

(19) **DANMARK**

(10) **DK/EP 3773291 T3**



(12) **Oversættelse af
europæisk patentskrift**

Patent- og
Varemærkestyrelsen

-
- (51) Int.Cl.: **A 61 B 18/12 (2006.01)** **A 61 N 1/32 (2006.01)**
- (45) Oversættelsen bekendtgjort den: **2021-11-15**
- (80) Dato for Den Europæiske Patentmyndigheds bekendtgørelse om meddelelse af patentet: **2021-10-27**
- (86) Europæisk ansøgning nr.: **19780002.2**
- (86) Europæisk indleveringsdag: **2019-08-22**
- (87) Den europæiske ansøgnings publiceringsdag: **2021-02-17**
- (86) International ansøgning nr.: **IB2019057091**
- (87) Internationalt publikationsnr.: **WO2020039390**
- (30) Prioritet: **2018-08-23 US 201862722100 P**
- (84) Designerede stater: **AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL PT RO RS SE SI SK SM TR**
- (73) Patenthaver: **Novocure GmbH, Park 6, 6039 Root D4, Schweiz**
- (72) Opfinder: **HAGEMANN Carsten, Calvadosstrasse 12, 97294 Unterpleichfeld, Tyskland**
LOHR Mario, Lohrenstrasse 1, 35685 Dillenburg, Tyskland
KESSLER Almuth F., Wirtsgasse 17a, 97234 Reichenberg, Tyskland
BUREK Malgorzata, Walther-von-Vogelweide-Str. 20, 97074 Wurzburg, Tyskland
FORSTER Carola, Plattnerstrasse 6, 97070 Wurzburg, Tyskland
BRAMI Catherine, Calanit 36, 45858 Matan, Israel
HERSHKOVICH Hadas Sara, Hachashmonaim 29/4, 2633730 Kiryat Motzkin, Israel
VOLOSHIN-SELA Tali, Kibbutz Gvat 1, 3657900 Kibbutz Gvat, Israel
- (74) Fuldmægtig i Danmark: **Plougmann Vingtoft A/S, Strandvejen 70, 2900 Hellerup, Danmark**
- (54) Benævnelse: **ANVENDELSE AF ALTERNERENDE ELEKTRISKE FELTER TIL AT FORØGE PERMEABILITET AF BLODHJERNEBARRIEREN**
- (56) Fremdragne publikationer:
EP-A1- 1 502 623
US-A1- 2003 191 506
US-A1- 2017 281 934

DESCRIPTION

BACKGROUND

[0001] Ordinarily, cerebral microvessels strictly regulate the transfer of substances between the blood and the brain tissue. This regulation by cerebral micro-vessels is called the blood-brain barrier (BBB), and is due to intercellular tight junctions (TJs) that form between brain capillary endothelial cells. In cerebral capillaries, TJs proteins are expressed 50-100 times more than in peripheral microvessels. TJs are formed by an intricate complex of transmembrane proteins (claudin and occludin) with cytoplasmic accessory proteins (ZO-1 and -2, cingulin, AF-6, and 7H6). By linking to the actin cytoskeleton, these proteins form a strong cell-cell connection. Brain endothelial cells, which form the endothelium of cerebral microvessels, are responsible for about 75-80% of the BBB's resistance to substances, and other cells such as astrocytes and pericytes provide the remainder of the resistance.

[0002] The BBB consists of tight junctions around the capillaries, and it ordinarily restricts diffusion of microscopic objects and large or hydrophilic molecules into the brain, while allowing for the diffusion of hydrophobic molecules (transcellular instead of paracellular transport).

[0003] In healthy people, the BBB serves a very important function because it prevents harmful substances (e.g. bacteria, viruses, and potentially harmful large or hydrophilic molecules) from entering the brain. There are, however, situations where the action of the BBB introduces difficulties. For example, it might be desirable to deliver large or hydrophilic drug molecules to treat a disease in the patient's brain. But when the BBB is operating normally, these drugs are blocked from entering the brain by the BBB.

[0004] US-A-2017/0281934 discloses an apparatus for inhibiting the spreading of cancer cells in a target region by imposing a first AC electric field having a frequency of 200 kHz \pm 10% in the target region for a first interval of time, and a second AC electric field having a frequency of 300 kHz \pm 10% in the target region for a second interval of time.

[0005] EP-A-1502623 discloses a transcranial electrostimulation apparatus which includes a generator of bipolar pulses of a first frequency, and a source of modulating control signals which produces an output at a second frequency, less than the first frequency, and causes the bipolar pulses to vary in amplitude in a predetermined asymmetrical pattern.

[0006] US-A-2003/0191506 discloses an apparatus for providing therapeutic electrical signals, which are generated by a signal generator applying a base wave within a frequency of about 150 kHz to 180 kHz at different frequencies between about 2 to 100 times in a second.

SUMMARY OF THE INVENTION

[0007] The present invention provides an apparatus for treating a tumor in a subject's body and facilitating delivery of a substance across a blood brain barrier of the subject's body according to claim 1.

BRIEF DESCRIPTION OF THE DRAWINGS

[0008]

FIG. 1 depicts an exemplary setup for in vitro experiments in which immortalized murine brain capillary endothelial cells (cerebEND) were grown on coverslips and transwell inserts to create an artificial in vitro version of the BBB.

FIGS. 2A and 2B depict the results of integrity and permeability testing, respectively, on the artificial BBB.

FIGS. 3A and 3B depict data showing that the increased permeability of the artificial BBB is not caused by cell death.

FIG. 4 depicts the positions at which rats' brains were sliced for an in vivo experiment.

FIG. 5 depicts the EB accumulation for this in vivo experiment in different sections of the rats' brains.

FIG. 6 depicts the average EB accumulation in the rat brain for this in vivo experiment, averaged over all sections.

FIG. 7 depicts the increase in BBB permeability in three different sections of rat brains that is induced by alternating electric fields in vivo, as determined using contrast enhancement MRIs.

FIG. 8 depicts the increase in BBB permeability in rat cortexes that is induced by alternating electric fields in vivo, as determined using contrast enhancement MRIs.

FIG. 9 depicts a suitable timing relationship between the application of the alternating electric field and the administration of the substance to the subject.

FIG. 10 is a block diagram of a dual-frequency apparatus that generates a first frequency for inducing BBB permeability and a second frequency for inducing cytotoxicity.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

[0009] This application describes a novel approach for temporarily increasing the permeability of the BBB using alternating electric fields so that substances that are ordinarily blocked by the

BBB will be able to cross the BBB.

[0010] A set of in vitro experiments was run in which immortalized murine brain capillary endothelial cells (cerebEND) were grown on coverslips and transwell inserts to create an artificial in vitro version of the BBB, and FIG. 1 depicts the setup for these experiments. The cells were then treated with alternating electric fields (100-300 kHz) for 24 h, 48 h, and 72 h. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions (i.e., 1 second in one direction followed by 1 second in the other direction, in a repeating sequence). The following effects were then analyzed: (a) Cell morphology (immunofluorescence staining of tight junction proteins Claudin 5 and ZO-1); (b) BBB integrity (using transendothelial electrical resistance (TEER)); and (c) BBB permeability (using fluoresceine isothiocyanate coupled to dextran (FITC) for flow cytometry).

[0011] A first set of experiments involved visualization of cell morphology and orientation, and visualization of the localization of stained proteins. This experiment was designed to ascertain how the frequency of the alternating electric field impacted the artificial BBB. Here, the cells were grown on coverslips, and alternating electric fields were applied for 72 hours at four different frequencies (100 kHz, 150 kHz, 200 kHz, and 300 kHz), with a field strength of 1.7 V/cm. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions. There was also a control in which alternating electric fields were not applied. Cell morphology images depicting the presence of Claudin 5, ZO-1, and 4,6-diamidino-2-phenylindole (DAPI) in (each of which was stained a different color) were then obtained. Claudin 5 and ZO-1 indicate the presence of an intact BBB. This set of cell morphology images revealed that alternating electric fields disturb the artificial BBB by delocalization of tight junction proteins from the cell boundaries to the cytoplasm, with the most dramatic effects at 100 kHz.

[0012] A second set of experiments also involved visualization of cell morphology. This experiment was designed to ascertain how the duration of time during which the alternating electric field was applied impacted the artificial BBB. Endothelial cells were grown on coverslips, and an alternating electric field at a frequency of 100 kHz was applied for three different durations (24 h, 48 h, 72 h) plus a control. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions. Cell morphology images depicting the presence of Claudin 5 and DAPI (each of which was stained a different color) were then obtained. This set of cell morphology images revealed that the phenomena discussed above in connection with the first set of experiments were already visible after 24 hours, and that the effects were most pronounced after 72 hours.

[0013] A third set of experiments also involved visualization of cell morphology. This experiment was similar to the second set of experiments, except that the endothelial cells were grown on transwell inserts instead of coverslips. The results were similar to the results of the second set of experiments. The delocalization of TJ proteins was visible after 24 hours and the effects were most pronounced after 72 hours. The three experiments described above support the conclusion that alternating electric fields cause structural changes in the cells, which might

be responsible for an increase in BBB permeability.

[0014] FIGS. 2A and 2B depict the results of integrity and permeability testing, respectively, on the artificial BBB after subjecting it to alternating electric fields at a frequency of 100 kHz for 72 hours (with the direction of the alternating electric fields switched every 1 second between two perpendicular directions), and for a control. More specifically, FIG. 2A depicts the results of a transendothelial electrical resistance (TEER) test, which reveals that alternating electric fields reduced the integrity of the artificial BBB to 35% of the control. FIG. 2B depicts the results of a fluoresceine isothiocyanate (FITC) permeability test, which reveals that the alternating electric fields increased the permeability of the artificial BBB to FITC-dextrans with a 4 kDa molecular weight to 110% of the control. These experiments further support the conclusion that alternating electric fields increase the permeability of the BBB to molecules that ordinarily cannot traverse a non-leaky BBB.

[0015] Collectively, these in vitro experiments reveal that applying alternating electric fields at certain frequencies for a sufficient duration of time causes the delocalization of tight junction proteins (Claudin 5, ZO-1) from the cell boundaries to the cytoplasm (with the most dramatic effects at 100 kHz), and increases the permeability of the BBB. The alternating electric fields' effects appear already after 24 h and are most prominent after 72 h. More specifically, after using the alternating electric fields to increase the permeability of the BBB, molecules of 4 kDa can pass through the BBB.

[0016] Additional in vitro experiments were then conducted to determine what happens to the BBB after the alternating electric fields were turned off. These experiments used visualization of cell morphology to show how the artificial BBB recovers after discontinuing the alternating electric fields. In these experiments, endothelial cells were grown on coverslips and treated with 100 kHz alternating electric fields at a field strength of 1.7 V/cm for 72 hours. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions. The alternating electric fields were then turned off, and the cells were followed for 96 hours after stopping the alternating electric field. Cell morphology images depicting the presence of Claudin 5 (stained) were obtained at 24 hours, 48 hours, 72 hours, and 96 hours. Those images revealed a progressive change in localization of Claudin between the cell boundaries and the cytoplasm on the 24 h, 48 h, 72 h, and 96 h images. Furthermore, a comparison of those four images to the respective images for the control (in which alternating electric fields were not applied during either the first 72 h or the last 96 h) revealed that the endothelial cell morphology was partially recovered 48 hours after stopping the alternating electric fields, and that the BBB was fully recovered (i.e., was comparable to the control) 96 hours after stopping the alternating electric fields.

[0017] FIGS. 3A and 3B depict the results of an in vitro experiment designed to determine whether the observed changes in the permeability of the artificial BBB described above might be attributable to cell death. This experiment tested cell division by comparing cell counts (a) when alternating electric fields were applied for 72 hours followed by no alternating electric fields for 96 hours with (b) a control in which alternating electric fields were never applied.

Endothelial cells were grown on coverslips and treated with 100 kHz alternating electric fields at a field strength of 1.7 V/cm for 72 hours. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions. The alternating electric fields were then turned off, and the cells were followed for 96 hours after stopping the alternating electric field. The number of cells per ml for the alternating electric fields and the control were counted, and the results are depicted in FIG. 3A and 3B (for control and for alternating electric fields, respectively). These results reveal that there was no statistically significant increase in the cell numbers during or after application of the alternating electric fields, which indicates that the changes in the BBB permeability noted above could not be attributed to cell death.

[0018] Another in vitro experiment used a TUNEL assay for apoptosis to determine whether the observed changes in the permeability of the artificial BBB described above might be attributable to cell death. In this experiment, endothelial cells were grown on coverslips and treated with 100 kHz alternating electric fields at a field strength of 1.7 V/cm for 72 hours. The direction of the alternating electric fields was switched every 1 second between two perpendicular directions. In the control, alternating electric fields were not applied. Cell morphology images depicting apoptosis (TUNEL) and Nuclei (DAPI) (each of which was stained a different color) were obtained after 24, 48, and 72 hours. None of those images revealed additional evidence of apoptosis, indicating that alternating electric fields did not cause cell death. This confirms that the changes in the BBB permeability noted above were not attributable to cell death.

[0019] A set of in vivo experiments on rats was also run to quantify the increase in vessel permeability caused by exposure to the alternating electric fields. These experiments used Evans Blue (EB) dye, which is an azo dye that has a very high affinity for serum albumin (molecule size ~ 69 kDa). Because of its large molecule size, serum albumin will ordinarily not be able to get past the BBB. But if the permeability of the BBB has been sufficiently increased, some of the serum albumin molecules (together with the EB dye that has been bound thereto) will make it across the BBB and can then be detected by looking for the EB in the rat's brain.

[0020] In this set of experiments, 100 kHz alternating electric fields were applied to the rat's brain for 72 hours, and the direction of the alternating electric fields was switched every 1 second between two perpendicular directions. This was accomplished by shaving each rat's head, positioning a first pair of capacitively coupled electrodes on the top and bottom of the rat's head, and positioning a second pair of capacitively coupled electrodes on the left and right sides of the rat's head. A 100 kHz AC voltage was then applied between the top and bottom electrodes for 1 second, followed by a 100 kHz AC voltage applied between the right and left electrodes for 1 second, in a repeating sequence.

[0021] Under the conditions indicated in Table 1 and for the times indicated on Table 1, EB was injected intravenously into the tail vein under anesthesia (Once injected, EB immediately binds to Albumin), and the EB was allowed to circulate for 2 hours in all cases. The following steps were then performed: (a) intracardiac perfusion with saline; (b) brains are sliced in four pieces with a brain slicer; (c) pieces were photographed to localize staining and weighted; (d) EB

extraction after tissue homogenization with TCA 50% (1:3) and centrifuge and (e) EB quantification at 610 nm. Results are given as μg EB per g tissue.

TABLE 1

Group #	Treatment	# of Rats	Time of EB Injection
1	72 hours 100 kHz fields	3	2h before the end of the 72 h period
2	72 hours 100 kHz fields + 2 hours rest	3	2h after the end of the 72 h period
3	72 hours heat using sham electrodes	3	2h before the end of heat
4	Control (no fields + no heat)	3	Same time as group 1

[0022] During the experiment, two animals from group 2 and one animal from group 4 were excluded (disrupted treatment, failure to inject EB into the tail vein). There were no differences between the animals treated with alternating electric fields (groups 1 and 2) and therefore these animals were grouped together. Similarly, there were no differences between sham heat and control animals (groups 3 and 4) and therefore these animals were grouped together.

[0023] The rats' brains were sliced into four pieces using a brain slicer at the positions shown in FIG. 4. EB accumulation in these four specific sections was then measured. In addition, a computer simulation was performed to determine the field strength in each of these four sections. Table 2 specifies the field strength obtained from the simulation in each of these four sections, with all values given in V/cm RMS.

TABLE 2

Section	1	2	3	4
Mean Field Strength	2.7 V/cm	3 V/cm	2.6 V/cm	1.6 V/cm
Media Field Strength	2.5 V/cm	2.6 V/cm	2.4 V/cm	1.6 V/cm

[0024] The results for EB accumulation in sections 1 through 4 are depicted in FIG. 5. The summary of these results is as follows: (1) a statistically significant increase was observed in sections 1, 2 (frontal cerebrum) where the field strength was highest; and a smaller increase (that was not statistically significant) was observed in the more posterior sections (3, 4) where the field strength was lower.

[0025] FIG. 6 depicts the average EB accumulation in the rat brain, averaged over all four sections 1-4. This result reveals higher accumulation of EB in the brains of rats treated with alternating electric fields for 72 hours, and this result was statistically significant ($p < 0.05$).

[0026] The in vivo experiments described above establish that: (1) alternating electric fields application permits the BBB passage of molecules of average molecular size of ~ 69 kDa to the

brain tissue; (2) the increase in permeability of the BBB is maintained 2 hours after terminating the alternating electric fields application; and (3) the increased permeability of the BBB varies between different sections of the brain. The latter may be the result of the different field strengths that were imposed in the various sections of the brain. These experiments further support our conclusion that alternating electric fields increase the permeability of the BBB to molecules that ordinarily cannot traverse a non-leaky BBB.

[0027] In another set of in vivo experiments, 5 rats were treated with alternating electric fields at 100 kHz for 72 h, and 4 control rats were not treated with alternating electric fields for the same period of time. At the end of the 72 hour period, the fluorescent compound TRITC-Dextran of 4 kDa was injected intravenously into the tail vein under anesthesia, and allowed to circulate for 2 minutes in all cases. The brains were then removed, frozen, sectioned and scanned with a fluorescent scanner. All slides were scanned with the same conditions. The resulting images revealed significantly higher levels of accumulation of the fluorescent 4 kDa TRITC-Dextran in the brain tissue of the rats that were subjected to alternating electric fields (as compared to the control), confirming yet again that alternating electric fields increase the permeability of the BBB.

[0028] Yet another set of in vivo experiments was performed using Dynamic Contrast Enhanced MRI (DCE-MRI) with intravenous injection of Gadolinium contrast agent (Gd-DTPA, Magnetol, MW 547). In these experiments, test rats were treated with 100 kHz alternating electric fields for 72 h, and control rats were not treated with alternating electric fields for the same period of time. After this 72 h period, the alternating electric field was turned off, the rats were anesthetized, and a series of 60 T1w MRI scans (each of the scans having a duration of 28 seconds) was acquired. The gadolinium contrast agent was injected into the rat's tail vein during the 7th of these 60 scans.

[0029] The image analysis for each rat included (1) determining a baseline for each voxel by calculating the mean of the first six T1w MRI scans for each voxel (i.e., the scans prior to the injection of the gadolinium); (2) computing voxel by voxel, the percent signal change (i.e., gadolinium accumulation) over time relative to the baseline; (3) segmenting the brain into anterior, middle, and posterior segments; (4) generating for each of the three segments the mean percent signal change with respect to the baseline over all the voxels in the respective segment and then (5) averaging 4 consecutive time points (i.e. 4 scans) together. Finally, the data from all of the rats within any given group were averaged together.

[0030] The results of this DCE-MRI experiment for each of the three segments of the brain (i.e. anterior, middle, and posterior) are depicted in FIG. 7. This data reveals that contrast agent accumulation in the brain tissue of rats that were treated with alternating electric fields (trace labeled TTFIELDS; n=6) was significantly higher than in the control rats (trace labeled control; n=3). Moreover, the distinction was most pronounced in the posterior brain, which is the portion of the brain where the alternating electric fields had the highest field strength. From this we can conclude that the alternating electric fields successfully increased the permeability of the BBB in vivo.

[0031] To test whether this increase in permeability of the BBB was temporary, the same test conditions were repeated, but followed with an additional 96 hours without alternating electric fields. After this 96 hour period, a series of 60 T1w MRI scans (each of the scans having a duration of 28 seconds) was acquired using the same procedure described above (including the gadolinium injection). The results of this portion of the DCE-MRI experiment for each of the three segments of the brain are also depicted in FIG. 8. This data reveals that contrast agent accumulation in the brain tissue of rats that were treated with alternating electric fields for 72 hours followed by 96 hours without alternating electric fields (trace labeled TTFIELDS+96h; n=7) was not significantly different from the control rats (trace labeled control+96h; n=3). From this we can conclude that the permeability of the BBB returns to normal after the alternating electric fields are discontinued.

[0032] An additional series of 60 T1w MRI scans (each of the scans having a duration of 28 seconds) was also acquired using the same procedure before the alternating electric field was applied to the rats (n=2). The results of this portion of the DCE-MRI experiment for each of the three segments of the brain (i.e. anterior, middle, and posterior) are also depicted in FIG. 8 (see the trace labeled "before").

[0033] FIG. 8 shows the average of all 3 segments of the brain (i.e. anterior, middle, and posterior) for 72 h of TTFIELDS (n=6) and a control of no TTFIELDS for 72 h (n=3), with standard deviation bars. A paired t test was used to compare between the two groups, and $p < 0.0001$.

[0034] We note that the upper size limit of molecules that can pass through the BBB after applying the alternating electric fields has not yet been determined. But based on (a) the in vitro experiments described herein using FITC-dextran with a 4 kDa molecular weight and (b) the in vivo experiments described herein using EB (which binds to serum albumin having a molecule size of ~ 69 kDa), the upper limit appears to be at least about 69 kDa, and is most certainly at least 4 kDa.

[0035] The implications of being able to reversibly increase the permeability of the BBB at will are far-reaching, because it now becomes possible to deliver many substances across the BBB of a subject, despite the fact that those substances have at least one characteristic that ordinarily impedes the substance from crossing a non-leaky BBB. Many of these implications involve delivering a substance including but not limited to treating agents and diagnostic agents across a blood brain barrier of a subject's brain.

[0036] Examples include but are not limited to the following: delivering chemotherapeutic agents across the BBB to treat cancer (in this context, it may be possible to lower the dosage of drugs for the treatment of brain tumors and metastases with severe side effects in other parts of the body based on the increased permeability of the drugs to the brain); delivering antibodies and/or cell-based therapies across the BBB for immunotherapy; delivering contrast agents dyes, reporters, and markers across the BBB for diagnostic purposes and for research (e.g. monitoring brain activity); delivering antibacterial agents across the BBB to treat infectious

diseases; delivering anti-viral agents or virus neutralizing antibodies across the BBB to treat viral infections; delivering anti-parasitic agents across the BBB to treat parasites; delivering agents to treat neurodegenerative and autoimmune disease across the BBB; delivering psychiatric drugs; delivering anti-epileptic drugs; delivering hydrocephalus drugs; delivering stroke intervention and recovery drugs; delivering compounds that are lacking in the brain across the BBB to treat conditions in which those compounds are lacking (e.g., for treating Parkinson's disease, etc.).

[0037] While the testing described above was done in vitro and in live rats, it is expected that similar results will be obtained with other animals and with humans.

[0038] The methods described herein can also be applied in the in vivo context by applying the alternating electric fields to a live subject's brain. Imposing the electric field in the subject's brain will increase the permeability of the BBB, which will enable molecules that are ordinarily blocked or impeded by the BBB to get through. This may be accomplished, for example, by positioning electrodes on or below the subject's skin so that application of an AC voltage between selected subsets of those electrodes will impose the alternating electric fields in the subject's brain.

[0039] For example, one pair of electrodes could be positioned on the front and back of the subject's head, and a second pair of electrodes could be positioned on the right and left sides of the subject's head. In some embodiments, the electrodes are capacitively coupled to the subject's body (e.g., by using electrodes that include a conductive plate and also have a dielectric layer disposed between the conductive plate and the subject's body). But in alternative embodiments, the dielectric layer may be omitted, in which case the conductive plates would make direct contact with the subject's body. In another embodiment, electrodes could be inserted subcutaneously below a patient's skin.

[0040] An AC voltage generator applies an AC voltage at a selected frequency (e.g., 100 kHz, or between 50 and 190 kHz) between the right and left electrodes for a first period of time (e.g. 1 second), which induces alternating electric fields where the most significant components of the field lines are parallel to the transverse axis of the subject's head. Then, the AC voltage generator applies an AC voltage at the same frequency (or a different frequency) between the front and back electrodes for a second period of time (e.g. 1 second), which induces alternating electric fields where the most significant components of the field lines are parallel to the sagittal axis of the subject's head. This two step sequence is then repeated for the duration of the treatment. Optionally, thermal sensors may be included at the electrodes, and the AC voltage generator can be configured to decrease the amplitude of the AC voltages that are applied to the electrodes if the sensed temperature at the electrodes gets too high. In some embodiments, one or more additional pairs of electrodes may be added and included in the sequence. In alternative embodiments, only a single pair of electrodes is used, in which case the direction of the field lines is not switched. Note that any of the parameters for this in vivo embodiment (e.g., frequency, field strength, duration, direction-switching rate, and the placement of the electrodes) may be varied as described above in connection with the in the

vitro embodiments. But care must be taken in the in vivo context to ensure that the electric field remains safe for the subject at all times.

[0041] A wide variety of applications for increasing the permeability of the BBB can be readily envisioned in the in vivo context. In one example, localized enhancement of drug uptake by tumor cells (e.g., glioblastoma cells) in the brain can be induced by applying alternating electric fields to the brain for a period of time (e.g., 72 hours or at least 24 hours) prior to and during administration of chemotherapies or other antineoplastic agents.

[0042] FIG. 9 depicts a suitable relationship in timing between the application of the alternating electric field and the administration of the substance to a live patient. Based on the data described above, and assuming that the substance is introduced or administered at a given time $t=0$, the alternating electric field can begin before the given time (e.g., 72 hours before $t=0$), and continue for an interval of time after the given time (e.g., until 12 hours following $t=0$). In this situation, the permeability of the BBB will begin to increase before the substance is administered and before the substance reaches the BBB. This will enable the substance to cross the BBB immediately upon its arrival. In the context of chemotherapy, this would correspond to starting application of the alternating electric fields, administering the chemotherapeutic agent 72 hours later, followed by application of the alternating electric fields for an additional interval of time (e.g., until 12 hours following the time at which the chemotherapeutic agent was administered).

[0043] Note that the intervals of time discussed above in connection with FIG. 9 can either be uninterrupted or can include breaks that are preferably short. For example, a 12 hour interval could be satisfied by a single uninterrupted block of 12 hours. Alternatively, the 12 hour interval could be satisfied by applying the alternating electric fields for 6 hours, followed by a 1 hour break, followed by applying the alternating electric fields for an additional 6 hours. Similar breaks may also optionally interrupt the 72 hour interval that precedes the administration of the substance. Note also that in the context of FIG. 9, when the substance is administered to a live patient, the administration of the substance may be performed using any of a variety of approaches including but not limited to intravenously, orally, subcutaneously, intrathecal, intraventricularly, and intraperitoneally.

[0044] In some preferred embodiments, the frequency of the alternating electric fields is less than 190 kHz (e.g., between 50 and 190 kHz or between 25 and 190 kHz). Based on the experiments discussed above, using a frequency of less than 190 kHz combined with a period of time of at least 24 hours will increase the change in permeability (as compared to operating outside of those ranges).

[0045] The methods described herein may be used to deliver a substance across the blood brain barrier of a subject's brain when the subject's brain includes a tumor. One existing approach to treating brain tumors (e.g., glioblastoma) is by applying alternating electric fields at frequencies between 50 and 500 kHz, preferably between 100 and 300 kHz to the tumor. For glioblastoma, 200 kHz is the most preferred frequency. Alternating electric fields at these

frequencies are referred to as TTFIELDS, and are described in US patents 6,868,289 and 7,565,205.

[0046] Briefly, those two applications describe disrupting dividing cells during mitosis. The effectiveness of TTFIELDS is improved when the direction of the electric field is periodically switched, when the strength of the field in at least a portion of the tumor is at least 1 V/cm, and when the fields are applied for long periods of time (e.g., weeks or months) with as few breaks as possible.

[0047] In patients with brain tumors, situations may arise where it will be desirable to treat the tumor with TTFIELDS and also deliver a substance across the same patient's blood brain barrier (e.g., to help get a therapeutically effective amount of a chemotherapy drug past the BBB to provide an additional line of attack against the tumor). In some situations, it may be possible to use a single frequency of an alternating electric field to both treat the tumor and increase the permeability of the BBB. In other situations, it may be desirable to use alternating electric fields with different frequencies: a first frequency that is selected to provide improved results for increasing the permeability of the BBB, and a second frequency that is selected to provide improved results for the anti-tumor action of the TTFIELDS.

[0048] FIG. 10 is a block diagram of an apparatus that generates a first frequency for inducing BBB permeability and a second frequency for inducing cytotoxicity. The apparatus includes an AC voltage generator 44 that is similar to the conventional Optune® field generator unit, but has the ability to operate at two different frequencies. The first frequency is between 50 and 190 kHz and the second frequency is between 50 and 500 kHz. In some embodiments, the first frequency is between 75 kHz and 125 kHz and the second frequency is between 150 kHz and 250 kHz.

[0049] The ability to operate at two different frequencies may be implemented, for example, using relays to switch either a first set of components or a second set of components into the conventional circuit that generates the AC voltage, and adjusting the operating frequency of an oscillator. The AC voltage generator 44 is configured to output either the first frequency or the second frequency depending on the state of a control input. When the control input is in a first state the AC voltage generator 44 outputs the first frequency, and when the control input is in a second state the AC voltage generator 44 outputs the second frequency. A controller 42 is programmed to place the control input in the second state so that the AC voltage generator 44 outputs the second frequency. The controller 42 is also programmed to accept a request to switch to the first frequency. In the embodiment depicted in FIG. 10, the request arrives via a user interface 40 that may be implemented using any of a variety of conventional approaches including but not limited to a pushbutton, a touch screen, etc. In alternative embodiments, the request may arrive via RF (e.g. Bluetooth, WiFi, etc.) from a tablet, smartphone, etc.

[0050] Upon receipt of the request, the controller 42 will place the control input in the first state so that the AC voltage generator 44 will output the first frequency for an interval of time (e.g., 72 hours). After the interval of time has elapsed, the controller 42 will place the control input in

the second state so that the AC voltage generator 44 reverts to outputting the second frequency.

[0051] Optionally, the AC voltage generator 44 may be configured to output one or more additional frequencies (e.g., a third frequency, a fourth frequency, etc.), depending on the state of the control input. Preferably each of these additional frequencies is selected to induce cytotoxicity. In these embodiments, the controller 42 is programmed to cycle the control input through the states that cause the AC voltage generator 44 to output the second frequency and the one or more additional frequencies before the request arrives. The controller 42 is also programmed to accept a request to switch to the first frequency. Upon receipt of the request, the controller 42 will place the control input in the first state so that the AC voltage generator 44 will output the first frequency for an interval of time (e.g., 72 hours). After the interval of time has elapsed, the controller 42 will revert to cycling the control input through the states that cause the AC voltage generator 44 to output the second frequency and the one or more additional frequencies.

[0052] The system depicted in FIG. 10 is particularly useful when a person has a tumor that is being treated by combination therapy that includes TTFIELDS and chemotherapy. In this situation, the system operates most of the time at the second frequency to provide the maximum cytotoxicity effect. But before a person visits a chemotherapy clinic for a dose of chemotherapy, healthcare personnel (or the user) actuates the user interface 40 to switch the system to the first frequency that promotes BBB permeability. In this situation, the actuation of the user interface could be done e.g., 72 hours before the expected start of the chemotherapy.

[0053] Alternatively, upon receipt of the request (e.g., from the user interface 40), the controller 42 can control the control input so that the AC voltage generator 44 will output the first frequency for an interval of time (e.g., 1 hour), then switch back and forth between the second frequency and the first frequency (e.g., switching every hour). Eventually (e.g., when the relevant substance has been exhausted from the patient's bloodstream), the controller 42 controls the control input so that the AC voltage generator 44 reverts to outputting the second frequency.

[0054] A set of electrodes (not shown) that are similar to the conventional electrodes used with Optune® are connected to the output of the AC voltage generator 44.

[0055] Note that in connection with any of the methods described above, the BBB should recover to its original low-permeability state after a sufficient amount of time has elapsed following the termination of the alternating electric field. This can be important in many contexts for the safety of the subject.

[0056] While the present invention has been disclosed with reference to certain embodiments, numerous modifications, alterations, and changes to the described embodiments are possible without departing from the scope of the present invention, as defined in the appended claim.

REFERENCES CITED IN THE DESCRIPTION

Cited references

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Patent documents cited in the description

- [US20170281934A \[0004\]](#)
- [EP1502623A \[0005\]](#)
- [US20030191506A \[0006\]](#)
- [US6868289B \[0045\]](#)
- [US7565205B \[0045\]](#)

Patentkrav

1. Apparat til at behandle en tumor i et individs krop og til at fremme levering af et stof over en blodhjernebarriere af individets krop, hvilket apparat omfatter:

- 5 en vekselspændingsgenerator (44) kan som fungere ved en første frekvens mellem 50 og 190 kHz og en anden frekvens mellem 50 og 500 kHz, idet den anden frekvens er forskellig fra den første frekvens, hvor vekselspændingsgeneratoren (44) har et kontrolinput og er konfigureret til at producere den første frekvens, når kontrolinputtet er i en første tilstand og producere den anden frekvens, når kontrolinputtet er i en anden
- 10 tilstand; og en styreenhed (42) programmeret til (a) at placere kontrolinputtet i den anden tilstand, således at vekselspændingsgeneratoren (44) producerer den anden frekvens, (b) at acceptere en anmodning om at omstille vekselspændingsgeneratoren (44) til at producere den første frekvens, (c)
- 15 efter modtagelse af anmodningen, at placere kontrolinputtet i den første tilstand, således at vekselspændingsgeneratoren (44) producerer den første frekvens i et tidsinterval, og (d) efter tidsintervallet er udløbet, at placere kontrolinputtet i den anden tilstand, således at vekselspændingsgeneratoren (44) producerer den anden frekvens;
- 20 **kendetegnet ved, at** tidsintervallet er mindst 72 timer.

DRAWINGS

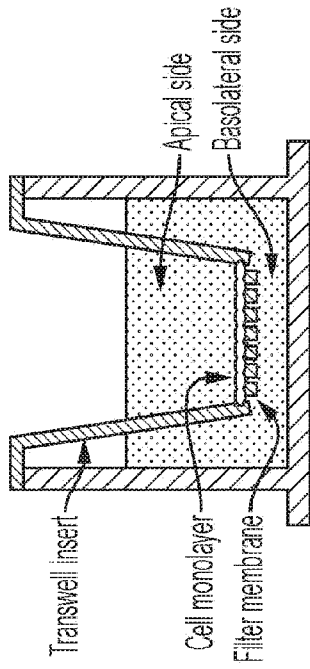


FIG. 1

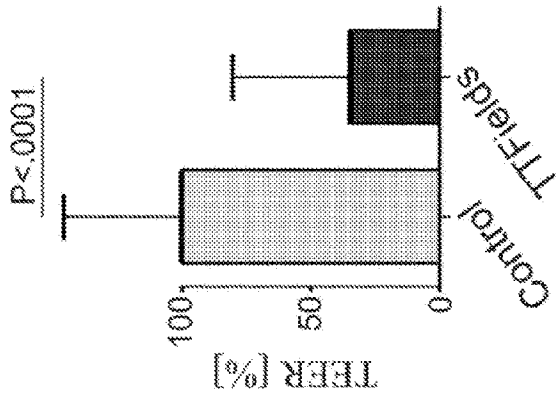


FIG. 2A

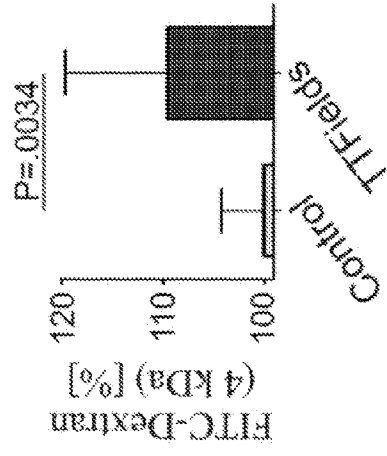


FIG. 2B

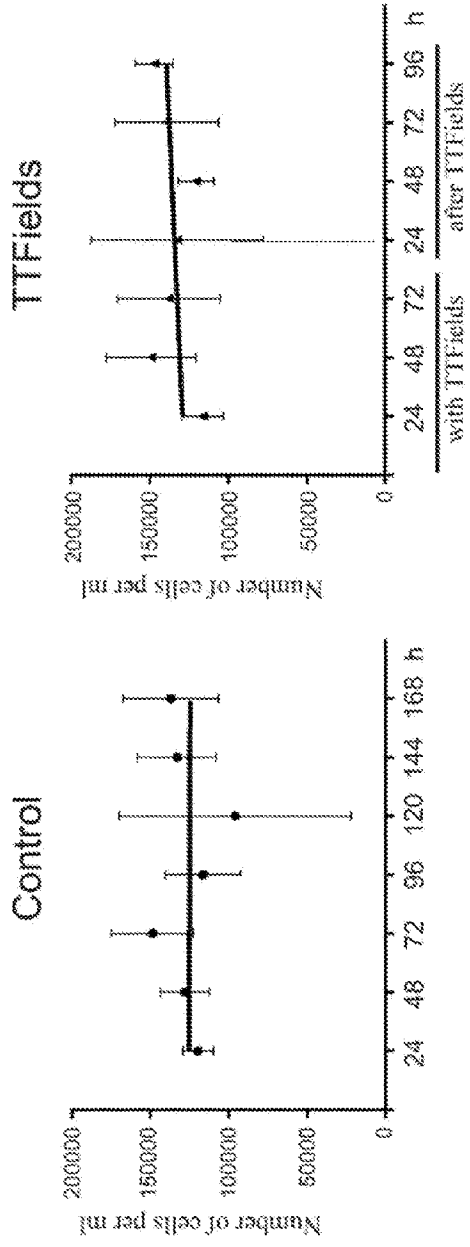


FIG. 3A

FIG. 3B

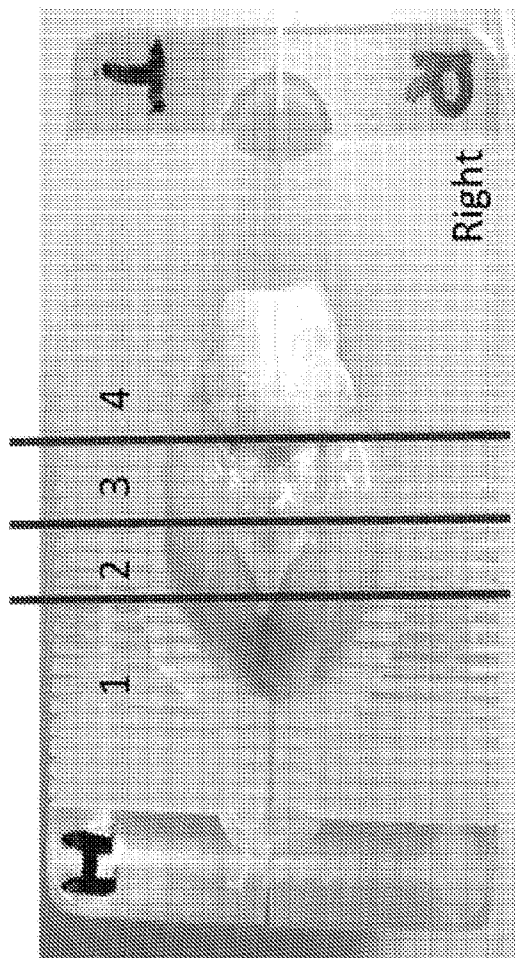


FIG. 4

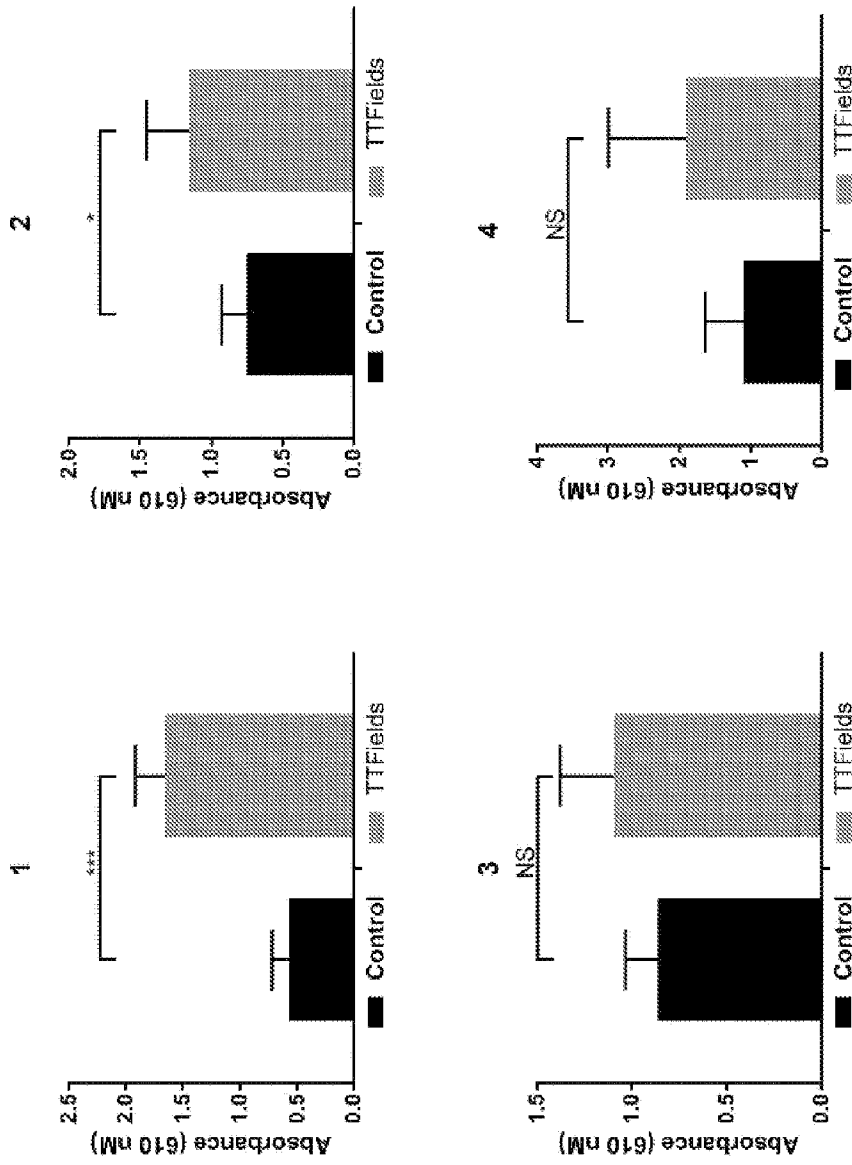


FIG. 5

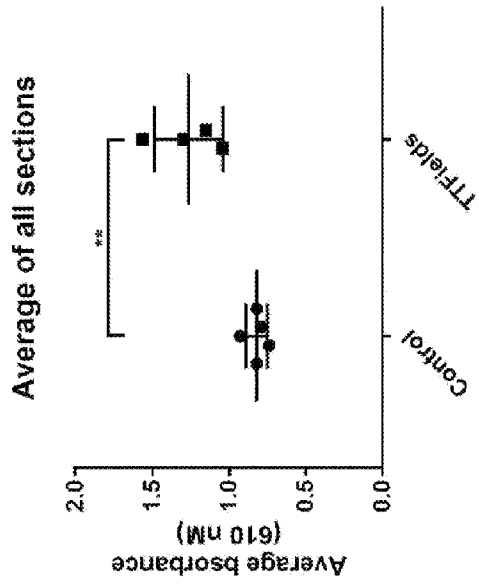


FIG. 6

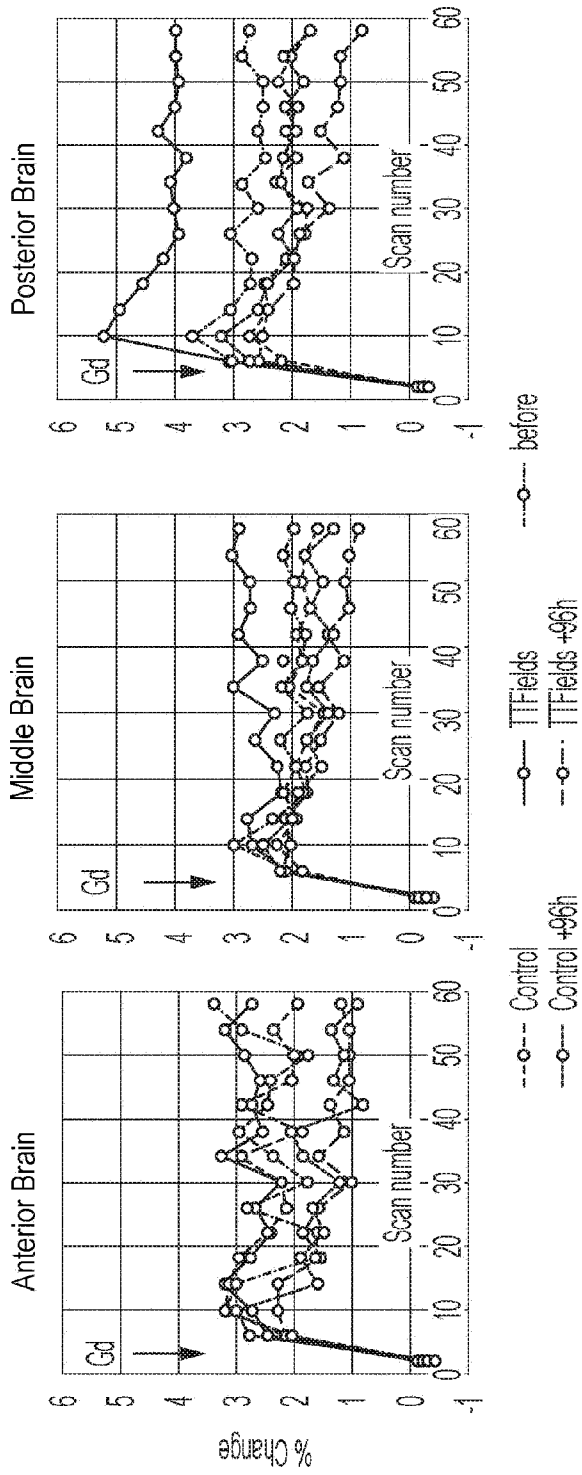


FIG. 7

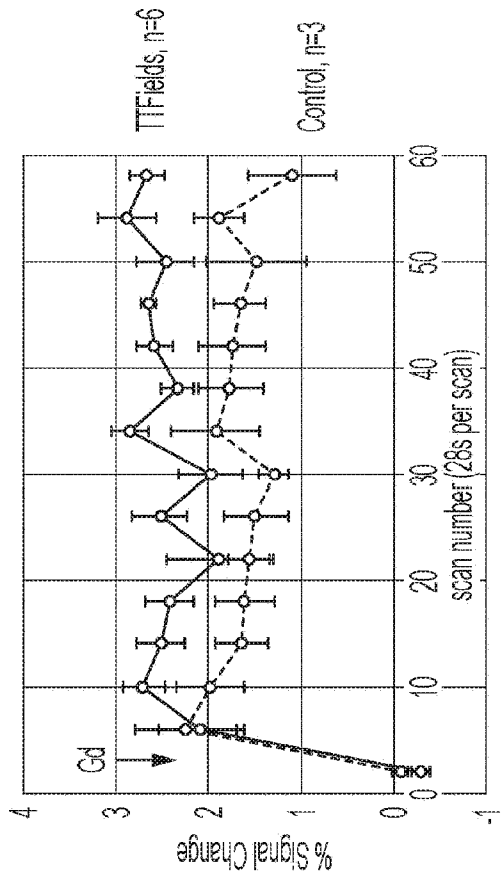


FIG. 8

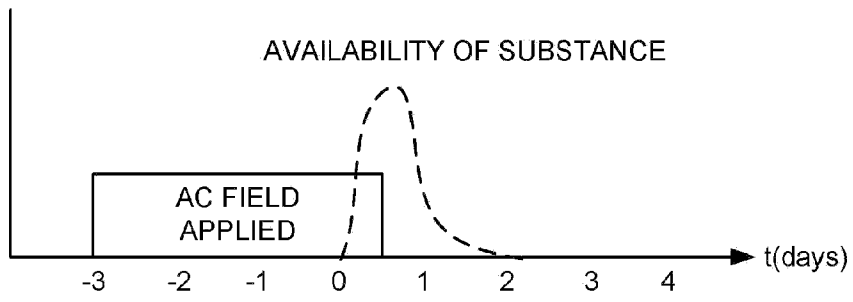


FIG. 9

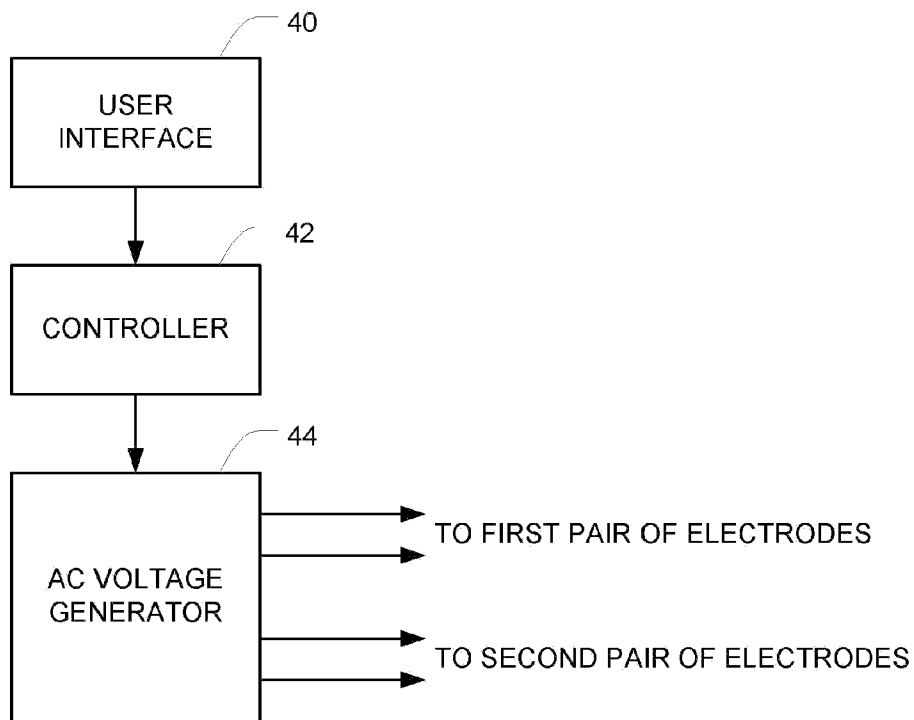


FIG. 10