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(54) Title: ENGINEERING PLANT GENOMES USING CRISPR/CAS SYSTEMS

(57) Abstract: Materials and methods for gene targeting using Clustered Regularly Interspersed Short Palindromic Repeats/CRISPR-associated (CRISPR/Cas) systems are provided herein.

# ENGINEERING PLANT GENOMES USING CRISPR/Cas SYSTEMS

## CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims benefit of priority from U.S. Provisional Application Serial No. 61/790,694, filed on March 15, 2013.

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## STATEMENT AS TO FEDERALLY SPONSORED RESEARCH

This invention was made with government support under GM 834720 awarded by the National Institutes of Health, and DBI0923827 awarded by the National Science Foundation. The government has certain rights in the invention.

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## TECHNICAL FIELD

This document relates to materials and methods for gene targeting in plants, and particularly to methods for gene targeting that include using CRISPR/Cas systems.

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## BACKGROUND

Technologies enabling the precise modification of DNA sequences within living cells can be valuable for both basic and applied research. Precise genome modification – either targeted mutagenesis or gene targeting (GT) – relies on the DNA-repair machinery of the target cell. With respect to targeted mutagenesis, sequence-specific nuclease (SSN)-mediated DNA double-strand breaks (DSBs) are frequently repaired by the error-prone non-homologous end joining (NHEJ) pathway, resulting in mutations at the break site. On the other hand, if a donor molecule is co-delivered with a SSN, the ensuing DSB can stimulate homologous recombination (HR) of sequences near the break site with sequences present on the donor molecule. Consequently, any modified sequence carried by the donor molecule will be stably incorporated into the genome (referred to as GT). Attempts to implement GT in plants often are plagued by extremely low HR frequencies. The majority of the time, donor DNA molecules integrate illegitimately via NHEJ. This process occurs regardless of the size of the homologous “arms;” increasing the length of homology to approximately 22 kb results in no significant enhancement in GT (Thykjaer

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et al., *Plant Mol Biol*, 35:523-530, 1997). However, introducing a DSB with a SSN can greatly increase the frequency of GT by HR (Shukla et al., *Nature* 459:437-441, 2009; and Townsend et al., *Nature* 459:442-445, 2009).

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## SUMMARY

This document is based in part on the discovery that the Clustered Regularly Interspersed Short Palindromic Repeats/CRISPR-associated (CRISPR/Cas) system can be used for plant genome engineering. The CRISPR/Cas system provides a relatively simple, effective tool for generating modifications in genomic DNA at selected sites. CRISPR/Cas systems can be used to create targeted DSBs or single-strand breaks, and can be used for, without limitation, targeted mutagenesis, gene targeting, gene replacement, targeted deletions, targeted inversions, targeted translocations, targeted insertions, and multiplexed genome modification through multiple DSBs in a single cell directed by co-expression of multiple targeting RNAs. This technology can be used to accelerate the rate of functional genetic studies in plants, and to engineer plants with improved characteristics, including enhanced nutritional quality, increased resistance to disease and stress, and heightened production of commercially valuable compounds.

In one aspect, this document features a method for modifying the genomic material in a plant cell. The method can include (a) introducing into the cell a nucleic acid comprising a crRNA and a tracrRNA, or a chimeric cr/tracrRNA hybrid, where the crRNA and tracrRNA, or the cr/tracrRNA hybrid, is targeted to a sequence that is endogenous to the plant cell; and (b) introducing into the cell a Cas9 endonuclease molecule that induces a double strand break at or near the sequence to which the crRNA and tracrRNA sequence is targeted, or at or near the sequence to which the cr/tracrRNA hybrid is targeted. The introducing steps can include delivering to the plant cell a nucleic acid encoding the Cas9 endonuclease and a nucleic acid encoding the crRNA and tracrRNA or the cr/tracrRNA hybrid, where the delivering is by a DNA virus (e.g., a geminivirus) or an RNA virus (e.g., a tobnavirus). The introducing steps can include delivering to the plant cell a T-DNA containing a nucleic acid sequence encoding the Cas9 endonuclease and a nucleic acid sequence encoding the crRNA and tracrRNA or the

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cr/tracrRNA hybrid, where the delivering is via *Agrobacterium* or *Ensifer*. The nucleic acid sequence encoding the Cas9 endonuclease can be operably linked to a promoter that is constitutive (e.g., a cauliflower mosaic virus 35S promoter), cell specific, inducible, or activated by alternative splicing of a suicide exon. The introducing steps can include  
5 microprojectile bombardment of nucleic acid encoding Cas9 and the crRNA and tracrRNA or the cr/tracrRNA hybrid. The nucleic acid sequence encoding the Cas9 endonuclease can be operably linked to a promoter that is constitutive, cell specific, inducible, or activated by alternative splicing of a suicide exon. The plant cell can be from a monocotyledonous plant (e.g., wheat, maize, rice, or *Setaria*), or from a  
10 dicotyledonous plant (e.g., tomato, soybean, tobacco, potato, cassava, or *Arabidopsis*). The method can further include screening the plant cell after the introducing steps to determine if a double strand break has occurred at or near the sequence targeted by the crRNA and tracrRNA or the cr/tracrRNA hybrid. The method also can include regenerating a plant from the plant cell, and in some embodiments, the method can  
15 include cross breeding the plant to obtain a genetically desired plant lineage.

In another aspect, this document features a plant cell containing a nucleic acid encoding a polypeptide having at least 80% sequence identity with SEQ ID NO:12, as well as a plant cell containing a nucleic acid encoding a polypeptide that includes an amino acid sequence having at least 80% sequence identity with amino acids 810 to 872  
20 of SEQ ID NO:12.

In another aspect, this document features a virus vector containing a nucleotide sequence that encodes a Cas9 polypeptide. The virus vector can contain a nucleotide sequence encoding a polypeptide with an amino acid sequence having at least 90% identity to SEQ ID NO:12. The virus vector can be from a tobnavirus or a geminivirus.  
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In another aspect, this document features a T-DNA containing a nucleic acid sequence encoding a polypeptide that has an amino acid sequence having at least 80% sequence identity with amino acids 810 to 872 of SEQ ID NO:12. This document also features an *Agrobacterium* strain containing the T-DNA.

In yet another aspect, this document features a method for expressing a Cas  
30 protein in a plant cell. The method can include providing an *Agrobacterium* or *Ensifer*

vector containing a T-DNA that includes a nucleic acid sequence encoding a polypeptide having an amino acid sequence with at least 80% sequence identity to amino acids 810 to 872 of SEQ ID NO:12, where the polypeptide-encoding sequence is operably linked to a promoter; bringing the *Agrobacterium* or *Ensifer* vector into contact with the plant cell;  
5 and expressing the nucleic acid sequence in the plant cell. The promoter can be an inducible promoter (e.g., an estrogen inducible promoter). The method can further include contacting the plant cell with a nucleic acid encoding a guide RNA that associates with the Cas protein. The plant cell can be a protoplast.

Unless otherwise defined, all technical and scientific terms used herein have the  
10 same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used to practice the invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the  
15 present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and from the claims.  
20

### DESCRIPTION OF DRAWINGS

FIG. 1 is a schematic of a pMDC32 plasmid (a standard T-DNA expression plasmid) containing a Cas9 coding sequence and a cr/tracrRNA hybrid sequence. The nucleotide sequence of the plasmid is set forth in SEQ ID NO:6.

25 FIG. 2 is a schematic of a pFZ19 plasmid (an estrogen-inducible T-DNA expression vector) containing a Cas9 coding sequence and a cr/tracrRNA hybrid sequence. The nucleotide sequence of the plasmid is set forth in SEQ ID NO:7.

FIG. 3 is a schematic of a pNJB121 plasmid (a geminivirus-replicon T-DNA vector) containing a Cas9 coding sequence and a cr/tracrRNA hybrid. The nucleotide  
30 sequence of the plasmid is set forth in SEQ ID NO:8.

FIGS. 4A-4D provide evidence of CRISPR/Cas function in plant cells in which a Cas9 coding sequence and a cr/tracrRNA hybrid were delivered by *Agrobacterium* or geminivirus replicons. FIG. 4A is an illustration of a T-DNA harboring a plant codon-optimized Cas9 sequence. The cr/tracrRNA hybrid (designated sgRNA) was placed downstream of the *Arabidopsis* AtU6-26 promoter (PU6). The “lollypops” indicate the long intergenic region (LIR) that is important for replication mediated by replicase (Rep). The gray box represents the short intergenic region (SIR) that also is important for replicon function. The unlabeled gray arrow is a 35S promoter that can drive Cas9 expression upon circularization of the replicon. Cas9 expression also can be driven by the LIR, which functions as a promoter. The entire construct depicted is referred to as an LSL T-DNA. FIG. 4B is a picture of an agarose gel containing PCR products, demonstrating circularization of the geminivirus replicon in plant cells. PCR primers (small arrows in FIG. 4A) were used to amplify DNA from cells infected with *Agrobacterium* T-DNA carrying the replicon. Only in the presence of the plasmid encoding the geminivirus replicase (pRep) did circularization and amplification of the replicon occur. FIG. 4C shows detection of Cas9-induced mutations at the *Nicotiana tabacum* *SurA/SurB* loci. Tobacco leaf tissue was syringe infiltrated with two strains of *Agrobacterium* containing pREP and the LSL T-DNA depicted in FIG. 4A; this was done to test for CRISPR/Cas9-mediated mutagenesis using geminivirus replicons. Alternatively, leaf tissue was infiltrated with single strain of *Agrobacterium* containing only the LSL T-DNA; this was done to test for CRISPR/Cas9-mediated mutagenesis by standard *Agrobacterium* T-DNA delivery. Five days post infiltration, genomic DNA was isolated and used as a template in a PCR reaction designed to amplify the Cas9 target site within *SurA/SurB*. The resulting amplicons were digested with AlwI, and bands were separated by gel electrophoresis. FIG. 4D shows sequences (SEQ ID NOS:1-5) that resulted from cleavage-resistant amplicons in the sample transformed with the LSL T-DNA and pREP T-DNA. PAM, protospacer adjacent motif.

FIG. 5 is a schematic of a reporter plasmid encoding a non-functional yellow fluorescent protein (YFP).

FIG. 6 is a graph plotting fluorescence levels as evidence of CRISPR/Cas function in protoplasts using a YFP reporter plasmid. Tobacco protoplasts were prepared and transformed with various constructs to test for targeted cleavage by CRISPR/Cas9, and YFP fluorescence was measured by flow cytometry. Column 1 shows levels of fluorescence observed from cells transformed with the YFP reporter and constructs expressing Cas9 and the cr/tracr RNA expressed from the AtU6-26 promoter. Column 2 shows levels of fluorescence observed from cells transformed with the reporter, Cas9 and the cr/tracr RNA expressed from the At7SL2-2 promoter. Column 3 shows fluorescence observed in cells transformed with the reporter only (negative control); column 4 shows fluorescence in cells transformed with a construct that expresses YFP (positive control).

#### DETAILED DESCRIPTION

Efficient genome engineering in plants can be enabled by introducing targeted double-strand breaks (DSBs) in a DNA sequence to be modified. The DSBs activate cellular DNA repair pathways, which can be harnessed to achieve desired DNA sequence modifications near the break site. Targeted DSBs can be introduced using sequence-specific nucleases (SSNs), a specialized class of proteins that includes transcription activator-liked (TAL) effector endonucleases, zinc-finger nucleases (ZFNs), and homing endonucleases (HEs). Recognition of a specific DNA sequence is achieved through interaction with specific amino acids encoded by the SSNs. Prior to the development of TAL effector endonucleases, a challenge of engineering SSNs was the unpredictable context dependencies between amino acids that bind to DNA sequence. While TAL effector endonucleases greatly alleviated this difficulty, their large size (on average, each TAL effector endonuclease monomer contains 2.5-3 kb of coding sequence) and repetitive nature may hinder their use in applications where vector size and stability is a concern (Voytas, *Annu Rev Plant Biol*, 64: 327-350, 2013).

This document is based in part on the discovery that the CRISPR/Cas system can be used as a simple, effective tool for plant genome engineering. CRISPR/Cas molecules are components of a prokaryotic adaptive immune system that uses RNA base pairing to direct DNA cleavage. Directing DNA DSBs requires two components: the Cas9 protein,



which functions as an endonuclease, and CRISPR RNA (crRNA) and tracrRNA (tracrRNA) sequences that aid in directing the Cas9/RNA complex to target DNA sequence (Makarova et al., *Nat Rev Microbiol*, 9(6):467-477, 2011). The modification of a single targeting RNA can be sufficient to alter the nucleotide target of a Cas protein. In  
5 some cases, crRNA and tracrRNA can be engineered as a single cr/tracrRNA hybrid to direct Cas9 cleavage activity (Jinek et al., *Science*, 337(6096):816-821, 2012). The CRISPR/Cas system can be used in bacteria, yeast, humans, and zebrafish, as described elsewhere (see, e.g., Jiang et al., *Nat Biotechnol*, 31(3):233-239, 2013; Dicarlo et al.,  
*Nucleic Acids Res*, doi:10.1093/nar/gkt135, 2013; Cong et al., *Science*, 339(6121):819-  
10 823, 2013; Mali et al., *Science*, 339(6121):823-826, 2013; Cho et al., *Nat Biotechnol*, 31(3):230-232, 2013; and Hwang et al., *Nat Biotechnol*, 31(3):227-229, 2013).

The utility of the CRISPR/Cas system in plants has not previously been demonstrated. The CRISPR/Cas system originates from prokaryotic cells with relatively small genomes, in which Cas9 is stably expressed in cells in the presence of significant  
15 RNase III activity. Thus, when the plant cell work described herein was initiated, there was uncertainty as to whether expression of a Cas9 transgene would be possible in plant cells, and whether Cas9 would properly cooperate with RNA-guides and RNase III activity in the plant context. In addition, expression of heterologous proteins in plant cells is generally challenging due to different codon usage. Further, some toxicity from  
20 Cas9 expression in plants was expected, as the large size of plant genomes increases the probability that nonspecific cleavage of genomic DNA may induce genotoxicity to the cells. The CRISPR/Cas9 system is reported to operate with specific recognition sequences comprising 10-20 nucleotides, which is less specific than most other rare-cutting endonuclease systems such as TAL effector endonucleases, meganucleases, and  
25 zinc finger nucleases.

As described herein, CRISPR/Cas systems can be used to create targeted DSBs or single-strand breaks, and can be used for, without limitation, targeted mutagenesis, gene targeting, gene replacement, targeted deletions, targeted inversions, targeted translocations, targeted insertions, and multiplexed genome modification through  
30 multiple DSBs in a single cell directed by co-expression of multiple targeting RNAs. This

technology can be used to accelerate the rate of functional genetic studies in plants, and to engineer plants with improved characteristics, including enhanced nutritional quality, increased resistance to disease and stress, and heightened production of commercially valuable compounds. Proof-of-concept experiments can be performed in plant leaf tissue  
5 by targeting DSBs to reporter genes and endogenous loci. The technology then can be adapted for use in protoplasts and whole plants, and in viral-based delivery systems. Finally, multiplex genome engineering can be demonstrated by targeting DSBs to multiple sites within the same genome.

In general, the systems and methods described herein include at least two  
10 components: the RNAs (crRNA and tracrRNA, or a single cr/tracrRNA hybrid) complementary (and thus targeted) to a particular sequence in a plant cell (e.g., in a plant genome, or in an extrachromosomal plasmid, such as a reporter), and a Cas9 endonuclease that can cleave the plant DNA at the target sequence. A representative Cas9 coding sequence is shown in nucleotides 9771 to 14045 of SEQ ID NO:6 (also  
15 nucleotides 4331 to 8605 of SEQ ID NO:7, and nucleotides 9487 to 13761 of SEQ ID NO:8). In some cases, a system also can include a nucleic acid containing a donor sequence targeted to a plant sequence. The endonuclease can to create targeted DNA double-strand breaks at the desired locus (or loci), and the plant cell can repair the double-strand break using the donor DNA sequence, thereby incorporating the  
20 modification stably into the plant genome.

The Cas9 protein includes two distinct active sites – a RuvC-like nuclease domain and a HNH-like nuclease domain, which generate site-specific nicks on opposite DNA strands (Gasiunas et al., *Proc Natl Acad Sci USA* 109(39):E2579-E2586, 2012). The RuvC-like domain is near the amino terminus of the Cas9 protein and is thought to cleave  
25 the target DNA noncomplementary to the crRNA, while the HNH-like domain is in the middle of the protein and is thought to cleave the target DNA complementary to the crRNA. A representative Cas9 sequence from *Streptococcus thermophilus* is set forth in SEQ ID NO:11 (*see, also, UniProtKB number Q03JI6*), and a representative Cas9 sequence from *S. pyogenese* is set forth in SEQ ID NO:12 (*see, also, UniProtKB number*  
30 *Q99ZW2*). Thus, the methods described herein can be carried out using a nucleotide

sequence encoding a Cas9 polypeptide having the sequence of SEQ ID NO:11 or SEQ ID NO:12. In some embodiments, however, the methods described herein can be carried out using a nucleotide sequence encoding a Cas9 functional variant having at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) sequence identity with SEQ ID NO:11 or SEQ ID NO:12. Further, Cas9 can be split into two portions, with one portion including the HNH domain and the other including the RuvC domain. The HNH domain may have some cleavage activity by itself in association with the RNA-guide, so this document also contemplates the use of Cas9 polypeptides containing an HNH domain with at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) sequence identity with the HNH domain within SEQ ID NO:11 (e.g., amino acids 828 to 879 of SEQ ID NO:11) or SEQ ID NO:12 (e.g., amino acids 810 to 872 of SEQ ID NO:12).

The percent sequence identity between a particular nucleic acid or amino acid sequence and a sequence referenced by a particular sequence identification number is determined as follows. First, a nucleic acid or amino acid sequence is compared to the sequence set forth in a particular sequence identification number using the BLAST 2 Sequences (Bl2seq) program from the stand-alone version of BLASTZ containing BLASTN version 2.0.14 and BLASTP version 2.0.14. This stand-alone version of BLASTZ can be obtained online at [fr.com/blast](http://fr.com/blast) or at [ncbi.nlm.nih.gov](http://ncbi.nlm.nih.gov). Instructions explaining how to use the Bl2seq program can be found in the readme file accompanying BLASTZ. Bl2seq performs a comparison between two sequences using either the BLASTN or BLASTP algorithm. BLASTN is used to compare nucleic acid sequences, while BLASTP is used to compare amino acid sequences. To compare two nucleic acid sequences, the options are set as follows: -i is set to a file containing the first nucleic acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second nucleic acid sequence to be compared (e.g., C:\seq2.txt); -p is set to blastn; -o is set to any desired file name (e.g., C:\output.txt); -q is set to -1; -r is set to 2; and all other options are left at their default setting. For example, the following command can be used to generate an output file containing a comparison between two sequences: C:\Bl2seq -i c:\seq1.txt -j c:\seq2.txt -p blastn -o c:\output.txt -q -1 -r 2. To compare two amino acid sequences, the options of Bl2seq are set as follows: -i is set to a file containing the first

amino acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second amino acid sequence to be compared (e.g., C:\seq2.txt); -p is set to blastp; -o is set to any desired file name (e.g., C:\output.txt); and all other options are left at their default setting. For example, the following command can be used to generate an output file  
5 containing a comparison between two amino acid sequences: C:\BI2seq -i c:\seq1.txt -j c:\seq2.txt -p blastp -o c:\output.txt. If the two compared sequences share homology, then the designated output file will present those regions of homology as aligned sequences. If the two compared sequences do not share homology, then the designated output file will not present aligned sequences.

10 Once aligned, the number of matches is determined by counting the number of positions where an identical nucleotide or amino acid residue is presented in both sequences. The percent sequence identity is determined by dividing the number of matches either by the length of the sequence set forth in the identified sequence (e.g., SEQ ID NO:11), or by an articulated length (e.g., 100 consecutive nucleotides or amino  
15 acid residues from a sequence set forth in an identified sequence), followed by multiplying the resulting value by 100. For example, an amino acid sequence that has 1300 matches when aligned with the sequence set forth in SEQ ID NO:11 is 93.7 percent identical to the sequence set forth in SEQ ID NO:11 (i.e.,  $1300 \div 1388 \times 100 = 93.7$ ). It is noted that the percent sequence identity value is rounded to the nearest tenth. For  
20 example, 75.11, 75.12, 75.13, and 75.14 is rounded down to 75.1, while 75.15, 75.16, 75.17, 75.18, and 75.19 is rounded up to 75.2. It also is noted that the length value will always be an integer.

As used herein, the term “functional variant” is intended to refer to a catalytically active mutant of a protein or a protein domain. Such a mutant can have the same level of  
25 activity, or a higher or lower level of activity as compared to the parent protein or protein domain.

The construct(s) containing the crRNA, tracrRNA, cr/tracrRNA hybrid, endonuclease coding sequence, and, where applicable, donor sequence, can be delivered to a plant, plant part, or plant cell using, for example, biolistic bombardment.

30 Alternatively, the system components can be delivered using *Agrobacterium*-mediated

transformation. In some embodiments, the system components can be delivered in a viral vector (e.g., a vector from a DNA virus such as, without limitation, geminivirus (e.g., cabbage leaf curl virus, bean yellow dwarf virus, wheat dwarf virus, tomato leaf curl virus, maize streak virus, tobacco leaf curl virus, or tomato golden mosaic virus) or nanovirus (e.g., Faba bean necrotic yellow virus), or a vector from an RNA virus such as, without limitation, tobnavirus (e.g., tobacco rattle virus, tobacco mosaic virus), potexvirus (e.g., potato virus X), or hordeivirus (e.g., barley stripe mosaic virus).

After a plant, plant part, or plant cell is infected or transfected with an endonuclease encoding sequence and a crRNA and a tracrRNA, or a cr/tracrRNA hybrid (and, in some cases, a donor sequence), any suitable method can be used to determine whether GT or targeted mutagenesis has occurred at the target site. In some embodiments, a phenotypic change can indicate that a donor sequence has been incorporated into the target site. PCR-based methods also can be used to ascertain whether a genomic target site contains targeted mutations or donor sequence, and/or whether precise recombination has occurred at the 5' and 3' ends of the donor. One method to detect targeted mutations, referred to herein as "PCR digest," is described by Zhang et al. (*Proc Natl Acad Sci USA* 107:12028-12033, 2010). Methods to detect precise recombination include southern blotting using a probe with homology to the donor sequence.

In some embodiments, the methods provided herein can include introducing into a plant, plant part, or plant cell a nucleic acid that includes a crRNA and a tracrRNA, or a chimeric cr/tracrRNA hybrid, where the crRNA and tracrRNA, or the cr/tracrRNA hybrid, is targeted to a nucleotide sequence that is endogenous to the plant cell, and also introducing into the plant, plant part, or plant cell a Cas9 endonuclease molecule (e.g., a Cas9 polypeptide or a portion thereof, such as a portion of a Cas9 polypeptide that includes the HNH domain, or a nucleic acid encoding a Cas9 polypeptide or a portion thereof), where the Cas9 endonuclease molecule induces a double strand break at or near the sequence to which the crRNA and tracrRNA sequences (or the cr/tracrRNA hybrid) are targeted.

The plants, plant parts, and plant cells used in the methods provided herein can be from any species of plant. In some embodiments, for example, the methods provided herein can utilize monocotyledonous plants, portions thereof, or cells therefrom. Exemplary monocotyledonous plants include, without limitation, wheat, maize, rice, orchids, onion, aloe, true lilies, grasses (e.g., *Setaria*), woody shrubs and trees (e.g., palms and bamboo), and food plants such as pineapple and sugar cane. Exemplary dicotyledonous plants include, without limitation, tomato, cassava, soybean, tobacco, potato, *Arabidopsis*, rose, pansy, sunflower, grape, strawberry, squash, bean, pea, and peanut.

In some embodiments, the methods described herein can include screening the plant, plant part, or plant cell to determine if a DSB has occurred at or near the sequence targeted by the crRNA and tracrRNA or the cr/tracrRNA hybrid. For example, the PCR-digest assay described by Zhang et al. (*supra*) can be used to determine whether a DSB has occurred. Other useful methods include, without limitation, the T7 assay, the Surveyor assay, and southern blotting (if a restriction enzyme binding sequence is present at or near the predicted cleavage site).

In addition, in some embodiments in which a plant part or plant cell is used, the methods provided herein can include regenerating a plant from the plant part or plant cell. The methods also can include breeding the plant (e.g., the plant into which the nucleic acids were introduced, or the plant obtained after regeneration of the plant part or plant cell used as a starting material) to obtain a genetically desired plant lineage. Methods for regenerating and breeding plants are well established in the art.

Also provided herein are plants, plant parts, and plant cells containing a nucleic acid that encodes a Cas9 polypeptide with an amino acid sequence that is at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) identical to the amino acid sequence set forth in SEQ ID NO:11 or SEQ ID NO:12, or a nucleic acid that encodes a Cas9 polypeptide containing an amino acid sequence that is at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) identical to amino acids 828 to 879 of SEQ ID NO:11, or amino acids 810 to 872 of SEQ ID NO:12.

This document also provides virus vectors that contain nucleotide sequences encoding Cas9 polypeptides. For example, a virus vector can include a nucleotide sequence encoding a polypeptide having an amino acid sequence that is at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) identical to the amino acid sequence set forth in SEQ ID NO:11 or SEQ ID NO:12. In some embodiments, a virus vector can have a nucleotide sequence encoding a Cas9 polypeptide that includes an amino acid sequence with at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) sequence identity to amino acids 828 to 879 of SEQ ID NO:11, or amino acids 810 to 872 of SEQ ID NO:12. The vector can be from any suitable type of virus, such as a tobnavirus or a geminivirus, for example.

Also provide herein are T-DNA molecules that contain a nucleic acid sequence encoding a Cas9 polypeptide having an amino acid sequence that is at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) identical to the amino acid sequence set forth in SEQ ID NO:11 or SEQ ID NO:12. In some embodiments, a T-DNA can include a nucleotide sequence encoding a Cas9 polypeptide that includes an amino acid sequence with at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) sequence identity to amino acids 828 to 879 of SEQ ID NO:11, or amino acids 810 to 872 of SEQ ID NO:12.

This document also provides *Agrobacterium* strains comprising a T-DNA as described herein.

In addition, this document provides methods for expressing a Cas protein in a plant, a plant part, or a plant cell. Such methods can include, for example, (a) providing an *Agrobacterium* or *Ensifer* vector containing a T-DNA that includes a nucleic acid sequence encoding a Cas9 polypeptide having an amino acid sequence with at least 80% (e.g., at least 85%, at least 90%, at least 95%, or at least 98%) sequence identity to SEQ ID NO:11 or SEQ ID NO:12, where the Cas9-encoding sequence is operably linked to a promoter, (b) bringing the *Agrobacterium* or *Ensifer* vector into contact with a plant, plant part, or plant cell, and (c) expressing the nucleic acid sequence in the plant, plant part, or plant cell. The promoter can be, for example, a constitutive promoter (e.g., a CaMV 35S promoter), an inducible promoter (e.g., an estradiol-induced XVE promoter;

Zuo et al., *Plant J* 24:265-273, 2000), a cell specific promoter, or a promoter that is activated by alternative splicing of a suicide exon. In some embodiments, such methods also can include contacting the plant, plant part, or plant cell with a nucleic acid encoding a guide RNA that associates with the Cas protein, and expressing the guide RNA.

5           The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

### EXAMPLES

#### Example 1 – Plasmids for expressing CRISPR/Cas components

10           To demonstrate functionality of the CRISPR/Cas systems for genome editing in plants, plasmids were constructed to encode Cas9, crRNA and tracrRNA, the cr/tracrRNA hybrid, and RNA polymerase III promoters (e.g., AtU6-26 or At7SL-2) from which to express the crRNA, tracrRNA, or cr/tracrRNA hybrid. Plant codon-optimized Cas9 coding sequence was synthesized and cloned into a MultiSite Gateway  
15           entry plasmid. Additionally, crRNA and tracrRNA, or cr/tracrRNA hybrid, driven by the RNA polymerase III (PolIII) promoters AtU6-26 and At7SL2-2, were synthesized and cloned into a second MultiSite Gateway entry plasmid. To enable efficient reconstruction of the crRNA sequences (serving to redirect CRISPR/Cas-mediated DSBs), inverted type-IIS restriction enzyme sites (e.g., *Bsa*I and *Esp*3I) were inserted within the crRNA  
20           nucleotide sequence. By digesting with the appropriate type-IIS restriction enzyme, target sequences can be efficiently cloned into the crRNA sequence using oligonucleotides. Entry plasmids for both Cas9 and the expression of the crRNA and tracrRNA or the cr/tracrRNA hybrid, from a RNA polymerase III promoter (AtU6-26 or At7SL2-2), were recombined into pMDC32 (a standard T-DNA expression plasmid with a 2x35S  
25           promoter; FIG. 1 and SEQ ID NO:6), pFZ19 (an estrogen-inducible T-DNA expression vector; FIG. 2 and SEQ ID NO:7; Zuo et al., *Plant J.* 24(2):265-273, 2000), and pNJB121 (a geminivirus-replicon T-DNA vector; FIG. 3 and SEQ ID NO:8).



### Example 2 – CRISPR/Cas activity in somatic plant tissue

To demonstrate the capacity for CRISPR/Cas systems to function as SSNs, the geminivirus-replicon T-DNA vector, pNJB121, was modified to encode both Cas9 and cr/tracrRNA hybrid sequences (FIG. 4A). Targeting RNA sequences (encoded by  
5 nucleotide sequence within the crRNA; responsible for directing Cas9 cleavage) were designed to be homologous to sequences within the endogenous *SuRA* and *SuRB* genes. The sequence of the targeting portion of the crRNA that matched the *SuR* loci was 5'-GUGGGAGGAUCGGUUCUAUA (SEQ ID NO:9; the 5' G does not match the *SuR* loci, but is needed for transcription by RNA polymerase III). Although pNJB121 is a  
10 geminivirus-replicon, in the absence of replicase (Rep), no amplification occurs. Therefore, pNJB121 in the absence of Rep is a standard T-DNA vector and no replicons are formed. The modified pNJB121 plasmid delivered to *Nicotiana tabacum* leaf tissue by syringe infiltration with *Agrobacterium tumefaciens*. Five days after infiltration, *SuRA/SuRB* sequences were assessed for Cas9-mediated mutations using PCR-digest  
15 (FIG. 4C). The presence of mutations at the corresponding target sequences indicated functionality of CRISPR/Cas systems in plant leaf cells.

### Example 3 – CRISPR/Cas activity in protoplasts

To further demonstrate the activity of CRISPR/Cas systems in plants, targeted  
20 mutagenesis of DNA sequence within *Arabidopsis thaliana* and *Nicotiana tabacum* protoplasts is assessed. Targeting crRNA sequences are redesigned to be homologous to sequences present within the endogenous *ADH1* or *TT4* genes (*Arabidopsis*), or the integrated *gus:nptII* reporter gene or *SuRA/SuRB* (*Nicotiana*). Protoplasts are isolated from *Arabidopsis* and *Nicotiana* leaf tissue and transfected with plasmids encoding Cas9  
25 and the *ADH1*- or *TT4*-targeting crRNAs, or Cas9 and the *gus:nptII*- or *SuRA/SuRB*-targeting crRNA, respectively. Genomic DNA is extracted 5-7 days post transfection and assessed for mutations at the corresponding target sequences. Detecting mutations within the *ADH1*, *TT4*, *gus:nptII* or *SuRA/SuRB* genes indicates the functionality of CRISPR/Cas systems to target endogenous genes in plant protoplasts.

In initial studies, the CRISPR/Cas system was assessed for the ability to cleave an extrachromosomal reporter plasmid, using methods similar to those described by Zhang et al. (*Plant Physiol* 161:20-27, 2013). The reporter plasmid encodes a non-functional yellow fluorescent protein (YFP; FIG. 5 and SEQ ID NO:10). YFP expression is  
5 disrupted by a direct repeat of internal coding sequence that flanks a target sequence for the Cas9/crRNA complex. The generation of targeted DSBs at the Cas9/crRNA target sequence results in recombination of the direct repeat sequences, thereby restoring YFP gene function. A sequence from the tobacco *SuRA/SuRB* loci was cloned into the YFP reporter between the direct repeats. A cr/tracrRNA hybrid construct that targets this site  
10 was then generated. The sequence of the portion of the crRNA that targets the *SuR* loci was 5'- GUGGGAGGAUCGGUUCUAUA (SEQ ID NO:9; again, the 5' G does not match the *SuR* loci, but it is needed for transcription by RNA polymerase III). *Nicotiana tabacum* protoplasts were transformed with plasmids encoding Cas9, a cr/tracrRNA hybrid, and the YFP reporter, and restoration of YFP expression as a result of  
15 CRISPR/Cas nuclease activity was monitored by flow cytometry. Using a positive control plasmid that encodes YFP, 94.7% of the cells were transformed and expressed YFP (FIG. 6, column 4). Cells transformed with the reporter alone gave activity levels barely above background (FIG. 6, column 3). When cells were transformed with constructs expressing Cas9 and a cr/tracr RNA, significant activity was observed,  
20 indicating the Cas9/crRNA complex cleaved the target. For the cr/tracrRNA expressed from the AtU6-26 promoter, 18.8% of the cells fluoresced (FIG. 6, column 1). When the cr/tracr RNA was expressed from the At7SL2-2 promoter, 20.7% of the cells were YFP positive (FIG. 6, column 2). Detection of YFP-expressing cells indicated the functionality of CRISPR/Cas systems in plant protoplasts.

#### Example 4 – Multiplex genome engineering in protoplasts using CRISPR/Cas systems

The ability of CRISPR/Cas systems to create multiple DSBs at different DNA sequences is assessed using plant protoplasts. To direct Cas9 nuclease activity to *TT4*, *ADH1*, and the extrachromosomal YFP reporter plasmid (within the same *Arabidopsis*  
30 protoplast), crRNA and tracrRNA or cr/tracrRNA hybrid plasmid is modified to express

multiple crRNA targeting sequences. These sequences are designed to be homologous to sequences present within *TT4*, *ADHI* and the YFP reporter plasmid. Following transfection with Cas9, crRNA, tracrRNA, or the cr/tracrRNA hybrid, and YFP reporter plasmids into *Arabidopsis* protoplasts, YFP-expressing cells are quantified and isolated, and genomic DNA is extracted. Observing mutations within the *ADHI* and *TT4* genes in YFP-expressing cells suggests that CRISPR/Cas can facilitate multiplex genome engineering in *Arabidopsis* cells.

To demonstrate multiplex genome engineering in *Nicotiana* protoplasts, plasmids containing multiple crRNA are modified to encode sequences that are homologous to the integrated *gus:nptII* reporter gene, *SuRA/SuRB*, and the YFP reporter plasmid. Similar to the methods described in *Arabidopsis* protoplasts, *Nicotiana* protoplasts are transfected with Cas9, crRNA, tracrRNA, or the cr/tracrRNA hybrid, and YFP reporter plasmids. YFP-expressing cells are quantified and isolated, and genomic DNA is extracted. Observing mutations within the integrated *gus:nptII* reporter gene and *SuRA/SuRB* in YFP-expressing cells suggests that CRISPR/Cas can facilitate multiplex genome engineering in tobacco cells.

#### Example 5 – CRISPR/Cas activity in planta

To demonstrate CRISPR/Cas activity in *planta*, pFZ19 T-DNA is modified to encode both Cas9 and the crRNA and tracrRNA, or the cr/tracrRNA hybrid sequences. Target DNA sequences are present within the endogenous *ADHI* or *TT4* genes. The resulting T-DNA is integrated into the *Arabidopsis thaliana* genome by floral dip using *Agrobacterium*. Cas9 expression is induced in primary transgenic plants by direct exposure to estrogen. Genomic DNA from somatic leaf tissue is extracted and assessed for mutations at the corresponding genomic locus by PCR-digest. Observing mutations within the *ADHI* or *TT4* genes demonstrates CRISPR/Cas activity in *planta*. Alternatively, CRISPR/Cas activity can be assessed by screening T2 seeds (produced from induced T1 patents) for heterozygous or homozygous mutations at the corresponding genomic locus. Furthermore, the capacity for CRISPR/Cas to carry out multiplex genome engineering is assessed by modifying plasmids containing multiple

crRNAs with homologous sequences to both *ADHI* and *TT4*. The resulting T-DNA plasmid is integrated into the *Arabidopsis* genome, Cas9 expression is induced in primary transgenic plants, and CRISPR/Cas activity is assessed by evaluating the *ADHI* and *TT4* genes in both T1 and T2 plants. Observing mutations in both the *ADHI* and *TT4* genes suggests CRISPR/Cas can facilitate multiplex genome engineering in *Arabidopsis* plants.

#### Example 6 – Viral delivery of CRISPR/Cas components

Plant viruses can be effective vectors for delivery of heterologous nucleic acid sequence, such as for RNAi reagents or for expressing heterologous proteins. Useful plant viruses include both RNA viruses (e.g., tobacco mosaic virus, tobacco rattle virus, potato virus X, and barley stripe mosaic virus) and DNA viruses (e.g., cabbage leaf curl virus, bean yellow dwarf virus, wheat dwarf virus, tomato leaf curl virus, maize streak virus, tobacco leaf curl virus, tomato golden mosaic virus, and Faba bean necrotic yellow virus; Rybicki et al., *Curr Top Microbiol Immunol*, 2011; and Gleba et al., *Curr Opin Biotechnol* 2007, 134-141). Such plant viruses can be modified for the delivery of CRISPR/Cas9 components. Proof-of-concept experiments were performed in *Nicotiana tabacum* leaf cells using DNA viruses (geminivirus replicons; Baltes et al., *Plant Cell* 26:151-163, 2014). To this end, crRNA sequences were modified to contain homology to the endogenous *SuRA/SuRB* loci. The resulting plasmids were cloned into pNJB121 (a T-DNA destination vector with *cis*-acting elements required for geminivirus replication (LSL T-DNA)) along with Cas9 (FIG. 4A). Co-delivery of LSL T-DNA along with T-DNA encoding replicase protein (Rep; REP T-DNA) by *Agrobacterium* resulted in the replicational release of geminiviral replicons (FIG. 4B). The T-DNA was delivered to tobacco leaf tissue by syringe infiltration with *Agrobacterium*. Five to seven days after infiltration, *SuRA/SuRB* sequences were assessed for Cas9-mediated mutations using PCR-digest (FIG. 4C). Digestion-resistant PCR amplicons were cloned and sequenced. The presence of mutations at the corresponding target sequences indicates that plant viruses are effective vectors for delivery of CRISPR/Cas components (FIG. 4D).

**OTHER EMBODIMENTS**

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended  
5 claims. Other aspects, advantages, and modifications are within the scope of the following claims.

## CLAIMS

1. A method for modifying the genomic material in a plant cell, comprising:
  - (a) introducing into the cell a nucleic acid comprising a clustered regularly interspersed short palindromic repeats- (CRISPR-) associated RNA(crRNA) and a trans-activating crRNA (tracrRNA), or a chimeric cr/tracrRNA hybrid, wherein the crRNA and tracrRNA, or the cr/tracrRNA hybrid, is targeted to a sequence that is endogenous to the plant cell; and
  - (b) introducing into the cell a CRISPR-associated (Cas9) endonuclease molecule that induces a double strand break at or near the sequence to which the crRNA and tracrRNA sequence is targeted, or at or near the sequence to which the cr/tracrRNA hybrid is targeted.
2. The method of claim 1, wherein the introducing steps comprise delivering to the plant cell a nucleic acid encoding the Cas9 endonuclease and a nucleic acid encoding the crRNA and tracrRNA or the cr/tracrRNA hybrid, and wherein the delivering is by a DNA or RNA virus.
3. The method of claim 2, wherein the delivering is by a DNA virus, and wherein the DNA virus is a geminivirus.
4. The method of claim 2, wherein the delivering is by an RNA virus, and wherein the RNA virus is a tobnavirus.
5. The method of claim 1, wherein the introducing steps comprise delivering to the plant cell a T-DNA containing a nucleic acid sequence encoding the Cas9 endonuclease and a nucleic acid sequence encoding the crRNA and tracrRNA or the cr/tracrRNA hybrid, and wherein the delivering is via *Agrobacterium* or *Ensifer*.
6. The method of claim 5, wherein the nucleic acid sequence encoding the Cas9 endonuclease is operably linked to a promoter that is constitutive, cell specific, inducible, or activated by alternative splicing of a suicide exon.

7. The method of claim 1, wherein the introducing steps comprise microprojectile bombardment of nucleic acid encoding Cas9 and the crRNA and tracrRNA or the cr/tracrRNA hybrid.
8. The method of claim 1, wherein the plant cell is from a monocotyledonous plant.
9. The method of claim 8, wherein the monocotyledonous plant is wheat, maize, rice, or *Setaria*.
10. The method of claim 1, wherein the plant cell is from a dicotyledonous plant.
11. The method of claim 10, wherein the dicotyledonous plant is tomato, soybean, tobacco, potato, cassava, or *Arabidopsis*.
12. The method of claim 1, further comprising screening the plant cell after the introducing steps to determine if a double strand break has occurred at or near the sequence targeted by the crRNA and tracrRNA or the cr/tracrRNA hybrid.
13. The method of claim 1, further comprising regenerating a plant from the plant cell.
14. The method of claim 13, further comprising cross breeding the plant to obtain a genetically desired plant lineage.
15. A plant cell comprising a nucleic acid encoding a polypeptide having at least 80% sequence identity with SEQ ID NO:12.
16. A plant cell comprising a nucleic acid encoding a polypeptide that comprises an amino acid sequence having at least 80% sequence identity with amino acids 810 to 872 of SEQ ID NO:12.
17. A virus vector comprising a nucleotide sequence that encodes a Cas9 polypeptide wherein the virus is a tobnavirus or a geminivirus.

18. The virus vector of claim 17, wherein the vector comprises a nucleotide sequence encoding a polypeptide with an amino acid sequence having at least 90% identity to SEQ ID NO:12.
19. A T-DNA comprising a nucleic acid sequence encoding a polypeptide that comprises an amino acid sequence having at least 80% sequence identity with amino acids 810 to 872 of SEQ ID NO:12.
20. An *Agrobacterium* strain comprising the T-DNA of claim 19.
21. A method for expressing a Cas protein in a plant cell, comprising:
  - providing an *Agrobacterium* or *Ensifer* vector containing a T-DNA that comprises a nucleic acid sequence encoding a polypeptide having an amino acid sequence with at least 80% sequence identity to amino acids 810 to 872 of SEQ ID NO:12, wherein the polypeptide-encoding sequence is operably linked to a promoter;
  - bringing the *Agrobacterium* or *Ensifer* vector into contact with the plant cell; and
  - expressing the nucleic acid sequence in the plant cell.
22. The method of claim 21, wherein the promoter is an inducible promoter.
23. The method of claim 22, wherein the inducible promoter is an estrogen inducible promoter.
24. The method of claim 21, further comprising contacting the plant cell with a nucleic acid encoding a guide RNA that associates with the Cas protein.
25. The method of claim 21, wherein the plant cell is a protoplast.



FIG. 1

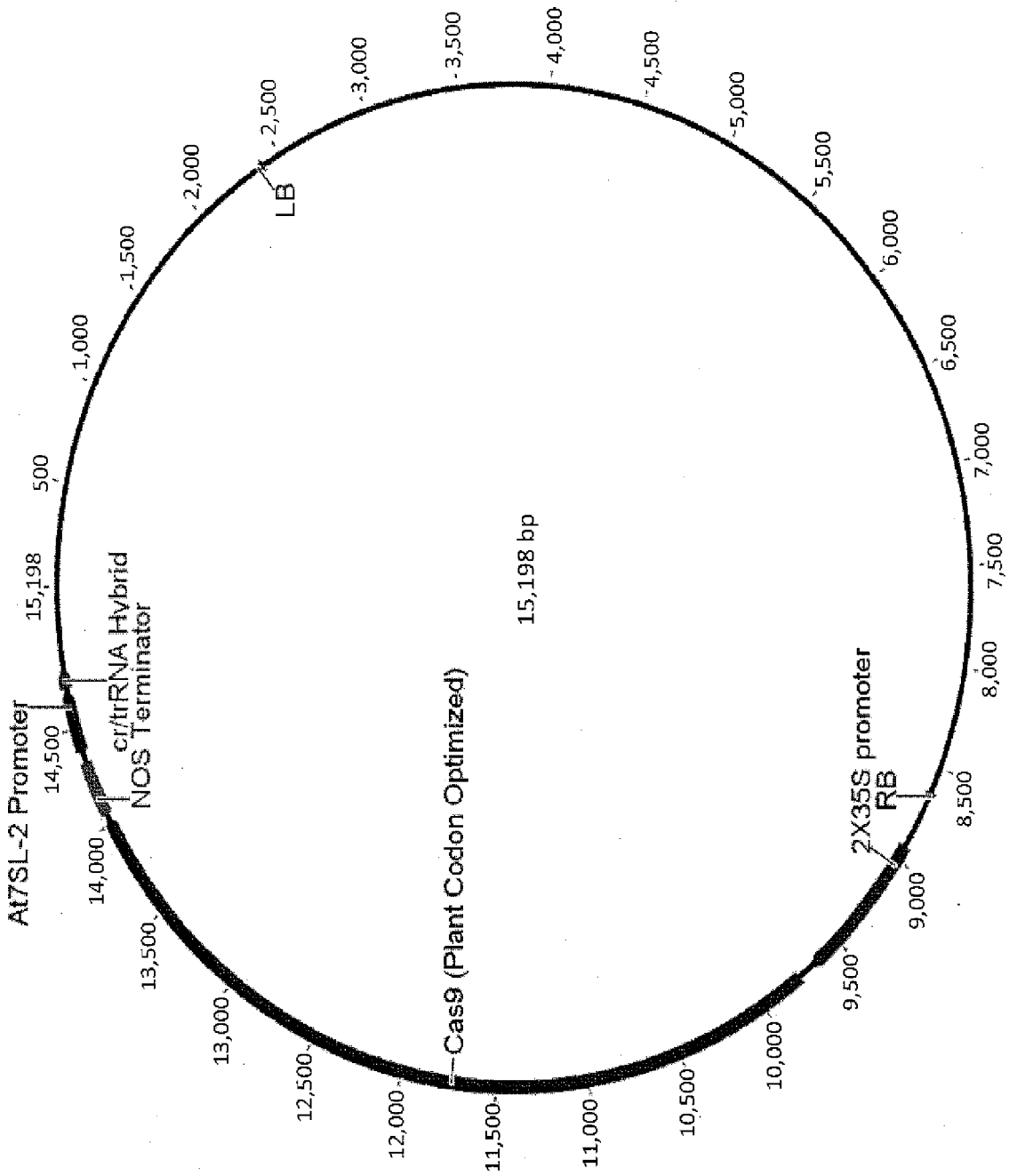
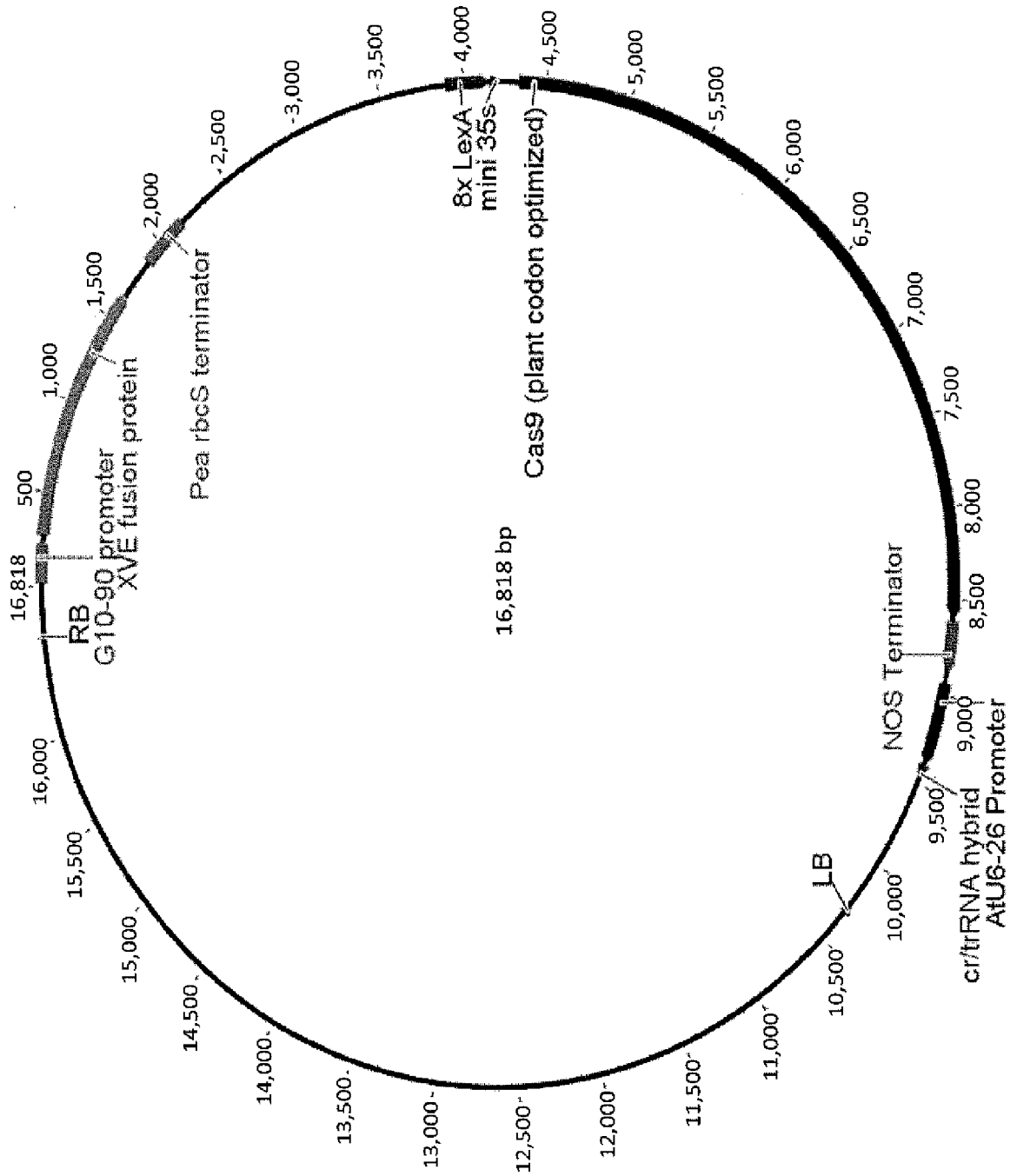


FIG. 2



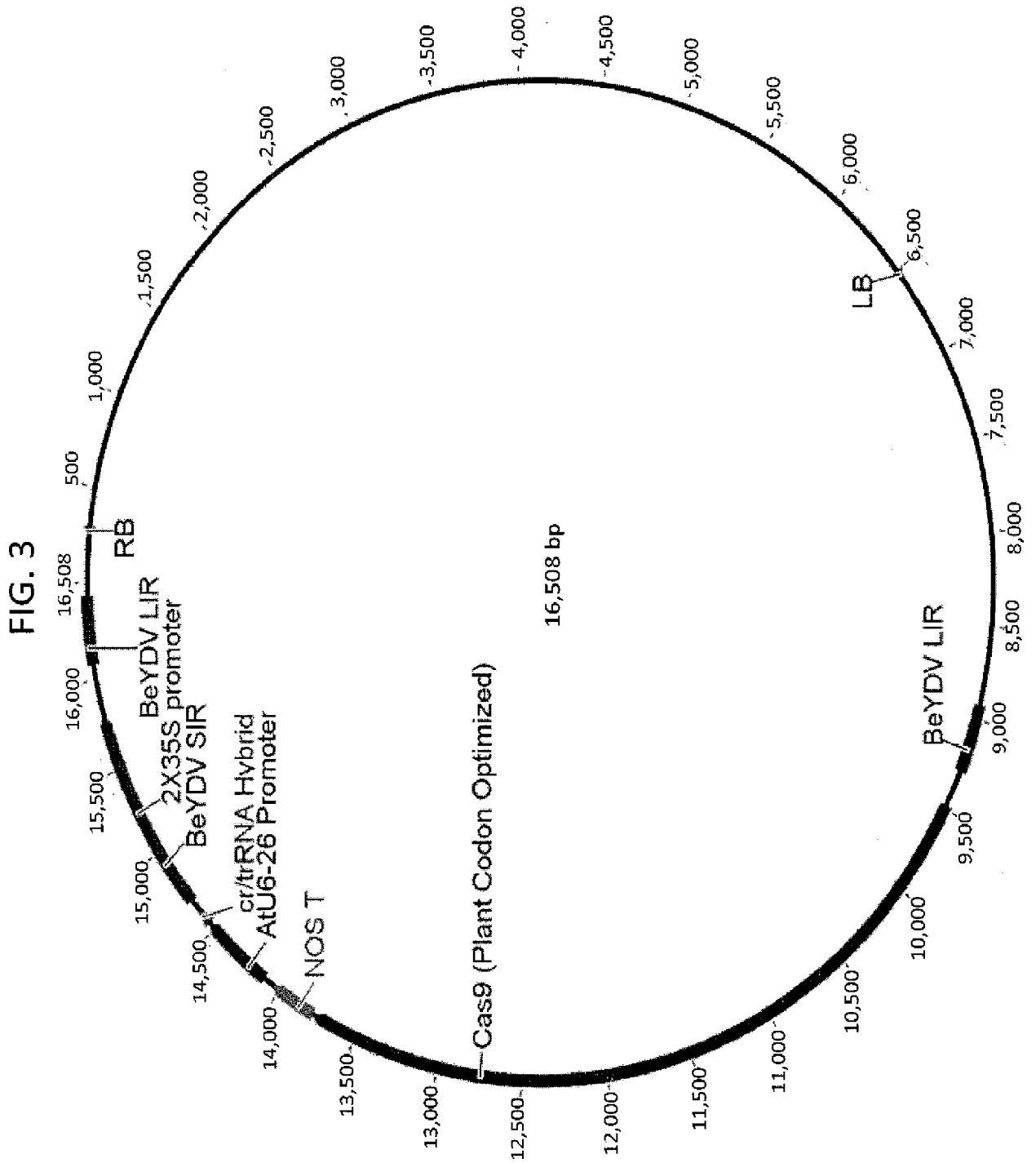


FIG. 4A

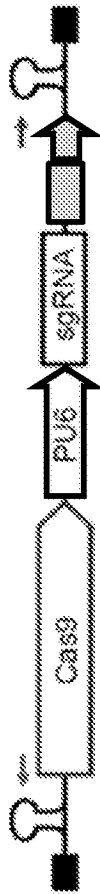


FIG. 4B

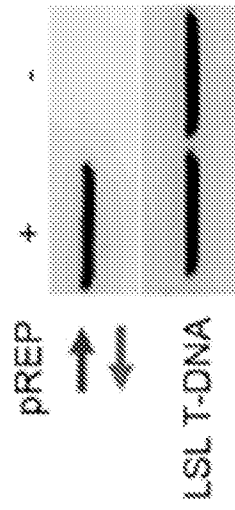


FIG. 4C

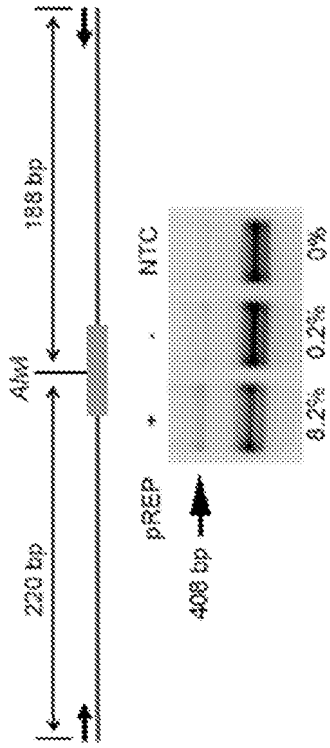


FIG. 4D

PAM	(SEQ ID NO: 1)
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GGT-----ATAAGGCTAACAGA	-20
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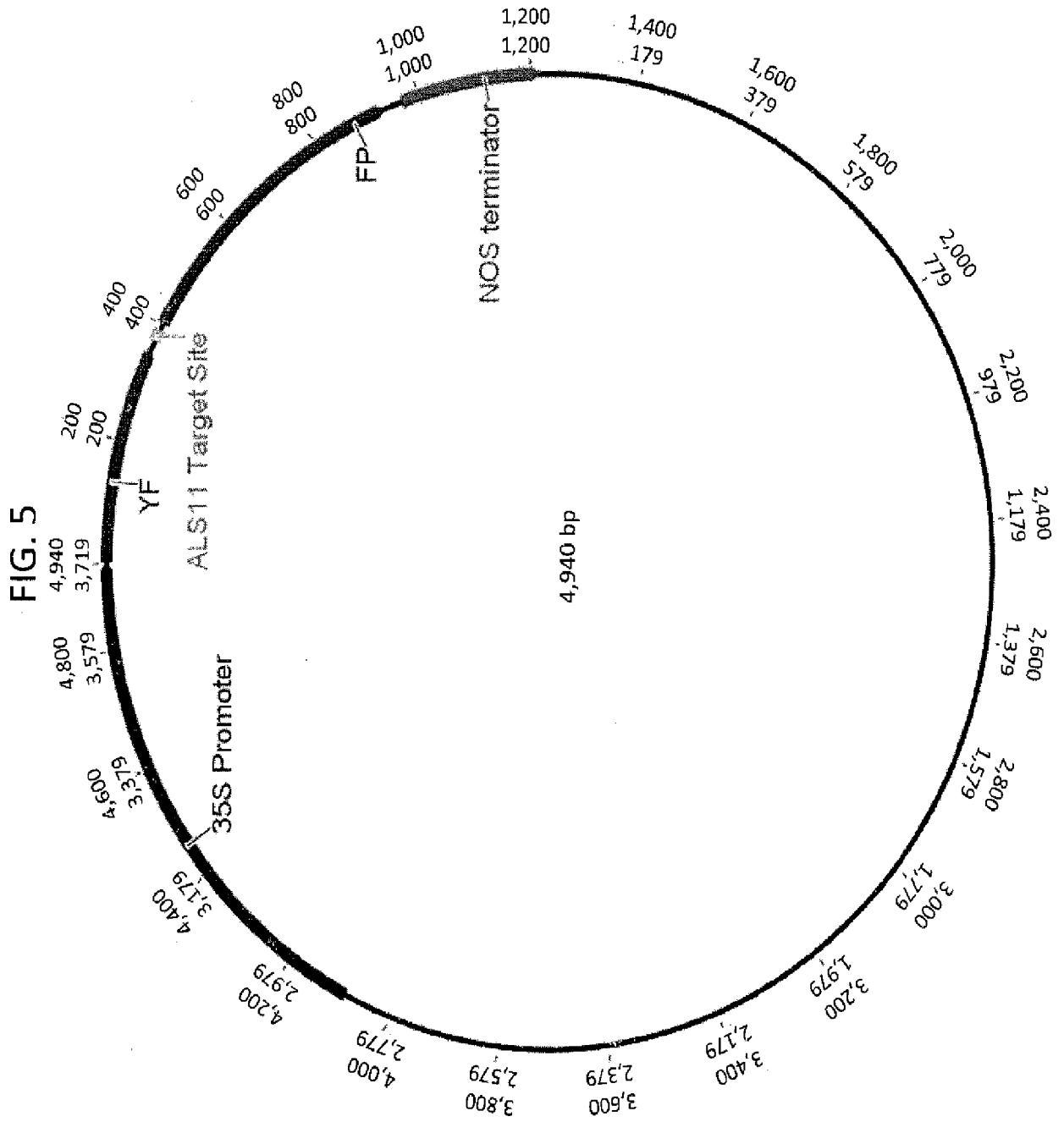
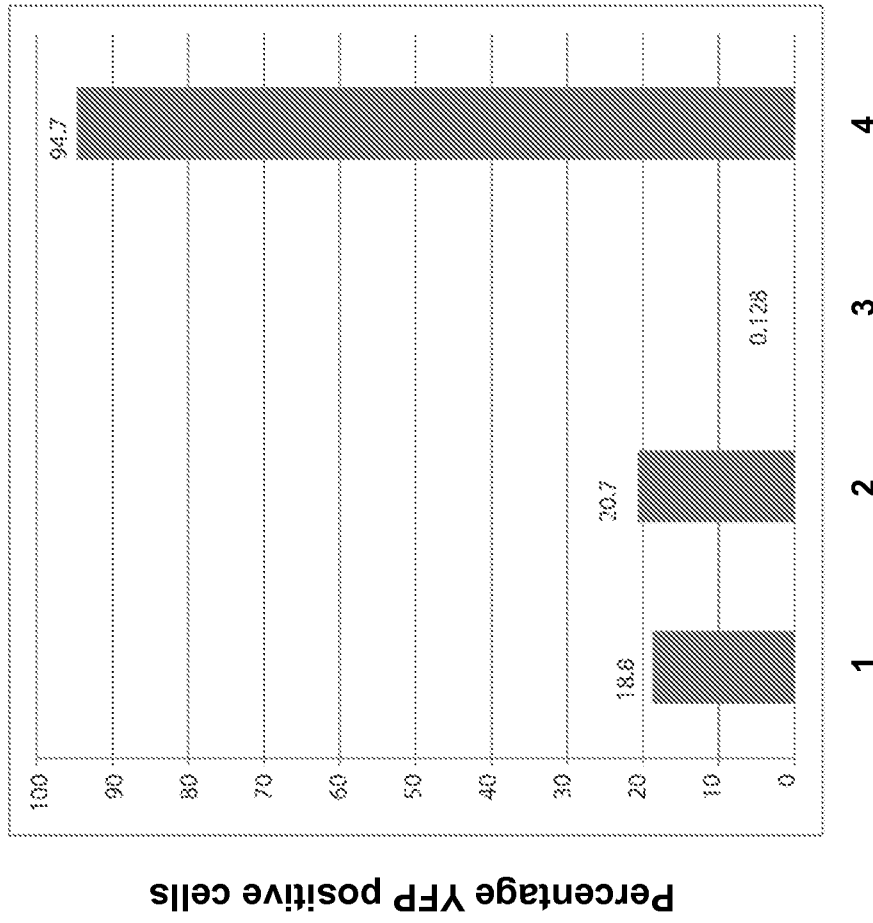


FIG. 6



SequenceListing.TXT

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CRISPR/Cas SYSTEMS

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<151> 2013-03-15

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SequenceListing.TXT

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SequenceListing.TXT

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SequenceListing.TXT

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SequenceListing.TXT

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SequenceListing.TXT

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tgaggacca	cttctgcgct	cggcccttcc	ggctggctgg	tttattgctg	ataaatctgg	2700
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ccgtatcgta	gttatctaca	cgacggggag	tcaggcaact	atggatgaac	gaaatagaca	2820
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agaccccgta	gaaaagatca	aaggatcttc	ttgagatcct	ttttttctgc	gcgtaatctg	3060
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aacacagaga	aagatatatt	tctcaagatc	agaagtaacta	ttccagtatg	gacgattcaa	4320
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acgtcttcaa	aagcaagtgg	attgatgtga	tatctccact	gacgtaaggg	gatgacgcac	4860
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agaacacggg	ggactctaga					4940

SequenceListing.TXT

<210> 11  
 <211> 1388  
 <212> PRT  
 <213> Streptococcus thermophilus

<400> 11

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			20					25					30		
Lys	Val	Leu	Gly	Asn	Thr	Ser	Lys	Lys	Tyr	Ile	Lys	Lys	Asn	Leu	Leu
		35					40					45			
Gly	Val	Leu	Leu	Phe	Asp	Ser	Gly	Ile	Thr	Ala	Glu	Gly	Arg	Arg	Leu
						55					60				
Lys	Arg	Thr	Ala	Arg	Arg	Arg	Tyr	Thr	Arg	Arg	Arg	Asn	Arg	Ile	Leu
65					70					75					80
Tyr	Leu	Gln	Glu	Ile	Phe	Ser	Thr	Glu	Met	Ala	Thr	Leu	Asp	Asp	Ala
				85					90					95	
Phe	Phe	Gln	Arg	Leu	Asp	Asp	Ser	Phe	Leu	Val	Pro	Asp	Asp	Lys	Arg
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Asp	Ser	Lys	Tyr	Pro	Ile	Phe	Gly	Asn	Leu	Val	Glu	Glu	Lys	Ala	Tyr
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His	Asp	Glu	Phe	Pro	Thr	Ile	Tyr	His	Leu	Arg	Lys	Tyr	Leu	Ala	Asp
	130					135						140			
Ser	Thr	Lys	Lys	Ala	Asp	Leu	Arg	Leu	Val	Tyr	Leu	Ala	Leu	Ala	His
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Met	Ile	Lys	Tyr	Arg	Gly	His	Phe	Leu	Ile	Glu	Gly	Glu	Phe	Asn	Ser
				165					170					175	
Lys	Asn	Asn	Asp	Ile	Gln	Lys	Asn	Phe	Gln	Asp	Phe	Leu	Asp	Thr	Tyr
			180					185						190	
Asn	Ala	Ile	Phe	Glu	Ser	Asp	Leu	Ser	Leu	Glu	Asn	Ser	Lys	Gln	Leu
		195					200					205			
Glu	Glu	Ile	Val	Lys	Asp	Lys	Ile	Ser	Lys	Leu	Glu	Lys	Lys	Asp	Arg
		210				215						220			
Ile	Leu	Lys	Leu	Phe	Pro	Gly	Glu	Lys	Asn	Ser	Gly	Ile	Phe	Ser	Glu
225					230					235					240
Phe	Leu	Lys	Leu	Ile	Val	Gly	Asn	Gln	Ala	Asp	Phe	Arg	Lys	Cys	Phe
				245					250					255	
Asn	Leu	Asp	Glu	Lys	Ala	Ser	Leu	His	Phe	Ser	Lys	Glu	Ser	Tyr	Asp
			260					265						270	
Glu	Asp	Leu	Glu	Thr	Leu	Leu	Gly	Tyr	Ile	Gly	Asp	Asp	Tyr	Ser	Asp
		275					280					285			
Val	Phe	Leu	Lys	Ala	Lys	Lys	Leu	Tyr	Asp	Ala	Ile	Leu	Leu	Ser	Gly
		290				295						300			
Phe	Leu	Thr	Val	Thr	Asp	Asn	Glu	Thr	Glu	Ala	Pro	Leu	Ser	Ser	Ala
305					310					315					320
Met	Ile	Lys	Arg	Tyr	Asn	Glu	His	Lys	Glu	Asp	Leu	Ala	Leu	Leu	Lys
				325						330					335
Glu	Tyr	Ile	Arg	Asn	Ile	Ser	Leu	Lys	Thr	Tyr	Asn	Glu	Val	Phe	Lys
			340					345						350	
Asp	Asp	Thr	Lys	Asn	Gly	Tyr	Ala	Gly	Tyr	Ile	Asp	Gly	Lys	Thr	Asn
		355					360					365			
Gln	Glu	Asp	Phe	Tyr	Val	Tyr	Leu	Lys	Lys	Leu	Leu	Ala	Glu	Phe	Glu
		370				375						380			
Gly	Ala	Asp	Tyr	Phe	Leu	Glu	Lys	Ile	Asp	Arg	Glu	Asp	Phe	Leu	Arg
385					390					395					400
Lys	Gln	Arg	Thr	Phe	Asp	Asn	Gly	Ser	Ile	Pro	Tyr	Gln	Ile	His	Leu
				405					410					415	
Gln	Glu	Met	Arg	Ala	Ile	Leu	Asp	Lys	Gln	Ala	Lys	Phe	Tyr	Pro	Phe
			420					425					430		
Leu	Ala	Lys	Asn	Lys	Glu	Arg	Ile	Glu	Lys	Ile	Leu	Thr	Phe	Arg	Ile
		435					440					445			
Pro	Tyr	Tyr	Val	Gly	Pro	Leu	Ala	Arg	Gly	Asn	Ser	Asp	Phe	Ala	Trp
	450					455					460				
Ser	Ile	Arg	Lys	Arg	Asn	Glu	Lys	Ile	Thr	Pro	Trp	Asn	Phe	Glu	Asp
465					470					475					480
Val	Ile	Asp	Lys	Glu	Ser	Ser	Ala	Glu	Ala	Phe	Ile	Asn	Arg	Met	Thr





SequenceListing.TXT

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 1045 1050 1055  
 Val Ile Glu Arg Pro Leu Ile Glu Val Asn Glu Glu Thr Gly Glu Ser  
 1060 1065 1070  
 Val Trp Asn Lys Glu Ser Asp Leu Ala Thr Val Arg Arg Val Leu Ser  
 1075 1080 1085  
 Tyr Pro Gln Val Asn Val Val Lys Lys Val Glu Glu Gln Asn His Gly  
 1090 1095 1100  
 Leu Asp Arg Gly Lys Pro Lys Gly Leu Phe Asn Ala Asn Leu Ser Ser  
 1105 1110 1115 1120  
 Lys Pro Lys Pro Asn Ser Asn Glu Asn Leu Val Gly Ala Lys Glu Tyr  
 1125 1130 1135  
 Leu Asp Pro Lys Lys Tyr Gly Gly Tyr Ala Gly Ile Ser Asn Ser Phe  
 1140 1145 1150  
 Thr Val Leu Val Lys Gly Thr Ile Glu Lys Gly Ala Lys Lys Lys Ile  
 1155 1160 1165  
 Thr Asn Val Leu Glu Phe Gln Gly Ile Ser Ile Leu Asp Arg Ile Asn  
 1170 1175 1180  
 Tyr Arg Lys Asp Lys Leu Asn Phe Leu Leu Glu Lys Gly Tyr Lys Asp  
 1185 1190 1195 1200  
 Ile Glu Leu Ile Ile Glu Leu Pro Lys Tyr Ser Leu Phe Glu Leu Ser  
 1205 1210 1215  
 Asp Gly Ser Arg Arg Met Leu Ala Ser Ile Leu Ser Thr Asn Asn Lys  
 1220 1225 1230  
 Arg Gly Glu Ile His Lys Gly Asn Gln Ile Phe Leu Ser Gln Lys Phe  
 1235 1240 1245  
 Val Lys Leu Leu Tyr His Ala Lys Arg Ile Ser Asn Thr Ile Asn Glu  
 1250 1255 1260  
 Asn His Arg Lys Tyr Val Glu Asn His Lys Lys Glu Phe Glu Glu Leu  
 1265 1270 1275 1280  
 Phe Tyr Tyr Ile Leu Glu Phe Asn Glu Asn Tyr Val Gly Ala Lys Lys  
 1285 1290 1295  
 Asn Gly Lys Leu Leu Asn Ser Ala Phe Gln Ser Trp Gln Asn His Ser  
 1300 1305 1310  
 Ile Asp Glu Leu Cys Ser Ser Phe Ile Gly Pro Thr Gly Ser Glu Arg  
 1315 1320 1325  
 Lys Gly Leu Phe Glu Leu Thr Ser Arg Gly Ser Ala Ala Asp Phe Glu  
 1330 1335 1340  
 Phe Leu Gly Val Lys Ile Pro Arg Tyr Arg Asp Tyr Thr Pro Ser Ser  
 1345 1350 1355 1360  
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 Glu Thr Arg Ile Asp Leu Ala Lys Leu Gly Glu Gly  
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 <212> PRT  
 <213> Streptococcus pyogenese

<400> 12  
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 Gly Trp Ala Val Ile Thr Asp Glu Tyr Lys Val Pro Ser Lys Lys Phe  
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 Lys Val Leu Gly Asn Thr Asp Arg His Ser Ile Lys Lys Asn Leu Ile  
 35 40 45  
 Gly Ala Leu Leu Phe Asp Ser Gly Glu Thr Ala Glu Ala Thr Arg Leu  
 50 55 60  
 Lys Arg Thr Ala Arg Arg Arg Tyr Thr Arg Arg Lys Asn Arg Ile Cys  
 65 70 75 80  
 Tyr Leu Gln Glu Ile Phe Ser Asn Glu Met Ala Lys Val Asp Asp Ser  
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 Phe Phe His Arg Leu Glu Glu Ser Phe Leu Val Glu Glu Asp Lys Lys  
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 His Glu Arg His Pro Ile Phe Gly Asn Ile Val Asp Glu Val Ala Tyr

SequenceListing.TXT

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Ser	Thr	Asp	Lys	Ala	Asp	Leu	Arg	Leu	Ile	Tyr	Leu	Ala	Leu	Ala	His
145					150					155					
Met	Ile	Lys	Phe	Arg	Gly	His	Phe	Leu	Ile	Glu	Gly	Asp	Leu	Asn	Pro
				165					170					175	
Asp	Asn	Ser	Asp	Val	Asp	Lys	Leu	Phe	Ile	Gln	Leu	Val	Gln	Thr	Tyr
			180					185					190		
Asn	Gln	Leu	Phe	Glu	Glu	Asn	Pro	Ile	Asn	Ala	Ser	Gly	Val	Asp	Ala
		195					200					205			
Lys	Ala	Ile	Leu	Ser	Ala	Arg	Leu	Ser	Lys	Ser	Arg	Arg	Leu	Glu	Asn
	210					215					220				
Leu	Ile	Ala	Gln	Leu	Pro	Gly	Glu	Lys	Lys	Asn	Gly	Leu	Phe	Gly	Asn
225					230					235					
Leu	Ile	Ala	Leu	Ser	Leu	Gly	Leu	Thr	Pro	Asn	Phe	Lys	Ser	Asn	Phe
				245					250					255	
Asp	Leu	Ala	Glu	Asp	Ala	Lys	Leu	Gln	Leu	Ser	Lys	Asp	Thr	Tyr	Asp
			260					265					270		
Asp	Asp	Leu	Asp	Asn	Leu	Leu	Ala	Gln	Ile	Gly	Asp	Gln	Tyr	Ala	Asp
		275					280					285			
Leu	Phe	Leu	Ala	Ala	Lys	Asn	Leu	Ser	Asp	Ala	Ile	Leu	Leu	Ser	Asp
	290					295					300				
Ile	Leu	Arg	Val	Asn	Thr	Glu	Ile	Thr	Lys	Ala	Pro	Leu	Ser	Ala	Ser
305					310					315					
Met	Ile	Lys	Arg	Tyr	Asp	Glu	His	His	Gln	Asp	Leu	Thr	Leu	Leu	Lys
				325					330					335	
Ala	Leu	Val	Arg	Gln	Gln	Leu	Pro	Glu	Lys	Tyr	Lys	Glu	Ile	Phe	Phe
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Asp	Gln	Ser	Lys	Asn	Gly	Tyr	Ala	Gly	Tyr	Ile	Asp	Gly	Gly	Ala	Ser
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Gln	Glu	Glu	Phe	Tyr	Lys	Phe	Ile	Lys	Pro	Ile	Leu	Glu	Lys	Met	Asp
	370					375					380				
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385					390					395					
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Gly	Glu	Leu	His	Ala	Ile	Leu	Arg	Arg	Gln	Glu	Asp	Phe	Tyr	Pro	Phe
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Leu	Lys	Asp	Asn	Arg	Glu	Lys	Ile	Glu	Lys	Ile	Leu	Thr	Phe	Arg	Ile
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Pro	Tyr	Tyr	Val	Gly	Pro	Leu	Ala	Arg	Gly	Asn	Ser	Arg	Phe	Ala	Trp
	450					455					460				
Met	Thr	Arg	Lys	Ser	Glu	Glu	Thr	Ile	Thr	Pro	Trp	Asn	Phe	Glu	Glu
465					470					475					
Val	Val	Asp	Lys	Gly	Ala	Ser	Ala	Gln	Ser	Phe	Ile	Glu	Arg	Met	Thr
				485				490						495	
Asn	Phe	Asp	Lys	Asn	Leu	Pro	Asn	Glu	Lys	Val	Leu	Pro	Lys	His	Ser
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Leu	Leu	Tyr	Glu	Tyr	Phe	Thr	Val	Tyr	Asn	Glu	Leu	Thr	Lys	Val	Lys
		515					520					525			
Tyr	Val	Thr	Glu	Gly	Met	Arg	Lys	Pro	Ala	Phe	Leu	Ser	Gly	Glu	Gln
	530				535						540				
Lys	Lys	Ala	Ile	Val	Asp	Leu	Leu	Phe	Lys	Thr	Asn	Arg	Lys	Val	Thr
545					550					555					
Val	Lys	Gln	Leu	Lys	Glu	Asp	Tyr	Phe	Lys	Lys	Ile	Glu	Cys	Phe	Asp
				565					570					575	
Ser	Val	Glu	Ile	Ser	Gly	Val	Glu	Asp	Arg	Phe	Asn	Ala	Ser	Leu	Gly
			580					585					590		
Thr	Tyr	His	Asp	Leu	Leu	Lys	Ile	Ile	Lys	Asp	Lys	Asp	Phe	Leu	Asp
		595					600					605			
Asn	Glu	Glu	Asn	Glu	Asp	Ile	Leu	Glu	Asp	Ile	Val	Leu	Thr	Leu	Thr
	610					615					620				
Leu	Phe	Glu	Asp	Arg	Glu	Met	Ile	Glu	Glu	Arg	Leu	Lys	Thr	Tyr	Ala
625					630					635					
His	Leu	Phe	Asp	Asp	Lys	Val	Met	Lys	Gln	Leu	Lys	Arg	Arg	Arg	Tyr
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Thr	Gly	Trp	Gly	Arg	Leu	Ser	Arg	Lys	Leu	Ile	Asn	Gly	Ile	Arg	Asp



SequenceListing.TXT

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Val	Asn	Phe	Leu	Tyr	Leu	Ala	Ser	His	Tyr	Glu	Lys	Leu	Lys	Gly	Ser
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Pro	Glu	Asp	Asn	Glu	Gln	Lys	Gln	Leu	Phe	Val	Glu	Gln	His	Lys	His
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Tyr	Leu	Asp	Glu	Ile	Ile	Glu	Gln	Ile	Ser	Glu	Phe	Ser	Lys	Arg	Val
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Ile	Leu	Ala	Asp	Ala	Asn	Leu	Asp	Lys	Val	Leu	Ser	Ala	Tyr	Asn	Lys
				1285					1290					1295	
His	Arg	Asp	Lys	Pro	Ile	Arg	Glu	Gln	Ala	Glu	Asn	Ile	Ile	His	Leu
				1300					1305					1310	
Phe	Thr	Leu	Thr	Asn	Leu	Gly	Ala	Pro	Ala	Ala	Phe	Lys	Tyr	Phe	Asp
				1315					1320					1325	
Thr	Thr	Ile	Asp	Arg	Lys	Arg	Tyr	Thr	Ser	Thr	Lys	Glu	Val	Leu	Asp
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Ala	Thr	Leu	Ile	His	Gln	Ser	Ile	Thr	Gly	Leu	Tyr	Glu	Thr	Arg	Ile
				1345					1350					1355	1360
Asp	Leu	Ser	Gln	Leu	Gly	Gly	Asp								
				1365											