The new vector is pCES5 and its sequence is given in Table 620. Having stuffers in place of the CDRs avoids the risk that a parental sequence would be over-represented in the library. The CDR1-2 stuffer contains restriction sites for *Bgl*II, *Bsu*36I, *Bcl*I, *Xcm*I, *Mlu*I, *Pvu*II, *Hpa*I, and *Hinc*II, the underscored sites being unique within the vector pCES5. The stuffer that replaces CDR3 contains the unique restriction endonuclease site *Rsr*II. The stuffer sequences are fragments from the penicillase gene of *E. coli*.

For the construction of the CDR1 and CDR2 diversity, 4 overlapping oligonucleotides (ON-vgC1, ON_Br12, ON_CD2Xba, and ON-vgC2, shown in Table 600 and Table 630) encoding CDR1/2, plus flanking regions, were designed. A mix of these 4 oligos was combined at a final concentration of 2.5uM in a 40ul PCR reaction. Two of the 4 oligos contained variegated sequences positioned at the CDR1 and the CDR2. The PCR mixture contained 200uM dNTPs, 2.5U Pwo DNA Polymerase (Roche), and 1X Pwo PCR buffer with 2mM MgSO₄. The PCR program consisted of 10 cycles at 94°C for 30s, 60°C for 30s, and 72°C for 60s. This assembled CDR1/2 DNA sequence was amplified, using 2.5ul of the mixture in 100ul PCR reaction. The PCR reaction contained 200uM dNTPs, 2.5U Pwo DNA Polymerase, 1X Pwo PCR Buffer with 2mM MgSO₄ and 2 outside primers at a concentration of 1uM. The PCR program consisted of 10 cycles at 94°C for 30s, 60°C for 30s, and 72°C for 60s. These variegated sequences were digested and cloned into the V3-23 framework in place of the CDR1/2 stuffer.

We obtained approximately 7×10^7 independent transformants. Into this diversity, we can clone CDR3 diversity either from donor populations or from synthetic DNA.

Furthermore, the present application describes the following items:

1. A method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:
 - (i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
 - (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.
2. A method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:
 - (i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and
 - (ii) cleaving the nucleic acid solely at the Type II-S cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.
3. In a method for displaying a member of a diverse family of peptides, polypeptides or proteins on the surface of a genetic package and collectively displaying at least a part of the diversity of the family, the improvement being characterized in that the displayed at least a part of peptide, polypeptide or protein is encoded at least in part by a nucleic acid that has been cleaved at a desired location by a method comprising the steps of:
 - (i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic

- acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.
4. In a method for displaying a member of a diverse family of peptides, polypeptides or proteins on the surface of a genetic package and collectively displaying at least a part of the diversity of the family, the improvement being characterized in that the displayed peptide, polypeptide or protein is encoded by a DNA sequence comprising a nucleic acid that has been cleaved at a desired location by
- (i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and
- (ii) cleaving the nucleic acid solely at the Type II-S cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide;
- the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.
5. A method for displaying a member of a diverse family of peptides, polypeptides or proteins on the surface of a genetic package and collectively displaying at least a part of the diversity of the family, the method comprising the steps of:
- (i) preparing a collection of nucleic acids that code at least in part for members of the diverse family;
- (ii) rendering the nucleic acids single-stranded;
- (iii) cleaving the single-stranded nucleic acids at a desired location by a method comprising the steps of;
- (a) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (b) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature; and
- (iv) displaying a member of the family of peptides, polypeptides or proteins coded, at least in part, by the cleaved nucleic acids on the surface of the genetic package and collectively displaying at least a portion of the diversity of the family.
6. A method for displaying a member of a diverse family of peptides, polypeptides or proteins on the surface of a genetic package and collectively displaying at least a portion of the diversity of the family, the method comprising the steps of:
- (i) preparing a collection of nucleic acids that code, at least in part, for members of the diverse family;
- (ii) rendering the nucleic acids single-stranded;
- (iii) cleaving the single-stranded nucleic acids at a desired location by a method comprising the steps of:
- (a) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at known distance from the recognition site; and
- (b) cleaving the nucleic acid solely at the Type II-S cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide;
- the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the restriction being carried out using a cleavage endonuclease that is active at the chosen temperature; and
- (iv) displaying a member of the family of peptides, polypeptides or proteins coded, at least in part, by the cleaved nucleic acids on the surface of the genetic package and collectively displaying at least a portion of the diversity of the family.
7. A library comprising a collection of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family, the library being produced using the methods of items 3, 4, 5 or 6.
8. A library comprising a collection of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and that collectively display at least a portion of the family, the displayed peptides, polypeptides or proteins being encoded by DNA sequences comprising at least in part sequences produced by cleaving single-stranded nucleic acid sequences at a desired location by a method comprising the steps of :
- (i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide; the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.
9. A library comprising a collection of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and that collectively display at least a portion of the diversity of the family of the displayed peptides, polypeptides or proteins being encoded by DNA sequences comprising at least in part sequences produced by cleaving single-stranded nucleic acid sequences at a desired location by a method comprising the steps of:
- (i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site where the cleavage of the nucleic acid is desired; and
- (ii) cleaving the nucleic acid solely at the Type II-S cleavage site formed by the complementation of the nucleic acid and the single-stranded

region of the oligonucleotide ;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

10. The methods according to any one of items 1 to 9, wherein the nucleic acids encode at least a portion of an immunoglobulin.

11. The methods according to item 10, wherein the immunoglobulin comprises a Fab or single chain Fv.

12. The methods according to item 10 or 11, wherein the immunoglobulin comprises at least portion of a heavy chain.

13. The methods according to item 12, wherein at least a portion of the heavy chain is human.

14. The methods according to item 10 or 11, wherein the immunoglobulin comprises at least a portion of FR1.

15. The methods according to item 14, wherein at least a portion of the FR1 is human.

16. The methods according to item 10 or 11, wherein the immunoglobulin at least a portion of a light chain.

17. The methods according to item 16, wherein at least a portion of the light chain is human.

18. The methods according to any one of items 1 to 9, wherein the nucleic acid sequences are at least in part derived from patients suffering from at least one autoimmune disease and/or cancer.

19. The methods according to item 18, wherein the autoimmune disease is selected from the group comprising lupus, erythematosus, systemic sclerosis, rheumatoid arthritis, antiphospholipid syndrome or vasculitis.

20. The methods according to item 18, wherein the nucleic acids are at least in part isolated from the group comprising peripheral blood cells, bone marrow cells spleen cells or lymph node cells.

21. The methods according to item 5 or 6 further comprising an nucleic acid amplification step between steps (i) and (ii), between steps (ii) and (iii) or between steps (iii) and (iv).

22. The methods according to item 21, wherein the amplification step uses geneRACE™.

23. The methods according to any one of items 1 to 9, wherein the temperature is between 45°C and 75°C.

24. The methods according to item 23, wherein the temperature is between 50°C and 60°C.

25. The methods according to item 24, wherein the temperature is between 55°C and 60° C.

26. The methods according to item 1, 3, 5 or 8, wherein the length of the single-stranded oligonucleotide is between 17 and 30 bases.

27. The methods according to item 26, wherein the length of the single-stranded oligonucleotide is between 18 and 24 based.

28. The methods according to item 1, 3, 5 or 8, wherein the restriction endonuclease is selected from the group comprising *MaeIII*, *Tsp4SI*, *HphI*, *BsaII*, *AluI*, *BlpI*, *DdeI*, *BglII*, *MslI*, *BsiEI*, *EaeI*, *EagI*, *HaeII*, *Bst4CI*, *HpyCH4III*, *Hinfl*, *MlyI*, *PleI*, *MnlI*, *HpyCH4V*, *BsmAI*, *BpmI*, *XmnI*, or *SacI*.

29. The methods according to item 28, wherein the restriction endonuclease is selected from the group comprising *Bst4CI*, *TaaI*, *HpyCH4III*, *BlpI*, *HpyCH4V* or *MslI*.

30. The methods according to item 2, 4, 6 or 9, wherein the length of the single-stranded region of the partially double-stranded oligonucleotide is between 14 and 22 bases.

31. The methods according to item 30, wherein the length of the single-stranded region of the partially double-stranded oligonucleotide is between 14 and 17 bases.

32. The methods according to item 31, wherein the length of the single-stranded region of the oligonucleotide is between 18 and 20 bases.

33. The methods according to item 2, 4, 6 or 9, wherein the length or the double-stranded region of the partially double-stranded oligonucleotide is between 10 and 14 base pairs formed by a stem and its palindrome.

34. The methods according to item 33 wherein, the partially double-stranded oligonucleotide comprises a loop of 3 to 8 bases between the stem and the palindrome.

35. The methods according to item 2, 4, 6 or 9, wherein the Type II-S restriction endonuclease is selected from the group comprising *AarICAC*, *AcerIII*, *Ebr7I*, *BbvI*, *BbvII*, *Bce83I*, *BceAI*, *BceFI*, *BciVI*, *BfiI*, *BinI*, *BscAI*, *BseRI*, *BsmFI*, *BspMI*, *EciI*, *Eco57I*, *FauI*, *FokI*, *GsuI*, *HgaI*, *HphI*, *MboII*, *MlyI*, *MmeI*, *MnlI*, *PleI*, *RleAI*, *SfaNI*, *SspD5I*, *Sth132I*, *StsI*, *TaqII*, *Tth111II*, or *UbaPI*.

36. The methods according to item 35, wherein the Type II-3 restriction endonuclease is *FokI*.

37. A method for preparing single-stranded nucleic acids for cloning into a vector, the method comprising the steps of:

(i) contacting a single-stranded nucleic acid sequence that has been cleaved with a restriction endonuclease with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region that remains after cleavage, the double-stranded region of the oligonucleotide including any sequences necessary to return the sequences that remain after cleavage into proper and original reading frame for expression and containing a restriction endonuclease recognition site 5' of those sequences; and

(ii) cleaving the partially double-stranded oligonucleotide sequence solely at the restriction endonuclease recognition site contained within the double-stranded region of the partially double-stranded oligonucleotide.

38. The method according to item 37, wherein the length of the single-stranded portion of the partially double-stranded oligonucleotide is between 2 and 15 bases.

39. The method according to item 38, wherein the length of the single-stranded portion of the partially double-stranded oligonucleotide is between 7 and 10 bases.

40. The method according to item 37, wherein the length of the double-stranded portion of the partially double-stranded oligonucleotide is between 12 and 100 base pairs.

41. The method according to item 40, wherein the length of the double-stranded portion of the partially double-stranded oligonucleotide is between 20 and 100 base pairs.

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Phe	Thr	Phe	Ser	Ser	Tyr	Ala	Met	Ser	Trp	Val	Arg	Gln	Ala	Pro	Gly
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Lys	Gly	Leu	Glu	Trp	Val	Ser	Ala	Ile	Ser	Gly	Ser	Gly	Gly	Ser	Thr
	50					55				60					
Tyr	Tyr	Ala	Asp	Ser	Val	Lys	Gly	Arg	Phe	Thr	Ile	Ser	Arg	Asp	Asn
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Ser	Lys	Asn	Thr	Leu	Tyr	Leu	Gln	Met	Asn	Ser	Leu	Arg	Ala	Glu	Asp
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Phe	Asp	Ile	Trp	Gly	Gln	Gly	Thr	Met	Val	Thr	Val	Ser	Ser	Ala	Ser
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20

25

30

Leu Val Gln Pro Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly
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 Phe Thr Phe Ser Ser Tyr Ala Met Ser Trp Val Arg Gln Ala Pro Gly
 50 55 60
 Lys Gly Leu Glu Trp Val Ser Ala Ile Ser Gly Ser Gly Gly Ser Thr
 65 70 75 80
 Tyr Tyr Ala Asp Ser Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn
 85 90 95
 Ser Lys Asn Thr Leu Tyr Leu Gln Met Asn Ser Leu Arg Ala Glu Asp
 100 105 110
 Thr Ala Val Tyr Tyr Cys Ala Lys Asp Tyr Glu Gly Thr Gly Tyr Ala
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 Phe Asp Ile Trp Gly Gln Gly Thr Met Val Thr Val Ser Ser Ala Ser
 130 135 140
 Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr
 145 150 155 160
 Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro
 165 170 175
 Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val
 180 185 190
 His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser
 195 200 205
 Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile
 210 215 220
 Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val
 225 230 235 240
 Glu Pro Lys Ser Cys Ala Ala Ala His His His His His His Ser Ala
 245 250 255
 Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Gly Ala Ala Asp Ile
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 Asn Asp Asp Arg Met Ala Ser Gly Ala Ala Glu Thr Val Glu Ser Cys
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 Leu Ala Lys Pro His Thr Glu Ile Ser Phe Thr Asn Val Trp Lys Asp
 290 295 300
 Asp Lys Thr Leu Asp Arg Tyr Ala Asn Tyr Glu Gly Cys Leu Trp Asn
 305 310 315 320
 Ala Thr Gly Val Val Val Cys Thr Gly Asp Glu Thr Gln Cys Tyr Gly
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 35 40 45
 Ile Gly Asp Val Ser Gly Leu Ala Asn Gly Asn Gly Ala Thr Gly Asp
 50 55 60
 Phe Ala Gly Ser Asn Ser Gln Met Ala Gln Val Gly Asp Gly Asp Asn
 65 70 75 80
 Ser Pro Leu Met Asn Asn Phe Arg Gln Tyr Leu Pro Ser Leu Pro Gln
 85 90 95
 Ser Val Glu Cys Arg Pro Phe Val Phe Ser Ala Gly Lys Pro Tyr Glu
 100 105 110
 Phe Ser Ile Asp Cys Asp Lys Ile Asn Leu Phe Arg Gly Val Phe Ala
 115 120 125
 Phe Leu Leu Tyr Val Ala Thr Phe Met Tyr Val Phe Ser Thr Phe Ala
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 Asn Ile Leu Arg Asn Lys Glu Ser
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 Val Gly Arg Phe Ala Lys Thr Pro Arg Val Leu Arg Ile Pro Asp Lys
 35 40 45
 Pro Ser Ile Ser Asp Leu Leu Ala Ile Gly Arg Gly Asn Asp Ser Tyr
 50 55 60
 Asp Glu Asn Lys Asn Gly Leu Leu Val Leu Asp Glu Cys Gly Thr Trp
 65 70 75 80
 Phe Asn Thr Arg Ser Trp Asn Asp Lys Glu Arg Gln Pro Ile Ile Asp
 85 90 95
 Trp Phe Leu His Ala Arg Lys Leu Gly Trp Asp Ile Ile Phe Leu Val
 100 105 110
 Gln Asp Leu Ser Ile Val Asp Lys Gln Ala Arg Ser Ala Leu Ala Glu
 115 120 125
 His Val Val Tyr Cys Arg Arg Leu Asp Arg Ile Thr Leu Pro Phe Val
 130 135 140
 Gly Thr Leu Tyr Ser Leu Ile Thr Gly Ser Lys Met Pro Leu Pro Lys
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 Leu His Val Gly Val Val Lys Tyr Gly Asp Ser Gln Leu Ser Pro Thr
 165 170 175
 Val Glu Arg Trp Leu Tyr Thr Gly Lys Asn Leu Tyr Asn Ala Tyr Asp
 180 185 190
 Thr Lys Gln Ala Phe Ser Ser Asn Tyr Asp Ser Gly Val Tyr Ser Tyr
 195 200 205
 Leu Thr Pro Tyr Leu Ser His Gly Arg Tyr Phe Lys Pro Leu Asn Leu
 210 215 220
 Gly Gln Lys Met Lys Leu Thr Lys Ile Tyr Leu Lys Lys Phe Ser Arg
 225 230 235 240
 Val Leu Cys Leu Ala Ile Gly Phe Ala Ser Ala Phe Thr Tyr Ser Tyr
 245 250 255
 Ile Thr Gln Pro Lys Pro Glu Val Lys Lys Val Val Ser Gln Thr Tyr
 260 265 270
 Asp Phe Asp Lys Phe Thr Ile Asp Ser Ser Gln Arg Leu Asn Leu Ser
 275 280 285
 Tyr Arg Tyr Val Phe Lys Asp Ser Lys Gly Lys Leu Ile Asn Ser Asp
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 Asp Leu Gln Lys Gln Gly Tyr Ser Leu Thr Tyr Ile Asp Leu Cys Thr
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  1           5           10           15
Ser Leu Arg Ala Glu Asp Thr Ala Val Tyr Tyr Cys Ala Lys
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 <211> 90
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 <213> Homo sapiens
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 acctgcactg tctctggtgg ctccatcagc 90
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 acctgcactg tctctggtgg ctccatcagc 90
 <210> 246
 <211> 90
 <212> DNA
 <213> Homo sapiens
 <400> 246

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acctgcgctg tctatggtgg gtccctcagt                                     90
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<212> DNA
<213> Homo sapiens
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acctgcactg tctctggtgg ctccatcagc                                     90
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acctgcactg tctctggtgg ctccatcagt                                     90
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<212> DNA
<213> Homo sapiens
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acctgcactg tctctggtgg ctccgtcagc                                     90
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<213> Homo sapiens
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acctgcgctg tctctggtta ctccatcagc                                     90
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<212> DNA
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gaggtgcagc tgggtgcagtc tggagcagag gtgaaaaagc ccggggagtc tctgaagatc 60
tctctgaagg gttctggata cagctttacc                                     90
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gaagtgcagc tgggtgcagtc tggagcagag gtgaaaaagc ccggggagtc tctgaggatc 60
tctctgaagg gttctggata cagctttacc                                     90
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caggtacagc tgcagcagtc aggtccagga ctggtgaagc cctcgcagac cctctcactc 60
acctgtgcca tctccgggga cagtgtctct                                     90
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caggtgcagc tgggtgcaatc tgggtctgag ttgaagaagc ctggggcctc agtgaagggt 60
tctctgaagg cttctggata caccttcactL                                     90
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<400> 255
agttctccct gcagctgaac tc                                     22
<210> 256
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<400> 256
cactgtatct gcaaatgaac ag                                     22

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<210> 257
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 <223> Description of Artificial Sequence: Probe
 <400> 257
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 <210> 258
 <211> 22
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 <400> 258
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 <400> 259
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 <210> 260
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 <400> 260
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 <210> 262
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 <223> Description of Artificial Sequence: Probe
 <400> 262
 ctgcctacct gcagtggagc ag 22
 <210> 263
 <211> 22
 <212> DNA
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 <223> Description of Artificial Sequence: Probe
 <400> 263
 tgcctatct gcaaatgaac ag 22
 <210> 264
 <211> 22
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 <400> 264
 acatggagct gaggcagcctg ag 22
 <210> 265
 <211> 22
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 <400> 265
 acatggagct gaggcagcctg ag 22
 <210> 266
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 <212> DNA

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 <210> 267
 < 211> 22
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 <400> 267
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 <210> 268
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 <400> 268
 atctgcaaat gaacagcctg aa 22
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 <400> 270
 atctgcaaat gaacagtctg ag 22
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 < 211> 22
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 <400> 271
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 <210> 272
 < 211> 22
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 <400> 272
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 <400> 273
 atcttcaaat gggcagcctg ag 22
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<400> 275
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 <210> 277
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 <400> 277
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 <210> 278
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 <400> 278
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 <400> 279
 ctgtgtatta ctgtgcgaga ga 22
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 ccatgtatta ctgtgcaaga ta 22
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 ccgtgtatta ctgtcggca ga 22
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< 211> 22
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<400> 287
ccttgatta ctgtgcaaaa ga 22
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< 211> 22
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<400> 288
ctgtgatta ctgtgcaaga ga 22
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ccgtgatta ctgtaccaca ga 22
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ccgtatatta ctgtgcgaaa ga 22
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ctgtgatta ctgtgcgaaa ga 22
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<400> 293
ccgtgatta ctgtactaga ga 22
<210> 294
< 211> 22
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< 213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 294
 ccgtgtatta ctgtgctaga ga 22
 <210> 295
 <211> 22
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 295
 ccgtgtatta ctgtactaga ca 22
 <210> 296
 <211> 22
 <212> DNA
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 <400> 296
 ctgtgtatta ctgtaagaaa ga 22
 <210> 297
 <211> 22
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 297
 ccgtgtatta ctgtgcgaga aa 22
 <210> 298
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 <400> 298
 ccgtgtatta ctgtgccaga ga 22
 <210> 299
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 <400> 299
 ctgtgtatta ctgtgcgaga ca 22
 <210> 300
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 <400> 300
 ccatgtatta ctgtgcgaga ca 22
 <210> 301
 <211> 22
 <212> DNA
 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 301
 ccatgtatta ctgtgcgaga aa 22
 <210> 302
 <211> 69
 <212> DNA
 <213> Unknown Organism
 <220>
 <223> Description of Unknown Organism: Kappa FR1 GLGs
 <400> 302
 gacatccaga tgacceagtc tccatccctcc ctgtctgcat ctgtaggaga cagagtcacc 60
 atcacttgc 69
 <210> 303
 <211> 66
 <212> DNA
 <213> Unknown Organism
 <220>
 <223> Description of Unknown Organism: Lambda FR1 GLG sequence

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<400> 303
cagtctgtgc tgactcagcc accctcggtg tctgaagccc ccaggcagag ggtcaccatc 60
tctctgt                                         66
<210> 304
<211> 668
<212> DNA
<213> Unknown Organism
<220>
<223> Description of Unknown Organism: h3401-h2
<400> 304
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agggccaccc tctcctgcag ggcagtcag agtgtagta acaacttagc ctggtaccag 120
cagaaacctg gccagggttc caggctcctc atctatggtg catccaccag ggccactgat 180
atcccagcca ggttcagtg ggtgggtct gggacagact tcaactctac catcagcaga 240
ctggagcctg aagattttgc agtgtattac tgtcagcggg atggtagctc accgggggtg 300
acgttcggcc aagggaccaa ggtggaaatc aaacgaactg tggctgcacc atctgtcttc 360
atcttcccgc catctgatga gcagttgaaa tctggaactg cctctgttgt gtgctgctg 420
aataacttct atcccagaga ggccaaagta cagtggaagg tggataacgc cctccaatcg 480
ggtaactccc aggagagtgt cacagagcag gacagcaagg acagcaccta cagcctcagc 540
agcacccctg acgtgagcaa agcagactac gagaaacaca aagtctacgc ctgccaagtc 600
acccatcagg gcctgagctc gcctgtcaca aagagcttca acaaaggaga gtgtaagggc 660
gaattcgc                                         668
<210> 305
<211> 223
<212> PRT
<213> Unknown Organism
<220>
<223> Description of Unknown Organism: h3401-h2
<400> 305
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Ser Pro Gly Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Ser Val
          20           25           30

Ser Asn Asn Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Val Pro Arg
          35           40           45

Leu Leu Ile Tyr Gly Ala Ser Thr Arg Ala Thr Asp Ile Pro Ala Arg
          50           55           60

Phe Ser Gly Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Arg
          65           70           75           80

Leu Glu Pro Glu Asp Phe Ala Val Tyr Tyr Cys Gln Arg Tyr Gly Ser
          85           90           95

Ser Pro Gly Trp Thr Phe Gly Gln Gly Thr Lys Val Glu Ile Lys Arg
          100          105          110

Thr Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln
          115          120          125

Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe Tyr
          130          135          140

Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser
          145          150          155          160

Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser Thr
          165          170          175

Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys
          180          185          190

His Lys Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser Pro
          195          200          205

Val Thr Lys Ser Phe Asn Lys Gly Glu Cys Lys Gly Glu Phe Ala
          210          215          220
<210> 306
<211> 700

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<212> DNA

<213> Unknown Organism

<220>

<223> Description of Unknown Organism: h3401-d8 KAPPA

<400> 306

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agagccaccc tctcctgcag ggcagtcag gtgtctccag gggaaagagc caccctctcc 120
tgcaatcttc tcagcaactt agcctggtag cagcagaaac ctggccaggc tcccaggctc 180
ctcatctatg gtgcttccac cggggccatt ggtatcccag ccaggttcag tggcagtggg 240
tctgggacag agttcactct caccatcagc agcctgcagt ctgaagattt tgcagtgtat 300
ttctgtcagc agtatggtac ctcaccgccc actttcggcg gagggaccaa ggtggagatc 360
aaacgaactg tggctgcacc atctgtcttc atcttcccgc catctgatga gcagttgaaa 420
tctggaactg cctctgttgt gtgccgctg aataacttct atcccagaga ggccaaagta 480
cagtggaagg tggataacgc cctccaatcg ggtaactccc aggagagtgt cacagagcag 540
gacaacaagg acagcaccta cagcctcagc agcacctga cgctgagcaa agtagactac 600
gagaaacacg aagtctacgc ctgcgaagtc acccatcagg gccttagctc gcccgtcacg 660
aagagcttca acaggggaga gtgtaagaaa gaattcgttt 700

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<210> 307

<211> 222

<212> PRT

<213> Unknown Organism

<220>

<223> Description of Unknown Organism: h3401-d8 KAPPA

<400> 307

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Ser Ala Gln Asp Ile Gln Met Thr Gln Ser Pro Ala Thr Leu Ser Val
  1                    5                    10                    15

Ser Pro Gly Glu Arg Ala Thr Leu Ser Cys Arg Ala Ser Gln Asn Leu
          20                    25                    30

Leu Ser Asn Leu Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ala Pro Arg
          35                    40                    45

Leu Leu Ile Tyr Gly Ala Ser Thr Gly Ala Ile Gly Ile Pro Ala Arg
  50                    55                    60

Phe Ser Gly Ser Gly Ser Gly Thr Glu Phe Thr Leu Thr Ile Ser Ser
  65                    70                    75                    80

Leu Gln Ser Glu Asp Phe Ala Val Tyr Phe Cys Gln Gln Tyr Gly Thr
          85                    90                    95

Ser Pro Pro Thr Phe Gly Gly Gly Thr Lys Val Glu Ile Lys Arg Thr
          100                    105                    110

Val Ala Ala Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln Leu
          115                    120                    125

Lys Ser Gly Thr Ala Ser Val Val Cys Pro Leu Asn Asn Phe Tyr Pro
          130                    135                    140

Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser Gly
          145                    150                    155                    160

Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Asn Lys Asp Ser Thr Tyr
          165                    170                    175

Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Val Asp Tyr Glu Lys His
          180                    185                    190

Glu Val Tyr Ala Cys Glu Val Thr His Gln Gly Leu Ser Ser Pro Val
          195                    200                    205

Thr Lys Ser Phe Asn Arg Gly Glu Cys Lys Lys Glu Phe Val
          210                    215                    220

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<210> 308

<211> 45

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic oligonucleotide

<400> 308

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 <211> 45
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 309
 gccgtgtatt actgtgag cacatccgtg ttgtcacgg atgtg 45
 <210> 310
 <211> 45
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 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 310
 gccgtatatt actgtgag cacatccgtg ttgtcacgg atgtg 45
 <210> 311
 <211> 45
 <212> DNA
 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 311
 gccgtgtatt actgtgag cacatccgtg ttgtcacgg atgtg 45
 <210> 312
 <211> 45
 <212> DNA
 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 312
 gccatgtatt actgtgag cacatccgtg ttgtcacgg atgtg 45
 <210> 313
 <211> 88
 <212> DNA
 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 313
 aatagtagac tgcagtgtcc tcagccctta agctgttcat ctgcaagtag agagtattct 60
 tagagttgtc tctagactta gtgaagcg 88
 <210> 314
 <211> 95
 <212> DNA
 <213> Artificial Sequence
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 314
 cgcttcacta agtctagaga caactcctaag aatactctct acttgcagat gaacagctta 60
 agggctgagg acactgcagt ctactattgt gcgag 95
 <210> 315
 <211> 95
 <212> DNA
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 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 315
 cgcttcacta agtctagaga caactcctaag aatactctct acttgcagat gaacagctta 60
 agggctgagg acactgcagt ctactattgt acgag 95
 <210> 316
 <211> 24
 <212> DNA
 <213> Artificial Sequence
 <220>
 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 316
 cgcttcacta agtctagaga caac 24
 <210> 317
 <211> 44
 <212> DNA
 <213> Artificial Sequence
 <220>
 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 317

cacatccgtg ttgtcacgg atgtgggagg atggagactg ggtc 44
 <210> 318
 <211> 44
 <212> DNA
 <213> Artificial Sequence
 <220>
 <223> Description of Artificial Sequence: Synthetic oligonucleotide
 <400> 318
 cacatccgtg ttgtcacgg atgtgggaga gtggagactg agtc 44
 <210> 319
 <211> 44
 <212> DNA
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<212> DNA

<213> Unknown Organism

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<223> Description of Unknown Organism: pCES5

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<212> PRT

<213> Unknown Organism

<220>

<223> Description of Unknown Organism: pCES5

<400> 354

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Asp Ala Glu Asp Gln Leu Gly Ala Arg Val Gly Tyr Ile Glu Leu Asp
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Leu Asn Ser Gly Lys Ile Leu Glu Ser Phe Arg Pro Glu Glu Arg Phe
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Pro Met Met Ser Thr Phe Lys Val Leu Leu Cys Gly Ala Val Leu Ser
 65          70          75          80

Arg Ile Asp Ala Gly Gln Glu Gln Leu Gly Arg Arg Ile His Tyr Ser
          85          90          95

Gln Asn Asp Leu Val Glu Tyr Ser Pro Val Thr Glu Lys His Leu Thr
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Asp Gly Met Thr Val Arg Glu Leu Cys Ser Ala Ala Ile Thr Met Ser
          115          120          125

Asp Asn Thr Ala Ala Asn Leu Leu Leu Thr Thr Ile Gly Gly Pro Lys
          130          135          140

Glu Leu Thr Ala Phe Leu His Asn Met Gly Asp His Val Thr Arg Leu
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Asp Arg Trp Glu Pro Glu Leu Asn Glu Ala Ile Pro Asn Asp Glu Arg
          165          170          175

Asp Thr Thr Met Pro Val Ala Met Ala Thr Thr Leu Arg Lys Leu Leu
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Thr Gly Glu Leu Leu Thr Leu Ala Ser Arg Gln Gln Leu Ile Asp Trp
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Ala Gly Trp Phe Ile Ala Asp Lys Ser Gly Ala Gly Glu Arg Gly Ser
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Arg Gly Ile Ile Ala Ala Leu Gly Pro Asp Gly Lys Pro Ser Arg Ile
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Val Val Ile Tyr Thr Thr Gly Ser Gln Ala Thr Met Asp Glu Arg Asn
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<400> 355

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 Leu Lys Ser Gly Thr Ala Ser Val Val Cys Leu Leu Asn Asn Phe Tyr
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 Pro Arg Glu Ala Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser
 65 70 75 80
 Gly Asn Ser Gln Glu Ser Val Thr Glu Gln Asp Ser Lys Asp Ser Thr
 85 90 95
 Tyr Ser Leu Ser Ser Thr Leu Thr Leu Ser Lys Ala Asp Tyr Glu Lys
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Val Thr Lys Ser Phe Asn Arg Gly Glu Cys
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<210> 356

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<212> PRT

<213> Unknown Organism

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<223> Description of Unknown Organism: pCES5

<400> 356

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Leu Val Gln Pro Gly Gly Ser Leu Arg Leu Ser Cys Ala Ala Ser Gly
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<211> 28

<212> PRT

<213> Unknown Organism

<220>

<223> Description of Unknown Organism: pCES5

<400> 357

Ser Arg Asp Asn Ser Lys Asn Thr Leu Tyr Leu Gln Met Asn Ser Leu
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Ser Leu Ser Ile Arg Ser Gly Gln His Ser Pro Thr
 20 25

<210> 358

<211> 129

<212> PRT

<213> Unknown Organism

<220>

<223> Description of Unknown Organism: pCES5

<400> 358

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys
 1 5 10 15

Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr
 20 25 30

Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser
 35 40 45

Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser
 50 55 60

Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr
 65 70 75 80

Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys
 85 90 95

Lys Val Glu Pro Lys Ser Cys Ala Ala Ala His His His His His His
 100 105 110

Gly Ala Ala Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu Asn Gly Ala
 115 120 125

Ala
 <210> 359
 <211> 404
 <212> PRT
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Thr Val Glu Ser Cys Leu Ala Lys Pro His Thr Glu Asn Ser Phe Thr
 1 5 10 15

Asn Val Trp Lys Asp Asp Lys Thr Leu Asp Arg Tyr Ala Asn Tyr Glu
 20 25 30

Gly Cys Leu Trp Asn Ala Thr Gly Val Val Val Cys Thr Gly Asp Glu
 35 40 45

Thr Gln Cys Tyr Gly Thr Trp Val Pro Ile Gly Leu Ala Ile Pro Glu
 50 55 60

Asn Glu Gly Gly Gly Ser Glu Gly Gly Gly Ser Glu Gly Gly Gly Ser
 65 70 75 80

Glu Gly Gly Gly Thr Lys Pro Pro Glu Tyr Gly Asp Thr Pro Ile Pro
 85 90 95

Gly Tyr Thr Tyr Ile Asn Pro Leu Asp Gly Thr Tyr Pro Pro Gly Thr
 100 105 110

Glu Gln Asn Pro Ala Asn Pro Asn Pro Ser Leu Glu Glu Ser Gln Pro
 115 120 125

Leu Asn Thr Phe Met Phe Gln Asn Asn Arg Phe Arg Asn Arg Gln Gly
 130 135 140

Ala Leu Thr Val Tyr Thr Gly Thr Val Thr Gln Gly Thr Asp Pro Val
 145 150 155 160

Lys Thr Tyr Tyr Gln Tyr Thr Pro Val Ser Ser Lys Ala Met Tyr Asp
 165 170 175

Ala Tyr Trp Asn Gly Lys Phe Arg Asp Cys Ala Phe His Ser Gly Phe
 180 185 190

Asn Glu Asp Pro Phe Val Cys Glu Tyr Gln Gly Gln Ser Ser Asp Leu
 195 200 205

Pro Gln Pro Pro Val Asn Ala Gly Gly Gly Ser Gly Gly Gly Ser Gly
 210 215 220
 Gly Gly Ser Glu Gly Gly Gly Ser Glu Gly Gly Gly Ser Glu Gly Gly
 225 230 235 240
 Gly Ser Glu Gly Gly Gly Ser Gly Gly Gly Ser Gly Ser Gly Asp Phe
 245 250 255
 Asp Tyr Glu Lys Met Ala Asn Ala Asn Lys Gly Ala Met Thr Glu Asn
 260 265 270
 Ala Asp Glu Asn Ala Leu Gln Ser Asp Ala Lys Gly Lys Leu Asp Ser
 275 280 285
 Val Ala Thr Asp Tyr Gly Ala Ala Ile Asp Gly Phe Ile Gly Asp Val
 290 295 300
 Ser Gly Leu Ala Asn Gly Asn Gly Ala Thr Gly Asp Phe Ala Gly Ser
 305 310 315 320
 Asn Ser Gln Met Ala Gln Val Gly Asp Gly Asp Asn Ser Pro Leu Met
 325 330 335
 Asn Asn Phe Arg Gln Tyr Leu Pro Ser Leu Pro Gln Ser Val Glu Cys
 340 345 350
 Arg Pro Tyr Val Phe Gly Ala Gly Lys Pro Tyr Glu Phe Ser Ile Asp
 355 360 365
 Cys Asp Lys Ile Asn Leu Phe Arg Gly Val Phe Ala Phe Leu Leu Tyr
 370 375 380
 Val Ala Thr Phe Met Tyr Val Phe Ser Thr Phe Ala Asn Ile Leu Arg
 385 390 395 400

Asn Lys Glu Ser

<210> 360

<211> 69

<212> DNA

<213> Unknown Organism

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<223> Description of Unknown Organism: Kappa FR1 GLGs

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<210> 361

<211> 69

<212> DNA

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<223> Description of Unknown. Organism: Kappa FR1 GLGs

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<210> 362

<211> 69

<212> DNA

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<223> Description of Unknown Organism: Kappa FR1 GLGs

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<210> 363

<211> 69

<212> DNA

<213> Unknown Organism

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<223> Description of Unknown Organism: Kappa FR1 GLGs

<400> 363

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<210> 365
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atcacttgt 69
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atcacttgt 69
<210> 368
<211> 69
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atcacttgc 69
<210> 369
<211> 69
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<400> 369
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atcacttgc 69
<210> 370
<211> 69
<212> DNA
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atcacttgt 69
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<211> 69
<212> DNA
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<210> 378
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atctctgac 69
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<211> 69
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atctcctgc 69
<210> 383
<211> 69
<212> DNA
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<210> 384
<211> 69
<212> DNA
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atctcctgc 69
<210> 385
<211> 69
<212> DNA
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atctcctgc 69
<210> 386
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<400> 386
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atctcctgc 69
<210> 387
<211> 69
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ctctcctgc 69
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ctctcctgc 69
<210> 390
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ctctcctgc 69
<210> 391
<211> 69
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ctctcctgc 69
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<211> 69
<212> DNA
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<400> 392
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ctctcctgc 69
<210> 393
<211> 69
<212> DNA
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<400> 393
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ctctcctgc 69
<210> 394
<211> 69
<212> DNA
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atcaactgc 69
<210> 395
<211> 69
<212> DNA
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<400> 395
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atctcctgc 69

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<212> DNA
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tcctgc 66
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<212> DNA
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tcctgc 66
<210> 406
<211> 66
<212> DNA
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<220>
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tcctgc 66
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tcctgc 66
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acctgc 66
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<211> 66
<212> DNA
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acctgt 66
<210> 410
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<210> 411
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<210> 412
<211> 66
<212> DNA
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acatgc 66
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acctgt 66
<210> 414
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acctgc 66
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acctgc 66
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acctgc 66
<210> 417
<211> 66
<212> DNA
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acctgc 66
<210> 418
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<212> DNA
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acctgc 66
<210> 419
<211> 66
<212> DNA
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acctgc 66
<210> 420
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acctgc 66
<210> 422
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acctgt 66
<210> 425
<211> 66
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<400> 425
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acctgt 66
<210> 426
<211> 66
<212> DNA
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acttgt 66
<210> 427
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cagcctgtgc tgactcagcc acctctctga tcagcctccc tgggagcctc ggtcacactc 60
acctgc 66
<210> 428
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<212> DNA
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<400> 428

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acctgc 66

Patentkrav

1. Fremgangsmåde til fremstilling af et bibliotek, der præsenterer en uensartet familie af peptider, polypeptider eller proteiner på overfladen af en genetisk pakke, hvilken fremgangsmåde omfatter følgende trin:

(i) spaltning af en nukleinsyre i en ønsket stilling ved en fremgangsmåde, som omfatter følgende trin:

(a) etablering af kontakt mellem en enkeltstrenget nukleinsyre og et enkeltstrenget oligonukleotid, hvilket enkeltstrengede oligonukleotid er komplementært til den enkeltstrengede nukleinsyre i den region, hvor spaltningen ønskes; hvor den enkeltstrengede nukleinsyre og det enkeltstrengede oligonukleotid associerer sig til dannelse af en lokalt dobbeltstrenget region af den enkeltstrengede nukleinsyre, hvor den lokalt dobbeltstrengede region omfatter et restriktionsendonuklease-genkendelsessted; og

(b) spaltning af nukleinsyren på restriktionsendonuklease-genkendelsesstedet, hvor spaltningen omfatter etablering af kontakt mellem en restriktions-endonuklease og den lokalt dobbeltstrengede region, hvor restriktions-endonukleasen er specifik for restriktionsendonuklease-genkendelsesstedet;

idet trinnet til etablering af kontakt og spaltningstrinet gennemføres ved en temperatur på mellem 45°C og 75°C, hvor den enkeltstrengede nukleinsyre og det enkeltstrengede oligonukleotid associerer sig til dannelse af en lokalt dobbeltstrenget region af den enkeltstrengede nukleinsyre, hvor resten af den enkeltstrengede nukleinsyre er enkeltstrenget, og hvor restriktionsendonukleasen er aktiv ved den pågældende temperatur; og

(ii) præsentation af et medlem af familien af peptider, polypeptider eller proteiner, som den spaltede nukleinsyre koder for, på overfladen af en genetisk pakke, og samlet præsentation af mindst en del af familiens diversitet.

2. Fremgangsmåde til fremstilling af et bibliotek, der præsenterer en uensartet familie af peptider, polypeptider eller proteiner på overfladen af en genetisk pakke, hvilken

fremgangsmåde omfatter følgende trin:

(i) spaltning af en nukleinsyre i en ønsket stilling ved en fremgangsmåde, som omfatter følgende trin:

5 (a) etablering af kontakt mellem en enkeltstrengt nukleinsyre og et delvist dobbeltstrengt oligonukleotid, hvor den enkeltstrengede region af oligonukleotidet er komplementær til den enkeltstrengede nukleinsyre i den region, hvor spaltningen ønskes, og den dobbeltstrengede region af oligonukleotidet har et restriktionsendonuklease-genkendelsessted af type II-S; 10 hvor den enkeltstrengede nukleinsyre og den enkeltstrengede region af oligonukleotidet associerer sig til dannelse af en lokalt dobbeltstrengt region af den enkeltstrengede nukleinsyre, hvor den lokalt dobbeltstrengede region omfatter et spaltningssted af type II-S; og

15 (b) spaltning af nukleinsyren på spaltningsstedet af type II-S, hvor spaltningen omfatter etablering af kontakt mellem en restriktionsendonuklease af type II-S og den lokalt dobbeltstrengede region af den enkeltstrengede nukleinsyre, hvor restriktionsendonukleasen af type II-S er specifik for 20 restriktionsendonuklease-genkendelsesstedet af type II-S;

idet trinnet til etablering af kontakt og spaltningstrinnet gennemføres ved en temperatur på mellem 45°C og 75°C, hvor den enkeltstrengede nukleinsyre og den enkeltstrengede region af oligonukleotidet associerer sig til dannelse af en lokalt 25 dobbeltstrengt region af den enkeltstrengede nukleinsyre, hvor resten af den enkeltstrengede nukleinsyre er enkeltstrengt, og hvor endonukleasen af type II-S er aktiv ved den pågældende temperatur; og

30 (ii) præsentation af et medlem af familien af peptider, polypeptider eller proteiner, som de spaltede nukleinsyrer koder for, på overfladen af den genetiske pakke, og samlet præsentation af mindst en del af familiens diversitet.

3. Fremgangsmåde ifølge krav 1 eller 2, hvor nukleinsyrerne 35 koder for mindst en del af et immunoglobulin.

4. Fremgangsmåde ifølge krav 3, hvor immunoglobulinet omfatter et Fab eller enkeltkædet Fv.

5. Fremgangsmåde ifølge krav 3 eller 4, hvor immunoglobulinet omfatter mindst en del af en tung kæde eller mindst en del af en let kæde.

5

6. Fremgangsmåde ifølge et hvilket som helst af kravene 1-5, hvor nukleinsyresekvenserne i det mindste delvist stammer fra patienter, som har mindst én af en autoimmun sygdom og en cancer.

10

7. Fremgangsmåde ifølge krav 1 eller 2, som endvidere omfatter et trin til nukleinsyreamplificering inden trin (i) eller mellem trin (i) og (ii).

15

8. Fremgangsmåde ifølge krav 1, hvor restriktionsendonukleasen er valgt fra gruppen omfattende *MaeIII*, *Tsp45I*, *HphI*, *BsaJI*, *AluI*, *BlpI*, *DdeI*, *BglII*, *MslI*, *BsiEI*, *EaeI*, *EagI*, *HaeIII*, *Bst4CI*, *HpyCH4III*, *HinfI*, *MlyI*, *PleI*, *MnlI*, *HpyCH4V*, *BsmAI*, *BpmI*, *XmnI* og *SacI*.

20

9. Fremgangsmåde ifølge krav 5, hvor mindst en del af den tunge kæde og/eller den lette kæde er human.

10. Fremgangsmåde ifølge krav 3 eller 4, hvor immunoglobulinet omfatter mindst en del af FR1.

25

11. Fremgangsmåde ifølge krav 10, hvor mindst en del af det pågældende FR1 er humant.

12. Fremgangsmåde ifølge krav 6, hvor den autoimmune sygdom er valgt fra gruppen bestående af lupus erythematosus, systemisk sklerose, reumatoid arthritis, antiphospholipid-syndrom og vaskulitis.

30

13. Fremgangsmåde ifølge krav 12, hvor nukleinsyrerne er i det mindste delvist isoleret fra gruppen bestående af perifere blodceller, knoglemarvsceller, miltceller og lymfeknudeceller.

35

14. Fremgangsmåde ifølge krav 1 eller 2, som endvidere omfatter ligering af en delvist dobbeltstrenget syntetisk DNA-adaptor til den spaltede enkeltstrengede nukleinsyre og kloning af nukleinsyren ind i en vektor.

5

15. Fremgangsmåde ifølge krav 14, hvor den delvist dobbeltstrengede syntetiske DNA-adaptor omfatter en dobbeltstrenget region med 12-100 nukleotider.

10 16. Fremgangsmåde ifølge krav 14, hvor den delvist dobbeltstrengede syntetiske DNA-adaptor omfatter et endonuklease-restriktionssted.

15 17. Fremgangsmåde ifølge krav 14, hvor den delvist dobbeltstrengede syntetiske DNA-adaptor omfatter en enkeltstrenget region med 2-15 baser.

20 18. Fremgangsmåde ifølge krav 1 eller 2, som endvidere omfatter fremstilling af en samling af nukleinsyrer, der, i det mindste delvist, koder for medlemmer af den uensartede familie.

19. Fremgangsmåde ifølge krav 1 eller 2, som endvidere omfatter at gøre nukleinsyren enkeltstrenget.

25

20. Fremgangsmåde ifølge krav 2, hvor restriktionsendonukleasen af type II-S er valgt fra gruppen omfattende *AarI*CAC, *AceIII*, *Bbr7I*, *BbvI*, *BbvII*, *Bce83I*, *BceAI*, *BcefI*, *BciVI*, *BfiI*, *BinI*, *BscAI*, *BseRI*, *BsmFI*, *BspMI*, *EciI*,
30 *Eco57I*, *RauI*, *FokI*, *GsuI*, *HgaI*, *HphI*, *MboII*, *MlyI*, *MmeI*, *MnlI*, *PleI*, *RleAI*, *SfaNI*, *SspD5I*, *Sth132I*, *StsI*, *TaqII*, *Tth1111I* og *UbaPI*.

AMPLIFY VH GENES WITHOUT
USING VH SEQUENCES

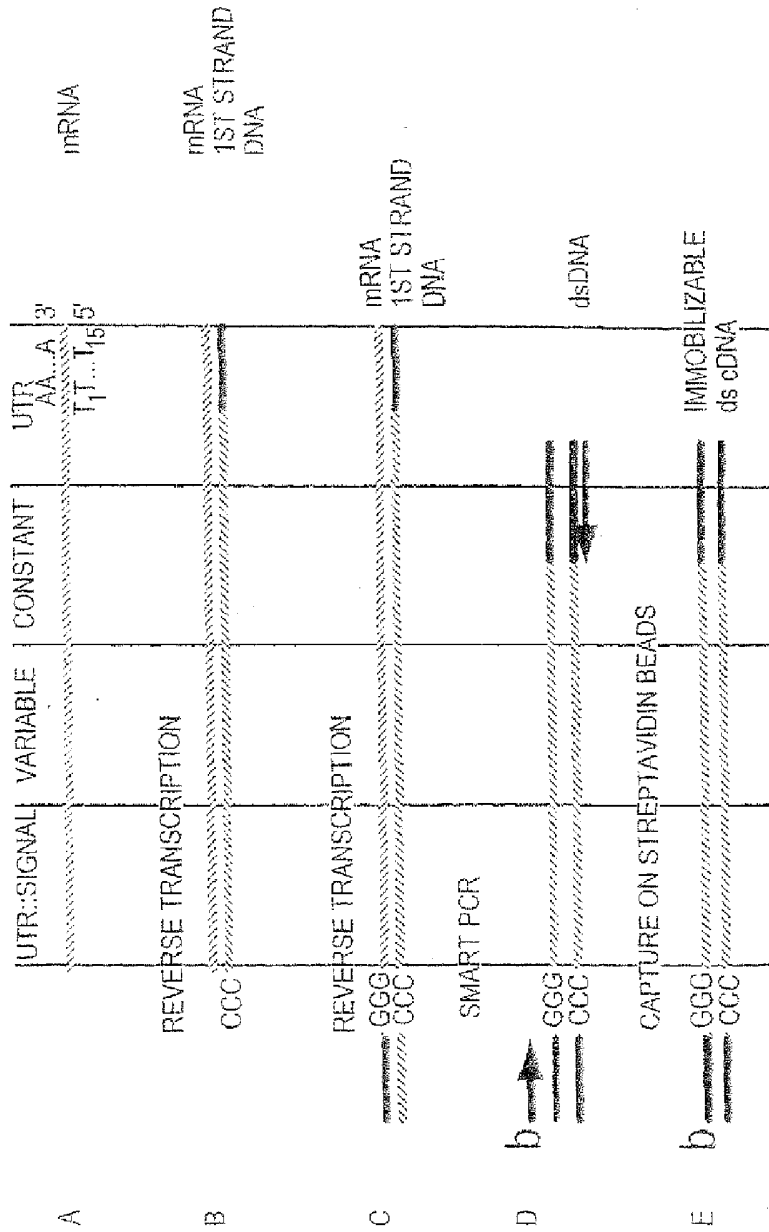
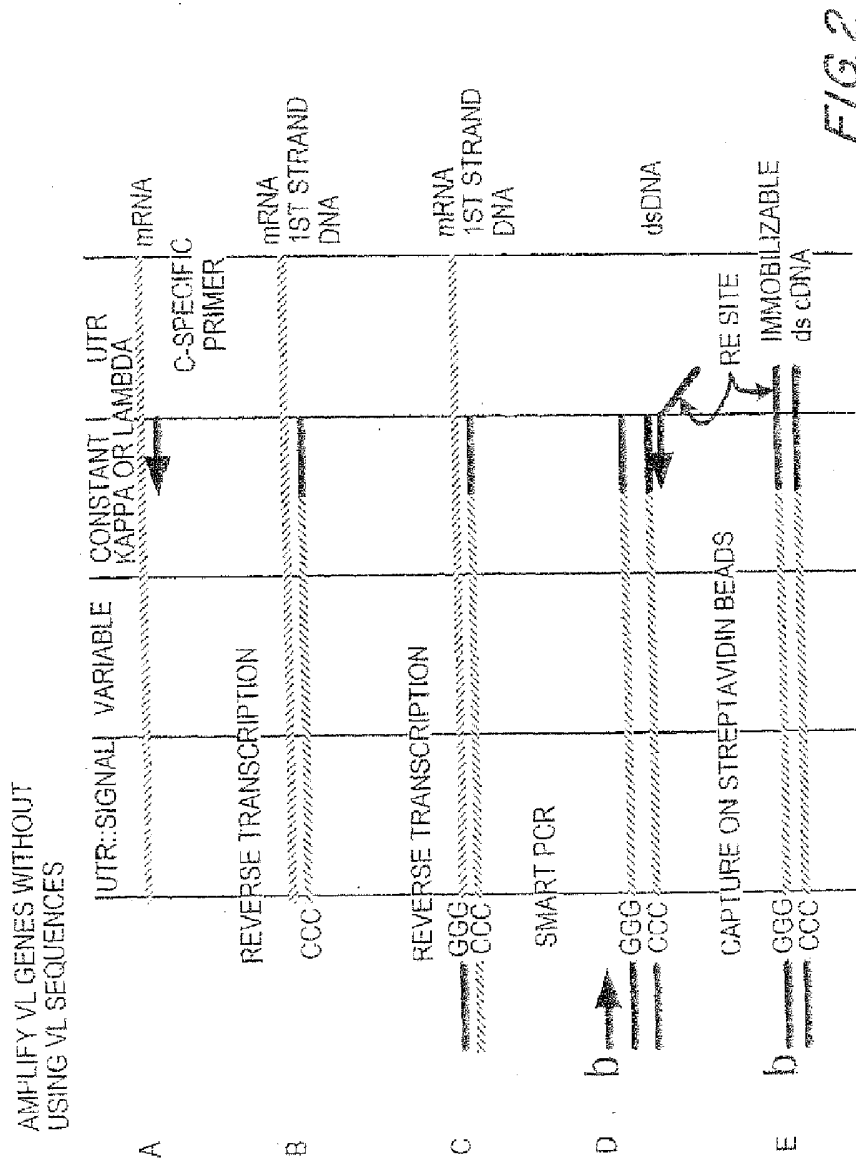


FIG. 1



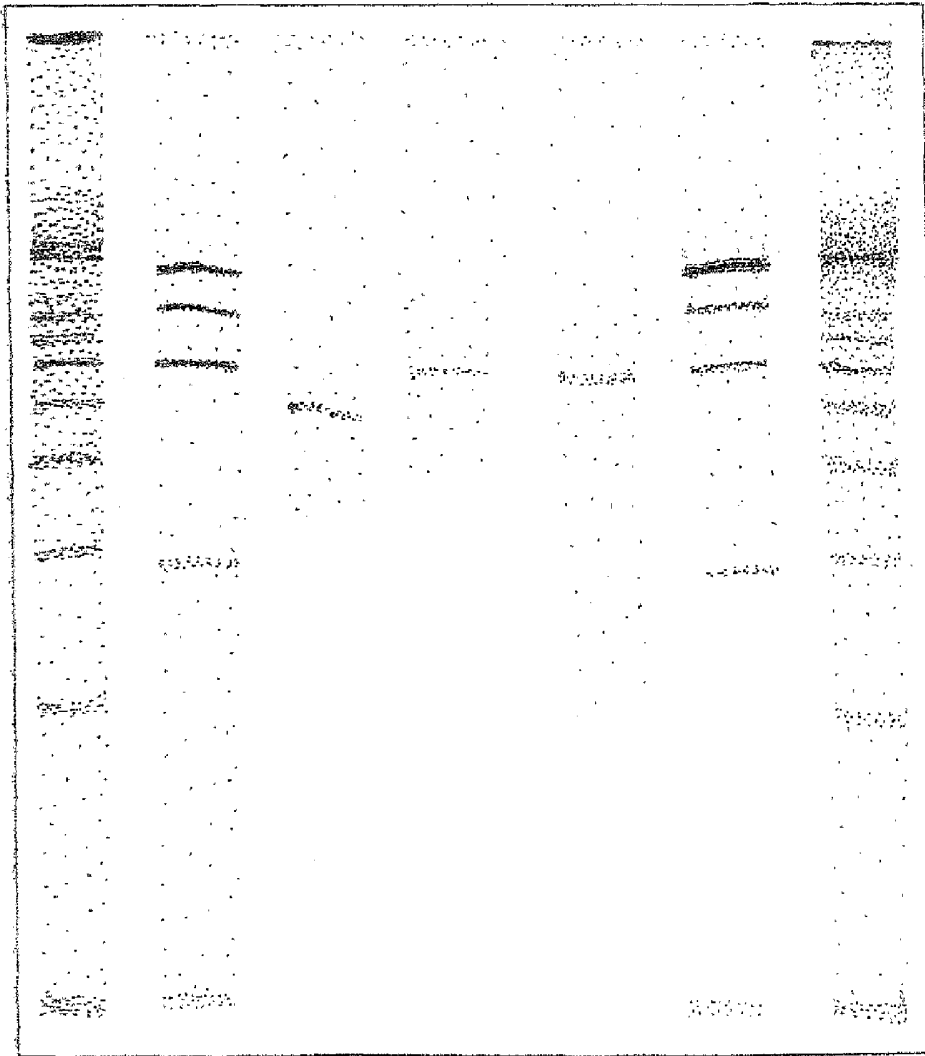


FIG. 3

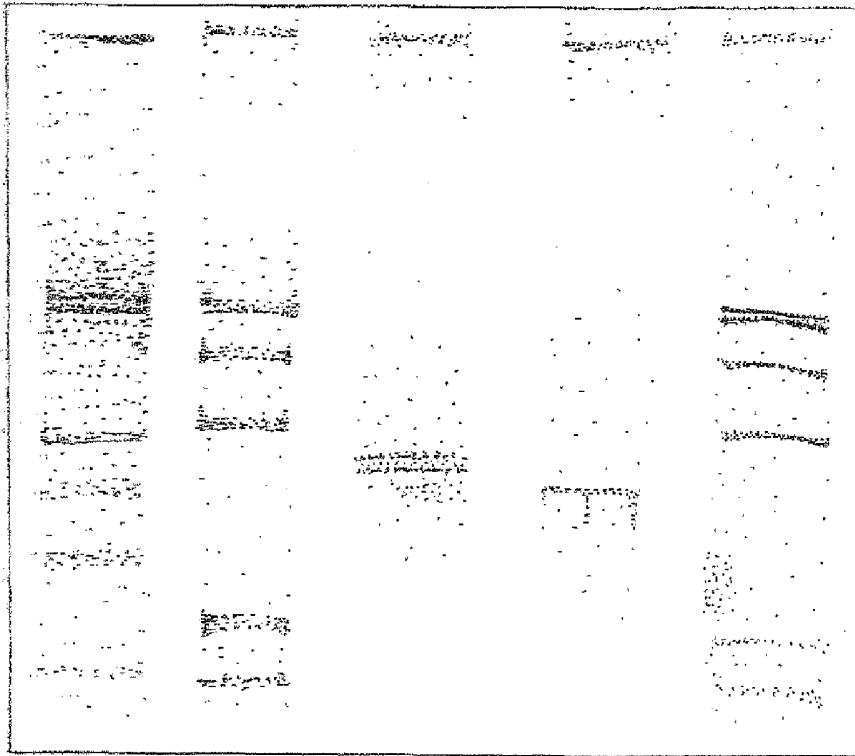


FIG. 4

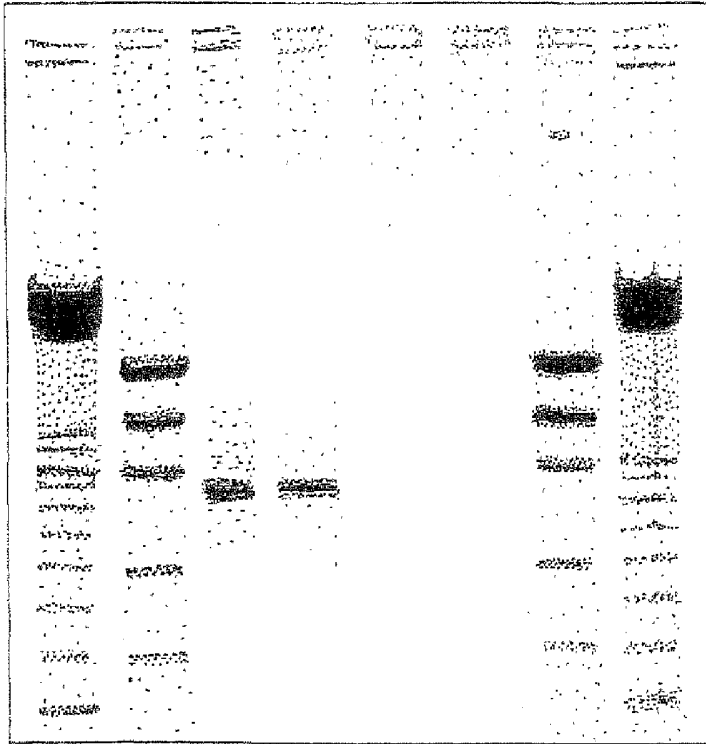


FIG. 5

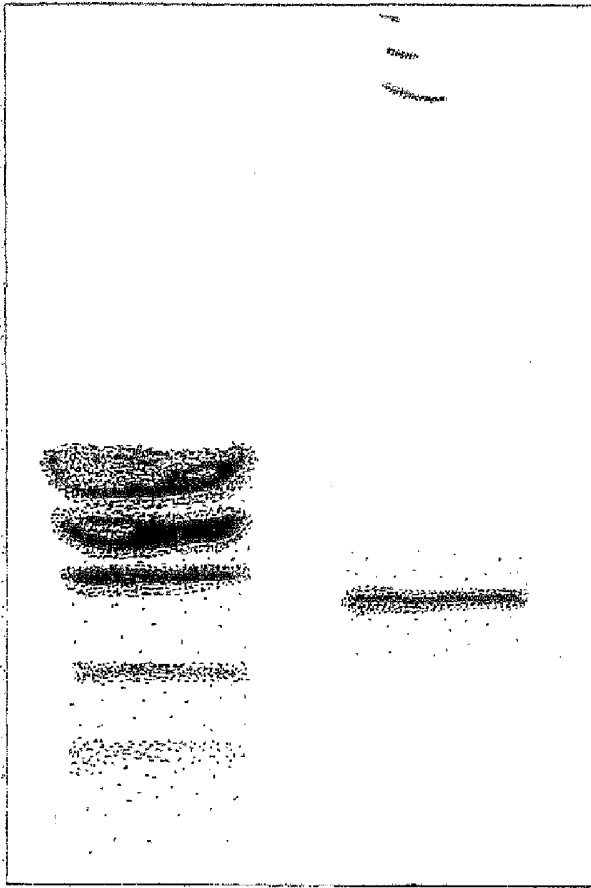


FIG. 6

Table 1: Cleavage of 75 human light chains.

Enzyme	Recognition*	Nch	Ns	Planned location of site
AfeI	AGCgct	0	0	
AflII	Cttaag	0	0	HC FR3
AgeI	Aacggc	0	0	
AecI	GGGgggca	0	0	After LC
BglII	Agatct	0	0	
BsiWI	Cgtacg	0	0	
BspDI	ATcgat	0	0	
BssHII	Gcgggc	0	0	
BstBI	TTcgaa	0	0	
DraIII	CACNNNgtc	0	0	
BaqI	Gggccg	0	0	
FseI	GGCCGcc	0	0	
FspI	TCCgca	0	0	
HpaI	GTTaac	0	0	
MfeI	Caatg	0	0	HC FR1
MluI	Acgogt	0	0	
NcoI	Ccaatg	0	0	Heavy chain signal
NheI	Gctagc	0	0	HC/anchor linker
NotI	GCggccgc	0	0	In linker after HC
NruI	TCCGca	0	0	
PacI	TTAATtaa	0	0	
PmaI	GTTTaaac	0	0	
PmlI	CACgtg	0	0	
PvuI	CGATcg	0	0	
SacII	CCGCGg	0	0	
SalI	Gtcgac	0	0	
SfiI	GGCCNNNNggccc	0	0	Heavy Chain signal
SgfII	GGGATcgc	0	0	
SnaBI	TACgta	0	0	
StuI	AGCctc	0	0	
XbaI	Tctaga	0	0	HC FR3
AatII	GACGTC	1	1	
AclI	AACgtt	1	1	
AseI	ATtaat	1	1	
BsmI	GAATGCN	1	1	
BspEI	Tccgga	1	1	HC FR1
BstXI	CCANNNNntgg	1	1	HC FR2
DrdI	GACNNNNngtc	1	1	
HindIII	Aacgtc	1	1	
PciI	Acatgt	1	1	
SapI	gaagagc	1	1	
ScaI	AGTact	1	1	
SexAI	Accwgg	1	1	
SpeI	Actagt	1	1	
TliI	Ctcgag	1	1	
XhoI	Ctcgag	1	1	
BcgI	cgannnnntgc	2	2	
ElpI	GCTnagc	2	2	
BssSI	Ctcgtg	2	2	
BstAP1	GCANNNNntgc	2	2	
EspI	GCTnagc	2	2	
KasI	Gggccc	2	2	
PELM1	CCANNNNntgg	2	2	
XmnI	GANNnnnttc	2	2	

ApaLI	Gtgcac	3	3	LC signal seq
NaeI	GCCggc	3	3	
NcoMI	Gccggc	3	3	
PvuII	CAGctg	3	3	
EsrII	CGgwcg	3	3	
BsrBI	GAGggg	4	4	
BsrDI	GCAATGNNn	4	4	
BstZ17I	GTAtac	4	4	
EcoRI	Gaattc	4	4	
SphI	GCATGc	4	4	
BspI	AATatt	4	4	
AccI	GTmkac	5	5	
BclI	Tgatca	5	5	
BsmBI	Nnnnnngagacg	5	5	
BsrGI	Tgtaca	5	5	
DraI	TTTaaa	6	6	
NdeI	CATatg	6	6	HC FR4
SwaI	ATTTaaat	6	6	
BamHI	Ggatcc	7	7	
SacI	GAGCTc	7	7	
BclVI	GTATCCNNNNNN	8	8	
BsaBI	GATNNnoatc	8	8	
NsiI	ATGCAI	8	8	
BspI20I	Gggccc	9	9	CHI
ApaI	GSGCCc	9	9	CHI
PspOoMI	Gggccc	9	9	
BspHI	Tcatga	9	11	
EcoRV	GATatc	9	9	
AndI	GACNNNngtc	11	11	
EbsI	GAAGAC	11	14	
PsiI	TTATAa	12	12	
BseI	GGTCTCNnnnn	13	15	
XmaI	Cccggg	13	14	
AvaI	Cycgfg	14	16	
BglI	GCCNNNnggc	14	17	
AlwNI	CAGNNNctg	16	16	
BspMI	ACCTGC	17	19	
XcmI	CCANNNNNNnnntgg	17	26	
BstEII	Ggtnacc	19	22	HC FR4
Sse8387I	CCDGCagg	20	20	
AvrII	Cctagg	22	22	
HincII	GTYtac	22	22	
BsgI	GTGCAG	27	29	
MscI	TGGcca	30	34	
BsaRI	NNnnnnnnnctcctc	32	35	
Bsu36I	Cctnagg	35	37	
PstI	CTGCAG	35	40	
BclI	nnnnnnnnntccgcc	38	40	
PpuMI	RGgwcy	41	50	
StyI	Ccwwgg	44	73	
BcoCI09I	RGgnccy	46	70	
Acc65I	Ggtacc	50	51	
EpnI	CGTACC	50	51	
BpmI	ctccag	53	82	
AvaII	Ggwoo	71	124	

* cleavage occurs in the top strand after the last upper-case base. For REs

that cut palindromic sequences, the lower strand is cut at the symmetrical site.

Table 2: Cleavage of 79 human heavy chains

Enzyme	Recognition	Nch	Ns	Planned location of site
AfeI	AGCgct	0	0	
AflII	Cttaag	0	0	HC FR3
AscI	GGGgggcc	0	0	After LC
BsiWI	Cgtacg	0	0	
BspBI	ATcgat	0	0	
BssHII	Gcgggc	0	0	
PseI	GGCCGGcc	0	0	
HpaI	GTTaac	0	0	
NheI	Gctagc	0	0	HC linker
NotI	GCggccgc	0	0	In linker, HC/anchor
NruI	TCGcga	0	0	
NsiI	ATGCAT	0	0	
PacI	TTAATtaa	0	0	
PclI	Acattg	0	0	
PmeI	GTTTaaac	0	0	
PvuI	CGATcg	0	0	
RsrII	CGgwcgg	0	0	
SapI	gaagagc	0	0	
SfiI	GGCCNNNNnggcc	0	0	HC signal seq
SgfI	GGCATcgc	0	0	
SwaI	ATTTaaat	0	0	
AclI	AACggt	1	1	
AgeI	Accggt	1	1	
AseI	ATtaat	1	1	
AvrII	Cctagg	1	1	
BsmI	GAATGCM	1	1	
BsrBI	GAGcgg	1	1	
BsrDI	GCATGNNn	1	1	
DraI	TCTaaa	1	1	
FspI	TGCgca	1	1	
HindIII	Aagctt	1	1	
MfeI	Caattg	1	1	HC FR1
NaeI	GCCggc	1	1	
NgoMI	Gccggc	1	1	
SpeI	Actagt	1	1	
Acc65I	Ggtacc	2	2	
BstBI	TTcgaa	2	2	
KpnI	GGTACC	2	2	
MluI	Acgcggt	2	2	
NcoI	Ccatgg	2	2	In HC signal seq
NdeI	CAtatg	2	2	HC FR4
EmlI	CACgctg	2	2	
XcmI	CCANNNNNnnntgg	2	2	
EcoGI	cgannnnntgc	3	3	
EclI	Tgatca	3	3	
EglI	GCCNNNNnggc	3	3	
BsaBI	GRTNNnnatc	3	3	
BsrGI	Tgtaca	3	3	
SnaBI	TACgta	3	3	
Sse8387I	CCTGCAGg	3	3	

ApaII	Gtgcac	4	4	LC Signal/FR1
BspHI	Tcatga	4	4	
BssSI	Ctcgtg	4	4	
PsiI	TTATAaa	4	5	
SphI	GCATGC	4	4	
AhdI	GACNNNngtgc	5	5	
BspEI	Tccgga	5	5	HC FR1
MscI	TGGcca	5	5	
SacI	GAGCTc	5	5	
ScaI	AGTact	5	5	
SexAI	Accwgg	5	6	
SspI	AATatu	5	5	
TliI	Ctcgag	5	5	
XhoI	Ctcgag	5	5	
BbsI	GAAGAC	7	8	
BstAPI	GCANNNNntgc	7	8	
BstZ17I	GTAtac	7	7	
EcoRV	GATatc	7	7	
EcoRI	Gaattc	8	8	
BlnI	GCtnagc	9	9	
Bsu36I	CCtnagg	9	9	
DraEII	CACNNNgtg	9	9	
EspI	GCtnagc	9	9	
StuI	AGGcct	9	13	
XbaI	Tctaga	9	9	HC FR3
Bsp120I	Gggccc	10	11	CHI
ApaI	GGGCCc	10	11	CHI
BspOOMI	Gggccc	10	11	
SciVI	GTATCCNNNNNN	11	11	
SalI	Gtcgac	11	12	
DrdI	GACNNNNngtgc	12	12	
KasI	Ggcgcc	12	12	
XraI	Cccggg	12	14	
BglII	Agatct	14	14	
HincII	GTYrac	16	18	
BamHI	Ggatcc	17	17	
FelMI	CCANNNNntgg	17	18	
BsmBI	Nnnnnggagacg	18	21	
BstXI	CCANNNNntgg	18	19	HC FR2
XmnI	GAANNnttc	18	18	
SacII	CCGCgg	19	19	
PstI	CTGCag	20	24	
FvuII	CAGctg	20	22	
AvaI	Cycgrg	21	24	
BaqI	Cggccc	21	22	
AatII	GACGtc	22	22	
BspMI	ACCTGC	27	33	
AccI	GTmkac	30	43	
StyI	Ccwggg	35	49	
AlwNI	CAGNNNctg	38	44	
BsaI	GGTCTCNannn	38	44	
PpuMI	RGgwccy	43	46	
BsgI	GTGCAG	44	54	
BseRI	NNnnnnnnnctcctc	48	80	
EciI	nnnnnnnnntccgcc	52	57	
BstEII	Ggtgacc	54	61	HC FR4, 47/79 have one
EcoO109I	RGgnccy	54	86	

BpmI	ctccag	60	121
AvaiI	Ggwcc	71	140

Table 5 (~~continued~~): Use of *FokI* as "Universal Restriction Enzyme"

FokI - for dsDNA, | represents sites of cleavage

 sites of cleavage
 5'-cacGGATGtg--nnnnnnn|nnnnnnn-3' (SEQ ID NO:15)
 3'-gtgCCTACac--nnnnnnnnnn|nnn-5' (SEQ ID NO:16)
 RECOG
 NITion of FokI

Case I

5'-...gtg|tatt-actgtgc..substrate....-3' (SEQ ID NO:17)
 3'-cac-ataa|tcacacc-
 gtGTACGcac\
 5'- cacATCCgtg/(SEQ ID NO:18)

Case II

5'-...gtgtatt|agac-tgc..substrate....-3' (SEQ ID NO:19)
 cacataa-tctg|acy-5'
 /gtgCCTACac
 \cacGGATctg-3' (SEQ ID NO:20)

Case III (Case I rotated 180 degrees)

/gtgCCTACac-5'
 \cacGGATGtg-
 gtgtatt|acag-tcc-3' Adapter (SEQ ID NO:21)
 3'-...cacagaa-tgto|agg..substrate....-5' (SEQ ID NO:22)

Case IV (Case II rotated 180 degrees)

```

          3'- gtGTAGGcaac\ (SEQ ID NO:23)
             rcaCATCCgtg/
5'-gag|cttc-actgagc
Substrate 3'-...ctc-agag|tgactcg...-5' (SEQ ID NO:24)

```

Improved FokI adapters

FokI - for dsDNA, | represents sites of cleavage

Case I

Stem 11, loop 5, stem 11, recognition 17

```

          5'-...catgtg|tatt-actgtgc..Substrate....-3'
          3'-gtacac-ataa|tgacacc-rT-
             gtGTAGGcaacG T
          5'- caCATCCgtgc C
             LTTJ

```

Case II

Stem 10, loop 5, stem 10, recognition 18

```

          5'-...gtgtatt,agac-tggtgcc..Substrate....-3'
          rT- rcaataa-tctg|agacgg-5'
T  gtgCCTACag
C  cacGCATGtg-3'
LTTJ

```

Case III (Case I rotated 180 degrees)

Stem 11, loop 5, stem 11, recognition 20

```

          r T-
T  TgtgCCTACac-3'
G  AcacGGATGtg-
LTTJ
          gtgtctt|acag-tccattctg-3' Adapter
          3'-...cacaga-tgtc|aggtagac..substrate....-5'

```

Case IV (Case II rotated 180 degrees)

Stem 11, loop 4, stem 11, recognition 17

```

          rT-
          3'- gtGTAGGcaac T
             rcaCATCCgtgg T
          5'-atcgag|cttc-actgagc
Substrate 3'-...tagctc-agag|tgactcg...-5'
             LTTJ

```

BseRI

```

! sites of cleavage
5'-cacGAGGAGnnnnnnnnnn|nnnn-3'
3'-gtggtcctcnnnnnnnn|nnnnn-5'
      RECOG
      NITion of BseRI

```

Stem 11, loop 5, stem 11, recognition 19

```

      3'-.....gaacat}cg-ttaagccagta.....5'
{T-T}      cttgta-gc|aatteggatcat-3'
C   GCTGAGGAGTC-|
T   cgactcctcag-5'  An adapter for BseRI to cleave the substrate above.
|g...|

```

Table 8: Matches to URE FR3 adapters in 79 human HC.

A. List of Heavy-chains genes sampled

AFC08566	af103343	HSA235676	HSU92452	HSZ93860
AFC35043	AF103367	HSA235675	HSU94412	HSZ93863
AF103026	AF103368	HSA235674	HSU94415	MCOMFRAA
af103033	AF103369	HSA235673	HSU94416	MCOMFRVA
AF103061	AF103370	HSA240559	HSU94417	S62745
AF103072	af103371	HSCB201	HSU94418	S62764
af103078	AF103372	HSIGGVHC	HSU96389	S83240
AF103099	AF158381	HSU44791	HSU96391	SABVH369
AF103102	E05213	HSU44793	HSU96392	SADEIGVH
AF103103	E05886	HSU82771	HSU96395	SAK2IGVH
AF103174	E05887	HSU82949	HSZ93849	SDA3IGVH
AF103186	HSA235661	HSU82950	HSZ93850	SIGVHTTD
af103187	HSA235664	HSU82952	HSZ93851	SUK4IGVH
AF103195	HSA235660	HSU82961	HSZ93853	
af103277	HSA235659	HSU86522	HSZ93855	
af103286	HSA235678	HSU86523	HSZ93857	
AF103309	HSA235677			

Table 8 B. Testing all distinct GLGs from bases 89.1 to 93.2 of the heavy variable domain

Id	Nb	0	1	2	3	4	Seq	SEQ ID NO:
1	38	15	11	10	0	2	Seq1 gtgtattactgtgc	25
2	19	7	6	4	2	0	Seq2 gtAtattactgtgc	26
3	1	0	0	1	0	0	Seq3 gtgtattactgtAA	27
4	7	1	5	1	0	0	Seq4 gtgtattactgtAc	28
5	0	0	0	0	0	0	Seq5 Ttgtattactgtgc	29
6	0	0	0	0	0	0	Seq6 TtgtatCactgtgc	30
7	3	1	0	1	1	0	Seq7 ACAtattactgtgc	31
8	2	0	2	0	0	0	Seq8 ACgtattactgtgc	32
9	9	2	2	4	1	0	Seq9 ATgtattactgtgc	33
Group		26	26	21	4	2		
Cumulative		26	52	73	77	79		

Table 8C Most important URE recognition seqs in FR3 Heavy

1	VHSzy1	GTGtattactgtgc	(ON_SHC103)	(SEQ ID NO:25)
2	VHSzy2	GTAtattactgtgc	(ON_SHC323)	(SEQ ID NO:26)
3	VHSzy4	GTGtattactgtac	(ON_SHC349)	(SEQ ID NO:28)
4	VHSzy9	ATGtattactgtgc	(ON_SHC5a)	(SEQ ID NO:33)

Table 8D, testing 79 human HC V genes with four probes

Number of sequences.....		79						
Number of bases.....		29143						
		Number of mismatches						
Id	Best	0	1	2	3	4	5	
1	39	15	11	10	1	2	0	Seq1 gtgtattactgtgc (SEQ ID NO:25)
2	22	7	6	5	3	0	1	Seq2 gtAtattactgtgc (SEQ ID NO:26)
3	7	1	5	1	0	0	0	Seq4 gtgtattactgtAc (SEQ ID NO:28)
4	11	2	4	4	1	0	0	Seq9 ATgtattactgtgc (SEQ ID NO:33)
Group		25	26	20	5	2		
Cumulative		25	51	71	76	78		

One sequence has five mismatches with sequences 2, 4, and 9; it is scored as best for 2.

Id is the number of the adapter.

Best is the number of sequence for which the identified adapter was the best available.

The rest of the table shows how well the sequences match the adapters. For example, there are 11 sequences that match VHSzy1 (Id=1) with 2 mismatches and are worse for all other adapters. In this sample, 90% come within 2 bases of one of the four adapters.

Table 130: PCR primers for amplification of human Ab genes

```

(HuIgMFOR)      5'-tgg aag agg cac gtt ctt ttc ttc-3'
!(HuIgMFORBtop)5'-aaa gaa aag aac gng cct ctt cca-3' = reverse complement
(HuChkFOR)      5'-aca ctc tcc ccg gtt gaa gct ctt-3'
(HuCL2FOR)      5'-tga nca ttc tgt agg ggc cac tg-3'
(HuCL7FOR)      5'-aga gca ttc tgc agg ggc cac tg-3'
! Kappa
(CKForeAsc)    5'-acc gcc tcc acc ggg cgc gcc tta tta aga ctc tcc cot gtt-
                gaa gct ctt-3'
(CL2ForeAsc)   5'-acc gcc tcc acc ggg cgc gcc tta tta tga aca ttc tgt-
                agg ggc cac tg-3'
(CL7ForeAsc)   5'-acc gcc tcc acc ggg cgc gcc tta tta aga gca ttc tgc-
                agg ggc cac tg-3'

```

Table 195: Human GLG FR3 sequences

```

! VH1
! 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80

```



```

agg gtc acc atg acc agg gac acg tcc atc agc aca gcc tac atg
: 81 82 82a 82b 82c 83 84 85 86 87 88 89 90 91 92
gag ctg agc agg ctg aga tct gac gac acg gcc gtg tat tac tgt
: 93 94 95
gcg aga ga ! 1-02# 1
aga gtc acc att acc agg gac aca tcc gcg agc aca gcc tac atg
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
gcg aga ga ! 1-03# 2
aga gtc acc atg acc agg aac acc tcc ata agc aca gcc tac atg
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
gcg aga gg ! 1-08# 3
aga gtc acc atg acc aca gac aca tcc acg agc aca gcc tac atg
gag ctg agg agc ctg aga tct gac gac acg gcc gtg tat tac tgt
gcg aga ga ! 1-18# 4
aga gtc acc atg acc gag gac aca tct aca gac aca gcc tac atg
gag ctg agc agc ctg aga tct gag gac aca gcc atg tat tac tgt
gca aca ga ! 1-24# 5
aga gtc acc att acc agg gac agg tct atg agc aca gcc tac atg
gag ctg agc agc ctg aga tct gag gac aca gcc atg tat tac tgt
gca aga ta ! 1-45# 6
aga gtc acc atg acc agg gac acg tcc acg agc aca gtc tac atg
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
gcg aga ga ! 1-46# 7

```

aga gtc acc att acc agg gac atg tcc aca agc aca gcc tac atg
 gag ctg agc agc ctg aga tcc gag gac acg gcc gtg tat tac tgt
 gcg gca ga ! 1-58# 8
 aga gtc acc att acc gcg gac gaa tcc acg agc aca gcc tac atg
 gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
 gcg aga ga ! 1-69# 9
 aga gtc acc att acc gcg gac aaa tcc acg agc aca gcc tac atg
 gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
 gcg aga ga ! 1-e# 10
 aga gtc acc ata acc gcg gac acg tct aca gac aca gcc tac atg
 gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt
 gca aca ga ! 1-f# 11
 ! VH2
 agg ctg acc atc acc aag gac acc tcc aaa aac cag gtg gtc ctt
 aca atg acc aac atg gac cct gtg gac aca gcc aca tat tac tgt
 gca cac aga c! 2-05# 12
 agg ctg acc atc tcc aag gac acc tcc aaa agc cag gtg gtc ctt
 acc atg acc aac atg gac cct gtg gac aca gcc aca tat tac tgt
 gca cgg ata c! 2-26# 13
 agg ctg acc atc tcc aag gac acc tcc aaa aac cag gtg gtc ctt
 acc atg acc aac atg gac cct gtg gac aca gcc aac tat tac tgt
 gca cgg ata c! 2-70# 14
 ! VH3
 cga ttc acc atc tcc aga gac aac gcc aag aac tca ctg tat ctg
 caa atg aac agc ctg aga gcc gag gac acg gcc gtg tat tac tgt
 gcg aga ga ! 3-07# 15
 cga ttc acc atc tcc aga gac aac gcc aag aac tca ctg tat ctg
 caa atg aac agt ctg aga gcc gag gac acg gcc ttg tat tac tgt
 gcc aaa gat a! 3-09#16
 cga ttc acc atc tcc agg gac aac gcc aag aac tca ctg tat ctg
 caa atg aac agc ctg aga gcc gag gac acg gcc gtg tat tac tgt
 gcg aga ga ! 3-11# 17
 cga ttc acc atc tcc aga gaa aat gcc aag aac tcc ttg tat ctg
 caa atg aac agc ctg aga gcc ggg gac acg gat gtg tat tac tgt
 gca aga ga ! 3-13# 18
 aga ttc acc atc tcc aga gat gat tca aaa aac aag ctg tat ctg
 caa atg aac agc ctg aaa acc gag gac aca gcc gtg tat tac tgt
 acc aca ga ! 3-15# 19
 cga ttc acc atc tcc aga gac aac gcc aag aac tca ctg tat ctg

caa atg aac agt ctg aga gcc gag gac acg gcc ttg tac cac tgt
gcg aga ga ! 3-20# 20
cga ttc acc atc tcc aga gac aat tcc aag aac tca ctg tat ctg
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt
gcg aga ga ! 3-21# 21
cgg ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gcc gag gac acg gcc gta tat tac tgt
gcg aaa ga ! 3-23# 22
cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt
gcg aaa ga ! 3-30# 23
cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt
gcg aga ga ! 3303# 24
cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt
gcg aaa ga ! 3305# 25
cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt
gcg aga ga ! 3-33# 26
cga ttc acc atc tcc aga gac aac agc aaa aac tcc ctg tat ctg
caa atg aac agt ctg aga act gag gac acc gcc ttg tat tac tgt
gca aaa gat a! 3-43#27
cga ttc acc atc tcc aga gac aat gcc aag aac tca ctg tat ctg
caa atg aac agc ctg aga gac gag gac acg gct gtg tat tac tgt
gcg aga ga ! 3-48# 28
aga ttc acc atc tcc aga gat ggt tcc aaa agc atc gcc tat ctg
caa atg aac agc ctg aaa acc gag gac aca gcc gtg tat tac tgt
act aga ga ! 3-49# 29
cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gcc gag gac acg gcc gtg tat tac tgt
gcg aga ga ! 3-53# 30
aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg gcc agc ctg aga gct gag gac atg gct gtg tat tac tgt
gcg aga ga ! 3-64# 31
aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt
ycg aga ga ! 3-66# 32
aga ttc acc atc tca aga gat gat tca aag aac tca ctg tat ctg

caa atg aac agc ctg aaa acc gag gac acg gcc gtg tat tac tgt
gct aga ga ! 3-72# 33

agg ttc acc atc tcc aga gat gat tca aag aac acg gcg tat ctg
caa atg aac agc ctg aaa acc gag gac acg gcc gtg tat tac tgt
act aga ca ! 3-73# 34

oga ttc acc atc tcc aga gac aac gcc aag aac acg ctg tat ctg
caa atg aac agt ctg aga gcc gag gac acg gct gtg tat tac tgt
gca aga ga ! 3-74# 35

aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg cat ctt
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt
aag aaa ga ! 3-d# 36

! V34

oga gtc acc ata tca gta gac aag tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gcc gcg gac acg gcc gtg tat tac tgt
gcg aga ga ! 4-04# 37

oga gtc acc atg tca gta gac acg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gcc gcg gac acg gcc gtg tat tac tgt
gcg aga aa ! 4-28# 38

oga gtt acc ata tca gta gac acg tct aag aac cag ttc tcc ctg
aag ctg agc tct gtg act gcc gcg gac acg gcc gtg tat tac tgt
gcg aga ga ! 4301# 39

oga gtc acc ata tca gta gac agg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gcc gcg gac acg gcc gtg tat tac tgt
gcc aga ga ! 4302# 40

oga gtt acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg act gcc gca gac acg gcc gtg tat tac tgt
gcc aga ga ! 4304# 41

oga gtt acc ata tca gta gac acg tct aag aac cag ttc tcc ctg
aag ctg agc tct gtg act gcc gcg gac acg gcc gtg tat tac tgt
gcg aga ga ! 4-31# 42

oga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gcc gcg gac acg gct gtg tat tac tgt
gcg aga ga ! 4-34# 43

oga gtc acc ata tcc gta gac acg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gcc gca gac acg gct gtg tat tac tgt
gcg aga ca ! 4-39# 44

oga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg
aag ctg agc tct gtg acc gct gcg gac acg gcc gtg tat tac tgt
gcg aga ga ! 4-59# 45

oga gtc acc ata tca gra gac acg tcc aag aac cag ttc tcc ctg
 aag ctg agc tct gtg acc got gcg gac acg gcc gtg tat tac tgt
 gcg aga ga ! 4-61# 46

oga gtc acc ata tca gra gac acg tcc aag aac cag ttc tcc ctg
 aag ctg agc tct gtg acc gcc gca gac acg gcc gtg tat tac tgt
 gcg aga ga ! 4-b# 47

! VII5

cag gtc acc atc tca gcc gac aag tcc atc agc acc gcc tac ctg
 cag tgg agc agc ctg aag gcc tcg gac acc gcc atg tat tac tgt
 gcg aga ca ! 5-51# 48

cag gtc acc atc tca gct gac aag tcc atc agc act gcc tac ctg
 cag tgg agc agc ctg aag gcc tcg gac acc gcc atg tat tac tgt
 gcg aga ! 5-a# 49

! VII6

oga ata acc atc aac cca gac acc tcc aag aac cag ttc tcc ctg
 cag ctg aac tct gtg act ccc gag gac acg gct gtg tat tac tgt
 gca aga ga ! 6-1# 50

! VII7

ogg ttt gtc ttc tcc ttg gac acc tct gtc agc acc gca tat ctg
 cag atc tgc agc cta aag gct gag gac act gcc gtg tat tac tgt
 gcg aga ga ! 74.1# 51

Table 250: REaptors, Extenders, and Bridges used for Cleavage and Capture of Human Heavy Chains in FR3.

A: HpyCH4V Probes of actual human HC genes

HpyCH4V in FR3 of human HC, bases 35-56; only those with TGca site

TGca;10,

RE recognition:tgca

of length 4 is expected at 10

1

6-1 agttctccctgcagctgaactc

2	3-11,3-07,3-21,3-72,3-48	cactgtatctgcaaatgaacag
3	3-09,3-43,3-20	ccctgtatctgcaaatgaacag
4	5-51	ccgcctaccctgcagtggagcag
5	3-15,3-30,3-30.5,3-30.3,3-74,3-23,3-33	cgctgtatctgcaaatgaacag
6	7-4.1	cgccatctctgcagatctgcag
7	3-73	cgccgtatctgcaaatgaacag
8	5-a	ctgcctaccctgcagtggagcag
9	3-49	tcgcatctctgcaaatgaacag

B: HpyCH4V REAdaptors, Extenders, and Bridges

B.1 REAdaptors

! Cutting HC lower strand:

! TmKeller for 100 mM NaCl, zero formamide

! Edaptors for cleavage

		T _m ^N	T _m ^F
(ON_HCFR36-1)	5'-agttctcccTGCAgctgaactc-3'	68.0	64.5
(ON_HCFR36-1A)	5'-ctctcccTGCAgctgaactc-3'	62.0	62.5
(ON_HCFR36-1B)	5'-ctctcccTGCAgctgaac-3'	56.0	59.9
(ON_HCFR33-15)	5'-cgctgtatcTGCAaatgaacag-3'	64.0	60.8
(ON_HCFR33-15A)	5'-ctgtatcTGCAaatgaacag-3'	56.0	56.2
(ON_HCFR33-15B)	5'-ctgtatcTGCAaatgaac-3'	50.0	53.1
(ON_HCFR33-11)	5'-cactgtatcTGCAaatgaacag-3'	62.0	53.9
(ON_HCFR35-51)	5'-ccgcctaccTGCAgtggagcag-3'	74.0	70.1

B.2 Segment of synthetic 3-23 gene into which captured CDR3 is to be cloned

```
!
!                               XbaI...
!B323*  cgcttcactaaag tcT aga gac aac tct aag aat act ctC taC
!       scab..... designed gene 3-23 gene.....
!
```

```
!   HpyCH4V
!   .. ..   PflII...
!   Ttg caG atg aac agc TAA agG . . .
!   .....
!
```

B.3 Extender and Bridges

! Extender (bottom strand):

```
(ON_HCHpyEx01)  5'-cAAgTAgAgAgTATTcTTAgAgTTgTcTcTAAgAcTTAgTgAAgcg-3'
```

! ON_HCHpyEx01 is the reverse complement of

```
! 5'-cgcttcactaaag tcT aga gac aac tct aag aat act ctC taC Ttg -3'
```

! Bridges (top strand, 9-base overlap):

```

!
! ON_HCHpyBr016-1) 5'-cgCttcacTaag tcT aga gac aaC tcT aag-
                aaT acT ctC taC Ttg CAgctgaac-3' (3'-term C is blocked)
!
! 3-15 et al. + 3-11
! ON_HCHpyBr023-15) 5'-cgCttcacTaag tcT aga gac aaC tcT aag-
                aaT acT ctC taC Ttg CAaatgaac-3' (3'-term C is blocked)
!
! 5-51
! ON_HCHpyBr045-51) 5'-cgCttcacTaag tcT aga gac aaC tcT aag-
                aaT acI ctC taC Ttg CAgtggaac-3' (3'-term C is blocked)
!
! PCR primer (top strand)
!
! ON_HCHpyPCR)      5'-cgCttcacTaag tcT aga gac-3'
!

```

C: BspI Probes from human HC GLGs

1	1-58, 1-03, 1-08, 1-69, 1-24, 1-45, 1-46, 1-5, 1-e	acatggaGCTGAGCagcctgag
2		1-02 acatggaGCTGACCagcctgag
3		1-18 acatggagctgaggagcctgag
4		5-51, 5-a acctgcagtggagcagcctgaa
5	3-15, 3-73, 3-49, 3-72	atctgcaaatgaacagcctgaa
6	3303, 3-33, 3-07, 3-11, 3-30, 3-21, 3-23, 3305, 3-48	atctgcaaatgaacagcctgag
7	3-20, 3-7a, 3-09, 3-43	atctgcaaatgaacagcctgag
8		74.1 atctgcagatctgcagcctaaa
9	3-66, 3-13, 3-53, 3-d	atcttcaaatgaacagcctgag
10		3-64 atcttcaaatgggcagcctgag
11	4301, 4-28, 4302, 4-34, 4304, 4-31, 4-34, 4-39, 4-59, 4-61, 4-b	ccctgaaGCTGAGCctctgtgac
12		6-1 ccttgcaactgaactctgtgac
13	2-79, 2-05	tccttacaatgaccaacatgga
14		2-26 tccttaccatgaccaacatgga

D: BspI REadaptors, Extenders, and Bridges

D.1 REadaptors

		T_m^*	T_m^K
(BspF3HC1-58)	5'-ac atg gag CTG AGC agc ctg ag-3'	70	66.4
(BspF3HC6-1)	5'-cc ctg aag ctg agc tct gct ac-3'	70	66.4

! BspF3HC6-1 matches 4-30.1, not 6-1.

D.2 Segment of synthetic 3-23 gene into which captured CDR3 is to be cloned


```

!
!                               BspI
!                               . . . . .
!D323+  cgCttcacTcaag TCT AGR gac aac tct aag aat act ctC taC Ttg caG atg aac
!
!                               AflIII...
!                               agC TTA AGG
!

```

D.3 Extender and Bridges

```

! Bridges
(BlpF3Br1) 5'-cgCttcacTcag tct aga gat aac AGT aAA aat act Ttg-
           taC Ttg caG Ctg a|5C agc ctg-3'
(BlpF3Br2) 5'-cgCttcacTcag tct aga gat aac AGT aAA aat act Ttg-
           taC Ttg caG Ctg a|gc tct ctg-3'
!
!                               | lower strand is cut here
! Extender
(BlpF3Ext) 5'-
TcAgcTgcAAgTAcAAAATATTTTTTAcTgTTATcTcTAgAcTgAgTgAAAgcg-3'
! BlpF3Ext is the reverse complement of:
! 5'-cgCttcacTcag tct aga gat aac AGT aAA aat act Ttg taC Ttg caG Ctg a-3'
!
(BlpF3PCR) 5'-cgCttcacTcag tct aga gat aac-3'

```

B: HpyCH4III Distinct GLG sequences surrounding site, bases 77-98

1	102#1, 116#1, 146#7, 169#9, 1e#10, 311#17, 353#30, 404#37, 4301	ccgtgtattactgtgagagaga
2	103#2, 307#15, 321#21, 3305#24, 333#26, 348#28, 364#31, 366#32	ctgtgtattactgtgagagaga
3		106#3 ccgtgtattactgtgagagag
4		124#5, 17#11 ccgtgtattactgtgcaacaga
5		145#6 ccctgtattactgtgcaagata
6		158#8 ccgtgtattactgtgagagaga
7		205#12 ccacatattactgtgcaacag
8		226#13 ccacatattactgtgcaagat
9		270#14 ccactgtattactgtgcaagat
10		309#16, 343#27 ccttgtattactgtgcaaaaga
11		313#18, 374#35, 61#50 cbtgtgtattactgtgcaagaga
12		315#19 ccgtgtattactgtgcaacaga
13		320#20 ccttgtattactgtgagagaga
14		323#22 ccgtgtattactgtgcaagaga
15		330#21, 3305#25 ctgtgtattactgtgagagaga
16		349#29 ccgtgtattactgtgcaagaga
17		372#33 ccgtgtattactgtgcaagaga
18		373#34 ccgtgtattactgtgcaagaga
19		3d#36 ctgtgtattactgtgcaagaga
20		428#38 ccgtgtattactgtgcaagaga
21		4302#40, 4304#41 ccgtgtattactgtgcaagaga
22		439#44 ctgtgtattactgtgcaagaga
23		351#46 ccctgtattactgtgcaagaga

24

5a#49 coagttattactgtggaga

F: HpyCH4III REadaptors, Extenders, and Bridges

F.1 REadaptors

! ONS for cleavage of HC(lower) in FR3(bases 77-97)

! For cleavage with HpyCH4III, Bst4CI, or TaaI

! cleavage is in lower chain before base 88.

	77	78	88	88	88	88	88	88	88	88	88	T_m^F	α_m^F
!	78	90	234	567	890	123	456	7					
(H43.77.97.1-02#1)	5'-cc	gtg	tat	tAC	TGT	gog	aga	g-3'		64		62.6	
(H43.77.97.1-03#2)	5'-c	gtg	tat	tAC	TGT	gog	aga	g-3'		52		60.6	
(H43.77.97.109#3)	5'-cc	gtg	tat	tAC	TGT	gog	aga	g-3'		64		62.6	
(H43.77.97.323#22)	5'-cc	gtg	tat	tac	tgt	gog	aga	g-3'		60		58.7	
(H43.77.97.330#23)	5'-c	gtg	tat	tac	tgt	gog	aga	g-3'		60		58.7	
(H43.77.97.439#44)	5'-c	gtg	tat	tac	tgt	gog	aga	g-3'		62		60.6	
(H43.77.97.551#48)	5'-cc	gtg	tat	tac	tgt	gog	aga	g-3'		62		60.6	
(H43.77.97.5a#49)	5'-cc	gtg	tat	tAC	TGT	gog	aga	g-3'		58		58.3	

F.2 Extender and Bridges

! XbaI and AflII sites in bridges are bunged

(H43.XABr1) 5'-gggtgtagtga-

|TCT|AGT|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

|aac|agC|TTT|AGG|gct|cag|gac|aCT|GCA|Gtc|tac|tat|tgt|gog|aga-3'

(H43.XABr2) 5'-gggtgtagtga-

|TCT|AGT|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

|aac|agC|TTT|AGG|gct|cag|gac|aCT|GCA|Gtc|tac|tat|tgt|gog|aaa-3'

(H43.XAExt) 5'-ATAgTAGAcT gcAgTgTccT cAgcccTAA gcTgTcAcT TgcAAgTAgA-

gAgTATTcTT AgAgTTgTcT cTAgATcAcT AcAcc-3'

!H43.XAExt is the reverse complement of

! 5'-gggtgtagtga-

! |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

! |aac|agC|TTT|AGG|gct|cag|gac|aCT|GCA|Gtc|tac|tat|-3'

(H43.XAPCR) 5'-gggtgtagtga |TCT|AGA|gac|aac-3'

! XbaI and AflII sites in bridges are bunged

(H43.ABr1) 5'-gggtgtagtga-

|aac|agC|TTT|AGG|gct|cag|gac|aCT|GCA|Gtc|tac|tat|tgt|gog|aga-3'

(H43.ABr2) 5'-gggtgtagtga-

|aac|agC|TTT|AGG|gct|cag|gac|aCT|GCA|Gtc|tac|tat|tgt|gog|aaa-3'

(H43.AExt) 5'-ATAgTAGAcTgcAgTgTccTcAgcccTAAgcTgTgTcAcTAcAcc-3'

!(H43.AEXT) is the reverse complement of 5'-gggtgtagtga-
! aaac|agC|TTA|AGc|gct|gag|gac|aCT|GCA|Gtc|tac|tat -3'
(H43.APCR) 5'-gggtgtagtga aaac|agC|TTA|AGc|gct|g-3'

Table 510

25 (E)KJact) 5'-spcAscgTg TtGTT cAsggAgtg-3'

(VH881) 5'-ANTAGAGAC TcAGTtTcc TcAgccctTA AgcTgTccT AgcTgTccT cTgcAAgAAG-
 AGAGTATtTc TAggTtTgc TcTAgAcTtTA gTgAAg-3',
 I note that VH881 is the reverse complement of the OR below
 I [RC] 5'-cgCttccAaag-
 I scab.....
 I Synthetic 3-23 as in Table 206
 I |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|tG|cag|atg|-
 I XbaI...
 I |aac|agc|TtA|AGg|gct|gag|gac|act|GCA|Gtc|tac|tat|t-3'
 I AllI...
 I (VH881) 5'-cgCttccAaag-
 I |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|tG|cag|atg|-
 I |aac|agc|TtA|AGg|gct|gag|gac|act|GCA|Gtc|tac|tat|tgt|gcg|ag-3'
 I (VH881) 5'-cgCttccAaag-

25

30

35

TCCTAGAGTgac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|
|aac|agc|TTA|AGg|ctc|gac|act|GCA|Gtc|Tact|tat|tgc|Acg ag-3'
(VR881PCR) 5'-cgcttccactaag|TCCTAGAGTgac|aac -3'


```

Sites to be varied-->      ***      ***      ***
-----FR1----->|...CDR1.....|-----FR2-----
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
A S G F T F S S Y A M S W V R
|ggt|TCC|GGA|ttc|act|ttc|tct|tCG|TAC|gcc|atg|tct|tgg|gtt|cgc| 143
|cga|agg|cct|aag|tga|aag|aga|agg|atg|cga|tac|aga|acc|caa|cgc|
|BspEI | | |BstXI.

          Sites to be varies--> ***      ***      ***
-----FR2----->|...CDR2.....|
61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
Q A P G K G L E W V S A I S G
|CAA|gct|ccT|GGT|aaa|ggt|ttg|cag|tgg|gtt|tct|gct|atc|tct|ggt| 188
|ggt|cga|gga|cca|ttt|cca|aac|ctc|acc|caa|aga|cga|tag|aga|cca|
...BstXI |

          ***      ***
...CDR2.....|-----FR3-----
76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
S G G S T Y Y A D S V K G R P
|tct|ggt|ggc|agt|act|tac|tat|gct|gac|tcc|gtt|aaa|ggt|cgc|tcc| 233
|aga|cca|ccg|cca|tga|atg|ata|cga|ctg|agg|caa|ttt|cca|gcg|aag|

-----FR3----->
91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
T I S R E N S K N T L Y L Q M
|act|acc|TCT|AGA|cac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg| 278
|tga|tag|aga|tct|ctg|ttg|aga|ttc|tta|tga|gag|atg|aac|ctc|tac|
|NbaI |

-----FR3----->|
106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
N S L R A E D T A V Y Y C A K
|aac|agC|TTA|AGg|gct|cag|cag|act|GCA|Gtc|tac|tat|cgc|gct|aaa| 323
|ttg|tcg|aat|tcc|cga|ctc|ctg|tga|cgt|cag|atg|ata|acc|cga|ttt|
|AflII | |PstI |

...CDR3.....|-----FR4----->
121 122 125 124 125 126 127 128 129 130 131 132 133 134 135
D Y E G T G Y A F D I W G Q G
|gac|tat|gaa|ggt|act|ggt|tat|ggt|ttc|gac|ATA|TCg|ggt|caa|ggt| 368
|ctg|ata|ctt|cca|tga|cca|ata|cga|aag|ctg|tat|acc|cca|ggt|cca|
|NdeI |

-----FR4----->|
136 137 138 139 140 141 142
T M V T V S S
|act|atG|GTC|ACC|gtc|tct|agt- 389
|tga|tac|cag|tgg|cag|aga|cca-
|BstEII |

          143 144 145 146 147 148 149 150 151 152
          A S T K G P S V F F
          gcc tcc acc aag GCC CCA tgg GTC TTC ccc-3' 419
          egg agg tga ttc ccc ggt agc gag aag cgg-5'
          Bsp120I. BbsI... (2/2)
          ApaI....

(SPPERMENT) 5'-ctg tct gaa cG GCC cag ccG-3'
(TOPFR1A) 5'-ctg tct gaa cG GCC cag ccG GCC atg gcc-
          gaa|gtt|CAA|TIG|tta|gag|tct|ggt|-
          |ggc|ggt|ctt|gtt|cag|cct|ggt|ggt|tct|tta-3'
(BOTFR1B) 3'-caa|gtc|gga|cca|cca|aga|aat|gca|gaa|aga|acg|cga|-
          |cga|agg|cct|aag|tga|aag-5' ! bottom strand
    
```

```

(BOTFR2:  3'-acc|caa|gog|-
          |gtt|cga|gga|cca|ttt|cca|aac|ctc|aac|caa|aga|-5' ! bottom strand
(BOTFR3:  3'- a|cga|ctg|agg|caa|ttt|cca|gog|aag|-
          |tga|tag|aga|tct|ctg|ttg|aga|ttc|tta|tga|gag|atg|aac|gtc|tcc|-
5 |ttg|tgg|aat|tcc|cga|ctc|ctg|tga-5'
(F06)    5'-gC|TTA|AGg|gct|gag|gac|ACT|CCA|Gtc|tac|tat|tgc|gct|aaa|-
          |gac|tat|gaa|ggt|act|ggt|tat|gct|ttc|gac|ATA|TCg|ggt|c-3'
(BOTFR4:  3'-cga|aag|ctg|tat|acc|cca|gtt|cca|-
          |tga|tac|cag|tgg|cag|aga|taa-
10 |agg|agg|tgg|ttc|cag|ggt|agc|cag|aag|ggg-5' ! bottom strand
(BOTFRCPRIM)  3'-gg|ttc|cag|ggt|agc|cag|aag|ggg-5'
|
| CDR1 diversity
15 |DN-vgC1)  5'-[gct|TCC|GGA|tcc|act|tcc|tct|<1>|TAC|<1>|atg|<1>]-
          |
          | CDR1 .....6859
          |tgg|ggt|cgc|CAa|gct|cct|GG-3'
|
| <1> stands for an equimolar mix of {ADEFGRHKLMNPQRSTVWY}; no C
20 | (this is not a sequence)
|
| CDR2 diversity
|
25 |DN-vgC2)  5'-ggg|tgg|gag|tgg|ggt|tct|<2>|abc|<2>|<3>|-
          | CDR2 .....
          |tct|ggt|ggc|<1>|act|<1>|tat|gct|gac|ccc|ggt|aaa|gg-3'
          | CDR2 .....
|
| <1> is an equimolar mixture of {ADEFGRHKLMNPQRSTVWY}; no C
| <2> is an equimolar mixture of {YRWVGS}; no ACDEFGRHKLMNPQT
30 | <3> is an equimolar mixture of {E}; no ACDEFGRHKLMNPQRSTVWY

```


Table 800 (new)

The following list of enzymes was taken from <http://rebase.neb.com/cgi-bin/asymmlist>.

I have removed the enzymes that a) cut within the recognition, b) cut on both sides of the recognition, or c) have fewer than 2 bases between recognition and closest cut site.

REBASE Enzymes
04/13/2001

Type II restriction enzymes with asymmetric recognition sequences:

Enzymes	Recognition Sequence	Isoschizomers	Suppliers
AarI	CACCTGCNNNN^NNNN	-	Y
AceIII	CAGCTCNNNNNNNN^NNNN	-	-
Bbr7I	GAAGACNNNNNNNN^NNNN	-	-
BbvI	GCAGCNNNNNNNN^NNNN	-	Y
BbvII	GAAGACNN^NNNN	-	-
Bce83I	CTTGAGNNNNNNNNNNNNNN^NN	-	-
BceAI	ACGGCNNNNNNNNNNNN^NN	-	Y
EcefI	ACGGCNNNNNNNNNNNN^N	-	-
EciVI	GTATCCNNNNK_N^	BfuI	Y
BfiI	ACTGGGNNNN_N^	BmrI	Y
BinI	GGATCNNNN^N	-	-
BscAI	GCATCNNNN^NN	-	-
BserI	GAGGACNNNNNNNN^NN	-	Y
BsmFI	GGGACNNNNNNNNNN^NNNN	BspLU11III	Y
BspMI	ACCTGCNNNN^NNNN	Acc36I	Y
EciI	GGCGGANNNNNNNNN^NN	-	Y
Eco57I	CTGAAGNNNNNNNNNNNN^NN	BspKT6I	Y
FauI	CCCGCNNNN^NN	BstFZ436I	Y
FokI	GGATGNNNNNNNNNN^NNNN	BstFZ418I	Y
GsuI	CTGGACNNNNNNNNNNNN^NN	-	Y
HqaI	GACGCHNNNN^NNNN	-	Y
HphI	GCTGANNNNNNN_N^	AsuHPI	Y
MboII	GAAGANNNNNN_N^	-	Y
MlyI	GAGTCNNNN^	SchI	Y
MneI	TCCRACNNNNNNNNNNNNNN^NN	-	-
MnlI	CCTCNNNNNN_N^	-	Y
PleI	GAGTCNNNN^N	PpsI	Y
RleAI	CCCACANNNNNNNNNN^NNN	-	-
SfaNI	GCATCNNNN^NNNN	BspST5I	Y
SspD5I	GGTGANNNNNNNK^	-	-
Sth132I	CCCGNNNN^NNNN	-	-
StsI	GGATGNNNNNNNNNN^NNNN	-	-
TaqII	GACCGANNNNNNNNNN^NN^, CACCCANNNNNNNNNN^NN^	-	-
Tth111III	CAARCANNNNNNNNN^NN^	-	-
UbaPI	CGAACC	-	-

The notation is ^ means cut the upper strand and _ means cut the lower strand. If the upper and lower strand are cut at the same place, then only ^ appears.

Table 120: MALLI3, annotated

! MALLI3 9532 bases

```

-----
1 aat get act act att agt aga att gat gcc acc ttt tca get cgc gcc
!   gene ii continued
49 cca aat gaa aat ata gct aaa cag gtt att gac cat ttg cga aat gta
97 tct aat ggt caa act aaa tct act cgt ccg cag aat cgg gaa tca act
145 gtt acc tgg aat gaa act tcc aga cac cgt act tta gtt gca tat tca
193 aaa cat gtt gag cta cag cac cag att cag caa tta agc tct aag cca
241 tcc gca aaa atg acc tct tat caa aag gag caa tta aag gta ctc tct
289 aat cct gac ctg ttg gag ttt gct tcc ggt cug gtt cgc ttt gaa get
337 cga att aaa acg cga tat ttg aag tct ttc ggg ctt cct ctt aat ctt
385 ttt get gca atc cgc ttt gct tct gac tat aat agt cag ggt aaa gac
433 ctg att ttt gat tta tgg tca ttc tcc ttt tct gaa cug ttt aaa gca
481 ttt gag ggg gat tca ATG aat att tat gac gat tcc gca gta ttg gac
!   RBS?..... Start gene X, ii continues
529 get atc cag tct aaa cat ttt act att acc ccc tct ggc aaa act tcc
577 ttt gca aaa gcc tct cgc tat ttt ggt ttt tat cgt cgt ctg gta aac
625 gag ggt tat gat agt gtt get ctt act atg cct cgt aat tcc ttt tgg
673 cgt tat gta tct gca tta gtt gaa tgt ggt att cct aaa tct caa ctg
721 atg aat ctt tct acc tgt aat aat gtt gtt ccg tta gtt cgt ttt aat
769 aac gta gat tct tct tcc caa cgt cct gac tgg tat aat gag cca gtt
817 ctt aaa atc gca TAA
!   End X & II
832 qgtaattca ca
!
!   M1           E5           Q10           T15
943 ATG att aaa gtt gaa att aaa cca tct caa gcc caa ttt act act cgt
!   Start gene Y
!
!   S17           S20           P25           E30
891 tct ggt gtt tct cgt cag gcc aag cct tat tca ctg aat gag cag ctt
!
!   V35           E40           V45
933 tgt tac gtt gat ttg ggt aat gaa tat ccg gtt ctt gtc aag att act
!
!   D50           A55           L60
987 ctt gat gaa ggt cag cca gcc tat gcc cct ggt ctg TAC Acc gtt cat
!   BsmGI...

```

```

!           L65           V70           S75           R80
1035 ctg tcc tct ttc aaa gtt ggt cag ttc ggt tcc ctt atg att gac cgt
!
!           P85       K87 end of V
1083 ctg cgc ctc gtt cag gct aag TAA C
!
1108 ATG gag cag gtc gcg gat ttc gac aca att tat cag gcg atg
!       Start gene VII
!
1150 ata caa atc tcc gtt gta ctt tgt ttc gcg ctt ggt ata atc
!
!           VII and IX overlap.
!           ..... S2  V3  L4  V5           S10
1192 gct ggg ggt caa agA TGA gt gtt tta gtg tat tct ttc gcc tct ttc gtt
!           End VII
!           |start IX
!           L13       W15           G20           T25           E29
1242 tta ggt tgg tgc ctt cgt agt gcc att aag tat ttt acc cgt tta atg gaa
!
1298 act tcc tc
!
!           ... stop of IX, IX and VIII overlap by four bases
1301 ACG aaa aag tct tta gtc ctc aaa gcc tct gta gcc gtt gct acc ctc
!       Start signal sequence of viii.
!
1349 gtt cag atg ctg tct ttc gct gct gag ggt gac gat ccc gca aaa gcg
!           mature VIII -->
1397 gcc ttt aac tcc ctg caa gcc tca gcg acc gaa tat atc ggt tat gcg
1445 tgg gcg aag gtt gti gtc att
1466 gtc gcc gca act atc ggt atc aag ctg ttt aag
1488 aaa ttc acc tgg aaa gca ! 1515
!           ..... -35 ..
!
1517       agc tga taaacogat acaattaag gctccttttg
!           ..... -10 ...
!
1552 gacccttttt tttGGAGAt ttt ! S.D. Underlined
!
!           <----- III signal sequence ----->

```

```

!           M K K L L F A I P L V
1575 caac GTG aaa aaa tta tta ttc gca att cct tta gtt ! 1611
!
!           V P F Y S E S A Q
1612 gtt cct ttc tat tct cac aGT gcA Cag tCT
!
!                                     ApaLI...
!
1642      GTC GTG ADE CAG CCG CCC TCA GTG TCT GGG GCC CCA GGG CAG
      AGG GTC ACC AUC TCC TGC ACT GGG AGC AGC TCC AAC ATC GGG CCA
!
!                                     EstEII...
1729      GGT TAT GAT GTA CAC TGG TAC CAG CAG CTT CCA GGA ACA GCC CCC AAA
1777      CTC CTC ATC TAT GGT AAC AGC AAT GGG CCC TCA GGG GTC CCT GAC CGA
1825      TTC TCT GGC TCC AAG TCT GGC ACC TCA GCC TCC CTG GCC ATC ACT
1870      GGG CTC CAG GCT GAG GAT GAG SCT GAT TAT
1900      TAC TGC CAG TCC TAT GAC AGC AGC CTG AGT
1930      GGC CIT TAT GTC TTC GGA ACT GGG ACC AAG GTC ACC GTC
!
!                                     BstEII...
1969      CTA GGT CAG CCC ABG GCC AAC CCC ACT GTC ACT
2002      CTG TTC CCG CCC TCC TCT GAG GAG CTC CAA GCC AAC AAG GCC ACA CTA
2050      GTG TGT CTG ATC AGT GAC TTC TAC CCG GGA GCT GTG ACA GTG CCC TGC
2098      AAG GCA GAT AGC AGC CCC GTC ABG GCG GGA GTG GAG ACC ACC ACA CCC
2146      TCC AAA CAA AGC AAC AAC AAG TAC GCG GCC AGC AGC TAT CTG AGC CTG
2194      ACC CCT GAG CAG TGG AAG TCC CAC AGA AGC TAC AGC TGC CAG GTC ACC
2242      CAT GAA GGG AGC ACC GTG GAG AAG ACA GTG GCC CCT ACA GAA TGT TCA
2290      TAA TAA ACCG CCTCCACCGG GCSCGCCAAT TCTATTTCAA GGAGACAGTC ATA
!
!                                     AscI.....
!
!           PstI signal----->
!           M K Y L L P T A A A G L L L L
2343      ATG AAA CAC CTA TTG CCT ACG GCA GCC GGT GGA TTG TTA TTA CTC
!
!           16 17 18 19 20      21 22
!           A A Q P A      M A
2386      gcG GCC cag ccG GCC.....ata gcc
!
!           SfiI.....
!           NcoMI... (1/2)
!           NcoI.....
!

```

```

!
!                                     FR1(DP47/V3-23)-----
!                                     23 24 25 26 27 28 29 30
!                                     E  V  Q  L  L  E  S  G
! 2409                               gaa|gtt|CAA|TTG|tta|gag|tct|ggt|
!                                     | MfeI |
!
! -----FR1-----
!      31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
!      G  G  L  V  Q  P  G  G  S  L  R  L  S  C  A
! 2433 |ggc|ggt|ctt|ggt|aac|ect|ggt|ggt|tct|tta|cgt|ctt|tct|tgc|ggt|
!
! -----FR1----->...CDR1.....|-----FR2-----
!      46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
!      A  S  G  F  T  F  S  S  Y  A  M  S  W  V  R
! 2479 |ggt|TCC|GGA|ttc|act|tcc|tct|tcg|TAC|Gct|atg|tct|tgg|ggt|cgc|
!      | BspEI |                | BsiWI |                | BstXI.
!
! -----FR2----->...CDR2.....
!      61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
!      Q  A  P  G  K  G  L  E  W  V  S  A  I  S  G
! 2523 |CAA|gct|ccT|GGt|aaa|ggt|ttg|gag|tgg|ggt|tct|gct|atc|tct|ggt|
!      ...BstXI
!
! .....CDR2.....|-----FR3-----
!      76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
!      S  G  G  S  T  Y  Y  A  D  S  V  K  G  F  F
! 2568 |tct|ggt|ggc|agt|act|tac|tat|gct|gac|tcc|gtt|aaa|ggt|cgc|tcc|
!
! -----FR3-----
!      91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
!      T  I  S  R  D  N  S  K  N  T  L  Y  L  Q  M
! 2613 |act|atc|TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|
!      | XbaI |
!
! -----FR3----->
!      106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
!      N  S  L  R  A  E  D  T  A  V  Y  Y  C  A  K
! 2658 |aac|agc|TAA|AGg|gct|gag|gac|act|GCA|Gtc|tac|tat|tgc|gct|aaa|

```

```

!           |AFLIII |           | PstI |
!
!           .....CDRS.....|----FR4-----
!           121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
!           D   Y   E   G   T   G   Y   A   E   D   E   W   G   Q   G
2703 |gac|tat|gaa|ggg|act|ggg|tat|gct|ttc|gac|ATA|TGg|ggg|caa|ggg.
!                                     | NdeI |(1/4)
!
!           -----FR4----->|
!           136 137 138 139 140 141 142
!           T   M   V   T   V   S   S
2748 |act|atG|GTC|ACC|gtc|tcc agt
!                                     | BstEII |
! From BstEII onwards, pV323 is same as pCES1, except as noted.
! BstEII sites may occur in light chains; not likely to be unique in final
! vector.
!
!           143 144 145 146 147 148 149 150 151 152
!           A   S   T   K   G   P   S   V   F   P
2769 |gac tcc acc aaG GGC Cca tgg GTC TTC ccc
!                                     BspI20I.      EbsI...(2/2)
!                                     ApaI...
!
!           153 154 155 156 157 158 159 160 161 162 163 164 165 166 167
!           L   A   P   S   S   K   S   T   S   G   E   T   A   A   L
2799 |ctg gca ccC TCC TCC aag agc acc tct ggg ggc aca gag gcc ctg
!                                     BseRI...(2/2)
!
!           168 169 170 171 172 173 174 175 176 177 178 179 180 181 182
!           G   C   L   V   K   D   Y   F   P   E   P   V   T   V   S
2844 |ggc tgc ctg GTC AAG GAC TAC TTC CCC gaA CCG GTg acg gtg tgg
!                                     AgeI....
!
!           183 184 185 186 187 188 189 190 191 192 193 194 195 196 197
!           W   N   S   S   A   L   T   S   G   V   H   T   F   P   A
2889 |tgg aac tca GGC GCC ctg acc agc ggc gtc cac acc ttc cgg gct
!                                     KasI...(1/4)
!
!           198 199 200 201 202 203 204 205 206 207 208 209 210 211 212

```

```

!       V L Q S S G L Y S L S S V V T
2934  gtc cta cag tCt agc GGa ctc tac tcc ctc agc agc gta gtg acc
!           (Bsu36I...) (knocked out)
!
!       213 214 215 216 217 218 219 220 221 222 223 224 225 226 227
!       V P S S S L G T Q P Y I C N V
2979  gtg ccC tCt tct agc tTC Ggc acc cag acc tac atc tgc aac gtg
!           (BstXI.....)N.B. destruction of BstXI & BpmI sites.
!
!       228 229 230 231 232 233 234 235 236 237 238 239 240 241 242
!       N H K P S N L K V D K K V E P
3024  aat cac aag ccc agc aac acc aag gtg gac aag aaa gtt gag ccc
!
!       243 244 245
!       K S C A A A H H H H H H S A
3069  aaa tct tgt GCG GCC GCT cat cac cac cat cat cac tct got
!           NotI.....
!
!       E Q K L I S E E D L N G A A
3111  gaa caa aaa ctc atc tca gaa gag gat ctg aat ggt gcc gca
!
!
!       D I N D D R M A S G A
3153  GAT ATC aac gat gat dgt atg gct AGC gcc gcc
!           rEK cleavage site..... NheI... KaeI...
!           EcoRV..
!
! Domain I -----
!       A E T V E S C L A
3183  gct gaa act gtt gaa agt tgt tta gca
!
!
!       K P H T E I S F
3210  aaa ccc cat aca gaa aat tca ttc
!
!       T N V W K D D K T
3234  aCT AAC GTC TGG AAA GAC GAC AAA ACT
!
!       L D R Y A N Y E G C D W N A T G V

```



```

3261 vta gat cgt tac ggt aac tat gag ggt tgt ctg tgc AAT GCT aca ggc gtc
!
!                               BsmI_____
!
!   V   V   C   T   E   D   E   T   Q   C   Y   G   T   W   V   P   I
3312 gta gtt tgt act ggc GAC GAA ACT CAG TGT TAC GGT ACA TGG GTT cct att
!
!   G   L   A   I   F   E   N
3363 ggg ctt gct atc cct gaa aat
!
! L1 linker -----
!   E   G   G   G   S   E   G   G   G   S
3384 gag ggt ggt ggc tct gag ggt ggc ggt tct
!
!   E   G   G   G   S   E   S   G   G   T
3414 gag ggt ggc ggt tct gag ggt ggc ggt act
!
! Domain 2 -----
3444 aaa cct cct gag tac ggt gat aca cct att ccg ggc tat act tat atc aac
3496 cct ctc gac ggc act tat ccg cct ggt act gag caa aac ccc gcl aat ccc
3546 aat cct tct ctt GAG GAG tct cag cct cct aat act ttc atg ttt cag aat
!
!                               BseRI_____
!
3597 aac agg ttc cga aat agg cag ggc gaa tca act gtt tat acg ggc act
3645 gtt act caa ggc act gac ccc gtt aaa act tat tac cag tac act cct
3693 gta tca tca aaa ggc atg tat gac gct tac tgy aac ggt aaa ttc AGA
!
!                               AlwNI
!
3741 GAC TGC gct ttc cat tct ggc ttt aat gaa gat cca ttc gtt tgt gaa
!
!                               AlwNI
!
3789 tat caa ggc caa tcg tct gac ctg cct caa cct cct gtc aac gct
!
3834 ggc ggc ggc tct
! start L2 -----
3846 ggt ggt ggt tct
3856 ggt ggc ggc tct
3870 gag ggt ggt ggc tct gag ggt ggc ggt tct
3900 gag ggt ggc ggc tct gag ggc ggc ggt tcc
3930 ggt ggt ggc tct ggt ! end L2
!
! Domain 3 -----
!   S   G   D   F   D   Y   E   K   M   A   N   A   N   K   G   A

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3945 tcc ggt gat ttt gat tat gaa aag atg gca aac gct aat aag ggg ggt
!
!   M T E R A D E N A L Q S D A K G
3993 atg acc gaa aat gcc gat gaa aac gcg cta cag tct gac gct aaa ggc
!
!   F L D S V A T D Y G A A I D G F
4041 aaa cct gat tct gtc gct aat gat tac ggt gct gct atc gat ggt ttc
!
!   I G D V S G L A K G N G A T G D
4089 att ggt gac gtc tcc ggc ctt gct aat ggt aat ggt gct aat ggt gat
!
!   F A G S N S Q M A Q V G D G D N
4137 ttt gct ggc tct aat tcc caa atg gct caa gtc ggt gac ggt gat aat
!
!   S P L M N N F R Q Y I P S L P Q
4185 tca cct tra atg aat aat ttc cgt caa tat tta cct tcc ctc cct caa
!
!   S V E C R P F V F S A G K P Y E
4233 tcc gtt gaa tgt cgc cct ttt gtc ttc agc gct ggt aaa cca tat gaa
!
!   F S I D C D K I N I F R
4281 ctt tct att gat tgt gac aaa ata aac tta ttc cgt
!
!                                     End Domain B
!
!   G V F A F I L Y V A T F M Y V F140
4327 ggc gcc ttt ggc ttt ctt tta tat gtt gcc acc ttt atg tat gta ttt
!   start transmembrane segment
!
!   S T F A N I L
4365 cct acg ttt gct aac ata atg
!
!   R N K E S
4386 cgt aat aag gag tct TAA ! stop of iii
!   Intracellular anchor.
!
!   M L P 2 V L L 5 G I P R L I C L R F L G 15
4404 tc ATG cca gtt ctt ttg ggt att ccg tta tta ttg cgt ttc ctc ggt
!   Start VI
!
!

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4451 ttc att ctg gta act ttg ttc ggc tat ctg ctg act ttt ctt aaa aag
4499 ggc ttc ggt aag ata gcc att gcc att tca ttg ttc ctt gcc ctt att
4547 att ggg ctt aac tca att ctg gtg ggt tat ctg tct gat att agc gcc
4595 caa tca ccc tct gac ttt gtt cag ggt gtt cag tta att ctg ccg tct
4643 aat ggc ctt ccc tgt ttt tat gtt att ctg tct gta aag gcc gcc att
4691 ttc att ttc gac gtt aaa caa aaa atc gtt tct tat ttg gat tgg gac
!
!
!           M1  A2  V3      P5           L10      G13
4739 aaa TAA t ATG gcc gtt tat ttt gta act ggc aaa tta ggc tct ggc
!
!   end VI   Start gene I
!
!
!   14  15  16  17  18  19  20  21  22  23  24  25  26  27  28
!   K   F   L   V   S   V   G   K   I   Q   D   K   I   V   A
4785 aag acg ctg gtt agc gtt ggt aag att cag gat aaa att gta gcc
!
!
!   29  30  31  32  33  34  35  36  37  38  39  40  41  42  43
!   G   C   K   I   A   T   N   L   D   L   R   L   Q   N   L
4830 ggg tgc aaa ata gca act aat ctt gat tta agg ctt caa aac ctg
!
!
!   44  45  46  47  48  49  50  51  52  53  54  55  56  57  58
!   F   Q   V   G   R   F   A   K   T   F   R   V   L   R   I
4875 ccg caa gtc ggg agg ttc gcc aaa agc act cgc gtt ctt aga ata
!
!
!   59  60  61  62  63  64  65  66  67  68  69  70  71  72  73
!   F   D   K   F   S   I   S   D   L   L   A   I   G   R   G
4920 ccg gat aag act tct ata tct gat ttg ctt gcc att ggc cgc ggt
!
!
!   74  75  76  77  78  79  80  81  82  83  84  85  86  87  88
!   N   D   S   Y   D   E   N   K   N   G   L   L   V   L   D
4965 aat gat tcc tac gat gaa aat aaa aac ggc ttg ctt gtt ctg gat
!
!
!   89  90  91  92  93  94  95  96  97  98  99 100 101 102 103
!   E   C   G   T   W   F   N   T   R   S   W   N   D   K   E
5010 gag tgc ggt act tgg ttt aat acc cgt tct tgg aat gat aag gaa
!
!
!   104 105 106 107 108 109 110 111 112 113 114 115 116 117 118
!   R   Q   P   I   I   D   W   F   L   H   A   R   K   L   G
5055 aga cag ccg att att gat tgg ttt cta cat gcc cgt aaa tta gga
!
!
!

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!      118 120 121 122 123 124 125 126 127 128 129 130 131 132 133
!      W  D  I  I  F  L  V  Q  D  L  S  I  V  D  K
5100 tgg gat att att ttt ctt gtt cag gac tta tct att gtt gat aaa
!
!      134 135 136 137 138 139 140 141 142 143 144 145 146 147 148
!      Q  A  R  S  A  L  A  E  H  V  V  Y  C  R  R
5145 cag gag cgt tct gca tta gct gaa cat gtt gth tct tgt cgt cgt
!
!      149 150 151 152 153 154 155 156 157 158 159 160 161 162 163
!      L  D  R  I  T  L  F  F  V  G  T  L  Y  S  L
5190 ctg gac aga att act tta cct ttt gtc ggt act tta tct tct ctt
!
!      164 165 166 167 168 169 170 171 172 173 174 175 176 177 178
!      I  T  G  S  K  M  P  L  P  K  L  H  V  G  V
5235 att act ggc tgg aaa atg cct ctg cct aaa tta cat gth ggc gtt
!
!      179 180 181 182 183 184 185 186 187 188 189 190 191 192 193
!      V  K  Y  G  D  S  Q  L  S  F  T  V  E  R  W
5280 gtt aaa tat ggc gat tct caa tta agc cct act gth gag cgt tgg
!
!      194 195 196 197 198 199 200 201 202 203 204 205 206 207 208
!      L  Y  T  G  K  N  L  Y  N  A  Y  D  T  K  Q
5325 ctt tat act ggt aag aat ttg tat aac gca tat gat act aaa cag
!
!      209 210 211 212 213 214 215 216 217 218 219 220 221 222 223
!      A  P  S  S  N  Y  D  S  G  V  Y  S  Y  L  T
5370 gct ttt tct agt aat tat gat tcc ggt gth tat tct tat tta aag
!
!      224 225 226 227 228 229 230 231 232 233 234 235 236 237 238
!      P  Y  L  S  H  G  R  Y  F  K  P  L  N  L  G
5415 cct tat tta tca cac ggt cgg tat ttc aaa cca tta aat tta ggt
!
!      239 240 241 242 243 244 245 246 247 248 249 250 251 252 253
!      Q  K  M  K  L  T  K  I  Y  L  K  K  F  S  R
5460 cag aag atg aaa tta act aac ata cat ttg aaa aag ttt tcc cgc
!
!      254 255 256 257 258 259 260 261 262 263 264 265 266 267 268
!      V  L  C  L  A  I  G  F  A  S  A  F  T  Y  S
5505 gtc ctt tgt ctt gag att gga ttt gca tca gca ttt aca tat agt

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!
!   269 270 271 272 273 274 275 276 277 278 279 280 281 282 283
!   Y I T Q P K P E V K K V V S Q
5550 tat ata acc caa cct aag cag gag gtt aaa aag gta gtc tcc cag
!
!   284 285 286 287 288 289 290 291 292 293 294 295 296 297 298
!   T Y D F D R F T I D S S Q S L
5595 acc tat gat ttt gat aaa ttc acc att gac tct tct cag cgt ctt
!
!   299 300 301 302 303 304 305 306 307 308 309 310 311 312 313
!   N L S Y R Y V F K D S K G K L
5640 aat cta agc tat cgc tat gtt ttc aag gat tct aag gga aaa TTA
!
!                                     PacI
!
!   314 315 316 317 318 319 320 321 322 323 324 325 326 327 328
!   I N S E D L Q K Q G Y S L T Y
5685 ATT AAT agc gac gat tta cag aag caa ggt tat tca ctc aca tat
!
!   PacI
!
!   329 330 331 332 333 334 335 336 337 338 339 340 341 342 343
!   i I D L C T V S I K K G N S N E
!   iv                                     M1 K
5730 att gat cta tgt act gtt tcc att aaa aaa ggt aat tca aAT Gaa
!
!                                     Start IV
!
!   344 345 346 347 348 349
!   I I V K C M .End of T
!   iv L3 L N5 V I7 N F.V10
5775 att gtt aaa tgt aat TAA T TTT GTT
! IV continued....
5800 ttc ttg atg ttt gtt tca tca tct tct ttt gct cag gta att gaa atg
5840 aat aat tgg cct ctg cgc gat tct gta act tgg tat tca aag caa tca
5896 ggc gaa tcc gtt att gtt tct ccc gat gta aaa ggt act gtt act gta
5944 tat tca tct gac gtt aaa cct gaa aat cta cgc aat ttc ttt att tct
5992 gtt tta cgt gct aat aat ttt gat atg gtt ggt tca att cct tcc aca
6040 att cag aag tat aat cca aac aat cag gat tat att gat gaa ttg cca
6080 tca tct gar aat cag gaa tat gat gat aat tcc gct cct tct ggt ggt
6136 ttc ttt gtt cag caa aat gat aat gtt act caa act ttt aaa att aat
6184 aac gtt cgg gca aag gat tta ata cga gtt gtc gaa ttg ttt gta aag

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6232  tct aat act tct aaa tcc tca aat gta tta tct att gac ggc tct aat
6280  cta tta gtt gtt TCT gca cct aaa gat att tta gat aac ctt cct caa
!
      ApaLI removed
6328  ttc ctz tct act gtt gat ttg cca act gac cag ata ttg att gag cgt
6376  ttg ata ttt gag gtt cag caa ggt gat gct tta gat ttt tca ttt gct
6424  gct ggc tct cag cgt ggc act gtt gca ggc ggt gtt aat act gac cgc
6472  ctc aac tct gtt tta tct tct gct ggt ggt tct ttc ggt att ttt aat
6520  ggc gat gtt tta ggc cta tca gtt cgc gca tta aag act aat agc cat
6568  tca aaa ata ttg tct ggc cca cgt att ctt acg ctt tca ggt cag aag
6616  ggt tct atc tct gTT GGC CAG aat gtc cct ttt att act ggt cgt gtg
!
      MsoI _____
6664  act ggt gaa tct gcc aat gta aat aat cca ttt cag acg att gag cgt
6712  caa aat gta ggt att tcc atg agc gtt ttt cct gtt gca atg gct ggc
6760  ggt aat att gtt ctg gat att acc agc aag gcc gat agt ttg agt tct
6808  tct act cag gca agt gat gtt att act aat caa aga agt att gct aca
6856  acg gtt aat ttg cgt gat gga cag act ctt tta ctc ggt ggc ctc act
6904  gat tat aaa aac act tct caa gat tct ggc gta cag ttc ctg tct aaa
6952  ctc cct tta atc ggc ctc ctg ttt agc tcc cgc tct gat tcc aac gag
7000  gaa ago acg tta tac ggc ctc gtc aaa gca acc ata gta cgc gcc ctg
7048  TAG cgggcatt
!
      End IV
7060  aagcggggcg ggtgtgggtg ttaegcgcg cgtgaccgct acacttgcca ggcacctaga
7120  gcccgctcct ttgcctttct tcccttcct tctcgccacg ttcGCCGcct ttcacctca
!
      NgoMI _____
7180  agctctaaat cggggggtcc ctttagggtt cagatttagt gotttaoggo acctcgacc
7240  caaaaaactt yattgggtg atcgtCACG TAGTGggcca tgcacctgat agcgggtttt
!
      DraIII _____
7300  tgcacctttG ACGTGGAGT Ccaagttctt taatagtgga ctcttgtrcc aaactggaac
!
      DndI _____
7360  aacactcaac cctatctcgg gctattcttt tgatttataa gggattttgc agatttcgga
7420  accaccatca aacaggattt tgcctcgtg gggcaaacca gcgtggaccg ctgctgcaa
7480  ctctctcagg gccaggcggg gaaggccaat CAGCTGttgc cCGTCTCact ggtgaaaaga
!
      PvuII.      BclI.
7540  aaaaaccacc cGGATCC AAGCTT
!
      BamBI      HindIII (1/2)
!
      Insert carrying bla gene
7563  gcaggtg gcacttttgc gggaaagtg cggggaacc
7600  ctattcgttt atttctctaa atacattcaa atatGTATCC gacctatgaga caataacct
!
      BclVI

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MISSING AT THE TIME OF PUBLICATION

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8790 CCTCAGG
!      Bsu36I_
8797      cccgat actgtctgtccg tcccccaaaa ctggccagatg
8832 caaggttacg atgcgcaccat ctacaccaac gtaacctatc ccattacggc caatccgcgc
8892 ttggttccca cggagaatcc gaagggttgc tactcgcctca cattaatgtc tyatgaaagc
8952 tggctacagg aaggccagac gogaattatt ttgatggcg ttccatattgg ttaaaaaatg
9012 agcngattca acaaaaattt aacgcgaatt ttaacaaaat attaacgctt acaATTAA
!
!                               SwaI...
9072 Tattcgttta tacaatcttc ctgcttttgg ggctttttctg attatcaacc GGGGTACac
!
!                               RBS?
9131 ATG att gac atg cta gtt tta cga tta ccg ttc atc gat tct cct gtt tgc
!      Start gene II
9182 tcc aqa ctc tca ggc aat gac ctg ata gcc ttt gtA GAT Ctc tca aaa ata
!
!                               BglII...
9233 gct acc ctc tcc ggc atg aat tta tca gct aga acg gtt gaa tat cat att
9284 gat ggt gat ctg act gtc tcc ggc ctt tct cac cct ttc gaa tct tta cct
9335 aca cat tac tca ggc atk gca ttt aaa ata tat gag ggt tct aaa aat ttt
9386 tat cat tgc gtt gaa ata aag gct tct ccc gca aaa gta cta cag ggt cat
9437 aat gtt ttt ggt aca acc gat tta gct tta tgc tct gag gct tta ttg ctt
9488 aat ttt gct aat tct ttg cct tgc ctg tar gat tta ttg gat gtt ! 9532
! gene II continues

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Table 129B: Sequence of MALIA3, condensed

LOCUS	MALIA3	9532	CIRCULAR			
ORIGIN						
1	AATGCTACTA	CTATTAGTAG	AATTGMPGCC	ACCTTETCAG	CTCGCGCCCC	AAATGAAAAT
61	ATAGCTAAAC	AGGTTATTGA	CCATTTCGCA	AATGATCTCA	ATGGTCAAAC	TAANTCTACT
121	CGTTCCGAGA	ATTGGGAATC	AACTGTTACA	TGGAATGAAA	CTTCCAGACA	CCCTACTTTA
181	GTTGCATATT	TAAAACATGT	TGAGCTACAG	CACCAGATTC	AGCAATTAAG	CTCTAAGCCA
241	TCGGCAAAA	TGACCTCTTA	TCAAAAGGAG	CAATTAAGG	TACTCTCTAA	TCCTGACCTG
301	TTGGASTTTG	CTTCCGGTCT	GGTTCGCTTT	GARGCTCGAA	TTAAAAGCGG	AAATTTGAAG
361	TCTTTCGGGC	TTCTCTCTAA	TCTTTTGGAD	GCAATCCGCT	TGCTCTCTCA	CTAATAATCT
421	CAGGGTAAAG	ACCTGATTTT	TGATTTATGG	TCATTCCTGT	TTTCTGAACT	GTTTAAAGCA
481	TTTGAGGGGG	ATTCAATGAA	TATTTATGAC	GATTCCCGAG	TATTGGACGC	TATCCAGTCT
541	AAACATTTTA	CTATTACCCC	CTCTGGCAAA	ACTTCTTTTG	CAAAAGCCTC	TCGCTATTTT
601	GGTTTTTATC	GTCTCTGGT	AAACGAGGGT	TACGATAGTG	TGCTCTCTAC	TATGCTCTGT
661	AATTCCTTTT	GGGCTAATGT	ATCTGCATTA	GTGATATGTG	GTATTCCTAA	ATCTCAACTG
721	ATGATCTCTT	CEACCTGTAA	TAATGTTGTT	CCCTTASTTC	GTTTTATTA	CTAGATTTT
781	TCTTCCCAAC	GTCTGACTG	GTATAATGAG	CCACTTCTTA	AAATCCGATA	AGGTAATPCA
841	CAATGATTA	AGTTGAAAT	AAACCATCTC	AAGCCCAAT	TACTACTCGT	TCGGGTGTTT
901	CTCCTCAGGG	CAAGCCTTAT	TCACTGAATG	AGCAGCTTTC	TTAGGTTGAT	TTGGCTAATG
961	AATATCCGCT	TCTTGTCAAG	ATTACTCTTG	ATGAGGTC	GCCAGCCTAT	GCCTCTGCTC
1021	TGTACACCCT	TCATCTGTCC	TCTTCTAAG	TTGGTCAGTT	CGGTTCCCTT	ATGATGACC
1081	GTCTCCGCTT	CGTCCGGCT	AAGTAACATG	GAGCAGGTCG	CGGATTTCTA	CACATTTTAT
1141	CAGGCGATGA	TACAAATCTC	CGTGTACTT	TGTTTCGGCC	TTGGTATAAT	CGCTGGGGGT
1201	CAAAGATGAG	TCTTTTATGT	IATTTCTTCC	CCTCTTTCTG	TTTAGGTTGC	TGCTCTCTTA
1261	GTGGCATTAC	GTATTTTACC	CGTTEAATGG	AAACTTCTTC	ATGAAAAAGT	CTTAACTCTT
1321	CAAAGCCTCT	GTAGCCTTGG	CTACCCTCTG	TCCGATCTTG	TCTTTCGCTG	CTGAGGGTGA
1381	CGATCCCGCA	AAAGCGGCTT	TTAACTCCCT	GCAAGCCTCA	GCGACCGAAT	ATATCGGTTA
1441	TCCCTGGGCG	ATGGTTCTTG	TCATTGTCTG	CGCAACTATC	GATATCAAGC	TGTTTAAGAA
1501	ATTCACTCTG	AAAGCAAGCT	GATAAACCCT	TACAATTAAT	GGCTCCTTTT	GGAGCCTTTT
1561	TTTTTGGAGA	TTTTCAAGCT	GAAAAAATA	TTATTCGCA	TTCTTTTATG	TGTTCTTTTC
1621	TATTCCTACA	GTGCACACTC	TGTCTGAGG	CAGCCGCCCT	CAGTCTCTTG	GGCCCCAGGG
1681	CAGAGGGTCA	CCATCTCTCT	CAGTGGGAGC	AGCTCCAACT	TGGGGGCGAG	TTATGATGTA
1741	CACTGGTACC	AGCAGCTTCC	AGCAACAGCC	CCCAACTCTC	TCATCTATGG	TACACGATAT
1801	CGGCCCTCAG	GGGTCCCTGA	CCGATCTCTT	GGTCCAGAT	CTGGCACCCT	AGCCTCCTCT
1861	GCCATCACTG	GGCTCCAGGC	TGAGGATGAG	GCTGATTAAT	ACTGCGAGTC	CTATGACAGC
1921	AGCCTGASTG	GCCTTTATGT	CTTCGGAAT	GGGACCAAGG	TCACCGTCTT	AGGTCAGCCC
1981	AAGGCCAACC	CCACTGCTAC	TCTGTTCCCG	CCCTCTCTTG	AGGAGCTCCA	AGCCACAAAG
2041	GCCACACTAG	TGTGTCTGAT	CAGTCACTTC	TACCCGGGAG	CTGTGACAGT	GGCCTGGGAG
2101	GCAGATAGCA	GCCCCCTCAA	GCCGGCACTG	SASACCACCA	CACCCCTCCA	ACAAASCAAC

2161 AACAAATACC CGCCACGACG CTAATCTGAGC CTGACGCGTG AGCACTGGAA GTCCACACGA
2221 AGCTACAGGT GCCAGGTCAC GCATGAAGGG AGCACCCTGG AGAAGACAGT GCGCCCTACA
2281 GAATGTTCAE AATAAACCGC CTCCACCGGG CGCGCCAAAT CTAATTCAGG GAGACAGTCA
2341 TAATGAAATA CCTATTCGCT ACGGCAGCG CTGGATTETT AFIACTCGGG GCCCAGCGGG
2401 CCATGGCCGA AGTTCARTG TTAGASTCTG GTGGCGGTCT TCTTCAGCCT GGTGGTTCTT
2461 TACGTCTTTC TTGCGCTGCT TCCGGATCA CTTTCTCTTC GTACGCTATG TCTTGGGTTG
2521 GCCAAGCTCC TGGTAARGGT TTGGAGTGGG TTTCCTGCTAT CTCTGGTCTT GGTGGCAGTA
2581 CTTACTATGC TGACTCCGTT AAAGGTGCGT TCACTATCTC TAGAGACAA CTAAGAATA
2641 CTCTCTACTT GCAGATGAAC AGCTTAAGGG CTGAGGACAC TGCAGTCTAC CATTCGGCTA
2701 AAGACTATGA AGTACTGGT TATGCTTTCG ACAAATGSGG TCAAGTACT ATGGTCACCG
2761 TCCCTAGTGC CTCCACCAAG GGCCTATCGG TCTTCCGCTT GGCACCCCTC TCCAAGAACA
2821 CCTCTGGGGG CACAGCGGGC CTGGGCTGCC TGGTCAAGGA CTACTTCCCC GAACCGSTGA
2881 CGGTCTCCTG GAACTCAGGC GCCCTGACCA GCGGCGTCCA CAOCTTCCCG GCTGTCCCTAC
2941 AGTCTAGCGG ACTCTACTCC CTCAGCAGCG TAGTGCAGCT GCGCTCTTCT AGCTTGGGCA
3001 CCCAGACCTA CATCTGCAAC GTGATTCACA AGCCACGCAA CACCAGGTC GACAAGAAAG
3061 TTGAGCCCAA ATCTTGTGGG GCGGCTCATC ACCACCATCA TCACITGCTT GACAAAAC
3121 TCTCTCAGA AGAGGATCTG AATGGTGGCG CAGATATCAA CCAATGCTGT ATGGCTGGCG
3181 CCGCTGAAC TGTTSAAAGT TGTPTAGCAA AACCCATAC AGAATATTC TTTACTAACG
3241 TCTGAAAGA CGACAAAAC TTAGATCGTT ACGCTAACTA TGAGGTTGT CTGTGGAATG
3301 CTAGAGCGT TGTAGTTTGT ACTGGTGACG AAACGAGTG TTACGGTACA TGGGTTCTTA
3361 TTGGGCTTGC TATCCCTGAA AATGAGGGTG GTGGCTCTGA GGGTGGCGGT TCTGAGGTTG
3421 GCGGTTCTGA GGGTGGCGGT ACTAAACCTC CTGASTAGCG TCATACACCT ATTCGGGCT
3481 ATACTTATAE CAACCCTCTC GACGGCACCT ATCCGCTTGG TACTGAGCAA AACCCGCTA
3541 ATCTTAATCC TTCTCTGAG GACTCTCAGC CTCTTAATAC TTTCTCTTTT CAGAAATAA
3601 GGTTCGAAA TAGGCAAGGG SCATTAAC TGTTATAGGG CACTGTTACT CAGGGCACTG
3661 ACCCGCTEAA AACTTATTAC CAGTACACTC CTGTATCATC AAAAGCCATG TATGAGGCTT
3721 ACTGSAACGG TAATTCAGA GACTCGGCTT TCCATCTTGG CTTTAAATGA GATCCATTCG
3781 TTTSTGAATA TCAAGGCCAA TCGTCTGACC TGGCTCAACC TCCCTCAAT GCTGGCGGGG
3841 GCTCTGGTGG TGGTCTTGGT GCGGGCTCTG AGGGTGGTGG CTCTGAGGGT GCGGGTCTG
3901 AGGGTGGGG CTCTGAGGGA GCGGTTCCG GTGGTGGCTC TGGTTCGGGT GATCTTGATT
3961 ATGAAAGAT GCCAAAGGCT AATAAGGGGG CTATGACCGA AAATGCCGAT GAAAACGGGC
4021 TACAGTCTGA CGCTAAGGC AACTTGATT CTCTCCCTAC TCAATACGGT GCTGCTATCG
4081 ATGCTTTCAT TGGTGACGTT TCCGGCCTTG CTAATGGHAA TGGTCTACT GCTGATTTTG
4141 CTGGCTCTAA TTCCCAAATG GCTCAAGTCC GTGACGGTGA TAATTCACCT TCAATGAATA
4201 AATTCGGTCA ATATTACCT TCCCTCCCTC AATCGGTTGA ATGTGGGCTT TTTGCTPTTA
4261 GCGGTGGTAA ACCAATGAA TTTCTATTG ATGTGACAA AATAACTTA TCCGGTGGTG
4321 TCTTTCGCTT TCTTCTATAT GTTCCACCT TCAATGATGT ATTTCTACG TTTGCTAACA
4381 TACTGGGTAA TAAGGACTCT TAATCATGCC AGTCTTTTG GGTATCCGT TATTATTCGG
4441 TTTCCCTCGST TTTCTCTG TAACPTTGT CCGCTATCTG CTTACTTTTC TTAARAGGG

4501 CTTCGGTAAG ATAGCATATG CTATTTCMT GTTTCCTGCT CTATTACTG GGCTTAACTC
4561 AATTCCTTGGG GGTATATCTCT CTGATATTAG CGCTCAATTA CCCTCTGACT TTGTTGAGGG
4621 TSTTCAGCTTA ATTCTCCCGT CTAATGCGCT TCCCTGTTTT TATGTIATTC TCTCTGTAAA
4681 GGCTGCTACT TTCATTPTTG ACGTTAAACA AAAAATCGTT TCTTATTGGG ATTGGGATAA
4741 ATAATATGGC TGTTIATPTT GTAAGTGGCA AATTAGGCTC TGGAAAGACG CTCGTTAGCG
4801 TTGGTAAGAT TCAGGATAAA ATTETAGCTG GGTGCAAAA ASCPACTAAT CTTGATTTAA
4861 GGCTTCAAAA CCTCCCGCAA GTCCGGAGGT TCGCTAAAAC GCCTCCCGTT CTTAGAAATC
4921 CGGATAAGCC TTCTATATCT GATTTGCTTG CTATTGGCGG CGGTAATGAT TCCTACGATG
4981 AAAATAAAAA CCGCTTCTCT GTTCTCGATG ACPGCGCTAC TCGGTTTAAI ACCCGTCTCT
5041 GGAATGATAA GGAAGACAG CCGATTATPG ATTGGTTTCT ACATCCCTCT AAATCAGGAT
5101 GGGATATTAT TTTCTTGTG CAGGACTTAT CTATTGTGA TAAACAGGGC CGTCTCGCAT
5161 TAGCTGAACA TGTPTTTTAT TGTCTGCTC TGGACAGAAI TACTTTACCT TTGCTCGGTA
5221 CTTTATATTC TCTIATTAAT GGCCTGAAAA TGCCCTCTGC TAAATACAT GTTGGCGTTG
5281 TTAATAATGG CGATTCTCAA TTAAGCCCTA CTGCTGACCG TTGGCTTTAT ACNGGTAAGA
5341 RITTGATAAA CGCAATAGAT ACTAAACAGG CTTTTTCTAG TAATATAGAT TCCGGTGTTC
5401 ATTCTTATTT AACGCCCTAT TTATCACAGG GTCCGTATTT CAAACCPATA AATTTAGGTC
5461 AGAAGATGAA ATTAACATAA ATATATTTGA AAAAGTTTT CCGCGTCTCT TGTCTGCGA
5521 TTGGATTTGG ATCAGCATTT ACATATAGTT ATATAACCCA ACCTAAGCCG GAGCTTAAAA
5581 AGGTAGTCTC TCAGACCTAT GATTTTGATA AATTCACTAT TGACTCTTCT CAGCGTCTIA
5641 ATCTAAGCTA TCGCTATGTT TTCAGGATTT CTAAGGGAAA ATTAATTAAT AGCGAGGAT
5701 TACAGAAGCA AGGTCTATCA CTCACAGATA TTGATTTATG TACTGTTTCC APTAAAAAAG
5761 GTAATTCAAA TGAATTTGTT AATGTAATTT AATTTGTTTT TCTGATGTT TGTTCATCA
5821 TCTTCTTTTG CTCAGGTAAT TGAATGAAI AATTCGCTC TCGCGATTT TGTAACTTGG
5881 TATTCAAAAG AATCAGGGCA ATCCGTTAAT GTTCTCTCCG ATGTAAAAGG TACTCTTACT
5941 CTATATTCAT CTGACGTTAA ACCTGAAAAT CTACGCAATP TCTTATTTTC TGTTTTACGT
6001 GCTAATAAAT TPGATAUGGT TGGTTCAATP COTTCCATAA TTCAGAGTA TAATCCAAAC
6061 AATCAGGATT ATATTGATGA APTGCCATCA TCTGATAATC AGGAATATGA TGATAATTTCC
6121 GCTCCTTCTG GTGTTTTCTT TGTCCGCAA AATGATAATG TTAATCAAC TTTTAAAPT
6181 AATAACGTTT GGGCAAGGA TTTAATACGA GTTGTGAAI TGTTTGTAAA GTCTAATACT
6241 TCTAATCTCT CAAATGTATT ATCTATTGAC GCTCTAATC TATTAGTTGT TTCTGCACCT
6301 AAAGATATTT TAGATAACCT TCTCAATTC CTTTCTACTG TTGATTTGCC AACIGACCAG
6361 ATATTGATTT ACGGTTTGT ATTGAGGTT CAGCAAGTTC ATGCTTTAGA TTTTCAATTT
6421 CTTGCTGCTT CTCAGCGTGG CACTGTTTCA GCGGCTGTTA ATACTGACCG CTTCACTCT
6481 GTTTTATCTT CTGCTGGTGG TTCSTTGGT APTTTTAAI GCGATTTTT AGGGCTATCA
6541 GTTCGCGCAT TAAAGACIAA TAGCCATTC AAAATATTT CTGTCGACG TATTCTTACG
6601 CTTCAGGTC AGAAGGGTTC TATCTCTCTT GGCCAGAAI TCCCTTTTAT TACTGCTCT
6661 GTGACTGTTG AATCTGCCAA TGTAAATAAT CCATTTGAGA CATTGAGCG TCAAAATGTA
6721 GSTATTTCCA TGAGCGTTTT TCTGTTTCA APGCTGCGG GTAATATPT TCTGGATATT
6781 ACCAGCAAGG CCGATACTTT GASTTCTCT ACTCAGSCAA GTGATGTTAT TACTAATCAA

6841 AGAAGTATTTG CTACAACGGT TAAATTGCGT GATGGACAGA CTCPTTTACT CGSTGGCCTC
 6901 ACTGATTATA AAAACACCTC TCAAGATTCT GCGGTACCGT TCCTGTCTAA AATCCCTTTA
 6961 ATCGGCCTCC TGTATTAGCTC CGCCTCTGAT TCCCAAGAGG AAAGCACGTT ATACGTGCTC
 7021 GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATA AGCGCGCGGG GTGTGGTGGT
 7081 TACGCGCAGC GTGACCGCTA CACTTGCCAG CGCCCTAGGG CCGCCTCCTT TCGCTTTCTT
 7141 CCGTTCTFTT CTGCGCACGT TCGCCGGCTT TCCCGTCAA GCTCTAATC GGGGGCTCCC
 7201 TTTAGGGTTC CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA
 7261 TGETTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCTTTGA CGTTGGAGTC
 7321 CAGTTCTTTT AATAGTGGAC TCTTGTCCA AACTGGAACA ACACTCAACC CTATCTCGGG
 7381 CTATTCTTTT GATTATAAAC CGATTTTGGC GATTTGGGAA CCACCATCAA ACAGGATTTT
 7441 CGCCTGCTGG GGCAAAACAG CGTGGACCGC TTGCTGCAAC TCTCTCAGGG CCAGCGGCTG
 7501 AAGGGCAATC AGCTGTGCTC CGTCTACTG GTGAAAAGAA AAACCACCTT GATCCAAGC
 7561 TTGCAGGTGG CACTTTTCCG GGAATCTGC GCGGAACCCC TATTTGTTTA TTTTCTAAA
 7621 TACATTCAAA TATGTATCCG CTGATGAGAC AATAACCCCTG AATAATGCTT CAATATATAT
 7681 GAAAAAGGAA GAGTATGAGT ATTCAACATT TCGGTGTCCG CCTTATTCCT TTTTTTGCGG
 7741 CATTTTCCCT TCCCTTTTTT GCTCACCCAG AAACGCTGCT GAAAGTAAA GATGCTGAG
 7801 ATCASATGGG CGCACCGATG GCTTACATCG AACTGGATCT CACACGCGGT AAGATCCTTG
 7861 AGAGTTTTCC CCGGAAGAA CTTTTTCAA TGATGAGCAC TTTTAAAGTT CTGCTATGTC
 7921 ATACACTATT ATCCCGTATT GACGCGGGGC AAGAGCAACT CGGTCCCGGG GCGCGGTATT
 7981 CTCAGATGTA CTTGGTTGAG TACTCACCAG TCACAGAAAA GCATCTTACG GATGGCATGA
 8041 CAGTAAGAGT ATTAGTCACT GCTGCCATAA CCATCAGTGA TAACACTGCG GCTAACTTAC
 8101 TTTGACAAAC GATCGGAGGA CCGAAGGAGC TAACCGCTTT TTTGCACAAC ATGGGGGATC
 8161 ATGTAAGTCC CCTGTATGCT TGGGAACCGG AGCTGATGA AGCCATACCA AACGACGAGC
 8221 GTGACACCAC GATGCTCTGA GCAATGCCAA CAACGTTCCG CAACCTATTA ACTGCGGAAC
 8281 TACTTACTCT AGCTTCCCGG CAACAATTAA TAGACTGGAT GGAGCGGAT AAAGTTGCAG
 8341 GACCAGTTCT GCGCTCGGCC CTTCCGCTG CTTGCTTAT TCTGATAAA TCTGGAGCCG
 8401 GTGAGCGTGG GTCTCGCGGT ATCATTGCAG CACTGGGGCC AGATGGTAG CCGTCCCGTA
 8461 TGGTAGTTAT CTACACGACG GGGATCAGG CAACTATGGA TGAACGAAT AGACAGATCG
 8521 CTGAGATAGG TGCCTCACTG ATTAAGCATT GGTAACTGTC AGACCAAGTT TACTCATATA
 8581 TACTTTAGAT TGATTAAAA CTTCATTTTT AATTTAAAG GATCTAGGTG AAGATCCTTT
 8641 TTGATAATCT CATGACCAAA ATCCCTTAC GTGAGTTTTT GTTCCACTGT ACCTAAGACC
 8701 CCCAAGCTTG TCGACTGAAAT GCGAATGGC GCTTTGCCTG GTTCCCGCA CCAGAAGCGG
 8761 TGCCGGAAG CTGGCTGGAG TCGATCTTC CTGAGGCCGA TACTGTCTCT GTCCCTCAA
 8821 ACTGGCAGAT GCACGGTTAC GATGCCCCCA TCTACACCAA CGTAACCTAT CCCATACGG
 8881 TCAATCCGCC GTTGTCTCCG ACGGAGAATC CGACGGGTTG TFACTGCTC ACATTTAATG
 8941 TTGATGAAAG CTGGCTACAG GAAGGCCAGA CCGAATTAT TTTTGATGGC GTTCTATTTG
 9001 GTTAAAAAAT GAGCTGATTT AACAAAATT TAACGGGAAT TTTAACAAA TATTAACGTT
 9061 TACAAATTAA ATATTTGCTT ATACAACTT CCTGTTTTTG GGGCTTTCT GATATPCAC
 9121 CGGGTACTAT ATGATTGACA TCTAGTTTTT ACGATTAACG TTCATCGATT CTCTGTTTG

9181 CTCCAGACTC TCAGGCAATG ACCTGATAGC CTTGTAGAT CTCTCAAAA TAGCTACCTT
9241 CTCCGGCATG AATTATCAG CTAGAACGGT TGAATATCAT ATTGATGGTG ATTTGACTGT
9301 CTCCGGCCTT TCTCACCTT TTGAATCTT ACCTACACAT TACTCAGGCA TTGCATTTAA
9361 AATATATGAG GGTCTAAAA ATTTTATCC TTGGTGGAA ATAAAGGCTT CTCCCGCAA
9421 AGTATTACAG GGTCTAATG TTTTGGTAC AACCGATTA GCTTTATGCT CTGAGGCTTT
9481 ATGGCTTAAI TTTGCTAAT CTTTGCCTG COTGTATGAT TTATGGATG TT

Table 20C: Enzymes that either cut 15 or more human GLGs or have 5+-base recognition in FR3

Typical entry:

RName	Recognition	#sites
GLGid#:base#	GLGid#:base#	GLGid#:base#.....
<hr/>		
BstEII	Ggtacc	2
1:	3	48: 3
There are 2 hits at base# 3		
MacIII	gtnac	36
1:	4	2: 4
3:	4	4: 4
5:	4	6: 4
7:	4	8: 4
9:	4	10: 4
11:	4	37: 4
37:	58	38: 4
38:	58	39: 4
39:	58	40: 4
40:	58	41: 4
41:	58	42: 4
42:	58	43: 4
43:	58	44: 4
44:	58	45: 4
45:	58	46: 4
46:	58	47: 4
47:	58	48: 4
49:	4	50: 58
There are 24 hits at base# 4		
Esp45I	gtsac	33
1:	4	2: 4
3:	4	4: 4
5:	4	6: 4
7:	4	8: 4
9:	4	10: 4
11:	4	37: 4
37:	58	38: 4
38:	58	39: 58
40:	4	40: 58
41:	58	42: 58
43:	4	43: 58
44:	4	44: 58
45:	4	45: 58
46:	4	46: 58
47:	4	47: 58
48:	4	49: 4
50:	58	
There are 21 hits at base# 4		
HpaI	tcacc	45
1:	5	2: 5
3:	5	4: 5
5:	5	6: 5
7:	5	8: 5
11:	5	12: 5
12:	5	13: 5
14:	5	15: 5
16:	5	17: 5
18:	5	19: 5
20:	5	21: 5
22:	5	23: 5
24:	5	25: 5
26:	5	27: 5
28:	5	29: 5
30:	5	31: 5
32:	5	33: 5
34:	5	35: 5
36:	5	37: 5
39:	5	40: 5
43:	5	44: 5
45:	5	46: 5
47:	5	48: 5
49:	5	
There are 44 hits at base# 5		

NlaIII CATG 26
 1: 9 1: 42 2: 42 3: 9 3: 42 4: 9
 4: 42 5: 9 5: 42 6: 42 6: 78 7: 9
 7: 42 8: 21 8: 42 9: 42 10: 42 11: 42
 12: 57 13: 48 13: 57 14: 57 31: 72 38: 9
 48: 78 49: 78
 There are 11 hits at base# 42
 There are 1 hits at base# 48 Could cause raggedness.

BsaJI Ccungg 37
 1: 14 2: 14 5: 14 6: 14 7: 14 8: 14
 6: 65 9: 14 10: 14 11: 14 12: 14 13: 14
 14: 14 15: 65 17: 14 17: 65 18: 65 19: 65
 20: 65 21: 65 22: 65 26: 65 29: 65 30: 65
 33: 65 34: 65 35: 65 37: 65 38: 65 39: 65
 40: 65 42: 65 43: 65 48: 65 49: 65 50: 65
 51: 14
 There are 23 hits at base# 65
 There are 14 hits at base# 14

AluI AGct 42
 1: 47 2: 47 3: 47 4: 47 5: 47 6: 47
 7: 47 8: 47 9: 47 10: 47 11: 47 16: 63
 23: 63 24: 63 25: 63 31: 63 32: 63 36: 63
37: 47 37: 52 38: 47 38: 52 39: 47 39: 52
40: 47 40: 52 41: 47 41: 52 42: 47 42: 52
43: 47 43: 52 44: 47 44: 52 45: 47 45: 52
46: 47 46: 52 47: 47 47: 52 49: 15 50: 47
 There are 23 hits at base# 47
There are 11 hits at base# 52 Only 5 bases from 47

BlnI GCTnagc 21
 1: 48 2: 48 3: 48 5: 48 6: 48 7: 48
 8: 48 9: 48 10: 48 11: 48 37: 48 38: 48
 39: 48 40: 48 41: 48 42: 48 43: 48 44: 48
 45: 48 46: 48 47: 48
 There are 21 hits at base# 48

MwoI GCNMMNNnngc 19
 1: 48 2: 28 19: 36 22: 36 23: 36 24: 36
 25: 36 26: 36 35: 36 37: 67 39: 67 40: 67
 41: 67 42: 67 43: 67 44: 67 45: 67 46: 67
 47: 67

There are 10 hits at base# 67

There are 7 hits at base# 36

DdeI Ctnag 71
 1: 49 1: 58 2: 49 2: 58 3: 49 3: 58
 3: 65 4: 49 4: 56 5: 49 5: 59 5: 65
 6: 49 6: 58 6: 65 7: 49 7: 53 7: 65
 8: 49 8: 58 9: 49 9: 58 9: 55 10: 49
10: 59 10: 65 11: 49 11: 58 11: 55 15: 58
16: 58 16: 65 17: 58 18: 58 20: 58 21: 58
22: 58 23: 58 23: 65 24: 58 24: 65 25: 58
26: 65 26: 58 27: 58 27: 65 28: 58 30: 58
31: 58 31: 65 32: 58 32: 65 35: 58 36: 58
36: 65 37: 49 38: 49 39: 26 39: 49 40: 49
 41: 49 42: 26 42: 49 43: 49 44: 49 45: 49
 46: 49 47: 49 48: 12 49: 12 51: 65

There are 25 hits at base# 58

There are 22 hits at base# 49 Only nine base from 58

There are 16 hits at base# 65 Only seven bases from 58

BglII Agatct 11
 1: 61 2: 61 3: 61 4: 61 5: 61 6: 61
 7: 61 9: 61 10: 61 11: 61 51: 47
 There are 10 hits at base# 61

HstYI Rgatcy 12
 1: 61 2: 61 3: 61 4: 61 5: 61 6: 61
 7: 61 8: 61 9: 61 10: 61 11: 61 51: 47
 There are 11 hits at base# 61

Hpy188I TCNGa 17
 1: 64 2: 64 3: 64 4: 64 5: 64 6: 64
 7: 64 8: 64 9: 64 10: 64 11: 64 16: 57
 20: 57 27: 57 35: 57 46: 67 49: 57
 There are 11 hits at base# 64
 There are 4 hits at base# 57
 There are 2 hits at base# 67 Could be ragged.

MslI CAYNNnARWG 44
 1: 72 2: 72 3: 72 4: 72 5: 72 6: 72
 7: 72 8: 72 9: 72 10: 72 11: 72 15: 72
 17: 72 18: 72 19: 72 21: 72 23: 72 24: 72
 25: 72 26: 72 28: 72 29: 72 30: 72 31: 72
 32: 72 33: 72 34: 72 35: 72 36: 72 37: 72
 39: 72 39: 72 40: 72 41: 72 42: 72 43: 72
 44: 72 45: 72 46: 72 47: 72 48: 72 49: 72
 50: 72 51: 72
 There are 44 hits at base# 72

BsiEI CGRYcg 23
 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74
 33: 74 34: 74 37: 74 38: 74 39: 74 40: 74
 41: 74 42: 74 45: 74 46: 74 47: 74
 There are 23 hits at base# 74

EaeI Yggccr 23
 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74
 33: 74 34: 74 37: 74 38: 74 39: 74 40: 74
 41: 74 42: 74 45: 74 46: 74 47: 74
 There are 23 hits at base# 74

EagI Cggcgg 23
 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74

33: 74 34: 74 37: 74 38: 74 39: 74 40: 74

41: 74 42: 74 45: 74 46: 74 47: 74

There are 23 hits at base# 74

HaeIII GGcc 27

1: 75 3: 75 4: 75 5: 75 7: 75 8: 75

9: 75 10: 75 11: 75 16: 75 17: 75 20: 75

22: 75 30: 75 33: 75 34: 75 37: 75 38: 75

39: 75 40: 75 41: 75 42: 75 45: 75 46: 75

47: 75 48: 63 49: 63

There are 25 hits at base# 75

Bst4CI ACNgt 65°C 63 Sites There is a third isoschizmer

1: 86 2: 86 3: 86 4: 86 5: 86 6: 86

7: 34 7: 86 8: 86 9: 86 10: 86 11: 86

12: 86 13: 86 14: 86 15: 36 15: 86 16: 53

16: 86 17: 36 17: 86 18: 86 19: 86 20: 53

20: 86 21: 36 21: 86 22: 0 22: 86 23: 86

24: 86 25: 86 26: 86 27: 53 27: 86 28: 36

28: 86 29: 86 30: 86 31: 86 32: 86 33: 36

33: 86 34: 86 35: 53 35: 86 36: 86 37: 86

38: 86 39: 86 40: 86 41: 86 42: 86 43: 86

44: 86 45: 86 46: 86 47: 86 48: 86 49: 86

50: 86 51: 0 51: 86

There are 51 hits at base# 86 All the other sites are well away

HpyCH4III ACNgt 63

1: 86 2: 86 3: 86 4: 86 5: 86 6: 86

7: 34 7: 86 8: 86 9: 86 10: 86 11: 86

12: 86 13: 86 14: 86 15: 36 15: 86 16: 53

16: 86 17: 36 17: 86 18: 86 19: 86 20: 53

20: 86 21: 36 21: 86 22: 0 22: 86 23: 86

24: 86 25: 86 26: 86 27: 53 27: 86 28: 36

28: 86 29: 86 30: 86 31: 86 32: 86 33: 36

33: 86 34: 86 35: 53 35: 86 36: 86 37: 86

38: 86 39: 86 40: 86 41: 86 42: 86 43: 86

44: 86 45: 86 46: 86 47: 86 48: 86 49: 86

50: 86 51: 0 51: 86

There are 51 hits at base# 86

HinfI Gantc 43

2: 2 3: 2 4: 2 5: 2 6: 2 7: 2
 8: 2 9: 2 9: 22 10: 2 11: 2 15: 2
 16: 2 17: 2 18: 2 19: 2 19: 22 20: 2
 21: 2 23: 2 24: 2 25: 2 26: 2 27: 2
 29: 2 29: 2 30: 2 31: 2 32: 2 33: 2
 33: 22 34: 22 35: 2 36: 2 37: 2 38: 2
 40: 2 43: 2 44: 2 45: 2 46: 2 47: 2
 50: 60

There are 38 hits at base# 2

MlyI GAGTCNNNNNn 10

2: 2 3: 2 4: 2 5: 2 6: 2 7: 2
 8: 2 9: 2 10: 2 11: 2 37: 2 38: 2
 40: 2 43: 2 44: 2 45: 2 46: 2 47: 2

There are 18 hits at base# 2

PleI gagtc 16

2: 2 3: 2 4: 2 5: 2 6: 2 7: 2
 8: 2 9: 2 10: 2 11: 2 37: 2 38: 2
 40: 2 43: 2 44: 2 45: 2 46: 2 47: 2

There are 18 hits at base# 2

AclI Cggc 24

2: 26 9: 14 10: 14 11: 14 27: 74 37: 62
37: 65 38: 62 39: 65 40: 62 40: 65 41: 65
 42: 65 43: 62 43: 65 44: 62 44: 65 45: 62
 46: 62 47: 62 47: 65 48: 65 48: 74 49: 74

There are 8 hits at base# 62

There are 8 hits at base# 65

There are 3 hits at base# 14

There are 3 hits at base# 74

There are 1 hits at base# 26

There are 1 hits at base# 35

-"- Gcgg 11
 8: 91 9: 16 10: 16 11: 16 37: 67 39: 67
 40: 67 42: 67 43: 67 45: 67 46: 67
 There are 7 hits at base# 67
 There are 3 hits at base# 16
 There are 1 hits at base# 91

BsiHKAI GWGCWc 20
 2: 30 4: 30 5: 30 7: 30 9: 30 10: 30
 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51
 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51
 46: 51 47: 51
 There are 11 hits at base# 51

Bsp1286I GDGCHc 20
 2: 30 4: 30 6: 30 7: 30 9: 30 10: 30
 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51
 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51
 46: 51 47: 51
 There are 11 hits at base# 51

HgiAI GWGCWc 20
 2: 30 4: 30 6: 30 7: 30 9: 30 10: 30
 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51
 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51
 46: 51 47: 51
 There are 11 hits at base# 51

BsoFI GCngc 26
 2: 53 3: 53 5: 53 6: 53 7: 53 8: 53
 8: 91 9: 53 10: 53 11: 53 31: 53 36: 36
 37: 64 39: 64 40: 64 41: 64 42: 64 43: 64
 44: 64 45: 64 46: 64 47: 64 48: 53 49: 53
 50: 45 51: 53
 There are 13 hits at base# 53
 There are 10 hits at base# 64

TseI Gcwgc 17
 2: 53 3: 53 5: 53 6: 53 7: 53 8: 53

9: 53 10: 53 11: 53 31: 53 36: 53 45: 64
 46: 64 48: 53 49: 53 50: 45 51: 53
 There are 13 hits at base# 53

MnlI gagg 34
 3: 67 3: 95 4: 51 5: 16 5: 67 6: 67
 7: 67 8: 67 9: 67 10: 67 11: 67 15: 67
 16: 67 17: 67 19: 67 20: 67 21: 67 22: 67
 23: 67 24: 67 25: 67 26: 67 27: 67 28: 67
 29: 67 30: 67 31: 67 32: 67 33: 67 34: 67
 35: 67 36: 67 50: 67 51: 67
 There are 31 hits at base# 67

HpyCH4V TGca 34
 5: 90 6: 90 11: 90 12: 90 13: 90 14: 90
 15: 44 16: 44 16: 90 17: 44 18: 90 19: 44
 20: 44 21: 44 22: 44 23: 44 24: 44 25: 44
 26: 44 27: 44 27: 90 28: 44 29: 44 33: 44
 34: 44 35: 44 35: 90 36: 38 48: 44 49: 44
 50: 44 50: 90 51: 44 51: 52
 There are 21 hits at base# 44
 There are 1 hits at base# 52

AccI GTmkac 15 5-base recognition
 7: 16 11: 24 37: 16 38: 16 39: 16 40: 16
 41: 16 42: 16 43: 16 44: 16 45: 16 46: 16
 47: 16
 There are 11 hits at base# 16

SacII CCGCgg 3 6-base recognition
 9: 14 10: 14 11: 14 37: 65 39: 65 40: 65
 42: 65 43: 65
 There are 5 hits at base# 65
 There are 3 hits at base# 14

EfiI Gawtc 24
 9: 22 15: 2 16: 2 17: 2 18: 2 19: 2
 19: 22 20: 2 21: 2 23: 2 24: 2 25: 2

26: 2 27: 2 28: 2 29: 2 30: 2 31: 2
 32: 2 33: 2 33: 22 34: 22 35: 2 36: 2
 There are 20 hits at base# 2

BsmAI Nnnmmgagac 19
 15: 11 16: 11 20: 11 21: 11 22: 11 23: 11
 24: 11 25: 11 26: 11 27: 11 28: 11 28: 56
 30: 11 31: 11 32: 11 35: 11 36: 11 44: 87
 48: 87
 There are 16 hits at base# 11

EpaI ctccaq 19
 15: 12 16: 12 17: 12 18: 12 20: 12 21: 12
 22: 12 23: 12 24: 12 25: 12 26: 12 27: 12
 28: 12 30: 12 31: 12 32: 12 34: 12 35: 12
 36: 12
 There are 19 hits at base# 12

XmnI GAANNnttc 12
 37: 30 38: 30 39: 30 40: 30 41: 30 42: 30
 43: 30 44: 30 45: 30 46: 30 47: 30 50: 30
 There are 12 hits at base# 30

BsrI NCCagt 12
 37: 32 38: 32 39: 32 40: 32 41: 32 42: 32
 43: 32 44: 32 45: 32 46: 32 47: 32 50: 32
 There are 12 hits at base# 32

EamII GRGcYc 11
 37: 51 38: 51 39: 51 40: 51 41: 51 42: 51
 43: 51 44: 51 45: 51 46: 51 47: 51
 There are 11 hits at base# 51

Bcl136I GAGctc 11
 37: 51 38: 51 39: 51 40: 51 41: 51 42: 51
 43: 51 44: 51 45: 51 46: 51 47: 51
 There are 11 hits at base# 51

SacI GAGCTc 11

37: 51 38: 51 39: 51 40: 51 41: 51 42: 51

43: 51 44: 51 45: 51 46: 51 47: 51

There are 11 hits at base# 51

Table 217: Human HC GLG FR1 Sequences

VH Exon - Nucleotide sequence alignment

VH1

1-02 CAG GTG CAG CTG CTG CAG TCT GGG GCT GAG GTG AAG AAG CCT GGG GCC TCA GTG AAG
GTC TCC TGC AAG GCT TCT GGA TAC ACC TTC ACC

1-03 cag gtc cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag
gtt tcc tgc aag gct tct gga tac acc ttc act

1-06 cag gtg cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag
gtc tcc tgc aag gct tct gga tac acc ttc acc

1-16 cag gtc cag ctg ctg cag tct gga gct gag gtg aag aag cct ggg gcc tca gtg aag
gtc tcc tgc aag gct tcc ggt tac acc ttt acc

1-24 cag gtc cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag
gtc tcc tgc aag gtt tcc gga tac acc ttc act

1-45 cag atg cag ctg ctg cag tct ggg gct gag gtg aag aag act ggg tcc tca gtg aag
gtt tcc tgc aag gct tcc gga tac acc ttc acc

1-46 cag gtg cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag
gtt tcc tgc aag gca tct gga tac acc ttc acc

1-58 caa atg cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg acc tca gtg aag
gtc tcc tgc aag gct tcc gga ttc acc ttt act

1-69 cag gtg cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg tcc tca gtg aag
gtc tcc tgc aag gct tcc gga gcc acc ttc acc

1-e cag gtg cag ctg ctg cag tct ggg gct gag gtg aag aag cct ggg tcc tca gtg aag
gtc tcc tgc aag gct tcc gga gcc acc ttc acc

1-f Gag gtc cag ctg gta cag tct ggg gct gag gtg aag aag cct ggg gct aca gtg aaa
Aac tcc tgc aag gtt tcc gga tac acc ttc acc

VH2

2-05 CAG ATC ACC TTG AAG GAG TCT GGT CCT ACG CTG CTG AAA CCC ACA CAG ACC CTC ACC
CTG ACC TGC ACC TTC TCT GGG TTC TCA CTC AGC

2-26 cag gtc acc ttg aag gag tct ggt cct ctg ctg gtg aaa ccc aca gag acc ctg acc
ctg acc tgc acc gtc tct ggg ttc tca ctg agc

2-70 cag gtc acc ttg aag gag tct ggt cct gag ctg gtg aaa ccc aca gag acc ctg acc
ctg acc tgc acc ttc tct ggg ttc tca ctg agc

VH3

3-07 GAG GTG CAG CTG CTG CAG TCT GGG GGA GGC TTC GTC CAG CCT GGC GGG TCC CTC ACA
CTC TCC TGT GCA GCC TCT GGA TTC ACC TTT AGT

3-09 gaA gtg cag ctg ctg gag tct ggg gga ggc ttg gta cag cct ggc agg tcc ctg aga
ctc tcc tgt gca gcc tct gga ttc acc ttt gat

3-11 Cag gtg cag ctg ctg gag tct ggg gga ggc ttg gtc Aag cct gga ggg tcc ctg aga
ctc tcc tgt gca gcc tct gga ttc acc ttc agt

3-13 gag gtg cag ctg ctg gag tct ggg gga ggc ttg gta cag cct ggg ggg tcc ctg aga
ctc tcc tgt gca gcc tct gga ttc acc ttc agt

3-15 gag gtg cag ctg ctg gag tct ggg gga ggc ttg gta Aag cct ggg ggg tcc ctT aga
ctc tcc tgt gca gcc tct gga ttc act ttc agt

3-20 gag gtg cag ctg ctg gag tct ggg gga ggt ctg gta cgg cct ggg ggg tcc ctg aga

ctc tcc tgt gca gcc tct gga ttc acc ttt GAc
 3-21 gag gtg cag ctg gtg gag tct ggg gga ggc Ctg gtc Aag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-23 gag gtg cag ctg Ttg gag tct ggg gga ggc ttg gTA cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agC
 3-30 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gcc cag cct ggg Agg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-30.3 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-30.5 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-33 Cag gtg cag ctg gtg gag tcc ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-43 gAA gtg cag ctg gtg gag tct ggg gga gTc Gtg gTA cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt GAc
 3-48 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gTA cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-49 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gTA cag cct ggg ggg tcc ctg aga
 ctc tcc tgt Aca gct tct gga ttc acc ttt Ggt
 3-53 gag gtg cag ctg gtg gag Acc gGA gGA ggc ttg Atc cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct ggg ttc acc GtC agt
 3-64 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-66 gag gTg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc GtC agt
 3-72 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct gGA ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-75 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aAA
 ctc tcc tgt gca gcc tct ggg ttc acc ttt agt
 3-74 gag gtg cag ctg gTg gag tcc ggg gga ggc tTA gTt cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc ttt agt
 3-8 gag gtg cag ctg gtg gag tct Ggg gga gTc ttg gTA cag cct ggg ggg tcc ctg aga
 ctc tcc tgt gca gcc tct gga ttc acc GtC agt
 3-14
 4-04 CAG GTG CAG CTG CAG GAG TCG GCG CCA GGA CTA GTS AAG CCI TCG GGG ACC CTG TCC
 CTC ACC TGC GCT GTC TCT GGT GGC TCC ATC AAT
 4-28 cag gtg cag ctg cag gag tcc ggg cca gga ctg gtc aag cct tcc gAC acc ctg tcc
 ctc acc tgc gct gtc tct ggt TAc tcc atc agc
 4-30.1 cag gtg cag ctg cag gag tcc ggg cca gga ctg gtc aag cct tca CAg acc ctg tcc
 ctc acc tgc Act gtc tct ggt ggc tcc atc agc
 4-30.2 cag Ctg cag ctg cag gag tcc ggg Tca gga ctg gtc aag cct tca CAg acc ctg tcc
 ctc acc tgc gct gtc tct ggt ggc tcc atc agc
 4-30.4 cag gtg cag ctg cag gag tcc ggg cca gga ctg gtc aag cct tca CAg acc ctg tcc
 ctc acc tgc Act gtc tct ggt ggc tcc atc agc

4-31 cag gtg cag ctg cag gag tgg ggc cca gga ctg gtg aag cct tca CAg acc ctg tcc
ctc acc tgc Act gtc tct ggt ggc tcc atc agc

4-34 cag gtg cag cta cag Cag tGg ygc Gca gga ctg Ttg aag cct tcc gAg acc ctg tcc
ctc acc tgc gct gtc tAt ggt ggc tcc Ttc agT

4-39 cag Ctg cag ctg cag gag tcc ggc cca gga ctg gtg aag cct tcc gAg acc ctg tcc
ctc acc tgc Act gtc tct ggt ggc tcc atc agc

4-59 cag gtg cag ctg cag gag tgg ggc cca gga ctg gtg aag cct tcc gAg acc ctg tcc
ctc acc tgc Act gtc tct ggt ggc tcc atc agT

4-61 cag gTg cag ctg cag gag tcc ggc cca gga ctg gtg aag cct tcc gAg acc ctg tcc
ctc acc tgc Act gtc tct ggt ggc tcc Gtc agc

4-b cag gtg cag ctg cag gag tcc ggc cca gga ctg gtg aag cct tcc gAg acc ctg tcc
ctc acc tgc gct gtc tct ggt TAc tcc atc agc

VH5

5-51 GAG GTG CAG CTG GTC CAG TCT GGA GCA GAG GTC AAA AAG CCC GCG GAG TCT CTC AAG
ATC TCC TGT AAG GGT TCT GGA TAC ACC TTT ACC

5-a gaA gtg cag ctg gtg cag tct ggc gca gag gtg aaa aag ccc ggc gag tct ctg aGg
atc tcc tgt aag ggt tct gga tac agc ttt acc

VH6

6-1 CAG GTA CAG CTG CAG CAG TCA GGT CCA GGA CTG GTG AAG CCC TCG CAG ACC CTC TCA
CTC ACC TGT GCC ATC TCC GGG GAG AGT GTC TCT

VH7

7-4.1 CAG GTG CAG CTG GTC CAA TCT GGG TCT GAG TTG AAG AAG CCT GGG GCC TCA GTG AAG
GTC TCC TGC AAG GCT TCT GGA TAC ACC TTC ACT

Table 220: RERS sites in Human HC GLG FRIs where there are at least 20 GLGs cut

```

BsqI GTGCAG          71 (cuts 16/14 bases to right)
  1:  4   1: 13   2: 13   3:  4   3: 13   4: 13
  6: 13   7:  4   7: 13   8: 13   9:  4   9: 13
 10:  4  10: 13  15:  4  15: 65  16:  4  16: 65
 17:  4  17: 65  18:  4  18: 65  19:  4  19: 65
 20:  4  20: 65  21:  4  21: 65  22:  4  22: 65
 23:  4  23: 65  24:  4  24: 65  25:  4  25: 65
 26:  4  26: 65  27:  4  27: 65  28:  4  28: 65
 29:  4  30:  4  30: 65  31:  4  31: 65  32:  4
 32: 65  33:  4  33: 65  34:  4  34: 65  35:  4
 35: 65  36:  4  36: 65  37:  4  38:  4  39:  4
 41:  4  42:  4  43:  4  45:  4  46:  4  47:  4
 48:  4  48: 13  49:  4  49: 13  51:  4

```

There are 39 hits at base# 4

There are 21 hits at base# 65

```

-"-  GTGCAG          9
 12: 63  13: 63  14: 63  39: 63  41: 63  42: 63
 44: 63  45: 63  46: 63

```

```

EbvI GCACC          65
  1:  6   3:  6   6:  6   7:  6   8:  6   9:  6
 10:  6  15:  6  15: 67  16:  6  16: 67  17:  6
 17: 67  18:  6  18: 67  19:  6  19: 67  20:  6
 20: 67  21:  6  21: 67  22:  6  22: 67  23:  6
 23: 67  24:  6  24: 67  25:  6  25: 67  26:  6
 26: 67  27:  6  27: 67  28:  6  28: 67  29:  6
 30:  6  30: 67  31:  6  31: 67  32:  6  32: 67
 33:  6  33: 67  34:  6  34: 67  35:  6  35: 67
 36:  6  36: 67  37:  6  38:  6  39:  6  40:  6
 41:  6  42:  6  43:  6  44:  6  45:  6  46:  6
 47:  6  48:  6  49:  6  50: 12  51:  6

```

There are 43 hits at base# 6 **Bolded sites very near sites**
listed below

There are 21 hits at base# 67

```

-"-  GGTGC          13
 37:  9  38:  9  39:  9  40:  3  40:  9  41:  9
 42:  9  44:  3  44:  9  45:  9  46:  9  47:  9

```

50: 9

There are 11 hits at base# 9

BsoFI GCngc 76

1: 6	3: 6	6: 6	7: 6	8: 6	9: 6
10: 6	15: 6	15: 67	16: 6	16: 67	17: 6
17: 67	18: 6	18: 67	19: 6	19: 67	20: 6
20: 67	21: 6	21: 67	22: 6	22: 67	23: 6
23: 67	24: 6	24: 67	25: 6	25: 67	26: 6
26: 67	27: 6	27: 67	28: 6	28: 67	29: 6
30: 6	30: 67	31: 6	31: 67	32: 6	32: 67
33: 6	33: 67	34: 6	34: 67	35: 6	35: 67
36: 6	36: 67	<u>37: 6</u>	<u>37: 9</u>	<u>38: 6</u>	<u>38: 9</u>
39: 6	39: 9	<u>40: 6</u>	<u>40: 6</u>	<u>40: 9</u>	41: 6
41: 9	42: 6	42: 9	43: 6	<u>44: 3</u>	<u>44: 6</u>
<u>44: 9</u>	<u>45: 6</u>	<u>45: 9</u>	<u>46: 6</u>	<u>46: 9</u>	<u>47: 6</u>
<u>47: 9</u>	48: 6	49: 6	50: 9	50: 12	51: 6

There are 43 hits at base# 6 These often occur together.

There are 11 hits at base# 9

There are 2 hits at base# 3

There are 21 hits at base# 67

TseI GCngc 78

1: 6	3: 6	6: 6	7: 6	8: 6	9: 6
10: 6	15: 6	15: 67	16: 6	16: 67	17: 6
17: 67	18: 6	18: 67	19: 6	19: 67	20: 6
20: 67	21: 6	21: 67	22: 6	22: 67	23: 6
23: 67	24: 6	24: 67	25: 6	25: 67	26: 6
26: 67	27: 6	27: 67	28: 6	28: 67	29: 6
30: 6	30: 67	31: 6	31: 67	32: 6	32: 67
33: 6	33: 67	34: 6	34: 67	35: 6	35: 67
36: 6	36: 67	<u>37: 6</u>	<u>37: 9</u>	<u>38: 6</u>	<u>38: 9</u>
39: 6	39: 9	<u>40: 3</u>	<u>40: 6</u>	<u>40: 9</u>	<u>41: 6</u>
<u>41: 9</u>	<u>42: 6</u>	<u>42: 9</u>	43: 6	<u>44: 3</u>	<u>44: 6</u>
<u>44: 9</u>	<u>45: 6</u>	<u>45: 9</u>	<u>46: 6</u>	<u>46: 9</u>	<u>47: 6</u>
<u>47: 9</u>	48: 6	49: 6	50: 9	50: 12	51: 6

There are 43 hits at base# 6 Often together.

There are 11 hits at base# 9

There are 2 hits at base# 3

There are 1 hits at base# 12

There are 21 hits at base# 67

MspAII CMGcckg 48

1: 7	3: 7	4: 7	5: 7	6: 7	7: 7
8: 7	9: 7	10: 7	11: 7	15: 7	16: 7
17: 7	18: 7	19: 7	20: 7	21: 7	22: 7
23: 7	24: 7	25: 7	26: 7	27: 7	28: 7
29: 7	30: 7	31: 7	32: 7	33: 7	34: 7
35: 7	36: 7	37: 7	38: 7	39: 7	<u>40: 1</u>
<u>40: 7</u>	41: 7	42: 7	<u>44: 1</u>	<u>44: 7</u>	45: 7
46: 7	47: 7	48: 7	49: 7	50: 7	51: 7

There are 46 hits at base# 7

EvaII CAGctg 48

1: 7	3: 7	4: 7	5: 7	6: 7	7: 7
8: 7	9: 7	10: 7	11: 7	15: 7	16: 7
17: 7	18: 7	19: 7	20: 7	21: 7	22: 7
23: 7	24: 7	25: 7	26: 7	27: 7	28: 7
29: 7	30: 7	31: 7	32: 7	33: 7	34: 7
35: 7	36: 7	37: 7	38: 7	39: 7	<u>40: 1</u>
<u>40: 7</u>	41: 7	42: 7	<u>44: 1</u>	<u>44: 7</u>	45: 7
46: 7	47: 7	48: 7	49: 7	50: 7	51: 7

There are 46 hits at base# 7

There are 2 hits at base# 1

AluI AGct 54

1: 8	2: 8	3: 8	4: 8	4: 24	5: 8
6: 8	7: 8	8: 8	9: 8	10: 8	11: 8
15: 8	16: 8	17: 8	18: 8	19: 8	20: 8
21: 8	22: 8	23: 8	24: 8	25: 8	26: 8
27: 8	28: 8	29: 8	29: 69	30: 8	31: 8
32: 8	33: 8	34: 8	35: 8	36: 8	37: 8
38: 8	39: 8	<u>40: 2</u>	<u>40: 8</u>	41: 8	42: 8
43: 8	<u>44: 2</u>	<u>44: 8</u>	45: 8	46: 8	47: 8
48: 8	48: 82	49: 8	49: 62	50: 8	51: 8

There are 48 hits at base# 8

There are 2 hits at base# 2

DdeI Cnnag 48

1: 26	1: 46	2: 26	2: 48	3: 26	3: 48
4: 26	4: 48	5: 26	5: 48	6: 26	6: 48
7: 26	7: 48	8: 26	8: 46	9: 26	10: 26
11: 26	12: 35	13: 35	14: 35	15: 52	16: 52
17: 52	18: 52	19: 52	20: 52	21: 52	22: 52
23: 52	24: 52	25: 52	26: 52	27: 52	28: 52
29: 52	30: 52	31: 52	32: 52	33: 52	35: 30
35: 52	36: 52	40: 24	48: 52	51: 26	51: 48

There are 22 hits at base# 52 52 and 48 never together.

There are 9 hits at base# 48

There are 12 hits at base# 26 26 and 24 never together.

HphI tcacc 42

1: 86	3: 86	6: 86	7: 86	8: 80	11: 86
12: 5	13: 5	14: 5	15: 80	16: 80	17: 80
19: 80	20: 80	21: 80	22: 80	23: 80	24: 80
25: 80	26: 80	27: 80	28: 80	29: 80	30: 80
31: 80	32: 80	33: 80	34: 80	35: 80	36: 80
37: 59	38: 59	39: 59	40: 59	41: 59	42: 59
43: 59	44: 59	45: 59	46: 59	47: 59	50: 59

There are 22 hits at base# 80 80 and 86 never together

There are 5 hits at base# 86

There are 12 hits at base# 59

BssKI Ncnnng 50

1: 39	2: 39	3: 39	4: 39	5: 39	7: 39
8: 39	9: 39	10: 39	11: 39	15: 39	16: 39
17: 39	18: 39	19: 39	20: 39	21: 29	21: 39
22: 39	23: 39	24: 39	25: 39	26: 39	27: 39
28: 39	29: 39	30: 39	31: 39	32: 39	33: 39
34: 39	35: 19	35: 39	36: 39	37: 24	38: 24
39: 24	41: 24	42: 24	44: 24	45: 24	46: 24
47: 24	<u>48: 39</u>	<u>48: 40</u>	<u>49: 39</u>	<u>49: 40</u>	50: 24
50: 73	51: 39				

There are 35 hits at base# 39 39 and 40 together twice.

There are 2 hits at base# 40

BsaOI CCnngg 47

1: 40	2: 40	3: 40	4: 40	5: 40	7: 40
8: 40	9: 40	9: 47	10: 40	10: 47	11: 40
15: 40	18: 40	19: 40	20: 40	21: 40	22: 40
23: 40	24: 40	25: 40	26: 40	27: 40	28: 40
29: 40	30: 40	31: 40	32: 40	34: 40	35: 20
35: 40	36: 40	37: 24	38: 24	39: 24	41: 24
42: 24	44: 24	45: 24	46: 24	47: 24	<u>48: 40</u>
<u>48: 41</u>	<u>49: 40</u>	<u>49: 41</u>	50: 74	51: 40	

There are 32 hits at base# 40 40 and 41 together twice
 There are 2 hits at base# 41
 There are 9 hits at base# 24
 There are 2 hits at base# 47

BstNI CCwgg 44

BspGI ccwgg

SacFI (\$M.HpaII) CCwgg

1: 40	2: 40	3: 40	4: 40	5: 40	7: 40
8: 40	9: 40	10: 40	11: 40	15: 40	16: 40
17: 40	18: 40	19: 40	20: 40	21: 30	21: 40
22: 40	23: 40	24: 40	25: 40	26: 40	27: 40
28: 40	29: 40	30: 40	31: 40	32: 40	33: 40
34: 40	35: 40	36: 40	37: 25	38: 25	39: 25
41: 25	42: 25	44: 25	45: 25	46: 25	47: 25
50: 25	51: 40				

There are 33 hits at base# 40

SacFI CCnngg 50

1: 40	2: 40	3: 40	4: 40	5: 40	7: 40
8: 40	9: 40	10: 40	11: 40	15: 40	16: 40
17: 40	18: 40	19: 40	20: 40	21: 30	21: 40
22: 40	23: 40	24: 40	25: 40	26: 40	27: 40
28: 40	29: 40	30: 40	31: 40	32: 40	33: 40
34: 40	35: 20	35: 40	36: 40	37: 25	38: 25
39: 25	41: 25	42: 25	44: 25	45: 25	46: 25
47: 25	48: 40	48: 41	49: 40	49: 41	50: 25
50: 74	51: 40				

There are 35 hits at base# 40

There are 2 hits at base# 41

EcoO109I RGGnccy 34
 1: 43 2: 43 3: 43 4: 43 5: 43 6: 43
 7: 43 8: 43 9: 43 10: 43 15: 46 16: 46
 17: 46 18: 46 19: 46 20: 46 21: 46 22: 46
 23: 46 24: 46 25: 46 26: 46 27: 46 28: 46
 30: 46 31: 46 32: 46 33: 46 34: 46 35: 46
 36: 46 37: 46 43: 79 51: 43

There are 22 hits at base# 46 46 and 43 never together

There are 11 hits at base# 43

NlaIV GGNnc 71
 1: 43 2: 43 3: 43 4: 43 5: 43 6: 43
 7: 43 8: 43 9: 43 9: 79 10: 43 10: 79
~~15: 46 15: 47 16: 47 17: 46 17: 47 18: 46~~
~~18: 47 19: 46 19: 47 20: 46 20: 47 21: 46~~
~~21: 47 22: 46 22: 47 23: 47 24: 47 25: 47~~
~~26: 47 27: 46 27: 47 28: 46 28: 47 29: 47~~
~~30: 46 30: 47 31: 46 31: 47 32: 46 32: 47~~
~~33: 46 33: 47 34: 46 34: 47 35: 46 35: 47~~
~~36: 46 36: 47 37: 21 37: 46 37: 47 37: 79~~
 38: 21 39: 21 39: 79 40: 79 41: 21 41: 79
 42: 21 42: 79 43: 79 44: 21 44: 79 45: 21
 45: 79 46: 21 46: 79 47: 21 51: 43

There are 23 hits at base# 47 46 & 47 often together

There are 17 hits at base# 46 There are 11 hits at base# 43

Sau96I Ggncc 70
 1: 44 2: 3 2: 44 3: 44 4: 44 5: 3 5: 44 6: 44
 7: 44 8: 22 8: 44 9: 44 10: 44 11: 3 12: 22 13: 22
 14: 22 15: 33 15: 47 16: 47 17: 47 18: 47 19: 47 20: 47
 21: 47 22: 47 23: 33 23: 47 24: 33 24: 47 25: 33 25: 47
 26: 33 26: 47 27: 47 28: 47 29: 47 30: 47 31: 33 31: 47
 32: 33 32: 47 33: 33 33: 47 34: 33 34: 47 35: 47 36: 47
~~37: 21 37: 22 37: 47 38: 21 38: 22 38: 21 38: 22 41: 21~~
 41: 22 42: 21 42: 22 43: 80 44: 21 44: 22 45: 21 45: 22
 46: 21 46: 22 47: 21 47: 22 50: 22 51: 44

There are 23 hits at base# 47 These do not occur together.

There are 11 hits at base# 44

There are 14 hits at base# 22 These do occur together.

There are 9 hits at base# 21

```
BsmAI GCTCCNnnnn          22
 1: 58   3: 58   4: 58   5: 58   8: 58   9: 58
10: 58  13: 70  36: 15  37: 70  38: 70  39: 70
40: 70  41: 70  42: 70  44: 70  45: 70  46: 70
47: 70  48: 48  49: 48  50: 83
There are 11 hits at base# 70
```

```
-"- Nnnnnngagac          27
13: 40  15: 48  16: 48  17: 48  18: 48  20: 48
21: 48  22: 48  23: 48  24: 48  25: 48  26: 48
27: 48  28: 48  29: 48  30: 10  30: 48  31: 48
32: 48  33: 48  35: 48  36: 48  43: 40  44: 40
45: 40  46: 40  47: 40
There are 20 hits at base# 48
```

```
AvaII Ggwcc          44
Sau96I($M.NaeIII) Ggwcc  44
 2: 3   5: 3   6: 44  8: 44   9: 44  10: 44
11: 3  12: 22  13: 22  14: 22  15: 33  13: 47
16: 47  17: 47  18: 47  19: 47  20: 47  21: 47
22: 47  23: 33  23: 47  24: 33  24: 47  25: 33
25: 47  26: 33  26: 47  27: 47  28: 47  29: 47
30: 47  31: 33  31: 47  32: 33  32: 47  33: 33
33: 47  34: 33  34: 47  35: 47  36: 47  37: 47
43: 80  50: 22
There are 23 hits at base# 47 44 & 47 never together
There are 4 hits at base# 44
```

```
EpuMI RGGwccy          27
 6: 43   8: 43   9: 43  10: 43  15: 46  16: 46
17: 46  18: 46  19: 46  20: 46  21: 46  22: 46
23: 46  24: 46  25: 46  26: 46  27: 46  28: 46
30: 46  31: 46  32: 46  33: 46  34: 46  35: 46
36: 46  37: 46  43: 79
There are 22 hits at base# 46 43 and 46 never occur together.
There are 4 hits at base# 43
```

BsmFI GGGAC 3
 8: 48 37: 46 50: 77
 -"- gtaac 33
 15: 48 16: 48 17: 48 1: 0 1: 0 20: 48
 21: 48 22: 48 23: 48 24: 48 25: 48 26: 48
 27: 48 28: 48 29: 48 30: 48 31: 48 32: 48
 33: 48 34: 48 35: 48 36: 48 37: 54 38: 54
 39: 54 40: 54 41: 54 42: 54 43: 54 44: 54
 45: 54 46: 54 47: 54
 There are 20 hits at base# 48
 There are 11 hits at base# 54

HinFI Gattc 80
 8: 77 12: 16 13: 16 14: 16 15: 16 15: 56
 15: 77 16: 16 16: 56 16: 77 17: 16 17: 56
 17: 77 18: 16 18: 56 18: 77 19: 16 19: 56
 19: 77 20: 16 20: 56 20: 77 21: 16 21: 56
 21: 77 22: 16 22: 56 22: 77 23: 16 23: 56
 23: 77 24: 16 24: 56 24: 77 25: 16 25: 56
 25: 77 26: 16 26: 56 26: 77 27: 16 27: 56
 27: 56 27: 77 28: 16 28: 56 28: 77 29: 16
 29: 56 29: 77 30: 56 31: 16 31: 56 31: 77
 32: 16 32: 56 32: 77 33: 16 33: 56 33: 77
 34: 16 35: 16 35: 56 35: 77 36: 16 36: 56
 36: 56 36: 77 37: 16 38: 16 39: 16 40: 16
 41: 16 42: 16 44: 16 45: 16 46: 16 47: 16
 48: 46 49: 46
 There are 34 hits at base# 16

RsiI Gattc 21
 8: 77 15: 77 16: 77 17: 77 18: 77 19: 77
 20: 77 21: 77 22: 77 23: 77 24: 77 25: 77
 26: 77 27: 77 28: 77 29: 77 31: 77 32: 77
 33: 77 35: 77 36: 77
 There are 21 hits at base# 77

MlyI GAGTC 36
 12: 16 13: 16 14: 16 15: 16 16: 16 17: 16
 18: 16 19: 16 20: 16 21: 16 22: 16 23: 16
 24: 16 25: 16 26: 16 27: 16 27: 26 28: 16
 29: 16 31: 16 32: 16 33: 16 34: 16 35: 16
 36: 16 36: 26 37: 16 38: 16 39: 16 40: 16
 41: 16 42: 16 44: 16 45: 16 46: 16 47: 16
 48: 46 49: 46
 There are 36 hits at base# 16

-"- GACTC 21
 15: 56 16: 56 17: 56 18: 56 19: 56 20: 56
 21: 56 22: 56 23: 56 24: 56 25: 56 26: 56
 27: 56 28: 56 29: 56 30: 56 31: 56 32: 56
 33: 56 35: 56 36: 56
 There are 21 hits at base# 56

PleI gagtc 36
 12: 16 13: 16 14: 16 15: 16 16: 16 17: 16
 18: 16 19: 16 20: 16 21: 16 22: 16 23: 16
 24: 16 25: 16 26: 16 27: 16 27: 26 28: 16
 29: 16 31: 16 32: 16 33: 16 34: 16 35: 16
 36: 16 36: 26 37: 16 38: 16 39: 16 40: 16
 41: 16 42: 16 44: 16 45: 16 46: 16 47: 16
 48: 46 49: 46
 There are 36 hits at base# 16

-"- gactc 21
 15: 56 16: 56 17: 56 18: 56 19: 56 20: 56
 21: 56 22: 56 23: 56 24: 56 25: 56 26: 56
 27: 56 28: 56 29: 56 30: 56 31: 56 32: 56
 33: 56 35: 56 36: 56
 There are 21 hits at base# 56

AlwNI CAGNNNctg 26
 15: 68 16: 68 17: 68 18: 68 19: 68 20: 68
 21: 68 22: 68 23: 68 24: 68 25: 68 26: 68
 27: 68 28: 68 29: 68 30: 68 31: 68 32: 68
 33: 68 34: 68 35: 68 36: 68 39: 46 40: 46
 41: 46 42: 46
 There are 22 hits at base# 68

Table 255: Analysis of frequency of matching MEdaptions in actual V genes

A: HpyCH4V in 6C at bases 35-56

Id	Number of mismatches										Number	Cut	Id	Probe										
	0	1	2	3	4	5	6	7	8	9					10									
1	510	5	11	274	92	61	25	22	11	1	3	5	443	6-1	agtttcccctcCAGctgaactc									
2	192	54	42	32	24	15	2	3	10	3	1	6	157	3-11	caactgtatctcCAaatgaacag									
3	58	19	7	17	6	5	1	0	1	0	2	0	54	3-09	ccctgtatctcCAaatgaacag									
4	267	42	33	9	8	8	82	43	22	8	11	1	100	5-51	ccgctaccctcCAGtgggacag									
5	250	111	59	41	24	7	5	1	0	0	2	0	242	3-15	cgctgtatctcCAaatgaacag									
6	7	0	2	0	1	0	0	0	0	0	4	0	3	7-4.1	eggcatatctcCAatctctcag									
7	7	0	2	2	0	0	2	1	0	0	0	0	4	3-73	ggggtalctcCAaatgaacag									
8	26	10	4	1	3	1	2	1	3	1	0	0	19	5-a	ctgctaccctcCAGtgggacag									
9	21	8	2	3	1	6	1	0	0	0	0	0	20	3-49	tgcctatctcCAaatgaacag									
											1338	249	162	379	149	103	120	71	47	13	23	12	1052	
											249	411	790	939	1162	1280	1316	1233	1293	1338				

Id	Probe	dotted probe
6-1	agtttcccctcCAGctgaactc	agtttcccctcCAGctgaactc
3-11	caactgtatctcCAaatgaacag	caac.g.at.....aa.....ag
3-09	ccctgtatctcCAaatgaacag	ccc.g.at.....aa.....ag
5-51	ccgctaccctcCAGtgggacag	ccgc..a.....tg.g.ag
3-15	cgctgtatctcCAaatgaacag	c.c.g.at.....aa.....ag
7-4.1	eggcatatctcCAatctctcag	c.gca.at.....a.ctg.ag
3-73	ggggtalctcCAaatgaacag	c.gcy.al.....aa.....ag
5-a	ctgctaccctcCAGtgggacag	ctgc..e.....tg.g.ag
3-49	tgcctatctcCAaatgaacag	tegc..at.....aa.....ag

Seqs with the expected RE site only.....1004
 (Counts only cases with 4 or fewer mismatches)
 Seqs with only an unexpected site..... 0
 Seqs with both expected and unexpected.... 48
 (Counts only cases with 4 or fewer mismatches)
 Seqs with no sites..... 0

B:RplmHC

Id	Ntot	0	1	2	3	4	5	6	7	8	Nout	Name	
1	133	73	16	11	13	6	9	1	4	0	119	1-58	acatggactcvtgacgactgag
2	14	11	1	0	0	0	1	0	1	1	12	1-02	acatggagctgacgactgag
3	34	17	8	2	6	1	0	0	0	0	1-18	acatggagctgacgactgag	
4	120	50	32	16	10	9	1	1	1	0	2	5-51	acctgactggagcaactgaa
5	55	13	11	10	7	3	1	0	0	0	0	3-15	atctgcaaatgacagactgaa
6	340	106	98	41	15	6	3	0	1	0	0	3303	atctgcaaatgacagactgag
7	82	25	16	25	12	1	3	0	0	0	0	3-20	atctgcaaatgacagactgag
8	3	0	2	0	1	0	0	0	0	0	0	74.1	atctgcaaatgacagactgaa
9	23	18	2	2	1	0	0	0	0	0	0	3-56	atcttcaaatgacagactgag
10	2	1	0	1	0	0	0	0	0	0	0	3-64	atcttcaaatgacagactgag
11	486	249	78	81	38	21	10	4	4	1	467	4301	ccctgaaactgactctgtgac
12	16	6	3	1	0	1	1	3	1	0	1	6-1	ccctgaaactgactctgtgac
13	28	15	8	2	2	1	0	0	0	0	0	2-70	tccttcaaatgacaaactgga
14	2	0	2	0	0	0	0	0	0	0	0	2-26	tccttcaaatgacaaactgga

Name	Full sequence	Dot mode
1-58	acatggagcctgacccagcctgag	acatggagcctgacccagcctgag
1-02	acatggagcctgagcaggctgagg.....
1-18	acatggagcctgagcaggctgagg.....
5-51	acatggagcctgagcaggctgaa	..c..c..tg.....a
3-15	atctgcaaatgcaacagcctgaa	.tc..c..aa...a.....a
3-30.3	atctgcaaatgcaacagcctgag	.tc..c..aa...a.....
3-20	atctgcaaatgcaacagcctgag	.tc..c..aa...a..t.....
7-4.1	atctgcaaatgcaacagcctgaa	.tc..s..a.ct.....a.a
3-66	atcttcaaatgcaacagcctgag	.tc..tc..aa...a.....
3-64	atcttcaaatgcaacagcctgag	.tc..tc..aa...g.....
4-30.1	ccctggaagcctgagcctctgag	c.c..a.....ctctg...c
6-1	ccctggaagcctgagcctctgag	c.c..c.....a.ctctg...c
2-70	tccttcaaatgcaacagcctgaa	t.c.tacaa...c..a.a..ga
2-26	tccttcaaatgcaacagcctgaa	t.c.tacaa...c..a.a..ga

Seqs with the expected RE site only..... 597 (counting sequences with 4 or fewer mismatches)

Seqs with only an unexpected site..... 2

Seqs with both expected and unexpected... 2

Seqs with no sites..... 686

C: HpyCH11, BstCI, or PstI in HC

In scoring whether the RE site of interest is present, only CIs that have 4 or fewer mismatches are counted.

Number of sequences..... 1617

Lot	0	1	2	3	4	5	6	7	8	Neut	Cont	Cont		
1	244	78	92	43	18	10	1	2	0	0	241	102#1,1	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
2	457	69	150	115	65	34	11	8	3	1	494	103#2,3	ctgtgtatlaactgtgtgagaga	ctgtgtatlaactgtgtgagaga
3	173	52	45	36	22	14	3	0	0	1	169	104#3	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
4	16	0	3	2	2	1	0	0	1	1	0	124#5,1	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
5	4	0	0	1	0	1	1	0	1	0	2	145#6	ccatgtattactctgtgtgagaga	ccatgtattactctgtgtgagaga
6	15	1	0	1	0	6	4	1	1	1	8	158#8	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
7	23	4	8	5	2	2	1	1	0	0	21	205#12	ccacatattactctgtgtgagaga	ccacatattactctgtgtgagaga
8	9	1	1	1	0	3	2	1	0	0	6	226#13	ccacatattactctgtgtgagaga	ccacatattactctgtgtgagaga
9	7	1	3	1	1	0	0	1	0	0	6	270#14	ccatgtattactctgtgtgagaga	ccatgtattactctgtgtgagaga
10	23	7	3	5	5	2	1	0	0	0	22	309#16	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
11	35	5	10	7	6	3	3	0	1	0	31	313#18	ctgtgtattactctgtgtgagaga	ctgtgtattactctgtgtgagaga
12	18	2	3	2	2	6	1	0	2	0	15	315#19	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
13	3	1	2	0	0	0	0	0	0	0	3	320#20	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
14	117	29	23	28	22	8	4	2	1	0	110	323#22	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
15	75	21	25	13	9	1	4	2	0	0	69	338#23	ctgtgtattactctgtgtgagaga	ctgtgtattactctgtgtgagaga
16	14	2	2	2	3	0	3	1	1	0	9	349#29	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
17	2	0	0	1	0	0	1	0	0	0	1	372#33	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
18	1	0	0	1	0	0	0	0	0	0	1	373#34	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
19	2	0	0	0	0	0	0	0	0	2	0	38#36	ctgtgtattactctgtgtgagaga	ctgtgtattactctgtgtgagaga
20	34	4	9	9	4	5	3	0	0	0	31	423#38	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
21	17	5	4	2	2	3	1	0	0	0	16	4302#40	ccgtgtattactctgtgtgagaga	ccgtgtattactctgtgtgagaga
22	75	15	17	24	7	19	1	1	0	0	73	439#44	ctgtgtattactctgtgtgagaga	ctgtgtattactctgtgtgagaga
23	40	14	15	4	5	1	0	1	0	0	39	551#48	ccatgtattactctgtgtgagaga	ccatgtattactctgtgtgagaga
24	213	26	56	60	42	20	7	2	0	0	204	58#49	ccatgtattactctgtgtgagaga	ccatgtattactctgtgtgagaga
Group		337	471	363	218	130	58	23	11	6				
Cumulative		337	808	1171	1389	1519	1577	1600	1611	1617				

Sacs with the expected SE site only.....1511
 Sacs with only an unexpected site..... 6

Table 255 15

Seqs with both expected and unexpected.... 8
 Seqs with no sites..... 6

Analysis repeated using only 8 best REadaptors

Id	Ntot	C	1	2	3	4	5	6	7	8+			
1	301	78	101	54	32	16	9	10	1	0	281	102#1	ccgtgtattactgtgcgagaga
2	493	69	155	125	73	37	14	11	3	6	459	103#2	ctgtgtattactgtgcgagaga
3	189	52	45	38	23	18	5	4	1	3	176	108#3	ccgtgtattactgtgcgagagg
4	127	29	23	28	24	10	6	5	2	0	114	323#22	ccgtatattactgtgcgaaaga
5	78	21	25	14	11	1	4	2	0	0	72	330#23	ctgtgtattactgtgcgaaaga
6	79	15	17	25	8	11	1	2	0	0	76	439#44	ctgtgtattactgtgcgagaca
7	43	14	15	5	5	3	0	1	0	0	42	551#48	ccatgtattactgtgcgagaca
8	307	26	63	72	51	38	24	14	13	6	250	5a#49	ccatgtattactgtgcgaga
1	102#1		ccgtgtattactgtgcgagaga		ccgtgtattactgtgcgagaga								
2	103#2		ctgtgtattactgtgcgagaga		.t.....								
3	108#3		ccgtgtattactgtgcgagagg	g								
4	323#22		ccgtatattactgtgcgaaaga		...a.....a...								
5	330#23		ctgtgtattactgtgcgaaaga		.t.....a...								
6	439#44		ctgtgtattactgtgcgagaca		.t.....c.								
7	551#48		ccatgtattactgtgcgagaca		..a.....c.								
8	5a#49		ccatgtattactgtgcgagaAA		..a.....AA								

Seqs with the expected RE site only.....1453 / 1617
 Seqs with only an unexpected site..... 0
 Seqs with both expected and unexpected.... 7
 Seqs with no sites..... 0

Table 200: Kappa FR1 CLGs

!	1	2	3	4	5	6	7	8	9	10	11	12	
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
!	13	14	15	16	17	18	19	20	21	22	23		
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	O12
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	.O2
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	O18
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	OS
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	A20
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	A30
	AAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	GCC	ATG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L14
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L1
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L15
	GCC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L4
	GCC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L18
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	TCC	GTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L5
	GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	TCT	GTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	I19
	GAC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TTC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L8
	GCC	ATC	CGG	ATG	ACC	CAG	TCT	CCA	TTC	TCC	CTG	TCT	
	GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L23
	GCC	ATC	CGG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	TTC	TCT	
	GCA	TCT	ACA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L9
	GTC	ATC	TGG	ATG	ACC	CAG	TCT	CCA	TCC	TTA	CTC	TCT	

GCA TCT ACA GGA GAC AGA GTC ACC ATC AGT TGT 1 L24
 GCC ATC CAG ATG ACC CAG TCT CCA TCC TCC CTG TCT
 GCA TCT GTA GGA GAC AGA GTC ACC ATC ACT TGC 1 L11
 GAC ATC CAG ATG ACC CAG TCT CCT TCC ACC CTG TCT
 GCA TCT GTA GGA GAC AGA GTC ACC ATC ACT TGC 1 L12
 GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC CTG CCC
 GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC 1 G11
 GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC CTG CCC
 GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC 1 G1
 GAT GTT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC
 GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC 1 A17
 GAT GTT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC
 GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC 1 A1
 GAT ATT GTG ATG ACC CAG ACT CCA CTC TCT CTG TCC
 GTC ACC CCT GGA CAG CCG GCC TCC ATC TCC TGC 1 A18
 GAT ATT GTG ATG ACC CAG ACT CCA CTC TCT CTG TCC
 GTC ACC CCT GGA CAG CCG GCC TCC ATC TCC TGC 1 A2
 GAT ATT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC
 GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC 1 A19
 GAT ATT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC
 GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC 1 A3
 GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC TCA CCT
 GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC 1 A23
 GAA ATT GTG TTG ACG CAG TCT CCA GCC ACC CTG TCT
 TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 A27
 GAA ATT GTG TTG ACG CAG TCT CCA GCC ACC CTG TCT
 TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 A11
 GAA ATA GTG ATG ACC CAG TCT CCA GCC ACC CTG TCT
 GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 L2
 GAA ATA GTG ATG ACC CAG TCT CCA GCC ACC CTG TCT
 GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 L16
 GAA ATT GTG TTG ACA CAG TCT CCA GCC ACC CTG TCT
 TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 L6
 GAA ATT GTG TTG ACA CAG TCT CCA GCC ACC CTG TCT
 TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC 1 L20
 GAA ATT GTA ATG ACA CAG TCT CCA GCC ACC CTG TCT

TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC	!	L25
GAC ATC GTG ATG ACC CAG TCT CCA GAC TCC CTG GCT		
GTG TCT CTG GGC GAG AGG GCC ACC ATC AAC TGC	!	E3
GAA ACC ACA CTC ACG CAG TCT CCA GCA TTC ATG TCA		
GCG ACT CCA GGA GAC AAA GTC AAC ATC TCC TGC	!	B2
GAA ATT GTG CTG ACT CAG TCT CCA GAC TTT CAG TCT		
GTG ACT CCA AAG GAG AAA GTC ACC ATC ACC TGC	!	A26
GAA ATT GTG CTG ACT CAG TCT CCA GAC TTT CAG TCT		
GTG ACT CCA AAG GAG ABA GTC ACC ATC ACC TGC	!	A10
GAT GTT GTG ATG ACA CAG TCT CCA GCT TTC CTC TCT		
GTG ACT CCA GGG GAG AAA GTC ACC ATC ACC TGC	!	A14

Table 302 RERS sites found in Human Kappa FR1 GLSs

	MslI	FokI	EcoRI	PstI	BamHI	MbII	HpyCH
		--> <-- -->					AV
G12	1-69	3	3	12 49	15 18	47	26
O2	101-169	103	123	112 149	115 118	147	136
O18	201-269	203	223	212 249	215 218	247	236
O8	301-369	303	323	312 349	315 318	347	336
A20	401-469	403	423	412 449	415 418	447	436
A30	501-569	503	523	512 549	515 518	547	536
L14	601-669	603	623	612 649	615 618	647	636
L1	701-769	703	723	712 749	715 718	747	736
L15	801-869	803	823	812 849	815 818	847	836
L4	901-969	-	903	912 949	915 918	947	936
L18	1001-1069	-	1003	1012 1049	1015 1018	1047	1036
L5	1101-1169	1103	-	1112 1149	1115 1118	1147	1136
L19	1201-1269	1203	1203	1212 1249	1215 1218	1247	1236
L8	1301-1369	-	1303	1312 1349	1315 1318	1347	1336
L23	1401-1469	1403	1408	1412 1449	1415 1418	1447	1436
L9	1501-1569	1503	1508 1523	1512 1549	1515 1518	1547	1536
L24	1601-1669	1603	1608 1623	1612 1649	1615 1618	1647	1636
L11	1701-1769	1703	1723	1712 1749	1715 1718	1747	1736
L12	1801-1869	1803	1803	1812 1849	1815 1818	1847	1836

	MnIT	FokI --> <-- --< >--	PflIT	BarI	PamAI	MnIT	HpyCH AV
OLL 1901-1959	-	-	-	-	-	1956	-
OL 2001-2059	-	-	-	-	-	2056	-
AL7 2101-2159	-	-	2112	-	2118	2156	-
AL 2201-2259	-	-	2212	-	2218	2256	-
ALB 2301-2359	-	-	-	-	-	2356	-
A2 2401-2459	-	-	-	-	-	2456	-
AL9 2501-2559	-	-	2512	-	2518	2556	-
A3 2601-2659	-	-	2612	-	2618	2656	-
A23 2701-2759	-	-	-	-	-	2729 2756	-

A27 2801-2859	-	-	2812	-	2818 2839	2860	-
AL1 2901-2959	-	-	2912	-	2918 2939	2960	-
L2 3001-3059	-	-	3012	-	3018 3039	3060	-
LL6 3101-3159	-	-	3112	-	3118 3139	3160	-
L6 3201-3259	-	-	3212	-	3218 3239	3260	-

	MslI	FokI --> <---	EcoRI --> <---	BamHI	BclI	BsuRI	HpaII	HpyCH 4V
L20 3301-3369	-	-	3312	3318 3339	-	-	-	-
L25 3401-3469	-	-	3412	3418 3439	-	-	-	-
B3 3501-3569	3503	-	3512	3518 3539	3515	-	-	-
B2 3601-3669	-	-	3649	3618	3647	-	-	-
A26 3701-3769	-	-	3712	3718	-	-	-	-
A10 3801-3869	-	-	3812	3818	-	-	-	-
A14 3901-3969	-	-	3912	3918	-	3930<	-	-

Table 302 RERS sites found in Human Kappa PK1 GIGS, continued

	SfaNI	SfiCI	HinfI	MluI --> <---	PacIII xx38 xx56 xx62	HpaII xx06 xx52
C12 1-69	37	41	53	55	56	-
C2 101-169	137	141	153	155	156	-
C18 201-269	237	241	253	255	256	-

	SfaNI	SfcFI	HinfI	MlyI --> ---> <---	MacIII Tsp45I same sites	HpbI xx38 xx56 xx62	HpaII MspI xx06 xx52
O8	301-369	341	353	353	355	356	-
A20	401-469	441	453	453	455	456	-
A30	501-569	541	553	553	555	556	-
L14	601-669	641	653	653	655	656	-
L1	701-769	741	753	753	755	756	-
L15	801-869	841	853	853	855	856	-
L4	901-969	941	953	953	955	956	-
L18	1001-1069	1041	1053	1053	1055	1056	-
L5	1101-1169	1141	1153	1153	1155	1156	-
L19	1201-1269	1241	1253	1253	1255	1256	-
L9	1301-1369	1341	1353	1353	1355	1356	-
L23	1401-1469	1441	1453	1453	1455	1456	1406
L9	1501-1569	1541	1553	1553	1555	1556	1506
L24	1601-1669	1641	1653	1653	1655	1656	-
L11	1701-1769	1741	1753	1753	1755	1756	-
L12	1801-1869	1841	1853	1853	1855	1856	-
VIG							
O11	1901-1969	-	1918	1918	1937	1930	1952
O1	2001-2069	-	2019	2019	2037	2038	2052
A17	2101-2169	-	2112	2112	2137	2138	2152
A1	2201-2269	-	2212	2212	2237	2238	2252

	SPART	SECT	Minfi	Maxfi --> --<	Maelll Tsp45T same sites	Hpft xx30 xx56 xx62	Hjaff NspT xx06 xx52
A18	2301-2369	-	2318	2318	2337	2336	2352
A2	2401-2469	-	2418	2418	2437	2438	2452
A19	2501-2569	-	2512	2512	2537	2538	2552
A3	2601-2669	-	2612	2612	2637	2638	2652
A23	2701-2769	-	2718	2718	2737	2731* 2738*	-
A27	2801-2869	-	-	-	-	-	-
A11	2901-2969	-	-	-	-	-	-
L2	3001-3069	-	-	-	-	-	-
L16	3101-3169	-	-	-	-	-	-
L6	3201-3269	-	-	-	-	-	-
L20	3301-3369	-	-	-	-	-	-
L25	3401-3469	-	-	-	-	-	-
R3	3501-3569	-	3525	3525	-	-	-
B2	3601-3669	-	3639	3639	-	-	-
A26	3701-3769	-	3712	3739	3737 3755	3756 3752	-
A10	3801-3869	-	3812	3839	3837 3855	3856 3852	-
A14	3901-3969	-	3939	3939	3937 3955	3956 3952	-

MISSING AT THE TIME OF PUBLICATION

Table 302 REBS sites found in Human Kappa FRL, continued

	BsaJI xx29 xx42 xx43	BssKI (NstEHI) xx22 xx30 xx43	EpmI xx20 xx41 xx44 --> --> <-- --> <--	DsrFI CacOI HaeI XgoMI V	HaeII E	Tsp509I
012	7-69	-	-	-	-	-
02	101-169	-	-	-	-	-
018	201-269	-	-	-	-	-
08	301-389	-	-	-	-	-
A20	401-469	-	-	-	-	-
A30	501-569	-	-	-	-	-
T14	601-669	-	-	-	-	-
L1	701-769	-	-	-	-	-
L15	801-869	-	-	-	-	-
L4	901-969	-	-	-	-	-
L18	1021-1069	-	-	-	-	-
L5	1101-1169	-	-	-	-	-
L19	1201-1269	-	-	-	-	-
L8	1301-1369	-	-	-	-	-
L23	1401-1469	-	-	-	-	-
L9	1501-1569	-	-	-	-	-
L24	1601-1669	-	-	-	-	-

	Bsaji xx29 xx42 xx43	Bs5KI (Hs5NI) xx22 xx30 xx43	HpmI xx20 xx41 xx44 --> --> <--	Bs5FI CacBL HacI Mg2OH V	HacII I	Tsp509I
L11 1701-1769	-	-	-	-	-	-
L12 1801-1869	-	-	-	-	-	-
3901						
O11 1901-1969	1942	1943	1944	1951	1954	-
O1 2001-2069	2042	2043	2044	2051	2054	-
A17 2101-2169	2142	-	-	2151	2154	-
A1 2201-2269	2242	-	-	2251	2254	-
A18 2301-2369	2342	2343	-	2351	2354	-
A2 2401-2469	2442	2443	-	2451	2454	-
A19 2501-2569	2542	2543	2544	2551	2554	-
A3 2601-2669	2642	2643	2644	2651	2654	-
A23 2701-2769	2742	-	-	2751	2754	-
3911						
A27 2801-2869	2843	2822	2820 2841	-	-	2803
A11 2901-2969	2943	2943	2920 2941	-	-	2903
I2 3001-3069	3043	3043	3041	-	-	-
L16 3101-3169	3143	3143	3120 3141	-	-	-
I6 3201-3269	3243	3243	3220 3241	-	-	3203
L20 3301-3369	3343	3343	3320 3341	-	-	3303

	BsaJI xx28 xx42 xx43	BssKI (NstUNT) xx22 xx36 xx43	EpaI xx20 xx41 xx44 --> --> <--	BsrFI CacRI HaeI NcoMI V	HaeII Y	Tsp509I
L25 3401-3469	3443	3443	3420 3441	-	-	3403
B3 3501-3569	3519	3530	3520	-	3554	
B2 3601-3669		3613	3620 3641	-	-	
A26 3701-3769			3720	-	-	3703
A10 3801-3869			3820	-	-	3803
A14 3901-3969	3943	3943	3920 3941	-	-	-

Table 400 Lambda FRI CLC sequences

! VL1

CAG TCT GTG CTG ACT CAG CCA CCC TCG GTG TCT GAA
 GCC CCC AGG CAG AGG GTC ACC ATC TCC TGT ! 1a
 cag tct gtg ctg act cag cca ccc tcA gtg tct gGG
 gcc cCA Ggg cag agg gtc acc atc tcc tgc ! 1e
 cag tct gtg ctg act cag cca ccc tcA gCg tcc gGG
 Acc ccc Ggg cag agg gtc acc atc tcT tgt ! 1c
 cag tct gtg ctg act cag cca ccc tcA gCg tct gGG
 Acc ccc Ggg cag agg gtc acc atc tcT tgt ! 1g
 cag tct gtg Ttg acG cag ccG ccc tcA gtg tct gCG
 gcc cCA GgA cag aAg gtc acc atc tcc tgc ! 1b

! VL2

CAG TCT GCC CTG ACT CAG CCT CCC TCC GCG TCC GGG
 TCT CCT GGA CAG TCA GTC ACC ATC TCC TGC ! 2c
 cag tct gcc ctg act cag cct ccG tcA gTg tcc ggg
 tct cct gga cag tcA gtc acc atc tcc tgc ! 2e
 cag tct gcc ctg act cag cct Gcc tcc gTg tcT ggg
 tct cct gga cag tcG Atc acc atc tcc tgc ! 2a2
 cag tct gcc ctg act cag cct ccc tcc gTg tcc ggg
 tct cct gga cag tcA gtc acc atc tcc tgc ! 2d
 cag tct gcc ctg act cag cct Gcc tcc gTg tcT ggg
 tct cct gga cag tcG Atc acc atc tcc tgc ! 2b2

! VL3

TCC TAT GAG CTG ACT CAG CCA CCC TCA GTG TCC GTG
 TCC CCA GGA CAG ACA GCC AGC ATC ACC TGC ! 3r
 tcc tat gag ctg act cag cca cTc tcA gtg tcA gtg
 Gcc cTG gga gag acG gcc agG atT acc tgt ! 3j
 tcc tat gag ctg acA cag cca ccc tcG gtg tcA gtc
 tcc cCA gga caA acG gcc agG atc acc tgc ! 3p
 tcc tat gag ctg acA cag cca ccc tcG gtg tcA gtg
 tcc cTa gga cag aTG gcc agG atc acc tgc ! 3a
 tcT tct gag ctg act cag CAC ccT GcT gtg tcT gtg
 Gcc TTG gga cag aca gTc agG atc acA tgc ! 3l

tcc tat gTg ctg act cag cca ccc tca gTg tca gTg
 gcc cca gga Aag acG gcc agG atT acc tGT ! 3h
 tcc tat gag ctg acA cag cTa ccc tcc gTg tca gTg
 tcc cca gga cag aca gcc agG atc acc tgc ! 3c
 tcc tat gag ctg atG cag cca ccc tcc gTg tca gTg
 tcc cca gga cag acG gcc agG atc acc tgc ! 3m
 tcc tat gag ctg acA cag cca Tcc tcc gTg tca gTg
 tct ccG gga cag aca gcc agG atc acc tgc ! V2-19

! VL4

CTG CCT GTG CTG ACT CAG CCC CCG TCT GCA TCT GCC
 TTC CTG GGA GCC TCG ATC AAG CTC ACC TGC ! 4c
 cAg cct gTg ctg act caA Tca Tcc tcc gcC tct gct
 tCC ctg gga Tcc tcc Gtc aag ctc acc tgc ! 4a
 cAg cTt gTg ctg act caA Tcc ccC tct gcC tct gcc
 tCC ctg gga gcc tcc Gtc aag ctc acc tgc ! 4b

! VL5

CAG CCT GTG CTG ACT CAG CCA CCT TCC TCC TCC GCA
 TCT CCT GGA GAA TCC GCC AGA CTC ACC TGC ! 5a
 cag Gct gTg ctg act cag ccG Gct tcc CTC tct gca
 tct cct gga gCa tca gcc agT ctc acc tgc ! 5c
 cag cct gTg ctg act cag cca Tct tcc CAT tct gca
 tct Tct gga gCa tca gTc aga ctc acc tgc ! 5b

! VL6

AAT TTT ATG CTG ACT CAG CCC CAC TCT GTG TCG GAG
 TCT CCG GGG AAG ACG GTA ACC ATC TCC TGC ! 6a

! VL7

CAG ACT GTG GTG ACT CAG GAG CCC TCA CTG ACT GTG
 TCC CCA GGA GGG ACA GTC ACT CTC ACC TGT ! 7a
 cag Gct gTg gTg act cag gag ccc tca ctg act gTg
 tcc cca gga ggg aca gTc act ctc acc tgt ! 7b

! VL8

CAG ACT GTG GTG ACC CAG GAG CCA TCG TTC TCA GTG
 TCC CCT GGA GGG ACA GTC ACA CTC ACT TGT ! 8a

! VL9

CAG CCT GTG CTG ACT CAG CCA CCT TCT GCA TCA GCC
TCC CTG RGA GCC TCG GTC ACA CTC ACC TGC ! 9a

! VL10

CAG GCA GGG CTG ACT CAG CCA CCC TCG GTG TCC AAG
GGC TTG AGA CAG ACC GCC ACA CTC ACC TGC ! 10a

Table 405 REERSs found in human lambda FRI GLGs

| There are 31 lambda GLGs

```
MlyI NnnnnnGACTC                25
  1: 6   3: 6   4: 6   6: 6   7: 6   8: 6
  9: 6  10: 6  11: 6  12: 6  15: 6  16: 6
 20: 6  21: 6  22: 6  23: 6  23: 50 24: 6
 25: 6  25: 50 26: 6  27: 6  28: 6  30: 6
 31: 6
There are 23 hits at base# 6
```

```
-"- GAGTCNNNNNn                1
 26: 34
```

```
MwoI GCNNNNNnngc                20
  1: 9   2: 9   3: 9   4: 9  11: 9  11: 56
 12: 9  13: 9  14: 9  16: 9  17: 9  18: 9
 19: 9  20: 9  23: 9  24: 9  25: 9  26: 9
 30: 9  31: 9
There are 19 hits at base# 9
```

```
HinfI Gantc                    27
  1: 12   3: 12   4: 12   6: 12   7: 12   8: 12
  9: 12  10: 12  11: 12  12: 12  15: 12  16: 12
 20: 12  21: 12  22: 12  23: 12  23: 46 23: 56
 24: 12  25: 12  25: 56 26: 12  26: 34 27: 12
 28: 12  30: 12  31: 12
There are 23 hits at base# 12
```

```
E1eI gactc                      25
  1: 12   3: 12   4: 12   6: 12   7: 12   8: 12
  9: 12  10: 12  11: 12  12: 12  15: 12  16: 12
 20: 12  21: 12  22: 12  23: 12  23: 56 24: 12
 25: 12  25: 56 26: 12  27: 12  28: 12  30: 12
 31: 12
There are 23 hits at base# 12
```

```
-"- gagtc                        1
 26: 34
```

DdeI Ctnag 32
 1: 14 2: 24 3: 14 3: 24 4: 14 4: 24
 5: 24 6: 14 7: 14 7: 24 8: 14 9: 14
 10: 14 11: 14 11: 24 12: 14 12: 24 15: 3
 15: 14 16: 14 16: 24 19: 24 20: 14 23: 14
 24: 14 25: 14 26: 14 27: 14 28: 14 29: 30
 30: 14 31: 14
 There are 21 hits at base# 14

BsaJI Ccnggg 38
 1: 23 1: 40 2: 39 2: 40 3: 39 3: 40
 4: 39 4: 40 5: 39 11: 39 12: 38 12: 39
 13: 23 13: 39 14: 23 14: 39 15: 38 16: 39
 17: 23 17: 39 18: 23 18: 39 21: 38 21: 39
 22: 47 22: 38 22: 39 22: 47 26: 40 27: 39
 28: 39 29: 14 29: 39 30: 38 30: 39 30: 47
 31: 23 31: 32
 There are 17 hits at base# 39
 There are 5 hits at base# 38
 There are 5 hits at base# 40 Makes cleavage ragged.

MnlI cctc 35
 1: 23 2: 23 3: 23 4: 23 5: 23 6: 19
 6: 23 7: 19 8: 23 9: 19 9: 23 10: 23
 11: 23 13: 23 14: 23 16: 23 17: 23 18: 23
 19: 23 20: 47 21: 23 21: 29 21: 47 22: 23
 22: 29 22: 35 22: 47 23: 26 23: 35 24: 27
 27: 23 28: 23 30: 35 30: 47 31: 23
 There are 21 hits at base# 23
 There are 3 hits at base# 19
 There are 3 hits at base# 29
 There are 1 hits at base# 26
 There are 1 hits at base# 27 These could make cleavage ragged.

-"- gagg 7
 1: 46 2: 46 3: 46 4: 46 27: 44 28: 44

29: 44

```

BssKI Ncngg                39
 1: 40   2: 39   3: 39   3: 40   4: 39   4: 40
 5: 39   6: 31   6: 39   7: 31   7: 39   8: 39
 9: 31   9: 39  10: 39  11: 39  12: 38  12: 52
13: 39  13: 52  14: 52  15: 39  16: 52  17: 39
17: 52  18: 39  18: 52  19: 39  19: 52  21: 38
22: 38  23: 39  24: 39  26: 39  27: 39  28: 39
29: 14  29: 39  30: 38

```

There are 21 hits at base# 39

There are 4 hits at base# 38

There are 3 hits at base# 31

There are 3 hits at base# 40 Ragged

```

BstNI CCwgg                30
 1: 41   2: 40   5: 40   6: 40   7: 40   8: 40
 9: 40  10: 40  11: 40  12: 39  12: 53  13: 40
13: 53  14: 53  16: 40  16: 53  17: 40  17: 53
18: 40  18: 53  19: 53  21: 39  22: 39  23: 40
24: 40  27: 40  28: 40  29: 15  29: 40  30: 39

```

There are 17 hits at base# 40

There are 7 hits at base# 53

There are 4 hits at base# 39

There are 1 hits at base# 41 Ragged

```

PspGI cwgg                30
 1: 41   2: 40   5: 40   6: 40   7: 40   8: 40
 9: 40  10: 40  11: 40  12: 39  12: 53  13: 40
13: 53  14: 53  16: 40  16: 53  17: 40  17: 53
18: 40  18: 53  19: 53  21: 39  22: 39  23: 40
24: 40  27: 40  28: 40  29: 15  29: 40  30: 39

```

There are 17 hits at base# 40

There are 7 hits at base# 53

There are 4 hits at base# 39

There are 1 hits at base# 41

ScrFI CCngg 39

1: 41	2: 40	3: 40	3: 41	4: 40	4: 41
5: 40	6: 32	6: 40	7: 32	7: 40	8: 40
9: 32	9: 40	10: 40	11: 40	12: 39	12: 53
13: 40	13: 53	14: 53	16: 40	16: 53	17: 40
17: 53	18: 40	18: 53	19: 40	19: 53	21: 39
22: 39	23: 40	24: 40	26: 40	27: 40	28: 40
29: 15	29: 40	30: 39			

There are 21 hits at base# 40

There are 4 hits at base# 39

There are 3 hits at base# 41

MaeIII gtnac 16

1: 52	2: 52	3: 52	4: 52	5: 52	6: 52
7: 52	9: 52	26: 52	27: 10	27: 52	28: 10
28: 52	29: 10	29: 52	30: 52		

There are 13 hits at base# 52

Tsp45I gtsac 15

1: 52	2: 52	3: 52	4: 52	5: 52	6: 52
7: 52	9: 52	27: 10	27: 52	28: 10	28: 52
29: 10	29: 52	30: 52			

There are 12 hits at base# 52

HphI tcacc 26

1: 53	2: 53	3: 53	4: 53	5: 53	6: 53
7: 53	8: 53	9: 53	10: 53	11: 59	13: 59
14: 59	17: 59	18: 59	19: 59	20: 59	21: 59
22: 59	23: 59	24: 59	25: 59	27: 59	28: 59
30: 59	31: 59				

There are 16 hits at base# 59

There are 10 hits at base# 53

BspMI ACCTGCNNNNn 14
11: 61 13: 61 14: 61 17: 61 18: 61 19: 61
20: 61 21: 61 22: 61 23: 61 24: 61 25: 61
30: 61 31: 61
There are 14 hits at base# 61 Goes into CDR1

Table 500: h3401-h2 captured Via C5 with BsmAI

```

! 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
! S A Q D I Q M T Q S P A T L S
! aGT GCA Caa gac atc cag atg acc cag tct cca gcc acc ctg tct
! ApaI... a gcc acc !
L25,L6,L20,L2,L16,A11
! Extender.....Bridge...

! 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
! V S P G E R A T L S C R A S Q
! gtg tct cca ggg gaa agg gcc acc ctg tcc tgc agg gcc agt cag

! 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
! S V S N N L A W Y Q Q K P G Q
! agt gtt agt aac aac tta gcc tgg tac cag cag aaa oct ggc cag

! 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
! V P R L L I Y G A S T R A T D
! gtt ccc agg ctg ctg atc tat ggt gca tcc acc agg gcc act gat

! 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
! I P A R F S G S G S G T D F T
! atc cca gcc agg ttc agt gcc agt ggg tct ggg aca gac ttc act

! 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
! L T I S R L E P E D F A V Y Y
! ctg acc atc agc aga ctg gag cct gaa gat ttt gca ggg tat tac

! 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
! C Q R Y G S S P G W T F G Q G
! tgt cag cgg tat ggt agc tca ccg ggg tgg acg ttc ggc caa ggg

! 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
! T K V E I K R T V A A P S Y F
! acc aag gtg gaa atc aaa cga act ctg gct gca cca tct gtc ttc

! 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
! I F P P S D E Q L K S G T A S
! atc ttc ccg cca tct gat gag cag ttg aaa tct gga act gcc tct

! 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
! V V C L L N N F Y P R E A K V
! gtt gtg tgc ctg ctg aat aac ttc tat ccc aga gag gcc aaa gta

```

```

! 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
! Q W K V D N A L Q S G N S Q E
  cag tgg aag gtg gat aac gcc ctc caa tog ggt aac tcc cag gag

! 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
! S V T E Q D S K D S T Y S L S
  agt gtc aca gag cag gac agc aag gac agc acc tac agc ctc agc

! 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
! S F L T L S K A D Y E K H K V
  agc acc ctg acg ctg agc aaa gca gac tac gag aaa cac aaa gtc

! 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
! Y A C E V T H Q G L S S P V T
  tac gcc tgc gaa gtc acc cat cag gcc ctg agc tog cct gtc aca

! 211 212 213 214 215 216 217 218 219 220 221 222 223
! K S F N K G E C K G E F A
  aag agc ttc aac aaa gga gag tgc aag gcc gaa ttc gc.....

```

Table 501: h3401-d8 KAPPA captured with CU and BsmAI

```

! 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
! S A Q D I Q M T Q S P A T L S
  aGT GCA Caa gac atc cag atg acc cag tct cct gcc acc ctg tct
! ApaI...Extender.....a gcc acc !
L25,L6,L20,L2,L16,A11
: A GCC ACC CTG TCT ! L2

! 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
! V S P G E R A T L S C R A S Q
  gtg tct cca ggt gaa aga gcc acc ctc tcc tgc agg gcc agt cag
! GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC ! L2

! 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
! N L L S N L A W Y Q Q K P G Q
  aat ctt ctc agc aac tta gcc tgg tac cag cag aaa cct gcc cag

! 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
! A P R L L I Y G A S T G A I G
  gct ccc agg ctc ctc atc tat ggt gct tcc acc ggg gcc att ggt

! 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
! I P A R F S G S G S G T E F T
  atc cca gcc agg ttc agt gcc agt ggg tct ggg aca gag ttc act

```

```

! 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
! L T I S S L Q S E D F A V Y F
! ctg acc atc agc agc ctg cag tct gaa gat ttt gca gtg tat ttc

! 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
! C Q Q Y G T S P P T F G G G T
! tgt cag cag tat ggt acc tca ccg ccc act ttc ggc gga ggg acc

! 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
! K V E I K R T V A A P S V F I
! aag gtg gag atc aaa cga act gtg gct gca cca tct gtc ttc atc

! 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
! F P P S D E Q L K S G T A S V
! ttc ccg cca tct gat gag cag ttg aaa tct gga act gcc tct gtt

! 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
! V C P L N N F Y P R E A K V Q
! gtg tgc ccg ctg aat aac ttc tat ccc aga gag gcc aaa gta cag

! 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
! W K V D N A L Q S G N S Q E S
! tgg aag gtg gat aac gcc ctg caa tcg ggt aac tcc cag gag agt

! 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
! V T E Q D N K D S T Y S L S S
! gtc aca gag cag gac aac aag gac agc acc tac agc ctg agc agc

! 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
! T L T L S K V D Y E K H E V Y
! acc ctg acg ctg agc aaa gta gac tac gag aaa cac gaa gtc tac

! 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
! A C E V T E Q G L S S P V T K
! gcc tgc gaa gtc acc cat cag ggc stt agc tcg ccc gtc acc aag

! 211 212 213 214 215 216 217 218 219 220 221 222 223
! S F N R G E C K K E F V
! agc ttc aac agg gga gag tgt aag aaa gaa ttc gtt t

```


{TCT}ZCA|gac|aac|tct|aag|laa|lact|ctc|tact|ttg|cag|atg|{-
 |aac|goc|TWA|Aag|gct|gag|gac|act|CCA|etc|aac|tca|tct|tct|Aag ag-3'
 (VH881P8) 5'-cgcttcacTaaG|TCT|ACA|gac|aac -3'

5 Table 512: kappa, bases 12-30

ID	Start	1	2	3	4	5	6	Name	Sequence	Dot Form
1	84	40	21	20	1	2	0	SK12012	gaccagttctccatctcttc	gaccagttctccatctcttc
2	32	19	3	6	2	1	0	SK12A17	gactcagttctccatctcttc	...L.....ct....
3	26	17	8	1	0	0	0	SK12A27	gaccagttctccagtcacc	...g.....gg.a...
4	49	21	18	1	0	0	0	SK12A11	gaccagttctccagtcacc	...g.....g.a...
182	97	50	28	3	3	0	1			
97	147	175	178	181	181	182				
URE adapters:										
(SZKB1230-Q12)									Stem..... Loop. Stem..... Recognition.....	
									5'-cAcATccgTg TTTgTt cAcgGATgTg gGAggATgGAgAcTggGTC-3'	
									[RC] 5'-gaccagttctccatctcttc cAcATccgTg AACAA cAcgGATgTg-3'	
									Recognition..... Stem..... Loop. Stem.....	
									FokI.	
(SZKB1230-A17)									Stem..... Loop. Stem..... Recognition.....	
									5'-cAcATccgTg TTTgTt cAcgGATgTg gGAggATgGAgAcTggGTC-3'	
									[RC] 5'-gactcagttctccatctcttc cAcATccgTg AACAA cAcgGATgTg-3'	
									Recognition..... Stem..... Loop. Stem.....	
									FokI.	
(SZKB1230-A27)									Stem..... Loop. Stem..... Recognition.....	
									5'-cAcATccgTg TTTgTt cAcgGATgTg gGAggATgGAgAcTggGTC-3'	
									[RC] 5'-gaccagttctccagtcacc cAcATccgTg AACAA cAcgGATgTg-3'	
									Recognition..... Stem..... Loop. Stem.....	
									FokI.	
(SZKB1230-A11)									Stem..... Loop. Stem..... Recognition.....	
									5'-cAcATccgTg TTTgTt cAcgGATgTg gGAggATgGAgAcTggGTC-3'	
									[RC] 5'-gaccagttctccagtcacc cAcATccgTg AACAA cAcgGATgTg-3'	
									Recognition..... Stem..... Loop. Stem.....	
									FokI.	

What happens in the upper strand:

```

152KB1230-012*)      5'-gac caa gtctcc a-ct ctc c-3'
|
|      | Site of cleavage in substrate
5 |-----|
|
152KB1230-A17*)      5'-gac tca gtctcc a-ct ctc c-3'
|
|
152KB1230-A27*)      5'-gac gca gtctcc a-gg cac c-3'
|
|
152KB1230-A11*)      5'-gac gca gtctcc a-gc cac c-3'
|
|
(kapextURE)          5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg-3'  |sense strand
Scab.....ApalI.
75 (kapextURUPUR)    5'-ccTctactctTgTcAcAgTg-3'
Scab.....
(kabRO1UR)           5'-ggAggATgG cTggAggtc TgtgcncTgt gAcAAgAgTA gAgg-3'
|
| [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-ct ctc c-3'  ON above is R.C. of this one
(kabRO2UR)           5'-ggAggATgG cTggAggtc TgtgcncTgt gAcAAgAgTA gAgg-3'
|
| [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-ct ctc c-3'  ON above is R.C. of this one
(kabRO3UR)           5'-ggTgcctggA cTggAggtc TgtgcncTgt gAcAAgAgTA gAgg-3'
|
| [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gg cac c-3'  ON above is R.C. of this one
(kabRO4UR)           5'-ggTgcctggA cTggAggtc TgtgcncTgt gAcAAgAgTA gAgg-3'
|
| [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gc cac c-3'  ON above is R.C. of this one
Scab.....ApalI.

```

(VH81PCP) 5'-cgcttcactcaag|TCT|AGA|gac|aac -3'
 |TCCT|AGA|gac|aac|TCT|aac|aat|aac|act|ctc|tac|ctg|cag|atg|~
 |aac|agc|TTA|AG|gcc|gag|gac|ctc|TCA|GCA|Gtc|tac|tat|tgt|Acg|ag-3'

Table 512: Kappa, bases 12-30

ID	Ntot	0	1	2	3	4	5	6	Name	Sequence	Dot Form
1	84	40	21	20	1	2	0	0	SK12012	gaccagctctccatctccc	gaccagctctccatctccc
2	32	19	3	6	2	1	0	1	SK12A17	gacccagctctccactctccc	...b.....ct.....
3	26	17	8	1	0	0	0	0	SK12A27	gacccagctctccaggcacc	...g.....gg.a..
4	40	21	18	1	0	0	0	0	SK12A11	gacccagctctccaggcacc	...g.....g.a..
182	97	50	28	3	3	0	1				
97	147	175	178	181	181	182					

IRE adapters:

(SZKB1230-012) Stem..... Loop. Stem..... Recognition.....
 5' cAcATccgTg TgTt cAcgATgTg gAggATgAgAcTggTc-3'
 [RC] 5'-gaccagctctccatctccc cAcATccgTg AAcAA cAcgATgTg-3'
 Recognition..... Stem..... Loop. Stem.....
 FokI.

(SZKB1230-A17) Stem..... Loop. Stem..... Recognition.....
 5'-cAcATccgTg TgTt cAcgATgTg gAggATgAgAcTgATc-3'
 [RC] 5'-gaccagctctccatctccc cAcATccgTg AAcAA cAcgATgTg-3'
 Recognition..... Stem..... Loop. Stem.....
 FokI.

(SZKB1230-A27) Stem..... Loop. Stem..... Recognition.....
 5'-cAcATccgTg TgTt cAcgATgTg gTgctTgAgAcTggTc-3'
 [RC] 5'-gaccagctctccaggcacc cAcATccgTg AAcAA cAcgATgTg-3'
 Recognition..... Stem..... Loop. Stem.....
 FokI.

(SZKB1230-A11) Stem..... Loop. Stem..... Recognition.....
 5'-cAcATccgTg TgTt cAcgATgTg gTgctTgAgAcTggTc-3'
 [RC] 5'-gaccagctctccaggcacc cAcATccgTg AAcAA cAcgATgTg-3'
 Recognition..... Stem..... Loop. Stem.....
 FokI.

What happens in the upper strand:

```

(SZRE1230-O12*)      5'-gac cca gtc|ccc a-tc ctc c-3'
|                    | Site of cleavage in substrate
(SZRH1230-A17*)      5'-gac tca gtc|ccc a-ct ctc c-3'
|
(SZRH1230-A27*)      5'-gac gca gtc|ccc a-gg cac c-3'
|
(SZRH1230-A11*)      5'-gac gca gtc|ccc a-gc cac c-3'
|
(kapextURE)          5' -ccTctactctTgTcAcAgTgcAGAA gAc ATc cAg-3' (sense strand
Scab.....ApalI.
(kapextUREPCR)       5' -ccTctactctTgTcAcAgTg-3'
Scab.....
(kaBRO1UR)           5' -ggAggATgca cTggATgTct TgTgcAcTgtT gAcAAgAgTA gAgg-3'
| [RC] 5' -ccTctactctTgTcAcAgTgcAGAA gAc ATc cAg tcc a-tc ctc c-3' ON above is R.C. of this one
(kaBRO2UR)           5' -ggAggTgga cTggATgTct TgTgcAcTgtT gAcAAgAgTA gAgg-3'
| [RC] 5' -ccTctactctTgTcAcAgTgcAGAA gAc ATc cAg tcc a-ct ctc c-3' ON above is R.C. of this one
(kaBRO3UR)           5' -ggTgcTgga cTggATgTct TgTgcAcTgtT gAcAAgAgTA gAgg-3'
| [RC] 5' -ccTctactctTgTcAcAgTgcAGAA gAc ATc cAg tcc a-gg cac c-3' ON above is R.C. of this one
(kaBRO4UR)           5' -ggTggtgga cTggATgTct TgTgcAcTgtT gAcAAgAgTA gAgg-3'
| [RC] 5' -ccTctactctTgTcAcAgTgcAGAA gAc ATc cAg tcc a-gc cac c-3' ON above is R.C. of this one
| Scab.....ApalI.

```

Table 515 Lambda URE adapters bases 13.3 to 19.3

Id	MFEQ	Number of mismatches								Name	Sequence	Dot form
		0	1	2	3	4	5	6	7			
1	58	7	1	0	0	2	2	1		VL133-2a2	gtctcttgacacgtcgatc	gtctcttgacacgtcgatc
2	16	10	0	1	0	1	1	0	2	VL133-31	ggctcttgacacacagtc	g.cttg.....a.ag..
3	17	6	0	0	4	1	1	5	0	VL133-2c	gtctcttgacacgtcgatcag..
4	37	3	0	10	4	4	3	7	4	VL133-1c	ggccccgggacaggggtc	.g.c.a..g...ag.g..
128	64	8	11	5	8	5	11	11	5			
	64	72	83	98	96	101	112	123	128			
(VL133-2a2)		Stem..... loop. Stem..... Recognition.....										
		5'-cAcAtccgTg TTgTT cAcgATgTg gATcATcTccAAGAc-3'										
[RC]		5'-gtctcttgacacgtcgatc cAcATccgTg AAcAA cAcgATgTg-3'										
		Recognition..... Stem..... Loop. Stem.....										
(VL133-31)		Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gATcATcTccAAGAc-3'										
[RC]		5'-ggctcttgacacacagtc cAcATccgTg AAcAA cAcgATgTg-3'										
		Recognition..... Stem..... Loop. Stem.....										
(VL133-2c)		Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gATcATcTccAAGAc-3'										
[RC]		5'-gtctcttgacacgtcgatc cAcATccgTg AAcAA cAcgATgTg-3'										
		Recognition..... Stem..... Loop. Stem.....										
(VL133-1c)		Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gATcATcTccAAGAc-3'										
[RC]		5'-ggccccgggacaggggtc cAcATccgTg AAcAA cAcgATgTg-3'										

What happens in the top strand:

```

!
!           ] site of cleavage in the upper strand
(VL133-2a2*) 5'-g tct cct g lga cag tcg atc
(VL133-3l*)  5'-g gcc ttg g lga cag aca gtc
(VL133-2c*)  5'-g tct cct g lga cag tca gtc
(VL133-1c*)  5'-g gcc cca g lgg cag agg gtc
!
! The following Extenders and Bridges all encode the AA sequence of 2a2 for
! codons 1-15
!
!           1
(ON_LamEx133) 5'-ccTcTgAcTgAgT gcA cAg -
!
!           2 3 4 5 6 7 8 9 10 11 12
!           AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-
!
!           13 14 15
!           tcC ccG g ! 2a2
!
!           1
(ON_LamB1-133) [RC] 5'-ccTcTgAcTgAgT gcA cAg -
!
!           2 3 4 5 6 7 8 9 10 11 12
!           AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-
!
!           13 14 15
!           tcC ccG g ga cag tcg at-3' ! 2a2 N.B. the actual seq is the
!           reverse complement of the
!           one shown.
(ON_LamB2-133) [RC] 5'-ccTcTgAcTgAgT gcA cAg -
!
!           2 3 4 5 6 7 8 9 10 11 12
!           AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-
!
!           13 14 15
!           tcC ccG g ga cag aca gt-3' ! 3l N.B. the actual seq is the
!           reverse complement of the
!           one shown.
(ON_LamB3-133) [RC] 5'-ccTcTgAcTgAgT gcA cAg -
!
!           2 3 4 5 6 7 8 9 10 11 12
!           AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-
!
!           13 14 15
!           tcC ccG g ga cag tca gt -3' ! 2c N.B. the actual seq is the
!           reverse complement of the
!           one shown.
(ON_LamB4-133) [RC] 5'-ccTcTgAcTgAgT gcA cAg -
!

```

```

!           2  3  4  5  6  7  8  9  10 11 12
!          AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-
!
!           13 14 15
!          tcC ccG g gg cag agg gt-3' ! lc N.B. the actual seq is the
!          reverse complement of the
!          one shown.
!
(OH_Lam133PCR) 5'-ccTcTgAcTgAgT gca cAg AGT gc-3'

```


Table 52's ONs used in Capture of kappa light chains using CJ method and *RvxAI*

All ONs are written 5' to 3'.

REaptors (6)
 ON_20SK15012 gggAggATggAgAcTgggTc
 ON_20SK15112 gggAgATggAgAcTgggTc
 ON_20SK15A17 gggAgATggAgAcTgAgTc
 ON_20SK15A27 gggTgccTggAgAcTgggTc
 ON_20SK15A11 gggTggcTggAgAcTgggTc
 ON_20SK15B3 gggAgTcTggAgAcTgggTc

Bridges (6)
 kapbr11012 gggAggATggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg
 kapbr11112 gggAgATggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg
 kapbr11A17 gggAgATggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg
 kapbr11A27 gggTgccTggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg
 kapbr11A11 gggTggcTggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg
 kapbr11B3 gggAgTcTggAgAcTgggTcATcTggATgTcTtTgAcTgTgAcAgAgg

Extender (5' biotinylated)
 kapext1bio ccTcTgTcAcAgTgcAcAagAcATccAgATgAcccAgTcTcc

Primers
 kapPCRfl ccTcTgTcAcAgTgcAcAagAc
 kapfor 5'-aca ctc tcc cct gLL gaa gct ctt-3'

Table 530
 PCR program for amplification of kappa DNA
 95°C 5 minutes
 95°C 15 seconds
 65°C 30 seconds

72°C 1 minute
72°C 7 minutes
4°C hold

Reagents (100 ul reaction):

Template 50 ng
10x turbo PCR buffer 1x
turbo Pfi 4U
dNTPs 200 µM each
kapCRT1 300 nM
kapfor 300 nM

Table 610: Staffer used in VII

1 TCCGGAGTTC CAGATCTGTT TCCCTTTTIG TGGGGTGGTG CAGATCSOGI TALEGAGATC
 61 GACCCNCIQC TWCASCAAAA GCCACGCTTA ACTGICIGATC AGGCAGSSGA TGTATTGSC
 121 CAWAGCAATC GTCAGBAICT TAACTCAGAG CTTTTIITAC CTACTCTSCA AGCAGGGACA
 181 TCTGGTTTGA CACAGAGCGA TCCCGCTCCP CAGTTGGTAG AATCATINAC ACCTTSGGAT
 241 GGCATPAAIT TGCYIAAIGA YGAFGGHAAA ACCGTGGCAGC AGCCAGGCTC TGCYIUCGIG
 301 NACGTTTGGC TGAACCACTAT GTTCAAGCCP ACGGTBRTGG CTGCGGUAAC TADGCCATII
 361 GATAAGTGGT ACRAGCGGAG TGGCTACGAA ACAAACCDAGG ACGGCCAAC TGGTTCGGTG
 421 ANTATAACTG TTGGAGGAAA AATTTTGTAT GAGGCGRTGC AGGGAGACA ATCACCAATC
 481 CCACAGGGGG TTAGCTGTT TGCYGGAAA CCACTAGTAG AGGTTGTFTT GGTGCGCTG
 541 GAAGAPACTE GGGAGACTCT TTCCAAACGC TATGGCAATA NITGGATAA CTGGAAACA
 601 CCTGCAATGG CCTTACGFT CCGGCAAT ATTTCTTIG GRTACCGGA GCGGCAAGG
 661 GNAGAAAGC GTUATCAGC GAGUATCAA AAGCGTGGAA CAGAAACGA TATGATGTI
 721 TPTCAACCAA CBAACAGGA TCGTCCCTGTG CTTCCCTGGG ATGTGRTGC ACCGGTCAG
 781 AGTGGGUA TTTCTGCGGA TGGAAAGTIT GAAAGCTCT ALGAAAGTCA GCGAAATAG
 841 TACGAAATTT TTGGCCGTAA CTCCCTCTGG YTAACGAGAC AGGATGAGA GGGCAATAG
 901 GAGTGGTCTA GA

Table 620: DNA sequence of pCES5
 pCES5 6880 bases ~ pCes4 with stuffers in CDR1-2 and CDR3 2000.12.13

```

1 Gene n 6880
1 Useful REs (cut MaxcII fewer than 3 times) 2000.06.05
1
1 Non-cutters
1 Acc65I Gctacc MfeI AGCgct AvrII Cctagg
1 SmaI GAPPNmatc BstXI Cgttag RseFI Mmmmmmmmmmmmmmmmmcc
1 BspI Tgtaac BstAPI GCANNNmngc BstXI Ttgaaa
1 BclII GTATac BstI CACgtg Ecl136I GAGctc
1 EcoRV GATatc PaeI GCGGgac KpnI GGTACC
1 SmaI TGGcca HspI TCGgga NsiI ATGcAt
1 PacI TTAATtae PseI GTTaaac PstI CACgtg
1 PvuII Rgnccy PshAI GACMhngtc ScaI GAGctc
1 SclI CCGCgg SbfI CTGCcAgg SmaI AccwggT
1 SfiI GCGVgac SmaI TACgta SpeI Actagt
1 SphI GATgc Sse8387I CTGCcAgg SstI ASgact
1 XbaI ATTTaat XbaI Cccggg
1
1 cutters
1
1 Enzymes that cut more than 3 times.
1 ALuNI CAGNNMctg 5
1 BspI ctgac 4
1 BstFI RCGgty 5
1 BstI CTCTCKmm 4
1 BstI RNNNNNGGg 10
1
1 Enzymes that cut from 1 to 3 times.
1
1 Eco109I Rgnccy 3 7 2636 4208
1 BssI Cctgtg 1 12
1 SmaI Cccgag 1 1703
1 BspHI TcaTga 3 43 148 1156
1 BstII GAGctc 1 65
1 BclVI GTATCCNNMNN 2 140 1667
1 Eco57I CTGAG 1 301
1 SmaI Ccttag 2 1349
1 XbaI Ccggg 3 319 2347 6137
1 BstIHZI GAGcwc 3 401 2321 4245
1 HpaI GAGcwc 3 401 2321 4245
1 BspI Gcannmmatcg 1 451
1 ScaI ASfract 1 506

```

!EvuI CGAtcg	3	616	3538	5926
!EpiI TGcGca	2	763	5946	
!EqlI GCCNNHhggc	3	864	2771	5952
!EpmI cTGGAG	1	898		
!E- cTccag	1	443		
!EsaI GcYcTCNnna	1	416		
!EtdI GAcTNNhagtc	1	903		
!EamI bSI GNCNNhngtc	1	903		
!ErdI GAcNNhngtc	3	1768	6197	6579
!EapiI gaagagc	1	1598		
!EpuI CAGctg	3	2054	3689	5996
!EpmI CCANNHhbgg	3	2293	3943	3991
!EindI I Aagctt	1	2235		
!EpaI I Gtcac	1	2321		
!EpmI Nmmmmhngcagct	1	2328		
!E- ACCGCGNNh	2	3460		
!EatiI cTGGAG	1	2335		
!EccI GfMkac	2	2341	2611	
!EincI I GfYfAc	2	2341	3740	
!EaliI Gtcgac	1	2341		
!EhiI Gtcgag	1	2347		
!EholI cTcgag	1	2347		
!EbeiI gctctc	2	2383	4219	
!EipiI cctnagc	1	2580		
!EepI cctnagc	1	2580		
!EgriI cGcggYg	1	2648		
!EgaiI Acaggt	2	2649	4302	
!EacI Gcgcgcc	1	2689		
!EashI Gcgcgc	1	2690		
!EfiI GCCNNHhggcc	1	2770	6349	
!EaelI cccgcc	2	2776	6349	
!NgomI V Gcgggc	2	2776	6349	
!EgtI CcYgg	3	2781	3553	5712
!EaalI CcYgg	3	2781	3553	5712
!EcofI Gcaltg	1	2781		
!EtyI Cawcgg	3	2781	4205	4472
!EfaI Ccatty	1	2795		
!EepI I Tcggca	1	2861		
!EgLI I gYctct	1	2872		
!EaliI Tgalea	1	2896		
!EsaI GcCnagc	3	3004	4143	4973
!EamI CCANNHhngtcg	1	3215		
!EduI Acagct	1	3527		

|PpaI GTTaac | 1 | 3730
 |KpaI Tctaga | 1 | 3757
 |
 |AflII Cttaaq | 1 | 3811
 |BsuI Hccatc | 1 | 3821
 |" " GAATcN | 1 | 4695
 |BstII Cggacc | 1 | 3827
 |NheI Cctaga | 1 | 4166
 |BspRI Cgtaac | 1 | 4182
 |BsmBI CctcNmnn | 2 | 4188 6625
 |" " Nnnnnngagc | 1 | 6673
 |PpaI Ggcgc | 1 | 4209
 |BamI Ggcgc | 3 | 4209 4482 6319
 |BspI20I Gggccc | 1 | 4209
 |PspOMI Gggccc | 1 | 4209
 |BseRI Hnnnnnnnctctc | 1 | 4226
 |" " GAGGAGGRRRRRRN | 1 | 4957
 |BcoNI CctNnnnnnagc | 1 | 4278
 |PflPI GAcNagtc | 1 | 4300
 |T6bIII GAcNagtc | 1 | 4308
 |KasI Ggcgc | 2 | 4327 5967
 |BstXI CCAMNNNngg | 1 | 5415
 |NotI Gcgccgc | 1 | 4507
 |EagI Gggcc | 1 | 4508
 |BamHI Ggatac | 1 | 5169
 |BspHI Acgat | 1 | 5776
 |NdeI Gatacg | 1 | 5672
 |EcoRI Gaatcc | 1 | 5806
 |PstI TTAaac | 1 | 6118
 |DraIII CAGNngtc | 1 | 6243
 |BsaAI YAcgLe | 1 | 6246
 |-----
 | 1 | yccgaagag cctcctgata cgcctattct tatagtttaa tgcctatgata ataatggctt
 | Bsst. (1/2)
 | 61 | cttaacGCTc agtgcgacct tttcggggaa atgtycgygg aacctcatt tgtttattt
 | AaIII.
 | 121 | tctaaataca ttcataatag TAUCGctca tggagacaata accttgataa atctctcaat
 | BcIV. (1 of 2)
 | 187 | aatattgaaa gaggaagadt
 | Base # 201 to 1061 = Apr gene from pUC119 with some Bb sites removed
 |
 | 1 | 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
 | | CM S I Q K F R V A L I F F F A

```

201 aty agt att caa cat ttc cgt gtc gcc ehl. att. ccc ttt ttt ggg
16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
A T C L E Y A E E F B T L Y K
246 gaa ttt tgc att cct gtt ttt gct cac caa gaa acg ctg gty aaa
31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
V K D A E D Q L G A R V G Y I
291 gla aaa gat gct gaa gat cag ttg ggt gcc cya gty ggt tac aac
46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
E L E L N S G K I L E S F R P
336 gaa ctg gat ctg aac agc dgt aag atc ctt gey agt ttt cgc ccc
61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
E S R F P M M S T F K V L L C
381 gaa gaa cgt ttt cca aty atg agc act tll. aaa gtt ctg cta tgt
76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
G A V L S R I D A G Q R Q L G
926 ggc ggy gba tta tcc cgt att gac gcc ggg caa gag CAa ctg ggt
91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
R R I H Y S Q N D L V K Y S F
471 cgc cgc ala cac tat tct cag aat gac ttg gll cag TAc Tca caa
106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
V T E K B L T D G M T V R E L
516 gtc aca gaa aag cat ctt acg gat gcc atg aca gta aga gaa tta
121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
C S A A I T M S D N T A A N L
561 tgc agt gct gcc ata acc atg agt gat aac act gcg gcc aac tta
136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
b L T Y I G C F K E L T A F E
606 ctt ctg aca acg ATC Gga Gga cgg aag gag cta acc gct ttt tly
151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
H N M G D H V T R I. D R W E F

```

RegI.....
ScaI.....

```

651   cac aac atg gag gat cat gta act cgc att gat cgt tgg gna ccg
|
|
166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
| E L N E A I P M D E R D T F T M
| gag ctg aal gaa gcc ata cca aac gcc gag cgt gcc acc acg atg
|
|
181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
| P V A H A T L R K L L T G E
| cct gta GCA AUC gca aca acg TTC CAC Aaa ata tta act ggc gaa
|   BSRDT..(1/2)   FSEI.... (1/2)
|
|
196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
| L L T L A S R Q Q L I D W M E
| ata ctt act cta gct tcc cgg caa caa tta ata gac tgg atg gag
|
|
211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
| A D K V A G P L L R S A L P A
| gcg gat aaa gtt gaa cca ctt ctg cgc tcc ggc att ccg gct
|
|
226 227 228 229 230 231 232 233 234 235 236 237 238 239 240
| G W F I A D K S G A G E R G S
| ggc tgg ttt att gct gat aat tCT CCA Gcc ggt gag cgt ggc TCT
|   BpaI....(1/2)
|
|
241 242 243 244 245 246 247 248 249 250 251 252 253 254 255
| R G I I A A I G P D G K F S R
| Ggc ggt abc ATY CCA gca ctg ggg cca gat ggt aag ccc tcc cgt
|   BsrBI....(2/2)
|
|
256 257 258 259 260 261 262 263 264 265 266 267 268 269 270
| E V V I I Y T G S Q A T M D E
| atc gta gtt atc tac aag TCG ggg aGT Cag gca act atg gat gaa
|   NdeI.....
|
|
271 272 273 274 275 276 277 278 279 280 281 282 283 284 285
| R N R Q I A E T G A S L F K H
| cga aat aag cag atc gct gag ata ggt gcc tca ctg att aag cat
|
|
286 287
| W .
| tgg taa
|
1056
|
1062
|
1081  catatatact ttgatttgt ttaaaacttc atttttaatt taaaaggatc taggtgaga
|   ctgtcaaac caagttact

```



```

1141  tcccttttga taatctctatg accaaatccc cttaccctga gttttctctc caactggcgt
1201  cadaccctgt aaaaagatc aaagatctt cttgagatccc ttttttcttg cggtaactct
1267  gctgcttgcg acaaaaaaa ccaaccgtac cagcgglygl ztgyttgcag gataaagagc
1321  taccacctct tttctcgag gtaactggct tcaagcagtc cgtgatacca aatactgctc
1381  tcttagtga ccccttagtta ggcaccact tcaagaactc tgtagcaccg cctacatacc
1441  tggctctgct aatctcttta ccaatggcty ctgcacagcg cgtaaagccg tgccttaccg
1501  gcttggactc aagcagatg ttaccggata aggcagcagc gtcgggctga acggygggtt
1561  cctgataca gcccaacttg gaggaaaga cctacacga actgagatcc ctacagcgtg
1621  agcaathyaga aadgcacag ctctcccgag gaaagagcgc ggaagcttct CCagtaagcg
      BstVI.. (2 of 2)

1661  gcaaggtccgg aacagagag cgcacgctagg agctttccagg ggaataaaccc tggatctctt
      BssSI (2/2)

1741  alaytctctg cgggtctctc cactcttgcg ttggagctag attttttga tctctctcag
1801  gggggcggag cctatggaaa aacgcagca accgycctt tttaagcttc ctggctttt
1861  gctggccttt tctctcctg Tctttctctg cyttatccc tgatctctg gataacgta
      PstI...

1921  ttaccgctt tyaatgagct gatccctctc gccagcagc gccagcagc agcagcagc
1981  cagttagcga ggaagcgaan gnaagccca lacgcaaac gccctctccc ccguyttggr
      SapI....

2041  cgalctatca atgcacgctg caagacaggt ttccgacty gaaagcgggc agtgagcga
      PvuII (1/3)

2101  acccaattaa tctgagttag ctccctcatt aggcaccaca ggcctttacc ttatgcttc
      ..-35..
      Flac
      ...-10.

2161  cggctcgtat gttctgtaga atgtgagcg gatacaatt tcaacacgca AACGACCTATG
      M13Rev_seq_P11mer

2221  ACCatgatta cgcacagcctt tcaagccttt tttctggaga ttctcaac
      PflMI.....
      Hind3.

signal::linker::Clight

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
FM K K L L F A I P L V V P F Y Y
gtg aaa aaa tta tta tta tta tta tta tta tta gtt gtt cct ttc tat

16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
S R S A Q V Q L Q V D L E I K
tct cac agt gca Cag gtc caa cys CAG CTC GAC CTC GAG abc aaa
ApaI.....
      BspMI....
      Sall....
      AccI... (1/2)
      HincII (1/2)

```

Light domains could be cloned in as Spall-XhoI fragments.
 W-CL(Kappa) segments can be cloned in as ApaI-AscI fragments. <----->

CKappa-----
 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
 R G W V A A P S V F Y F P P S

2359 cgt gga act gtg gat gca cca tct gtc TTC atc ttc ccg cac tct
 bbsI... (1/2)

46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
 D E Q L K S G T A S V Y C L L
 gat gag cag ttg aaa tct gga act gcc tct gtt gtg tgc ctg ctg

2404 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
 R N F Y P R E A K V Q W K V D
 aat aac ttc tat ccc aga gag gcc aaa gta cag tgg aag gtg gat

76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
 N A L Q S G N S Q E S V T E Q
 aac gcc ctc caa tgg ggt aac tcc cag gag agt gtc aca gag cag

2494 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
 D S K D S T Y S L S S T L T L
 gag agc aag gac agr acc tac agc ctc agc acc ctg acc ctg

2539 gag agc agc agc agc agc agc agc agc agc agc agc agc agc agc agc
 EspI...

106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
 S K A E Y E K H K V Y A C E V
 Acc aaa gca gcc tac gag aaa cac aaa gtc TTC gcc tgc gaa gtc
 ...EspI....
 AccI... (2/2)

121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
 F H Q G L S S P V T K S F N R
 acc cat cag ggc ctg agt tca ccc ctg aca aag agc ttc aac agg
 AgeI... (1/2)

136 137 138 139 140
 G S C . .
 gga gag tgt taa taa GG CGCCGCaatt
 AscI.....
 BssHII.

```

2701      ctatttcag gacacagta ta
|
| PeIR::3-23(stuffed)::CHI::III fusion gene
|
|   1  2  3  4  5  6  7  8  9 10 11 12 13 14 15
|   M  K  Y  L  I  P  T  A  A  A  G  I  L  L  L
| 2723 atg aaa tac.cta ttg cct acg gca gcc gct gga ttg tta tta ctc
|
|-----
|   16 17 18 19 20 21 22
|   A  A  Q  P  A  M  A
| 2768 gcc gcc gag gcc gcc atg gcc
|   SELL.....
|   NgMEV..(1/2)
|   NcoI....
|
|-----
|   23 24 25 26 27 28 29 30
|   E  V  Q  L  L  E  S  G
| 2789 aaatgt|CAA|TGG|ctatgag|ctct|ggt|
|   | MFEI |
|
|-----
|   31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
|   G  G  L  V  Q  P  G  G  S  L  R  L  S  C  A
| 2813 |ggc|ggt|ctt|ggt|cag|ccl|ggt|ggt|tat|tca|cgt|ctt|ctt|tct|tct|tct|ggt|
|
|-----FR1-----
|   46 47 48
|   A  S  G
| 2858 |gct|TCC|GSA|
|   | DspBI |
|
|-----
|   Stuffer for CDR1, FR2, and CDR2----->
|   There are no stop codons in this stuffer.
|
|-----
|   2867      gcttcgagtc tqtlttgcctt
|   2887      ttctgtggt agcgcagtc gcgttacga gctgcacga ctgctlgagc aaaagccacg
|   2917      cttaactgt GATCAGccat gggatgtat tggccaaac agtcgtccgg atcttaacct
|   3097      gaggctttt ctacctacc tccagcagc gacatctggt ttgacacaga gcgatccgcg
|   3067      tcttcagttg atagaaacat taccacgttg ggtatggacc aatttgtta atgatgatgg

```

```

3127 taaaacctgg cagcagcag gctctgccc atcagaacctt tggcagcca ctatgtttgaa
3187 gctaacgta gtggctgucg taactatgac atttggaleag tggctacagc ccaagtgcta
      xhoI.....
3247 cgaacaacc cagcagggcc caactggtc gctgatatc atgtttggag caaaaatttt
3307 gtatggagg gtgcagggag acaaatcaac aatccacac gctgttgatc tgtttgtggg
3367 gaaccacag caggggtttg tgttggctgc gctggagat aorttggaga cttttccaa
3427 acgtatggc aataatgtga gtaactggaa aacactgca atggacttaa cyttccgggc
3487 aataanttc ttgttgtag ccagggccc agcgaagaa agcctcacc agggggagta
      MluI..
3547 tcaaacctg ggaacagaa acgatatgat tgttttca caaagacaa gggatcgtcc
3607 tggcttggc tggatgltg tggcaccgg tcaagtggg ttatgtgac ccgatggac
3667 agttagaag tactatgag atcagctgaa aatgtagaa aattttggcc gaaagtcgt
      PvuII.
3727 ctgctttagc aagcaggatg tggggcgca taaggagtcg
      HpaI..
      BlnCI(2/2)
.....FR3.....
4 5 6 7 8 9 10 11 12 13 14 15 16
93 94 95 96 97 98 99 100 101 102 103 104 105
S R D N S K N R L Y L Q M
|CTTACN|gac|aac|tct|aag|aat|act|ctc|tan|ttg|cag|tatg|
| XbaI |
.....FR3.....>|
17 18 19 20
106 107 108 109
N S L s i s i r s g
|aac|agc|ttn|ac|t|ctg|agc|att|cgg|tcc|g
|AFLII |
      y h s p t
3834 gg caa cat tct cca aac tga ccagcaga cacaaagggc
3872 ttacgttaa tccggcgcat gggatggtaa agagggggc tctttgttgg cctggaactca
3932 taagatgag gcaaaaatt ggcagggatg gaacagcag gtagggccc aagcactgac
3992 catcaatgg tactatgctg atgtaaacg caaatattggt talgttcata ctggcttta
4052 tccagttgt caatcagccc atgatacgg ataccggtt ccgggtacgg gaaaatggga
4112 ctggaaaggy ctattgcat ttgaaatga cctaaaggtt tataacccc ag
      as GTCAGC ctggcgcttc
      MheI..
4182 GTCCTAACI gtc tca agc
      | BstEII |

```

```

136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
A S T K G P S V F P E A P S S
gac tac acc aag ggc cca tgg ttc ccc ctg gaa ccc tcc tcc
151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
K S T S G G T A A L G C L V K
aag agc acc tct ggg ggc aca ggc ctg ggc tgc ctg gtc aag
166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
D Y F P E P V T V S W N S G A
gac tac ttc ccc gaa ccc gtc agc gtc tgg acc tca ggc ggc
181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
L T S G V H T F P A V L Q S S
ctg acc agc ggc gtc cac acc ttc ccc gct gtc cta cag tcc tca
196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
G L Y S L S V V T V P S S S
gga ctc tac tcc ctc agc agc gta gtc acc gtc ccc tcc agc agc
211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
L G T G T Y I C N V U H K P S
tgg ggc acc cag acc tac atc tgc aac gtc aat cac aag ccc agc
226 227 228 229 230 231 232 233 234 235 236 237 238
N T K V D K K V E P K S C
aac acc aag gtc gac aag AAA GGT GAG CCC AAA TCT TGT
      ON-TORCF019.....
      Poly His Kinase
139 140 141 142 143 144 145 146 147 148 149 150
A A A F H H H H G A A
GCC GCC CCA cat cat cat cac cat cac ggg gcc qud
      NAFI.....
      BagI....
151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
E D K L I S E E D L N G A A
gaa caa aaa ctg atc tca gaa gag gat ctg aat ggg gcc gca tgg
Mature III-----
166 167 168 169 170 171 172 173 174 175 176 177 178 179 180

```

I 4588 T V B S C L A K P H T E M S F
 act gtt gaa agt tgt tta gca aaa cct cat aca gaa aat tca ttt
 I 181 192 183 184 185 186 187 188 189 190 191 192 193 194 195
 T N Y W K D D K T L D R Y A N
 I 4633 act aac gtc tgg aaa gac gac ana act tta gat agt cgt tac gct aac
 I 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
 Y E C C L W N A T G V V C T
 I 4678 tat gag ggc tgt ctg tgc aat gct aca ggc gtt gtt gtt tgt act
 I 211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
 G D E T Q C Y G T W V F I G L
 I 4723 ggt gac gaa act cag tgt tac ggt aca tgg gtt cct att ggg ctt
 I 226 227 228 229 230 231 232 233 234 235 236 237 238 239 240
 A J P E W E G G C S E G G S
 I 4768 gct atc cct gaa aat gag ggt ggt ggc tct gag ggt ggc ggt tct
 I 241 242 243 244 245 246 247 248 249 250 251 252 253 254 255
 E G G C S W C G T K P E Y
 I 4813 gag ggt ggc ggt tct gag ggt ggc act aaa cct cct gag tac
 I 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270
 G D T P I P G Y T Y I N P L D
 I 4858 ggt gat aca cct att cag ggc tat act tat atc aac cct ctc gac
 I 271 272 273 274 275 276 277 278 279 280 281 282 283 284 285
 G T Y P P G T E Q N P A N P M
 I 4903 ggc act tat cag cct ggt act gag aca aac ccc gct aat cct aat
 I 296 297 298 299 300 291 292 293 294 295 296 297 298 299 300
 P S L E E S Q E L N T F N F Q
 I 4948 cct tct ctt gaa gag tct cag cct ctt aat act ttc atg ttt cag
 I 301 302 303 304 305 306 307 308 309 310 311 312 313 314 315
 N N R T R N R Q G A L T V Y T
 I 4983 aat aat agg ttc gga aat agg cag ggt gca tta act gtt tat acg
 I 316 317 318 319 320 321 322 323 324 325 326 327 328 329 330
 G T V T Q G T D E V K T Y Q

Rsmn!....

BserI..(2/2)

5038 ggc act ggt act caa ggc act gac ccc gtt aaa act tat tac cag
 331 332 333 334 335 336 337 338 339 340 341 342 343 344 345
 Y T S Y S S K A M Y D A Y W N
 5039 tac act act gta tca aaa gcc atg tat gac gct tac tgg aac
 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360
 G K F R D C A F H S G F N E D
 5128 ggt aaa ttc aga gac tgc gct ttc ctt tct gcc ttt aat gag GAT
 Rambl...
 361 362 363 364 365 366 367 368 369 370 371 372 373 374 375
 P F Y C E Y Q G Q S S D L R Q
 5173 cca ttc gct tgl gaa tat caa ggc caa tgg tct gac ctc Oct caa
 Rambl... BspMT... (2/2)
 376 377 378 379 380 381 382 383 384 385 386 387 388 389 390
 P P Y N A G G G S G G G S G G
 5218 cct cct gtc aat gct gcc ggc ggc tct ggt ggt ggt tct ggt ggc
 391 392 393 394 395 396 397 398 399 400 401 402 403 404 405
 G S R G G S R G G G G G S G G
 5260 ggc tct gag ggt gcc ggc tct gag gtl gcc ggc ggt tct gag ggt ggc
 406 407 408 409 410 411 412 413 414 415 416 417 418 419 420
 G S E G G G S G G G S G S G D
 5308 ggc tct gag ggt gcc ggt tct ggt gcc gcc tcc gcc tcc gcc ggt gct
 421 422 423 424 425 426 427 428 429 430 431 432 433 434 435
 K D Y S K M A N A N K G A M Y
 5353 ttt gat tat gaa aaa atg gca aac gct aat aag ggg gct atg aac
 436 437 438 439 440 441 442 443 444 445 446 447 448 449 450
 F N A H F N A T Q S D A K G K
 5398 gaa aat gcc gat gaa aac gcc cta cag tct gcc gct aag gcc aaa
 451 452 453 454 455 456 457 458 459 460 461 462 463 464 465
 L D S V A T D Y G A A T D G E
 5447 ctt gct tct gtc gct act gat tac ggt gct acc gat ggt ttc
 Rambl...
 466 467 468 469 470 471 472 473 474 475 476 477 478 479 480
 I G D V S G L A R G N G A T G

Table 630: Oligonucleotides used to clone CDRI/2 diversity

All sequences are 5' to 3'.

1) ON_CD1Bsf, 30 bases

A C C T C A C T G G C T T C C G G A
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

T T C A C T T T C T C T
 19 20 21 22 23 24 25 26 27 28 29 30

2) ON_Bcl2, 42 bases

A G A A A C C C A C T C C A A A C C
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

T T T A C C A G G A G C T T G C G
 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36

A A C C C A
 37 38 39 40 41 42

3) ON_CD2Xba, 51 bases

G G A A G G C A G T G A T C T A G A
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

G A T A G T G A A G C G A C C T T T
 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36

A A C G G A G T C A G C A T A
 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51

4) ON_BolXba, 23 bases

9 9 A A 9 9 C A 9 T 9 A T 9 A 9 A
i 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

9 A T A 9
19 20 21 22 23

End Tables