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**Methods of Constructing Display Libraries of Genetic Packages for Members of a Diverse Family of Peptides**

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2007211861 21 Aug 2007

## ABSTRACT

Methods useful in constructing libraries that collectively display members of diverse families of peptides, polypeptides or proteins and the libraries produced using those methods. Methods of screening those libraries and the peptides, polypeptides or proteins identified by such screens.

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**COMPLETE SPECIFICATION  
FOR A STANDARD PATENT**

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Invention Title: Method of Constructing Display Libraries of  
Genetic Packages for Members of a Diverse  
Family of Peptides

The following statement is a full description of the invention, including the best method of performing it, known to us:

The present invention relates to constructing libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family. In a preferred embodiment, the displayed polypeptides are human Fabs.

5 More specifically, the invention is directed to the methods of cleaving single-stranded nucleic acids at chosen locations, the cleaved nucleic acids encoding, at least in part, the peptides, polypeptides or proteins displayed on the genetic packages of the libraries of the invention. In a preferred embodiment, the genetic packages are filamentous phage or phagemids.

10 The present invention further relates to methods of screening the libraries of genetic packages that display useful peptides, polypeptides and proteins and to the peptides, polypeptides and proteins identified by such screening.

#### BACKGROUND OF THE INVENTION

15 It is now common practice in the art to prepare libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family. In many common libraries, the displayed peptides, polypeptides or proteins are related to antibodies. Often, they are Fabs or single chain antibodies.

In general, the DNAs that encode members of the families to be displayed must be amplified before they are cloned and used to display the desired member on the surface of a genetic package. Such amplification typically makes use of forward and backward primers.

20 Such primers can be complementary to sequences native to the DNA to be amplified or complementary to oligonucleotides attached at the 5' or 3' ends of that DNA. Primers that are complementary to sequences native to the DNA to be amplified are disadvantaged in that they bias the members of the families to be displayed. Only those members that contain a sequence in the native DNA that is substantially complementary to the primer will be amplified. Those that do not will be  
25 absent from the family. For those members that are amplified, any diversity within the primer region will be suppressed.

30 For example, in European patent 368,684 B1, the primer that is used is at the 5' end of the V<sub>H</sub> region of an antibody gene. It anneals to a sequence region in the native DNA that is said to be "sufficiently well conserved" within a single species. Such primer will bias the members amplified to those having this "conserved" region. Any diversity within this region is extinguished.

It is generally accepted that human antibody genes arise through a process that involves a combinatorial selection of V and J or V, D, and J followed by somatic mutations. Although most diversity occurs in the Complementary Determining Regions (CDRs), diversity also occurs in the more conserved Framework Regions (FRs) and at least some of this diversity confers or enhances specific binding to antigens (Ag). As a consequence, libraries should contain as much of the CDR and FR diversity as possible.

To clone the amplified DNAs for display on a genetic package of the peptides, polypeptides or proteins that they encode, the DNAs must be cleaved to produce appropriate ends for ligation to a vector. Such cleavage is generally effected using restriction endonuclease recognition sites carried on the primers. When the primers are at the 5' end of DNA produced from reverse transcription of RNA, such restriction leaves deleterious 5' untranslated regions in the amplified DNA. These regions interfere with expression of the cloned genes and thus the display of the peptides, polypeptides and proteins coded for by them.

#### SUMMARY OF THE INVENTION

It is an object of this invention to provide novel methods for constructing libraries of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and collectively display at least a portion of the diversity of the family. These methods are not biased toward DNAs that contain native sequences that are complementary to the primers used for amplification. They also enable any sequences that may be deleterious to expression to be removed from the amplified DNA before cloning and displaying.

It is another object of this invention to provide a method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:

- (i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate

such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

It is a further object of this invention to provide an alternative method for cleaving single-stranded nucleic acid sequences at a desired location, the method comprising the steps of:

- 5 (i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and
- 10 (ii) cleaving the nucleic acid solely at the cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such

15 that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

It is another objective of the present invention to provide a method of capturing DNA molecules that comprise a member of a diverse family of DNAs and collectively comprise at least a portion of the diversity of the family. These DNA molecules in single-stranded form have been

20 cleaved by one of the methods of this invention. This method involves ligating the individual single-stranded DNA members of the family to a partially duplex DNA complex. The method comprises the steps of:

- 25 (i) contacting a single-stranded nucleic acid sequence that has been cleaved with a restriction endonuclease with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region that remains after cleavage, the double-stranded region of the oligonucleotide including any sequences necessary to return the sequences that remain after cleavage into proper reading frame for expression and containing a restriction endonuclease recognition site 5' of those sequences; and
- 30 (ii) cleaving the partially double-stranded oligonucleotide sequence solely at the restriction endonuclease recognition site contained within the double-stranded region of the partially double-stranded oligonucleotide.

It is another object of this invention to prepare libraries, that display a diverse family of peptides, polypeptides or proteins and collectively display at least part of the diversity of the family, using the methods and DNAs described above.

It is an object of this invention to screen those libraries to identify useful peptides, polypeptides and proteins and to use those substances in human therapy.

A definition of the specific embodiment of the invention as claimed herein follows.

In a broad embodiment of the invention, there is provided a library comprising a collection of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and that collectively display at least a portion of the family, the displayed peptides, polypeptides or proteins being encoded by DNA sequences comprising sequences encoding

- (a) a CDR1 having an amino acid sequence according to the formula  $-X_1-Y-X_2-M-X_3-$ , wherein  $X_1$ ,  $X_2$ , and  $X_3$  are independently selected from the group consisting of A, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, and Y, and
- (b) a CDR2 having an amino acid sequence according to the formula  $X_4-I-X_5-X_6-S-G-G-X_7-T-X_8-Y-A-D-S-V-K-G-$ , wherein  $X_4$  and  $X_5$  are independently selected from the group consisting of Y, R, W, V, G, and S,  $X_6$  is selected from the group consisting of P and S, and  $X_7$  and  $X_8$  are independently selected from the group consisting of A, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, and Y.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a schematic of various methods that may be employed to amplify VH genes without using primers specific for VH sequences.

FIG. 2 is a schematic of various methods that may be employed to amplify VL genes without using VL sequences.

FIG. 3 depicts gel analysis of cleaved kappa DNA from Example 2.

FIG. 4 depicts gel analysis of cleaved kappa DNA from Example 2.

FIG. 5 depicts gel analysis of amplified kappa DNA from Example 2.

FIG. 6 depicts gel purified amplified kappa DNA from Example 2.

TERMS

In this application, the following terms and abbreviations are used:

	Sense strand	The upper strand of ds DNA as usually written. In the sense strand, 5'-ATG-3' codes for Met.
5	Antisense strand	The lower strand of ds DNA as usually written. In the antisense strand, 3'-TAC-5' would correspond to a Met codon in the sense strand.
10	Forward primer:	A "forward" primer is complementary to a part of the sense strand and primes for synthesis of a new antisense-strand molecule. "Forward primer" and "lower-strand primer" are equivalent.
	Backward primer:	A "backward" primer is complementary to a part of the antisense strand and primes for synthesis of a new sense-strand molecule. "Backward primer" and "top-strand primer" are equivalent.
15	Bases	Bases are specified either by their position in a vector or gene as their position within a gene by codon and base. For example, "89.1" is the first base of codon 89, 89.2 is the second base of codon 89.
	Sv	Streptavidin
	Ap	Ampicillin
	$\varphi^R$	A gene conferring ampicillin resistance
20	RE	Restriction endonuclease
	URE	Universal restriction endonuclease
	Functionally Complementary	Two sequences are sufficiently complementary so as to anneal under the chosen conditions.
25	RERS	Restriction endonuclease recognition site
	AA	Amino acid
	PCR	Polymerization chain reaction
	GLGs	Germline genes



2007211861 21 Aug 2007

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Ab Antibody: an immunoglobulin. The term also covers any protein having a binding domain which is homologous to an immunoglobulin binding domain. A few examples of antibodies within this definition are, *inter alia*, immunoglobulin isotypes and the Fab, F(ab<sup>1</sup>)<sub>2</sub>, scFv, Fv, dAb and Fd fragments.

Fab Two chain molecule comprising an Ab light chain and part of a heavy-chain.

scFv A single-chain Ab comprising either VH::linker::VL or VL::linker::VH

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wt. Wild type

HC Heavy chain

LC Light chain

VK A variable domain of a Kappa light chain.

VH A variable domain of a heavy chain.

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VL A variable domain of a lambda light chain.

In this application, all references referred to are specifically incorporated by reference.

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The nucleic acid sequences that are useful in the methods of this invention, i.e., those that encode at least in part the individual peptides, polypeptides and proteins displayed on the genetic packages of this invention, may be naturally occurring, synthetic or a combination thereof. They may 20 be mRNA, DNA or cDNA. In the preferred embodiment, the nucleic acids encode antibodies. Most preferably, they encode Fabs.

The nucleic acids useful in this invention may be naturally diverse, synthetic diversity may be introduced into those naturally diverse members, or the diversity may be entirely synthetic. For 25 example, synthetic diversity can be introduced into one or more CDRs of antibody genes.

Synthetic diversity may be created, for example, through the use of TRIM technology (U.S. 5,869,644). TRIM technology allows control over exactly which amino-acid types are allowed at variegated positions and in what proportions. In TRIM technology, codons to be diversified are

synthesized using mixtures of trinucleotides. This allows any set of amino acid types to be included in any proportion.

Another alternative that may be used to generate diversified DNA is mixed oligonucleotide synthesis. With TRIM technology, one could allow Ala and Trp. With mixed oligonucleotide synthesis, a mixture that included Ala and Trp would also necessarily include Ser and Gly. The amino-acid types allowed at the variegated positions are picked with reference to the structure of antibodies, or other peptides, polypeptides or proteins of the family, the observed diversity in germline genes, the observed somatic mutations frequently observed, and the desired areas and types of variegation.

In a preferred embodiment of this invention, the nucleic acid sequences for at least one CDR or other region of the peptides, polypeptides or proteins of the family are cDNAs produced by reverse transcription from mRNA. More preferably, the mRNAs are obtained from peripheral blood cells, bone marrow cells, spleen cells or lymph node cells (such as B-lymphocytes or plasma cells) that express members of naturally diverse sets of related genes. More preferable, the mRNAs encode a diverse family of antibodies. Most preferably, the mRNAs are obtained from patients suffering from at least one autoimmune disorder or cancer. Preferably, mRNAs containing a high diversity of autoimmune diseases, such as systemic lupus erythematosus, systemic sclerosis, rheumatoid arthritis, antiphospholipid syndrome and vasculitis are used.

In a preferred embodiment of this invention, the cDNAs are produced from the mRNAs using reverse transcription. In this preferred embodiment, the mRNAs are separated from the cell and degraded using standard methods, such that only the full length (i.e., capped) mRNAs remain. The cap is then removed and reverse transcription used to produce the cDNAs.

The reverse transcription of the first (antisense) strand can be done in any manner with any suitable primer. See, *e.g.*, HJ de Haard et al., *Journal of Biological Chemistry*, 274 (26) :18218-30 (1999). In the preferred embodiment of this invention where the mRNAs encode antibodies, primers that are complementary to the constant regions of antibody genes may be used. Those primers are useful because they do not generate bias toward subclasses of antibodies. In another embodiment, poly-dT primers may be used (and may be preferred for the heavy-chain genes).

Alternatively, sequences complementary to the primer may be attached to the termini of the antisense strand.

In one preferred embodiment of this invention, the reverse transcriptase primer may be biotinylated, thus allowing the cDNA product to be immobilized *on* streptavidin (Sv) beads.

Immobilization can also be effected using a primer labeled at the 5' end with one of a) free amine group, b) thiol, c) carboxylic acid, or d) another group not found in DNA that can react to form a strong bond to a known partner on an insoluble medium. If, for example, a free amine (preferably primary amine) is provided at the 5' end of a DNA primer, this amine can be reacted with  
5 carboxylic acid groups on a polymer bead using standard amide-forming chemistry. If such preferred immobilization is used during reverse transcription, the top strand RNA is degraded using well-known enzymes, such as a combination of RNaseH and RNaseA, either before or after immobilization.

The nucleic acid sequences useful in the methods of this invention are generally amplified  
10 before being used to display the peptides, polypeptides or proteins that they encode. Prior to amplification, the single-stranded DNAs may be cleaved using either of the methods described before. Alternatively, the single-stranded DNAs may be amplified and then cleaved using one of those methods.

Any of the well known methods for amplifying nucleic acid sequences may be used for  
15 such amplification. Methods that maximize, and do not bias, diversity are preferred. In a preferred embodiment of this invention where the nucleic acid sequences are derived from antibody genes, the present invention preferably utilizes primers in the constant regions of the heavy and light chain genes and primers to a synthetic sequence that are attached at the 5' end of the sense strand. Priming at such synthetic sequence avoids the use of sequences within the variable regions of the  
20 antibody genes. Those variable region priming sites generate bias against V genes that are either of rare subclasses or that have been mutated at the priming sites. This bias is partly due to suppression of diversity within the primer region and partly due to lack of priming when many mutations are present in the region complementary to the primer. The methods disclosed in this invention have the advantage of not biasing the population of amplified antibody genes for particular V gene types.

25 The synthetic sequences may be attached to the 5' end of the DNA strand by various methods well known for ligating DNA sequences together. RT CapExtension is one preferred method.

In RT CapExtension (derived from Smart PCR<sup>TM</sup>) a short overlap (5' -...GGG-3' in the upper-strand primer (USP-GGG) complements 3'-CCC...5' in the lower strand) and reverse transcriptases are used so that the reverse complement of the upper-strand primer is attached to the lower strand.

30 In a preferred embodiment of this invention, the upper strand or lower strand primer may be also biotinylated or labeled at the 5' end with *one* of a) free amino group, b) thiol, c) carboxylic acid and d) another group not found in DNA that can react to form a strong bond to a known partner as an

insoluble medium. These can then be used to immobilize the labeled strand after amplification. The immobilized DNA can be either single or double-stranded.

FIG. 1 shows a schematic of the amplification of VH genes. FIG. 1, Panel A shows a primer specific to the poly-dT region of the 3' UTR priming synthesis of the first, lower strand. Primers that bind in the constant region are also suitable. Panel B shows the lower strand extended at its 3' end by three Cs that are not complementary to the mRNA. Panel C shows the result of annealing a synthetic top-strand primer ending in three GGGs that hybridize to the 3' terminal CCCs and extending the reverse transcription extending the lower strand by the reverse complement of the synthetic primer sequence. Panel D shows the result of PCR amplification using a 5' biotinylated synthetic top-strand primer that replicates the 5' end of the synthetic primer of panel C and a bottom-strand primer complementary to part of the constant domain. Panel E shows immobilized double-stranded (ds) cDNA obtained by using a 5'-biotinylated top-strand primer.

FIG. 2 shows a similar schematic for amplification of VL genes. FIG. 2, Panel A shows a primer specific to the constant region at or near the 3' end priming synthesis of the first, lower strand. Primers that bind in the poly-dT region are also suitable. Panel B shows the lower strand extended at its 3' end by three Cs that are not complementary to the mRNA. Panel C shows the result of annealing a synthetic top-strand primer ending in three GGGs that hybridize to the 3' terminal CCCs and extending the reverse transcription extending the lower strand by the reverse complement of the synthetic primer sequence. Panel D shows the result of PCR amplification using a 5' biotinylated synthetic top-strand primer that replicates the 5' end of the synthetic primer of panel C and a bottom-strand primer complementary to part of the constant domain. The bottom-strand primer also contains a useful restriction endonuclease site, such as *AscI*. Panel E shows immobilized ds cDNA obtained by using a 5'-biotinylated top-strand primer.

In FIGs. 1 and 2, each V gene consists of a 5' untranslated region (UTR) and a secretion signal, followed by the variable region, followed by a constant region, followed by a 3' untranslated region (which typically ends in poly-A). An initial primer for reverse transcription may be complementary to the constant region or to the poly A segment of the 3'-UTR. For human heavy-chain genes, a primer of 15 T is preferred. Reverse transcriptases attach several C residues to the 3' end of the newly synthesized DNA. RT CapExtension exploits this feature. The reverse transcription reaction is first run with only a lower-strand primer. After about 1 hour, a primer ending in GGG (USP-GGG) and more RTase are added. This causes the lower-strand cDNA to be extended by the reverse complement of the USP-GGG up to the final GGG. Using one primer

identical to part of the attached synthetic sequence and a second primer complementary to a region of known sequence at the 3' end of the sense strand, all the V genes are amplified irrespective of their V gene subclass.

After amplification, the DNAs of this invention are rendered single-stranded. For example, the strands can be separated by using a biotinylated primer, capturing the biotinylated product on streptavidin beads, denaturing the DNA, and washing away the complementary strand. Depending on which end of the captured DNA is wanted, one will choose to immobilize either the upper (sense) strand or the lower (antisense) strand.

To prepare the single-stranded amplified DNAs for cloning into genetic packages so as to effect display of the peptides, polypeptides or proteins encoded, at least in part, by those DNAs, they must be manipulated to provide ends suitable for cloning and expression. In particular, any 5' untranslated regions and mammalian signal sequences must be removed and replaced, in frame, by a suitable signal sequence that functions in the display host. Additionally, parts of the variable domains (in antibody genes) may be removed and replaced by synthetic segments containing synthetic diversity. The diversity of other gene families may likewise be expanded with synthetic diversity.

According to the methods of this invention, there are two ways to manipulate the single-stranded amplified DNAs for cloning. The first method comprises the steps of:

- (i) contacting the nucleic acid with a single-stranded oligonucleotide, the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired and including a sequence that with its complement in the nucleic acid forms a restriction endonuclease recognition site that on restriction results in cleavage of the nucleic acid at the desired location; and
- (ii) cleaving the nucleic acid solely at the recognition site formed by the complementation of the nucleic acid and the oligonucleotide;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature. In this first method, short oligonucleotides are annealed to the single-stranded DNA so that restriction endonuclease recognition sites formed within the now locally double-stranded regions of the DNA

can be cleaved. In particular, a recognition site that occurs at the same position in a substantial fraction of the single-stranded DNAs is identical.

For antibody genes, this can be done using a catalog of germline sequences. See, e.g., "http://www.mrc-cpe.cam.ac.uk/imt-doc/restricted/ok.htm 1." Updates can be obtained from this site under the heading "Amino acid and nucleotide sequence alignments." For other families, similar comparisons exist and may be used to select appropriate regions for cleavage and to maintain diversity.

For example, Table 195 depicts the DNA sequences of the FR3 regions of the 51 known human VH germline genes. In this region, the genes contain restriction endonuclease recognition sites shown in Table 200. Restriction endonucleases that cleave a large fraction of germline genes at the same site are preferred over endonucleases that cut at a variety of sites. Furthermore, it is preferred that there be only one site for the restriction endonucleases within the region to which the short oligonucleotide binds on the single-stranded DNA, e.g., about 10 bases *on* either side of the restriction endonuclease recognition site.

An enzyme that cleaves downstream in FR3 is also more preferable because it captures fewer mutations in the framework. This may be advantageous in some *cases*. However, it is well known that framework mutations exist and confer and enhance antibody binding. The present invention, by choice of appropriate restriction site, allows all or part of FR3 diversity to be captured. Hence, the method also allows extensive diversity to be captured.

Finally, in the methods of this invention restriction endonucleases that are active between about 45° and about 75°C are used. Preferably enzymes that are active above 50°C, and more preferably active about 55°C, are used. Such temperatures maintain the nucleic acid sequence to be cleaved in substantially single-stranded form.

Enzymes shown in Table 200 that cut many of the heavy chain FR3 germline genes at a single position include: *MaeIII*(24@4), *Tsp45I*(21@4), *HphI*(44@5), *BsaII*(23@65), *AluI*(23@47), *BlnI*(21@48), *DdeI*(29@58), *BglII*(10@61), *MsiI*(44@72), *BsiEI*(23@74), *EaeI*(23@74), *EagI*(23@74), *HaeIII*(25@75), *Bst4CI*(51@86), *HpyCH4III*(51@86), *HinfI*(38@2), *MlyI*(18@2), *PleI*(18@2), *MnlI*(31@67), *HpyCE4V*(21@44), *BsmAI*(16@11), *BpmI*(19@12), *XmnI*(12@30), and *SacI*(11@51). (The notation used means, for example, that *BsmAI* cuts 16 of the FR3 germline genes with a restriction endonuclease recognition site beginning at base 11 of FR3.)

For cleavage of human heavy chains in FR3, the preferred restriction endonucleases are: *Bst4CI* (or *Taal* or *HpyCH4III*), *BlnI*, *HpyCH4V*, and *MsiI*. Because ACNGT (the restriction

endonuclease recognition site for *Bst4CI*, *Taal*, and *HpyCH4III*) is found at a consistent site in all the human FR3 germline genes, one of those enzymes is the most preferred for capture of heavy chain CDR3 diversity. *BlpI* and *HpyCH4V* are complementary. *BlpI* cuts most members of the VH1 and VH4 families while *HpyCH4V* cuts most members of the VH3, VH5, VH6, and VH7 families. Neither enzyme cuts VH2s, but this is a very small family, containing only three members. Thus, these enzymes may also be used in preferred embodiments of the methods of this invention.

The restriction endonucleases *HpyCH4III*, *Bst4CI*, and *Taal* all recognize 5'-ACnGT-3' and cut upper strand DNA after n and lower strand DNA before the base complementary to n. This is the most preferred restriction endonuclease recognition site for this method on human heavy chains because it is found in all germline genes. Furthermore, the restriction endonuclease recognition region (ACnGT) matches the second and third bases of a tyrosine codon (tay.) and the following cysteine codon (tgy) as shown in Table 206. These codons are highly conserved, especially the cysteine in mature antibody genes.

Table 250 E shows the distinct oligonucleotides of length 22 (except the last one which is of length 20) bases. Table 255 C shows the analysis of 1617 actual heavy chain antibody genes. Of these, 1511 have the site and match one of the candidate oligonucleotides to within 4 mismatches. Eight oligonucleotides account for most of the matches and are given in Table 250 F.I. The 8 oligonucleotides are very similar so that it is likely that satisfactory cleavage will be achieved with only one oligonucleotide (such as H43.77.97.1-02#1) by adjusting temperature, pH, salinity, and the like. One or two oligonucleotides may likewise suffice whenever the germline gene sequences differ very little and especially if they differ very little close to the restriction endonuclease recognition region to be cleaved. Table 255 D shows a repeat analysis of 1617 actual heavy chain antibody genes using only the 8 chosen oligonucleotides. This shows that 1463 of the sequences match at least one of the oligonucleotides to within 4 mismatches and have the site as expected. Only 7 sequences have a second *FpyCH4III* restriction endonuclease recognition region in this region.

Another illustration of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of human heavy chains. Cleavage in FR1 allows capture of the entire CDR diversity of the heavy chain.

The germline genes for human heavy chain FR1 are shown in Table 217. Table 220 shows the restriction endonuclease recognition sites found in human germline genes FR1s. The preferred sites are *BsgI*(GTGCAG;39@4), *BsoFI*(GCngc;43@6, 11@9, 2@3, 1@12), *TseI*(Gcwg;43@6, 11@9, 2@3, 1@12), *MspAII*(CMGckg;46@7, 2@1), *PvuII*(CAGctg;46@7, 2@1), *AluI*(AGct;48@8, 2@2),

*DdeI*(Ctnag;22@52, 9@48), *HphI*(tcacc;22@80), *BssKI*(Nccngg;35@39, 2@40), *BsaII*(Ccnngg;32@40, 2@41), *BstNI*(CCwgg; 33@40), *ScrFI*(CCngg;35@40, 2@41), *EcoO109I*(RGgnccy;22@46, 11@43), *Sau96I*(Ggncc;23@47, 11@44), *AvaII*(Ggwcc;23@47, 4@44), *PpuMI*(RGgwccy;22@46, 4@43), *BsmFI*(gtccc;20@48), *HinII*(Gantc;34@16, 21@56, 21@77), *TfiI* (21@77), *MlyI* (GAGTC;34@16), *MlyI*(gactc;21@56), and *AlwNI*(CAGnnctg;22@68). The more preferred sites are *MspAI* and *PvuII*. *MspAI* and *PvuII* have 46 sites at 7-12 and 2 at 1-6. To avoid cleavage at both sites, oligonucleotides are used that do not fully cover the site at 1-6. Thus, the DNA will not be cleaved at that site. We have shown that DNA that extends 3, 4, or 5 bases beyond a *PvuII*-site can be cleaved efficiently.

Another illustration of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of human kappa light chains. Table 300 shows the human kappa FR1 germline genes and Table 302 shows restriction endonuclease recognition sites that are found in a substantial number of human kappa FR1 germline genes at consistent locations. Of the restriction endonuclease recognition sites listed, *BsmAI* and *PfI* are the most preferred enzymes. *BsmAI* sites are found at base 18 in 35 of 40 germline genes. *PfI* sites are found in 35 of 40 germline genes at base 12.

Another example of choosing an appropriate restriction endonuclease recognition site involves cleavage in FR1 of the human lambda light chain. Table 400 shows the 31 known human lambda FR1 germline gene sequences. Table 405 shows restriction endonuclease recognition sites found in human lambda FR1 germline genes. *HinII* and *DdeI* are the most preferred restriction endonucleases for cutting human lambda chains in FR1.

After the appropriate site or sites for cleavage are chosen, one or more short oligonucleotides are prepared so as to functionally complement, alone or in combination, the chosen recognition site. The oligonucleotides also include sequences that flank the recognition site in the majority of the amplified genes. This flanking region allows the sequence to anneal to the single-stranded DNA sufficiently to allow cleavage by the restriction endonuclease specific for the site chosen.

The actual length and sequence of the oligonucleotide depends on the recognition site and the conditions to be used for contacting and cleavage. The length must be sufficient so that the oligonucleotide is functionally complementary to the single-stranded DNA over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and solely at the desired location.



Typically, the oligonucleotides of this preferred method of the invention are about 17 to about 30 nucleotides in length. Below about 17 bases, annealing is too weak and above 30 bases there can be a loss of specificity. A preferred length is 18 to 24 bases.

Oligonucleotides of this length need not be identical complements of the germline genes. Rather, a few mismatches taken may be tolerated. Preferably, however, no more than 1-3 mismatches are allowed. Such mismatches do not adversely affect annealing of the oligonucleotide to the single-stranded DNA. Hence, the two DNAs are said to be functionally complementary.

The second method to manipulate the amplified single-stranded DNAs of this invention for cloning comprises the steps of:

- (i) contacting the nucleic acid with a partially double-stranded oligonucleotide, the single-stranded region of the oligonucleotide being functionally complementary to the nucleic acid in the region in which cleavage is desired, and the double-stranded region of the oligonucleotide having a Type II-S restriction endonuclease recognition site, whose cleavage site is located at a known distance from the recognition site; and
- (ii) cleaving the nucleic acid solely at the cleavage site formed by the complementation of the nucleic acid and the single-stranded region of the oligonucleotide;

the contacting and the cleaving steps being performed at a temperature sufficient to maintain the nucleic acid in substantially single-stranded form, the oligonucleotide being functionally complementary to the nucleic acid over a large enough region to allow the two strands to associate such that cleavage may occur at the chosen temperature and at the desired location, and the cleavage being carried out using a restriction endonuclease that is active at the chosen temperature.

This second method employs Universal Restriction Endonucleases ("URE"). UREs are partially double-stranded oligonucleotides. The single-stranded portion or overlap of the URE consists of a DNA adapter that is functionally complementary to the sequence to be cleaved in the single-stranded DNA. The double-stranded portion consists of a type II-S restriction endonuclease recognition site.

The URE method of this invention is specific and precise and can tolerate some (e.g., 1-3) mismatches in the complementary regions, i.e., it is functionally complementary to that region. Further, conditions under which the URE is used can be adjusted so that most of the genes that are amplified can be cut, reducing bias in the library produced from those genes.

The sequence of the single-stranded DNA adapter or overlap portion of the URE typically consists of about 14-22 bases. However, longer or shorter adapters may be used. The size depends

on the ability of the adapter to associate with its functional complement in the single-stranded DNA and the temperature used for contacting the URE and the single-stranded DNA at the temperature used for cleaving the DNA with the type II-S enzyme. The adapter must be functionally complementary to the single-stranded DNA over a large enough region to allow the two strands to associate such that the cleavage may occur at the chosen temperature and at the desired location. We prefer single-stranded or overlap portions of 14-17 bases in length, and more preferably 18-20 bases in length.

The site chosen for cleavage using the URE is preferably one that is substantially conserved in the family of amplified DNAs. As compared to the first cleavage method of this invention, these sites do not need to be endonuclease recognition sites. However, like the first method, the sites chosen can be synthetic rather than existing in the native DNA. Such sites may be chosen by references to the sequences of known antibodies or other families of genes. For example, the sequences of many germline genes are reported at <http://www.mrc-cpe.cam.ac.uk/imt-doc/restricted/ok.html>. For example, one preferred site occurs near the end of FR3 -- codon 89 through the second base of codon 93. CDR3 begins at codon 95.

The sequences of 79 human heavy-chain genes are also available at <http://www.ncbi.nlm.nih.gov/entre2/nucleotide.html>. This site can be used to identify appropriate sequences for URE cleavage according to the methods of this invention. See, e.g., Table 8B.

Most preferably, one or more sequences are identified using these sites or other available sequence information. These sequences together are present in a substantial fraction of the amplified DNAs. For example, multiple sequences could be used to allow for known diversity in germline genes or for frequent somatic mutations. Synthetic degenerate sequences could also be used. Preferably, a sequence(s) that occurs in at least 65% of genes examined with no more than 2-3 mismatches is chosen URE single-stranded adapters or overlaps are then made to be complementary to the chosen regions. Conditions for using the UREs are determined empirically. These conditions should allow cleavage of DNA that contains the functionally complementary sequences with no more than 2 or 3 mismatches but that do not allow cleavage of DNA lacking such sequences.

As described above, the double-stranded portion of the URE includes a Type II-S endonuclease recognition site. Any Type II-S enzyme that is active at a temperature necessary to maintain the single-stranded DNA substantially in that form and to allow the single-stranded DNA adapter portion of the URE to anneal long enough to the single-stranded DNA to permit cleavage at the desired site may be used.

The preferred Type II-S enzymes for use in the URE methods of this invention provide asymmetrical cleavage of the single-stranded DNA. Among these are the enzymes listed in Table 800. The most preferred Type II-S enzyme is FokI.

When the preferred FokI containing URE is used, several conditions are preferably used to effect cleavage:

- (i) Excess of the URE over target DNA should be present to activate the enzyme. URE present only in equimolar amounts to the target DNA would yield poor cleavage of ssDNA because the amount of active enzyme available would be limiting.
- (ii) An activator may be used to activate part of the FokI enzyme to dimerize without causing cleavage. Examples of appropriate activators are shown in Table 510.
- (iii) The cleavage reaction is performed at a temperature between 45°-75°C, preferably above 50°C and most preferably above 55°C.

The UREs used in the prior art contained a 14-base single-stranded segment, a 10-base stem (containing a *FokI* site), followed by the palindrome of the 10-base stem. While such UREs may be used in the methods of this invention, the preferred UREs of this invention also include a segment of three to eight bases (a loop) between the *FokI* restriction endonuclease recognition site containing segments. In the preferred embodiment, the stem (containing the *FokI* site) and its palindrome are also longer than 10 bases. Preferably, they are 10-14 bases in length. Examples of these "lollipop" URE adapters are shown in Table 5.

One example of using a URE to cleave an single-stranded DNA involves the FR3 region of human heavy chain. Table 508 shows an analysis of 840 full-length mature human heavy chains with the URE recognition sequences shown. The vast majority (718/840=0.85) will be recognized with 2 or fewer mismatches using five UREs (VHS881-1.1, VHS881-1.2, VHS881-2.1, VHS881-4.1, and VHS881-9.1). Each has a 20-base adaptor sequence to complement the germline gene, a ten-base stem segment containing a *FokI* site, a five base loop, and the reverse complement of the first stem segment. Annealing those adapters, alone or in combination, to single-stranded antisense heavy chain DNA and treating with *FokI* in the presence of, *e.g.*, the activator FOKIact, will lead to cleavage of the antisense strand at the position indicated.

Another example of using a URE(s) to cleave a single-stranded DNA involves the FR1 region of the human Kappa light chains. Table 512 shows an analysis of 182 full-length human kappa chains for matching by the four 19-base probe sequences shown. Ninety-six percent of the sequences match one of the probes with 2 or fewer mismatches. The URE adapters shown in Table

512 are for cleavage of the sense strand of kappa chains. Thus, the adaptor sequences are the reverse complement of the germline gene sequences. The URE consists of a ten-base stem, a five base loop, the reverse complement of the stem and the complementation sequence. The loop shown here is TTGTT, but other sequences could be used. Its function is to interrupt the palindrome of the stems so that formation of a lollypop monomer is favored over dimerization. Table 512 also shows where the sense strand is cleaved.

Another example of using a URE to cleave a single-stranded DNA involves the human lambda light chain. Table 515 shows analysis of 128 human lambda light chains for matching the four 19-base probes shown. With three or fewer mismatches, 88 of 128 (69%) of the chains match one of the probes. Table 515 also shows URE adapters corresponding to these probes.

Annealing these adapters to upper-strand ssDNA of lambda chains and treatment with *FokI* in the presence of FOKIact at a temperature at or above 45°C will lead to specific and precise cleavage of the chains. The conditions under which the short oligonucleotide sequences of the first method and the UREs of the second method are contacted with the single-stranded DNAs may be empirically determined. The conditions must be such that the single-stranded DNA remains in substantially single-stranded form.

More particularly, the conditions must be such that the single-stranded DNA does not form loops that may interfere with its association with the oligonucleotide sequence or the URE or that may themselves provide sites for cleavage by the chosen restriction endonuclease.

The effectiveness and specificity of short oligonucleotides (first method) and UREs (second method) can be adjusted by controlling the concentrations of the URE adapters/oligonucleotides and substrate DNA, the temperature, the pH, the concentration of metal ions, the ionic strength, the concentration of chaotropes (such as urea and formamide), the concentration of the restriction endonuclease (e.g., *FokI*), and the time of the digestion. These conditions can be optimized with synthetic oligonucleotides having:

- 1) target germline gene sequences,
- 2) mutated target gene sequences, or
- 3) somewhat related non-target sequences.

The goal is to cleave most of the target sequences and minimal amounts of non-targets.

In the preferred embodiment of this invention, the single-stranded DNA is maintained in substantially that form using a temperature between 45°C to 75°C. More preferably, a temperature between 50°C and 60°C, most preferably between 55°C and 60°C, is used. These temperatures are

employed both when contacting the DNA with the oligonucleotide or URE and when cleaving the DNA using the methods of this invention.

The two cleavage methods of this invention have several advantages. The first method allows the individual members of the family of single-stranded DNAs to be cleaved solely at one substantially conserved endonuclease recognition site. The method also does not require an endonuclease recognition site to be built in to the reverse transcription or amplification primers. Any native or synthetic site in the family can be used.

The second method has both of these advantages. In addition, the URE method allows the single-stranded DNAs to be cleaved at positions where no endonuclease recognition site naturally occurs or has been synthetically constructed.

Most importantly, both cleavage methods permit the use of 5' and 3' primers so as to maximize diversity and then cleavage to remove unwanted or deleterious sequences before cloning and display.

After cleavage of the amplified DNAs using one of the methods of this invention, the DNA is prepared for cloning. This is done by using a partially duplexed synthetic DNA adapter, whose terminal sequence is based on the specific cleavage site at which the amplified DNA has been cleaved.

The synthetic DNA is designed such that when it is ligated to the cleaved single-stranded DNA, it allows that DNA to be expressed in the correct reading frame so as to display the desired peptide, polypeptide or protein on the surface of the genetic package.

Preferably, the double-stranded portion of the adapter comprises the sequence of several codons that encode the amino acid sequence characteristic of the family of peptides, polypeptides or proteins up to the cleavage site. For human heavy chains, the amino acids of the 3-23 framework are preferably used to provide the sequences required for expression of the cleaved DNA. Preferably, the double-stranded portion of the adapter is about 12 to 100 bases in length. More preferably, about 20 to 100 bases are used. The double-stranded region of the adapter also preferably contains at least one endonuclease recognition site useful for cloning the DNA into a suitable display vector (or a recipient vector used to archive the diversity). This endonuclease restriction site may be native to the germline gene sequences used to extend the DNA sequence. It may be also constructed using degenerate sequences to the native germline gene sequences. Or, it may be wholly synthetic.

The single-stranded portion of the adapter is complementary to the region of the cleavage in the single-stranded DNA. The overlap can be from about 2 bases up to about 15 bases. The longer

the overlap, the more efficient the ligation is likely to be. A preferred length for the overlap is 7 to 10. This allows some mismatches in the region so that diversity in this region may be captured.

The single-stranded region or overlap of the partially duplexed adapter is advantageous because it allows DNA cleaved at the chosen site, but not other fragments to be captured. Such fragments would contaminate the library with genes encoding sequences that will not fold into proper antibodies and are likely to be non-specifically sticky.

One illustration of the use of a partially duplexed adaptor in the methods of this invention involves ligating such adaptor to a human FR3 region that has been cleaved, as described above, at 5'-ACnGT- 3' using *HpyCH4III*, *Bst4CI* or *Taal*.

Table 250 F.2 shows the bottom strand of the double-stranded portion of the adaptor for ligation to the cleaved bottom-strand DNA. Since the *HpyCH4III*-site is so far to the right (as shown in Table 206), a sequence that includes the *AflII*-site as well as the *XbaI* site can be added. This bottom strand portion of the partially-duplexed adaptor, H43.XAExt, incorporates both *XbaI* and *AflII*-sites. The top strand of the double-stranded portion of the adaptor has neither site (due to planned mismatches in the segments opposite the *XbaI* and *AflII*-sites of H43.XAExt), but will anneal very tightly to H43.XAExt. H43AExt contains only the *AflII*-site and is to be used with the top strands H43.ABr1 and H43.ABr2 (which have intentional alterations to destroy the *AflII*-site).

After ligation, the desired, captured DNA can be PCR amplified again, if desired, using in the preferred embodiment a primer to the downstream constant region of the antibody gene and a primer to part of the double-strand region of the adapter. The primers may also carry restriction endonuclease sites for use in cloning the amplified DNA.

After ligation, and perhaps amplification, of the partially double-stranded adapter to the single-stranded amplified DNA, the composite DNA is cleaved at chosen 5' and 3' endonuclease recognition sites.

The cleavage sites useful for cloning depend on the phage or phagemid into which the cassette will be inserted and the available sites in the antibody genes. Table 1 provides restriction endonuclease data for 75 human light chains. Table 2 shows corresponding data for 79 human heavy chains. In each Table, the endonucleases are ordered by increasing frequency of cutting. In these Tables, Nch is the number of chains cut by the enzyme and Ns is the number of sites (some chains have more than one site).

From this analysis, *SfiI*, *NotI*, *AflIII*, *ApaLI*, and *AscI* are very suitable. *SfiI* and *NotI* are preferably used in pCESI to insert the heavy-chain display segment. *ApaLI* and *AscI* are preferably used in pCESI to insert the light-chain display segment.

*BstEII*-sites occur in 97% of germ-line JH genes. In rearranged V genes, only 54/79 (68%) of heavy-chain genes contain a *J3stEII*-Site and 7/61 of these contain two sites. Thus, 47/79 (59%) contain a single *BstEII*-Site. An alternative to using *BstEII* is to cleave *via* UREs at the end of JH and ligate to a synthetic oligonucleotide that encodes part of CHI.

One example of preparing a family of DNA sequences using the methods of this invention involves capturing human CDR 3 diversity. As described above, mRNAs from various autoimmune patients is reverse transcribed into lower strand cDNA. After the top strand RNA is degraded, the lower strand is immobilized and a short oligonucleotide used to cleave the cDNA upstream of CDR3. A partially duplexed synthetic DNA adapter is then annealed to the DNA and the DNA is amplified using a primer to the adapter and a primer to the constant region (after FR4). The DNA is then cleaved using *BstEII* (in FR4) and a restriction endonuclease appropriate to the partially double-stranded adapter (e.g., *XbaI* and *AflIII* (in FR3)). The DNA is then ligated into a synthetic VH skeleton such as 3-23.

One example of preparing a single-stranded DNA that was cleaved using the URE method involves the human Kappa chain. The cleavage site in the sense strand of this chain is depicted in Table 512. The oligonucleotide kapextURE is annealed to the oligonucleotides (kaBROIUR, kaBR02UR, kaBR03UR, and kaBR04UR) to form a partially duplex DNA. This DNA is then ligated to the cleaved soluble kappa chains. The ligation product is then amplified using primers kapextUREPCR and CKForeAsc (which inserts a *AscI* site after the end of C kappa). This product is then cleaved with *ApaLI* and *AscI* and ligated to similarly cut recipient vector.

Another example involves the cleavage illustrated in Table 515. After cleavage, an extender (ON\_LamExi33) and four bridge oligonucleotides (oN\_LamBi-133, ON\_LamB2-133, ON\_LamB3-133, and ON\_LamB4-133) are annealed to form a partially duplex DNA. That DNA is ligated to the cleaved lambda-chain sense strands. After ligation, the DNA is amplified with ON\_Lami33PCR and a forward primer specific to the lambda constant domain, such as CL2ForeAsc or CL7ForeAsc (Table 130). In human heavy chains, one can cleave almost all genes in FR4 (downstream, i.e. toward the 3' end of the sense strand, of CDR3) at a *BstEII*-Site that occurs at a constant position in a very large fraction of human heavy-chain V genes. One then needs a site in FR3, if only CDR3 diversity is to

2007211861 21 Aug 2007  
be captured, in FR2, if CDR2 and CDR3 diversity is wanted, or in FR1, if all the CDR diversity is wanted. These sites are preferably inserted as part of the partially double-stranded adaptor.

5 The preferred process of this invention is to provide recipient vectors having sites that allow cloning of either light or heavy chains. Such vectors are well known and widely used in the art. A preferred phage display vector in accordance with this invention is phage MALIA3. This displays in gene III. The sequence of the phage MALIA3 is shown in Table 120A (annotated) and Table 120B (condensed).

10 The DNA encoding the selected regions of the light or heavy chains can be transferred to the vectors using endonucleases that cut either light or heavy chains only very rarely. For example, light chains may be captured with ApaI and AscI. Heavy-chain genes are preferably cloned into a recipient vector having SfiI, NcoI, XbaI, AflII, BstEII, ApaI, and NotI sites. The light chains are preferably moved into the library as ApaI-AscI fragments. The heavy chains are preferably moved into the library as SfiI-NotI fragments.

15 Most preferably, the display is had on the surface of a derivative of M13 phage. The most preferred vector contains all the genes of M13, an antibiotic resistance gene, and the display cassette. The preferred vector is provided with restriction sites that allow introduction and excision of members of the diverse family of genes, as cassettes. The preferred vector is stable against rearrangement under the growth conditions used to amplify phage.

20 In another embodiment of this invention, the diversity captured by the methods of the present invention may be displayed in a phagemid vector (e.g., pCESI) that displays the peptide, polypeptide or protein on the III protein. Such vectors may also be used to store the diversity for subsequent display using other vectors or phage.

25 In another embodiment, the mode of display may be through a short linker to three possible anchor domains. One anchor domain being the final portion of M13 III ("IIIstump"), a second anchor being the full length III mature protein, and the third being the M13 VIII mature protein.

30 The IIIstump fragment contains enough of M13 III to assemble into phage but not the domains involved in mediating infectivity. Because the w.t. III and VIII proteins are present, the phage is unlikely to delete the antibody genes and phage that do delete these segments receive only a very small growth advantage. For each of the anchor domains, the DNA encodes the w.t. AA sequence, but differs from the w.t. DNA sequence to a very high extent. This will greatly reduce the potential for homologous recombination between the display anchor and the w.t. gene that is also present.



Most preferably, the present invention uses a complete phage carrying an antibiotic-resistance gene (such as an ampicillin-resistance gene) and the display cassette. Because the w.t. iii and viii genes are present, the w.t. proteins are also present. The display cassette is transcribed from a regulatable promoter (e.g., PLac'z) • Use of a regulatable promoter allows control of the ratio of the fusion display gene to the corresponding w.t. coat protein. This ratio determines the average number of copies of the display fusion per phage (or phagemid) particle.

Another aspect of the invention is a method of displaying peptides, polypeptides or proteins (and particularly Fabs) on filamentous phage. In the most preferred embodiment this method displays FABs and comprises:

- a) obtaining a cassette capturing a diversity of segments of DNA encoding the elements:

Preg::RBS1::SSI::VL::CL::stop::RBS2::SS2::VH::CHI::linker::anchor::stop::,  
 where Preg is a regulatable promoter, RBS1 is a first ribosome binding site, SSI is a signal sequence operable in the host strain, VL is a member of a diverse set of light-chain variable regions, CL is a light-chain constant region, stop is one or more stop codons, RBS2 is a second ribosome binding site, SS2 is a second signal\_sequence operable in the host strain, VH is a member of a diverse set of heavy-chain variable regions, CHI is an antibody heavy-chain first constant domain, linker is a sequence of amino acids of one to about 50 residues, anchor is a protein that will assemble into the filamentous phage particle and stop is a second example of one or more stop codons; and

- b) positioning that cassette within the phagegenome to maximize the viability of the phage and to minimize the potential for deletion of the cassette or parts thereof.

The DNA encoding the anchor protein in the above preferred cassette should be designed to encode the same (or a closely related) amino acid sequence as is found in one of the coat proteins of the phage, but with a distinct DNA sequence. This is to prevent unwanted homologous recombination with the w.t. gene. In addition, the cassette should be placed in the intergenic region. The positioning and orientation of the display cassette can influence the behavior of the phage.

In one embodiment of the invention, a transcription terminator may be placed after the second stop of the display cassette above (e.g., Trp). This, will reduce interaction between the display cassette and other genes in the phage antibody display vector (PADV).

In another embodiment of the methods of this, invention, the phage or phagemid can display proteins other than Fab, by replacing the Fab portions indicated above, with other protein genes.

Various hosts can be used for growth of the display phage or phagemids of this invention. Such hosts are well known in the art. In the preferred embodiment, where Fabs are being displayed, the preferred host should grow at 30°C and be RecA- (to reduce unwanted genetic recombination) and EndA- (to make recovery of RF DNA easier). It is also preferred that the host strain be easily transformed by electroporation.

XLI-Blue MRF' satisfies most of these preferences, but does not grow well at 30°C. XLI-Blue MRF' does grow slowly at 38°C and thus is an acceptable host. TG-1 is also an acceptable host although it is RecA+ and EndA+. XLI-Blue MRF' is more preferred for the intermediate host used to accumulate diversity prior to final construction of the library.

After display, the libraries of this invention may be screened using well known and conventionally used techniques. The selected peptides, polypeptides or proteins may then be used to treat disease. Generally, the peptides, polypeptides or proteins for use in therapy or in pharmaceutical compositions are produced by isolating the DNA encoding the desired peptide, polypeptide or protein from the member of the library selected. That DNA is then used in conventional methods to produce the peptide, polypeptides or protein it encodes in appropriate host cells, preferably mammalian host cells, e.g., CHO cells. After isolation, the peptide, polypeptide or protein is used alone or with pharmaceutically acceptable compositions in therapy to treat disease.

### EXAMPLES

#### **Example 1: Capturing kappa chains with BsmAI:**

A repertoire of human-kappa chain mRNAs was prepared by treating total or poly(A+) RNA isolated from a collection of patients having various autoimmune diseases with calf intestinal phosphatase to remove the 5'-phosphate from all molecules that have them, such as ribosomal RNA, fragmented mRNA, tRNA and genomic DNA. Full length mRNA (containing a protective 7-methyl cap structure) is unaffected. The RNA is then treated with tobacco acid pyrophosphatase to remove the cap structure from full length mRNAs leaving a 5'-monophosphate group.

Full length mRNA's were modified with an adaptor at the 5' end and then reversed transcribed and amplified using the GeneRACE™ method and kit (Invitrogen). A 5' biotinylated

primer complementary to the adaptor and a 3' primer complementary to a portion of the construct region were used.

Approximately 2 micrograms (ug) of human kappa-chain (Ilgkappa) gene RACE material with biotin attached to 5'-end of upper strand was immobilized on 200 microliters (uL) of Seradyn magnetic beads. The lower strand was removed by washing the DNA with 2 aliquots 200 uL of 0.1 M NaOH (pH 13) for 3 minutes for the first aliquot followed by 30 seconds for the second aliquot. The beads were neutralized with 200 uL of 10 mM Tris (pH 7.5) 100 mM NaCl. The short oligonucleotides shown in Table 525 were added in 40 fold molar excess in 100 uL of NEB buffer 2 (50 mM NaCl, 10 mM Tris-HCl, 10 mM MgCl<sub>2</sub>, 1 mM dithiothreitol pH 7.9) to the dry beads. The mixture was incubated at 95°C for 5 minutes then cooled down to 55°C over 30 minutes. Excess oligonucleotide was washed away with 2 washes of NEB buffer 3 (100 mM NaCl, 50 mM Tris-HCl, 10 mM MgCl<sub>2</sub>, 1 mM dithiothreitol pH 7.9). Ten units of BsmAI (NEB) were added in NEB buffer 3 and incubated for 1 h at 55°C. The cleaved downstream DNA was collected and purified over a Qiagen PCR purification column (FIGs. 3 and 4).

A partially double-stranded adaptor was prepared using the oligonucleotide shown in Table 525. The adaptor was added to the single-stranded DNA in 100 fold molar excess along with 1000 units of T4 DNA ligase (NEB) and incubated overnight at 16°C. The excess oligonucleotide was removed with a Qiagen PCR purification column. The ligated material was amplified by PCR using the primers kapPCRtl and kapfor shown in Table 525 for 10 cycles with the program shown in Table 530.

The soluble PCR product was run on a gel and showed a band of approximately 700 n, as expected (FIGs. 5 and 6). The DNA was cleaved with enzymes *Apa*LI and *As*cl, gel purified, and ligated to similarly cleaved vector pCESI. The presence of the correct size insert was checked by PCR in several clones as shown in FIG. 15.

Table 500 shows the DNA sequence of a kappa light chain captured by this procedure. Table 501 shows a second sequence captured by this procedure. The closest bridge sequence was complementary to the sequence 5'-agccacc-3', but the sequence captured reads 5'-Tgccacc-3', showing that some mismatch in the overlapped region is tolerated.

### Example 2: Construction of Synthetic CDR1 and CDR2 Diversity in V-3-23 VH

#### Framework

A synthetic Complementary Determinant Region (CDR) 1 and 2 diversity was constructed in the 3-23 VH framework in a two step process: first, a vector containing the 3-23 VH framework was constructed, and then, a synthetic CDR 1 and 2 was assembled and cloned into this vector.

For construction of the V3-23 framework, 8 oligos and two PCR primers (long oligonucleotides: TOPFR1A, BOTFR1B, BOTFR2, BOTFR3, F06, BOTFR4, ON-vgCl, and ON-vgC2 and primers: SFPRMET and BOTPCRPRIM, shown in Table 600) that overlap were designed based on the Genbank sequence of V323 VH. The design incorporated at least one useful restriction site in each framework region, as shown in Table 600. In Table 600, the segments that were synthesized are shown as bold, the overlapping regions are underscored, and the PCR priming regions at each end are underscored. A mixture of these 8 oligos was combined at a final concentration of 2.5uM in a 20ul Polymerase Chain Reaction (PCR) reaction. The PCR mixture contained 200uM dNTPs, 2.5mM MgCl<sub>2</sub>, 0.02U *Pfu Turbo*<sup>TM</sup> DNA Polymerase, 1U Qiagen HotStart Taq DNA Polymerase, and IX Qiagen PCR buffer. The PCR program consisted of 10 cycles of 94°C for 30s, 55°C for 30s, and 72°C for 30s. The assembled V3-23 DNA sequence was then amplified, using 2.5ul of a 10-fold dilution from the initial PCR in 100ul PCR reaction. The PCR reaction contained 200uM dNTPs, 2.5mM MgCl<sub>2</sub>, 0.02U *Pfu Turbo*<sup>TM</sup> DNA Polymerase, 1U Qiagen HotStart Taq DNA Polymerase, IX Qiagen PCR Buffer and 2 outside primers (SFPRMET and BOTPCRPRIM) at a concentration of 1uM. The PCR program consisted of 23 cycles at 94°C for 30s, 55°C for 30s, and 72°C for 60s. The V3-23 VH DNA sequence was digested and cloned into pCES1 (phagemid vector) using the *Sfi*I and *Bst*EII restriction endonuclease sites (All restriction enzymes mentioned herein were supplied by New England BioLabs, Beverly, MA and used as per manufacturer's instructions).

Stuffer sequences (shown in Table 610 and Table 620) were introduced into pCES1 to replace CDR1/CDR2 sequences (900 bases between *Bsp*EI and *Xba*I RE sites) and CDR3 sequences (358 bases between *Afl*III and *Bst*EII), prior to cloning the CDR1/CDR2 diversity. The new vector is pCES5 and its sequence is given in Table 620. Having stuffers in place of the CDRs avoids the risk that a parental sequence would be over-represented in the library. The CDR1-2 stuffer contains restriction sites for *Bgl*II, *Bsu*36I, *Bcl*I, *Xcm*I, *Mlu*I, *Pvu*II, *Hpa*I, and *Hinc*II, the underscored sites being unique within the vector pCES5. The stuffer that replaces CDR3 contains the unique restriction endonuclease site *Rsr*II. The stuffer sequences are fragments from the penicillase gene of *E. coli*.

For the construction of the CDR1 and CDR2 diversity, 4 overlapping oligonucleotides (ON-vgCl, 0N\_Brl2, ON\_CD2Xba, and ON-vgC2, shown in Table 600 and Table 630) encoding CDR1/2, plus flanking regions, were designed. A mix of these 4 oligos was combined at a final concentration of 2.5uM in a 40ul PCR reaction. Two of the 4 oligos contained variegated sequences positioned at the CDR1 and the CDR2. The PCR mixture contained 200uM dNTPs, 2.5U Pwo DNA Polymerase (Roche), and IX Pwo PCR buffer with 2mM MgSO<sub>4</sub>. The PCR program consisted of 10 cycles at 94°C for 30s, 60°C for 30s, and 72°C for 60s. This assembled CDR1/2 DNA sequence was amplified, using 2.5ul of the mixture in 100ul PCR reaction. The PCR reaction contained 200uM dNTPs, 2.5U Pwo DNA Polymerase, IX Pwo PCR Buffer with 2mM MgSO<sub>4</sub> and 2 outside primers at a concentration of 1uM. The PCR program consisted of 10 cycles at 94°C for 30s, 60°C for 30s, and 72°C for 60s. These variegated sequences were digested and cloned into the V3-23 framework in place of the CDR1/2 stuffer.

We obtained approximately  $7 \times 10^7$  independent transformants. Into this diversity, we can clone CDR3 diversity either from donor populations or from synthetic DNA.

It will be understood that the foregoing is only illustrative of the principles of this invention and that various modifications can be made by those skilled in the art without departing from the scope of and spirit of the invention.

The term "comprise" and variants of the term such as "comprises" or "comprising" are used herein to denote the inclusion of a stated integer or stated integers but not to exclude any other integer or any other integers, unless in the context or usage an exclusive interpretation of the term is required.

Any reference to publications cited in this specification is not an admission that the disclosures constitute common general knowledge in Australia.

The claims defining the invention are as follows:

1. A library comprising a collection of genetic packages that display a member of a diverse family of peptides, polypeptides or proteins and that collectively display at least a portion of the family, the displayed peptides, polypeptides or proteins being encoded by DNA sequences  
5 comprising sequences encoding

(a) a CDR1 having an amino acid sequence according to the formula  $-X_1-Y-X_2-M-X_3-$ , wherein  $X_1$ ,  $X_2$ , and  $X_3$  are independently selected from the group consisting of A, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, and Y, and

(b) a CDR2 having an amino acid sequence according to the formula  $X_4-I-X_5-X_6-S-G-G-X_7-T-X_8-Y-A-D-S-V-K-G-$ , wherein  $X_4$  and  $X_5$  are independently selected from the group consisting of Y, R, W, V, G, and S,  $X_6$  is selected from the group consisting of P and S, and  $X_7$  and  $X_8$  are independently selected from the group consisting of A, D, E, F, G, H, I, K, L, M, N, P, Q, R, S, T, V, W, and Y.

2. The library according to claim 1, wherein said DNA sequences further comprise  
15 sequences encoding the framework regions of VH 3-23.

3. The library according to claim 1 or claim 2, wherein said genetic packages are M13 phage.

4. The library according to claim 2, wherein said DNA sequences are in a phage vector.

5. The library according to claim 2, wherein said DNA sequences are in a phagemid  
20 vector.

6. The library according to any one of claims 1 to 5, wherein said displayed peptides, polypeptides, or proteins are displayed through a short linker to the final portion of M13 gene III.

7. A library as defined in claim 1 and substantially as hereinbefore described with  
25 reference to the examples.

Date: 22 August 2007

AMPLIFY VH GENES WITHOUT USING VH SEQUENCES

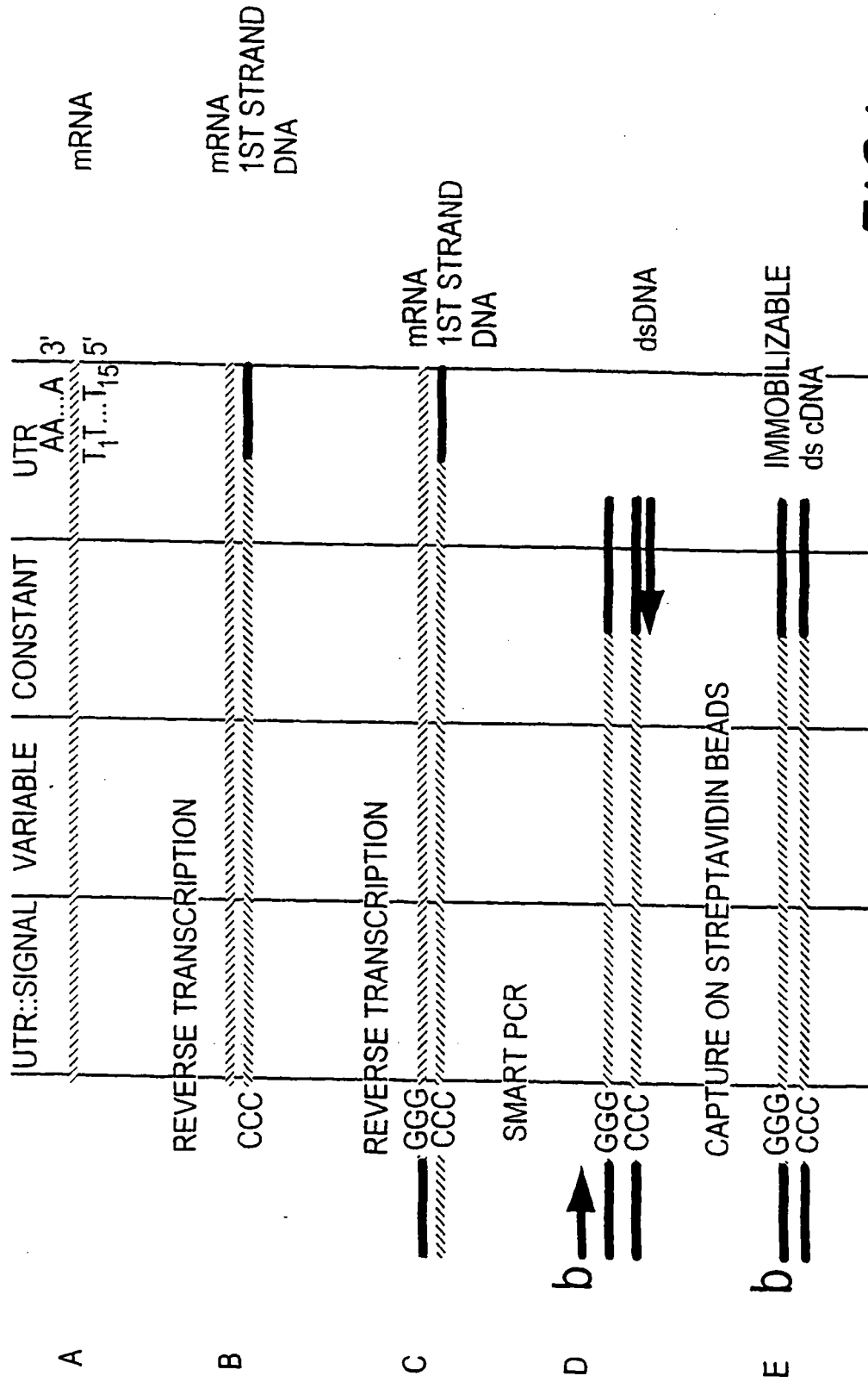


FIG. 1

AMPLIFY VL GENES WITHOUT USING VL SEQUENCES

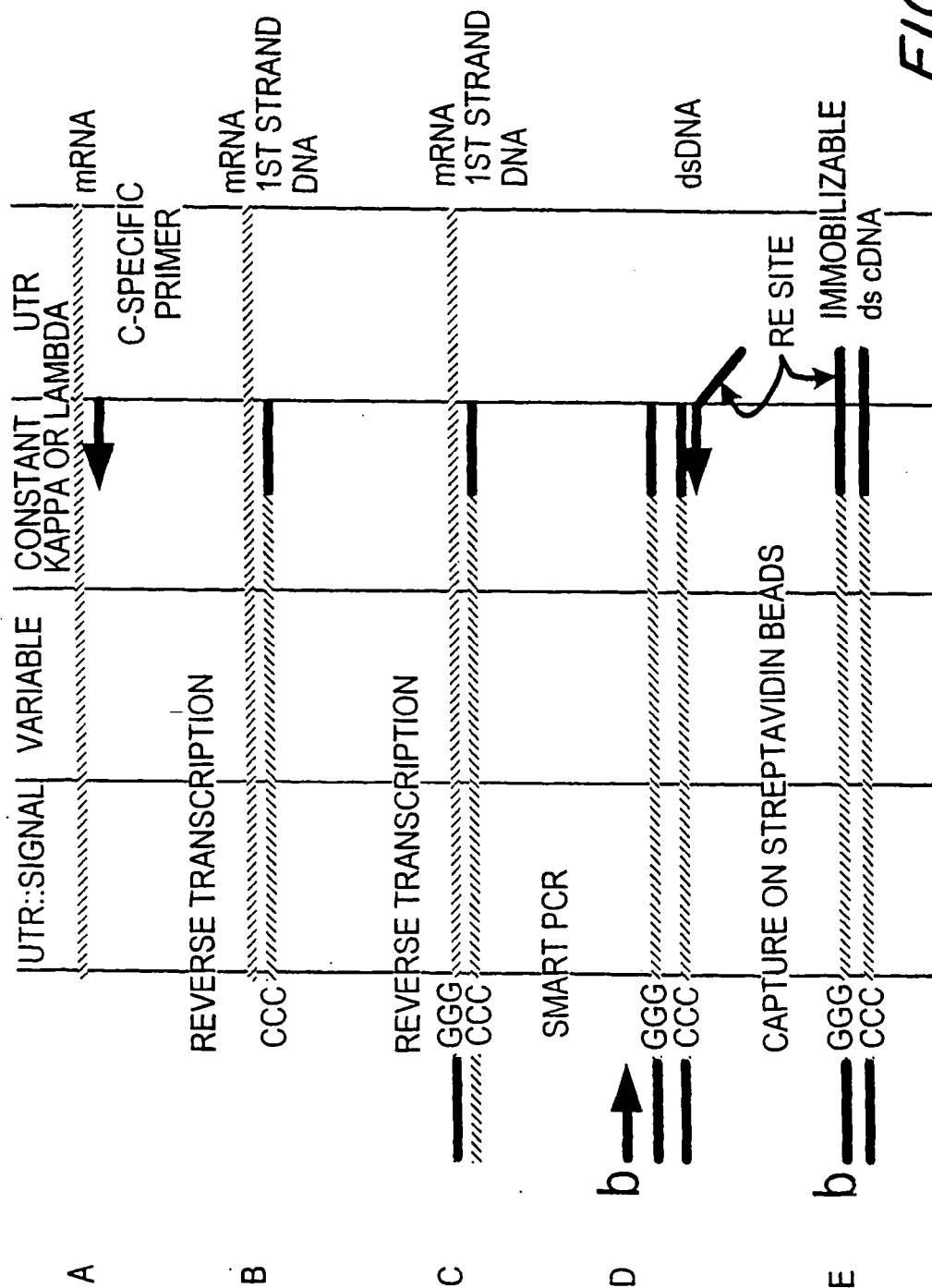


FIG. 2



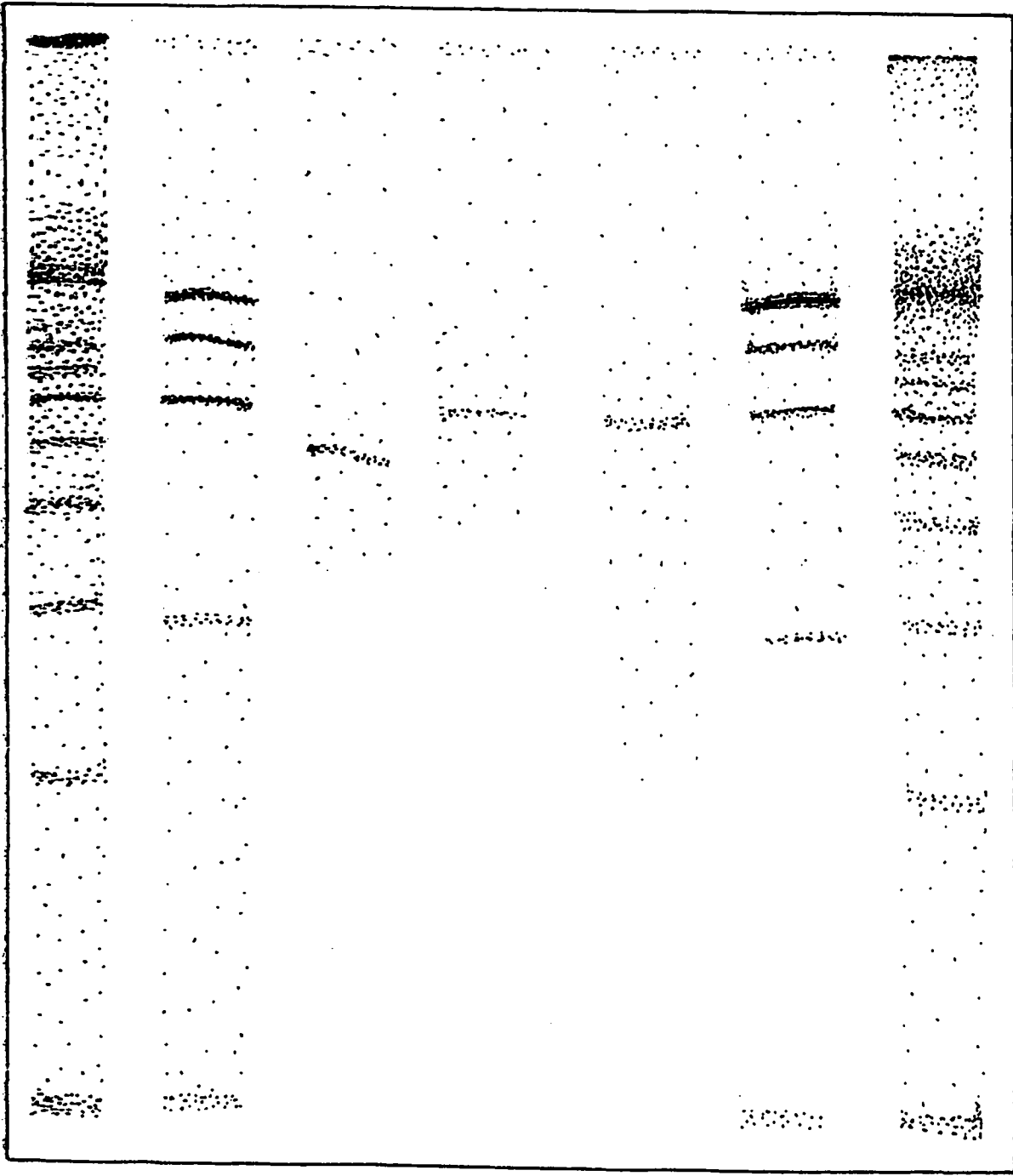


FIG. 3

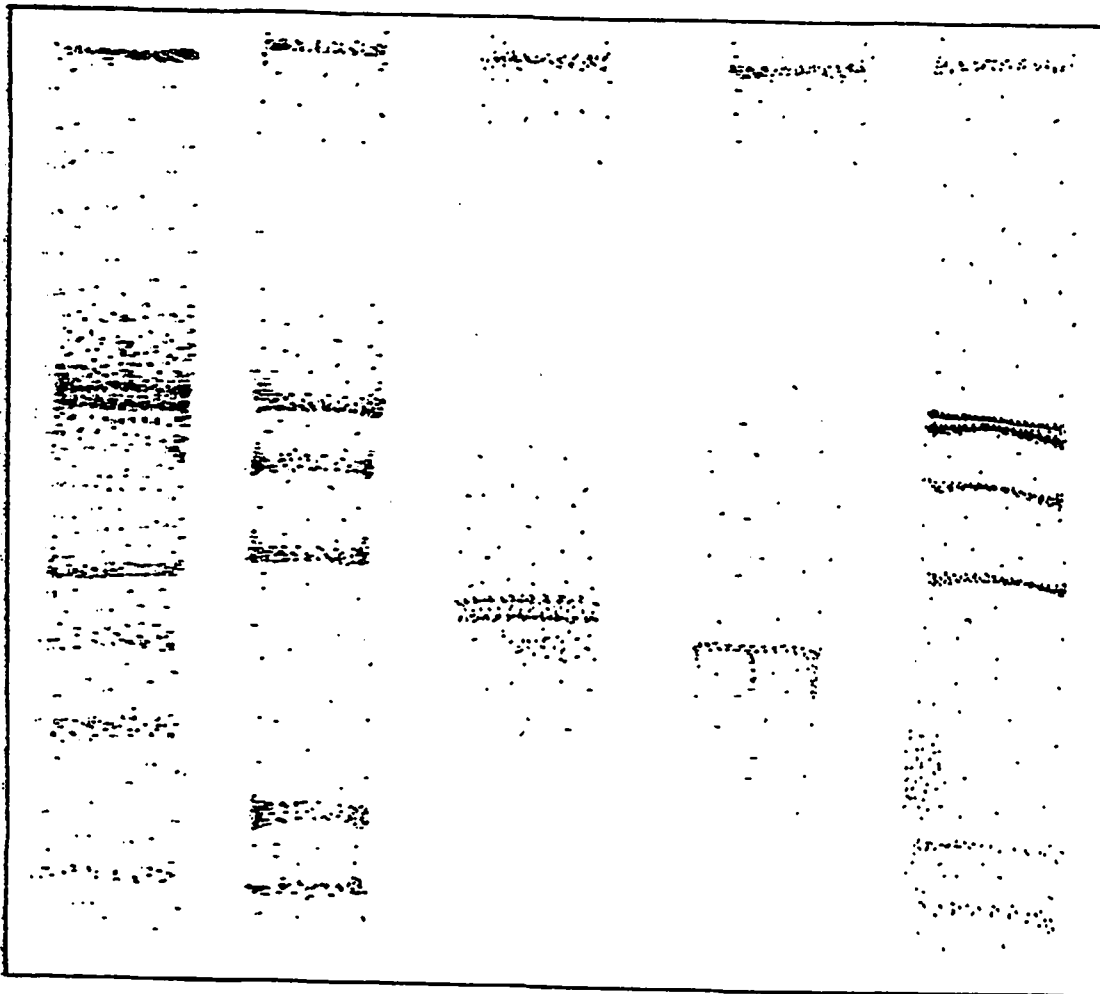


FIG. 4

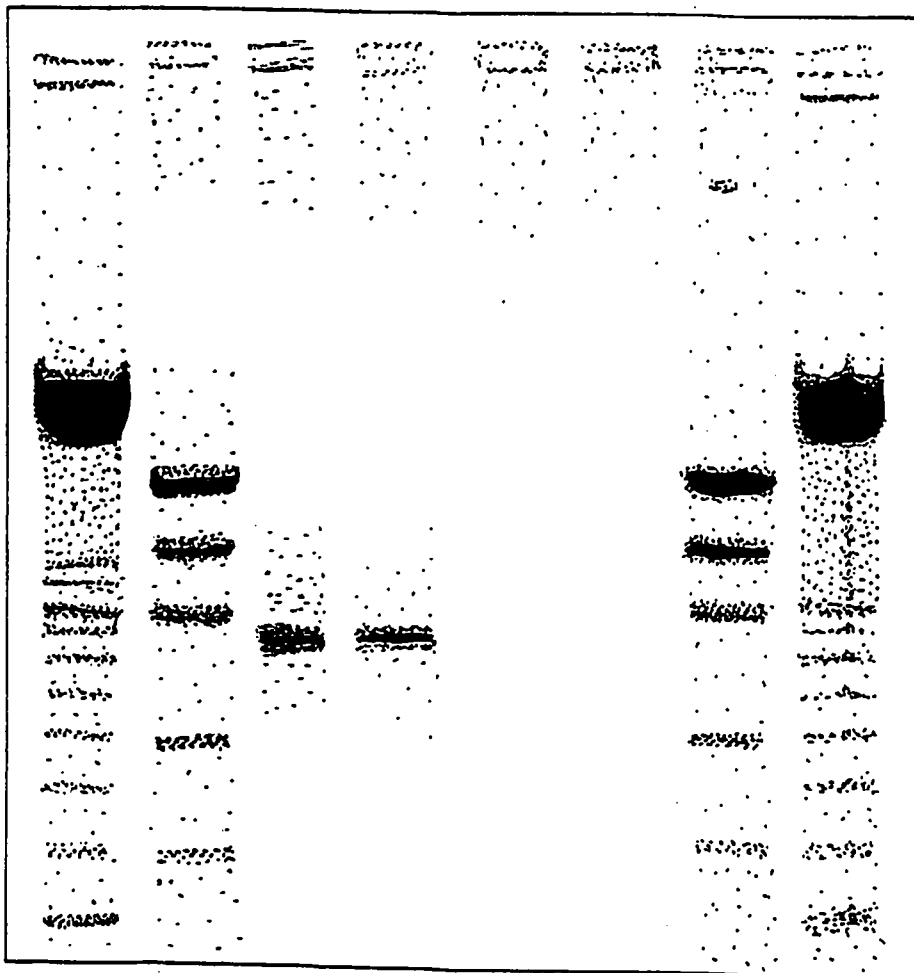
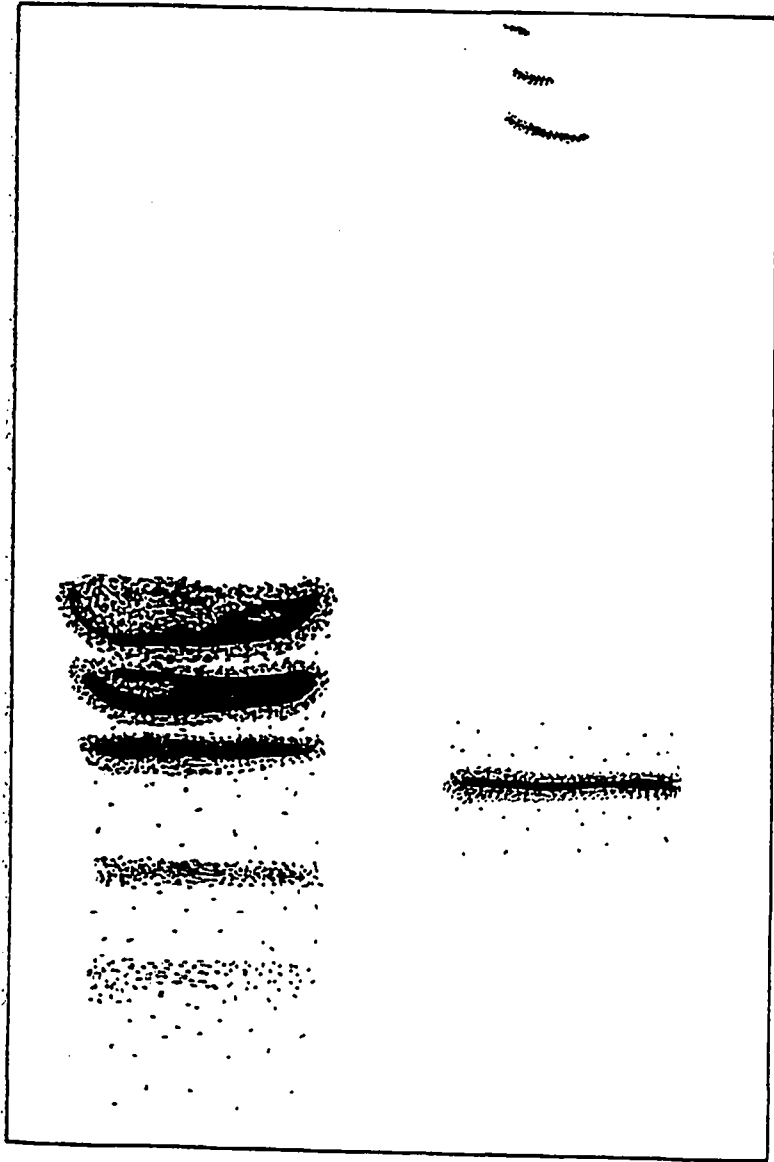


FIG. 5



*FIG. 6*

Table 1: Cleavage of 75 human light chains.

Enzyme	Recognition*	Nch	Ns	Planned location of site
AfeI	AGCgct	0	0	
AflIII	Cttaag	0	0	HC FR3
AgeI	Accggt	0	0	
AscI	GGcgcgcc	0	0	After LC
BglII	Agatct	0	0	
BsiWI	Cgtacg	0	0	
BspDI	ATcgat	0	0	
BssHII	Gcgcg	0	0	
BstBI	TTcgaa	0	0	
DraIII	CACNNNgtg	0	0	
EagI	Cggccg	0	0	
FseI	GGCCGGcc	0	0	
FspI	TGCgca	0	0	
HpaI	GTTaac	0	0	
MfeI	Caattg	0	0	HC FR1
MluI	Acgagt	0	0	
NcoI	Caatgg	0	0	Heavy chain signal
NheI	Gctagc	0	0	HC/anchor linker
NotI	GCggccgc	0	0	In linker after HC
NruI	TCGcga	0	0	
PacI	TTAATtaa	0	0	
PmeI	GTTTaaac	0	0	
PmlI	CACgtg	0	0	
PvuI	CGATcg	0	0	
SacII	CCGCgg	0	0	
SalI	Gtcgac	0	0	
SfiI	GGCCNNNNnggcc	0	0	Heavy Chain signal
SgfI	GCGATcgc	0	0	
SnaBI	TACgta	0	0	
StuI	AGGcct	0	0	
XbaI	Tctaga	0	0	HC FR3
AatII	GACGTC	1	1	
AclI	AACggt	1	1	
AseI	ATtaat	1	1	
BsmI	GAATGCN	1	1	
BspEI	Tccgga	1	1	HC FR1
BstXI	CCANNNNntgg	1	1	HC FR2
DrdI	GACNNNNngtc	1	1	
HindIII	Aagctt	1	1	
PciI	Acatgt	1	1	
SapI	gaagagc	1	1	
ScaI	AGTact	1	1	
SexAI	Accwgt	1	1	
SpeI	Actagt	1	1	
TliI	Ctcgag	1	1	
XhoI	Ctcgag	1	1	
BcgI	cgannnnntgc	2	2	
BlpI	GCTnagc	2	2	
BssSI	Ctcgtg	2	2	
BstAPI	GCANNNNntgc	2	2	
EspI	GCTnagc	2	2	
KasI	Gcgcc	2	2	
PflMI	CCANNNNntgg	2	2	
XmnI	GAANNnttc	2	2	

<b>ApaI</b>	<b>Gtgcac</b>	<b>3</b>	<b>3</b>	<b>LC signal seq</b>
NaeI	GCCggc	3	3	
NgoMI	Gccggc	3	3	
PvuII	CAGctg	3	3	
RsrII	CGgwccg	3	3	
BsrBI	GAGcgg	4	4	
BsrDI	GCAATGNNn	4	4	
BstZ17I	GTAtac	4	4	
EcoRI	Gaattc	4	4	
SphI	GCATGc	4	4	
SspI	AATatt	4	4	
AccI	GTmkac	5	5	
BclI	Tgatca	5	5	
BsmBI	Nnnnnngagacg	5	5	
BsrGI	Tgtaca	5	5	
DraI	TTTaaa	6	6	
<b>NdeI</b>	<b>CAtatg</b>	<b>6</b>	<b>6</b>	<b>HC FR4</b>
SwaI	ATTTaaat	6	6	
BamHI	Ggatcc	7	7	
SacI	GAGCTc	7	7	
BclVI	GTATCCNNNNNN	8	8	
BsaBI	GATNNnnatc	8	8	
NsiI	ATGCAt	8	8	
<b>Bsp120I</b>	<b>Gggccc</b>	<b>9</b>	<b>9</b>	<b>CH1</b>
<b>ApaI</b>	<b>GGGCCc</b>	<b>9</b>	<b>9</b>	<b>CH1</b>
PspOoMI	Gggccc	9	9	
BspHI	Tcatga	9	11	
EcoRV	GATatc	9	9	
AhdI	GACNNNnngtc	11	11	
BbsI	GAAGAC	11	14	
PsiI	TTAtaa	12	12	
BsaI	GGTCTCNnnnn	13	15	
XmaI	Cccggg	13	14	
AvaI	Cycgrg	14	16	
BglI	GCCNNNNnggc	14	17	
AlwNI	CAGNNNctg	16	16	
BspMI	ACCTGC	17	19	
XcmI	CCANNNNNnnnntgg	17	26	
<b>BstEII</b>	<b>Ggtnacc</b>	<b>19</b>	<b>22</b>	<b>HC FR4</b>
Sse8387I	CCTGCagg	20	20	
AvrII	Cctagg	22	22	
HincII	GTYrac	22	22	
BsgI	GTGCAG	27	29	
MscI	TGGcca	30	34	
BseRI	NNnnnnnnnnctcctc	32	35	
Bsu36I	CCtnagg	35	37	
PstI	CTGCAG	35	40	
EciI	nnnnnnnnntccgcc	38	40	
PpuMI	RGgwccy	41	50	
StyI	Ccwwgg	44	73	
EcoO109I	RGgnccy	46	70	
Acc65I	Ggtacc	50	51	
KpnI	GGTACc	50	51	
BpmI	ctccag	53	82	
AvaII	Ggwcc	71	124	

\* cleavage occurs in the top strand after the last upper-case base. For REs

that cut palindromic sequences, the lower strand is cut at the symmetrical site.

Table 2: Cleavage of 79 human heavy chains

Enzyme	Recognition	Nch	Ns	Planned location of site
AfeI	AGCgct	0	0	
AflII	Cttaag	0	0	HC FR3
AscI	GGGcgcc	0	0	After LC
BsiWI	Cgtacg	0	0	
BspDI	ATcgat	0	0	
BssHII	Gcgcgc	0	0	
FseI	GGCCGGcc	0	0	
HpaI	GTTaac	0	0	
NheI	Gctagc	0	0	HC Linker
NotI	GCggcgc	0	0	In linker, HC/anchor
NruI	TCGcga	0	0	
NsiI	ATGCAT	0	0	
PacI	TTAATtaa	0	0	
PciI	Acatgt	0	0	
PmeI	GTTTaaac	0	0	
PvuI	CGATcg	0	0	
RsrII	CGgwccg	0	0	
SapI	gaagagc	0	0	
SfiI	GGCCNNNNnggcc	0	0	HC signal seq
SgfI	GCGATcgc	0	0	
SwaI	ATTTaaat	0	0	
AclI	AACgtt	1	1	
AgeI	Accggt	1	1	
AseI	ATtaat	1	1	
AvrII	Cctagg	1	1	
BsmI	GAATGCN	1	1	
BsrBI	GAGcgg	1	1	
BsrDI	GCAATGNNn	1	1	
DraI	TTTaaa	1	1	
FspI	TGCgca	1	1	
HindIII	Aagctt	1	1	
MfeI	Caattg	1	1	HC FR1
NaeI	GCCggc	1	1	
NgoMI	Gccggc	1	1	
SpeI	Actagt	1	1	
Acc65I	Ggtacc	2	2	
BstBI	TTcgaa	2	2	
KpnI	GGTACc	2	2	
MluI	Acgcgt	2	2	
NcoI	Ccatgg	2	2	In HC signal seq
NdeI	CATatg	2	2	HC FR4
PmlI	CACgtg	2	2	
XcmI	CCANNNNNnnntgg	2	2	
BcgI	cgannnnntgc	3	3	
BclI	Tgatca	3	3	
BglI	GCCNNNNnggc	3	3	
BsaBI	GATNNnnatc	3	3	
BsrGI	Tgtaca	3	3	
SnaBI	TACgta	3	3	
Sse8387I	CCTGCAgg	3	3	

<b>ApaI</b>	<b>Gtgcac</b>	<b>4</b>	<b>4</b>	<b>LC Signal/FR1</b>
<b>BspHI</b>	<b>Tcatga</b>	<b>4</b>	<b>4</b>	
<b>BssSI</b>	<b>Ctcgtg</b>	<b>4</b>	<b>4</b>	
<b>PsiI</b>	<b>TTATAa</b>	<b>4</b>	<b>5</b>	
<b>SphI</b>	<b>GCATGc</b>	<b>4</b>	<b>4</b>	
<b>AhdI</b>	<b>GACNNNnngtc</b>	<b>5</b>	<b>5</b>	
<b>BspEI</b>	<b>Tccgga</b>	<b>5</b>	<b>5</b>	<b>HC FR1</b>
<b>MscI</b>	<b>TGGcca</b>	<b>5</b>	<b>5</b>	
<b>SacI</b>	<b>GAGCTc</b>	<b>5</b>	<b>5</b>	
<b>ScaI</b>	<b>AGTact</b>	<b>5</b>	<b>5</b>	
<b>SexAI</b>	<b>Accwgg</b>	<b>5</b>	<b>6</b>	
<b>SspI</b>	<b>AATatt</b>	<b>5</b>	<b>5</b>	
<b>TliI</b>	<b>Ctcgag</b>	<b>5</b>	<b>5</b>	
<b>XhoI</b>	<b>Ctcgag</b>	<b>5</b>	<b>5</b>	
<b>BbsI</b>	<b>GAAGAC</b>	<b>7</b>	<b>8</b>	
<b>BstAPI</b>	<b>GCANNNNntgc</b>	<b>7</b>	<b>8</b>	
<b>BstZ17I</b>	<b>GTAtac</b>	<b>7</b>	<b>7</b>	
<b>EcoRV</b>	<b>GATatc</b>	<b>7</b>	<b>7</b>	
<b>EcoRI</b>	<b>Gaattc</b>	<b>8</b>	<b>8</b>	
<b>BlpI</b>	<b>GCtnagc</b>	<b>9</b>	<b>9</b>	
<b>Bsu36I</b>	<b>CCtnagg</b>	<b>9</b>	<b>9</b>	
<b>DraIII</b>	<b>CACNNNgtg</b>	<b>9</b>	<b>9</b>	
<b>EspI</b>	<b>GCtnagc</b>	<b>9</b>	<b>9</b>	
<b>StuI</b>	<b>AGGcct</b>	<b>9</b>	<b>13</b>	
<b>XbaI</b>	<b>Tctaga</b>	<b>9</b>	<b>9</b>	<b>HC FR3</b>
<b>Bsp120I</b>	<b>Gggccc</b>	<b>10</b>	<b>11</b>	<b>CH1</b>
<b>ApaI</b>	<b>GGGCCc</b>	<b>10</b>	<b>11</b>	<b>CH1</b>
<b>PspOoMI</b>	<b>Gggccc</b>	<b>10</b>	<b>11</b>	
<b>BciVI</b>	<b>GTATCCNNNNNN</b>	<b>11</b>	<b>11</b>	
<b>SalI</b>	<b>Gtgcac</b>	<b>11</b>	<b>12</b>	
<b>DrdI</b>	<b>GACNNNnngtc</b>	<b>12</b>	<b>12</b>	
<b>KasI</b>	<b>Ggcgcc</b>	<b>12</b>	<b>12</b>	
<b>XmaI</b>	<b>Cccggg</b>	<b>12</b>	<b>14</b>	
<b>BglII</b>	<b>Agatct</b>	<b>14</b>	<b>14</b>	
<b>HincII</b>	<b>GTYrac</b>	<b>16</b>	<b>18</b>	
<b>BamHI</b>	<b>Ggatcc</b>	<b>17</b>	<b>17</b>	
<b>PflMI</b>	<b>CCANNNNntgg</b>	<b>17</b>	<b>18</b>	
<b>BsmBI</b>	<b>Nnnnnngagacg</b>	<b>18</b>	<b>21</b>	
<b>BstXI</b>	<b>CCANNNNntgg</b>	<b>18</b>	<b>19</b>	<b>HC FR2</b>
<b>XmnI</b>	<b>GAANNnttc</b>	<b>18</b>	<b>18</b>	
<b>SacII</b>	<b>CCGCgg</b>	<b>19</b>	<b>19</b>	
<b>PstI</b>	<b>CTGCAg</b>	<b>20</b>	<b>24</b>	
<b>PvuII</b>	<b>CAGctg</b>	<b>20</b>	<b>22</b>	
<b>AvaI</b>	<b>Cycgrg</b>	<b>21</b>	<b>24</b>	
<b>EagI</b>	<b>Cggccg</b>	<b>21</b>	<b>22</b>	
<b>AatII</b>	<b>GACGTc</b>	<b>22</b>	<b>22</b>	
<b>BspMI</b>	<b>ACCTGC</b>	<b>27</b>	<b>33</b>	
<b>AccI</b>	<b>GTmkac</b>	<b>30</b>	<b>43</b>	
<b>StyI</b>	<b>Ccwwgg</b>	<b>36</b>	<b>49</b>	
<b>AlwNI</b>	<b>CAGNNNctg</b>	<b>38</b>	<b>44</b>	
<b>BsaI</b>	<b>GGTCTCnann</b>	<b>38</b>	<b>44</b>	
<b>PpuMI</b>	<b>RGgwccy</b>	<b>43</b>	<b>46</b>	
<b>BsgI</b>	<b>GTGCAG</b>	<b>44</b>	<b>54</b>	
<b>BseRI</b>	<b>NNnnnnnnnctcctc</b>	<b>48</b>	<b>60</b>	
<b>EciI</b>	<b>nnnnnnnnntccgcc</b>	<b>52</b>	<b>57</b>	
<b>BstEII</b>	<b>Ggtnacc</b>	<b>54</b>	<b>61</b>	<b>HC Fr4, 47/79 have one</b>
<b>EcoO109I</b>	<b>RGgnccy</b>	<b>54</b>	<b>86</b>	
<b>BpmI</b>	<b>ctccag</b>	<b>60</b>	<b>121</b>	
<b>AvaII</b>	<b>Ggwcc</b>	<b>71</b>	<b>140</b>	





2007211861 21 Aug 2007

12/128

3'- gtGTAGGcac\ (SEQ ID NO:23)

└caCATCCgtg/

5'-gag|tctc-actgagc

Substrate 3'-...ctc-agag|tgactcg...-5' (SEQ ID NO:24)

Improved FokI adapters

FokI - for dsDNA, | represents sites of cleavage

Case I

Stem 11, loop 5, stem 11, recognition 17

5'-...catgtg|tatt-actgtgc..Substrate....-3'

3'-gtacac-ataa|tgacacg┐  
gtGTAGGcacG T  
5'- caCATCCgtgc C  
└TT┘

Case II

Stem 10, loop 5, stem 10, recognition 18

5'-...gtgtatt|agac-tgctgcc..Substrate....-3'

┐T┘  
┐cacataa-tctg|acgacgg-5'  
T gtgCCTACac  
C cacGGATGtg-3'  
└TT┘

Case III (Case I rotated 180 degrees)

Stem 11, loop 5, stem 11, recognition 20

┐ T┘  
T TgtgCCTACac-5'  
G AcacGGATGtg┐  
└TT┘  
gtgtctt|acag-tccattctg-3' Adapter  
3'-...cacagaa-tgtc|aggtaagac..substrate....-5'

Case IV (Case II rotated 180 degrees)

Stem 11, loop 4, stem 11, recognition 17

3'- gtGTAGGcacc T  
┐caCATCCgtgg T  
5'-atcgag|tctc-actgagc┐  
Substrate 3'-...tagctc-agag|tgactcg...-5'  
└TT┘

## BseRI

| sites of cleavage  
 5'-cacGAGGAGnnnnnnnnnn|nnnnn-3'  
 3'-gtgctcctcnnnnnnnnn|nnnnnn-5'

RECOG

NITion of *BseRI*

Stem 11, loop 5, stem 11, recognition 19

3'-.....gaacat|cg-ttaagccagta.....5'  
 cttgta-gc|aattcggtcat-3'  
 [T-T]  
 C GCTGAGGAGTC-]  
 T cgactcctcag-5' An adapter for *BseRI* to cleave the substrate above.  
 [T-]

Table 8: Matches to URE FR3 adapters in 79 human HC.

## A. List of Heavy-chains genes sampled

AF008566	af103343	HSA235676	HSU92452	HSZ93860
AF035043	AF103367	HSA235675	HSU94412	HSZ93863
AF103026	AF103368	HSA235674	HSU94415	MCOMFRAA
af103033	AF103369	HSA235673	HSU94416	MCOMFRVA
AF103061	AF103370	HSA240559	HSU94417	S82745
Af103072	af103371	HSCB201	HSU94418	S82764
af103078	AF103372	HSIGGVHC	HSU96389	S83240
AF103099	AF158381	HSU44791	HSU96391	SABVH369
AF103102	E05213	HSU44793	HSU96392	SADEIGVH
AF103103	E05886	HSU82771	HSU96395	SAH2IGVH
AF103174	E05887	HSU82949	HSZ93849	SDA3IGVH
AF103186	HSA235661	HSU82950	HSZ93850	SIGVHTTD
af103187	HSA235664	HSU82952	HSZ93851	SUK4IGVH
AF103195	HSA235660	HSU82961	HSZ93853	
af103277	HSA235659	HSU86522	HSZ93855	
af103286	HSA235678	HSU86523	HSZ93857	
AF103309	HSA235677			

Table 8 B. Testing all distinct GLGs from bases 89.1 to 93.2 of the heavy variable domain

Id	Nb	0	1	2	3	4		SEQ ID NO:
1	38	15	11	10	0	2	Seq1	gtgtattactgtgc 25
2	19	7	6	4	2	0	Seq2	gtAtattactgtgc 26
3	1	0	0	1	0	0	Seq3	gtgtattactgtAA 27
4	7	1	5	1	0	0	Seq4	gtgtattactgtAc 28
5	0	0	0	0	0	0	Seq5	Ttgtattactgtgc 29
6	0	0	0	0	0	0	Seq6	TtgtatCactgtgc 30
7	3	1	0	1	1	0	Seq7	ACAtattactgtgc 31
8	2	0	2	0	0	0	Seq8	ACgtattactgtgc 32
9	9	2	2	4	1	0	Seq9	ATgtattactgtgc 33
Group		26	26	21	4	2		
Cumulative		26	52	73	77	79		

Table 8C Most important URE recognition seqs in FR3 Heavy

1	VHSzy1	GTGtattactgtgc	(ON_SHC103)	(SEQ ID NO:25)
2	VHSzy2	GTAtattactgtgc	(ON_SHC323)	(SEQ ID NO:26)
3	VHSzy4	GTGtattactgtac	(ON_SHC349)	(SEQ ID NO:28)
4	VHSzy9	ATGtattactgtgc	(ON_SHC5a)	(SEQ ID NO:33)

Table 8D, testing 79 human HC V genes with four probes

Number of sequences..... 79  
 Number of bases..... 29143

Id	Best	Number of mismatches							
		0	1	2	3	4			
1	39	15	11	10	1	2	0	Seq1	gtgtattactgtgc (SEQ ID NO:25)
2	22	7	6	5	3	0	1	Seq2	gtAtattactgtgc (SEQ ID NO:26)
3	7	1	5	1	0	0	0	Seq4	gtgtattactgtAc (SEQ ID NO:28)
4	11	2	4	4	1	0	0	Seq9	ATgtattactgtgc (SEQ ID NO:33)
Group		25	26	20	5	2			
Cumulative		25	51	71	76	78			

One sequence has five mismatches with sequences 2, 4, and 9; it is scored as best for 2.

Id is the number of the adapter.

Best is the number of sequence for which the identified adapter was the best available.

The rest of the table shows how well the sequences match the adapters. For example, there are 11 sequences that match VHSzy1(Id=1) with 2 mismatches and are worse for all other adapters. In this sample, 90% come within 2 bases of one of the four adapters.

Table 130: PCR primers for amplification of human Ab genes

(HuIgMFOR) 5'-tgg aag agg cac gtt ctt ttc ttt-3'

30 ! (HuIgMFOREtop) 5'-aaa gaa aag aac gtg cct ctt cca-3' = reverse complement

(HuCkFOR) 5'-aca ctc tcc cct gtt gaa gct ctt-3'

(HuCL2FOR) 5'-tga aca ttc tgt agg ggc cac tg-3'

(HuCL7FOR) 5'-aga gca ttc tgc agg ggc cac tg-3'

! Kappa

35 (CKForeAsc) 5'-acc gcc tcc acc ggg cgc gcc tta tta aca ctc tcc cct gtt-  
gaa gct ctt-3'

(CL2ForeAsc) 5'-acc gcc tcc acc ggg cgc gcc tta tta tga aca ttc tgt-  
agg ggc cac tg-3'

(CL7ForeAsc) 5'-acc gcc tcc acc ggg cgc gcc tta tta aga gca ttc tgc-  
40 agg ggc cac tg-3'

Table 195: Human GLG FR3 sequences

45 ! VH1

! 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80

agg gtc acc atg acc agg gac acg tcc atc agc aca gcc tac atg  
! 81 82 82a 82b 82c 83 84 85 86 87 88 89 90 91 92  
gag ctg agc agg ctg aga tct gac gac acg gcc gtg tat tac tgt  
! 93 94 95  
5 gcg aga ga ! 1-02# 1  
aga gtc acc att acc agg gac aca tcc gcg agc aca gcc tac atg  
gag ctg agc agc ctg aga tct gaa gac acg gct gtg tat tac tgt  
gcg aga ga ! 1-03# 2  
10 aga gtc acc atg acc agg aac acc tcc ata agc aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gcg aga gg ! 1-08# 3  
aga gtc acc atg acc aca gac aca tcc acg agc aca gcc tac atg  
gag ctg agg agc ctg aga tct gac gac acg gcc gtg tat tac tgt  
gcg aga ga ! 1-18# 4  
15 aga gtc acc atg acc gag gac aca tct aca gac aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gca aca ga ! 1-24# 5  
aga gtc acc att acc agg gac agg tct atg agc aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac aca gcc atg tat tac tgt  
20 gca aga ta ! 1-45# 6  
aga gtc acc atg acc agg gac acg tcc acg agc aca gtc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gcg aga ga ! 1-46# 7

18/128

aga gtc acc att acc agg gac atg tcc aca agc aca gcc tac atg  
gag ctg agc agc ctg aga tcc gag gac acg gcc gtg tat tac tgt  
gcg gca ga ! 1-58# 8

5  
aga gtc acg att acc gcg gac gaa tcc acg agc aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gcg aga ga ! 1-69# 9

aga gtc acg att acc gcg gac aaa tcc acg agc aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gcg aga ga ! 1-e# 10

10  
aga gtc acc ata acc gcg gac acg tct aca gac aca gcc tac atg  
gag ctg agc agc ctg aga tct gag gac acg gcc gtg tat tac tgt  
gca aca ga ! 1-f# 11

! VH2

15  
agg ctc acc atc acc aag gac acc tcc aaa aac cag gtg gtc ctt  
aca atg acc aac atg gac cct gtg gac aca gcc aca tat tac tgt  
gca cac aga c! 2-05# 12

agg ctc acc atc tcc aag gac acc tcc aaa agc cag gtg gtc ctt  
acc atg acc aac atg gac cct gtg gac aca gcc aca tat tac tgt  
gca cgg ata c! 2-26# 13

20  
agg ctc acc atc tcc aag gac acc tcc aaa aac cag gtg gtc ctt  
aca atg acc aac atg gac cct gtg gac aca gcc acg tat tac tgt  
gca cgg ata c! 2-70# 14

! VH3

25  
cga ttc acc atc tcc aga gac aac gcc aag aac tca ctg tat ctg  
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3-07# 15

cga ttc acc atc tcc aga gac aac gcc aag aac tcc ctg tat ctg  
caa atg aac agt ctg aga gct gag gac acg gcc ttg tat tac tgt  
gca aaa gat a! 3-09#16

30  
cga ttc acc atc tcc agg gac aac gcc aag aac tca ctg tat ctg  
caa atg aac agc ctg aga gcc gag gac acg gcc gtg tat tac tgt  
gcg aga ga ! 3-11# 17

cga ttc acc atc tcc aga gaa aat gcc aag aac tcc ttg tat ctt  
caa atg aac agc ctg aga gcc ggg gac acg gct gtg tat tac tgt  
gca aga ga ! 3-13# 18

35  
aga ttc acc atc tca aga gat gat tca aaa aac acg ctg tat ctg  
caa atg aac agc ctg aaa acc gag gac aca gcc gtg tat tac tgt  
acc aca ga ! 3-15# 19

cga ttc acc atc tcc aga gac aac gcc aag aac tcc ctg tat ctg



caa atg aac agt ctg aga gcc gag gac acg gcc ttg tat cac tgt  
gcg aga ga ! 3-20# 20

cga ttc acc atc tcc aga gac aac gcc aag aac tca ctg tat ctg  
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3-21# 21

cgg ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg  
caa atg aac agc ctg aga gcc gag gac acg gcc gta tat tac tgt  
gcg aaa ga ! 3-23# 22

cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg  
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt  
gcg aaa ga ! 3-30# 23

cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg  
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3303# 24

cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg  
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt  
gcg aaa ga ! 3305# 25

cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctg  
caa atg aac agc ctg aga gcc gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3-33# 26

cga ttc acc atc tcc aga gac aac agc aaa aac tcc ctg tat ctg  
caa atg aac agt ctg aga act gag gac acc gcc ttg tat tac tgt  
gca aaa gat a! 3-43#27

cga ttc acc atc tcc aga gac aat gcc aag aac tca ctg tat ctg  
caa atg aac agc ctg aga gac gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3-48# 28

aga ttc acc atc tca aga gat ggt tcc aaa agc atc gcc tat ctg  
caa atg aac agc ctg aaa acc gag gac aca gcc gtg tat tac tgt  
act aga ga ! 3-49# 29

cga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctt  
caa atg aac agc ctg aga gcc gag gac acg gcc gtg tat tac tgt  
gcg aga ga ! 3-53# 30

aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctt  
caa atg ggc agc ctg aga gct gag gac atg gct gtg tat tac tgt  
gcg aga ga ! 3-64# 31

aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg tat ctt  
caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt  
gcg aga ga ! 3-66# 32

aga ttc acc atc tca aga gat gat tca aag aac tca ctg tat ctg

caa atg aac agc ctg aaa acc gag gac acg gcc gtg tat tac tgt  
gct aga ga ! 3-72# 33

agg ttc acc atc tcc aga gat gat tca aag aac acg gcg tat ctg  
caa atg aac agc ctg aaa acc gag gac acg gcc gtg tat tac tgt

act aga ca ! 3-73# 34

cga ttc acc atc tcc aga gac aac gcc aag aac acg ctg tat ctg

caa atg aac agt ctg aga gcc gag gac acg gct gtg tat tac tgt

gca aga ga ! 3-74# 35

aga ttc acc atc tcc aga gac aat tcc aag aac acg ctg cat ctt

caa atg aac agc ctg aga gct gag gac acg gct gtg tat tac tgt

aag aaa ga ! 3-d# 36

! VH4

cga gtc acc ata tca gta gac aag tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gcc gcg gac acg gcc gtg tat tac tgt

gcg aga ga ! 4-04# 37

cga gtc acc atg tca gta gac acg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gcc gtg gac acg gcc gtg tat tac tgt

gcg aga aa ! 4-28# 38

cga gtt acc ata tca gta gac acg tct aag aac cag ttc tcc ctg

aag ctg agc tct gtg act gcc gcg gac acg gcc gtg tat tac tgt

gcg aga ga ! 4301# 39

cga gtc acc ata tca gta gac agg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gcc gcg gac acg gcc gtg tat tac tgt

gcc aga ga ! 4302# 40

cga gtt acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg act gcc gca gac acg gcc gtg tat tac tgt

gcc aga ga ! 4304# 41

cga gtt acc ata tca gta gac acg tct aag aac cag ttc tcc ctg

aag ctg agc tct gtg act gcc gcg gac acg gcc gtg tat tac tgt

gcg aga ga ! 4-31# 42

cga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gcc gcg gac acg gct gtg tat tac tgt

gcg aga ga ! 4-34# 43

cga gtc acc ata tcc gta gac acg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gcc gca gac acg gct gtg tat tac tgt

gcg aga ca ! 4-39# 44

cga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg

aag ctg agc tct gtg acc gct gcg gac acg gcc gtg tat tac tgt

gcg aga ga ! 4-59# 45

21/128

cga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg  
aag ctg agc tct gtg acc gct gcg gac acg gcc gtg tat tac tgt  
gcg aga ga ! 4-61# 46

cga gtc acc ata tca gta gac acg tcc aag aac cag ttc tcc ctg  
aag ctg agc tct gtg acc gcc gca gac acg gcc gtg tat tac tgt  
gcg aga ga ! 4-b# 47

! VH5

cag gtc acc atc tca gcc gac aag tcc atc agc acc gcc tac ctg  
cag tgg agc agc ctg aag gcc tcg gac acc gcc atg tat tac tgt  
gcg aga ca ! 5-51# 48

cac gtc acc atc tca gct gac aag tcc atc agc act gcc tac ctg  
cag tgg agc agc ctg aag gcc tcg gac acc gcc atg tat tac tgt  
gcg aga ! 5-a# 49

! VH6

cga ata acc atc aac cca gac aca tcc aag aac cag ttc tcc ctg  
cag ctg aac tct gtg act ccc gag gac acg gct gtg tat tac tgt  
gca aga ga ! 6-1# 50

! VH7

cgg ttt gtc ttc tcc ttg gac acc tct gtc agc acg gca tat ctg  
cag atc tgc agc cta aag gct gag gac act gcc gtg tat tac tgt  
gcg aga ga ! 74.1# 51

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21 Aug 2007

2007211861

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Table 250: REaptors, Extenders, and Bridges used for Cleavage and Capture of Human Heavy Chains in FR3.

**A: HpyCH4V Probes of actual human HC genes**

!HpyCH4V in FR3 of human HC, bases 35-56; only those with TGca site

TGca;10,

RE recognition:tgca

of length 4 is expected at 10

1

6-1 agttctccctgcagctgaactc

2007211861 21 Aug 2007

```

2          3-11,3-07,3-21,3-72,3-48  cactgtatctgcaaatgaacag
3          3-09,3-43,3-20          ccctgtatctgcaaatgaacag
4          5-51                    cgcctacctgcagtgaggagcag
5  3-15,3-30,3-30.5,3-30.3,3-74,3-23,3-33  cgctgtatctgcaaatgaacag
6          7-4.1                   cggcatatctgcagatctgcag
7          3-73                    cggcgtatctgcaaatgaacag
8          5-a                      ctgcctacctgcagtgaggagcag
9          3-49                    tcgcctatctgcaaatgaacag

```

10 **B: HpyCH4V REadaptors, Extenders, and Bridges**

**B.1 REadaptors**

```

! Cutting HC lower strand:
! TmKeller for 100 mM NaCl, zero formamide
! Edapters for cleavage

```

		$T_m^W$	$T_m^K$
15	(ON_HCFR36-1) 5'-agttctcccTGCAgctgaactc-3'	68.0	64.5
	(ON_HCFR36-1A) 5'-ttctcccTGCAgctgaactc-3'	62.0	62.5
	(ON_HCFR36-1B) 5'-ttctcccTGCAgctgaac-3'	56.0	59.9
	(ON_HCFR33-15) 5'-cgctgtatcTGCAaatgaacag-3'	64.0	60.8
	(ON_HCFR33-15A) 5'-ctgtatcTGCAaatgaacag-3'	56.0	56.3
20	(ON_HCFR33-15B) 5'-ctgtatcTGCAaatgaac-3'	50.0	53.1
	(ON_HCFR33-11) 5'-cactgtatcTGCAaatgaacag-3'	62.0	58.9
	(ON_HCFR35-51) 5'-ccgcctaccTGCAgtggagcag-3'	74.0	70.1

**B.2 Segment of synthetic 3-23 gene into which captured CDR3 is to be cloned**

```

25 ! XbaI...
! D323* cgCttcacTaag tcT aqa gac aaC tcT aag aaT acT ctC taC
! scab..... designed gene 3-23 gene.....
!

```

```

30 ! HpyCH4V
! .. .. AflIII...
! Ttg caG atg aac agc TtA aqG . . .
! .....
!

```

**B.3 Extender and Bridges**

35 ! Extender (bottom strand):

(ON\_HCHpyEx01) 5' -cAAGTAGAgAgTATTcTTAgAgTTgTcTcTAGAcTTAgTgAAgcg-3'

! ON\_HCHpyEx01 is the reverse complement of

! 5'-cgCttcacTaag tcT aqa gac aaC tcT aag aaT acT ctC taC Ttg -3'

40 !

! Bridges (top strand, 9-base overlap):

!  
 (ON\_HCHpyBr016-1) 5'-cgCttcacTaag tcT aqa gac aaC tcT aag-  
 aaT acT ctC taC Ttg CAgctgaac-3' (3'-term C is blocked)

5 ! 3-15 et al. + 3-11

(ON\_HCHpyBr023-15) 5'-cgCttcacTaag tcT aqa gac aaC tcT aag-  
 aaT acT ctC taC Ttg CAaatgaac-3' (3'-term C is blocked)

! 5-51

10 (ON\_HCHpyBr045-51) 5'-cgCttcacTaag tcT aqa gac aaC tcT aag-  
 aaT acT ctC taC Ttg CAgtgaggac-3' (3'-term C is blocked)

! PCR primer (top strand)

15 (ON\_HCHpyPCR) 5'-cgCttcacTaag tcT aqa gac-3'

C: B1pI Probes from human HC GLGs

1	1-58, 1-03, 1-08, 1-69, 1-24, 1-45, 1-46, 1-f, 1-e	acatggaGCTGAGCagcctgag
20 2		1-02 acatggaGCTGAGCaggctgag
3		1-18 acatggagctgaggagcctgag
4		5-51, 5-a acctgcagtggagcagcctgaa
5	3-15, 3-73, 3-49, 3-72	atctgcaaatgaacagcctgaa
6	3303, 3-33, 3-07, 3-11, 3-30, 3-21, 3-23, 3305, 3-48	atctgcaaatgaacagcctgag
25 7	3-20, 3-74, 3-09, 3-43	atctgcaaatgaacagctctgag
8		74.1 atctgcagatctgcagcctaaa
9	3-66, 3-13, 3-53, 3-d	atcttcaaatgaacagcctgag
10		3-64 atcttcaaatgggcagcctgag
11	4301, 4-28, 4302, 4-04, 4304, 4-31, 4-34, 4-39, 4-59, 4-61, 4-b	ccctgaaGCTGAGCtctgtgac
30 12		6-1 ccctgcagctgaactctgtgac
13	2-70, 2-05	tccttacaatgaccaacatgga
14		2-26 tccttaccatgaccaacatgga

D: B1pI REaptors, Extenders, and Bridges

35 D.1 REaptors

		$T_m^W$	$T_m^K$
(BlpF3HC1-58)	5'-ac atg gaG CTG AGC agc ctg ag-3'	70	66.4
(BlpF3HC6-1)	5'-cc ctg aag ctg agc tct gtg ac-3'	70	66.4

! BlpF3HC6-1 matches 4-30.1, not 6-1.

40

D.2 Segment of synthetic 3-23 gene into which captured CDR3 is to be cloned



**F: HpyCH4III REaptors, Extenders, and Bridges****F.1 REaptors**

! ONs for cleavage of HC(lower) in FR3(bases 77-97)

! For cleavage with HpyCH4III, Bst4CI, or TaaI

! cleavage is in lower chain before base 88.

	77	78	88	88	88	889	999	999	9	$T_m^W$	$T_m^K$
(H43.77.97.1-02#1)	5'-cc	gtg	tat	tAC	TGT	gcg	aga	g-3'		64	62.6
(H43.77.97.1-03#2)	5'-c	gtg	tat	tAC	TGT	gcg	aga	g-3'		62	60.6
(H43.77.97.108#3)	5'-cc	gtg	tat	tAC	TGT	gcg	aga	g-3'		64	62.6
(H43.77.97.323#22)	5'-cc	gt	tat	tac	tgt	gcg	aga	g-3'		60	58.7
(H43.77.97.330#23)	5'-c	gtg	tat	tac	tgt	gcg	aga	g-3'		60	58.7
(H43.77.97.439#44)	5'-c	gtg	tat	tac	tgt	gcg	aga	g-3'		62	60.6
(H43.77.97.551#48)	5'-cc	gtg	tat	tac	tgt	gcg	aga	g-3'		62	60.6
(H43.77.97.5a#49)	5'-cc	gtg	tat	tAC	TGT	gcg	aga	g-3'		58	58.3

**F.2 Extender and Bridges**

! XbaI and AflII sites in bridges are bunged

(H43.XABr1) 5'-ggtgtagtga-

|TCT|AGT|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

|aac|agC|TTT|AGG|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt|gcg|aga-3'

(H43.XABr2) 5'-ggtgtagtga-

|TCT|AGT|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

|aac|agC|TTT|AGG|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt|gcg|aaa-3'

(H43.XAExt) 5'-ATAgTAGAcT gcAgTgTccT cAgcccTTAA gcTgTTcATc TgcAAgTAGA-  
gAgTATTcTT AgAgTTgTcT cTAgATcAcT AcAcc-3'

!H43.XAExt is the reverse complement of

! 5'-ggtgtagtga-

! |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

! |aac|agC|TTA|AGG|gct|gag|gac|aCT|GCA|Gtc|tac|tat|-3'

(H43.XAPCR) 5'-ggtgtagtga |TCT|AGA|gac|aac-3'

! XbaI and AflII sites in bridges are bunged

(H43.ABr1) 5'-ggtgtagtga-

|aac|agC|TTT|AGG|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt|gcg|aga-3'

(H43.ABr2) 5'-ggtgtagtga-

|aac|agC|TTT|AGG|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt|gcg|aaa-3'

(H43.AEExt) 5'-ATAgTAGAcTgcAgTgTccTcAgcccTTAAgcTgTTcAcTAcAcc-3'



2007211861 21 Aug 2007

27/128

!(H43.AExt) is the reverse complement of 5'-ggtgtagtga-

! |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat -3'

(H43.APCR) 5'-ggtgtagtga |aac|agC|TTA|AGg|gct|g-3'

Table 510

(FOKIact) 5'-cAcATccgTg TTgTT cAcgqATgTg-3'

(VHEX881) 5'-AATAgTAgAc TgcAgTgTcc TcAgcccTTA AgcTgTTcAT cTgcAAgTAG-  
AgAgTATTcT TAGAgTTgTc TcTAGAcTTA gTgAAgcg-3'

! note that VHEX881 is the reverse complement of the ON below

! [RC] 5'-cgCttcacTaag-

! Scab.....

! Synthetic 3-23 as in Table 206

! |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-

! XbaI...

! |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|t-3'

! AflII...

(VHBA881) 5'-cgCttcacTaag-  
|TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-  
|aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt gcg ag-3'

(VHBB881) 5'-cgCttcacTaag-  
|TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|-  
|aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt Acg ag-3'

(VH881PCR) 5'-cgCttcacTaag|TCT|AGA|gac|aac -3'



(BOTFR2) 3'-acc|caa|gcg|-  
 |ggt|cga|gga|cca|ttt|cca|aac|ctc|acc|caa|aga|-5' ! bottom strand

(BOTFR3) 3'- a|cga|ctg|agg|caa|ttt|cca|gcg|aag|-  
 |tga|tag|aga|tct|ctg|ttg|aga|ttc|tta|tga|gag|atg|aac|gtc|tac|-  
 5 |ttg|tcg|aat|tcc|oga|ctc|ctg|tga-5'

(F06) 5'-gC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgc|gct|aaa|-  
 |gac|tat|gaa|ggg|act|ggt|tat|gct|ttc|gaC|ATA|TGg|ggt|c-3'

(BOTFR4) 3'-cga|aag|ctg|tat|acc|cca|ggt|cca|-  
 |tga|tac|cag|tgg|cag|aga|tca-  
 10 |cgg|agg|tgg|ttc|cgg|ggt|agc|cag|aag|ggg-5' ! bottom strand

(BOTPRCPRIM) 3'-gg|ttc|cgg|ggt|agc|cag|aag|ggg-5'

! CDR1 diversity

15 (ON-vgC1) 5'-|gct|TCC|GGA|ttc|act|ttc|tct|<1>|TAC|<1>|atg|<1>|-  
 CDR1.....6859  
|tgg|ggt|cgC|CAa|gct|cct|GG-3'

!<1> stands for an equimolar mix of {ADEFHGHIKLMNPQRSTVWY}; no C  
 (this is not a sequence)

! CDR2 diversity

25 (ON-vgC2) 5'-ggg|ttg|gag|tgg|ggt|tct|<2>|atc|<2>|<3>|-  
 CDR2.....  
 |tct|ggt|ggc|<1>|act|<1>|tat|gct|gac|tcc|ggt|aaa|gg-3'

! CDR2.....

! <1> is an equimolar mixture of {ADEFHGHIKLMNPQRSTVWY}; no C  
 ! <2> is an equimolar mixture of {YRWVGS}; no ACDEFHGHIKLMNPQT  
 30 ! <3> is an equimolar mixture of {PS}; no ACDEFHGHIKLMNPQRSTVWY



Table 800

The following list of enzymes was taken from  
<http://rebase.neb.com/cgi-bin/asymmlist>.

I have removed the enzymes that a) cut within the recognition, b) cut on both sides of the recognition, or c) have fewer than 2 bases between recognition and closest cut site.

REBASE Enzymes  
 04/13/2001

Type II restriction enzymes with asymmetric recognition sequences:

Enzymes	Recognition Sequence	Isoschizomers	Suppliers
AarI	CACCTGCNNNN <sup>^</sup> NNNN <sub>-</sub>	-	Y
AceIII	CAGCTCNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	-	-
Bbr7I	GAAGACNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	-	-
BbvI	GCAGCNNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	-	Y
BbvII	GAAGACNN <sup>^</sup> NNNN <sub>-</sub>	-	-
Bce83I	CTTGAGNNNNNNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	-
BceAI	ACGGCNNNNNNNNNNNNNN <sup>^</sup> NN <sub>-</sub>	-	Y
BceFI	ACGGCNNNNNNNNNNNNNN <sup>^</sup> N <sub>-</sub>	-	-
BciVI	GTATCCNNNNN <sub>-</sub> N <sup>^</sup>	BfuI	Y
BfiI	ACTGGGNNNN <sub>-</sub> N <sup>^</sup>	BmrI	Y
BinI	GGATCNNNN <sup>^</sup> N <sub>-</sub>	-	-
BscAI	GCATCNNNN <sup>^</sup> NN <sub>-</sub>	-	-
BseRI	GAGGAGNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	Y
BsmFI	GGGACNNNNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	BspLU11III	Y
BspMI	ACCTGCNNNN <sup>^</sup> NNNN <sub>-</sub>	Acc36I	Y
EciI	GGCGGANNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	Y
Eco57I	CTGAAGNNNNNNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	BspKT5I	Y
FauI	CCCGCNNNN <sup>^</sup> NN <sub>-</sub>	BstFZ438I	Y
FokI	GGATGNNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	BstPZ418I	Y
GsuI	CTGGAGNNNNNNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	Y
HgaI	GACGCNNNN <sup>^</sup> NNNNN <sub>-</sub>	-	Y
HphI	GGTGANNNNNN <sub>-</sub> N <sup>^</sup>	AsuHPI	Y
MboII	GAAGANNNNNN <sub>-</sub> N <sup>^</sup>	-	Y
MlyI	GAGTCNNNN <sup>^</sup> <sub>-</sub>	SchI	Y
MmeI	TCCRACNNNNNNNNNNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	-
MnlI	CCTCNNNNNN <sub>-</sub> N <sup>^</sup>	-	Y
PleI	GAGTCNNNN <sup>^</sup> N <sub>-</sub>	PpsI	Y
RleAI	CCCACANNNNNNNNNN <sub>-</sub> NNN <sup>^</sup>	-	-
SfaNI	GCATCNNNN <sup>^</sup> NNNN <sub>-</sub>	BspST5I	Y
SspD5I	GGTGANNNNNNN <sup>^</sup> <sub>-</sub>	-	-
Sth132I	CCCGNNNN <sup>^</sup> NNNN <sub>-</sub>	-	-
StsI	GGATGNNNNNNNNNN <sup>^</sup> NNNN <sub>-</sub>	-	-
TaqII	GACCGANNNNNNNNNN <sub>-</sub> NN <sup>^</sup> , CACCCANNNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	-
Tth111III	CAARCANNNNNNNNN <sub>-</sub> NN <sup>^</sup>	-	-
UbaPI	CGAACG	-	-

The notation is <sup>^</sup> means cut the upper strand and <sub>-</sub> means cut the lower strand. If the upper and lower strand are cut at the same place, then only <sup>^</sup> appears.

Table 120: MALIA3, annotated

! MALIA3 9532 bases

```

-----
1 aat gct act act att agt aga att gat gcc acc ttt tca gct cgc gcc
5 ! gene ii continued
49 cca aat gaa aat ata gct aaa cag gtt att gac cat ttg cga aat gta
97 tct aat ggt caa act aaa tct act cgt tcg cag aat tgg gaa tca act
145 gtt aca tgg aat gaa act tcc aga cac cgt act tta gtt gca tat tta
193 aaa cat gtt gag cta cag cac cag att cag caa tta agc tct aag cca
10 241 tcc gca aaa atg acc tct tat caa aag gag caa tta aag gta ctc tct
289 aat cct gac ctg ttg gag ttt gct tcc ggt ctg gtt cgc ttt gaa gct
337 cga att aaa acg cga tat ttg aag tct ttc ggg ctt cct ctt aat ctt
385 ttt gat gca atc cgc ttt gct tct gac tat aat agt cag ggt aaa gac
433 ctg att ttt gat tta tgg tca ttc tcg ttt tct gaa ctg ttt aaa gca
15 481 ttt gag ggg gat tca ATG aat att tat gac gat tcc gca gta ttg gac
! RBS?..... Start gene x, ii continues
529 gct atc cag tct aaa cat ttt act att acc ccc tct ggc aaa act tct
577 ttt gca aaa gcc tct cgc tat ttt ggt ttt tat cgt cgt ctg gta aac
625 gag ggt tat gat agt gtt gct ctt act atg cct cgt aat tcc ttt tgg
20 673 cgt tat gta tct gca tta gtt gaa tgt ggt att cct aaa tct caa ctg
721 atg aat ctt tct acc tgt aat aat gtt gtt ccg tta gtt cgt ttt att
769 aac gta gat ttt tct tcc caa cgt cct gac tgg tat aat gag cca gtt
817 ctt aaa atc gca TAA
! End X & II
25 832 ggtaattca ca
!
! M1 E5 Q10 T15
843 ATG att aaa gtt gaa att aaa cca tct caa gcc caa ttt act act cgt
! Start gene V
30 !
! S17 S20 P25 E30
891 tct ggt gtt tct cgt cag ggc aag cct tat tca ctg aat gag cag ctt
!
! V35 E40 V45
35 939 tgt tac gtt gat ttg ggt aat gaa tat ccg gtt ctt gtc aag att act
!
! D50 A55 L60
987 ctt gat gaa ggt cag cca gcc tat gcg cct ggt cTG TAC Acc gtt cat
! BsrGI...

```

2007211861 21 Aug 2007

34/128

```
!           L65                V70                S75                R80
1035 ctg tcc tct ttc aaa gtt ggt cag ttc ggt tcc ctt atg att gac cgt
!
!
!           P85           K87 end of V
5 1083 ctg cgc ctc gtt ccg gct aag TAA C
!
!
1108 ATG gag cag gtc gcg gat ttc gac aca att tat cag gcg atg
!       Start gene VII
!
!
10 1150 ata caa atc tcc gtt gta ctt tgt ttc gcg ctt ggt ata atc
!
!
!           VII and IX overlap.
!           ..... S2 V3 L4 V5                S10
1192 gct ggg ggt caa agA TGA gt gtt tta gtg tat tct ttc gcc tct ttc gtt
15 !
!           End VII
!           |start IX
!           L13           W15                G20                T25                E29
1242 tta ggt tgg tgc ctt cgt agt ggc att acg tat ttt acc cgt tta atg gaa
!
!
20 1293 act tcc tc
!
!
!           .... stop of IX, IX and VIII overlap by four bases
1301 ATG aaa aag tct tta gtc ctc aaa gcc tct gta gcc gtt gct acc ctc
!       Start signal sequence of viii.
25 !
!
1349 gtt ccg atg ctg tct ttc gct gct gag ggt gac gat ccc gca aaa gcg
!
!           mature VIII --->
1397 gcc ttt aac tcc ctg caa gcc tca gcg acc gaa tat atc ggt tat gcg
1445 tgg gcg atg gtt gtt gtc att
30 1466 gtc ggc gca act atc ggt atc aag ctg ttt aag
1499 aaa ttc acc tcg aaa gca ! 1515
!
!           ..... -35 ..
!
!
1517         agc tga taaaccgat acaattaaag gctccttttg
35 !
!           ..... -10 ...
!
!
1552 gagccttttt ttttGGAGAt ttt ! S.D. underlined
!
!
!           <----- III signal sequence ----->
```



2007211861 21 Aug 2007

35/128

! M K K L L F A I P L V  
1575 caac GTG aaa aaa tta tta ttc gca att cct tta gtt ! 1611

! V P F Y S H S A Q  
5 1612 gtt cct ttc tat tct cac aGT gcA Cag tCT  
! ApaLI...

! 1642 GTC GTG ACG CAG CCG CCC TCA GTG TCT GGG GCC CCA GGG CAG  
AGG GTC ACC ATC TCC TGC ACT GGG AGC AGC TCC AAC ATC GGG GCA  
10 ! BstEII...

1729 GGT TAT GAT GTA CAC TGG TAC CAG CAG CTT CCA GGA ACA GCC CCC AAA  
1777 CTC CTC ATC TAT GGT AAC AGC AAT CGG CCC TCA GGG GTC CCT GAC CGA  
1825 TTC TCT GGC TCC AAG TCT GGC ACC TCA GCC TCC CTG GCC ATC ACT  
1870 GGG CTC CAG GCT GAG GAT GAG GCT GAT TAT  
15 1900 TAC TGC CAG TCC TAT GAC AGC AGC CTG AGT  
1930 GGC CTT TAT GTC TTC GGA ACT GGG ACC AAG GTC ACC GTC

! BstEII...  
1969 CTA GGT CAG CCC AAG GCC AAC CCC ACT GTC ACT  
2002 CTG TTC CCG CCC TCC TCT GAG GAG CTC CAA GCC AAC AAG GCC ACA CTA  
20 2050 GTG TGT CTG ATC AGT GAC TTC TAC CCG GGA GCT GTG ACA GTG GCC TGG  
2098 AAG GCA GAT AGC AGC CCC GTC AAG GCG GGA GTG GAG ACC ACC ACA CCC  
2146 TCC AAA CAA AGC AAC AAC AAG TAC GCG GCC AGC AGC TAT CTG AGC CTG  
2194 ACG CCT GAG CAG TGG AAG TCC CAC AGA AGC TAC AGC TGC CAG GTC ACG  
2242 CAT GAA GGG AGC ACC GTG GAG AAG ACA GTG GCC CCT ACA GAA TGT TCA  
25 2290 TAA TAA ACCG CCTCCACCGG GCGCGCCAAT TCTATTTCAA GGAGACAGTC ATA

! AscI.....

! PelB signal----->  
! M K Y L L P T A A A G L L L L  
30 2343 ATG AAA TAC CTA TTG CCT ACG GCA GCC GCT GGA TTG TTA TTA CTC

! 16 17 18 19 20 21 22  
! A A Q P A M A  
2388 gcG GCC cag ccG GCC atg gcc

35 ! SfiI.....  
! NgoMI... (1/2)  
! NcoI.....  
!

2007211861 21 Aug 2007

36/128

```

!                                     FR1 (DP47/V3-23)-----
!                                     23 24 25 26 27 28 29 30
!                                     E  V  Q  L  L  E  S  G
2409  gaa|ggt|CAA|TTG|tta|gag|tct|ggt|
5   | MfeI |
!
! -----FR1-----
!   31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
!   G  G  L  V  Q  P  G  G  S  L  R  L  S  C  A
! 2433 |ggc|ggt|ctt|ggt|cag|cct|ggt|ggt|tct|tta|cgt|ctt|tct|tgc|gct|
!
! -----FR1----->|...CDR1.....|-----FR2-----
!   46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
!   A  S  G  F  T  F  S  S  Y  A  M  S  W  V  R
5   2478 |gct|TCC|GGA|ttc|act|ttc|tct|tCG|TAC|Gct|atg|tct|tgg|ggt|cgC|
!   | BspEI | | BsiWI | | BstXI.
!
! -----FR2----->|...CDR2.....
!   61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
!   Q  A  P  G  K  G  L  E  W  V  S  A  I  S  G
10  2523 |CAa|gct|ccT|GGt|aaa|ggt|ttg|gag|tgg|ggt|tct|gct|atc|tct|ggt|
!   ...BstXI |
!
! .....CDR2.....|-----FR3-----
5   !   76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
!   S  G  G  S  T  Y  Y  A  D  S  V  K  G  R  F
! 2568 |tct|ggt|ggc|agt|act|tac|tat|gct|gac|tcc|ggt|aaa|ggt|cgc|ttc|
!
! -----FR3-----
!   91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
!   T  I  S  R  D  N  S  K  N  T  L  Y  L  Q  M
! 2613 |act|atc|TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|
!   | XbaI |
5   !
! -----FR3----->|
!   106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
!   N  S  L  R  A  E  D  T  A  V  Y  Y  C  A  K
! 2658 |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgc|gct|aaa|

```

37/128

2007211861 21 Aug 2007

```

!           |AflIII |           | PstI |
!
!           .....CDR3.....|----FR4-----
!           121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
5 !           D   Y   E   G   T   G   Y   A   F   D   I   W   G   Q   G
2703 |gac|tat|gaa|ggt|act|ggt|tat|gct|ttc|gaC|ATA|TGg|ggt|caa|ggt|
!                                     | NdeI |(1/4)

```

```

!           -----FR4----->|
!           136 137 138 139 140 141 142
!           T   M   V   T   V   S   S
0 2748 |act|atG|GTC|ACC|gtc|tct|agt
!           | BstEII |

```

```

! From BstEII onwards, pV323 is same as pCES1, except as noted.
5 ! BstEII sites may occur in light chains; not likely to be unique in final
! vector.

```

```

!           143 144 145 146 147 148 149 150 151 152
!           A   S   T   K   G   P   S   V   F   P
0 2769 gcc tcc acc aaG GGC CCa tcg GTC TTC ccc
!                                     Bsp120I.      BbsI...(2/2)
!                                     ApaI....

```

```

!           153 154 155 156 157 158 159 160 161 162 163 164 165 166 167
5 !           L   A   P   S   S   K   S   T   S   G   G   T   A   A   L
2799 ctg gca ccC TCC TCc aag agc acc tct ggg ggc aca gcg gcc ctg
!           BseRI...(2/2)

```

```

!           168 169 170 171 172 173 174 175 176 177 178 179 180 181 182
!           G   C   L   V   K   D   Y   F   P   E   P   V   T   V   S
) 2844 ggc tgc ctg GTC AAG GAC TAC TTC CCc gaA CCG GTg acg gtg tcg
!                                     AgeI....

```

```

!           183 184 185 186 187 188 189 190 191 192 193 194 195 196 197
!           W   N   S   G   A   L   T   S   G   V   H   T   F   P   A
5 2889 tgg aac tca GGC GCC ctg acc agc ggc gtc cac acc ttc ccg gct
!           KasI...(1/4)

```

```

!           198 199 200 201 202 203 204 205 206 207 208 209 210 211 212

```

2007211861 21 Aug 2007

38/128

! V L Q S S G L Y S L S S V V T  
2934 gtc cta cag tCt agc GGa ctc tac tcc ctc agc agc gta gtg acc  
! (Bsu36I...) (knocked out)  
!

5 ! 213 214 215 216 217 218 219 220 221 222 223 224 225 226 227  
! V P S S S L G T Q T Y I C N V  
2979 gtg ccC tCt tct agc tTG Ggc acc cag acc tac atc tgc aac gtg  
! (BstXI.....)N.B. destruction of BstXI & BpmI sites.  
!

10 ! 228 229 230 231 232 233 234 235 236 237 238 239 240 241 242  
! N H K P S N T K V D K K V E P  
3024 aat cac aag ccc agc aac acc aag gtg gac aag aaa gtt gag ccc  
!

15 ! 243 244 245  
! K S C A A A H H H H H H S A  
3069 aaa tct tgt GCG GCC Gct cat cac cac cat cat cac tct gct  
! NotI.....  
!

20 ! E Q K L I S E E D L N G A A  
3111 gaa caa aaa ctc atc tca gaa gag gat ctg aat ggt gcc gca  
!

! D I N D D R M A S G A  
3153 GAT ATC aac gat gat cgt atg gct AGC ggc gcc  
25 ! rEK cleavage\_site..... NheI... KasI...  
! EcoRV..  
!

! Domain 1 -----

30 ! A E T V E S C L A  
3183 gct gaa act gtt gaa agt tgt tta gca  
!

! K P H T E I S F  
3210 aaa ccc cat aca gaa aat tca ttt  
!

35 ! T N V W K D D K T  
3234 aCT AAC GTC TGG AAA GAC GAC AAA ACT  
!

! L D R Y A N Y E G C L W N A T G V

21 Aug 2007

2007211861

3261 tta gat cgt tac gct aac tat gag ggt tgt ctg tgG AAT Gct aca ggc gtt  
! BsmI\_\_\_\_\_

5 V V C T G D E T Q C Y G T W V P I  
3312 gta gtt tgt act ggt GAC GAA ACT CAG TGT TAC GGT ACA TGG GTT cct att

G L A I P E N  
3363 ggg ctt gct atc cct gaa aat

10 ! L1 linker -----

E G G G S E G G G S  
3384 gag ggt ggt ggc tct gag ggt ggc ggt tct

15 E G G G S E G G G T  
3414 gag ggt ggc ggt tct gag ggt ggc ggt act

! Domain 2 -----

20 3444 aaa cct cct gag tac ggt gat aca cct att ccg ggc tat act tat atc aac  
3495 cct ctc gac ggc act tat ccg cct ggt act gag caa aac ccc gct aat cct  
3546 aat cct tct ctt GAG GAG tct cag cct ctt aat act ttc atg ttt cag aat

! BseRI\_\_\_\_\_

25 3597 aat agg ttc cga aat agg cag ggg gca tta act gtt tat acg ggc act  
3645 gtt act caa ggc act gac ccc gtt aaa act tat tac cag tac act cct  
3693 gta tca tca aaa gcc atg tat gac gct tac tgg aac ggt aaa ttC AGA

! AlwNI

3741 GAC TGc gct ttc cat tct ggc ttt aat gaa gat cca ttc gtt tgt gaa

! AlwNI

3789 tat caa ggc caa tcg tct gac ctg cct caa cct cct gtc aat gct

30 3834 ggc ggc ggc tct

! start L2 -----

3846 ggt ggt ggt tct  
3858 ggt ggc ggc tct  
3870 gag ggt ggt ggc tct gag ggt ggc ggt tct  
15 3900 gag ggt ggc ggc tct gag gga ggc ggt tcc  
3930 ggt ggt ggc tct ggt ! end L2

! Domain 3 -----

S G D F D Y E K M A N A N K G A

21 Aug 2007

2007211861

```

3945 tcc ggt gat ttt gat tat gaa aag atg gca aac gct aat aag ggg gct
!
!   M   T   E   N   A   D   E   N   A   L   Q   S   D   A   K   G
3993 atg acc gaa aat gcc gat gaa aac gcg cta cag tct gac gct aaa ggc
5 !
!   K   L   D   S   V   A   T   D   Y   G   A   A   I   D   G   F
4041 aaa ctt gat tct gtc gct act gat tac ggt gct gct atc gat ggt_ttc
!
!   I   G   D   V   S   G   L   A   N   G   N   G   A   T   G   D
10 4089 att ggt gac gtt tcc ggc ctt gct aat ggt aat ggt gct act ggt gat
!
!   F   A   G   S   N   S   Q   M   A   Q   V   G   D   G   D   N
4137 ttt gct ggc tct aat tcc caa atg gct caa gtc ggt gac ggt gat aat
!
15 !   S   P   L   M   N   N   F   R   Q   Y   L   P   S   L   P   Q
4185 tca cct tta atg aat aat ttc cgt caa tat tta cct tcc ctc cct caa
!
!   S   V   E   C   R   P   F   V   F   S   A   G   K   P   Y   E
20 4233 tcg gtt gaa tgt cgc cct ttt gtc ttt agc gct ggt aaa cca tat gaa
!
!   F   S   I   D   C   D   K   I   N   L   F   R
4281 ttt tct att gat tgt gac aaa ata aac tta ttc cgt
!
!                                     End Domain 3
!
15 !   G   V   F   A   F   L   L   Y   V   A   T   F   M   Y   V   F140
4317 ggt gtc ttt gcg ttt ctt tta tat gtt gcc acc ttt atg tat gta ttt
!
!   start transmembrane segment
!
!   S   T   F   A   N   I   L
10 4365 tct acg ttt gct aac ata ctg
!
!   R   N   K   E   S
4386 cgt aat aag gag tct TAA ! stop of iii
!
!   Intracellular anchor.
15 !
!
!   M1 P2 V L L5 G I P L L10 L R F L G15
4404 tc ATG cca gtt ctt ttg ggt att ccg tta tta ttg cgt ttc ctc ggt
!
!   Start VI
!
```

21 Aug 2007

2007211861

```

4451 ttc ctt ctg gta act ttg ttc ggc tat ctg ctt act ttt ctt aaa aag
4499 ggc ttc ggt aag ata gct att gct att tca ttg ttt ctt gct ctt att
4547 att ggg ctt aac tca att ctt gtg ggt tat ctc tct gat att agc gct
4595 caa tta ccc tct gac ttt gtt cag ggt gtt cag tta att ctc ccg tct
5 4643 aat gcg ctt ccc tgt ttt tat gtt att ctc tct gta aag gct gct att
4691 ttc att ttt gac gtt aaa caa aaa atc gtt tct tat ttg gat tgg gat
!
!           M1  A2  V3      F5                L10      G13
4739 aaa TAA t ATG gct gtt tat ttt gta act ggc aaa tta ggc tct gga
10 !       end VI   Start gene I
!
!       14  15  16  17  18  19  20  21  22  23  24  25  26  27  28
!       K  T  L  V  S  V  G  K  I  Q  D  K  I  V  A
4785 aag acg ctc gtt agc gtt ggt aag att cag gat aaa att gta gct
15 !
!       29  30  31  32  33  34  35  36  37  38  39  40  41  42  43
!       G  C  K  I  A  T  N  L  D  L  R  L  Q  N  L
4830 ggg tgc aaa ata gca act aat ctt gat tta agg ctt caa aac ctc
!
!       44  45  46  47  48  49  50  51  52  53  54  55  56  57  58
!       P  Q  V  G  R  F  A  K  T  P  R  V  L  R  I
4875 ccg caa gtc ggg agg ttc gct aaa acg cct cgc gtt ctt aga ata
!
!       59  60  61  62  63  64  65  66  67  68  69  70  71  72  73
25 !       P  D  K  P  S  I  S  D  L  L  A  I  G  R  G
4920 ccg gat aag cct tct ata tct gat ttg ctt gct att ggg cgc ggt
!
!       74  75  76  77  78  79  80  81  82  83  84  85  86  87  88
!       N  D  S  Y  D  E  N  K  N  G  L  L  V  L  D
30 4965 aat gat tcc tac gat gaa aat aaa aac ggc ttg ctt gtt ctc gat
!
!       89  90  91  92  93  94  95  96  97  98  99 100 101 102 103
!       E  C  G  T  W  F  N  T  R  S  W  N  D  K  E
5010 gag tgc ggt act tgg ttt aat acc cgt tct tgg aat gat aag gaa
35 !
!       104 105 106 107 108 109 110 111 112 113 114 115 116 117 118
!       R  Q  P  I  I  D  W  F  L  H  A  R  K  L  G
5055 aga cag ccg att att gat tgg ttt cta cat gct cgt aaa tta gga
!

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2007211861 21 Aug 2007

42/128

! 119 120 121 122 123 124 125 126 127 128 129 130 131 132 133  
! W D I I F L V Q D L S I V D K  
5100 tgg gat att att ttt ctt gtt cag gac tta tct att gtt gat aaa  
!  
5 ! 134 135 136 137 138 139 140 141 142 143 144 145 146 147 148  
! Q A R S A L A E H V V Y C R R  
5145 cag gcg cgt tct gca tta gct gaa cat gtt gtt tat tgt cgt cgt  
!  
10 ! 149 150 151 152 153 154 155 156 157 158 159 160 161 162 163  
! L D R I T L P F V G T L Y S L  
5190 ctg gac aga att act tta cct ttt gtc ggt act tta tat tct ctt  
!  
15 ! 164 165 166 167 168 169 170 171 172 173 174 175 176 177 178  
! I T G S K M P L P K L H V G V  
5235 att act ggc tcg aaa atg cct ctg cct aaa tta cat gtt ggc gtt  
!  
20 ! 179 180 181 182 183 184 185 186 187 188 189 190 191 192 193  
! V K Y G D S Q L S P T V E R W  
5280 gtt aaa tat ggc gat tct caa tta agc cct act gtt gag cgt tgg  
!  
25 ! 194 195 196 197 198 199 200 201 202 203 204 205 206 207 208  
! L Y T G K N L Y N A Y D T K Q  
5325 ctt tat act ggt aag aat ttg tat aac gca tat gat act aaa cag  
!  
30 ! 209 210 211 212 213 214 215 216 217 218 219 220 221 222 223  
! A F S S N Y D S G V Y S Y L T  
5370 gct ttt tct agt aat tat gat tcc ggt gtt tat tct tat tta acg  
!  
35 ! 224 225 226 227 228 229 230 231 232 233 234 235 236 237 238  
! P Y L S H G R Y F K P L N L G  
5415 cct tat tta tca cac ggt cgg tat ttc aaa cca tta aat tta ggt  
!  
! 239 240 241 242 243 244 245 246 247 248 249 250 251 252 253  
! Q K M K L T K I Y L K K F S R  
5460 cag aag atg aaa tta act aaa ata tat ttg aaa aag ttt tct cgc  
!  
! 254 255 256 257 258 259 260 261 262 263 264 265 266 267 268  
! V L C L A I G F A S A F T Y S  
5505 gtt ctt tgt ctt gcg att gga ttt gca tca gca ttt aca tat agt



2007211861 21 Aug 2007

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!
!   269 270 271 272 273 274 275 276 277 278 279 280 281 282 283
!   Y   I   T   Q   P   K   P   E   V   K   K   V   V   S   Q
5550 tat ata acc caa cct aag ccg gag gtt aaa aag gta gtc tct cag
5
!
!   284 285 286 287 288 289 290 291 292 293 294 295 296 297 298
!   T   Y   D   F   D   K   F   T   I   D   S   S   Q   R   L
5595 acc tat gat ttt gat aaa ttc act att gac tct tct cag cgt ctt
!
!
10 !   299 300 301 302 303 304 305 306 307 308 309 310 311 312 313
!   N   L   S   Y   R   Y   V   F   K   D   S   K   G   K   L
5640 aat cta agc tat cgc tat gtt ttc aag gat tct aag gga aaa TTA
!
!
!
15 !
!   314 315 316 317 318 319 320 321 322 323 324 325 326 327 328
!   I   N   S   D   D   L   Q   K   Q   G   Y   S   L   T   Y
5685 ATT AAt agc gac gat tta cag aag caa ggt tat tca ctc aca tat
!
!   PacI
!
!
20 !
!   329 330 331 332 333 334 335 336 337 338 339 340 341 342 343
!   i   I   D   L   C   T   V   S   I   K   K   G   N   S   N   E
!   iv
!
5730 att gat tta tgt act gtt tcc att aaa aaa ggt aat tca aAT Gaa
!
!
!
25 !
!
!   344 345 346 347 348 349
!   i   I   V   K   C   N   .End of I
!   iv   L3 L   N5 V   I7 N   F   V10
5775 att gtt aaa tgt aat TAA T TTT GTT
!
30 !   IV continued.....
5800 ttc ttg atg ttt gtt tca tca tct tct ttt gct cag gta att gaa atg
5848 aat aat tcg cct ctg cgc gat ttt gta act tgg tat tca aag caa tca
5896 ggc gaa tcc gtt att gtt tct ccc gat gta aaa ggt act gtt act gta
5944 tat tca tct gac gtt aaa cct gaa aat cta cgc aat ttc ttt att tct
35 5992 gtt tta cgt gct aat aat ttt gat atg gtt ggt tca att cct tcc ata
6040 att cag aag tat aat cca. aac aat cag gat tat att gat gaa ttg cca
6088 tca tct gat aat cag gaa tat gat gat aat tcc gct cct tct ggt ggt
6136 ttc ttt gtt ccg caa aat gat aat gtt act caa act ttt aaa att aat
6184 aac gtt cgg gca aag gat tta ata cga gtt gtc gaa ttg ttt gta aag

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7660 gataaatgct tcaataatat tgaaaaAGGA AGAgT

RBS.?...

Start bla gene

7695 ATG agt att caa cat ttc cgt gtc gcc ctt att ccc ttt ttt gcg gca ttt

7746 tgc ctt cct gtt ttt gct cac cca gaa acg ctg gtg aaa gta aaa gat gct

7797 gaa gat cag ttg ggC gCA CGA Gtg ggt tac atc gaa ctg gat ctc aac agc

BssSI...

ApaLI removed

7848 ggt aag atc ctt gag agt ttt cgc ccc gaa gaa cgt ttt cca atg atg agc

7899 act ttt aaa gtt ctg cta tgt cat aca cta tta tcc cgt att gac gcc ggg

7950 caa gaG CAA CTC GGT CGc cgg gcg cgg tat tct cag aat gac ttg gtt gAG

BcgI\_\_\_\_\_

ScaI

8001 TAC Tca cca gtc aca gaa aag cat ctt acg gat ggc atg aca gta aga gaa

ScaI\_

8052 tta tgc agt gct gcc ata acc atg agt gat aac act gcg gcc aac tta ctt

8103 ctg aca aCG ATC Gga gga ccg aag gag cta acc gct ttt ttg cac aac atg

PvuI\_\_\_\_\_

8154 ggg gat cat gta act cgc ctt gat cgt tgg gaa ccg gag ctg aat gaa gcc

8205 ata cca aac gac gag cgt gac acc acg atg cct gta gca atg cca aca acg

8256 tTG CGC Aaa cta tta act ggc gaa cta ctt act cta gct tcc cgg caa caa

FspI....

8307 tta ata gac tgg atg gag gcg gat aaa gtt gca gga cca ctt ctg cgc tgc

8358 GCC ctt ccG Gct ggc tgg ttt att gct gat aaa tct gga gcc ggt gag cgt

BglI\_\_\_\_\_

8409 gGG TCT Cgc ggt atc att gca gca ctg ggg cca gat ggt aag ccc tcc cgt

BsaI\_\_\_\_\_

8460 atc gta gtt atc tac acG ACg ggg aGT Cag gca act atg gat gaa cga aat

AhdI\_\_\_\_\_

8511 aga cag atc gct gag ata ggt gcc tca ctg att aag cat tgg TAA ctgt

stop

8560 cagaccaagt ttactcatat atactttaga ttgatttaaa acttcatttt taatttataaa

8620 ggatctaggt gaagatcctt tttgataatc tcatgaccaa aatcccttaa cgtgagtttt

8680 cgttccactg tacgtaagac cccc

35 8704 AAGCTT GTCGAC tgaa tggcgaatgg cgctttgcct

HindIII SalI..

(2/2) HincII

8740 ggtttccggc accagaagcg gtgccggaaa gctggetgga gtgcgatctt

8790 CCTGAGG

! Bsu36I\_

8797 cccgat actgtcgtcg tcccctcaaa ctggcagatg

8832 cacggttacg atgcgcccac ctacaccaac gtaacctatc ccattacggt caatccgccc

5 8892 tttgttccca cggagaatcc gacgggttgt tactcgctca catttaatgt tgatgaaagc

8952 tggctacagg aaggccagac gcgaattatt tttgatggcg ttcctattgg ttaaaaaatg

9012 agctgattta acaaaaattt aacgcgaatt ttaacaaaat attaacgttt acaATTTAAA

! SwaI...

9072 Tatttgctta tacaatcttc ctgtttttgg ggcttttctg attatcaacc GGGGTAcac

10 ! RBS?

9131 ATG att gac atg cta gtt tta cga tta ccg ttc atc gat tct ctt gtt tgc

! Start gene II

9182 tcc aga ctc tca ggc aat gac ctg ata gcc ttt gtA GAT CTc tca aaa ata

! BglII...

15 9233 gct acc ctc tcc ggc atg aat tta tca gct aga acg gtt gaa tat cat att

9284 gat ggt gat ttg act gtc tcc ggc ctt tct cac cct ttt gaa tct tta cct

9335 aca cat tac tca ggc att gca ttt aaa ata tat gag ggt tct aaa aat ttt

9386 tat cct tgc gtt gaa ata aag gct tct ccc gca aaa gta tta cag ggt cat

9437 aat gtt ttt ggt aca acc gat tta gct tta tgc tct gag gct tta ttg ctt

20 9488 aat ttt gct aat tct ttg cct tgc ctg tat gat tta ttg gat gtt ! 9532

! gene II continues

2007211861 21 Aug 2007

Table 120B: Sequence of MALIA3, condensed

LOCUS	MALIA3	9532	CIRCULAR				
ORIGIN							
	1	AATGCTACTA	CTATTAGTAG	AATTGATGCC	ACCTTTTCAG	CTCGCGCCCC	AAATGAAAAT
5	61	ATAGCTAAAC	AGGTTATTGA	CCATTTGCGA	AATGTATCTA	ATGGTCAAAC	TAAATCTACT
	121	CGTTCGCAGA	ATTGGGAATC	AACTGTTACA	TGGAATGAAA	CTCCAGACA	CCGTACTTTA
	181	GTTGCATATT	TAAAACATGT	TGAGCTACAG	CACCAGATTC	AGCAATTAAG	CTCTAAGCCA
	241	TCCGCAAAAA	TGACCTCTTA	TCAAAGGAG	CAATTAAAGG	TACTCTCTAA	TCCTGACCTG
10	301	TTGGAGTTTG	CTTCCGGTCT	GGTTCGCTTT	GAAGCTCGAA	TTAAAACGCG	ATATTTGAAG
	361	TCTTTCGGGC	TTCCTCTTAA	TCTTTTTGAT	GCAATCCGCT	TTGCTTCTGA	CTATAATAGT
	421	CAGGGTAAAG	ACCTGATTTT	TGATTTATGG	TCATTCTCGT	TTTCTGAACT	GTTTAAAGCA
	481	TTTGAGGGGG	ATTCAATGAA	TATTTATGAC	GATTCGCGAG	TATTGGACGC	TATCCAGTCT
	541	AAACATTTTA	CTATTACCCC	CTCTGGCAA	ACTTCTTTTG	CAAAAGCCTC	TCGCTATTTT
15	601	GGTTTTTATC	GTCGTCTGGT	AAACGAGGGT	TATGATAGTG	TTGCTCTTAC	TATGCCTCGT
	661	AATTCCTTTT	GGCGTTATGT	ATCTGCATTA	GTTGAATGTG	GTATTCCTAA	ATCTCAACTG
	721	ATGAATCTTT	CTACCTGTAA	TAATGTTGTT	CCGTTAGTTC	GTTTTATTAA	CGTAGATTTT
	781	TCTTCCCAAC	GTCCTGACTG	GTATAATGAG	CCAGTTCTTA	AAATCGCATA	AGGTAATTCA
	841	CAATGATTAA	AGTTGAAATT	AAACCATCTC	AAGCCCAATT	TACTACTCGT	TCTGGTGTTT
	901	CTCGTCAGGG	CAAGCCTTAT	TCACTGAATG	AGCAGCTTTG	TTACGTTGAT	TTGGGTAATG
20	961	AATATCCGGT	TCTTGTC AAG	ATTACTCTTG	ATGAAGGTCA	GCCAGCCTAT	GCGCCTGGTC
	1021	TGTACACCGT	TCATCTGTCC	TCTTTC A AAG	TTGGTCAGTT	CGGTTCCCTT	ATGATTGACC
	1081	GTCTGCGCCT	CGTTCGGGCT	AAGTAACATG	GAGCAGGTCG	CGGATTTCGA	CACAATTTAT
	1141	CAGGCGATGA	TACAAATCTC	CGTTGACTTT	TGTTTTCGCGC	TTGGTATAAT	CGCTGGGGGT
	1201	CAAAGATGAG	TGTTTTAGTG	TATTCTTTTCG	CCTCTTTTCGT	TTTAGGTTGG	TGCCTTCGTA
25	1261	GTGGCATTAC	GTATTTTACC	CGTTTAATGG	AAACTTCCTC	ATGAAAAAGT	CTTTAGTCTT
	1321	CAAAGCCTCT	GTAGCCGTTG	CTACCCTCGT	TCCGATGCTG	TCTTTTCGCTG	CTGAGGGTGA
	1381	CGATCCCGCA	AAAGCGGCCT	TTAACTCCCT	GCAAGCCTCA	GCGACCGAAT	ATATCGGTTA
	1441	TGCGTGGGCG	ATGGTTGTTG	TCATTGTCCG	CGCAACTATC	GGTATCAAGC	TGTTTAAGAA
	1501	ATTCACCTCG	AAAGCAAGCT	GATAAACCGA	TACAATTAAA	GGCTCCTTTT	GGAGCCTTTT
30	1561	TTTTTGGAGA	TTTTCAACGT	GAAAAAATTA	TTATTCGCAA	TTCCTTTAGT	TGTTCCTTTC
	1621	TATTCTCACA	GTGCACAGTC	TGTCGTGACG	CAGCCGCCCT	CAGTGTCTGG	GGCCCCAGGG
	1681	CAGAGGGTCA	CCATCTCCTG	CACTGGGAGC	AGCTCCAACA	TCGGGGCAGG	TTATGATGTA
	1741	CACTGGTACC	AGCAGCTTCC	AGGAACAGCC	CCCAAACCTC	TCATCTATGG	TAACAGCAAT
	1801	CGGCCCTCAG	GGGTCCCTGA	CCGATTCTCT	GGCTCCAAGT	CTGGCACCTC	AGCCTCCCTG
35	1861	GCCATCACTG	GGCTCCAGGC	TGAGGATGAG	GCTGATTATT	ACTGCCAGTC	CTATGACAGC
	1921	AGCCTGAGTG	GCCTTTATGT	CTTCGGAACT	GGGACCAAGG	TCACCGTCCT	AGGTCAGCCC
	1981	AAGGCCAACC	CCACTGTAC	TCTGTTCCCG	CCCTCCTCTG	AGGAGCTCCA	AGCCAACAAG
	2041	GCCACACTAG	TGTGTCTGAT	CAGTGACTTC	TACCCGGGAG	CTGTGACAGT	GGCCTGGAAG
	2101	GCAGATAGCA	GCCCCGTCAA	GGCGGGAGTG	GAGACCACCA	CACCCTCAA	ACAAAGCAAC

21 Aug 2007

2007211861

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2161 AACAAAGTACG CGGCCAGCAG CTATCTGAGC CTGACGCCTG AGCAGTGGAA GTCCACAGA  
 2221 AGCTACAGCT GCCAGGTCAC GCATGAAGGG AGCACCGTGG AGAAGACAGT GGCCCTACA  
 2281 GAATGTTCAT AATAAACCGC CTCCACCGGG CGCGCCAATT CTATTTCAAG GAGACAGTCA  
 2341 TAATGAAATA CCTATTGCCT ACGGCAGCCG CTGGATTGTT ATTACTCGCG GCCCAGCCGG  
 2401 CCATGGCCGA AGTTCAATTG TTAGAGTCTG GTGGCGGTCT TGTTCAGCCT GGTGGTTCTT  
 2461 TACGTCTTTC TTGCGCTGCT TCCGGATTCA CTTTCTCTTC GTACGCTATG TCTTGGGTTT  
 2521 GCCAAGCTCC TGGTAAAGGT TTGGAGTGGG TTTCTGCTAT CTCTGGTTCT GGTGGCAGTA  
 2581 CTTACTATGC TGA CTCCGTT AAAGGTCGCT TCACTATCTC TAGAGACAAC TCTAAGAATA  
 2641 CTCTCTACTT GCAGATGAAC AGCTTAAGGG CTGAGGACAC TGCAGTCTAC TATTGCGCTA  
 2701 AAGACTATGA AGGTA CTGGT TATGCTTTCG ACATATGGGG TCAAGSTACT ATGGTCACCG  
 2761 TCTCTAGTGC CTCCACCAAG GGCCCATCGG TCTTCCCCCT GGCACCCTCC TCCAAGAGCA  
 2821 CCTCTGGGGG CACAGCGGCC CTGGGCTGCC TGGTCAAGGA CTA CTTCCCC GAACCGGTGA  
 2881 CGGTGTCGTG GAACTCAGGC GCCCTGACCA GCGGCGTCCA CACCTTCCCG GCTGTCTTAC  
 2941 AGTCTAGCGG ACTCTACTCC CTCAGCAGCG TAGTGACCGT GCCCTCTTCT AGCTTGGGCA  
 3001 CCCAGACCTA CATCTGCAAC GTGAATCACA AGCCCAGCAA CACCAAGGTG GACAAGAAA  
 3061 TTGAGCCCAA ATCTTGTGCG GCCGCTCATC ACCACCATCA TCACTCTGCT GAACAAAAAC  
 3121 TCATCTCAGA AGAGGATCTG AATGGTGCCG CAGATATCAA CGATGATCGT ATGGCTGGCG  
 3181 CCGCTGAAAC TGTTGAAAGT TGTTTAGCAA AACCCCATAC AGAAAATTCA TTTACTAACG  
 3241 TCTGGAAAGA CGACAAA ACT TTAGATCGTT ACGCTAACTA TGAGGGTTGT CTGTGGAATG  
 3301 CTACAGGCGT TGTA GTTTGT ACTGGTGACG AA ACTCAGTG TTACGGTACA TGGGTTCCCTA  
 3361 TTGGGCTTGC TATCCCTGAA AATGAGGGTG GTGGCTCTGA GGGTGGCGGT TCTGAGGGTG  
 3421 GCGGTTCTGA GGGTGGCGGT ACTAAACCTC CTGAGTACGG TGATACACCT ATTCCGGGCT  
 3481 ATACTTATAT CAACCCCTCTC GACGGCACTT ATCCGCTGG TACTGAGCAA AACCCGCTA  
 3541 ATCCTAATCC TTCTCTTGAG GAGTCTCAGC CTCTTAATAC TTTCATGTTT CAGAATAATA  
 3601 GGTTCCGAAA TAGGCAGGGG GCATTA ACTG TTTATACGGG CACTGTTACT CAAGGCACTG  
 3661 ACCCCGTTAA AACTTATTAC CAGTACACTC CTGTATCATC AAAAGCCATG TATGACGCTT  
 3721 ACTGGAACGG TAAATT CAGA GACTGCGCTT TCCATTCTGG CTTTAATGAA GATCCATT CG  
 3781 TTTGTGAATA TCAAGGCCAA TCGTCTGACC TGCCTCAACC TCCTGTCAAT GCTGGCGGGC  
 3841 GCTCTGGTGG TGGTCTGTT GCGGCTCTG AGGGTGGTGG CTCTGAGGGT GCGGTTCTG  
 3901 AGGGTGGCGG CTCTGAGGGA GCGGTTCCG GTGGTGGCTC TGGTCCGGT GATTTTGATT  
 3961 ATGAAAAGAT GGCAAACGCT AATAAGGGGG CTATGACCGA AAATGCCGAT GAAAACGCGC  
 4021 TACAGTCTGA CGCTAAAGGC AA ACTTGATT CTGTGCTAC TGATTACGGT GCTGCTATCG  
 4081 ATGGTTTCAT TGGTGACGTT TCCGGCCTTG CTAATGGTAA TGGTGTACT GGTGATTTTG  
 4141 CTGGCTCTAA TTCCCAAATG GCTCAAGTCG GTGACGGTGA TAATTACCT TTAATGAATA  
 4201 ATTTCCGTCA ATATTTACCT TCCCTCCCTC AATCGGTTGA ATGTCGCCCT TTTGTCTTTA  
 4261 GCGCTGGTAA ACCATATGAA TTTTCTATTG ATTGTGACAA AATAAACTTA TTCCGTGGTG  
 4321 TCTTTGCGTT TCTTTTATAT GTTGCCACCT TTATGTATGT ATTTTCTACG TTTGCTAACA  
 4381 TACTGCGTAA TAAGGAGTCT TAATCATGCC AGTTCTTTTG GGTATTCCGT TATTATTGCG  
 4441 TTTCCCTCGGT TTCCTTCTGG TAACTTTGTT CGGCTATCTG CTTACTTTTC TTA AAAAGGG

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4501 CTTGCGTAAG ATAGCTATTG CTATTTTCATT GTTTCTTGCT CTTATTATTG GGCTTAACTC  
 4561 AATTCTTGTG GGTTATCTCT CTGATATTAG CGCTCAATTA CCCTCTGACT TTGTTGAGGG  
 4621 TGTTGAGTTA ATTCTCCCCT CTAAATGCGCT TCCCTGTTTT TATGTTATTG TCTCTGTAAA  
 4681 GGCTGCTATT TTCATTTTTG ACGTTAAACA AAAAATCGTT TCTTATTGG ATTGGGATAA  
 4741 ATAATATGGC TGTTTATTTT GTAACGGCA AATTAGGCTC TGGAAAGACG CTCGTTAGCG  
 4801 TTGGTAAGAT TCAGGATAAA ATTGTAGCTG GGTGCAAAAT AGCAACTAAT CTTGATTTAA  
 4861 GGCTTCAAAA CCTCCCACAA GTCGGGAGGT TCGCTAAAAC GCCTCGCGTT CTTAGAATAC  
 4921 CGGATAAGCC TTCTATATCT GATTTGCTTG CTATTGGGCG CGGTAATGAT TCCTACGATG  
 4981 AAAATAAAAA CGGCTTGCTT GTTCTCGATG AGTGCGGTAC TTGGTTTAAAT ACCCGTTCTT  
 5041 GGAATGATAA GGAAAGACAG CCGATTATTG ATTGGTTTCT ACATGCTCGT AAATTAGGAT  
 5101 GGGATATTAT TTTTCTTGTT CAGGACTTAT CTATTGTTGA TAAACAGGCG CGTTCTGCAT  
 5161 TAGCTGAACA TGTTGTTTAT TGTCGTCGTC TGGACAGAAT TACTTTACCT TTTGTCGGTA  
 5221 CTTTATATTC TCTTATTACT GGCTCGAAAA TGCCTCTGCC TAAATTACAT GTTGGCGTTG  
 5281 TTAAATATGG CGATTCTCAA TTAAGCCCTA CTGTTGAGCG TTGGCTTTAT ACTGGTAAGA  
 5341 ATTTGTATAA CGCATATGAT ACTAAACAGG CTTTTTCTAG TAATTATGAT TCCGGTGT  
 5401 ATTCTTATTT AACGCCTTAT TTATCACACG GTCGGTATTT CAAACCATTA AATTTAGGTC  
 5461 AGAAGATGAA ATTAACATAA ATATATTTGA AAAAGTTTTT TCGCGTTCTT TGTCTTGCGA  
 5521 TTGGATTGTC ATCAGCATT ACATATAGTT ATATAACCCA ACCTAAGCCG GAGGTTAAAA  
 5581 AGGTAGTCTC TCAGACCTAT GATTTTGATA AATTCACTAT TGAATCTCT CAGCGTCTTA  
 5641 ATCTAAGCTA TCGCTATGTT TTCAAGGATT CTAAGGGAAA ATTAATTAAT AGCGACGATT  
 5701 TACAGAAGCA AGGTTATTCA CTCACATATA TTGATTTATG TACTGTTTCC ATTAATAAAG  
 5761 GTAATTCAAA TGAAATTGTT AAATGTAATT AATTTTGTTT TCTTGATGTT TGTTTCATCA  
 5821 TCTTCTTTTG CTCAGGTAAT TGAAATGAAT AATTCGCCTC TCGCGGATT TGTAACCTGG  
 5881 TATTCAAAGC AATCAGGCGA ATCCGTTATT GTTTCTCCCG ATGTAAAAGG TACTGTTACT  
 5941 GTATATTCAT CTGACGTAA ACCTGAAAT CTACGCAAT TCTTTATTTT TGTTTTACGT  
 6001 GCTAATAAAT TTGATATGGT TGGTTCAATT CCTTCCATAA TTCAGAAGTA TAATCCAAAC  
 6061 AATCAGGATT ATATTGATGA ATTGCCATCA TCTGATAATC AGGAATATGA TGATAATTCC  
 6121 GCTCCTTCTG GTGGTTTCTT TGTTCCGCAA AATGATAATG TTAATCAAAC TTTTAAAT  
 6181 AATAACGTTT GGGCAAAGGA TTTAATACGA GTTGTGCAAT TGTTTGATAA GTCTAATACT  
 6241 TCTAAATCCT CAAATGTATT ATCTATTGAC GGCTCTAATC TATTAGTTGT TTCTGCACCT  
 6301 AAAGATATTT TAGATAACCT TCCTCAATTC CTTTCTACTG TTGATTTGCC AACTGACCAG  
 6361 ATATTGATTG AGGGTTTGAT ATTTGAGGTT CAGCAAGGTG ATGCTTTAGA TTTTTCATTT  
 6421 GCTGCTGGCT CTCAGCGTGG CACTGTTGCA GCGGGTGTTA ATACTGACCG CCTCACCTCT  
 6481 GTTTTATCTT CTGCTGGTGG TTCGTTCCGT ATTTTAAATG GCGATGTTTT AGGCTATCA  
 6541 GTTCGCGCAT TAAAGACTAA TAGCCATTCA AAAATATTGT CTGTGCCACG TATTCTTACG  
 6601 CTTTCAGGTC AGAAGGGTTC TATCTCTGTT GGCCAGAATG TCCCTTTTAT TACTGGTCTG  
 6661 TGACTGGTG AATCTGCCAA TGTAATAAAT CCATTTGAGA CGATTGAGCG TCAAATGTA  
 6721 GGTATTTCCTA TGAGCGTTTT TCCTGTTGCA ATGGCTGGCG GTAATATTGT TCTGGATATT  
 6781 ACCAGCAAGG CCGATAGTTT GAGTCTTCT ACTCAGGCAA GTGATGTTAT TACTAATCAA

21 Aug 2007

2007211861

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6841 AGAAGTATTG CTACAACGGT TAATTTGCGT GATGGACAGA CTCTTTTACT CGGTGGCCTC  
 6901 ACTGATTATA AAAACACTTC TCAAGATTCT GGCGTACCGT TCCTGTCTAA AATCCCTTTA  
 6961 ATCGGCCTCC TGTTTAGCTC CCGCTCTGAT TCCAACGAGG AAAGCACGTT ATACGTGCTC  
 7021 GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT AAGCGCGCGG GTGTGGTGGT  
 7081 TACGCGCAGC GTGACCGCTA CACTTGCCAG CGCCCTAGCG CCCGCTCCTT TCGCTTTCTT  
 7141 CCCTTCCTTT CTCGCCACGT TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC  
 7201 TTTAGGGTTC CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA  
 7261 TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTTGA CGTTGGAGTC  
 7321 CACGTTCTTT AATAGTGGAC TCTTGTTCCA AACTGGAACA AACTCAACC CTATCTCGGG  
 7381 CTATTCCTTT GATTTATAAG GGATTTTGCC GATTTCGGAA CCACCATCAA ACAGGATTTT  
 7441 CGCCTGCTGG GGCAAACCAG CGTGGACCGC TTGCTGCAAC TCTCTCAGGG CCAGGCGGTG  
 7501 AAGGGCAATC AGCTGTTGCC CGTCTCACTG GTGAAAAGAA AAACCACCCT GGATCCAAGC  
 7561 TTGCAGGTGG CACTTTTCGG GGAAATGTGC GCGGAACCCC TATTTGTTTA TTTTCTAAA  
 7621 TACATTCAA TATGTATCCG CTCATGAGAC AATAACCCTG ATAAATGCTT CAATAATATT  
 7681 GAAAAAGGAA GAGTATGAGT ATCAACATT TCCGTGTCGC CTTATTCCC TTTTTGCGG  
 7741 CATTTGCCT TCCTGTTTTT GCTCACCCAG AAACGCTGGT GAAAGTAAA GATGCTGAAG  
 7801 ATCAGTTGGG CGCACGAGTG GGTTACATCG AACTGGATCT CAACAGCGGT AAGATCCTTG  
 7861 AGAGTTTTCG CCCCAGAGAA CGTTTTCCAA TGATGAGCAC TTTTAAAGTT CTGCTATGTC  
 7921 ATACACTATT ATCCCGTATT GACGCCGGGC AAGAGCAACT CGGTGCGCGG GCGCGGTATT  
 7981 CTCAGAATGA CTTGGTTGAG TACTCACCAG TCACAGAAAA GCATCTTACG GATGGCATGA  
 8041 CAGTAAGAGA ATTATGCAGT GCTGCCATAA CCATGAGTGA TAACACTGCG GCCAACTTAC  
 8101 TTCTGACAAC GATCGGAGGA CCGAAGGAGC TAACCGCTTT TTTGCACAAC ATGGGGGATC  
 8161 ATGTAACCTG CCTTGATCGT TGGGAACCGG AGCTGAATGA AGCCATACCA AACGACGAGC  
 8221 GTGACACCAC GATGCCTGTA GCAATGCCAA CAACGTTGCG CAACTATTA ACTGGCGAAC  
 8281 TACTTACTCT AGCTTCCCGG CAACAATTA TAGACTGGAT GGAGGCGGAT AAAGTTGACG  
 8341 GACCACTTCT GCGCTCGGCC CTTCCGGCTG GCTGGTTTAT TGCTGATAAA TCTGGAGCCG  
 8401 GTGAGCGTGG GTCTCGCGGT ATCATTGCAG CACTGGGGCC AGATGGTAAG CCCTCCCGTA  
 8461 TCGTAGTTAT CTACACGACG GGGAGTCAGG CAACTATGGA TGAACGAAAT AGACAGATCG  
 8521 CTGAGATAGG TGCCCTACTG ATTAAGCATT GGTAACGTC AGACCAAGTT TACTCATATA  
 8581 TACTTTAGAT TGATTTAAA CTTCAATTTT AATTTAAAAG GATCTAGGTG AAGATCCTTT  
 8641 TTGATAATCT CATGACCAA ATCCCTTAAC GTGAGTTTTT GTTCCACTGT ACGTAAGACC  
 8701 CCAAAGCTTG TCGACTGAAT GGCGAATGGC GCTTTGCCTG GTTCCGGCA CCAGAAGCGG  
 8761 TGCCGGAAAG CTGGCTGGAG TGCATCTTC CTGAGGCCGA TACTGTCGTC GTCCCCTCAA  
 8821 ACTGGCAGAT GCACGGTTAC GATGCGCCA TCTACACCAA CGTAACCTAT CCCATTACGG  
 8881 TCAATCCGCC GTTTGTTCCC ACGGAGAATC CGACGGGTTG TTA CTGCTC ACATTTAATG  
 8941 TTGATGAAAG CTGGCTACAG GAAGGCCAGA CGCGAATTAT TTTTGATGGC GTTCTATTG  
 9001 GTTAAAAAAT GAGCTGATTT AACAAAAATT TAACGCGAAT TTTAACAAA TATTAACGTT  
 9061 TACAATTTAA ATATTTGCTT ATACAATCTT CCTGTTTTTG GGGCTTTTCT GATTATCAAC  
 9121 CGGGGTACAT ATGATTGACA TGCTAGTTTT ACGATTACCG TTCATCGATT CTCTTGTTTG



2007211861 21 Aug 2007

5

51/128

9181 CTCCAGACTC TCAGGCAATG ACCTGATAGC CTTTGTAGAT CTCTCAAAAA TAGCTACCCT  
9241 CTCCGGCATG AATTTATCAG CTAGAACGGT TGAATATCAT ATTGATGGTG ATTTGACTGT  
9301 CTCCGGCCTT TCTCACCCCTT TTGAATCTTT ACCTACACAT TACTCAGGCA TTGCATTTAA  
9361 AATATATGAG GGTCTAAAA ATTTTTATCC TTGCGTTGAA ATAAAGGCTT CTCCCGCAAA  
9421 AGTATTACAG GGCATAATG TTTTTGGTAC AACCGATTTA GCTTTATGCT CTGAGGCTTT  
9481 ATTGCTTAAT TTTGCTAATT CTTTGCCTTG CCTGTATGAT TTATTGGATG TT

2007211861 21 Aug 2007

52/128

Table 200: Enzymes that either cut 15 or more human GLGs or have 5+-base recognition in FR3

Typical entry:

Rename	Recognition	#sites
GLGid#:base#	GLGid#:base#	GLGid#:base#.....

5

---

BstEII Ggtnacc 2  
 1: 3 48: 3  
 There are 2 hits at base# 3

10

MaeIII gtnac 36  
 1: 4 2: 4 3: 4 4: 4 5: 4 6: 4  
 7: 4 8: 4 9: 4 10: 4 11: 4 37: 4  
 37: 58 38: 4 38: 58 39: 4 39: 58 40: 4  
 40: 58 41: 4 41: 58 42: 4 42: 58 43: 4  
 43: 58 44: 4 44: 58 45: 4 45: 58 46: 4  
 46: 58 47: 4 47: 58 48: 4 49: 4 50: 58  
 There are 24 hits at base# 4

15

Tsp45I gtsac 33  
 1: 4 2: 4 3: 4 4: 4 5: 4 6: 4  
 7: 4 8: 4 9: 4 10: 4 11: 4 37: 4  
 37: 58 38: 4 38: 58 39: 58 40: 4 40: 58  
 41: 58 42: 58 43: 4 43: 58 44: 4 44: 58  
 45: 4 45: 58 46: 4 46: 58 47: 4 47: 58  
 48: 4 49: 4 50: 58  
 There are 21 hits at base# 4

20

25

HphI tcacc 45  
 1: 5 2: 5 3: 5 4: 5 5: 5 6: 5  
 7: 5 8: 5 11: 5 12: 5 12: 11 13: 5  
 14: 5 15: 5 16: 5 17: 5 18: 5 19: 5  
 20: 5 21: 5 22: 5 23: 5 24: 5 25: 5  
 26: 5 27: 5 28: 5 29: 5 30: 5 31: 5  
 32: 5 33: 5 34: 5 35: 5 36: 5 37: 5  
 38: 5 40: 5 43: 5 44: 5 45: 5 46: 5  
 47: 5 48: 5 49: 5  
 There are 44 hits at base# 5

30

35

2007211861 21 Aug 2007

53/128

NlaIII CATG 26  
1: 9 1: 42 2: 42 3: 9 3: 42 4: 9  
4: 42 5: 9 5: 42 6: 42 6: 78 7: 9  
7: 42 8: 21 8: 42 9: 42 10: 42 11: 42  
5 12: 57 13: 48 13: 57 14: 57 31: 72 38: 9  
48: 78 49: 78

There are 11 hits at base# 42

There are 1 hits at base# 48 Could cause raggedness.

10 BsaJI Ccngg 37  
1: 14 2: 14 5: 14 6: 14 7: 14 8: 14  
8: 65 9: 14 10: 14 11: 14 12: 14 13: 14  
14: 14 15: 65 17: 14 17: 65 18: 65 19: 65  
20: 65 21: 65 22: 65 26: 65 29: 65 30: 65  
15 33: 65 34: 65 35: 65 37: 65 38: 65 39: 65  
40: 65 42: 65 43: 65 48: 65 49: 65 50: 65  
51: 14

There are 23 hits at base# 65

There are 14 hits at base# 14

20 AluI AGct 42  
1: 47 2: 47 3: 47 4: 47 5: 47 6: 47  
7: 47 8: 47 9: 47 10: 47 11: 47 16: 63  
23: 63 24: 63 25: 63 31: 63 32: 63 36: 63  
25 37: 47 37: 52 38: 47 38: 52 39: 47 39: 52  
40: 47 40: 52 41: 47 41: 52 42: 47 42: 52  
43: 47 43: 52 44: 47 44: 52 45: 47 45: 52  
46: 47 46: 52 47: 47 47: 52 49: 15 50: 47

There are 23 hits at base# 47

30 There are 11 hits at base# 52 Only 5 bases from 47

35 BspI GCtnagc 21  
1: 48 2: 48 3: 48 5: 48 6: 48 7: 48  
8: 48 9: 48 10: 48 11: 48 37: 48 38: 48  
39: 48 40: 48 41: 48 42: 48 43: 48 44: 48  
45: 48 46: 48 47: 48

There are 21 hits at base# 48

54/128

MwoI GCNNNNNngc 19  
 1: 48 2: 28 19: 36 22: 36 23: 36 24: 36  
 25: 36 26: 36 35: 36 37: 67 39: 67 40: 67  
 41: 67 42: 67 43: 67 44: 67 45: 67 46: 67  
 5 47: 67

There are 10 hits at base# 67  
 There are 7 hits at base# 36

DdeI Ctnag 71  
 10 1: 49 1: 58 2: 49 2: 58 3: 49 3: 58  
 3: 65 4: 49 4: 58 5: 49 5: 58 5: 65  
 6: 49 6: 58 6: 65 7: 49 7: 58 7: 65  
 8: 49 8: 58 9: 49 9: 58 9: 65 10: 49  
10: 58 10: 65 11: 49 11: 58 11: 65 15: 58

15 16: 58 16: 65 17: 58 18: 58 20: 58 21: 58  
 22: 58 23: 58 23: 65 24: 58 24: 65 25: 58  
25: 65 26: 58 27: 58 27: 65 28: 58 30: 58  
31: 58 31: 65 32: 58 32: 65 35: 58 36: 58  
36: 65 37: 49 38: 49 39: 26 39: 49 40: 49  
 20 41: 49 42: 26 42: 49 43: 49 44: 49 45: 49  
 46: 49 47: 49 48: 12 49: 12 51: 65

There are 29 hits at base# 58  
There are 22 hits at base# 49 Only nine base from 58  
There are 16 hits at base# 65 Only seven bases from 58

25 BglII Agatct 11  
 1: 61 2: 61 3: 61 4: 61 5: 61 6: 61  
 7: 61 9: 61 10: 61 11: 61 51: 47

There are 10 hits at base# 61

30 BstYI Rgatcy 12  
 1: 61 2: 61 3: 61 4: 61 5: 61 6: 61  
 7: 61 8: 61 9: 61 10: 61 11: 61 51: 47

There are 11 hits at base# 61

35

Hpy188I TCNga 17  
 1: 64 2: 64 3: 64 4: 64 5: 64 6: 64  
 7: 64 8: 64 9: 64 10: 64 11: 64 16: 57  
 20: 57 27: 57 35: 57 48: 67 49: 67

5 There are 11 hits at base# 64

There are 4 hits at base# 57

There are 2 hits at base# 67 Could be ragged.

MslI CAYNNnnRTG 44  
 '0 1: 72 2: 72 3: 72 4: 72 5: 72 6: 72  
 7: 72 8: 72 9: 72 10: 72 11: 72 15: 72  
 17: 72 18: 72 19: 72 21: 72 23: 72 24: 72  
 25: 72 26: 72 28: 72 29: 72 30: 72 31: 72  
 32: 72 33: 72 34: 72 35: 72 36: 72 37: 72  
 '5 38: 72 39: 72 40: 72 41: 72 42: 72 43: 72  
 44: 72 45: 72 46: 72 47: 72 48: 72 49: 72  
 50: 72 51: 72

There are 44 hits at base# 72

'0 BsiEI CGRYcg 23  
 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74  
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74  
 33: 74 34: 74 37: 74 38: 74 39: 74 40: 74  
 41: 74 42: 74 45: 74 46: 74 47: 74

'5 There are 23 hits at base# 74

EaeI Yggccr 23  
 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74  
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74  
 '0 33: 74 34: 74 37: 74 38: 74 39: 74 40: 74  
 41: 74 42: 74 45: 74 46: 74 47: 74

There are 23 hits at base# 74

EagI Cggccg 23  
 '5 1: 74 3: 74 4: 74 5: 74 7: 74 8: 74  
 9: 74 10: 74 11: 74 17: 74 22: 74 30: 74

2007211861 21 Aug 2007

56/128

33: 74 34: 74 37: 74 38: 74 39: 74 40: 74  
41: 74 42: 74 45: 74 46: 74 47: 74

There are 23 hits at base# 74

5 HaeIII GGcc 27  
1: 75 3: 75 4: 75 5: 75 7: 75 8: 75  
9: 75 10: 75 11: 75 16: 75 17: 75 20: 75  
22: 75 30: 75 33: 75 34: 75 37: 75 38: 75  
39: 75 40: 75 41: 75 42: 75 45: 75 46: 75  
10 47: 75 48: 63 49: 63

There are 25 hits at base# 75

Bst4CI ACNgt 65°C 63 Sites There is a third isoschismer

15 1: 86 2: 86 3: 86 4: 86 5: 86 6: 86  
7: 34 7: 86 8: 86 9: 86 10: 86 11: 86  
12: 86 13: 86 14: 86 15: 36 15: 86 16: 53  
16: 86 17: 36 17: 86 18: 86 19: 86 20: 53  
20: 86 21: 36 21: 86 22: 0 22: 86 23: 86  
24: 86 25: 86 26: 86 27: 53 27: 86 28: 36  
20 28: 86 29: 86 30: 86 31: 86 32: 86 33: 36  
33: 86 34: 86 35: 53 35: 86 36: 86 37: 86  
38: 86 39: 86 40: 86 41: 86 42: 86 43: 86  
44: 86 45: 86 46: 86 47: 86 48: 86 49: 86  
50: 86 51: 0 51: 86

15 There are 51 hits at base# 86 All the other sites are well away

HpyCH4III ACNgt 63

10 1: 86 2: 86 3: 86 4: 86 5: 86 6: 86  
7: 34 7: 86 8: 86 9: 86 10: 86 11: 86  
12: 86 13: 86 14: 86 15: 36 15: 86 16: 53  
16: 86 17: 36 17: 86 18: 86 19: 86 20: 53  
20: 86 21: 36 21: 86 22: 0 22: 86 23: 86  
24: 86 25: 86 26: 86 27: 53 27: 86 28: 36  
28: 86 29: 86 30: 86 31: 86 32: 86 33: 36  
15 33: 86 34: 86 35: 53 35: 86 36: 86 37: 86  
38: 86 39: 86 40: 86 41: 86 42: 86 43: 86

2007211861 21 Aug 2007

57/128

44: 86 45: 86 46: 86 47: 86 48: 86 49: 86  
50: 86 51: 0 51: 86  
There are 51 hits at base# 86

5 HinFI Gantc 43  
2: 2 3: 2 4: 2 5: 2 6: 2 7: 2  
8: 2 9: 2 9: 22 10: 2 11: 2 15: 2  
16: 2 17: 2 18: 2 19: 2 19: 22 20: 2  
21: 2 23: 2 24: 2 25: 2 26: 2 27: 2  
'0 28: 2 29: 2 30: 2 31: 2 32: 2 33: 2  
33: 22 34: 22 35: 2 36: 2 37: 2 38: 2  
40: 2 43: 2 44: 2 45: 2 46: 2 47: 2  
50: 60

There are 38 hits at base# 2

'5 MlyI GAGTCNNNNNn 18  
2: 2 3: 2 4: 2 5: 2 6: 2 7: 2  
8: 2 9: 2 10: 2 11: 2 37: 2 38: 2  
40: 2 43: 2 44: 2 45: 2 46: 2 47: 2

'0 There are 18 hits at base# 2

PleI gagtc 18  
2: 2 3: 2 4: 2 5: 2 6: 2 7: 2  
8: 2 9: 2 10: 2 11: 2 37: 2 38: 2  
'5 40: 2 43: 2 44: 2 45: 2 46: 2 47: 2

There are 18 hits at base# 2

AciI Ccgc 24  
2: 26 9: 14 10: 14 11: 14 27: 74 37: 62  
37: 65 38: 62 39: 65 40: 62 40: 65 41: 65  
0 42: 65 43: 62 43: 65 44: 62 44: 65 45: 62  
46: 62 47: 62 47: 65 48: 35 48: 74 49: 74

There are 8 hits at base# 62

There are 8 hits at base# 65

There are 3 hits at base# 14

'5 There are 3 hits at base# 74

There are 1 hits at base# 26

There are 1 hits at base# 35

21 Aug 2007

2007211861

58/128

--- Gcgg 11  
 8: 91 9: 16 10: 16 11: 16 37: 67 39: 67  
 40: 67 42: 67 43: 67 45: 67 46: 67

There are 7 hits at base# 67

5 There are 3 hits at base# 16

There are 1 hits at base# 91

BsiHKAI GWGCWc 20  
 2: 30 4: 30 6: 30 7: 30 9: 30 10: 30  
 10 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51  
 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51  
 46: 51 47: 51

There are 11 hits at base# 51

15 Bsp1286I GDGCHc 20  
 2: 30 4: 30 6: 30 7: 30 9: 30 10: 30  
 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51  
 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51  
 46: 51 47: 51

20 There are 11 hits at base# 51

HgiAI GWGCWc 20  
 2: 30 4: 30 6: 30 7: 30 9: 30 10: 30  
 12: 89 13: 89 14: 89 37: 51 38: 51 39: 51  
 25 40: 51 41: 51 42: 51 43: 51 44: 51 45: 51  
 46: 51 47: 51

There are 11 hits at base# 51

BsoFI GCngc 26  
 10 2: 53 3: 53 5: 53 6: 53 7: 53 8: 53  
 8: 91 9: 53 10: 53 11: 53 31: 53 36: 36  
 37: 64 39: 64 40: 64 41: 64 42: 64 43: 64  
 44: 64 45: 64 46: 64 47: 64 48: 53 49: 53  
 50: 45 51: 53

15 There are 13 hits at base# 53

There are 10 hits at base# 64

TseI Gcwgc 17  
 2: 53 3: 53 5: 53 6: 53 7: 53 8: 53



2007211861 21 Aug 2007

59/128

9: 53 10: 53 11: 53 31: 53 36: 36 45: 64  
46: 64 48: 53 49: 53 50: 45 51: 53

There are 13 hits at base# 53

5

**MnlI gagg 34**

3: 67 3: 95 4: 51 5: 16 5: 67 6: 67  
7: 67 8: 67 9: 67 10: 67 11: 67 15: 67  
16: 67 17: 67 19: 67 20: 67 21: 67 22: 67  
23: 67 24: 67 25: 67 26: 67 27: 67 28: 67  
29: 67 30: 67 31: 67 32: 67 33: 67 34: 67  
35: 67 36: 67 50: 67 51: 67

There are 31 hits at base# 67

10

**HpyCH4V TGca 34**

5: 90 6: 90 11: 90 12: 90 13: 90 14: 90  
15: 44 16: 44 16: 90 17: 44 18: 90 19: 44  
20: 44 21: 44 22: 44 23: 44 24: 44 25: 44  
26: 44 27: 44 27: 90 28: 44 29: 44 33: 44  
34: 44 35: 44 35: 90 36: 38 48: 44 49: 44  
50: 44 50: 90 51: 44 51: 52

There are 21 hits at base# 44

There are 1 hits at base# 52

20

**AccI GTmkac 13 5-base recognition**

7: 37 11: 24 37: 16 38: 16 39: 16 40: 16  
41: 16 42: 16 43: 16 44: 16 45: 16 46: 16  
47: 16

There are 11 hits at base# 16

25

30

**SacII CCGCgg 8 6-base recognition**

9: 14 10: 14 11: 14 37: 65 39: 65 40: 65  
42: 65 43: 65

There are 5 hits at base# 65

There are 3 hits at base# 14

35

**TfiI Gawtc 24**

9: 22 15: 2 16: 2 17: 2 18: 2 19: 2  
19: 22 20: 2 21: 2 23: 2 24: 2 25: 2

60/128

26: 2 27: 2 28: 2 29: 2 30: 2 31: 2  
 32: 2 33: 2 33: 22 34: 22 35: 2 36: 2

There are 20 hits at base# 2

5 BsmAI Nnnnnngagac 19  
 15: 11 16: 11 20: 11 21: 11 22: 11 23: 11  
 24: 11 25: 11 26: 11 27: 11 28: 11 28: 56  
 30: 11 31: 11 32: 11 35: 11 36: 11 44: 87  
 48: 87

10 There are 16 hits at base# 11

BpmI ctccag 19  
 15: 12 16: 12 17: 12 18: 12 20: 12 21: 12  
 22: 12 23: 12 24: 12 25: 12 26: 12 27: 12  
 15 28: 12 30: 12 31: 12 32: 12 34: 12 35: 12  
 36: 12

There are 19 hits at base# 12

20 XmnI GAANNnnttc 12  
 37: 30 38: 30 39: 30 40: 30 41: 30 42: 30  
 43: 30 44: 30 45: 30 46: 30 47: 30 50: 30

There are 12 hits at base# 30

25 BsrI NCcagt 12  
 37: 32 38: 32 39: 32 40: 32 41: 32 42: 32  
 43: 32 44: 32 45: 32 46: 32 47: 32 50: 32

There are 12 hits at base# 32

30 BanII GRGCYc 11  
 37: 51 38: 51 39: 51 40: 51 41: 51 42: 51  
 43: 51 44: 51 45: 51 46: 51 47: 51

There are 11 hits at base# 51

35 Ecl136I GAGctc 11  
 37: 51 38: 51 39: 51 40: 51 41: 51 42: 51  
 43: 51 44: 51 45: 51 46: 51 47: 51

There are 11 hits at base# 51

SacI GAGCTc 11

2007211861 21 Aug 2007

61/128

37: 51 38: 51 39: 51 40: 51 41: 51 42: 51  
43: 51 44: 51 45: 51 46: 51 47: 51

There are 11 hits at base# 51

---

5



Table 217: Human HC GLG FR1 Sequences

VH Exon - Nucleotide sequence alignment

VH1

5  
1-02 CAG GTG CAG CTG GTG CAG TCT GGG GCT GAG GTG AAG AAG CCT GGG GCC TCA GTG AAG  
GTC TCC TGC AAG GCT TCT GGA TAC ACC TTC ACC

1-03 cag gtC cag ctT gtg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag  
gtT tcc tgc aag gct tct gga tac acc ttc acT

1-08 cag gtg cag ctg gtg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag  
gtc tcc tgc aag gct tct gga tac acc ttc acc

'0  
1-18 cag gtT cag ctg gtg cag tct ggA gct gag gtg aag aag cct ggg gcc tca gtg aag  
gtc tcc tgc aag gct tct ggT tac acc ttT acc

1-24 cag gtC cag ctg gtA cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag  
gtc tcc tgc aag gTt tcc gga tac acc Ctc acT

5  
1-45 cag Atg cag ctg gtg cag tct ggg gct gag gtg aag aag Act ggg Tcc tca gtg aag  
gtT tcc tgc aag gct tcc gga tac acc ttc acc

1-46 cag gtg cag ctg gtg cag tct ggg gct gag gtg aag aag cct ggg gcc tca gtg aag  
gtT tcc tgc aag gcA tct gga tac acc ttc acc

1-58 caA Atg cag ctg gtg cag tct ggg Cct gag gtg aag aag cct ggg Acc tca gtg aag  
gtc tcc tgc aag gct tct gga tTc acc ttT acT

'0  
1-69 cag gtg cag ctg gtg cag tct ggg gct gag gtg aag aag cct ggg Tcc tcG gtg aag  
gtc tcc tgc aag gct tct gga GGc acc ttc aGc

1-e cag gtg cag ctg gtg cag tct ggg gct gag gtg aag aag cct ggg Tcc tcG gtg aag  
gtc tcc tgc aag gct tct gga GGc acc ttc aGc

5  
1-f Gag gtC cag ctg gtA cag tct ggg gct gag gtg aag aag cct ggg gcT Aca gtg aaA  
Atc tcc tgc aag gTt tct gga tac acc ttc acc

VH2

2-05 CAG ATC ACC TTG AAG GAG TCT GGT CCT ACG CTG GTG AAA CCC ACA CAG ACC CTC ACG  
CTG ACC TGC ACC TTC TCT GGG TTC TCA CTC AGC

0  
2-26 cag Gtc acc ttg aag gag tct ggt cct GTg ctg gtg aaa ccc aca Gag acc ctc acg  
ctg acc tgc acc Gtc tct ggg ttc tca ctc agc

2-70 cag Gtc acc ttg aag gag tct ggt cct Gcg ctg gtg aaa ccc aca cag acc ctc acA  
ctg acc tgc acc ttc tct ggg ttc tca ctc agc

VH3

5  
3-07 GAG GTG CAG CTG GTG GAG TCT GGG GGA GGC TTG GTC CAG CCT GGG GGG TCC CTG AGA  
CTC TCC TGT GCA GCC TCT GGA TTC ACC TTT AGT

3-09 gaA gtg cag ctg gtg gag tct ggg gga ggc ttg gtA cag cct ggC Agg tcc ctg aga  
ctc tcc tgt gca gcc tct gga ttc acc ttt GAt

3-11 Cag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc Aag cct ggA ggg tcc ctg aga  
ctc tcc tgt gca gcc tct gga ttc acc ttC agt

0  
3-13 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtA cag cct ggg ggg tcc ctg aga  
ctc tcc tgt gca gcc tct gga ttc acc ttC agt

3-15 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtA Aag cct ggg ggg tcc ctT aga  
ctc tcc tgt gca gcc tct gga ttc acT ttC agt

3-20 gag gtg cag ctg gtg gag tct ggg gga ggT Gtg gtA cGg cct ggg ggg tcc ctg aga

ctc tcc tgt gca gcc tct gga ttc acc ttt GAt  
 3-21 gag gtg cag ctg gtg gag tct ggg gga ggc Ctg gtc Aag cct ggg ggg. tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 5 3-23 gag gtg cag ctg Ttg gag tct ggg gga ggc ttg gtA cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttt agC  
 3-30 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 3-30.3 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 10 3-30.5 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 3-33 Cag gtg cag ctg gtg gag tct ggg gga ggc Gtg gtc cag cct ggg Agg tcc ctg aga  
 ctc tcc tgt gca gcG tct gga ttc acc ttC agt  
 15 3-43 gaA gtg cag ctg gtg gag tct ggg gga gTc Gtg gtA cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttt GAt  
 3-48 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtA cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 3-49 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtA cag ccA ggg Cgg tcc ctg aga  
 ctc tcc tgt Aca gcT tct gga ttc acc ttt Ggt  
 20 3-53 gag gtg cag ctg gtg gag Act ggA gga ggc ttg Atc cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct ggG ttc acc GtC agt  
 3-64 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 25 3-66 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc GtC agt  
 3-72 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggA ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 3-73 gag gtg cag ctg gtg gag tct ggg gga ggc ttg gtc cag cct ggg ggg tcc ctg aAa  
 ctc tcc tgt gca gcc tct ggG ttc acc ttC agt  
 30 3-74 gag gtg cag ctg gtg gag tcC ggg gga ggc ttA gtT cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc ttC agt  
 3-d gag gtg cag ctg gtg gag tct Cgg gga gTc ttg gtA cag cct ggg ggg tcc ctg aga  
 ctc tcc tgt gca gcc tct gga ttc acc GtC agt  
 VH4  
 35 4-04 CAG GTG CAG CTG CAG GAG TCG GGC CCA GGA CTG GTG AAG CCT TCG GGG ACC CTG TCC  
 CTC ACC TGC GCT GTC TCT GGT GGC TCC ATC AGC  
 4-28 cag gtg cag ctg cag gag tct ggc cca gga ctg gtg aag cct tct gAC acc ctg tcc  
 ctc acc tgc gct gtc tct ggt TAc tcc atc agc  
 4-30.1 cag gtg cag ctg cag gag tct ggc cca gga ctg gtg aag cct tcA CAg acc ctg tcc  
 40 ctc acc tgc Act gtc tct ggt ggc tcc atc agc  
 4-30.2 cag Ctg cag ctg cag gag tcC ggc Tcà gga ctg gtg aag cct tcA CAg acc ctg tcc  
 ctc acc tgc gct gtc tct ggt ggc tcc atc agc  
 4-30.4 cag gtg cag ctg cag gag tct ggc cca gga ctg gtg aag cct tcA CAg acc ctg tcc  
 ctc acc tgc Act gtc tct ggt ggc tcc atc agc

4-31 cag gtg cag ctg cag gag tcg ggc cca gga ctg gtg aag cct tcA CAg acc ctg tcc  
 ctc acc tgc Act gtc tct ggt ggc tcc atc agc  
 4-34 cag gtg cag ctA cag Cag tGg ggc Gca gga ctg Ttg aag cct tcg gAg acc ctg tcc  
 ctc acc tgc gct gtc tAt ggt ggG tcc Ttc agT  
 5 4-39 cag Ctg cag ctg cag gag tcg ggc cca gga ctg gtg aag cct tcg gAg acc ctg tcc  
 ctc acc tgc Act gtc tct ggt ggc tcc atc agc  
 4-59 cag gtg cag ctg cag gag tcg ggc cca gga ctg gtg aag cct tcg gAg acc ctg tcc  
 ctc acc tgc Act gtc tct ggt ggc tcc atc agT  
 0 4-61 cag gtg cag ctg cag gag tcg ggc cca gga ctg gtg aag cct tcg gAg acc ctg tcc  
 ctc acc tgc Act gtc tct ggt ggc tcc Gtc agc  
 4-b cag gtg cag ctg cag gag tcg ggc cca gga ctg gtg aag cct tcg gAg acc ctg tcc  
 ctc acc tgc gct gtc tct ggt TAc tcc atc agc  
 VH5  
 5 5-51 GAG GTG CAG CTG GTG CAG TCT GGA GCA GAG GTG AAA AAG CCC GGG GAG TCT CTG AAG  
 ATC TCC TGT AAG GGT TCT GGA TAC AGC TTT ACC  
 5-a gaA gtg cag ctg gtg cag tct gga gca gag gtg aaa aag ccc ggg gag tct ctg aGg  
 atc tcc tgt aag ggt tct gga tac agc ttt acc  
 VH6  
 0 6-1 CAG GTA CAG CTG CAG CAG TCA GGT CCA GGA CTG GTG AAG CCC TCG CAG ACC CTC TCA  
 CTC ACC TGT GCC ATC TCC GGG GAC AGT GTC TCT  
 VH7  
 7-4.1 CAG GTG CAG CTG GTG CAA TCT GGG TCT GAG TTG AAG AAG CCT GGG GCC TCA GTG AAG  
 GTT TCC TGC AAG GCT TCT GGA TAC ACC TTC ACT

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2007211861 21 Aug 2007

66/128

Table 220: RERS sites in Human HC GLG FR1s where there are at least 20 GLGs cut

BsgI GTGCAG 71 (cuts 16/14 bases to right)

1: 4	1: 13	2: 13	3: 4	3: 13	4: 13
6: 13	7: 4	7: 13	8: 13	9: 4	9: 13
10: 4	10: 13	15: 4	15: 65	16: 4	16: 65
17: 4	17: 65	18: 4	18: 65	19: 4	19: 65
20: 4	20: 65	21: 4	21: 65	22: 4	22: 65
23: 4	23: 65	24: 4	24: 65	25: 4	25: 65
26: 4	26: 65	27: 4	27: 65	28: 4	28: 65
29: 4	30: 4	30: 65	31: 4	31: 65	32: 4
32: 65	33: 4	33: 65	34: 4	34: 65	35: 4
35: 65	36: 4	36: 65	37: 4	38: 4	39: 4
41: 4	42: 4	43: 4	45: 4	46: 4	47: 4
48: 4	48: 13	49: 4	49: 13	51: 4	

There are 39 hits at base# 4

There are 21 hits at base# 65

-"- ctgcac	9				
12: 63	13: 63	14: 63	39: 63	41: 63	42: 63
44: 63	45: 63	46: 63			

BbvI GCAGC	65				
1: 6	3: 6	6: 6	7: 6	8: 6	9: 6
10: 6	15: 6	15: 67	16: 6	16: 67	17: 6
17: 67	18: 6	18: 67	19: 6	19: 67	20: 6
20: 67	21: 6	21: 67	22: 6	22: 67	23: 6
23: 67	24: 6	24: 67	25: 6	25: 67	26: 6
26: 67	27: 6	27: 67	28: 6	28: 67	29: 6
30: 6	30: 67	31: 6	31: 67	32: 6	32: 67
33: 6	33: 67	34: 6	34: 67	35: 6	35: 67
36: 6	36: 67	37: 6	38: 6	39: 6	40: 6
41: 6	42: 6	43: 6	44: 6	45: 6	46: 6
47: 6	48: 6	49: 6	50: 12	51: 6	

There are 43 hits at base# 6 Boded sites very near sites listed below

There are 21 hits at base# 67

-"- gctgc	13				
37: 9	38: 9	39: 9	40: 3	40: 9	41: 9
42: 9	44: 3	44: 9	45: 9	46: 9	47: 9



67/128

50: 9

There are 11 hits at base# 9

BsoFI GCngc

78

5	1: 6	3: 6	6: 6	7: 6	8: 6	9: 6
	10: 6	15: 6	15: 67	16: 6	16: 67	17: 6
	17: 67	18: 6	18: 67	19: 6	19: 67	20: 6
	20: 67	21: 6	21: 67	22: 6	22: 67	23: 6
	23: 67	24: 6	24: 67	25: 6	25: 67	26: 6
10	26: 67	27: 6	27: 67	28: 6	28: 67	29: 6
	30: 6	30: 67	31: 6	31: 67	32: 6	32: 67
	33: 6	33: 67	34: 6	34: 67	35: 6	35: 67
	36: 6	36: 67	<u>37: 6</u>	<u>37: 9</u>	<u>38: 6</u>	<u>38: 9</u>
	39: 6	39: 9	<u>40: 3</u>	<u>40: 6</u>	<u>40: 9</u>	41: 6
15	41: 9	42: 6	42: 9	43: 6	<u>44: 3</u>	<u>44: 6</u>
	<u>44: 9</u>	<u>45: 6</u>	<u>45: 9</u>	<u>46: 6</u>	<u>46: 9</u>	<u>47: 6</u>
	<u>47: 9</u>	48: 6	49: 6	50: 9	50: 12	51: 6

There are 43 hits at base# 6 These often occur together.

There are 11 hits at base# 9

20 There are 2 hits at base# 3

There are 21 hits at base# 67

TseI Gcwgc

78

	1: 6	3: 6	6: 6	7: 6	8: 6	9: 6
25	10: 6	15: 6	15: 67	16: 6	16: 67	17: 6
	17: 67	18: 6	18: 67	19: 6	19: 67	20: 6
	20: 67	21: 6	21: 67	22: 6	22: 67	23: 6
	23: 67	24: 6	24: 67	25: 6	25: 67	26: 6
	26: 67	27: 6	27: 67	28: 6	28: 67	29: 6
30	30: 6	30: 67	31: 6	31: 67	32: 6	32: 67
	33: 6	33: 67	34: 6	34: 67	35: 6	35: 67
	36: 6	36: 67	<u>37: 6</u>	<u>37: 9</u>	<u>38: 6</u>	<u>38: 9</u>
	<u>39: 6</u>	<u>39: 9</u>	<u>40: 3</u>	<u>40: 6</u>	<u>40: 9</u>	<u>41: 6</u>
	<u>41: 9</u>	<u>42: 6</u>	<u>42: 9</u>	43: 6	<u>44: 3</u>	<u>44: 6</u>
35	<u>44: 9</u>	<u>45: 6</u>	<u>45: 9</u>	<u>46: 6</u>	<u>46: 9</u>	<u>47: 6</u>
	<u>47: 9</u>	48: 6	49: 6	<u>50: 9</u>	<u>50: 12</u>	51: 6

There are 43 hits at base# 6 Often together.

There are 11 hits at base# 9

68/128

There are 2 hits at base# 3

There are 1 hits at base# 12

There are 21 hits at base# 67

5 MspAII CMGckg 48

	1: 7	3: 7	4: 7	5: 7	6: 7	7: 7
	8: 7	9: 7	10: 7	11: 7	15: 7	16: 7
	17: 7	18: 7	19: 7	20: 7	21: 7	22: 7
	23: 7	24: 7	25: 7	26: 7	27: 7	28: 7
10	29: 7	30: 7	31: 7	32: 7	33: 7	34: 7
	35: 7	36: 7	37: 7	38: 7	39: 7	<u>40: 1</u>
	<u>40: 7</u>	41: 7	42: 7	<u>44: 1</u>	<u>44: 7</u>	45: 7
	46: 7	47: 7	48: 7	49: 7	50: 7	51: 7

There are 46 hits at base# 7

15 PvuII CAGctg 48

	1: 7	3: 7	4: 7	5: 7	6: 7	7: 7
	8: 7	9: 7	10: 7	11: 7	15: 7	16: 7
	17: 7	18: 7	19: 7	20: 7	21: 7	22: 7
20	23: 7	24: 7	25: 7	26: 7	27: 7	28: 7
	29: 7	30: 7	31: 7	32: 7	33: 7	34: 7
	35: 7	36: 7	37: 7	38: 7	39: 7	<u>40: 1</u>
	<u>40: 7</u>	41: 7	42: 7	<u>44: 1</u>	<u>44: 7</u>	45: 7
	46: 7	47: 7	48: 7	49: 7	50: 7	51: 7

25 There are 46 hits at base# 7

There are 2 hits at base# 1

AluI AGct 54

	1: 8	2: 8	3: 8	4: 8	4: 24	5: 8
30	6: 8	7: 8	8: 8	9: 8	10: 8	11: 8
	15: 8	16: 8	17: 8	18: 8	19: 8	20: 8
	21: 8	22: 8	23: 8	24: 8	25: 8	26: 8
	27: 8	28: 8	29: 8	29: 69	30: 8	31: 8
	32: 8	33: 8	34: 8	35: 8	36: 8	37: 8
35	38: 8	39: 8	<u>40: 2</u>	<u>40: 8</u>	41: 8	42: 8
	43: 8	<u>44: 2</u>	<u>44: 8</u>	45: 8	46: 8	47: 8
	48: 8	48: 82	49: 8	49: 82	50: 8	51: 8

There are 48 hits at base# 8

There are 2 hits at base# 2

DdeI Ctnag 48

1: 26	1: 48	2: 26	2: 48	3: 26	3: 48
4: 26	4: 48	5: 26	5: 48	6: 26	6: 48
7: 26	7: 48	8: 26	8: 48	9: 26	10: 26
11: 26	12: 85	13: 85	14: 85	15: 52	16: 52
17: 52	18: 52	19: 52	20: 52	21: 52	22: 52
23: 52	24: 52	25: 52	26: 52	27: 52	28: 52
29: 52	30: 52	31: 52	32: 52	33: 52	35: 30
35: 52	36: 52	40: 24	49: 52	51: 26	51: 48

There are 22 hits at base# 52 52 and 48 never together.

There are 9 hits at base# 48

There are 12 hits at base# 26 26 and 24 never together.

HphI tcacc 42

1: 86	3: 86	6: 86	7: 86	8: 80	11: 86
12: 5	13: 5	14: 5	15: 80	16: 80	17: 80
18: 80	20: 80	21: 80	22: 80	23: 80	24: 80
25: 80	26: 80	27: 80	28: 80	29: 80	30: 80
31: 80	32: 80	33: 80	34: 80	35: 80	36: 80
37: 59	38: 59	39: 59	40: 59	41: 59	42: 59
43: 59	44: 59	45: 59	46: 59	47: 59	50: 59

There are 22 hits at base# 80 80 and 86 never together

There are 5 hits at base# 86

There are 12 hits at base# 59

BssKI Nccngg 50

1: 39	2: 39	3: 39	4: 39	5: 39	7: 39
8: 39	9: 39	10: 39	11: 39	15: 39	16: 39
17: 39	18: 39	19: 39	20: 39	21: 29	21: 39
22: 39	23: 39	24: 39	25: 39	26: 39	27: 39
28: 39	29: 39	30: 39	31: 39	32: 39	33: 39
34: 39	35: 19	35: 39	36: 39	37: 24	38: 24
39: 24	41: 24	42: 24	44: 24	45: 24	46: 24
47: 24	<u>48: 39</u>	<u>48: 40</u>	<u>49: 39</u>	<u>49: 40</u>	50: 24
50: 73	51: 39				

There are 35 hits at base# 39 39 and 40 together twice.

There are 2 hits at base# 40

BsaJI CCnngg

47

5  
 1: 40 2: 40 3: 40 4: 40 5: 40 7: 40  
 8: 40 9: 40 9: 47 10: 40 10: 47 11: 40  
 15: 40 18: 40 19: 40 20: 40 21: 40 22: 40  
 23: 40 24: 40 25: 40 26: 40 27: 40 28: 40  
 29: 40 30: 40 31: 40 32: 40 34: 40 35: 20  
 35: 40 36: 40 37: 24 38: 24 39: 24 41: 24  
 42: 24 44: 24 45: 24 46: 24 47: 24 48: 40  
 10 48: 41 49: 40 49: 41 50: 74 51: 40

There are 32 hits at base# 40 40 and 41 together twice

There are 2 hits at base# 41

There are 9 hits at base# 24

There are 2 hits at base# 47

15

BstNI CCwgg

44

PspGI ccwgg

ScrFI(\$M.HpaII) CCwgg

20  
 1: 40 2: 40 3: 40 4: 40 5: 40 7: 40  
 8: 40 9: 40 10: 40 11: 40 15: 40 16: 40  
 17: 40 18: 40 19: 40 20: 40 21: 30 21: 40  
 22: 40 23: 40 24: 40 25: 40 26: 40 27: 40  
 28: 40 29: 40 30: 40 31: 40 32: 40 33: 40  
 34: 40 35: 40 36: 40 37: 25 38: 25 39: 25  
 25 41: 25 42: 25 44: 25 45: 25 46: 25 47: 25  
 50: 25 51: 40

There are 33 hits at base# 40

ScrFI CCnngg

50

30  
 1: 40 2: 40 3: 40 4: 40 5: 40 7: 40  
 8: 40 9: 40 10: 40 11: 40 15: 40 16: 40  
 17: 40 18: 40 19: 40 20: 40 21: 30 21: 40  
 22: 40 23: 40 24: 40 25: 40 26: 40 27: 40  
 28: 40 29: 40 30: 40 31: 40 32: 40 33: 40  
 35  
 34: 40 35: 20 35: 40 36: 40 37: 25 38: 25  
 39: 25 41: 25 42: 25 44: 25 45: 25 46: 25  
 47: 25 48: 40 48: 41 49: 40 49: 41 50: 25  
 50: 74 51: 40

There are 35 hits at base# 40

2007211861 21 Aug 2007

71/128

There are 2 hits at base# 41

EcoO109I RGgnccy 34

1: 43	2: 43	3: 43	4: 43	5: 43	6: 43
7: 43	8: 43	9: 43	10: 43	15: 46	16: 46
17: 46	18: 46	19: 46	20: 46	21: 46	22: 46
23: 46	24: 46	25: 46	26: 46	27: 46	28: 46
30: 46	31: 46	32: 46	33: 46	34: 46	35: 46
36: 46	37: 46	43: 79	51: 43		

There are 22 hits at base# 46 46 and 43 never together

There are 11 hits at base# 43

NlaIV GGNncc 71

1: 43	2: 43	3: 43	4: 43	5: 43	6: 43
7: 43	8: 43	9: 43	9: 79	10: 43	10: 79
<u>15: 46</u>	<u>15: 47</u>	16: 47	<u>17: 46</u>	<u>17: 47</u>	<u>18: 46</u>
<u>18: 47</u>	<u>19: 46</u>	<u>19: 47</u>	<u>20: 46</u>	<u>20: 47</u>	<u>21: 46</u>
<u>21: 47</u>	<u>22: 46</u>	<u>22: 47</u>	23: 47	24: 47	25: 47
26: 47	<u>27: 46</u>	<u>27: 47</u>	<u>28: 46</u>	<u>28: 47</u>	29: 47
<u>30: 46</u>	<u>30: 47</u>	<u>31: 46</u>	<u>31: 47</u>	<u>32: 46</u>	<u>32: 47</u>
<u>33: 46</u>	<u>33: 47</u>	<u>34: 46</u>	<u>34: 47</u>	<u>35: 46</u>	<u>35: 47</u>
<u>36: 46</u>	<u>36: 47</u>	37: 21	<u>37: 46</u>	<u>37: 47</u>	37: 79
38: 21	39: 21	39: 79	40: 79	41: 21	41: 79
42: 21	42: 79	43: 79	44: 21	44: 79	45: 21
45: 79	46: 21	46: 79	47: 21	51: 43	

There are 23 hits at base# 47 46 & 47 often together

There are 17 hits at base# 46

There are 11 hits at base# 43

Sau96I Ggncc 70

1: 44	2: 3	2: 44	3: 44	4: 44	5: 3	5: 44	6: 44
7: 44	8: 22	8: 44	9: 44	10: 44	11: 3	12: 22	13: 22
14: 22	15: 33	15: 47	16: 47	17: 47	18: 47	19: 47	20: 47
21: 47	22: 47	23: 33	23: 47	24: 33	24: 47	25: 33	25: 47
26: 33	26: 47	27: 47	28: 47	29: 47	30: 47	31: 33	31: 47
32: 33	32: 47	33: 33	33: 47	34: 33	34: 47	35: 47	36: 47
<u>37: 21</u>	<u>37: 22</u>	37: 47	<u>38: 21</u>	<u>38: 22</u>	39: 21	39: 22	41: 21
41: 22	42: 21	42: 22	43: 80	44: 21	44: 22	45: 21	45: 22
46: 21	46: 22	47: 21	47: 22	50: 22	51: 44		

There are 23 hits at base# 47 These do not occur together.

There are 11 hits at base# 44

72/128

There are 14 hits at base# 22 These do occur together.  
 There are 9 hits at base# 21

5 BsmAI GTCTCn<sup>n</sup>n<sup>n</sup>n 22  
 1: 58 3: 58 4: 58 5: 58 8: 58 9: 58  
 10: 58 13: 70 36: 18 37: 70 38: 70 39: 70  
 40: 70 41: 70 42: 70 44: 70 45: 70 46: 70  
 47: 70 48: 48 49: 48 50: 85

There are 11 hits at base# 70

10

-"- Nnnnngagac 27  
 13: 40 15: 48 16: 48 17: 48 18: 48 20: 48  
 21: 48 22: 48 23: 48 24: 48 25: 48 26: 48  
 27: 48 28: 48 29: 48 30: 10 30: 48 31: 48  
 15 32: 48 33: 48 35: 48 36: 48 43: 40 44: 40  
 45: 40 46: 40 47: 40

There are 20 hits at base# 48

20 AvaII Ggwcc 44  
 Sau96I (\$M.HaeIII) Ggwcc 44  
 2: 3 5: 3 6: 44 8: 44 9: 44 10: 44  
 11: 3 12: 22 13: 22 14: 22 15: 33 15: 47  
 16: 47 17: 47 18: 47 19: 47 20: 47 21: 47  
 22: 47 23: 33 23: 47 24: 33 24: 47 25: 33  
 25 25: 47 26: 33 26: 47 27: 47 28: 47 29: 47  
 30: 47 31: 33 31: 47 32: 33 32: 47 33: 33  
 33: 47 34: 33 34: 47 35: 47 36: 47 37: 47  
 43: 80 50: 22

There are 23 hits at base# 47 44 & 47 never together

30 There are 4 hits at base# 44

PpuMI RGgwccy 27  
 6: 43 8: 43 9: 43 10: 43 15: 46 16: 46  
 17: 46 18: 46 19: 46 20: 46 21: 46 22: 46  
 35 23: 46 24: 46 25: 46 26: 46 27: 46 28: 46  
 30: 46 31: 46 32: 46 33: 46 34: 46 35: 46  
 36: 46 37: 46 43: 79

There are 22 hits at base# 46 43 and 46 never occur together.

There are 4 hits at base# 43

BsmFI GGGAC 3

8: 43 37: 46 50: 77

-"- gtccc 33

5 15: 48 16: 48 17: 48 1: 0 1: 0 20: 48  
 21: 48 22: 48 23: 48 24: 48 25: 48 26: 48  
 27: 48 28: 48 29: 48 30: 48 31: 48 32: 48  
 33: 48 34: 48 35: 48 36: 48 37: 54 38: 54  
 39: 54 40: 54 41: 54 42: 54 43: 54 44: 54  
 10 45: 54 46: 54 47: 54

There are 20 hits at base# 48

There are 11 hits at base# 54

HinfI Gantc 80

15 8: 77 12: 16 13: 16 14: 16 15: 16 15: 56  
 15: 77 16: 16 16: 56 16: 77 17: 16 17: 56  
 17: 77 18: 16 18: 56 18: 77 19: 16 19: 56  
 19: 77 20: 16 20: 56 20: 77 21: 16 21: 56  
 21: 77 22: 16 22: 56 22: 77 23: 16 23: 56  
 20 23: 77 24: 16 24: 56 24: 77 25: 16 25: 56  
 25: 77 26: 16 26: 56 26: 77 27: 16 27: 26  
 27: 56 27: 77 28: 16 28: 56 28: 77 29: 16  
 29: 56 29: 77 30: 56 31: 16 31: 56 31: 77  
 32: 16 32: 56 32: 77 33: 16 33: 56 33: 77  
 25 34: 16 35: 16 35: 56 35: 77 36: 16 36: 26  
 36: 56 36: 77 37: 16 38: 16 39: 16 40: 16  
 41: 16 42: 16 44: 16 45: 16 46: 16 47: 16  
 48: 46 49: 46

There are 34 hits at base# 16

30

TfiI Gawtc 21

8: 77 15: 77 16: 77 17: 77 18: 77 19: 77  
 20: 77 21: 77 22: 77 23: 77 24: 77 25: 77  
 26: 77 27: 77 28: 77 29: 77 31: 77 32: 77  
 35 33: 77 35: 77 36: 77

There are 21 hits at base# 77

2007211861 21 Aug 2007

74/128

MlyI GAGTC 38  
12: 16 13: 16 14: 16 15: 16 16: 16 17: 16  
18: 16 19: 16 20: 16 21: 16 22: 16 23: 16  
24: 16 25: 16 26: 16 27: 16 27: 26 28: 16  
5 29: 16 31: 16 32: 16 33: 16 34: 16 35: 16  
36: 16 36: 26 37: 16 38: 16 39: 16 40: 16  
41: 16 42: 16 44: 16 45: 16 46: 16 47: 16  
48: 46 49: 46

There are 34 hits at base# 16

10  
-- GACTC 21  
15: 56 16: 56 17: 56 18: 56 19: 56 20: 56  
21: 56 22: 56 23: 56 24: 56 25: 56 26: 56  
27: 56 28: 56 29: 56 30: 56 31: 56 32: 56  
15 33: 56 35: 56 36: 56

There are 21 hits at base# 56

PleI gagtc 38  
12: 16 13: 16 14: 16 15: 16 16: 16 17: 16  
20 18: 16 19: 16 20: 16 21: 16 22: 16 23: 16  
24: 16 25: 16 26: 16 27: 16 27: 26 28: 16  
29: 16 31: 16 32: 16 33: 16 34: 16 35: 16  
36: 16 36: 26 37: 16 38: 16 39: 16 40: 16  
41: 16 42: 16 44: 16 45: 16 46: 16 47: 16  
25 48: 46 49: 46 -

There are 34 hits at base# 16

-- gactc 21  
15: 56 16: 56 17: 56 18: 56 19: 56 20: 56  
21: 56 22: 56 23: 56 24: 56 25: 56 26: 56  
30 27: 56 28: 56 29: 56 30: 56 31: 56 32: 56  
33: 56 35: 56 36: 56

There are 21 hits at base# 56

AlwNI CAGNNNctg 26  
15: 68 16: 68 17: 68 18: 68 19: 68 20: 68  
35 21: 68 22: 68 23: 68 24: 68 25: 68 26: 68  
27: 68 28: 68 29: 68 30: 68 31: 68 32: 68  
33: 68 34: 68 35: 68 36: 68 39: 46 40: 46  
41: 46 42: 46

There are 22 hits at base# 68



Table 255: Analysis of frequency of matching REaptors in actual V genes

A: HpyCH4V in HC at bases 35-56

Id	Ntot	Number of mismatches.....										Number	Cut	Id	Probe
		0	1	2	3	4	5	6	7	8	9				
5	510	5	11	274	92	61	25	22	11	1	3	5	443	6-1	agttctcccTGCAgctgaactc
	192	54	42	32	24	15	2	3	10	3	1	6	167	3-11	cactgtatcTGCAaatgaacag
	58	19	7	17	6	5	1	0	1	0	2	0	54	3-09	ccctgtatcTGCAaatgaacag
	267	42	33	9	8	8	82	43	22	8	11	1	100	5-51	ccgcctaccTGCAgtggagcag
	250	111	59	41	24	7	5	1	0	0	2	0	242	3-15	cgctgtatcTGCAaatgaacag
10	7	0	2	0	1	0	0	0	0	0	4	0	3	7-4.1	cggcatatcTGCAgatctgcag
	7	0	2	2	0	0	2	1	0	0	0	0	4	3-73	cggcgtatcTGCAaatgaacag
	26	10	4	1	3	1	2	1	3	1	0	0	19	5-a	ctgcctaccTGCAgtggagcag
	21	8	2	3	1	6	1	0	0	0	0	0	20	3-49	tcgcctatcTGCAaatgaacag
	1338	249	162	379	149	103	120	71	47	13	23	12	1052		
15	249	411	790	939	1162	1280	1316	1042	1233	1293	1338				

Id	Probe	dotted probe
6-1	agttctcccTGCAgctgaactc	agttctcccTGCAgctgaactc
3-11	cactgtatcTGCAaatgaacag	cac.g.at.....aa.....ag
3-09	ccctgtatcTGCAaatgaacag	ccc.g.at.....aa.....ag
5-51	ccgcctaccTGCAgtggagcag	ccgc..a.....tg..g.ag
3-15	cgctgtatcTGCAaatgaacag	c.c.g.at.....aa.....ag
7-4.1	cggcatatcTGCAgatctgcag	c.gca.at.....a.ctg.ag
3-73	cggcgtatcTGCAaatgaacag	c.gcg.at.....aa.....ag
5-a	ctgcctaccTGCAgtggagcag	ctgc..a.....tg..g.ag
3-49	tcgcctatcTGCAaatgaacag	tcgc..at.....aa.....ag

Seqs with the expected RE site only.....1004

(Counts only cases with 4 or fewer mismatches)

Seqs with only an unexpected site..... 0

Seqs with both expected and unexpected..... 48

(Counts only cases with 4 or fewer mismatches)

Seqs with no sites..... 0

5

B: BpI in HC

Id	Ntot	0	1	2	3	4	5	6	7	8	Ncut	Name	
10	1	133	73	16	11	13	6	9	1	4	0	119	1-58
												<b>acatgggAGCTGAGCagcctgag</b>	
	2	14	11	1	0	0	0	1	0	1	12	1-02	
												<b>acatgggagctgagcaggctgag</b>	
	3	34	17	8	2	6	1	0	0	0	0	1-18	
												<b>acatgggagctgaggagcctgag</b>	
	4	120	50	32	16	10	9	1	1	1	0	2	5-51
												<b>acctgcagtggagcagcctgaa</b>	
	5	55	13	11	10	17	3	1	0	0	0	0	3-15
												<b>atctgcaaatgaaacagcctgaa</b>	
15	6	340	186	88	41	15	6	3	0	1	0	0	3303
												<b>atctgcaaatgaaacagcctgag</b>	
	7	82	25	16	25	12	1	3	0	0	0	0	3-20
												<b>atctgcaaatgaaacagtctgag</b>	
	8	3	0	2	0	1	0	0	0	0	0	0	74.1
												<b>atctgcagatctgcagcctaaa</b>	
	9	23	18	2	2	1	0	0	0	0	0	0	3-66
												<b>atcttcaaatgaaacagcctgag</b>	
	10	2	1	0	1	0	0	0	0	0	0	0	3-64
												<b>atcttcaaatgggcagcctgag</b>	
20	11	486	249	78	81	38	21	10	4	4	1	467	4301
												<b>ccctgaagctgagctctgtgac</b>	
	12	16	6	3	1	0	1	1	3	1	0	1	6-1
												<b>ccctgcagctgaactctgtgac</b>	
	13	28	15	8	2	2	1	0	0	0	0	0	2-70
												<b>tccttaaatgaccaacatgga</b>	
	14	2	0	2	0	0	0	0	0	0	0	0	2-26
												<b>tccttaccatgaccaacatgga</b>	

601

25

Name	Full sequence	Dot mode
1-58	acatggaCTGAGCagcctgag	acatggaCTGAGCagcctgag
1-02	acatggagctgagcaggctgag	.....g.....
1-18	acatggagctgagcctgag	.....g.....
5-51	acctgcagtggagcagcctgaa	..c..c..tg.....a
3-15	atctgcaaatgaacagcctgaa	.tc..c.aa...a.....a
3-30.3	atctgcaaatgaacagcctgag	.tc..c.aa...a.....
3-20	atctgcaaatgaacagtctgag	.tc..c.aa...a...t.....
7-4.1	atctgcagatctgcagcctaaa	.tc..c..a.ct.....a.a
3-66	atcttcaaatgaacagcctgag	.tc.tc.aa...a.....
3-64	atcttcaaatggcagcctgag	.tc.tc.aa..g.....
4-30.1	ccctgaagctgagctctgtgac	c.c.a.....tctg...c
6-1	ccctgcagctgaactctgtgac	c.c..C.....a.tctg...c
2-70	tccttacaatgaccaacatgga	t.c.tacaa...C..a.a..ga
2-26	tccttaccatgaccaacatgga	t.c.tacca...c..a.a..ga

Seqs with the expected RE site only..... 597 (counting sequences with 4 or fewer mismatches)  
 Seqs with only an unexpected site..... 2  
 Seqs with both expected and unexpected.... 2  
 Seqs with no sites..... 686

C: HpyCH4III, Bst4CI, or TaaI in HC

In scoring whether the RE site of interest is present, only ONs that have 4 or fewer mismatches are counted.

25 Number of sequences..... 1617

Id	Ntot	0	1	2	3	4	5	6	7	8	Neut	acngt	acngt
1	244	78	92	43	18	10	1	2	0	0	241	102#1,1	ccgtgtattACNeTgcgagaga ccgtgtattactgtgcgagaga
2	457	69	150	115	66	34	11	8	3	1	434	103#2,3	ctgtgtattactgtgcgagaga .t.....
3	173	52	45	36	22	14	3	0	0	1	169	108#3	ccgtgtattactgtgcgagagg .....
5	4	0	0	3	2	2	1	6	0	1	8	124#5,1	ccgtgtattactgtgcaacaga .....a.c....
	4	0	0	1	0	1	1	0	1	0	2	145#6	ccatgtattactgtgcaagata ..a.....a...t.
6	15	1	0	1	0	6	4	1	1	1	8	158#8	ccgtgtattactgtgcggcaga .....
7	23	4	8	5	2	2	1	1	0	0	21	205#12	ccacataattactgtgcacacag ..aca.....acacag
8	9	1	1	1	0	3	2	1	0	0	6	226#13	ccacataattactgtgcacggat ..aca.....ac.gat
10	23	7	3	5	5	2	1	0	0	0	6	270#14	ccacgtattactgtgcacggat ..ac.....ac.gat
9	7	1	3	1	1	0	0	1	0	0	6	309#16,	ccttgtattactgtgcaaaaga ..t.....a.a...
11	35	5	10	7	6	3	3	0	1	0	31	313#18,	ctgtgtattactgtgcaagaga .t.....a.....
12	18	2	3	2	2	6	1	0	2	0	15	315#19	ccgtgtattactgtaccacaga .....
13	3	1	2	0	0	0	0	0	0	0	3	320#20	ccttgtaccactgtgcgagaga ..t.....C.....
14	117	29	23	28	22	8	4	2	1	0	110	323#22	ccgtattactgtgcgaaaga .....
15	75	21	25	13	9	1	4	2	0	0	69	330#23,	ctgtgtattactgtgcgaaaga .t.....a.....
16	14	2	2	2	3	0	3	1	1	0	9	349#29	ccgtgtattactgtactagaga .....
17	2	0	0	1	0	0	1	0	0	0	1	372#33	ccgtgtattactgtgctagaga .....
18	1	0	0	1	0	0	0	0	0	0	1	373#34	ccgtgtattactgtactagaca .....
19	2	0	0	0	0	0	0	0	0	2	0	3d#36	ctgtgtattactgtgaaaga .t.....aa.a...
20	34	4	9	9	4	5	3	0	0	0	31	428#38	ccgtgtattactgtgcgagaaa .....
21	17	5	4	2	2	3	1	0	0	0	16	4302#40	ccgtgtattactgtgccagaga .....
22	75	15	17	24	7	10	1	1	0	0	73	439#44	ctgtgtattactgtgcgagaca .t.....C.....
23	40	14	15	4	5	1	0	1	0	0	39	551#48	ccatgtattactgtgcgagaca ..a.....C.....
24	213	26	56	60	42	20	7	2	0	0	204	5a#49	ccatgtattactgtgcgagaaa ..a.....AA
Group		337	471	363	218	130	58	23	11	6			
Cumulative		337	808	1171	1389	1519	1577	1600	1611	1617			

Seqs with the expected RE site only.....1511  
 Seqs with only an unexpected site..... 0

2007211861 21 Aug 2007

Table 255 D

79/128

Seqs with both expected and unexpected.... 8  
 Seqs with no sites..... 0

Analysis repeated using only 8 best REaptors

5	Id	Ntot	0	1	2	3	4	5	6	7	8+		
	1	301	78	101	54	32	16	9	10	1	0	281	102#1 ccgtgtattactgtgcgagaga
	2	493	69	155	125	73	37	14	11	3	6	459	103#2 ctgtgtattactgtgcgagaga
	3	189	52	45	38	23	18	5	4	1	3	176	108#3 ccgtgtattactgtgcgagagg
	4	127	29	23	28	24	10	6	5	2	0	114	323#22 ccgtatattactgtgcgaaaga
0	5	78	21	25	14	11	1	4	2	0	0	72	330#23 ctgtgtattactgtgcgaaaga
	6	79	15	17	25	8	11	1	2	0	0	76	439#44 ctgtgtattactgtgcgagaca
	7	43	14	15	5	5	3	0	1	0	0	42	551#48 ccatgtattactgtgcgagaca
	8	307	26	63	72	51	38	24	14	13	6	250	5a#49 ccatgtattactgtgcgagaga
	1	102#1	ccgtgtattactgtgcgagaga				ccgtgtattactgtgcgagaga						
5	2	103#2	ctgtgtattactgtgcgagaga				.t.....						
	3	108#3	ccgtgtattactgtgcgagagg				.....g						
	4	323#22	ccgtatattactgtgcgaaaga				....a.....a...						
	5	330#23	ctgtgtattactgtgcgaaaga				.t.....a...						
	6	439#44	ctgtgtattactgtgcgagaca				.t.....c.						
0	7	551#48	ccatgtattactgtgcgagaca				..a.....c.						
	8	5a#49	ccatgtattactgtgcgagaa				..a.....AA						

Seqs with the expected RE site only.....1463 / 1617

Seqs with only an unexpected site..... 0

5 Seqs with both expected and unexpected.... 7

Seqs with no sites..... 0

Table 300: Kappa FR1 GLGs

	!	1	2	3	4	5	6	7	8	9	10	11	12	
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
	!	13	14	15	16	17	18	19	20	21	22	23		
5		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	O12
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	.O2
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	O18
10		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	O8
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	A20
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
15		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	A30
		AAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	GCC	ATG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L14
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L1
20		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L15
		GCC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L4
		GCC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TCC	CTG	TCT	
25		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L18
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	TCC	GTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L5
		GAC	ATC	CAG	ATG	ACC	CAG	TCT	CCA	TCT	TCT	GTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L19
30		GAC	ATC	CAG	TTG	ACC	CAG	TCT	CCA	TCC	TTC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L8
		GCC	ATC	CGG	ATG	ACC	CAG	TCT	CCA	TTC	TCC	CTG	TCT	
		GCA	TCT	GTA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGC	!	L23
		GCC	ATC	CGG	ATG	ACC	CAG	TCT	CCA	TCC	TCA	TTC	TCT	
35		GCA	TCT	ACA	GGA	GAC	AGA	GTC	ACC	ATC	ACT	TGT	!	L9
		GTC	ATC	TGG	ATG	ACC	CAG	TCT	CCA	TCC	TTA	CTC	TCT	

21 Aug 2007

2007211861

81/128

	GCA TCT ACA GGA GAC AGA GTC ACC ATC AGT TGT !	L24
	GCC ATC CAG ATG ACC CAG TCT CCA TCC TCC CTG TCT	
	GCA TCT GTA GGA GAC AGA GTC ACC ATC ACT TGC !	L11
	GAC ATC CAG ATG ACC CAG TCT CCT TCC ACC CTG TCT	
5	GCA TCT GTA GGA GAC AGA GTC ACC ATC ACT TGC !	L12
	GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC CTG CCC	
	GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC !	.011
	GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC CTG CCC	
	GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC !	01
10	GAT GTT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC	
	GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC !	A17
	GAT GTT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC	
	GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC !	A1
	GAT ATT GTG ATG ACC CAG ACT CCA CTC TCT CTG TCC	
15	GTC ACC CCT GGA CAG CCG GCC TCC ATC TCC TGC !	A18
	GAT ATT GTG ATG ACC CAG ACT CCA CTC TCT CTG TCC	
	GTC ACC CCT GGA CAG CCG GCC TCC ATC TCC TGC !	A2
	GAT ATT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC	
	GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC !	A19
20	GAT ATT GTG ATG ACT CAG TCT CCA CTC TCC CTG CCC	
	GTC ACC CCT GGA GAG CCG GCC TCC ATC TCC TGC !	A3
	GAT ATT GTG ATG ACC CAG ACT CCA CTC TCC TCA CCT	
	GTC ACC CTT GGA CAG CCG GCC TCC ATC TCC TGC !	A23
	GAA ATT GTG TTG ACG CAG TCT CCA GCC ACC CTG TCT	
25	TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	A27
	GAA ATT GTG TTG ACG CAG TCT CCA GCC ACC CTG TCT	
	TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	A11
	GAA ATA GTG ATG ACG CAG TCT CCA GCC ACC CTG TCT	
	GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	L2
30	GAA ATA GTG ATG ACG CAG TCT CCA GCC ACC CTG TCT	
	GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	L16
	GAA ATT GTG TTG ACA CAG TCT CCA GCC ACC CTG TCT	
	TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	L6
	GAA ATT GTG TTG ACA CAG TCT CCA GCC ACC CTG TCT	
35	TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	L20
	GAA ATT GTA ATG ACA CAG TCT CCA GCC ACC CTG TCT	

2007211861 21 Aug 2007

82/128

TTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC !	L25
GAC ATC GTG ATG ACC CAG TCT CCA GAC TCC CTG GCT	
GTG TCT CTG GGC GAG AGG GCC ACC ATC AAC TGC !	B3
GAA ACG ACA CTC ACG CAG TCT CCA GCA TTC ATG TCA	
GCG ACT CCA GGA GAC AAA GTC AAC ATC TCC TGC !	B2
GAA ATT GTG CTG ACT CAG TCT CCA GAC TTT CAG TCT	
GTG ACT CCA AAG GAG AAA GTC ACC ATC ACC TGC !	A26
GAA ATT GTG CTG ACT CAG TCT CCA GAC TTT CAG TCT	
GTG ACT CCA AAG GAG AAA GTC ACC ATC ACC TGC !	A10
GAT GTT GTG ATG ACA CAG TCT CCA GCT TTC CTC TCT	
GTG ACT CCA GGG GAG AAA GTC ACC ATC ACC TGC !	A14

5

0



Table 302 RERS sites found in Human Kappa FR1 GLGs

	MslI	FokI --> <--< -->	PflFI	BsrI	BsmAI	MnlI	HpyCH 4V
<i>VKI</i>							
O12	3	3 23	12 49	15	18 47	26	36
O2	103	103 123	112 149	115	118 147	126	136
O18	203	203 223	212 249	215	218 247	226	236
O8	303	303 323	312 349	315	318 347	326	336
A20	403	403 423	412 449	415	418 447	426	436
A30	503	503 523	512 549	515	518 547	526	536
L14	603	603	612 649	615	618 647	-	636
L1	703	703 723	712 749	715	718 747	726	736
L15	803	803 823	812 849	815	818 847	826	836
L4	-	903 923	912 949	906 915	918 947	926	936
L18	-	1003	1012 1049	1006 1015	1018 1047	1026	1036
L5	1103	-	1112 1149	1115	1118 1147	-	1136
L19	1203	1203	1212 1249	1215	1218 1247	-	1236
L8	-	1303 1323	1312 1349	1306 1315	1318 1347	-	1336
L23	1403	1403 1408	1412 1449	1415	1418 1447	-	1436
L9	1503	1503 1508 1523	1512 1549	1515	1518 1547	1526	1536
L24	1603	1608 1623	1612 1649	1615	1618 1647	-	1636
L11	1703	1703 1723	1712 1749	1715	1718 1747	1726	1736
L12	1803	1803	1812 1849	1815	1818 1847	-	1836

5

10

15

20

	MslI	FokI --> <--&dashrightarrow	PflFI	BsrI	BsmAI	MnlI	HpyCH 4V
VKII							
O11	1901-1969	-	-	-	-	1956	-
O1	2001-2069	-	-	-	-	2056	-
A17	2101-2169	-	2112	-	2118	2156	-
A1	2201-2269	-	2212	-	2218	2256	-
A18	2301-2369	-	-	-	-	2356	-
A2	2401-2469	-	-	-	-	2456	-
A19	2501-2569	-	2512	-	2518	2556	-
A3	2601-2669	-	2612	-	2618	2656	-
A23	2701-2769	-	-	-	-	2729 2756	-
VKIII							
A27	2801-2869	-	2812	-	2818 2839	2860	-
A11	2901-2969	-	2912	-	2918 2939	2960	-
L2	3001-3069	-	3012	-	3018 3039	3060	-
L16	3101-3169	-	3112	-	3118 3139	3160	-
L6	3201-3269	-	3212	-	3218 3239	3260	-

5

10

15

	MslI	FokI --> <--> -->	PflFI	BsrI	BsmAI	MnlI	HpyCH 4V
L20 3301-3369	-	-	3312	-	3318 3339	3360	-
L25 3401-3469	-	-	3412	-	3418 3439	3460	-
VRIV							
B3 3501-3569	3503	-	3512	3515	3518 3539	3551<	-
VRV							
B2 3601-3669	-	-	3649	-	3618 3647	-	-
VRVI							
A26 3701-3769	-	-	3712	-	3718	-	-
A10 3801-3869	-	-	3812	-	3818	-	-
A14 3901-3969	-	-	3912	-	3918	3930>	-

5

10

Table 302 RERS sites found in Human Kappa FR1 GLGs, continued

	SfaNI	SfcI	HinfI	MlyI --> --> <-->	MaeIII Tsp45I same sites	HphI xx38 xx56 xx62	HpaII MspI xx06 xx52
VRV							
O12 1-69	37	41	53	53	55	56	-
O2 101-169	137	141	153	153	155	156	-
O18 201-269	237	241	253	253	255	256	-

15

	SfaNI	SfcI	HinfI	MlyI --> ---> <--	MaeIII Tsp45I same sites	HphI xx38 xx56 xx62	HpaII MspI xx06 xx52
O8	301-369	341	353	353	355	356	-
A20	401-469	441	453	453	455	456	-
A30	501-569	541	553	553	555	556	-
L14	601-669	637	641	653	655	656	-
L1	701-769	737	741	753	755	756	-
L15	801-869	837	841	853	855	856	-
L4	901-969	937	941	953	955	956	-
L18	1001-1069	1037	1041	1053	1055	1056	-
L5	1101-1169	1137	1141	1153	1155	1156	-
L19	1201-1269	1237	1241	1253	1255	1256	-
L8	1301-1369	1337	1341	1353	1355	1356	-
L23	1401-1469	1437	1441	1453	1455	1456	1406
L9	1501-1569	1537	1541	1553	1555	1556	1506
L24	1601-1669	1637	1641	1653	1655	1656	
L11	1701-1769	1737	1741	1753	1755	1756	
L12	1801-1869	1837	1841	1853	1855	1856	
PKI							
O11	1901-1969	-	1918	1918	1937	1938	1952
O1	2001-2069	-	2018	2018	2037	2038	2052
A17	2101-2169	-	2112	2112	2137	2138	2152
A1	2201-2269	-	2212	2212	2237	2238	2252

5

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20

	SfaNI	SfcI	HinfI	MlyI --> --> <--	MaeIII Tsp45I same sites	HphI xx38 xx56 xx62	HpaII MspI xx06 xx52
A18 2301-2369	-	-	2318	2318	2337	2338	2352
A2 2401-2469	-	-	2418	2418	2437	2438	2452
A19 2501-2569	-	-	2512	2512	2537	2538	2552
A3 2601-2669	-	-	2612	2612	2637	2638	2652
A23 2701-2769	-	-	2718	2718	2737	2731* 2738*	-
VKII							
A27 2801-2869	-	-	-	-	-	-	-
A11 2901-2969	-	-	-	-	-	-	-
L2 3001-3069	-	-	-	-	-	-	-
L16 3101-3169	-	-	-	-	-	-	-
L6 3201-3269	-	-	-	-	-	-	-
L20 3301-3369	-	-	-	-	-	-	-
L25 3401-3469	-	-	-	-	-	-	-
VKIV							
B3 3501-3569	-	-	3525	3525	-	-	-
VKV							
B2 3601-3669	-	-	3639	3639	-	-	-
VKV							
A26 3701-3769	-	-	3712	3739	3737 3755	3756 3762	-
A10 3801-3869	-	-	3812	3839	3837 3855	3856 3862	-
A14 3901-3969	-	-	3939	3939	3937 3955	3956 3962	-

5

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20



	BsaJI xx29 xx42 xx43	BssKI (NstNI) xx22 xx30 xx43	BpmI xx20 xx41 xx44 --> --> <--	BsrFI Cac8I NaeI NgoMI V	HaeII I	Tsp509I
L11 1701-1769	-	-	-	-	-	-
L12 1801-1869	-	-	-	-	-	-
VIGI						
O11 1901-1969	1942	1943	1944	1951	1954	-
O1 2001-2069	2042	2043	2044	2051	2054	-
A17 2101-2169	2142	-	-	2151	2154	-
A1 2201-2269	2242	-	-	2251	2254	-
A18 2301-2369	2342	2343	-	2351	2354	-
A2 2401-2469	2442	2443	-	2451	2454	-
A19 2501-2569	2542	2543	2544	2551	2554	-
A3 2601-2669	2642	2643	2644	2651	2654	-
A23 2701-2769	2742	-	-	2751	2754	-
VIGI						
A27 2801-2869	2843	2822 2843	2820 2841	-	-	2803
A11 2901-2969	2943	2943	2920 2941	-	-	2903
L2 3001-3069	3043	3043	3041	-	-	-
L16 3101-3169	3143	3143	3120 3141	-	-	-
L6 3201-3269	3243	3243	3220 3241	-	-	3203
L20 3301-3369	3343	3343	3320 3341	-	-	3303

5

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	BsaJI xx29 xx42 xx43	BssKI (NstNI) xx22 xx30 xx43	BpmI xx20 xx41 xx44 --> --> <--<	BsrFI Cac8I NaeI NgoMI V	HaeII I	Tsp509I
L25 3401-3469	3443	3443	3420 3441	-	-	3403
VICV						
B3 3501-3569	3529	3530	3520	-	3554	
GRV						
B2 3601-3669		3643	3620 3641	-	-	
VKVJ						
A26 3701-3769		-	3720	-	-	3703
A10 3801-3869		-	3820	-	-	3803
A14 3901-3969	3943	3943	3920 3941	-	-	-



21 Aug 2007

2007211861

91/128

Table 400 Lambda FR1 GLG sequences

! VL1

5  
 CAG TCT GTG CTG ACT CAG CCA CCC TCG GTG TCT GAA  
 GCC CCC AGG CAG AGG GTC ACC ATC TCC TGT ! 1a  
 cag tct gtg ctg acG cag ccG ccc tcA gtg tct gGG  
 gcc ccA Ggg cag agg gtc acc atc tcc tgC ! 1e  
 cag tct gtg ctg act cag cca ccc tcA gCg tct gGG  
 Acc ccc Ggg cag agg gtc acc atc tcT tgt ! 1c  
 0  
 cag tct gtg ctg act cag cca ccc tcA gCg tct gGG  
 Acc ccc Ggg cag agg gtc acc atc tcT tgt ! 1g  
 cag tct gtg Ttg acG cag ccG ccc tcA gtg tct gCG  
 gcc ccA GgA cag aAg gtc acc atc tcc tgC ! 1b

! VL2

5  
 CAG TCT GCC CTG ACT CAG CCT CCC TCC GCG TCC GGG  
 TCT CCT GGA CAG TCA GTC ACC ATC TCC TGC ! 2c  
 cag tct gcc ctg act cag cct cGc tcA gTg tcc ggg  
 tct cct gga cag tca gtc acc atc tcc tgc! 2e  
 cag tct gcc ctg act cag cct Gcc tcc gTg tcT ggg  
 0  
 tct cct gga cag tcG Atc acc atc tcc tgc ! 2a2  
 cag tct gcc ctg act cag cct ccc tcc gTg tcc ggg  
 tct cct gga cag tca gtc acc atc tcc tgc ! 2d  
 cag tct gcc ctg act cag cct Gcc tcc gTg tcT ggg  
 tct cct gga cag tcG Atc acc atc tcc tgc ! 2b2

! VL3

5  
 TCC TAT GAG CTG ACT CAG CCA CCC TCA GTG TCC GTG  
 TCC CCA GGA CAG ACA GCC AGC ATC ACC TGC! 3r  
 tcc tat gag ctg act cag cca cTc tca gtg tcA gtg  
 Gcc cTG gga cag acG gcc agG atT acc tgT ! 3j  
 0  
 tcc tat gag ctg acA cag cca ccc tcG gtg tcA gtg  
 tcc cca gga caA acG gcc agG atc acc tgc! 3p  
 tcc tat gag ctg acA cag cca ccc tcG gtg tcA gtg  
 tcc cTa gga cag aTG gcc agG atc acc tgc ! 3a  
 tcT tCt gag ctg aet cag GAC ccT GcT gtg tcT gtg  
 Gcc TTG gga cag aca gTc agG atc acA tgc ! 3l

92/128

```

tcc tat gTg ctg act cag cca ccc tca gtg tcA gtg
Gcc cca gga Aag acG gcc agG atT acc tgT ! 3h
tcc tat gag ctg acA cag cTa ccc tcG gtg tcA gtg
tcc cca gga cag aca gcc agG atc acc tgc ! 3e
5 tcc tat gag ctg aTG cag cca ccc tcG gtg tcA gtg
tcc cca gga cag acG gcc agG atc acc tgc ! 3m
tcc tat gag ctg acA cag cca Tcc tca gtg tcA gtg
tcT ccG gga cag aca gcc agG atc acc tgc ! V2-19
! VL4
10 CTG CCT GTG CTG ACT CAG CCC CCG TCT GCA TCT GCC
TTG CTG GGA GCC TCG ATC AAG CTC ACC TGC ! 4c
cAg cct gtg ctg act caA TcA TcC tct gcC tct gcT
tCC ctg gga Tcc tcg Gtc aag ctc acc tgc ! 4a
cAg cTt gtg ctg act caA TcG ccC tct gcC tct gcc
15 tCC ctg gga gcc tcg Gtc aag ctc acc tgc ! 4b
! VL5
CAG CCT GTG CTG ACT CAG CCA CCT TCC TCC TCC GCA
TCT CCT GGA GAA TCC GCC AGA CTC ACC TGC ! 5e
cag Gct gtg ctg act cag ccG Gct tcc CTc tcT gca
20 tct cct gga gCa tcA gcc agT ctc acc tgc ! 5c
cag cct gtg ctg act cag cca Tct tcc CAT tcT gca
tct Tct gga gCa tcA gTc aga ctc acc tgc ! 5b
! VL6
AAT TTT ATG CTG ACT CAG CCC CAC TCT GTG TCG GAG
25 TCT CCG GGG AAG ACG GTA ACC ATC TCC TGC ! 6a
! VL7
CAG ACT GTG GTG ACT CAG GAG CCC TCA CTG ACT GTG
TCC CCA GGA GGG ACA GTC ACT CTC ACC TGT ! 7a
cag Gct gtg gtg act cag gag ccc tca ctg act gtg
30 tcc cca gga ggg aca gtc act ctc acc tgt ! 7b
! VL8
CAG ACT GTG GTG ACC CAG GAG CCA TCG TTC TCA GTG
TCC CCT GGA GGG ACA GTC ACA CTC ACT TGT ! 8a

```

2007211861 21 Aug 2007

93/128

! VL9

CAG CCT GTG CTG ACT CAG CCA CCT TCT GCA TCA GCC  
TCC CTG GGA GCC TCG GTC ACA CTC ACC TGC ! 9a

! VL10

CAG GCA GGG CTG ACT CAG CCA CCC TCG GTG TCC AAG  
GGC TTG AGA CAG ACC GCC ACA CTC ACC TGC ! 10a

5

2007211861 21 Aug 2007

94/128

Table 405 RERSs found in human lambda FR1 GLGs

! There are 31 lambda GLGs

MlyI NnnnnnGACTC 25

1: 6	3: 6	4: 6	6: 6	7: 6	8: 6
9: 6	10: 6	11: 6	12: 6	15: 6	16: 6
20: 6	21: 6	22: 6	23: 6	23: 50	24: 6
25: 6	25: 50	26: 6	27: 6	28: 6	30: 6
31: 6					

There are 23 hits at base# 6

0 -"- GAGTCNNNNNn 1

26: 34

MwoI GCNNNNNngc 20

1: 9	2: 9	3: 9	4: 9	11: 9	11: 56
12: 9	13: 9	14: 9	16: 9	17: 9	18: 9
19: 9	20: 9	23: 9	24: 9	25: 9	26: 9
30: 9	31: 9				

There are 19 hits at base# 9

0 HinfI Gantc 27

1: 12	3: 12	4: 12	6: 12	7: 12	8: 12
9: 12	10: 12	11: 12	12: 12	15: 12	16: 12
20: 12	21: 12	22: 12	23: 12	23: 46	23: 56
24: 12	25: 12	25: 56	26: 12	26: 34	27: 12
28: 12	30: 12	31: 12			

There are 23 hits at base# 12

PleI gactc 25

1: 12	3: 12	4: 12	6: 12	7: 12	8: 12
9: 12	10: 12	11: 12	12: 12	15: 12	16: 12
20: 12	21: 12	22: 12	23: 12	23: 56	24: 12
25: 12	25: 56	26: 12	27: 12	28: 12	30: 12
31: 12					

There are 23 hits at base# 12

5 -"- gagtc 1

26: 34

DdeI Ctnag 32

	1: 14	2: 24	3: 14	3: 24	4: 14	4: 24
	5: 24	6: 14	7: 14	7: 24	8: 14	9: 14
5	10: 14	11: 14	11: 24	12: 14	12: 24	15: 5
	15: 14	16: 14	16: 24	19: 24	20: 14	23: 14
	24: 14	25: 14	26: 14	27: 14	28: 14	29: 30
	30: 14	31: 14				

There are 21 hits at base# 14

10

BsaJI Ccnngg 38

	1: 23	1: 40	2: 39	2: 40	3: 39	3: 40
	4: 39	4: 40	5: 39	11: 39	12: 38	12: 39
	13: 23	13: 39	14: 23	14: 39	15: 38	16: 39
15	17: 23	17: 39	18: 23	18: 39	21: 38	21: 39
	21: 47	22: 38	22: 39	22: 47	26: 40	27: 39
	28: 39	29: 14	29: 39	30: 38	30: 39	30: 47
	31: 23	31: 32				

There are 17 hits at base# 39

20 There are 5 hits at base# 38

There are 5 hits at base# 40 Makes cleavage ragged.

MnlI cctc 35

	1: 23	2: 23	3: 23	4: 23	5: 23	6: 19
	6: 23	7: 19	8: 23	9: 19	9: 23	10: 23
25	11: 23	13: 23	14: 23	16: 23	17: 23	18: 23
	19: 23	20: 47	21: 23	21: 29	21: 47	22: 23
	22: 29	22: 35	22: 47	23: 26	23: 29	24: 27
	27: 23	28: 23	30: 35	30: 47	31: 23	

There are 21 hits at base# 23

30 There are 3 hits at base# 19

There are 3 hits at base# 29

There are 1 hits at base# 26

There are 1 hits at base# 27 These could make cleavage ragged.

-"- gagg 7

35	1: 48	2: 48	3: 48	4: 48	27: 44	28: 44
----	-------	-------	-------	-------	--------	--------

29: 44

BssKI Nccngg 39

	1: 40	2: 39	3: 39	3: 40	4: 39	4: 40
5	5: 39	6: 31	6: 39	7: 31	7: 39	8: 39
	9: 31	9: 39	10: 39	11: 39	12: 38	12: 52
	13: 39	13: 52	14: 52	16: 39	16: 52	17: 39
	17: 52	18: 39	18: 52	19: 39	19: 52	21: 38
	22: 38	23: 39	24: 39	26: 39	27: 39	28: 39
10	29: 14	29: 39	30: 38			

There are 21 hits at base# 39  
 There are 4 hits at base# 38  
 There are 3 hits at base# 31  
 There are 3 hits at base# 40 Ragged

15

BstNI CCwgg 30

	1: 41	2: 40	5: 40	6: 40	7: 40	8: 40
	9: 40	10: 40	11: 40	12: 39	12: 53	13: 40
	13: 53	14: 53	16: 40	16: 53	17: 40	17: 53
20	18: 40	18: 53	19: 53	21: 39	22: 39	23: 40
	24: 40	27: 40	28: 40	29: 15	29: 40	30: 39

There are 17 hits at base# 40  
 There are 7 hits at base# 53  
 There are 4 hits at base# 39  
 25 There are 1 hits at base# 41 Ragged

PspGI ccwgg 30

	1: 41	2: 40	5: 40	6: 40	7: 40	8: 40
	9: 40	10: 40	11: 40	12: 39	12: 53	13: 40
30	13: 53	14: 53	16: 40	16: 53	17: 40	17: 53
	18: 40	18: 53	19: 53	21: 39	22: 39	23: 40
	24: 40	27: 40	28: 40	29: 15	29: 40	30: 39

There are 17 hits at base# 40  
 There are 7 hits at base# 53  
 35 There are 4 hits at base# 39

2007211861 21 Aug 2007

97/128

There are 1 hits at base# 41

ScrFI CCngg 39

1: 41	2: 40	3: 40	3: 41	4: 40	4: 41
5: 40	6: 32	6: 40	7: 32	7: 40	8: 40
9: 32	9: 40	10: 40	11: 40	12: 39	12: 53
13: 40	13: 53	14: 53	16: 40	16: 53	17: 40
17: 53	18: 40	18: 53	19: 40	19: 53	21: 39
22: 39	23: 40	24: 40	26: 40	27: 40	28: 40
29: 15	29: 40	30: 39			

There are 21 hits at base# 40

There are 4 hits at base# 39

There are 3 hits at base# 41

15 MaeIII gtnac 16

1: 52	2: 52	3: 52	4: 52	5: 52	6: 52
7: 52	9: 52	26: 52	27: 10	27: 52	28: 10
28: 52	29: 10	29: 52	30: 52		

There are 13 hits at base# 52

20 Tsp45I gtsac 15

1: 52	2: 52	3: 52	4: 52	5: 52	6: 52
7: 52	9: 52	27: 10	27: 52	28: 10	28: 52
29: 10	29: 52	30: 52			

25 There are 12 hits at base# 52

HphI tcacc 26

1: 53	2: 53	3: 53	4: 53	5: 53	6: 53
7: 53	8: 53	9: 53	10: 53	11: 59	13: 59
14: 59	17: 59	18: 59	19: 59	20: 59	21: 59
22: 59	23: 59	24: 59	25: 59	27: 59	28: 59
30: 59	31: 59				

There are 16 hits at base# 59

There are 10 hits at base# 53

15

21 Aug 2007

2007211861

98/128

BspMI ACCTGCNNNNn

14

11: 61 13: 61 14: 61 17: 61 18: 61 19: 61

20: 61 21: 61 22: 61 23: 61 24: 61 25: 61

30: 61 31: 61

5 There are 14 hits at base# 61 Goes into CDR1

---



Table 500: h3401-h2 captured Via CJ with BsmAI

! 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15  
 ! S A Q D I Q M T Q S P A T L S  
 aGT GCA Caa gac atc cag atg acc cag tct cca gcc acc ctg tct  
 5 ! ApaLI... a gcc acc !  
 L25,L6,L20,L2,L16,A11  
 ! Extender.....Bridge...

! 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30  
 ! V S P G E R A T L S C R A S Q  
 gtg tct cca ggg gaa agg gcc acc ctc tcc tgc agg gcc agt cag

! 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45  
 ! S V S N N L A W Y Q Q K P G Q  
 15 agt gtt agt aac aac tta gcc tgg tac cag cag aaa cct ggc cag

! 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60  
 ! V P R L L I Y G A S T R A T D  
 20 gtt ccc agg ctc ctc atc tat ggt gca tcc acc agg gcc act gat

! 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75  
 ! I P A R F S G S G S G T D F T  
 atc cca gcc agg ttc agt ggc agt ggg tct ggg aca gac ttc act

! 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90  
 ! L T I S R L E P E D F A V Y Y  
 25 ctc acc atc agc aga ctg gag cct gaa gat ttt gca gtg tat tac

! 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105  
 ! C Q R Y G S S P G W T F G Q G  
 30 tgt cag cgg tat ggt agc tca ccg ggg tgg acg ttc ggc caa ggg

! 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120  
 ! T K V E I K R T V A A P S V F  
 35 acc aag gtg gaa atc aaa cga act gtg gct gca cca tct gtc ttc

! 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135  
 ! I F P P S D E Q L K S G T A S  
 40 atc ttc ccg cca tct gat gag cag ttg aaa tct gga act gcc tct

! 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150  
 ! V V C L L N N F Y P R E A K V  
 gtt gtg tgc ctg ctg aat aac ttc tat ccc aga gag gcc aaa gta

21 Aug 2007

2007211861

21 Aug 2007

2007211861

100/128

```

! 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
! Q W K V D N A L Q S G N S Q E
cag tgg aag gtg gat aac gcc ctc caa tcg ggt aac tcc cag gag

5 ! 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
! S V T E Q D S K D S T Y S L S
agt gtc aca gag cag gac agc aag gac agc acc tac agc ctc agc

10 ! 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
! S T L T L S K A D Y E K H K V
agc acc ctg acg ctg agc aaa gca gac tac gag aaa cac aaa gtc

! 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
! Y A C E V T H Q G L S S P V T
15 tac gcc tgc gaa gtc acc cat cag ggc ctg agc tcg cct gtc aca

! 211 212 213 214 215 216 217 218 219 220 221 222 223
! K S F N K G E C K G E F A
aag agc ttc aac aaa gga gag tgt aag ggc gaa ttc gc.....

!0

```

Table 501: h3401-d8 KAPPA captured with CJ and BsmAI

```

! 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
! S A Q D I Q M T Q S P A T L S
aGT GCA Caa gac atc cag atg acc cag tct cct gcc acc ctg tct
! ApaLI...Extender.....a gcc acc !
L25,L6,L20,L2,L16,A11

! A GCC ACC CTG TCT ! L2

10 ! 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
! V S P G E R A T L S C R A S Q
gtg tct cca ggt gaa aga gcc acc ctc tcc tgc agg gcc agt cag
! GTG TCT CCA GGG GAA AGA GCC ACC CTC TCC TGC ! L2

15 ! 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
! N L L S N L A W Y Q Q K P G Q
aat ctt ctc agc aac tta gcc tgg tac cag cag aaa cct ggc cag

!0 ! 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60
! A P R L L I Y G A S T G A I G
gct ccc agg ctc ctc atc tat ggt gct tcc acc ggg gcc att ggt

! 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
! I P A R F S G S G S G T E F T
15 atc cca gcc agg ttc agt ggc agt ggg tct ggg aca gag ttc act

```

## 101/128

21 Aug 2007

2007211861

```

! 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
! L T I S S L Q S E D F A V Y F
ctc acc atc agc agc ctg cag tct gaa gat ttt gca gtg tat ttc

5 ! 91 92 93 94 95 96 97 98 99 100 101 102 103 104 105
! C Q Q Y G T S P P T F G G G T
tgt cag cag tat ggt acc tca ccg ccc act ttc ggc gga ggg acc

0 ! 106 107 108 109 110 111 112 113 114 115 116 117 118 119 120
! K V E I K R T V A A P S V F I
aag gtg gag atc aaa cga act gtg gct gca cca tct gtc ttc atc

5 ! 121 122 123 124 125 126 127 128 129 130 131 132 133 134 135
! F P P S D E Q L K S G T A S V
ttc ccg cca tct gat gag cag ttg aaa tct gga act gcc tct gtt

0 ! 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
! V C P L N N F Y P R E A K V Q
gtg tgc ccg ctg aat aac ttc tat ccc aga gag gcc aaa gta cag

! 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
! W K V D N A L Q S G N S Q E S
tgg aag gtg gat aac gcc ctc caa tcg ggt aac tcc cag gag agt

5 ! 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
! V T E Q D N K D S T Y S L S S
gtc aca gag cag gac aac aag gac agc acc tac agc ctc agc agc

0 ! 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
! T L T L S K V D Y E K H E V Y
acc ctg acg ctg agc aaa gta gac tac gag aaa cac gaa gtc tac

5 ! 196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
! A C E V T H Q G L S S P V T K
gcc tgc gaa gtc acc cat cag ggc ctt agc tcg ccc gtc acg aag

! 211 212 213 214 215 216 217 218 219 220 221 222 223
! S F N R G E C K K E F V
agc ttc aac agg gga gag tgt aag aaa gaa ttc gtt t

0

```

Table 508 Human heavy chains bases 88.1 to 94.2

Number of sequences..... 840

Id	Ntot	Number of Mismatches.....							Name	Sequence.....	Probe	Dot form.....
		0	1	2	3	4	5	6				
1	364	152	97	76	26	7	4	2	0	VHS881-1.1	gctgtgtattactgtgcgag	gctgtgtattactgtgcgag
2	265	150	60	33	13	5	4	0	0	VHS881-1.2	gccgtgtattactgtgcgag	..c.....
3	96	14	34	16	10	5	7	9	1	VHS881-2.1	gccgtataattactgtgcgag	..c..a.....
4	20	0	3	4	9	2	2	0	0	VHS881-4.1	gccgtgtattactgtgcgag	..c.....a.....
5	95	25	36	18	11	2	2	0	1	VHS881-9.1	gccatgtattactgtgcgag	..Ca.....
840		341	230	147	69	21	19	11	2			
341		571	718	787	808	827	838	840				

15 88 89 90 91 92 93 94 95 Codon number as in Table 195

Recognition..... Stem..... Loop. Stem.....  
 (VHS881-1.1) 5'-gctgtgtat|tact-gtgcgag cACATccgTg TTgTT cAcggATgTg-3'  
 (VHS881-1.2) 5'-gccgtgtat|tact-gtgcgag cACATccgTg TTgTT cAcggATgTg-3'  
 (VHS881-2.1) 5'-gccgtatat|tact-gtgcgag cACATccgTg TTgTT cAcggATgTg-3'  
 (VHS881-4.1) 5'-gccgtgtat|tact-gtgcgag cACATccgTg TTgTT cAcggATgTg-3'  
 (VHS881-9.1) 5'-gccatgtat|tact-gtgcgag cACATccgTg TTgTT cAcggATgTg-3'  
 | site of substrate cleavage

20 (FOK)act) 5'-cACATccgTg TTgTT cAcggATgTg-3'

(VHEX881) 5'-AATAGTAGAc TgcAgTgTcc TcAgcccTTA AgcTgTTcAT cTgcAAgTAG-  
 AgAgTATTcT TAGAgTTgTc TcTAGAcTTA gTgAAgcg-3'

! note that VHEX881 is the reverse complement of the ON below

! [RC] 5'-cgCttcacTaag-

! Scab.....

! Synthetic 3-23 as in Table 206

! |TCT|AGA|gac|aac|tct|aac|aat|act|ctc|tac|ttg|cag|atg|/-

! XbaI...

! |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|t-3'

! AfIII...

(VHBA881) 5'-cgCttcacTaag-

! |TCT|AGA|gac|aac|tct|aac|aat|act|ctc|tac|ttg|cag|atg|/-

! |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt gcg ag-3'

(VHBB881) 5'-cgCttcacTaag-

(VH881PCR) 5'-cgCttcacTaag|TCT|AGA|gac|aac -3'  
 |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|ttg|cag|atg|  
 |aac|agC|TTA|AGg|gct|gag|gac|aCT|GCA|Gtc|tac|tat|tgt|Acg|ag-3'

5 Table 512: Kappa, bases 12-30

ID	Ntot	0	1	2	3	4	5	6	Name	Sequence.....	Dot Form.....
1	84	40	21	20	1	2	0	0	SK12012	gaccagtcctccatctcc	gaccagtcctccatctcc
2	32	19	3	6	2	1	0	1	SK12A17	gactcagtcctccactctcc	...t.....ct....
3	26	17	8	1	0	0	0	0	SK12A27	gacgcagtcctccaggcacc	...g.....gg.a..
4	40	21	18	1	0	0	0	0	SK12A11	gacgcagtcctccaggcacc	...g.....g...a..
182	97	50	28	3	3	0	1				
97	147	175	178	181	181	182					

15 URE adapters:

Stem..... Loop. Stem..... Recognition.....  
 (SzKB1230-012) 5'-cAcATccgTg TTgTT cAcgATgTg ggAggATggAgAcTggTc-3'  
 [RC] 5'-gaccagtcctccatctcc cAcATccgTg AACAA cAcgATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

20

Stem..... Loop. Stem..... Recognition.....  
 (SzKB1230-A17) 5'-cAcATccgTg TTgTT cAcgATgTg ggAggATggAgAcTgAgTc-3'  
 [RC] 5'-gactcagtcctccactctcc cAcATccgTg AACAA cAcgATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

25

Stem..... Loop. Stem..... Recognition.....  
 (SzKB1230-A27) 5'-cAcATccgTg TTgTT cAcgATgTg ggTgccTggAgAcTgcgTc-3'  
 [RC] 5'-gacgcagtcctccaggcacc cAcATccgTg AACAA cAcgATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

30

Stem..... Loop. Stem..... Recognition.....  
 (SzKB1230-A11) 5'-cAcATccgTg TTgTT cAcgATgTg ggTggcTggAgAcTgcgTc-3'  
 [RC] 5'-gacgcagtcctccaggcacc cAcATccgTg AACAA cAcgATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

35

What happens in the upper strand:

(SzKB1230-O12\*)  
 5'-gac cca gtc|tcc a-tc ctc c-3'  
 | Site of cleavage in substrate  
 5  
 (SzKB1230-A17\*)  
 5'-gac tea gtc|tcc a-ct ctc c-3'  
 (SzKB1230-A27\*)  
 5'-gac gca gtc|tcc a-gg cac c-3'  
 10 (SzKB1230-A11\*)  
 5'-gac gca gtc|tcc a-gc cac c-3'  
 (kapextURE) 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg-3' Isense strand  
 Scab.....ApaLI.  
 15 (kapextUREPCR) 5'-ccTctactctTgTcAcAgTg-3'  
 Scab.....  
 (kaBRO1UR) 5'-ggAggATggA cTggATgTcT TgTgcActTgT gAcAAGAgTA gAgg-3'  
 [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-tc ctc c-3' ON above is R.C. of this one  
 20 (kaBRO2UR) 5'-ggAggATggA cTggATgTcT TgTgcActTgT gAcAAGAgTA gAgg-3'  
 [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-ct ctc c-3' ON above is R.C. of this one  
 (kaBRO3UR) 5'-ggTgccTggA cTggATgTcT TgTgcActTgT gAcAAGAgTA gAgg-3'  
 [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gg cac c-3' ON above is R.C. of this one  
 (kaBRO4UR) 5'-ggTgccTggA cTggATgTcT TgTgcActTgT gAcAAGAgTA gAgg-3'  
 [RC] 5'-ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gc cac c-3' ON above is R.C. of this one  
 25 | Scab.....ApaLI.

(VH881PCR) 5'-cgCttcacTaaG|TCT|AGA|gac|aac -3'  
 |TCT|AGA|gac|aac|tct|aat|act|ctc|tac|ttg|cag|atg|-  
 |aac|agC|TTA|AGg|gct|gag|gac|act|GCA|Gtc|tac|tat|tgt|Acg|ag-3'

5 Table 512: Kappa, bases 12-30

ID	Ntot	0	1	2	3	4	5	6	Name	Sequence	Dot Form
1	84	40	21	20	1	2	0	0	SK12012	gaccagtcctccatcctcc	gaccagtcctccatcctcc
2	32	19	3	6	2	1	0	1	SK12A17	gactcagtcctccactctcc	...t.....ct....
3	26	17	8	1	0	0	0	0	SK12A27	gacgcagtcctccaggcacc	...g.....gg.a...
4	40	21	18	1	0	0	0	0	SK12A11	gacgcagtcctccaggcacc	...g.....g.a...

15 URE adapters:

(SzKB1230-O12) Stem..... Loop. Stem..... Recognition.....  
 5'-cAcATccgTg TTgTT cAcggATgTg ggAggATggAgAcTgggTc-3'  
 [RC] 5'-gaccagtcctccactctcc cAcATccgTg AACAA cAcggATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

(SzKB1230-A17) Stem..... Loop. Stem..... Recognition.....  
 5'-cAcATccgTg TTgTT cAcggATgTg ggAggATggAgAcTgAgTc-3'  
 [RC] 5'-gactcagtcctccactctcc cAcATccgTg AACAA cAcggATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

(SzKB1230-A27) Stem..... Loop. Stem..... Recognition.....  
 5'-cAcATccgTg TTgTT cAcggATgTg ggTgccTggAgAcTgTc-3'  
 [RC] 5'-gacgcagtcctccaggcacc cAcATccgTg AACAA cAcggATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

(SzKB1230-A11) Stem..... Loop. Stem..... Recognition.....  
 5'-cAcATccgTg TTgTT cAcggATgTg ggTggcTggAgAcTgTc-3'  
 [RC] 5'-gacgcagtcctccaggcacc cAcATccgTg AACAA cAcggATgTg-3'  
 Recognition..... Stem..... loop. Stem.....  
 FokI.

What happens in the upper strand:

(SzKB1230-O12\*)  
 !  
 !  
 5' -gac cca gtc|tcc a-tc ctc c-3'  
 | Site of cleavage in substrate  
 (SzKB1230-A17\*)  
 !  
 !  
 5' -gac tca gtc|tcc a-ct ctc c-3'  
 (SzKB1230-A27\*)  
 !  
 !  
 5' -gac gca gtc|tcc a-gg cac c-3'  
 (SzKB1230-A11\*)  
 !  
 !  
 5' -gac gca gtc|tcc a-gc cac c-3'

(kapextURE) 5' -ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg-3' |sense strand  
 Scab.....ApaLI.

15 (kapextUREPCR) 5' -ccTctactctTgTcAcAgTg-3'  
 Scab.....

(kaBRO1UR) 5' -ggAggATggA cTggATgTcT TgTgcAcTgT gAcAAgAgTA gAgg-3'  
 ! [RC] 5' -ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-tc ctc c-3' ON above is R.C. of this one  
 (kaBRO2UR) 5' -ggAggATggA cTggATgTcT TgTgcAcTgT gAcAAgAgTA gAgg-3'  
 ! [RC] 5' -ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-ct ctc c-3' ON above is R.C. of this one  
 (kaBRO3UR) 5' -ggTgccTggA cTggATgTcT TgTgcAcTgT gAcAAgAgTA gAgg-3'  
 ! [RC] 5' -ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gg cac c-3' ON above is R.C. of this one  
 (kaBRO4UR) 5' -ggTggcTggA cTggATgTcT TgTgcAcTgT gAcAAgAgTA gAgg-3'  
 ! [RC] 5' -ccTctactctTgTcAcAgTgcAcAA gAc ATc cAg tcc a-gc cac c-3' ON above is R.C. of this one  
 Scab.....ApaLI.



Table 515 Lambda URE adapters bases 13.3 to 19.3

Id	Ntot	Number of mismatches.....								Name	Sequence.....	Dot form.....
		0	1	2	3	4	5	6	7			
5	58	45	7	1	0	0	2	2	1	VL133-2a2	gtctcctggacagtcgatac	gtctcctggacagtcgatac
	16	10	1	0	1	0	1	0	2	VL133-31	ggccttgggacagacagtc	.g.cttg.....a.ag..
	17	6	0	0	0	4	1	1	5	VL133-2c	gtctcctggacagtcagtc	.....ag..
	37	3	0	10	4	4	3	7	4	2	VL133-1c	ggccccagggcagagggtc
128	64	8	11	5	8	5	11	11	5			
64	72	83	88	96	101	112	123	128				
15	(VL133-2a2)	Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gATcgAcTgTccAggAgAc-3'										
		[RC] 5'-gtctcctggacagtcagtc <u>cAcATccgTg</u> AACAA <u>cAcgATgTg</u> -3'										
		Recognition..... Stem..... Loop. Stem.....										
20	(VL133-31)	Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gAcTgTcTgTcccAagcc-3'										
		[RC] 5'-ggccttgggacagacagtc <u>cAcATccgTg</u> AACAA <u>cAcgATgTg</u> -3'										
		Recognition..... Stem..... Loop. Stem.....										
25	(VL133-2c)	Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gAcTgAcTgTccAggAgAc-3'										
		[RC] 5'-gtctcctggacagtcagtc <u>cAcATccgTg</u> AACAA <u>cAcgATgTg</u> -3'										
		Recognition..... Stem..... Loop. Stem.....										
30	(VL133-1c)	Stem..... loop. Stem..... Recognition.....										
		5'-cAcATccgTg TTgTT cAcgATgTg gAcctTgcccTggggcc-3'										
		[RC] 5'-ggccccagggcagagggtc <u>cAcATccgTg</u> AACAA <u>cAcgATgTg</u> -3'										

2007211861 21 Aug 2007

108/128

What happens in the top strand:

! site of cleavage in the upper strand

5 (VL133-2a2\*) 5'-g tct cct g|ga cag tcg atc  
 !  
 (VL133-3l\*) 5'-g gcc ttg g|ga cag aca gtc  
 !  
 (VL133-2c\*) 5'-g tct cct g|ga cag tca gtc  
 10 (VL133-1c\*) 5'-g gcc cca g|gg cag agg gtc

! The following Extenders and Bridges all encode the AA sequence of 2a2 for codons 1-15

15 (ON\_LamEx133) 5'-ccTcTgAcTgAgT <sup>1</sup>gcA cAg -  
 !  
 2 3 4 5 6 7 8 9 10 11 12  
 AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-  
 !  
 20 13 14 15  
 tcC ccG g ! 2a2

(ON\_LamB1-133) [RC] 5'-ccTcTgAcTgAgT <sup>1</sup>gcA cAg -  
 25 !  
 2 3 4 5 6 7 8 9 10 11 12  
 AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-  
 !  
 13 14 15  
 tcC ccG g ga cag tcg at-3' ! 2a2

30 ! N.B. the actual seq is the reverse complement of the one shown.

(ON\_LamB2-133) [RC] 5'-ccTcTgAcTgAgT <sup>1</sup>gcA cAg -  
 35 !  
 2 3 4 5 6 7 8 9 10 11 12  
 AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-  
 !  
 13 14 15  
 tcC ccG g ga cag aca gt-3' ! 3l

40 ! N.B. the actual seq is the reverse complement of the one shown.

(ON\_LamB3-133) [RC] 5'-ccTcTgAcTgAgT <sup>1</sup>gcA cAg -  
 45 !  
 2 3 4 5 6 7 8 9 10 11 12  
 AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-  
 !  
 13 14 15  
 tcC ccG g ga cag tca gt -3' ! 2c

50 ! N.B. the actual seq is the reverse complement of the one shown.

(ON\_LamB4-133) [RC] 5'-ccTcTgAcTgAgT <sup>1</sup>gcA cAg -  
 55 !

!

2007211861 21 Aug 2007

109/128

! 2 3 4 5 6 7 8 9 10 11 12  
AGt gcT TtA acC caA ccG gcT AGT gtT AGC ggT-

! 13 14 15  
! 5 tCC ccG g gg cag agg gt-3' ! 1c

N.B. the actual seq is the reverse complement of the one shown.

(ON\_Laml33PCR) 5'-ccTcTgAcTgAgT gca cAg AGt gc-3'

Table 525 ONs used in Capture of kappa light chains using CJ method and BsmAI

All ONs are written 5' to 3'.

5	REdapters (6)	
	ON_20SK15012	gggAggATggAgAcTgggTc
	ON_20SK15L12	gggAAGATggAgAcTgggTc
	ON_20SK15A17	gggAgAgTggAgAcTgAgTc
	ON_20SK15A27	gggTgccTggAgAcTgcTc
10	ON_20SK15A11	gggTggcTggAgAcTgcTc
	ON_20SK15B3	gggAgTcTggAgAcTgggTc
	Bridges (6)	
15	kapbr11012	gggAggATggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
	kapbr11L12	gggAAGATggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
	kapbr11A17	gggAgAgTggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
	kapbr11A27	gggTgccTggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
	kapbr11A11	gggTggcTggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
20	kapbr11B3	gggAgTcTggAgAcTgggTcATcTggATgTcTTgTgcAcTgTgAcAgAgg
	Extender (5' biotinylated)	
	kapext1bio	ccTcTgTcAcAgTgcAcAAGAcATccAgATgAcccAgTcTcc
	Primers	
25	kaPCRt1	ccTcTgTcAcAgTgcAcAAGAc
	kapfor	5'-aca ctc tcc cct gtt gaa gct ctt-3'

Table 530

PCR program for amplification of kappa DNA

- 95°C 5 minutes
- 95°C 15 seconds
- 65°C 30 seconds

72°C 1 minute  
72°C 7 minutes  
4°C hold

5 Reagents (100 ul reaction):

Template	50 ng
10x turbo PCR buffer	1x
turbo Pfu	4U
dNTPs	200 µM each
10 kaPCRt1	300 nM
kapfor	300 nM

Table 610: Stuffer used in VH

1 TCCGGAGCTT CAGATCTGTT TGCCTTTTTG TGGGGTGGTG CAGATCGCGT TACGGAGATC  
 61 GACCGACTGC TTGAGCAAAA GCCACGCTTA ACTGCTGATC AGGCATGGGA TGTATTTCCG  
 121 CAATCCAGTC GTCAGGATCT TAACCTGAGG CTTTTTTTAC CTACTCTGCA AGCAGCGACA  
 181 TCTGGTTTTGA CACAGAGCGA TCCGCGTGGT CAGTTGGTAG AAACATTAAC ACGTTGGGAT  
 241 GGCATCAATT TGCITTAATGA TGATGGTAAA ACCTGGCAGC AGCCAGGCTC TGCCATCCTG  
 301 AACGTTGGC TGACCAGTAT GTTGAAGCGT ACCGTAGTGG CTGCCGTACC TATGCCAATTT  
 361 GATPAAAGTG ACAGCGCCAG TGGCTACGAA AATTTTGTAT GAGCGGTGC AGGGAGACAA ATCACCNAATC  
 421 AATATAAGTG TTGGAGCAAA AATTTTGTAT TGCTGGGAAA CCACAGCAGG AGGTTGTGTT GGCTGGCGTGT  
 481 CCACATACCT GGGAGACTCT TTCCAAAACGC TATGGCAATA ATGTGAGTAA CTGGAAACA  
 541 GAAGATACCT CTTAACCCTT CCGGGCAAAAT AATTTCTTTG GTGTACCCTCA GCGCCGACGG  
 601 CCTGCAATGG CTTAACCCTT CCGGGCAAAAT AATTTCTTTG GTGTACCCTCA GCGCCGACGG  
 661 GAAGAAACGC GTCATCAGGC GGAGTATCAA AACCGTGGAA CAGAAAACCGA TATGATTTGTT  
 721 TTCTCACCAA CGACAAGCGA TCGTCCCTGTG CTTGCCCTGGG ATGTGGTCCG ACCCGGTCCAG  
 781 AGTGGGTTTTA TTGCTCCCGA TGGAAACAGTT GATPAAAGCACT ATGAAGATCA GCTGAAAATG  
 841 TACGAAAAAAT TTGGCCGTAA GTCGCTCTGG TTAACGAAAGC AGGATGTGGA GCGCCATPAA  
 901 GAGTCGTCTA GA

Table 620: DNA sequence of pCESS  
 ! pCESS 6680 bases = pCes4 with stuffers in CDR1-2 and CDR3 2000.12.13

```

! Ngene = 6680
! Useful REs (cut MnoLI fewer than 3 times) 2000.06.05
!
! Non-cutters
!Acc65I Ggtacc      AfeI AGCgct      AvrII Cctagg
!BsaBI GATNnnnatac BsiWI Cgtacc     BsmFI Nnnnnnnnnnnngtccc
!BsrGI Tgtaca      BstAPI GCANNntgc BstBI Ttcgaa
!BstZ17I GTAtac   Btri CACgtg     Ecl136I GAGctc
!EcoRV GAtatc     Fsei GGCCGgccc KpnI GGTACC
!MscI TGGcca      NruI TCGcga     NsiI ATGCAT
!PacI TTAATtaa    PmeI GTTaaac   PmlI CACgtg
!PpuMI RGGwccy    PshAI GACNNngtc SacI GAGCTC
!SacII CCGCgg     SbfI CTGCAgg   SexAI Accwgg
!SgfI CCGATcgc   SnaBI TACgta   SpeI Actagt
!SphI GCATGC     Sse8387I CCTGCagg StuI AGGcct
!SwaI ATTaaat    XmaI Cccggg
!
! cutters
! Enzymes that cut more than 3 times.
!AlwNI CAGNNctg   5
!BsgI ctgcac     4
!BsrFI Rccggy    5
!EarI CTCTTCNnnh 4
!FauI nNNNNNGCGG 10
!
! Enzymes that cut from 1 to 3 times.
!EcoO109I Rggnccy 3 7 2636 4208
!BssSI Ctcgtg    1 12
!"- Caccgag      1 1703
!BspHI Tcatga    3 43 148 1156
!AatII GACGTC    1 65
!BciVI GTATCCNNNN 2 140 1667
!Eco57I CTGAAG   1 301
!"- cttcag       2 1349
!AvaI Cycgrg     3 319 2347 6137
!BsiHKAI GWGCWc  3 401 2321 4245
!HgiAI GWGCWc    3 401 2321 4245
!BcgI gcannnnntcg 1 461
!ScaI AGTact     1 505

```

5	!PvuI CGATcg	3	616	3598	5926
	!FspI TCGcga	2	763	5946	
	!BglI GCCNNNnggc	3	864	2771	5952
	!BpmI CTGGAG	1	898		
	!-"- ctccag	1	4413		
	!BsaI GGTCTCnlnnn	1	916		
	!AhdI GACNNNngtc	1	983		
	!Eam1105I GACNNNngtc	1	983		
10	!DrdI GACNNNngtc	3	1768	6197	6579
	!SapI gaagagc	1	1998		
	!PvuII CAGctg	3	2054	3689	5896
	!PflMI CCANNNntgg	3	2233	3943	3991
	!HindIII Aagctt	1	2235		
	!ApaLI Gtgcac	1	2321		
	!BspMI Nnnnnnnngcaggt	1	2328		
15	!-"- ACCTGCNNNn	2	3460		
	!PstI CTGCAG	1	2335		
	!AccI Gtmkac	2	2341	2611	
	!HincII GtYrac	2	2341	3730	
20	!SalI Gtcgac	1	2341		
	!TliI Ctcgag	1	2347		
	!XhoI Ctcgag	1	2347		
	!BbsI gtcttc	2	2383	4219	
	!BlpI Gctnagc	1	2580		
25	!EspI Gctnagc	1	2580		
	!SgrAI CRccggyg	1	2648		
	!AgeI Accggt	2	2649	4302	
	!AscI Ggcgcgcc	1	2689		
	!BssHII Gcgcgc	1	2690		
30	!BfII GGCCNNNnggccc	1	2770		
	!NaeI GCCggc	2	2776	6349	
	!NgomIV Gccggc	2	2776	6349	
	!BtgI Ccrygg	3	2781	3553	5712
	!DsaI Ccrygg	3	2781	3553	5712
35	!NcoI Coatgg	1	2781		
	!StyI Ccwwgg	3	2781	4205	4472
	!MfeI Caattg	1	2795		
	!BspEI Tccgga	1	2861		
	!BglII Agatct	1	2872		
40	!BclI Tgatca	1	2956		
	!Bsu36I CCTnagc	3	3004	4143	4373
	!XcmI CCANNNNnnntgg	1	3215		
	!MluI Acgpgt	1	3527		



```

!HpaI GTTaaC      1 3730
!XbaI Tctaga      1 3767
!
!AflII Cttaag     1 3811
!BsmI NGcattc     1 3821
!"- GAATGCN       1 4695
!RsrII CGyccg     1 3827
!NheI Gotagc      1 4166
!BstEII Ggtnacc   1 4182
!SmbI CGTCTCnmmn  2 4188 6625
!"- Nnnnnngagacg  1 6673
!ApaI GGGCCc      1 4209
!BamII GRGcYc     3 4209 4492 6319
!Bsp120I Gggccc   1 4209
!PspOMI Gggccc    1 4209
!BseRI Nnnnnnnnctcctc 1 4226
!"- GAGGAGNnnnnnnnn 1 4957
!EcoNI CCTNnnnagg 1 4278
!PflFI GACNngtc   1 4308
!Tth111I GACNngtc 1 4308
!KasI Ggcgcc      2 4327 5967
!BstXI CCANNNNntgg 1 4415
!NotI GCggcccg    1 4507
!EagI Cggccg      1 4508
!BamHI Ggatcc     1 5169
!BspDI ATcgat     1 5476
!NdeI CATatg      1 5672
!EcoRI Gaattc     1 5806
!PsiI TTAtaa      1 6118
!DraIII CACNNngtg 1 6243
!BsaI YACgtr      1 6246
!-----
! 1 gacgaaaggg cCTCGTGata cgcctatttt tataggttaa tgtcatgata ataatggttt
! BssSI.(1/2)
! 61 cttaGACGTC aggtggcaact ttctggggaa atgtgcgcg aaccctatt tgtttatttt
! AatII.
! 121 tctaaataca ttcaaatatG TATCCGctca tgagacaata accctgataa atgcttcaat
! BciVI..(1 of 2)
! 181 aatattgaaa aaggaagagt
! Base # 201 to 1061 = Apr gene from pUC119 with some RE sites removed
!
! 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
! FM S I Q H F R V A L I P F A

```



```

651 cac aac atg ggg gat cat gta act cgc ctt gat cgt tgg gaa ccg
!
!
!
5 166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
! E L N E A I P N D E R D T M
! gag ctg aat gaa gcc ata cca aac gac gag cgt gac acc acg atg
!
!
!
10 181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
! P V A M A T L R K L L T G E
! cct gta GCA ATG gca aca acg tTG CGC Aaa cta tta act ggc gaa
! BsrDI...(1/2) FspI.... (1/2)
!
!
!
15 196 197 198, 199 200 201 202 203 204 205 206 207 208 209 210
! L L T L A S R Q Q L I D W M E
! cta ctt act cta gct tcc cgg caa caa tta ata gac tgg atg gag
!
!
!
20 211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
! A D K V A G P L L R S A L P A
! gcg gat aaa gtt gca gga cca ctt ctg cgc tcg gcc ctt ccg gct
!
!
!
25 226 227 228 229 230 231 232 233 234 235 236 237 238 239 240
! G W F I A D K S G A G E R G S
! ggc tgg ttt att gct gat aaa tCT GGA Gcc ggt gag cgt GGG TCT
! BpmI....(1/2) BsaI.....
!
!
!
30 241 242 243 244 245 246 247 248 249 250 251 252 253 254 255
! R G I I A A L G P D G K P S R
! Cgc ggt atC ATT Gca gca ctg ggg cca gat ggt aag ccc tcc cgt
! BsaI..... BsrDI...(2/2)
!
!
!
35 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270
! I V V I Y T T G S Q A T M D E
! atc gta gtt atc tac acG ACg ggg aGT Cag gca act atg gat gaa
! Ahdi.....
!
!
!
40 271 272 273 274 275 276 277 278 279 280 281 282 283 284 285
! R N R Q I A E I G A S L I K H
! cga aat aga cag atc gct gag ata ggt gcc tca ctg att aag cat
!
!
!
286 287
! W .
! tgg taa
!
!
1056
1062
1081 catatatact ttagattgat ttaaacttc atttttaatt taaaggatc tagtgaaga
! ctgtcagac caagttact

```

```

1141 tccttttga taatctcatg accaaaaatcc cttaacgtga gttttcgttc cactgagcgt
1201 cagaccccggt agaaaagatc aaaggatctt cttgagatcc tttttttctg cgcgtaactc
1261 gctgcttgca aacaaaaaaa ccaccgctac cagcgggtgt ttgtttgccg gatcaagagc
1321 taceactct ttttccgaag gtaactggct tcagcagagc gcagatacca aatactgtcc
1381 ttctagtga gccgtagtta gcccaccact tcaagaactc thtagcaccg cctacatacc
1441 tgcctctgct aatcctgtta ccagtggctg ctgccagtgg cgataagtgc tgtctttaccg
1501 ggttgactc aagacgatag ttaccggata aggcgcagcg gtcgggctga acgggggggtt
1561 cgtgcataca gccacgcttg gagcgaacga cctacaccca actgagatac ctacagcgtg
1621 agcattgaga aagcgcacag ctttcccgaag ggagaaaagc ggacagGTAT CCggtaaagc
      BciVI.. (2 of 2)
1681 gcaggggtcgg aacagggagag cgCACGAGgg agcttccagg gggaaaacgcc tggatatcttt
      BssSI.(2/2)
1741 atagtcctgt cgggtttcgc cacctctgac ttgagcgtcg atttttgtga tgctcgtcag
1801 gggggcggag cctatggaaa aacgccagca acgcggcctt tttacggttc ctggcctttt
1861 gctggcctt tgctCACATG Ttctttctg cgttatcccc tgattctgtg gataaccgta
      PciI...
1921 ttaccgcctt tgagtgagct gatacgcctc gccgcagccg aacgaccgag cgcagcaggt
1981 cagtgagcga ggaagcggAA GAGCGccaa tacgcaaac ccctctcccc gcgcgttggc
      SapI....
2041 cgattcatta atGCAGCTGg cagcagcaggt ttccccactg gaaagcgggc agtgagcgc
      PvuII.(1/3)
2101 acgcaatTAA TGTgagttag ctcactcatt aggcacccca ggCTTTACAC tttatgcttc
      ...-35.. Plac
2161 cggctcgtat gttgtgtgga atttgagcg gataacaatt tcacaCAGGA AACAGCTATG
      M13Rev_seq_primer
2221 Accatgatta cgCCAAGCTT TGGagccttt tttttggaga ttttcaac
      PflMI.....
      Hind3.
signal::linker::CLight
30 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
fm K K L L F A I P L V V P F Y
gtg aaa aaa tta tta ttc gca att cct tta gtt gtt cct ttc tat

35 Linker..... End of FR4
16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
S H S A Q V Q L Q V D L E I K
tct cac aGT GCA Cag gtc caa CTG CAG GTC GAC CTC GAG atc aaa
      ApaI..... PstI... XhoI...
      BspMI... Sali...
40 ACCI...(1/2)
      HincII.(1/2)

```



```

2701 ctatttcaag gagacagtca ta
|
| PeIB::3-23(stuffed)::CH1::III fusion gene
|
5 | 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
| M K Y L L P T A A A G L L L L
| 2723 atg aaa tac.cta ttg cct acg gca gcc gct gga ttg tta tta ctc
|-----
|
| 16 17 18 19 20 21 22
| A A Q P A M A
| 2768 gcG GCC cag ccG GCC atg gcc
| SfiI.....
| NgoMIV..(1/2)
| NcoI....
|
| 2789
|
|-----FR1-----
| 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45
| G G L V Q P G G S L R L S C A
| 2813 |ggc|ggg|ctt|gtt|cag|cct|ggg|ggg|tct|tta|cgt|cct|tct|tgc|gct|
|-----FR1-----
| 46 47 48
| A S G
| 2858 |gct|TCC|GGA|
| | BspEI |
|
| 35 | Stuffer for CDR1, FR2, and CDR2----->
| | There are no stop codons in this stuffer.
|
| 2867 gcttcAGATC Tgtttgcctt
| | BglII..
| 2887 tttgtggggt ggtgcagatc gcgttacgga gatcgaccga ctgcttgagc aaaagccacg
| 2947 cttaactgCT GATCAGgcat gggatgttat tcgccaacc agtcgacagg atcttaacct
| | BclI....
| 3007 gagcgttttt ttacctactc tgaagcagc gacatctggt ttgacacaga gcgatccgcg
| 3067 tcgtcagttg gtagaacat taacacgctg ggatggcacc aatttgctta atgatgatgg

```

```

3127 taaaacctgg cagcagccag gctctgccat cctgaacggtt tggctgacca gstatgtgaa
3187 gcgtaccgta gtggctgccc tacctatgCC Attgataag TGGtacagcg ccagtggcta
      XcmI.....
3247 cgaaacaacc caggcggcc caactggttc gctgaatata agtgttggag caaaaatttt
3307 gtatgagcgg gtgcagggag acaaatcacc aatcccacag cgggttgatc tgtttgctgg
3367 gaaaccacag caggaggttg tgttggctgc gctggaagat acctgggaga ctcttccaa
3427 acgctatggc aataatgtga gtaactggaa acaactgca atggccttaa cgttccgggc
3487 aaataatttc tttggtgtac cgcaggccc agcggaagaa ACGCGTcatc aggcggagta
      MluI..
3547 tcaaaaacct ggaacagaaa acgatattgat tgttttctca ccaacgacaa gcgatcgtcc
3607 tgtgcttgcc tgggatgtgg tcgcaccccg tcagagtggg tttattgctc ccgatggaac
3667 agttgataag cactatgaag atcagctgaa aatgtacgaa aattttggcc gtaagtctct
      PvuII.
3727 ctgGTTAACg aagcaggatg tggaggcgcga taaggagctcg
      HpaI..
      HincII(2/2)
-----FR3-----
      4 5 6 7 8 9 10 11 12 13 14 15 16
      93 94 95 96 97 98 99 100 101 102 103 104 105
      . S R D N S K N T L Y L Q M
3767 |TCT|AGA|gac|aac|tct|aag|aat|act|ctc|tac|tgg|cag|atg|
      | XbaI |
-----FR3-----
      17 18 19 20
      106 107 108 109
      N S L S l s i r s g
3806 |aac|agC|TTA|AG t ctg agc att CGG TCC G
      |AFIII |
      |RsrII..
      q h s p t .
3834 gg caa cat tct cca aac tga ccagacga cacaaacggc
3872 ttacgctaaa tcccgcgat gggatgtaa agagtgggcg tctttgctgg cctggactca
3932 tcagatgaag gccaaaaatt ggcaggagtg gacacagcag gcagcgaac aagcactgac
3992 catcaactgg tactatctg atgtaaacgg caatatgggt tatgttcata ctggtgctta
4052 tccagatcgt caatcaggcc atgatcccgc attaccctgt cctggtacgg gaaaatggga
4112 ctggaaaagg ctattgctt ttgaaatgaa ccctaagggtg tataaccccc ag
4164 aa GCTAGC ctgcggcttc
      NheI..
4182 G|GTC|ACC|
      | BstEII |
      gtc tca agc

```

```

5 136 137 138 139 140 141 142 143 144 145 146 147 148 149 150
   A S T K G P S V F P L A P S S
   gcc tcc acc aag ggc cca tcg gtc ttc ccc ctg gca ccc tcc tcc

4198
151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
   K S T S G G T A A L G C L V K
   aag agc acc tct ggg ggc aca gcg gcc ctg ggc tgc ctg gtc aag

4243
166 167 168 169 170 171 172 173 174 175 176 177 178 179 180
   D Y F P E P V T V S W N S G A
   gac tac ttc ccc gaa ccg gta acg gtc gtc tgg aac tca ggc gcc

4288
181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
   L T S G V H T F P A V L Q S S
   ctg acc agc ggc gtc cac acc ttc ccg gct gtc cta cag tcc tca

4333
196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
   G L Y S L S S V V T V P S S S
   gga ctc tac tcc ctc agc agc gta gtc acc gtc ccc tcc agc agc

4378
211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
   L G T Q T Y I C N V N H K P S
   ttg ggc acc cag acc tac atc tgc aac gtc aat cac aag ccc agc

4423
226 227 228 229 230 231 232 233 234 235 236 237 238
   N T K V D K K V E P K S C
   aac acc aag gtc gac aag AAA GTT GAG CCC AAA TCT TGT
   ON-TQHCforw.....

30 Poly His linker
   139 140 141 142 143 144 145 146 147 148 149 150
   A A A H H H H H H G A A
   GCG GCC GCA cat cat cac cat cac ggg gcc gca

35 NotI.....
   EagI.....

40 151 152 153 154 155 156 157 158 159 160 161 162 163 164 165
   E Q K L I S E E D L N G A A
   gaa caa aaa ctc atc tca gaa gag gat ctg aat ggg gcc gca tag

Mature III----->....
166 167 168 169 170 171 172 173 174 175 176 177 178 179 180

```



```

!   T V E S C L A K P H T E N S F
4588 act gtt gaa agt tgt tta gca aaa cct cat aca gaa aat tca ttt
!
!   181 182 183 184 185 186 187 188 189 190 191 192 193 194 195
5 !   T N V W K D D K T L D R Y A N
!   act aac gtc tgg aaa gac gac aaa act tta gat cgt tac gct aac
!
!   196 197 198 199 200 201 202 203 204 205 206 207 208 209 210
10 !   Y E G C L W N A T G V V C T
!   tat gag ggc tgt ctg tgg AAT GCt aca ggc gtt gtg gtt tgt act
!           BsmI....
!
!   211 212 213 214 215 216 217 218 219 220 221 222 223 224 225
15 !   G D E T Q C Y G T W V P I G L
!   ggt gac gaa act cag tgt tac ggt aca tgg gtt cct att ggg ctt
!
!   226 227 228 229 230 231 232 233 234 235 236 237 238 239 240
!   A I P E N E G G G S E G G S
!   gct atc cct gaa aat gag ggt ggt ggc tct gag ggt ggc ggt tct
!
!   241 242 243 244 245 246 247 248 249 250 251 252 253 254 255
!   E G G G S E G G G T K P P E Y
!   gag ggt ggc ggt tct gag ggt ggc ggt act aaa cct cct gag tac
!
!   256 257 258 259 260 261 262 263 264 265 266 267 268 269 270
25 !   G D T P I P G Y T Y I N P L D
!   ggt gat aca cct att ccg ggc tat act tat atc aac cct ctc gac
!
!   271 272 273 274 275 276 277 278 279 280 281 282 283 284 285
30 !   G T Y P P G T E Q N P A N P N
!   ggc act tat ccg cct ggt act gag caa aac ccc gct aat cct aat
!
!   286 287 288 289 290 291 292 293 294 295 296 297 298 299 300
!   P S L E E S Q P L N T F M F Q
35 !   cct tct ctt GAG GAG tct cag cct ctt aat act ttc atg ttt cag
!           BseRI.. (2/2)
!
!   301 302 303 304 305 306 307 308 309 310 311 312 313 314 315
!   N N R F R N R Q G A L T V Y T
!   aat aat agg ttc cga aat agg cag ggt gca tta act gtt tat acg
!
!   316 317 318 319 320 321 322 323 324 325 326 327 328 329 330
40 !   G T V T T Q G T D P V K T Y Q

```

```

5038 ggc act gtt act caa ggc act gac ccc gtt aaa act tat tac cag
|
|
|
5 331 332 333 334 335 336 337 338 339 340 341 342 343 344 345
| Y T P V S S K A M Y D A Y W N
| tac act cct gta tca tca aaa gcc atg tat gac gct tac tgg aac
|
|
|
10 346 347 348 349 350 351 352 353 354 355 356 357 358 359 360
| G K F R D C A F H S G F N E D
| 99t aaa ttc aga gac tgc gct ttc cat tct ggc ttt aat gag GAT
| BamHI..
|
|
|
15 361 362 363 364 365 366 367 368 369 370 371 372 373 374 375
| P F V C E Y Q G Q S S D L P Q
| 5173 Cca ttc gtt tgt gaa tat caa ggc caa tcg tct gAC CTG Cct caa
| BspMI...(2/2)
|
|
|
20 376 377 378 379 380 381 382 383 384 385 386 387 388 389 390
| P P V N A G G G S G G S G G S G G
| 5218 cct cct gtc aat gct ggc ggc tct ggt ggc tct ggt tct ggt ggc
|
|
|
25 391 392 393 394 395 396 397 398 399 400 401 402 403 404 405
| G S E G G G S E G G G S E G G
| 5263 ggc tct gag ggt ggc ggc tct gag ggt ggc tct gag ggt ggc
|
|
|
30 406 407 408 409 410 411 412 413 414 415 416 417 418 419 420
| G S E G G G S G G G S G S G S G D
| 5308 ggc tct gag ggt ggc ggt tcc ggt ggc ggc tcc ggt tcc ggt gat
|
|
|
35 421 422 423 424 425 426 427 428 429 430 431 432 433 434 435
| F D Y E K M A N A N K G A M T
| 5353 ttt gat tat gaa aaa atg gca aac gct aat aag ggg gct atg acc
|
|
|
40 436 437 438 439 440 441 442 443 444 445 446 447 448 449 450
| E N A D E N A L Q S D A K G K
| 5398 gaa aat gcc gat gaa aac gcg cta cag tct gac gct aaa ggc aaa
|
|
|
45 451 452 453 454 455 456 457 458 459 460 461 462 463 464 465
| L D S V A T D Y G A A I D G F
| 5443 ctt gat tct gtc gct act gat tac ggt gct gct ATC GAT ggt ttc
| BspDI..
|
|
|
50 466 467 468 469 470 471 472 473 474 475 476 477 478 479 480
| I G D V S G G L A N G N G A T G

```



6291 gtaaagcact aaatcggaac cctaaagga gcccccgatt tagagcttga cggggaaaGC  
! CGCCgaacgt ggcgagaaaag gaagggaaga aagcgaagag agcgggcgct agggcgctgg  
! ..NgmIV. (2/2) NgoMIV..  
5 6411 caagtgtagc ggtcacgctg cgcgtaacca ccacaccgc cgcgcttaat gcgccgctac  
6471 agggcgcgta ctatggttgc tttagcgggt gcagtcacag tacaatctgc tctgatgccg  
6531 catagttaag ccagccccga caccgcctga caccgcctga cgggcttgtc  
6591 tgctccccgc atccgcttac agacaagctg tgaccgtctc cgggagctgc atgtgtcaga  
6651 ggtttcacc gtcatcaccg aaacgcgcga

Table 630: Oligonucleotides used to clone CDR1/2 diversity

All sequences are 5' to 3'.

5 1) ON\_CD1Bsp, 30 bases  
 A C C T C A C T g g c T T c c g g A  
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

10 T T C A C T T T c T c T  
 19 20 21 22 23 24 25 26 27 28 29 30

2) ON\_Br12, 42 bases

15 A g A A A C C C A C T C C A A A C C  
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

20 T T T A C C A g g A g c T T g g c g  
 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36

A A C C C A  
 37 38 39 40 41 42

25 3) ON\_CD2Xba, 51 bases

g g A A g g c A g T g A T c T A g A  
 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

30 g A T A g T g A A g c g A c c T T T  
 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36

A A C g g A g T c A g c A T A  
 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51

35 4) ON\_BotXba, 23 bases

g g A A g g c A g T g A T c T A g A  
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18

g A T A g  
5 19 20 21 22 23

10 End Tables