

(19) United States

(12) Patent Application Publication (10) Pub. No.: US 2002/0151026 A1 **Briskin**

Oct. 17, 2002 (43) Pub. Date:

(54) MAMMALIAN HYALURONAN SYNTHASES, NUCLEIC ACIDS AND USES THEREOF

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10/042,523 (21)Appl. No.:

(22) Filed: Oct. 19, 2001

Related U.S. Application Data

Division of application No. 08/635,552, filed on Apr. 22, 1996.

Publication Classification

(51) **Int. Cl.**⁷ **C12P 19/26**; C12N 9/24; C12P 21/02; C12N 5/06; C08B 37/00 (52) **U.S. Cl.** **435/200**; 435/69.1; 435/320.1; 435/325; 536/23.2; 435/84; 536/53

(57)ABSTRACT

The present invention relates to an isolated or recombinant nucleic acid which encodes a mammalian hyaluronan synthase (e.g. human). The present invention also relates to a host cell comprising the nucleic acid encoding mammalian hyaluronan synthase. The present invention also relates to a method for producing a mammalian hyaluronan synthase comprising introducing into a host cell a nucleic acid construct comprising a nucleic acid which encodes a mammalian hyaluronan synthase, whereby a recombinant host cell is produced having said coding sequence operably linked to at least one expression control sequence; and maintaining the host cells produced in a suitable medium under conditions whereby the nucleic acid is expressed. The present invention also relates to an antibody or functional portion thereof which binds mammalian hyaluronan synthase. The present invention also relates to a method of detecting mammalian hyaluronan synthase in a sample comprising contacting a sample with an antibody which binds hyaluronan synthase under conditions suitable for specific binding of said antibody to the mammalian hyaluronan synthase; and detecting antibody-mammalian hyaluronan synthase. The invention further relates to a method of using hyaluronan synthase to make hyaluronan.

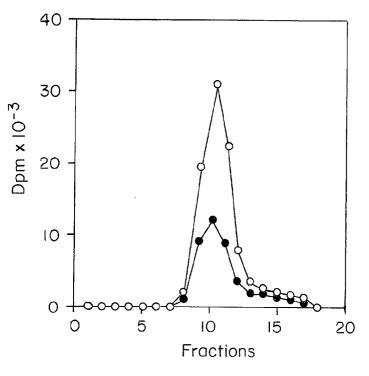


FIG. IA

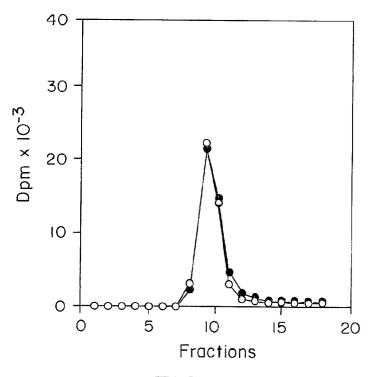


FIG. 1B

CGGAGAGAGAGAGAGCCCGGCCAGACCCACTGCGATGAGACAGCAGGACGCGCCCAAGCCCCACTCCTGCAGCCCGCCGC	80
M R Q Q D A P K P T P A A R R	
TGCTCCGGCCTGGCCCGGAGGGTGCTGACCATCGCCTTCGCCCTGCTCATCCTGGGCCTCATGACCTGGGCCTACGCCGC © S G L A R R V L T I A F A L L I L G L M T W A Y A A	160
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	240
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	320
AGTGTGGCGCTGACCATCTCCGCCTACCAGGAGGACCCCGCGCGTACCTGCGCCAGTGCCTGGCGTCCGCCCGC	_
GTACCCGCGCGCGCGCGCTCCTCATGGTGGTGGATGGCAACCGCGCCGAGGACCTCTACATGGTCGACATGTTCC Y P R A R L R V L M V V D G N R A E D L Y M V D M F	480
GCGAGGTCTTCGCTGACGAGGACCCCGCCACGTACGTGTGGGACGGCAACTACCACCAGCCCTGGGAACCCGCGGCGGCGCGCGC	560
GGCGCGGTGGGCGCCCGGAGCCTATCGGGAGGTGGAGGCGGAGGATCCTGGGCGCTGGCAGTGGAGGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGTGAGGACGCGCTGGAGGAGCGCTGGAGGACGCGCTGGAGGACGCGCTGGAGGAGCGCTGGAGGAGCGCTGGAGGACGCGCTGGAGGAGACCGCTGGAGGAGACCGCTGGAGGAGACCGCTGGAGGAGGACGCGCTGGAGGAGACGCGCTGGAGGAGACCGCTGGAGGAGACGCGCTGGAGGAGACCGCTGGAGGAGACCGCTGGAGGAGACCGCTGGAGGAGACACACAC	640
TCGCAGGTGCGTGCGCGCAGCGCTGGGGCGGCAAGCGCGAGGTCATGTACACAGCCTTCAAGGCGCTCGGAGATT R R CO V CO V A Q R W G G K R E V M Y T A F K A L G D	720
CGGTGGACTACGTGCAGGTCTGTGACTCGGACACAAGGTTGGACCCCATGGCACTGCTGGAGCTCGTGCGGGTACTGGAC S V D Y V Q V C D S D T R L D P M A L L E L V R V L D	800
GAGGACCCCCGGGTAGGGGGCTGTTGGTGGGGACGTGCGGATCCTTAACCCTCTGGACTCCTGGGTCAGCTTCCTAAGCAG E D P R V G A V G G D V R I L N P L D S W V S F L S S	
CCTGCGATACTGGGTAGCCTTCAATGTGGAGCGGGCTTGTCAGAGCTACTTCCACTGTGTATCCTGCATCAGCGGTCCTC L R Y W V A F N V E R A (C) Q S Y F H (C) V S (C) I S G P	960
TAGGCCTATATAGGAATAACCTCTTGCAGCAGTTTCTTGAGGCCTGGTACAACCAGAAGTTCCTGGGTACCCACTGTACT L G L Y R N N L L Q Q F L E A W Y N Q K F L G T H C T	1040
TTTGGGGATGACCGGCACCTCACCAACCGCATGCTCAGCATGGGTTATGCTACCAAGTACACCTCCAGGTCCCGCTGCTAFGDDDRHLTNRMLSMGYATKYTSRSRCC)Y	1120
CTCAGAGACGCCCTCGTCCTTCCTGCGGTGGCTGAGCCAGCAGACACGCTGGTCCAAGTCGTACTTCCGTGAGTGGCTGT SETPSSFLRWLSQQTRWSKSYFREWL	1200
ACAACGCGCTCTGGTGGCACCGCACCATGCGTGGATGACCTACGAGGCGGTGGTCTCCGGCCTGTTCCCCTTCTTCGTG Y N A L W W H R H H A W M T Y E A V Y S G L F P F F V	1280
GCGGCCACTGTGCTGCGTCTGTTCTACGCGGGCCCCCTTGGGCGCTGCTGTGGGTGCTGCTGTGCGTGCAGGGCGTGGCAAATVLRLFYAGRPWALLWVLLCVQQCGTGC	1360
ACTGGCCAAGGCGGCCTTCGCGGCCTGGCTGCGGGGCTGCCTGC	1440
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1520
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1600
ACACGAGGCCAGGGCCGACTGGAGCGGCCCTTCCCGCGCAGCCGAGGCCTACCACTTGGCCGCGGGGGCCCGGCGCCTACG H E A R A D W S G P S R A A E A Y H L A A G A G A Y	1680
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1760
GTCCAGGTGTGAGTCCAGCCACGCGGATGCCGCCTCAAGGGTCTTCAGGGGAGGCCAGAGGAGAGCTGCTGGGCCCCGAG V Q V	1840
CCACGAACTTGCTGGGTGGTTCTCTGGGCCTCAGTTTCCCTCCTCTGCCAAACGAGGGGGTCAGCCCAAGATTCTTCAGT	1920
CTGGACTATATTGGGACTGGGACTTCTGGGTCTCCAGGGAGGG	2000
GGGAAAGGGTCCTGCTTCTCCCCGTTCTTATTTAATCTCCATTTCTACTGTGTGATCAGGATGT <u>AATAAA</u> GAATTTTAT	2080

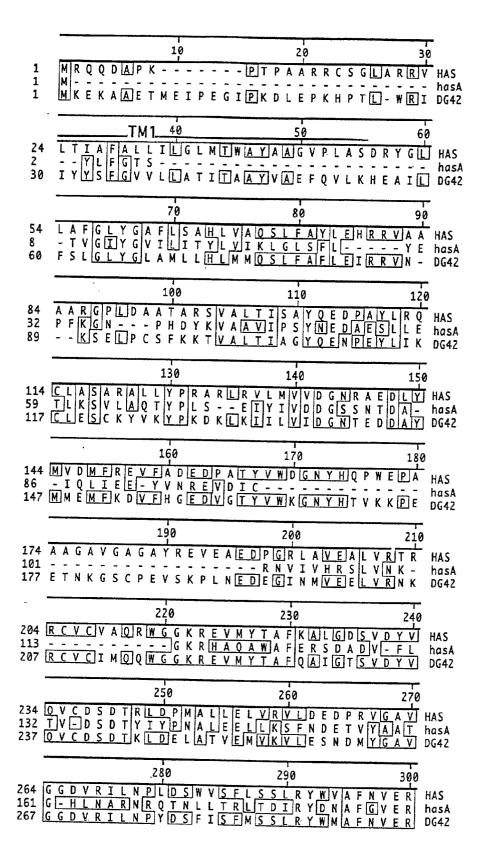


FIG. 3A

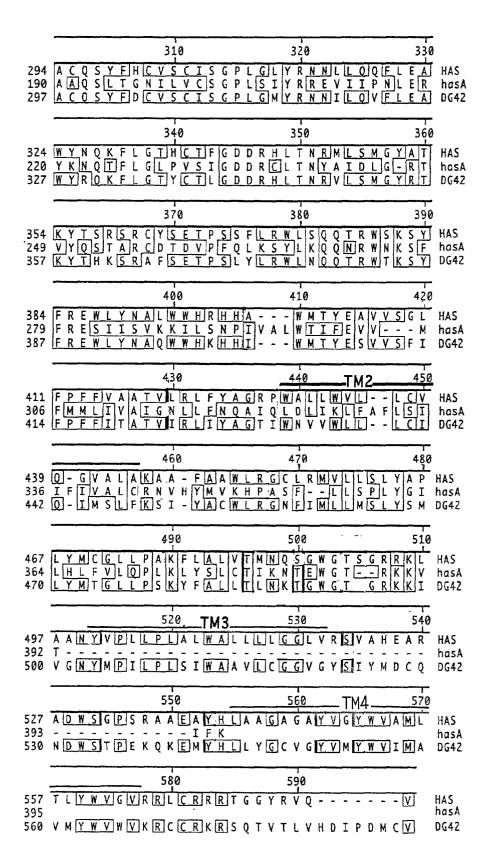
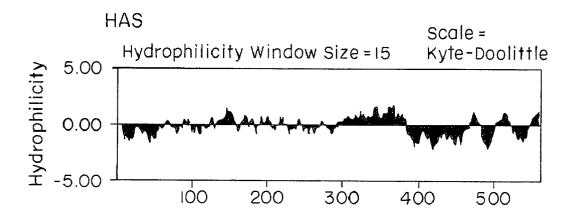
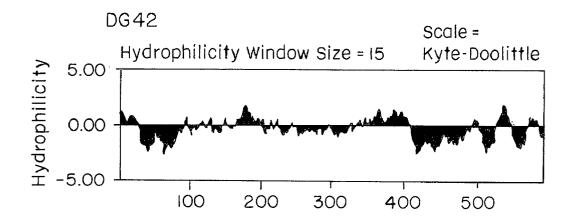
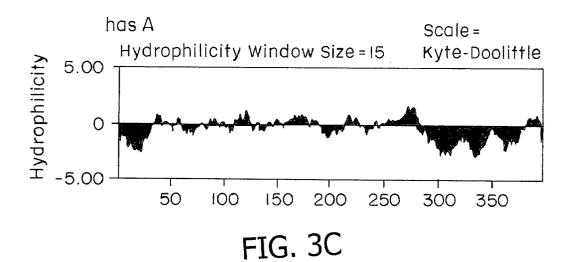
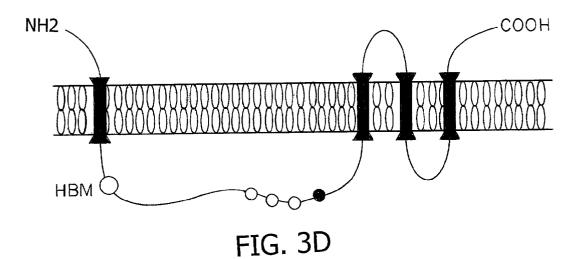


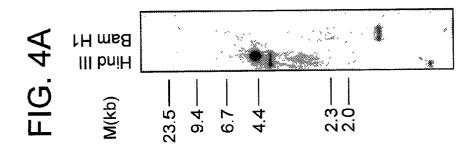
FIG. 3B

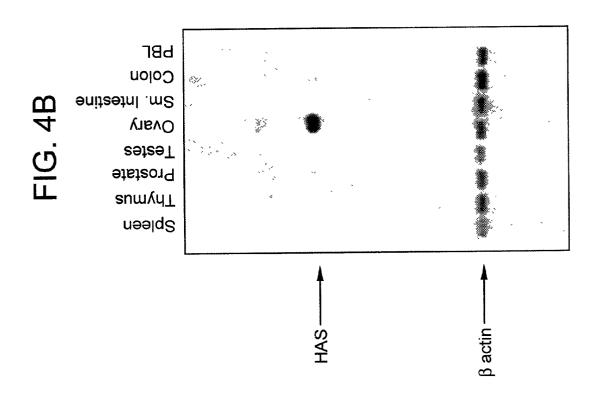












MAMMALIAN HYALURONAN SYNTHASES, NUCLEIC ACIDS AND USES THEREOF

BACKGROUND

[0001] Hyaluronan is a constituent of the extracellular matrix of connective tissue, and is actively synthesized during wound healing and tissue repair to provide a framework for ingrowth of blood vessels and fibroblasts. Changes in the serum concentration of hyaluronan are associated with inflammatory and degenerative arthropathies such as rheumatoid arthritis. In addition, hyaluronan has been implicated as an important substrate for migration of adhesion of leukocytes during inflammation.

[0002] Hyaluronan (hyaluronic acid, HA) is a high molecular mass polysaccharide that has ubiquitous distribution in the extracellular matrix, with highest concentrations in soft connective tissue. It is a linear polysaccharide comprising alternating glucuronic acid and N-acetylglucosamine residues linked by β -1-3 and β -1-4 glycosidic bonds (Laurent, T. C. et al. (1986), "The properties and turnover of hyaluronan." Functions of proteoglycans (Symposium, C.F., Ed. 124, Chichester, England). By interacting with other matrix molecules, such as chondroitin sulfate proteoglycans, hyaluronan provides stability and elasticity to the extracellular matrix. Hyaluronan has several physiochemical and biological functions such as space filling, lubrication, and providing a hydrated matrix through which cells can migrate (Toole, B. P. et al., Hyaluronate-cell interactions. The role of the extracellular matrix in development, (Trelstad, R.L., Ed., Alan R. Liss, New York (1984); Laurent, T. C. et al., Faseb J. 6:2397-2404 (1992)). Interaction of hyaluronan with the leukocyte cell surface receptor CD44 has been shown to contribute to organ specific leukocyte homing and migration (Jalkanen, S. T. et al., J. Cell. Biol., 105:893-990 (1987); Aruffo, A., et al., Cell 61:1303-1313 (1990); Culty, M. et al., J. Cell. Biol., 111:2765-2774 (1990); Miyake, K. et al., J. Exp. Med. 172:69-75 (1990); Sherman, L. et al., Current Opinions in Cell Biology, 6:726-733 (1994)). Hyaluronan synthesis has been suggested to be required for cellular proliferation (Brecht, M. et al., Biochem. J. 239:445-450 (1986); Hronowski, L. and Anastassiades, T. P., J. Biol. Chem. 255:9210-9217 (1980); Matuoka, K. et al., J. Cell Biol. 104:1105-1115 (1987); Mian, N., Biochem. J. 237:333-342 (1986); Tomida, M. et al., J. Cell Physiol. 86:121-130 (1975)), and over-expression of receptors for hyaluronan, including a receptor for hyaluronan mediated motility (RHAMM) and CD44, correlates with increased levels of tumor metastasis (Gunthert, U., Curr. Topics Microbiol. Immunol. 184:47-63 (1993); Hall, C. L. et al., Cell 82:19-28 (1995); Turley, E. A., Cancer and Metastasis Reviews 11:1233-1241 (1992)). Purified preparations of hyaluronan exhibit unique viscoelastic properties, and as a consequence of these characteristics have been used in viscoelastic surgery and viscosupplementation (Balazs, E. A., and Denninger, J. L., Clinical uses of hyaluronan, The biology of hyaluronan, Ciba foundation symposium, Wiley, Chichester, England (1989)). Hyaluronan is synthesized mainly by mesenchymal cells and the accumulation of HA is an early event in tissue repair. The serum level of hyaluronan is elevated in inflammatory settings such as rheumatoid arthritis, osteoarthritis, liver cirrhosis, Werner's syndrome, renal failure and psoriasis (Laurent, T. C. et al., Faseb J. 6:2397-2404 (1992); Laurent, T. C. Annals of Medicine 28:in press (1996)).

[0003] Hyaluronan is synthesized by a membrane bound synthase; monosaccharide and disaccharide residues are added to the reducing end of the polysaccharide as it protrudes through the plasma membrane (Prehm, P., *Biochem. J.* 211:181-189 (1983); Prehm, P., *Biochem. J.* 220:597-600 (1984)). Regulation of hyaluronan biosynthesis has been studied in several tissue culture systems. Factors involved in tissue growth and repair such as different isoforms of platelet derived growth factor (PDGF-AA, PDGF-BB), epidermal growth factor (EGF), basic fibroblast growth factor (bFGF), and transforming growth factor $\beta(TGF-\beta)$, all exhibit stimulatory activity on hyaluronan biosynthesis (Heldin, P. et al., *Biochem. J.* 258, 919-922 (1992)).

[0004] A cDNA encoding a bacterial hyaluronan synthase has been cloned from Streptococcus pyogenes (hasA) (DeAngelis, J. P. et al., *J. Biol. Chem.* 268, 19181-19184 (1993)). Other related genes with N-acetylglucosaminyl transferase activity have been isolated from the nitrogen fixing bacteria Rhizobium (nodC) and chitin synthases (Chs) from Saccharomyces (DeAngelis, P. L. et al., *Biochem. Biophys. Res. Comm.* 199:1-10 (1994)). A putative vertebrate homolog, (DG42), was cloned from Xenopus laevis and has also been speculated to be a glycosaminoglycan synthetase (Rosa, F. et al., *Develop. Biol.* 129:114-123 (1988)). To date, however, a mammalian hyaluronan synthase gene has not been identified.

SUMMARY OF THE INVENTION

[0005] The present invention relates to isolated and/or recombinant nucleic acids which encode a mammalian hyaluronan synthase (e.g., human). In one embodiment, the nucleic acid of the present invention comprises SEQ ID NO: 1. In another embodiment, the invention relates to a nucleic acid wherein said nucleic acid hybridizes under stringent conditions with a second nucleic acid having a nucleotide sequence of SEQ ID NO: 1.

[0006] The present invention also relates to a host cell comprising a nucleic acid encoding mammalian hyaluronan synthase. In a particular embodiment, the host cell comprises nucleic acid encoding mammalian hyaluronan synthase which is operably linked to an expression control sequence, whereby mammalian hyaluronan synthase is expressed when the host cell is maintained under conditions suitable for expression.

[0007] The present invention also relates to a method for producing a mammalian hyaluronan synthase comprising introducing into a host cell a nucleic acid construct comprising a nucleic acid which encodes a mammalian hyaluronan synthase, whereby a recombinant host cell is produced having said coding sequence operably linked to an (i.e., at least one) expression control sequence; and maintaining the host cells produced in a suitable medium under conditions whereby the nucleic acid is expressed.

[0008] The present invention also relates to an antibody or functional portion thereof (e.g., an antigen binding portion such as an Fv, Fab, Fab', or F(ab')₂ fragment) which binds mammalian hyaluronan synthase.

[0009] The present invention also relates to a method of detecting mammalian hyaluronan synthase in a sample comprising contacting a sample with an antibody which binds hyaluronan synthase under conditions suitable for specific

binding of said antibody to the mammalian hyaluronan synthase; and detecting antibody-mammalian hyaluronan synthase.

[0010] The invention further relates to a method of using hyaluronan synthase to make hyaluronan.

BRIEF DESCRIPTION OF THE FIGURES

[0011] FIG. 1A is a graph illustrating that CHO cells transfected with human hyaluronan synthase cDNA synthesize hyaluronic acid; media and cell lysates were combined and then incubated overnight in the absence (○ - - - ○) or presence (● - - - ●) of 10U Streptomyces hyaluronidase/ml and subjected to chromatography on Sephadex G-50 columns; Streptomyces hyaluronidase-sensitive radioactivity represents synthesized hyaluronan.

[0012] FIG. 1B is a graph illustrating that CHO cells not transfected with human hyaluronan synthase cDNA produce very little high molecular weight streptomyces hyaluronidase-sensitive material.

[0013] FIG. 2 is an illustration of the nucleotide sequence (SEQ ID NO: 1) and deduced protein sequence (SEQ ID NO: 2) determined from human hyaluronan synthase cDNA clone 30C; cysteine residues are circled and a conserved motif, B(X₇)B, believed to be important for binding hyaluronan is lightly outlined; consensus phosphorylation sequences for protein kinase C (RHLT, KYT and RWLS) and cAMP dependent protein kinases (RWS) are outlined in bold; also shown with a bold underline at position 2066 is a consensus polyadenylation signal, AATAAA. (Standard single letter amino acid codes are used.)

[0014] FIG. 3A is an amino acid alignment of the human hyaluronan synthase protein sequence (SEQ ID NO: 2) with the DG42 sequence from Xenopus laevis (SEQ ID NO: 3) and has A sequence of *Streptococcus pyogenes* (SEQ ID NO: 4) prepared using the DNAStar program and the Clustal method with default parameters for gap penalties.

[0015] FIG. 3B is a comparison of Kyte-Doolittle hydrophilicity profiles of human hyaluronan synthase, DG42 and hasA.

[0016] FIG. 3C is a proposed structure of human hyaluronan synthase, indicating approximate boundaries of transmembrane regions and intra- and extracellular loops; a hyaluronan binding motif (HBM), B(X₇)B, is indicated at the amino portion of a large predicted intracellular loop; approximate locations of protein kinase C consensus sites are indicated by open circles, while a single cAMP dependent kinase site is shown as a filled circle.

[0017] FIG. 4A is a Northern blot probed with the full length insert of the human hyaluronan synthase cDNA clone 30C; the blot was subsequently stripped and reprobed with a M-actin cDNA as a control.

[0018] FIG. 4B is a Southern blot initially hybridized with full-length human hyaluronan synthase cDNA, washed at 50° C., and exposed overnight; a considerable amount of background was seen although specific bands could be detected; subsequently the blot was stripped and probed with a 450 bp Sac II fragment encompassing the 3' end of the cDNA; this probe gave a similar pattern with less background (likely due to a lower GC content).

DETAILED DESCRIPTION OF THE INVENTION

[0019] Proteins and Peptides

[0020] The present invention relates to isolated and/or recombinant (including, e.g., essentially pure) proteins or polypeptides designated mammalian hyaluronan synthase and variants of mammalian hyaluronan synthase. In a preferred embodiment, the isolated and/or recombinant proteins of the present invention have at least one property, activity or function characteristic of a mammalian hyaluronan synthase (as defined herein), such as activity in the synthesis of hyaluronan and/or ability to confer of cell adhesion by the lymphocyte receptor CD44 (i.e., human CD44 or a mammalian homolog thereof).

[0021] Proteins or polypeptides referred to herein as "isolated" are proteins or polypeptides purified to a state beyond that in which they exist in mammalian cells. "Isolated" proteins or polypeptides include proteins or polypeptides obtained by methods described herein, similar methods or other suitable methods, including essentially pure proteins or polypeptides, proteins or polypeptides produced by chemical synthesis (e.g., synthetic peptides), or by combinations of biological and chemical methods, and recombinant proteins or polypeptides which are isolated. The proteins can be obtained in an isolated state of at least about 50% by weight, preferably at least about 75% by weight, and more preferably, in essentially pure form. Proteins or polypeptides referred to herein as "recombinant" are proteins or polypeptides produced by the expression of recombinant nucleic acids.

[0022] As used herein "mammalian hyaluronan synthase" refers to naturally occurring or endogenous mammalian hyaluronan synthase proteins, to proteins having an amino acid sequence which is the same as that of a naturally occurring or endogenous corresponding mammalian hyaluronan synthase (e.g., recombinant proteins), and to functional variants of each of the foregoing (e.g., functional fragments and/or mutants produced via mutagenesis and/or recombinant techniques). Accordingly, as defined herein, the term includes mature mammalian hyaluronan synthase, glycosylated or unglycosylated mammalian hyaluronan synthase proteins, polymorphic or allelic variants, and other isoforms of mammalian hyaluronan synthase (e.g., produced by alternative splicing or other cellular processes), and functional fragments.

[0023] Naturally occurring or endogenous mammalian hyaluronan synthase proteins include wild type proteins such as mature mammalian hyaluronan synthase, polymorphic or allelic variants and other isoforms which occur naturally in mammals (e.g., primate, preferably human, murine, bovine). Such proteins can be recovered from a source which naturally produces mammalian hyaluronan synthase, for example. These mammalian proteins having the same amino acid sequence as naturally occurring or endogenous corresponding mammalian hyaluronan synthase, are referred to by the name of the corresponding mammal. For example, as described herein, where the corresponding mammal is human, the protein is designated as a human hyaluronan synthase (HAS), such as recombinant human hyaluronan synthase produced in a suitable host cell.

[0024] "Functional variants" of mammalian hyaluronan synthase include functional fragments, functional mutant

proteins, and/or functional fusion proteins. Generally, fragments or portions of mammalian hyaluronan synthase encompassed by the present invention include those having a deletion (i.e., one or more deletions) of an amino acid (i.e., one or more amino acids) relative to the mature mammalian hyaluronan synthase (such as N-terminal, C-terminal or internal deletions). Fragments or portions in which only contiguous amino acids have been deleted or in which non-contiguous amino acids have been deleted relative to mature mammalian hyaluronan synthase are also envisioned.

[0025] Generally, mutants or derivatives of mammalian hyaluronan synthase, encompassed by the present invention include natural or artificial variants differing by the addition, deletion and/or substitution of one or more contiguous or non-contiguous amino acid residues, or modified polypeptides in which one or more residues is modified, and mutants comprising one or more modified residues. Preferred mutants are natural or artificial variants of mammalian hyaluronan synthase differing by the addition, deletion and/or substitution of one or more contiguous or non-contiguous amino acid residues.

[0026] A "functional fragment or portion", "functional mutant" and/or "functional fusion protein" of a mammalian hyaluronan synthase refers to an isolated and/or recombinant protein or oligopeptide which has at least one property, activity and/or function characteristic of a mammalian hyaluronan synthase, such as activity or function characteristic of a mammalian hyaluronan synthase (as defined herein), such as activity in the synthesis of hyaluronan and/or ability to confer cell adhesion by the lymphocyte receptor CD44.

[0027] Suitable fragments or mutants can be identified by screening. For example, the N-terminal, C-terminal, or internal regions of the protein can be deleted in a step-wise fashion and the resulting protein or polypeptide can be screened using a suitable binding or adhesion assay. Where the resulting protein displays activity in the assay, the resulting protein ("fragment") is functional. Information regarding the structure and function of other hyaluronan synthases (e.g., hasA, DG42), and of HAS as shown herein, provides a basis for dividing HAS into functional domains.

[0028] The term variant also encompasses fusion proteins, comprising a mammalian hyaluronan synthase (e.g., mature mammalian hyaluronan synthase) as a first moiety, linked to a second moiety not occurring in the mammalian hyaluronan synthaseas found in nature. Thus, the second moiety can be an amino acid, oligopeptide or polypeptide. The first moiety can be in an N-terminal location, C-terminal location or internal to the fusion protein. In one embodiment, the fusion protein comprises a mammalian hyaluronan synthase or portion thereof as the first moiety, and a second moiety comprising a linker sequence and affinity ligand (e.g., an enzyme, an antigen, epitope tag).

[0029] Examples of "mammalian hyaluronan synthase" proteins include proteins having an amino acid sequence as set forth or substantially as set forth in FIG. 2 (SEQ ID NO: 2) and functional portions thereof. In a preferred embodiment, a mammalian hyaluronan synthase or variant has an amino acid sequence which has at least about 50% identity, more preferably at least about 75% identity, and still more preferably at least about 90% identity, to the protein shown in FIG. 2 (SEQ ID NO: 2).

[0030] Method of Producing Recombinant Proteins

[0031] Another aspect of the invention relates to a method of producing a mammalian hyaluronan synthase or variant (e.g., portion) thereof. Recombinant protein can be obtained, for example, by the expression of a recombinant DNA molecule encoding a mammalian hyaluronan synthase or variant thereof in a suitable host cell, for example.

[0032] Constructs suitable for the expression of a mammalian hyaluronan synthase or variant thereof are also provided. The constructs can be introduced into a suitable host cell, and cells which express a recombinant mammalian hyaluronan synthase or variant thereof, can be produced and maintained in culture. Such cells are useful for a variety of purposes, and can be used in the production of protein for characterization, isolation and/or purification, (e.g., affinity purification), and as immunogens, for instance. Suitable host cells can be procaryotic, including bacterial cells such as E. coli, B. subtilis and or other suitable bacteria (e.g., Streptococci) or eucarvotic, such as fungal or yeast cells (e.g., Pichia pastoris, Aspergillus species, Saccharomyces cerevisiae, Schizosaccharomyces pombe, Neurospora crassa) or other lower eucaryotic cells, and cells of higher eucaryotes such as those from insects (e.g., Sf9 insect cells) or mammals (e.g., Chinese hamster ovary cells (CHO), COS cells, HuT 78 cells, 293 cells). (See, e.g., Ausubel, F. M. et al., eds. Current Protocols in Molecular Biology, Greene Publishing Associates and John Wiley & Sons Inc., (1993)).

[0033] Host cells which produce a recombinant mammalian hyaluronan synthase or variants thereof can be produced as follows. For example, a nucleic acid encoding all or part of the coding sequence for the desired protein can be inserted into a nucleic acid vector, e.g., a DNA vector, such as a plasmid, virus or other suitable replicon for expression. A variety of vectors are available, including vectors which are maintained in single copy or multiple copy, or which become integrated into the host cell chromosome.

[0034] The transcriptional and/or translational signals of a mammalian hyaluronan synthase gene can be used to direct expression. Alternatively, suitable expression vectors for the expression of a nucleic acid encoding all or part of the coding sequence of the desired protein are available. Suitable expression vectors can contain a number of components, including, but not limited to one or more of the following: an origin of replication; a selectable marker gene; one or more expression control elements, such as a transcriptional control element (e.g., a promoter, an enhancer, terminator), and/or one or more translation signals; a signal sequence or leader sequence for membrane targeting or secretion (of mammalian origin or from a heterologous mammal or non-mammalian species). In a construct, a signal sequence can be provided by the vector, the mammalian hyaluronan synthase coding sequence, or other source.

[0035] A promoter can be provided for expression in a suitable host cell. Promoters can be constitutive or inducible. The promoter is operably linked to a nucleic acid encoding the mammalian hyaluronan synthase or variant thereof, and is capable of directing expression of the encoded polypeptide in the host cell. A variety of suitable promoters for procaryotic (e.g., lac, tac, T3, T7 promoters for *E. coli*) and eucaryotic (e.g., yeast alcohol dehydrogenase (ADH1), SV40, CMV) hosts are available.

[0036] In addition, the expression vectors typically comprise a selectable marker for selection of host cells carrying

the vector, and in the case of a replicable expression vector, an origin of replication. Genes encoding products which confer antibiotic or drug resistance are common selectable markers and may be used in procaryotic (e.g., β-lactamase gene (ampicillin resistance), Tet gene for tetracycline resistance) and eucaryotic cells (e.g., neomycin (G418 or geneticin), gpt (mycophenolic acid), ampicillin, or hygromycin resistance genes). Dihydrofolate reductase marker genes permit selection with methotrexate in a variety of hosts. Genes encoding the gene product of auxotrophic markers of the host (e.g., LEU2, URA3, HIS3) are often used as selectable markers in yeast. Use of viral (e.g., baculovirus) or phage vectors, and vectors which are capable of integrating into the genome of the host cell, such as retroviral vectors, are also contemplated. The present invention also relates to cells carrying these expression vectors.

[0037] For example, a nucleic acid encoding a mammalian hyaluronan synthase or variant thereof can be incorporated into a vector, operably linked to one or more expression control elements, and the construct can be introduced into host cells which are maintained under conditions suitable for expression, whereby the encoded polypeptide is produced. The construct can be introduced into cells by a method appropriate to the host cell selected (e.g., transformation, transfection, electroporation, infection). For production of a protein, host cells comprising the construct are maintained under conditions appropriate for expression, (e.g., in the presence of inducer, suitable media supplemented with appropriate salts, growth factors, antibiotic, nutritional supplements, etc.). The encoded protein (e.g., human hyaluronan synthase) can be isolated from the host cells or medium.

[0038] Fusion proteins can also be produced in this manner. For example, some embodiments can be produced by the insertion of a mammalian hyaluronan synthase cDNA or portion thereof into a suitable expression vector, such as Bluescript®II SK+/-(Stratagene), pGEX-4T-2 (Pharmacia), pcDNA-3 (Invitrogen) and pET-15b (Novagen). The resulting construct can then be introduced into a suitable host cell for expression. Upon expression, fusion protein can be isolated or purified from a cell lysate by means of a suitable affinity matrix (see e.g., Current Protocols in Molecular Biology (Ausubel, F. M. et al., eds., Vol. 2, Suppl. 26, pp. 16.4.1-16.7.8 (1991)). In addition, affinity labels provide a means of detecting a fusion protein. For example, the cell surface expression or presence in a particular cell fraction of a fusion protein comprising an antigen or epitope affinity label can be detected by means of an appropriate antibody.

[0039] Nucleic Acids, Constructs and Vectors

[0040] The present invention relates to isolated and/or recombinant (including, e.g., essentially pure) nucleic acids (e.g., polynucleotides) having sequences which encode a mammalian hyaluronan synthase or variant thereof as described herein.

[0041] Nucleic acids referred to herein as "isolated" are nucleic acids separated away from the nucleic acids of the genomic DNA or cellular RNA of their source of origin (e.g., as it exists in cells or in a mixture of nucleic acids such as a library), and may have undergone further processing. "Isolated" nucleic acids include nucleic acids obtained by methods described herein, similar methods or other suitable methods, including essentially pure nucleic acids, nucleic

acids produced by chemical synthesis, by combinations of biological and chemical methods, and recombinant nucleic acids which are isolated (see e.g., Daugherty, B. L. et al., Nucleic Acids Res., 19(9):2471-2476 (1991); Lewis, A. P. and J. S. Crowe, Gene, 101: 297-302 (1991)). Nucleic acids referred to herein as "recombinant" are nucleic acids which have been produced by recombinant DNA methodology, including those nucleic acids that are generated by procedures which rely upon a method of artificial recombination, such as the polymerase chain reaction (PCR) and/or cloning into a vector using restriction enzymes. "Recombinant" nucleic acids are also those that result from recombination events that occur through the natural mechanisms of cells, but are selected for after the introduction to the cells of nucleic acids designed to allow and make probable a desired recombination event.

[0042] In one embodiment, the nucleic acid or portion thereof encodes a protein or polypeptide having at least one property, activity or function characteristic of a mammalian hyaluronan synthase (as defined herein), such as activity or function characteristic of a mammalian hyaluronan synthase (as defined herein), such as activity in the synthesis of hyaluronan and/or ability to mediate cell adhesion by the lymphocyte receptor CD44.

[0043] The present invention also relates more specifically to isolated and/or recombinant nucleic acids or a portion thereof having sequences which encode mammalian hyaluronan synthase or variants thereof.

[0044] The invention relates to isolated and/or recombinant nucleic acids that are characterized by:

[0045] (1) their ability to hybridize to (a) a nucleic acid encoding a mammalian hyaluronan synthase, such as a nucleic acid having a nucleotide sequence as set forth or substantially as set forth in FIG. 2 (SEQ ID NO: 1); (b) the complement of (a); or (c) portions of either of the foregoing (e.g., a portion comprising the open reading frame); or

[0046] (2) by their ability to encode a polypeptide having the amino acid sequence of a mammalian hyaluronan synthase (e.g., SEQ ID NO: 2); or

[0047] (3) by both characteristics.

[0048] In one embodiment, the nucleic acid shares at least about 50% nucleotide sequence similarity to the nucleotide sequences shown in FIG. 2 (SEQ ID NO: 1). More preferably, the nucleic acid shares at least about 75% nucleotide sequence similarity, and still more preferably, at least about 90% nucleotide sequence similarity, to the sequence shown in FIG. 2 (SEQ ID NO: 1).

[0049] Isolated and/or recombinant nucleic acids meeting these criteria comprise nucleic acids having sequences identical to sequences of naturally occurring mammalian hyaluronan synthase or variants of the naturally occurring sequences. Such variants include mutants differing by the addition, deletion or substitution of one or more residues, modified nucleic acids in which one or more residues are modified (e.g., DNA or RNA analogs), and mutants comprising one or more modified residues.

[0050] A nucleic acid of the present invention may be in the form of RNA or in the form of DNA (e.g., cDNA, genomic DNA, and synthetic DNA). The DNA may be double-stranded or single-stranded, and if single stranded may be the coding strand or non-coding (anti-sense) strand. The coding sequence which encodes the mature polypeptide may be identical to the coding sequence shown in **FIG. 2** (SEQ ID NO: 1) or that of the cDNA in clone 30C or may be a different coding sequence which coding sequence, as a result of the redundancy or degeneracy of the genetic code, encodes the same, mature polypeptides as the DNA of **FIG. 2** (SEQ ID NO: 2) or the cDNA in clone 30C.

[0051] The polynucleotide which encodes a mature polypeptide encoded by the cDNA of clone 30C may include: only the coding sequence of a mature polypeptide; the coding sequence for a mature polypeptide and additional coding sequence such as a leader or secretory sequence; the coding sequence for a mature polypeptide (and optionally additional coding sequence) and non-coding sequence, such as introns or non-coding sequence 5' and/or 3' of the coding sequence.

[0052] Nucleic acids of the present invention, including those which hybridize to a selected nucleic acid as described above, can be detected or isolated under high stringency conditions or moderate stringency conditions, for example. "High stringency conditions" and "moderate stringency conditions" for nucleic acid hybridizations are explained at pages 2.10.1-2.10.16 (see particularly 2.10.8-11) and pages 6.3.1-6 in Current Protocols in Molecular Biology (Ausubel, F. M. et al., eds., Vol. 1, Suppl. 26, 1991), the teachings of which are hereby incorporated by reference. Factors such as probe length, base composition, percent mismatch between the hybridizing sequences, temperature and ionic strength influence the stability of nucleic acid hybrids. Thus, high or moderate stringency conditions can be determined empirically, and depend in part upon the characteristics of the known nucleic acid (e.g., DNA) and the other nucleic acids to be assessed for hybridization thereto.

[0053] Isolated and/or recombinant nucleic acids that are characterized by their ability to hybridize (e.g., under high or moderate stringency conditions) to (a) a nucleic acid encoding a mammalian hyaluronan synthase (for example, the nucleic acid depicted in FIG. 2 (SEQ ID NO: 1); (b) the complement of the nucleic acids of (a), (c) or a portion thereof, can also encode a protein or polypeptide having at least one property, activity or function characteristic of a mammalian hyaluronan synthase (as defined herein), such as activity in the synthesis of hyaluronan and/or ability to mediate cell adhesion by the lymphocyte receptor CD44, and in a preferred embodiment encode polypeptides which retain substantially the same biological function or activity as the mature polypeptide encoded by the cDNA of FIG. 2 (SEQ ID NO: 1) or the cDNA of clone 30C.

[0054] Nucleic acids of the present invention can be used in the production of proteins or polypeptides. For example, a nucleic acid (e.g., DNA) encoding a mammalian hyaluronan synthase can be incorporated into various constructs and vectors created for further manipulation of sequences or for production of the encoded polypeptide in suitable host cells as described above.

[0055] A further embodiment of the invention is antisense nucleic acid, which is complementary, in whole or in part, to a target molecule comprising a sense strand, and can hybridize with the target molecule. The target can be DNA, or its RNA counterpart (i.e., wherein T residues of the DNA are U

residues in the RNA counterpart). When introduced into a cell, antisense nucleic acid can inhibit the expression of the gene encoded by the sense strand. Antisense nucleic acids can be produced by standard techniques.

[0056] In a particular embodiment, the antisense nucleic acid is wholly or partially complementary to and can hybridize with a target nucleic acid, wherein the target nucleic acid can hybridize to a nucleic acid having the sequence of the complement of the strand shown in FIG. 2 (SEQ ID NO: 1). For example, antisense nucleic acid can be complementary to a target nucleic acid having the sequence shown as the open reading frame in FIG. 2 (SEQ ID NO: 1) or to a portion thereof sufficient to allow hybridization. In another embodiment, the antisense nucleic acid is wholly or partially complementary to and can hybridize with a target nucleic acid which encodes a mammalian hyaluronan synthase.

[0057] The nucleic acids can also be used as probes (e.g., in in situ hybridization) to assess associations between inflammatory settings (e.g., rheumatoid arthritis, osteoarthritis, liver cirrhosis, Werner's syndrome, renal failure and psoriasis) and increased expression of mammalian hyaluronan synthase in affected tissues or serum. The nucleic acids can also be used as probes to detect and/or isolate (e.g., by hybridization with RNA or DNA) polymorphic or allelic variants, for example, in a sample (e.g., inflamed tissue) obtained from a host (e.g. mammalian). Moreover, the presence or frequency of a particular variant in a sample(s) obtained from one or more affected hosts, as compared with a sample(s) from normal host(s), can be indicative of an association between an inflammatory setting and a particular variant, which in turn can be used in the diagnosis of the condition.

[0058] As described in the exemplification, functional expression cloning was used to identify a cDNA encoding human hyaluronan synthase, and it was demonstrated that this gene can confer activity both in the synthesis of hyaluronan and as a mediator of cell adhesion by the lymphocyte receptor CD44. A human hyaluronan synthase (HAS) cDNA was isolated by a functional expression cloning approach. Transfection of CHO cells conferred hyaluronidase sensitive adhesiveness of a mucosal T cell line via the lymphocyte hyaluronan receptor, CD44, as well as increased hyaluronan levels in the cultures of transfected cells. The HAS amino acid sequence shows homology to the hasA gene product of Streptococcus pyogenes and a putative glycosaminoglycan synthetase from xenopus laevis. Expression of HAS message parallels tissues where high levels of hyaluronan synthesis occur, indicating that transcription of synthase mRNA is a critical component of hyaluronate synthesis.

[0059] Utilities

[0060] Mammalian hyaluronan synthases of the present invention can be used to produce hyaluronan. Hyaluronan has a variety of uses, including use in cosmetics and pharmaceuticals (see e.g., EPO,443,043 B1 and U.S. Pat. No. 5,015,577 the teachings of which are each incorporated herein by reference). Hyaluronan or pharmaceutical compositions comprising hyaluronan are useful for treating wounds or surgical incisions and can reduce or prevent hypertrophic scars and keloid formation, and in eye surgery as a replacement for vitreous fluid, for example.

[0061] For example, a mammalian hyaluronan synthase can be expressed in a suitable host cell under conditions

appropriate for production of hyaluronan to occur (e.g., in suitable medium comprising any required precursors). Isolated or purified hyaluronan synthase can also be used to prepare hyaluronan from precursors (e.g., UDP-glucuronic acid and UDP-N-aceytl-glucosamine).

[0062] The present invention also provides antibodies which (1) can bind a "mammalian hyaluronan synthase" in vitro and/or in vivo; and/or (2) can inhibit an activity or function characteristic of a "mammalian hyaluronan synthase", such as hyaluronan synthesis. Preferably the antibodies are capable of selective binding of mammalian hyaluronan synthase in vitro and/or in vivo (e.g., bind selectively to mammalian hyaluronan synthase expressed in ovary and/or spleen, thymus, prostate, etc. (e.g., as assessed immunohistologically)).

[0063] Preferably, the antibodies can bind a mammalian (e.g. human) hyaluronan synthase with high affinity (for example, a Ka in the range of about 1-10 nM, or a Kd in the range of about $133 \ 10^{-8}$ to $1 \times 10^{-10} \ \text{mol}^{-1}$).

[0064] The antibodies of the present invention are useful in a variety of applications, including processes, research, diagnostic and therapeutic applications. For instance, they can be used to isolate and/or purify mammalian hyaluronan synthase or variants thereof (e.g., by affinity purification or other suitable methods), and to study mammalian hyaluronan synthase structure (e.g., conformation) and function.

[0065] The antibodies of the present invention can also be used to modulate mammalian hyaluronan synthase function in diagnostic (e.g., in vitro) or therapeutic applications. For instance, antibodies can act as inhibitors of (reduce or prevent) hyaluronan synthesis, thereby inhibiting process mediated by hyaluronan such as cell adhesion and metastasis.

[0066] In addition, antibodies of the present invention can be used to detect and/or measure the level of a mammalian hyaluronan synthase in a sample (e.g., tissues or body fluids, such as an inflammatory exudate, blood, serum, bowel fluid, or on cells transfected with a nucleic acid of the present invention). For example, a sample (e.g., tissue and/or fluid) can be obtained from a host (e.g., mammalian) and a suitable immunological method can be used to detect and/or measure mammalian hyaluronan synthase levels, including methods such as enzyme-linked immunosorbent assays (ELISA), including chemiluminescence assays, radioimmunoassay, and immunohistology. In one embodiment, a method of detecting a selected mammalian hyaluronan synthase in a sample is provided, comprising contacting a sample with an antibody which binds an isolated mammalian hyaluronan synthase under conditions suitable for specific binding of said antibody to the selected mammalian hyaluronan synthase, and detecting antibody-mammalian hyaluronan synthase complexes which are formed.

[0067] In an application of the method, antibodies reactive with a mammalian hyaluronan synthase can be used to analyze normal versus inflamed tissues in mammals for mammalian hyaluronan synthase reactivity and/or expression (e.g., immunohistologically). Thus, the antibodies of the present invention permit immunological methods of assessment of expression of primate (e.g., human mammalian hyaluronan synthase) in normal versus inflamed tissues, through which the presence of disease, disease progress

and/or the efficacy of anti-mammalian hyaluronan synthase therapy in inflammatory disease can be assessed.

[0068] An antibody can be administered in an effective amount which inhibits mammalian hyaluronan synthase activity. For therapy, an effective amount will be sufficient to achieve the desired therapeutic and/or prophylactic effect (such as an amount sufficient to reduce or prevent mammalian hyaluronan synthase-mediated hyaluronan synthesis). The antibody can be administered in a single dose or multiple doses. The dosage can be determined by methods known in the art and is dependent, for example, upon the individual's age, sensitivity, tolerance and overall well-being. Suitable dosages for antibodies can be from 0.1-1.0 mg/kg body weight per treatment.

[0069] According to the method, an antibody can be administered to an individual (e.g., a human) alone or in conjunction with another agent (administered before, along with or subsequent to administration of the additional agent).

[0070] A variety of routes of administration are possible including, but not necessarily limited to parenteral (e.g., intravenous, intraarterial, intramuscular, subcutaneous injection), oral (e.g., dietary), topical, inhalation (e.g., intrabronchial, intranasal or oral inhalation, intranasal drops), or rectal, depending on the disease or condition to be treated. Parenteral administration is a preferred mode of administration.

[0071] Formulation will vary according to the route of administration selected (e.g., solution, emulsion, capsule). An appropriate composition comprising the antibody to be administered can be prepared in a physiologically acceptable vehicle or carrier. For solutions or emulsions, suitable carriers include, for example, aqueous or alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles can include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's or fixed oils. Intravenous vehicles can include various additives, preservatives, or fluid, nutrient or electrolyte replenishers (See, generally, Remington's Pharmaceutical Science, 16th Edition, Mack, Ed. 1980). For inhalation, the compound can be solubilized and loaded into a suitable dispenser for administration (e.g., an atomizer, nebulizer or pressurized aerosol dispenser).

Exemplification

[0072] Plasmids, Monoclonal Antibodies and Cell Lines

[0073] The following plasmids were used as controls in expression cloning and for functional adhesion assays: pSV-SPORT-1 (GIBCO, Gaithersburg, Md.) or pcDNA3 (Invitrogen, San Diego, Calif.) controls and murine MAdCAM-1 in pCDM8 (pCDMAD-7 (Briskin, M. J., *Nature* 363:461-464 (1993)). Monoclonal antibodies used were anti-murine CD-44 TJB1.7 (a gift from T. Yoshino and E. Butcher, Stanford, Calif.); anti-murine MAdCAM-1 MECA-367 (Streeter, P. R. et al., *Nature* 331:41-46 (1988)); anti-human VCAM-1 2G7 (Graber, N. *J. Immunol.* (145):819 (1990)); anti-murine β7 FIB 504 (Andrew, D. P. et al., *J. Immunol.* 153:3847-3861 (1994)); and anti-murine α4 PS/2 (Miyake, K. *J. Exp. Med.* 173:599-607 (1991)). Cell lines used for expression cloning and functional adhesion assays were: CHO/P (Heffernan, M. and Dennis, J. D. *Nucl. Acids Res.*

19:85 (1991)) and the murine T cell lymphoma TK1 (Butcher, E. C. et al., Eur. J. Immunol. 10:556-561 (1980)).

[0074] cDNA Synthesis and Library Construction

[0075] mRNA was isolated from human lymph nodes using standard procedures previously described (Briskin, M. J., *Nature* 363:461-464 (1993)). cDNA was synthesized using the Superscript™ lambda system in conjunction with the pSV-SPORT-1 vector (Gibco, Gaithersburg, Md.) essentially using the manufacturer's protocol. The highest molecular weight fractions (>1.5 kb) of cDNA were ligated into the pSV-SPORT-1 vector and plated in pools at a density of 5,000 clones/plate on 100 LB agar plates with ampicillin (50 µg/ml). After incubation overnight, plasmid DNAs were purified from each plate individually by use of QIAprep spin columns (QIAGEN, Chatsworth, Calif.) according to manufacturer's instructions.

[0076] Expression Cloning

[0077] CHO/P cells were seeded into 24 well plates approximately 24 hours prior to transfection at a density of 40,000 cells/well. DNAs were transiently transfected using the LipofectAMINE™ reagent (GIBCO, Gaithersburg, Md.) as recently described (Shyjan, A. M. et al., *J. Immunol.*, 156:2851-2857 (1996)).

[0078] For the adhesion assays in the expression cloning screen, TK1 cells are resuspended at a density of 2×10⁶/ml in a cell binding assay buffer previously described (Shyjan, A. M. et al., J. Immunol. in press (1996)). After incubation at 4° C. for 15 minutes, 0.25 ml of the TK1 cell suspension $(5\times10^5 \text{ TK1 cells})$ was added to each well and incubation on a rocking platform was continued for an additional 30 minutes at 4° C. Plates were washed by gently inverting in a large beaker of phosphate buffered saline (PBS) followed by inversion in a beaker of PBS with 1.5% glutaraldehyde for fixation for a minimum of 1 hour. Wells were then examined microscopically (10x objective) for rosetting of TK1 cells mediated by the pools of cDNA clones. Pools yielding one or more TK1 rosettes were further subfractionated three times until individual colonies could be assayed and the clones conferring adhesion of the TK1 cells were identified.

[0079] Functional Adhesion Assays

[0080] Assays with purified clones were similar to those performed in expression cloning with the following exception: as several wells were to be transfected for antibody inhibition studies, a master liposome mix with multiples of the wells to be transfected was first made for each plasmid. On the day of the assay monoclonal antibodies were incubated with cells at $20 \,\mu\text{g/ml}$ or supernatants (undiluted) at 4° C. for 15 minutes prior to the start of the assay.

[0081] For adhesion assays with hyaluronan, human umbilical cord hyaluronan (Calbiochem, San Diego, Calif.) was diluted to 5 mg/ml in PBS. Streptomyces hyaluronidase (Calbiochem, San Diego, Calif.) was diluted to 20 TRU/ml in HBSS. TK1 cells were resuspended in HBSS containing 2 mM CaCl₂, 2 mM MgCl₂, 2% serum and 20 mM HEPES at 10⁶ cells/ml. Wells of 24-well plates were coated with 200 μ l of hyaluronan and stored at 4° C. overnight. Wells were rinsed with 0.5 ml PBS three times, and were treated with 0.25 ml Streptomyces hyaluronidase at final concentrations of 0, 5, 10 and 20 TRU/ml for 1 hour at 37° C. Wells were

rinsed three times with 0.5 ml PBS, blocked with 0.5 ml serum for 1 hour on ice and then rinsed three times with 0.5 ml PBS. TK1 cells (0.5 ml) were added to each well and plates were incubated with shaking at 4° C. for 20 minutes.

[0082] For assessment of hyaluronate mediated binding to CHO/P cells, the transfectants were rinsed with 0.5 ml PBS three times. Individual wells were treated with 250 μ l Streptomyces hyaluronidase at 0, 5, 10 and 20 TRU/ml (final concentrations) for 1 hour at 37° C. Transfectants were rinsed three times with 0.5 ml PBS. TK1 cells (0.5 ml in the same buffer as described above) were added to each well and plates were incubated with shaking at 4° C. for 30 minutes. Wells were rinsed with 0.5 ml PBS three times and viewed under the light microscope. Assays were fixed as described above and analyzed by examination of multiple fields and counting both lymphocytes and CHO cells at 10× magnification.

[0083] Measurement of Hyaluronic Acid Biosynthesis in CHO Cell Transfectants

[0084] 0.5×10^6 CHO cells seeded in 100 mm plates were transfected with Lipofectamine reagent according to manufacaturer's instructions. Transections utilized 20 μ g of HAS cDNA in pcDNA3 (Invitrogen, San Diego, Calif.) and 160 μ l of lipofectamine reagent. Clone 30C was digested with EcoRI and NotI and the insert released thereby was cloned into the ECORI and NotI sites of pcDNA3. Transformants of E. coli XL-1 Blue (Stratagene) or DH10B (Gibco) containing the resulting construct were obtained. Approximately 72 hours after transfection, 440 µg/ml of G418 was added in fresh media. After the transfected and control (non transfected) cells had reached subconfluency, the media was replaced with fresh complete media containing 5 mCi/ml D-[6-3H] glucosamine hydrochloride (New England Nuclear, Boston, Mass., specific activity 33.3 ci/ml, concentration 1 mCi/ml), a precursor of sulfated glucosaminoglycans such as hyaluronan. The amounts of synthesized hyaluronan in transfected and control CHO cells were determined after 48 hours of incubation at 37° C. as follows. Media was collected and the cell layers were combined with the corresponding media. Aliquots from each sample were incubated overnight at 37° C. in the presence or absence of Streptomyces hyaluronidase. Then the samples were applied on sephadex G-50 superfine columns (100×100 mm) which were equilibrated with 0.05 M sodium acetate, pH 6.0 containing 0.2M NaCl. Newly synthesized [3H] hyaluronan was determined as the Streptomyces sensitive radioactivity.

[0085] DNA Sequencing

[0086] Plasmids were sequenced on both strands using oligonucleotide primers and the sequenase™ 7-deaza-dGTP DNA sequencing kit with sequenase version 2.0 T7 DNA polymerase (United States Biochemical, Cleveland, Ohio) and ³⁵SdCTP (Amersham Life Science, Arlington Heights, Ill. and New England Nuclear, Boston, Mass.) using manufacturer's instructions.

[0087] Northern and Southern Blot Analysis

[0088] Northern blots used were human multiple tissue northerns I and II (Clontech, Palo Alto, Calif.). Hybridization was performed with ExpressHyb (Clontech) solution, using manufacturer's instructions except that a final wash at high stringency (0.1×SSC, 0.1% SDS, 65° C.) for 30 min was added. A commercially prepared southern blot (Human

GENO-BLOT) (Clontech, Palo Alto, Calif.) was hybridized as described for the Northern blot with the exception that an initial wash at 50° C. was exposed and then the blot was subsequently washed at 65° C. and exposed again. cDNA's were labelled with α^{32} P-dCTP by priming with random hexamers. After washing, filters were exposed to Kodak XAR film with an intensifying screen.

[0089] Results and Discussion

[0090] An expression cloning system was developed to isolate cDNA clones that encode proteins that confer adhesion of the murine T cell lymphoma TK1 (Butcher, E. C. et al., Eur. J. Immunol. 10:556-561 (1980)). A human mesenteric lymph node expression library was constructed that, upon transfection into CHO/P cells, yielded a cDNA clone, called 30C, that mediated rosetting of TK1 cells to some of the CHO/P transfectants. Transformants of E. coli XL-1 Blue (Stratagene) containing Clone 30C were obtained. In order to understand the nature of the observed interaction, the adhesion assay after pre-incubation of the TK1 line with several antibodies to adhesion receptors known to be expressed on TK1 cells was repeated. Binding could be completely inhibited by pre-incubation of TK1 cells with an antibody to CD44 (Table 1), while other antibodies (anti-α4 and anti-β7 integrins (Andrew, D. P. et al., J. Immunol. 153:3847-3861 (1994); Miyake, K. J. Exp. Med. 173:599-607 (1991)) had no effect.

TABLE 1

<u> </u>	Adhesion of TK	1 cells to clone 30	C transfectar	ıts.
Cells/Matrix	Tk1 Cell	TK1 Binding after hyaluronidase	Tk1 Binding after anti-CD44 MAb TJB1.7	TK1 Binding after anti-a4 MAb PS/2
HAS	+++	_	_	+++
Transfectants Mock Transfectants	_	_	_	_
Hyaluronate	+++	_	_	+++

TK1 cells bind to CHO/P cells transiently transfected with clone 30C. Binding is blocked by pretreatment of the transfectants with hyaluronidase or pretreatment of TK1 cells with anti-CD44 MAb TJB1.7. Similar results are seen with binding to immobilized hyaluronate, while TK1 cells do not bind mock transfectants. A score of "—" indicates that no TK1 cells (above controls) were observed in those wells while "+++" indicates TK1 rosetting on transfectants (>5 TK1 cells/CHO/P transfectant) or a monolayer of cells binding to immobilized hyaluronate. Assays were all repeated three times with similar results.

[0091] As CD44 is known to be a hyaluronan receptor (Aruffo, A., et al., *Cell* 61:1303-1313 (1990); Culty, M. et al., *J. Cell. Biol.*, 111:2765-2774 (1990); Miyake, K. et al., *J. Exp. Med.* 172:69-75 (1990)), it was investigated whether the isolated cDNA encoded a novel CD44 ligand or, alternatively, was involved in de novo synthesis of hyaluronan. Hyalurondase pretreatment completely abrogated TK1 binding to the transfectants as well as to hyaluronan controls (Table 1), indicating that the cloned cDNA mediated synthesis of HA. Finally, CHO cells were stably transfected with the 30C cDNA and assessed for their ability to mediate hyaluronan biosynthesis (FIG. 1A,B). Whereas, untranfected cells produced very little high molecular weight Streptomyces hyaluronaidase-sensitive material (FIG. 1B),

cell cultures transfected with 30C cDNA produced a substantial amount of hyaluronan (FIG. 1A).

[0092] The cDNA encoding clone 30C is 2116 nucleotides in length (FIG. 2) with a short 5' untranslated region of 35 bp and a longer 3' untranslated region of 347 bp. From the first ATG, a predicted open reading frame of 1734 bp yielding a protein of 578 amino acid residues is present. Genbank searches of the nucleotide and protein sequences revealed significant homology with the hasA gene of Streptococcus pyogenes (DeAngelis, J. P. a. P. H. W., J. Biol. Chem. 268:19181-19184 (1993)), which was reported to be a hyaluronan synthase (FIG. 3A,B) and a sequence from Xenopus laevis called DG42 (FIG. 3A,B) which has also been speculated to be a glycosaminoglycan synthetase (Rosa, F. et al., Develop. Biol. 129:114-123 (1988)). Amino acid sequence identities between the predicted protein and these sequences were 22% and 54%, respectively. Significant similarity was also observed with other membrane associated proteins with N-acetylyglucosylamino transferase activity including NodC from Rhizobium and three chitin synthases from Saccharomyces (Chs) (DeAngelis, P. L. et al., Biochem. and Biophys. Res. Comm. 199:1-10 (1994)). The similarities observed, coupled with the functional adhesion indicate that clone 30C encodes a human homolog of hyaluronan synthase (HS). Using nomenclature based on the streptococcus gene locus, this human gene encoding hyaluronan synthase is designated HAS.

[0093] The predicted molecular mass of the HAS protein is 64,793 daltons. Hydrophilicity (Kyte-Doolitle) analysis predicts a membrane protein with several hydrophobic regions that would be predicted to span the cell membrane at least four times (FIG. 3A-C). This prediction is in agreement with labeling studies which suggested that hyaluronan synthase is associated with the plasma membrane (Prehm, P., Biochem. J. 220:597-600 (1984); Phillipson, L. H. and Schwartz, N. B. J. Biol. Chem. 259:5017-5023 (1984); Klewes, L. et al., Biochem J. 290:791-795 (1993); O'Regan, M. et al., Int. J. Biol. Macromol. 16:283-286 (1994)). Conservation of secondary structure between hasA, DG42 and HAS, is indicated by similar hydrophilicity plots. The approximate locations of these regions, with respect to HAS, are shown in the alignment in FIG. 3A and their representative hydrophilicity plots are shown in FIG. 3B.

[0094] The estimated number of transmembrane segments would suggest a structure with a small N-terminal extracellular domain followed by a long intracellular loop and then three more transmembrane regions to yield one more small extracellular loop, a small intracellular loop followed by a C-terminal extracellular extension (FIG. 3C). Such a model, with the predominant portion of the protein located intracellularly would be consistent with studies indicating that hyaluronan biosynthesis occurs at the inner surface of the plasma membrane (Prehm, P. Biochem. J. 220:597-600 (1984); Phillipson, L. H., and Schwartz, N. B. J. Biol. Chem. 259:5017-5023 (1984)). This predicted large intercellular loop, is more highly conserved than the overall protein at 70% (vs 54%) when compared with DG42, which would imply conservation of a functional domain. Within the amino terminal portion of this domain lies a motif, designated B(X₇)B (FIG. 2, 3C), where B is a basic amino acid (e.g., R, K) and X is any non-acidic residue. This motif has been found in both RHAMM, link protein and CD44, and mutagenesis studies has shown that this sequence is required

for binding hyaluronan (Yang, B., et al., *EMBO* 13:286-296 (1994)). The presence of this putative hyaluronan binding motif (HBM) in HAS raises the possibility of a requirement of binding hyaluronan during its synthesis and prior to transport out of the cell.

[0095] Northern blots probed with the entire human cDNA, revealed a major transcript of 2.4 kb that was most highly expressed in ovary and also expressed at significant levels in spleen, thymus, prostate, testes and large intestine (FIG. 4A). In addition, a less abundant transcript of approximately 7 kb was also observed in these tissues and in addition to a faint 9 kb species only expressed in ovary. Extremely weak expression was observed in small intestine while peripheral blood leukocytes (PBL) were negative under the conditions used. Moderate expression was also observed in heart. The larger transcript observed might be a related gene in these tissues although a southern blot probed first with both full length and then a 3' region of HAS cDNA and washed at several temperatures shows a simple banding pattern suggestive of a single copy gene (FIG. 4B). It is therefore likely that these larger species represent unprocessed nuclear precursors, as opposed to related genes. The expression pattern observed is consistent with high levels of hyaluronan that are observed in lymphoid tissues, preovulatory follicles and in perivascular connective tissue and vessel walls of both atrium and ventricle (Edelstrom, G. A. B. et al., Histochem. Cytochem., 39:1131-1135 (1991); Laurent, C. et al., Cell Tissue Res., 263: 201-205 (1991)) and would indicate that synthesis of hyaluronan is at least partially regulated by transcriptional mechanisms. Interestingly, however, expression of HAS RNA was barely detectable in skeletal muscle under the conditions used, although histochemical analysis has shown ubiquitous distribution of hyaluronan in connective tissue and the septum dividing muscle fibers (Edelstrom, G. A. B. et al., Histochem. Cytochem 39:1131-1135 (1991); Laurent, C. et al., Cell Tissue Res. 263: 201-205 (1991)). This may indicate that turnover rates of hyaluronan may display great variation in different tissues.

[0096] Induction of synthase activity by growth factors has been shown to require protein synthesis and is mediated by a signaling pathway involving tyrosine phosphorylation and/or activation of protein kinase C (Heldin, P. et al., Biochem. J. 258, 919-922 (1992); Suzuki, M. et al., Biochem. J. 307:817-821 (1995)) as both PMA and inhibitors of phosphotyrosine phosphatases can induce hyaluronan synthesis. Serum alone can also induce synthase activity and this induction was blocked by protein kinase C inhibitors and cycloheximide. cAMP has also been implicated in activation and phosphorlyation of the synthase itself may play a key role in regulation of its activity (Klewes, L. and Prehm, P., J. of Cell. Physiol. 160:539-544 (1994)). Examination of hydrophilic regions of HAS reveals several conserved motifs which are potential substrates for protein kinase C and cAMP dependent kinases (FIGS. 2, 3C.) and are likely targets for future mutagenesis studies (Pearson, R. B. Studies of protein kinase/phosphatase specificity using synthetic peptides. Protein phosphorylation: A practical approach (Hardie, D. G., Ed.), Oxford University Press, Oxford (1993)). As observed, increased expression of the HAS gene in tissues that are known to produce large quantities of hyaluronan, it is likely that the regulation of hyaluronan synthesis is mediated by regulation of HAS gene transcription, in addition to complex regulatory circuits which involve both alterations in phosphorylation of the synthase or proteins associated with HAS.

[0097] Previously, a 52 kDa protein was isolated from a mouse/hamster hybridoma (B6 cells) that was initially reported to be a mammalian hyaluronan synthase (Klewes, L. et al., Biochem J. 290:791-795 (1993)). This protein was incapable of binding UDP-Glucuronic acid (UDP-[14C] GlcA) and UDP-N-acetyl glucosamine (UDP-[3H] GlcNAc) unless complexed to a 60 kDa protein, which may be the hyaluronan receptor (RHAMM) recently implicated in fibroblast migration and tumor metastasis (Turley, E. A. et al., J. Cell Biol., 112:1041-1047 (1991)). This protein crossreacted with antibodies against a putative synthase from Streptococcus equisimilis. The gene encoding this protein was cloned from a streptoccal library and shown to be related to proteins involved in oligopeptide processing and transport and showed no homology to the hasA gene sequence (O'Regan, M. et al., Int. J. Biol. Macromol. 16:283-286 (1994); Lansing, M. et al., Biochem. J. 289:179-184 (1993)). It is likely that the 52 kd protein isolated from the B6 line is a homolog to the streptococcal transport protein and not the synthase itself. The human hyaluronan synthase cDNA is therefore the first example of a mammalian gene responsible for synthesis of hyaluronan.

[0098] Studies in streptococci show that the machinery responsible for synthesis of hyaluronan is encoded in the has operon which consists of three genes has A, B and C (Dougherty, B. P., and van de Rijn, I. J. Biol. Chem. 269:169-175 (1994); Dougherty, B. P., and van de Rijn, I. J. Biol. Chem. 268:7118-7124 (1993); Crater, D. L., and van de Rijn, I. J. Biol. Chem. 270:18452-18458 (1995)). It has been demonstrated that HAS is homologous to hasA which encodes hyaluronan synthase, along with a recently cloned cDNA encoding the murine synthase (Has) as well. The hasB and C loci encode UDP:Glc dehydrogenase and UDP-GLc pyrophosphorylase respectively (Dougherty, B. P., and van de Rijn, I. J. Biol. Chem. 269:169-175 (1994); Dougherty, B. P., and van de Rijn, I. J. Biol. Chem. 268:7118-7124 (1993); Crater, D. L., and van de Rijn, I. J. Biol. Chem. 270:18452-18458 (1995)). Also demonstrated herein is that transfection of the HAS cDNA into CHO cells is sufficient to mediate de novo synthesis of hyaluronan, which indicates that all of the other factors necessary for hyaluronan biosynthesis such as those encoded by hasB and C are possibly expressed in CHO cells. Recent data suggests that hyaluronan can also be synthesized upon transfection of the synthase into COS cells and a murine preB lymphoma which suggests that these backgrounds have endogenous UDP-GLc dehydrogenase and UDP-GLc phosphorylase and expression of HAS is then the most significant factor in regulating hyaluronan synthesis in mammalian cells. The identification of this cDNA will therefore assist further characterization of the molecular events resulting in synthesis of hyaluronan and its relationship to cellular migration in wound healing, tumor metastasis and leukocyte migration.

[0099] Equivalents

[0100] Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described specifically herein. Such equivalents are intended to be encompassed in the scope of the following claims.

SEQUENCE LISTING

(1)	GENERAL	INFORMATION:
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(iii) NUMBER OF SEQUENCES: 4

(2) INFORMATION FOR SEQ ID NO: 1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2116 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 36..1769
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

	(xi) SEC	QUEN	CE DI	ESCR	IPTIC	ON: S	SEQ :	ID NO): 1	:				
CGG	AGAGA	AAG 1	AGAG	AGCC	CG GO	CCAG	ACCC	A CTO			AGA (53
	AAG Lys													•	101
	CTG Leu														149
	TAC Tyr 40													;	197
	TTC Phe													;	245
	CTC Leu													:	293
	CCG Pro													;	341
	TAC Tyr													;	389
	GCC Ala 120														437
	GGC Gly														485
	TTC Phe													!	533
	CAG Gln													!	581
	CGG Arg													•	629

_								 	
			CGC Arg						677
L			ATG Met						725
			GTC Val 235						773
			GTG Val						821
			GTG Val						869
			CTG Leu						917
С			TTC Phe						965
			AAC Asn 315						1013
			GGT Gly						1061
			CTC Leu						1109
			TCA Ser						1157
G			TGG Trp						1205
			CAC His 395						1253
			TTC Phe						1301
			CGC Arg						1349
			CTG Leu						1397
С			GTG Val						1445
			GCC Ala 475						1493
			TCG Ser						1541

12

-continued

CTG CTG CCC CTG GCG CTC TGG GCG CTG CTG	1589
CGC AGC GTA GCA CAC GAG GCC AGG GCC GAC TGG AGC GGC CCT TCC CGC Arg Ser Val Ala His Glu Ala Arg Ala Asp Trp Ser Gly Pro Ser Arg 520 525 530	1637
GCA GCC GAG GCC TAC CAC TTG GCC GCG GGG GCC GGC GCC TAC GTG GGC Ala Ala Glu Ala Tyr His Leu Ala Ala Gly Ala Gly Ala Tyr Val Gly 535 540 550	1685
TAC TGG GTG GCC ATG TTG ACG CTG TAC TGG GTG GGC GTG CGG AGG CTT Tyr Trp Val Ala Met Leu Thr Leu Tyr Trp Val Gly Val Arg Arg Leu 555 560 565	1733
TGC CGG CGG CGG ACC GGG GGC TAC CGC GTC CAG GTG TGAGTCCAGC Cys Arg Arg Thr Gly Gly Tyr Arg Val Gln Val 570 575	1779
CACGCGGATG CCGCCTCAAG GGTCTTCAGG GGAGGCCAGA GGAGAGCTGC TGGGCCCCGA	1839
GCCACGAACT TGCTGGGTGG TTCTCTGGGC CTCAGTTTCC CTCCTCTGCC AAACGAGGGG	1899
GTCAGCCCAA GATTCTTCAG TCTGGACTAT ATTGGGACTG GGACTTCTGG GTCTCCAGGG	1959
AGGGTATTTA TTGGTCAGGA TGTGGGATTT GAGGAGTGGA GGGGAAAGGG TCCTGCTTTC	2019
TCCTCGTTCT TATTTAATCT CCATTTCTAC TGTGTGATCA GGATGTAATA AAGAATTTTA	2079
TTTATTTCA AAAAAAAAA AAAAAAAAA AAAAAAA	2116

(2) INFORMATION FOR SEQ ID NO: 2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 578 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Arg Gln Gln Asp Ala Pro Lys Pro Thr Pro Ala Ala Arg Arg Cys $1 \hspace{1.5cm} 1 \hspace{1.5cm} 5 \hspace{1.5cm} 15$

Ser Gly Leu Ala Arg Arg Val Leu Thr Ile Ala Phe Ala Leu Leu Ile $20 \ \ 25 \ \ 30$

Leu Gly Leu Met Thr Trp Ala Tyr Ala Ala Gly Val Pro Leu Ala Ser \$35\$

Ala His Leu Val Ala Gln Ser Leu Phe Ala Tyr Leu Glu His Arg Arg 65 70 75 80

Gln Cys Leu Ala Ser Ala Arg Ala Leu Leu Tyr Pro Arg Ala Arg Leu 115 120 125

Arg Val Leu Met Val Val Asp Gly Asn Arg Ala Glu Asp Leu Tyr Met 130 135 140

Val Asp Met Phe Arg Glu Val Phe Ala Asp Glu Asp Pro Ala Thr Tyr 145 150 155 160

Val Trp Asp Gly Asn Tyr His Gln Pro Trp Glu Pro Ala Ala Ala Gly 165 170 175

Ala	Val	Gly	Ala 180	Gly	Ala	Tyr	Arg	Glu 185	Val	Glu	Ala	Glu	Asp 190	Pro	Gly
Arg	Leu	Ala 195	Val	Glu	Ala	Leu	Val 200	Arg	Thr	Arg	Arg	C y s 205	Val	Сув	Val
Ala	Gln 210	Arg	Trp	Gly	Gly	Lys 215	Arg	Glu	Val	Met	Tyr 220	Thr	Ala	Phe	Lys
Ala 225	Leu	Gly	Asp	Ser	Val 230	Asp	Tyr	Val	Gln	Val 235	Сув	Asp	Ser	Asp	Thr 240
Arg	Leu	Asp	Pro	Met 245	Ala	Leu	Leu	Glu	Leu 250	Val	Arg	Val	Leu	A sp 255	Glu
Asp	Pro	Arg	Val 260	Gly	Ala	Val	Gly	Gly 265	Asp	Val	Arg	Ile	Leu 270	Asn	Pro
Leu	Asp	Ser 275	Trp	Val	Ser	Phe	Leu 280	Ser	Ser	Leu	Arg	Tyr 285	Trp	Val	Ala
Phe	Asn 290	Val	Glu	Arg	Ala	C y s 295	Gln	Ser	Tyr	Phe	His 300	Cys	Val	Ser	Cys
Ile 305	Ser	Gly	Pro	Leu	Gly 310	Leu	Tyr	Arg	Asn	Asn 315	Leu	Leu	Gln	Gln	Phe 320
Leu	Glu	Ala	Trp	Tyr 325	Asn	Gln	Lys	Phe	Leu 330	Gly	Thr	His	Сув	Thr 335	Phe
Gly	Asp	Asp	Arg 340	His	Leu	Thr	Asn	Arg 345	Met	Leu	Ser	Met	Gly 350	Tyr	Ala
Thr	Lys	Tyr 355	Thr	Ser	Arg	Ser	Arg 360	Cys	Tyr	Ser	Glu	Thr 365	Pro	Ser	Ser
Phe	Leu 370	Arg	Trp	Leu	Ser	Gln 375	Gln	Thr	Arg	Trp	Ser 380	Lys	Ser	Tyr	Phe
Arg 385	Glu	Trp	Leu	Tyr	Asn 390	Ala	Leu	Trp	Trp	His 395	Arg	His	His	Ala	Trp 400
Met	Thr	Tyr	Glu	Ala 405	Val	Val	Ser	Gly	Leu 410	Phe	Pro	Phe	Phe	Val 415	Ala
Ala	Thr	Val	Leu 420	Arg	Leu	Phe	Tyr	Ala 425	Gly	Arg	Pro	Trp	Ala 430	Leu	Leu
Trp	Val	Leu 435	Leu	Сув	Val	Gln	Gly 440	Val	Ala	Leu	Ala	Lys 445	Ala	Ala	Phe
Ala	Ala 450	Trp	Leu	Arg	Gly	Cys 455	Leu	Arg	Met	Val	Leu 460	Leu	Ser	Leu	Tyr
Ala 465	Pro	Leu	Tyr	Met	Cys 470	Gly	Leu	Leu	Pro	Ala 475	Lys	Phe	Leu	Ala	Leu 480
Val	Thr	Met	Asn	Gln 485	Ser	Gly	Trp	Gly	Thr 490	Ser	Gly	Arg	Arg	Lys 495	Leu
Ala	Ala	Asn	Tyr 500	Val	Pro	Leu	Leu	Pro 505	Leu	Ala	Leu	Trp	Ala 510	Leu	Leu
Leu	Leu	Gly 515	Gly	Leu	Val	Arg	Ser 520	Val	Ala	His	Glu	Ala 525	Arg	Ala	Asp
Trp	Ser 530	Gly	Pro	Ser	Arg	Ala 535	Ala	Glu	Ala	Tyr	His 540	Leu	Ala	Ala	Gly
Ala 545	Gly	Ala	Tyr	Val	Gl y 550	Tyr	Trp	Val	Ala	Met 555	Leu	Thr	Leu	Tyr	Trp 560

Val Gly Val Arg Arg Leu Cys Arg Arg Arg Thr Gly Gly Tyr Arg Val

Gln Val

- (2) INFORMATION FOR SEQ ID NO: 3:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 587 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: <Unknown>
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

Met Lys Glu Lys Ala Ala Glu Thr Met Glu Ile Pro Glu Gly Ile Pro 1 5 10 15

Lys Asp Leu Glu Pro Lys His Pro Thr Leu Trp Arg Ile Ile Tyr Tyr
20 25 30

Ser Phe Gly Val Val Leu Leu Ala Thr Ile Thr Ala Ala Tyr Val Ala 35 404045

Glu Phe Gln Val Leu Lys His Glu Ala Ile Leu Phe Ser Leu Gly Leu 50 60

Tyr Gly Leu Ala Met Leu Leu His Leu Met Met Gln Ser Leu Phe Ala 65 70 75 80

Phe Leu Glu Ile Arg Arg Val Asn Lys Ser Glu Leu Pro Cys Ser Phe 85 90 95

Lys Lys Thr Val Ala Leu Thr Ile Ala Gly Tyr Gln Glu Asn Pro Glu 100 105 110

Tyr Leu Ile Lys Cys Leu Glu Ser Cys Lys Tyr Val Lys Tyr Pro Lys 115 120 125

Asp Lys Leu Lys Ile Ile Leu Val Ile Asp Gly Asn Thr Glu Asp Asp 130 135 140

Ala Tyr Met Met Glu Met Phe Lys Asp Val Phe His Gly Glu Asp Val 145 150155155

Glu Thr Asn Lys Gly Ser Cys Pro Glu Val Ser Lys Pro Leu Asn Glu 180 185 190

Asp Glu Gly Ile Asn Met Val Glu Glu Leu Val Arg Asn Lys Arg Cys $195 \hspace{1.5cm} 200 \hspace{1.5cm} 205 \hspace{1.5cm}$

Val Cys Ile Met Gln Gln Trp Gly Gly Lys Arg Glu Val Met Tyr Thr 210 215 220

Ala Phe Gln Ala Ile Gly Thr Ser Val Asp Tyr Val Gln Val Cys Asp 225 230 235 240

Ser Asp Thr Lys Leu Asp Glu Leu Ala Thr Val Glu Met Val Lys Val 245 250 255

Leu Glu Ser Asn Asp Met Tyr Gly Ala Val Gly Gly Asp Val Arg Ile $260 \hspace{1.5cm} 265 \hspace{1.5cm} 270 \hspace{1.5cm}$

Leu Asn Pro Tyr Asp Ser Phe Ile Ser Phe Met Ser Ser Leu Arg Tyr 275 280 285

Trp Met Ala Phe Asn Val Glu Arg Ala Cys Gln Ser Tyr Phe Asp Cys 290 295 300

Val Ser Cys Ile Ser Gly Pro Leu Gly Met Tyr Arg Asn Asn Ile Leu Gln Val Phe Leu Glu Ala Trp Tyr Arg Gln Lys Phe Leu Gly Thr Tyr 330 Cys Thr Leu Gly Asp Asp Arg His Leu Thr Asn Arg Val Leu Ser Met 340 345 350Gly Tyr Arg Thr Lys Tyr Thr His Lys Ser Arg Ala Phe Ser Glu Thr 355 360 365 Pro Ser Leu Tyr Leu Arg Trp Leu Asn Gln Gln Thr Arg Trp Thr Lys $_{\rm 370}$ $_{\rm 375}$ Ser Tyr Phe Arg Glu Trp Leu Tyr Asn Ala Gln Trp Trp His Lys His His Ile Trp Met Thr Tyr Glu Ser Val Val Ser Phe Ile Phe Pro Phe 410 Asn Val Val Trp Leu Leu Cys Ile Gln Ile Met Ser Leu Phe Lys 435 440 445Ser Leu Tyr Ser Met Leu Tyr Met Thr Gly Leu Leu Pro Ser Lys Tyr Phe Ala Leu Leu Thr Leu Asn Lys Thr Gly Trp Gly Thr Gly Arg Lys $485 \hspace{1cm} 490 \hspace{1cm} 495 \hspace{1cm}$ Ala Val Leu Cys Gly Gly Val Gly Tyr Ser Ile Tyr Met Asp Cys Gln 515 520 525 Asn Asp Trp Ser Thr Pro Glu Lys Gln Lys Glu Met Tyr His Leu Leu 530 540Tyr Gly Cys Val Gly Tyr Val Met Tyr Trp Val Ile Met Ala Val Met 545 550555 550 560 Tyr Trp Val Trp Val Lys Arg Cys Cys Arg Lys Arg Ser Gln Thr Val $565 \ \ 570 \ \ 575$ Thr Leu Val His Asp Ile Pro Asp Met Cys Val

- (2) INFORMATION FOR SEQ ID NO: 4:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 395 amino acids (B) TYPE: amino acid

 - (C) STRANDEDNESS: <Unknown>
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Tyr Leu Phe Gly Thr Ser Thr Val Gly Ile Tyr Gly Val Ile Leu 1 5 10 15

Ile Thr Tyr Leu Val Ile Lys Leu Gly Leu Ser Phe Leu Tyr Glu Pro $20 \hspace{1cm} 25 \hspace{1cm} 30 \hspace{1cm}$

Phe Lys Gly Asn Pro His Asp Tyr Lys Val Ala Ala Val Ile Pro Ser 35 40 45

Tyr	Asn 50	Glu	Asp	Ala	Glu	Ser 55	Leu	Leu	Glu	Thr	Leu 60	Lys	Ser	Val	Leu
Ala 65	Gln	Thr	Tyr	Pro	Leu 70	Ser	Glu	Ile	Tyr	Ile 75	Val	Asp	Asp	Gly	Ser 80
Ser	Asn	Thr	Asp	Ala 85	Ile	Gln	Leu	Ile	Glu 90	Glu	Tyr	Val	Asn	Arg 95	Glu
Val	Asp	Ile	C y s 100	Arg	Asn	Val	Ile	Val 105	His	Arg	Ser	Leu	Val 110	Asn	Lys
Gly	Lys	Arg 115	His	Ala	Gln	Ala	Trp 120	Ala	Phe	Glu	Arg	Ser 125	Asp	Ala	Asp
Val	Phe 130	Leu	Thr	Val	Asp	Ser 135		Thr	Tyr	Ile	Tyr 140	Pro	Asn	Ala	Leu
Glu 145	Glu	Leu	Leu	Lys	Ser 150	Phe	Asn	Asp	Glu	Thr 155	Val	Tyr	Ala	Ala	Thr 160
Gly	His	Leu	Asn	Ala 165	Arg	Asn	Arg	Gln	Thr 170	Asn	Leu	Leu	Thr	A rg 175	Leu
Thr	Asp	Ile	Arg 180	Tyr	Asp	Asn	Ala	Phe 185	Gly	Val	Glu	Arg	Ala 190	Ala	Gln
Ser	Leu	Thr 195	Gly	Asn	Ile	Leu	Val 200	Cys	Ser	Gly	Pro	Leu 205	Ser	Ile	Tyr
Arg	Arg 210	Glu	Val	Ile	Ile	Pro 215	Asn	Leu	Glu	Arg	Tyr 220	Lys	Asn	Gln	Thr
Phe 225	Leu	Gly	Leu	Pro	Val 230	Ser	Ile	Gly	Asp	Asp 235	Arg	Cys	Leu	Thr	Asn 240
Tyr	Ala	Ile	Asp	Leu 245	Gly	Arg	Thr	Val	Tyr 250	Gln	Ser	Thr	Ala	A rg 255	Cys
Asp	Thr	Asp	Val 260	Pro	Phe	Gln	Leu	L y s 265	Ser	Tyr	Leu	Lys	Gln 270	Gln	Asn
Arg	Trp	Asn 275	Lys	Ser	Phe	Phe	Arg 280	Glu	Ser	Ile	Ile	Ser 285	Val	Lys	Lys
Ile	Leu 290	Ser	Asn	Pro	Ile	Val 295	Ala	Leu	Trp	Thr	Ile 300	Phe	Glu	Val	Val
Met 305	Phe	Met	Met	Leu	Ile 310	Val	Ala	Ile	Gly	Asn 315	Leu	Leu	Phe	Asn	Gln 320
Ala	Ile	Gln	Leu	Asp 325	Leu	Ile	Lys	Leu	Phe 330	Ala	Phe	Leu	Ser	Ile 335	Ile
Phe	Ile		Ala 340		Сув		Asn			Tyr			Lys 350		Pro
Ala	Ser	Phe 355	Leu	Leu	Ser	Pro	Leu 360	Tyr	Gly	Ile	Leu	His 365	Leu	Phe	Val
Leu	Gln 370	Pro	Leu	Lys	Leu	Tyr 375		Leu	Cys	Thr	Ile 380	Lys	Asn	Thr	Glu
Trp 385	Gly	Thr	Arg	Lys	Lys 390	Val	Thr	Ile	Phe	L y s 395					

We claim:

- 1. An isolated or recombinant nucleic acid which encodes a mammalian hyaluronan synthase.
- 2. The nucleic acid of claim 1 wherein the hyaluronan synthase is human.
 - 3. The nucleic acid of claim 1 comprising SEQ ID NO: 1.
- **4**. The nucleic acid of claim 1 wherein said nucleic acid hybridizes under stringent conditions with a second nucleic acid having a nucleotide sequence of SEQ ID NO: 1.
- 5. The nucleic acid of claim 1 wherein the nucleic acid encodes the amino acid sequence of SEQ ID NO: 2.
- **6**. A recombinant nucleic acid construct comprising a nucleic acid of claim 1.

- 7. The recombinant nucleic acid construct of claim 6 comprising SEQ ID NO: 1.
- **8**. The recombinant nucleic acid construct of claim 6 wherein the nucleic acid encodes the amino acid sequence of SEQ ID NO: 2.
- **9**. The recombinant nucleic acid construct of claim 6 wherein the nucleic acid is operably linked to an expression control sequence.
 - 10. A host cell comprising the nucleic acid of claim 1.
- 11. The host cell of claim 10 wherein the nucleic acid is operably linked to an expression control sequence, whereby mammalian hyaluronan synthase is expressed when the host cell is maintained under conditions suitable for expression.
- 12. A method for producing a mammalian hyaluronan synthase comprising:
 - a) introducing into a host cell a nucleic acid construct comprising a nucleic acid which encodes a mammalian hyaluronan synthase, whereby a recombinant host cell is produced having said coding sequence operably linked to at least one expression control sequence; and

- b) maintaining the host cells produced in step a) under conditions whereby the nucleic acid is expressed.
- 13. An antibody or functional portion thereof which binds mammalian hyaluronan synthase.
- **14.** A method of detecting mammalian hyaluronan synthase in a sample comprising:
 - a) contacting a sample with an antibody which binds hyaluronan synthase under conditions suitable for specific binding of said antibody to the mammalian hyaluronan synthase; and
 - b) detecting an antibody-mammalian hyaluronan synthase complex.
- 15. A method of producing hyaluronan comprising maintaining a host cell of claim 10 under conditions whereby hyaluronan is produced.
- 16. The method of claim 15, comprising isolating hyaluronan thereby produced.

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