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(54) **PROTECTIVE SURFACE COATINGS FOR FLOW CELLS**

(57) An example of a method includes adding surface chemistry to a portion of a flow cell substrate. This example of the method further includes applying a water-soluble protective coating on at least the surface chemistry. Examples of flow cells incorporating examples of the water-soluble protective coating are also disclosed herein.

PROTECTIVE SURFACE COATINGS FOR FLOW CELLS

BACKGROUND

[0001] Biological arrays are among a wide range of tools used to detect and analyze molecules, including deoxyribonucleic acid (DNA) and ribonucleic acid (RNA). In these applications, the arrays are engineered to include probes for nucleotide sequences present in genes of humans and other organisms. In certain applications, for example, individual DNA and RNA probes may be attached at small locations in a geometric grid (or randomly) on an array support. A test sample, e.g., from a known person or organism, may be exposed to the grid, such that complementary fragments hybridize to the probes at the individual sites in the array. The array can then be examined by scanning specific frequencies of light over the sites to identify which fragments are present in the sample, by fluorescence of the sites at which the fragments hybridized.

[0002] Biological arrays may be used for genetic sequencing. In general, genetic sequencing involves determining the order of nucleotides or nucleic acids in a length of genetic material, such as a fragment of DNA or RNA. Increasingly longer sequences of base pairs are being analyzed, and the resulting sequence information may be used in various bioinformatics methods to logically fit fragments together so as to reliably determine the sequence of extensive lengths of genetic material from which the fragments were derived. Automated, computer-based examination of characteristic fragments have been developed, and have been used in genome mapping, identification of genes and their function, evaluation of risks of certain conditions and disease states, and so forth. Beyond these applications, biological arrays may be used for the detection and evaluation of a wide range of molecules, families of molecules, genetic expression levels, single nucleotide polymorphisms, and genotyping.

SUMMARY

[0003] In an example of a method disclosed herein, surface chemistry is added to a portion of a flow cell substrate, and a water-soluble protective coating is applied on at least the surface chemistry.

[0004] In another example of a method disclosed herein, a silane or a silane derivative is attached to a surface of a patterned substrate including depressions separated by interstitial regions, to form silanized depressions and silanized interstitial regions. A functionalized coating layer is formed in the silanized depressions and on the silanized interstitial regions. The functionalized coating layer is polished from the silanized interstitial regions. A primer is grafted to the functionalized coating layer in the silanized depressions to form functionalized depressions. A water-soluble protective coating is formed on the functionalized depressions and at least a portion of the patterned substrate.

[0005] An example of a flow cell disclosed herein includes a patterned substrate. The patterned substrate includes depressions separated by interstitial regions, and surface chemistry positioned in the depressions. A lid is bonded to a bonding region of the patterned substrate, wherein the lid at least partially defines a flow channel that is in selective communication with the depressions. A water-soluble protective coating covers the surface chemistry in the depressions and at least a portion of the patterned substrate.

[0006] Another example of a flow cell disclosed herein includes a non-patterned substrate, and a lid bonded to a bonding region of the non-patterned substrate, wherein the lid and the non-patterned substrate at least partially define a flow channel. Surface chemistry is positioned on the non-patterned substrate and in the flow channel. A water-soluble protective coating covers the surface chemistry.

BRIEF DESCRIPTION OF THE DRAWINGS

[0007] Features of examples of the present disclosure will become apparent by reference to the following detailed description and drawings, in which like reference numerals correspond to similar, though perhaps not identical, components. For the sake of brevity, reference numerals or features having a previously described function may or may not be described in connection with other drawings in which they appear.

[0008] Fig. 1 is a flow diagram illustrating an example of the methods disclosed herein;

[0009] Fig. 2 is a flow diagram illustrating more detailed examples of the method shown in Fig. 1;

[0010] Fig. 3 is a cross-sectional view of an example flow cell formed by the methods shown in Fig. 2;

[0011] Fig. 4 is a flow diagram illustrating another example of the methods disclosed herein;

[0012] Figs. 5A through 5H are cross-sectional views which together illustrate an example of the method shown in Fig. 4;

[0013] Figs. 5A through 5D and 5I through 5L are cross-sectional views which together illustrate another example of the method shown in Fig. 4;

[0014] Figs. 6A through 6E are cross-sectional views which together illustrate another example of the methods disclosed herein;

[0015] Fig. 7 is a bar graph depicting the Hairpin-TET (HP-TET) retention rate for flow cells after storage for 7 days at 30°C with different examples of the protective coating thereon;

[0016] Fig. 8 is a plot depicting the HP-TET retention for flow cells having had an example of the protective coating thereon and for an uncoated control;

[0017] Fig. 9 is a plot of the fluorescence intensity after a first or initial sequencing cycle for flow cells with example flow lanes and comparative flow lanes packaged in different humidity conditions;

[0018] Fig. 10 is a plot of the fluorescence intensity after the first sequencing cycle for a flow cell with example flow lanes and comparative flow lanes exposed to a temperature ramp profile for different time periods ranging from 2 days to 19 days;

[0019] Fig. 11A is a plot of the read 1 (R1) fluorescence intensity after the first initial sequencing cycle (C1) for an example flow cell and a comparative flow cell aged at 60°C for 6 days;

[0020] Fig. 11B includes a plot of the percentage of clusters passing filter (top) and a plot of the read 1 (R1) fluorescence intensity after the first sequencing cycle (C1) for an example flow cell and a comparative flow cell aged at room temperature for 7 days;

[0021] Fig. 12A is a plot of the median intensity for an example flow cell incubated at different temperatures for one hour;

[0022] Fig. 12B is a plot of the median intensity for a comparative flow cell incubated at different temperatures for one hour;

[0023] Fig. 13 is a plot of the read 1 (R1) fluorescence intensity after the first sequencing cycle for flow cells with example flow lanes and comparative flow lanes stored at different temperatures for different time periods;

[0024] Fig. 14 is a graph depicting a distance across a protective coating on a patterned wafer in mm (x-axis) versus a distance of the coating thickness in μm (y-axis) (i.e., coating thickness (μm) as a function of distance across the wafer surface (mm));

[0025] Fig. 15 is a plot of the fluorescence intensity after a surface primer accessibility test (CFR assay) for an example flow cell and a comparative flow cell after storage at room temperature for 28 days, 35 days, and 71 days;

[0026] Fig. 16 is a plot of the fluorescence intensity after the first sequencing cycle for an example flow cell after dry storage at 30°C for 0 weeks, 2 weeks, and 6 weeks, for a comparative flow cell after dry storage at 30°C for 2 weeks and 6 weeks, and for an example flow cell after wet storage at 4°C for 6 weeks;

[0027] Fig. 17 is a plot depicting the results of a Cal Fluor Red (CFR) retention assay after 11 days of storage at different storage conditions, where CFR retention means the CFR signal at day 0 versus the CFR signal after storage;

[0028] Fig. 18 is a bar graph depicting the CFR retention (%, ratio of CFR signal at day X / signal at day 0)) at time 0 (T0) and at day 1 and day 2 for an example flow cell and a comparative example flow cell;

[0029] Fig. 19 is a plot of intensity percent increase for example flow cell portions compared to a dry control with respect to a thickness of the protective coating of respective example flow cell portions;

[0030] Fig. 20 is a plot of the fluorescence intensity of two reads after the first sequencing cycle (C1) for a flow cell including example lanes and comparative example lanes;

[0031] Fig. 21 is a graph of the CFR (relative signal) with respect to a percentage of the protective coating that was formed simultaneously with primer grafting; and

[0032] Fig. 22 is a graph of intensity (in arbitrary fluorescence units, AFU) versus concentration for an example protective coating and several comparative example protective coatings.

INTRODUCTION

[0033] In a first aspect, a method comprises adding surface chemistry to a portion of a flow cell substrate, and applying a water-soluble protective coating on at least the surface chemistry.

[0034] In an example of this first aspect of the method, adding the surface chemistry involves forming a functionalized coating layer, and grafting a primer to the functionalized coating layer; and the water-soluble protective coating is applied after the primer is grafted. Also in this example of this first aspect, the water-soluble protective coating is patterned to define a bonding region of the flow cell substrate after the water-soluble protective coating is formed; and the method further comprises bonding a lid to the defined bonding region of the flow cell substrate to form a flow channel. Still further in this example of this first aspect, the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

[0035] In another example of this first aspect of the method, adding the surface chemistry involves forming a functionalized coating layer, and the water-soluble protective coating is applied after the functionalized coating layer is formed. Also in this example of this first aspect, the water-soluble protective coating is selected from the group consisting of a polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, polyacrylamide, and polyethylene glycol. Still further in this example of this first aspect, the water-soluble protective coating is patterned to define a bonding region of the flow cell substrate after the water-soluble protective coating is formed; and the method further comprises bonding a lid to the defined bonding region of the flow cell substrate to form a flow channel. After bonding, the method further comprises removing the

water-soluble protective coating, thereby exposing the functionalized coating layer and another portion of the substrate; grafting a primer to the functionalized coating layer; and forming a second water-soluble protective coating on the primer, the functionalized coating layer and the other portion of the flow cell substrate. In an example of this aspect, removing the water-soluble protective coating involves a dissolution process. Also in this aspect, the second water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

[0036] In an example of this first aspect of the method, the water-soluble protective coating is selected from the group consisting of a polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, dextran, polyacrylamide, glycols, ethylenediaminetetraacetic acid sodium salt, tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, and saline sodium citrate.

[0037] In an example of this first aspect of the method, applying the water-soluble protective coating involves applying an aqueous solution including up to about 15% (mass to volume) of a water-soluble material.

[0038] In an example of this first aspect of the method, applying the water-soluble protective coating involves flow through deposition, dip coating, spin coating, spray coating, ultrasonic spray coating, doctor blade coating, aerosol printing, or inkjet printing.

[0039] In an example of this first aspect of the method, the surface chemistry includes a primer, and the method further comprises at least partially removing the water-soluble protective coating, and performing a dye-based assay to detect any degradation of the primer.

[0040] In an example of this first aspect of the method, the method further comprises bonding a lid to a bonding region of the flow cell substrate to form a flow

channel, and then adding the surface chemistry and applying the water-soluble protective coating.

[0041] It is to be understood that any features of this first aspect of the method may be combined together in any desirable manner and/or configuration.

[0042] In a second aspect, the method comprises attaching a silane or a silane derivative to a surface of the patterned substrate including depressions separated by interstitial regions, thereby forming silanized depressions and silanized interstitial regions; forming a functionalized coating layer in the silanized depressions and on the silanized interstitial regions; polishing the functionalized coating layer from the silanized interstitial regions; grafting a primer to the functionalized coating layer in the silanized depressions to form functionalized depressions; and forming a water-soluble protective coating on the functionalized depressions and at least a portion of the patterned substrate.

[0043] In an example of this second aspect of the method, the water-soluble protective coating is formed after the primer is grafted; the water-soluble protective coating is patterned such that a bonding region of the patterned substrate remains exposed after the water-soluble protective coating is formed; and the method further comprises bonding a lid to the bonding region of the patterned substrate to form a flow channel that is in selective fluid communication with at least some of the functionalized depressions. In this example of the second aspect, the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

[0044] In an example of this second aspect of the method, after the functionalized coating layer is polished and before i) the primer is grafted and ii) the water-soluble protective coating is formed, the method further comprises: patterning an initial water-soluble protective coating on the functionalized coating layer and on the patterned substrate such that a bonding region of the patterned substrate remains exposed; bonding a lid to the bonding region of the patterned substrate to form a flow channel

that is in selective fluid communication with at least some of the depressions; and removing the initial water-soluble protective coating.

[0045] It is to be understood that any features of this second aspect of the method may be combined together in any desirable manner. Moreover, it is to be understood that any combination of features of this second aspect of the method and/or of the first aspect of the method may be used together, and/or that any features from either or both of these aspects may be combined with any of the examples disclosed herein.

[0046] A first aspect of a flow cell comprises a patterned substrate, including depressions separated by interstitial regions and surface chemistry positioned in the depressions; a lid bonded to a bonding region of the patterned substrate, wherein the lid at least partially defines a flow channel that is in selective communication with the depressions; and a water-soluble protective coating covering the surface chemistry in the depressions and at least a portion of the patterned substrate.

[0047] In an example of this first aspect of the flow cell, the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

[0048] In an example of this first aspect of the flow cell, a thickness of the water-soluble protective coating is at least about 50 nm.

[0049] In an example of this first aspect of the flow cell, the surface chemistry includes a functionalized coating layer and a primer grafted to the functionalized coating layer.

[0050] It is to be understood that any features of the first aspect of the flow cell may be combined together in any desirable manner. Moreover, it is to be understood that any combination of features from either or both of the methods and/or from the flow cell may be used together, and/or that any features from any or all of these aspects may be combined with any of the features of the examples disclosed herein.

[0051] A second aspect of a flow cell comprises a non-patterned substrate; a lid bonded to a bonding region of the non-patterned substrate, wherein the lid and the non-patterned substrate at least partially define a flow channel; surface chemistry positioned on the non-patterned substrate and in the flow channel; and a water-soluble protective coating covering the surface chemistry.

[0052] It is to be understood that any features of the second aspect of the flow cell may be combined together in any desirable manner. Moreover, it is to be understood that any combination of features from either or both of the methods and/or from the flow cells may be used together, and/or that any features from any or all of these aspects may be combined with any of the features of the examples disclosed herein.

DETAILED DESCRIPTION

[0053] Examples of the method disclosed herein involve the application of a protective coating during flow cell manufacturing workflows. The protective coating may be applied directly on chemically and/or biologically active surface chemistry (e.g., a functionalized coating layer, a primer) that is deposited on a substrate. The protective coating may protect the surface chemistry during subsequent processing techniques (e.g., assembly techniques, such as lid bonding, wafer dicing, etc.) and/or during shipping of the flow cell and/or during short and/or long term storage of the flow cell. In an example, the storage period may be up to 120 days, or longer. In another example, the storage period may range from about 1 day to about 75 days. While examples have been provided, it is to be understood that the storage period may range from any time after the protective coating has been applied until it is desirable to use the flow cell. As an example, the protective coating may protect the surface chemistry from debris and/or contamination that may otherwise contact the surface chemistry during lid bonding or other assembly processes. As another example, the protective coating may protect the surface chemistry from scratches or other handling related defects that may result during shipping. As still another example, the protective coating may protect the surface chemistry from environmental factors (e.g., temperature, humidity, etc.) during manufacturing, shipping, and/or short and/or long term storage (e.g., at a temperature

ranging from about 4°C to about 80°C, or in some instances lower temperatures, down to about -25°C). The protective coating helps to maintain the inherent stability of the surface chemistry, and thus improves the shelf life, temperature tolerance, durability, and ambient storage capability of the flow cell. The stabilization of the surface chemistry is efficient and stable over time.

[0054] The method(s) disclosed herein may be performed entirely at the wafer level, entirely at the die level, in part at the wafer level, and/or in part at the die level. As an example of performing the method partially at the wafer and die levels, the method may be initiated using a wafer, which is then diced to form several dies, and the method may continue using each of the dies. The ability to perform open wafer processing, at least in some examples, enables a variety of metrology/analytical techniques to be used for quality control and characterization. Prior to being bonded to form a flow cell, the patterned and surface modified wafer/substrate may be exposed to, for example, atomic force microscopy (AFM), scanning electron microscopy (SEM), ellipsometry, goniometry, scatterometry, and/or fluorescence techniques. Alternatively, the bonded flow cell may be exposed to these techniques. At the die level, the method(s) may be performed on an open faced die, or on an assembled flow cell (with an enclosed flow channel).

[0055] It is to be understood that terms used herein will take on their ordinary meaning in the relevant art unless specified otherwise. Several terms used herein and their meanings are set forth below.

[0056] The singular forms “a”, “an”, and “the” include plural referents unless the context clearly dictates otherwise.

[0057] The terms comprising, including, containing and various forms of these terms are synonymous with each other and are meant to be equally broad.

[0058] The terms top, bottom, lower, upper, on, etc. are used herein to describe the flow cell and/or the various components of the flow cell. It is to be understood that these directional terms are not meant to imply a specific orientation, but are used to designate relative orientation between components. The use of directional terms should not be interpreted to limit the examples disclosed herein to any specific orientation(s).

[0059] As used herein, "alkyl" refers to a straight or branched hydrocarbon chain that is fully saturated (i.e., contains no double or triple bonds). The alkyl group may have 1 to 20 carbon atoms. Example alkyl groups include methyl, ethyl, propyl, isopropyl, butyl, isobutyl, tertiary butyl, pentyl, hexyl, and the like. As an example, the designation "C1-4 alkyl" indicates that there are one to four carbon atoms in the alkyl chain, i.e., the alkyl chain is selected from the group consisting of methyl, ethyl, propyl, iso-propyl, n-butyl, isobutyl, sec-butyl, and t-butyl.

[0060] As used herein, "alkenyl" refers to a straight or branched hydrocarbon chain containing one or more double bonds. The alkenyl group may have 2 to 20 carbon atoms. Example alkenyl groups include ethenyl, propenyl, butenyl, pentenyl, hexenyl, and the like. The alkenyl group may be designated as, for example, "C2-4 alkenyl," which indicates that there are two to four carbon atoms in the alkenyl chain.

[0061] As used herein, "alkynyl" refers to a straight or branched hydrocarbon chain containing one or more triple bonds. The alkynyl group may have 2 to 20 carbon atoms. The alkynyl group may be designated, for example, as "C2-4 alkynyl," which indicates that there are two to four carbon atoms in the alkynyl chain.

[0062] An "amino" functional group refers to an $-NR_aR_b$ group, where R_a and R_b are each independently selected from hydrogen, C1-6 alkyl, C2-6 alkenyl, C2-6 alkynyl, C3-7 carbocyclyl, C6-10 aryl, 5-10 membered heteroaryl, and 5-10 membered heterocyclyl, as defined herein.

[0063] As used herein, "aryl" refers to an aromatic ring or ring system (i.e., two or more fused rings that share two adjacent carbon atoms) containing only carbon in the ring backbone. When the aryl is a ring system, every ring in the system is aromatic. The aryl group may have 6 to 18 carbon atoms, which may be designated as C6-18. Examples of aryl groups include phenyl, naphthyl, azulenyl, and anthracenyl.

[0064] As used herein, the term "attached" refers to the state of two things being joined, fastened, adhered, connected or bound to each other. For example, a nucleic acid can be attached to a functionalized coating layer by a covalent or non-covalent bond. A covalent bond is characterized by the sharing of pairs of electrons between atoms. A non-covalent bond is a physical bond that does not involve the sharing of

pairs of electrons and can include, for example, hydrogen bonds, ionic bonds, van der Waals forces, hydrophilic interactions and hydrophobic interactions.

[0065] An "azide" or "azido" functional group refers to $-N_3$.

[0066] As used herein, "carbocyclyl" means a non-aromatic cyclic ring or ring system containing only carbon atoms in the ring system backbone. When the carbocyclyl is a ring system, two or more rings may be joined together in a fused, bridged or spiro-connected fashion. Carbocyclyls may have any degree of saturation, provided that at least one ring in a ring system is not aromatic. Thus, carbocyclyls include cycloalkyls, cycloalkenyls, and cycloalkynyls. The carbocyclyl group may have 3 to 20 carbon atoms (i.e., C3-20). Examples of carbocyclyl rings include cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cyclohexenyl, 2,3-dihydro-indene, bicyclo[2.2.2]octanyl, adamantyl, and spiro[4.4]nonanyl.

[0067] As used herein, the term "carboxylic acid" or "carboxyl" as used herein refers to -C(O)OH.

[0068] As used herein, "cycloalkyl" means a fully saturated carbocyclyl ring or ring system. Examples include cyclopropyl, cyclobutyl, cyclopentyl, and cyclohexyl.

[0069] As used herein, "cycloalkylene" means a fully saturated carbocyclyl ring or ring system that is attached to the rest of the molecule via two points of attachment.

[0070] As used herein, "cycloalkenyl" or "cycloalkane" means a carbocyclyl ring or ring system having at least one double bond, wherein no ring in the ring system is aromatic. Examples include cyclohexenyl or cyclohexene and norbornene or norbornenyl. Also as used herein, "heterocycloalkenyl" or "heterocycloalkene" means a carbocyclyl ring or ring system with at least one heteroatom in ring backbone, having at least one double bond, wherein no ring in the ring system is aromatic.

[0071] As used herein, "cycloalkynyl" or "cycloalkyne" means a carbocyclyl ring or ring system having at least one triple bond, wherein no ring in the ring system is aromatic. An example is cyclooctyne. Another example is bicyclononyne. Also as used herein, "heterocycloalkynyl" or "heterocycloalkyne" means a carbocyclyl ring or ring system with at least one heteroatom in ring backbone, having at least one triple bond, wherein no ring in the ring system is aromatic.

[0072] The term “depositing,” as used herein, refers to any suitable application technique, which may be manual or automated, and results in modification of the surface properties. Generally, depositing may be performed using vapor deposition techniques, coating techniques, grafting techniques, or the like. Some specific examples include chemical vapor deposition (CVD), spray coating (e.g., ultrasonic spray coating), spin coating, dunk or dip coating, doctor blade coating, puddle dispensing, flow through coating, aerosol printing, inkjet printing, or the like.

[0073] As used herein, the term “depression” refers to a discrete concave feature in a patterned substrate having a surface opening that is completely surrounded by interstitial region(s) of the patterned substrate surface. Depressions can have any of a variety of shapes at their opening in a surface including, as examples, round, elliptical, square, polygonal, star shaped (with any number of vertices), etc. The cross-section of a depression taken orthogonally with the surface can be curved, square, polygonal, hyperbolic, conical, angular, etc. As an example, the depression can be a well. Also as used herein, a “functionalized depression” refers to the discrete concave feature where the functionalized coating layer and primer(s) are attached.

[0074] The term “each,” when used in reference to a collection of items, is intended to identify an individual item in the collection, but does not necessarily refer to every item in the collection. Exceptions can occur if explicit disclosure or context clearly dictates otherwise.

[0075] As used herein, the term "flow cell" is intended to mean a vessel having a chamber (i.e., flow channel) where a reaction can be carried out, an inlet for delivering reagent(s) to the chamber, and an outlet for removing reagent(s) from the chamber. In some examples, the chamber enables the detection of the reaction that occurs in the chamber. For example, the chamber can include one or more transparent surfaces allowing for the optical detection of arrays, optically labeled molecules, or the like, in the chamber.

[0076] As used herein, a “flow channel” may be an area defined between two bonded components, which can selectively receive a liquid sample. In some examples, the flow channel may be defined between a patterned substrate and a lid, and thus may be in fluid communication with one or more depressions defined in the patterned

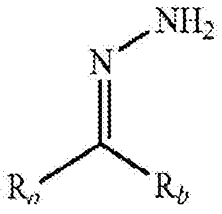
substrate. In other examples, the flow channel may be defined between a non-patterned substrate and a lid.

[0077] The "functionalized coating layer" referred to herein is intended to mean a semi-rigid material that is permeable to liquids and gases. The functionalized coating layer may be a hydrogel that can swell when liquid is taken up and that can contract when liquid is removed by drying. In an example, the functionalized coating layer is poly(N-(5-azidoacetamidylpentyl) acrylamide-co-acrylamide) (PAZAM). The functionalized coating layer may also include other molecules, which include one or more functional groups selected from the group consisting of optionally substituted alkenyl, azide/azido, optionally substituted amino, carboxyl, optionally substituted hydrazone, optionally substituted hydrazine, hydroxyl, optionally substituted tetrazole, optionally substituted tetrazine, nitrile oxide, nitrone, or thiol.

[0078] As used herein, "heteroaryl" refers to an aromatic ring or ring system (i.e., two or more fused rings that share two adjacent atoms) that contain(s) one or more heteroatoms, that is, an element other than carbon, including but not limited to, nitrogen, oxygen and sulfur, in the ring backbone. When the heteroaryl is a ring system, every ring in the system is aromatic. The heteroaryl group may have 5-18 ring members.

[0079] As used herein, "heterocyclyl" means a non-aromatic cyclic ring or ring system containing at least one heteroatom in the ring backbone. Heterocyclyls may be joined together in a fused, bridged or spiro-connected fashion. Heterocyclyls may have any degree of saturation provided that at least one ring in the ring system is not aromatic. In the ring system, the heteroatom(s) may be present in either a non-aromatic or aromatic ring. The heterocyclyl group may have 3 to 20 ring members (i.e., the number of atoms making up the ring backbone, including carbon atoms and heteroatoms). The heterocyclyl group may be designated as "3-6 membered heterocyclyl" or similar designations. In some examples, the heteroatom(s) are O, N, or S.

[0080] The term "hydrazine" or "hydrazinyl" as used herein refers to a $-NHNH_2$ group.

[0081] As used herein, the term "hydrazone" or "hydrazonyl" as used herein refers to a  group in which R_a and R_b are defined herein.

[0082] As used herein, "hydroxy" or "hydroxyl" refers to an –OH group.

[0083] As used herein, the term "interstitial region" refers to an area in a substrate or on a surface that separates depressions. For example, an interstitial region can separate one feature of an array from another feature of the array. The two features that are separated from each other can be discrete, i.e., lacking physical contact with each other. In another example, an interstitial region can separate a first portion of a feature from a second portion of a feature. In many examples, the interstitial region is continuous whereas the features are discrete, for example, as is the case for a plurality of wells defined in an otherwise continuous surface. The separation provided by an interstitial region can be partial or full separation. Interstitial regions may have a surface material that differs from the surface material of the features defined in the surface. For example, features of an array can have an amount or concentration of the coating layer and primer(s) that exceeds the amount or concentration present at the interstitial regions. In some examples, the coating layer and primer(s) may not be present at the interstitial regions.

[0084] "Nitrile oxide," as used herein, means a "$R_aC{\equiv}N^+O^-$" group in which R_a is defined herein. Examples of preparing nitrile oxide include *in situ* generation from aldoximes by treatment with chloramide-T or through action of base on imidoyl chlorides [RC(Cl)=NOH].

[0085] "Nitrone," as used herein, means a "$R_aR_bC{\equiv}NR_c^+O^-$" group in which R_a and R_b are defined herein and R_c is selected from C1-6 alkyl, C2-6 alkenyl, C2-6 alkynyl, C3-7 carbocyclyl, C6-10 aryl, 5-10 membered heteroaryl, and 5-10 membered heterocyclyl, as defined herein.

[0086] As used herein, a "nucleotide" includes a nitrogen containing heterocyclic base, a sugar, and one or more phosphate groups. Nucleotides are monomeric units of a nucleic acid sequence. In RNA, the sugar is a ribose, and in DNA, the sugar is a

deoxyribose, i.e. a sugar lacking a hydroxyl group that is present at the 2' position in ribose. The nitrogen containing heterocyclic base (i.e., nucleobase) can be a purine base or a pyrimidine base. Purine bases include adenine (A) and guanine (G), and modified derivatives or analogs thereof. Pyrimidine bases include cytosine (C), thymine (T), and uracil (U), and modified derivatives or analogs thereof. The C-1 atom of deoxyribose is bonded to N-1 of a pyrimidine or N-9 of a purine.

[0087] The term "open wafer processing," as used herein, refers to a series of sequential processes used to modify the surface of a substrate wafer with surface chemistry and protective coating, prior to any assembly (e.g., bonding) process.

[0088] The term "flow cell substrate" or "substrate" refers to a support upon which the surface chemistry may be added. The term "patterned substrate" refers to a support in which or on which depressions are defined. The term "non-patterned substrate" refers to a substantially planar support. The substrate may be a wafer, a panel, a rectangular sheet, a die, or any other suitable configuration. The substrate is generally rigid and is insoluble in an aqueous liquid. The substrate may be inert to a chemistry that is used to modify the depressions. For example, a substrate can be inert to chemistry used to form the functionalized coating layer, to form the protective coating, to attach the primer(s) to the functionalized coating layer, etc. Examples of suitable substrates include epoxy siloxane, glass and modified or functionalized glass, plastics (including acrylics, polystyrene and copolymers of styrene and other materials, polypropylene, polyethylene, polybutylene, polyurethanes, polytetrafluoroethylene (such as TEFLON® from Chemours), cyclic olefins/cyclo-olefin polymers (COP) (such as ZEONOR® from Zeon), polyimides, etc.), nylon, ceramics/ceramic oxides, silica, fused silica, or silica-based materials, aluminum silicate, silicon and modified silicon (e.g., boron doped p+ silicon), silicon nitride (Si_3N_4), silicon oxide (SiO_2), tantalum pentoxide (TaO_5) or other tantalum oxide(s) (TaO_x), hafnium oxide (HaO_2), carbon, metals, inorganic glasses, or the like. The substrate may also be glass or silicon, with a coating layer of tantalum oxide or another ceramic oxide at the surface.

[0089] As used herein, "plasma ashing" refers to a process of removing organic matter from a substrate by an oxygen plasma. The products that result from plasma

ashing may be removed with a vacuum pump/system. Plasma ashing can activate the substrate by introducing reactive hydroxyl groups.

[0090] As used herein, the "primer" is defined as a single stranded nucleic acid sequence (e.g., single strand DNA or single strand RNA) that serves as a starting point for DNA or RNA synthesis. The 5' terminus of the primer may be modified to allow a coupling reaction with the functionalized coating layer. The primer length can be any number of bases long and can include a variety of non-natural nucleotides. In an example, the sequencing primer is a short strand, ranging from 20 to 40 bases.

[0091] As used herein, the term "protective coating" refers to a water-soluble material in the form of a solid (e.g., a thin film), a liquid, or a gel that is applied on the surface chemistry, for example in a depression of a patterned substrate or on flow channel area(s) of an at least substantially planar substrate (i.e., a non-patterned substrate). The protective coating may be any water-soluble material that does not deleteriously affect the underlying surface chemistry or substrate and that serves to protect and/or preserve the functionality of the surface chemistry. For example, the protective coating may swell the functionalized layer and at least substantially prevent the layer from undergoing deleterious changes during processing and/or shipping and/or storage. For another example, the protective coating may preserve the accessibility of the primer and at least substantially prevent degradation of the functionalized layer. As examples, the water-soluble protective coating may be selected from the group consisting of a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

[0092] Some specific examples of water-soluble non-cationic synthetic polymers include water-soluble and non-cationic: polyacrylamides, poly(acrylic acid), poly(methacrylic acid), poly(vinyl pyrrolidone), poly(vinyl alcohol), poly(methacrylamide), poly(N-alkyl acrylamide)s, poly(N-dialkyl acrylamide)s, poly(N-(2-hydroxypropyl) methacrylamide), poly(divinyl ether-maleic anhydride), poly(phosphates), poly(2-alkyl-2-oxazolines), poly(hydroxyethyl methacrylate), poly(sulfobetaine methacrylate), silicones,

polyacrylates (e.g., sodium polyacrylate), polyethers (e.g., polyvinyl ethers, polyethylene glycol, poly(ethylene oxide), etc.), poly(vinyl ether-maleic acid), poly(2-hydroxyethyl acrylate), hydroxyl functional polymers, polypeptides (e.g., poly(glutamic acid) and its salts), or derivatives of the aforementioned polymers including, for example, random, block and graft copolymers and branched analogues. An example of a suitable copolymer is a polyvinyl alcohol/polyethylene glycol graft copolymer (one example of which includes KOLLICOAT® IR, available from BASF Corp.). An example of a suitable hydroxyl functional polymer is commercially available from BASF Corp. under the tradename KOLLICOAT® PR. Any of the water-soluble non-cationic synthetic polymers that include acid groups may be used in their alkali metal salt form.

[0093] Some specific examples of the natural polysaccharides or the derivatives thereof include starch, carboxymethylcellulose, xanthan gum, pectin, dextran, carrageenan, guar gum, cellulose, hydroxypropylmethyl cellulose (HPMC) hydroxypropyl cellulose (HPC), hydroxyethyl cellulose (HEC), methyl cellulose, carboxymethylhydroxyethyl cellulose (CMHEC), hyaluronic acid, starch phosphate, hydroxypropyl starch, hydroxyethyl starch, agarose, agar, and alginate.

[0094] Some specific examples of the natural proteins or the derivatives thereof include casein and albumin.

[0095] Some specific examples of water soluble salts include sodium and potassium salts of chloride, bromide, sulfate, phosphate, carbonate, acetate, or citrate. One example includes saline sodium citrate.

[0096] As mentioned above, the small molecule compounds may be selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin. Some specific examples of the water-soluble surfactant include anionic and nonionic surfactants, such as sodium dodecyl sulfate, alkyl ethoxylates, or ethoxylated oils and fats and sulfosuccinates. An example of a suitable sugar is sucrose. Examples of suitable chelators include ethylenediaminetetraacetic acid, sodium salt (i.e., EDTA), tris(3-hydroxypropyltriazolylmethyl)amine, (tris(2-carboxyethyl)phosphine) (which may also be considered a reducing agent), or bathophenanthrolinedisulfonic acid disodium salt. Examples of suitable buffers include tris(hydroxymethyl)aminomethane (i.e., Tris Base)

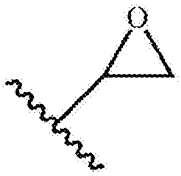
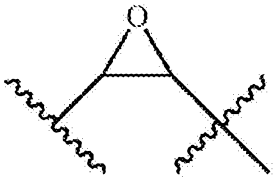
with ethylenediaminetetraacetic acid, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES), 3-[N-Tris(hydroxymethyl)methylamino]-2-hydroxypropanesulfonic acid (TAPSO), tricine, (3-(N-morpholino)propanesulfonic acid) (MOPS), and 3-(N,N-Bis[2-hydroxyethyl]amino)-2-hydroxypropanesulfonic acid (DIPSO).

[0097] In some of the examples disclosed herein, when applied after primer grafting, the protective coating may be a polyvinyl alcohol/polyethylene glycol graft copolymer (e.g., KOLLICOAT® IR, both available from BASF Corp.), sucrose, dextran (e.g., molecular weight of 200,000 Da), polyacrylamide (e.g., molecular weight of 40,000 Da, 200,000 Da, etc.), polyethylene glycol, ethylenediaminetetraacetic acid sodium salt (i.e., EDTA), tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, or saline sodium citrate. In some other of the examples disclosed herein, when applied after functionalized coating layer formation, the protective coating may be a polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, polyacrylamide, or polyethylene glycol. For at least sucrose and polyacrylamide, it is to be understood that the functionalized coating layer should be cured prior to applying the protective coating of these materials.

[0098] While several examples have been provided, it is to be understood that other water-soluble, non-cationic synthetic polymers and small molecules may be used to form the protective coating, as long as the selected material does not deleteriously affect the underlying surface chemistry or substrate. Moreover, it may also be desirable to select a material that can readily be removed, that does not interfere with subsequently performed process(es) (e.g., primer attachment, washing, etc.), and/or does not interfere with subsequently performed sequencing technique(s). In an example, the protective coating is not an auto-fluorescent material.

[0099] In an example, the thickness or depth of the water-soluble protective coating is at least about 25 nm. In an example, the thickness or depth of the water-soluble protective coating ranges from about 50 nm to about 100 nm. In another example, the thickness or depth of the water-soluble protective coating ranges from about 1 μm to about 15 μm. In yet another example, the thickness or depth of the

water-soluble protective coating ranges from about 1.5 μm to about 12 μm. The upper limit on the thickness may depend, at least in part, upon the architecture and dimensions of the flow channel and the flow cell that is formed. In an example, the upper end of the thickness range may range from about 10 μm to about 15 μm.

[0100] As used herein, the terms "silane" and "silane derivative" refer to an organic or inorganic compound containing one or more silicon atoms. An example of an inorganic silane compound is SiH_4, or halogenated SiH_4 where hydrogen is replaced by one or more halogen atoms. An example of an organic silane compound is $X\text{-}R^B\text{-}Si(OR^C)_3$, wherein X is a nonhydrolyzable organic group, such as amino, vinyl, epoxy ( or ), methacrylate, sulfur, alkyl, alkenyl, alkynyl; R^B is a spacer, for example $-(CH_2)_n-$, wherein n is 0 to 1000; R^C is selected from hydrogen, optionally substituted alkyl, optionally substituted alkenyl, optionally substituted alkynyl, optionally substituted carbocyclyl, optionally substituted aryl, optionally substituted 5-10 membered heteroaryl, and optionally substituted 5-10 membered heterocyclyl, as defined herein. As used herein, the terms "silane" and "silane derivative" can include mixtures of different silane and/or silane derivative compounds.

[0101] In some examples, the silane or silane derivative includes an unsaturated moiety that is capable of reacting with a functional group of the functionalized coating layer. As used herein, the term "unsaturated moiety" refers to a chemical group which includes cycloalkenes, cycloalkynes, heterocycloalkenes, heterocycloalkynes, or optionally substituted variants thereof including at least one double bond or one triple bond. The unsaturated moieties can be mono-valent or di-valent. When the unsaturated moiety is mono-valent, cycloalkene, cycloalkyne, heterocycloalkene, and heterocycloalkyne are used interchangeably with cycloalkenyls, cycloalkynyls, heterocycloalkenyl, and heterocycloalkynyl, respectively. When the unsaturated moiety is di-valent, cycloalkene, cycloalkyne, heterocycloalkene, and heterocycloalkyne are

used interchangeably with cycloalkenylene, cycloalkynylene, heterocycloalkenylene, and heterocycloalkynylene, respectively.

[0102] The unsaturated moiety can be covalently attached either directly to the silicon atoms of the silane or silane derivative, or indirectly attached via linkers. Examples of suitable the linkers include optionally substituted alkylenes (i.e., bivalent saturated aliphatic radicals (such as ethylene) regarded as being derived from an alkene by opening of the double bond or from an alkane by removal of two hydrogen atoms from different carbon atoms), substituted polyethylene glycols, or the like.

[0103] A "spacer layer," as used herein refers to a material that bonds two components together. In some examples, the spacer layer can be a radiation-absorbing material that aids in bonding, or can be put into contact with a radiation-absorbing material that aids in bonding.

[0104] The term "surface chemistry," as used herein refers to chemically and/or biologically active component(s) that are incorporated into the chamber of the flow cell. Examples of the surface chemistry disclosed herein include the functionalized coating layer attached to at least a portion of a surface of the substrate and/or and the primer(s) attached to at least a portion of the functionalized coating layer.

[0105] A "thiol" functional group refers to -SH.

[0106] As used herein, the terms "tetrazine" and "tetrazinyl" refer to six-membered heteroaryl group comprising four nitrogen atoms. Tetrazine can be optionally substituted.

[0107] "Tetrazole," as used herein, refer to five-membered heterocyclic group including four nitrogen atoms. Tetrazole can be optionally substituted.

[0108] As used herein, the term "YES method" refers a chemical vapor deposition process developed by Illumina, Inc. which uses the chemical vapor deposition tool provided by Yield Engineering Systems ("YES"). The tool includes three different vapor deposition systems. The automated YES-VertaCoat silane vapor system is designed for volume production with a flexible wafer handling module that can accommodate 200 mm or 300 mm wafers. The manual load YES-1224P Silane Vapor System is designed for versatile volume production with its configurable large capacity chambers. Yes-LabKote is a low-cost, tabletop version that is ideal for feasibility studies and for R&D.

[0109] The aspects and examples set forth herein and recited in the claims can be understood in view of the above definitions.

[0110] Examples of the flow cell including the protective coating and methods for making and using the same will now be described in reference to the figures.

[0111] An example of the method 100 is depicted in Fig. 1. The method 100 includes adding surface chemistry to a portion of a substrate (as shown at reference numeral 102), and applying a water-soluble protective coating at least on the surface chemistry (as shown at reference numeral 104). In an example, the method 100 includes adding surface chemistry to a portion of a flow cell substrate, wherein the surface chemistry is a sequencing primer, or a functionalized coating layer to attach the sequencing primer, or the sequencing primer and the functionalized coating layer; and applying a water-soluble protective coating on at least the surface chemistry. Two examples of the method 100 are shown in Fig. 2, as methods 100' and 100".

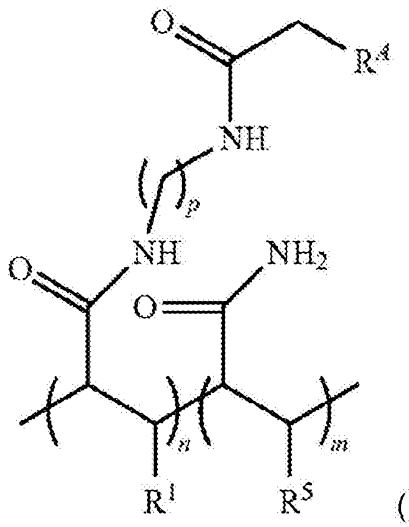
[0112] The methods 100', 100" may involve the patterned substrate, as shown at reference numeral 101 in Fig. 2, or a non-patterned substrate. Another example of the method 100 involving the non-patterned substrate 12' is further described in reference to Figs. 6A through 6E. The patterned substrate may be a patterned wafer or a patterned die or any of the other patterned substrates disclosed herein. Any example of the substrate described herein may be used. The patterned substrate (shown as at reference numeral 12 in Fig. 3) includes depressions defined on or in an exposed layer or surface of the substrate, and interstitial regions separating adjacent depressions. The depressions may be fabricated in or on the substrate using a variety of techniques, including, for example, photolithography, nanoimprint lithography, stamping techniques, embossing techniques, molding techniques, microetching techniques, printing techniques, etc. As will be appreciated by those in the art, the technique used will depend on the composition and shape of the substrate. Many different layouts of the depressions may be envisaged, as is discussed below in reference to Fig. 5A.

[0113] While not shown in Fig. 2, prior to adding the surface chemistry, each of the methods 100', 100" may involve exposing the patterned substrate to a cleaning process and/or to another process that prepares the surface (e.g., depressions and, in some instances, adjacent interstitial regions) of the patterned substrate for the

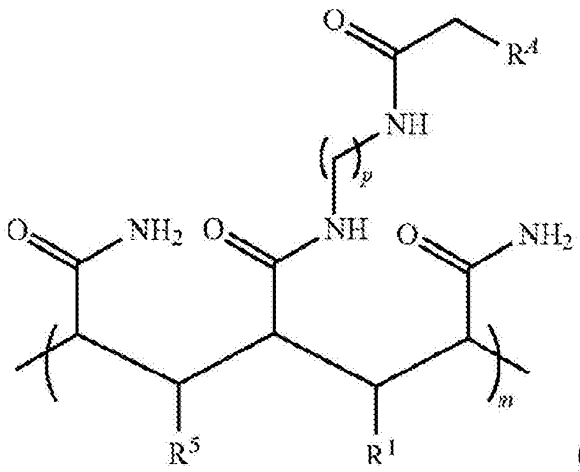
subsequent deposition of the surface chemistry. Examples of the cleaning process and surface preparation process(es) are discussed further below in reference to Figs. 4 and 5A through 5L.

[0114] In the example method 100', the surface chemistry may be both the functionalized coating layer and the primer(s), and a single protective coating may be used. In this example, adding the surface chemistry involves forming the functionalized coating layer in the depression(s) (reference numeral 102') and grafting the primer to the functionalized coating layer (reference numeral 102"), and the water-soluble protective coating is formed after the primer is grafted (reference numeral 104').

[0115] At reference numeral 102' in method 100', functionalized coating layer formation takes place. An example of the functionalized coating layer includes poly(N-(5-azidoacetamidylpentyl)acrylamide-co-acrylamide, PAZAM. PAZAM is represented by Formula (I) or Formula (II):



(I)



(II)

wherein:

R^1 is H or optionally substituted alkyl;

R^A is selected from the group consisting of azido, optionally substituted amino, optionally substituted alkenyl, optionally substituted hydrazone, optionally substituted hydrazine, carboxyl, hydroxy, optionally substituted tetrazole, optionally

substituted tetrazine, nitrile oxide, nitrone, and thiol; R^5 is selected from the group consisting of H and optionally substituted alkyl;

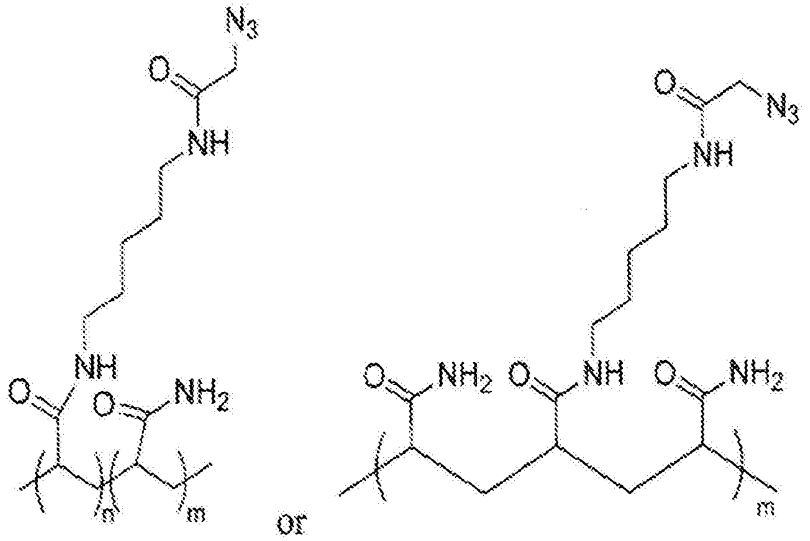
each of the $-(CH_2)_p-$ can be optionally substituted;

p is an integer in the range of 1 to 50;

n is an integer in the range of 1 to 50,000; and

m is an integer in the range of 1 to 100,000.

[0116] Specific examples of PAZAM are represented by:



wherein n is an integer in the range of 1-20,000, and m is an integer in the range of 1-100,000.

[0117] The molecular weight of the PAZAM may range from about 10 kDa to about 1500 kDa, or may be, in a specific example, about 312 kDa.

[0118] In some examples, PAZAM is a linear polymer. In some other embodiments, PAZAM is a lightly cross-linked polymer.

[0119] It is to be understood that other functionalized molecules may be used to form the functionalized coating layer, as long as they are functionalized to interact with the patterned substrate and the subsequently applied primer(s). Other examples of suitable molecules for forming the functionalized coating layer include those having a colloidal structure, such as agarose; or a polymer mesh structure, such as gelatin; or a cross-linked polymer structure, such as polyacrylamide polymers and copolymers, silane free acrylamide (SFA), or an azidolyzed version of SFA. Examples of suitable

polyacrylamide polymers may be synthesized from acrylamide and an acrylic acid or an acrylic acid containing a vinyl group, or from monomers that form [2+2] photo-cycloaddition reactions.

[0120] The functionalized molecule (e.g., PAZAM) may be deposited on the surface of the patterned substrate using spin coating, or dipping or dip coating, or flow of the functionalized molecule under positive or negative pressure, or other suitable techniques. The functionalized molecule may be present in a solution. In an example, the solution includes PAZAM in an ethanol and water mixture.

[0121] After being coated, the functionalized molecule may also be exposed to a curing process to form the functionalized coating layer across the entire patterned substrate (i.e., on depression(s) and interstitial region(s)). In an example, curing the functionalized molecule may take place at a temperature ranging from room temperature (e.g., about 25°C) to about 60°C for a time ranging from about 5 minutes to about 2 hours. In some instances, the functionalized coating layer is cured prior to the application of the protective coating.

[0122] To form the functionalized coating layer in the depression(s) and not on the interstitial region(s) of the patterned substrate, the functionalized coating layer may be polished off of the interstitial regions using i) a basic, aqueous slurry having a pH ranging from about 7.5 to about 11 and including an abrasive particle or ii) a polishing pad and a solution free of the abrasive particle.

[0123] In this example of the method 100', the primer is then grafted to the functionalized coating layer remaining in the depression(s), as shown as primer attachment at reference numeral 102''. Examples of suitable primers include forward amplification primers or reverse amplification primers. Specific examples of suitable primers include P5 or P7 primers, which are used on the surface of commercial flow cells sold by Illumina Inc. for sequencing on HiSeq®, HiSeqX®, MiSeq®, MiSeqX®, NextSeq®, NovaSeq™, Genome Analyzer®, and other instrument platforms.

[0124] Grafting may be accomplished by dunk coating, spray coating, puddle dispensing, or by another suitable method that will attach the primer(s) to the functionalized coating layer in at least some of the depressions. Each of these example

techniques may utilize a primer solution or mixture, which may include the primer(s), water, a buffer, and a catalyst.

[0125] Dunk coating may involve submerging the patterned substrate (having the functionalized coating layer in the depression(s) thereof) into a series of temperature controlled baths. The baths may also be flow controlled and/or covered with a nitrogen blanket. The baths may include the primer solution or mixture. Throughout the various baths, the primer(s) will attach to the functionalized coating layer in at least some of the depression(s). In an example, the coated and polished patterned substrate will be introduced into a first bath including the primer solution or mixture where a reaction takes place to attach the primer(s), and then the patterned substrate will be moved to additional baths for washing. The patterned substrate may be moved from bath to bath with a robotic arm or manually. A drying system may also be used in dunk coating.

[0126] Spray coating may be accomplished by spraying the primer solution or mixture directly onto the coated and polished patterned substrate. The spray coated wafer may be incubated for a time ranging from about 4 minutes to about 60 minutes at a temperature ranging from about 0°C to about 70°C. After incubation, the primer solution or mixture may be diluted and removed using, for example, a spin coater.

[0127] Puddle dispensing may be performed according to a pool and spin off method, and thus may be accomplished with a spin coater. The primer solution or mixture may be applied (manually or via an automated process) to the coated and polished patterned substrate. The applied primer solution or mixture may be applied to or spread across the entire surface of the coated and polished patterned substrate. The primer coated patterned substrate may be incubated for a time ranging from about 2 minutes to about 60 minutes at a temperature ranging from about 0°C to about 80°C. After incubation, the primer solution or mixture may be diluted and removed using, for example, the spin coater.

[0128] In one example, after the primer is grafted to the functionalized coating layer in the depression(s), this example of the method 100' further includes applying the water-soluble protective coating on the surface chemistry and on at least a portion of the substrate (as shown as protective coating formation at reference numeral 104'). In another example, the primer solution or mixture may include a water-soluble, film

forming material that forms the protective coating, and thus simultaneous primer grafting and protective coating formation may take place.

[0129] When primer grafting and protective coating formation are separate processes, the water-soluble protective coating may be selectively deposited, or patterned, such that the surface chemistry (in this example the functionalized coating layer and the primer(s) thereon) is covered and such that a bonding region of the patterned substrate remains exposed. The bonding region of the patterned substrate is generally located on some of the interstitial region(s) of the patterned substrate where a lid will be bonded to the patterned substrate. When the patterned substrate is a wafer, the bonding region may define the boundaries of several flow cells that are being formed from the wafer. When the patterned substrate is a die, the bonding region may define the outer boundaries of one flow cell that is being formed. It is to be understood that other portion(s) of the patterned substrate that are not part of the bonding region may be coated with the water-soluble protective coating.

[0130] In this example of the method 100', selectively depositing or patterning the water-soluble protective coating may be accomplished via dip coating, spin coating, spray coating, ultrasonic spray coating, doctor blade coating, aerosol printing, or inkjet printing. A mask may be used to cover the bonding region of the patterned substrate so that the water-soluble protective coating is not applied on the bonding region.

[0131] Each of the example selective deposition or patterning techniques for the water-soluble protective coating may utilize an aqueous solution, which may include the water and up to about 15% (mass to volume) of a water-soluble material. In some examples, the water-soluble material makes up 15% or less of the aqueous solution. In other examples, the aqueous solution includes from about 2% to about 13% of the water-soluble material, or from about 2.5% to about 10% of the water-soluble material. It is to be understood that the concentration of the aqueous solution may vary depending upon the flow cell architecture (e.g., the dimensions of the flow channel, input and output ports, etc.). For example, when flow through deposition is utilized, the concentration may be selected so that the aqueous solution can flow through the flow cell without clogging the port(s), flow channel, etc. As such, the concentration may also be greater than about 15%. To obtain a desirable thickness, the lower limit of the

concentration may be about 2 wt% (mass to volume). The water-soluble material (and the resulting protective coating), in this example of the method 100' may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin). In an example, the water-soluble material may be a polyvinyl alcohol/polyethylene glycol graft copolymer (an example of this is commercially available as KOLLICOAT® IR, available from BASF Corp.), sucrose, dextran, polyacrylamide, polyethylene glycol, ethylenediaminetetraacetic acid disodium salt (EDTA), tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, glycerol, or saline sodium citrate.

[0132] The aqueous solution may also include additives, such as water-soluble co-solvents, antioxidants, dyes, ultraviolet light stabilizers, processing aids, or the like. These additives may be included in the aqueous solution in amounts that do not deleteriously affect the flowability or the solution or the film forming ability of the water-soluble, film forming material. For example, a co-solvent, such as ethanol, may be present in an amount ranging from about 1% to about 10%, or from about 2.5% to about 7.5%. In another example, the aqueous solution may include about 5% of the water-soluble material, about 5% of the co-solvent, and a balance (about 90%) of water.

[0133] After the aqueous solution is applied, it may be dried to form the water-soluble protective coating in solid or gel form. Drying may be accomplished via air exposure, nitrogen exposure, vacuum, heating (e.g., in an oven), or spin coating (i.e., spinning until dry).

[0134] Some examples of the protective coating may remain in liquid form, such as sodium chloride-sodium citrate (SSC). The liquid form of the protective coating constitutes wet storage because the solution may at least substantially fill a flow channel and cover the surface chemistry. The liquid form of the protective coating may be used, for example, when the flow channel is formed prior to protective coating

application (e.g., at step 104''' in Fig. 2 or in Figs. 6A-6E), so that the liquid can be contained therein.

[0135] This example of the method 100' then includes assembly, as shown at reference numeral 106. The protective coating protects the surface chemistry during any assembly processes. In an example, assembly involves bonding a lid to the bonding region of the patterned substrate to form a flow channel that is in selective fluid communication with the depression(s). When the patterned substrate is a wafer, different areas of the lid may at least partially define respective flow channels that are being formed using the wafer. When the patterned substrate is a die, the lid may define the one or more flow channels that is/are being formed.

[0136] The lid may be any material that is transparent to an excitation light that is directed toward the surface chemistry in the depression(s). As examples, the lid may be glass (e.g., borosilicate, fused silica, etc.), plastic, or the like. A commercially available example of a suitable borosilicate glass is D 263®, available from Schott North America, Inc. Commercially available examples of suitable plastic materials, namely cyclo olefin polymers, are the ZEONOR® products available from Zeon Chemicals L.P.

[0137] In some examples, the lid may be integrally formed with sidewall(s) that correspond with the shape of bonding region, and that will be bonded to the bonding region. For example, a recess may be etched into a transparent block to form a substantially planar portion and sidewall(s) extending from the substantially planar portion. When the etched block is mounted to the bonding region of the patterned substrate, the recess may become the flow channel.

[0138] In other examples, the sidewall(s) and the lid may be separate components that are coupled to each other. For example, the lid may be a substantially rectangular block having an at least substantially planar exterior surface and an at least substantially planar interior surface that defines a portion (e.g., a top portion) of the flow channel (once bonded to the patterned substrate). The block may be mounted onto (e.g., bonded to) the sidewall(s), which are bonded to the bonding region of the patterned substrate and form sidewall(s) of the flow channel. In this example, the sidewall(s) may include any of the materials set forth herein for the spacer layer (described below).

[0139] The lid may be bonded to the bonding region of the patterned substrate using any suitable technique, such as laser bonding, diffusion bonding, anodic bonding, eutectic bonding, plasma activation bonding, glass frit bonding, or others methods known in the art. In an example, a spacer layer may be used to bond the lid to the bonding region of the patterned substrate. The spacer layer may be any material that will seal at least some of the interstitial regions (e.g., the bonding region) of the patterned substrate and the lid together.

[0140] In one example, the spacer layer may be a radiation-absorbing material that absorbs radiation at a wavelength that is transmitted by the lid and/or the patterned substrate. The absorbed energy, in turn, forms the bond between the spacer layer and the lid and between the spacer layer and the patterned substrate. An example of this radiation-absorbing material is black KAPTON® (polyimide containing carbon black) from DuPont (USA), which absorbs at about 1064 nm. It is to be understood that polyimide could be used without the addition of carbon black, except that the wavelength would have to be altered to one that is significantly absorbed by the natural polyimide material (e.g., 480 nm). As another example, polyimide CEN JP can be bonded when irradiated with light at 532 nm. When the spacer layer is the radiation-absorbing material, the spacer layer may be positioned at an interface between the lid and the patterned substrate so that the spacer layer contacts the desired bonding region. Compression may be applied (e.g., approximately 100 PSI of pressure) while laser energy at a suitable wavelength is applied to the interface (i.e., the radiation-absorbing material is irradiated). The laser energy may be applied to the interface both from the top and from the bottom in order to achieve suitable bonding.

[0141] In another example, the spacer layer may include a radiation-absorbing material in contact therewith. The radiation-absorbing material may be applied at the interface between the spacer layer and the lid as well as at the interface between the spacer layer and the patterned substrate. As an example, the spacer layer may be polyimide and the separate radiation-absorbing material may be carbon black. In this example, the separate radiation-absorbing material absorbs the laser energy that forms the bonds between the spacer layer and the lid and between the spacer layer and the patterned substrate. In this example, compression may be applied at the respective

interfaces while laser energy at a suitable wavelength is applied to the interfaces (i.e., the radiation-absorbing material is irradiated).

[0142] When the patterned substrate is a wafer, the spacer layer and sidewalls (of or connected to the lid) may physically separate one flow channel from an adjacent flow channel and may be located at the periphery of the wafers. When the patterned substrate is a die and the flow cell that is being formed is to include a single flow channel or lane, the spacer layer and sidewalls (of or connected to the lid) may be located at the periphery of the die to define the flow channel and seal the flow cell. When the patterned substrate is a die and the flow cell that is being formed is to include multiple isolated flow channels (e.g., eight flow channels or four flow lanes), the spacer layer and sidewalls (of or connected to the lid) may physically separate one flow channel/lane from an adjacent flow channel/lane and may be located at the periphery of the die. It is to be understood, however, that the spacer layer and sidewalls may be located in any desired region depending on the implementation.

[0143] When the patterned substrate is a die, the assembly 106 of the method 100' involves the bonding of the lid, which forms the flow cell. When the patterned substrate is a wafer, the assembly 106 of the method 100' may involve additional processing, such as dicing. In one example, the lid may be bonded to the patterned wafer and dicing forms individual flow cells. As mentioned herein, on a wafer, the sidewalls may physically separate one flow channel from an adjacent flow channel, and thus dicing may take place through at least some of the sidewalls, so that each individual flow cell includes a desirable number of flow channels, each of which has a portion of the original sidewall surrounding its periphery. In another example, the patterned wafer may be diced to form non-lidded dies, which can have respective lids bonded thereto to form individual flow cells.

[0144] In one example, after the assembly 106, one or more flow cells are formed.

[0145] Referring now to the example method 100" in Fig. 2, the surface chemistry may be both the functionalized coating layer and the primer(s), and two different protective coatings may be used. Like the method 100', adding the surface chemistry during the method 100" involves forming the functionalized coating layer in the

depression(s) (reference numeral 102') and grafting the primer to the functionalized coating layer (reference numeral 102'''). However, unlike the method 100', during the method 100'' an initial water-soluble protective coating is formed after the functionalized coating layer is formed (reference numeral 104'') and another water-soluble protective coating is formed after the primer is grafted (reference numeral 104''').

[0146] At reference numeral 102' and 104'' in method 100'', adding the surface chemistry involves forming the functionalized coating layer in the depression (102'), and the initial water-soluble protective coating is formed after the functionalized coating layer is formed (104''). It is to be understood that materials and methods for forming the functionalized coating layer as described herein may be used in the method 100''. In an example, it may be desirable to deposit the initial water-soluble protective coating within about 5 hours of polishing the functionalized coating layer. This time frame may decrease the surface decay rate of the functionalized coating layer and may also provide a larger thermal processing window.

[0147] The initial water-soluble protective coating may be selectively deposited, or patterned, such that the surface chemistry (in this example, the functionalized coating layer) is covered and such that a bonding region of the patterned substrate remains exposed. The bonding region is generally located on some of the interstitial region(s) of the patterned substrate where a lid will be bonded to the patterned substrate.

[0148] Selective deposition or patterning of the water-soluble protective coating may be accomplished via dip coating, spin coating, spray coating, ultrasonic spray coating, doctor blade coating, aerosol printing, or inkjet printing. As mentioned above, a mask may be used to cover the bonding region of the patterned substrate so that the initial water-soluble protective coating is not applied on the bonding region.

[0149] Each of the example selective deposition or patterning techniques for the initial water-soluble protective coating may utilize an aqueous solution, which may include the water and up to about 15% (mass to volume) of the water-soluble material. As mentioned herein, the concentration of the aqueous solution may vary depending upon the flow cell architecture (e.g., the dimensions of the flow channel, input and output ports, etc.), and thus may be greater than 15% or less than 15%. To obtain a desirable thickness, the lower limit of the concentration may be about 2 wt% (mass to

volume). The water-soluble material (and the resulting protective coating), in this example of the method 100", may be the polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, polyacrylamide, or polyethylene glycol. In this example, the aqueous solution may also include additives, such as antioxidants, dyes, ultraviolet light stabilizers, processing aids, or the like.

[0150] As shown at reference numeral 106', the method 100" involves assembly of the flow cell after the initial protective coating is formed. In this example, the initial protective coating protects the functionalized coating layer during any assembly processes. When the patterned substrate is a wafer, assembly involves bonding a lid to the bonding region to form respective flow channels that are in selective fluid communication with respective sets of the depressions, and then dicing the bonded lid and substrate to form respective flow cells. When the patterned substrate is a die, assembly involves bonding a lid to the bonding region to form a single flow cell with one or more flow channels. The bonding and/or dicing processes may be performed as described herein.

[0151] After the assembly 106' (i.e., bonding, or bonding and dicing), one or more flow cells are formed. The method 100' then involves removing the initial water-soluble protective coating, thereby exposing the functionalized coating layer and the at least the portion of the patterned substrate (reference numeral 108, shown as IPC removal); grafting a primer to the functionalized coating layer (reference numeral 102"', shown as primer attachment); and forming a second water-soluble protective coating on the primer, the functionalized coating layer and the at least the portion of the patterned substrate (reference numeral 104"', shown as protective coating formation).

[0152] The processes 108, 102"', and 104"' may take place in each of, or any one or more of the flow cells formed as a result of the assembly 106'.

[0153] Because the initial protective coating is water soluble, its removal may involve a dissolution process. Water and a buffer, or water, or an aqueous buffer may be introduced into the flow channel(s) of the flow cell(s) through respective input port(s) formed in the lid or the patterned substrate, may be maintained in the flow channel(s) for a time sufficient to dissolve the initial protective coating, and then may be removed from respective output port(s) formed in the lid or the patterned substrate. As such, the

dissolution may be performed as a flow through process. When the protective coating dissolves, the solution that is formed will have a concentration of the water-soluble material that is relatively low (in some instances, e.g., about 15% or less, mass to volume), which enables the flow channel(s) and port(s) to remain unclogged. Upon dissolution of the initial protective coating and removal of the solution from the flow cell(s), the functionalized coating layer and any of the patterned substrate that had been coated by the initial protective coating are exposed.

[0154] In this example of the method 100", the primer is then grafted to the functionalized coating layer in the depression(s), as represented by reference numeral 102"'. Any of the primers described herein may be used. In this example of the method 100", grafting may be accomplished by a flow through process. In the flow through process, the primer solution or mixture described herein may be introduced into the flow channel(s) of the flow cell(s) through respective input port(s), may be maintained in the flow channel(s) for a time sufficient (i.e., an incubation period) for the primer to attach to the functionalized coating layer in one or more of the depressions, and then may be removed from respective output port(s). After primer attachment, the additional fluid(s) may be directed through the flow channel(s) to wash the now functionalized depressions and the flow channel(s).

[0155] In an example, after the primer is grafted to the functionalized coating layer in the depression(s), this example of the method 100" further includes forming the (second) water-soluble protective coating on the surface chemistry (in this example, the functionalized coating layer having the primer(s) attached thereto) and on at least a portion of the patterned substrate (reference numeral 104"').

[0156] In this example method 100", the (second) water-soluble protective coating may be deposited by a flow through process. In the flow through process, the aqueous solution (including water and, in some instances, up to about 15% (mass to volume) of the water-soluble, material) may be introduced into the flow channel(s) of the flow cell(s) through respective input port(s) and may be maintained in the flow channel(s). Enough of the aqueous solution may be introduced to cover the surface chemistry and any exposed surfaces of the patterned substrate within the flow channel. While the solution is in the flow channel(s), the flow cell(s) may be exposed to a dry

down process where air, nitrogen, or vacuum is flushed through the input port for a set amount of time to dry the (second) water-soluble protective coating on the surface chemistry and any exposed portions of the substrate. In other example, the drying process may not be performed, and a liquid protective coating may be formed in the flow cell flow channel.

[0157] In this example method 100'', the (second) water-soluble protective coating may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin). In an example, the water-soluble material may be the polyvinyl alcohol/polyethylene glycol graft copolymer (e.g., KOLLICOAT® IR, available from BASF Corp.), sucrose, dextran, polyacrylamide, glycols, ethylenediaminetetraacetic acid sodium salt, tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, or saline sodium citrate.

[0158] In another example, primer grafting and (second) water-soluble protective coating on the surface chemistry 20, 22 may occur simultaneously via a flow through process. In this example, a single solution includes the primer and the water-soluble material.

[0159] In one example, after the protective coating formation 104''', one or more flow cells are formed.

[0160] An example of the flow cell 10 formed by the method 100' or 100'' is shown in Fig. 3. The flow cell 10 includes the patterned substrate 12, which may be a die that has been exposed to the processes of the method 100' or 100'', or a wafer that has been that has been exposed to the processes of the method 100' or 100'' and has been diced as a part of the assembly 106 or 106'.

[0161] Generally, the patterned substrate 12 includes depressions 14 separated by interstitial regions 16, and surface chemistry 20, 22 positioned in the depressions 14.

The surface chemistry includes the functionalized coating layer 20 and the primers 22. While not shown, it is to be understood that the depressions 14 may also have surface preparation chemistry (e.g., silane or a silane derivative) positioned between the substrate 12 and the functionalized coating layer. This same surface preparation chemistry may also be positioned on the interstitial regions 16.

[0162] The flow cell 10 also includes the lid 26 bonded to bonding region(s) 25 of the patterned substrate 12, wherein the lid 26 at least partially defines a flow channel 30A, 30B, etc. that is in selective communication with the depressions 14. In the example shown in Fig. 3, the lid 26 includes a top portion 27 that is connected to several sidewalls 29, and these components 27, 29 define a portion of each of the six flow channels 30A, 30B, 30C, 30D, 30E, 30F. The respective sidewalls 29 isolate one flow channel 30A, 30B, 30C, 30D, 30E, 30F from each adjacent flow channel 30A, 30B, 30C, 30D, 30E, 30F, each flow channel 30A, 30B, 30C, 30D, 30E, 30F is in selective fluid communication with a respective set of the depressions 14.

[0163] While not shown, the 26 or the patterned substrate 12 may include inlet and outlet ports that are to fluidically engage other ports (not shown) for directing fluid(s) into the respective flow channels 30A, 30B, 30C, 30D, 30E, 30F (e.g., from a reagent cartridge or other fluid storage system) and out of the flow channel (e.g., to a waste removal system).

[0164] The water-soluble protective coating 24 covers the surface chemistry 20, 22 in the depressions 14, and at least a portion of the patterned substrate 12 (e.g., those interstitial regions 16 that are not also bonding regions 25). In the example flow cell 10, the protective coating 24 has been formed by the process shown at reference numeral 104' in method 100' or the process shown at reference numeral 104''' in method 100''. As such, the protective coating 24 may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin).

[0165] The flow cell 10 can be shipped, stored, etc. with the protective coating 24 in place. When it is desirable to utilize the flow cell 10 in an application (e.g., a sequencing operation), the protective coating may be at least partially removed via the dissolution process described in reference to reference numeral 108. Removal may take place as part of a sequencing operation. Alternatively, removal may not take place in order for sequencing to proceed.

[0166] The flow cell 10 shown in Fig. 3 could be formed using a non-patterned substrate instead of a patterned substrate. With a non-patterned substrate, a continuous surface would include the same surface chemistry 20, 22 that is found in the wells 14' of Fig. 3. An example is shown and further described in reference to Figs. 6A through 6E.

[0167] Another example of the method 200 is depicted in Fig. 4. This example of the method 200 is a variation of the method 100, and describes in detail some of the other processes that may be involved, such as process(s) that prepare the surface (i.e., depressions 14 and, in some instances, adjacent interstitial regions 16) of the patterned substrate 12 for the subsequent deposition of the surface chemistry 20, 22.

[0168] The method 200 includes attaching a silane or a silane derivative to a surface of the patterned substrate including depressions separated by interstitial regions, thereby forming silanized depressions and silanized interstitial regions (as shown at reference numeral 202); forming a functionalized coating layer in the silanized depressions and on the silanized interstitial regions (as shown at reference numeral 204); polishing the functionalized coating layer from the silanized interstitial regions (as shown at reference numeral 206); grafting a primer to the functionalized coating layer in the silanized depressions to form functionalized depressions (as shown at reference numeral 208); and forming a water-soluble protective coating on the functionalized depressions and at least a portion of the patterned substrate (as shown at reference numeral 210).

[0169] Examples of the method 200 will be further described in reference to Figs. 5A through 5H (which is similar to method 100'), or Figs. 5A through 5D and 5I through 5L (which is similar to method 100''). Any details of the method 200 may also be combined with or included in the methods 100' or 100''.

[0170] Fig. 5A is a cross-sectional view of the patterned substrate 12. The patterned substrate 12 may be a patterned wafer or a patterned die or any other patterned substrate (e.g., panel, rectangular sheet, etc.). Any example of the substrate 12 described herein may be used. The patterned wafer may be used to form several flow cells, and the patterned die may be used to form a single flow cell. In an example, the substrate may have a diameter ranging from about 2 mm to about 300 mm, or a rectangular sheet or panel having its largest dimension up to 10 feet (~ 3 meters). In an example, the substrate wafer has a diameter ranging from about 200 mm to about 300 mm. In another example, the substrate die has a width ranging from about 0.1 mm to about 10 mm. While example dimensions have been provided, it is to be understood that substrates with any suitable dimensions may be used.

[0171] The patterned substrate 12 includes depressions 14 defined on or in an exposed layer or surface of the substrate 12, and interstitial regions 16 separating adjacent depressions 14. In the examples disclosed herein, the depressions 14 become functionalized with surface chemistry (e.g., 20, 22), while the interstitial regions 16 may be used for bonding but will not have primer(s) (shown in Figs. 5E and 5K) present thereon.

[0172] The depressions 14 may be fabricated in or on the substrate 12 using a variety of techniques, including, for example, photolithography, nanoimprint lithography, stamping techniques, embossing techniques, molding techniques, microetching techniques, printing techniques, etc. As will be appreciated by those in the art, the technique used will depend on the composition and shape of the substrate 12.

[0173] Many different layouts of the depressions 14 may be envisaged, including regular, repeating, and non-regular patterns. In an example, the depressions 14 are disposed in a hexagonal grid for close packing and improved density. Other layouts may include, for example, rectilinear (i.e., rectangular) layouts, triangular layouts, and so forth. In some examples, the layout or pattern can be an x-y format of depressions 14 that are in rows and columns. In some other examples, the layout or pattern can be a repeating arrangement of depressions 14 and/or interstitial regions 16. In still other examples, the layout or pattern can be a random arrangement of depressions 14 and/or interstitial regions 16. The pattern may include spots, pads, wells, posts, stripes, swirls,

lines, triangles, rectangles, circles, arcs, checks, plaids, diagonals, arrows, squares, and/or cross-hatches.

[0174] The layout or pattern may be characterized with respect to the density of the depressions 14 (i.e., number of depressions 14) in a defined area. For example, the depressions 14 may be present at a density of approximately 2 million per mm^2. The density may be tuned to different densities including, for example, a density of at least about 100 per mm^2, about 1,000 per mm^2, about 0.1 million per mm^2, about 1 million per mm^2, about 2 million per mm^2, about 5 million per mm^2, about 10 million per mm^2, about 50 million per mm^2, or more. Alternatively or additionally, the density may be tuned to be no more than about 50 million per mm^2, about 10 million per mm^2, about 5 million per mm^2, about 2 million per mm^2, about 1 million per mm^2, about 0.1 million per mm^2, about 1,000 per mm^2, about 100 per mm^2, or less. It is to be further understood that the density of depressions 14 on the substrate 12 can be between one of the lower values and one of the upper values selected from the ranges above. As examples, a high density array may be characterized as having depressions 14 separated by less than about 100 nm, a medium density array may be characterized as having depressions 14 separated by about 400 nm to about 1 μm, and a low density array may be characterized as having depressions 14 separated by greater than about 1 μm. While example densities have been provided, it is to be understood that substrates with any suitable densities may be used.

[0175] The layout or pattern may also or alternatively be characterized in terms of the average pitch, i.e., the spacing from the center of the depression 14 to the center of an adjacent interstitial region 16 (center-to-center spacing). The pattern can be regular, such that the coefficient of variation around the average pitch is small, or the pattern can be non-regular in which case the coefficient of variation can be relatively large. In either case, the average pitch can be, for example, at least about 10 nm, about 0.1 μm, about 0.5 μm, about 1 μm, about 5 μm, about 10 μm, about 100 μm, or more. Alternatively or additionally, the average pitch can be, for example, at most about 100 μm, about 10 μm, about 5 μm, about 1 μm, about 0.5 μm, about 0.1 μm, or less. The average pitch for a particular pattern of sites 16 can be between one of the lower values and one of the upper values selected from the ranges above. In an example, the

depressions 14 have a pitch (center-to-center spacing) of about 1.5 μm. While example average pitch values have been provided, it is to be understood that other avareage pitch values may be used.

[0176] In the examples shown in Figs. 5A through 5L, the depressions 14 are wells 14', and thus the patterned substrate 12 includes an array of wells 14' in a surface thereof. The wells 14' may be micro wells or nanowells. Each well 14' may be characterized by its volume, well opening area, depth, and/or diameter.

[0177] Each well 14' can have any volume that is capable of confining a liquid. The minimum or maximum volume can be selected, for example, to accommodate the throughput (e.g. multiplexity), resolution, analyte composition, or analyte reactivity expected for downstream uses of the flow cell 10 (see Figs. 5G and 5K). For example, the volume can be at least about 1×10^{-3} μm^3, about 1×10^{-2} μm^3, about 0.1 μm^3, about 1 μm^3, about 10 μm^3, about 100 μm^3, or more. Alternatively or additionally, the volume can be at most about 1×10^{4} μm^3, about 1×10^{3} μm^3, about 100 μm^3, about 10 μm^3, about 1 μm^3, about 0.1 μm^3, or less. It is to be understood that the functionalized coating layer can fill all or part of the volume of a well 14'. The volume of the coating layer in an individual well 14' can be greater than, less than or between the values specified above.

[0178] The area occupied by each well opening on a surface can be selected based upon similar criteria as those set forth above for well volume. For example, the area for each well opening on a surface can be at least about 1×10^{-3} μm^2, about 1×10^{-2} μm^2, about 0.1 μm^2, about 1 μm^2, about 10 μm^2, about 100 μm^2, or more. Alternatively or additionally, the area can be at most about 1×10^{3} μm^2, about 100 μm^2, about 10 μm^2, about 1 μm^2, about 0.1 μm^2, about 1×10^{-2} μm^2, or less. The area occupied by each well opening can be greater than, less than or between the values specified above.

[0179] The depth of each well 14' can be at least about 0.1 μm, about 1 μm, about 10 μm, about 100 μm, or more. Alternatively or additionally, the depth can be at most about 1×10^{3} μm, about 100 μm, about 10 μm, about 1 μm, about 0.1 μm, or less. The depth of each well 14' can be greater than, less than or between the values specified above.

[0180] In some instances, the diameter of each well 14' can be at least about 50 nm, about 0.1 μm, about 0.5 μm, about 1 μm, about 10 μm, about 100 μm, or more.

Alternatively or additionally, the diameter can be at most about 1×10^3 μm, about 100 μm, about 10 μm, about 1 μm, about 0.5 μm, about 0.1 μm, or less (e.g., about 50 nm). The diameter of each well 14' can be greater than, less than or between the values specified above.

[0181] The patterned substrate 12 may be exposed to a series of processes in order to add the surface chemistry 20, 22 in the depression(s) 14 and to form the water-soluble protective coating 24 on the surface chemistry 20, 22 and on at least a portion of the patterned substrate 12. Figs. 5B through 5H together illustrate an example in which the surface chemistry 20, 22 is added before the protective coating 24 is formed; and Figs. 5B through 5D and 5I through 5L together illustrate an example in which several protective coatings 24', 24 are formed in order to protect different surface chemistry 20, 22 at different stages of the method 200.

[0182] While not shown, it is to be understood that the patterned substrate 12 may be exposed to a plasma ashing in order to clean and activate the surface. For example, the plasma ashing process may remove organic material and introduce surface hydroxyl groups. Other suitable cleaning processes may be used to clean the substrate 12, depending, in part, on the type of substrate 12. For example, chemical cleaning may be performed with oxidizing agents or caustic solutions.

[0183] The patterned substrate 12 (shown in Fig. 5A) may then be exposed to a process that will prepare the surface 12 for deposition of the functionalized coating layer 20 (Fig. 5C), which is one example of the surface chemistry disclosed herein. In an example, the patterned substrate 12 may be exposed to silanization, which attaches a silane or the silane derivative 18 (Fig. 5B) to the patterned wafer surface. Silanization introduces the silane or the silane derivative 18 across the surface, including in the depression 14, 14' (e.g., on the bottom surface and along the side walls) and on the interstitial regions 16.

[0184] Silanization may be accomplished using any silane or silane derivative 18. The selection of the silane or silane derivative 18 may depend, in part, upon the functionalized molecule that is to be used to form the functionalized coating layer 20 (shown in Fig. 5C), as it may be desirable to form a covalent bond between the silane or silane derivative 18 and the functionalized coating layer 20. The method used to attach

the silane or silane derivative 18 to the substrate 12 may vary depending upon the silane or silane derivative 18 that is being used. Several examples are set forth herein.

[0185] In an example, the silane or silane derivative 18 is (3-aminopropyl)triethoxysilane (APTES) or 3-aminopropyl)trimethoxysilane (APTMS) (i.e., $X\text{-}R^{B}\text{-}Si(OR^{C})_3$, wherein X is amino, R^{B} is $-(CH_2)_3-$, and R^{C} is ethyl or methyl). In this example, the substrate 12 surface may be pre-treated with the (3-aminopropyl)triethoxysilane (APTES) or 3-aminopropyl)trimethoxysilane (APTMS) to covalently link silicon to one or more oxygen atoms on the surface (without intending to be held by mechanism, each silicon may bond to one, two or three oxygen atoms). This chemically treated surface is baked to form an amine group monolayer. The amine groups are then reacted with Sulfo-HSAB to form an azido derivative. UV activation at 21°C with 1 J/cm^2 to 30 J/cm^2 of energy generates an active nitrene species, which can readily undergo a variety of insertion reactions with PAZAM (e.g., the functionalized molecule).

[0186] Other silanization methods may also be used. Examples of suitable silanization methods include vapor deposition, the YES method, spin coating, or other deposition methods. Some examples of methods and materials that may be used to silanize the substrate 12 are described herein, although it is to be understood that other methods and materials may be used.

[0187] In an example utilizing the YES CVD oven, the patterned substrate 12 is placed in the CVD oven. The chamber may be vented and then the silanization cycle started. During cycling, the silane or silane derivative vessel may be maintained at a suitable temperature (e.g., about 120°C for norbornene silane), the silane or silane derivative vapor lines be maintained at a suitable temperature (e.g., about 125°C for norbornene silane), and the vacuum lines be maintained at a suitable temperature (e.g., about 145°C).

[0188] In another example, the silane or silane derivative 18 (e.g., liquid norbornene silane) may be deposited inside a glass vial and placed inside a glass vacuum desiccator with a patterned substrate 12. The desiccator can then be evacuated to a pressure ranging from about 15 mTorr to about 30 mTorr, and placed inside an oven at a temperature ranging from about 60°C to about 125°C. Silanization

is allowed to proceed, and then the desiccator is removed from the oven, cooled and vented in air.

[0189] Vapor deposition, the YES method and/or the vacuum desiccator may be used with a variety of silane or silane derivative 18, such as those silane or silane derivatives 18 including examples of the unsaturated moieties disclosed herein. As examples, these methods may be used when the silane or silane derivative 18 includes a cycloalkene unsaturated moiety, such as norbornene, a norbornene derivative (e.g., a (hetero)norbornene including an oxygen or nitrogen in place of one of the carbon atoms), transcyclooctene, transcyclooctene derivatives, transcyclopentene, transcycloheptene, trans-cyclononene, bicyclo[3.3.1]non-1-ene, bicyclo[4.3.1]dec-1 (9)-ene, bicyclo [4.2.1]non-1(8)-ene, and bicyclo[4.2.1]non-1-ene. Any of these cycloalkenes can be substituted, for example, with an R group, such as hydrogen, alkyl, alkenyl, alkynyl, cycloalkyl, cycloalkenyl, cycloalkynyl, aryl, heteroaryl, heteroalicyclyl, aralkyl, or (heteroalicyclyl)alkyl. An example of the norbornene derivative includes [(5-bicyclo[2.2.1]hept-2-enyl)ethyl]trimethoxysilane. As other examples, these methods may be used when the silane or silane derivative 18 includes a cycloalkyne unsaturated moiety, such as cyclooctyne, a cyclooctyne derivative, or bicyclononynes (e.g., bicyclo[6.1.0]non-4-yne or derivatives thereof, bicyclo[6.1.0]non-2-yne, or bicyclo[6.1.0]non-3-yne). These cycloalkynes can be substituted with any of the R groups described herein.

[0190] As shown in Fig. 5B, the attachment of the silane or silane derivative 18 forms a silanized patterned substrate, including silanized depressions and silanized interstitial regions.

[0191] The silanized patterned wafer may then be exposed to a process that will form the functionalized coating layer 20 on the silanized depressions and silanized interstitial regions.

[0192] As described herein, examples of the functionalized coating layer 20 include PAZAM, or any other molecule that is functionalized to interact with the patterned wafer 12 and the subsequently applied primer(s) 22. The functionalized coating layer 20 may be formed on the surface of the silanized patterned wafer (i.e., onto the silanized depressions and the silanized interstitial regions) using any of the

techniques described in reference to reference numeral 102'. The resulting coating layer 20 is shown in Fig. 5C.

[0193] The attachment of the functionalized coating layer 20 to the silanized depressions and silanized interstitial regions (i.e., 18) may be through covalent bonding. The covalent linking of the functionalized coating layer 20 to the silanized depressions is helpful for maintaining the functionalized coating layer 20 in the depressions 14, 14' throughout the lifetime of the ultimately formed flow cell during a variety of uses. The following are some examples of reactions that can take place between the silane or silane derivative 18 and the functionalized coating layer 20.

[0194] When the silane or silane derivative 18 includes norbornene or a norbornene derivative as the unsaturated moiety, the norbornene or a norbornene derivative can: i) undergo a 1,3-dipolar cycloaddition reaction with an azide/azido group of PAZAM; ii) undergo a coupling reaction with a tetrazine group attached to PAZAM; undergo a cycloaddition reaction with a hydrazone group attached to PAZAM; undergo a photo-click reaction with a tetrazole group attached to PAZAM; or undergo a cycloaddition with a nitrile oxide group attached to PAZAM.

[0195] When the silane or silane derivative 18 includes cyclooctyne or a cyclooctyne derivative as the unsaturated moiety, the cyclooctyne or cyclooctyne derivative can: i) undergo a strain-promoted azide-alkyne 1,3-cycloaddition (SPAAC) reaction with an azide/azido of PAZAM, or ii) undergo a strain-promoted alkyne-nitrile oxide cycloaddition reaction with a nitrile oxide group attached to PAZAM.

[0196] When the silane or silane derivative 18 includes a bicyclononyne as the unsaturated moiety, the bicyclononyne can undergo similar SPAAC alkyne cycloaddition with azides or nitrile oxides attached to PAZAM due to the strain in the bicyclic ring system.

[0197] While not shown, it is to be understood that in some examples of the method, the patterned substrate 12 may not be exposed to silanization. Rather, the patterned substrate 12 may be exposed to plasma ashing, and then the functionalized coating layer 20 may be directly spin coated (or otherwise deposited) on the plasma ashed patterned substrate 12. In this example, plasma ashing may generate surface-activating agent(s) (e.g., -OH groups) that can adhere the functionalized coating layer

20 to the patterned substrate 12. In these examples, the functionalized coating layer 20 is selected so that it reacts with the surface groups generated by plasma ashing.

[0198] Also while not shown, it is to be understood that the silanized and coated patterned substrate (shown in Fig. 5C) may be exposed to a cleaning process. This process may utilize a water bath and sonication. The water bath may be maintained at a relatively low temperature ranging from about 22°C to about 45°C. In another example the water bath temperature ranges from about 25°C to about 30°C.

[0199] The silanized and coated patterned substrate is then exposed to polishing to remove portion(s) of the functionalized coating layer 20 from the silanized interstitial regions. The silanized, coated, and polished patterned substrate is shown in Fig. 5D. The portions of the silane or silane derivative 18 that are adjacent to the interstitial regions 16 may or may not be removed as a result of polishing. As such, in Figs. 5D through 5L, the portions of the silane or silane derivative 18 that are adjacent to the interstitial regions 16 are shown in phantom, as they may at least partially remain after polishing or they may be removed after polishing. When these silanized portions are completely removed, it is to be understood that the underlying substrate 12 is exposed. As such, in some examples, the spacer layer 28 may directly contact the substrate 12 at the bonding regions 25 (e.g., in Figs. 5G and 5J) and the protective coatings 24', 24 may directly contact the substrate 12 at one or more interstitial region(s) 16 (e.g., in Figs. 5F, 5J and 5L). When these silanized portions at least partially remain after polishing, the subsequently bonded lid and the subsequently formed protective coatings 24, 24' directly contact the silane or silane derivative 18 at the bonding regions 25 and the interstitial regions 16.

[0200] The polishing process may be performed with a gentle chemical slurry (including an abrasive) which can remove the thin functionalized coating layer 20, and in some instances, at least part of the silane or silane derivative 18, from the interstitial regions 16 without deleteriously affecting the underlying substrate 12 at those regions. Alternatively, polishing may be performed with a solution that does not include the abrasive particles.

[0201] The gentle chemical slurry is a basic, aqueous slurry having a pH ranging from about 7.5 to about 11 and including an abrasive particle. Examples of the abrasive

particle include calcium carbonate ($CaCO_3$), agarose, graphite, poly(methyl methacrylate) (PMMA), silica, aluminum oxide (i.e., alumina), ceria, polystyrene, and combinations thereof. In some examples, the abrasive particle is selected from the group consisting of calcium carbonate ($CaCO_3$), agarose, and graphite. The average particle size of the abrasive particles may range from about 15 nm to about 5 μm, and in one example is about 700 nm.

[0202] In addition to the abrasive particles, the basic, aqueous slurry may also include a buffer, a chelating agent, a surfactant, and/or a dispersant. An example of the buffer includes tris base (i.e., tris(hydroxymethyl)aminomethane), which may be present in a solution having a pH of about 9. An example of the chelating agent is ethylenediaminetetraacetic acid (EDTA), which may be present in a solution having a pH of about 8. An example of the surfactant is an anionic surfactant, such as sodium dodecyl sulfate. Polyacrylate dispersants having different molecular weights may be used. An example of the dispersant is poly(acrylic acid sodium salt). The dispersant may help to maintain the size of, and at least substantially prevent settling of the abrasive particles.

[0203] The basic, aqueous slurry may be used in a chemical mechanical polishing system to polish the surface of the silanized and coated patterned substrate shown in Fig. 5C. The polishing head(s)/pad(s) or other polishing tool(s) is/are capable of polishing the functionalized coating layer 20 from the interstitial regions 16 while leaving the functionalized coating layer 20 in the depressions 14, 14' and leaving the underlying substrate 12 at least substantially intact. As an example, the polishing head may be a Strasbaugh ViPRR II polishing head.

[0204] As mentioned above, polishing may be performed with a polishing pad and a solution without any abrasive. For example, the polish pad may be utilized with a solution free of the abrasive particle (i.e., a solution that does not include abrasive particles).

[0205] Polishing removes portion(s) of the functionalized coating layer 20 (and in some instances at least part of the silane or silane derivative 18) from the interstitial regions 16 and leaves portion(s) of the functionalized coating layer 20 in the silanized depressions, as shown in Fig. 5D. Also as mentioned above, the interstitial region(s) 16

may remain silanized after polishing is complete. In other words, the silanized interstitial regions may remain intact after the polishing. Alternatively (as indicated by the phantom portions of 18), the silane or silane derivative 18 may be removed from the interstitial region(s) 16 as a result of polishing.

[0206] While not shown, it is to be understood that the silanized, coated, and polished patterned substrate (shown in Fig. 5D) may be exposed to a cleaning process. This process may utilize a water bath and sonication. The water bath may be maintained at a relatively low temperature ranging from about 22°C to about 30°C. The silanized, coated, and polished patterned substrate may also be spin dried, or dried via another suitable technique.

[0207] The silanized, coated, and polished patterned substrate shown in Fig. 5D may then be exposed to the processes shown in Figs. 5E through 5H or to the processes shown in Figs. 5I through 5L. In Figs. 5E through 5H, a single water-soluble protective coating 24 is formed, and in Figs. 5I through 5L, multiple water-soluble protective coatings 24', 24 are formed.

[0208] The example shown in Figs. 5E though 5H will now be described. In Fig. 5E, the primer 22 is grafted to the functionalized coating layer 20 in the depression(s) 14, 14'. Any of the primers described herein may be used. In this example, grafting may be accomplished by dunk coating, spray coating, puddle dispensing, or by another suitable method that will attach the primer(s) to the functionalized coating layer 20 in at least some of the depressions 14, 14'. Each of these example techniques may utilize the primer solution or mixture described herein, which may include the primer(s), water, a buffer, and a catalyst.

[0209] As shown in Fig. 5F, after the primer 22 is grafted to the functionalized coating layer 20 in the depressions 14, 14', the water-soluble protective coating 24 is formed on the surface chemistry 20, 22 and on at least a portion of the patterned substrate. The water-soluble protective coating 24 may be formed on the exposed surface of the patterned substrate 12 that is not part of a bonding region 25. In this example, the water-soluble protective coating 24 is selectively deposited or patterned on the interstitial regions 16 between adjacent depressions 14, 14', but not at the edge/periphery of the patterned substrate 12 where the bonding region 25 is located.

The selective deposition/patterning of the water-soluble protective coating 24 may be accomplished using the aqueous solution, as described herein. In this example, the material in the aqueous solution may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin). In an example, the water-soluble material may be. After the aqueous solution is deposited, it may be dried to form the water-soluble protective coating 24.

[0210] As depicted in Fig. 5G, the lid 26 may then be bonded to the bonding region 25. The lid 26 may be any of the materials and in any of the configurations described herein. The lid 26 may also be bonded to the bonding region 25 via any of the techniques described herein.

[0211] In the example shown in Fig. 5G, the lid 26 includes a top portion 27 integrally formed with sidewall(s) 29. The sidewall(s) 29 are bonded to the bonding region 25 of the patterned substrate 12 through the spacer layer 28.

[0212] Together, the lid 26 and the patterned substrate 12 (with the surface chemistry 20, 22) define the flow channel 30, which is in selective fluid communication with the depressions 14, 14'. The flow channel 30 may serve to, for example, selectively introduce fluid to the protective coating 24 in order to remove the coating 24, and to selectively introduce reaction components or reactants to the surface chemistry 20, 22 (after the protective coating 24 is removed) in order initiate designated reactions in/at the depressions 14, 14'.

[0213] When the lid 26 is bonded to the silanized, coated, polished, and grafted patterned substrate, an example of the flow cell 10' is formed, as shown in Fig. 5G. In this example, the protective 24 remain in place on the surface chemistry 20, 22 and some of the patterned substrate surface. The flow cell 10' can be shipped, stored, etc. with the protective coating 24 in place.

[0214] When it is desirable to utilize the flow cell 10' in an application (e.g., a sequencing operation), the protective coating 24 may be removed via the dissolution

process described in reference to reference numeral 108. The water solubility of the protective coating 24 enables it to be removed via dissolution in the aqueous solution, which is not deleterious to the underlying surface chemistry 20, 22 or patterned substrate 12. Fig. 5H depicts the flow cell 10' after the protective coating 24 is removed.

[0215] The example shown in Figs. 5I though 5L will now be described. In Fig. 5I, the initial water-soluble protective coating 24' may be selectively deposited, or patterned, such that the functionalized coating layer 20 is covered and such that the bonding region 25 of the patterned substrate 12 remains exposed. In this example, the initial water-soluble protective coating 24' is selectively deposited or patterned on the interstitial regions 16 between adjacent depressions 14, 14', but not at the edge/periphery of the patterned substrate 12 where the bonding region 25 is located. The selective deposition/patterning of the initial water-soluble protective coating 24' may be accomplished as described herein using the aqueous solution described in reference to reference numeral 104'' (which includes water and, in some instances, up to about 15% (mass to volume) of the polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, or polyethylene glycol).

[0216] The initial protective coating 24 protects the functionalized coating layer 20 during any assembly processes that are subsequently performed. As depicted in Fig. 5J, the assembly process may include bonding the lid 26 to the bonding region 25. While not shown, when the substrate wafer is used, the assembly process may include bonding and dicing. The lid 26 may be any of the materials and may have any of the configurations described herein. The lid 26 may be bonded to the bonding region 25 via any of the techniques described herein.

[0217] In the example shown in Fig. 5J, the lid 26 includes a top portion 27 integrally formed with sidewall(s) 29. The sidewall(s) 29 are bonded to the bonding region 25 of the patterned substrate 12 through the spacer layer 28. After the lid 26 is bonded, the flow channel 30 is formed between the lid 26 and the patterned substrate 12. The flow channel 30 may serve to selectively introduce various fluids to the flow cell 10'.

[0218] The initial water-soluble protective coating 24' then may be removed, as shown in Fig. 5K. Because the initial protective coating is water soluble, its removal may involve aqueous dissolution, as described in reference to reference numeral 108. Upon dissolution of the initial protective coating 24' and removal of the solution from the flow channel 30, the functionalized coating layer 20 and any of the patterned substrate 12 (e.g., interstitial regions 16 not bonded to the lid 26) that had been coated by the initial protective coating 24' are exposed.

[0219] Also shown in Fig. 5K, after the initial protective coating 24' is removed, the primer 22 may be grafted to the functionalized coating layer 20 in the depression(s) 14, 14'. Any of the primers 22 described herein may be used. In this example, grafting may be accomplished using the flow through process (reference numeral 102'''), and the primer solution or mixture described herein, which may include the primer(s), water, a buffer, and a catalyst. After the primer 22 is grafted, the flow cell 10' is formed.

[0220] If the flow cell 10' is to be shipped, or stored for some period of time, the (second) water-soluble protective coating 24 may be applied to the surface chemistry 20, 22 and on at least a portion of the patterned substrate 12 (i.e., interstitial regions 16 exposed within the channel 30), as shown in Fig. 5L. The deposition of the (second) water-soluble protective coating 24 may be accomplished using the flow through process and the aqueous solution described in reference to reference numeral 104'''. To reiterate, the water-soluble material in the aqueous solution in this example may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin). In an example, the water-soluble material may be the polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, dextran, polyacrylamide, glycols, ethylenediaminetetraacetic acid sodium salt, tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, or saline sodium citrate.

[0221] Still another example of the method (e.g., method 100 or 200) involves a non-patterned substrate 12', as shown in Figs. 6A through 6E.

[0222] Any of the substrates disclosed herein, and non-patterned substrate 12' does not include depressions 14 or interstitial regions 16. In this example method, the lid 26 is bonded to the non-patterned substrate 12' at the outset to form the flow channel(s) 30. The lid 26 may be any of the materials and in any of the configurations described herein. The lid 26 may also be bonded to the non-patterned substrate 12' via any of the techniques described herein.

[0223] In the example shown in Fig. 6B, the lid 26 includes a top portion 27 integrally formed with sidewall(s) 29. The sidewall(s) 29 are bonded to a bonding region of the non-patterned substrate 12' through the spacer layer 28. The bonding region may be at a periphery of the non-patterned substrate 12', or at any areas where it is desirable to form a boundary of a flow channel 30. In other examples, the spacer layer 28 may form the sidewall(s) and may be attached to an at least substantially planar lid 26.

[0224] Together, the lid 26 (including the sidewall(s) 29) and the non-patterned substrate 12 define the flow channel 30. The flow channel 30 may serve to, for example, selectively introduce fluids in order to form the surface chemistry 20, 22 and the protective coating 24, to remove the coating 24, and to selectively introduce reaction components or reactants to the surface chemistry 20, 22 (after the protective coating 24 is removed) in order initiate designated reactions within the flow channel 30.

[0225] Prior to forming the functionalized coating layer 20 (shown in Fig. 6C), the method may involve exposing the non-patterned substrate (via a flow through process) to a cleaning process and/or to another process (e.g., silanization) that prepares the exposed surface of the non-patterned substrate for the subsequent deposition of the functionalized molecule.

[0226] Silanization of the non-patterned substrate 12' is shown in Fig. 6B. In this example, silanization attaches the silane or the silane derivative 18 to the exposed portions of the non-patterned wafer surface 12' that are present in the flow channel 30.

[0227] Silanization may be accomplished using any silane or silane derivative 18. The selection of the silane or silane derivative 18 may depend, in part, upon the

functionalized molecule that is to be used to form the functionalized coating layer 20 (shown in Fig. 6C), as it may be desirable to form a covalent bond between the silane or silane derivative 18 and the functionalized coating layer 20. The method used to attach the silane or silane derivative 18 to the substrate 12' may be a flow through process.

[0228] As shown in Fig. 6C, in this example, the functionalized coating layer 20 is then formed on the silane or silane derivative 18, or on other chemistry that has been deposited to prepare the exposed surface of the non-patterned substrate 12' within the flow channel 30.

[0229] Any of the functionalized molecules described herein may be used. In this example, functionalized coating layer formation may be accomplished by a flow through process. In the flow through process, the functionalized molecule may be introduced into the flow channel(s) 30 through respective input port(s) and may be cured. The functionalized coating layer 20 will form on the exposed surface of the non-patterned substrate 12' and polishing does not take place.

[0230] As shown in Fig. 6D, the primer 22 is grafted to the functionalized coating layer 20 in the flow channel 30. Any of the primers described herein may be used. In this example, grafting may be accomplished by a flow through process. In the flow through process, the primer solution or mixture described herein may be introduced into the flow channel(s) 30 through respective input port(s), may be maintained in the flow channel(s) for a time sufficient (i.e., an incubation period) for the primer to attach to the functionalized coating layer 20, and then may be removed from respective output port(s). After primer attachment, the additional fluid(s) may be directed through the flow channel(s) to wash the now functionalized flow channel(s) 30.

[0231] The resulting flow cell 10'' in this example is shown in Fig. 6D. This flow cell 10'' may be used in a sequencing operation, or may be coated with the protective coating 24 for shipping and/or storage.

[0232] As shown in Fig 6E, in an example, after the primer 22 is grafted to the functionalized coating layer 20 in the flow channel 30, the water-soluble protective coating 24 is formed on the surface chemistry 20, 22. In this example, water-soluble protective coating 24 formation may be accomplished by a flow through process. In the flow through process, the aqueous solution (including water and, in some instances, up

to about 15% (mass to volume) of a water-soluble, film forming material) may be introduced into the flow channel(s) 30 through respective input port(s) and may be maintained in the flow channel(s). Enough of the aqueous solution may be introduced to cover the surface chemistry 20, 22 within the flow channel 30. While in the flow channel(s) 30, the flow cell(s) may be exposed to a dry down process where air, nitrogen, or vacuum is flushed through the input port for a set amount of time to dry the (second) water-soluble protective coating on the surface chemistry 20, 22.

[0233] In this example, the water-soluble material in the aqueous solution may be any of the examples disclosed herein (i.e., a water-soluble non-cationic synthetic polymer; a water-soluble natural polysaccharide or a derivative thereof; a water-soluble natural protein or a derivative thereof; a water-soluble salt; a water-soluble small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol; or a cyclodextrin). In an example, the water-soluble material may be any of the polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, dextran, polyacrylamide, glycols, ethylenediaminetetraacetic acid sodium salt, tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, or saline sodium citrate.

[0234] It is to be understood that primer grafting and water-soluble protective coating 24 formation may occur simultaneously in some examples.

[0235] When it is desirable to utilize the flow cell 10" (having the protective coating 24 on the surface chemistry 20, 22) in an application (e.g., a sequencing operation or a genotyping operation), the protective coating 24 may be removed via the dissolution process described in reference to reference numeral 108. The water solubility of the protective coating 24 enables it to be removed via dissolution in the aqueous solution, which is not deleterious to the underlying surface chemistry 20, 22.

[0236] Moreover, the methods disclosed herein may involve performing a quality control assay. In an example, the assay may be a CFR or a HP-TET assay or another suitable dye-based assay. The assay may be performed prior to introducing the protective coating 24 (but after primer grafting), and then again after the protective

coating 24 is removed (and the primers are re-exposed). The assay data may indicate whether any primer degradation has occurred. In an example, the method involves removing the water-soluble protective coating; and performing a dye-based assay to detect any degradation of the primer. In an example, flow cell 10, 10', 10'' disclosed herein may exhibit less than a 15% drop in CFR retention after 2 days of storage with the protective coating thereon, and in another example, less than a 10% drop in CFR retention after 2 days of storage with the protective coating thereon. In still another example, the flow cell 10, 10', 10'' may not exhibit a drop in CFR retention, but rather, may exhibit an increased CFR retention (ranging, for example, from about 1% to about 25%, or for another example, from about 5% to about 20%).

[0237] The examples disclosed herein illustrate that the protective coating 24 is not formed on bonding regions 25. In other examples, however, the protective coating 24 may be formed on the entire substrate 12, 12' surface (i.e., on the surface chemistry 20, 22, in some examples on interstitial regions 16, and on the bonding regions 25), and the lid 26 may be bonded to the substrate 12, 12' through the protective coating 24.

[0238] While not shown, it is to be understood that some examples of the flow cell 10, 10', 10'' may be affixed directly to, and thus be in physical contact with, a detection device (not shown) through one or more securing mechanisms (e.g., adhesive, bond, fasteners, and the like). The detection device may include a CMOS device (which includes a plurality of stacked layers including, for example, silicon layer(s), dielectric layer(s), metal-dielectric layer(s), metal layer(s), etc.) and optical components. The optical components may be arranged such that an optical sensor of the detection device is at least substantially aligned with, and thus is operatively associated with, a single optical waveguide of the detection device and the surface chemistry 20, 22 within a single depression 14, 14' of the flow cell 10, 10', 10''.

[0239] Also while not shown, it is to be understood that instead of being bonded to a lid 26, a functionalized substrate (with surface chemistry, 20, 22 thereon or in depression(s) 14 thereof) may be bonded to another functionalized substrate with surface chemistry, 20, 22 thereon on in depression(s) thereof. The two functionalized surfaces can face each other and can have a flow channel defined therebetween. A

spacer layer and suitable bonding method may be used to bond two of the functionalized substrates together.

[0240] The flow cells 10, 10', 10'' disclosed herein may be used in a variety of sequencing approaches or technologies, including techniques often referred to as sequencing-by-synthesis (SBS), cyclic-array sequencing, sequencing-by-ligation, pyrosequencing, and so forth. With any of these techniques and in examples using a patterned substrate, since the functional molecule layer 20 and attached sequencing primer(s) 22 are present in the functionalized depressions (i.e., 14, 14' with surface chemistry 20, 22 thereon) and not on the interstitial regions 16, amplification will be confined to the functionalized depressions. In other examples, amplification can take place across an entire flow cell lane.

[0241] As one example, a sequencing by synthesis (SBS) reaction may be run on a system such as the HiSeq®, HiSeqX®, MiSeq®, NovaSeq™, or NextSeq® sequencer systems from Illumina (San Diego, CA). In SBS, extension of a nucleic acid primer along a nucleic acid template is monitored to determine the sequence of nucleotides in the template. The underlying chemical process can be polymerization (e.g., catalyzed by a polymerase enzyme) or ligation (e.g., catalyzed by a ligase enzyme). In a particular polymerase-based SBS process, fluorescently labeled nucleotides are added to the primer 22 (thereby extending the primer 22) in a template dependent fashion such that detection of the order and type of nucleotides added to the primer 22 can be used to determine the sequence of the template. For example, to initiate a first SBS cycle, one or more labeled nucleotides, DNA polymerase, etc., may be delivered into/through the flow channel 30, 30A, etc. that houses an array of primers 22. The functionalized depressions (i.e., 14, 14' with surface chemistry 20, 22 thereon), where primer extension causes a labeled nucleotide to be incorporated, can be detected through an imaging event. During an imaging event, an illumination system (not shown) may provide an excitation light to the functionalized depressions (i.e., 14, 14' with surface chemistry 20, 22 thereon).

[0242] In some examples, the nucleotides can further include a reversible termination property that terminates further primer extension once a nucleotide has been added to the primer 22. For example, a nucleotide analog having a reversible

terminator moiety can be added to the primer 22 such that subsequent extension cannot occur until a deblocking agent is delivered to remove the moiety. Thus, for examples that use reversible termination, a deblocking reagent can be delivered to the flow channel 30, 30A, etc. (before or after detection occurs).

[0243] Wash(es) may take place between the various fluid delivery steps. The SBS cycle can then be repeated n times to extend the primer 22 by n nucleotides, thereby detecting a sequence of length n.

[0244] While SBS has been described in detail, it is to be understood that the flow cells 10, 10', 10'' described herein may be utilized with other sequencing protocol, for genotyping, or in other chemical and/or biological applications.

[0245] To further illustrate the present disclosure, examples are given herein. It is to be understood that these examples are provided for illustrative purposes and are not to be construed as limiting the scope of the disclosure.

[0246] In some of these examples, the protective coatings are dried and some of the protective coatings are wet. The wet coatings are referred to as "wet storage" or "wet stored". While the coatings that were exposed to a drying process were visibly dry and appeared solid, it is to be understood that traces of moisture may have been present. The wet coatings were in liquid form.

NON-LIMITING WORKING EXAMPLES

[0247] ***Example 1***

[0248] The example flow cells (1A and 1B) included several flow channels/lanes defined on a patterned silicon and/or a tantalum oxide substrate, where each lane was in fluid communication with a plurality of wells. A PAZAM layer was formed in each well, and 1 μm primers were grafted on the PAZAM layer. Protective coatings were ultimately formed on the surface chemistry (as described below). Some of the protective coatings were dried and one of the protective coatings was wet.

[0249] A comparative flow cell was tested. The surface chemistry was the same as the example flow cells. No protective coating was formed on the surface chemistry of the comparative flow cell.

[0250] A first HP-TET quality control assay was performed in each of the lanes of each of the example and comparative flow cell before the protective coatings were added to the example flow cells. HP or hairpin defines the secondary structure part of the DNA molecule used to probe the primers on the grafted flowcell surface, and TET (or TET+DNA) is a dye labeled oligonucleotide having complementary sequence to the primers used. TET was hybridized to the primers, the excess TET was washed away, and the fluorescence of the attached dye was measured by fluorescence detection.

[0251] After the first HP-TET assay, several aqueous solutions of a polyvinyl alcohol/polyethylene glycol graft copolymer (in this example KOLLICOAT® IR) were prepared. Each solution had a different concentration of the copolymer ranging from 0.10 % to 10% (mass to volume). A flow through process was used to introduce one of the aqueous solutions into one of the lanes of each of the flow cells, and onto the PAZAM layers and primers within the respective lanes. The aqueous solution was dried to form a protective coating. Each of the dried protective coatings had a different concentration of the copolymer. These example cells and the comparative cell were dried using nitrogen gas.

[0252] The dried example flow cells were then dry stored for 3 days at 60°C (equivalent to dry storage for 1 month at 25°C or ambient conditions) with the protective coating in place.

[0253] The comparative uncoated flow cell ("uncoated") was exposed to the same dry storage conditions.

[0254] Another protective coating was formed by introducing a liquid sodium chloride-sodium citrate (SSC) buffer using a flow through process. Thus, this example flow cell was exposed to wet storage conditions in which the sodium chloride-sodium citrate (SSC) buffer at 4°C was left to soak on the flow cell surface chemistry.

[0255] After storage, the protective coating was removed from the lanes of the respective example flow cells via aqueous dissolution during washing, and the wet stored flow cell was rinsed out. Another HPTET quality control assay was performed in each of the lanes of each of the example and comparative flow cells.

[0256] The HP-TET retention rate results were calculated using the before coating and after storage HP-TET results. The retention rates are shown in Fig. 7. As

depicted, the HP-TET retention generally increased with increasing copolymer concentrations for both flow cells 1A and 1B. At or below 1%, the copolymer coating was not as effective as the comparative example or the wet storage. This example illustrates that at copolymer concentrations ranging from about 2.5% to about 10%, the protective coating improves the dry storage stability of the surface chemistry compared to uncoated surface chemistry stored at the same dry conditions and approaches the stability of the wet storage conditions.

[0257] ***Example 2***

[0258] The example flow cells (two of each of 2A through 2H) included one lane defined on a tantalum oxide coated silicon substrate. A PAZAM layer was formed in the lane, and 1 μm primers were grafted on the PAZAM layer. Protective coatings were ultimately formed on the surface chemistry. As described below, two example flow cells were tested for each type of protective coating, and the results in Fig. 8 are the average for the two flow cells having the same type of protective coating. Some of the protective coatings were dried (2A through 2H) and one of the protective coatings was wet.

[0259] A comparative flow cell was tested. The surface chemistry was the same as the example flow cells. No protective coating was formed on the surface chemistry of the comparative flow cell.

[0260] A first HP-TET quality control assay was performed in each of the lanes of the example and comparative flow cells before the protective coatings were added to the example flow cells.

[0261] After the first HP-TET assay, several aqueous solutions of different potential protective coating materials were prepared. The solutions were prepared with ethylenediaminetetraacetic acid sodium salt (EDTA) (0.1 wt%), glycerol (1 wt%), hydroquinone (0.1 wt%), KOLLICOAT® IR (1%), polyethylene glycol 3000 (1%), (tris(2-carboxyethyl)phosphine) (TCEP) (0.1 wt%), Tris Base (pH 7-8) (100 mM), and TWEEN® 20 (a nonionic emulsifying agent) (1%).

[0262] A flow through process was used to introduce one of the aqueous solutions into two of the example flow cells (i.e., the EDTA solution was in two flow cells (collectively referred to as 2A), the glycerol solution was in another two flow cells

(collectively referred to as 2B), the hydroquinone solution was in two more flow cells (collectively referred to as 2C), the KOLLICOAT® IR solution was in two other flow cells (collectively referred to as 2D), the polyethylene glycol 3000 solution was in two additional flow cells (collectively referred to as 2E), the TCEP solution was in two more flow cells (collectively referred to as 2F), the Tris Base solution was in still two more flow cells (collectively referred to as 2G), and the TWEEN® 20 solution was in two other flow cells (collectively referred to as 2H)), and onto the PAZAM layers and primers within the respective flow cells. The aqueous solutions were dried to form protective coatings. Drying of the example and comparative cells was performed using nitrogen gas.

[0263] The example flow cells were then dry stored for 3 days at 60°C (equivalent to dry storage for 1 month at 25°C or ambient conditions) with the protective coating in place.

[0264] The comparative uncoated flow cell (C1) was exposed to the same dry storage conditions.

[0265] Another protective coating was formed by introducing a liquid sodium chloride-sodium citrate (SSC) buffer using a flow through process. Thus, this example flow cell (WC) was exposed to wet storage conditions in which the sodium chloride-sodium citrate (SSC) buffer at 4°C was left to soak on the flow cell surface chemistry.

[0266] After storage, the protective coating was removed from the lanes of the example flow cell via aqueous dissolution during washing, and the wet stored flow cell (WC) was rinsed out. Another HP-TET quality control assay was performed in each of the lanes of each of the example and comparative flow cells.

[0267] The HP-TET retention results were calculated using the before coating and after storage HP-TET results. The retention data are shown in Fig. 8. As depicted, the HP-TET retention for the flow cells with coatings of glycerol (2B), the copolymer (2D), the polyethylene glycol (2E), and the TCEP (2F) were comparable with or better than the dry stored comparative example (C1) stored at similar conditions. The results for coatings 2A, 2C, 2G and 2H indicate that these coatings, at these concentrations and/or applied by a flow through process, may hinder the accessibility of the primers and/or may not prevent degradation of the surface chemistry as well as the coatings 2B

and 2D-2F. It is believed that the coatings 2A, 2C, 2G, and 2H may perform better at different concentrations and/or when applied by a different coating process.

[0268] ***Example 3***

[0269] The flow cells used in this Example each included a non-patterned glass substrate with 4 lanes defined thereon. In one of the flow cells, two of the lanes (collectively referred to as 3A) were used as example lanes, and two of the lanes (collectively referred to C2A) were used as comparative example lanes. In another of the flow cells, two of the lanes (collectively referred to as 3B) were used as example lanes, and two of the lanes (collectively referred to C2B) were used as comparative example lanes. In a third of the flow cells, two of the lanes (collectively referred to as 3C) were used as example lanes, and two of the lanes (collectively referred to as C2C) were used as comparative example lanes.

[0270] A PAZAM layer was formed in each lane, and primers were grafted on the PAZAM layer. In the example lanes (3A, 3B, 3C), KOLLICOAT® IR protective coatings were formed on the surface chemistry using a flow through process. In the comparative example lanes (C2A, C2B, C2C), no protective coating was formed. Drying of the example lanes and the comparative example lanes was performed by flowing nitrogen gas through each lane for 30 seconds.

[0271] The flow cell with example flow lanes 3A and comparative flow lanes C2A were stored with 84% relative humidity in the packaging for 7 days and 14 days. The flow cell with example flow lanes 3B and comparative flow lanes C2B were stored with 5% relative humidity in the packaging for 7 days and 14 days.

[0272] These flow cells were removed from storage and were used for sequencing. The example lanes 3A, 3B and the comparative example lanes C2A, C2B were exposed to wash steps during sequencing. It is believed that washing at least partially removes the protective coating from the example lanes 3A, 3B. However, sequencing may also be performed without removing the protective coating.

[0273] Fig. 9 shows a plot of the fluorescence intensity for the lanes (3A, 3B, C2A, C2B) after the first sequencing cycle. The data show that the cycle 1 intensities were minimally impacted by the humidity storage conditions when the protective coating

was in place, compared to when the protective coating was not used (i.e., compare 3A with C2A and compare 3B with C2B). The cycle 1 intensities for the uncoated lanes C2A, C2B indicate that the surface chemistry decayed as a result of the humidity exposure. The effect on intensity of 5% humidity (flow lanes 3B) was also much less than the effect observed at 84% humidity (flow lanes 3A).

[0274] The flow cell with example flow lanes 3CA and comparative flow lanes C2C were stored under external temperature fluctuations for between 2 days and 19 days. The temperature profile ramp ranged from about -23°C to about 60°C during each 24 hour period over the respective time period. From hour 0 to about hour 8, the temperature was maintained between about -23°C to -25°C. The temperature was then ramped up to about 60°C and maintained for about 8 hours. The temperature was then ramped back down to between about -23°C to -25°C, and from about hour 18 to hour 24, the temperature was maintained between about -23°C to -25°C.

[0275] This flow cell was removed from storage and was used for sequencing. The example lanes 3C and the comparative example lanes C2C were exposed to wash steps during sequencing, and these steps are believed to at least partially remove the protective coating from the example lanes 3C (although sequencing can be performed before without protective coating removal).

[0276] Fig. 10 shows a plot of the fluorescence intensity for the lanes (3C, C2C) after the first sequencing cycle. The data show that the cycle 1 intensity was minimally impacted by external temperature fluctuations when the protective coating was in place, compared to when the protective coating was not used (i.e., compare 3C with C2C. The cycle 1 intensity for the uncoated lanes C2C indicates that the surface chemistry decayed as a result of the external temperature fluctuations.

[0277] ***Example 4***

[0278] Two open flow cells (i.e., no lids were attached) were used in this example. Each open flow cell included a single lane (4A, 4B) defined on a patterned silicon substrate, where each lane was in fluid communication with a plurality of wells. A PAZAM layer was formed in each well, and 1 μm primers were grafted on the PAZAM layer.

[0279] Prior to forming any protective coatings, the two open flow cells were exposed to sequencing.

[0280] KOLLICOAT® IR protective coatings were then formed on half of the surface chemistry of each open flow cell using a dip coating process. The open flow cells were dried in nitrogen gas. As such, in this example, the coated halves are referred to as examples 4A and 4B and the uncoated halves are referred to as comparative examples C3 and C4.

[0281] After the protective coatings were formed, the sample flow cell including coated side 4A and comparative uncoated side C3 was exposed to 6 days of open storage at 60°C. In open storage, the protective coating and the exposed surface chemistry of the uncoated side were directly exposed to the 60°C temperature. The sample flow cell including coated side 4B and comparative uncoated side C4 was exposed to 7 days of open storage at room temperature.

[0282] The entire flow cells (i.e., previously coated halves 4A and 4B) and uncoated halves (C3 and C4) were removed from storage used for sequencing. The protective coatings were believed to be at least partially removed from the coated halves 4A and 4B via aqueous dissolution during washing step(s).

[0283] Fig. 11A shows a plot of the fluorescence intensity after the first sequencing cycle for the flow cell including 4A and C3 at time T0 (i.e., pre-coating on 4A) and after storage at time T1. The data show that the cycle 1 intensity was minimally impacted by the open storage at 60°C when the protective coating was in place, compared to when the protective coating was not used (i.e., compare 4A with C3). The cycle 1 intensity for the uncoated half C3 indicates that the surface chemistry decayed as a result of the open storage.

[0284] Fig. 11B shows two plots – the top illustrates the percentage of clusters passing through a filter (%passing filter (PF)) and the bottom illustrates the fluorescence intensity after the first sequencing cycle for the flow cell including 4B and C4 at time T0 (i.e., pre-coating on 4B) and two runs taken after storage at time T1. %Passing filter (PF) is the metric used to describe clusters which pass a chastity threshold and are used for further processing and analysis of sequencing data. Higher %passing filter result in increased yield of unique clusters used for sequencing data. The data in Fig.

11B shows that the %passing filter was improved when the protective coating was used (compare 4B to C4 at times T0 and both T1 data points). The data in Fig. 11B also shows that cycle 1 intensity was minimally impacted by the open storage at room temperature when the protective coating was in place, compared to when the protective coating was not used (i.e., compare 4B with C4 at times T0 and both T1 data points). The cycle 1 intensity for the uncoated half C4 indicates that the surface chemistry decayed as a result of the open storage, even at room temperature.

[0285] ***Example 5***

[0286] An open patterned glass wafer had a PAZAM layer formed in each of its wells. After polishing, the wafer was diced, and a KOLLICOAT® IR protective coating was spray coated on one of the diced cells. A comparative diced cell was formed in a similar manner (with a PAZAM coating), except that the KOLLICOAT® IR protective coating was not formed.

[0287] The example and comparative diced cells were stored for 1 hour at 20°C, 60°C, and 80°C. After storage, the protective coating was removed from the example diced cell via aqueous dissolution during washing. 1 µm primers were then grafted on the PAZAM layers of the example and comparative diced cells to form, respectively an example flow cell and a comparative flow cell.

[0288] A CFR assay was performed to determine whether grafting of the primers was impacted by the different storage conditions. During a CFR assay, primer grafted surfaces are exposed to fluorescently tagged (Cal Fluor Red) complementary oligos in a buffer solution. These oligos bind to surface bound primers and excess CFR is washed off. The surface is then scanned in a fluorescent detector to measure CFR intensity on the surface to provide a quantitative measure of primers' concentration and health on the surface. After measurement, the oligos are removed with a mild base solution and surfaces are rescanned to confirm all CFR was removed.

[0289] The median intensity results for the example flow cell are shown in Fig. 12A and the median intensity results for the comparative flow cell are shown in Fig. 12B. Clearly, the ability of the uncoated PAZAM layer of the comparative flow cell to graft the primers was deleteriously affected at exposure to 60°C or higher temperatures

(Fig. 12B). The coated PAZAM layer of the example flow cell was able to graft the primers better at all tested temperatures than the uncoated PAZAM layer of the comparative diced cell (compare Figs. 12A and 12B). While median intensity decreased for both the example and comparative flow cells at 80°C, the example flow cell seemed to degrade slower compared to the comparative flow cell.

[0290] ***Example 6***

[0291] The flow cells used in this Example each included a non-patterned glass substrate with 4 lanes defined thereon. In each of the flow cells, two of the lanes were example lanes having the protective coating formed thereon, and two of the lanes were comparative lanes that did not have the protective coating formed thereon.

[0292] In each lane of each cell, a PAZAM layer was formed, and primers were grafted on the PAZAM layer using a flow through process. In each of the two example lanes of each flow cell, a KOLLICOAT® IR protective coating was formed on the surface chemistry using a flow through process. In each of the two comparative example lanes of each flow cell, the protecting coating was not formed. Drying of the example lanes and the comparative lanes was performed by flowing nitrogen gas through each lane for 30 seconds.

[0293] Each flow cell (including two example lanes and two comparative example lanes) was stored at a particular temperature, namely 25°C, 40°C, 60°C, or 80°C. The number of days for which the cells were stored varied from 1 day for up to 120 days.

[0294] The flow cells were removed from storage and were used for sequencing. The example lanes and the comparative example lanes were exposed to wash steps during sequencing, and these steps are believed to at least partially remove the protective coatings from the example lanes (although sequencing can be performed without such removal).

[0295] Fig. 13 shows a plot of the average fluorescence intensity for the sets of lanes (example (i.e., 6) or comparative (i.e., C5)) after the first sequencing cycle. The intensity data is shown for example lanes and comparative example lanes, and the data is plotted by the temperature at which the cell was stored and the number of days for which the cell was stored. The data clearly show that the cycle 1 intensities were

minimally impacted by the elevated temperature storage conditions when the protective coating was in place, regardless of the number of storage days, compared to when the protective coating was not used (i.e., compare data labeled "6" with comparative data labeled "C5". The cycle 1 intensities for the uncoated comparative example lanes indicate that the surface chemistry decayed as a result of the temperature exposure.

[0296] ***Example 7***

[0297] KOLLICOAT® IR was spray coated onto a patterned CMOS wafer having a TaO_x surface. The number of spray passes varied across the patterned CMOS wafer to determine the effect on the thickness of the resulting coating. The results are shown in Fig. 14 as the distance across the protective coating on the wafer in mm (x-axis) versus the distance of the protective coating thickness in µm (y-axis). The thickness of the copolymer coating increased slightly as the number of passes was increased across the wafer, and then decreased slightly as the number of passes was decreased. This data indicates that spray coating may be used to obtain a protective coating with a controlled thickness.

[0298] ***Example 8***

[0299] An open patterned CMOS wafer had a PAZAM layer formed in each of its wells. After polishing, the wafer was diced to 700 nm cells, and a KOLLICOAT® IR protective coating was spray coated on three of the diced cells. Comparative diced cells were formed in a similar manner (with a PAZAM coating), except that the KOLLICOAT® IR protective coating was not formed.

[0300] One example diced cell and one comparative diced cell were stored for 28 days, or 35 days, or 71 days at room temperature (i.e., ~25°C). After storage, the protective coatings were removed from the example diced cell via aqueous dissolution during washing. 1 µm primers were then grafted on the PAZAM layers of the example and comparative diced cells to form, respectively example flow cells (9A (28 day storage), 9B (35 day storage), 9C (71 day storage)) and comparative flow cells (C7A (28 day storage), C7B (35 day storage), C7C (71 day storage)).

[0301] Sequencing was then performed on each of the cells. The wash step(s) removed the protective coating from the example flow cells.

[0302] Fig. 15 shows a plot of the fluorescence intensity after the first sequencing cycle. At each of the storage periods, the comparative flow cells C7A, C7B, C7C (stored without the protective coating) exhibited reduced intensity when compared to the example flow cells 9A, 9B, 9C (stored with the protective coating). The cycle 1 intensities for the uncoated flow cells C7A, C7B, C7C indicate that the surface chemistry decayed as a result of the exposure, even at room temperature.

[0303] ***Example 9***

[0304] An open patterned silicon wafer had a PAZAM layer formed in each wells. After polishing, the wafer was diced to form cells, and an initial KOLLICOAT® IR protective coating was selectively spray coated on two of the diced cells, so that a bonding region remained exposed. Comparative diced cells were formed in a similar manner (with a PAZAM coating), except that the KOLLICOAT® IR protective coating was not formed. Lids were bonded to the respective bonding regions of the example and comparative diced cells to form flow cells. The initial KOLLICOAT® IR protective coatings were then removed from the PAZAM layers within the example flow cells via aqueous dissolution using a flow through process.

[0305] Primers were then grafted to each of the PAZAM layers within each of the example and comparative example flow cells using a flow through process.

[0306] After primer grafting, another KOLLICOAT® IR protective coating was formed on the primers and PAZAM layers within the example flow cells via a flow through process and drying.

[0307] The example flow cells (10A and 10B) were then dry stored for 2 weeks or 6 weeks at 30°C with the protective coating in place. Two comparative uncoated flow cells (C8A, C8B) were exposed to the same dry storage conditions (2 week storage or 6 week storage at 30°C), and the another example flow cell (WC2) was exposed to wet storage conditions for 6 weeks, during which sodium chloride-sodium citrate (SSC) buffer was left to soak at 4°C on the flow cell surface chemistry.

[0308] After storage, sequencing was then performed on each of the cells. During washing step(s), the protective coating was believed to be at least partially removed from the example flow cells 10A, 10B, and the SSC buffer was rinsed from the wet stored flow cell WC2.

[0309] Fig. 16 shows a plot of the fluorescence intensity after the first sequencing cycle. At each of the storage periods, the comparative flow cells C8A, C8B (stored at dry conditions without the protective coating) exhibited reduced intensity when compared to the example flow cells 10A, 10B (stored with the protective coating). At 6 weeks, the dry protective coating provided as good of protection for the surface chemistry as the wet storage protective coating (compare 10B with WC2).

[0310] ***Example 10***

[0311] Open faced patterned tantalum oxide coated substrates had a PAZAM layer formed in each of the wells, and primers were grafted on the PAZAM layers using a flow through process. Comparative flow cells were also tested. The surface chemistry was the same as the example flow cells. No protective coatings were formed on the surface chemistry of the comparative flow cells.

[0312] A first CFR assay was performed prior to any protective coatings being formed on the example flow cells.

[0313] After the first CFR assay, several aqueous solutions of different potential protective coating materials were prepared. The solutions were prepared with ethylenediaminetetraacetic acid sodium salt (EDTA), polyethylene glycol, (tris(2-carboxyethyl)phosphine) (TCEP), tris(3-hydroxypropyltriazolylmethyl)amine (THPTA), tris(hydroxymethyl)aminomethane with EDTA (TE buffer), and bathophenanthrolinedisulfonic acid disodium salt (Bath). Each solution had a concentration as shown in Table 1. The aqueous solutions were applied by puddling the solutions on the surfaces of the example flow cells and the solutions were allowed to evaporate to form the coatings. The example flow cells were then dry stored for 11 days at 4°C or at room temperature. Another example flow cell was a wet storage cell. The following Table 1 identifies the various cells tested and the conditions at which they were tested.

TABLE 1

		Storage Condition	
Identifier	**Protective Coating**	**4°C**	**Room Temp. (RT)**
WC3	wet storage	yes	no
C11 (comparative example)	none	yes	no
11A	0.5M EDTA	yes	no
11B	4.4 wt% PEG	yes	no
11C	0.8 wt% TCEP	yes	no
11D	1.0 wt% THPTA	yes	no
11E	1x TE buffer	yes	no
C12 (comparative example)	none	no	yes
11F	1x TE buffer	no	yes
11G	1.6 wt% Bath	no	yes
11H	0.5M EDTA	no	yes
11I	4.4 wt% PEG	no	yes
11J	0.8 wt% TCEP	no	yes
11K	1.0 wt% THPTA	no	yes

[0314] The comparative uncoated flow cell C11 and C12 exposed to the same dry storage conditions, respectively, as the example cells stored at dry 4°C and at dry room temperature. The other flow cell WC3 was exposed to wet storage conditions in which sodium chloride-sodium citrate (SSC) buffer was left to soak at 4°C on the flow cell surface chemistry.

[0315] After storage at the respective conditions, the protective coating was removed from each of the example flow cells (11A through 11K) via aqueous dissolution during washing, and the wet stored flow cell WC3 was rinsed out. Another CFR quality control assay was performed in each of the example and comparative flow cells.

[0316] The results shown in Fig. 17 are the 11 day CFR results divided by the T0 CFR results for each of the examples and comparative examples. As depicted, the CFR retention for the flow cells with each of the dry protective coatings were better than the dry stored comparative examples stored at similar conditions, and were on par with the wet storage conditions.

[0317] ***Example 11***

[0318] Open faced patterned silicon substrates had a PAZAM layer formed in each of its wells. After polishing, 1 μm primers were then grafted on the PAZAM layers. A comparative example was formed in a similar manner.

[0319] A first CFR assay was performed prior to any protective coatings being formed on the example open faced flow cells. This data is shown in Fig. 18 as T0.

[0320] After the first CFR assay, an aqueous solution of KOLLICOAT® IR was prepared and used to form spray coated protective coatings on the example open faced cells.

[0321] The example open face flow cells were then dry stored for 1 day or 2 days at 60°C.

[0322] Sequencing was then performed on each of the open faced cells after their respective storage periods, and the protective coatings were removed during washing step(s).

[0323] Fig. 18 shows a bar graph of the CFR retention (%, ratio of the CFR signal at day X (X = 1 or 2) divided by the signal at day 0) after the first sequencing cycle was performed following 1 storage day and 2 storage days. At each of the storage periods, the comparative flow cells C13 (stored without the protective coating) exhibited significantly reduced CFR retention when compared to the example flow cells 12 (stored with the protective coating). After one day of storage, the CFR of the comparative flow cell dropped 40%.

[0324] ***Example 12***

[0325] A patterned CMOS wafer had a PAZAM layer formed in each of its wells. After polishing, KOLLICOAT® IR coatings of different thicknesses were spray coated on the patterned CMOS wafer and in respective wells (and the PAZAM layers therein) of the patterned CMOS wafer. The solution used to selectively spray the KOLLICOAT® IR coatings included about 5% KOLLICOAT® IR, about 5% ethanol, and about 90% water. The thicknesses of the various coatings ranged from 1.708 μm to 11.73 μm. Another portion of the wafer remained uncoated, and was used as a comparative example.

[0326] The wafer (including example portions and comparative example portion) was then dry stored at 40°C for 2 days.

[0327] The respective coatings were removed from the wafer via aqueous dissolution, and primers were then dunk grafted on the PAZAM layers in the wells. A CFR assay was then performed.

[0328] The % intensity increase, with respect to the comparative example portion, is shown for each of the example portions of the wafer that had been coated with a KOLLICOAT® IR coating. The results are shown in Fig. 19 and are labeled by the thickness of the KOLLICOAT® IR coating that has been used. The results indicate that the thickness of the protective coating does not seem to affect the functionality in preserving the performance of the PAZAM layer to graft primers after the protective coating is removed.

[0329] ***Example 13***

[0330] The example flow cell included eight lanes defined on a patterned silicon substrate, where each lane was in fluid communication with a plurality of wells. A PAZAM layer was formed in each well, and 1 μm primers were grafted on the PAZAM layer.

[0331] Protective coatings were ultimately formed on the surface chemistry in four of the eight lanes. More particularly, four of the eight lanes were coated with a solution including about 5% KOLLICOAT® IR, about 5% ethanol, and about 90% water. The cell was dried using vacuum followed by 30 seconds of nitrogen gas exposure. The flowcell was stored for 2 days at 60°C.

[0332] The other four lanes were comparative flow lanes. The surface chemistry in the comparative flow lanes was the same as the example flow lanes, except that o protective coatings were formed on the surface chemistry of the comparative flow lanes.

[0333] After storage, sequencing was performed in each of the example flow lanes (collectively referred to as 13) and the comparative example flow lanes (collectively referred to as C14). The protective coating was removed using an aqueous wash as part of the sequencing.

[0334] Fig. 20 shows a plot of the average fluorescence intensity for the example lanes (13) and the comparative example lanes (C14) for two reads after the first sequencing cycle. As depicted, the protective coating protected the example lanes of the flow cell against a significant decline in C1 intensity.

[0335] ***Example 14***

[0336] The example flow cell included eight lanes defined on a patterned silicon substrate, where each lane was in fluid communication with a plurality of wells. A PAZAM layer was formed in each well, and 1 μm primers were grafted on the PAZAM layer.

[0337] A primer graft mix was prepared with varying amounts of KOLLICOAT® IR ranging from 5% to 0.1%, and was randomized among the eight lanes on the flow cell. This graft mix was allowed to incubate and then the cell was dried using vacuum followed by 30 seconds of nitrogen gas exposure in each hole/port.

[0338] The protective coatings were removed via aqueous dissolution, and a CFR assay was performed.

[0339] Fig. 21 shows a plot of the relative CFR versus the percentage of KOLLICOAT® IR. The relatively consistent CFR data indicates that the primers may be grafted at the same time that the protective coating is deposited. This example provides evidence that the coating does not deleteriously affect the primer deposition chemistry used in this example.

[0340] ***Example 15***

[0341] The three flow cells in this Example included eight lanes defined on a patterned silicon substrate, where each lane was in fluid communication with a plurality of wells. A PAZAM layer was formed in each well, and 1 μm primers were grafted on the PAZAM layer.

[0342] Polyacrylamide protective coatings were formed on the surface chemistry in one of the eight lanes. Comparative coatings (different water-soluble cationic polymers) were formed on the surface chemistry in five of the eight lanes, and one of the eight lanes was left uncoated. The comparative coatings were different cationic

polymers. The following Table 2 identifies the various cells tested and the concentrations at which they were tested.

TABLE 2

		Concentration (%)		
Identifier	**Protective Coating**	**FC1**	**FC2**	**FC3**
No Coating (comparative example)	None	NA	NA	NA
14	Polyacrylamide	0.01	0.1	1
C15A (comparative example)	Luviquat® HOLD	0.01	0.1	1
C15B (comparative example)	Luviquat® FC 370	0.01	0.1	1
C15C (comparative example)	Luviquat® FC 550	0.01	0.1	1
C15D (comparative example)	Polyquaternium-10	0.01	0.1	1
C15E (comparative example)	Poly(ethylene imine) (PEI)	0.01	0.1	1

[0343] The coatings were formed via a flow through process, and were dried using 30 seconds of nitrogen gas exposure. The cells were washed.

[0344] Clustering was performed in each of the flow cell lanes. A sequencing library was loaded into each flow lane, and fragments were captured by the complementary primers. Each fragment was amplified into distinct clonal clusters. The intensity was measured, and Fig. 22 illustrates the results as arbitrary fluorescence units, which quantifies the amount of double stranded DNA present on the flow cell lane surfaces. As depicted, the protective coating protected the example lane, while the cationic polymers hindered clustering (compare 14 to each of C15A through C15E). The cationic polymers were likely bound to the PAZAM and/or the primer surface chemistry.

Additional Notes

[0345] It should be appreciated that all combinations of the foregoing concepts and additional concepts discussed in greater detail below (provided such concepts are not mutually inconsistent) are contemplated as being part of the inventive subject matter

disclosed herein. In particular, all combinations of claimed subject matter appearing at the end of this disclosure are contemplated as being part of the inventive subject matter disclosed herein. It should also be appreciated that terminology explicitly employed herein that also may appear in any disclosure incorporated by reference should be accorded a meaning most consistent with the particular concepts disclosed herein.

[0346] Reference throughout the specification to "one example", "another example", "an example", and so forth, means that a particular element (e.g., feature, structure, and/or characteristic) described in connection with the example is included in at least one example described herein, and may or may not be present in other examples. In addition, it is to be understood that the described elements for any example may be combined in any suitable manner in the various examples unless the context clearly dictates otherwise.

[0347] It is to be understood that the ranges provided herein include the stated range and any value or sub-range within the stated range. For example, a range from about 200 mm to about 300 mm, should be interpreted to include not only the explicitly recited limits of from about 200 mm to about 300 mm, but also to include individual values, such as about 208 mm, about 245 mm, about 275.5 mm, etc., and sub-ranges, such as from about 225 mm to about 290 mm, from about 235 mm to about 280 mm, etc. Furthermore, when "about" and/or "substantially" are/is utilized to describe a value, they are meant to encompass minor variations (up to +/- 10%) from the stated value.

[0348] While several examples have been described in detail, it is to be understood that the disclosed examples may be modified. Therefore, the foregoing description is to be considered non-limiting.

What is claimed is:

1. A method, comprising:

adding surface chemistry to a portion of a flow cell substrate; and

applying a water-soluble protective coating on at least the surface chemistry.

2. The method as defined in claim 1 wherein:

adding the surface chemistry involves:

forming a functionalized coating layer; and

grafting a primer to the functionalized coating layer; and

the water-soluble protective coating is applied after the primer is grafted.

3. The method as defined in claim 2 wherein:

the water-soluble protective coating is patterned to define a bonding region of the flow cell substrate after the water-soluble protective coating is applied; and

the method further comprises bonding a lid to the defined bonding region of the flow cell substrate to form a flow channel.

4. The method as defined in claim 3 wherein the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

5. The method as defined in claim 1 wherein:

adding the surface chemistry involves forming a functionalized coating layer; and

the water-soluble protective coating is applied after the functionalized coating layer is formed.

6. The method as defined in claim 5 wherein the water-soluble protective coating is selected from the group consisting of a polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, polyacrylamide, and polyethylene glycol.

7. The method as defined in claim 5 wherein:

the water-soluble protective coating is patterned to define a bonding region of the flow cell substrate after the water-soluble protective coating is formed; and

the method further comprises bonding a lid to the bonding region of the flow cell substrate to form a flow channel.

8. The method as defined in claim 7 wherein after bonding, the method further comprises:

removing the water-soluble protective coating, thereby exposing the functionalized coating layer and an other portion of the flow cell substrate;

grafting a primer to the functionalized coating layer; and

forming a second water-soluble protective coating on the primer, the functionalized coating layer and the other portion of the patterned flow cell substrate.

9. The method as defined in claim 8 wherein removing the water-soluble protective coating involves a dissolution process.

10. The method as defined in claim 8 wherein the second water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

11. The method as defined in claim 1 wherein the water-soluble protective coating is selected from the group consisting of a polyvinyl alcohol/polyethylene glycol graft copolymer, sucrose, dextran, polyacrylamide, glycols, ethylenediaminetetraacetic

acid sodium salt, tris(hydroxymethyl)aminomethane with ethylenediaminetetraacetic acid, (tris(2-carboxyethyl)phosphine), tris(3-hydroxypropyltriazolylmethyl)amine, bathophenanthrolinedisulfonic acid disodium salt, hydroxyl functional polymers, glycerol, and saline sodium citrate.

12. The method as defined in claim 1 wherein applying the water-soluble protective coating involves applying an aqueous solution including up to about 15% (mass to volume) of a water-soluble material.

13. The method as defined in claim 1 wherein applying the water-soluble protective coating involves flow through deposition, dip coating, spin coating, spray coating, ultrasonic spray coating, doctor blade coating, aerosol printing, or inkjet printing.

14. The method as defined in claim 1 wherein the surface chemistry includes a primer, and wherein the method further comprises:

at least partially removing the water-soluble protective coating; and

performing a dye-based assay to detect any degradation of the primer.

15. The method as defined in claim 1, further comprising:

bonding a lid to a bonding region of the flow cell substrate to form a flow channel; and

then adding the surface chemistry and applying the water-soluble protective coating.

16. A method, comprising:

attaching a silane or a silane derivative to a surface of a patterned substrate including depressions separated by interstitial regions, thereby forming silanized depressions and silanized interstitial regions;

forming a functionalized coating layer in the silanized depressions and on the silanized interstitial regions;

polishing the functionalized coating layer from the silanized interstitial regions;

grafting a primer to the functionalized coating layer in the silanized depressions to form functionalized depressions; and

forming a water-soluble protective coating on the functionalized depressions and at least a portion of the patterned substrate.

17. The method as defined in claim 16 wherein:

the water-soluble protective coating is formed after the primer is grafted;

the water-soluble protective coating is patterned such that a bonding region of the patterned substrate remains exposed after the water-soluble protective coating is formed; and

the method further comprises bonding a lid to the bonding region of the patterned substrate to form a flow channel that is in selective fluid communication with at least some of the functionalized depressions.

18. The method as defined in claim 17 wherein the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

19. The method as defined in claim 16 wherein after the functionalized coating layer is polished and before i) the primer is grafted and ii) the water-soluble protective coating is formed, the method further comprises:

patterning an initial water-soluble protective coating on the functionalized coating layer and on the patterned substrate such that a bonding region of the patterned substrate remains exposed;

bonding a lid to the bonding region of the patterned substrate to form a flow channel that is in selective fluid communication with at least some of the depressions; and

removing the initial water-soluble protective coating.

20. A flow cell, comprising:

a patterned substrate, including:

depressions separated by interstitial regions; and

surface chemistry positioned in the depressions;

a lid bonded to a bonding region of the patterned substrate, wherein the lid at least partially defines a flow channel that is in selective communication with the depressions; and

a water-soluble protective coating covering the surface chemistry in the depressions and at least a portion of the patterned substrate.

21. The flow cell as defined in claim 20 wherein the water-soluble protective coating is selected from the group consisting of a non-cationic synthetic polymer; a natural polysaccharide or a derivative thereof; a natural protein or a derivative thereof; a water-soluble salt; a small molecule compound selected from the group consisting of a water-soluble surfactant, a sugar, an antioxidant, a chelator, a buffer, a glycol, and a cyclodextrin; and combinations thereof.

22. The flow cell as defined in claim 20 wherein a thickness of the water-soluble protective coating is at least about 50 nm.

23. The flow cell as defined in claim 20 wherein the surface chemistry includes a functionalized coating layer and a primer grafted to the functionalized coating layer.

24. A flow cell, comprising:

a non-patterned substrate;

a lid bonded to a bonding region of the non-patterned substrate, wherein the lid and the non-patterned substrate at least partially define a flow channel;

surface chemistry positioned on the non-patterned substrate and in the flow channel; and

a water-soluble protective coating covering the surface chemistry.

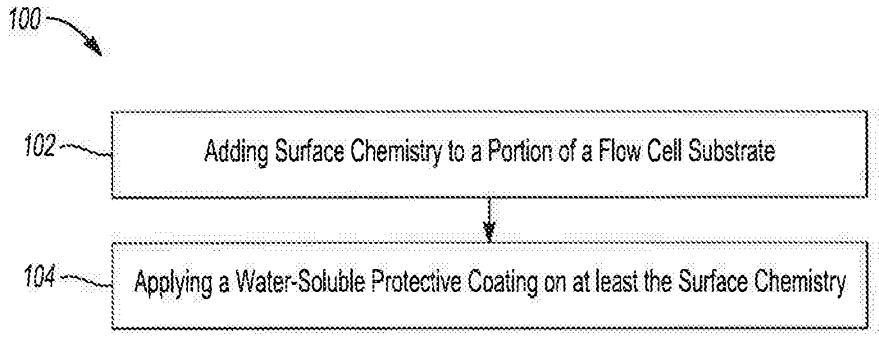

100
102
Adding Surface Chemistry to a Portion of a Flow Cell Substrate
104
Applying a Water-Soluble Protective Coating on at least the Surface Chemistry

Fig-1

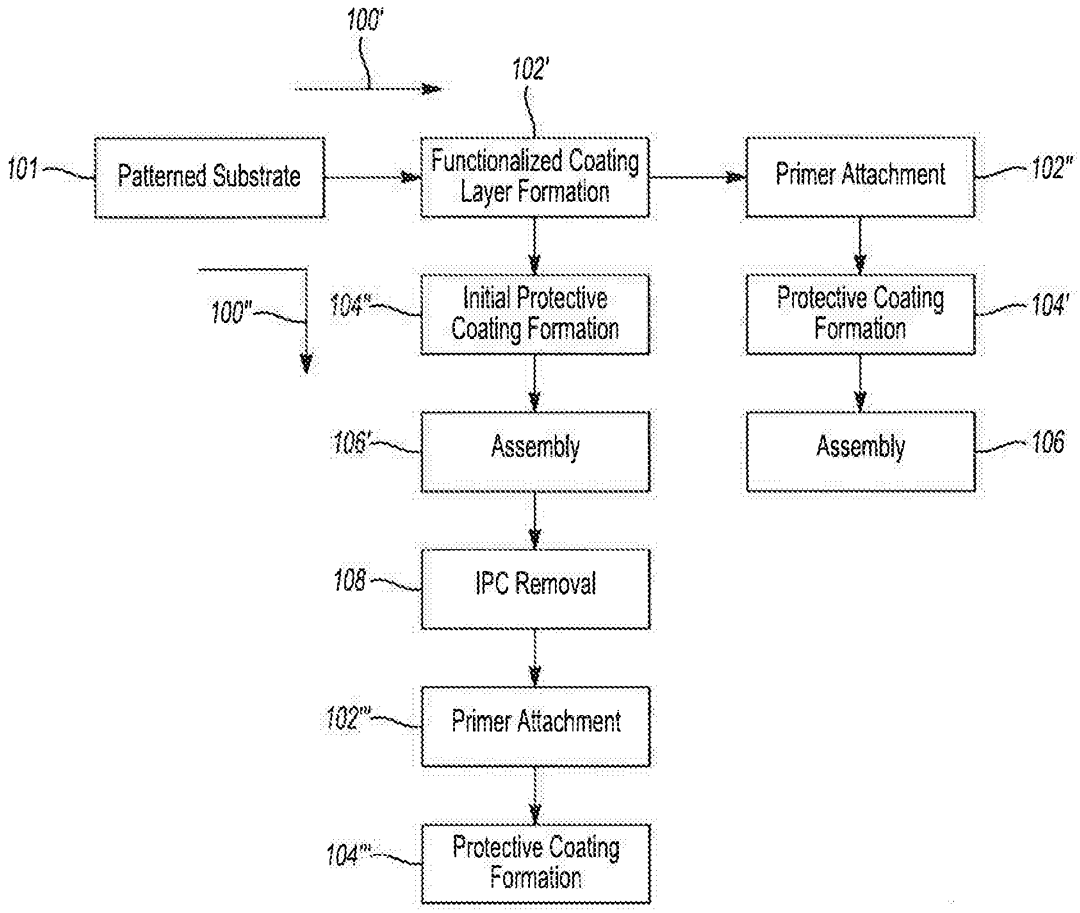

100'
102'
101
Patterned Substrate
Functionalized Coating Layer Formation
Primer Attachment
102"
100"
104"
Initial Protective Coating Formation
Protective Coating Formation
104'
106'
Assembly
Assembly
106
108
IPC Removal
102'"
Primer Attachment
104'"
Protective Coating Formation

Fig-2

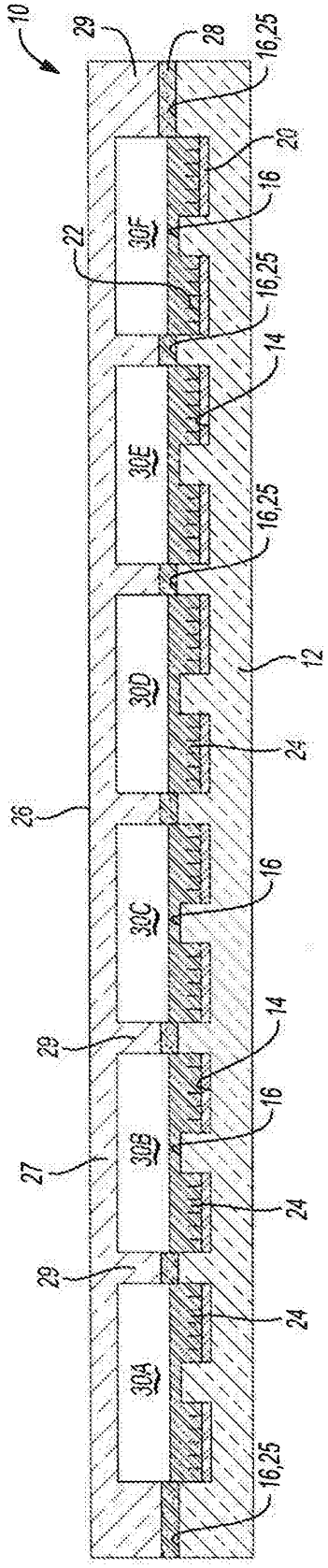

10
29
28
16,25
20
16
22
30F
16,25
14
30E
16,25
12
30D
24
26
16
30C
14
29
16
27
30B
24
29
24
30A
16,25

Fig-3

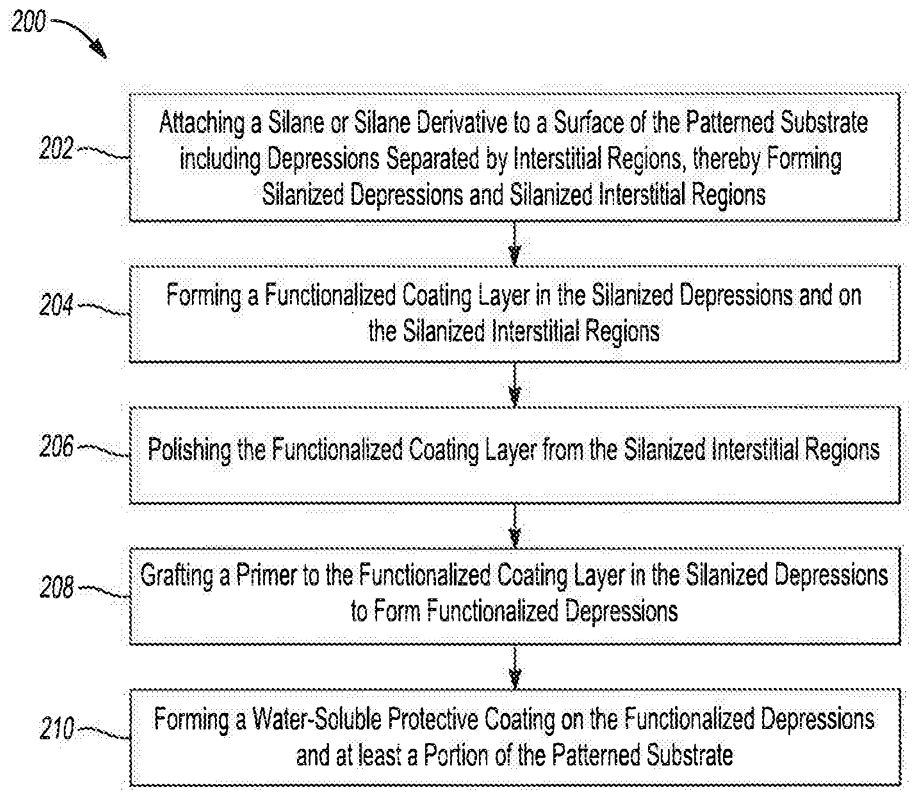

200
202
Attaching a Silane or Silane Derivative to a Surface of the Patterned Substrate including Depressions Separated by Interstitial Regions, thereby Forming Silanized Depressions and Silanized Interstitial Regions
204
Forming a Functionalized Coating Layer in the Silanized Depressions and on the Silanized Interstitial Regions
206
Polishing the Functionalized Coating Layer from the Silanized Interstitial Regions
208
Grafting a Primer to the Functionalized Coating Layer in the Silanized Depressions to Form Functionalized Depressions
210
Forming a Water-Soluble Protective Coating on the Functionalized Depressions and at least a Portion of the Patterned Substrate

Fig-4

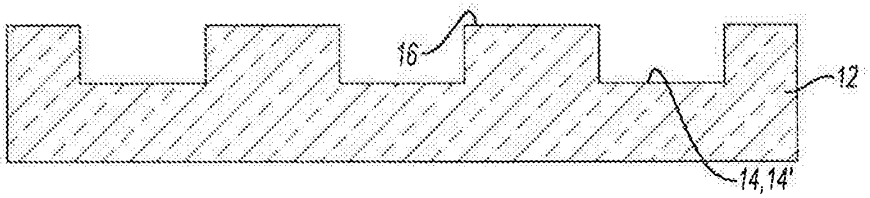


Fig-5A

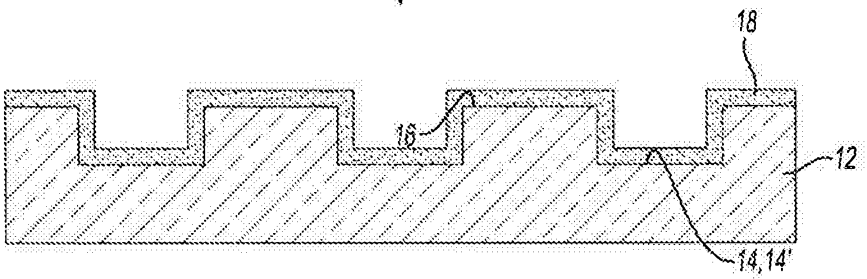


Fig-5B

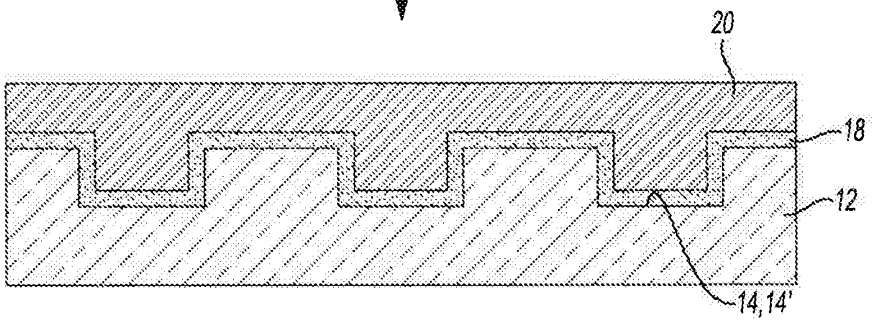


Fig-5C

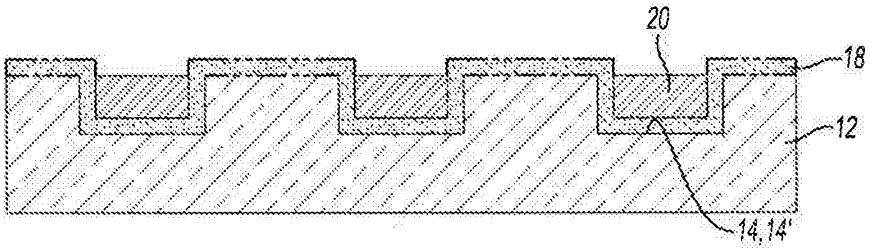


Fig-5D

To Fig 5E

To Fig 5I

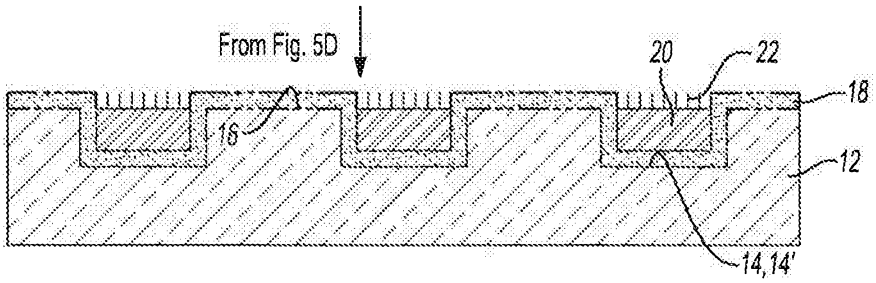


Fig-5E

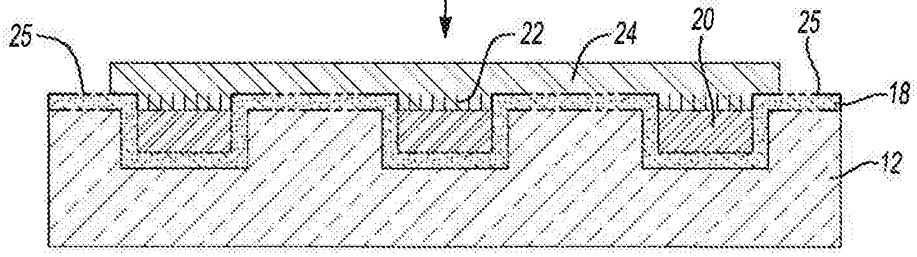


Fig-5F

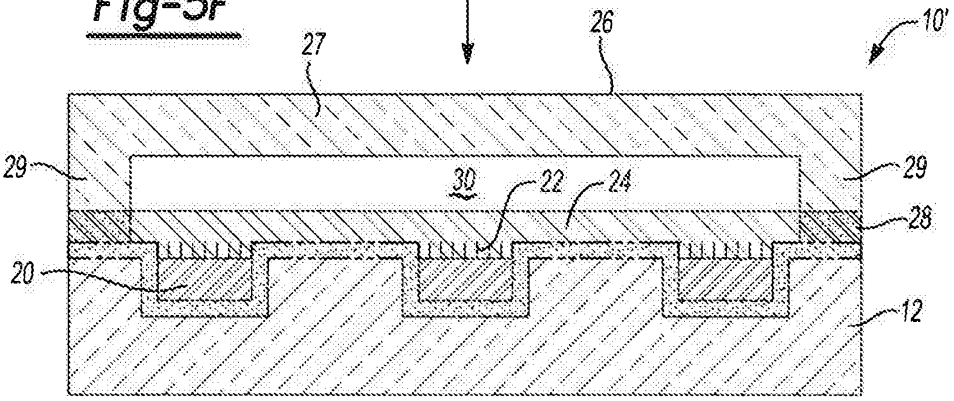


Fig-5G

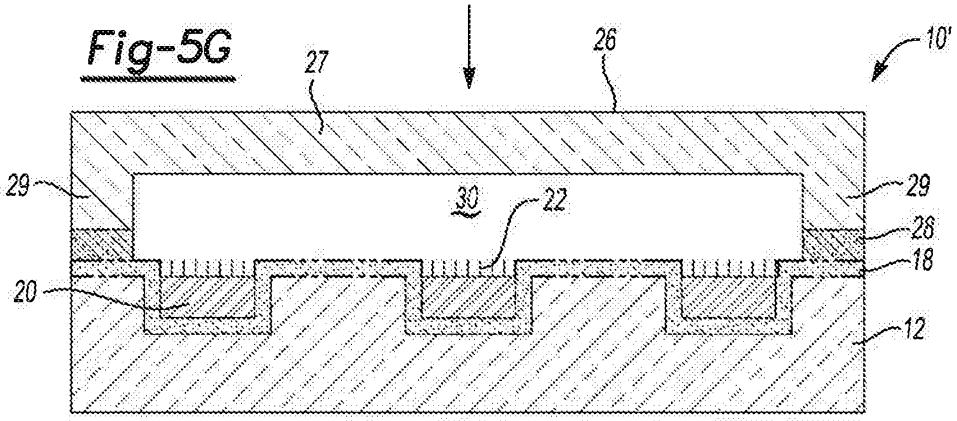


Fig-5H

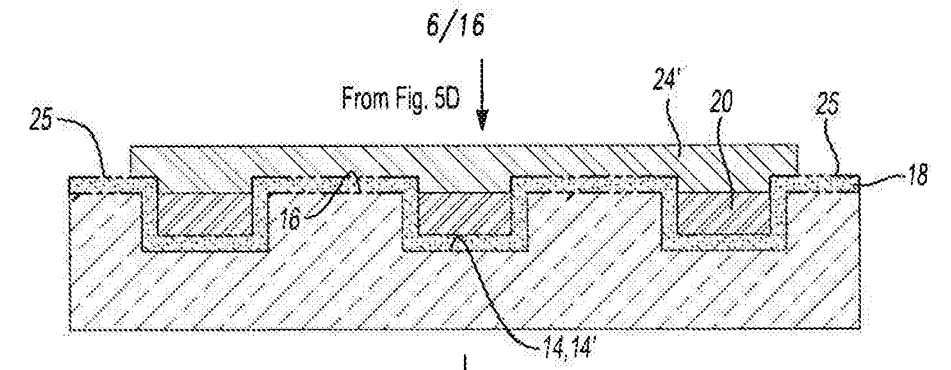


Fig-5I

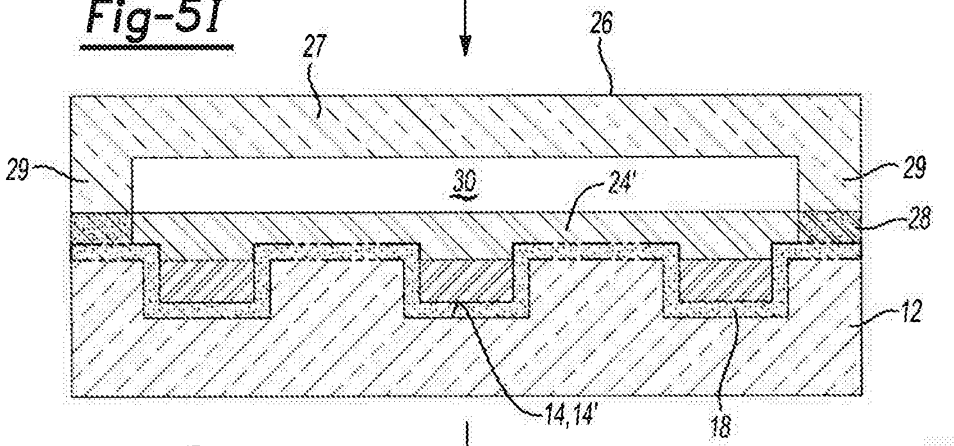


Fig-5J

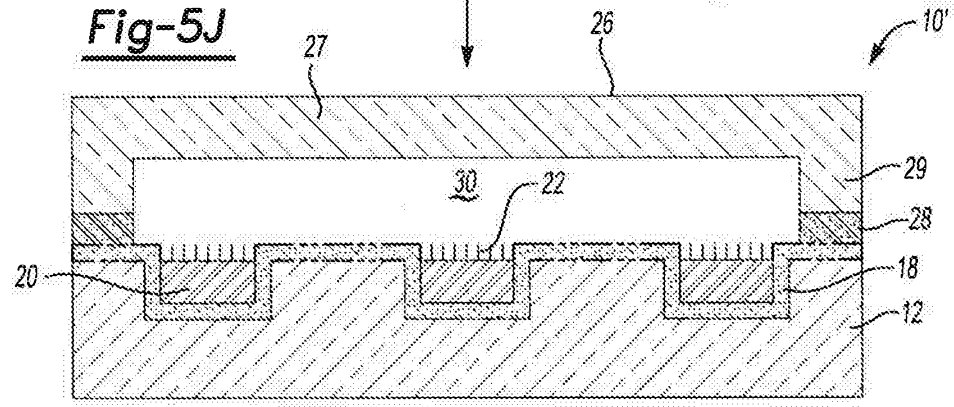


Fig-5K

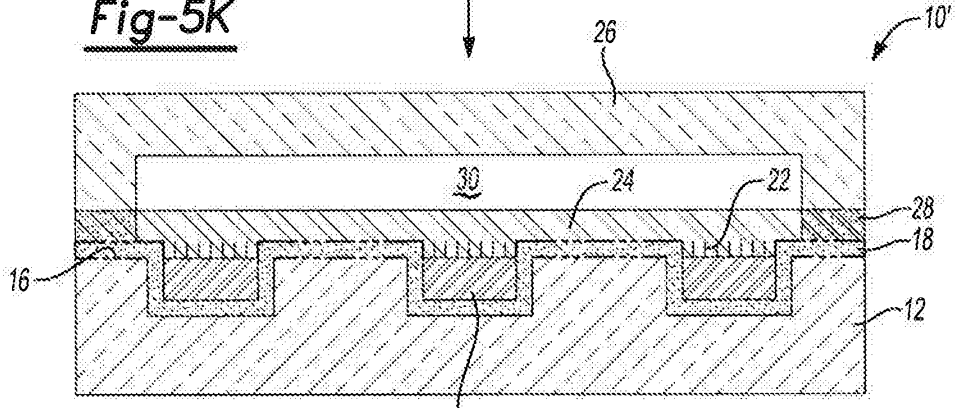


Fig-5L

Fig-6A

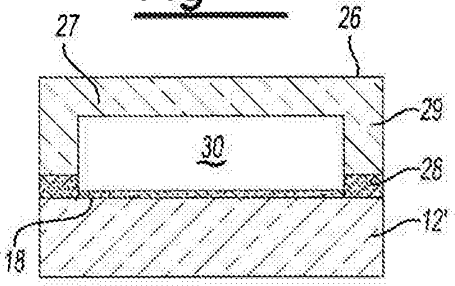


Fig-6B

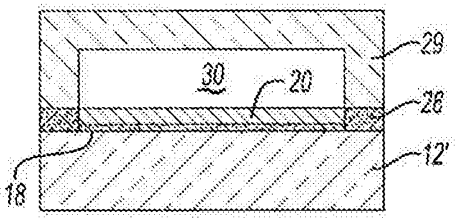


Fig-6C

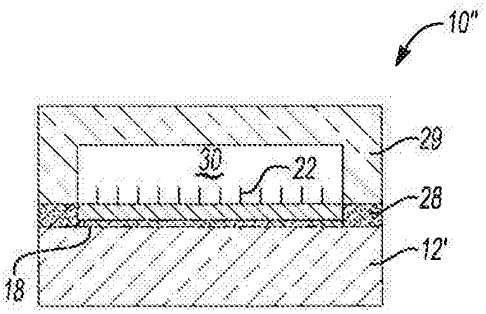


Fig-6D

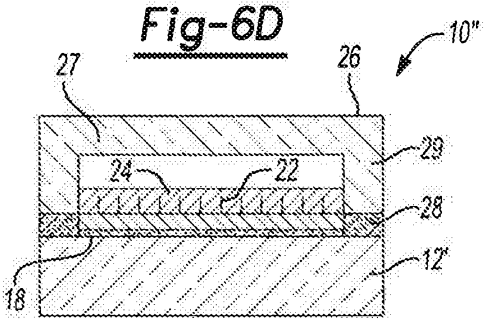


Fig-6E

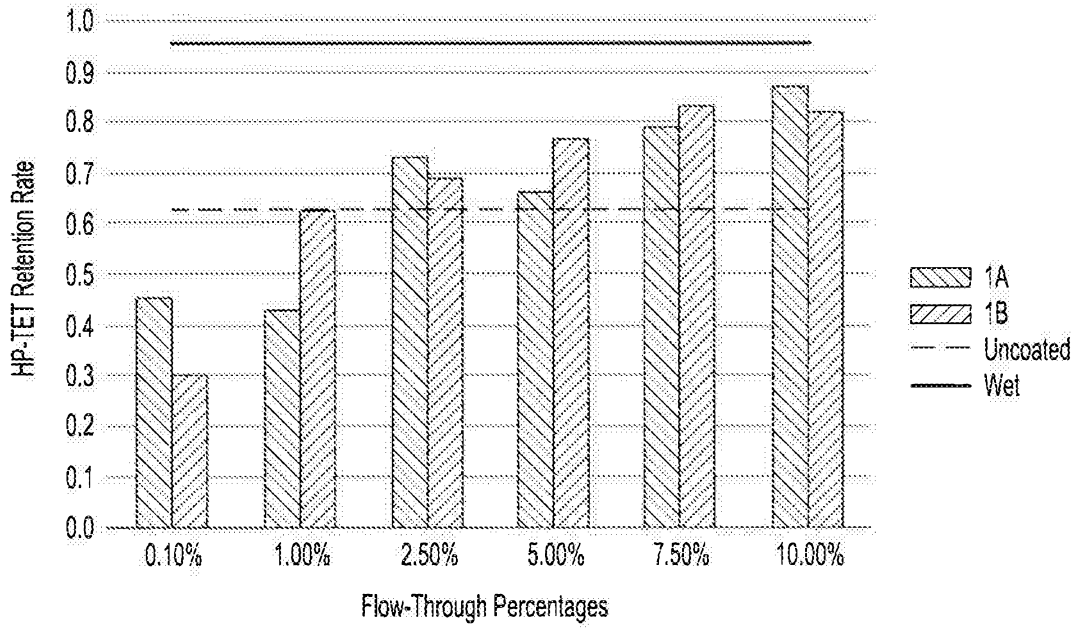


Fig-7

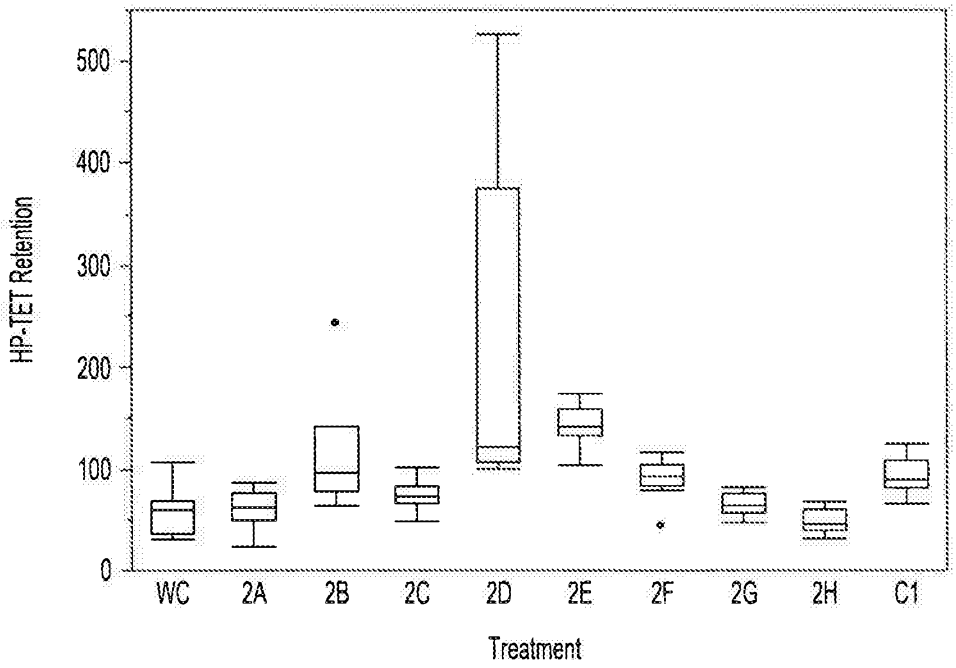

HP-TET Retention
500
400
300
200
100
0
WC
2A
2B
2C
2D
2E
2F
2G
2H
C1
Treatment

Fig-8

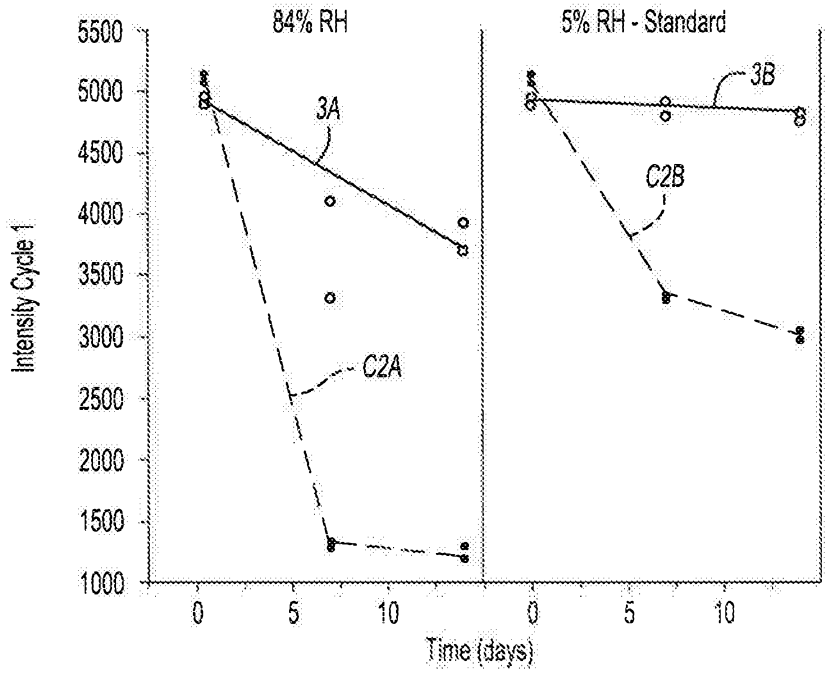

84% RH
5% RH - Standard
Intensity Cycle 1
5500
5000
4500
4000
3500
3000
2500
2000
1500
1000
3A
C2A
3B
C2B
0
5
10
0
5
10
Time (days)

Fig-9

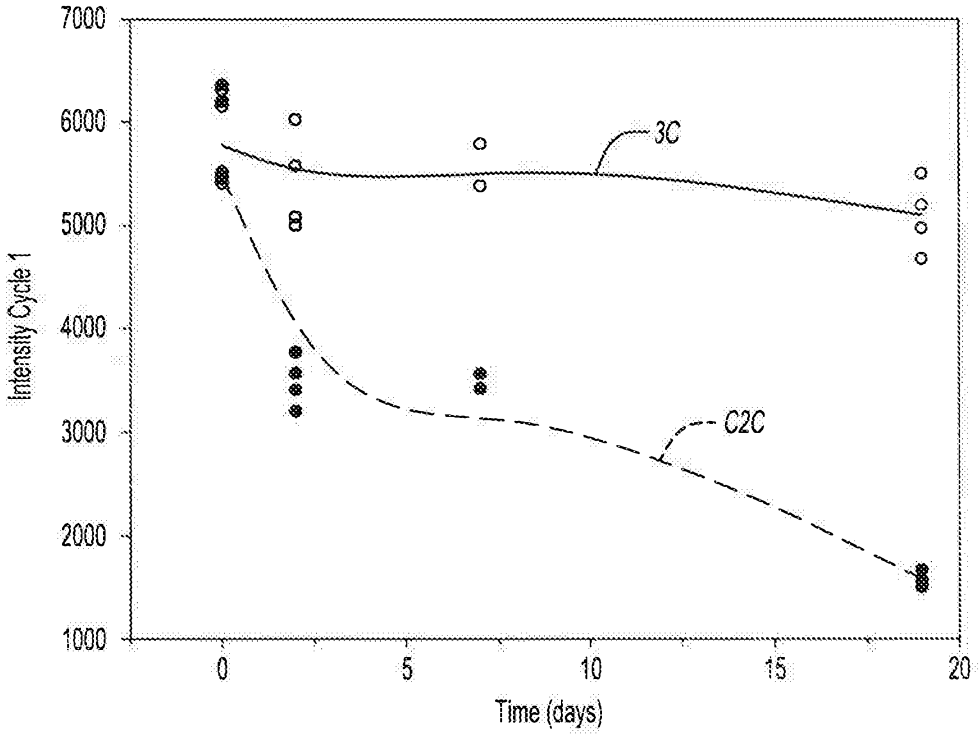


Fig-10

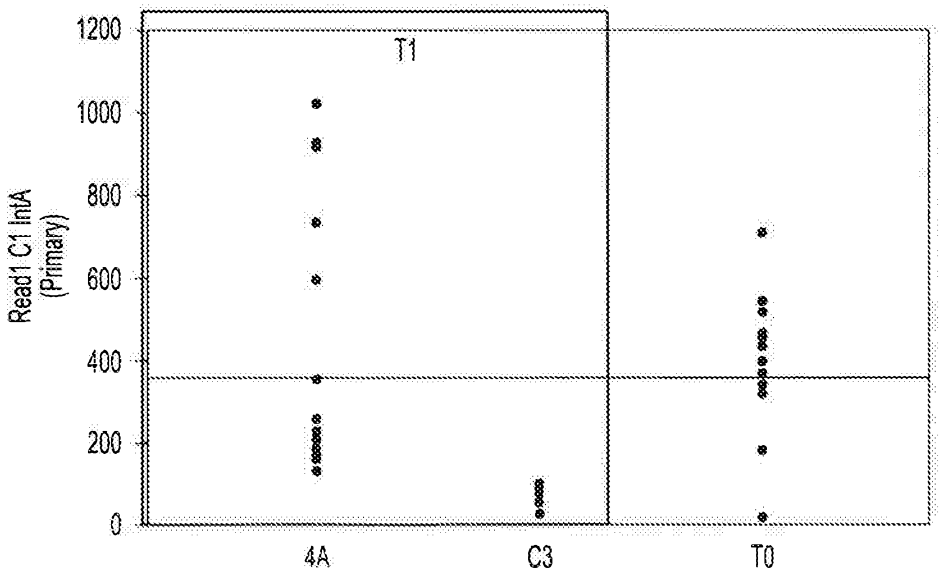


Fig-11A

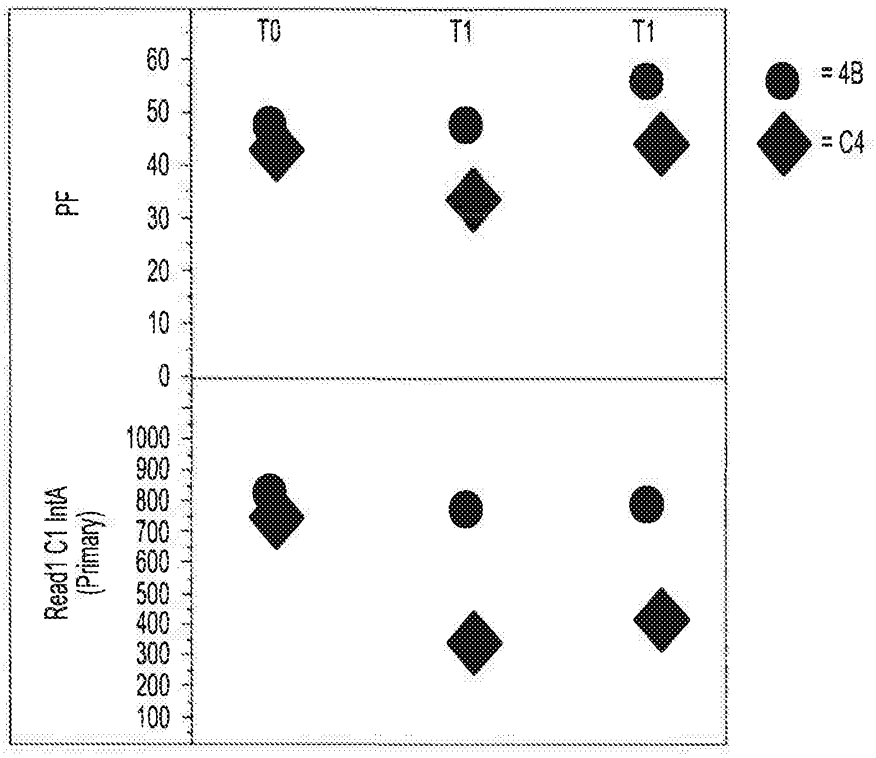


Fig-11B

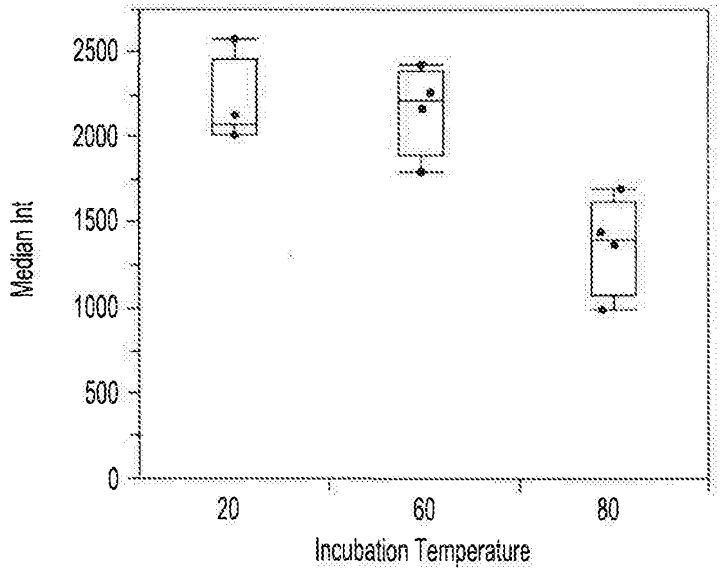


Fig-12A

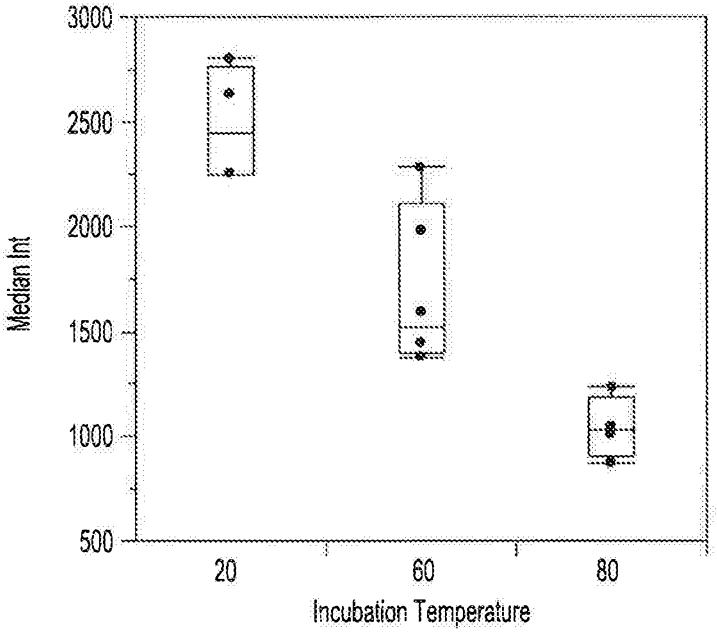


Fig-12B

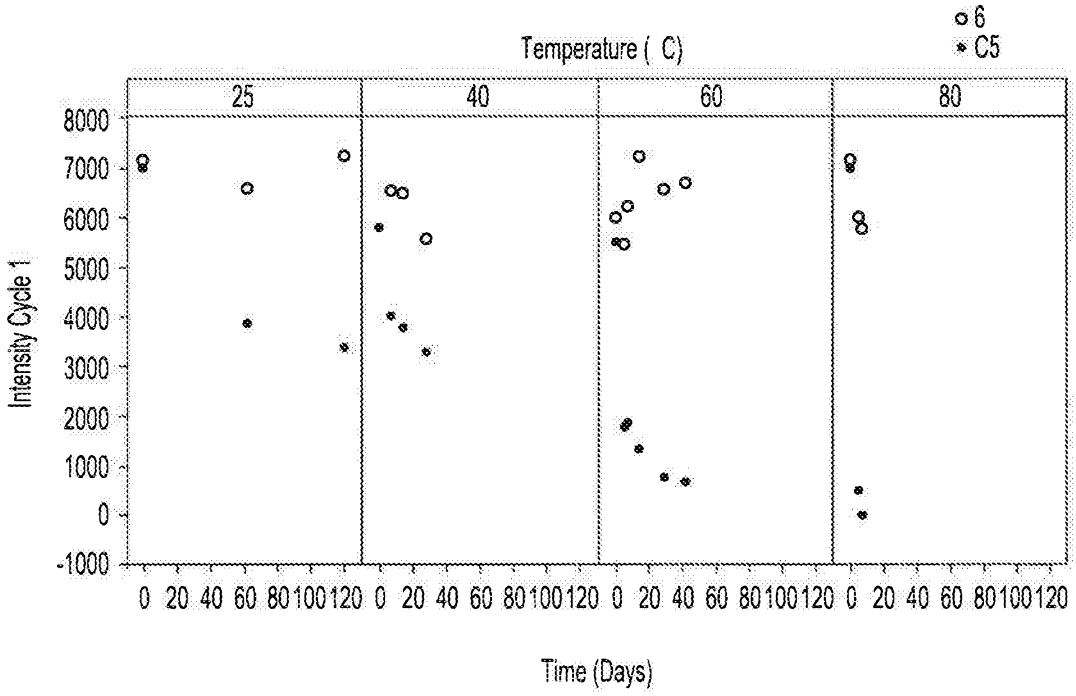


Fig-13

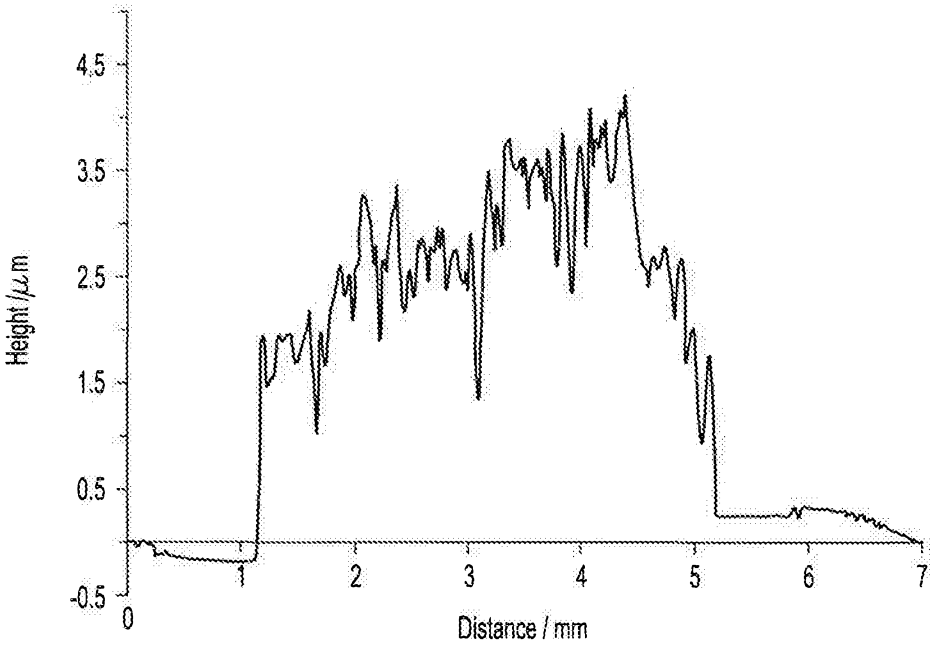

Height /μm
4.5
3.5
2.5
1.5
0.5
-0.5
0
1
2
3
4
5
6
7
Distance / mm

Fig-14

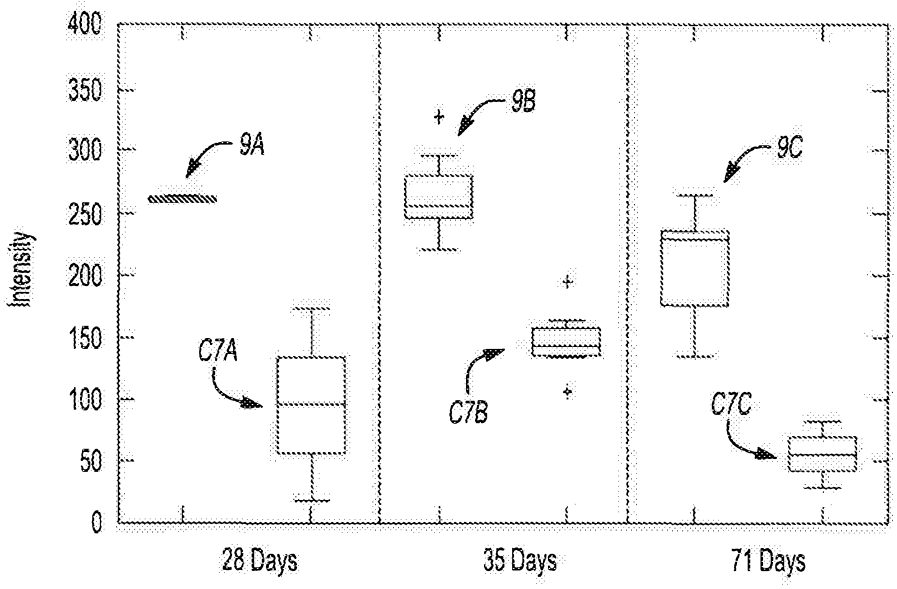

Intensity
400
350
300
250
200
150
100
50
0
9A
9B
9C
C7A
C7B
C7C
28 Days
35 Days
71 Days

Fig-15

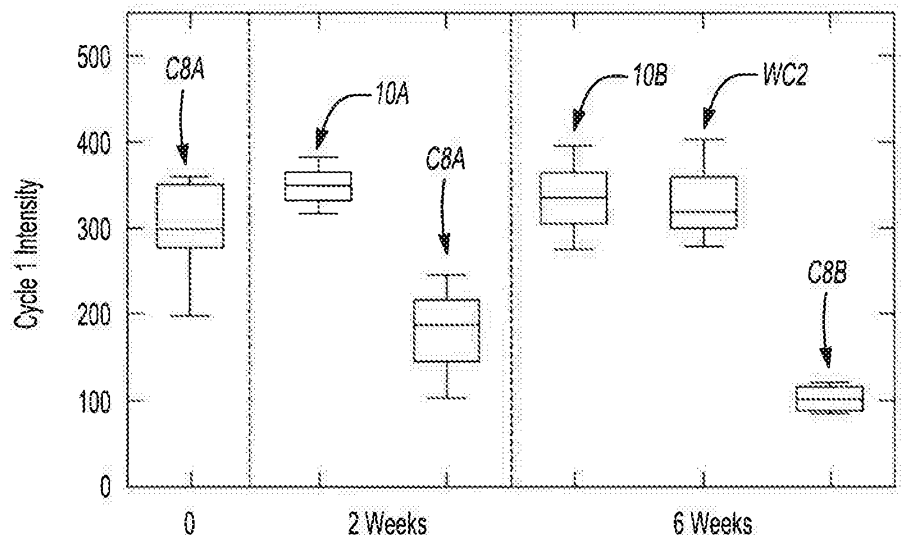

Cycle 1 Intensity
500
400
300
200
100
0
C8A
10A
C8A
10B
WC2
C8B
0
2 Weeks
6 Weeks

Fig-16

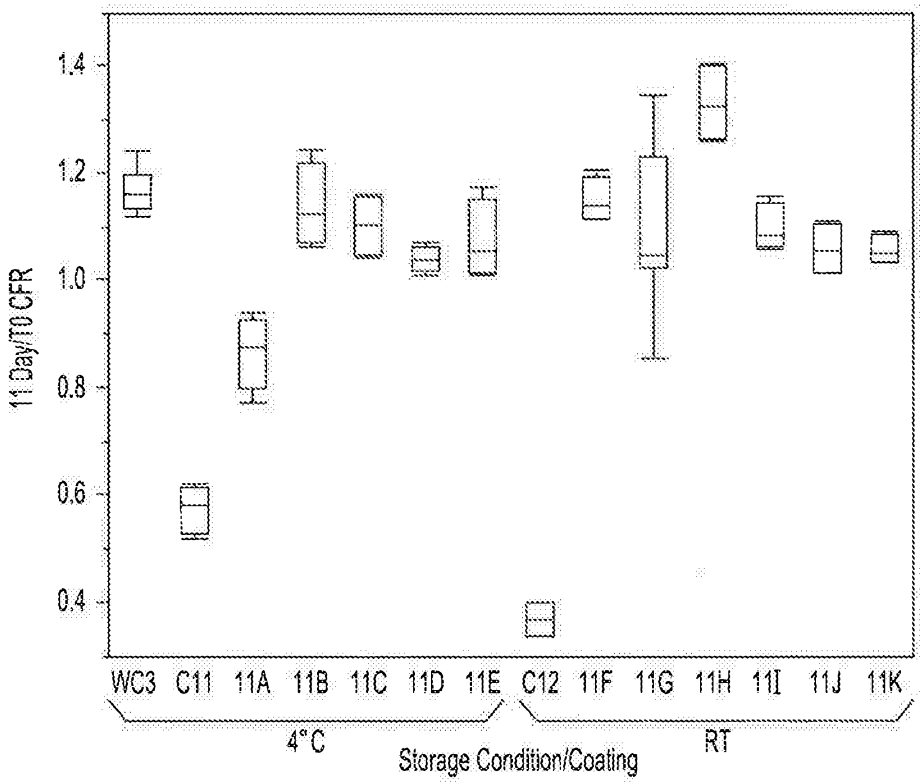

11 Day/T0 CFR
1.4
1.2
1.0
0.8
0.6
0.4
WC3
C11
11A
11B
11C
11D
11E
C12
11F
11G
11H
11I
11J
11K
4°C
RT
Storage Condition/Coating

Fig-17

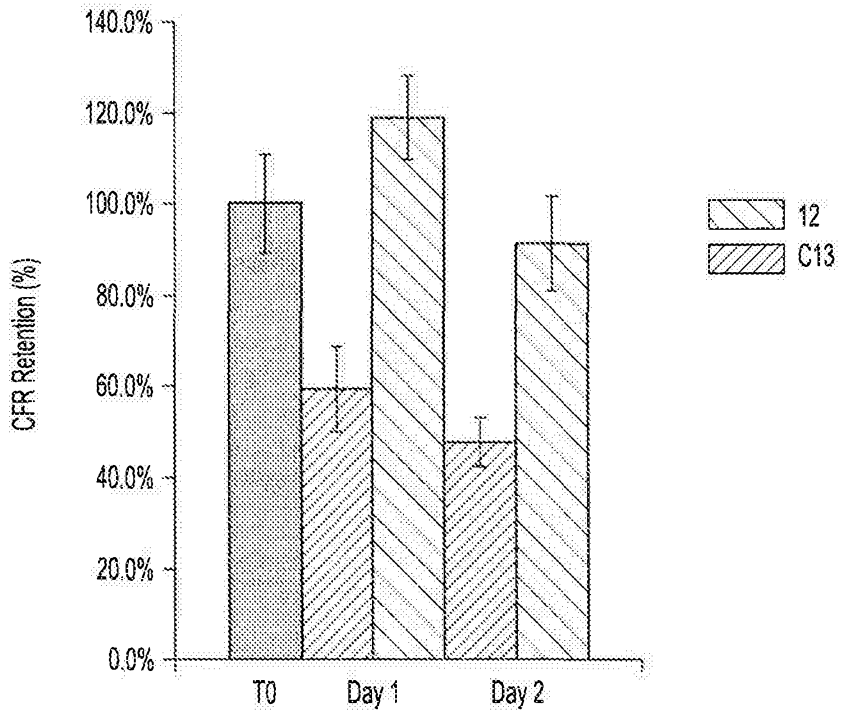

140.0%
120.0%
100.0%
80.0%
60.0%
40.0%
20.0%
0.0%
CFR Retention (%)
T0
Day 1
Day 2
12
C13

Fig-18

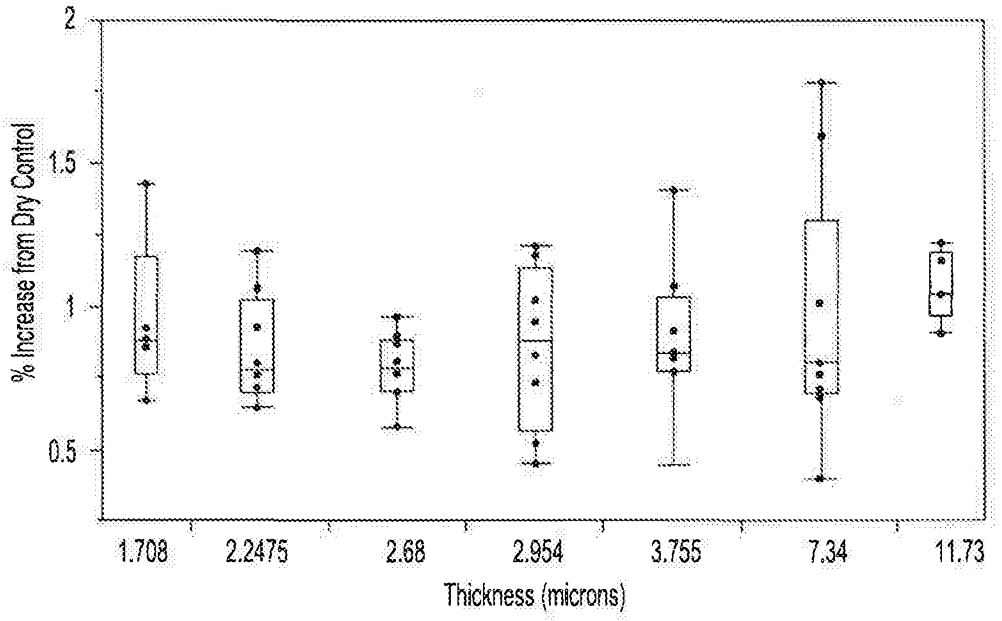

2
1.5
1
0.5
% Increase from Dry Control
1.708
2.2475
2.68
2.954
3.755
7.34
11.73
Thickness (microns)

Fig-19

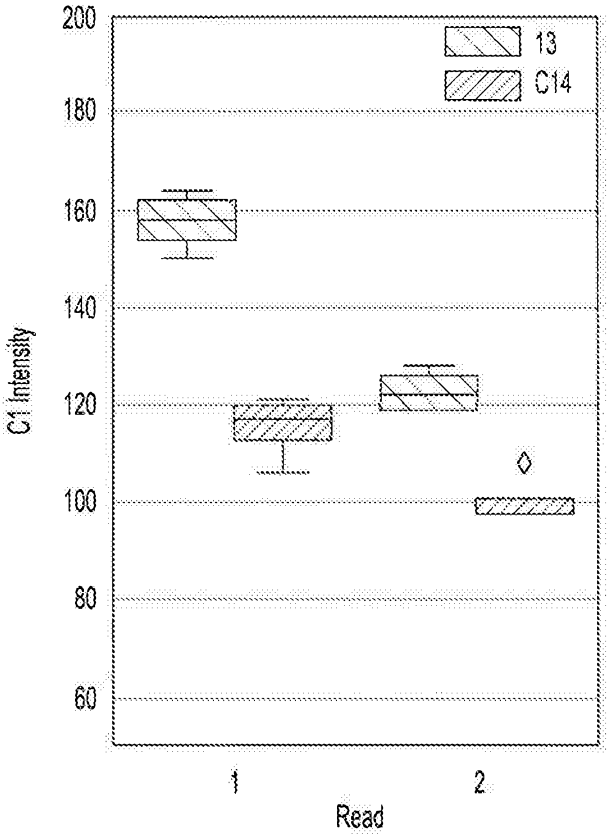

200
180
160
140
120
100
80
60
C1 Intensity
13
C14
1
2
Read

Fig-20

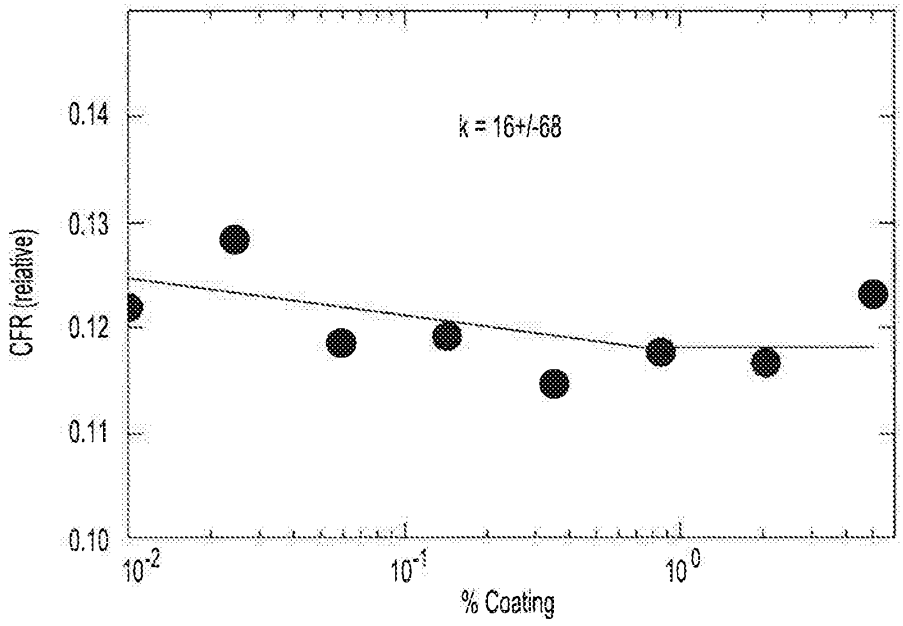

0.14
0.13
0.12
0.11
0.10
CFR (relative)
k = 16+/-68
10-2
10-1
100
% Coating

Fig-21

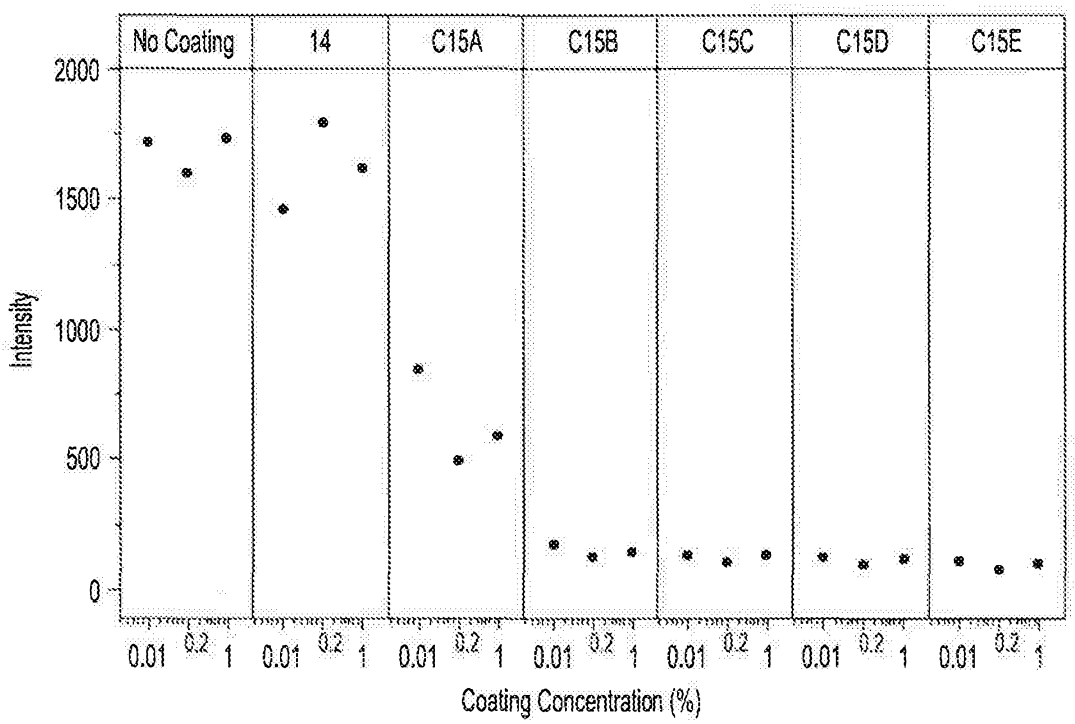

No Coating
14
C15A
C15B
C15C
C15D
C15E
2000
1500
1000
500
0
Intensity
0.01
0.2
1
Coating Concentration (%)

Fig-22

PROTECTIVE SURFACE COATINGS FOR FLOW CELLS

ABSTRACT

An example of a method includes adding surface chemistry to a portion of a flow cell substrate. This example of the method further includes applying a water-soluble protective coating on at least the surface chemistry. Examples of flow cells incorporating examples of the water-soluble protective coating are also disclosed herein.

SAMENWERKINGSVERDRAG (PCT)

RAPPORT BETREFFENDE NIEUWHEIDSONDERZOEK VAN INTERNATIONAAL TYPE

IDENTIFICATIE VAN DE NATIONALE AANVRAGE	KENMERK VAN DE AANVRAGER OF VAN DE GEMACHTIGDE
	P188105.NL.01/RDE
Nederlands aanvraag nr. **2019044**	Indieningsdatum **09-06-2017**
	Ingeroepen voorrangsdatum
Aanvrager (Naam) **Illumina, Inc.**	
Datum van het verzoek voor een onderzoek van internationaal type **12-08-2017**	Door de Instantie voor Internationaal Onderzoek aan het verzoek voor een onderzoek van internationaal type toegekend nr. **SN69483**

I. CLASSIFICATIE VAN HET ONDERWERP (bij toepassing van verschillende classificaties, alle classificatiesymbolen opgeven)

Volgens de internationale classificatie (IPC)

B01L3/00

II. ONDERZOCHTE GEBIEDEN VAN DE TECHNIEK

Onderzochte minimumdocumentatie

Classificatiesysteem	Classificatiesymbolen
IPC	**B01J;B01L**

Onderzochte andere documentatie dan de minimum documentatie, voor zover dergelijke documenten in de onderzochte gebieden zijn opgenomen

III. [ ] **GEEN ONDERZOEK MOGELIJK VOOR BEPAALDE CONCLUSIES** (opmerkingen op aanvullingsblad)

IV. [X] **GEBREK AAN EENHEID VAN UITVINDING** (opmerkingen op aanvullingsblad)

ONDERZOEKSRAPPORT BETREFFENDE HET RESULTAAT VAN HET ONDERZOEK NAAR DE STAND VAN DE TECHNIEK VAN HET INTERNATIONALE TYPE

Nummer van het verzoek om een onderzoek naar de stand van de techniek
NL 2019044

A. CLASSIFICATIE VAN HET ONDERWERP
INV. B01L3/00
ADD.

Volgens de Internationale Classificatie van octrooien (IPC) of zowel volgens de nationale classificatie als volgens de IPC.

B. ONDERZOCHTE GEBIEDEN VAN DE TECHNIEK

Onderzochte minimum documentatie (classificatie gevolgd door classificatiesymbolen)
B01J B01L

Onderzochte andere documentatie dan de minimum documentatie, voor dergelijke documenten, voor zover dergelijke documenten in de onderzochte gebieden zijn opgenomen

Tijdens het onderzoek geraadpleegde elektronische gegevensbestanden (naam van de gegevensbestanden en, waar uitvoerbaar, gebruikte trefwoorden)
EPO-Internal, WPI Data

C. VAN BELANG GEACHTE DOCUMENTEN

Categorie *	Geciteerde documenten, eventueel met aanduiding van speciaal van belang zijnde passages	Van belang voor conclusie nr.
	EENHEID VAN UITVINDING ONTBREEKT zie aanvullingsblad B -----	
X A	US 2009/054264 A1 (UGOLIN NICOLAS [FR] ET AL) 26 februari 2009 (2009-02-26) * alinea [0193] - alinea [0202]; figuren 11-17 * * alinea [0208] - alinea [0212]; figuur 8 * * conclusies 64,65 * -----	1-13,15, 20-23 14
X	US 6 589 778 B1 (HAWKINS GEORGE W [US]) 8 juli 2003 (2003-07-08) * kolom 8, regel 51 - regel 60; figuur 1 * -----	1,11,13, 15,20,24
X	WO 2017/024271 A1 (LIA DIAGNOSTICS INC [US]) 9 februari 2017 (2017-02-09) * alinea [0182]; figuur 4 * -----	1,11,20, 24
	-/--	

[X] Verdere documenten worden vermeld in het vervolg van vak C.

[X] Leden van dezelfde octrooifamilie zijn vermeld in een bijlage

* Speciale categorieën van aangehaalde documenten

"A" niet tot de categorie X of Y behorende literatuur die de stand van de techniek beschrijft

"D" in de octrooiaanvraag vermeld

"E" eerdere octrooi(aanvraag), gepubliceerd op of na de indieningsdatum, waarin dezelfde uitvinding wordt beschreven

"L" om andere redenen vermelde literatuur

"O" niet-schriftelijke stand van de techniek

"P" tussen de voorrangsdatum en de indieningsdatum gepubliceerde literatuur

"T" na de indieningsdatum of de voorrangsdatum gepubliceerde literatuur die niet bezwarend is voor de octrooiaanvraag, maar wordt vermeld ter verheldering van de theorie of het principe dat ten grondslag ligt aan de uitvinding

"X" de conclusie wordt als niet nieuw of niet inventief beschouwd ten opzichte van deze literatuur

"Y" de conclusie wordt als niet inventief beschouwd ten opzichte van de combinatie van deze literatuur met andere geciteerde literatuur van dezelfde categorie, waarbij de combinatie voor de vakman voor de hand liggend wordt geacht

"&" lid van dezelfde octrooifamilie of overeenkomstige octrooipublicatie

Datum waarop het onderzoek naar de stand van de techniek van internationaal type werd voltooid	Verzenddatum van het rapport van het onderzoek naar de stand van de techniek van internationaal type
21 december 2017	

Naam en adres van de instantie	De bevoegde ambtenaar
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Veefkind, Victor

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ONDERZOEKSRAPPORT BETREFFENDE HET RESULTAAT VAN HET ONDERZOEK NAAR DE STAND VAN DE TECHNIEK VAN HET INTERNATIONALE TYPE

Nummer van het verzoek om een onderzoek naar de stand van de techniek
NL 2019044

C.(Vervolg). VAN BELANG GEACHTE DOCUMENTEN

Categorie *	Geciteerde documenten, eventueel met aanduiding van speciaal van belang zijnde passages	Van belang voor conclusie nr.
X	US 2011/008776 A1 (WARTHOE PETER [DK] ET AL) 13 januari 2011 (2011-01-13) * alinea [0162] - alinea [0168] * -----	1,11,12, 15,20,24

Formulier PCT/ISA/201 (vervolg tweede blad) (januari 2004)

GEBREK AAN EENHEID VAN UITVINDING

Octrooiaanvrage Nr.:
SN 69483
NL 2019044

AANVULLINGSBLAD B

De instantie belast met het uitvoeren van het onderzoek naar de stand van de techniek heeft vastgesteld dat deze aanvrage meerdere uitvindingen bevat, te weten:

1. conclusies: 1-15, 20-24

   Method and flow cells

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2. conclusies: 16-19

   Method

Het vooronderzoek werd tot het eerste onderwerp beperkt.

ONDERZOEKSRAPPORT BETREFFENDE HET RESULTAAT VAN HET ONDERZOEK NAAR DE STAND VAN DE TECHNIEK VAN HET INTERNATIONALE TYPE

Informatie over leden van dezelfde octrooifamilie

Nummer van het verzoek om een onderzoek naar de stand van de techniek: NL 2019044

In het rapport genoemd octrooigeschrift	Datum van publicatie	Overeenkomend(e) geschrift(en)	Datum van publicatie
US 2009054264 A1	26-02-2009	CN 101389408 A	18-03-2009
		EP 1989000 A2	12-11-2008
		FR 2897858 A1	31-08-2007
		JP 2009528513 A	06-08-2009
		US 2009054264 A1	26-02-2009
		WO 2007096535 A2	30-08-2007
US 6589778 B1	08-07-2003	US 6589778 B1	08-07-2003
		US 2004018523 A1	29-01-2004
WO 2017024271 A1	09-02-2017	GEEN	
US 2011008776 A1	13-01-2011	EP 2214822 A1	11-08-2010
		EP 2214823 A1	11-08-2010
		JP 2011504591 A	10-02-2011
		JP 2011504592 A	10-02-2011
		US 2011008776 A1	13-01-2011
		US 2011045505 A1	24-02-2011
		WO 2009068583 A2	04-06-2009
		WO 2009068584 A1	04-06-2009
		WO 2009068585 A1	04-06-2009

WRITTEN OPINION

File No.	Filing date *(day/month/year)*	Priority date *(day/month/year)*	Application No.
SN69483	09.06.2017		NL2019044

International Patent Classification (IPC)
INV. B01L3/00

Applicant
Illumina, Inc.

This opinion contains indications relating to the following items:

- ☒ Box No. I Basis of the opinion
- ☐ Box No. II Priority
- ☒ Box No. III Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
- ☒ Box No. IV Lack of unity of invention
- ☒ Box No. V Reasoned statement with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement
- ☐ Box No. VI Certain documents cited
- ☐ Box No. VII Certain defects in the application
- ☒ Box No. VIII Certain observations on the application

Examiner
Veefkind, Victor

Application number

WRITTEN OPINION

NL2019044

Box No. I Basis of this opinion

1. This opinion has been established on the basis of the latest set of claims filed before the start of the search.

2. With regard to any **nucleotide and/or amino acid sequence** disclosed in the application and necessary to the claimed invention, this opinion has been established on the basis of:

   a. type of material:

   ☐ a sequence listing

   ☐ table(s) related to the sequence listing

   b. format of material:

   ☐ on paper

   ☐ in electronic form

   c. time of filing/furnishing:

   ☐ contained in the application as filed.

   ☐ filed together with the application in electronic form.

   ☐ furnished subsequently for the purposes of search.

3. ☐ In addition, in the case that more than one version or copy of a sequence listing and/or table relating thereto has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that in the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

4. Additional comments:

WRITTEN OPINION

Box No. III Non-establishment of opinion with regard to novelty, inventive step and industrial applicability

The questions whether the claimed invention appears to be novel, to involve an inventive step, or to be industrially applicable have not been examined in respect of

☐ the entire application

☒ claims Nos. 16-19

because:

☐ the said application, or the said claims Nos. relate to the following subject matter which does not require a search *(specify)*:

☐ the description, claims or drawings *(indicate particular elements below)* or said claims Nos. are so unclear that no meaningful opinion could be formed *(specify)*:

☐ the claims, or said claims Nos. are so inadequately supported by the description that no meaningful opinion could be formed *(specify)*:

☒ no search report has been established for the whole application or for said claims Nos. 16-19

☐ a meaningful opinion could not be formed as the sequence listing was either not available, or was not furnished in the international format (WIPO ST25).

☐ a meaningful opinion could not be formed without the tables related to the sequence listings; or such tables were not available in electronic form.

☐ See Supplemental Box for further details.

Box No. IV Lack of unity of invention

1. The requirement of unity of invention is not complied with for the following reasons:

   see separate sheet

2. This report has been established in respect of the following parts of the application:

   ☐ all parts.

   ☒ the parts relating to claims Nos. (see Search Report)

WRITTEN OPINION

Box No. V Reasoned statement with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty	Yes:	Claims	2-4, 14, 22, 23
	No:	Claims	1, 5-13, 15, 20, 21, 24
Inventive step	Yes:	Claims	14
	No:	Claims	1-13, 15, 20-24
Industrial applicability	Yes:	Claims	1-15, 20-24
	No:	Claims	

2. Citations and explanations

see separate sheet

Box No. VIII Certain observations on the application

see separate sheet

1 **Re Item IV**

Lack of unity of invention

1.1 It is considered that there are 2 inventions covered by the claims indicated as follows:

1. conclusies: 1-15, 20-24
Method and flow cells

2. conclusies: 16-19
Method

1.2 The reasons for which the inventions are not so linked as to form a single general inventive concept, are as follows:

1.3 Unity of invention is firstly to be considered in view of the independent claims. The common matter between all independent claims is the application of a "surface chemistry" to a flow cell substrate and the application of a water-soluble protection coating (i.e. essentially the subject-matter of claim 1). This is well-known, see e.g. D1-D4 and the passages cited therein, in combination with the analysis of these documents below.

1.4 Two groups of inventions have been identified:

1. independent claims 1, 20 and 24, and their dependent claims

2. independent claim 16, and its dependent claim.

1.5 There are no real distinguishing features between independent claims 1, 20, 24 and D1

The distinguishing features of claim 16 appear to be the specific "surface chemistry" (silane chemistry) and polishing of the coating layer, which have no equivalents in any of the other claims.

1.6 As claims 1, 20 and 24 are not new, there are no special technical features, and, hence, these can neither be common, nor corresponding with the features of claim 16.

1.7 As there is no common matter which is not known, and as there are no special technical features, which are common or corresponding, the two groups of inventions identified above, are non-unitary.

2 **Re Item V**

Reasoned statement with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

2.1 Reference is made to the following documents:

D1	US 2009/054264 A1 (UGOLIN NICOLAS [FR] ET AL) 26 februari 2009 (2009-02-26)
D2	US 6 589 778 B1 (HAWKINS GEORGE W [US]) 8 juli 2003 (2003-07-08)
D3	WO 2017/024271 A1 (LIA DIAGNOSTICS INC [US]) 9 februari 2017 (2017-02-09)
D4	US 2011/008776 A1 (WARTHOE PETER [DK] ET AL) 13 januari 2011 (2011-01-13)

2.2 D1 (claims 64 and 65) discloses fixing chemical or biological molecules ("surface chemistry") on a support plate, and covering them with a suitable protective material, which can be polyacrylamide (preferred protective layer of the application, see e.g. claim 11).
D1 also discloses ([0202] – [0212]) spotting of molecules on a flow cell substrate which are protected by a layer of EDTA during fabrication of the flow cell. The EDTA layer can later be dissolved. EDTA is a chelate former and one of the exemplary water-soluble coatings of the application, see e.g. claim 11.
Hence D1 discloses the subject-matter of independent claim 1, as well as that of claims 5, 11, 13. it also discloses the subject-matter of independent claim 20, 21.

Also (see [0196] – [0202] , the EDTA is patterned and used to define bonding regions, which are filled with another polymer (e.g. PMMA), and later an adhesive on top of that to eventually bond the top plate to the substrate. The EDTA form channels from which the EDTA is later dissolved out.
D1, therefore, also seems to disclose the subject-matter of claims 5-10.

2.3 D2 discloses (see figure 1 and column 8, lines 51-60) a microfluidic substrate (12) with an array (17) on it ("surface chemistry"), covered with a plate (16). A protective layer of a water soluble compound is formed on it, preferably filling the entire volume 14. The water soluble compound preferably is PEG (600). PEG also is a preferred water soluble coating of the present application (see e.g. claims 2,4,6 or 11).
D2 therefore discloses the subject-matter of independent claims 1, 20 and 24, as well as that of claims 11, 13 and 15.

2.4 D3 discloses (see e.g. [0184]) applying a protective coating on reagents which were supplied to a flow cell surface. Preferred coatings include water-soluble PVA films.
Hence, claim 1 lacks novelty over D3 also.

2.5 D4 discloses a flow cell substrate with a capillary channel which was corona treated to increase hydrophilicity (i.e. "surface chemistry" was added) and then coated with magnetic particles in 5% sucrose (water-soluble coating) and then dried for storage.
Hence, claims 1, 11, 12 and 15 lack novelty over D4.

2.6 The subject matter of the other searched dependent claims, if at all new, is considered to lack an inventive step over D1, which discloses applying several coatings and patterning and to which the additional features of the not new dependent claims are merely foreseeable variations.

2.7 Claim 14 is new, and seems to be non-obvious over the cited prioor art.

3 **Re Item VIII**

Certain observations on the application

3.1 The claims lack clarity, the essential feature of the invention appears to be that the protective coating is water-soluble. Some preferred polymers mentioned, such as e.g. silicones, are not normally considered "water-soluble" once they have formed a coating. This means that the skilled person will have to determine which silicone is "water-soluble" and which one isn't. Absent a definition of "water-soluble", i.e. to which extent should it be soluble, the boundaries of many claims are unclear.

3.2 The difference between claim 20 and 24 is unclear, especially since the "pattern" can be formed by walls on a substrate, which can either be considered to be part of the substrate, or as separate walls.

3.3 Claim 14 is not clear. It is not clear what is meant with "in which the surface chemistry comprises a ground layer". It had been understood that the surface chemistry by definition would form some sort of layer, which can be called a "ground layer". The distinction between "surface chemistry" and "ground layer" is not clear, and because of it, the boundaries of the claims are ill-defined.