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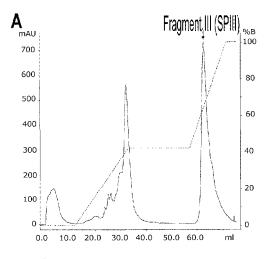
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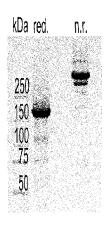


Fig. 1

(57) Abstract: A composition comprising a complex of Factor VIII and one or more Von Willebrand Factor peptides, wherein the Von Willebrand Factor peptides comprise at least the amino acids 764 to 1035 and 1691 to 1905 of SEQ ID No. 1 but not amino acids 2255 to 2645 of SEQ ID No. 1.

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Preparation comprising Factor VIII and Von Willebrand factor peptides

The present invention relates to pharmaceutical preparations for treating bleeding disorders.

Any discussion of the prior art throughout the specification should in no way be considered as an admission that such prior art is widely known or forms part of common general knowledge in the field.

Background of the invention

Factor VIII ("FVIII") is a blood plasma glycoprotein of about 280 kDa molecular mass. It is involved in the cascade of coagulation reactions that lead to blood clotting. The most common bleeding disorder is caused by a deficiency of functional Factor VIII, called haemophilia A. It is treated with replacement of Factor VIII, either plasma derived or recombinant. Factor VIII is used for acute 15 and prophylactic treatment of bleedings in haemophilia A patients.

The amino acid sequence of Factor VIII is organized into three structural domains: a triplicated A domain of 330 amino acids, a single B domain of 980 amino acids, and a duplicated C domain of 150 amino acids. The B domain has no homology to other proteins and provides 18 of the 25 potential asparagine(N)-linked glycosylation sites of this protein. The B domain has apparently no function in coagulation. B-domain deleted Factor VIII molecules have unchanged procoagulant activity compared to full-length Factor VIII. Some recombinant Factor VIII (rFVIII) preparations are B-domain deleted.

In plasma, Factor VIII is stabilized by association with Von Willebrand Factor protein ("vWF"), which appears to inhibit clearance of Factor VIII e.g. by proteolysis or receptor-mediated clearance via the LRP-receptor. In circulation, Von Willebrand Factor is present in a 50-fold molar excess relative to Factor VIII under normal physiological conditions.

Von Willebrand Factor is a multimeric adhesive glycoprotein present in the plasma of mammals, which has multiple physiological functions. During primary hemostasis, Von Willebrand Factor acts as a mediator between specific receptors on the platelet surface and components of the extracellular matrix such as collagen. Moreover, Von Willebrand Factor serves as a carrier and stabilizing protein for procoagulant Factor VIII. Von Willebrand Factor is synthesized in endothelial cells and megakaryocytes as a 2813 amino acid precursor molecule. The precursor polypeptide, pre-pro-Von Willebrand Factor, consists of a 22residue signal peptide, a 741 - residue pro-peptide and the 2050-residue polypeptide found in mature plasma Von Willebrand Factor (Fischer et al., FEBS Lett. 351: 345-348, 1994). Upon secretion into plasma, Von Willebrand Factor circulates in the form of various species with different molecular sizes. These Von Willebrand Factor molecules consist of oligo- and multimers of the mature subunit of 2050 amino acid residues. Von Willebrand Factor can be usually found in plasma as multimers ranging in size approximately from 500 to 20.000 kDa (Furlan, Ann Hematol. 1996 Jun; 72(6): 341-8)

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The average in vivo half-life of human Factor VIII in the human circulation is approximately 12 hours. Von Willebrand Factor might decrease possible immunoreactions against Factor VIII when in complex with Factor VIII by shielding FVIII from known potential inhibitor antibody sites on the heavy chain (A2 domain) and the light chain (A3/C2 domain) (Ragni, J Thromb.

Haemost. 10: 2324-2327, 2012) or on other potential antibody inhibitor sites on the Factor VIII molecule.

A further bleeding disorder in humans is Von Willebrand's disease (vWD). Depending on the severity of the bleeding symptoms, vWD can be treated by replacement therapy with concentrates containing Von Willebrand Factor, in general derived from plasma but recombinant Von Willebrand Factor also is under development. Von Willebrand Factor is known to stabilize Factor VIII in vivo and, thus, plays a crucial role to regulate plasma levels of Factor VIII and as a consequence is a central factor to control primary and secondary haemostasis.

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Until today, the standard treatment of Haemophilia A and vWD involves frequent intravenous infusions of preparations of Factor VIII and Factor VIII/Von Willebrand Factor concentrates. These replacement therapies are generally effective, however, for example in severe haemophilia A patients undergoing prophylactic treatment Factor VIII has to be administered intravenously (i.v.) about 3 times per week due to the short plasma half life of Factor VIII of about 12 hours. Already by achieving Factor VIII levels above 1% of normal human plasma corresponding to a raise of Factor VIII levels by 0.01 U/ml, severe haemophilia A is turned into moderate haemophilia A. In prophylactic therapy, the dosing regime is designed such that the levels of Factor VIII activity do not fall below levels of 2-3% of the Factor VIII activity of non-haemophiliacs.

The administration of a Factor VIII via intravenous administration (i.v.) is cumbersome, associated with pain and entails the risk of an infection especially as this is mostly done in home treatment by the patients themselves or by the parents of children being diagnosed for haemophilia A. In addition, frequent intravenous injections inevitably result in scar formation, interfering with future infusions. Still, i.v. treatment might be needed in emergency situation or surgery, i.e. when a high Factor VIII-level is needed immediately.

Subcutaneous administration (s.c.) has been proposed for Factor VIII, e.g. in WO 95/01804 A1 and WO 95/026750 A1. However, very high doses of Factor VIII had to be administered to achieve an acceptable bioavailability.

Another approach to improve the bioavailability upon non-intravenous administration has been to use albumin-fused Factor VIII (WO 2011/020866 A2).

WO 2013/057167 A1 proposes to administer Factor VIII in combination with sulphated glycosaminoglycans via non-intravenous administration, optionally together with Von Willebrand Factor.

40 WO 2008/151817 A1 describes the general use of uncleaved Von Willebrand Factor multimers for stabilisation of Factor VIII, plasma derived or recombinant (full-length and deletion mutants) intended for extravascular treatment.

WO 2013/160005 A1 describes the general use of recombinant Von Willebrand Factor or recombinant Von Willebrand Factor-fragments to improve bioavailability after s.c. treatment for very specific Factor VIII molecules, wherein the said Factor VIII molecules comprise a truncated B domain at a

size of 100-400 amino acids. According to WO 2013/160005 A1 Factor VIII molecules with truncated B domains between 100 and 400 amino acids have a higher Factor VIII bioavailability compared to Factor VIII having the entire B domain or B domain truncated Factor VIII molecules having no or only a few amino acids.

There is still a need for Factor VIII preparations showing improved bioavailability, stability and/or lower risk for antibody generation thereby avoiding drawbacks of prior art.

It is an object of the present invention to overcome or ameliorate at least one of the disadvantages of the prior art, or to provide a useful alternative.

Unless the context clearly requires otherwise, throughout the description and the claims, the words "comprise", "comprising", and the like are to be construed in an inclusive sense as opposed to an exclusive or exhaustive sense; that is to say, in the sense of "including, but not limited to".

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According to a first aspect, the present invention provides a composition comprising a complex of Factor VIII and one or more Von Willebrand Factor peptides, wherein the Von Willebrand Factor peptides comprise at least the amino acids 764 to 1035 and 1691 to 1905 of SEQ ID No. 1 but not amino acids 2255 to 2645 of SEQ ID No. 1.

According to a second aspect, the present invention provides use of the composition of the invention in the manufacture of a medicament for the treatment or prevention of a bleeding disorder.

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According to a third aspect, the present invention provides a method of treating or preventing a bleeding disorder comprising the administration of the composition of the invention.

30 According to a fourth aspect, the present invention provides a method for virus reduction during preparation of the composition of the invention comprising the step of nanofiltrating the Von Willebrand Factor peptides prior to or after combination with Factor VIII, whereby porcine parvovirus, if present, is reduced by at least a factor of 100.

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Provided are alternative Factor VIII preparations. Preferably, these preparations should show improved stability, improved bioavailability and/or reduced risk for immunological reactions.

In one embodiment, provided is a composition comprising a complex of Factor 40 VIII and one or more Von Willebrand Factor peptides, wherein the Von Willebrand Factor peptides comprise at least the amino acids 764 to 1035 and 1691 to 1905 of SEQ ID No. 1 but not amino acids 2255 to 2645 of SEQ ID NO 1.

According to the present invention, a Factor VIII preparation comprising Von Willebrand Factor peptides is provided. Factor VIII form a complex with the comprising Von Willebrand Factor peptides.

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Factor VIII as used herein covers full-length Factor VIII, B domain deleted Factor VIII or a Factor VIII wherein the B domain has been replaced by an artificial linker or a fragment of the natural B domain or a combination of both, i.e. the Bdomain has a different size compared to full-length Factor VIII. It also covers Factor VIII with a limited number of modifications having insertion, deletion or substitutions, especially Factor VIII adapted to haplotypes as described in K.R. Viel, et al. New England J Med 2009; 360:1618-1627. Preferably, the sequence homology to Factor VIII (as defined in amino acids 20-2351 of P00451 of SwissProt July 21, 1986) but disregarding the homology in the B-Domain of 99% according to FASTA as implemented in FASTA version 36, based on W. R. Pearson (1996) "Effective protein sequence comparison" Meth. Enzymol. 266:227-258. In other words, when calculating a sequence homology, the Bdomain is not included in the comparison of both proteins. Also covered is modified Factor VIII, like HES-Factor VIII or PEG Factor VIII or Factor VIII Fc fusion proteins and Factor VIII albumin fusion proteins as described in Oldenburg, Haemophilia (2014), 20 (Suppl. 4), 23-28.

The Factor VIII of the present invention may be plasma derived or recombinant Factor VIII. When recombinant Factor VIII is used, it is preferably expressed in a human cell line to mimic human glycosylation pattern (Casademunt, Eur J Haematol. 2012; 89:165-76) or as described in WO 2010/020690.

Von Willebrand Factor peptides as used herein are peptides comprising at least amino acids 764 to 1035 of SEQ ID No. 1 and 1691 to 1905 of SEQ ID No. 1 in a single amino acid chain. These amino acids may be part of a longer sequence comprising both of these sequences together. In other words, the Von Willebrand peptides of the invention comprise both SEQ ID No. 5 and SEQ ID No. 6. They may comprise further parts of Von Willebrand Factor, excluding all the amino acids 2255 to 2645 (SEQ ID No. 7). The Von Willebrand peptides

may comprise other sequences that are part of SEQ ID No. 1 or sequences that are not part of SEQ ID No. 1, e.g. amino acid linkers or the like. Preferably, the total amount of amino acids that are not part of SEQ ID No. 1 ist not more than 50, not more than 20 or not more than 10 amino acids.

One important aspect of the invention is that amino acids 2255 to 2645 of SEQ ID No. 1 are not part of the Von Willebrand Factor peptides. In other words, the Von Willebrand Factor peptides do not comprise any sequence that has at least 90 % homology to SEQ ID No. 7 according to FASTA, described below.

SEQ ID No. 1 is sequence P04275 of Swiss Prot database as of January 11, 2011.

The Von Willebrand Factor peptides in the composition of the present invention may be peptides having the same sequence or may be a mixture of peptides having sequences as defined above.

Typically a molecular ratio of Factor VIII and Von Willebrand Factor peptides will be between 1:1 and 1:20, preferably 1:2 to 1:10. If the Von Willebrand factor peptides are in the form of dimers or multimers, the molecular ratio is calculated on a single amino acid chain, i.e. a complex of a Factor VIII molecule with a dimer of Von Willebrand factor peptides will have a ratio of 1:2.

A complex, as used herein refers to a non-covalent binding of Factor VIII to one or more Von Willebrand Factor peptides.

In a preferred embodiment of the invention, the Von Willebrand Factor peptides are fragments of Von Willebrand Factor, i.e. N-terminal and/or C-terminal truncated forms of Von Willebrand Factor.

In one embodiment, the fragments comprise amino acids 764 to 1905 of SEQ ID No. 1.

A further embodiment of the invention is a composition comprising a complex of Factor VIII and one or more Von Willebrand Factor peptides that are fragments of Von Willebrand Factor and have an amino acid sequence that corresponds to the amino acid sequence of SEQ ID NO 1 starting form amino acid 764 and ending between amino acid 1905 and 2153 with up to 20, or up to 10 modifications selected from amino acid deletions, amino acid insertions or amino acid substitutions.

Preferred Von Willebrand Factor peptides are:

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- Peptides having the sequence 764 to 1905 of SEQ ID No. 1
 Peptides having the sequence 764 to 1906 of SEQ ID No. 1
 Peptides having the sequence 764 to 1907 of SEQ ID No. 1
 Peptides having the sequence 764 to 1908 of SEQ ID No. 1
 Peptides having the sequence 764 to 1909 of SEQ ID No. 1
- Peptides having the sequence 764 to 1910 of SEQ ID No. 1
 Peptides having the sequence 764 to 1911 of SEQ ID No. 1
 Peptides having the sequence 764 to 1912 of SEQ ID No. 1
 Peptides having the sequence 764 to 1913 of SEQ ID No. 1
 Peptides having the sequence 764 to 1914 of SEQ ID No. 1
- Peptides having the sequence 764 to 1915 of SEQ ID No. 1 Peptides having the sequence 764 to 1916 of SEQ ID No. 1

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Peptides having the sequence 764 to 1917 of SEO ID No. 1 Peptides having the sequence 764 to 1918 of SEQ ID No. 1 Peptides having the sequence 764 to 1919 of SEQ ID No. 1 Peptides having the sequence 764 to 1920 of SEQ ID No. 1 5 Peptides having the sequence 764 to 1921 of SEQ ID No. 1 Peptides having the sequence 764 to 1922 of SEQ ID No. 1 Peptides having the sequence 764 to 1923 of SEQ ID No. 1 Peptides having the sequence 764 to 1924 of SEQ ID No. 1 Peptides having the sequence 764 to 1925 of SEQ ID No. 1 10 Peptides having the sequence 764 to 1926 of SEQ ID No. 1 Peptides having the sequence 764 to 1927 of SEQ ID No. 1 Peptides having the sequence 764 to 1928 of SEQ ID No. 1 Peptides having the sequence 764 to 1929 of SEQ ID No. 1 Peptides having the sequence 764 to 1930 of SEQ ID No. 1 15 Peptides having the sequence 764 to 1931 of SEQ ID No. 1 Peptides having the sequence 764 to 1932 of SEQ ID No. 1 Peptides having the sequence 764 to 1933 of SEQ ID No. 1 Peptides having the sequence 764 to 1934 of SEQ ID No. 1 Peptides having the sequence 764 to 1935 of SEQ ID No. 1 20 Peptides having the sequence 764 to 1936 of SEQ ID No. 1 Peptides having the sequence 764 to 1937 of SEQ ID No. 1 Peptides having the sequence 764 to 1938 of SEQ ID No. 1 Peptides having the sequence 764 to 1939 of SEO ID No. 1 Peptides having the sequence 764 to 1940 of SEQ ID No. 1 25 Peptides having the sequence 764 to 1941 of SEQ ID No. 1 Peptides having the sequence 764 to 1942 of SEQ ID No. 1 Peptides having the sequence 764 to 1943 of SEO ID No. 1 Peptides having the sequence 764 to 1944 of SEQ ID No. 1 Peptides having the sequence 764 to 1945 of SEQ ID No. 1 Peptides having the sequence 764 to 1946 of SEQ ID No. 1 30 Peptides having the sequence 764 to 1947 of SEQ ID No. 1 Peptides having the sequence 764 to 1948 of SEQ ID No. 1 Peptides having the sequence 764 to 1949 of SEQ ID No. 1 Peptides having the sequence 764 to 1950 of SEQ ID No. 1 Peptides having the sequence 764 to 1951 of SEO ID No. 1 35 Peptides having the sequence 764 to 1952 of SEO ID No. 1 Peptides having the sequence 764 to 1953 of SEQ ID No. 1 Peptides having the sequence 764 to 1954 of SEQ ID No. 1 Peptides having the sequence 764 to 1955 of SEQ ID No. 1 40 Peptides having the sequence 764 to 1956 of SEQ ID No. 1 Peptides having the sequence 764 to 1957 of SEQ ID No. 1 Peptides having the sequence 764 to 1958 of SEQ ID No. 1 Peptides having the sequence 764 to 1959 of SEQ ID No. 1 Peptides having the sequence 764 to 1960 of SEQ ID No. 1 45 Peptides having the sequence 764 to 1961 of SEQ ID No. 1 Peptides having the sequence 764 to 1962 of SEQ ID No. 1 Peptides having the sequence 764 to 1963 of SEQ ID No. 1 Peptides having the sequence 764 to 1964 of SEQ ID No. 1 Peptides having the sequence 764 to 1965 of SEQ ID No. 1 50 Peptides having the sequence 764 to 1966 of SEQ ID No. 1

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Peptides having the sequence 764 to 2017 of SEO ID No. 1 Peptides having the sequence 764 to 2018 of SEQ ID No. 1 Peptides having the sequence 764 to 2019 of SEQ ID No. 1 Peptides having the sequence 764 to 2020 of SEQ ID No. 1 5 Peptides having the sequence 764 to 2021 of SEQ ID No. 1 Peptides having the sequence 764 to 2022 of SEQ ID No. 1 Peptides having the sequence 764 to 2023 of SEQ ID No. 1 Peptides having the sequence 764 to 2024 of SEQ ID No. 1 Peptides having the sequence 764 to 2025 of SEQ ID No. 1 10 Peptides having the sequence 764 to 2026 of SEQ ID No. 1 Peptides having the sequence 764 to 2027 of SEQ ID No. 1 Peptides having the sequence 764 to 2028 of SEQ ID No. 1 Peptides having the sequence 764 to 2029 of SEQ ID No. 1 Peptides having the sequence 764 to 2030 of SEQ ID No. 1 15 Peptides having the sequence 764 to 2031 of SEQ ID No. 1 Peptides having the sequence 764 to 2032 of SEQ ID No. 1 Peptides having the sequence 764 to 2033 of SEQ ID No. 1 Peptides having the sequence 764 to 2034 of SEQ ID No. 1 Peptides having the sequence 764 to 2035 of SEQ ID No. 1 20 Peptides having the sequence 764 to 2036 of SEQ ID No. 1 Peptides having the sequence 764 to 2037 of SEQ ID No. 1 Peptides having the sequence 764 to 2038 of SEQ ID No. 1 Peptides having the sequence 764 to 2039 of SEO ID No. 1 Peptides having the sequence 764 to 2040 of SEQ ID No. 1 25 Peptides having the sequence 764 to 2041 of SEQ ID No. 1 Peptides having the sequence 764 to 2042 of SEQ ID No. 1 Peptides having the sequence 764 to 2043 of SEO ID No. 1 Peptides having the sequence 764 to 2044 of SEQ ID No. 1 Peptides having the sequence 764 to 2045 of SEQ ID No. 1 30 Peptides having the sequence 764 to 2046 of SEQ ID No. 1 Peptides having the sequence 764 to 2047 of SEQ ID No. 1 Peptides having the sequence 764 to 2048 of SEQ ID No. 1 Peptides having the sequence 764 to 2049 of SEQ ID No. 1 Peptides having the sequence 764 to 2050 of SEQ ID No. 1 Peptides having the sequence 764 to 2051 of SEO ID No. 1 35 Peptides having the sequence 764 to 2052 of SEO ID No. 1 Peptides having the sequence 764 to 2053 of SEQ ID No. 1 Peptides having the sequence 764 to 2054 of SEQ ID No. 1 Peptides having the sequence 764 to 2055 of SEQ ID No. 1 40 Peptides having the sequence 764 to 2056 of SEQ ID No. 1 Peptides having the sequence 764 to 2057 of SEQ ID No. 1 Peptides having the sequence 764 to 2058 of SEQ ID No. 1 Peptides having the sequence 764 to 2059 of SEQ ID No. 1 Peptides having the sequence 764 to 2060 of SEQ ID No. 1 45 Peptides having the sequence 764 to 2061 of SEQ ID No. 1 Peptides having the sequence 764 to 2062 of SEQ ID No. 1 Peptides having the sequence 764 to 2063 of SEQ ID No. 1 Peptides having the sequence 764 to 2064 of SEQ ID No. 1 Peptides having the sequence 764 to 2065 of SEQ ID No. 1 50 Peptides having the sequence 764 to 2066 of SEQ ID No. 1

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Peptides having the sequence 764 to 2067 of SEO ID No. 1 Peptides having the sequence 764 to 2068 of SEQ ID No. 1 Peptides having the sequence 764 to 2069 of SEQ ID No. 1 Peptides having the sequence 764 to 2070 of SEQ ID No. 1 5 Peptides having the sequence 764 to 2071 of SEQ ID No. 1 Peptides having the sequence 764 to 2072 of SEQ ID No. 1 Peptides having the sequence 764 to 2073 of SEQ ID No. 1 Peptides having the sequence 764 to 2074 of SEQ ID No. 1 Peptides having the sequence 764 to 2075 of SEQ ID No. 1 10 Peptides having the sequence 764 to 2076 of SEQ ID No. 1 Peptides having the sequence 764 to 2077 of SEQ ID No. 1 Peptides having the sequence 764 to 2078 of SEQ ID No. 1 Peptides having the sequence 764 to 2079 of SEQ ID No. 1 Peptides having the sequence 764 to 2080 of SEQ ID No. 1 15 Peptides having the sequence 764 to 2081 of SEQ ID No. 1 Peptides having the sequence 764 to 2082 of SEQ ID No. 1 Peptides having the sequence 764 to 2083 of SEQ ID No. 1 Peptides having the sequence 764 to 2084 of SEQ ID No. 1 Peptides having the sequence 764 to 2085 of SEQ ID No. 1 20 Peptides having the sequence 764 to 2086 of SEQ ID No. 1 Peptides having the sequence 764 to 2087 of SEQ ID No. 1 Peptides having the sequence 764 to 2088 of SEQ ID No. 1 Peptides having the sequence 764 to 2089 of SEO ID No. 1 Peptides having the sequence 764 to 2090 of SEQ ID No. 1 25 Peptides having the sequence 764 to 2091 of SEQ ID No. 1 Peptides having the sequence 764 to 2092 of SEQ ID No. 1 Peptides having the sequence 764 to 2093 of SEO ID No. 1 Peptides having the sequence 764 to 2094 of SEQ ID No. 1 Peptides having the sequence 764 to 2095 of SEQ ID No. 1 Peptides having the sequence 764 to 2096 of SEQ ID No. 1 30 Peptides having the sequence 764 to 2097 of SEQ ID No. 1 Peptides having the sequence 764 to 2098 of SEQ ID No. 1 Peptides having the sequence 764 to 2099 of SEQ ID No. 1 Peptides having the sequence 764 to 2100 of SEQ ID No. 1 Peptides having the sequence 764 to 2101 of SEO ID No. 1 35 Peptides having the sequence 764 to 2102 of SEO ID No. 1 Peptides having the sequence 764 to 2103 of SEQ ID No. 1 Peptides having the sequence 764 to 2104 of SEQ ID No. 1 Peptides having the sequence 764 to 2105 of SEQ ID No. 1 40 Peptides having the sequence 764 to 2106 of SEQ ID No. 1 Peptides having the sequence 764 to 2107 of SEQ ID No. 1 Peptides having the sequence 764 to 2108 of SEQ ID No. 1 Peptides having the sequence 764 to 2109 of SEO ID No. 1 Peptides having the sequence 764 to 2110 of SEQ ID No. 1 45 Peptides having the sequence 764 to 2111 of SEQ ID No. 1 Peptides having the sequence 764 to 2112 of SEQ ID No. 1 Peptides having the sequence 764 to 2113 of SEQ ID No. 1 Peptides having the sequence 764 to 2114 of SEQ ID No. 1 Peptides having the sequence 764 to 2115 of SEQ ID No. 1 50 Peptides having the sequence 764 to 2116 of SEQ ID No. 1 - 9 –

Peptides having the sequence 764 to 2117 of SEO ID No. 1 Peptides having the sequence 764 to 2118 of SEQ ID No. 1 Peptides having the sequence 764 to 2119 of SEQ ID No. 1 Peptides having the sequence 764 to 2120 of SEQ ID No. 1 5 Peptides having the sequence 764 to 2121 of SEQ ID No. 1 Peptides having the sequence 764 to 2122 of SEQ ID No. 1 Peptides having the sequence 764 to 2123 of SEQ ID No. 1 Peptides having the sequence 764 to 2124 of SEQ ID No. 1 Peptides having the sequence 764 to 2125 of SEQ ID No. 1 10 Peptides having the sequence 764 to 2126 of SEQ ID No. 1 Peptides having the sequence 764 to 2127 of SEQ ID No. 1 Peptides having the sequence 764 to 2128 of SEQ ID No. 1 Peptides having the sequence 764 to 2129 of SEQ ID No. 1 Peptides having the sequence 764 to 2130 of SEQ ID No. 1 15 Peptides having the sequence 764 to 2131 of SEQ ID No. 1 Peptides having the sequence 764 to 2132 of SEQ ID No. 1 Peptides having the sequence 764 to 2133 of SEQ ID No. 1 Peptides having the sequence 764 to 2134 of SEQ ID No. 1 Peptides having the sequence 764 to 2135 of SEQ ID No. 1 20 Peptides having the sequence 764 to 2136 of SEQ ID No. 1 Peptides having the sequence 764 to 2137 of SEQ ID No. 1 Peptides having the sequence 764 to 2138 of SEQ ID No. 1 Peptides having the sequence 764 to 2139 of SEO ID No. 1 Peptides having the sequence 764 to 2140 of SEQ ID No. 1 25 Peptides having the sequence 764 to 2141 of SEQ ID No. 1 Peptides having the sequence 764 to 2142 of SEQ ID No. 1 Peptides having the sequence 764 to 2143 of SEO ID No. 1 Peptides having the sequence 764 to 2144 of SEQ ID No. 1 Peptides having the sequence 764 to 2145 of SEQ ID No. 1 30 Peptides having the sequence 764 to 2146 of SEQ ID No. 1 Peptides having the sequence 764 to 2147 of SEQ ID No. 1 Peptides having the sequence 764 to 2148 of SEQ ID No. 1 Peptides having the sequence 764 to 2149 of SEQ ID No. 1 Peptides having the sequence 764 to 2150 of SEQ ID No. 1 Peptides having the sequence 764 to 2151 of SEO ID No. 1 35 Peptides having the sequence 764 to 2152 of SEO ID No. 1 Peptides having the sequence 764 to 2153 of SEQ ID No. 1

A further embodiment of the invention is a composition comprising a complex of Factor VIII with one or more Von Willebrand Factor peptides, wherein

- 40 the Von Willebrand factor peptides are fragments of Von Willebrand Factor
 - the complex of Factor VIII and the fragments of Von Willebrand Factor show a reduced binding to phospholipid membranes compared to Factor VIII alone
- the complex of Factor VIII and the fragments of Von Willebrand Factor show
 a reduced binding to collagen III compared to the complex of Factor VIII and full length Von Willebrand Factor
 - the complex of Factor VIII and the fragments of Von Willebrand Factor show a reduced binding to heparin compared to the complex of Factor VIII and full length Von Willebrand Factor.

Preferably, the Von Willebrand Factor peptides have a molecular weight < 500 kD, preferably < 400 kD. As the Von Willebrand Factor often forms oligomers or multimers, also the peptides of the present invention may be in the form of multimers or oligomers.

- In a preferred embodiment the peptides of the present invention have at least one property selected from the group consisting of
 - (i) an affinity binding constant for heparin of $K_D > 1\,$ nM, preferably \geq 2,43 nM
- (ii) an affinity binding constant for collagen III of $K_D > 5$ nM, preferably \geq 10 17.02 nM
 - (iii) an affinity binding constant for Factor VIII of $K_D\!\!<100$ nM or $<\!10$ nM, preferably ≤ 6.19 nM and
 - (iv) an inhibition of Factor VIII phospolipid binding binding of at least 70%, preferably at least 80% or at least 90%.
- 15 The Von Willebrand factor peptides of the invention show preferably a reduced binding to heparin, a lower affinity for collagen (like collagen III), a lower affinity to phospholipids but still a high binding to Factor VIII.
 - Surprisingly, low binding to phospolipids and collagen improves release rate in case of non-intravenous administration, especially subcutaneous.
- The measurement of the respective affinity binding constants is described in the experimental part.
 - In one embodiment, the Von Willebrand Factor peptides are derived from Von Willebrand Factor by proteolytic or chemical cleavage. If proteolytic cleavage is used, *S. aureus* V-8 protease is especially preferred.
- 25 Preferably, the composition of the present invention has at least one of the following properties:
 - (i) the Von Willebrand Factor peptides shield Factor VIII from antibody binding to minimize inhibitor formation in a patient
 - (ii) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 12 month in a frozen liquid form at 70 °C

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- (iii) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 24 month in a freeze-dried form at 5 °C
- (iv) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 12 month in a freeze-dried form at 25 °C
 - (v) prolonges half-life of Factor VIII in-vivo by at least 20 % and
- (vi) reduces inhibitor formation in previously untreated patients to less than 20 %, preferably less than 10 %after treatment with the composition for 6 months.
- Surprisingly, the Von Willebrand Factor peptides seem to increase stability of Factor upon storage (shelf-life) and/or reduce inhibitor formation in patients. Inhibitor formation is one of the major problems in the treatment of chronic bleeding disorders.

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The composition of the present invention is especially useful in the treatment or prevention of a bleeding disorder.

Therefore, a further embodiment of the invention is a method of treating a bleeding disorder comprising administering to a patient in need thereof an effective amount of the composition of the present invention.

The amount depends on the disease or condition to be treated and may be selected by a person skilled in the art. For long term treatment, amounts of 20 to 40 IU/kg bodyweight per application are typically suitable. In an emergency situation, the amount may be about 10 to 50 IU/kg bodyweight.

10 The composition of the invention may be applied by intravenous administration or non-intravenous administration. The non-intravenous administration may be a subcutaneous injection, an intradermal injection or an intramuscular administration.

One advantage of the method of the present invention is the possibility to use nano filtration for virus removal. Von Willebrand Factor, because of its size, may not be nanofiltrated with a nanofilter with a small pore size to remove viruses. Because the Von Willebrand Factor peptides are much smaller in size than the full length Von Willebrand Factor molecule, nanofiltration with small pore sizes becomes possible. Nanofiltration is done at a pore size and conditions that reduces the concentration of one of the smallest known viruses porcine parvovirus by a least a factor of 100 (2 log), preferably by at least a factor 1000 (3 log) and most preferably to a concentration below detection limit of the parvovirus assay, optionally using one or more nanofilters in series. For this test, porcine parvovirus is spiked in a sample and analysed after filtration.

Therefore, a further embodiment of the invention is a method for virus reduction comprising the step of nanofiltrating the Von Willebrand Factor peptides prior or after a combination with Factor VIII, whereby porcine parvovirus would be reduced by at least 2 log.

30 A preferred puffer for administration of the composition of the invention comprises melizitose, preferably in an amount of up to 1,000 mM particularly from about 10 mM to about 200 mM, in particular from about 10 mM to about 100 mM.

A further embodiment of the invention is a method of preparing Von 35 Willebrand Factor peptides comprising the following steps:

- Incubating Von Willebrand Factor with S. aureus V-8 protease for 2 to 16 hours at an enzyme to Von Willebrand Factor weight/weight ratio of 1:5 to 1:100
- Binding and purifying on an anion exchanger and collecting the desired purified vWF peptides in a fraction coming from the anion exchanger by applying an increased amount of salt concentration.

Brief description of drawings

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Figure 1 shows purification of the fragment III (SPIII) from pdVWF digested by S.aureus V8 protease. A- MonoQ chromatogram of elution profile of the

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fragment III (indicated by an arrow). B- SDS-PAGE gel of the purified fragment; red.- reduced; n.r.- non-reduced.

Figure 2 shows purification of the fragment I (III-T4) from fragment III digested by trypsin. MonoQ chromatogram of elution profile of the fragment I (indicated by an arrow). The non-reducing SDS-PAGE picture of the purified fragment is shown in the insert.

Figure 3 shows purification of the fragment II from fragment III after second S.aureus V8 protease digestion. A- MonoQ chromatogram of elution profile of the fragment II (indicated by an arrow), the second cleavage product as well as the V8 protease are also indicated. B- Chromatogram of the second MonoQ chromatography required for complete removal of the protease. The reducing SDS-PAGE picture of the purified fragment is shown in the insert.

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Figure 4 shows binding of pdVWF, fragment II and III to rFVIII. A, B, C-Binding sensorgrams (grey curves), and curve alignment (black curves) representative for the interaction between immobilized rFVIII and pdVWF/purified fragments II and III. The concentrations and sample type are indicated on the diagram. C- Dissociation constants (K_D) expressed as mean and SEM; n=8.

Figure 5 shows binding of rFVIII to phospholipid monolayer in SPR and inhibition by pdVWF. A-binding sensorgrams of rFVIII and rFVIII in the presence of either 108 nM BSA (bovine serum albumin) or 47.6 nM pdVWF; each sample in triplicate. B- Mean and SD of binding levels measured 120 sec after end of analyte injection expressed as percentage of rFVIII binding; n=3.

Figure 6 shows inhibition of the rFVIII-phospholipid interaction by Von Willebrand Factor-derived fragments measured in SPR. rFVIII binding to phospholipid monolayer was performed in the presence of three different concentrations of the three Von Willebrand Factor-derived fragments (concentrations and fragment type are indicated on graph). Graph represents mean and SD of binding levels measured 120 sec after end of analyte injection expressed as percentage of rFVIII binding; n=3.

Figure 7 shows concentration dependent inhibition of rFVIII binding to phospholipid monolayer by fragment III. A-binding sensorgrams of rFVIII to phospholipid monolayer in the presence of different concentrations of the fragment III (concentrations are indicated on graph), each sample in triplicate. B- Mean and SD of binding levels measured 120 sec after end of analyte injection expressed as percentage of rFVIII binding; n=3.

Figure 8 shows binding of pdVWF and fragment III to collagen type III. A, B-Binding sensorgrams (grey curves), and curve alignment (black curves) representative for the interaction between immobilized collagen type III and pdVWF/purified fragment III. The concentrations and sample type are indicated on the diagram. C- Dissociation constants (K_D) expressed as mean and SEM; n=9.

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Figure 9 shows binding of pdVWF and fragment III to heparin. A, B- Binding sensorgrams (grey curves), and curve alignment (black curves) representative for the interaction between immobilized heparin and pdVWF/purified fragment III. The concentrations and sample type are indicated on the diagram. C- Dissociation constants (K_D) expressed as mean and SEM; n=6.

Figure 10 shows a comparison of whole blood clotting time (WBCT) values measured in blood samples from haemophilia A dogs treated s.c. with FVIII alone or in combination with VWF fragment III. WBCT obtained after s.c. application of FVIII in combination with five-fold molar excess of VWF fragment III applied at 200 IU FVIII / kg BW. Horizontal dashed line marks upper limit of clotting time in normal dogs (12 minutes).

Figure 11 shows FVIII activity measured with chromogenic FVIII activity assay in haemophilia A dogs plasma samples obtained after application of FVIII or FVIII in combination with VWF fragment III. A- FVIII or FVIII with five fold molar excess of VWF fragment III was applied subcutaneously at 200 IU FVIII /kg BW; the area under the curve (AUC) for the FVIII sample alone was 2.867, and for FVIII in combination with VWF fragment III- 4.917. B- FVIII or FVIII with five fold molar excess of VWF fragment III was applied intravenously at 200 IU FVIII /kg BW. The AUC for the FVIII sample alone was 27.69, and for FVIII in combination with VWF fragment III- 45.72.

Figure 12 shows binding of recombinant fragment III monomer, recombinant fragment III dimer and plasma derived VWF (flVWF) to rFVIII. A, B, C- Binding sensorgrams (grey curves), and curve alignment (black curves) representative for the interaction between immobilized VWF or recombinant VWF-fragments and FVIII. The sample type is indicated on the diagram. The concentration of applied FVIII was 0, 0.2, 0.6, 1.7, 5, 15, 45 and 135 nM. D- Dissociation constants (K_D) expressed as mean and SD; n=4.

Figure 13 shows stabilisation of FVIII by VWF fragment III. FVIII activity of FVIII alone or FVIII in complex with VWF fragment III incubated at 40°C measured at different time points.

Figure 14 shows Heparin binding using heparin affinity chromatography of two VWF fragments as described in Example 9.

Examples

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40 The invention is further explained by the following, non-limiting examples.

Example 1

Production and purification of fragments derived from plasmatic Von Willebrand Factor.

<u>Production and purification of fragment III (SPIII, res. 764-2128)</u> (According to Marti *et al.* Identification of disulfide-bridged substructures within human von Willebrand factor. Biochemistry 1987; 26:8099-8109 with modifications) (SEQ. ID. No. 2):

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SLSCRPPMVKLVCPADNLRAEGLECTKTCQNYDLECMSMGCVSGCLCPPGMVRHENRCVA LERCPCFHOGKEYAPGETVKIGCNTCVCQDRKWNCTDHVCDATCSTIGMAHYLTFDGLKY LFPGECQYVLVQDYCGSNPGTFRILVGNKGCSHPSVKCKKRVTILVEGGEIELFDGEVNV KRPMKDETHFEVVESGRYIILLLGKALSVVWDRHLSISVVLKQTYQEKVCGLCGNFDGIQ 5 NNDLTSSNLQVEEDPVDFGNSWKVSSQCADTRKVPLDSSPATCHNNIMKQTMVDSSCRIL TSDVFQDCNKLVDPEPYLDVCIYDTCSCESIGDCACFCDTIAAYAHVCAQHGKVVTWRTA TLCPQSCEERNLRENGYECEWRYNSCAPACQVTCQHPEPLACPVQCVEGCHAHCPPGKIL DELLQTCVDPEDCPVCEVAGRRFASGKKVTLNPSDPEHCQICHCDVVNLTCEACQEPGGL VVPPTDAPVSPTTLYVEDISEPPLHDFYCSRLLDLVFLLDGSSRLSEAEFEVLKAFVVDM 10 MERLRISQKWVRVAVVEYHDGSHAYIGLKDRKRPSELRRIASQVKYAGSQVASTSEVLKY TLFQIFSKIDRPEASRITLLLMASQEPQRMSRNFVRYVQGLKKKKVIVIPVGIGPHANLK QIRLIEKQAPENKAFVLSSVDELEQQRDEIVSYLCDLAPEAPPPTLPPDMAQVTVGPGLL GVSTLGPKRNSMVLDVAFVLEGSDKIGEADFNRSKEFMEEVIQRMDVGQDSIHVTVLQYS YMVTVEYPFSEAOSKGDILORVREIRYOGGNRTNTGLALRYLSDHSFLVSOGDREOAPNL 15 VYMVTGNPASDEIKRLPGDIOVVPIGVGPNANVOELERIGWPNAPILIODFETLPREAPD LVLQRCCSGEGLQIPTLSPAPDCSQPLDVILLLDGSSSFPASYFDEMKSFAKAFISKANI GPRLTQVSVLQYGSITTIDVPWNVVPEKAHLLSLVDVMQREGGPSQIGDALGFAVRYLTS EMHGARPGASKAVVILVTDVSVDSVDAAADAARSNRVTVFPIGIGDRYDAAQLRILAGPA GDSNVVKLQRIEDLPTMVTLGNSFLHKLCSGFVRICMDEDGNEKRPGDVWTLPDQCHTVT 20 CQPDGQTLLKSHRVNCDRGLRPSCPNSQSPVKVEETCGCRWTCPCVCTGSSTRHIVTFDG QNFKLTGSCSYVLFQNKEQDLEVILHNGACSPGARQGCMKSIEVKHSALSVELHSDMEVT VNGRLVSVPYVGGNMEVNVYGAIMHEVRFNHLGHIFTFTPQNNEFQLQLSPKTFASKTYG LCGICDENGANDFMLRDGTVTTDWKTLVQEWTVQRPGQTCQPILE

25 Fragment III is prepared by digestion of plasma derived Von Willebrand Factor (pdVWF) with *S. aureus* V-8 protease. The digestion is carried out for 3 hours at 37 °C in a 50 mM Tris-HCl, 150 mM NaCl pH 7.8 buffer at a 1:40 enzyme to protein weight ratio.

The purification of the fragment is carried out using a strong anion exchange column (MonoQ). The running buffer is a 20 mM Tris-HCl pH 7.4, and the elution buffer (buffer B) is 20 mM Tris-HCl, 500 mM NaCl pH 7.4. The *S. aureus* V-8 protease elutes from the anion exchange column at ca. 22 mS/cm (ca. 40 % buffer B), therefore long washing step at 42 % prior to elution of the fragment is required to wash out the protease. Alternatively an SEC step on Superose 6 10/300 GL can be conducted for protease removal.

The fragment III purification and the product obtained are depicted on Fig. 1. The sequence defined by Marti *et al.* 1987 has been confirmed by MS analysis.

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<u>Production and purification of fragment I (III-T4, res. 764-1035)</u> (According to Marti *et al.* 1987 with modifications) (SEQ. ID. No. 3):

SLSCRPPMVKLVCPADNLRAEGLECTKTCQNYDLECMSMGCVSGCLCPPGMVRHENRCVA LERCPCFHQGKEYAPGETVKIGCNTCVCQDRKWNCTDHVCDATCSTIGMAHYLTFDGLKY LFPGECQYVLVQDYCGSNPGTFRILVGNKGCSHPSVKCKKRVTILVEGGEIELFDGEVNV KRPMKDETHFEVVESGRYIILLLGKALSVVWDRHLSISVVLKQTYQEKVCGLCGNFDGIQ NNDLTSSNLQVEEDPVDFGNSWKVSSQCADTR

Fragment I is prepared from fragment III (SPIII) by trypsin (TPCK treated from bovine) digestion. The digestion is carried out for 1.5 hours in a 100 mM NH₄HCO₃ pH 8.0 buffer at a 1:100 enzyme to protein weight ratio. The digestion was terminated by the addition of soybean trypsin inhibitor. The purification of the fragment I is carried out using a strong anion exchange column (MonoQ) followed by SEC on Superose 6, 10/300 GL. The running buffer for the anion exchange column is 20 mM Tris-HCl pH 7.4, and the

elution buffer (buffer B) is 20 mM Tris-HCl, 500 mM NaCl pH 7.4. The running buffer for the SEC is PBS (phosphate buffered saline) pH 7.0. The fragment I purification and the product obtained is depicted on Fig. 2. The

The fragment I purification and the product obtained is depicted on Fig. 2. The sequence defined by Marti *et al.* 1987 has been confirmed by MS analysis.

Production and purification of fragment II (res. 764-1673) (SEQ. ID. No. 4):
SLSCRPPMVKLVCPADNLRAEGLECTKTCQNYDLECMSMGCVSGCLCPPGMVRHENRCVA
LERCPCFHQGKEYAPGETVKIGCNTCVCQDRKWNCTDHVCDATCSTIGMAHYLTFDGLKY
LFPGECQYVLVQDYCGSNPGTFRILVGNKGCSHPSVKCKKRVTILVEGGEIELFDGEVNV

KRPMKDETHFEVVESGRYIILLLGKALSVVWDRHLSISVVLKQTYQEKVCGLCGNFDGIQ
NNDLTSSNLQVEEDPVDFGNSWKVSSQCADTRKVPLDSSPATCHNNIMKQTMVDSSCRIL
TSDVFQDCNKLVDPEPYLDVCIYDTCSCESIGDCACFCDTIAAYAHVCAQHGKVVTWRTA
TLCPQSCEERNLRENGYECEWRYNSCAPACQVTCQHPEPLACPVQCVEGCHAHCPPGKIL
DELLQTCVDPEDCPVCEVAGRRFASGKKVTLNPSDPEHCQICHCDVVNLTCEACQEPGGL
VVPPTDAPVSPTTLYVEDISEPPLHDFYCSRLLDLVFLLDGSSRLSEAEFEVLKAFVVDM

MERLRISQKWVRVAVVEYHDGSHAYIGLKDRKRPSELRRIASQVKYAGSQVASTSEVLKY
TLFQIFSKIDRPEASRITLLLMASQEPQRMSRNFVRYVQGLKKKKVIVIPVGIGPHANLK
QIRLIEKQAPENKAFVLSSVDELEQQRDEIVSYLCDLAPEAPPPTLPPDMAQVTVGPGLL
GVSTLGPKRNSMVLDVAFVLEGSDKIGEADFNRSKEFMEEVIQRMDVGQDSIHVTVLQYS
YMVTVEYPFSEAQSKGDILQRVREIRYQGGNRTNTGLALRYLSDHSFLVSQGDREQAPNL

VYMVTGNPASDEIKRLPGDIOVVPIGVGPNANVOELERIGWPNAPILIODFETLPREAPD

LVLQRCCSGE

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Fragment II is prepared from fragment III by second S. aureus V8 protease digestion. The digestion is carried out for 21 hours in a 50 mM Tris-HCl, 150 mM NaCl pH 7.8 buffer in a 1:10 enzyme to protein weight ratio.

The purification of the fragment II is carried out using a strong anion exchange column (MonoQ). The running buffer is a 20 mM Tris-HCl pH 7.4, and the elution buffer (buffer B) is 20 mM Tris-HCl, 500 mM NaCl pH 7.4. A second MonoQ purification with a long washing step at 42 % B was required to remove the protease.

The fragment II purification and the product obtained are depicted on Fig. 3. 10 The second V8 cleavage site between Glu¹⁶⁷³-Gly¹⁶⁷⁴ was determined by Fretto et al. 1986 and confirmed by MS analysis.

Example 2

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Determination of Factor VIII binding affinity.

15 The analysis was carried out using Biacore 2000 instrument (GE Healthcare) according to McCormick et al. 2004 with modifications. Briefly rFVIII was covalently coupled to CM5 Sensor Chip resulting in a ~200 RU coating level. Subsequently the Von Willebrand Factor-fragments as well as full length Von Willebrand Factor (flvWF) were injected over the sensor chip surface. The 20 running buffer was 20 mM HEPES, 150 mM NaCl, 5 mM CaCl₂, 0.02 % Tween 20. The dissociation affinity constants were determined for flVWF as well as for fragments II and III, there was no significant binding of fragment I to Factor VIII therefore the K_D was not determined. Binding sensorgrams and the calculated K_D values are depicted in Fig. 4. The flVWF bound to rFVIII with K_D 25 of 0.67 nM, fragment III bound with lower affinity (K_D of 6.18 nM), the affinity was further decreased for fragment II (KD of 154.60 nM)

Example 3

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Determination of Factor VIII binding to phospolipid-monolayer and inhibition by Von Willebrand Factor and Von Willebrand Factor-derived fragments.

The analysis was carried out using Biacore 2000 instrument (GE Healthcare) according to Saenko et al. 1999 with modifications. Briefly, phospholipid-(1,2-Dioleoyl-sn-glycerol-3were vesicles prepared from DOPC phosphocholine) and DOPS (1,2-Dioleoyl-sn-glycerol-3-phospho-L-serine). Unilamellar vesicles were prepared according to MacDonal et al. 1991 using an extruder and coated on a HPA sensor chip. Subsequently the componds of interest were injected over the PCPS surface and the binding level 120 s after injection end was evaluated.

- 40 Negative controls; Von Willebrand Factor and BSA did not bind to the PSPC surface (not shown), in contrast, a high binding level of rFVIII was shown. This binding could be completely inhibited with Von Willebrand Factor, in contrast addition of high BSA concentration had no effect on the binding (Fig.
- 45 To evaluate, if the fragments obtained by limited digestion were able to inhibit PSPC binding similar to flVWF, the fragments I, II and III were injected over the sensor chip surface. Only fragment III was able to inhibit the interaction between rFVIII and phospholipid monolayer (Fig. 6). This effect was dose dependent with almost complete inhibition at 2.5 x excess of fragment III over 50 the rFVIII (Fig. 7).

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Example 4

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Determination of collagen III binding affinity of flVWF and fragment III.

The analysis was carried out using Biacore 2000 instrument (GE Healthcare) according to Romjin et al. 2003 with modifications. Briefly human pepsin-digested collagen type III was covalently bound to the surface of a CM5 sensor chip. Subsequently the samples were injected over the sensor chip surface. The running buffer was 10 mM HEPES, 150 mM NaCl, 3.4 mM EDTA, 0.005 % Tween 20. The flVWF bound to collagen III with very high affinity (0.75 nM), the binding of the fragment III was significantly decreased to 17.02 nM (Fig. 8).

Example 5

15 Determination of heparin binding affinity of flVWF and fragment III.

The analysis was carried out using Biacore T200 instrument (GE Healthcare) according to Sarafanov *et al.* 2001. Briefly, heparin from porcine intestinal mucosa was biotinylated using NHS-biotin reagent kit, and bound to the surface of a SA sensor chip. The reference flow cell was coated with biotin. Subsequently the samples were injected over the sensor chip surface. The running buffer was 150 mM HEPES, 150 mM NaCl, 5 mM CaCl₂, 0.05 % Tween 20. The flVWF bound to heparin with an affinity of 0.65 nM, the binding affinity of the fragment III was significantly decreased to 2.43 nM (Fig. 9).

25 **Example 6**

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Determination of FVIII or FVIII/VWF Fragment III complex recovery and half life in circulation in haemophilia A dogs.

- Two haemophilia A dogs were subjected to s.c. and subsequent i.v. injection of recombinant B-domain-deleted FVIII alone or in combination with five-fold molar excess of VWF Fragment III. Dog 1 received 200 IU/kg BW of FVIII alone and Dog 2 received 200 IU/kg BW FVIII in complex with VWF Fragment III. Blood samples were collected at 0.5, 1, 2, 4, 8, 12, 24, 32, 48, 72 and 96 hours after each s.c. or i.v. drug administration. Samples were analyzed for whole blood clotting time (WBCT) and activity in chromogenic FVIII activity assay. The subcutaneous administration of VWF Fragment III in complex with FVIII resulted in 1.4-fold increase in time required to exceed a clotting time for a normal dog comparing with s.c. administration of FVIII alone (Fig. 10). The administration of VWF Fragment III with FVIII resulted also in increased FVIII activity in dog plasma over time and in nearly doubled area under the
- FVIII activity in dog plasma over time and in nearly doubled area under the curve (AUC) values for both, s.c. and i.v. application compared to administration of FVIII alone (Fig. 11).

45 **Example 7**

Determination of FVIII binding affinity of recombinant fragment III monomer and dimer.

Recombinant fragment III was transiently expressed in HEK293 cell line with a C-terminal Strep-Tag and purified by Strep-tactin affinity chromatography.

The fragment III monomers and dimers were separated by size exclusion chromatography (SEC). The analysis was carried out using Biacore 2000 instrument. The fragment III monomers and dimers were immobilized on CM5 and FVIII concentration series was injected over the sensor chip surface. Plasma derived full length VWF was used as control. The running buffer was 150 mM HEPES, 150 mM NaCl, 5 mM CaCl₂, 0.05 % Tween 20. FVIII bound to fragment III dimer with an affinity constant of 1.9 nM. The affinity of FVIII to the monomeric Fragment III was significantly lower ($K_D = 14.3 \text{ nM}$) (Fig. 12).

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10 Example 8

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Stabilisation of rFVIII in solution by VWF Fragment III.

2000 IU of recombinant FVIII (Nuwig®) was reconstituted in 2.5 ml water, with or without addition of five-fold molar excess VWF Fragment III. Both preparations were incubated at 40°C and aliquots were taken at 48, 96, 192, 384, 408 and 672 hours. Samples were analysed for FVIII activity in a chromogenic FVIII activity assay. VWF Fragment III contributed to significant longer activity of FVIII at 40°C (Fig. 13).

20 Example 9

Comparison of heparin binding between recombinant fragment III and NovoSeg21 fragment.

Recombinant fragment III and NovoSeg21 (SEQ ID No 21 from 25 WO2013/160005A1) fragment were transiently expressed in HEK293 cell line a C-terminal Strep-Tag and purified by Strep-tactin Heparin chromatography. binding was tested usina heparin chromatography. Both recombinant fragments were bound to heparin column (HiTrap Heparin HP 1ml, GE Healthcare) and eluted with linear salt gradient ranging from 0-500 mM NaCl. Both fragments were run in triplicates, see Fig. 30 14. The mean elution peak for the NovoSeg21 fragment was at 15.57 \pm 0.04 min which corresponds to 285.381 mM NaCl, and for the fragment III at 15.47 ± 0.02 min which corresponds to 282.051 mM NaCl. This indicates higher heparin affinity for the NovoSeq21 fragment.

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Analytical methods

Description of analytical methods

FVIII: C, Screening method based on Coatest

The method is based on the two-stage principle, and was performed using micro plate technique. In stage one, activated factor X (Xa) is generated via 40 the intrinsic pathway where FVIII: C acts as a co-factor. In stage two, Factor Xa is then determined by the use of a synthetic chromogenic substrate, S-2222 in the presence of a thrombin inhibitor I-2581 to prevent hydrolysis of the substrate by thrombin. The reaction is stopped with acid, and the VIII: C 45 activity, which is proportional to the release of pNA (para-nitroaniline), is determined photo metrically at 405 nm against a reagent blank.

- 19 -

The method complies with the requirements in the European Pharmacopoeia. The unit of FVIII: C is expressed in international units (IU) as defined in the current International Concentrate Standard (IS) established by the World Health Organization (WHO). The routine using buffer containing 1 % BSA 5 instead of severe hemophilic plasma for predilutions has been validated. See also literature references (European Pharmacopoeia Supplement 2000, general Methods, 2.7.4. Assay of Blood Coagulation FVIII; Rosén S (1984) Assay of FVIII: C with a Chromogenic Substrate. J, Haematol, Suppl 40, vol 33, 139-145, 1984; Carlebjörk G, Oswaldsson U, Rosén S (1987) A simple and accurate micro plate assay for the determination of FVIII activity. Thrombosis Research 47; 5-14, 1987; Mire-Sluis AR, Gerrard T, Gaines das R, Padilla A and Thorpe R. Biological assays: Their Role in the development and quality Control of Recombinant Biological Medicinal Products. Biological, 24, 351-362 (1996)).

15 Determination of total protein according to Bradford

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Protein determination according to Bradford is based on the observation that the absorbance maximum for an acidic solution of Coomassie Brilliant Blue G-250 shifts from 465 nm to 595 nm when binding to protein occurs. Both hydrophobic and ionic interactions stabilize the anionic form of the dye, causing a visible colour change. The assay is useful since the extinction coefficient of a dye-albumin complex solution is constant over a 10-fold concentration range. See also reference Bradford, MM. A rapid and sensitive method for the quantisation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry 72: 248-254. 1976. 25 for further information.

Determination of total protein according to amino acid analysis (AAA)

Before the AAA all proteins are hydrolyzed by 6 M HCI for 24 h at 110 °C. The amino acids are separated by cation-exchange chromatography sulphonated polystyrene resins and detected continuously in the eluent. The detection is based on post-column ninhydrin derivatisation using a dual photometer for simultaneous measurement at 440 nm for proline and hydroxyproline and 570 nm for all other amino acids. The amino acids asparagine and glutamine are both deamidated during AAA and are determined as aspartic acid and glutamic acid, respectively. Thus, the results of aspartic acid and glutamic acid represent the sum of aspartic acid/asparagine (Asx) and glutamic acid/glutamine (Glx), respectively, in the original sample. Tryptophan is not generating a distinct response using this method, and, thus, is not quantified by the AAA. Cysteine is destroyed during the hydrolysis and is not quantified. The AAA is further described in reference: Total protein AAA analytical method. Spackman, D. H., Stein, W. H., and Moore, S. (1958) Anal. Biochem. 30: 1190-1206.

Purity or specific activity(FVIII:C/Total protein)

The purity (or also called specific activity) for a sample, is calculated taking the value achieved from the FVIII:C analysis and divide it with the value achieved from the analysis of total protein.

- 20 -

SDS-PAGE (Molecular weight distribution)

SDS polyacrylamide gel electrophoresis (SDS-PAGE) involves the separation of proteins based on their size. This method describes the SDS-PAGE of proteins, which is run under reduced conditions. By heating the sample under 5 denaturing and reducing conditions, proteins become unfolded and coated with anionic detergent sodium dodecyl sulphate (SDS), acquiring a high net negative charge that is proportional to the length of the polypeptide chain. When loaded onto a polyacrylamide gel matrix and placed in an electric field, the negatively charged protein molecules migrate towards the positively 10 charged electrode and are separated by a molecular sieving effect, i.e. by their molecular weight. Polyacrylamide gels restrain larger molecules from migrating as fast as smaller molecules. Because the charge-to-mass ratio is nearly the same among SDS-denatured polypeptides, the final separation of proteins is dependent almost entirely on the differences in relative molecular mass of polypeptides. In a gel of uniform density the relative migration 15 distance of a protein (R_f) is negatively proportional to the log of its mass. If proteins of known mass are run simultaneously with the unknowns, the relationship between Rf and mass can be plotted, and the masses of unknown proteins estimated. The protein bands separated by electrophoresis are 20 visualized by silver staining. Evaluation is done visually by judging the appearances of the standards, reference (control sample) and analysed samples.

Factor VIII antigen content (FVIII:Ag)

The amount of Factor VIII antigen content (FVIII:Ag) is measured with a ELISA kit (ASSERACHROM® VIII:Ag, enzyme immunoassay for Factor VIII, kit, Diagnostica Stago (France), as further described⁽¹⁸⁾ with replacement of the provided kit buffer with Tris-NaCl buffer + 1% bovine serum albumin for sample dilutions.

Size exclusion chromatography (SEC)

30 Monomer, aggregate and fragment is measured using a size exclusion chromatography (SEC-HPLC) analytical column (Superdex 200, 10/300 GL, GE Healthcare) processed under native buffer conditions (25mM HEPES, 0.5M NaCl, 0.3M arginine, 50mM CaCl₂, 0.02% Polysorbate 80, pH 7.5). Sample load is approximately 1% of the size exclusion column and the Factor VIII:C concentration is approximately 1000 IU/ml.

Western blot against Factor VIII

Factor VIII degeneration product based on size is measured using FVIII Western Blot. FVIII molecular mass distribution proteins and peptides in factor VIII preparations are separated according to molecular mass by sodium dodecyl sulphate (SDS) polyacrylamide gel electrophoresis (PAGE) under reducing conditions. Thereafter, the proteins are transferred electrophoretically from the gel matrix to a nitrocellulose membrane which is subsequently incubated with a blocking agent. Commercial available polyclonal sheep antibodies directed to the whole human factor VIII molecule is then

- 21 -

added followed by a secondary enzyme-labelled antibody as a probe. As a third step a chemiluminescent substrate is added and when combined with the enzyme, light is produced as a by-product. The light output is captured as a real time image using a cooled Charge-Coupled Device camera. The intensity of the signal is correlated with the abundance of the antigen (FVIII) on the blotting membrane.

2D-PAGE

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2D-Electrophoresis with Silver Staining was carried out in order to study the electrophoretic band pattern of the Factor VIII protein chain. Isoelectric focusing was performed as the first dimension run using a linear pH gradient of pH 3 to 10. The second dimension SDS-PAGE was run using Tris-Acetate (3-8%) gels. The gels were stained with silver-stain following the second dimension run.

15 Total protein (Bradford)

Protein determination according to Bradford is based on the observation that the absorbance maximum for an acidic solution of Coomassie Brilliant Blue G-250 shifts from 465 nm to 595 nm when binding to protein occurs. Both hydrophobic and ionic interactions stabilize the anionic form of the dye, causing a visible colour change. The assay is useful since the extinction coefficient of a dye-albumin complex solution is constant over a 10-fold concentration range. See also reference Bradford, MM. A rapid and sensitive method for the quantisation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry 72:* 248-254. 1976.

All references cited herein are incorporated by reference to the full extent to which the incorporation is not inconsistent with the express teachings herein.

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Claims

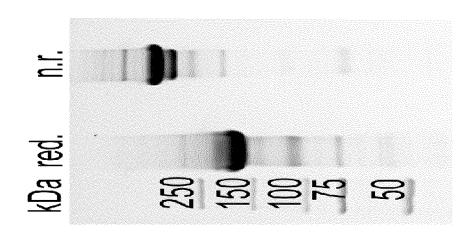
- A composition comprising a complex of Factor VIII and one or more Von 1. Willebrand Factor peptides, wherein the Von Willebrand Factor peptides comprise at least the amino acids 764 to 1035 and 1691 to 1905 of SEQ ID No. 1 but not amino acids 2255 to 2645 of SEQ ID No. 1.
- The composition of claim 1, wherein the Von Willebrand Factor peptides are 2. fragments of Von Willebrand factor.
- 3. The composition of claim 2, wherein the fragments of Von Willebrand Factor comprise amino acids 764 to 1905 of SEQ ID No. 1.
 - 4. The composition of claim 2, wherein the fragments of Von Willebrand factor have an amino acid sequence that corresponds to the amino acid sequence of SEQ ID No. 1 starting form amino acid 764 and ending between amino acid 1905 and 2153 with up to 10 deletions, insertions or substitutions.
- 15 5. The composition of any one of claims 1 to 4 wherein the Von Willebrand Factor peptide has a molecular weight of < 500 kD.
 - The composition of any one of claims 1 to 5, wherein the Von Willebrand 6. Factor peptides has a molecular weight of <400 KD.
- 7. The composition of any one of claims 1 to 6, wherein the Von Willebrand 20 Factor peptides have at least one property selected from the group consisting of:
 - (i) an affinity binding constant for heparin of $K_D > 1$ nM, preferably \geq 2.43 nM;
- (ii) an affinity binding constant for collagen III of $K_D > 5$ nM, preferably \geq 25 17.02 nM:
 - (iii) an affinity binding constant for Factor VIII of K_D < 100 nM, or < 10 nM, preferably ≤ 6.19 nM; and
 - (iv) an inhibition of Factor VIII phospholipid binding of at least 70%, preferably at least 80%.
- 30 8. The composition of any one of claims 1 to 7, wherein the Von Willebrand Factor peptides are derived from Willebrand Factor by proteolytic cleavage or chemical cleavage.
 - 9. The composition of claim 8, wherein the proteolytic cleavage is proteolytic cleavage with S. aureus V-8 protease.
- 35 The composition of any one of claims 1 to 9, wherein Factor VIII is a full length Factor VIII, a B-domain deleted Factor VIII or a Factor VIII- where the B-domain has been replaced by an artificial linker or a fragment of the natural B-domain or a combination thereof.
- 11. The composition of any one of claims 1 to 10, wherein Factor VIII is plasma derived Factor VIII or recombinant Factor VIII. 40
 - 12. The composition of claim 11, wherein the recombinant Factor VIII is expressed in a human cell line.

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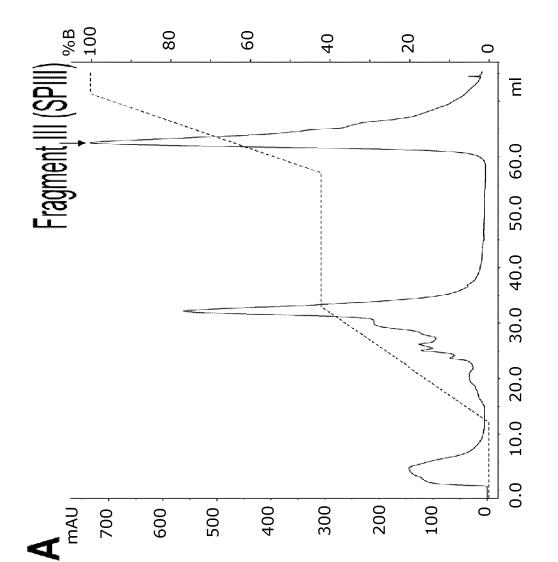
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- 13. The composition of any one of claims 1 to 12, wherein the composition has at least one of the properties selected from the group consisting of:
 - (i) the Von Willebrand Factor peptides shield Factor VIII from antibody binding to minimize inhibitor formation in a patient;
 - (ii) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 12 months in a frozen liquid form at - 70 °C;
 - (iii) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 24 months in a freeze-dried form at 5 °C;
 - (iv) stabilises Factor VIII to provide a remaining Factor VIII activity of at least 90% after storage for 12 months in a freeze-dried form at 25 °C;
 - (v) prolongs half-life of Factor VIII in-vivo by at least 20%; and
 - (vi) reduces inhibitor formation in previously untreated patients to less than 20% after treatment with the composition for 6 months.
- 14. Use of the composition of any one of claims 1 to 13 in the manufacture of a 15 medicament for the treatment or prevention of a bleeding disorder.
 - 15. The use of claim 14, wherein the medicament is adapted for nonintravenous administration.
 - The use of claim 15, wherein the non-intravenous administration is a subcutaneous injection.
- 20 17. A method of treating or preventing a bleeding disorder comprising the administration of the composition of any one of claims 1 to 13.
 - 18. The method of claim 17, wherein the administration is a non-intravenous administration.
- The method of claim 18, wherein the non-intravenous administration is a 25 subcutaneous injection.
 - 20. A method for virus reduction during preparation of the composition of any one of claims 1 to 13 comprising the step of nanofiltrating the Von Willebrand Factor peptides prior to or after combination with Factor VIII, whereby porcine parvovirus, if present, is reduced by at least a factor of 100.



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<u>-ig. 1</u>

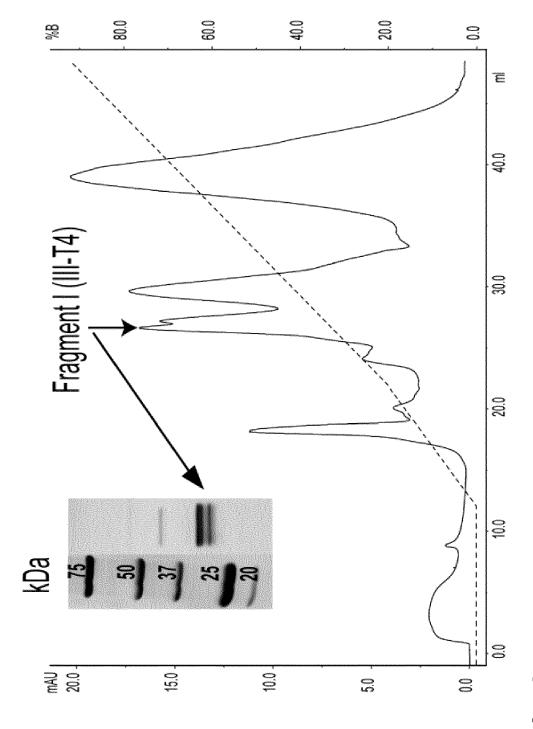


Fig. 2

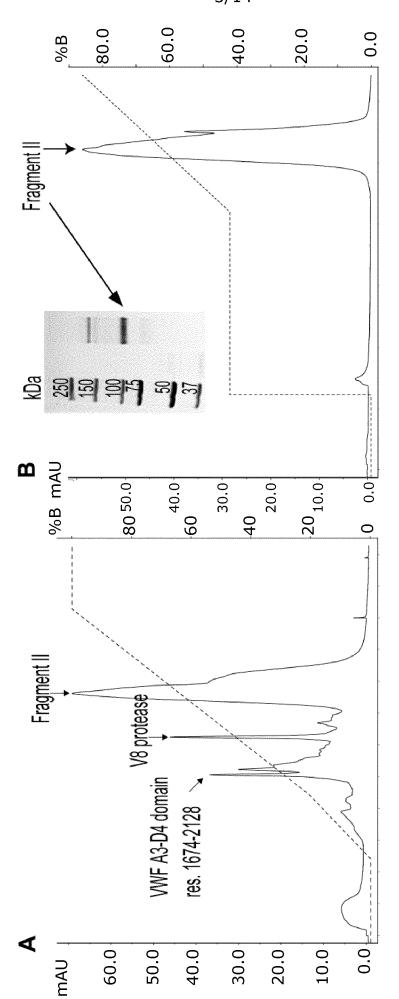
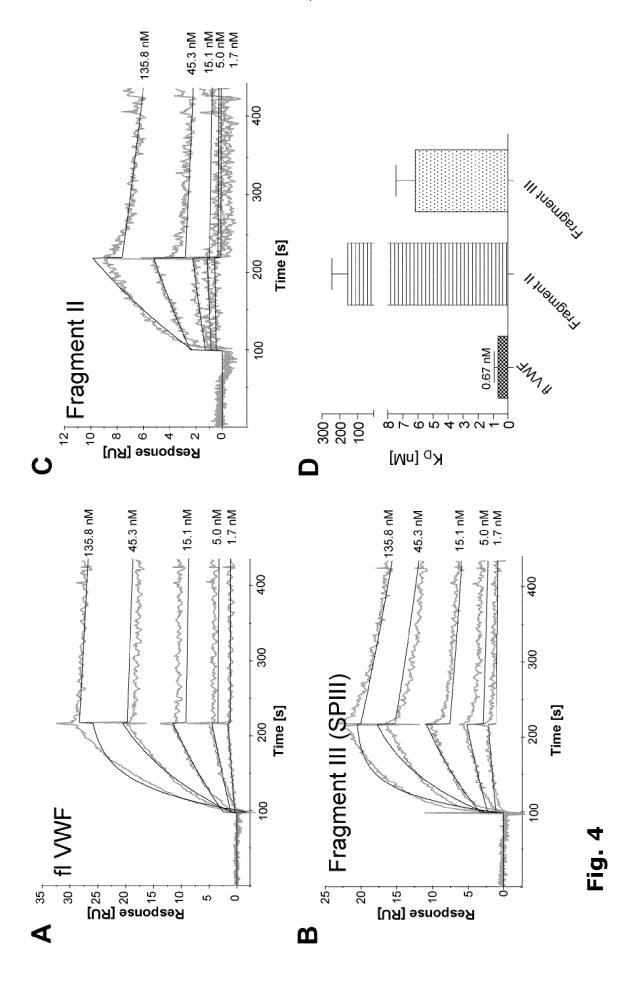
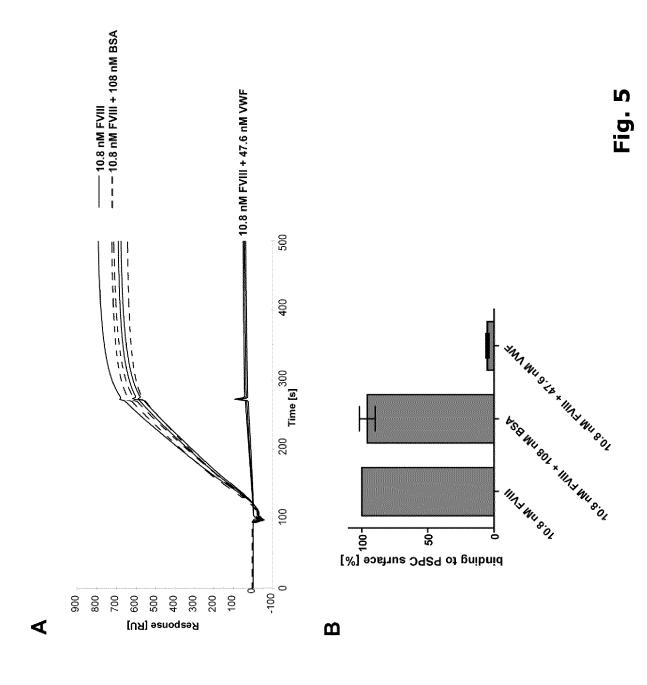


Fig. 3





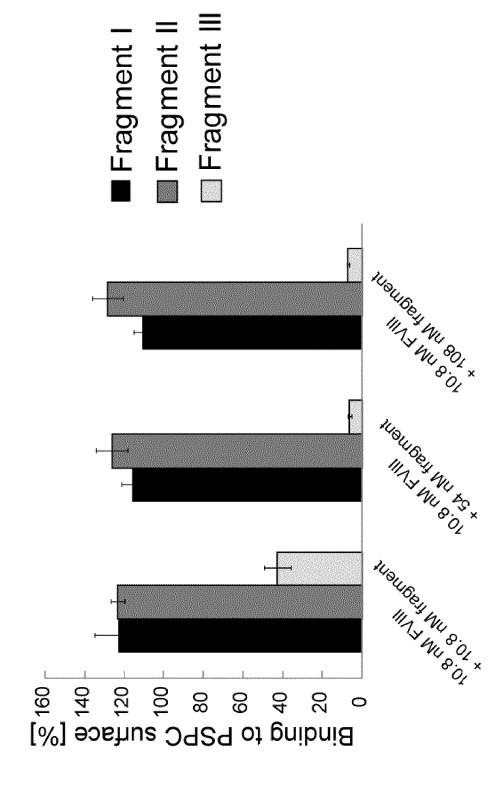


Fig. (

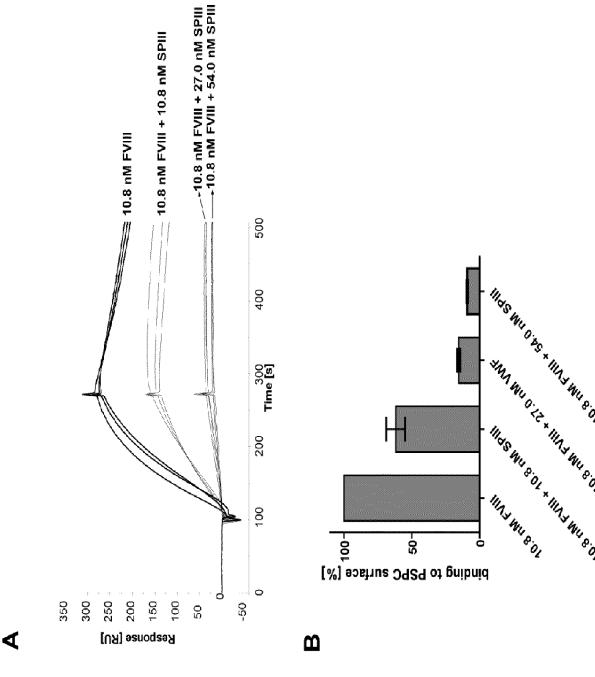
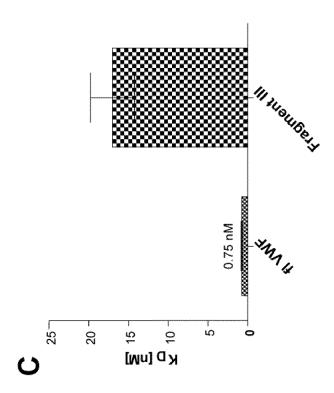
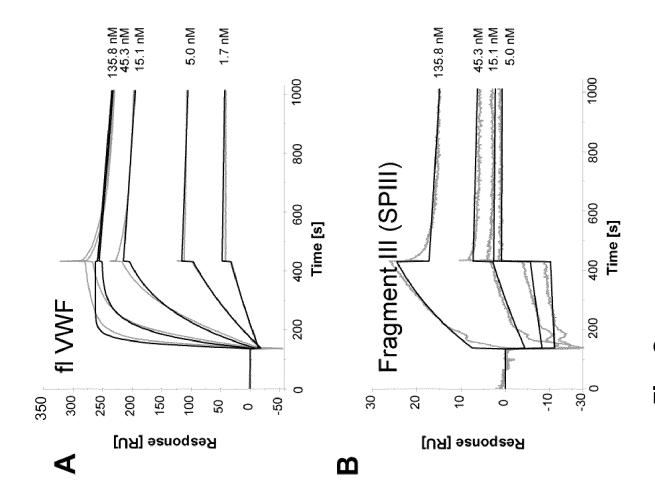


Fig. 7

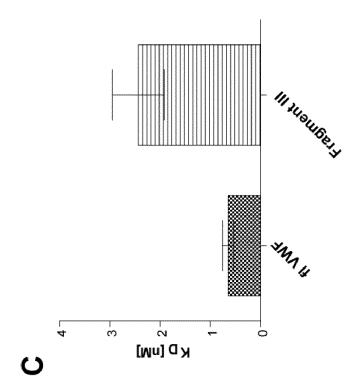
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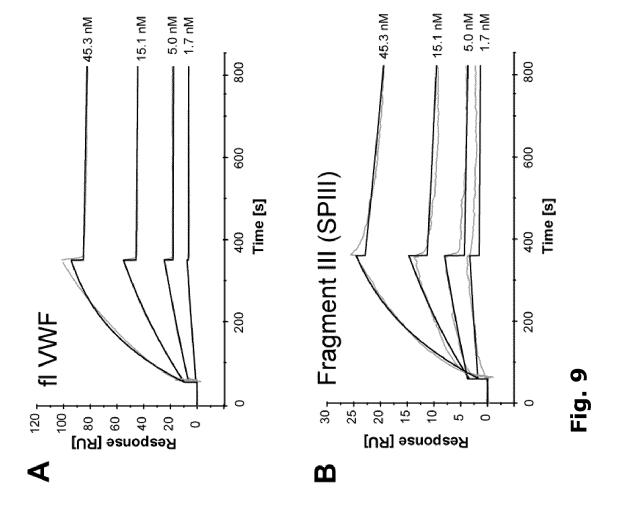




<u>Fig. 8</u>

WO 2015/185758 PCT/EP2015/062730 -9/14-





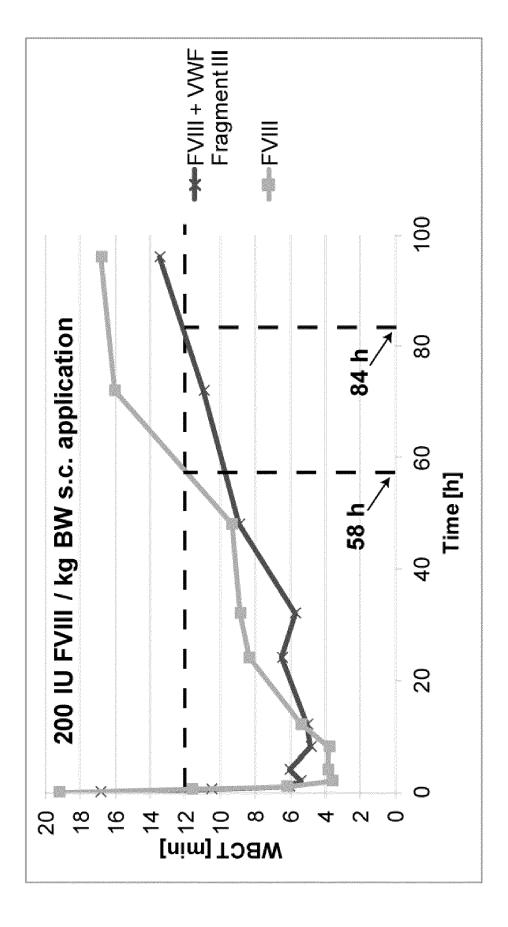
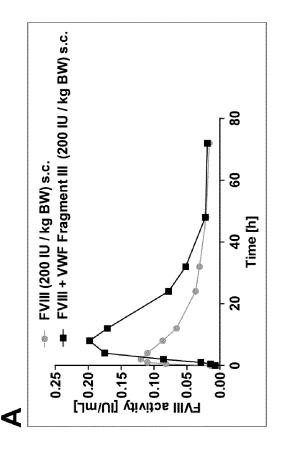
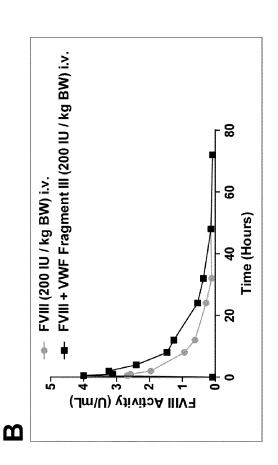


FIG. 1(

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Fig. 11





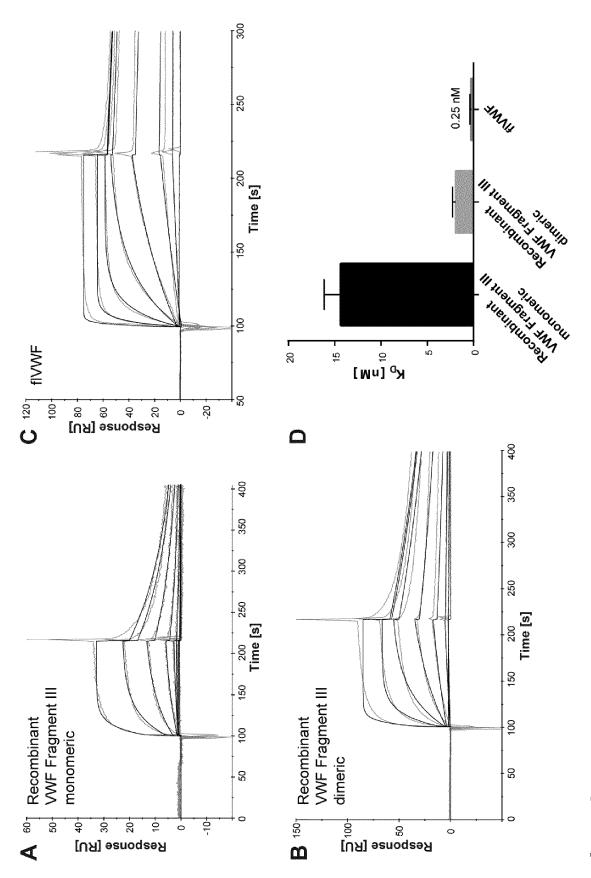


Fig. 12

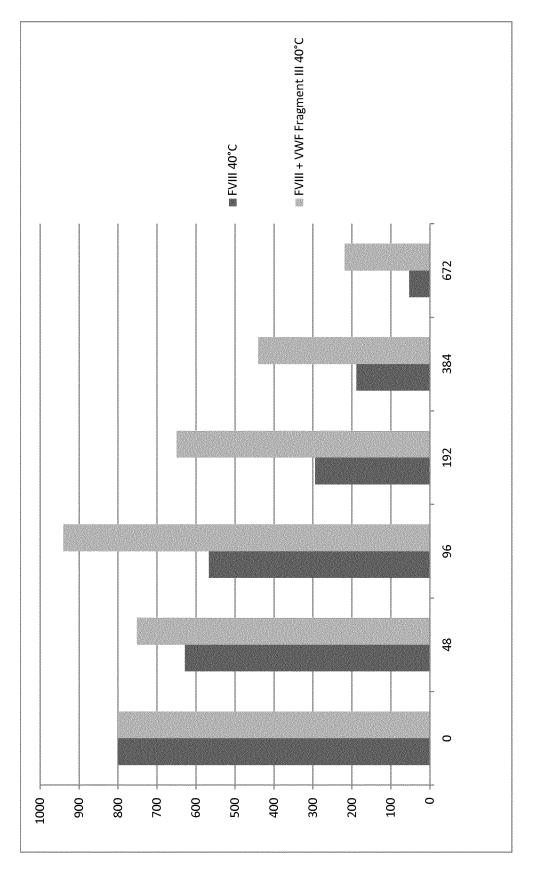
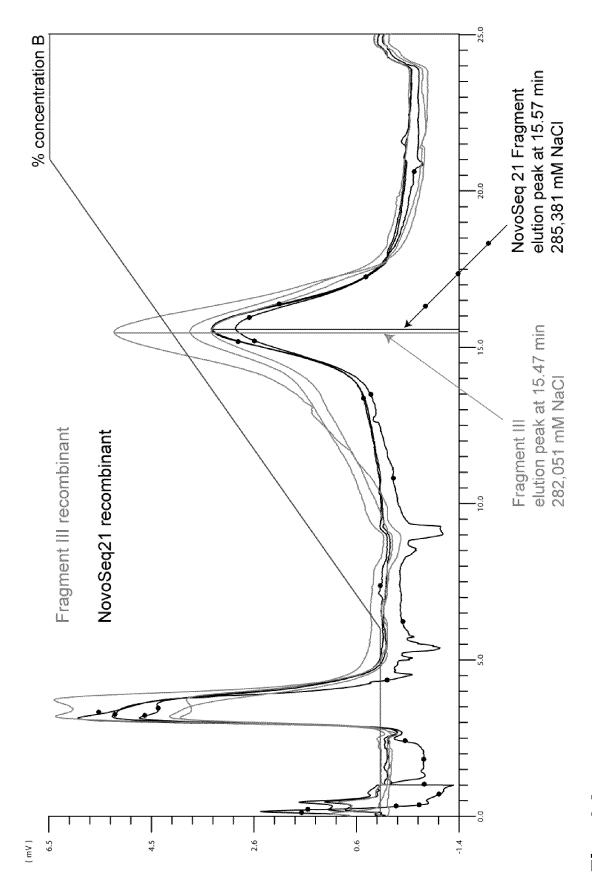


Fig. 13



-ig. 14

eolf-seql SEQUENCE LISTING

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Homo sapiens

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Pro Cys Phe His Gln Gly Lys Glu Tyr Ala Pro Gly Glu Thr Val Lys 65 70 75 80

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Leu Val Gly Asn Lys Gly Cys Ser His Pro Ser Val Lys Cys Lys 145 150 160

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Glu Val Asn Val Lys Arg Pro Met Lys Asp Glu Thr His Phe Glu Val 180 185 190

Val Glu Ser Gly Arg Tyr IIe IIe Leu Leu Gly Lys Ala Leu Ser 195 200 205

Val Val Trp Asp Arg His Leu Ser IIe Ser Val Val Leu Lys Gln Thr Page 12

eol f-seql 220 210 215

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Lys Val Pro Leu Asp Ser Ser Pro Ala Thr Cys His Asn Asn Ile Met

Lys Gln Thr Met Val Asp Ser Ser Cys Arg IIe Leu Thr Ser Asp Val 290 295 300

Phe GIn Asp Cys Asn Lys Leu Val Asp Pro GIu Pro Tyr Leu Asp Val 305 310 315

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Phe Cys Asp Thr IIe Ala Ala Tyr Ala His Val Cys Ala Gln His Gly

Lys Val Val Thr Trp Arg Thr Ala Thr Leu Cys Pro Gln Ser Cys Glu

Glu Arg Asn Leu Arg Glu Asn Gly Tyr Glu Cys Glu Trp Arg Tyr Asn 370 380

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Val Val Pro Pro Thr Asp Ala Pro Val Ser Pro Thr Thr Leu Tyr Val Page 13

eol f-seql 490

485

495

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Page 14

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Gly Ala Arg Pro Gly Ala Ser Lys Ala Val Val IIe Leu Val Thr Page 15

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Asn	Val 1085	Val	Lys	Leu	GI n	Arg 1090	Пе	GI u	Asp	Leu	Pro 1095	Thr	Met	Val
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Arg	Leu 1265	Val	Ser	Val	Pro	Tyr 1270	Val	GI y	GI y	Asn	Met 1275	GI u	Val	Asn
Val	Tyr	GI y	Al a	Пe	Met	Hi s	GI u		Arg age		Asn	Hi s	Leu	GI y

eol f-seql

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Pro Cys Phe His Gln Gly Lys Glu Tyr Ala Pro Gly Glu Thr Val Lys 65 70 75 80

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eol f-seql

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Thr II e Asp Val Pro Trp Asn Val Val Pro Glu Lys Ala His Leu Leu 50 60

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Lys Val Asn Cys Thr Thr Gln Pro Cys Pro Thr Ala Lys Ala Pro Thr 35 40 45

Cys Gly Leu Cys Glu Val Ala Arg Leu Arg Gln Asn Ala Asp Gln Cys 50 60 Cys Pro Glu Tyr Glu Cys Val Cys Asp Pro Val Ser Cys Asp Leu Pro 65 70 75 80 Pro Val Pro His Cys Glu Arg Gly Leu Gln Pro Thr Leu Thr Asn Pro 85 90 95 Gly Glu Cys Arg Pro Asn Phe Thr Cys Ala Cys Arg Lys Glu Glu Cys 100 105 110 Lys Arg Val Ser Pro Pro Ser Cys Pro Pro His Arg Leu Pro Thr Leu 115 120 125 Arg Lys Thr Gln Cys Cys Asp Glu Tyr Glu Cys Ala Cys Asn Cys Val Asn Ser Thr Val Ser Cys Pro Leu Gly Tyr Leu Ala Ser Thr Ala Thr 145 150 155 160 Asn Asp Cys Gly Cys Thr Thr Thr Thr Cys Leu Pro Asp Lys Val Cys
165 170 175 Val His Arg Ser Thr IIe Tyr Pro Val Gly Gln Phe Trp Glu Glu Gly 180 185 190 Cys Asp Val Cys Thr Cys Thr Asp Met Glu Asp Ala Val Met Gly Leu 195 200 205 Arg Val Ala Gln Cys Ser Gln Lys Pro Cys Glu Asp Ser Cys Arg Ser 210 220 Gly Phe Thr Tyr Val Leu His Glu Gly Glu Cys Cys Gly Arg Cys Leu 225 230 235 240 Pro Ser Ala Cys Glu Val Val Thr Gly Ser Pro Arg Gly Asp Ser Gln 245 250 255 Ser Ser Trp Lys Ser Val Gly Ser Gln Trp Ala Ser Pro Glu Asn Pro 260 265 270 Cys Leu IIe Asn Glu Cys Val Arg Val Lys Glu Glu Val Phe IIe Gln 275 280 285 Gln Arg Asn Val Ser Cys Pro Gln Leu Glu Val Pro Val Cys Pro Ser 290 295 300 Gly Phe Gln Leu Ser Cys Lys Thr Ser Ala Cys Cys Pro Ser Cys Arg 305 310 315 320Page 24

eol f-seql

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Gly Lys Thr Val Met IIe Asp Val Cys Thr Thr Cys Arg Cys Met Val 340 $$ 345 $$ Thr Thr Cys Arg Cys Met Val $$

Gln Val Gly Val IIe Ser Gly Phe Lys Leu Glu Cys Arg Lys Thr Thr $355 \hspace{1.5cm} 360 \hspace{1.5cm} 365$

Cys Asn Pro Cys Pro Leu Gly Tyr Lys Glu Glu Asn Asn Thr Gly Glu 370 375 380

Cys Cys Gly Arg Cys Leu 385 390