

(19) **DANMARK**



Patent- og
Varemærkestyrelsen

(10) **DK/EP 2937359 T3**

(12) **Oversættelse af
europæisk patentskrift**

-
- (51) Int.Cl.: **C 07 K 1/18 (2006.01)** **C 07 K 14/765 (2006.01)**
- (45) Oversættelsen bekendtgjort den: **2018-09-17**
- (80) Dato for Den Europæiske Patentmyndigheds bekendtgørelse om meddelelse af patentet: **2018-06-13**
- (86) Europæisk ansøgning nr.: **13864364.8**
- (86) Europæisk indleveringsdag: **2013-05-09**
- (87) Den europæiske ansøgnings publiceringsdag: **2015-10-28**
- (86) International ansøgning nr.: **CN2013075405**
- (87) Internationalt publikationsnr.: **WO2014094406**
- (30) Prioritet: **2012-12-21 CN 201210559390**
- (84) Designerede stater: **AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL PT RO RS SE SI SK SM TR**
- (73) Patenthaver: **WUHAN HEALTHGEN BIOTECHNOLOGY CORP, No.666 Gaoxin Avenue, East Lake High-Tech Development Zone, Wuhan, Hubei 430079, Kina**
- (72) Opfinder: **YANG, Daichang, 666 Gaoxin Avenue, East Lake High-Tech Development Zone, , Wuhan, Hubei 430079, Kina**
SHI, Bo, 666 Gaoxin Avenue, East Lake High-Tech Development Zone, Wuhan, Hubei 430079, Kina
SHI, Qianni, 666 Gaoxin Avenue, East Lake High-Tech Development Zone, Wuhan, Hubei 430079, Kina
OU, Jiquan, 666 Gaoxin Avenue, East Lake High-Tech Development Zone, Wuhan, Hubei 430079, Kina
LIU, Jingru, 666 Gaoxin Avenue, East Lake High-Tech Development Zone, , Wuhan, Hubei 430079, Kina
- (74) Fuldmægtig i Danmark: **HØIBERG P/S, Adelgade 12, 1304 København K, Danmark**
- (54) Benævnelse: **Kromatografisk fremgangsmåde til isolering og oprensning af rekombinant humant serumalbumin med høj renhed**
- (56) Fremdragne publikationer:
WO-A1-90/15617
WO-A1-2005/003152
WO-A1-2012/083580
CN-A- 101 260 145
CN-A- 101 665 798
CN-A- 102 127 164
CN-A- 102 190 722
CN-A- 102 532 254
US-A- 5 917 022
LIN M F ET AL: "Removal of lipopolysaccharides from protein-lipopolysaccharide complexes by nonflammable solvents", JOURNAL OF CHROMATOGRAPHY B: BIOMEDICAL SCIENCES & APPLICATIONS, ELSEVIER, AMSTERDAM, NL, vol. 816, no. 1-2, 25 February 2005 (2005-02-25), pages 167-174, XP027626884, ISSN: 1570-

Fortsættes ...

0232 [retrieved on 2005-02-25]

BURNOUF T ET AL: "Affinity chromatography in the industrial purification of plasma proteins for therapeutic use", JOURNAL OF BIOCHEMICAL AND BIOPHYSICAL METHODS, AMSTERDAM, NL, vol. 49, no. 1-3, 30 October 2001 (2001-10-30), pages 575-586, XP002353092, ISSN: 0165-022X, DOI: 10.1016/S0165-022X(01)00221-4
BELEW M ET AL: "Purification of Recombinant Human Serum Albumin (rHSA) Produced by Genetically Modified Pichia Pastoris", SEPARATION SCIENCE AND TECHNOLOGY, DEKKER, NEW YORK, NY, US, vol. 43, no. 13, 31 August 2008 (2008-08-31), pages 3134-3153, XP008169071, ISSN: 0149-6395, DOI: 10.1080/01496390802221857 [retrieved on 2008-08-01]

DESCRIPTION

Field of the Invention

[0001] The invention belongs to the field of biotechnology, and in particular relates to a method for isolating and purifying high-purity recombinant human serum albumin (OsrHSA) for clinical application.

Background of the Invention

[0002] Human serum albumin (HSA) is a non-glycosylated single chain protein consisting of 585 amino acids, having a molecular weight of 66.5 kD and an isoelectric point between 4.7-4.9. It is the most abundant protein in human blood plasma, making up about 60% of the total plasma proteins. There is about 40 g of HSA in per liter of human blood. Besides being present in the plasma, HSA is also found in tissues and body secretions, skins and lymph cavities. Under normal physiological conditions of human beings, HSA has an effect of maintaining plasma colloid osmotic pressure, nourishing, accelerating the concrescence of wounds, and as a carrier, participating in transportation of many hydrophobic biological molecules such as hormones, biological active substances and drugs in the blood. Therefore, HSA is an important medical protein that is mainly used clinically for the treatment of hypoproteinemia caused by loss of blood, burn, scald, plastic surgery and brain lesion, as well as for the treatment of liver cirrhosis, hydronephrosis and so on.

[0003] At present, HSA for clinical use is mainly prepared by extracting and isolating from human plasma. However, this preparation approach has the following disadvantages: on one hand, the source of plasma is insufficient, i.e. the limited blood supply is unable to meet the demands of production of HSA and the relevant preparations thereof; on the other hand, blood itself may potentially be a risk factor, for example, it may contain dangerous infectious pathogens such as hepatitis virus, human immunodeficiency virus (HIV) and so on, which causes enormously concerns about the application of HSA extracted from plasma.

[0004] With the development of modern DNA recombinant and synthesis techniques, the researchers take a profound interest in the production and application of recombinant human serum albumin (OsrHSA). So far, people have tried to use various expression systems for mass production of OsrHSA. For example, prokaryotes such as *E. coli* (Latta, M. et al., *Bio/Technology*, 5:1309-1314,(1987)), *Bacillus subtilis* (Saunders, C. W. et al, *J. Bacteriol.* 169: 2917-2925, (1987)), eukaryotes such as yeasts (WO 00/44772, EP0683233A2, US 5612196) and cultivation of animal cells have been used for the production of OsrHSA. However, such approaches are not suitable for industrial production due to low expression level or high production cost.

[0005] Chinese patent application No. 201010606635.8 and WO12083580 of the present inventors discloses a method for extracting OsrHSA from rice. Based on the method, the present invention further studies the process for removing endotoxin from OsrHSA and improving the protein purity >99.9999%, thereby obtaining this novel technical solution of the present invention. Lin et al, Journ. Chrom. B: Biomed. Sc. and Appl., 2005 discloses use of alcohol for removal of Lipopolysaccharide (LPS) in method for purification of recombinant proteins. WO05003152 also discloses use of alcohol for removal of LPS using anion exchange chromatography. Burnouf et al, Journ. Biomed. Biophys meth., 2001 discloses purification of plasma proteins, such as albumin, and states that albumin can be obtained by combining ethanol fractionation and chromatography.

Summary of the Invention

[0006] It is an object to provide a chromatography method for isolating and purifying high-purity recombinant human serum albumin from crude protein extract of transgenic rice seeds. The purity of the obtained recombinant human serum albumin can reach 99.9999%. The content of endotoxin meets the standards of human serum albumin stipulated in Chinese pharmacopoeia.

[0007] The invention is set out in the appended set of claims. The technical solution of the present invention involves:

A chromatography method for isolating and purifying high-purity recombinant human serum albumin (OsrHSA), comprising the following steps of:

1. 1) subjecting crude extract of recombinant human serum albumin to cation exchange chromatography, adding an alcohol in an buffer to remove endotoxin, to obtain primary product I;
2. 2) subjecting the primary product I to anion exchange chromatography, to obtain secondary product II;
3. 3) subjecting the secondary product II to hydrophobic interaction chromatography, to obtain the target product, high-purity recombinant human serum albumin.

[0008] The hydrophobic interaction chromatography is performed on a resin selected from Phenyl Sepharose HP, Phenyl Sepharose FF, Phenyl Bestarose HP or Phenyl Bestarose FF.

[0009] Specifically, the alcohol according to the method of the present invention is ethanol.

[0010] Specifically, the buffer according to the method of the present invention comprises a wash buffer, an equilibrium buffer I and an equilibrium buffer II; wherein the wash buffer comprises 10-20% anhydrous ethanol by volume, the equilibrium buffer I comprises 0-10% anhydrous ethanol by volume, the equilibrium buffer II comprises 5-15% anhydrous ethanol by

volume.

[0011] Preferably, the wash buffer comprises 15% anhydrous ethanol, and the equilibrium buffer II comprises 10% anhydrous ethanol. More preferably, the wash buffer comprises sodium acetate anhydrous 2 g/L, 15% anhydrous ethanol; the conductivity is adjusted with NaCl to 83 mS/cm, and pH is adjusted to 4.6-5.0 with acetic acid. The equilibrium buffer I comprises: sodium acetate anhydrous 2 g/L, NaCl 15 g/L, pH is adjusted to 4.2-4.8 with acetic acid. The equilibrium buffer II comprises: sodium acetate anhydrous 2 g/L, NaCl 15 g/L, 10% anhydrous ethanol, pH is adjusted to 4.2-4.8 with acetic acid.

Brief Description of the Drawings

[0012]

Fig.1 shows purification efficiency and loading capacity of Capto-MMC chromatography under three different conditions;

wherein, M: molecular mark, L: extract sample, M1: control group, M2: test group 1, M3: test group 2; FT: flow-through fraction of the loaded sample; M3FT: flow-through fraction of M3 test group, 600, 660 and 730 corresponding to the flow-through fraction of the loaded sample 600mL, 660 mL and 730 mL, respectively; wash: impurity-washing fraction containing OsrHSA; Elution: Elution fraction containing OsrHSA.

Fig.2 shows the comparison of purity of primary purification performed on Capto MMC and Bestarose Diamond MMC;

wherein, GE-MMC: Capto-MMC; Best-MMC: Bestarose Diamond MMC; Elu: Elution fraction from primary purification performed on the two resins; M: molecular mark; FT: flow-through fraction of the loaded sample; wash: impurity-washing fraction containing OsrHSA; Elu: Elution fraction containing OsrHSA; S: extract sample; CIP1: Elution fraction of resin regeneration 1; CIP2: Elution fraction of resin regeneration 2.

Fig.3 shows the comparison of purification efficiency of UNO Sphere S chromatography with or without adding alcohol;

wherein, CK: control group; UE: test group (10% ethanol was added in the sample and the equilibrium buffer); M: molecular mark; Load: sample; FT: flow-through fraction of the loaded sample; Elution: Collected elution fraction containing target protein under two conditions; CIP: resin regeneration.

Fig.4 shows the comparison of SDS-PAGE of the secondary purification performed on Capto-Adhere and Bestarose Diamond MMA under flow-through conditions;

wherein, M: molecular mark; MMC: sample; FT: flow-through fraction containing OsrHSA, CIP: resin regeneration.

Fig.5 shows the comparison of the secondary purification performed on Capto-Adhere and Bestarose Diamond MMA under binding conditions;

wherein M: molecular mark; L: sample; FT: flow-through fraction containing OsrHSA; Elu: elution fraction containing OsrHSA; C: resin regeneration.

Fig.6 shows the comparison of hydrophobic interaction chromatography performed on Phenyl Sepharose HP and Phenyl Bestarose HP as the final purification; wherein, M: molecular mark; Ad: loaded sample; FT: flow-through fraction containing OsrHSA; CIP: resin regeneration.

Fig.7 shows the image of impurity detection before and after the protein purification in the rice seeds;

wherein, Left: comparison image of SDS-PAGE of impurity detection before and after purification of human serum albumin in the rice seeds; Right: Western Blotting detection image of hybridization with full impurity antibody of rice seeds before and after purification of human serum albumin in the rice seeds; M: molecular mark.

Fig.8 shows the SDS-PAGE image of the purified OsrHSA according to one example of the present invention;

wherein, A: Capto-MMC chromatography; B: Capto-Adhere chromatography; C: Phenyl HP chromatography; L: loaded sample; FT: flow-through fraction; W: impurity-washing fraction; Elu: elution fraction; C: CIP of resin.

Fig.9 shows the SDS-PAGE image (Left) and the corresponding Western Blot image (Right) of the purified OsrHSA according to another example of the present invention;

wherein, A: Capto-MMC chromatography; B: Capto-Adhere chromatography; C: Phenyl HP chromatography; M: molecular mark; L: loaded sample; FT: flow-through fraction; W: impurity-washing fraction; E: elution fraction; C: CIP of resin.

Detailed Description of the Invention

[0013] The characteristics and advantages of the present disclosure will be described in detail in conjunction with the accompanying drawings. Unless otherwise specified, the materials and reagents used in the following examples were ordinary commercially available.

[Example 1] Preparation of OsrHSA Extract

[0014] Transgenic rice containing OsrHSA was prepared according to the method of Chinese patent NO.200510019084.4 and OsrHSA was extracted from the transgenic rice seeds according to the method of Chinese patent NO.201010606635.8 to obtain clear OsrHSA extract.

[Example 2] Selection of Conditions for Cation Exchange Chromatography as Primary

Purification

[0015] Method: referring to the method of Chinese Patent Application NO. 201010606635.8;
Groups: test group and control group

1. Cation exchange chromatography performed on Capto MMC as primary purification and process for removing endotoxin with alcohol

Capto MMC resin was used to perform cation exchange chromatography as primary chromatography and ethanol was added into the buffer of the test group to remove endotoxin, while no ethanol was added into the control group (M1).

Test group 1(M2): The equilibrium buffer and the sample were added with 10% ethanol by volume, and the particular process was:

Column-packing: About 30 mL of Capto MMC resin was packed in an XK16/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid) until the pH reached 4.5 and was on the base line;

Sample-loading: The clear extract sample containing OsrHSA was added with 10% (v/v) anhydrous ethanol and 11g/L NaCl, and NaOH was used to adjust pH to 4.5; the sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 20-21 mS/cm, pH was 4.5;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 100 mL of equilibrium buffer (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, NaCl 58.5g/L, pH 4.8 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Test group 2 (M3): The equilibrium buffer and the sample were added with 10% ethanol, the impurity wash buffer was added with 20% ethanol, and the particular process was:

Column-packing: About 30 mL of Capto MMC resin was packed in an XK16/400 mm chromatography column (bed height: 21cm), and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer (sodium acetate

anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: The clear extract sample containing OsrHSA was added with 10% (v/v) anhydrous ethanol and 11g/L NaCl, pH 4.5 adjusted with NaOH. The sample was loaded at a flow rate of 600 cm/h. The conductivity of the sample was 20-21 mS/cm, and pH was 4.5;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 100 mL of equilibrium buffer (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, NaCl 120 g/L, pH 4.8 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Results: The determination results of purity and loading capacity are shown in Fig.1. There was no obvious difference in the purity between the control group and the two experimental groups. The addition of alcohol in the wash buffer was favorable to remove the 17-26KD purity bands. The addition of alcohol in the wash buffer did not influence the loading capacity of Capto-MMC. The FT of the three groups showed that the loading capacity of each group was above 30mL extract/mL resin.

Endotoxin: As shown in Table 1, the addition of alcohol in the wash buffer of Capto-MMC chromatography had good removal efficiency on endotoxin. When the equilibrium buffer and extraction buffer were added with 10% (v/v) ethanol and the wash buffer was added with 20% ethanol, Capto-MMC chromatography had the best efficiency for removal of endotoxin, which was 3.6 times of the control group.

Table 1 Comparison of endotoxin contents of Capto-MMC chromatography containing an alcohol in the wash buffer

Group	Extract (EU/mL)	Total EU of loaded sample	Collected fraction (EU/mL)	Total EU of collected fraction	n-fold decrease of total EU
Control group (M1)		3×10^5	400-600	1.25×10^4	24
Test group 1 (M2)	1500-2000	3.3×10^5	100-200	4.5×10^3	76
Test group 2 (M3)		3.7×10^5	100-200	4.3×10^3	87

Group	Extract (EU/mL)	Total EU of loaded sample	Collected fraction (EU/mL)	Total EU of collected fraction	n-fold decrease of total EU
-------	-----------------	---------------------------	----------------------------	--------------------------------	-----------------------------

2. Optimization of process for removing endotoxin by Capto MMC chromatography with an alcohol added in the buffer

2.1 The removal efficiency of endotoxin with or without adding an alcohol in the extract sample was compared, and the particular process was:

Control group 1 (M1): The equilibrium buffer and the extract sample were added with 10% ethanol, and the wash buffer was added with 20% ethanol, the particular process was:

Column-packing: About 29 mL of Capto MMC resin was packed in a BioRad 15/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer I (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: The clear extract solution sample containing OsrHSA was added with 10% (v/v) anhydrous ethanol and 11g/L NaCl, pH 4.5 adjusted with NaOH; 870 mL of sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 20-21 mS/cm, pH was 4.5; the content of endotoxin in the sample was 1000-2000EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 100 mL of equilibrium buffer II (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, 20% (v/v) anhydrous ethanol, a conductivity of 83 mS/cm adjusted with NaCl, pH 4.8 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Test group (M2): Neither the extract sample nor the equilibrium buffer I were added with ethanol. After loading sample, the chromatography column was equilibrated with 150 mL of equilibrium buffer II containing 10% ethanol (about 5CV), the wash buffer was added with 20% ethanol; the other conditions were the same as M1 group.

The results are shown in Table 2. The decrease of the total EU was basically identical in the two parallel test groups. The extract sample and equilibrium buffer with or without alcohol has no obvious effect on endotoxin removal. There was little difference in the concentration and

volume of the collected fraction between the two groups, and the contents of endotoxin were the same. This further showed that the same efficiency of endotoxin removal can be achieved even there was no alcohol in the sample and equilibrium buffer I.

Table 2: Comparison of endotoxin of Capto-MMC chromatography when the sample and equilibrium buffer I had or had no alcohol

Test group	Loaded sample (mL)	Total EU of loaded sample	Collected fraction (mL)	Endotoxin of collected fraction (EU/mL)	Total EU of collected fraction	n-fold decrease
M1	870	1.31×10^6	95	200-400	2.4×10^4	55
M2	870	1.31×10^6	100	200-300	2.5×10^4	53

2.2 Comparison of endotoxin removal efficiency when different amounts of an alcohol were added in the wash buffer

Control group 1 (M1):

Column-packing: About 29 mL of Capto MMC resin was packed in a BioRad 15/400 mm chromatography column, and then the column was washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer I (sodium acetate anhydrous 2g/L, NaCl 15g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: The clear extract sample containing OsrHSA was added with 11g/L NaCl, pH was adjusted to 4.5 with NaOH; 650 mL of sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 20-21 mS/cm, pH was 4.5, the content of endotoxin in the sample was 1000-2000 EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 150 mL of equilibrium buffer II (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2 g/L, 20% (v/v) anhydrous ethanol, a conductivity of 83 mS/cm adjusted with NaCl, pH 4.8 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Test group 1 (M2): Except that 15% (v/v) anhydrous ethanol was added in the wash buffer, the conditions were the same as M1 group.

Test group 2 (M3): Except that 10% (v/v) anhydrous ethanol was added in the wash buffer, the conditions were the same as M1 group.

The results are shown in Table 3.

Table 3 Comparison of endotoxin content when different amounts of alcohol were added in the wash buffer

Test group	Total EU of loaded sample	Collected fraction (mL)	Endotoxin of collected fraction (EU/mL)	Total EU of collected fraction	n-fold decrease
M1	1.17×10^6	78	200-300	1.9×10^4	62
M2	1.17×10^6	90	200-300	2.25×10^4	52
M3	1.17×10^6	88	300-400	3.08×10^4	38

As shown in Table 3, the total EU of endotoxin in M3 group was 38-fold decrease, and had significant difference compared with M1 group (control group); 15% ethanol was added in M2 group, the total EU of endotoxin in M2 group was 52-fold decrease; the content of endotoxin of the collected fraction in M2 group was the same as that of M1 group and has no obvious difference compared with M1 group (control group). The concentration of ethanol in the wash buffer of MMC can be decreased to about 15%, which had no obvious influence on endotoxin removal efficiency.

2.3 Endotoxin removal efficiency when the wash buffer contained isopropyl alcohol

Control group (CK): Column-packing: About 29 mL of Capto MMC resin was packed in a BioRad 15/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer I (sodium acetate anhydrous 2g/L, NaCl 15g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: 900 mL of sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 5-8 mS/cm, pH was 4.5, the content of endotoxin in the sample was 1000-2000EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 150 mL of equilibrium buffer II (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.6-4.7 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, a conductivity of 82-89 mS/cm adjusted with NaCl, 15% (v/v) anhydrous ethanol, pH 4.7-4.9 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Test group 1(M1): Column packing: About 29 mL of Capto MMC resin was packed in a BioRad 15/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer I (sodium acetate anhydrous 2g/L, NaCl 15g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: 900 mL of sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 5-8 mS/cm, pH was 4.5, the content of endotoxin in the sample was 1000-2000EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 150 mL of equilibrium buffer II (sodium acetate anhydrous 2g/L, NaCl 15g/L, 4-6% (v/v) isopropyl alcohol, pH 4.6-4.7 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, 10-11% (v/v) isopropyl alcohol, a conductivity of 82-89 mS/cm adjusted with NaCl, pH 4.7-4.9 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Test group 2 (M2): Except that the equilibrium buffer II was added with 10-11% (v/v) isopropyl alcohol instead of 4-6% (v/v) isopropyl alcohol, the wash buffer was added with 15-16% (v/v) isopropyl alcohol instead of 10-11% (v/v) isopropyl alcohol, and the conductivity was reduced to 75-76mS/cm, the conditions were the same as that of M1 group.

The results are shown in Table 4.

Table 4 Comparison of endotoxin removal efficiency when isopropyl alcohol was added in the wash buffer

Group	Total EU of loaded sample	Collected fraction (mL)	Endotoxin of collected fraction (EU/mL)	Total EU of collected fraction	n-fold decrease
CK	8.9×10^5	110	100-200	1.7×10^4	52
M1	7.2×10^5	110	10-20	1.6×10^3	450
M2	8.0×10^5	112	5-10	1.1×10^3	710

As shown in Table 4, the total EU of endotoxin in the control group was 52-fold decrease, and that of M1 group and M2 group were 450-fold and 710-fold decrease, respectively. The removal efficiency of endotoxin was significantly improved compared to the process that ethanol was added in wash buffer. In M2 group, 15% (v/v) isopropyl alcohol was added, and the content of endotoxin was 710-fold decrease, the endotoxin removal efficiency of which was comparable to that of M1 group. This demonstrated that the amount of isopropyl alcohol added in the equilibrium buffer could be 4-11% (v/v), the amount of isopropyl alcohol added in the wash buffer could be 10-16% (v/v), and there was no obvious effect on endotoxin removal when the conductivity was in the range of 75-89mS/cm.

3. Selection of chromatography resin for cation exchange chromatography and the removal efficiency of endotoxin with an alcohol added in the wash buffer

3.1 Bestarose Diamond MMC was used to perform cation exchange chromatography, and endotoxin was removed by adding an alcohol in the buffer. The particular process was:

Column-packing: About 33 mL of Bestarose Diamond MMC resin was packed in a BioRad 15/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 200 mL of equilibrium buffer I (sodium acetate anhydrous 2g/L, NaCl 15g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was on the base line;

Sample-loading: The extract solution sample containing OsrHSA was added with 11g/L NaCl, pH was adjusted to 4.5 with NaOH; 600 mL of sample was loaded at a flow rate of 600 cm/h, and the conductivity of the sample was 20-21 mS/cm, pH was 4.5, the content of endotoxin in the sample was 1500-2000EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 150 mL of equilibrium buffer II (sodium acetate anhydrous 2g/L, NaCl 15g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300 cm/h;

Impurity-washing impurity: 200 mL of wash buffer (sodium acetate anhydrous 2g/L, 15% (v/v) anhydrous ethanol, a conductivity 83 mS/cm adjusted with NaCl, pH 4.8 adjusted with acetic acid, prepared with water for injection) was used to elute the impurities at a flow rate of 300 cm/h;

Elution: Elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Results: As shown in Fig.2, purity was no obvious difference between Bestarose Diamond MMC and Capto-MMC resins. This demonstrated that Bestarose Diamond MMC can be replaced of Capto-MMC for the primary purification of OsrHSA.

Endotoxin: After the extract sample was subjected to primary purification on Bestarose

Diamond MMC resin with an alcohol added in the buffer, the total EU of endotoxin was 62-fold decrease, achieving the same endotoxin removal efficiency as Capto-MMC resin.

3.2 Cation exchange chromatography performed on UNO Sphere S and process of endotoxin removal with an alcohol added in the buffer

Control group (CK): referring to the method of the Chinese Patent CN 201010606635.8 of the present inventors. The loaded sample was 400 mL.

[0016] Test group (UE), the particular process was:

Column-packing: About 18 mL of UNO Sphere S resin was packed in a BioRad 15/200 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens;

Equilibration: The column was equilibrated with 100 mL of equilibrium buffer (sodium acetate anhydrous 2g/L, 10% (v/v) anhydrous ethanol, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h until the pH reached 4.5 and was constant;

Sample-loading: The extract solution sample containing rHSA was added with 10% (v/v) anhydrous ethanol, pH was adjusted to 4.5 with acetic acid; 400 mL of the sample was loaded at a flow rate of 300 cm/h, and the content of endotoxin in the sample was 800-1000EU/mL;

Re-equilibration: After sample-loading, the chromatography column was re-equilibrated with 60 mL of equilibrium buffer at a flow rate of 300 cm/h;

Elution: Elution buffer (anhydrous sodium acetate 2g/L, NaCl 14.6 g/L, pH 5.5 adjusted with acetic acid, prepared with water for injection) was used to elute the target protein, obtaining the fraction containing OsrHSA.

Results: As shown in Fig. 3, purity was no obvious difference between UNO Sphere S control group and the alcohol addition group. That is, UNO Sphere S resin chromatography by adding an alcohol can be used for the primary purification.

Endotoxin: After the extraction sample was subjected to primary purification on UNO Sphere S resin with alcohol addition, the decrease of the total EU of endotoxin was the same as that of the control group. This demonstrated that the alcohol in the wash buffer cannot enable UNO Sphere S to have stronger endotoxin removal efficiency. This may be relevant to the matrix of UNO Sphere S resin itself, and both Capto-MMC and Bestarose Diamond MMC resins have complex multifunctional ligands and have weak cation exchange and hydrophobic property.

[Example 3] Selection of conditions for anion exchange chromatography as intermediate purification

[0017]

1. Under OsrHSA flow-through conditions (i.e., under the conditions of the conductivity of 40mS/cm and pH 7.0, OsrHSA flowed through instead of binding to the resin, while the impurities bond to the resin, and thus OsrHSA was separated from the impurities). Anion exchange chromatography was performed on Capto-Adhere and Bestarose Diamond MMA.

About 40mL of Capto-Adhere resin was packed in an XK16×400mm chromatography column, and the packing height was 20.5cm. The column was equilibrated with about 8 CV of equilibrium buffer (25mM PB, NaCl 23.4g/L, pH7.0) until UV, pH and conductivity reached the baseline. The elution buffer sample obtained from Capto-MMC or Bestarose Diamond MMC chromatography was adjusted to pH 7.0, and 288mL of the sample was loaded at a flow rate of 300cm/h. When the ultraviolet absorption value of flow-through fraction was >20mAU, the target fraction containing OsrHSA was collected. The sample was taken from each 20mL of fraction to monitor the loading capacity and the total amount of the sample when the purity of the flow-through fraction was not obviously changed; the actual loading capacity of each milliliter of Capto-Adhere resin at a flow rate of 300cm/h was calculated.

Under the same OsrHSA flow-through conditions as Capto-Adhere, the parallel test was performed on Bestarose Diamond MMA resin to compare the purification effect and loading capacity.

Under the OsrHSA flow-through conditions, after the secondary purification with the two anion resins, HPLC purity of the target protein can reach above 98%. As shown in Fig.4, there was no significant difference on purity between the two resins.

2. Anion change chromatography performed on Capto-Adhere resin under OsrHSA-binding conditions

OsrHSA-binding conditions mean that the target protein first bond to the resin and part of impurities flowed through the column, and then the target protein was eluted through particular salt and pH, to achieve the effect of separation and purification, and enrichment of target product. The conditions included: the target protein bond to the anion exchange resin under the conditions of the conductivity of 20mS/cm and pH of 7.0-7.5, and then the target protein was eluted under the conditions of the conductivity of 40mS/cm and pH of 7.0-7.2 to remove the impurities and enrich the target protein.

About 22 mL of Capto-Adhere resin was packed in a BioRad 15/200 mm chromatographic column, and then the column was equilibrated with 220mL of equilibrium buffer (sodium dihydrogen phosphate 1.0g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 5.0 g/L, pH 7.5-7.6) at a flow rate of 300cm/h until the pH reached 7.5-7.6 and stable. The elution fraction sample from Capto-MMC or Bestarose Diamond MMC chromatography was diluted with water to reach a conductivity of 19-21mS/cm, and the pH was adjusted to 7.0 with NaOH. 150 mL of the sample was loaded at a flow rate of 300cm/h. The column was re-equilibrated with 100mL of equilibrium buffer. The target protein was eluted with the elution buffer (sodium dihydrogen phosphate 1.5g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 23.4 g/L, pH 7.1-7.2) at a flow rate of 300cm/h, collecting the fraction containing OsrHSA. The content of endotoxin in the

collected fraction was 100-200EU/mL.

Under the same OsrHSA-binding conditions as Capto-Adhere resin, the parallel test was performed on Bestarose Diamond MMA resin to compare the purification effect and loading capacity.

Under the OsrHSA-binding conditions, after secondary purification with Capto-Adhere resin or Bestarose Diamond MMA resin, the purity of the target protein could reach above 99.9% (ELISA detection results). Bestarose Diamond MMA resin and Capto-Adhere resin have the same purification efficiency. The electrophoretograms are shown in Fig. 5.

3.3. Selection of chromatography conditions for anion exchange chromatography performed on Capto-Adhere and Bestarose Diamond MMA

By the parallel comparison of two resins of Bestarose Diamond MMA and Capto-Adhere, it was found that under the same conditions, the two resins had the substantially identical purification efficiency and loading capacity on the elution fraction obtained from the secondary purification performed on Capto-MMC or Bestarose Diamond MMC resin, demonstrating that both the two anion exchange chromatography resins can be used for the secondary purification.

[0018] The loading capacity of the two resins was higher under OsrHSA flow-through conditions than that of OrsHSA-binding conditions. The loading capacity of the former was about 25mg sample/ mL resin, while the loading capacity of the latter was about 20mg sample / mL resin. However, the purification efficiency under OrsHSA-binding conditions was better than that under OsrHSA flow-through conditions. The purity of the former can reach above 99.9%, and the purity of the latter was only 98-99%.

[0019] Under OrsHSA-binding conditions, the two resins had better endotoxin removal efficiency than flow-through conditions. Under OsrHSA flow-through conditions, most of the endotoxin flowed through the column together with OsrHSA, resulting in a poorer endotoxin removal efficiency. But under OrsHSA-binding conditions, part of free endotoxin flowed through, and another part of endotoxin bond to the resin tightly so that OsrHSA can be selectively eluted, resulting in a better endotoxin removal efficiency.

[0020] Taking various factors into consideration, it is preferable to use the OrsHSA-binding conditions as the chromatography conditions of the two anion exchange resins, Capto-Adhere and Bestarose Diamond MMA.

[Example 4] Selection of Conditions for Hydrophobic Chromatography as Final Purification

1. Hydrophobic chromatography performed on Phenyl Sepharose HP

[0021] Referring to the method of Chinese patent CN201010606635.8 of the present applicant, about 28mL of Phenyl sepharose HP resin was packed in a 20 XK16/200 mm chromatography column, and the column was equilibrated with 150mL of equilibration buffer (anhydrous acetic acid sodium 2.32g/L, sodium dihydrogen phosphate 2.81g/L, sodium octanoate 2g/L, ammonium sulfate, 66g/L, pH6.5) at a flow rate of 100cm/h. 100mL of the elution fraction containing OsrHSA obtained from Capto-Adhere or Bestarose Diamond MMA chromatography was added with ammonium sulfate to adjust the conductivity to 75mS/cm, and then added with 0.15g of sodium caprylate, and then adjust pH to 6.5 with hydrochloric acid. It was loaded on the column at a flow rate of 100cm/h. The flow-through was collected to obtain the fraction containing OsrHSA. The fraction samples were taken to determine the loading capacity and calculate the actual loading capacity of the sample from secondary purification per millilitre resin. The electrophoretogram is shown in Fig. 6.

2. Hydrophobic chromatography performed on Phenyl Bestarose HP

[0022] About 24mL of Phenyl Bestarose HP resin was packed in an XK16/200 mm chromatography column. Parallel test was carried out according to the above method mentioned in 1 of this example. The electrophotograms are shown in Figs. 6 and 7.

Results and analysis:

[0023] After the fraction sample obtained from secondary purification with Capto-Adhere or Bestarose Diamond MMA was subjected to hydrophobic interaction chromatography with Phenyl Sepharose HP or Phenyl Bestarose HP, the purity of target protein could reach 99.9999% (by ELISA). The impurity proteins in the rice seed were not detected. The two hydrophobic resins had no significant difference on purity. The endotoxin contents of the fractions containing OsrHSA obtained from hydrophobic interaction chromatography performed on the two resins were less than 0.06EU/mg. The results showed that both Phenyl Sepharose HP and Phenyl Bestarose HP hydrophobic chromatography resin can be used for the final purification of OsrHSA, and can achieve a better purification and endotoxin removal.

[Example 5] Purification of Recombinant Human Serum Albumin

[0024] Cation exchange chromatography as primary purification: About 400mL of Capto MMC resin was packed in an XK50/400 mm chromatography column and then washed with 0.5N NaOH for 30 min to kill the pyrogens. The chromatography column was equilibrated with 2L of equilibration buffer I (anhydrous acetic acid sodium 2 g/L, NaCl 11g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h, until pH reached 5.0 and was stable. 10L of sample was loaded on the column, and the sample had a conductivity of 20.3 mS/cm and pH of 4.8. After

sample-loading, the chromatography column was equilibrated with 2L of equilibration buffer II (anhydrous acetic acid sodium 2 g/L, NaCl 15 g/L, 10-11% (v/v) anhydrous ethanol, pH 4.8 adjusted with acetic acid, prepared by water for injection) at a flow rate of 300 cm/h. The impurities were eluted with 3000 mL of wash buffer (anhydrous acetic acid sodium 2 g/L, 16% (v/v) anhydrous ethanol, a conductivity of 83.5mS/cm adjusted with NaCl, pH 5.0 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300cm/h. The target protein was eluted with the elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with injectable water) to obtain the fraction containing OsrHSA. The electrophotograms are shown in Fig. 8A.

[0025] Anion/hydrophobic composite resin used for exchange chromatography as secondary purification: About 68mL of Cpto-Adhere resin was packed in an XK26/200 mm chromatography column. The column was equilibrated with 600 mL of equilibration buffer (sodium dihydrogen phosphate 1.0 g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 5.0 g/L, pH 7.5-7.6) at a flow rate of 300cm/h, until the pH reach 7.5-7.6 and was stable. 500mL of sample was loaded on the column at a flow rate of 300cm/h. The column was re-equilibrated with about 200 mL of the equilibration buffer, and then eluted with elution buffer (sodium dihydrogen phosphate 1.5 g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 23.4 g/L, pH7.1-7.2) at a flow rate of 300cm/h, to collect the fraction containing OsrHSA. The content of endotoxin of the collected fraction was 100-200 EU/mL. The electrophotograms are shown in Fig. 8B.

[0026] Hydrophobic chromatography as final purification: About 15mL of Phenyl sepharose HP resin was packed in an XK16/200 mm chromatography column. The column was equilibrated with 100mL of equilibration buffer (anhydrous acetic acid sodium 2.32 g/L, sodium dihydrogen phosphate 2.81 g/L, sodium caprylate 2g/L, ammonium sulfate 66 g/L, pH 6.5) at a flow rate of 100cm/h. 100 mL of the elution fraction containing OsrHSA obtained from Cpto-Adhere chromatography was added with ammonium sulfate to adjust the conductivity to 75 mS/cm, then added with 0.15 g of sodium caprylate, hydrochloric acid to adjust pH to 6.5. The sample was loaded at a flow rate of 100cm/h. The flow-through was collected as the fraction containing OsrHSA. The content of endotoxin was less than 0.08 EU/mg, and the purity of the target protein was more than 99.9999%. The electrophotograms are shown in Fig. 8C.

[Example 6] Purification of Recombinant Human Serum Albumin

[0027] About 400 mL of Cpto MMC resin was packed in an XK50/400 mm chromatography column, and then washed with 0.5N NaOH for 30 min to kill the pyrogens. The chromatography column was equilibrated with 2000 mL of equilibration buffer I (anhydrous acetic acid sodium 2 g/L, NaCl 11 g/L, pH 4.5 adjusted with acetic acid) at a flow rate of 300cm/h, until pH reached 4.5 and was stable. 10000 mL of sample with a conductivity of 5.5-8.0 mS/cm and a pH of 4.5 was loaded on the column. After sample-loading, the chromatography column was re-equilibrated with 2000 mL of equilibration buffer II (anhydrous acetic acid sodium 2 g/L, NaCl 15g/L, 10-11% (v/v) isopropyl alcohol, pH 4.5 adjusted with acetic acid, prepared by injectable

water) at a flow rate of 300 cm/h. The impurities were eluted with 3000 mL of wash buffer (anhydrous acetic acid sodium 2 g/L, 15-16% (v/v) isopropyl alcohol, a conductivity of 82-89 mS/cm adjusted with NaCl, pH 4.8-4.9 adjusted with acetic acid, prepared with water for injection) at a flow rate of 300cm/h. The target protein was eluted with elution buffer (sodium dihydrogen phosphate 2.67 g/L, disodium hydrogen phosphate 2.82 g/L, NaCl 23.4 g/L, pH 6.3-6.4, prepared with water for injection) to obtain the fraction containing OsrHSA. The electrophotograms are shown in Fig. 9A.

[0028] Anion /hydrophobic composite resin used for chromatography as secondary purification: About 68mL of Capto-Adhere resin was packed in an XK26/200 mm chromatography column. The column was equilibrated with 600mL of equilibration buffer (sodium dihydrogen phosphate 1.0 g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 5.0 g/L, pH 7.5-7.6) at a flow rate of 300cm/h, until the pH reach 7.5-7.6 and was stable. 500 mL of sample was loaded on the column at a flow rate of 300cm/h. The column was re-equilibrated with about 200 mL of the equilibration buffer, and then eluted with wash buffer (sodium dihydrogen phosphate 1.5 g/L, disodium hydrogen phosphate 5.0 g/L, NaCl 23.4 g/L, pH 7.1-7.2) at a flow rate of 300cm/h, to collect the fraction containing OsrHSA. The content of endotoxin of the collected fraction was 100-200 EU/mL. The electrophotograms are shown in Fig. 9B.

[0029] Hydrophobic chromatography as final purification: About 15mL of Phenyl sepharose HP resin was packed in an XK16/200 mm chromatography column. The column was equilibrated with 100 mL of equilibration buffer (anhydrous acetic acid sodium 2.32g/L, sodium dihydrogen phosphate 2.81 g/L, sodium caprylate 2g/L, ammonium sulfate 66g/L, pH 6.5) at a flow rate of 100cm/h. 100mL of the elution fraction containing OsrHSA obtained from Capto-Adhere chromatography was added with ammonium sulfate to adjust the conductivity to 75 mS/cm, then added with 0.15g of sodium caprylate, pH 6.5 adjusted with hydrochloric acid, loaded at a flow rate of 100cm/h. The flow-through was collected as the fraction containing OsrHSA. The content of endotoxin was less than 0.08 EU/mg, and the purity of the target protein was more than 99.9999%. The electrophotograms are shown in Fig. 9C.

REFERENCES CITED IN THE DESCRIPTION

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Patent documents cited in the description

- [WO0044772A](#) [0004]
- [EP0683233A2](#) [0004]
- [US5612196A](#) [0004]
- [CN201010606635](#) [0005] [0014] [0015] [0015] [0021]
- [WO12083580A](#) [0005]
- [WO05003152A](#) [0005]
- [CN200510019084](#) [0014]

Non-patent literature cited in the description

- **LATTA, M. et al.** Bio/Technology, 1987, vol. 5, 1309-1314 [0004]
- **SAUNDERS, C. W. et al.** J. Bacteriol., 1987, vol. 169, 2917-2925 [0004]
- **LIN et al.** Journ. Chrom. B: Biomed. Sc. and Appl., 2005, [0005]
- **BURNOUF et al.** Journ. Biomed. Biophys meth., 2001, [0005]

PATENTKRAV

1. Fremgangsmåde til isolering og oprensning af rekombinant humant serumalbumin med høj renhed, som sekventielt omfatter følgende trin:

- 5 a) udsættelse af rå ekstrakt af rekombinant humant serumalbumin for kation-
bytningskromatografi, tilsætning af ethanol i buffere for at fjerne endotoxin for at
opnå primærprodukt I; nævnte buffere omfatter en vaskebuffer, en ækvilibre-
ringsbuffer I og en ækvilibreringsbuffer II; vaskebufferen omfatter 10-20% vand-
fri ethanol efter volumen, ækvilibreringsbufferen I omfatter 0-10% vandfri etha-
10 nol efter volumen, ækvilibreringsbufferen II omfatter 5-15% vandfri ethanol efter
volumen;
ækvilibrerung af kromatografisøjlen med ækvilibreringsbuffer I, påføring af prø-
ven, ækvilibrerung af kromatografisøjlen med ækvilibreringsbuffer II, eluering af
urenhederne med vaskebuffer, eluering af målprotein med elueringsbufferen;
15 idet kationbytningskromatografi udføres på en kation/hydrofob kompositresin
valgt blandt Capto-MMC eller Bestarose Diamond MMC;
- b) udsættelse af primærproduktet I for anionbytningskromatografi for at opnå
sekundært produkt II;
idet anionbytningskromatografi udføres på en anion/hydrofob kompositresin
20 valgt blandt Capto-Adhere eller Bestarose Diamond MMA;
- c) udsættelse af det sekundære produkt II for hydrofob kromatografi for at opnå
rekombinant humant serumalbumin med høj renhed;
- idet den hydrofobe kromatografi udføres på en resin valgt blandt Phenyl Sepha-
rose HP, Phenyl Sepharose FF, Phenyl Bestarose HP eller Phenyl Bestarose
25 FF.

2. Fremgangsmåde ifølge krav 1, hvor vaskebufferen omfatter 15% vandfri ethanol
efter volumen, og ækvilibreringsbufferen II omfatter 10% vandfri ethanol efter volumen.

3. Fremgangsmåde ifølge krav 1, hvor vaskebufferen omfatter: vandfrit natriumacetat
30 2 g/l, 15% vandfri ethanol efter volumen; idet konduktiviteten deraf justeres med NaCl
til 83 mS/cm, og pH deraf indstilles til 4,6-5,0 med eddikesyre.

4. Fremgangsmåde ifølge krav 1, hvor ækvilibreringsbufferen I omfatter: vandfrit natriumacetat 2 g/l, NaCl 15 g/l, og pH deraf indstilles til 4,2-4,8 med eddikesyre.

5. Fremgangsmåde ifølge krav 1, hvor ækvilibreringsbufferen II omfatter: vandfrit natriumacetat 2 g/l, NaCl 15 g/l, 10% vandfri ethanol efter volumen, og pH deraf indstilles til 4,2-4,8 med eddikesyre.

DRAWINGS

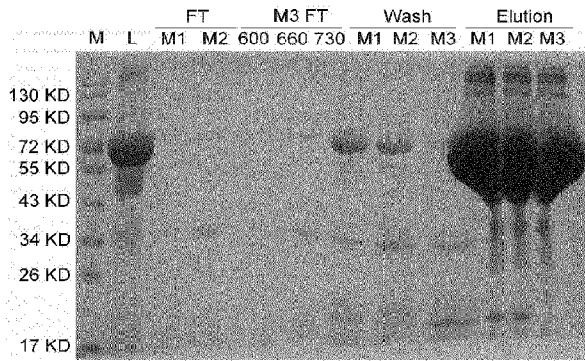


Fig. 1

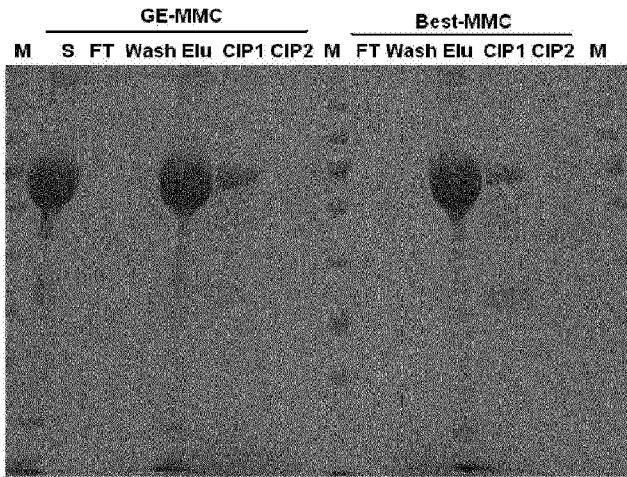


Fig. 2

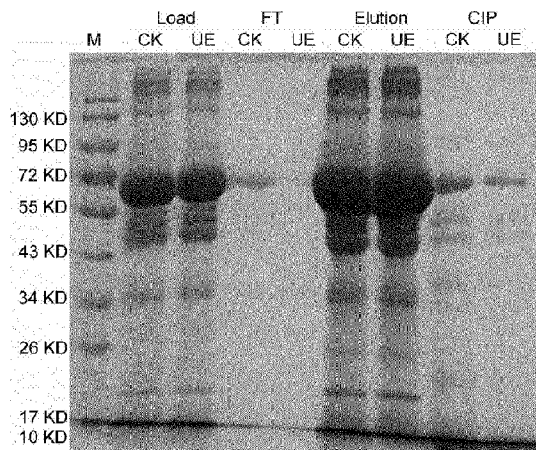


Fig. 3

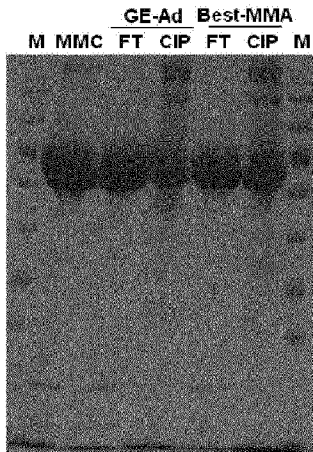


Fig. 4

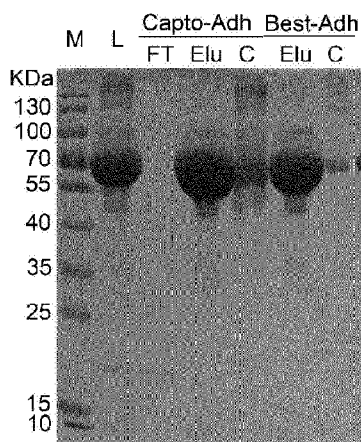


Fig. 5

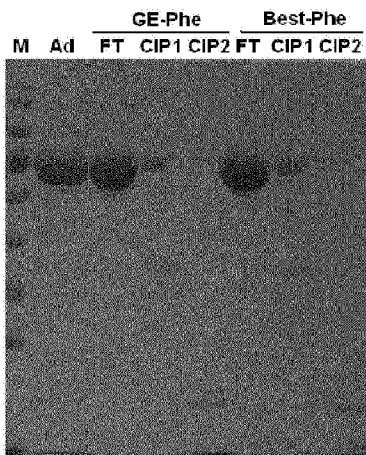


Fig. 6

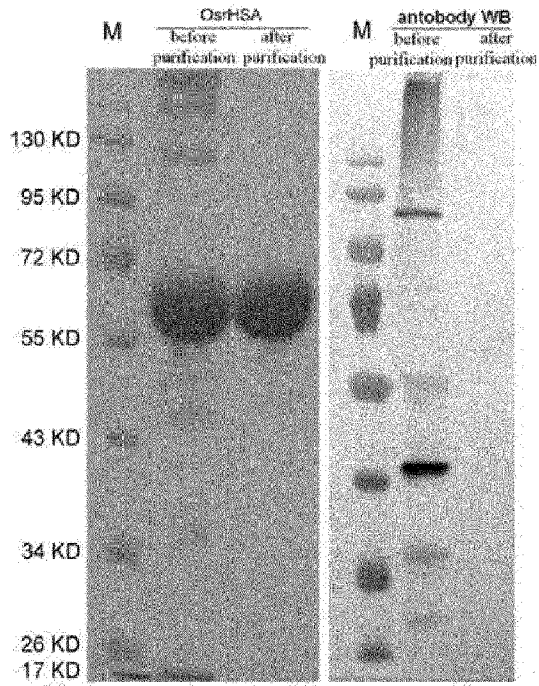


Fig. 7

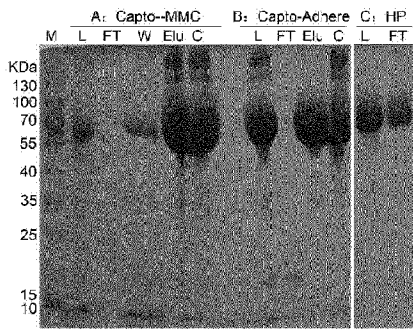


Fig. 8

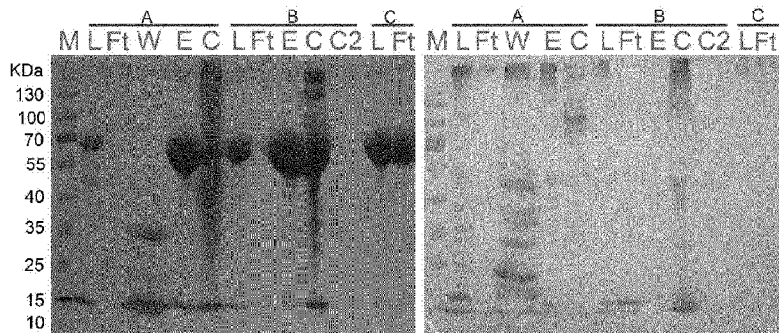


Fig. 9