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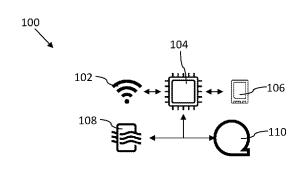


FIG. 1A

(57) Abstract: In one aspect, a system for generating and monitoring an antimicrobial is provided, the system including: a microprocessor and/or a microcontroller; an external communications device; a computational system; an antimicrobial sensor and/or an environmental sensor, and an antimicrobial generator, wherein the external communications device, the computational system, the antimicrobial generator, and the antimicrobial sensor and/or the environmental sensor are operatively connected to the microprocessor and/or the microcontroller. The system may further include a separate sensor sub-system comprising: a sensor sub-system microprocessor and/or a sensor sub-system microcontroller; a sensor sub-system external communications device; a sensor sub-system antimicrobial sensor and/or a sensor subsystem environmental sensor; and a sensor sub-system computational system. The system may further include a separate generation sub-system comprising: a generation sub-system microprocessor and/or a generation sub-system microcontroller; a generation sub-system external communications device; and a generation sub- system antimicrobial generator.

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SYSTEMS, METHODS, AND APPARATUSES FOR DISINFECTION AND DECONTAMINATION

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority from U.S. Provisional Patent Application No. 63/036,412, filed on June 8, 2020, U.S. Provisional Patent Application No. 63/049,524, filed on July 8, 2020, U.S. Provisional Patent Application No. 63/049,541, filed on July 8, 2020, U.S. Provisional Patent Application No. 63/049,919, filed on July 9, 2020, U.S. Provisional Patent Application No. 63/081,459, filed on September 22, 2020, U.S. Provisional Patent Application No. 63/126,734, filed on December 17, 2020, and U.S. Provisional Patent Application No. 63/157,368, filed on March 5, 2021, each of which is incorporated by reference herein in its entirety.

BACKGROUND

[0002] Infectious diseases such as human immunodeficiency virus and acquired immune deficiency syndrome (HIV/AIDS), tuberculosis (TB), severe acute respiratory syndrome (SARS-CoV-1), Ebola virus disease (EVD), and coronavirus disease 2019 (COVID-19) are contagious diseases transmissible through direct contact from person to person, through indirect contact by breathing airborne droplets spread from an infected person, and through contact with surfaces of contaminated objects.

[0003] With the current COVID-19 pandemic outbreak, facemask or respirator wearing and practicing social distancing may mitigate airborne droplets spread by potential neighboring human carriers. Nevertheless, both of these practices are defensive actions that do not destroy or disinfect the germs or viruses in the airborne droplets. Currently, methods that are used to generate antimicrobial gases or vapor are large and impractical for general household or office use or for personal use in a limited localized space, and methods of generating ClO₂ from liquid and solid precursor chemicals are slow and/or generate low quality ClO₂ solutions.

[0004] Antimicrobial gas, such as chlorine dioxide (ClO₂), has demonstrated capability as an antimicrobial or inactivator for pathogens on hard surfaces. In gas form, ClO₂ has demonstrated capability to disinfect hard surfaces and porous materials within three-dimensional spaces. ClO₂ gas has also been shown to kill or otherwise inactivate airborne pathogens, and even protect against airborne contagion.

[0005] ClO₂ gas is also currently used as a deodorizer in vehicles, rooms, and other enclosed spaces. Typical products used for enclosed space odor removal include placing a cup or container housing one or more dry solid chemical constituents (typically consisting of a chlorite

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salt and an activator), adding water to activate the ClO₂ generation process, enclosing the ClO₂ generation materials in the space for an extended period of time before opening up the space, removing the spent ClO₂ solution, and allowing the space to air out to reduce ClO₂ concentration to safe levels.

The present disclosure relates to a safe and effective system and method for quickly and safely generating antimicrobial gas (e.g., ClO₂ gas). Antimicrobial gas may be generated from small amounts of concentrated liquid and solid precursor chemicals and actively dispersing the antimicrobial gas into an enclosed three-dimensional space. Additionally, the present disclosure relates to a safe and effective system and method for monitoring antimicrobial gas concentration in the enclosed three-dimensional space and generating additional antimicrobial gas as necessary to maintain the desired concentration in the space. When used at higher concentrations, the resultant antimicrobial gas will sanitize or disinfect the air and contact surfaces within the enclosed space. At low concentrations (e.g., < 0.1 ppm), the antimicrobial gas can be used to decrease or otherwise inactivate airborne pathogens and actively protect persons against airborne contagions.

[0007] The present disclosure also relates to a safe and effective system and method of generating and monitoring the concentration of antimicrobial gas on demand.

SUMMARY

[8000]In one aspect, a system for generating and monitoring an antimicrobial gas is provided, the system including: a microprocessor and/or a microcontroller; an external communications device; a computational system; an antimicrobial sensor and/or an environmental sensor; and an antimicrobial generator, wherein the external communications device, the computational system, the antimicrobial generator, and the antimicrobial sensor and/or the environmental sensor are operatively connected to the microprocessor and/or the microcontroller. The system may further include a separate sensor sub-system comprising: a sensor sub-system microprocessor and/or a sensor sub-system microcontroller; a sensor sub-system external communications device; a sensor sub-system antimicrobial sensor and/or a sensor sub-system environmental sensor; and a sensor sub-system computational system. The system may further include a separate generation sub-system comprising: a generation sub-system microprocessor and/or a generation sub-system microcontroller; a generation sub-system external communications device; and a generation sub-system antimicrobial generator. In another aspect, a network of these systems for generating and monitoring an antimicrobial gas is provided.

[0009] In another aspect, a system for generating and monitoring ClO₂ gas is provided, the system comprising: a device housing including an inlet; a microcontroller; one or more reagent

containers containing a reagent; a microfluidic liquid dispensing and metering system; a microfluidic device for generating a ClO₂ gas from the reagent(s); a device for separation of ClO₂ gas and post-generator waste in communication with the air pump air duct and an air duct to one or more outlets; on-device or in-device waste storage prior to disposal; and one or more sensing system for either ClO₂ gas or the environment in which the device is installed.

In another aspect, a ClO₂ gas generator is provided, comprising: a base including a pressure generator; one or more reagent containers holding liquid reagent(s), the containers being pressurized by the pressure generator; a chamber passage in communication with the pressure chamber and the reagent container; one or more control valves in communication with the pressure generator and reagent container; one or more control valves in communication with the chamber passage and a microfluidic chip; a sensor system for determining the quantity, mass, or volume of the reagents transiting the chamber passage; a microfluidic chip having a generation chamber in communication with a second chamber passage; a second chamber passage in communication with a CLO₂ gas-liquid separation chamber; and, a waste container for storage and/or inactivation of post-CLO₂ generator waste products.

[0011] In another aspect, a network of systems for generating and monitoring ClO₂ gas, is provided, the network comprising: a plurality of systems for generating and monitoring ClO₂ gas, including: a device housing including an inlet; a microcontroller; one or more reagent containers containing a reagent; a microfluidic device for generating a ClO₂ gas from the reagent; and a sensing system; wherein the microcontroller includes a communication device capable of communication between the plurality of systems, and wherein the communication device establishes distributed control of each system's microcontroller, wherein the microcontroller is controlled by machine learning algorithms to alter system performance.

[0012] In another aspect, a network of systems for generating and monitoring an antimicrobial gas is provided, the system including: a microprocessor and/or a microcontroller; an external communications device; a computational system; an antimicrobial sensor and/or an environmental sensor; and an antimicrobial generator, wherein the external communications device, the computational system, the antimicrobial generator, and the antimicrobial sensor and/or the environmental sensor are operatively connected to the microprocessor and/or the microcontroller. The system may further include a separate sensor sub-system comprising: a sensor sub-system microprocessor and/or a sensor sub-system antimicrobial sensor and/or a sensor sub-system environmental sensor; and a sensor sub-system computational system. The system may further include a separate generation sub-system comprising: a generation sub-system

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microprocessor and/or a generation sub-system microcontroller; a generation sub-system external communications device; and a generation sub-system antimicrobial generator.

In another aspect, the microcontroller of the system will have the computational and local data storage ability to enable closed-loop control of the ClO₂ generation system, including but not limited to: local storage and microcontroller operations on data from sensor systems for ClO₂ levels to space environment variables like barometric pressure, humidity, temperature, occupancy, or sounds that may be used to alter generator system performance automatically or via user intervention; measurement, local storage, and microcontroller operations on data from microfluidic subsystems such as mass/volume sensors of reagents, pressure generator performance, microfluidic chip-borne sensors, valve status to any other electronic subsystem to provide control as well as storage of system performance data for maintenance, alert, troubleshooting, inactive modes of operation, active modes of operation, and local setup.

[0014] In another aspect, the system has a communication device connected to the microcontroller and/or electronic components such that data from any electronic component within, on, or connected to the housing can be gathered, locally stored, operated on by the microcontroller, and transmitted to external data gathering systems on mobile to fixed devices.

[0015] In another aspect, machine learning and/or artificial intelligence algorithms can be incorporated into the system microcontroller to alter system performance automatically or by user interactions. An example of local control includes alteration of system performance for detection of a virus or bacteria in the ambient air, altitude, temperature, air changes in the local space measured by changes in concentration in the air of spaces containing ClO₂, changes in occupancy by living beings, alterations for user preference, prediction of cycles of occupancy/vacancy, alerts as to normal or abnormal performance of the system, and the like.

[0016] In another aspect, a plurality of systems within a plurality of spaces which are arranged into a group connected via communication devices described above to each other for distributed control via coordination of each system's microcontroller, centralized unit control, and/or a combination of both local and distributed control.

[0017] In another aspect, machine learning and/or artificial intelligence algorithms can be incorporated into the distributed network of systems by the aspects described above; example of distributed control include adjusting individual systems to achieve uniform and/or deliberately non-uniform distribution of ClO₂ in each individual generator's location across an entire building floor, multiple floors, or the entire building due to changes in ClO₂ concentration from HVAC, consumption or self-dissipation of ClO₂ gas, control of day/night generation cycles, sensing patterns across time, three-dimensional volumes, seasonal variations, to previously unknown

factors which can be sensed either directly by the sensor systems in/on the system, inferred or traced to the signal measured, or directly traceable to the variations observed in ClO₂ concentrations across a collection of systems installed across distinctly separate to varying interconnection of real world spaces in which control of infectious species is desired.

[0018] In another aspect, the system for distribution and monitoring of ClO2 gas in a three-dimensional space will be designed for a plurality of operating modes; the first operating mode is designed for occupied spaces, the second mode is designed for un-occupied spaces and may include one or more sub-modes to achieve desired outcomes; future user or engineered modes may be added. These modes may be changed by authorized users on the unit, via connected mobile devices, and/or by a centralized distributed control system connected to a plurality of units.

BRIEF DESCRIPTION OF THE DRAWINGS

[0019] FIG. 1A illustrates a schematic of an example system 100 for generating and monitoring an antimicrobial gas.

[0020] FIG. 1B illustrates a schematic of system 100 oriented within a volume under treatment 124.

[0021] FIG. 1C illustrates a schematic of system 100 oriented within a volume under treatment 124.

[0022] FIG. 2A illustrates a schematic of an example complimentary sensing sub-system 120 and generation sub-system 122 for generating and monitoring an antimicrobial gas.

[0023] FIG. 2B illustrates a schematic of sensing sub-system 120 and generation subsystem 122 within a volume under treatment 124.

[0024] FIG. 2C illustrates a schematic of sensing sub-system 120 and generation subsystem 122 used in conjunction with an HVAC system.

[0025] FIG. 2D illustrates a schematic of sensing sub-system 120 within a volume under treatment 124 engaging with generation sub-system 122 outside of the volume under treatment 124.

[0026] FIG. 2E illustrates a schematic of generation sub-system 122 within a volume under treatment 124 engaging with sensing sub-system 120 outside of the volume under treatment 124.

[0027] FIG. 3 illustrates a schematic of an example system 300 for generating and monitoring an antimicrobial gas.

[0028] FIG. 4 illustrates a schematic of an example system 400 for generating and monitoring an antimicrobial gas.

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[0029] FIG. 5 illustrates an example blueprint of a network 500 of antimicrobial gas systems 300 and sensors distributed in rooms and spaces within a floor of a building.

[0030] FIG. 6 illustrates a schematic of an example system 600 for generating and monitoring an antimicrobial gas.

[0031] FIG. 7 illustrates a schematic of an example system 700 for generating and monitoring an antimicrobial gas.

[0032] FIG. 8 illustrates a cutaway perspective view of a system 800 generating antimicrobial vapor within a sealed environment 810 for disinfecting items therein.

[0033] FIG. 9 illustrates a system 900 for generation of an antimicrobial gas and/or solution.

[0034] FIG. 12 illustrates a system 1200 for generation of an antimicrobial gas and/or solution.

[0035] FIG. 13 illustrates a system 1300 for generation of an antimicrobial gas and/or solution.

[0036] FIG. 14 illustrates a system 1400 for generation of an antimicrobial gas and/or solution.

[0037] FIG. 15 illustrates a system 1500 for generation of an antimicrobial gas and/or solution.

[0038] FIG. 16 illustrates a system 1600 for generation of an antimicrobial gas and/or solution.

[0039] FIG. 17 illustrates a system 1700 for generation of an antimicrobial gas and/or solution.

[0040] FIG. 18A illustrates a plan view of a reactor 1800 for generating an antimicrobial gas.

[0041] FIG. 18B illustrates a front perspective view of reactor 1800 for generating an antimicrobial gas.

[0042] FIG. 18C illustrates a top perspective view of reactor 1800 for generating an antimicrobial gas.

[0043] FIG. 18D illustrates a front perspective view of reactor 1800 for generating an antimicrobial gas.

[0044] FIG. 19A illustrates a side perspective view of a reactor 1900 for generating an antimicrobial gas.

[0045] FIG. 19B illustrates a side elevational view of reactor 1900 for generating an antimicrobial gas.

[0046] FIG. 19C illustrates a plan view of reactor 1900 for generating an antimicrobial gas.

[0047] FIG. 19D illustrates an exploded side perspective view of reactor 1900 for generating an antimicrobial gas.

[0048] FIG. 19E illustrates an exploded front perspective view of reactor 1900 for generating an antimicrobial gas.

[0049] FIG. 19F illustrates a front perspective view of reactor 1900 for generating an antimicrobial gas.

[0050] FIG. 19G illustrates a rear perspective view of reactor 1900 for generating an antimicrobial gas.

[0051] FIG. 19H illustrates a side perspective view of reactor input mechanism 1962 in a first position.

[0052] FIG. 19I illustrates a side perspective view of reactor input mechanism 1962 in a second position.

[0053] FIG. 20A illustrates a plan view of a reactor 2000 for generating an antimicrobial gas.

[0054] FIG. 20B illustrates a front perspective view of reactor 2000 for generating an antimicrobial gas.

[0055] FIG. 20C illustrates a top perspective view of reactor 2000 for generating an antimicrobial gas.

[0056] FIG. 20D illustrates a front perspective view of reactor 2000 for generating an antimicrobial gas.

[0057] FIG. 21A illustrates a plan view of a reactor 1200 for generating an antimicrobial gas.

[0058] FIG. 21B illustrates a top perspective view of reactor 1200 for generating an antimicrobial gas.

[0059] FIG. 21C illustrates a rear perspective view of reactor 1200 for generating an antimicrobial gas.

[0060] FIG. 21D illustrates a top perspective view of reactor 1200 for generating an antimicrobial gas.

[0061] FIG. 21E illustrates a rear perspective view of reactor 1200 for generating an antimicrobial gas.

[0062] FIG. 22A illustrates a plan view of a reactor 2200 for generating an antimicrobial gas.

[0063] FIG. 22B illustrates a top perspective view of reactor 2200 for generating an antimicrobial gas.

[0064] FIG. 22C illustrates a rear perspective view of reactor 2200 for generating an antimicrobial gas.

[0065] FIG. 22D illustrates a rear perspective view of reactor 2200 for generating an antimicrobial gas.

[0066] FIG. 22E illustrates a front perspective view of reactor 2200 for generating an antimicrobial gas.

[0067] FIG. 23A illustrates a sectional view of an example antimicrobial gas generator 2300.

[0068] FIG. 23B illustrates a partial sectional view of antimicrobial gas generator 2300.

[0069] FIG. 23C illustrates a sectional view of antimicrobial gas generator 2300.

[0070] FIG. 23D illustrates a partial sectional view of antimicrobial gas generator 2300.

[0071] FIG. 23E illustrates a sectional view of antimicrobial gas generator 2300.

[0072] FIG. 23F illustrates a partial sectional view of antimicrobial gas generator 2300.

[0073] FIG. 23G illustrates a partial sectional view of antimicrobial gas generator 2300.

[0074] FIG. 24 illustrates a sectional view of an example antimicrobial gas generator 2400.

[0075] FIG. 25A illustrates an elevation view of an example antimicrobial gas generator and sensor device 2500.

[0076] FIG. 25B illustrates a perspective view of antimicrobial gas generator and sensor device 2500.

[0077] FIG. 25C illustrates a sectional view of antimicrobial gas generator and sensor device 2500.

[0078] FIG. 26A illustrates a perspective view of an example antimicrobial gas generator and sensor device 2600.

[0079] FIG. 26B illustrates a sectional view of antimicrobial gas generator and sensor device 2600.

[0080] FIG. 27A illustrates an elevation view of an example antimicrobial gas generator and sensor device 2700.

[0081] FIG. 27B illustrates a perspective view of antimicrobial gas generator and sensor device 2700.

[0082] FIG. 28A illustrates an elevation view of an example portable antimicrobial gas reactor 2800.

[0083] FIG. 28B illustrates a schematic view of portable antimicrobial gas reactor 2800.

[0084] FIG. 29A illustrates a plan view of an example packaged antimicrobial gas generator solution and packaged activator solution.

[0085] FIG. 29B illustrates a sectional view of packaged antimicrobial gas generator solution and packaged activator solution.

[0086] FIG. 30A illustrates an elevation view of an example of an antimicrobial gas generator 3000 in the form of a card shape or a sheet.

[0087] FIG. 30B illustrates a sectional view antimicrobial gas generator 3000 containing an antimicrobial generating compound.

[0088] FIG. 31 illustrates an example of an antimicrobial generator 3100 in the form of a pouch with optional addition of water internal to the pouch.

[0089] FIG. 32A illustrates an example of an antimicrobial generator 3200 in the form of a solution treated single or multi-ply porous material.

[0090] FIG. 32B illustrates an example of antimicrobial generator 3200 with liquid reactants absorbed or adsorbed on substrates and blended with a porous matrix material with optional addition of an exterior film to control release.

[0091] FIG. 32C illustrates an example of antimicrobial generator 3200 with solid reactants blended in a porous material and optional addition of an exterior film to control release.

[0092] FIG. 32D illustrates an example of antimicrobial generator 3200 in the form of a perforated pouch.

[0093] FIG. 32E illustrates an example of antimicrobial generator 3200 where reactant materials of FIGS. 32A-32C are configured side by side with optional materials to support activation and control release.

[0094] FIG. 33A illustrates a cutaway view of an aerosol container 3386 including an interrupted dip tube 3390.

[0095] FIG. 33B illustrates a cutaway view of aerosol container 3386 including reactor 2100, 2200 engaged with dip tube 3390.

[0096] FIG. 33C illustrates a cutaway view of aerosol container 3386 including reactor 2100 engaged with dip tube 3390.

[0097] FIG. 33D illustrates a cutaway view of aerosol container 3386 including reactor 2200 engaged with dip tube 3390.

[0098] FIG. 34 illustrates a cutaway view of an aerosol container 3486 including a flexible bladder 3492 connected to a dip tube 3490.

[0099] FIG. 35A illustrates a cutaway view of an aerosol container 3586 including a plurality of flexible bladders 3594.

[00100] FIG. 35B illustrates a cutaway view of aerosol container 3586 including a plurality of flexible bladders 3594.

[00101] FIG. 36 illustrates a schematic diagram of an apparatus 3600 for generating antimicrobial gas or vapor external to a sealed environment for disinfecting items therein.

[00102] FIG. 37 illustrates a schematic diagram of a system 3700 generating antimicrobial gas or vapor external to a sealed environment for disinfecting items therein.

[00103] FIG. 38A illustrates a schematic diagram of a system 3800 generating antimicrobial gas or vapor within a sealed environment for disinfecting items in the sealed environment.

[00104] FIG. 38B illustrates a schematic diagram of apparatus 3800 generating antimicrobial gas or vapor within a sealed environment for disinfecting items in the sealed environment.

[00105] FIG. 39A illustrates an apparatus 3900 generating antimicrobial vapor within a sealed environment for disinfecting items therein.

[00106] FIG. 39B illustrates apparatus 3900 generating antimicrobial vapor within a sealed environment for disinfecting items therein.

[00107] FIG. 39C illustrates apparatus 3900 generating antimicrobial vapor within a sealed environment for disinfecting items therein.

[00108] FIG. 40 illustrates methods of generating antimicrobial gas or vapor within a sealed environment or external to the sealed environment to disinfect items within the sealed environment.

[00109] FIG. 41A illustrates ClO₂ efficacy test data on controlled samples.

[00110] FIG. 41B illustrates ClO₂ efficacy test data on controlled samples.

[00111] FIG. 42 illustrates an example of an apparatus 4200 that generates antimicrobial gas or vapor for disinfecting items in three-dimensional space.

[00112] FIG. 43 illustrates an example of an apparatus 4300 that generates antimicrobial gas or vapor for disinfecting items in three-dimensional space.

[00113] FIG. 44 illustrates an example procedure 4400 for the use of apparatus 4300 in FIG. 43 to generate antimicrobial gas.

[00114] FIG. 45 illustrates a table showing temperature effects to solubility of ClO₂ gas in water and in air and required amount of ClO₂ gas for a defined room size.

[00115] FIG. 46 illustrates a uniformity of ClO₂ gas concentration distributed within a room.

- [00116] FIG. 47 illustrates gas concentration profiles in room setting with furniture.
- [00117] FIG. 48 illustrates relative humidity and generated ClO₂ gas concentration from a ClO₂ solution.
- [00118] FIG. 49 illustrates a correlation of increase in disinfection efficacy with elevated humidity.
- [00119] FIG. 50 illustrates a method for generating an antimicrobial gas and dispersing the gas via an apparatus.
- [00120] FIG. 51A illustrates the time (minutes) to equilibrium for a target concentration of 0.1 ppm of ClO₂ to air.
- [00121] FIG. 51B illustrates the concentration (ppm of ClO₂ to air) measured at five ports over time (minutes).
- [00122] FIG. 52A illustrates the time (minutes) to equilibrium for a target concentration of 350 ppm of ClO₂ to air.
- [00123] FIG. 52B illustrates the concentration (ppm of ClO₂ to air) measured at 12 ports over time (minutes).
- [00124] FIG. 53A illustrates a diagram of an example system 5300 for generating ClO₂ vapor from small volumes of high concentration liquid precursors.
- [00125] FIG. 53B illustrates a diagram of example system 5300 for generating ClO₂ vapor from small volumes of high concentration liquid precursors.
- [00126] FIG. 54A illustrates results of ClO₂ generation using system 5300 or similar systems.
- [00127] FIG. 54B illustrates results of ClO₂ generation using system 5300 or similar systems.
- [00128] FIG. 54C illustrates requirements for ClO₂ generation using system 5300 or similar systems.
- **FIG. 55A** illustrates the mean D-values (hours) from replicate tests per organism performed at the range of 0.11 ± 0.04 ppmv.
- [00130] FIG. 55B illustrates the mean D-values (hours) from replicate tests per organism performed at the range of 5.3 ± 2.4 ppmv.

DETAILED DESCRIPTION

Closed Loop Antimicrobial Concept

[00131] A system is provided including an interconnection of platform component elements described below. FIGS. 1A-1C illustrate a system 100 for generating and monitoring an antimicrobial gas. FIGS. 2A-2E illustrate a system 200 for generating and monitoring an antimicrobial gas. Platform component elements may be used individually or in combination to implement a system and device to create, maintain, optimize and/or document the presence of a concentration of an antimicrobial agent in a volume under treatment 124.

The system (such as systems 100, 200) is capable of maintaining an antimicrobial agent in the atmosphere of volume under treatment 124, and may include: (1) controlled release of antimicrobials to maintain a target antimicrobial concentration in volume under treatment 124; (2) at least one type of sensor 108, within volume under treatment 124, and possibly several sensors 108 or several types of sensors 108, are used to sense the concentration of the antimicrobial; (3) a computational system 106 that can compare the measured difference between the antimicrobial concentration sensing and a target antimicrobial concentration in volume under treatment 124; (4) an antimicrobial generator 110 (which may be connected to computational system 106) capable of initial establishment and maintenance of a target antimicrobial concentration in volume under treatment 124; (5) where a computed difference between a target antimicrobial concentration and a sensed antimicrobial concentration is determined, a target control may adjust antimicrobial generation to maintain the target antimicrobial concentration; (6) at least one base safety assurance implementation at the physical components of system 100, 200, electronic hardware, and firmware to software levels of the product.

[00133] System 100, 200 may be designed for modes of operation to prevent transmission or infection between humans in occupied spaces, as well as modes of operation wherein unoccupied rooms can be treated. To maintain target antimicrobial concentrations, system 100, 200 may separate the durable reusable components from disposable components to maintain refill and physical-digital control across deployed system elements.

[00134] Regarding the antimicrobials, the self-degradation kinetics and kinetics of inactivation to log-kill microbes may depend upon more than just the concentration of the antimicrobials in the volume under treatment 124. Thus, system 100, 200 may include a broad spectrum of environment sensing to enable system 100, 200 to use machine learning and artificial intelligence, including for example, enhanced target control, automated volume estimation, humidity measurement, and programmatic antimicrobial cycles.

[00135] Antimicrobial generator 110 designs may use matter displacement (including positive displacement pumps) to activate systems, many of which may have an electronic signal that can be harvested to enable enhanced safety assurance utilizing signals collected by a microprocessor/microcontroller 104 that may be part of computational system 106.

[00136] System 100, 200 may use external communication 102 to form a connectivity network designed to utilize distributed system data of the aforementioned variables of interest to enable the network coordination of distributed product nodes, and the correspondingly required strategy of spatial and temporal identification constants durably and/or variably assigned to system 100, 200 products.

[00137] System 100, 200 may use a combination of platform components, to create an antimicrobial dashboard system. System 100, 200 may provide real-time as well as historical data on infection control, either for safety and health in a user's own spaces, or in high requirements markets such as healthcare facilities. The antimicrobial dashboard system may be used to map a data lake of environment sensing, target antimicrobial concentrations, and use of the connectivity network to deliver distributed system data on the distributed product nodes, which may be identified by unique spatial and temporal identifications, and combine all of this data into human-meaningful information.

[00138] Distributing the intelligence (e.g., computational system 106), sensing (e.g., sensor 108), and generation (e.g., antimicrobial generator 110) may enable the development of a digital twin of space for antimicrobial control. This concept may enable additional network safety assurance implementations and may contain all of the information required to develop and deploy proactive strategies in system 100, 200 products such as a predictive antimicrobial control.

[00139] System 100, 200 includes the ability to combine platform components in multiple ways to achieve product implementation options that are designed specifically for rooms in buildings and provide digital control to low-concentration of an airborne antimicrobial. This antimicrobial may be used to fight transmission and infection caused by microbe-emitting beings and microbes that are circulated through the air currents in rooms, adjacent rooms via open infiltration/exfiltration passages, and shared HVAC systems.

[00140] As illustrated in FIGS. 1A-1C, system 100 may include platform components including external communications 102 devices, microprocessors/microcontrollers 104, computational system 106, antimicrobial and/or environmental sensors 108, and antimicrobial generator 110. System 100 may be entirely contained inside of volume under treatment 124. Optionally, system 100 may be contained within volume under treatment 124 and supplemented

with one or more additional sensor sub-systems 120 configured to provide additional data, including antimicrobial concentration and/or environmental data.

[00141] As illustrated in FIGS. 2A-2E, system 200 may include both a sensor sub-system 120 and a generation sub-system 122. Sensor sub-system 120 may include external communication 102 devices, a microprocessor/microcontroller 104A, computational system 106, and antimicrobial and/or environmental sensors 108. Generation sub-system 122 may include external communication 102 devices, a microprocessor/microcontroller 104B, and antimicrobial generator 110. System 200 may be entirely contained inside of volume under treatment 124.

[00142] As illustrated in FIG. 2C, system 200 may, in a first aspect, include sensor subsystem 120 within volume under treatment 124, and generation sub-system 122 outside of volume under treatment 124. In the first aspect, generation sub-system 122 generates an antimicrobial and via a fluid connection to an HVAC air supply 126, directs antimicrobial into the interior of volume under treatment 124. The concentration of antimicrobial within volume under treatment 124 is sensed by sensor sub-system 120.

[00143] As illustrated in FIG. 2C, system 200 may, in a second aspect, include generation sub-system 122 within volume under treatment 124, and sensor sub-system 120 outside of volume under treatment 124. In the second aspect, generation sub-system 122 generates an antimicrobial within volume under treatment, and via a fluid connection to an HVAC air return 128, sensor subsystem 120 senses the concentration of antimicrobial within volume under treatment 124.

[00144] As illustrated in **FIG. 2D**, system 200 may include a sensor sub-system 120 within volume under treatment 124, and generation sub-system 122 outside of volume under treatment 124, wherein generation sub-system 122 is in fluid communication within volume under treatment 124 to place generated antimicrobial within volume under treatment 124.

[00145] As illustrated in FIG. 2E, system 200 may include a sensor sub-system 120 outside of volume under treatment 124, and generation sub-system 122 within volume under treatment 124, wherein sensor sub-system 120 is in fluid communication within volume under treatment 124 to sense generated antimicrobial concentrations within volume under treatment 124.

[00146] Volume under treatment 124 is conceptually a volume in which a user seeks to distribute an antimicrobial. The volume may be sealed permanently or temporarily to isolate the volume for a period of time. The volume may have openings through which atmosphere can be allowed to infiltrate and/or exfiltrate before, during, or after distributing an antimicrobial. The infiltration/exfiltration can be a characteristic of the volume that is either an uncontrolled variable due to consequence of the volume configuration, or active control strategies of infiltration/exfiltration of atmosphere.

[00147] The target volumes may be the living spaces where human beings gather for work, activities, entertainment, and/or their domiciles. Therefore, product targets may be volumes that can be termed rooms, with groups of rooms forming floorplans, collections of floorplans that form a building, and collections of buildings that comprise a facility.

[00148] Modular platform components may be extended into other volumes under treatment 124, including for example: (1) mobile vehicles such as the interiors of cars, trains, subways, airplanes, recreational vehicles, ride share vehicles, autonomous vehicles, cabins in ships, and the like; (2) leisure spaces such as restaurants, nightclubs, bars, churches, community centers, libraries, and the like; (3) hospital spaces such as hospital rooms, operating rooms, procedure rooms, patient examination rooms, vivariums, morgues, and the like; and (4) business spaces such as offices, conference rooms, hallways, cafeterias, coffee and lounge areas, and the like.

[00149] Target antimicrobial concentrations may be a setpoint desired for antimicrobial release into a volume under treatment 124. The concentration of an antimicrobial in the air can be expressed in relative ratios such as percentages, parts-per-million ("ppm"), or parts-per-billion ("ppb"), and similar terms. As the term is used herein, ppm and ppb are based upon volume.

International standard terms are often used to describe antimicrobial concentrations similar to how industrial chemicals are regulated. Important to system 100, 200 product designs is to treat the air in rooms where people live, work, and play. Regulatory terminology for antimicrobial concentrations in the air in volume under treatment 124 include: (1) recommended/permitted exposure limit, abbreviated "REL/PEL," are concentration and time exposure limits safe for human occupation based upon historical studies and evidence; (2) immediately dangerous to life or health, abbreviated "IDLH," is a concentration at which human exposure can begin to quickly cause an adverse reaction; (3) lethal concentration with 50% mortality, abbreviated "LC-50," is a concentration at which a time-based exposure to an airborne concentration shown to have a mortality rate of 50% in animals exposed in a trial of time at concentration; and (4) lethal dose with 50% mortality, abbreviated "LD-50," is an immediate dose extrapolated from animal trials where a mortality rate of 50% is observed from a single large dose, including air measured as near-immediate mortality at an airborne concentration.

[00151] The first target antimicrobial concentrations include:

[00152] (1) prevention mode in occupied volumes: simple target number typically predicated upon, but not necessarily constrained to, known and published REL/PEL from regulatory bodies. The objective of the prevention mode is to maintain a known-safe concentration of an antimicrobial in the air in which humans can occupy for a meaningful length

of time, typically defined by safety regulators in the context of a "work shift" between 8 to 10 hours. The objective of the concentration is to limit and/or eliminate the transmission potential and/or infection potential of microbes that are already present in a room, or are being emitted into the room by other living beings or room systems like HVAC;

[00153] (2) decontamination mode in unoccupied volumes: simple target number typically predicated on, but not necessarily constrained to, known and published IDLH from regulatory bodies. One objective of decontamination mode is to enable the use of higher concentration levels of an airborne antimicrobial that can shorten the time required to inactivate/kill microbes that need elimination faster, are more difficult to kill organisms (such as spores) or are typically easier to kill but that are partially protected in nutrient rich soils, fluids in obvious to hidden locations, and are suspected or confirmed in a specific volume under treatment 124. Targeting the range near to or below the IDLH includes a likelihood that a person who accidently or purposefully walks into volume under treatment 124 will notice effects associated with the IDLH such as watery eyes, nasal irritation, and other immediately dangerous but not lethal concentrations;

[00154] (3) emergency decontamination volumes: target number potentially selected where a highly dangerous concentration of and/or highly resistant species of microbe require an emergency decontamination of volume under treatment 124. Once volume under treatment 124 is isolated and evacuated, system 100, 200 products could be set by authorized users to perform higher concentration "civil defense mode" concentrations that are at or exceed the LC-50 and LD-50, therefore requiring a degree of user interaction and implementing physical safety safeguards that such a mode will not be an automated mode.

[00155] Sensors 108 and sensor sub-system 120 can include a broad range of sensing technologies to determine the concentration of the antimicrobial in volume under treatment 124.

[00156] Any one or a combination of these sensing technologies may be utilized for many different species of antimicrobials, including for example ClO₂, which is part of the class of oxidizing antimicrobials, which may additionally include: hydrogen peroxide, dry hydrogen peroxide, ozone, nitric oxide.

[00157] System 100, 200 may incorporate any combination of the following sensors 108 to achieve digital control: (1) electrochemical sensors that utilize a depletable chemical which reacts with the antimicrobial, and an electrical circuit that measures the effect of this chemical reaction using measures of charge, voltage, current, conductivity, resistivity, and the like to provide a signal that is in proportion to the known capable range of the sensors. An example of electrochemical sensors for ClO₂ include sensors from Analytical Technologies, Inc.; (2) MOx sensors (metal oxide semiconductor sensors) are widely used in air quality measurement, typically

for airborne pollutants such as H2S, volatile organic compounds, and are known to work to sense gaseous oxidizing species. Two examples of these MOx sensors include the Sensirion SGP40 and the Renesas ZMOD4410 family of sensors.

[00158] Advantages of MOx sensors over electrochemical sensors may include: (a) 10-year lifetimes with no chemicals to deplete; (b) calibration and training values last the lifetime of the sensor; (c) sensors can be "trained" to gas species of interest. The number of gases the sensor can be trained to is not limited by choices of chemical species in the sensor, therefore, as opposed to electrochemical sensors, one MOx sensor can be used to sense multiple antimicrobial species of interest, as well and complementary and potentially interfering gases, without requiring use of different chemicals, membranes, or other interaction/barrier methods to provide species specificity.

Alternative sensing solutions may be able to sense an antimicrobial species to the parts-per-billion to parts-per-trillion levels of concentration expected in the prevention mode in occupied volumes. These alternatives may include: (1) Colorimetry: using a chemical "dye" that interacts with the antimicrobial species of interest and causes a reaction that can be observed be electronic color sensors. The "color" can be in the spectrum of visible, infrared, UV, and other wavelengths of light. The fundamental output of such a system would be an electronic signal that is proportional to the "color change" expected for known chemical interactions that underpin such sensing technologies; (2) Fluorescence: if the antimicrobial species fluoresces, or can be bound to a chemical species that is selective and can be sensed via fluorescence, the magnitude of the fluorescence can be sensed and calibrated to known sources to translate fluorescence levels sensed into and electronic signal that is proportional to said fluorescence.

[00160] Electronic and/or computational controls (computational system 106) act as the "heart" and "brains" of a system 100, 200 product. While there are electronic analog, field-programmable gate array ("FPGA"), and discrete circuitry methods that may work for control, the digital solutions designed for low power battery-powered connected products are particularly beneficial for wireless system 100, 200 products.

[00161] Microprocessors or microcontrollers 104 may form the control intelligence backbone of system 100, 200 products. Microprocessors may be used as these may be required for the embodiments of certain simple safety assurance systems.

[00162] Microprocessor unit 104 may be the central processing core electrically connected to all of the elements of the system 100, 200 platform components.

[00163] Antimicrobial generator 110 is any of a variety of subcomponents responsible for the generation and/or dispersion of an antimicrobial into volume under treatment 124. A large variety of antimicrobial generators 110 is discussed herein, including:

[00164] (1) compressed matter release: an antimicrobial stored in a compressed state is released by a pressure reducing regulator. For example, a canister of antimicrobial gas connected to a pressure reducing regulator, which when opened, allows compressed antimicrobial gas to flow out of the canister to an uncompressed state. A mass flow controller in the path of the matter being transformed from a compressed to an uncompressed state can provide quantitative measurement of the quantity of antimicrobial released;

[00165] (2) two or more chemical activation: two or more precursors are combined to cause a chemical reaction that generates the desired antimicrobial. The two or more precursors can be mixed in passive or active structures, including microfluidic structures to accelerate reaction kinetics. Examples of systems contained herein utilizing this concept include, without limitation: reactors 1800, 1900, 2000, 2100, and 2200; gas generators 2300, 2400, and 3000; gas reactor 2800; antimicrobial generators 3100 and 3200; and aerosol containers 3486 and 3586;

[00166] (3) electrochemical activation: voltage potential and/or current can be varied to control species release and kinetics of antimicrobial generation. In one aspect, termed a flow-through electrochemical cell, NaClO₂ can be flowed over electrodes and recycled until depleted by the electrochemical cleaving of Na from NaClO₂. In another aspect, the precursor material can be contained in a static volume into which electrodes are co-located to generate the electrochemical cleaving of Na from NaClO₂ until the bulk fluid is depleted. In another aspect, ClO₂ is electrochemically generated from a solution of NaClO₂ as the anolyte that is separated from a catholyte by a membrane. Each anolyte and catholyte is in communication with at least one electrode, and a membrane plays an active role in increasing the yield or desired species of antimicrobial (e.g., ClO₂) while sequestering undesired species in the catholyte like Na (in this example for ClO₂). In another aspect, a thin layer of sodium chlorite is flowed in a closed, open, or one-sided membrane channel where material could be introduced to an electrochemical cell designed to generate ClO₂ only from the small quantity of NaClO₂, after which the depleted precursor is transferred to a waste container and the processed is repeated.

[00167] Systems 100, 200 may be a platform that includes durable reusable components and disposable components. The disposable components may include refill cartridges. The refill cartridges may include precursors or direct antimicrobial in a concentrated form. Refill cartridges may include a reservoir. Refill cartridges may include platform components that are prone to failure from wear, including for example, pumps, sensors, and the like.

[00168] Systems 100, 200 may include digital and physical signals to achieve the changing of modes as described above. System 100, 200 may be used either in occupied volumes under treatment 124.

[00169] As such, one class of refill cartridges can incorporate a mechanism (physical, electrical/digital, or both) to limit the base unit into which it is installed to operate in an occupied volume under treatment 124 mode (e.g., prevention mode in occupied volumes) and REL/PEL concentration levels.

[00170] Another class of refill cartridges can incorporate a mechanism (physical, electrical/digital, or both) to allow the base unit into which it is installed to operate in a decontamination mode (e.g., decontamination mode in unoccupied volumes). Additional features may include a mechanism limiting installation to a subset of users who are authorized to install the decontamination mode cartridge. These features may include a requirement to enter an appropriate electronic or digital authorization (e.g., a code, swipe a keycard, enter a biometric pass, or the like) to unlock a decontamination mode that would be inappropriate for occupied spaces. Such a decontamination mode may utilize IDLH or higher concentration levels and may be suitable for regular or exceptional "deep-clean" scenarios.

[00171] System 100, 200 may use a combination of platform components, to create an antimicrobial dashboard system. The dashboard system may combine distributed intelligence, distributed data across the system, and other platform components to enable beneficial system features, including for example: (1) a room, floor, building control dashboard for antimicrobial treatment; (2) provide notifications to phones that are nearby a base unit; (3) system 100, 200 coordination in physically adjacent volumes under treatment 124; (4) antimicrobial output coordination of multiple units in a single contiguous volume (e.g., a large open space such as a concert hall); (5) data portability for integration into building management systems, such as hospital command centers; and (6) civil defense alert network for biological threats or attacks.

[00172] Each system 100, 200 unit may securely connect (IoT connections), for purposes of data collection and storage, software and firmware updates, and/or user interactions, to assign: (1) unique identifiers for each hardware unit; (2) unique identifiers for each refill unit; (3) and/or two different types of refill units (one for when in low-concentration occupied mode, and another for authorized user to change to unoccupied decontamination mode).

[00173] Additionally, each system 100, 200 unit may securely connect to each other (via external communication 102) and may pass identification validation data as well as recorded operational performance data along to a data gathering point. Each unit may record its own data, and if necessary for redundancy and safety, neighboring unit data. In one aspect, each unit may

connect to a WiFi hub to achieve interconnectivity. In another aspect, all units may be required to connect to a central identification validation and data gathering point.

[00174] Computational system 106 may include local storage mediums for each system 100, 200 unit. Alternatively, one unit can have storage capability and may act as an accumulator for multiple units in a logical grouping. All units or all accumulators may be required to report up into a central data gathering point, which may also be a point of connection into cloud data.

In one aspect, system 100, 200 units may have safety features including: (1) input received from each unit, including location, environmental sensor suite, antimicrobial sensor data, quantity of antimicrobial generated, and/or corresponding time stamps; (2) output to user and/or system controls including possible safety signal generation based upon: (i) operational parameters that do not make sense and thus that particular unit may be malfunctioning, (ii) recognition that a neighboring unit has experienced an error can initiate "alert status" among a local group of units, and a group or region of units could be powered off if airflow-dictated interactions between two local units cause interference, (iii) client and/or host operations control: the control system will watch for signals of parameters that do not make sense, across the entire installation of units.

[00176] System 100, 200 may include machine learning algorithms. For example, machine learning algorithms may use a multi-sensor suite to both measure and classify at least two fundamental characteristics of airborne microbial concentration in volume under treatment 124.

[00177] System 100, 200 may include the capability to automatically measure the volume of any given volume under treatment 124. Sensor array 108 may be utilized to automatically measure room volume so that generator 110 closed-loop performance can be translated from a concentration in the air to a value of required make up antimicrobial that will move the concentration from a measured value to the target concentration within volume under treatment 124. System 100, 200 units may generate and emit a known test quantity of the antimicrobial upon initialization. The unit may initiate continuous antimicrobial sensor 108 readings while generator 110 is kept idle for a period of time between 1 min to 4 hours. On-unit computation capability measures peak concentration and uses machine learning aspect 1 ("ML1") to measure Understanding that concentration = mass (derived from sensed dispensed room kinetics. antimicrobial volume, directly or indirectly sensed precursor utilization, mass flow measurement of antimicrobial gas, or any other value that can be traced back to quantity) of antimicrobial divided by volume of volume under treatment 124. The volume of volume under treatment 124 is determined by using the measured quantity of antimicrobial generated and antimicrobial concentration reading at a time appropriate to the room kinetics measured with ML1. System 100, 200 may iterate with each antimicrobial gas emission to update ML1 room kinetics estimates,

while cataloging changes by time stamp. As machine learning aspect 2 ("ML2") "learns" from the data lake or direct verification experiments, future algorithms may be designed to provide input data to the generator to predict the specific quantity of antimicrobial needed to achieve the desired concentration based upon environment conditions within volume under treatment 124. Alternatively, or as a backup method, system 100, 200 may use a three-dimensional laser measuring system, or a tone emitter and microphone on units to ping the volume of volume under treatment 124 with a CHIRP acoustic signal. Measuring time of flight and collision of sound waves, system 100, 200 may build a characteristic volume estimate of volume under treatment 124.

[00178] FIGS. 3 and 4 illustrate schematics of example systems 300 and 400 for generating and monitoring an antimicrobial gas (including a disinfection gas and/or decontamination gas). The antimicrobial gas may be a ClO₂ gas. Systems 300 and 400 may include a microfluidic liquid dispensing and metering system. Systems 300 and 400 may be used to both generate antimicrobial gas (e.g., ClO₂ gas) and dispense the antimicrobial gas (e.g., ClO₂ gas) to the ambient environment, and to sample the ambient air to identify antimicrobial gas concentration therein and generate more or less antimicrobial gas as necessary to maintain a desired antimicrobial gas concentration. Systems 300 and 400 may be used to test air in a particular environment (e.g., a three-dimensional enclosed space) to determine the concentration of antimicrobial gas (e.g., ClO₂ gas) in parts per billion ("ppb") of air. Systems 300 and 400 may be used to maintain a desired antimicrobial gas concentration in ambient air surrounding devices housing systems 300 and 400 by regularly sampling the ambient air, determining the concentration of antimicrobial gas in the ambient air, and via closed-loop control of the device, generating more or less antimicrobial gas to maintain the desired antimicrobial gas concentration in the ambient air.

[00179] Systems 300 and 400 may include wired connections to a computer network, cloud storage, or the like. Systems 300 and 400 may include wireless connections to a computer network, cloud storage, or the like. Systems 300 and 400 may document time-based tracking of system use, product maintenance, target concentration performance, and environmental parameters of interest. This documentation may be in the form of files, logs, or other records stored locally within a device housing system 300 and/or 400 or transmitted via wired connection or wirelessly to a computer network, cloud storage, or the like. Systems 300 and 400 may have cloud and/or IoT connectivity to enable user personas to effectively set up, train, manage, and maintain devices housing systems 300 and/or 400 in the three-dimensional enclosed spaces under treatment, view real-time and stored performance and environment data, and/or export data to compare validation tests such as animal and human exposure trials.

[00180] Systems 300 and 400 may be used to decontaminate (that is, to inactivate or destroy pathogens) a three-dimensional enclosed space (e.g., a hospital room) through high concentrations of antimicrobial gas (e.g., ClO₂ gas) (when unoccupied by humans), or through low concentrations of antimicrobial gas (e.g., ClO₂ gas) (when occupied by humans). In one aspect, systems 300 and 400 generate antimicrobial gas in a concentration of 1,000 ppb to 5,000 ppb or 50,000 to 300,000 ppb to decontaminate an unoccupied three-dimensional enclosed space. Systems 300 and 400 may destroy the COVID-19 within a three-dimensional enclosed space.

[00181] Systems 300 and 400 may be used to prevent the spread and/or survival of a virus in a three-dimensional enclosed space (e.g., a hospital room) through low concentrations of antimicrobial gas (e.g., ClO₂ gas) (whether occupied by humans or not). In one aspect, systems 300 and 400 generate antimicrobial gas in a concentration of less than 100ppb, for example 50 ppb, to prevent the spread and/or survival of a virus in an occupied three-dimensional enclosed space. Systems 300 and 400 may reduce aerosolized virus transmission and infection of viruses including COVID-19. Systems 300 and 400 may inactivate and/or kill airborne pathogens, and even protect against airborne contagions.

[00182] System 300 and/or 400 may be contained within a device housing 304. Ambient air 302 may enter one or more inlet in device housing 304. Ambient air 302 may pass through a particulate filter within device housing 304. The particulate filter may not exclude any atmospheric molecules.

[00183] Ambient air 302 passes from device housing 304 into one or all of air pumps 308A, 308B, and 308C via one or more air ducts. System 300 includes air pumps 308A, 308B, and 308C, while system 400 only includes air pumps 308A and 308C, as will be further explained below.

[00184] A microcontroller 306 may control all on-board functions of system 300 and 400. Microcontroller 306 includes software that can be written to change system 300 and 400's functions where necessary. Microcontroller 306 is operatively connected to various elements (described further below) of systems 300 and 400 via wired or wireless connection.

[00185] Microcontroller 306 is connected to air pumps 308A, 308B, and 308C as illustrated, and controls the function of 308A, 308B, and 308C, including one or more of start, stop, velocity, flow rate, pressure, and the like. Air pumps 308A, 308B, and 308C may be disc pumps. In one aspect, air pumps 308A, 308B, and 308C may be capable of producing pressure in excess of 270 mbar, flow rates in excess of 0.55 L/min, and vacuum in excess of 220 mbar. Air pumps 308A, 308B, and 308C may include separate motor control units. Air pumps 308A, 308B,

and **308C** may include integrated motor control units. It is understood that system **400** does not include air pump **308B**.

[00186] Air pumps 308A and 308B in system 300 are connected to pressure relief valves 310A and 310B, such that excess or unnecessary pressure produced by air pumps 308A and 308B may be routed out of system 300. System 400 likewise includes a pressure relief valve 310A having the same function but does not include a pressure relief valve 310B. Alternatively, as illustrated in FIGS. 6 and 7, systems 600 and 700 eliminate at least air pumps 308A and 308B as reagent containers 312A and 312B may be pressurized prior to assembly of systems 600 and 700, and thus air pumps 308A and 308B are unnecessary.

[00187] System 300 includes reagent containers 312A and 312B, each containing a different liquid reagent 314A and 314B. Reagents 314A and 314B may be combined within a microfluidic mixer 320 to generate ClO₂ gas. One of reagents 314A and 314B may be a liquid precursor such as NaClO₂ (sodium chlorite). The other of reagents 314A and 314B may be a liquid activator such as an acid/H+ activator.

[00188] With respect to system 300, air pump 308A pressurizes reagent container 312A, thus causing reagent 314A to travel from reagent container 312A, through a passage into an electronically operated normally closed valve 316A (which is connected to a controlled by microcontroller 306). From valve 316A reagent 314A travels through a microfluidic flow sensor 318A (which is used for closed loop control signals and is connected to and provides data to microcontroller 306), and into microfluidic mixer 320. It is contemplated that any pressure generator may be used in lieu of air pump 308A to pressurize reagent container 312A. In one aspect, reagent container 312A may be pressurized by an external source during assembly of system 300, and a valve connected to reagent container 312A (e.g., valve 316A) may open to permit the passage of a quantity of pressurized reagent to exist reagent container 312A and proceed into microfluidic mixer 320 as described above. Such a system is illustrated in FIG. 6.

[00189] With respect to system 300, air pump 308B pressurizes reagent container 312B, thus causing reagent 314B to travel from reagent container 312B, through a passage into an electronically operated normally closed valve 316B (which is connected to and controlled by microcontroller 306). From valve 316B reagent 314B travels through a microfluidic flow sensor 318B (which is used for closed loop control signals and is connected to and provides data to microcontroller 306), and into microfluidic mixer 320. It is contemplated that any pressure generator may be used in lieu of air pump 308B to pressurize reagent container 312B. In one aspect, reagent container 312B may be pressurized by an external source during assembly of system 300, and a valve connected to reagent container 312B (e.g., valve 316B) may open to

permit the passage of a quantity of pressurized reagent to exist reagent container 312B and proceed into microfluidic mixer 320 as described above. Such a system is illustrated in FIG. 6.

[00190] Microfluidic mixer 320 may be a planar chape designed for low dead space volume and effective mixing to increase reaction kinetics of precursors.

The mixture of reagents 314A and 314B in microfluidic mixer 320 creates an antimicrobial gas, including for example, ClO₂ gas. Antimicrobial gas may pass via a passage into an off-gas and waste chamber 322. Chamber 322 may include an absorber material, an evaporator, or the like. Within chamber 322, any waste from the creation of antimicrobial gas may be absorbed in an absorber material. Chamber 322 may include a membrane. Antimicrobial gas may exit chamber 322 into the ambient atmosphere. In one aspect, antimicrobial gas exits chamber 322 through the membrane. System 300 and 400 may include a device for separation of antimicrobial gas (e.g., ClO₂ gas) and post-generator waste in communication with one or more air pumps and air ducts to one or more outlets, and on-device or in-device waste storage prior to disposal. Chamber 322 may act as the device for separation of antimicrobial gas and post-generator waste. Chamber 322 may act as both the device for separation of antimicrobial gas and post-generator waste and the device for on-device waste storage prior to disposal.

[00192] With respect to system 400, system 400 does not include an air pump 308B, pressure relief valve 310B, reagent container 312B, reagent 314B, valve 316B, or a microfluidic flow sensor 318B. Further, system 400 substitutes microfluidic mixer 320 with a microfluidic electrochemical generator 434. In system 400, air pump 308A pressurizes reagent container 312A, thus causing reagent 314A to travel from reagent container 312A, through a passage into an electronically operated normally closed valve 316A (which is connected to and controlled by microcontroller 306). From valve 316A reagent 314A travels through a microfluidic flow sensor 318A (which is used for closed loop control signals and is connected to and provides data to microcontroller 306), and into microfluidic electrochemical generator 434. An electrical current, provided by and controlled by microcontroller 306 within microfluidic electrochemical generator 434 that produces an antimicrobial gas, such as ClO₂ gas. Antimicrobial gas passes from microfluidic electrochemical generator 434 into chamber 322 and ultimately into the ambient environment as described with respect to system 300.

[00193] It is contemplated that any pressure generator may be used in lieu of air pump 308A to pressurize reagent container 312A. In one aspect, reagent container 312A may be pressurized by an external source during assembly of system 300, and a valve connected to reagent container

312A (e.g., valve **316A**) may open to permit the passage of a quantity of pressurized reagent to exist reagent container **312A** and proceed into microfluidic mixer **320** as described above. Such a system is illustrated in **FIG. 7**.

[00194] Electronically operated normally closed valve 316A, 316B may be controlled by microcontroller 306, and may be oriented such that when no power is provided, valve 316A, 316B is open.

[00195] Microfluidic flow sensor 318A, 318B may sense the flow of reagent 314A, 314B, respectively, and may provide data regarding that flow to microcontroller 306. Such data may include flow rate, flow volume, flow time, mass, and the like.

may sense the pressure within the three-dimensional enclosed space that systems 300 and 400 operate. Upon sensing a negative pressure (indicating that a HVAC return system is pulling air out of the room, a door or window is open, or the like), barometric sensor 328 may communicate the negative pressure via its connection with microcontroller 306, upon which microcontroller 306 may pause antimicrobial gas (e.g., ClO₂ gas) generation until a neutral and/or positive pressure is sensed by barometric sensor 328. Upon sensing a neutral or positive pressure, barometric sensor 328 may communicate the neutral or positive pressure to microcontroller 306, at which point microcontroller 306 may once again initiate gas generation (e.g., ClO₂ gas).

[00197] Systems 300 and 400 may include an air quality sensor 330. Air quality sensor 330 may sense any of a variety of ambient air 302's characteristics, including for example, humidity, temperature, and the like. Data regarding air quality may be recorded for evaluating the effectiveness of systems 300 and 400. Alternatively, as antimicrobial gas (e.g., ClO₂ gas) may be more effective at destroying pathogens in more humid environments, humidity data, for example, may be communicated via air quality sensor 330's connection with microcontroller 306, upon which microcontroller 306 may adjust the target concentration of antimicrobial gas in ambient air 302 based upon humidity readings.

[00198] The above-described aspects, methods, and processes of systems 300 and 400 demonstrate the generation of antimicrobial gas by each of systems 300 and 400. Below is described the aspects of systems 300 and 400 that sample ambient air 302 to determine the concentration of antimicrobial gas (e.g., ClO₂ gas) within ambient air 302.

[00199] In both systems 300 and 400, ambient air 302 may be ducted to air pump 308C, which causes a sample of ambient air 302 to enter a concentrator 324. Concentrator 324 may separate antimicrobial gas (e.g., ClO₂ gas) from the mostly diamagnetic other components of ambient air 302. One aspect of a concentrator is illustrated in FIGS. 8A and 8B. Concentrator

324 may separate and concentrate a very low concentration of antimicrobial gas (e.g., ClO₂ gas) so that a more accurate measurement of its concentration may be obtained. Concentrator 324 may utilize magnets to separate diamagnetic gases from antimicrobial gas, thus permitting the testing of a concentrated and amplified level of antimicrobial gas. Diamagnetic gases may be passed back into the ambient environment after separation. In one aspect, antimicrobial gas may be amplified at least 100 times prior to further concentration testing.

[00200] Systems 300 and 400 may include a sensing system 326. Sensing system 326 may sense the concentration of antimicrobial gas (e.g., ClO₂ gas) (which may be amplified 100 times or more following processing in concentrator 324). Sensing system 326 may measure a time weighted average of the concentration of antimicrobial gas (e.g., ClO₂) in ambient air 302. Data regarding the concentration is passed to microcontroller 306, and if necessary, microcontroller 306 causes system 300 or 400 to generate more or less antimicrobial gas based upon the concentration measured in sensing system 326.

[00201] After sensing in sensing system 326, the sampled gas passes via a passage to offgas and waste chamber 322 and is ultimately passed into the ambient atmosphere with the generated antimicrobial gas.

[00202] Thus, systems 300 and 400 may measure the concentration of antimicrobial gas (e.g., ClO₂ gas) in ambient air 302, and if the concentration is below the target concentration, microcontroller 306 can cause system 300 or 400 to generate more antimicrobial gas to raise the concentration of antimicrobial gas (e.g., ClO₂ gas) in ambient air 302 until the sampled ambient air 302 meets the target concentration threshold.

All microcontrollers referenced herein (including microcontroller 306), may have the computational ability and local data storage ability to enable closed-loop control of the antimicrobial gas generation system (including systems 300, 400, 600, and 700), including but not limited to: (1) local storage and microcontroller operations on data from sensor systems for antimicrobial gas (e.g., ClO₂) levels to the space environment variables such as barometric pressure, humidity, temperature, occupancy, or sounds that may be used to alter generation system (including systems 300, 400, 600, and 700) performance automatically or via user intervention; (2) measurement, local storage, and microcontroller operations on data from microfluidic subsystems such as mass/volume sensors of reagents, pressure generator performance, microfluidic chip-borne sensors, valve status and/or any other electronic subsystem to provide control as well as storage of system performance data for maintenance, alert, troubleshooting, inactive modes of operation, active modes of operation, and local setup.

[00204] In another aspect, the system (including systems 300, 400, 600, and 700) has a communication device connected to the microcontroller and/or electronic components such that data from any electronic component within, on, or connected to the systems (including systems 300, 400, 600, and 700) or housing 304 can be gathered, locally stored, operated on by the microcontroller, and transmitted to external data gathering systems on mobile and/or fixed devices.

[00205] In another aspect, machine learning and/or artificial intelligence algorithms can be incorporated into the system (including systems 300, 400, 600, and 700) microcontroller (including microcontroller 306) to alter system performance automatically or by user interactions. An example of local control includes alteration of system performance for detection of a virus or bacteria in the ambient air, altitude, temperature, air changes in the local space measured by changes in antimicrobial gas (e.g., ClO₂) concentration in the air of spaces containing antimicrobial gas (e.g., ClO₂), changes in occupancy by living beings, alterations for user preference, prediction of cycles of occupancy/vacancy, alerts as to normal or abnormal performance of the system, and the like. In one aspect, microcontroller 306 is controlled by machine learning algorithms to alter system performance. In another aspect, microcontroller 306 is controlled by artificial intelligence algorithms to alter system performance. Microcontroller 306 may alter system performance automatically. Microcontroller 306 may alter system performance by control by a user. Microcontroller 306 may alter the system performance based upon at least one of: a detection of a virus or bacteria in the ambient air; an altitude of the system; a temperature of the system; changes in the ambient air measured by changes in a concentration of antimicrobial gas (e.g., ClO₂) in ambient air; changes in occupancy by living beings of an area containing the system; alterations for a user's preferences; prediction of cycles of occupancy and vacancy by living beings of the area containing the system; and a diagnosis of normal or abnormal performance of the system.

[00206] In another aspect, the system (including systems 300, 400, 600, and 700) for distribution and monitoring of antimicrobial gas (e.g., ClO2 gas) in a three-dimensional space will be designed for a plurality of operating modes. A first operating mode may be designed for occupied spaces, while a second operating mode may be designed for un-occupied spaces. Future user or engineered operating modes may be added. These operating modes may be changed by authorized users on the system (including systems 300, 400, 600, and 700) network (e.g., network 300) connected to a plurality of system (including systems 300, 400, 600, and 700).

[00207] FIG. 5 illustrates an example blueprint of a network 500 of disinfecting gas (e.g., ClO₂ gas) generator systems 300 and sensors 536 distributed in rooms and spaces in a floor of a

building. Network **500** illustrates a floor of a building bounded by exterior walls **538** and divided by interior walls **540**. Gas generator systems **300** may operate with the configuration and method of systems **300** or **400** described above, or **600** or **700** described below, and thus may include disinfecting gas (e.g., ClO₂ gas) concentration sensors. As illustrated, gas generator systems **300** may be oriented in each individual room of the floor, as well as in open spaces between the individual rooms. Standalone sensors **536** (configured simply to sense the concentration of disinfecting gas, such as ClO₂ gas, in the ambient air) supplement network **500** to ensure that the target concentration is achieved throughout network **500**.

[00208] The various gas generator systems 300 may operate to generate disinfecting gas (e.g., ClO₂ gas) independent of one another, and at different concentration target values depending upon the desired function of a particular gas generator systems 300.

[00209] For example, where a room is occupied by a patient (e.g., in a hospital or nursing facility), employee (e.g., in an office), a guest (e.g., in a hotel), or the like, the gas generator system 300 in that particular room may have a target disinfecting gas (e.g., ClO₂ gas) concentration of about 50 ppb. After the room is no longer occupied (e.g., patient is moved from the room for a set period of time, employee is gone for the night, guest checks out, etc.), the gas generator system 300 in that room may increase its target disinfecting gas (e.g., ClO₂ gas) concentration to about 1,000 ppb to about 5,000 ppb for a set period of time. In this manner, the room can be decontaminated (1,000 ppb to 5,000 ppb concentration level, or 50,000 ppb to 300,000 ppb concentration level for extreme pathogens) between its use by particular individuals, or on a regular time schedule, and maintain a lower safe (to humans) concentration of 50 ppb for prevention or mitigation of virus spreading while occupied.

[00210] In another aspect, a plurality of systems 300 within a plurality of spaces which are arranged into network 500 can be connected via communication devices (as described above) to each other for distributed control via coordination of each system's microcontroller (e.g., microcontroller 306), centralized unit control, and/or a combination of both local and distributed control.

[00211] In another aspect, machine learning and/or artificial intelligence algorithms can be incorporated into the distributed network 500 of systems 300 by the aspects described above. Examples of distributed control include adjusting individual systems 300 to achieve uniform and/or deliberately non-uniform distribution of disinfecting gas (e.g., ClO₂) in each individual generator system 300's location across an entire building floor to the entire building due to changes in disinfecting gas (e.g., ClO₂) concentration from HVAC, consumption or self-dissipation of disinfecting gas, control of day/night generation cycles, sensing patterns across

time, three-dimensional volumes, seasonal variations, and/or previously unknown factors that can be sensed either directly by the sensor systems **300** in/on the network **500**, inferred or traced to the signal measured, or directly traceable to the variations observed in disinfecting gas (e.g., ClO₂) concentrations across a collection of systems **300** installed across distinctly separate and/or varying interconnection of real world spaces in which control of infectious species is desired.

[00212] FIGS. 6 and 7 illustrate schematics of example systems 600 and 700 for generating and monitoring antimicrobial gas (e.g., ClO₂ gas). Systems 600 and 700 are substantially similar to systems 300 and 400, respectively, except that air pumps 308A and 308B and pressure relief valves 310A and 310B are replaced with check valves 609A and 609B. These check valves are one-way, directional flow valves that permit the passage of fluid through check valves 609A, 609B toward reagent containers 312A, 312B, but prevent the passage of fluid away from reagent containers 312A, 312B through check valves 609A, 609B.

[00213] Such an arrangement may be used where reagent containers 312A, 312B are pressurized by an external source before or during assembly of systems 600, 700. Thus, reagent containers 312A, 312B may be pressurized containers housing reagents 314A, 314B, and as such do not need air pumps 308A, 308B to cause reagent 314A, 314B to flow to microfluidic mixer 320. The flow of pressurized reagent 314A, 314B may be controlled by a valve, such as valves 316A, 316B. When valves 316A, 316B are opened, pressurized reagent 314A, 314B may flow from pressurized reagent containers 312A, 312B, through microfluidic flow sensors 318A, 318B, and into microfluid mixer 320.

Antimicrobial Generation Systems and Devices

FIG. 9 illustrates a schematic for a system **900** for generating a disinfecting gas and/or solution. The disinfecting gas and/or solution may be ClO₂ gas and/or solution. System **900** may be used to generate a pure disinfecting gas (e.g., ClO₂ gas). System **900** may create a disinfecting gas (e.g., ClO₂ gas), which may be immediately vented, swept, or evacuated out of a reaction chamber **906** and either used as a disinfecting gas **908** in an end use application or dissolved into water to create a pure disinfecting solution (e.g., ClO₂) **910**.

[00215] System 900 may use a concentrated liquid precursor 902. Liquid precursor 902 may be NaClO₂ (sodium chlorite).

[00216] System 900 may include an activator 904. Activator 904 may be an acid/H+ activator. Activator 904 may be a concentrated liquid activator.

[00217] Liquid precursor 902 and activator 904 may be brought into contact with one another in reaction chamber 906. At least one of liquid precursor 902 and activator 904 may be conveyed into reaction chamber 906 via at least one of a gravity feed, a metered gravity feed,

pressurization via a pump, pressurization via a syringe, pressurization via any mechanism, vacuum/low pressure, microfluidics, or the like. At least one of liquid precursor **902** and activator **904** are conveyed into reaction chamber **906** in correct proportions and rates to meet antimicrobial gas and/or solution (e.g., ClO₂) production requirements.

Disinfecting gas (e.g., ClO₂ gas) **908** may be created in reaction chamber **906** and allowed to escape into the environment within an enclosed space (e.g., a room in a building) for treatment of the air and/or surfaces within the enclosed space; optionally, a polymer membrane may be used to control the rate of antimicrobial (e.g., ClO₂) release. Antimicrobial gas **908** may be dissolved into a liquid to create pure antimicrobial solution (e.g., ClO₂ solution) **910**.

[00219] Antimicrobial solution (e.g., ClO₂ solution) 910 may be at least one of: (a) transferred to a dispensing device 912 (e.g., a spray bottle), which may be diluted with water 914, or (b) transferred to a waste liquid container 916. Solution 910 transferred to a dispensing device 912 may be diluted with another liquid 914, including for example, liquid water. Where solution 910 is not desired (e.g., a user of system 900 only desires the creation of antimicrobial gas (e.g., ClO₂ gas) 908), or more solution 910 than desired is produced, all or some of solution 910 is moved to waste liquid container 916.

[00220] Additionally, any waste liquid created in reaction chamber 906 may likewise be ducted directly to waste liquid container 916. Waste liquid can be removed from system 900 and properly disposed of via a waste liquid outlet 920.

[00221] FIG. 10 illustrates a system 1000 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1000 is substantially similar to system 900 described above, where like reference numbers indicate like elements. System 1000 additionally includes a neutralizing agent 1018 that is ducted to waste liquid container 916. Neutralizing agent or process 1018 may be any agent or process capable of neutralizing the antimicrobial (such as ClO₂) or the reaction between NaClO₂ and the activator that is combined in reaction chamber 906. Neutralizing agent 1018 may include, for example, a chemical agent or a physical process wherein ultraviolet light, carbon, or the like is used for neutralizing an antimicrobial (such as ClO₂) or the reaction between NaClO₂ and the activator. Neutralizing agent 1018 may be used to neutralize an antimicrobial solution (such as ClO₂ solution) (either solution 910 or from reaction chamber 906) to render the solution safe for disposal via waste liquid outlet 920.

[00222] FIG. 11 illustrates a system 1100 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1100 is substantially similar to system 900 described above, where like reference numbers indicate like elements. System 1100 additionally includes an air pressurization device 1122 (e.g., a fan or blower) and an optional air meter 1124.

Air pressurization device 1122 and air meter 1124 may control a flow of air into reaction chamber 906 to safely evacuate antimicrobial gas (such as ClO₂ gas) 908 from reaction chamber 906, or both. Air meter 1124 may control the volume of air fed into reaction chamber 906, the amount of time that air is fed into reaction chamber 906, or both. Air meter 1124 may include a valve to control air flow.

[00223] FIG. 12 illustrates a system 1200 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1200 is substantially similar to systems 1000 and 1100 described above, where like reference numbers indicate like elements. System 1200 is a combination of systems 1000 and 1100, including a neutralizing agent or process 1018, an air pressurization device 1122, and an air meter 1124.

FIG. 13 illustrates a system 1300 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1300 is substantially similar to system 900 described above, where like reference numbers indicate like elements. However, liquid activator 904 is replaced with a solid activator 1330. Activator 1330 may include an acidic chemical powder like citric acid or sodium persulfate, a bed of cationic ion exchange resin/polymer with optional metal-oxide catalyst. Liquid precursor 902 may be exposed to solid activator 1330 via gravity feed, active pumping, etc., as discussed above with respect to system 900. Liquid precursor 902 may flow through solid activator 1330.

[00225] FIG. 14 illustrates a system 1400 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1400 is substantially similar to system 1300 described above, where like reference numbers indicate like elements. System 1400 includes a neutralizing agent or process 1018 as described above in system 1000.

[00226] FIG. 15 illustrates a system 1500 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1500 is substantially similar to system 1300 described above, where like reference numbers indicate like elements. System 1500 additionally includes an air pressurization device 1122 and an air meter 1124 as described above in system 1100.

[00227] FIG. 16 illustrates a system 1600 for generation of an antimicrobial gas and/or solution (e.g., ClO₂ gas and/or solution). System 1600 is substantially similar to systems 1400 and 1500 described above, where like reference numbers indicate like elements. System 1600 is a combination of systems 1400 and 1500, including a neutralizing agent or process 1018, an air pressurization device 1122, and an air meter 1124.

[00228] FIG. 17 illustrates a system 1700 that is substantially similar to system 900 illustrated in FIG. 9 and described above, where like reference numbers indicate like elements.

System 1700 additionally includes pressurized aerosol containers 1786A and 1786B. Aerosol container 1786A may contain liquid precursor 902, while aerosol container 1786B may contain liquid activator 904. Liquid precursor 902 and liquid activator 904 may be delivered to a reaction chamber 906 using the pressure within containers 1786A and 1786B to cause the delivery. Such an embodiment may eliminate the need for electrically powered components for antimicrobial generation and dispersal.

[00229] FIGS. 18A-18D illustrate a reactor 1800 for generating an antimicrobial gas (e.g., ClO₂ gas). Reactor 1800 may be a prepackaged device loaded with a liquid precursor 902 and a liquid activator 904 within containers inside reactor 1800. Liquid precursor 902 may be sealed within its container and may require pressurized air to flow into reaction chamber 906. Liquid activator 904 may be sealed within its container and may require pressurized air to flow into reaction chamber 906.

[00230] Reactor 1800 may include a housing 1840 containing liquid precursor 902, activator 904, a reaction chamber 906, and a waste liquid container 916, wherein device elements are machined or otherwise formed out of a housing material, as in common in the production of microfluidic devices. Reactor 1800 may be a microfluidic device.

[00231] Reactor 1800 may include a pressure input 1841 capable of applying an air pressure to liquid precursor 902 and activator 904 to break seals within their respective containers and/or cause them to travel to reaction chamber 906. Pressure input 1841 may receive pressure from a pump, a syringe, or the like.

[00232] Reaction chamber 906 may include a capillary filter 1842 that permits waste liquid to travel into waste liquid container 916 via capillary action. Waste liquid container 916 may include an inactivator, neutralizing agent, or the like capable of rendering waste liquid from reaction chamber 906 into a safe state.

[00233] Reaction chamber 906 may include a gas permeable membrane 1844, which allows an antimicrobial gas (e.g., ClO₂ gas) created in reaction chamber 906 to pass through membrane 1844 at a controlled rate but prevents a waste liquid from reaction chamber 906 from passing through membrane 1844. Antimicrobial gas (e.g., ClO₂ gas) may exit reactor 1800 via a gas outlet 1843. Gas outlet 1843 may permit antimicrobial gas (e.g., ClO₂ gas) to exit reactor 1800 and enter the surrounding area, including for example an enclosed space (e.g., a room within a building).

[00234] As illustrated in FIG. 18D, reactor 1800 may include a cover 1846 that seals the above-referenced contents (e.g., liquid precursor 902, activator 904, reaction chamber 906, and waste liquid container 916) within housing 1840.

[00235] FIGS. 19A–19I illustrate a reactor 1900 for generating a disinfecting gas (e.g., ClO₂ gas). Reactor 1900 may include a housing 1950 having an upper surface 1952. Housing 1950 may include a container 1970 and a reaction chamber 1972 connected to one another by a chamber duct 1976, wherein device elements are machined or otherwise formed out of a housing material, as in common in the production of microfluidic devices. Reactor 1900 may be a microfluidic device.

[00236] Container 1970 may include a solid container cover 1954 oriented on or near upper surface 1952. Reaction chamber 1972 may include a reaction chamber cover 1956 oriented on or near upper surface 1952. Reaction chamber cover 1956 may include an outlet 1960 having an aperture 1958 in fluid communication with the interior of reaction chamber 1972. Aperture 1958 may include a gas permeable membrane that allows a gas (e.g., ClO₂) to pass through, but prevents a liquid (e.g., waste liquid) from passing through.

[00237] Chamber duct 1976 may include a valve 1980. Valve 1980 may be a check valve, backflow valve, seal, or the like that prevents the contends of container 1970 and the contents of reaction chamber 1972 from coming into contact with one another until a user selectively causes the contents of container 1970 to be transferred to reaction chamber 1972.

[00238] Container 1970 may contain a liquid activator as described above, while reaction chamber 1972 may contain a liquid or solid precursor (e.g., liquid NaClO₂ or solid NaClO₂). Alternatively, container 1970 may contain a liquid precursor (e.g., NaClO₂) as described above, while reaction chamber 1972 contains a solid activator or liquid activator.

Pressurization device may include any device capable of pressurizing the contents of container 1970, thereby causing the contents of container 1970 to overcome and pass valve 1980 and enter reaction chamber 1972. Pressurization device 1962 may include a plunger device including a hollow body 1966 extending from end 1964 and in fluid communication with container 1970 via a pressurization duct 1974, and a plunger 1968 extending into hollow body 1966. As illustrated in FIGS. 19H and 19I, plunger 1968 may be actuated by a user and pressed into hollow body 1966, thus causing pressurization of the contents of container 1970, which overcome and pass valve 1980 and flow into reaction chamber 1972. Antimicrobial gas (e.g., ClO₂ gas) is allowed to escape aperture 1958 via an optional gas permeable membrane, while waste liquid is contained within reaction chamber 1972 until reactor 1900 is cleaned and recharged (fresh precursor and activator is added).

[00240] Pressurization device 1962 may be removable. Alternatively, pressurization device 1962 may be entirely separate from housing 1950 and may be applied to housing 1950 by

a user only when the user desires to activate reactor 1900. Pressurization duct 1974 may likewise include a valve 1980, which may be a check valve, backflow valve, seal, or the like.

[00241] FIGS. 20A-20D illustrate a reactor 2000 for generating an antimicrobial gas (e.g., ClO₂ gas). Reactor 2000 may be a prepackaged device loaded with a liquid precursor 902, and a solid activator 1330 within containers inside reactor 2000. Liquid precursor 902 may be sealed within its container and may require pressurized air to flow into reaction chamber 906. Solid activator 1330 may be sealed within the reaction chamber 906.

[00242] Reactor 2000 may include a housing 2040 containing liquid precursor 902, activator 1330, a reaction chamber 906, and a waste liquid container 916, wherein device elements are machined or otherwise formed out of a housing material, as in common in the production of microfluidic devices. Reactor 2000 may be a microfluidic device.

[00243] Reactor 2000 may include a pressure input 2041 capable of applying an air pressure to liquid precursor 902 to break a seal within its container and/or cause liquid precursor 902 to travel to reaction chamber 906. Pressure input 2041 may receive pressure from a pump, a syringe, or the like.

[00244] Reaction chamber 906 may include a capillary element 2042 that permits waste liquid to travel into waste liquid container 916 via capillary action. Waste liquid container 916 may include an inactivator, neutralizing agent, or the like capable of rendering waste liquid from reaction chamber 906 into a safe state.

Reaction chamber 906 may include a gas permeable membrane 2044, which allows antimicrobial gas (e.g., ClO₂ gas) created in reaction chamber 906 to pass through membrane 2044 at a controlled rate but prevents a waste liquid from reaction chamber 906 from passing through membrane 2044. Antimicrobial gas (e.g., ClO₂ gas) may exit reactor 2000 via a gas outlet 2043. Gas outlet 2043 may permit antimicrobial gas (e.g., ClO₂ gas) to exit reactor 2000 and enter the surrounding area, including for example an enclosed space (e.g., a room within a building).

[00246] As illustrated in FIG. 20D, reactor 2000 may include a cover 2046 that seals the above-referenced contents (e.g., liquid precursor 902, reaction chamber 906, and waste liquid container 916) within housing 2040.

[00247] FIG. 21A-21E illustrate a reactor 2100 for generating an antimicrobial gas (e.g., ClO₂ gas). Reactor 2100 may be a prepackaged device loaded with a liquid precursor 902 and a liquid activator 904 within containers inside reactor 2100. Reactor 2100 may include a housing 2140 containing liquid precursor 902, activator 904, a reaction chamber 906, and a waste liquid container 916, wherein device elements are machined or otherwise formed out of a housing

material, as in common in the production of microfluidic devices. Reactor 2100 may be a microfluidic device.

Liquid precursor 902 may be sealed within its container and may require pressurized air to flow into reaction chamber 906. Liquid activator 904 may be sealed within its container and may require pressurized air to flow into reaction chamber 906. Reactor 2100 may include seals 2184 within channels connecting liquid precursor 902's container and liquid activator 904's container with reaction chamber 906. Seals 2184 may include a foil, membrane, check valve, backflow valve, or the like, which may be breached with the application of an adequate pressure to permit the passage of liquid precursor 902 and/or liquid activator 904 past seals 2184.

[00249] Reactor 2100 may include a pressure input 2141 capable of receiving a fluid pressure and directing the pressure to liquid precursor 902 and activator 904 to break seals 2184 within, before, or after their respective containers and/or cause them to travel to reaction chamber 906. Pressure input 2141 may receive pressure from a fluid surrounding reactor 2100, including for example, a pressurized gas. Reactor 2100 may include seals 2184 between pressure input 2141 and liquid precursor 902's container and liquid activator 904's container.

[00250] Reaction chamber 906 may include a capillary filter (not shown) that permits waste liquid to travel into waste liquid container 916 via capillary action. Waste liquid container 916 may include an inactivator, neutralizing agent, or the like capable of rendering waste liquid from reaction chamber 906 into a safe state.

[00251] Reaction chamber 906 may include a gas permeable membrane (not shown), which allows antimicrobial gas (e.g., ClO₂ gas) created in reaction chamber 906 to pass through the membrane at a controlled rate but prevents a waste liquid from reaction chamber 906 from passing through the membrane. Antimicrobial gas (e.g., ClO₂ gas) may exit reactor 2100 via an outlet 2143. Outlet 2143 may permit antimicrobial gas (e.g., ClO₂ gas) to exit reactor 2100 and enter a dip tube tee 2182. Dip tube tee 2182 may be configured to receive the ends of an interrupted (discontinuous) dip tub as described more fully herein. Tee 2182 includes a hollow bore 2183 through which the desired contents of reaction chamber 906 (a gas, a liquid, or both) may pass and enter a dip tube. While tee 2182 is illustrated having a straight portion extending along the end of reactor 2100 with a perpendicular extending into outlet 2143, it is understood that tee 2182 could take on other shapes and configurations capable of fluidically connecting reactor 2100 to the interior of a dip tube.

[00252] As illustrated in FIGS. 21A, 21D, and 21E reactor 2100 may include a cover 2146 that seals the above-referenced contents (e.g., liquid precursor 902, activator 904, reaction chamber 906, and waste liquid container 916) within housing 2140.

[00253] FIG. 22A-22E illustrate a reactor 2200 for generating an antimicrobial gas (e.g., ClO₂ gas). Reactor 2200 may be a prepackaged device loaded with a liquid precursor 902 and a solid activator 1330 within containers inside reactor 2200. Reactor 2200 may include a housing 2240 containing liquid precursor 902, a reaction chamber 906, and a waste liquid container 916, wherein device elements are machined or otherwise formed out of a housing material, as in common in the production of microfluidic devices. Solid activator 1330 may be sealed within reaction chamber 906. Reactor 2200 may be a microfluidic device.

[00254] Liquid precursor 902 may be sealed within its container and may require pressurized air to flow into reaction chamber 906. Reactor 2200 may include seals 2284 within channels connecting liquid precursor 902's container with reaction chamber 906. Seals 2284 may include a foil, membrane, check valve, backflow valve, or the like, which may be breached with the application of an adequate pressure to permit the passage of liquid precursor 902 past seals 2284.

[00255] Reactor 2200 may include a pressure input 2241 capable of receiving a fluid pressure and directing the pressure to liquid precursor 902 to break seals 2284 within, before, or after their respective containers and/or cause them to travel to reaction chamber 906. Pressure input 2241 may receive pressure from a fluid surrounding reactor 2200, including for example, a pressurized gas. Reactor 2200 may include seals 2284 between pressure input 2241 and liquid precursor 902's container.

[00256] Reaction chamber 906 may include a capillary filter (not shown) that permits waste liquid to travel into waste liquid container 916 via capillary action. Waste liquid container 916 may include an inactivator, neutralizing agent, or the like capable of rendering waste liquid from reaction chamber 906 into a safe state.

[00257] Reaction chamber 906 may include a gas permeable membrane (not shown), which allows antimicrobial gas (e.g., ClO₂ gas) created in reaction chamber 906 to pass through the membrane at a controlled rate but prevents a waste liquid and/or solid activator 1330 from reaction chamber 906 from passing through the membrane. Antimicrobial gas (e.g., ClO₂ gas) may exit reactor 2200 via an outlet 2243. Outlet 2243 may permit antimicrobial gas (e.g., ClO₂ gas) to exit reactor 2200 and enter a dip tube tee 2282. Dip tube tee 2282 may be configured to receive the ends of an interrupted (discontinuous) dip tub as described more fully herein. Tee 2282 includes a hollow bore 2283 through which the desired contents of reaction chamber 906 (a gas, a liquid,

or both) may pass and enter a dip tube. While tee 2282 is illustrated having a straight portion extending along the end of reactor 2200 with a perpendicular extending into outlet 2243, it is understood that tee 2282 could take on other shapes and configurations capable of fluidically connecting reactor 2200 to the interior of a dip tube.

[00258] As illustrated in FIGS. 22A, 22D, and 22E reactor 2200 may include a cover 2246 that seals the above-referenced contents (e.g., liquid precursor 902, reaction chamber 906, and waste liquid container 916) within housing 2240.

[00259] Reactors 2100, 2200 may be substantially similar in function and layout to reactors 1800, 2000, with the addition of dip tube tee 2182, 2282.

[00260] FIGS. 23A-23G, and FIG. 24 illustrate an example antimicrobial gas (e.g., ClO₂ gas) generator 2300, 2400, respectively. Generator 2400 is substantially similar to generator 2300, but includes a second reagent container 2356, a second pressure generator 2366, and all associated ducts and passages to permit the second reagent to flow to a generation chamber 2374.

[00261] Antimicrobial gas (e.g., ClO₂ gas) generator 2300, 2400 may include a base 2354, at least one reagent container 2356 holding a liquid reagent 2358, and a reagent container lid 2360 with air permeable seal configured to prevent escape of liquid reagent 2358.

[00262] Within base 2354, at least one pressure chamber 2362 is oriented below at least one reagent container 2356, with at least one chamber passage 2364 in communication with pressure chamber 2362 and reagent container 2356. At least one pressure generator 2366 is oriented in communication with both pressure chamber 2362 and passage 2364, such that pressure generator 2366 can selectively block or unblock the entrance of chamber passage 2364 into pressure chamber 2362. Pressure generator 2366 is biased into a position by at least one biasing device 2368. Biasing device 2368 may be a common biasing device such as a spring. Biasing device 2368 may bias pressure generator 2366 into an open position. Biasing device 2368 may bias pressure generator 2366 into a closed position.

At least one fluid duct 2370 extends through base 2354 from pressure chamber 2362 to a microfluidic chip 2372. Microfluidic chip 2372 may include a generation chamber 2374. A generation duct 2376 may extend partly through base 2354 and partly through the wall of an off-gas and waste chamber 2378. Chamber 2378 may include an absorber material, an evaporator, or the like. Chamber 2378 may include an absorber material for absorbing spent reagent waste. Chamber 2378 may include an inactivator for waste. Chamber 2378 may include a gas permeable lid 2380 configured to allow the passage of antimicrobial gas (e.g., ClO₂ gas) out of chamber 2378. Lid 2380 may be a gas permeable membrane.

[00264] Each of the reagent container 2356, microfluidic chip 2372, and off-gas and waste chamber 2378 may be attached to and supported upon base 2354.

In operation, pressure generator 2366 begins in its closed position, as illustrated in [00265] FIGS. 23A and 23B. In this position, pressure generator 2366 seals chamber passage 2364, such that no liquid reagent 2358 may enter chamber 2362. Upon an instruction to generate antimicrobial gas (such as ClO₂ gas) (e.g., from a microcontroller such as microcontroller 306), pressure generator 2366 moves to its open position, as illustrated in FIGS. 23C and 23D. Liquid reagent 2358 passes out of reagent container 2356 and into pressure chamber 2362. Pressure chamber 2362 may be sized and shaped to permit a specific desired volume of liquid reagent 2358 to fill pressure chamber 2362 for transfer to microfluidic chip 2372. Finally, pressure generator 2366 moves back to its closed position, as illustrated in FIGS. 23E-23G, at once sealing chamber passage 2364 to prevent further introduction of liquid reagent 2358 from reagent container 2356, and pressurizing the liquid reagent within chamber 2362, so as to force the liquid reagent through the remainder of the system. Specifically, the liquid reagent is forced through fluid duct 2370, through generation chamber 2374, through generation duct 2376, and into off-gas and waste chamber 2378. Here, liquid waste 2382 is captured within chamber 2378, while antimicrobial gas (e.g., ClO₂ gas) 2384 passes through gas permeable lid 2380 and into the ambient environment.

[00266] It is understood that more than one cycle of pressure generator 2366 from its closed position, to its open position, and back to its closed position, may be required to push reagent 2358 completely through the system. Particularly, when antimicrobial gas (e.g., ClO₂ gas) generator 2300 is new, a few cycles of pressure generator 2366 may be required to begin generating antimicrobial gas (e.g., ClO₂ gas).

[00267] As illustrated in FIGS. 23A-23G, generator 2300 may include a single liquid reagent 2358 that enters generation chamber 2374 to generate an antimicrobial gas (e.g., ClO₂ gas). Thus, microfluidic chip 2372 may utilize a microfluidic electrochemical generator as described above.

[00268] Alternatively, as illustrated in FIGS. 23A-23G, generator 2300 may include a single liquid reagent 2358 comprising NaClO₂ that enters generation chamber 2374 where a solid activator is contained, thus generating an antimicrobial gas, such as ClO₂ gas.

[00269] Alternatively, as illustrated in **FIG. 24**, generator **2400** may include two separate reagent containers **2356**, with two separate pressure chambers **2362**, two separate pressure generators **2366**, and two separate fluid ducts **2370**, such that the two separate liquid reagents enter generation chamber **2374** separately where they combine and mix to generate antimicrobial gas (e.g., ClO₂ gas).

[00270] Pressure generator 2366 may be actuated via a connection to an actuator (not shown), including for example an electric motor, including an electric step motor, or the like. Pressure generator 2366 may be a plunger and may generate pressure via translation fore and aft (longitudinally).

[00271] Pressure generator 2366 may be, and translate in a direction, coaxial with chamber 2362 and fluid duct 2370. Pressure generator 2366 may be oriented at, and translate in a direction at, an angle to chamber passage 2364. In one aspect, pressure generator 2366 translates along an axis that is at a right angle (90 degrees) from the axis of chamber passage 2364.

[00272] Generator 2300, 2400 may include one or more control valves (not shown) in communication with pressure generator(s) 2366 and/or reagent container(s) 2356. Likewise, one or more control valve (not shown) may be in communication with chamber passage(s) 2364 and microfluidic chip 2372. These valves may selectively permit, prevent, or otherwise control the flow of reagents into generation chamber 2374. Generator 2300, 2400 may include a sensor system for determining the quantity, mass, volume, or the like of reagents transiting chamber passage(s) 2364.

[00273] As such, while these alternative embodiments are not illustrated, they are contemplated and as such, the figures are not intended to be limiting.

Antimicrobial Distribution Systems and Devices

[00274] FIGS. 25A-25C illustrate an example antimicrobial gas (e.g., ClO₂ gas) generator and sensor device 2500. Device 2500 may include a device housing 2504, a reservoir 2542, a nozzle 2544, a cartridge 2546, an indicator light 2548, an air sampler intake 2550, and a base 2552.

[00275] Housing 2504 may generally contain the remainder of device 2500. Reservoir 2542 may contain one or more liquid reagent for use in the generation of antimicrobial gas (e.g., ClO₂ gas). One or more reagent from reservoir 2542 may be directed into cartridge 2546, which includes the consumable elements of system 300, 400 described above. Antimicrobial gas (e.g., ClO₂ gas) is generated in cartridge 2546 and directed out of device 2500 via nozzle 2544.

[00276] Base 2552 may contain the electrical elements of device 2500, including for example, a microcontroller, one or more air pumps, sensors, and the like as described in system 300, 400. Indicator light 2548 may act to communicate information to a user regarding the state of device 2500, such as generating, low battery, connecting to a wireless network, cartridge replacement or reservoir replacement is necessary, and the like.

Base 2552 may include the air sampler intake 2550, which pulls ambient air into device 2500 for sampling to determine the antimicrobial gas (e.g., ClO₂ gas) concentration in the ambient air, as described in system 300, 400.

[00278] FIGS. 26A and 26B illustrate an example antimicrobial gas (e.g., ClO₂ gas) generator and sensor device 2600. Device 2600 may include a device housing 2604, a reservoir 2642, a nozzle 2644, a cartridge 2646, an indicator light 2648, an air sampler intake 2650, and a base 2652.

Housing 2604 may generally contain the remainder of device 2600. Reservoir 2642 may contain one or more liquid reagent for use in the generation of antimicrobial gas (e.g., ClO₂ gas). One or more reagent from reservoir 2642 may be directed into cartridge 2646, which includes the consumable elements of system 300, 400 described above. Antimicrobial gas (e.g., ClO₂ gas) is generated in cartridge 2646 and directed out of device 2600 via nozzle 2644.

[00280] Base 2652 may contain the electrical elements of device 2600, including for example, a microcontroller, one or more air pumps, sensors, and the like as described in system 300, 400. Indicator light 2648 may act to communicate information to a user regarding the state of device 2600, such as generating, low battery, connecting to a wireless network, cartridge replacement or reservoir replacement is necessary, and the like.

[00281] Base 2652 may include the air sampler intake 2650, which pulls ambient air into device 2600 for sampling to determine the antimicrobial gas (e.g., ClO₂ gas) concentration in the ambient air, as described in system 300, 400.

[00282] FIGS. 27A and 27B illustrate an example antimicrobial gas (e.g., ClO₂ gas) generator and sensor device 2700. Device 2700 may include a device housing 2704, a reservoir 2742, a nozzle 2744, a cartridge 2746, an indicator light 2748, an air sampler intake 2750, and a base 2752.

[00283] Housing 2704 may generally contain the remainder of device 2700. Reservoir 2742 may contain one or more liquid reagent for use in the generation of antimicrobial gas (e.g., ClO₂ gas). One or more reagent from reservoir 2742 may be directed into cartridge 2746, which includes the consumable elements of system 300, 400 described above. Antimicrobial gas (e.g., ClO₂ gas) is generated in cartridge 2746 and directed out of device 2700 via nozzle 2744.

Base 2752 may contain the electrical elements of device 2700, including for example, a microcontroller, one or more air pumps, sensors, and the like as described in system 300, 400. Indicator light 2748 may act to communicate information to a user regarding the state of device 2700, such as generating, low battery, connecting to a wireless network, cartridge replacement or reservoir replacement is necessary, and the like.

Base 2752 may include the air sampler intake 2750, which pulls ambient air into device 2700 for sampling to determine the antimicrobial gas (e.g., ClO₂ gas) concentration in the ambient air, as described in system 300, 400.

[00286] Any of devices 2500, 2600, 2700 may use system 300 or 400 described above for the generation and sensing of antimicrobial gas (e.g., ClO₂ gas).

[00287] FIGS. 28A and 28B illustrate an example portable antimicrobial gas reactor 2800.

[00288] Portable reactor 2800 includes: a reactor body 2802; a base 2803; a lid 2804 removably connected to one or both of reactor body 2802 or a snap on fan and pierce assembly 2806; a sidewall 2805; a fan 2808; impeller blades 2809; a power switch 2810; an optional power cord 2812; a package holder 2814; a funnel tip 2816; piercers 2818; an antimicrobial reactant package 2820; an activator package 2830; an antimicrobial liquid 2840; and antimicrobial gas 2842.

[00289] Portable reactor 2800 mixes an antimicrobial gas generating solution (via antimicrobial reactant package 2820) with an activator solution (via activator package 2830) to generate a safe level of concentration of antimicrobial gas 2842 such as chlorine dioxide (ClO₂) or hydrogen peroxide (H₂O₂) gas to disinfect localized ambient air in an indoor environment or in a limited enclosed space.

[00290] Portable reactor 2800 may be a cup formed from reactor body 2802 having a base 2803 and one or more sidewall 2805, with a funnel-shaped package holder 2814 and a pair of piercers 2818 to pierce the packages 2820, 2830 and mix the solutions to generate antimicrobial gas 2842.

[00291] The generated antimicrobial gas 2842 in reactor 2800 may be ventilated to ambient air through forced convection by an electric powered fan 2808 disposed at a cup opening, opposite base 2803. Electric powered fan 2808 may be powered through an external power source (e.g., power cord 2812), or through a built-in battery (not shown), where the battery may be rechargeable.

[00292] Reactor 2800 may have a built-in stirrer (not shown) and a wall baffle (not shown) to facilitate agitation of the mixture in the cup to facilitate completeness of chemical reactions. These elements may be included at or near base 2803 where antimicrobial liquid 2840 pools following piercing of packages 2820 and 2830.

[00293] In one aspect, previously unpierced packages 2820 and 2830 are placed inside funnel-shaped package holder 2814. Snap-on fan and pierce assembly 2806 is placed onto the top of body 2802, engaging with a lip 2807, and causing piercers 2818 to pierce packages 2820 and 2830. This piercing allows an antimicrobial reactant stream 2822 and an activator solution stream

2832 to flow downwardly (via funnel tip 2816) and form the antimicrobial liquid 2840 pool. The reactant and activator mix to form both antimicrobial liquid 2840 and antimicrobial gas 2842, which flows out of body 2802 via a pathway defined by one or more sidewall 2805, and into the ambient air. Optionally, fan 2808 via impeller blades 2809 may draw antimicrobial gas 2842 upwardly and out of reactor 2800.

[00294] The packaged antimicrobial reactant 2820 and the packaged activator solution 2830 may be individually packaged or come as a dual packages pair ready for mixing.

Portable reactor **2800** may alternatively use an anhydrous solid powder form of the antimicrobial gas generator compound, instead of an antimicrobial gas generating solution (vina antimicrobial reactant package **2820**). The antimicrobial generating compound may comprise of an anhydrous powder form a chlorite containing compound and an anhydrous powder of a chemical activator which, when exposed to water, serves as an acid or a proton donor to chemically react with the chlorite containing compound to generate chlorine dioxide (ClO₂) as the antimicrobial gas. The antimicrobial generating compound may also be an anhydrous powder of a urea hydrogen peroxide, borax, perborate, or a percarbonate compound to generate hydrogen peroxide H₂O₂ as the antimicrobial gas in the presence of water.

[00296] In another example, an anhydrous solid mixture includes an antimicrobial generating powder and an activator powder which may chemically react by addition of water, alcohol, or a solvent.

[00297] FIGS. 29A and 29B illustrate an example packaged antimicrobial gas generator solution and packaged activator solution. FIG. 29A illustrates packaged antimicrobial reactant 2820 and packaged activator solution 2830. FIG. 29B illustrates a sectional view of package 2820 taken about section A-A.

[00298] FIGS. 30A and 30B illustrate an antimicrobial gas generator 3000 in the form of a card shape or a sheet containing an antimicrobial generating compound. Antimicrobial card generator 3000 may include a cavity 3001; a second surface 3002; a lid 3004 opposite second surface 3002; a surrounding sealed wall or circumferential seam 3006; an opening 3008; a semi-permeable membrane 3010; a seal 3012; an adhesive layer 3014; and an anhydrous solid mixture 3030. Card generator 3000 may be carried in a card carrier 3020 having a strap 3016; a clip 3018; and an opening 3022.

[00299] The antimicrobial generator 3000 in various structures, may be fabricated into items comprising: a card, a badge, a face mask, a respirator, a blanket, a note pad, a deodorant card, a fragrant releasing card, a pouch, a packaging box, grocery bags, wipes, air filters, decorative items, greeting cards, bookmarks, and paper products. In another application, an

antimicrobial solution may be applied directly onto the antimicrobial gas generator or to refill the solid form of antimicrobial compound mixture for a recharge or to extend a slow release of low concentration antimicrobial gas. Antimicrobial gas generator **3000** may be used in tandem with a device to deliver antimicrobial gas into a three-dimensional space. Antimicrobial gas from antimicrobial generator **3000** or the antimicrobial generator materials may be added to water to produce an antimicrobial solution.

[00300] The antimicrobial may be a chlorine dioxide (ClO₂) or hydrogen peroxide (H₂O₂) anhydrous solid mixture, which includes: an antimicrobial generating compound and an activator that chemically reacts in presence of moisture, liquid, or solvent. The generated antimicrobial gas released from the item may be sufficient to disinfect within a close proximity of surrounding ambient air, by destroying airborne viruses, germs and when coming into contacts with bacteria or certain insects or pests.

[00301] Some configurations of antimicrobial generating compounds which may be impregnated into the absorbent materials of the multi-ply sheets structure are disclosed below

[00302] Preparation of reactant plies: starting with liquid precursors, reactant plies may be formed by absorbing antimicrobial (e.g., ClO₂) generating liquids (sodium chlorite and any form of activator) into a medium; drying medium to leave ClO₂ generating salts in medium; exposing medium to water or humidity; causing interaction with H₂O mobilizing salts/reagents; and reagents chemically interacting to form ClO₂.

[00303] Preparation of reactant plies: starting with solid precursors, reactant materials may be formed by blending solid reactants (sodium chlorite and any form of activator) into a water permeable medium; solidifying medium and reactants into a heterogeneous phase; exposing medium to water or humidity; interaction with H₂O mobilizing salts/reagents; and reagents chemically interacting to form ClO₂. A solution bath may be water, solvent, alcohol, or blends of these.

[00304] The solidification process may be physical, thermal, or chemical.

[00305] The medium can be anything that is permeable to water or water vapor (e.g., natural or synthetic fibers/papers or polymers). Alternatively, the medium may be a material that is soluble in water.

[00306] In constructing antimicrobial generator 3000, one or more plies of each component must be in contact for reaction to occur when activated. Plies may be physically, thermally, or chemically bonded. A binder may be applied separately as a tie layer, including for example, an adhesive applied via spraying, dipping, and the like. A binder may be included in one or both of

the solutions used to soak plies. A binder may be an emulsion adhesive, naturally derived like a starch, a polymer, a water absorbent polymer, and the like.

[00307] The plies should be in close proximity for reaction to occur; one on top or another or side by side. Additional plies may be added to as follows: polymer films may be incorporated into the construction to limit the rate of reaction and/or release of ClO₂; films may be between reactant plies to regulate reagent transport; films may be applied on the outside of the reagent plies to regulate ClO₂ release and/or water absorption; and other plies may be used to enhance water absorption.

[00308] Salt solutions may be used to make plies that absorb water to drive the reaction. Hydrophilic polymers may be used in a film or coated form or blends of the two. Hydrophilic fibers (natural or synthetic) may be sued to absorb or wick water. Masking plies may be incorporated to effectively reduce the surface area of reaction or release of the antimicrobial gas.

[00309] Adhesive (e.g., peel and stick) layers may be added to support application, including to permit a user to attach antimicrobial generator 3000 to a surface, a garment, or the like.

[00310] Removable non-permeable layers may be added to prevent premature generation via the sealing of reactants away from moisture or the like. Acid scavenging materials may be integrated into/between plies to inhibit premature reaction process, including for example AHTC (activated hydrotalcite).

[00311] Antioxidants, retardants, or polymers may be added to reduce flammability of antimicrobial generator 3000.

[00312] Controlling generation: the amount of antimicrobial gas (e.g., ClO₂) generated, rate, and duration of generation may be controlled by: the number of reactant plies; concentration of solutions or materials used to make reactant plies; absorption rate of materials used to make reactant plies; residence time or reactant ply materials in solution; thickness of plies; number of plies; and surface area of plies. To control water absorption: control water absorption rate of materials used to make reactant plies; adjust the number, type, and placement of plies used to enhance water absorption; and adjust the number, type, and placement of plies used to control (retard) water absorption or transport between plies. To control ClO₂ release: adjust type and thickness of materials used to regulate ClO₂ release and/or water absorption; address packaging of antimicrobial generator 3000 by ensuring plies stored in non-permeable packaging materials prior to use, desiccating materials may be used to eliminate moisture. Ply papers may change color to indicate stages of use, including for example not activated, activated, and spent.

[00313] The aforementioned concept may be used to generate other chemical solutions or reactions., including for example generating sodium chlorite as a disinfecting agent by itself.

[00314] FIG. 31 illustrates an example of an antimicrobial generator 3100 in the form of a pouch with optional addition of water internal to the pouch. Pouch generator 3100 may include: a cavity 3101; a bottom surface 3102; a top surface 3104; a surrounding sealed wall or circumferential seam 3106; an opening 3108; a semi-permeable membrane 3110; a seal 3112; an adhesive layer 3114; a spout 3124; and an anhydrous solid mixture 3130.

Pouch generator 3100 may be designed, optimized, and used in the same manner as card generator 3000. Semi-permeable membrane 3110 may be ClO₂ permeable (where the antimicrobial gas is ClO₂), or air permeable only. Seal 3112 may be a vapor barrier film that contacts adhesive layer 3114 and is removable prior to activation and use of pouch generator 3100. Spout 3124 may be closable to add water for higher ClO₂ generation, and/or may be humidity activated for lower concentration of ClO₂. Anhydrous solid mixture 3130 may be ClO₂ generating powder that is contained in a smaller pouch, a loose powder, or formed in a solid block.

[00316] FIGS. 32A illustrates an example of an antimicrobial generator 3200 in the form of a solution treated single or multi-ply porous material.

[00317] FIG. 32B illustrates an example of antimicrobial generator 3200 with liquid reactants absorbed or adsorbed on substrates and blended with a porous matrix material with optional addition of an exterior film to control release.

[00318] FIG. 32C illustrates an example of antimicrobial generator 3200 with solid reactants blended in a porous material and optional addition of an exterior film to control release.

[00319] FIG. 32D illustrates an example of antimicrobial generator 3200 in the form of a perforated pouch.

[00320] FIG. 32E illustrates an example of antimicrobial generator 3200 where reactant materials of FIGS. 32A-32C are configured side by side with optional materials to support activation and control release.

[00321] Antimicrobial generator 3200 may be designed, optimized, and used in the same manner as card generator 3000.

[00322] Antimicrobial generator 3200 may include an antimicrobial sheet 3210. Antimicrobial sheet 3210 may include a top sheet 3212, an intermediate sheet 3214, and a bottom sheet 3216. Top sheet 3212 may be paper or another absorbent material dip-coated in a sodium chlorite solution and dried. Bottom sheet 3216 may be paper or other absorbent materials dip-coated in an activator solution and dried. Intermediate sheet 3214 may act as a tie layer.

[00323] Antimicrobial generator 3200 may include an antimicrobial sheet 3220. Antimicrobial sheet 3220 may include a seal or package 3222 and an anhydrous solid mixture 3224. Anhydrous solid mixture 3224 may be formed by absorbing liquid sodium chlorite and an activator into dry powders or fibers, redry the powders or fibers, and press form the powder or fibers into a solid with an optional binder. Seal or package 3222 may be made of, or may include, optional films to control release characteristics of antimicrobial generator 3200.

[00324] Antimicrobial generator 3200 may include an anhydrous solid mixture 3230. Anhydrous solid mixture 3230 may include a seal or package 3223 and an antimicrobial compound and activator mixture 3234. Anhydrous solid mixture 3230 may be formed by pressing sodium chlorite and citric acid into a matrix of synthetic or natural fibers (LLDPE, paper, or the like).

[00325] Antimicrobial generator 3200 may include a pouch 3240. Pouch 3240 may include a seal 3242, an anhydrous solid mixture 3244, an adhesive layer 3246, and an opening 3248. Anhydrous solid mixture 3244 may include LLDPE film to hold a powder in a desired shape or configuration.

Antimicrobial generator 3200 may include an antimicrobial sheet 3250. Antimicrobial sheet 3250 may include a top sheet 3252, a control release film or coating 3254, a first anhydrous solid mixture 3256, and a second anhydrous solid mixture 3258. Top sheet 3252 may include a wicking or water absorbent material configured to activate first and second anhydrous solid mixtures 3256, 3258 when top sheet 3252 is exposed to moisture/water. Control release film or coating 3254 may be optional and added to one or more sides to control release characteristics of generator 3200. First anhydrous solid mixture 3256 may include sodium chlorite made pursuant to the method and configuration described with respect to FIGS. 32A-32C. Second anhydrous solid mixture 3258 may include an activator made pursuant to the method and configuration described with respect to FIGS. 32A-32C.

[00327] FIGS. 33A-33D illustrate an aerosol container 3386. It is understood that while container 3386 is illustrated in a cutaway manner in FIGS. 33A-33D, container 3386 is an enclosed container, and may be similar to a common aerosol can, such as a spray paint can, with an interior defined by a wall and an optional liner for chemical isolation. Container 3386 may include a nozzle 3388 and a hollow dip tube 3390. Dip tube 3390 may be oriented within the interior of container 3386. Dip tube 3390 may include a distal end 3391 open to the interior of container 3386. Dip tube 3390 as illustrated in FIG. 33A is interrupted (discontinuous) with a cutout portion permitting attachment to a dip tube tee 2182, 2282 of a reactor 2100, 2200. Dip tube 3390 may include a proximal end fluidically connected to nozzle 3388, such that contents of

container **3386** may be directed through the interior of dip tube **3390** and out nozzle **3388** and into the surrounding environment.

[00328] As dip tube 3390 is discontinuous, dip tube 3390 includes a proximal (upper, as illustrated) portion and a distal (lower, as illustrated) portion, with two ends abutting the interrupted portion of dip tube 3390. These ends of dip tube 3390 are inserted into opposing ends of dip tube tee 2182, 2282, such that the interior of reactor 2100, 2200 is fluidically connected to the interior of dip tube 3390.

[00329] FIG. 33C illustrates an arrangement wherein reactor 2100 may be fluidically connected to the interior of dip tube 3390, while FIG. 3D illustrates an arrangement wherein reactor 2200 may be fluidically connected to the interior of dip tube 3390.

[00330] Reactor 2100, 2200 may generate an antimicrobial gas (e.g., ClO₂ gas), which exits outlet 2143, 2243 and enters dip tube tee 2182, 2282. The antimicrobial gas may be drawn into dip tube 3390 and exit container 3386 via nozzle 3388. Container 3386 may include a pressurized propellant gas contained within the interior of container 3386 outside of reactor 2100, 2200. The antimicrobial gas may be mixed with a carrier fluid or carrier gas that is contained within the interior of container 3386 outside of reactor 2100, 2200. The carrier fluid/gas may enter dip tube 3390 via distal end 3391. The carrier fluid/gas may carry antimicrobial gas out of nozzle 3388 and into the surrounding environment. The carrier fluid/gas may carry antimicrobial gas out of nozzle 3388 using pressure provided by the pressurized propellant gas. In one aspect, container 3386 may contain a pressurized propellant gas and a carrier liquid. In another aspect, container 3386 may contain a pressurized carrier gas and no pressurized propellant gas.

The propellant gas and/or carrier fluid/gas may be pre-pressurized at a pressure above atmospheric pressure outside of container 3386, and thus may flow (or the propellant gas may cause the carrier fluid/gas to flow) into dip tube 3390 via distal end 3391 upon opening of nozzle 3388. The carrier fluid/gas may draw antimicrobial gas into dip tube 3390 via outlet 2143, 2243 by way of a venturi effect due to reactor 2100, 2200 being open to the interior pressure of container 3386 at pressure input 2141, 2241. With reference to reactor 2100, this venturi effect may create a negative pressure at outlet 2143 drawing liquid precursor 902 and liquid activator 904 in reactor 2100 into reaction chamber 906, where liquid precursor 902 and liquid activator 904 react to create antimicrobial gas, which is then drawn into outlet 2143. With reference to reactor 2200, the venturi effect may create a negative pressure at outlet 2243, drawing liquid precursor 902 into reaction chamber 906 where it reacts with solid activator 1330 to create ClO₂ gas, which is then drawn into outlet 2243.

Nozzle 3388 may include any of a variety of valves, including a spring-loaded valve that may be manually opened by a user (e.g., by pressing down on the nozzle, pulling a trigger, and the like), and which returns to a closed position upon release by a user. Nozzle 3388 may also be automated to open and/or close by an actuator. In this manner, only a desired amount of the contents of container 3386 are discharged into the environment as the valve is selectively opened and closed. Nozzle 3388 may be manipulated by a user, actuator, or the like, and locked into place, such that when opened the contents of container 3386 are discharged into the surrounding environment until the internal pressure of container 3386 is equal to the pressure of the surrounding environment (e.g., atmospheric pressure).

[00333] Any waste liquids created in the generation of antimicrobial gas (e.g., ClO₂ gas) may be maintained within reactor 2100, 2200. Seals 2184, 2284 may be designed so as to prevent liquid precursor 902, liquid activator 904, and/or waste liquids created in reaction chamber 906 from flowing backward (that is, away from outlet 2143, 2243) and thus maintains these liquids within reactor 2100, 2200.

While reactors 2100, 2200 are described as generating antimicrobial gas (e.g., ClO₂ gas), it is understood that reactors 2100, 2200 may be used to generate any multi-component liquid or gas by simply altering the precursor and/or activator contained within reactors 2100, 2200. Reactors 2100, 2200 may be used to produce any multi-component liquid or gas from materials that may react or otherwise be incompatible (immediately or over time) when mixed, and thus cannot be mixed during the initial packaging of container 3386. For example, container 3386 may be used with two-component paints or adhesives, activated hydrogen peroxide products, activated peracetic acid products, and the like.

[00335] In one alternative aspect, reactor 2100, 2200 does not contain any activator, and alternatively, a solid activator is contained within dip tube 3390 in a proximal (upper) portion of drip tube 3390, between reactor 2100, 2200 and nozzle 3388.

[00336] Any of reactors 1800, 1900, and 2000 may be adapted for use in place of reactors 2100, 2200 in aerosol can 3386.

[0001] FIG. 34 illustrates an aerosol container 3486 including a flexible bladder 3492 connected to a dip tube 3490. It is understood that while container 3486 is illustrated in a cutaway manner in FIG. 34, container 3486 is an enclosed container, and may be similar to a common aerosol can, such as a spray paint can, with an interior defined by a wall and an optional liner. Container 3486 may include a nozzle 3488 and a hollow dip tube 3490. Dip tube 3490 may be oriented within the interior of container 3486. Dip tube 3490 may include a distal end 3491 open to the interior of container 3486. Dip tube 3490 may include a proximal end fluidically connected

to nozzle **3488**, such that contents of container **3486** may be directed through the interior of dip tube **3490** and out nozzle **3488** and into the surrounding environment.

[0002] A flexible bladder 3492 is contained within container 3486, and fluidically connected to dip tube 3490 by a fitting 3493. That is, the interior of flexible bladder 3492 is fluidically connected to the interior of dip tube 3490, such that the contents of flexible bladder 3492 may be conveyed into the interior of dip tube 3490.

[0003] Flexible bladder 3492 may be made of any of a variety of materials, including for example a rubber. Flexible bladder 3492 may contain within its interior a secondary liquid reactant, including for example a precursor, an activator, or the like. Additionally, a primary liquid reactant that is different from the liquid reactant within flexible bladder 3492 may be contained within the interior of container 3486, but outside of flexible bladder 3492. Additionally, a pressurized propellant gas may be contained within container 3486.

In practice, the pressure of the secondary liquid reactant within flexible bladder 3492 may be maintained at the same pressure as the primary liquid reactant and/or pressurized propellant gas (if present). In this state, an equilibrium exists within container 3486 that keeps the secondary reactant within flexible bladder 3492. However, a check valve, backflow valve, seal, or the like (not shown) may be fluidically connected to the interior of flexible bladder 3492 (e.g., within fitting 3493) that helps keep secondary liquid within flexible bladder 3492 until a user operates nozzle 3488.

Nozzle 3488 may include any of a variety of nozzles, including a spring-loaded valve that may be manually opened by a user (e.g., by pressing down on the nozzle, pulling a trigger, and the like), and which returns to a closed position upon release by a user. Nozzle 3488 may also be automated to open and/or close by an actuator. In this manner, only a desired amount of the contents of container 3486 are discharged into the environment. Nozzle 3488 may be manipulated by a user, actuator, or the like, and locked into place, such that when opened the contents of container 3486 are discharged into the surrounding environment until the internal pressure of container 3486 is equal to the pressure of the surrounding environment (e.g., atmospheric pressure).

[0006] When nozzle 3488 is opened, primary liquid reactant travels into distal end 3491 of dip tube 3490, and out nozzle 3488. This in turn causes the pressure of primary liquid reactant within container 3486 to drop, which in turn causes secondary liquid reactant within flexible bladder 3492 to overcome a seal, backflow valve, or the like (referenced above) and flow from flexible bladder 3492, through fitting 3493, into dip tube 3490, and out nozzle 3488. A backflow valve fluidically connected to bladder 3492 may prevent the primary liquid reactant from entering

flexible bladder **3492** and keep the secondary liquid reactant traveling in the direction of nozzle **3488**. The primary liquid precursor and the secondary liquid precursor may mix and react within dip tube **3490**, thus directing the product of the reaction out of nozzle **3488**.

[0007] In one aspect, the product is antimicrobial gas, such as ClO₂ gas. However, it is understood that this arrangement may be used to generate any multi-component liquid or gas by simply altering the primary and second liquid reactants. The product may be any multi-component liquid or gas formed from materials that may react or otherwise be incompatible (immediately or over time) when mixed, and thus cannot be mixed during the initial packaging of container 3486. For example, container 3486 may be used with two-component paints or adhesives, activated hydrogen peroxide products, activated peracetic acid products, and the like.

[0008] While only one flexible bladder 3492 is illustrated, it is contemplated that more than one flexible bladder 3492 may be used where additional reactants are required for the desired product.

[0009] It is contemplated that where a solid activator is desired, the solid activator may be oriented within dip tube **3490** so as to react with one or both of the primary and secondary liquid reactants on the way out of dip tube **3490** and nozzle **3488**.

[0010] The volume and pressure of the flexible bladder 3492 and container 3486, diameter of fitting 3493 and dip tube 3490, liquid reactant viscosities, and/or the use of restrictor valves may be adjusted and optimized to create the desired product from the reactants, at the desired rate of discharge from nozzle 3488.

[0011] FIGS. 35A and 35B illustrate an aerosol container 3586 including a plurality of flexible bladders 3596. It is understood that while container 3586 is illustrated in a cutaway manner in FIGS. 35A and 35B, container 3586 is an enclosed container, and may be similar to a common aerosol can, such as a spray paint can, with an interior defined by a wall. Container 3586 may include a nozzle 3588 and a hollow mixing tube 3595. Mixing tube 3595 may be oriented within the interior of container 3586. Mixing tube 3595 may include a tee with distal ends fluidically connected to the interior of flexible bladders 3594. Mixing tube 3595 may include a proximal end fluidically connected to nozzle 3588, such that contents of flexible bladders 3594 may be directed through the interior 3596 of mixing tube 3595 and out nozzle 3588 and into the surrounding environment.

[0012] In practice, the flexible bladders 3594 contain liquid reactants necessary for the generation of a desired product. For example, a first flexible bladder 3594 may contain a primary liquid reactant while a second flexible bladder 3594 may contain a secondary liquid reactant. Each flexible bladder 3594 is pressurized to a pressure greater than the atmospheric pressure outside of

container **3586**. When nozzle **3588** is opened, the pressure within flexible bladders **3594** forces the contents of flexible bladders **3594** into the interior **3596** of mixing tube **3595** where the primary and secondary liquid reactants mix and react to create a product, which is directed out nozzle **3588**.

Nozzle 3588 may include any of a variety of nozzles, including a spring-loaded valve that may be manually opened by a user (e.g., by pressing down on the nozzle, pulling a trigger, and the like), and which returns to a closed position upon release by a user. Nozzle 3588 may also be automated to open and/or close by an actuator. In this manner, only a desired amount of the contents of container 3586 are discharged into the environment. Nozzle 3588 may be manipulated by a user, actuator, or the like, and locked into place, such that when opened the contents of container 3586 are discharged into the surrounding environment until the internal pressure of container 3586 is equal to the pressure of the surrounding environment (e.g., atmospheric pressure).

[0014] In one aspect, the product is an antimicrobial gas, such as ClO₂ gas. However, it is understood that this arrangement may be used to generate any multi-component liquid or gas by simply altering the primary and second liquid reactants. The product may be any multi-component liquid or gas formed from materials that may react or otherwise be incompatible (immediately or over time) when mixed, and thus cannot be mixed during the initial packaging of container 3586. For example, container 3586 may be used with two-component paints or adhesives, activated hydrogen peroxide products, activated peracetic acid products, and the like.

[0015] While only two flexible bladders 3594 are illustrated, it is contemplated that more than one flexible bladder 3594 may be used where additional reactants are required for the desired product.

[0016] It is contemplated that where a solid activator is desired, the solid activator may be oriented within mixing tube **3595** so as to react with one or both of the primary and secondary liquid reactants on the way through mixing tube **3595** and nozzle **3588**.

[0017] The volume and pressure of flexible bladders 3594 and container 3586, diameter of mixing tub 3595, liquid reactant viscosities, and/or the use of restrictor valves may be adjusted and optimized to create the desired product from the reactants, at the desired rate of discharge from nozzle 3588.

[0018] In one aspect, nozzle 3588 may include a shroud or baffle (not shown) upon which the liquid product exiting nozzle 3588 impinges, causing liquid to fall into a containment reservoir (not shown), while pure gas product is permitted to continue out of nozzle 3588 and into the surrounding environment. In this manner, liquid product can be captured and retained while gas

product is permitted to be dispensed. The containment reservoir may be within container **3586**, outside container **3586**, or in one possible arrangement, between an inner container **3586** and an outer container (not shown) within which container **3586** is contained.

[00337] Reactors 1800, 1900, 2000, 2100, 2200 described herein may be small and portable and may create an antimicrobial gas (e.g., ClO₂ gas) rapidly for convenient use to disinfect threedimensional spaces. For example, reactors 1800, 1900, 2000, 2100, and/or 2200 may be approximately 1.0 in. (2.54 cm) wide and 3.0 in. (7.62 cm) long. Reactor 1800, 1900, 2000, 2100, and/or 2200 may be reusable/rechargeable. Reactor 1800, 1900, 2000, 2100, and/or 2200 may be sized to create an amount of antimicrobial gas (e.g., ClO₂ gas) optimal for disinfecting a threedimensional space (e.g., a room) of a particular size or range of sizes, such that reactor 1800, 1900, 2000, 2100, and/or 2200 may be larger or smaller for larger or smaller spaces. Alternatively, use of reactor 1800, 1900, 2000, 2100, and/or 2200 in spaces larger than intended may require the use of more than one reactor 1800, 1900, 2000, 2100, and/or 2200. Additionally, aerosol containers 3386, 3486, and/or 3586 may be sized, shaped, and designed for treatment of three-dimensional spaces of a particular size or range of sizes. Aerosol containers 3386, 3486, and/or 3586 may be designed to be recharged by accessing the interior of aerosol containers 3386, 3486, and/or 3586 and replacing, reusing, or recharging the reactor. Alternatively, aerosol containers 3386, 3486, and/or 3586 may be designed for a one-time use.

FIG. 36 illustrates an apparatus 3600 for generating antimicrobial gas or vapor external to a sealed environment for disinfecting items therein. FIG. 37 illustrates a system 3700 generating antimicrobial gas or vapor external to a sealed environment for disinfecting items therein. FIGS. 38A and 38B illustrate a system 3800 generating antimicrobial gas or vapor within a sealed environment for disinfecting items in the sealed environment. FIGS. 39A-39C illustrate an apparatus 3900 generating antimicrobial vapor within a sealed environment for disinfecting items therein. FIG. 40 illustrates methods of generating antimicrobial gas or vapor within a sealed environment or external to the sealed environment to disinfect items within the sealed environment. FIGS. 41A and 41B illustrate ClO₂ efficacy test data on controlled samples.

[00339] As used herein, the term "items" may include both personal protective equipment ("PPE") and non-PPE items, such as personal items, garments, medical equipment, apparel, garments, shoes, personal electronic devices, furniture, office supplies, built-in structures, drapes, fabrics, utensils, fixtures, decorative items, food, and plants. In addition, the term "sealed environment" may include any of: a sealable bag, a tent, a storage container, a drum, a tumbler drum, a chamber, a room, an office, a store, a warehouse, a home, a hospital, a floor of a multi-

level building, a cabin, an aircraft cabin, a vehicle cabin, a shipping container, a surface vessel cabin, an underwater vessel cabin, public transportation vehicles.

[00340] Apparatuses/systems 3600, 3700, 3800, and 3900 utilize an antimicrobial gas, such as chlorine dioxide (ClO₂) gas, for disinfecting PPE including, without limitation, N95 respirators, surgical masks, protective suits, goggles, and helmets, making them safe for reuse by healthcare professionals and patients, as well as personal items and facilities in office and home settings. In addition, the disclosed methods and apparatuses may be applicable to general decontamination of contained spaces and items.

[00341] The technology has been shown effective against an Ebola surrogate on common hard surface and porous household materials and as a broad-spectrum chemical and biological decontaminant for sensitive equipment. The feasibility of disinfecting and reusing N95 masks has previously been demonstrated using hydrogen peroxide gas or vapor, but the need for specialized (and expensive) equipment requires moving used/contaminated N95 masks to a single location for treatment.

With the proposed approach, used N95s may be placed in a sealable chamber [00342] (FIGS. 8 and 39A-39C) and exposed to the headspace of an antimicrobial gas (e.g., ClO₂ solution, generated on-site and at the time of use). After a short antimicrobial gas generation period, dissolved antimicrobial gas is off-gassed within the chamber, allowing the antimicrobial gas to penetrate and disinfect the respirators. After a sufficient disinfection period, the liquid and gas disinfection solution (e.g., ClO₂) are neutralized before opening the chamber to retrieve the disinfected N95s. Neutralization may be performed by adding a small quantity of neutralizing agent, such as a non-hazardous dry chemical packaged with the kit. The spent disinfection solution and any packaging materials are then disposed as non-hazardous waste. The system design is very scalable, from a single item construct for small batches (approx. 1 - 20 respirators) to a room-size chambers or dedicated rooms for large batches (hundreds or thousands of N95s) for use at treatment facilities, forward operating bases, or hospitals to treat large numbers of N95 masks and other equipment. The method requires no electricity, and the decontamination kit (including the reactive ingredients and a container, such as a plastic bag) can be easily transported with other field equipment. Optionally, gas dispersion units ("GDU") within large room-size chambers for dedicated rooms for large batches may include fans or blowers to accelerate the liberation of ClO₂ while forcing ClO₂ out into the enclosure for faster and more uniform distribution. The GDU may require very little power (e.g., may be operated with a battery).

Preliminary efficacy testing (FIGS. 41A and 41B) had been conducted by contaminating nine coupons cut from an N95 with 7.7-logs of Phi6 bacteriophage (surrogate for Coronavirus and Ebola) prepared in an organic test soil. Coupons were exposed to ClO₂ gas generated from 6 liters of a 180 ppm ClO₂ solution in an 82-L container. A small fan was directed across the surface of the ClO₂ solution to aid in off-gassing and mixing. Coupons were removed after 1.50, 2.25, and 3.00 hours; the exposure times correspond to three treatment levels: 1500, 2200, and 2800 ppm-hours, respectively. Control coupons were held under ambient conditions for the duration of the experiment (3 hours). After treatment, the coupons were assayed; for all three treatment levels, no virions were recovered from the coupons treated with ClO₂ gas. For all three treatment levels, a 6-log reduction of virons was observed. FIGS. 41A and 41B show the recovered virions and log reduction observed for each treatment level. The limit of detection (LOD) for this assay was 1.7-logs. Based on this initial test, ClO₂ gas was effective against the Phi6 surrogate and 6-log reduction was achieved in less than 90 minutes.

FIG. 36 is a schematic diagram of an apparatus 3600 generating antimicrobial gas or vapor 3636 external to a sealed environment 3601 for disinfecting items 3608 (e.g., PPE, clothing, and the like) therein. Apparatus 3600 may include a gas or vapor generator 3630 coupled to a blower or pump 3603 which provides antimicrobial gas or vapor 3636 to disinfect items 3608 contained within sealed environment 3601. Gas or vapor generator 3630 may be a tank in which a chemical reaction may take place by reacting a chlorite containing compound (sodium chlorite NaClO₂) with a proton donor 3632 (e.g., a mild acid solution such as oxalic acid or citric acid) to generate antimicrobial gas or vapor 3636, such as chlorine dioxide ClO₂. The chlorite containing compound may be a permeable membrane or a sachet 3634 containing sodium chlorite NaClO₂ powder.

Antimicrobial vapor 3636 may be externally pumped into sealed environment 3601 containing items 3608 which are to be disinfected. As shown in FIG. 36, sealed environment 3601 may be a tumbling drum that turns by a motor 3604 through belts or pulleys 3607, to rotate the tumbling drum about a central axis to ensure uniformity in mixing and tumbling items 3608 inside. The tumbling drum may be similar to a dryer having inlet 3611 (first passage) and an exhaust outlet 3610 (second passage), such that antimicrobial vapor 3636 may be recirculated back to the vapor generator 3630. Items 3608 for decontamination may include one or more of: healthcare PPE, respirators, surgical masks, helmets, medical gloves, medical gowns, protective suits, goggles, shoe, and the like. Disinfecting of items 3608 contained in sealed environment

3601 may achieve destruction of one or more of: microbial organisms, bacteria, viruses, fungi, pests, toxins, germs, mites, bed bugs, and the like.

[00346] FIG. 37 is another schematic diagram of a system 3700 including an apparatus 3710 generating antimicrobial gas or vapor external to a sealed environment 3750 for disinfecting items therein. System 3700 may include a gas or vapor generator apparatus 3710 coupled to a heating ventilation and air conditioning ("HVAC") system or a humidifier system to pass humidified disinfecting gas or vapor (e.g., ClO₂ gas or vapor) from a concentrated chlorine dioxide solution 3702, and the humidified disinfecting gas or vapor 3730 (e.g., ClO₂ gas or vapor) may be further evaporated through a heater 3720 that may be recirculated within the sealed environment for disinfecting items therein.

System 3700 may additionally include a pump 3704 for pumping ClO₂ solution 3702 to generator apparatus 3710. A water line 3706 may provide water to generator apparatus 3710. A generator controller 3708 may act to control, permit user input into, or both, generator apparatus 3710. A ClO₂ sensor 3734 may be oriented within sealed environment 3750 and may be in communication with (wired or wireless) a process controller 3736. Process controller 3736 may ultimately control all antimicrobial gas or vapor generation of system 3700, including receiving data from sensor 3734 regarding the concentration of disinfecting gas or vapor within sealed environment 3750. Process controller 3736 may cause the generation of more or less disinfecting gas or vapor 3730 to achieve a desired antimicrobial gas or vapor concentration, based upon data received from sensor 3734.

[00348] The items for disinfection may include one or more of: healthcare personal protective equipment ("PPE"), medical equipment, apparel, garments, shoes, personal electronic devices, furniture, office supplies, built-in structures, drapes, fabrics, utensils, fixtures, decorative items, plants, and packaged or unpackaged food.

[00349] Sealed environment 3750 may be any of: a sealable bag, a tent, a container, a drum, a tumbler drum, a chamber, a room, an office, a store, a warehouse, a home, a floor of a multi-level building, a cabin, an aircraft cabin, a vehicle cabin, a surface vessel cabin, an underwater vessel cabin. Disinfecting the items contained in sealed environment 3750 may achieve destruction of one or more of: microbial organisms, bacteria, viruses, fungi, pests, toxins, germs, mites, bed bugs, and the like.

[00350] FIGS. 38A and 38B illustrate a system 3800 for generating antimicrobial gas or vapor within a sealed environment for disinfecting items in the sealed environment. System 3800 may include apparatuses 3810 and 3860 for generating antimicrobial vapor within a sealed environment for disinfecting items in the sealed environment (3820, 3830, 3840, 3850).

In FIG. 38A, system 3800 includes an apparatus 3810 (e.g., shop vacuum) used to evacuate 3812 (via a suction side 3813) a sealed environment 3820 (e.g., large bag) through a single passage 3822. After the evacuation, sealed environment 3820 may be back filled 3814 (via an exhaust side 3815) with antimicrobial gas or vapor through single passage 3822 for disinfecting items 3824 therein.

In FIG. 38B, apparatus 3860 may be used to evacuate (via a suction side 3813) and a plurality of sealed environments 3830, 3840, 3850, and backfill (via an exhaust side 3815) the plurality of sealed environments 3830, 3840, 3850 with antimicrobial gas or vapor through respective passages 3832, 3842, 3852 for disinfecting items therein. Alternatively, as illustrated in FIG. 38B, apparatus 3860 may be used as a gas or vapor generator within a sealed environment 3870. Sealed environment 3870 may be anyone of: a sealable bag, a tent, a container, a drum, a tumbler drum, a chamber, a room, an office, a store, a warehouse, a home, a floor of a multi-level building, a cabin, an aircraft cabin, a vehicle cabin, a surface vessel cabin, an underwater vessel cabin.

[00353] FIGS. 39A-39C illustrate a system 3900 apparatus 3930 for generating antimicrobial gas or vapor within a sealed environment 3910 for disinfecting items 3920 therein. FIG. 39A shows that items 3920 to be disinfected may first be put into a sealed environment 3910 (e.g., a bag). Items 3920 may include PPE, medical equipment, apparel, garments, shoes, personal electronic devices, goggles, helmets, drapes, fabrics, utensils, decorative items, and the like. FIG. 39B shows that antimicrobial gas or vapor (e.g., ClO₂) may be generated using a gas or vapor generator 3930. Gas or vapor generator 3930 may be a cup, holder, or a container having a lid to keep the content from spilling out, where antimicrobial gas or vapor (e.g., ClO₂) may be generated by mixing reactants (e.g., a sodium chlorite package 3932 mixed with a mild acid 3934) and placing gas or vapor generator 3930 inside sealed environment 3910. FIG. 39C shows that the items 3920 inside the sealed environment 3910 may be disinfected by the antimicrobial gas or vapor (e.g., ClO₂) 3934 after a defined time period.

[00354] FIGS. 8 and 39A-39C illustrate example systems 800 and 3900 for applying an antimicrobial to a sealed environment 810 and 3910, respectively. Gas or vapor generator 830 and 3930 may be placed within sealed environment 810 and 3910, respectively, to treat items 3920 (not shown in FIG. 8). Neutralization may be performed by adding a small quantity of neutralizing agent, such as a non-hazardous dry chemical packaged with the kit. The spent disinfection solution and any packaging materials are then disposed as non-hazardous waste. The system design is very scalable, from a single item construct for small batches (approx. 1 - 20

respirators) to a room-size chambers or dedicated rooms for large batches (hundreds or thousands of N95s) for use at treatment facilities, forward operating bases, or hospitals to treat large numbers of N95 masks and other equipment. The method requires no electricity, and the decontamination kit (including the reactive ingredients and a container, such as a plastic bag) can be easily transported with other field equipment.

[00355] FIG. 40 illustrates a method 4000 for generating antimicrobial gas or vapor within a sealed environment or external to the sealed environment to disinfect items within the sealed Method 4000 includes: providing an antimicrobial vapor to an enclosed environment. environment (step 4002); generating the antimicrobial vapor within the enclosed environment through one of: (step 4004) (a) pumping a controlled concentration level of liquid chlorine dioxide solution into a humidifier system to produce a stream of chlorine dioxide antimicrobial vapor, (b) directly pumping a controlled concentration level of chlorine dioxide antimicrobial vapor to an ambient of the enclosed environment, (c) generating ambient chlorine dioxide antimicrobial vapor by dissolving solid chlorine dioxide reactant reagent into a container filled with water; and placing the container that releases the ambient chlorine dioxide antimicrobial vapor into the enclosed environment, (step 4006); generating the antimicrobial vapor external to the enclosed environment through one of: (step 4008) (a) pumping a controlled concentration level of liquid chlorine dioxide solution into a humidifier system to produce a stream of chlorine dioxide antimicrobial vapor, (b) directly pumping a controlled concentration level of liquid chlorine dioxide antimicrobial vapor to ingress the enclosed environment through a single passage of the enclosed environment, and (c) directly pumping a controlled concentration level of chlorine dioxide antimicrobial vapor to ingress the enclosed environment through a first passage of the enclosed environment (step 4010).

[00356] As illustrated, method 4000 optionally performs step 4002 followed by steps 4004 and 4006, or performs step 4002 followed by steps 4008 and 4010.

[00357] FIG. 42 illustrates an apparatus 4200 that generates antimicrobial gas or vapor for disinfecting items in three-dimensional space. FIG. 43 illustrates an apparatus 4300 that generates antimicrobial gas or vapor for disinfecting items in three-dimensional space. FIG. 44 illustrates a procedure 4400 for the use of apparatus 4300 in FIG. 43 to generate antimicrobial gas. FIG. 45 illustrates a table showing temperature effects to solubility of ClO₂ gas in water and in air and required amount of ClO₂ gas for a defined room size. FIG. 46 illustrates a uniformity of ClO₂ gas concentration distributed within a room. FIG. 47 illustrates gas concentration profiles in room setting with furniture. FIG. 48 illustrates relative humidity and generated ClO₂ gas concentration from a ClO₂ solution. FIG. 49 illustrates a correlation of increase in disinfection efficacy with

elevated humidity. **FIG. 50** illustrates a method **5000** for generating an antimicrobial gas and dispersing the gas via an apparatus.

[00358] FIGS. 42 and 50 illustrate an example of a mobile apparatus 4200 performing a computer implemented method 5000 to generate and disperse an antimicrobial gas 4220 to a defined volume of space. Method 5000 includes performing the following steps: measuring a volume of space to be disinfected (step 5002); setting by a controller 4231, disinfection parameters based on the measured volume of space 4250 to be disinfected (step 5004), wherein the disinfection parameters comprising at least the following: (a) determining an optimal location of the apparatus 4200 in the volume of space for uniform dispersion of the antimicrobial gas 4220, (b) a time duration of disinfection cycle, (c) a minimum amount of antimicrobial gas required to be generated, (d) a flow rate of antimicrobial gas generation, a volume of antimicrobial solution, and a concentration of antimicrobial solution 4205 to meet the required flow rate of antimicrobial gas 4220, (e) a range of antimicrobial gas relative humidity to be used during a disinfection cycle (step 5006). Afterwards, activating the apparatus 4200 to run the disinfection cycle until completion; discharging through a plurality of nozzles 4210 which are mounted on an oscillating head 4212, the antimicrobial gas 4220 to volume of space 4250.

[00359] Method 5000 may further include: monitoring periodically, a reading of antimicrobial gas concentration at a plurality of remote locations (by a plurality of remote sensors 4242-4248) within volume of space 4250 during the disinfection cycle and adjusting one or more of: the antimicrobial gas flow rate and the antimicrobial gas concentration for uniform antimicrobial gas dispersion in volume of space 4250 (step 5010).

[00360] Measuring of the volume of space may be performed by an integrated on-board laser beam scanner 4214. The method may include oscillating along an axis, the plurality of nozzles 4210 mounted on the oscillating head 4212 in a full circle or less than a half circle. The method may include: in response to the monitored reading of the antimicrobial gas concentration at each of the plurality of locations 4242-4248, configuring one or more respective nozzles 4210 mounted on the oscillating head 4212 to perform one or a combination of the following to offset concentration differences of the antimicrobial gas at the plurality of locations: adjusting a vertical angle of the nozzle, adjusting a discharge flow rate of the antimicrobial gas, and adjusting a discharge pressure of the antimicrobial.

[00361] In response to the monitored reading of the antimicrobial gas concentration at each of the plurality of locations 4242-4248, the method may include varying a fan speed of a first blower 4206 which sucks the antimicrobial gas 4208 released from an antimicrobial solution contained in a reactor 4204. The antimicrobial gas may be released from the antimicrobial

solution in vapor phase at the range of relative humidity (RH) according the setting of the controller 4231, wherein the range of relative humidity of the vapor phase is correlated to a temperature of the antimicrobial solution 4205.

[00362] The antimicrobial gas or vapor 4220 may be one of: chlorine dioxide (ClO_2) gas or vapor and hydrogen peroxide (H_2O_2) gas or vapor. The ClO_2 gas or vapor may be generated by chemically reacting a chlorite containing compound with an activator and the H_2O_2 gas or vapor is generated by chemically reacting a urea hydrogen peroxide, borax, perborate, or percarbonate compound with the activator, wherein the activator includes an acid or a proton donating solvent. The chlorite or peroxide containing compound and the activator are separately packaged as anhydrous powder or separately packaged as concentrated solution packages which are to be mixed together in the reactor 4206 to form the antimicrobial solution 4205.

[00363] Upon completion of the disinfection cycle, an aeration cycle may be started for a defined duration of time to adsorb ambient antimicrobial gas in the volume of space. Alternately, the aeration may also take place during the antimicrobial dispersing cycle to facilitate homogeneity of the antimicrobial gas 4220 in ambient. More specifically, the aeration cycle may be performed by drawing and recirculating by a second blower 4207, ambient air through a carbon/HEPA filter 4224 disposed at an inlet 4209 of the apparatus, and venting filtered air at an outlet 4211 of the apparatus, wherein the second blower 4207 is physically disposed below and away from the first blower, such that the antimicrobial gas or vapor in the ambient is adsorbed by the carbon/HEPA filter 4224.

The mobile apparatus **4200** may be mounted on wheels **4228** to provide mobility. The method may include sending a warning signal from the mobile apparatus in a situation including one or a combination of: (1) when the antimicrobial gas or vapor **4250** in the ambient air exceeds a defined unsafe level, (2) malfunctioning of either the first blower **4206** or the second blower **4207**, or (3) depletion of antimicrobial solution **4205** in the reactor **4206**. The warning signal may be visual, audible, transmitted wirelessly to a remote device (e.g., phone), or any combination thereof.

[00365] FIGS. 43 and 44 illustrate an example of a mobile apparatus 4300 performing a computer implemented method 4400 to generate and disperse an antimicrobial gas or vapor 4310 for disinfecting items in three-dimensional space. Apparatus 4300 may include a humidifier unit 4302, a main unit 4304, a support element 4306, a filtered air outlet 4308, a carbon filter intake 4312, main unit fans 4314, a gas sensor 4316, a removable remote and data readout tablet 4318, an oscillating tower fan 4322 with fan outlet 4320, under cart storage and portable battery

placement area **4324**, and a cycle indicator light **4326**. Apparatus **4300** may be mobile and placed upon wheels for easy transport into and from a three-dimensional space to be disinfected.

Method 4400 may include turning a cap 4436 of a concentrated solution bottle 4432, 4434 to "prime" and let sit for at 2 hours; place bottles 4432, 4434 into container unit 4402 (of humidifier unit 4302); turn caps 4436 of bottles 4432, 4434 to "in use" and close lid down to lock bottles 4432, 4434 into the system's pump; place container unit 4402 into mobile apparatus 4300 adjacent to main unit 4404; remove remote 4318 from apparatus 4300 and leave the room to be treated, and when yellow light 4441 illuminates, apparatus 4300 will begin pumping fluid into main unit 4404; when green light 4442 illuminates, apparatus 4300 is ready to begin disinfection; when blue light 4443 illuminates, apparatus 4300 has begun its cycle and humidifier 4302 and fans 4314, 4322 are activated; when the disinfection cycle is complete, a deactivation command will appear, and UV lights 4446 and chemical release will cause the system to deactivate; when red light 4444 illuminates, apparatus 4300's cycle is complete, it is safe for the user to return to the disinfected room, and the fluid has been pumped out of main unit 4304, 4404 and into the original bottles 4432, 4434; container unit 4402 will unlock upon replacement of remote 4318 back upon apparatus 4300, bottles 4432, 4434 may be inspected and caps 4436 may be set to "dispose" and discarded.

[00367] In another aspect, the process of generating antimicrobial gas may include the following steps:

[00368] 1. <u>Steps in Process</u>

[00369] a. User inputs room identifying information (or automated via RFID tag) and selects disinfection routine via touch screen, Bluetooth or WIFI communication.

[00370] i. Disinfection cycle may vary based on need, e.g., short-cycle sanitization, between patient turnover, known contagion in room, deodorizing, and the like.

[00371] ii. Laser scanner to calculate room dimensions/volume.

[00372] iii. Disinfection cycle parameters (e.g., RH, ramp up, concentration, time, aeration) based on data models for targeted level of disinfection and room size.

[00373] 1. Actual data will be recorded for each cycle for each room and used to refine models, in general and for specific rooms/spaces.

[00374] iv. User provided feedback giving estimated time to run disinfection cycle.

[00375] v. Warning light activated indicating cycle about to start (occupants should leave the room).

[00376] b. Device executes user selected disinfection routine based on data model.

[00377] i. Warning light changes color indicating cycle has started; user is notified via remote monitoring app.

[00378] ii. Conditions the room to target RH value (based on remote sensor feedback).

[00379] iii. Calculates rate of antimicrobial gas generation needed to reach target concentration (ppm) during pre-determined time window for initial ramp up.

[00380] iv. Generates and dispenses antimicrobial gas at calculated rate.

[00381] v. Uses forced air flow and directional and/or rotating nozzles to dispense gas into room volume.

[00382] vi. CFM is in excess of gas generation uptake rate and enough to uniformly mix gas in room volume.

[00383] vii. Intake air for gas antimicrobial mixing is HEPA filtered.

[00384] viii. Adjusts rate of antimicrobial gas generation to hit targeted rate during ramp up based on feedback from remote chemical sensors to adjust gas generation.

[00385] 1. Required rate of antimicrobial gas generation is impacted by amount of equipment, furniture, etc. in the room (taking up calculated volume space), the uptake of antimicrobial gas by porous items in the room, and the natural decay of antimicrobial gas concentration.

[00386] ix. Automatically adjusts rate of antimicrobial gas generation based on feedback from remote chemical sensors to maintain target concentration for duration of disinfection cycle.

[00387] x. Updates user on time to end of cycle once steady-state conditions are met.

[00388] xi Continuously monitors and records sensor data, creating a record that disinfection process parameters were maintain throughout cycle.

[00389] xii. Terminates gas generation at end of program cycle.

[00390] xiii. Aeration cycle is initiated.

[00391] 1. Lower blower system turns over room in air until chemical antimicrobial is no longer detectable (plus factor of safety).

[00392] 2. Intake air is filtered through carbon filter and HEPA filter to remove antimicrobial and contaminants from air.

[00393] xiv. Warning light changes color indicating cycle has ended; user is notified via remote monitoring app that it is safe to enter the room.

[00394] c. Reporting and data analytics.

[00395] i. Report file generated and uploaded to central data collection system for documentation purposes.

[00396] ii. Process data added to model training data set to continuously refine disinfection models, both generally and for that specific room.

[00397] 2. Chlorine Dioxide Gas

[00398] a. Pure chlorine dioxide gas can be generated from any number of source materials; preferably, generation materials produce a high level of chlorine dioxide.

[00399] b. Method of generating ClO₂ needs to be capable of enough ClO₂ for at least one antimicrobial cycle.

[00400] c. Method of generating ClO₂ needs to be capable of producing ClO₂ fast enough to reach target room concentration levels within about 15 minutes.

[00401] d. Method of ClO₂ generation may be batch process generation or just-in-time production.

[00402] e. Generation materials are preferably provided in a form that does not require human contact, here introduction/integration process with equipment support chemical feed, and feed rate can be controlled to control the rate of ClO₂ production.

[00403] f. Pure ClO₂ gas can be separated from liquid using any method, e.g., stirring/mixing; aeration; surface fans/blower; water tower with countercurrent air; airflow over/through water flow or spray; thin film evaporation; vacuum; piezoelectric; heating; and the like.

[00404] g. Liquid byproducts from ClO₂ generation process may be neutralized by any number of chemical reaction processes to destroy residual ClO₂. Alternatively, generation liquids can be recirculated through the system during the aeration cycle to remove residual ClO₂.

[**00405**] 3. Configuration

[00406] a. The ClO₂ gas disinfection system can be configured as a fully automated unit, with full process control and documentation features, as described.

[00407] b. Manual configurations without process automation and control may be configured for use by properly trained personnel.

Example 1:

As illustrated in **FIG. 8**, gas or vapor generator **830** may be oriented within sealed environment **810**. The use of micro devices (e.g., gas or vapor generator **830**) to generate ClO₂ can generate low target concentrations in large volumes (e.g., 1,300 cubic feet/36.8 cubic meters) while requiring low raw materials. In Example 1, a dose of 22 μL of 0.75 g/mL NaClO₂ and 36 μL of 12 M HCl were dispensed using two syringe pumps into a PVC tube applicator with 1.5 L/min flow rate of air blowing across the liquid drop as it was dispensed. The syringe pumps and PVC tube applicator were located in the center of an ISO shipping container (e.g., sealed

environment 810). The blower blew the air within the enclosed, and sealed, ISO shipping container, which had an internal volume of 1,300 cubic feet/36.8 cubic meters. FIG. 51A illustrates the time (minutes) to equilibrium for a target concentration of 0.1 ppm of ClO₂ to air. As illustrated, equilibrium was reached in a matter of minutes using this very small setup.

[00409] After the ambient air inside the ISO shipping container reached an equilibrium of 0.08 ppm, the syringe pumps were turned on to a rate of 1 μL/min. and the ClO₂ concentration was measured at five different ports in the ISO shipping container walls, the ports being spread at different locations around the ISO shipping container. **FIG. 51B** illustrates the concentration (ppm of ClO₂ to air) measured at each of the five ports over time (minutes). The concentration measured at each port was substantially similar over the test time, as illustrated in **FIG. 51B**.

Example 2:

The use of micro devices (e.g., gas or vapor generator 830) to generate ClO₂ can generate low target concentrations in large volumes (e.g., 1,300 cubic feet/36.8 cubic meters) while requiring low raw materials. In Example 2, a dose of 125 mL of 0.75 g/mL NaClO₂ and 632 mL of 0.50 g/mL Na₂S₂O₈ were dispensed into a unit with fans blowing down onto the ClO₂ solution. The unit was located in the center of an ISO shipping container (e.g., sealed environment 810). The fans blew the air within the enclosed, and sealed, ISO shipping container, which had an internal volume of 1,300 cubic feet/36.8 cubic meters. FIG. 52A illustrates the time (minutes) to equilibrium for a target concentration of 350 ppm of ClO₂ to air. As illustrated, equilibrium was reached in about 60 minutes.

[00411] After the ambient air inside the ISO shipping container reached a concentration of about 350 ppm, ClO₂ production was ceased, and a PortaSens device was used to read the concentration at 12 different ports in the ISO shipping container walls, the ports being spread at different locations around the ISO shipping container. **FIG. 52B** illustrates the concentration (ppm of ClO₂ to air) measured at each of the 12 ports over time (minutes). The concentration measured at each port was substantially similar over the test time, as illustrated in **FIG. 52B**.

Example 3:

[00412] FIGS. 53A and 53B illustrate diagrams of an example system 5300 for generating ClO₂ vapor from small volumes of high concentration liquid precursors. System 5300 includes a sodium chlorite concentrate 5302 and an activator concentrate 5304. Sodium chlorite concentrate 5302 is fluidically connected to a pump 5306, while activator concentrate 5304 is fluidically connected to a pump 5308. A controller 5310 is operatively connected to both of pumps 5306 and 5308. Controller 5310 controls the operation of pumps 5306 and 5308, including at least volume of fluid pumped, flow rate, timing of pump activation, and the like.

[00413] As illustrated in FIG. 53A, each of pumps 5306 and 5308 are fluidically connected to a t-mixing chamber 5312, where sodium chlorite concentrate 5302 and activator concentrate 5304 are combined to generate ClO₂ vapor. As illustrated in FIG. 53B, each of pumps 5306 and 5308 are fluidically connected to a microfluidic mixing chip 5318, where sodium chlorite concentrate 5302 and activator concentrate 5304 are combined to generate ClO₂ vapor.

ClO₂ vapor is diffused into the ambient air at diffuser 5314. A ClO₂ sensor 5316 senses the concentration of ClO₂ in the ambient air and is operatively connected to controller 5310. If the concentration of ClO₂ in the ambient air is lower than desired, controller 5310 causes pumps 5306 and 5308 to generate more ClO₂, or to generate ClO₂ at a greater rate, as necessary to achieve the desired concentration of ClO₂. If the concentration of ClO₂ in the ambient air is greater than desired, controller 5310 causes pumps 5306 and 5308 to generate less ClO₂, or to generate ClO₂ at a lesser rate, or to cease the generation of ClO₂ for a desired time to allow the concentration of ClO₂ to fall to a desired level, as necessary to achieve the desired concentration of ClO₂.

[00415] Pumps, such as pumps 5306, 5308, 308A, 308B, and 308C, may be positive displacement pumps. Positive displacement pumps may provide a benefit in that for each rotation/reciprocation of the pump, the volume of fluid pumped is known. In this arrangement, a mass flow controller or flow sensor (such as flow sensors 318A and 318B) may be eliminated from the system. Positive displacement pumps may allow a closed loop independent sensor (e.g., an encoder) on the pump's rotation/reciprocation means, which further allows the system to yield an independent measure of the pump's movement and/or the volume of fluid pumped. When not reciprocating or rotating, the pumping action may maintain a normally-closed configuration to eliminate leakage flow, which is critical to the control of microvolumes (e.g., microliters), and may eliminate one or more secondary valves, including for example one or more of a leak control valve and a check valve.

[00416] In one aspect, the matter transport system of antimicrobial generators must be designed to minimize post-pump to generator-release "dead volume," which pertains to how much material is left between a pump and downstream active/passive fluidic and/or generator elements. In one aspect, a target may be less than 1X, or less than 0.5X of minimum generator cycle volume of precursors consumed as same dead space.

FIGS. 54A-C illustrate results of ClO₂ generation using system 5300 or similar systems. The results illustrated in FIGS. 54A-C correspond to generation of 0.1 ppm of ClO₂ vapor from small volumes of high concentration precursors. FIG. 54A illustrates results obtained from a two-component concentrated liquid generation of ClO₂. FIG. 54B illustrates results obtained from an electrochemical generation of ClO₂ from concentrated liquid NaClO₂. FIG.

54C illustrates projections for chemical use necessary for a 1,000 cubic foot (28.3 cubic meter) room including various activators, both for initial treatment and after 30 days of continuous operation.

Example 4:

The efficacy of ClO₂ at ranges of approximately 0.1 ppmv and 5 ppmv was assessed against clinically-relevant infectious bacteria including Klebsiella pneumonia (Kp), Pseudomonas aeruginosa (PA), Staphylococcus aureus (Sa), and Salmonella enterica (Se), as well as bacteriophage Phi6 and MS2 (representing enveloped and non-enveloped virus, respectively). The microorganisms were prepared in phosphate buffered saline (PBS), dispensed onto replicate glass coupons (five 10 μL droplets; equivalent to 5-6 log cells or virions per coupon), placed into a room-scale test chamber conditioned with ClO₂ and 50-60% relative humidity (RH) and operated at ambient temperatures ranging from 18 to 21 °C.

[00419] To assess efficacy, the concentration viable bacteria or infective virions recovered from ClO₂ treated coupons versus untreated control coupons versus time were measured. Per test, replicate coupons (duplicates or triplicates) were removed from the test chamber at various time intervals and assayed (extracted and enumerated) to determine the total quantity of organisms recovered. The results were plotted as kill curves (expressed as log organisms recovered versus time). The kill curves were then used to calculate the D-values of gas treatment, representing the time required to achieve a 90% reduction (or 1 log reduction) of viable/infective organisms at a given test condition. The area of the kill curve in which linear decay was observed was used to determine the D-value (calculated as the negative inverse of the linear decay slope).

[00420] FIGS. 55A and 55B illustrate the mean D-values (hours) from replicate tests per organism performed at the range of 0.11 ± 0.04 ppmv (FIG. 55A) and 5.3 ± 2.4 ppmv (FIG. 55B).

[00421] The results demonstrate that at 0.1 ppmv (FIG. 55A) a reduction of 90% of all organisms was rapid and comparable ranging from 0.5 to 1.2 hours (or 31 to 70 minutes).

[00422] As would be expected, the efficacy increased with treatment at 5 ppmv (FIG. 55B) with D-values ranging from 0.2 to 0.3 hours (or 13 to 19 minutes).

[00423] Based on this data, the time to achieve a 99.9 % or 3-log reduction at 0.1 ppmv and 5 ppmv correlates to 1.5 to 3.6 hours and 0.6 to 0.9 hours, respectively.

[00424] A system for generating and monitoring an antimicrobial, is provided, the system comprising: a computational system; an antimicrobial sensor; and an antimicrobial generator, wherein the computational system, the antimicrobial generator, and the antimicrobial sensor are operatively connected. The computational system may be at least one of a microprocessor and a microcontroller. The system may further include an external communication device. The system

may include a separate sensor sub-system comprising: at least one of a sensor sub-system microprocessor and a sensor sub-system microcontroller; a sensor sub-system external communications device; at least one of a sensor sub-system antimicrobial sensor and a sensor sub-system environmental sensor; and a sensor sub-system computational system. The system may include a separate generation sub-system comprising: at least one of a generation sub-system microprocessor and a generation sub-system microcontroller; a generation sub-system external communications device; and a generation sub-system antimicrobial generator. The external communications device, the computational system, the antimicrobial generator, and the at least one of an antimicrobial sensor and an environmental sensor may be oriented within an enclosed volume under treatment. At least one sensor sub-system and/or generation sub-system may be oriented within an enclosed volume under treatment.

A system for generating and monitoring an antimicrobial is provided, the system [00425] comprising: a sensor sub-system comprising: at least one of a sensor sub-system microprocessor and a sensor sub-system microcontroller, a sensor sub-system external communications device, at least one of a sensor sub-system antimicrobial sensor and a sensor sub-system environmental sensor, and a sensor sub-system computational system; a generation sub-system comprising: at least one of a generation sub-system microprocessor and a generation sub-system microcontroller, a generation sub-system external communications device, and a generation sub-system antimicrobial generator; and an enclosed space forming a volume under treatment. The sensor sub-system and the generation sub-system may be oriented within the enclosed volume under treatment. The sensor sub-system may be oriented within the enclosed volume under treatment and the generation sub-system may be oriented outside of the enclosed volume under treatment. The generation sub-system may be oriented within the enclosed volume under treatment and the sensor sub-system may be oriented outside of the enclosed volume under treatment. The system may include an HVAC air supply fluidically connected to the interior of the enclosed volume under treatment, the sensor sub-system may be oriented within the enclosed volume under treatment, the generation sub-system may be oriented outside of the enclosed volume under treatment, and the generation sub-system may be fluidically connected to the HVAC air supply. The system may include an HVAC air return fluidically connected to the interior of the enclosed volume under treatment, the generation sub-system may be oriented within the enclosed volume under treatment, the sensor sub-system may be oriented outside of the enclosed volume under treatment, and the sensor sub-system may be fluidically connected to the HVAC air return.

[00426] A system for generating and monitoring ClO₂ is provided, the system comprising: a device housing including an inlet; a microcontroller or microprocessor; a reagent container

containing a reagent; a device for generating a ClO₂ from the reagent; and a sensing system. The system may include two reagent containers, and each reagent container may contain a different reagent. The device for generating the ClO₂ may be a microfluidic mixer, and the two reagents may mix in the microfluidic mixer to generate the ClO₂. The device for generating the ClO₂ may be an electrochemical generator. The sensing system may measure a concentration of ClO₂ in ambient air introduced via the inlet. The measurement of concentration of ClO₂ in the ambient air may be communicated to the microcontroller or microprocessor, and the microcontroller or microprocessor may cause the system to generate the ClO₂ if the ClO₂ concentration is below a target value. The system may include one reagent container and one reagent, the device for generating the ClO₂ may be an electrochemical generator, and the electrochemical generator may use an electrical potential to cause a reaction with the reagent that generates the ClO₂. The electrochemical generator may be a microfluidic device. The system may include a barometric sensor to sense a pressure of ambient air introduced via the inlet, the pressure may be communicated to the microcontroller or microprocessor, and a negative pressure may cause the microcontroller or microprocessor to pause ClO₂ generation until a neutral and/or positive pressure is sensed by the barometric sensor. The system may include an off-gas and waste chamber having a membrane, waste from the generation of the ClO₂ may be absorbed in an absorber material, and ClO₂ may exit the off-gas and waste chamber through the membrane and into an ambient atmosphere. The system may include an air pump electrically connected to the microcontroller or microprocessor and fluidically connected to the inlet via an air duct. The microcontroller or microprocessor is controlled by machine learning algorithms to alter system performance. The microcontroller or microprocessor may be controlled by artificial intelligence algorithms to alter system performance. The microcontroller or microprocessor may alter system performance automatically. The microcontroller or microprocessor may alter system performance by control by a user. The microcontroller or microprocessor may alter the system performance based upon at least one of: a detection of a virus in ambient air containing the system; a detection of bacteria in ambient air containing the system; an altitude of the system; a temperature of the system; changes in ambient air measured by changes in a concentration of ClO₂ in ambient air; changes in occupancy by living beings of an area containing the system; alterations for a user's preferences; prediction of cycles of occupancy and vacancy by living beings of the area containing the system; and a diagnosis of normal or abnormal performance of the system.

[00427] A network of systems for generating and monitoring ClO₂ is provided, the network of systems comprising: a plurality of systems for generating and monitoring ClO₂, including: a device housing including an inlet; a microcontroller; a reagent container containing a reagent; a

microcontroller includes a communication device capable of communication between the plurality of systems, wherein the communication device establishes distributed control of each system's microcontroller, and wherein the microcontroller is controlled by machine learning algorithms to alter system performance. The distributed control may include at least one of: adjusting individual systems to achieve a uniform or deliberately non-uniform distribution of ClO₂ in each individual sensor's location within a specified space; consumption of ClO₂; control of day and/or night generation cycles; using the sensing system to sense patterns across time, three-dimensional volumes, seasonal variations; sending patterns that are inferred or traced to a signal measured; and sensing patterns that are directly traceable to variations observed in ClO₂ concentrations across the network of systems installed across distinct spaces.

A network of systems for generating and monitoring ClO₂ concentration is [00428] provided, the network of systems comprising: a plurality of systems for generating and monitoring ClO₂, including: a device housing including an inlet; a microcontroller; a reagent container containing a reagent; a microfluidic device for generating a ClO₂ from the reagent; and a sensing system; wherein the microcontroller includes a communication device capable of communication between the plurality of systems, wherein the communication device establishes distributed control of each system's microcontroller, and wherein the microcontroller is controlled by artificial intelligence algorithms to alter system performance. The distributed control may include at least one of: adjusting individual systems to achieve a uniform or deliberately nonuniform distribution of ClO₂ in each individual sensor's location within a specified space; consumption of ClO₂; control of day and/or night generation cycles; using the sensing system to sense patterns across time, three-dimensional volumes, seasonal variations; sending patterns that are inferred or traced to a signal measured; and sensing patterns that are directly traceable to variations observed in ClO₂ concentrations across the network of systems installed across distinct spaces.

To the extent that the term "includes" or "including" is used in the specification or the claims, it is intended to be inclusive in a manner similar to the term "comprising" as that term is interpreted when employed as a transitional word in a claim. Furthermore, to the extent that the term "or" is employed (e.g., A or B) it is intended to mean "A or B or both." When the applicants intend to indicate "only A or B but not both" then the term "only A or B but not both" will be employed. Thus, use of the term "or" herein is the inclusive, and not the exclusive use. See Bryan A. Garner, A Dictionary of Modern Legal Usage 624 (2d. Ed. 1995). Also, to the extent that the terms "in" or "into" are used in the specification or the claims, it is intended to additionally mean

"on" or "onto." To the extent that the term "substantially" is used in the specification or the claims, it is intended to take into consideration the degree of precision available in manufacturing. To the extent that the term "selectively" is used in the specification or the claims, it is intended to refer to a condition of a component wherein a user of the apparatus may activate or deactivate the feature or function of the component as is necessary or desired in use of the apparatus. To the extent that the term "operatively connected" is used in the specification or the claims, it is intended to mean that the identified components are connected in a way to perform a designated function. As used in the specification and the claims, the singular forms "a," "an," and "the" include the plural. Finally, where the term "about" is used in conjunction with a number, it is intended to include \pm 10 % of the number. In other words, "about 10" may mean from 9 to 11.

[00430] As stated above, while the present application has been illustrated by the description of aspects thereof, and while the aspects have been described in considerable detail, it is not the intention of the applicants to restrict or in any way limit the scope of the appended claims to such detail. Additional advantages and modifications will readily appear to those skilled in the art, having the benefit of the present application. Therefore, the application, in its broader aspects, is not limited to the specific details, illustrative examples shown, or any apparatus referred to. Departures may be made from such details, examples, and apparatuses without departing from the spirit or scope of the general inventive concept.

CLAIMS

What is claimed is:

1. A system for generating and monitoring an antimicrobial, comprising:

a computational system;

an antimicrobial sensor; and

an antimicrobial generator,

wherein the computational system, the antimicrobial generator, and the antimicrobial sensor are operatively connected.

- 2. The system of claim 1, wherein the computational system is at least one of a microprocessor and a microcontroller.
 - 3. The system of claim 1, further comprising an external communication device.
- 4. The system of claim 1, further comprising a separate sensor sub-system comprising:
 - at least one of a sensor sub-system microprocessor and a sensor sub-system microcontroller;
 - a sensor sub-system external communications device;
 - at least one of a sensor sub-system antimicrobial sensor and a sensor sub-system environmental sensor; and
 - a sensor sub-system computational system.
- 5. The system of claim 1, further comprising a separate generation sub-system comprising:
 - at least one of a generation sub-system microprocessor and a generation subsystem microcontroller;
 - a generation sub-system external communications device; and
 - a generation sub-system antimicrobial generator.
- 6. The system of claim 3, wherein the external communications device, the computational system, the antimicrobial generator, and the at least one of an antimicrobial sensor and an environmental sensor is oriented within an enclosed volume under treatment.
- 7. The system of claim 4, wherein at least one sensor sub-system is oriented within an enclosed volume under treatment.
- 8. The system of claim 5, wherein at least one generation sub-system is oriented within an enclosed volume under treatment.
 - A system for generating and monitoring an antimicrobial, comprising:
 a sensor sub-system comprising:

at least one of a sensor sub-system microprocessor and a sensor sub-system microcontroller,

- a sensor sub-system external communications device,
- at least one of a sensor sub-system antimicrobial sensor and a sensor subsystem environmental sensor, and
- a sensor sub-system computational system;
- a generation sub-system comprising:
- at least one of a generation sub-system microprocessor and a generation sub-system microcontroller,
 - a generation sub-system external communications device, and
 - a generation sub-system antimicrobial generator; and
- an enclosed space forming a volume under treatment.
- 10. The system of claim 9, wherein the sensor sub-system and the generation sub-system are oriented within the enclosed volume under treatment.
- 11. The system of claim 9, wherein the sensor sub-system is oriented within the enclosed volume under treatment and wherein the generation sub-system is oriented outside of the enclosed volume under treatment.
- 12. The system of claim 9, wherein the generation sub-system is oriented within the enclosed volume under treatment and wherein the sensor sub-system is oriented outside of the enclosed volume under treatment.
- 13. The system of claim 9, further comprising an HVAC air supply fluidically connected to the interior of the enclosed volume under treatment, wherein the sensor sub-system is oriented within the enclosed volume under treatment, wherein the generation sub-system is oriented outside of the enclosed volume under treatment, and wherein the generation sub-system is fluidically connected to the HVAC air supply.
- 14. The system of claim 9, further comprising an HVAC air return fluidically connected to the interior of the enclosed volume under treatment, wherein the generation subsystem is oriented within the enclosed volume under treatment, wherein the sensor sub-system is oriented outside of the enclosed volume under treatment, and wherein the sensor sub-system is fluidically connected to the HVAC air return.
 - 15. A system for generating and monitoring ClO₂, comprising:
 - a device housing including an inlet;
 - a microcontroller or microprocessor;
 - a reagent container containing a reagent;

- a device for generating a ClO₂ from the reagent; and a sensing system.
- 16. The system of claim 15, wherein the system comprises two reagent containers, and wherein each reagent container contains a different reagent.
- 17. The system of claim 16, wherein the device for generating the ClO₂ is a microfluidic mixer, and wherein the two reagents mix in the microfluidic mixer to generate the ClO₂.
- 18. The system of claim 15, wherein the device for generating the ClO₂ is an electrochemical generator.
- 19. The system of claim 15, wherein the sensing system measures a concentration of ClO₂ in ambient air introduced via the inlet.
- 20. The system of claim 19, wherein the measurement of concentration of ClO₂ in the ambient air is communicated to the microcontroller or microprocessor, and wherein the microcontroller or microprocessor causes the system to generate the ClO₂ if the ClO₂ concentration is below a target value.
- 21. The system of claim 15, wherein the system comprises one reagent container and one reagent, wherein the device for generating the ClO₂ is a electrochemical generator, and wherein the electrochemical generator uses an electrical potential to cause a reaction with the reagent that generates the ClO₂.
- 22. The system of claim 21, wherein the electrochemical generator is a microfluidic device.
- 23. The system of claim 15, further comprising a barometric sensor to sense a pressure of ambient air introduced via the inlet, wherein the pressure is communicated to the microcontroller or microprocessor, and wherein a negative pressure causes the microcontroller or microprocessor to pause ClO₂ generation until a neutral and/or positive pressure is sensed by the barometric sensor.
- 24. The system of claim 15, further comprising an off-gas and waste chamber having a membrane, wherein waste from the generation of the ClO₂ is absorbed in an absorber material, and wherein ClO₂ exits the off-gas and waste chamber through the membrane and into an ambient atmosphere.
- 25. The system of claim 15, further comprising an air pump electrically connected to the microcontroller or microprocessor and fluidically connected to the inlet via an air duct.
- 26. The system of claim 15, wherein the microcontroller or microprocessor is controlled by machine learning algorithms to alter system performance.

27. The system of claim 15, wherein the microcontroller or microprocessor is controlled by artificial intelligence algorithms to alter system performance.

- 28. The system of claim 26 or 27, wherein the microcontroller or microprocessor alters system performance automatically.
- 29. The system of claim 26 or 27, wherein the microcontroller or microprocessor alters system performance by control by a user.
- 30. The system of claim 26 or 27, wherein the microcontroller or microprocessor alters the system performance based upon at least one of:
 - a detection of a virus in ambient air containing the system;
 - a detection of bacteria in ambient air containing the system;
 - an altitude of the system;
 - a temperature of the system;

changes in ambient air measured by changes in a concentration of ClO₂ in ambient air;

changes in occupancy by living beings of an area containing the system; alterations for a user's preferences;

prediction of cycles of occupancy and vacancy by living beings of the area containing the system; and

- a diagnosis of normal or abnormal performance of the system.
- A network of systems for generating and monitoring ClO₂, comprising: a plurality of systems for generating and monitoring ClO₂, including:
 - a device housing including an inlet;
 - a microcontroller;
 - a reagent container containing a reagent;
 - a microfluidic device for generating a ClO₂ from the reagent; and
 - a sensing system;

wherein the microcontroller includes a communication device capable of communication between the plurality of systems,

wherein the communication device establishes distributed control of each system's microcontroller, and

wherein the microcontroller is controlled by machine learning algorithms to alter system performance.

32. The network of systems of claim 31, wherein the distributed control includes at least one of:

adjusting individual systems to achieve a uniform or deliberately non-uniform distribution of ClO_2 in each individual sensor's location within a specified space;

consumption of ClO₂;

control of day and/or night generation cycles;

using the sensing system to sense patterns across time, three-dimensional volumes, seasonal variations;

sending patterns that are inferred or traced to a signal measured; and sensing patterns that are directly traceable to variations observed in ClO₂ concentrations across the network of systems installed across distinct spaces.

33. A network of systems for generating and monitoring ClO₂ concentration, comprising:

a plurality of systems for generating and monitoring ClO₂, including:

- a device housing including an inlet;
- a microcontroller;
- a reagent container containing a reagent;
- a microfluidic device for generating a ClO₂ from the reagent; and
- a sensing system;

wherein the microcontroller includes a communication device capable of communication between the plurality of systems,

wherein the communication device establishes distributed control of each system's microcontroller, and

wherein the microcontroller is controlled by artificial intelligence algorithms to alter system performance.

34. The network of systems of claim 33, wherein the distributed control includes at least one of:

adjusting individual systems to achieve a uniform or deliberately non-uniform distribution of ClO_2 in each individual sensor's location within a specified space;

consumption of ClO₂;

control of day and/or night generation cycles;

using the sensing system to sense patterns across time, three-dimensional volumes, seasonal variations;

sending patterns that are inferred or traced to a signal measured; and sensing patterns that are directly traceable to variations observed in ClO₂ concentrations across the network of systems installed across distinct spaces.

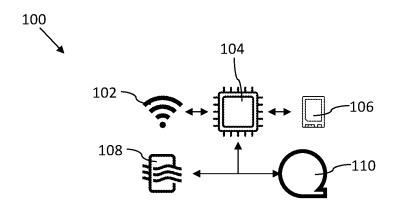


FIG. 1A

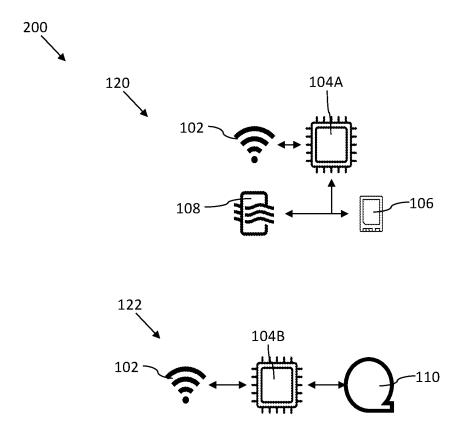
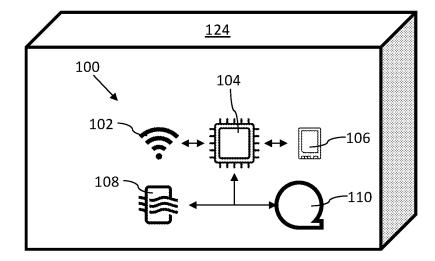


FIG. 2A



PCT/US2021/036501

FIG. 1B

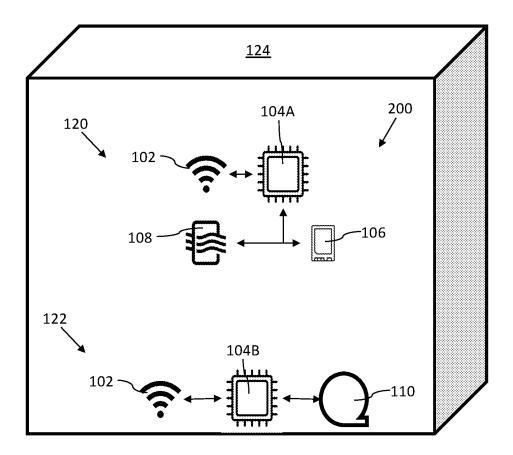


FIG. 2B

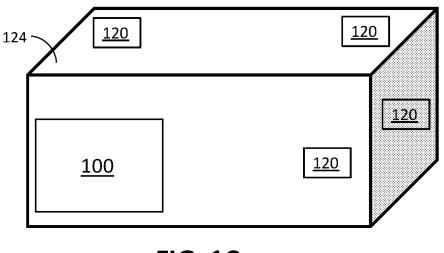


FIG. 1C

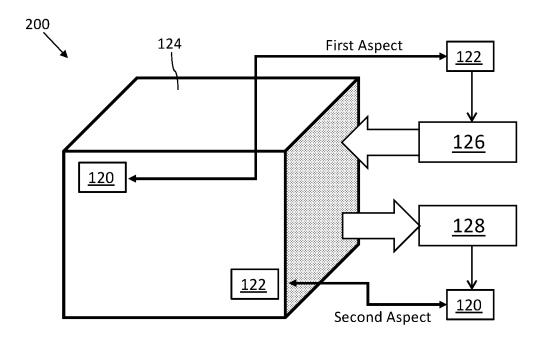


FIG. 2C

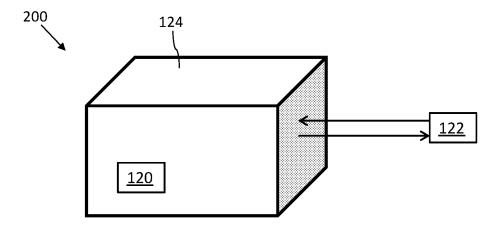


FIG. 2D

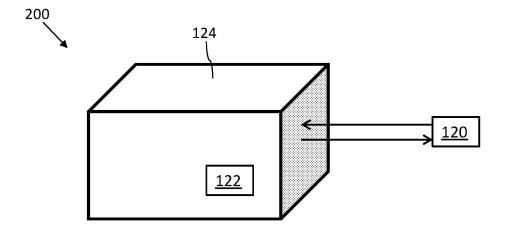
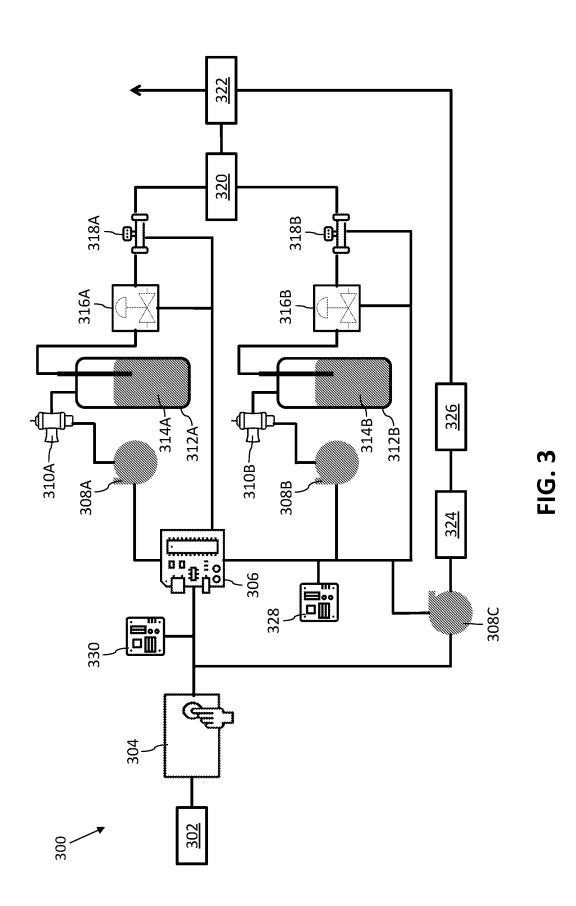
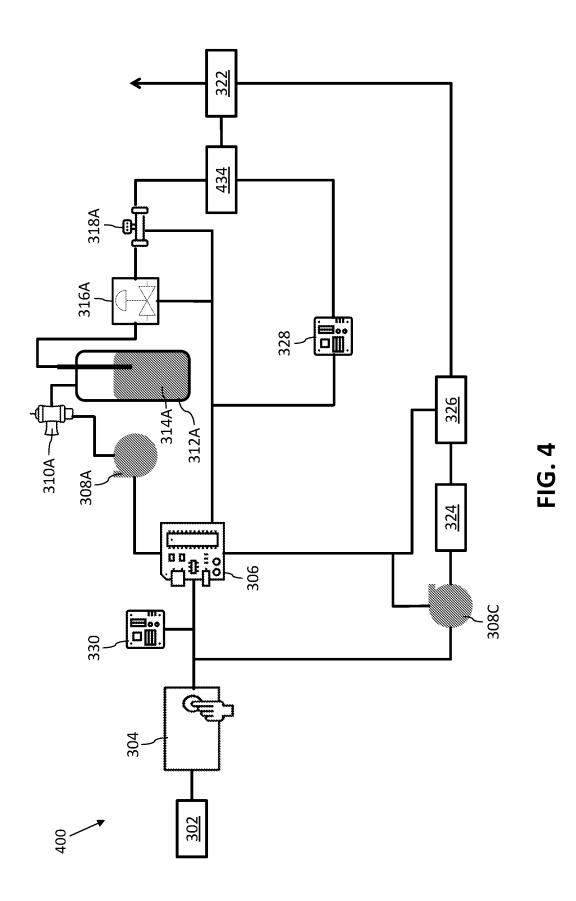
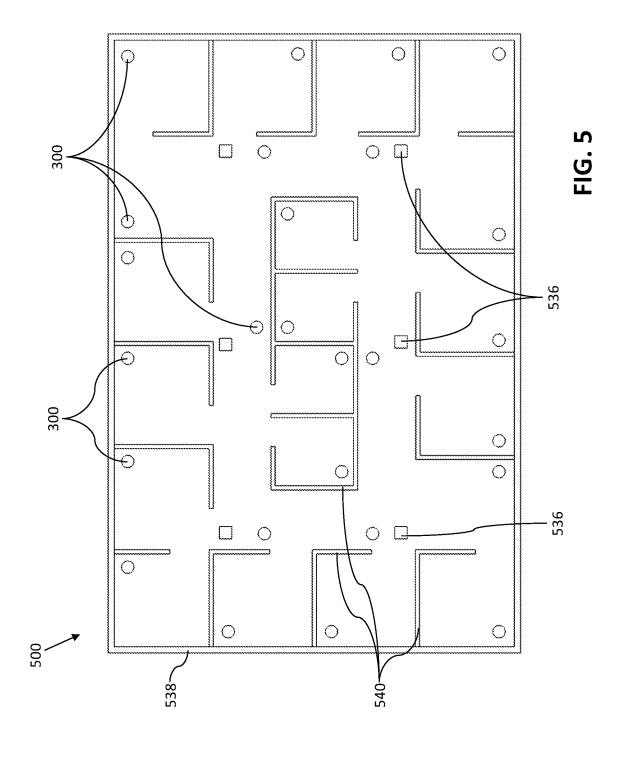
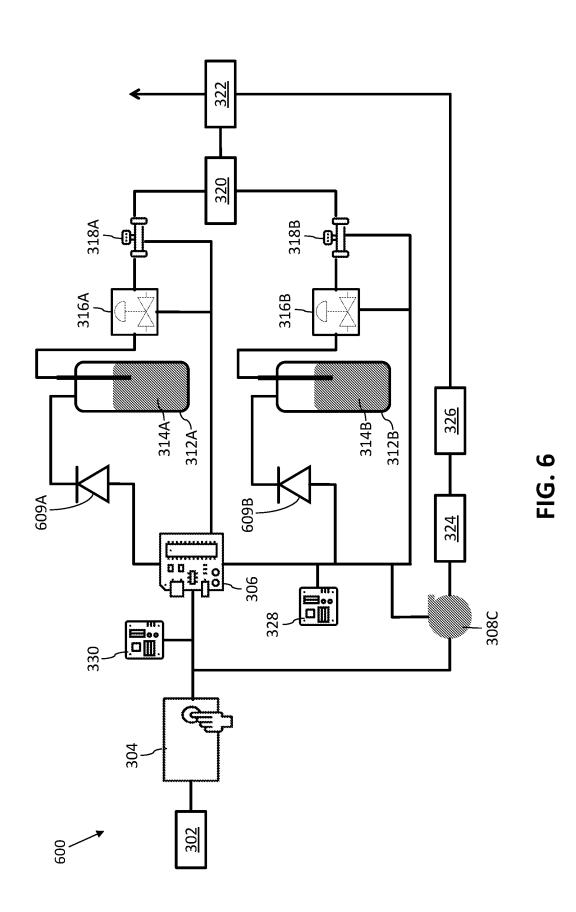


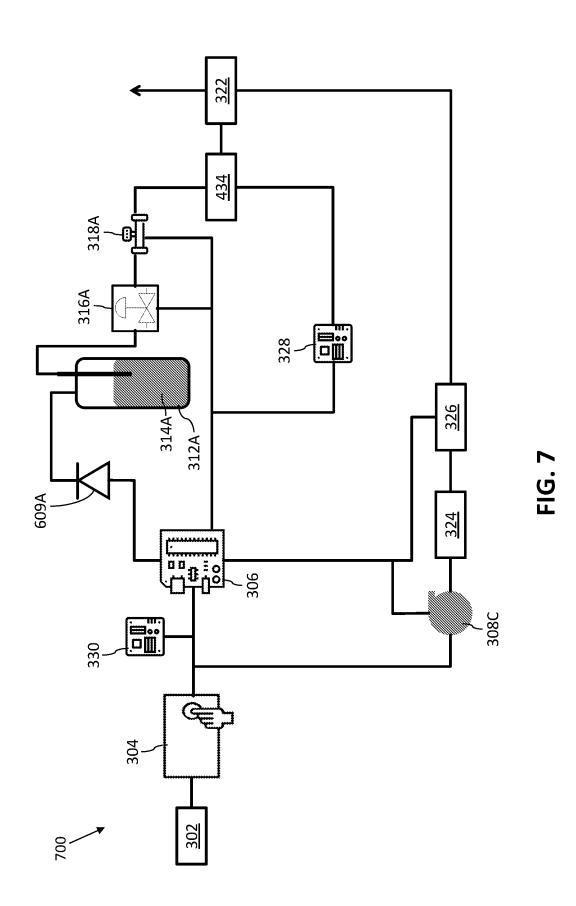
FIG. 2E











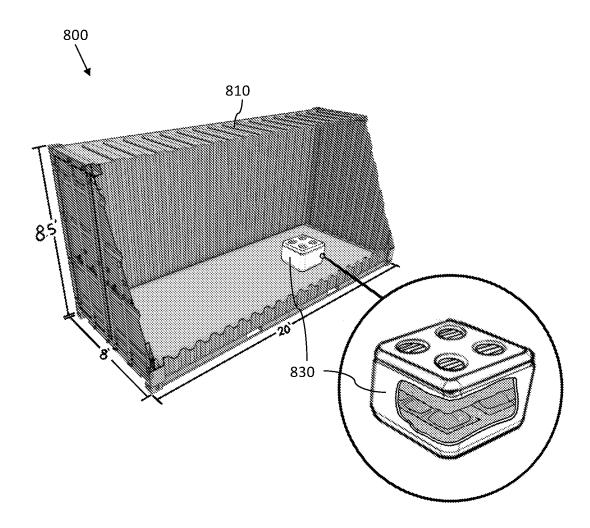


FIG. 8



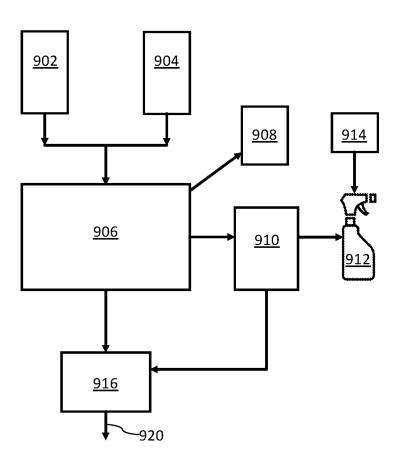


FIG. 9



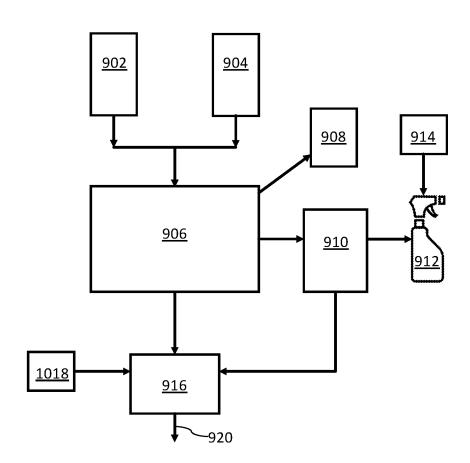


FIG. 10



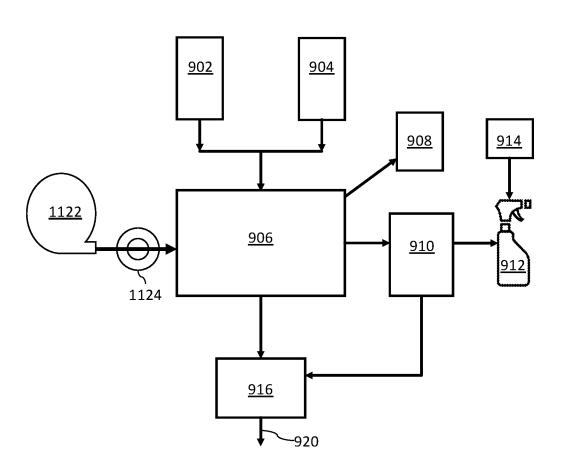


FIG. 11



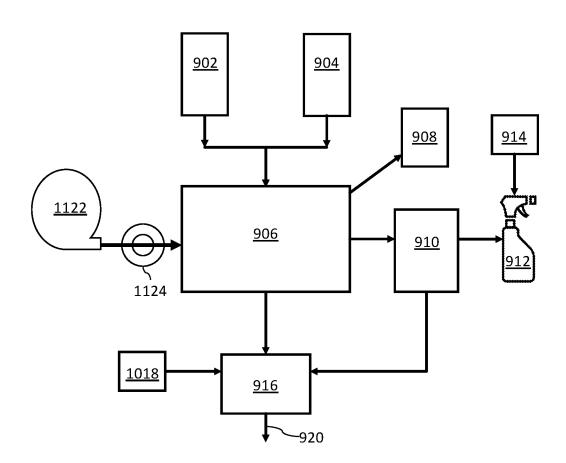


FIG. 12

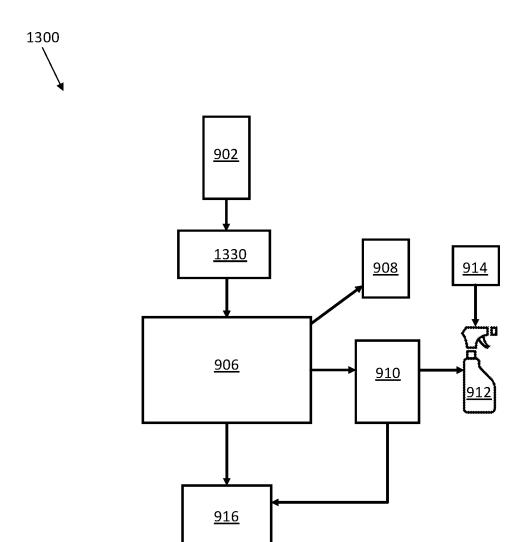


FIG. 13

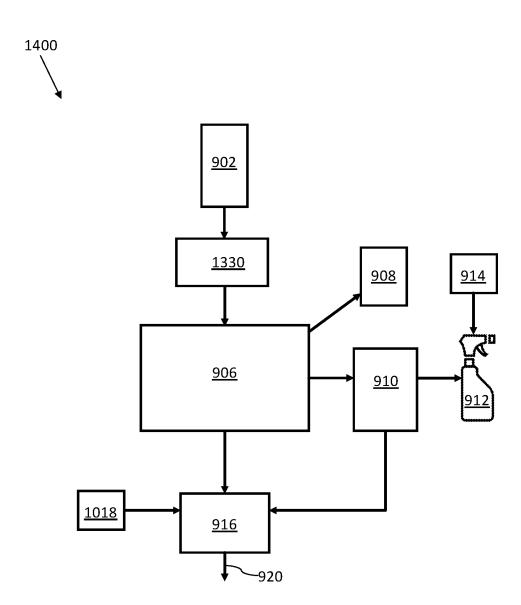


FIG. 14

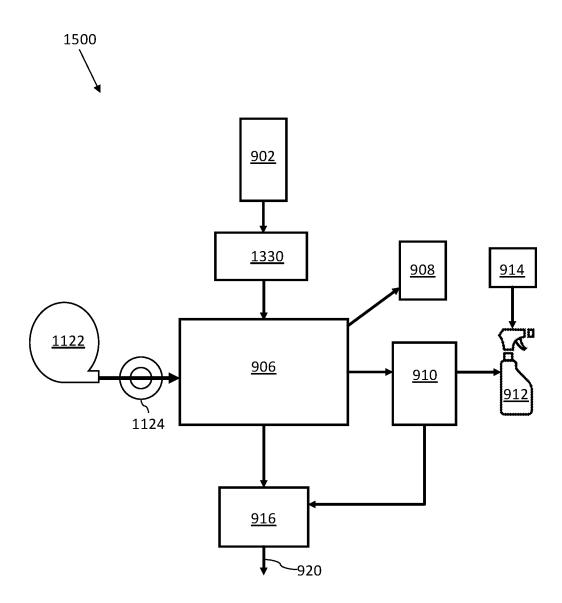


FIG. 15

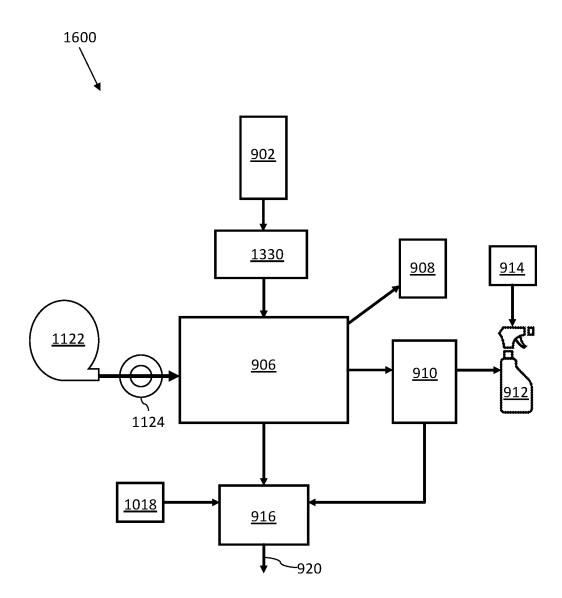


FIG. 16



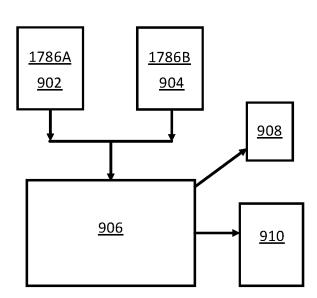


FIG. 17

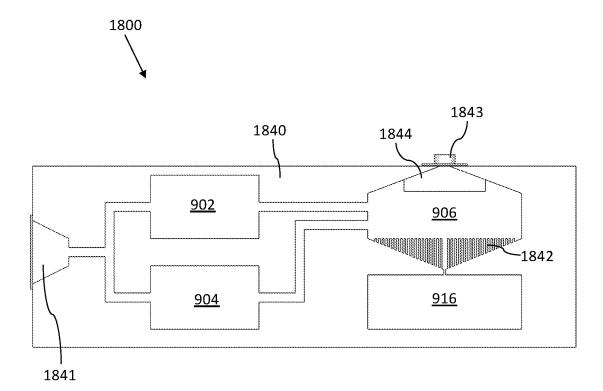


FIG. 18A

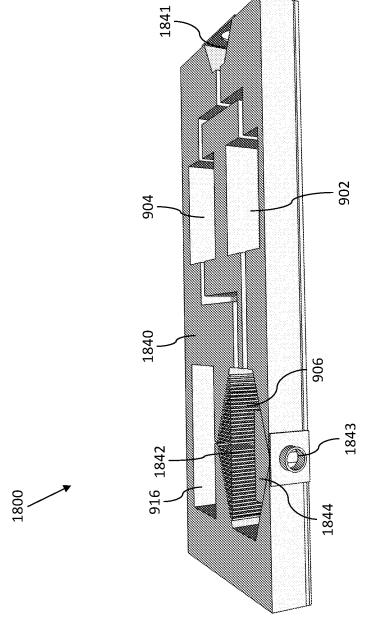


FIG. 18B

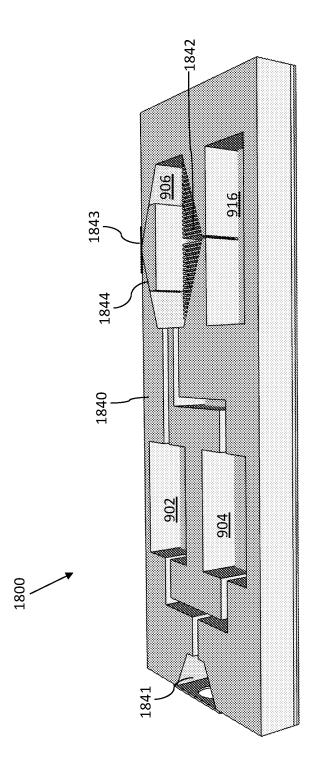


FIG. 18C

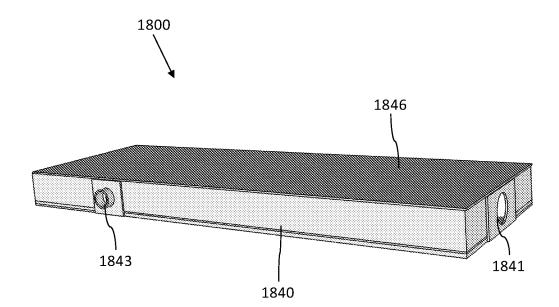


FIG. 18D



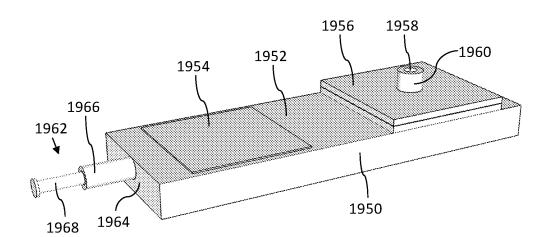


FIG. 19A

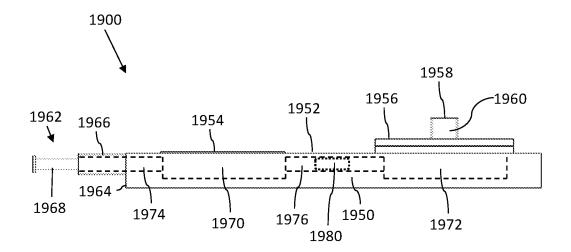


FIG. 19B

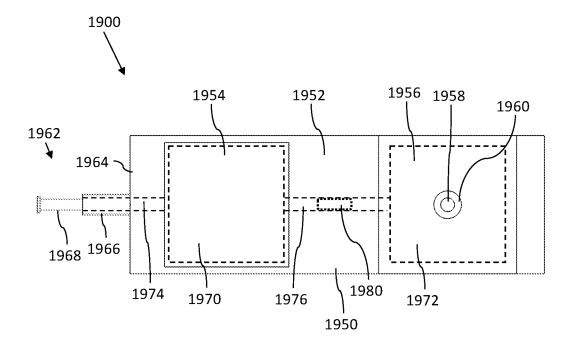


FIG. 19C

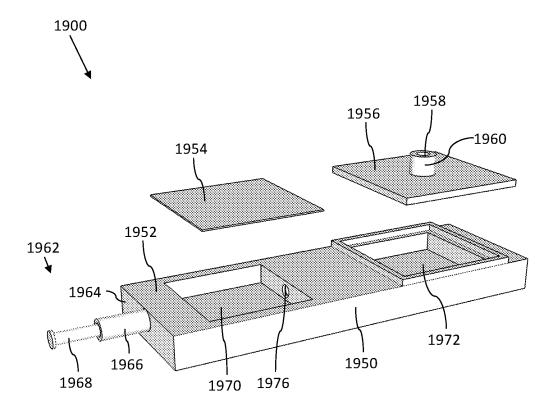


FIG. 19D

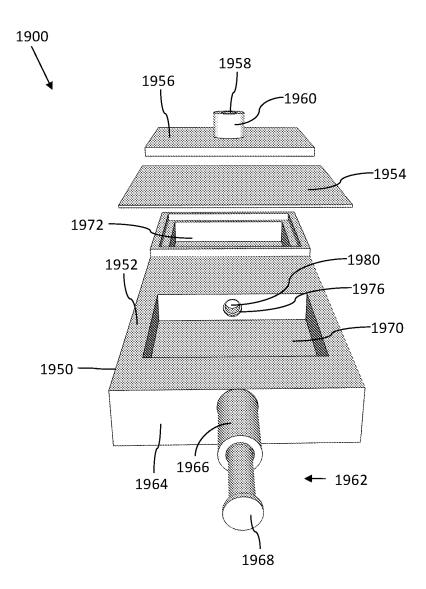


FIG. 19E

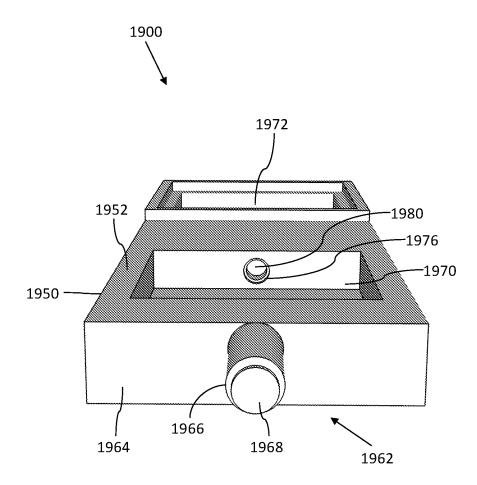


FIG. 19F

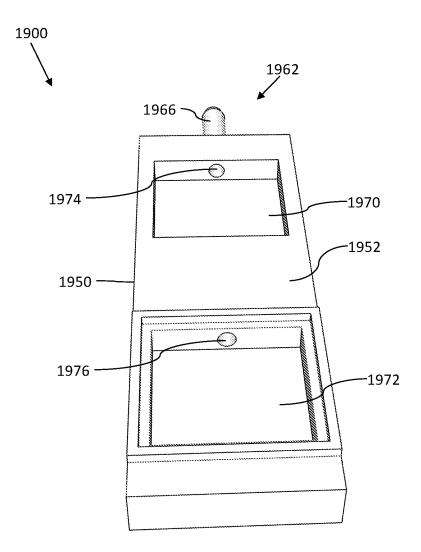


FIG. 19G

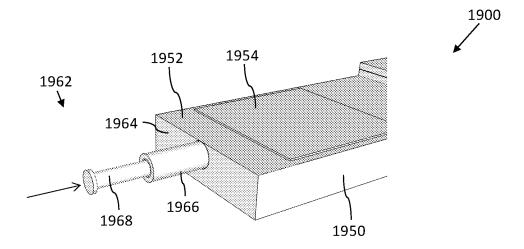


FIG. 19H

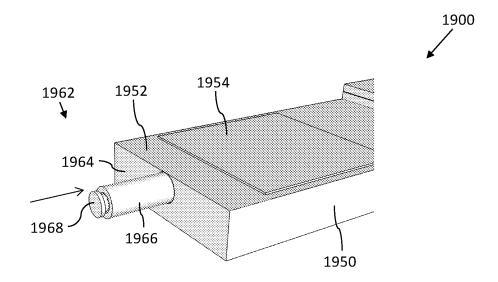


FIG. 191



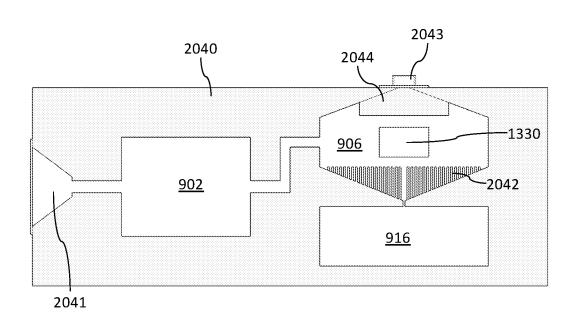


FIG. 20A

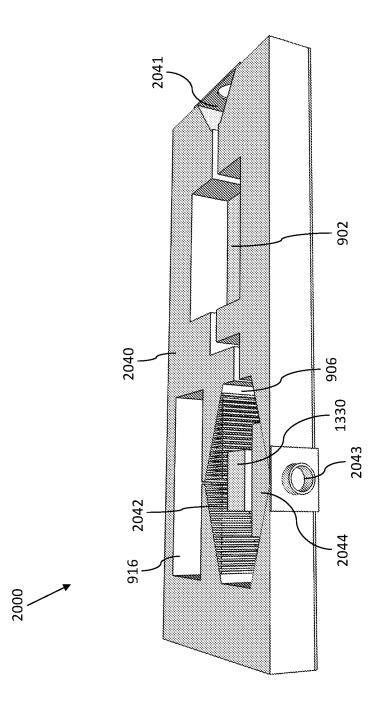
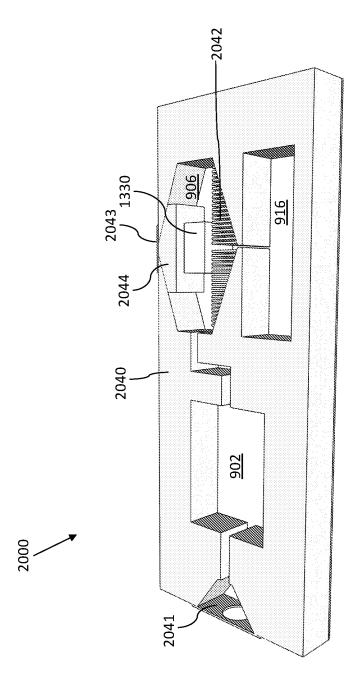


FIG. 20B



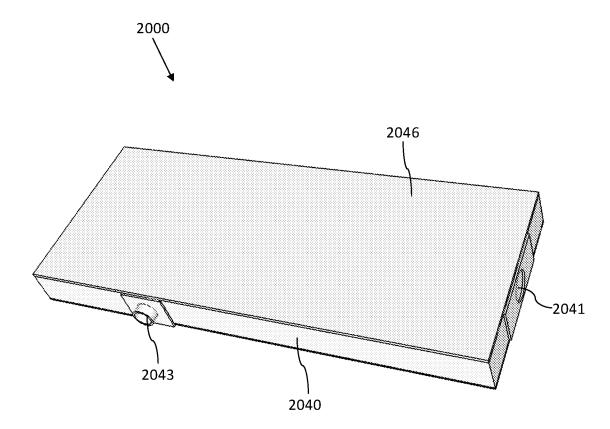


FIG. 20D

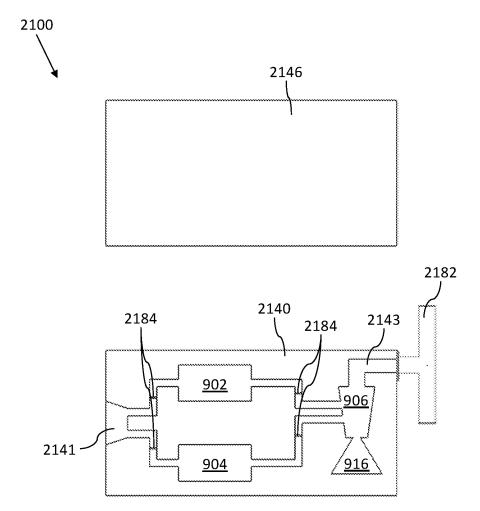


FIG. 21A

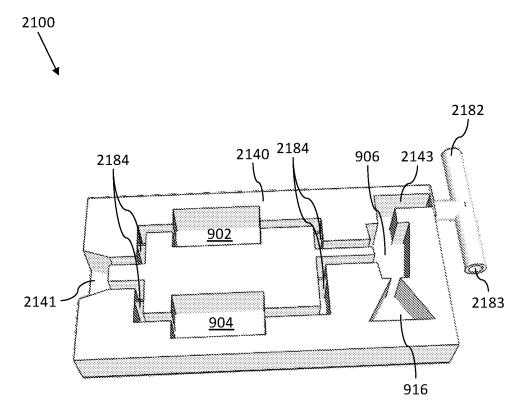


FIG. 21B



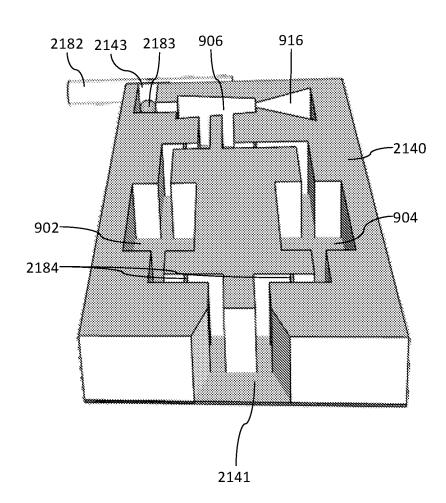


FIG. 21C

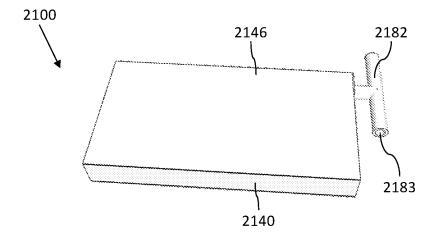


FIG. 21D

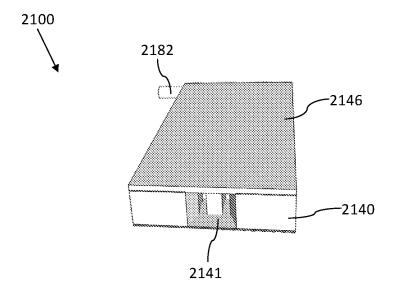


FIG. 21E

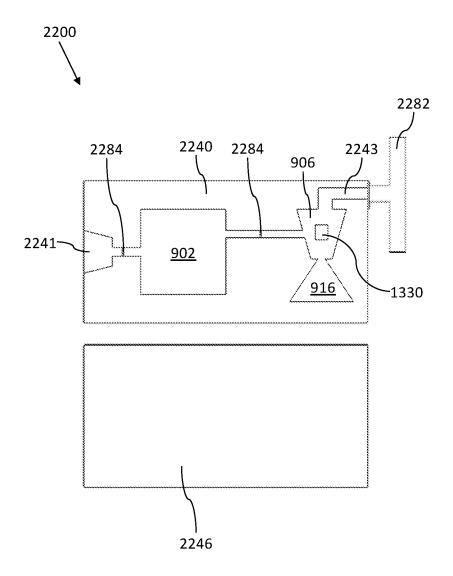


FIG. 22A



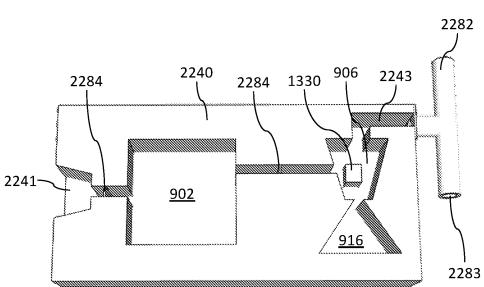


FIG. 22B

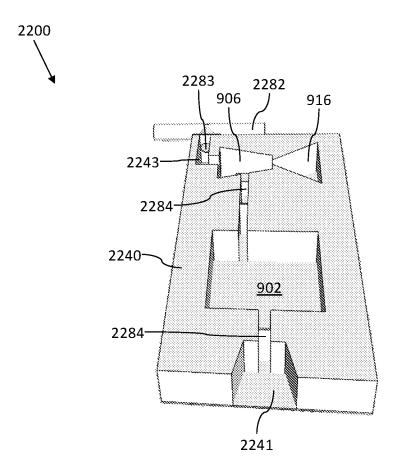


FIG. 22C

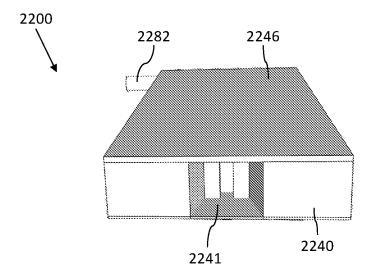


FIG. 22D

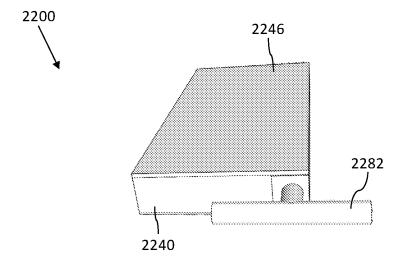


FIG. 22E

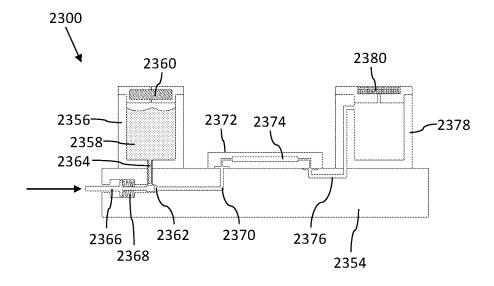


FIG. 23A

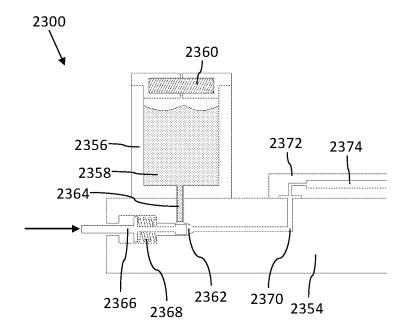


FIG. 23B

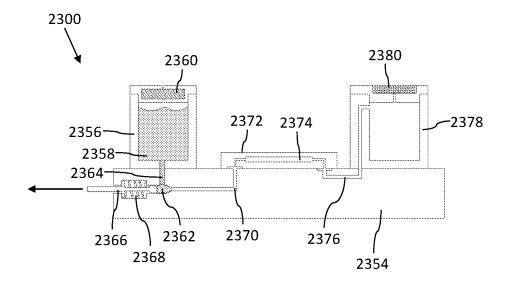


FIG. 23C

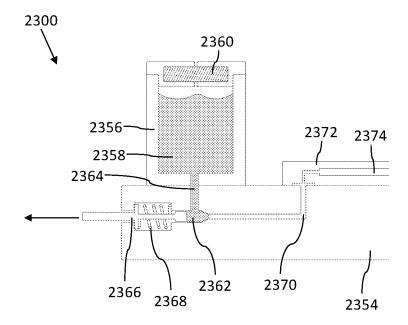


FIG. 23D

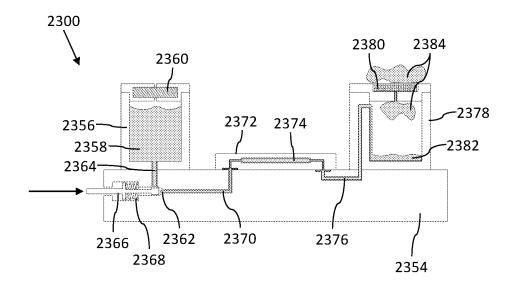


FIG. 23E

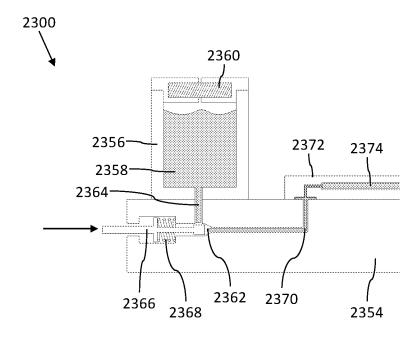


FIG. 23F

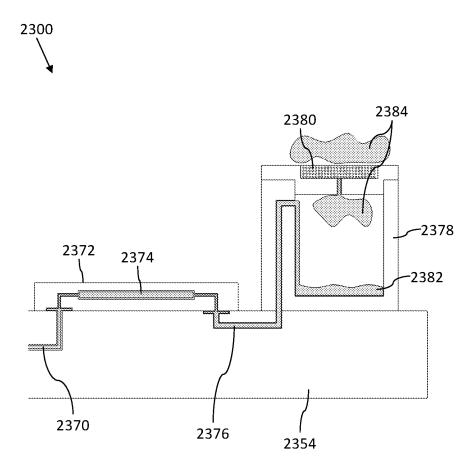


FIG. 23G

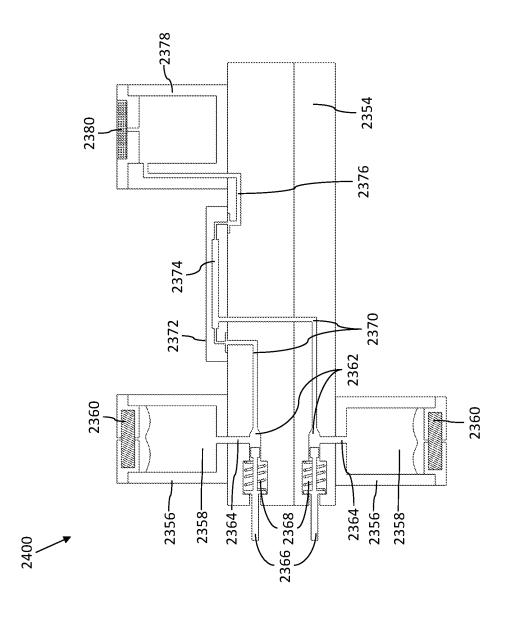
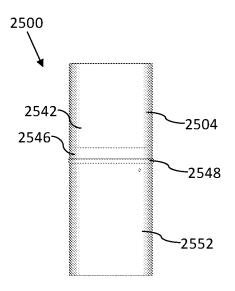


FIG. 24



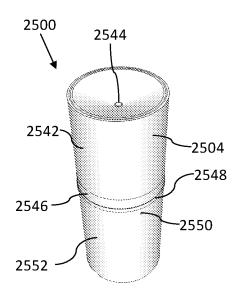


FIG. 25A

FIG. 25B

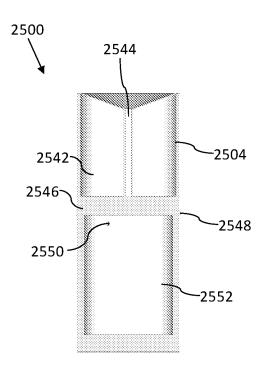


FIG. 25C

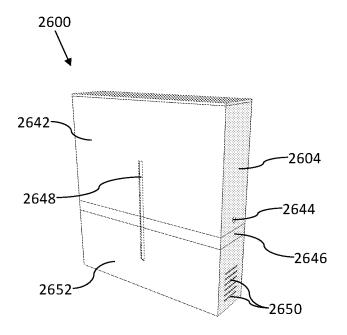


FIG. 26A

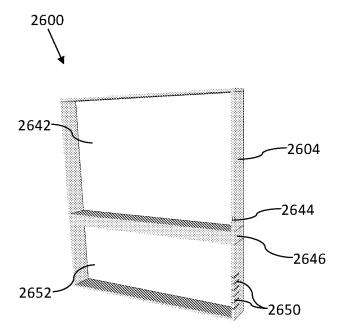


FIG. 26B

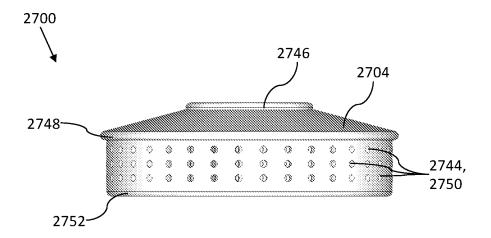


FIG. 27A

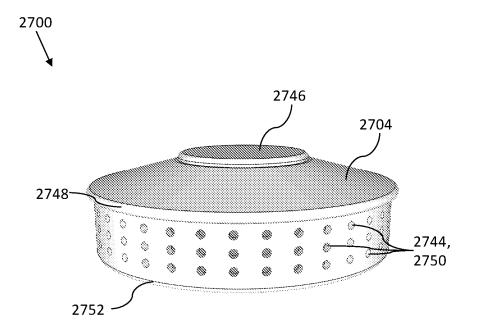


FIG. 27B

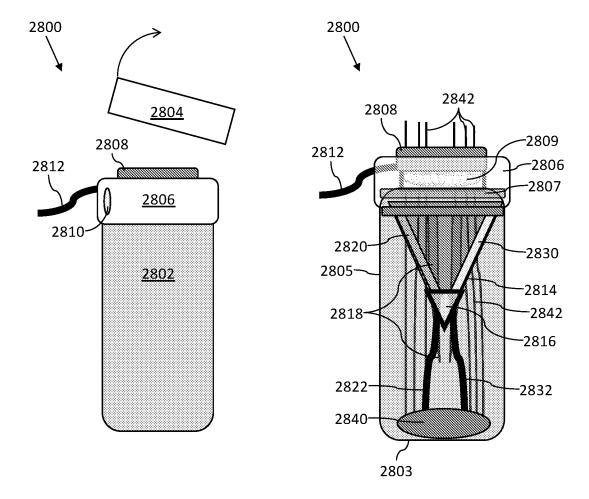


FIG. 28A

FIG. 28B

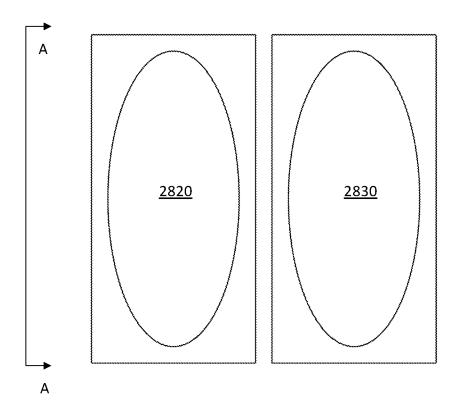


FIG. 29A

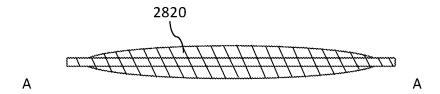


FIG. 29B

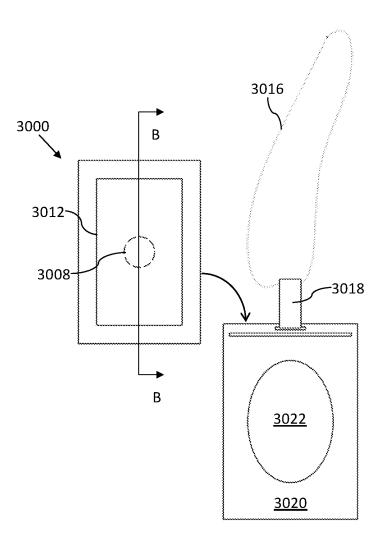
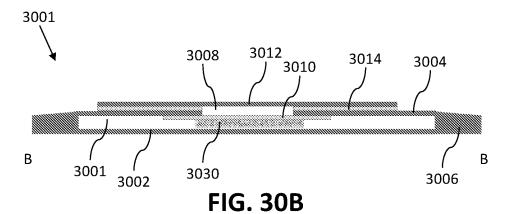


FIG. 30A



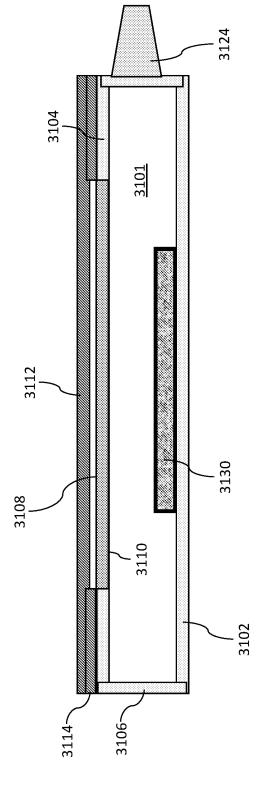
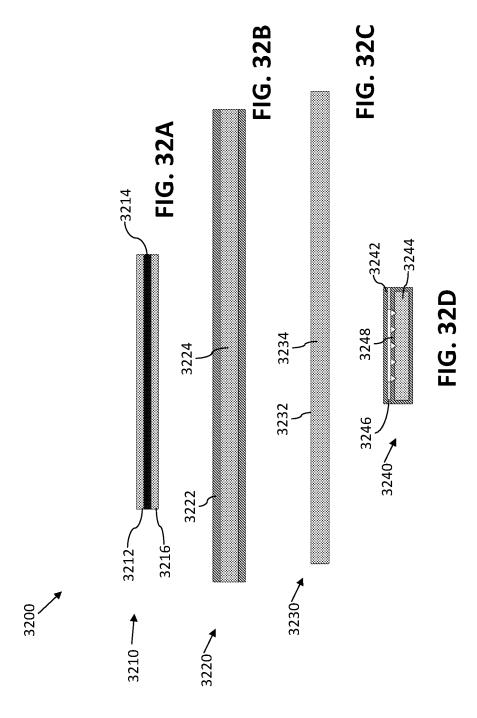
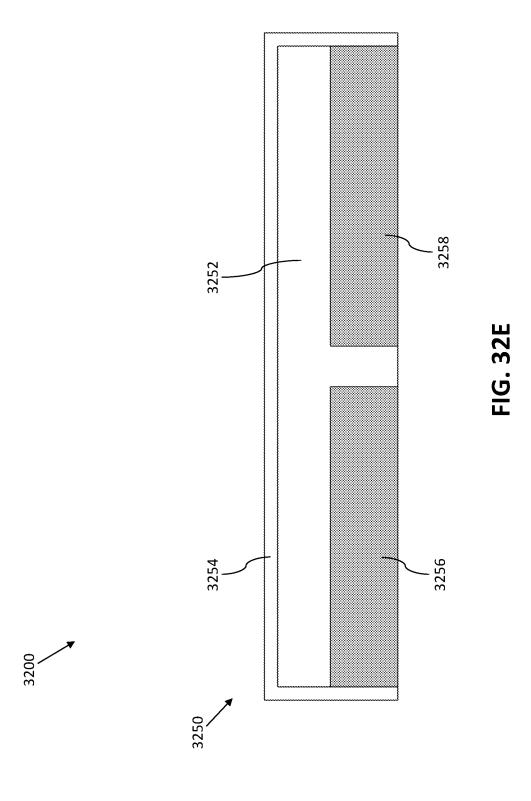


FIG. 31





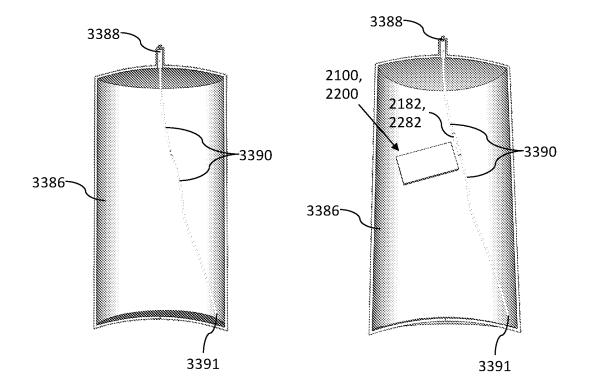


FIG. 33A

FIG. 33B

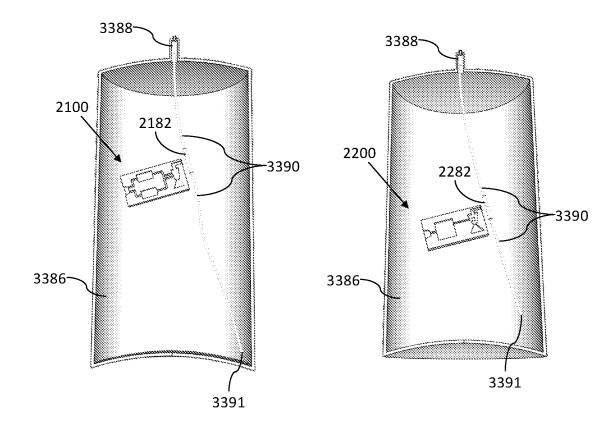


FIG. 33C

FIG. 33D

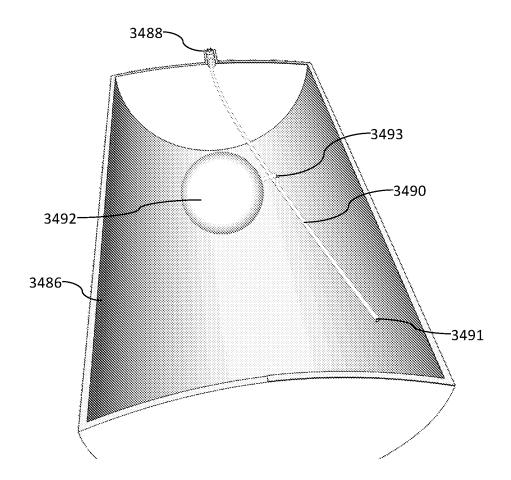


FIG. 34

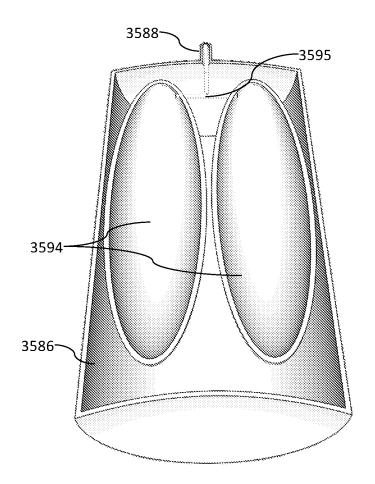


FIG. 35A

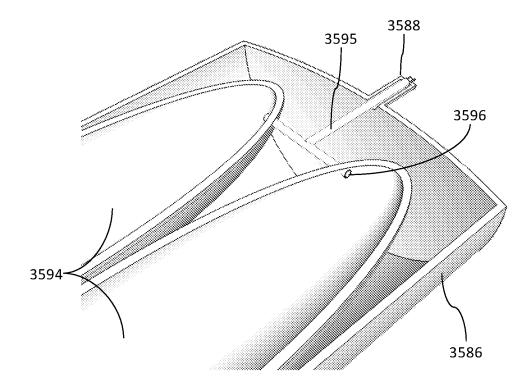
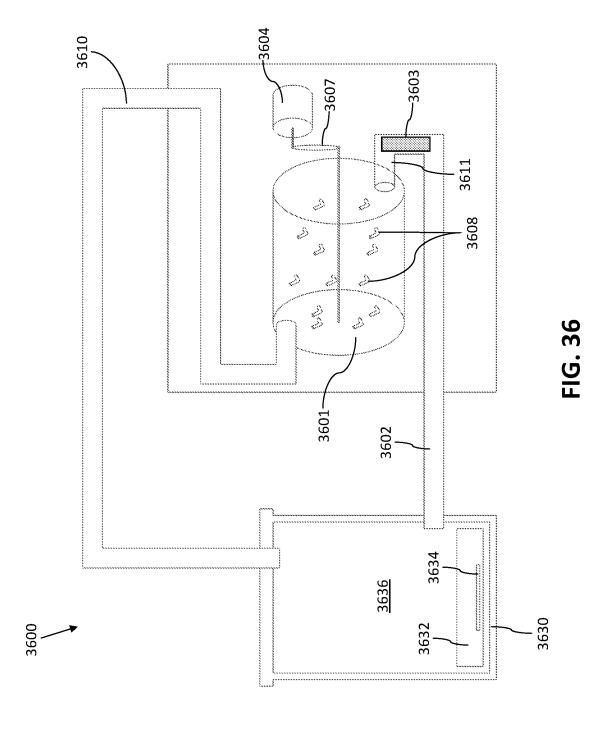
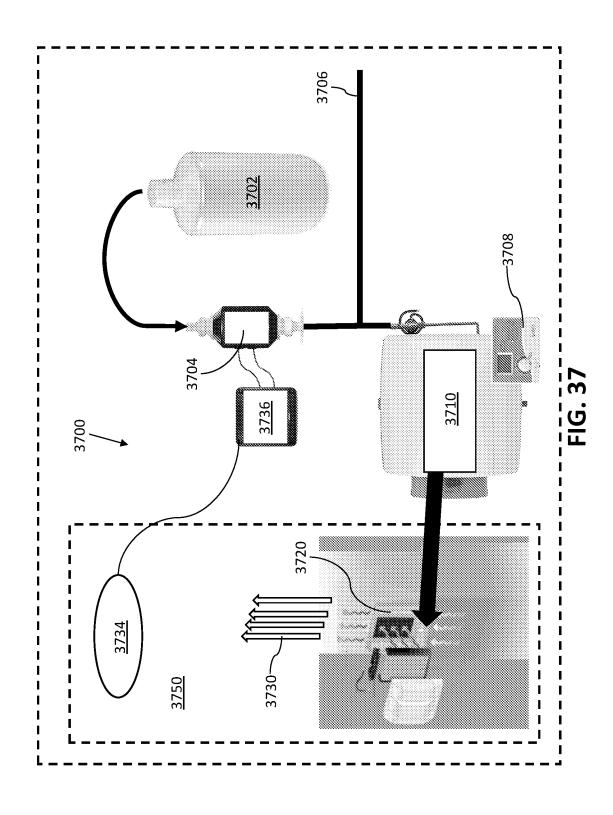


FIG. 35B





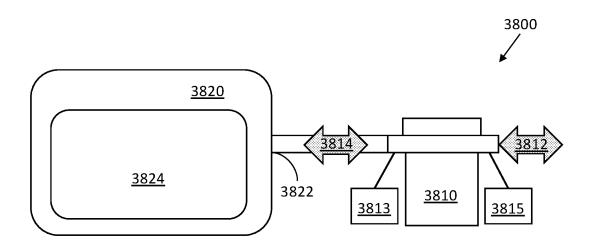


FIG. 38A

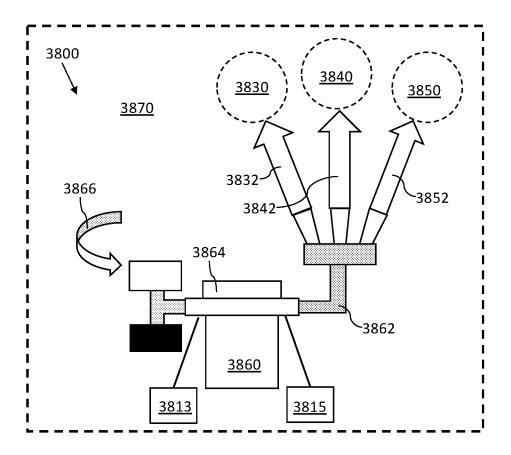


FIG. 38B

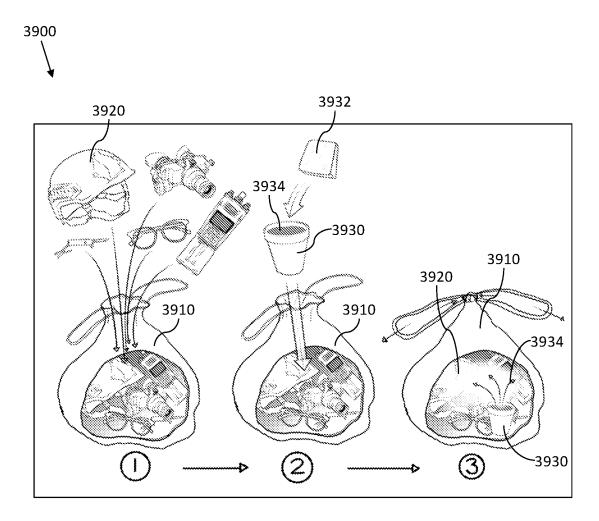


FIG. 39A FIG. 39B FIG. 39C

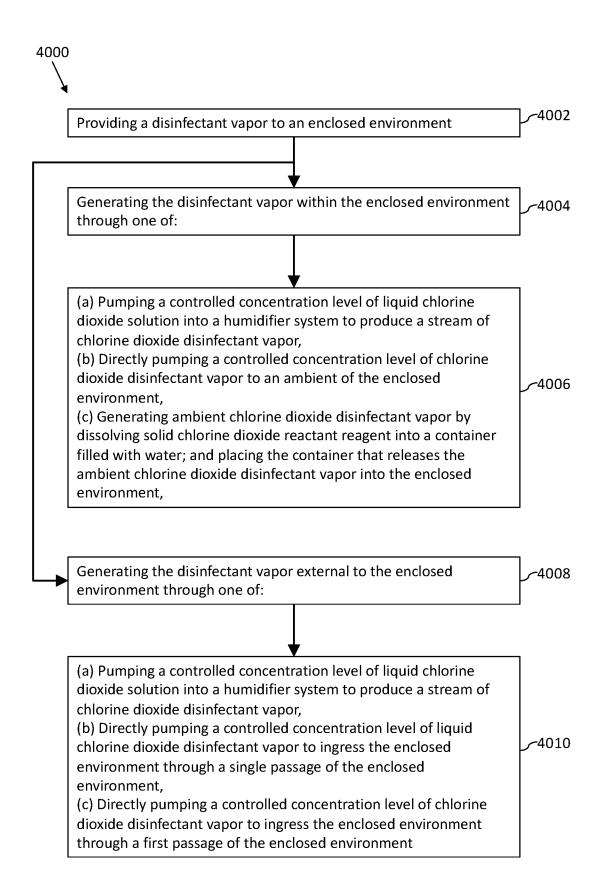


FIG. 40

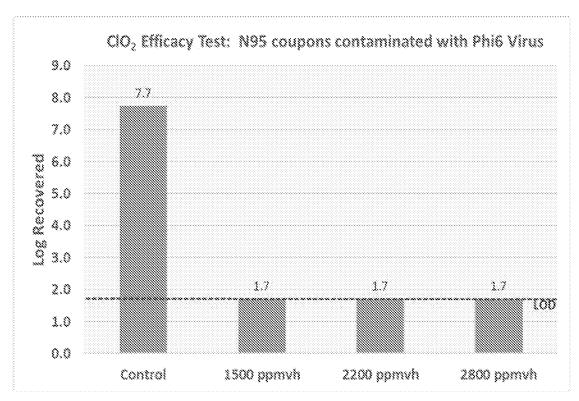


FIG. 41A

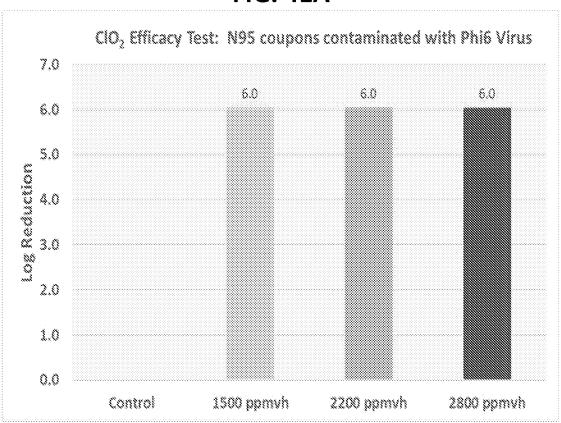


FIG. 41B



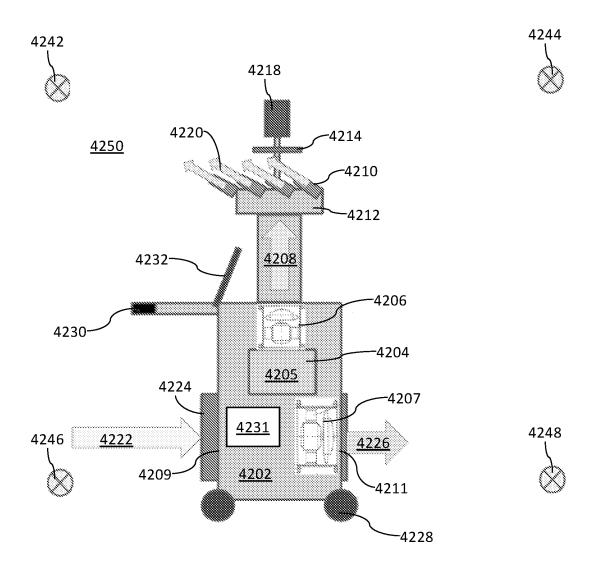


FIG. 42

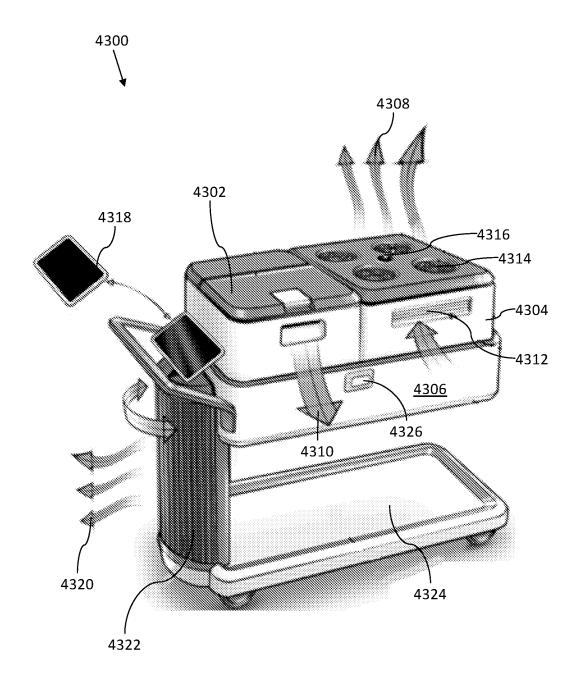


FIG. 43



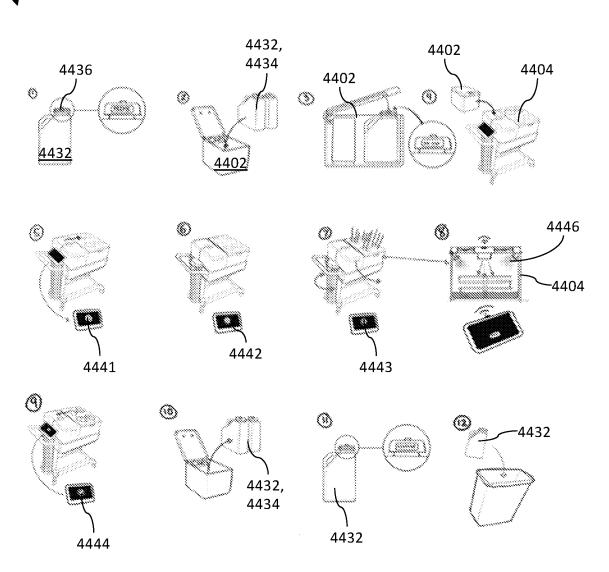


FIG. 44

Chlorine Dioxide: Prediction of Gas in Water and Air

***************************************	W(#) H(#)	7.7 7.4	18,000 Exers
	***	11.2	Vokane =

Volume of water	*#	Liters	000 mt 0002	67.451
Volume of air	18,001	Liters		
Start CICL2 consc. in HZO	305	18068		
Mass of 0.02				
produced	333	22	Total moles CICL	0.029651

şu.	78541
Gallans	

ma OKO2 m and	1975.1342	1979,7008	1963.3110	1366.1856	1988,4905	1990, 3509	1991.8624	1993,0990	1994,1141	1995,6535	1996,7269
magin3 in air	1.80	110.8	68	## E	118.5	120.8	(M)	100 100 100 100 100 100 100 100 100 100	200 000	310.8	110.5
13 20 20 81		37.19	37.93	88	833	40.08	40.78	41,48	42.17	43.55	44.92
	\$22	5.07	200. 200.	3,46	2.88	2.4	2.03	173	#. \$4.	**************************************	23
V, mol frac in cas	0.0000%4	0.0000372	0.0000379	0.0000387	0.0000394	0.0000401	0.0000408	0.0000415	0.0000422	0.0000436	0.0000449
X and frac in H20	1,658925-06	1.35426E-06	1112416-06	9.21628E-07	7.67860E-07	6.43740E-07	5.42903E-07	4.60470E-07	3,92679£-07	2.89977E-07	2.18233E-87
8	22.3983	22,8983	23,2483	23.6263	24.03%3	24,4483	24,8583	25,2683	25.6783	26.4983	27,3183
S	21.36	27.46	36.26	4:34	51.27	62.28	75.41	90 G6	107.40	150.19	205.86
Terro (C)	æ	ಚು	\$	70	R	×	×	88	40	83	88

Henry's law states that the solubility of a gas in a liquid is directly proportional to the partial pressure of the gas above the liquid. In a mixture of gases, each gas has a partial pressure which is the hypothetical pressure of that gas if it alone occupied the volume of the mixture at the same temperature. The total pressure of an ideal gas mixture is the sum of the partial pressures of each individual gas in the mixture.

a constant with the dimensions of pressure divided by concentration. The constant, known as the Henry's law constant, depends on the solute, the solvent and the partiel pressure of the solute in the gas above the solution (atm); kH temperature, (L*atm/mol), c — concentration of the solute (mol/L)

FIG 45

TEST T13 - Gas Uniformity Test

Date: 2/11/20

Furnished room w/ fan and elevated RH (80% set point

(max))

Yellow Jacket with one 126

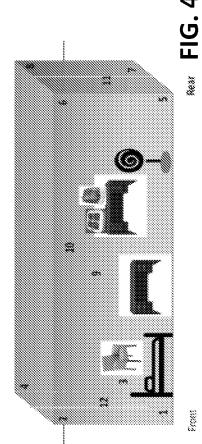
renow water mill olic 120 Portaxens datalogger inedvertantly not activated for

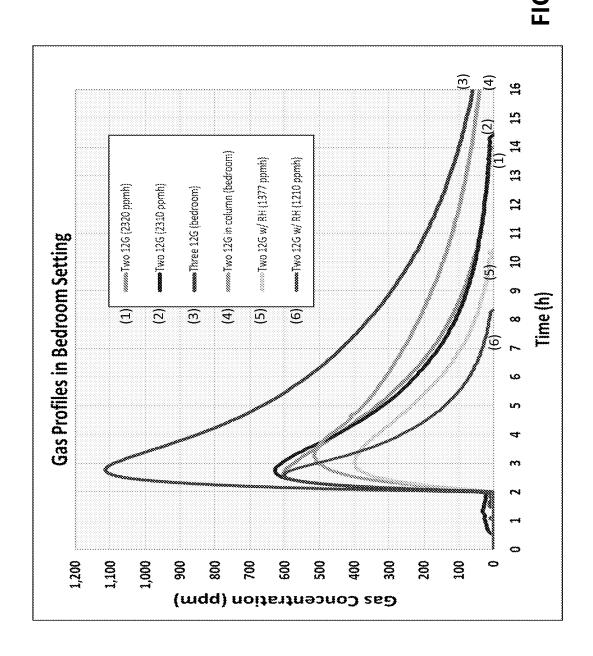
ž ž

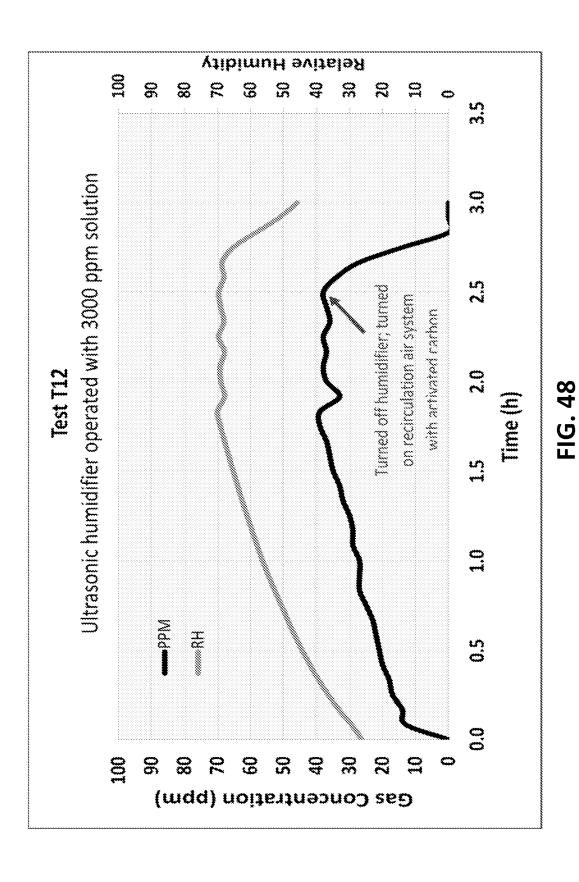
		Concentration	entration Measurements (pp.	*ents (ppm)	
Sortio					ğ
	Round	7 70008	Round 3	Mean	3600
ç.	318	186	156	385	23
3	218	136	132	185	64 86
24.5	211	321	153	130	22
~₹	310	378	151	180	23
u)	224	171	181	382	24
ğ	222	161	162	193	23
ž	194	164	145	168	2.7
82	301	170	148	273	85
ಏ	223	183	192	1365	22
22	302	173	131	178	23
23	230	184	163	192	24
*** \$74	134	180	157	133	44 \$4.5
Mean	213	821	154	281	8/4
300	۰.	٥	Œ	¢	×

When making rounds, I tried to follow the same patiern. I started with the door (port 11), moved dockwise and started at the top (port 6), then bottom (port 5), continued dockwise (port 9), and continued this way around the container. However, I did not follow this pattern perfectly (ex. In the first round, forgetting port 12 until I had done the rest).

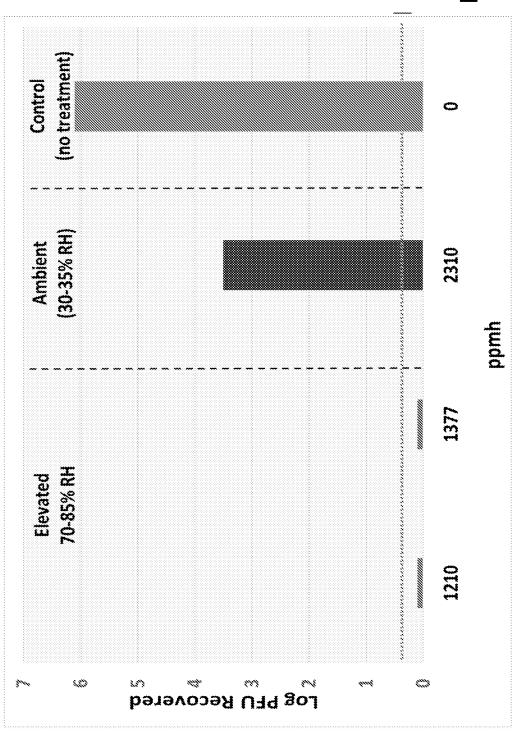
"Port"S, round 2 was observed to fluctuate between 160 and 180 ppm, never settling at a specific value for very long. This could have been caused by the position of the fan in the room, however it did not fluctuate like this in round 1 or 3.











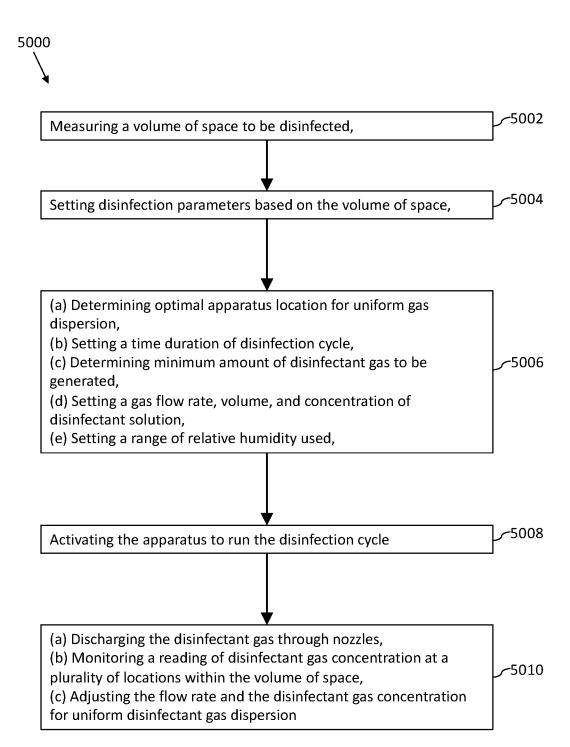


FIG. 50

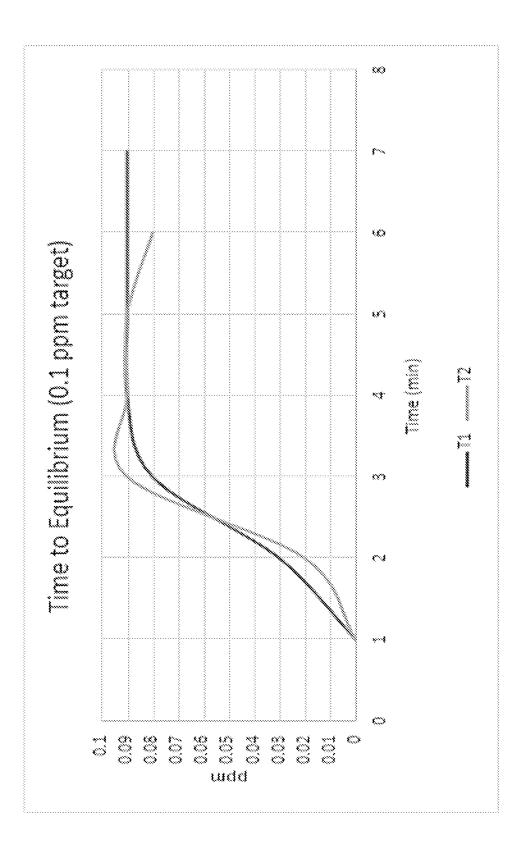


FIG. 51A

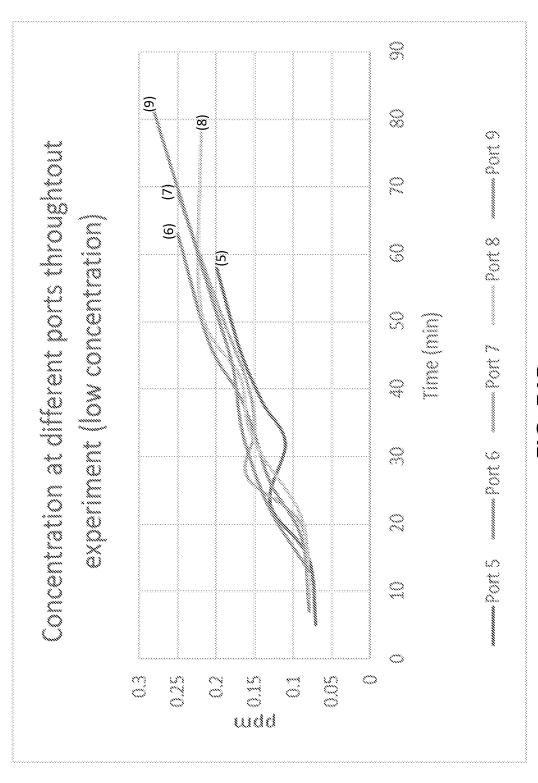


FIG. 51B

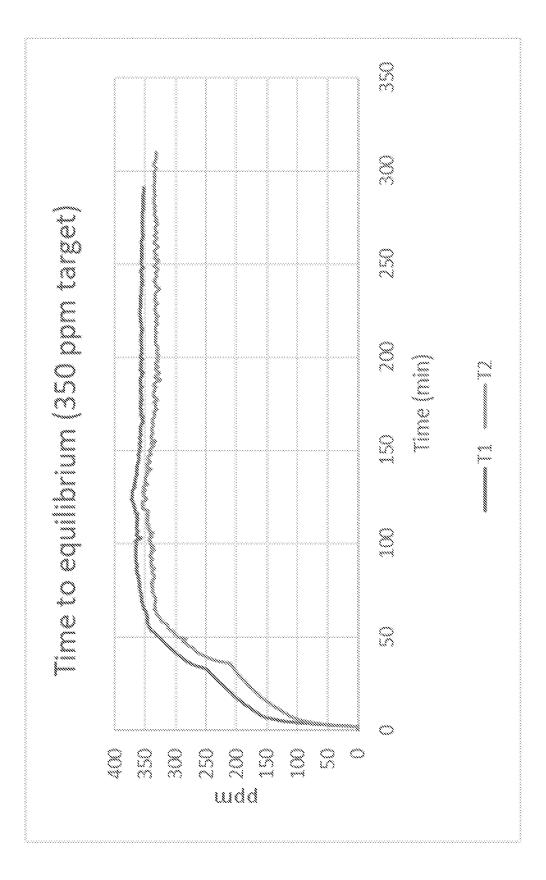


FIG. 52A

Concentration at different ports throught experiment (high concentraion)

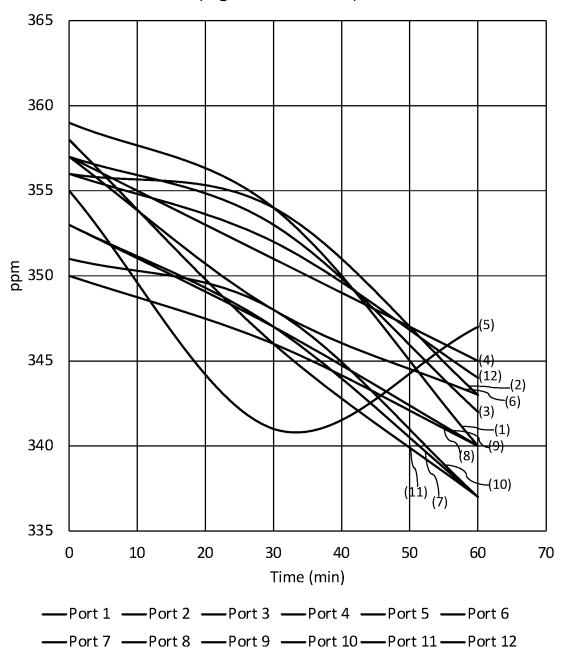


FIG. 52B

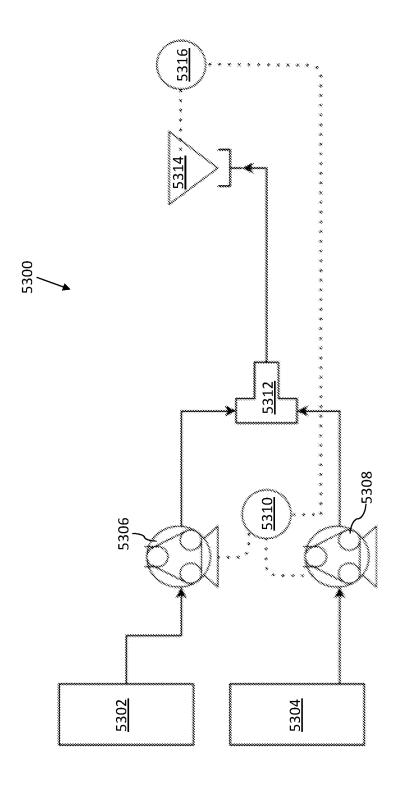


FIG. 53A

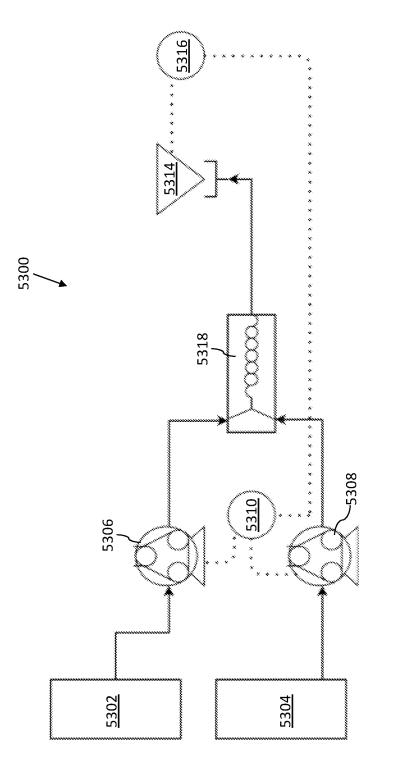


FIG. 53B

	*****		,
Measured Efficiency	103%	117%	115%
Measured (mg ClO2)	10.15	6776	11.28
Complete Conversion (mg CIO2)	9.84	88'.	9 86
CIOZ Conc. In Tote ppm	48.6	44.3	P S
Molar Ratio [NaClO2: Activator]	2	3	9
Volume Activator ul	139	36	33
Activator Conc.	0.5 g/mL	N 21	%001
Activator	22 Na2S208	22 HCI	22 CH202
Volume NaCiO2 Solution uf			
NaClO2 Solution (80%) g/ml	0.75	0.75	0.75
Acid Generation	***	2	3

FIG. 54A

	356	26	38
Measured Efficiency	왕	55	떲
de sured Efficiency			
2 5			
CO2 Measured (mg)	81	5.00	9.00
-	23	#3	ಯ
2			
33			

8			
~			
	24.99	16.50	16.50
COZ (mg) Generated	25	₩	33
- B			
8			
*	=======================================	523	4
302 Conc. In Tate ppm			
H B			
.5 =			
≅			
8			
		- SQ	2
Energy (I)	E	425	v2S
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22			
	ΥĢ	<u>``</u>	C
Power (W)	ນ		00
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25			
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Current [A] Time On (min)			
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3 🙀			

	0.25	0.50	0.50
# OF	C	c i	ော်
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* 8			
¥ 5			
Analyte: Nat102 NaNO3 Solution Solution (80%) g/ml. g/ml			
~ 8			
Electrolysis Generation Cell Type	٠	ایرا	No Membrane
	Membrane	Membrane	-22
-	<u> </u>	يظ	a a
-	<u>5</u>	<u></u>	-co
- X E	,		~
一带带			
llectrolysis Seneration			
3 3			
	- 1	1	

FIG. 54B

Chemica	Use - 100	Chemical Use - 1000 ft3 room			3 7	Single Dose - 0.10 ppm in 1000 ft3	pm in 1000 ft.		1000 ft3 30 days continuous operation 5 air exchanges per hr 250% makeup air	1000 ft3 30 days continuous operation 5 air exchanges per hr 100% makeup air
Generation Method	Activator	Molar Ratio	Volume Ratio	Efficiency	NaClO2 Activator Volume [ul] Volume [ul]		otal Volume (u)	Total Volume (mi)	Total Volume [mi]	Total Volume (mil)
	Na2S208	7	6.32	100%	17.46				114.96	459.82
2	P	8	1.64	100%	17.46	28.95	46.41	90'0	41.77	167.08
m	CH202	9	1.50	100%	17.46	26.22	43.68	0.04	39.31	157.23
4	Electrolysis	N/A	N/A	25%	31.74	None	31.74	0.03	78.57	114.27
5	Membrane	N/A	N/A	100%	17.46	None	17.46	70'0	15.71	. 62.85

FIG. 54C

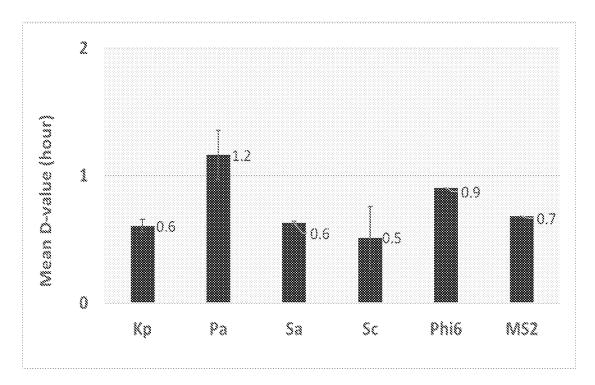


FIG. 55A

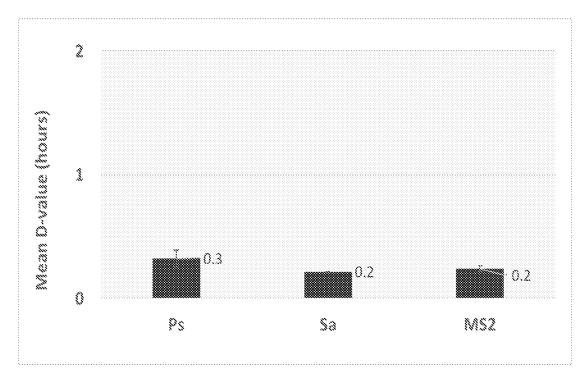


FIG. 55B

Form PCT/ISA/210 (second sheet) (July 2019)

International application No. PCT/US2021/036501

A. CLASSIFICATION OF SUBJECT MATTER IPC(8) - A61B 5/00; A61B 5/01; A61B 5/145; A61B 5/0CPC - C11D 3/48; A61L 2202/25; A61B 5/00; A61E)			
According to International Patent Classification (IPC) or to both	national classification and IPC				
B. FIELDS SEARCHED					
Minimum documentation searched (classification system followed by see Search History document	y classification symbols)				
Documentation searched other than minimum documentation to the e see Search History document	xtent that such documents are included in the	fields searched			
Electronic data base consulted during the international search (name of	of data base and, where practicable, search te	rms used)			
see Search History document	,, ,				
C. DOCUMENTS CONSIDERED TO BE RELEVANT					
Category* Citation of document, with indication, where app	propriate, of the relevant passages	Relevant to claim No.			
X US 2018/0271450 A1 (DEXCOM INC.) 27 September	r 2018 (27.09.2018) entire document	1-3			
 Y		18, 21, 22			
 A		 4-14, 32, 34			
X US 2006/0051285 A1 (HAWKER et al) 09 March 200	6 (09.03.2006) entire document	15, 16			
 Y					
A		4-14, 20, 23, 24			
Y US 2005/0233198 A1 (NUZZO et al) 20 October 2005	5 (20.10.2005) entire document	17, 22, 31, 33			
 A		32, 34			
Y US 2016/0251219 A1 (CHEMTREAT INC.) 01 Septer	mber 2016 (01.09.2016) entire document	19			
 A		20			
Y US 4,852,613 A (TIPPETTS et al) 01 August 1989 (01	1.08.1989) entire document	25			
Y US 2016/0110657 A1 (SKYTREE INC.) 21 April 2016	(21.04.2016) entire document	26-31, 33			
 A		32, 34			
Further documents are listed in the continuation of Box C.	See patent family annex.				
* Special categories of cited documents: "A" document defining the general state of the art which is not considered date and not in conflict with the application but cited to understand					
to be of particular relevance the principle or theory underlying the invention "D" document cited by the applicant in the international application "E" earlier application or patent but published on or after the international considered novel or cannot be considered to involve an inventive step					
fling date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Entire application or patient but published on or after the international when the document is taken alone when the document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination					
"O" document referring to an oral disclosure, use, exhibition or other means document published prior to the international filing date but later than the priority date claimed					
Date of the actual completion of the international search	Date of mailing of the international search	ch report			
13 October 2021	NOV 18 2021				
Name and mailing address of the ISA/US	Authorized officer				
Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, VA 22313-1450	Harry Kim				
Facsimile No. 571-273-8300	Telephone No. PCT Helpdesk: 571-27	72-4300			

International application No.
PCT/US2021/036501

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
Y 	US 2012/0121731 A1 (PETERS et al) 17 May 2012 (17.05.2012) entire document	31, 33 9-14, 24, 32, 34
Α	US 2008/0167650 A1 (JOSHI et al) 10 July 2008 (10.07.2008) entire document	23

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Box No. II	Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)
This intern	ational search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
t t	Claims Nos.: Decause they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.	Claims Nos.: secause they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box No. II	I Observations where unity of invention is lacking (Continuation of item 3 of first sheet)
	ational Searching Authority found multiple inventions in this international application, as follows:
See extra s	sileet(s).
1. A	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable laims.
	As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of dditional fees.
	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. N	To required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on	The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee. The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation. No protest accompanied the payment of additional search fees.

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Continued from Box No. III Observations where unity of invention is lacking

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claims 1-14, is drawn to a system for generating and monitoring an antimicrobial, comprising: a computational system.

Group II, claims 15-34, is drawn to a system for generating and monitoring Cl02, comprising: a device housing including an inlet

The inventions listed as Groups I-II do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: the special technical feature of the Group I invention: a computational system; an antimicrobial sensor; and an antimicrobial generator, wherein the computational system, the antimicrobial generator, and the antimicrobial sensor are operatively connected as claimed therein is not present in the invention of Group II. The special technical feature of the Group II invention: a device housing including an inlet; a microcontroller or microprocessor; a reagent containing a reagent; a device for generating a Cl02 from the reagent as claimed therein is not present in the invention of Group I.

Groups I and II lack unity of invention because even though the inventions of these groups require the technical feature of a system for generating and monitoring a material, comprising a sensing system; a communication device, this technical feature is not a special technical feature as it does not make a contribution over the prior art.

Specifically, US 2018/0271450 to DexCom, Inc. teaches a system for generating and monitoring a material, comprising a sensing system; a communication device (Paras. [0236], [0379-0380], [0394], [0412]).

Since none of the special technical features of the Group I or II inventions are found in more than one of the inventions, unity of invention is lacking.