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[Continued on next page]

(54) Title: CYCLOBUTYL SULFONES AS NOTCH SPARING GAMMA SECRETASE INHIBITORS

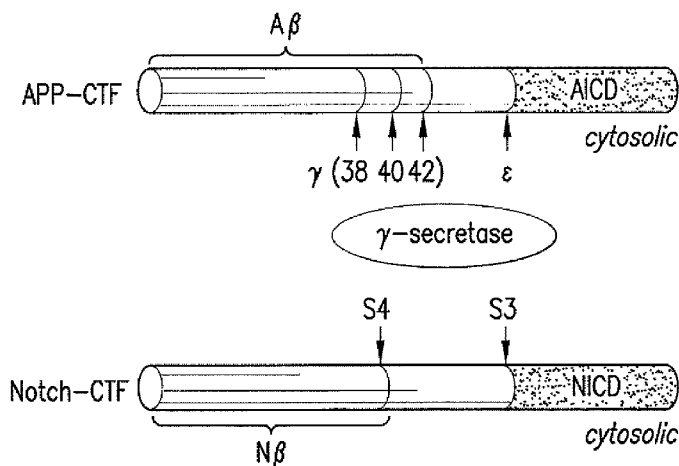


FIG. 1

(57) Abstract: The invention encompasses a novel class of cyclobutyl sulfone derivatives which inhibit the processing of APP by the putative γ -secretase while sparing Notch signaling pathway, and thus are useful in the treatment or prevention of Alzheimer's disease without the development of Notch inhibition mediated gastrointestinal issues. Pharmaceutical compositions and methods of use are also included.

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TITLE OF THE INVENTION

CYCLOBUTYL SULFONES AS NOTCH SPARING GAMMA SECRETASE INHIBITORS.

BACKGROUND OF THE INVENTION

5 The present invention relates to a novel class of compounds, their salts, pharmaceutical compositions comprising them and their use in therapy of the human body. In particular, the invention relates to novel cyclobutyl sulfone derivatives which inhibit the processing of APP by γ -secretase, and hence are useful in the treatment or prevention of Alzheimer's disease. The compounds of the invention also spare the Notch signaling pathway.

10 As such, the compounds of the invention are believed to halt or potentially reverse the progression of Alzheimer's disease without the development of toxicities mediated by Notch inhibition.

 Alzheimer's disease (AD) is the most prevalent form of dementia. Although primarily a disease of the elderly, affecting up to 10% of the population over the age of 65, AD

15 also affects significant numbers of younger patients with a genetic predisposition. It is a neurodegenerative disorder, clinically characterized by a progressive loss of memory and cognitive function, and pathologically characterized by the deposition of extracellular proteinaceous plaques in the cortical and associative brain regions of sufferers. These plaques are mainly comprised of fibrillar aggregates of β -amyloid peptide ($A\beta$) (Glennner G G and Wong

20 C W (1984) Alzheimer's disease: Initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochemical and Biophysical research Communications*. 120(3);885-890). The role of secretases, including that of γ -secretase, in the processing of amyloid precursor protein (APP) to form $A\beta$ is well documented in the literature. $A\beta$ is generated by proteolytic processing of APP by two enzymes, β -amyloid cleavage enzyme

25 (BACE) and γ -secretase (Figure 1; Selkoe D J (2001) Alzheimer's disease: genes, proteins, and therapy. *Physiological Review*. 81(2):741-766). γ -Secretase is a complex comprised of four proteins: presenilin (presenilin-1 or -2), nicastrin, APH-1 and PEN-2 (Takasugi N, Tomita T, Hayashi I, Tsuruoka M, Niimura M, Takahashi Y, Thinakaran G, Iwatsubo T (2003) The role of presenilin cofactors in the gamma-secretase complex. *Nature*. 422(6930):438-441; Kimberly W

30 T, LaVoie M J, Ostaszewski B L, Ye W, Wolfe M S, Selkoe D J (2003) Gamma-secretase is a membrane protein complex comprised of presenilin, nicastrin, Aph-1, and Pen-2. *Proceedings of the National Academy of Sciences*. 100(11):6382-6387; Edbauer D, Winkler E, Regula J T,

Pesold B, Steiner H, Haass C (2003) Reconstitution of gamma-secretase activity. *Nature Cell Biology*. 5(5):486-488. Presenilin-1 and -2 contain transmembrane aspartyl residues that have been shown to be essential for the catalytic activity of the complex (Wolfe M S, Xia W, Ostaszewski B L, Diehl T S, Kimberly W T, Selkoe D J (1999) Two transmembrane aspartates in presenilin-1 required for presenilin endoproteolysis and gamma-secretase activity. *Nature*. 1999 398(6727):513-517). The majority of the mutations linked to the early onset, familial form of AD (FAD) are associated with either PS-1 or PS-2 (Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird T D, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad E, Viitanen M, Peskind E, Poorkaj P, Schellenberg G, Tanzi R, Wasco W, Lannfelt L, Selkoe D, Younkin S (1996) Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nature Medicine*. 2(8):864-870; Duff K, Eckman C, Zehr C, Yu X, Prada C M, Perez-tur J, Hutton M, Buee L, Harigaya Y, Yager D, Morgan D, Gordon M N, Holcomb L, Refolo L, Zenk B, Hardy J, Younkin S (1996) Increased amyloid-beta42(43) in brains of mice expressing mutant presenilin 1. *Nature*. 383(6602):710-713; Lemere CA, Lopera F, Kosik K S, Lendon C L, Ossa J, Saido T C, Yamaguchi H, Ruiz A, Martinez A, Madrigal L, Hincapie L, Arango J C, Anthony D C, Koo E H, Goate A M, Selkoe D J, Arango J C (1996) The E280A presenilin 1 Alzheimer mutation produces increased A beta 42 deposition and severe cerebellar pathology. *Nature Medicine*. 2(10):1146-1150; Citron M, Westaway D, Xia W, Carlson G, Diehl T, Levesque G, Johnson-Wood K, Lee M, Seubert P, Davis A, Kholodenko D, Motter R, Sherrington R, Perry B, Yao H, Strome R, Lieberburg I, Rommens J, Kim S, Schenk D, Fraser P, St George Hyslop P, Selkoe D J (1997) Mutant presenilins of Alzheimer's disease increase production of 42-residue amyloid beta-protein in both transfected cells and transgenic mice. *Nature Medicine*. 3(1):67-72). γ -Secretase processes a number of other type I membrane proteins that have undergone a prerequisite ectodomain shedding (Lleó A (2008) Activity of gamma-secretase on substrates other than APP. *Current Topics in Medicinal Chemistry*. 8(1):9-16).

In addition to processing APP, γ -secretase cleaves the Notch family of receptors. Genetic evidence indicates that γ -secretase activity is critically required for Notch signaling and functions (Shen J, Bronson R T, Chen D F, Xia W, Selkoe D J, Tonegawa S (1997) Skeletal and CNS defects in Presenilin-1-deficient mice. *Cell*. 89(4):629-639; Wong P C, Zheng H, Chen H, Becher M W, Sirinathsinghji D J, Trumbauer M E, Chen H Y, Price D L, Van der Ploeg L H, Sisodia S S (1997) Presenilin 1 is required for Notch1 and Dll1 expression in the paraxial mesoderm. *Nature*. 387(6630):288-292). Notch is an evolutionarily conserved and widely

expressed single-span type I transmembrane receptor that plays a prominent role in regulating cell fate decisions in the developing embryo (Artavanis-Tsakonas S, Rand M D, Lake R J (1999) Notch signaling: cell fate control and signal integration in development. *Science*. 284(5415):770-776). The role of Notch in the adult remains unclear but Notch proteins are expressed in various adult tissues and are thought to play a role in regulating stem cell differentiation. Four Notch genes have been identified in mammals (Notch 1-4); all four Notch proteins are cleaved by γ -secretase (Saxena M T, Schroeter E H, Mumm J S, Kopan R (2001) Murine notch homologs (N1-4) undergo presenilin-dependent proteolysis. *Journal of Biological Chemistry*. 276(43):40268-40273). Notch activation is induced by binding, in trans, to the Delta/Serrate/LAG2 family of transmembrane ligands. Notch signal transduction is mediated by three cleavage events: (a) cleavage at Site 1 in extracellular domain; (b) cleavage at Site 2 just N-terminal to the extracellular/transmembrane domain boundary following ligand binding; and (c) cleavage at Site 3 (S3) within the transmembrane near the transmembrane/cytoplasmic domain boundary. Site 3 cleavage is required for release of Notch intracellular domain (NICD) and is mediated by γ -secretase (Schroeter E H, Kisslinger J A, Kopan R (1998) Notch-1 signalling requires ligand-induced proteolytic release of intracellular domain. *Nature*. 393(6683):382-386). NICD activates transcription mediated by the (CSL) CBF1/Serrate/LAG-1 family of DNA binding proteins and induces expression of various genes. NICD-regulated transcription is thought to be a key component of Notch-mediated signal transduction.

The development of γ -secretase inhibitors to block APP cleavage and A β generation is hampered by the potential for mechanism-based toxicity due to inhibition of Notch processing. Notch-related toxicities have been observed in studies where animals have been dosed subchronically with γ -secretase inhibitors. Intestinal goblet cell metaplasia is consistently observed following three or more days of treatment (Searfoss G H, Jordan W H, Calligaro D O, Galbreath E J, Schirtzinger L M, Berridge B R, Gao H, Higgins M A, May P C, Ryan T P (2003) Adipsin, a biomarker of gastrointestinal toxicity mediated by a functional gamma-secretase inhibitor. *Journal of Biological Chemistry*. 278(46):46107-46116; Wong G T, Manfra D, Poulet F M, Zhang Q, Josien H, Bara T, Engstrom L, Pinzon-Ortiz M, Fine J S, Lee H J, Zhang L, Higgins G A, Parker E M (2004) Chronic treatment with the gamma-secretase inhibitor LY-411,575 inhibits beta-amyloid peptide production and alters lymphopoiesis and intestinal cell differentiation. *Journal of Biological Chemistry*. 279(13):12876-12882; Milano J, McKay J, Dagenais C, Foster-Brown L, Pognan F, Gadiant R, Jacobs R T, Zacco A, Greenberg B, Ciaccio P J (2004) Modulation of notch processing by gamma-secretase inhibitors causes intestinal

goblet cell metaplasia and induction of genes known to specify gut secretory lineage differentiation. *Toxicological Sciences*. 82(1):341-358; van Es J H, van Gijn M E, Riccio O, van den Born M, Vooijs M, Begthel H, Cozijnsen M, Robine S, Winton D J, Radtke F, Clevers H (2005) Notch/gamma-secretase inhibition turns proliferative cells in intestinal crypts and adenomas into goblet cells. *Nature*. 435(7044):959-963). In addition, Notch function appears to be critical for the proper differentiation of T and B lymphocytes (Hadland B K, Manley N R, Su D, Longmore G D, Moore C L, Wolfe M S, Schroeter E H, Kopan R (2001) Gamma -secretase inhibitors repress thymocyte development. *Proceedings of the National Academy of Sciences*. 98(13):7487-7491; Doerfler P, Shearman M S, Perlmutter R M (2001) Presenilin-dependent gamma-secretase activity modulates thymocyte development. *Proceedings of the National Academy of Sciences*. 98(16):9312-9317). Thus, pharmacologically targeting γ -secretase activity requires agents that selectively block A β while minimally inhibiting activity towards Notch.

The present invention provides a novel class of cyclobutyl sulfone derivatives which inhibit the processing of APP by the putative γ -secretase while sparing Notch signaling pathway, and thus are useful in the treatment or prevention of AD.

SUMMARY OF THE INVENTION

The invention encompasses a novel class of cyclobutyl sulfone derivatives which inhibit the processing of APP by the putative γ -secretase while sparing Notch signaling pathway, and thus are useful in the treatment or prevention of Alzheimer's disease without the development of Notch inhibition mediated gastrointestinal issues. Pharmaceutical compositions and methods of use are also included.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. γ -Secretase cleaves APP and Notch at two main positions approximately in the middle of the membrane (γ /S4-cleavage) and at the cytosolic face of the membrane (ϵ /S3-cleavage). The ϵ /S3-cleavage is a critical processing event since it liberates the intracellular domain (ICD) of the substrate from the membrane: AICD and NICD, respectively. This step is prerequisite for ICD translocation to the nucleus and its subsequent function as transcriptional modulator. On the other hand, the γ /S4-cleavage leads to the release of A β peptides and A β -like peptides (N β) from APP and Notch, respectively. The latter cleavage has mainly generated attention since it produces the C terminus of the A β peptide, which is believed to be the disease-causing agent for AD.

Figure 2. Results of MRK-560 and Example 2 tested in the transactivation assay described in this application to examine the effects on the initial cleavage of other γ -secretase substrates. HEK cells were transiently co-transfected with the chimeric substrate along with a UAS promoter driven luciferase and β -galactosidase, and then treated with 1 μ M of each compound for 48 hours. MRK-560 inhibited ICD release of all examined substrates, whereas Example 2 retained ICD release and subsequent translocation of the ICD-GVP construct to allow for reporter activation.

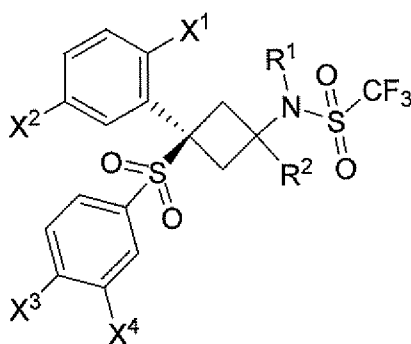
Figure 3. HEK293 cells stably overexpressing each substrate, Notch Δ E, E-cadherin, SCN2b and CD43, were treated with MRK-560 or Example 2 at titrated concentrations along with TPA to induce the shedding. Immunoblot analysis of cell lysates revealed that MRK-560 inhibited NICD generation and caused accumulation of CTFs, direct substrates of γ -secretase, in a similar manner to a traditional γ -secretase inhibitor, DAPT. In contrast, Example 2 allowed for NICD generation and no or less CTF accumulation was observed compared MRK-560.

Figure 4. The results of MRK-560 and Example 2 tested in the compound binding assay described in this application. Tritiated GSI tracers L-458 (transition state, red) or L-881 (non transition state, blue) were incubated with semi-purified γ -secretase complex and increasing concentrations of the respective compounds. A-MRK-560 showed full and partial displacement of at L-881 and L-458 sites, respectively. B-Example 2 was able to fully displace L-881 but not L-458. The results demonstrate that notch sparing compounds such as Example 2 have a shifted binding site as compared to traditional inhibitors such as MRK-560. This

deregulates enzymatic cleavage in a manner that spares ϵ /S3 (AICD/NICD release) while potently inhibiting all γ -cleavage sites.

DETAILED DESCRIPTION OF THE INVENTION

5 The invention encompasses a genus of compounds according to Formula I

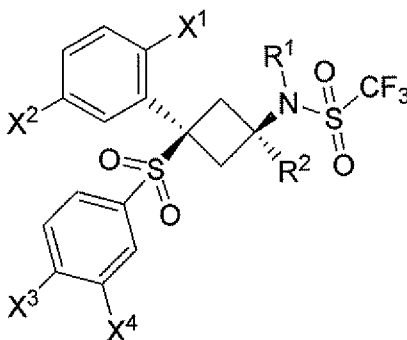


I

or pharmaceutically acceptable salt thereof, wherein:

- 10 X¹ is selected from the group consisting of: F and CN;
 X² is selected from the group consisting of: F, Cl and CN;
 X³ is selected from the group consisting of: F, Br, Cl, CN, CF₃, OCF₃, C(O)-OCH₃ and S-CH₃;
 X⁴ is selected from the group consisting of: H, F and Cl;
 R¹ is selected from the group consisting of:
- 15 (a) H,
 (b) CH₃,
 (c) -(CH₂)_n-OR³;
 (d) -(CH₂)_n-C(O)-OR⁴ and
 (e) -SO₂-CF₃;
- 20 R² is H or CH₃ when the compound of formula I is in the *cis* configuration, otherwise R² is H;
 R³ is a five- or six-membered non-aromatic heterocycle having one oxygen heteroatom;
 R⁴ is H or CH₃; and
 n is 1 to 4.

Within the genus, the invention encompasses a first sub-genus of compounds of Formula Ia



Ia

5

or a pharmaceutically acceptable salt thereof.

Within the first subgenus, the invention encompasses a first class of compounds of formula Ia wherein:

X¹ is F and

10 X⁴ is H.

Within the first class, the invention encompasses a sub-class of compounds of formula Ia wherein:

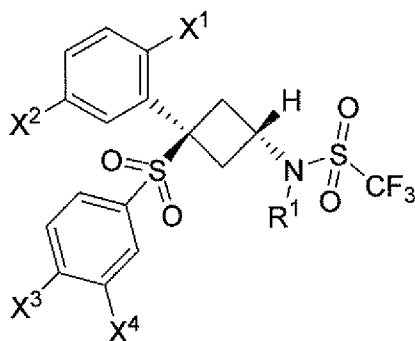
X² is F and

X³ is Cl.

15 Within the sub-class, the invention encompasses a group of compounds of formula Ia wherein R² is H.

Also within the group, the invention encompasses a second sub-group of compounds of formula Ia wherein R¹ is $-(CH_2)_n-C(O)-OR^4$.

20 Within the genus, the invention encompasses a second sub-genus of compounds of Formula Ib



Ib

or a pharmaceutically acceptable salt thereof.

5 Within the second sub-genus, the invention encompasses a second class of compounds of formula Ib wherein:

X¹ and X² are F;

X³ is Cl; and

X⁴ is H.

The invention also encompasses any of the examples that follow.

10 The invention also encompasses a pharmaceutical composition comprising a compound according to formula I or a pharmaceutically acceptable salt thereof and a pharmaceutically acceptable carrier.

15 The invention also encompasses a method of treatment of a subject suffering or prone to a condition associated with the deposition of β -amyloid which comprises administering to that subject an effective amount of a compound according to formula I or a pharmaceutically acceptable salt thereof.

The invention also encompasses the use of a compound according to formula I or a pharmaceutically acceptable salt thereof for the manufacture of a medicament for treating or preventing Alzheimer's disease.

20 Where a variable occurs more than once in formula I or in a substituent thereof, the individual occurrences of that variable are independent of each other, unless otherwise specified.

For use in medicine, the compounds of formula I may be in the form of pharmaceutically acceptable salts. Other salts may, however, be useful in the preparation of the compounds of formula I or of their pharmaceutically acceptable salts. Suitable pharmaceutically acceptable salts of the compounds of this invention include acid addition salts which may, for example, be formed by mixing a solution of the compound according to the invention with a

solution of a pharmaceutically acceptable acid such as hydrochloric acid, sulphuric acid, methanesulphonic acid, benzenesulphonic acid, fumaric acid, maleic acid, succinic acid, acetic acid, benzoic acid, oxalic acid, citric acid, tartaric acid, carbonic acid or phosphoric acid.

Alternatively, where the compound of the invention carries an acidic moiety, a pharmaceutically acceptable salt may be formed by neutralisation of said acidic moiety with a suitable base. Examples of pharmaceutically acceptable salts thus formed include alkali metal salts such as sodium or potassium salts; ammonium salts; alkaline earth metal salts such as calcium or magnesium salts; and salts formed with suitable organic bases, such as amine salts (including pyridinium salts) and quaternary ammonium salts.

It is to be emphasized that the invention, for each compound in accordance with formula I, encompasses both enantiomeric forms, either as homochiral compounds or as mixtures of enantiomers in any proportion. In an embodiment of the invention, the compound of formula I is a homochiral compound of formula Ia or formula Ib, or a pharmaceutically acceptable salt thereof. Where the compounds according to the invention possess two or more asymmetric centres, they may additionally exist as diastereoisomers. It is to be understood that all such isomers and mixtures thereof in any proportion are encompassed within the scope of the present invention.

It will also be appreciated that where more than one isomer can be obtained from a reaction then the resulting mixture of isomers can be separated by conventional means. Where the processes for the preparation of the compounds according to the invention gives rise to mixtures of stereoisomers, these isomers may be separated by conventional techniques such as preparative chromatography. The novel compounds may be prepared in racemic form, or individual enantiomers may be prepared either by enantiospecific synthesis or by resolution. The novel compounds may, for example, be resolved into their component enantiomers by standard techniques such as preparative HPLC, or the formation of diastereomeric pairs by salt formation with an optically active acid, such as (-)-di-p-toluoyl-d-tartaric acid and/or (+)-di-p-toluoyl-l-tartaric acid, followed by fractional crystallization and regeneration of the free base. The novel compounds may also be resolved by formation of diastereomeric esters or amides, followed by chromatographic separation and removal of the chiral auxiliary. Alternatively, such techniques may be carried out on racemic synthetic precursors of the compounds of interest.

The compounds of the present invention have an activity as inhibitors of γ secretase.

The invention also provides pharmaceutical compositions comprising one or more compounds of this invention and a pharmaceutically acceptable carrier. Preferably these compositions are in unit dosage forms such as tablets, pills, capsules, powders, granules, sterile parenteral solutions or suspensions, metered aerosol or liquid sprays, drops, ampoules, 5 transdermal patches, auto-injector devices or suppositories; for oral, parenteral, intranasal, sublingual or rectal administration, or for administration by inhalation or insufflation. The principal active ingredient typically is mixed with a pharmaceutical carrier, e.g. conventional tableting ingredients such as corn starch, lactose, sucrose, sorbitol, talc, stearic acid, magnesium stearate and dicalcium phosphate, or gums, dispersing agents, suspending agents or surfactants 10 such as sorbitan monooleate and polyethylene glycol, and other pharmaceutical diluents, e.g. water, to form a homogeneous preformulation composition containing a compound of the present invention, or a pharmaceutically acceptable salt thereof. When referring to these preformulation compositions as homogeneous, it is meant that the active ingredient is dispersed evenly throughout the composition so that the composition may be readily subdivided into equally 15 effective unit dosage forms such as tablets, pills and capsules. This preformulation composition is then subdivided into unit dosage forms of the type described above containing from 0.1 to about 500 mg of the active ingredient of the present invention. Typical unit dosage forms contain from 1 to 100 mg, for example 1, 2, 5, 10, 25, 50 or 100 mg, of the active ingredient. Tablets or pills of the novel composition can be coated or otherwise compounded to provide a 20 dosage form affording the advantage of prolonged action. For example, the tablet or pill can comprise an inner dosage and an outer dosage component, the latter being in the form of an envelope over the former. The two components can be separated by an enteric layer which serves to resist disintegration in the stomach and permits the inner component to pass intact into the duodenum or to be delayed in release. A variety of materials can be used for such enteric 25 layers or coatings, such materials including a number of polymeric acids and mixtures of polymeric acids with such materials as shellac, cetyl alcohol and cellulose acetate.

The liquid forms in which the novel compositions of the present invention may be incorporated for administration orally or by injection include aqueous solutions, liquid- or gel-filled capsules, suitably flavoured syrups, aqueous or oil suspensions, and flavoured emulsions 30 with edible oils such as cottonseed oil, sesame oil or coconut oil, as well as elixirs and similar pharmaceutical vehicles. Suitable dispersing or suspending agents for aqueous suspensions include synthetic and natural gums such as tragacanth, acacia, alginate, dextran, sodium

carboxymethylcellulose, methylcellulose, poly(ethylene glycol), poly(vinylpyrrolidone) or gelatin.

The present invention also provides a compound of the invention or a pharmaceutically acceptable salt thereof for use in a method of treatment of the human body.

5 Preferably the treatment is for a condition associated with the deposition of β -amyloid.

Preferably the condition is a neurological disease having associated β -amyloid deposition such as Alzheimer's disease.

The present invention further provides the use of a compound of the present invention or a pharmaceutically acceptable salt thereof in the manufacture of a medicament for
10 treating or preventing Alzheimer's disease.

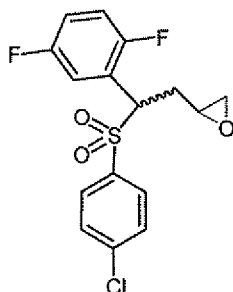
Also disclosed is a method of treatment of a subject suffering from or prone to Alzheimer's disease which comprises administering to that subject an effective amount of a compound according to the present invention or a pharmaceutically acceptable salt thereof.

For treating or preventing Alzheimer's disease, a suitable dosage level is about
15 0.01 to 250 mg/kg per day, preferably about 0.01 to 100 mg/kg per day, more preferably about 0.05 to 50 mg/kg of body weight per day, and for the most preferred compounds, about 0.1 to 10 mg/kg of body weight per day. The compounds may be administered on a regimen of 1 to 4 times per day. In some cases, however, a dosage outside these limits may be used.

The following examples illustrate the present invention. Where they are not
20 commercially available, the starting materials and reagents used in the synthetic schemes may be prepared by conventional means. The invention also encompasses a compound selected from the examples that follow.

During any of the synthetic sequences it may be necessary and/or desirable to protect sensitive or reactive groups on any of the molecules concerned. This may be achieved by
25 means of conventional protecting groups, such as those described in Protective Groups in Organic Chemistry, ed. J.F.W. McOmie, Plenum Press, 1973; and T.W. Greene & P.G.M. Wuts, Protective Groups in Organic Synthesis, John Wiley & Sons, 1991. The protecting groups may be removed at a convenient subsequent stage using methods known from the art.

EXAMPLES

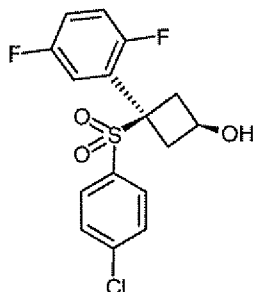
Intermediate A**2-[2-[(4-chlorophenyl)sulfonyl]-2-(2,5-difluorophenyl)ethyl]oxirane**

5 4-chlorophenyl-2,5-difluorobenzylsulfone was prepared as described in WO 02/081435 (Intermediate 1) from 4-chlorothiophenol and 2,5-difluorobenzyl bromide in two steps.

4-chlorophenyl-2,5-difluorobenzylsulfone (12 g, 39.6 mmol) in THF (99 ml) was treated with ⁿBuLi (19 ml, 2.5 M in hexane, 47.6 mmol) at 0°C for 10 min followed by addition
 10 of epichlorohydrin (3.73 ml, 47.6 mmol). The reaction was slowly warmed to room temperature for 14 h, quenched with water (100 ml) and diluted with EtOAc (300 ml). The organic phase was separated, dried (Na₂SO₄) and evaporated to dryness to give an oil. This material was chromatographed on silica, eluting with 10-45% ethyl acetate in hexanes to afford 9.8 g of the desired product as off-white solid. ¹H NMR (600 MHz, CDCl₃) two diastereomers (~1/1) δ 7.52 (m, 2H/2H), 7.38 (m, 2H/2H), 7.30-7.22 (m, 1H/1H), 6.99 (m, 1H/1H), 6.86-6.80 (m, 1H/1H),
 15 4.76-4.70 (m, 1H/1H), 3.03-2.48 (m, 4H/4H), 2.37 (m, 1H), 2.20 (m, 1H). MS calculated 359.0 (MH⁺), exp 358.9 (MH⁺).

Intermediate B

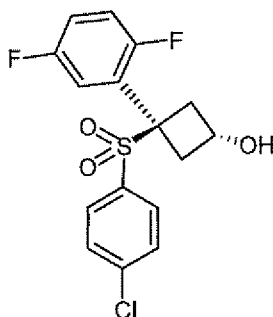
20



***cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol**

To 2-[2-[(4-chlorophenyl)sulfonyl]-2-(2,5-difluorophenyl)ethyl]oxirane (100 mg, 0.279 mmol) in THF (2.7 ml) was added MeMgBr (279 μ L, 3M in ether, 0.836 mmol) at -78°C . The reaction was warmed to room temperature over 1h then quenched with sat. NH_4Cl (3 ml) and diluted with EtOAc (10 ml). The organic phase was washed with brine (10 ml), dried (Na_2SO_4) and evaporated to dryness to afford the desired product (100 mg). ^1H NMR (600 MHz, CDCl_3) δ 7.66 (d, $J = 8.4$ Hz, 2H), 7.57 (d, $J = 8.4$ Hz, 2H), 6.99 (m, 1H), 6.82 (m, 1H), 6.74 (m, 1H), 4.28 (m, 1H), 3.47 (d, $J = 11.4$ Hz, 1H, OH), 3.13 (m, 4H). MS calculated 422.0 ($\text{MNa}^+ + \text{CH}_3\text{CN}$), exp 421.9 ($\text{MNa}^+ + \text{CH}_3\text{CN}$).

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Intermediate C***trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol**

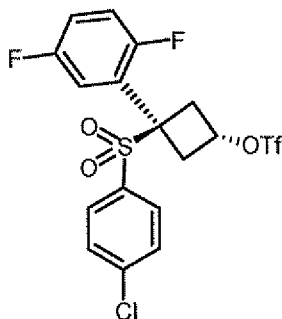
To PPh_3 (8.19 g, 31.2 mmol) in THF (100 ml) was added DIAD (6.31g, 31.2 mmol); the resulting mixture was stirred at room temperature for 0.5 h. The mixture was cooled to -50°C and *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol (8g, 22.3 mmol) in THF (10 ml) was added. The reaction was stirred for 20 min followed by addition of solid 4-nitrobenzoic acid (5.22 g, 31.2 mmol). The resulting mixture was warmed to room temperature and allowed to stir at room temperature for 20 h. The reaction was then cooled to 0°C to which was added NaOMe (134 ml, 0.5 M in MeOH, 66.9 mmol). After 40 min the reaction was quenched with Sat. NH_4Cl (100 ml) and diluted with EtOAc (300 ml). The organic phase was separated, dried (Na_2SO_4) and evaporated to dryness to afford an oil. This material was chromatographed on silica, eluting with ether in hexanes to give 8 g of the title product as white solid containing $\sim 15\%$ of *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol (starting material). ^1H NMR (600 MHz, CDCl_3) major product, δ

25

7.35 (m, 4H), 6.96 (m, 1H), 6.80-6.72 (m, 2H), 4.84 (m, 1H), 3.49 (m, 2H), 2.59 (m, 2H). MS calculated 422.0 (MNa⁺+CH₃CN), exp 421.9 (MNa⁺+CH₃CN).

Intermediate D

5

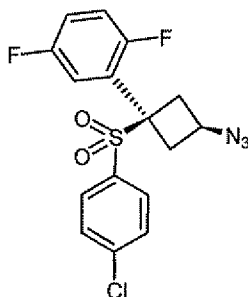


***trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl trifluoromethanesulfonate**

To *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol (2 g, 5 mmol) in DCM (27.9 ml) was added trifluoromethanesulfonic anhydride (1.13 ml, 6.69 mmol) and pyridine (0.902 ml, 11.15 mmol) at 0°C. The reaction was stirred for 30 min, quenched with Sat. NH₄Cl (30 ml) and diluted with EtOAc (150 ml). The organic phase was separated, dried (Na₂SO₄) and evaporated to dryness to give an oil. This material was chromatographed on silica, eluting with 0-25% ethyl acetate in hexanes to afford 2.7 g of the desired product as white solid, 10
15
¹H NMR (600 MHz, CDCl₃) δ 7.36 (m, 4H), 7.02 (m, 1H), 6.79 (m, 2H), 5.72 (m, 1H), 3.66 (broad s, 2H), 3.02 (dd, *J*=14.4, 6.6 Hz, 2H). MS calculated 554.0 (MNa⁺+CH₃CN), exp 553.8 (MNa⁺+CH₃CN).

Intermediate E

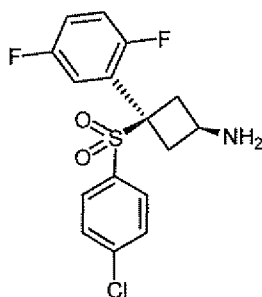
20



***cis*-3-azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone**

A mixture of *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl trifluoromethanesulfonate (3.9 g, 7.95 mmol) and sodium azide (5.17g, 79 mmol) in ethanol (39.7 ml) and water (39.7 ml) was heated at 85°C for 2h. The mixture was cooled to room temperature and diluted with water (100 ml) and EtOAc (150 ml). The organic phase was separated, dried (Na₂SO₄) and evaporated to dryness to afford an oil. This material was chromatographed on silica, eluting with 0-30% ethyl acetate in hexanes to afford the desired 2.3 g product as white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.35 (m, 4H), 7.02 (m, 1H), 6.91 (m, 1H), 6.80 (m, 1H), 3.81 (m, 1H), 3.30 (m, 2H), 3.02 (m, 2H).

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Intermediate F***cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine**

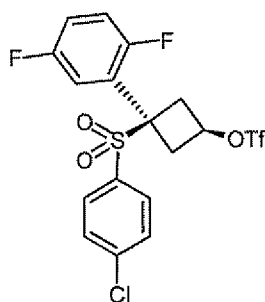
cis-3-Azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone (2.07 g, 5.39 mmol) and palladium (0.861 g, 10 % on carbon, 0.809 mmol) in MeOH (27 ml) was stirred under H₂ balloon for 4h. The crude mixture was filtered through a silica gel pack and washed with 5:1 DCM/MeOH (100 ml) to remove palladium residue. The solvent was removed to give the product which was used directly in next transformation without further purification. ¹H NMR (600 MHz, CDCl₃) δ 7.34 (m, 4H), 6.97 (m, 1H), 6.84 (m, 1H), 6.77 (m, 1H), 3.45 (m, 1H), 3.05-2.94 (m, 4H). MS calculated 358.0 (MH⁺), exp 358.0 (MH⁺).

Method (b)

To a solution of *cis*-3-azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone (9.5g, 24.75 mmol) in ethanol/THF stirred at RT was added zinc (3.24 g, 49.5 mmol), followed by ammonium formate (3.12 g, 49.5 mmol). Reaction was stirred at room temperature

for 1 h. The mixture was filtered through Celite and the solvents were removed. To the resulting residue was added 100 ml of sat'd NaHCO₃ solution and the products were extracted with EtOAc (2 X 100 ml). Combined organics were washed with brine and dried over anhydrous sodium sulfate, and filtered through Celite. The filtrate was concentrated to afford the desired product

Intermediate G



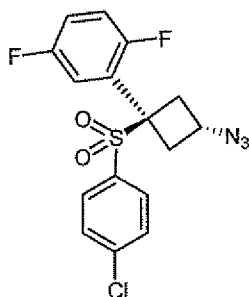
10 *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl trifluoromethanesulfonate

Prepared as for *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl trifluoromethanesulfonate, using *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanol:

15 ¹H NMR (600 MHz, CDCl₃) δ 7.35 (m, 4H), 7.04 (m, 1H), 6.88 (m, 1H), 6.81 (m, 1H), 5.08 (m, 1H), 3.60 (m, 2H), 3.23 (m, 2H). MS calculated 554.0 (MNa⁺+CH₃CN), exp 553.8 (MNa⁺+CH₃CN).

Intermediate H

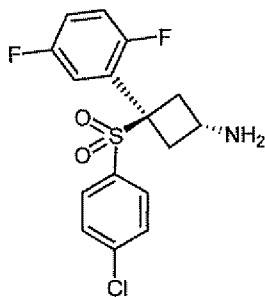
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***trans*-3-azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone**

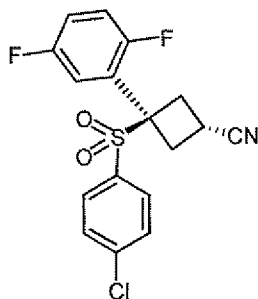
Prepared as for *cis*-3-azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone, using *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl trifluoromethanesulfonate.

- 5 ^1H NMR (600 MHz, CDCl_3) δ 7.35 (m, 4H), 6.98 (m, 1H), 6.76 (m, 2H), 4.56 (m, 1H), 3.49 (m, 2H), 2.66 (m, 2H).

Intermediate I***trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine**

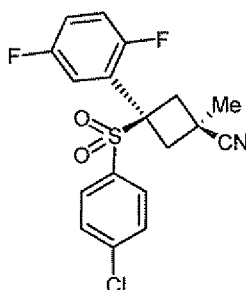
Prepared as for *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine, using *trans*-3-azido-1-(2,5-difluorophenyl)cyclobutyl 4-chlorophenyl sulfone.

- 15 ^1H NMR (600 MHz, CD_3OD) δ 7.48-7.41 (m, 4H), 7.07 (m, 1H), 6.88 (m, 1H), 6.79 (m, 1H), 3.88 (m, 1H), 3.41 (m, 2H), 2.50 (m, 2H). MS calculated 358.0 (MH^+), exp 358.0 (MH^+).

Intermediate J

trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanecarbonitrilecis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl

trifluoromethanesulfonate (4 g, 8.15 mmol) and tetrabutylammonium cyanide (5.47 g, 20.37 mmol) in DMSO (54 ml) was heated at 80°C for 45 min. The resulting mixture was cooled to room temperature and diluted with water (200 ml) and EtOAc (250 ml). The organic phase was washed with water, brine, separated, dried (Na₂SO₄) and evaporated to dryness, to afford an oil. This material was chromatographed on silica, eluting with 0-50% ethyl acetate in hexanes to give the desired product (2.9 g) as off-white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.38-7.33 (m, 4H), 7.02 (m, 1H), 6.80-6.74 (m, 2H), 3.74 (m, 1H), 3.51 (m, 2H), 3.02 (m, 2H). MS calculated 431.0 (MNa⁺+CH₃CN), exp 431.0 (MNa⁺+CH₃CN).

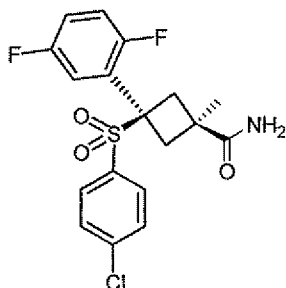
Intermediate K

15

***cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanecarbonitrile**

To *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanecarbonitrile (600 mg, 1.63 mmol) in THF (8 mL) was added LiHMDS (2.45 ml, 1 M in THF, 2.45 mmol) at -78°C. After 10 min MeI (306 μl, 4.89 mmol) was introduced to reaction mixture. The reaction was stirred for 2h with the temperature slowly increasing to 0°C. The reaction was then quenched with water and extracted with EtOAc. The organic phase was separated, dried (Na₂SO₄) and evaporated to dryness to afford an oil. This material was chromatographed on silica, eluting with 0-50% ethyl acetate in hexanes to give the product (260 mg) as a single diastereomeric products. ¹H NMR (600 MHz, CD₃OD) δ 7.36 (s, 4H), 7.02 (m, 1H), 6.85-6.78 (m, 2H), 3.79 (d, *J* = 14.4 Hz, 2H), 2.72 (d, *J* = 14.4 Hz, 2H), 1.44 (s, 3H). MS calculated 785.1 (2M+Na⁺), exp 785.0 (2M+Na⁺).

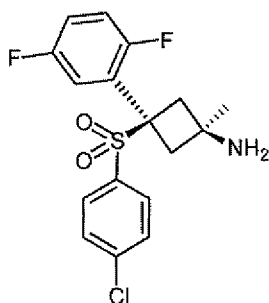
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Intermediate L***cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanecarboxamide**

5 To *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanecarbonitrile (500 mg, 1.31 mmol) and K₂CO₃ (362 mg, 2.62 mmol) in DMSO (6.5 ml) was added H₂O₂ (1.15 ml, 35 % in water, 13.1 mmol) dropwise and the reaction was stirred vigorously for 2h. The mixture was diluted with water (50 ml) and EtOAc (50 ml). The organic phase was washed with water, brine, separated, dried (Na₂SO₄) and evaporated to

10 dryness to give *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanecarboxamide (500 mg) as off-white solid which was used in the next reaction without further purification. ¹H NMR (600 MHz, CDCl₃) δ 7.35 (m, 4H), 7.02 (m, 1H), 6.87-6.79 (m, 2H), 6.54 (broad s, 1H), 6.40 (broad s, 1H), 3.67 (d, *J* = 14.4 Hz, 2H), 2.62 (d, *J* = 14.4 Hz, 2H), 1.28 (s, 3H). MS calculated 400.0 (MH⁺), exp 400.0 (MH⁺).

15

Intermediate M

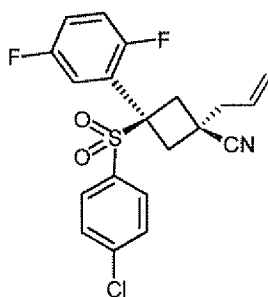
20 ***cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanamine**

cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-

methylcyclobutanecarboxamide (400 mg, 1 mmol) and PIFA (473 mg, 1.1 mmol) in acetonitrile (2.5 ml) and water (2.5 ml) was stirred at 0°C and the mixture was slowly warmed up to room temperature over 27 h. The reaction was then quenched with Sat. NaHCO₃ and extracted with EtOAc. The organic phase was separated, dried (Na₂SO₄) and evaporated to dryness to give an oil. This material was chromatographed on silica, eluting with 0-40% MeOH in DCM to give *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanamine (380 mg) as product. ¹H NMR (600 MHz, CD₃OD) δ 7.41-7.36 (m, 4H), 7.02 (m, 1H), 6.81 (m, 1H), 6.74 (m, 1H), 3.58 (d, *J* = 15.6 Hz, 2H), 2.89 (d, *J* = 15.6 Hz, 2H), 1.58 (s, 3H). MS calculated 372.1 (MH⁺), exp 372.0 (MH⁺).

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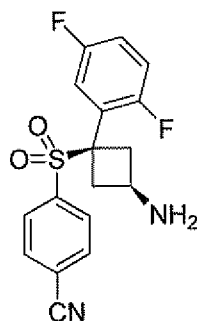
Intermediate N



***cis*-1-allyl-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanecarbonitrile**

15 Prepared as for *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanecarbonitrile, using *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanecarbonitrile and allyl bromide.

20 ¹H NMR (600 MHz, CDCl₃) δ 7.36 (s, 4H), 7.03 (m, 1H), 6.83-6.78 (m, 2H), 5.70 (m, 1H), 5.18 (dd, *J* = 10.2, 1.2 Hz, 1H), 5.07 (dd, 1H, *J* = 16.8, 1.2 Hz, 1H), 3.70 (d, *J* = 15.0 Hz, 2H), 2.77 (d, *J* = 15.0 Hz, 2H), 2.32 (d, *J* = 6.6 Hz, 2H). MS calculated 837.1 (2M+Na⁺), exp 837.0 (2M+Na⁺).

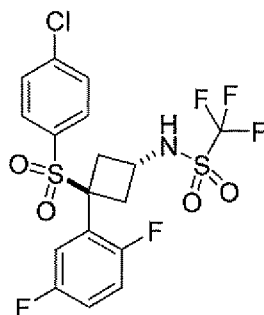
Intermediate O

5

4-[[*cis*-3-amino-1-(2,5-difluorophenyl)cyclobutyl]sulfonyl]benzonitrile

Zinc cyanide (0.295g, 2.52 mmol), Pd₂(dba)₃ (0.384g, 0.419mmol), Zinc (0.030g, 0.461 mmol), DPPF (0.465g, 0.838 mmol) and *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine (1.5g, 4.19 mmol) were taken up in DMA and stirred in a 25 mL Schlenk tube under an argon environment at 135 °C for 16 hours. Water was then added and the mixture was extracted with EtOAc. The organic layer was washed with a saturated NaHCO₃ solution and brine then dried over MgSO₄, filtered and concentrated. The residue was purified via silica column chromatography (0->8% MeOH/DCM) to give the title compound. MS: cal'd 349 (MH⁺), exp 349 (MH⁺)

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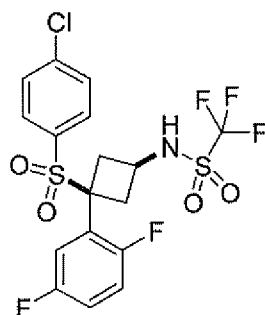
Example 1

20 ***N*-[*trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide**

To *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine (1.5 g, 4.19 mmol) in DCM (27.9 ml) was added triethylamine (1.169 ml, 8.38 mmol) and trifluoromethanesulfonic anhydride (0.850 ml, 5.03 mmol) at 0°C and the mixture was stirred for 2h.

5 The mixture diluted with water (50 ml) and EtOAc (100 ml). The organic phase was washed with brine, separated, dried (Na₂SO₄) and evaporated to dryness to afford an oil. This material was chromatographed on silica, eluting with ethyl acetate in hexanes to give the desired product (1.45 g as white solid). ¹H NMR (600 MHz, CDCl₃) δ 7.39 (d, *J* = 9.0 Hz, 2H), 7.34 (d, *J* = 9.0 Hz, 2H), 7.02 (m, 1H), 6.77 (m, 2H), 4.22 (m, 1H), 3.23 (m, 4H). MS calculated
10 553.0 (MNa⁺+CH₃CN), exp 553.0 (MNa⁺+CH₃CN).

Example 2

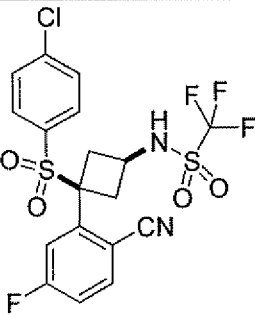
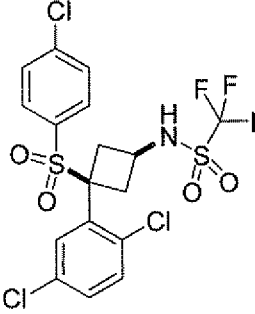
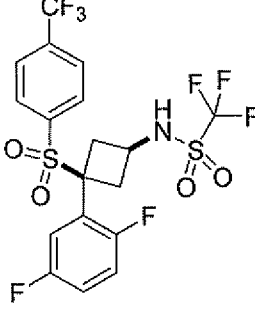


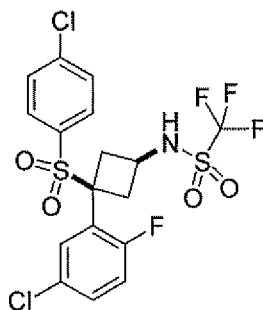
15 ***N*-[*cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide**

Prepared as for Example 1, using *trans*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine.

¹H NMR (600 MHz, CDCl₃) δ 7.33 (m, 4H), 6.98 (m, 1H), 6.74 (m, 2H), 5.35 (d, *J* = 7.8 Hz, 1H, NH), 4.67 (m, 1H), 3.59 (m, 2H), 2.68 (m, 2H). MS calculated 553.0 (MNa⁺+CH₃CN), exp
20 552.8 (MNa⁺+CH₃CN).

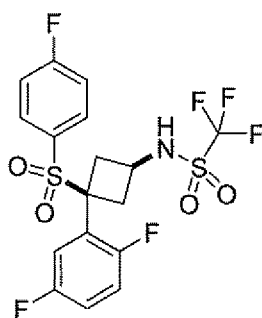
The following were prepared by similar procedures:

#	Structure	Name	MS	Salt form
3		<i>N</i> -[<i>cis</i> -3-[(4-chlorophenyl)sulfonyl]-3-(2-cyano-5-fluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide	Cal'd 519.0(M Na+), exp519.0	Free base
4		<i>N</i> -[<i>cis</i> -3-[(4-chlorophenyl)sulfonyl]-3-(2,5-dichlorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide	Cal'd 543.9(M Na+), exp 543.9.	Free base
5		<i>N</i> -(<i>cis</i> -3-(2,5-difluorophenyl)-3-[[4-(trifluoromethyl)phenyl]sulfonyl]cyclobutyl)-1,1,1-trifluoromethanesulfonamide	Cal'd 546(MNa +), exp 546.	Free base

Example 6

N-{*cis*-3-(5-chloro-2-fluorophenyl)-3-[(4-chlorophenyl)sulfonyl]cyclobutyl}-1,1,1-trifluoromethanesulfonamide: MS Cal'd 506 (MNa⁺), exp 529; ¹H NMR (400MHz, CDCl₃) δ 7.41 ~ 7.43 (d, *J* = 8.8 Hz, 2 H), 7.34 ~ 7.36 (d, *J* = 8.4 Hz, 2 H), 7.28 ~ 7.33 (m, 1 H), 6.98 ~ 7.01 (m, 1 H), 6.75 ~ 6.83 (m, 2 H), 4.20 ~ 4.29 (m, 1 H), 3.20 ~ 3.32 (m, 4 H).

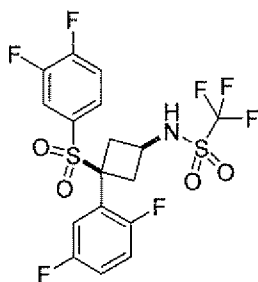
Example 7



10

N-{*cis*-3-(2,5-difluorophenyl)-3-[(4-fluorophenyl)sulfonyl]cyclobutyl}-1,1,1-trifluoromethanesulfonamide: ¹H NMR (400MHz, CDCl₃): δ 7.45 ~ 7.49 (m, 2 H), 7.12 ~ 7.29 (m, 2 H), 7.04 ~ 7.10 (m, 1 H), 6.80 ~ 6.85 (m, 3 H), 4.24 ~ 4.30 (m, 1 H), 3.24 ~ 3.35 (m, 4 H).

Example 8

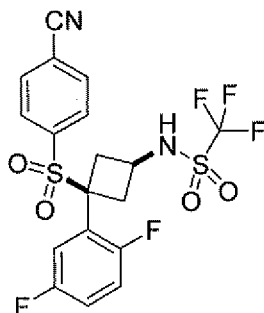


N-{*cis*-3-(2,5-difluorophenyl)-3-[(3,4-difluorophenyl)sulfonyl]cyclobutyl}-1,1,1-trifluoromethanesulfonamide: MS Cal'd 514 (MH⁺), exp 514; ¹H NMR (400MHz, CDCl₃): δ

20

7.15 ~ 7.19 (m, 3 H), 6.97~ 7.04 (m, 1 H), 6.72 ~ 6.80 (m, 2 H), 6.63 ~ 6.65 (d, J = 10.4 Hz, 1 H), 4.13 ~ 4.23 (m, 1 H), 3.14 ~ 3.26 (m, 4 H).

Example 9



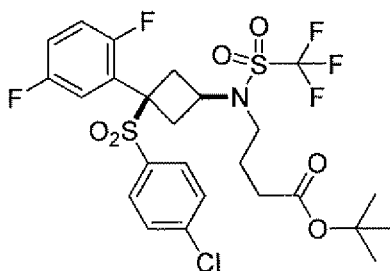
5

N-[*cis*-3-[(4-cyanophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide

MS: cal'd 502 (MNa⁺), exp 502 (MNa⁺). ¹H NMR (CDCl₃, 600MHz) 7.67 (d, J = 4.1 Hz, 2H), 7.58 (d, J = 4.1, 2H), 7.08 (bm, 1H), 6.84 (m, 1H), 6.68 (m, 1H), 6.60 (d, 1H), 4.24 (bm, 1H), 3.4-3.2 (bm, 4H)

10

Intermediate P



15

Tert-Butyl 4-[[3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]amino]butanoate

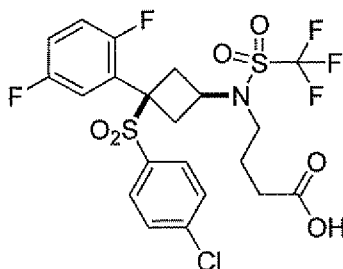
20

N-[*cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide (190 mg, 0.388 mmol) was added to DMF (1.1 mL) and treated

with potassium carbonate (59 mg, 0.427 mmol), tert -butyl 4-bromobutanoate (95 mg, 0.427 mmol). The mixture was heated to 80 °C and stirred for 16 hours. The reaction was cooled to ambient temperature, diluted with ethyl acetate, and washed with 1/2 saturated brine solution twice. The organic layer was dried over anhydrous magnesium sulfate, filtered then
 5 concentrated in vacuo. The residue was purified by MPLC (0-30 % EtOAc:Hept) to give the title compound. MS: cal'd 654 (M Na+), exp 654 (M Na+)

Example 10

10



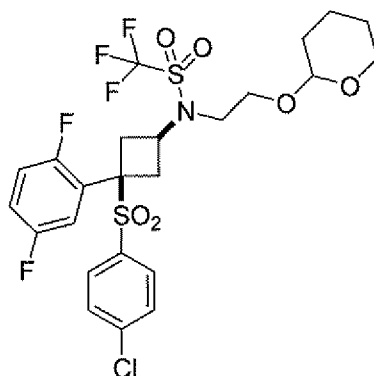
4-({ [cis -3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]amino}butanoic acid

15

Tert-Butyl 4-{{3-[(4-chlorophenyl)sulfonyl]-3-(2,5

difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]amino}butanoate (144 mg, 0.228 mmol) was added to 1:1 DCM:TFA (1.1 mL) and stirred at ambient temperature for 40 minutes. The reaction mixture was concentrated in vacuo. The title compound was isolated as a white solid after trituration with heptane. ¹H NMR (DMSO D₆, 600MHz) δ 12.25 (s, 1H), 7.58 (d, J=8.2 Hz, 2H), 7.43 (d, J=8.2 Hz, 2H), 7.25-7.31 (m, 1H), 7.07-7.16 (m, 2H), 4.18-4.28 (m, 1H), 3.53 (s br, 2H), 3.30-3.42 (m, 2H), 3.08 (s br, 2H), 2.26-2.38 (m, 2H), 1.80-1.90 (m, 2H). MS: cal'd 598 (M Na+), exp 598 (M Na+).

20

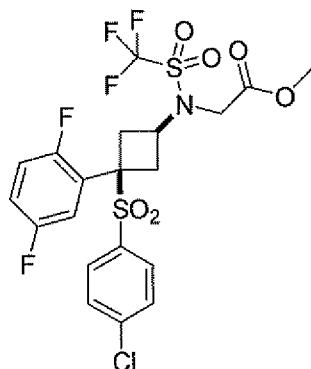
Example 11

5 **N- [cis -3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoro- N- [2-(tetrahydro-2 -pyran-2-yloxy)ethyl]methanesulfonamide**

N-[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide (50 mg, 0.102 mmol) was dissolved in anhydrous DMF (0.3 mL) and to this stirring solution was added potassium carbonate (49 mg, 0.357 mmol) followed by 2-(2-bromoethoxy)tetrahydro-2 H-pyran (53 mg, 0.255 mmol). The resulting mixture was stirred at 80 °C for 16 hours. The mixture was cooled to ambient temperature, diluted with water, and extracted with EtOAc. The organic layer was again washed with saturated aqueous bicarbonate solution, then dried over anhydrous magnesium sulfate and concentrated in vacuo. The extract was purified by MPLC (0-45% EtOAc/ DCM) to give the title compound. R_f = 0.71 in 40% EtOAc/ DCM. ¹H NMR (CDCl₃, 600MHz) δ 7.31-7.38 (m, 4H), 7.00-7.07 (m, 2H), 6.88-6.93 (m, 1H), 6.78-6.84 (m, 1H), 4.65-4.68 (m, 1H), 4.06-4.08 (m, 1H), 3.50-4.00 (m, 7H), 2.80-3.10 (m, 2H), 1.20-1.90 (m, 7H). MS: cal'd 640 (M Na⁺), exp 640 (M Na⁺)

10

15

Example 12

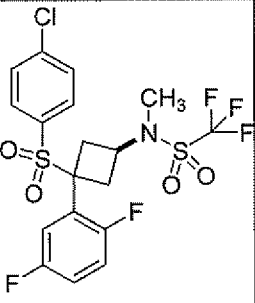
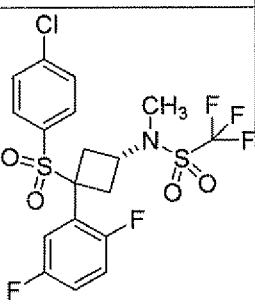
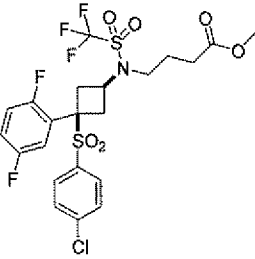
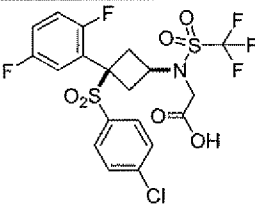
5 **Methyl {[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]amino}acetate**

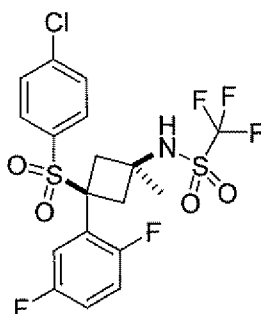
Anhydrous THF (0.8 mL) was added to N-[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide (200 mg, 0.408 mmol) and the reaction was then cooled to 0 °C. Sodium hydride (49 mg, 1.225 mmol) was added in one
 10 portion and the mixture was stirred at 0 °C for 15 minutes. The mixture effervesced and stirred as an off white suspension. After 15 minutes methyl bromoacetate (187 mg, 1.225 mmol) was added and the mixture effervesced again as it turned yellow. The mixture was stirred for 16 hours then quenched with saturated aq ammonium chloride and extracted with EtOAc. The organic layer was again washed with saturated aq ammonium chloride then dried over anhydrous
 15 magnesium sulfate and concentrated in vacuo. The extract was purified by MPLC (0-45% EtOAc:Hept) to give the title compound. R_f=0.6 in 40% EtOAc:Hept. ¹H NMR (CDCl₃, 600MHz) δ 7.36 (d, J=7.0 Hz, 2H), 7.43 (d, J=7.0 Hz, 2H), 7.02-7.08 (m, 1H), 6.86-6.90 (m, 1H), 6.78-6.84 (m, 1H), 4.30-4.60 (m, 3H), 3.85 (s, 3H), 3.38-3.34 (m, 2H), 3.04 (s br, 2H). MS: cal'd 584 (M Na⁺), exp 584 (M Na⁺)

20

The following list of compounds was prepared by similar procedures:

25

#	Structure	Name	MS	Salt form
13		<i>N</i> -[3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoro- <i>N</i> -methylmethanesulfonamide	Cal'd 526.0(M Na+), exp 525.8 (MNa+)	Free base
14		<i>N</i> -[3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoro- <i>N</i> -methylmethanesulfonamide	Cal'd 567.0 (MNa ⁺⁺ MeCN), exp 566.8 (MNa ⁺⁺ MeCN).	Free base
15		methyl 4-[[<i>cis</i> -3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]amino}butanoate	Cal'd 612.0 (MNa+), exp 612.0	Free base
16		<i>N</i> -[<i>cis</i> -3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]- <i>N</i> -[(trifluoromethyl)sulfonyl]glycine	Cal'd 570 (MNa+), exp 570.	Free base

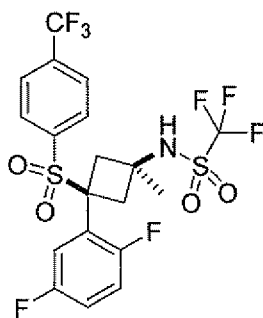
Example 17

***N*-[*cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutyl]-1,1,1-trifluoromethanesulfonamide**

5 Prepared as for Example 1, using *cis*-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)-1-methylcyclobutanamine.

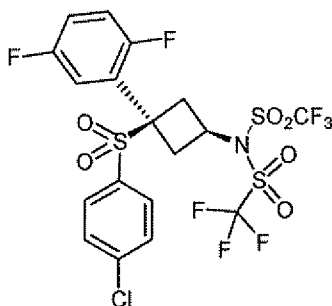
¹H NMR (600 MHz, CDCl₃) δ 7.33 (m, 4H), 7.00 (m, 1H), 6.90 (s, 1H, NH), 6.82-6.75 (m, 2H), 3.54 (d, *J* = 14.4 Hz, 2H), 2.87 (d, *J* = 14.4 Hz, 2H), 1.44 (s, 3H). MS calculated 526.0 (MNa⁺), exp 525.9 (MNa⁺).

10

Example 18

15 ***N*-[*cis*-3-(2,5-difluorophenyl)-1-methyl-3-{4-(trifluoromethyl)phenyl}sulfonyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide**

Prepared using procedures similar to example 17. Calcd (2M+Na)⁺: 1097.0, Found: 1096.5.

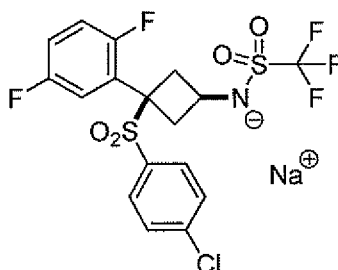
Example 19

5 ***N*-[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoro-*N*-[(trifluoromethyl)sulfonyl]methanesulfonamide**

Prepared in Example 1 synthesis as byproduct.

¹H NMR (600 MHz, CDCl₃) δ 7.38-7.33 (m, 4H), 7.07 (m, 1H), 6.95 (m, 1H), 6.84 (m, 1H), 4.44 (m, 1H), 3.82 (m, 2H), 3.15 (m, 2H). MS calculated 684.9 (MNa⁺+CH₃CN), exp 684.9 (MNa⁺+CH₃CN).

10

Example 20

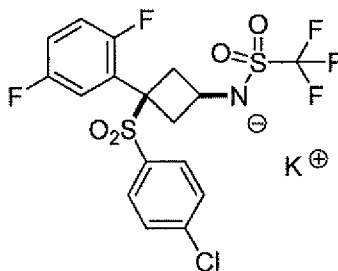
15 **Sodium [cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]azanide**

Sodium hydride was suspended in hexane and cooled to 0 °C. *N*-[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide (200 mg, 0.408 mmol) in THF (1 mL) was added dropwise to the sodium hydride suspension. The resulting mixture was stirred at 0 °C for 15 minutes then at ambient temperature for 30 minutes. At which time, the reaction mixture was concentrated in *vacuo*. A dry white powder was scraped out of the flask, placed in a glass fritted funnel and washed with ice cold pentane (45 mL). The powder was then placed under high vacuum for 16 hours. ¹H NMR (DMSO D₆,

600MHz) δ 7.56 (d, $J=8.5$ Hz, 2H), 7.33 (d, $J=8.5$ Hz, 2H), 7.20-7.26 (m, 1H), 7.00-7.12 (m, 2H), 3.42-3.52 (m, 1H), 2.66-2.80 (m, 4H).

Example 21

5

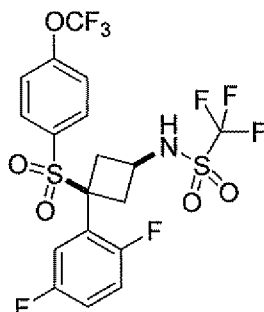


Potassium [cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl][(trifluoromethyl)sulfonyl]azanide

10 N-[cis-3-[(4-chlorophenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide (1.2 g, 2.55 mmol) was stirred in anhydrous THF (25.5 mL) at 0 °C and then treated with potassium tert-butoxide (0.29 g, 2.55 mmol). The reaction mixture was stirred at 0 °C for 15 minutes then warmed to ambient temperature and stirred for another 45

15 minutes. After the reaction was concentrated in *vacuo*, the resultant white powder was recrystallized from a minimal amount of 3:1 IPA:Toluene (400 mL) stirring at 100 °C. Once in solution the mixture was filtered through paper and allowed to sit undisturbed at 4 °C for 20

20 hours. Crystals were harvested by filtration through a glass frit, and washed with cold pentane three times. Residual solvent was removed under vacuum. ¹H NMR (DMSO D₆, 600MHz) δ 7.56 (d, $J=8.5$ Hz, 2H), 7.33 (d, $J=8.5$ Hz, 2H), 7.20-7.26 (m, 1H), 7.00-7.12 (m, 2H), 3.42-3.52 (m, 1H), 2.66-2.80 (m, 4H).

Example 22

***N*-[*cis*-3-[(4-trifluoromethoxyphenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutyl]-1,1,1-trifluoromethanesulfonamide**

Prepared as for Example 2, using *cis*-3-[(4-trifluoromethoxyphenyl)sulfonyl]-3-(2,5-difluorophenyl)cyclobutanamine.

¹H NMR (600 MHz, CDCl₃) δ 7.56 (m, 2H), 7.27 (m, 2H) 7.088 (m, 1H), 6.78 (m, 2H), 6.68 (d, *J* = 10.6 Hz, 1H, NH), 4.30 (m, 1H), 3.31 (m, 2H), 3.18 (m, 2H). MS calculated 603.46

(MNa⁺+CH₃CN), exp 603.0 (MNa⁺+CH₃CN).

BIOLOGICAL ACTIVITY

Assays to determine the biological activity of the compounds of the invention are described as follows:

APP processing (Assay quantitates secreted Aβ analytes from cell lines) :

The effect of compounds on the abundance of Aβ₄₀ and Aβ₄₂ peptides generated from SH-SY5Y cells expressing amyloid β protein (SP4CT cells) was determined by an AlphaLisa™ assay. Analogous to an ELISA assay, generation of signal in this AlphaLisa™ assay requires "donor" and "acceptor" beads to be brought in close proximity by specific antibody recognition of either Aβ₄₀ or Aβ₄₂ peptides. The assay was accomplished by removing media from compound-treated SP4CT cells to two different microplates, followed by the addition of donor beads conjugated with streptavidin binding a biotinylated anti-amyloid β monoclonal antibody (clone 4G8). Acceptor beads directly conjugated with anti-Aβ₄₀ monoclonal antibody (G210) were added to one microplate and anti-Aβ₄₂ monoclonal antibody (12F4) acceptor beads were added to the other. Abundance of Aβ₄₀ and Aβ₄₂ was directly proportional to the luminescent signal generated following excitation of donor beads by laser light.

Notch processing: (Assay quantitates Notch intracellular domain release in cell lines):

A "split-luciferase" assay is used to measure inhibition of gamma secretase-dependent cleavage of the Notch protein. In this assay, HeLa cells were made to express a Notch protein lacking its extracellular domain (Notch Δ E) fused to an N-terminal fragment of luciferase. The same cells also expressed a C-terminal fragment of luciferase fused to the immunoglobulin J kappa recombination signal sequence binding protein (RBP). Upon Notch Δ E cleavage by gamma secretase, a Notch intracellular domain (NICD)-N terminal luciferase protein is generated which translocates to the nucleus and binds the RBP-C terminal luciferase fusion, bringing two independently nonfunctional halves of luciferase together to form a functional luciferase enzyme. The activity of luciferase in these cells is directly proportional to the amount of gamma secretase-cleaved Notch. Luciferase activity is determined by the standard techniques of luciferin addition to lysed cells and measurement of total luminescence.

PXR assay description

The CYP3A4-SEAP transactivation assay (PCSTA) effectively and rapidly evaluates compounds for their potential to induce cytochrome P450 CYP3A (human CYP3A4 or rat CYP3A1). The reporter construct contains regulatory regions from the CYP3A4 gene positioned just upstream of a secreted alkaline phosphatase (SEAP) gene. The human PXR nuclear receptor has been modified at the 5'-end so that methionine is the initiating amino acid replacing leucine found in the wild-type sequence. HEP G2 cells are transfected with the PXR plasmid (human or rat) and the reporter plasmid. Read-out for the induction of CYP3A4 consists of a SEAP colorimetric assay with para-nitro-phenyl phosphate (pNPP) as the substrate. Five point dose-response curves in duplicate are then generated with each point corresponding to the rate of conversion of pNPP to pNP by SEAP. For the human PCSTA, rifampicin is used as the positive control and 100% induction is based upon the maximum induction produced by rifampicin at 10 μ M.

ICD Transactivation (Assay quantitates intracellular domain release of a panel of γ -secretase substrates in cell lines)

A Firefly luciferase based transactivation assay is used to measure inhibition of ϵ /S3-site cleavage of γ -secretase substrates. This assay involves the use of chimeric substrates harboring a GAL4/VP16 (GVP) transactivation domain fused to the intracellular domain (ICD): APP-GVP, Notch Δ E-GVP, E-cadherin-GVP and CD44-GVP. Upon cleavage and release of ICDs, the GVP domain drives the expression of the luciferase gene under the control of the UAS promoter. In this assay, HEK cells were transiently co-transfected with the chimeric substrate along with a UAS promoter driven luciferase and β -galactosidase (transfection control). Upon

cleavage by γ -secretase, the released ICD-GVP translocates to the nucleus to drive the expression of the UAS-luciferase gene. The activity of luciferase in these cells is directly proportional to the amount of γ -secretase-cleaved ICDs. Luciferase activity is determined by the standard techniques of luciferin addition to lysed cells and measurement of total luminescence. In addition, to account for the differences in transfection efficiencies an absorbance based β -galactosidase enzyme assay is performed to normalize the luminescence read-out.

Assessing full length γ -secretase substrates (Assay qualitatively assesses the processing of a panel of γ -secretase substrates)

To examine the effect of compounds on γ -secretase activity against other substrates, four HEK 293 stable cell lines over-expressing one of the following type I membrane proteins: CD43, CD44, E-Cadherin and SCN2b with a C-terminal V5 tag, were generated. Cells are plated and treated overnight with titrated compound and the phorbol ester, TPA. Since all of the proteins undergo regulated membrane proteolysis characterized by an initial ectodomain shedding event followed by the intramembraneous cleavage of the C-terminal fragment (CTF) by γ -secretase, TPA induces the initial cleavage event producing the substrate for γ -secretase. The effect of compounds on γ -secretase activity in relation to these substrates is measured by tracking the processing of the V5 tagged CTFs by Western blot analysis. Accumulation of the CTFs indicates inhibition of γ -secretase activity.

ICD Transactivation (Assay quantitates intracellular domain release of a panel of γ -secretase substrates in cell lines)

A Firefly luciferase based transactivation assay is used to measure inhibition of ϵ /S3-site cleavage of γ -secretase substrates. This assay involves the use of chimeric substrates harboring a GAL4/VP16 (GVP) transactivation domain fused to the intracellular domain (ICD): APP-GVP, Notch Δ E-GVP, E-cadherin-GVP and CD44-GVP. Upon cleavage and release of ICDs, the GVP domain drives the expression of the luciferase gene under the control of the UAS promoter. In this assay, HEK cells were transiently co-transfected with the chimeric substrate along with a UAS promoter driven luciferase and β -galactosidase (transfection control). Upon cleavage by γ -secretase, the released ICD-GVP translocates to the nucleus to drive the expression of the UAS-luciferase gene. The activity of luciferase in these cells is directly proportional to the amount of γ -secretase-cleaved ICDs. Luciferase activity is determined by the standard techniques of luciferin addition to lysed cells and measurement of total luminescence. In addition, to account for the differences in transfection efficiencies an absorbance based β -galactosidase enzyme assay is performed to normalize the luminescence read-out.

***In vitro* APP processing (Assay quantitates A β analytes generated from a recombinant APPC100Flag substrate incubated with semi-purified γ -secretase)**

The effect of compounds on the abundance of A β 40 and A β 42 peptides generated from exogenous C100Flag substrate by semi-purified γ -secretase was determined by MESO Scale ELISA. Generation of signal in this MESO Scale assay requires an anti-amyloid monoclonal antibody (clone 4G8) conjugated with streptavidin to bind to a biotin-coated plate. Specific [Ru(bpy)₃]²⁺-labeled monoclonal antibodies for either A β 40 (G210) or A β 42 (12F4) subsequently generate an electrochemiluminescence signal upon electrochemical stimulation. The assay was accomplished by incubating compound, C100Flag substrate and CHAPSO-solubilized P2 membranes from HeLa cells or brains of mouse, rat, or dog. The reaction was then transferred to two different biotinylated microplates for detection of either A β 40 or A β 42.

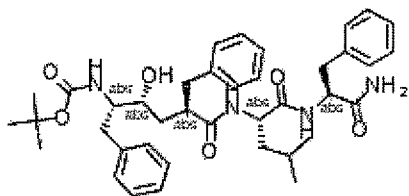
***In vitro* Notch processing (Assay qualitatively assess Notch intracellular domain generation from recombinant Notch Δ E100Flag substrate incubated with semi-purified γ -secretase)**

In an analogous manner, Notch processing can be monitored using the same method as the C100Flag *in vitro* assay but by substituting substrate for N100Flag. A polyclonal biotin-conjugated anti-DYKDDDDK antibody was used as capture antibody while a polyclonal [Ru(bpy)₃]²⁺-labeled cleaved Notch1 antibody was used to detect NICD.

Compound Binding: (Assay quantitates *in vitro* displacement of bound γ -secretase inhibitor tracers)

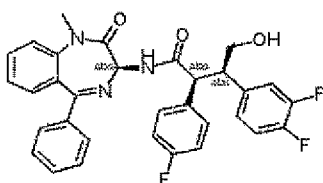
All radioligand binding experiments are performed using CHAPSO-solubilized HEK293 (gammaNRCF8) P2 membranes stably over expressing recombinant gamma secretase. For radioligand binding, solubilized enzyme is incubated in the presence of tritiated inhibitors. Nonspecific binding is determined by adding an excess of unlabeled inhibitor to the reaction, and serial dilutions of the tritiated ligands are used to obtain saturation binding isotherm. Bound ligand is separated from free ligand by adsorption of the enzyme complex to polyethyleneimine-coated glass fiber filter plates and rapid filtration in a cell harvester followed by washing. After drying plates, scintillant is added, and the plates are read on a Microplate Scintillation counter. Binding competition assay is performed by incubating serial dilutions of various inhibitors in the presence of 1nM ³H-labeled compound Reference Example L-458 (transition state inhibitor) or 4nM ³H-labeled compound Reference Example L-881 (non-transition state inhibitor). To determine the antagonist competitiveness of various inhibitors, respective ³H tracer dose-response curves are analyzed in presence of different concentrations of these compounds.

Reference Example L-458:



Reference Example L-881:

5



Results

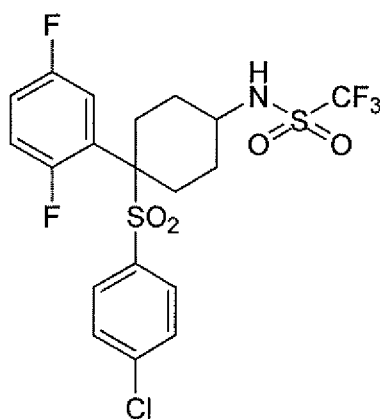
10 The examples herein were tested in the APP and Notch processing cell based functional assays described above. The tested compounds demonstrated *in vitro* inhibition of APP processing while sparing Notch signaling pathway as shown in the following table. The data is based on an average of at least >4 replicates.

Ex. No.	AB40 IC50 AVG (nM)	AB42 IC50 AVG (nM)	Notch IC50 AVG (nM)
1	151.2	117.3	50000
2	42.84	38.37	3485
3	70.08	66.08	3208
4	70.84	78.23	1940

Ex. No.	AB40 IC50 AVG (nM)	AB42 IC50 AVG (nM)	Notch IC50 AVG (nM)
5	100.4	90.21	3808
6	129.9	105.6	5645
7	68.37	57.57	3081
8	79.17	62.84	3142
9	260.5	230.6	8896
10	35.19	30.74	8315
11	43.42	36.93	2538
12	97.71	116.9	13180
13	18.35	18.9	4812
14	89.87	97.09	9865
15	51.78	51.76	2834
16	96.65	67.91	32040
17	37.28	37	5214

Ex. No.	AB40 IC50 AVG (nM)	AB42 IC50 AVG (nM)	Notch IC50 AVG (nM)
18	60.76	65.44	4470
19	62.57	48.7	5677
20	61.24	53.2	1769
21	64.93	70.93	2130
22	250.2	237.3	5204

WO 02/081435 A1, published October 17, 2002, discloses sulfone derivatives that modulate the activity of gamma secretase. WO 2004/031139, published April 15, 2004, discloses cyclohexyl sulfone derivatives as gamma secretase inhibitors. Example 47 of WO
 5 2004/031139 has the following structure:



The above compound is also described as MRK-560 and disclosed in Best et al., *J. Pharmacol. Exp. Ther.*, 317:786-790, 2006 and Best et al., *J. Pharmacol. Exp. Ther.*, 320:552-558, 2007.
 10 Although the literature reports little or no separation for MRK-560 between the *in vitro* inhibition of the APP and Notch processing pathway, the compound did demonstrate *in vivo*

beneficial effects on amyloid plaque deposition in the absence of toxicity related to changes in the Notch signaling pathway in the Tg2576 mouse.

MRK-560 and Example 2 were tested in a covalent protein binding assay which is predictive of drug toxicity. The potential of drug candidates to cause covalent binding to proteins is evaluated by incubation of the radiolabeled version of the compound in question with liver microsomes. A semi-automated method based on Brandel Harvester technique is then used to measure the formation of covalent adducts of the test compound to liver proteins binding (Ref. Day et. al, *J. Pharmacol Toxicol Mehtods.* 52, 278-85, 2005). The results are shown in the table below.

10

Ex. No.	covalent protein binding (human microsomes) pmol/mg
MK-560	1308
2	380

MRK-560 and the examples disclosed herein were tested for their ability to bind to and/or activate the pregnane X receptor (PXR), which is predictive of drug-drug interactions. The results are shown in the following table.

15

Ex. No.	PXR EC50 AVG (nM)	PXR % Activation @ 10uM
MK-560	617	
1	1219	
2	3433	55.5

Ex. No.	PXR EC50 AVG (nM)	PXR % Activation @ 10uM
3	713.6	92.3
4	1956	70.7
5	5201	64.4
6	1989	79.2
7	766.3	113.2
8	1256	82.3
9	8358	54.9
10	1236	76.7
11	826	86.5
12	1603	
13	1261	

Ex. No.	PXR EC50 AVG (nM)	PXR % Activation @ 10uM
14	828.8	
15	1206	
16	1011	100
17	848	74.6
18	3093	75.5
19	1246	89.1
20	2891	71.9
21	3383	64.4
22	9647	50.6

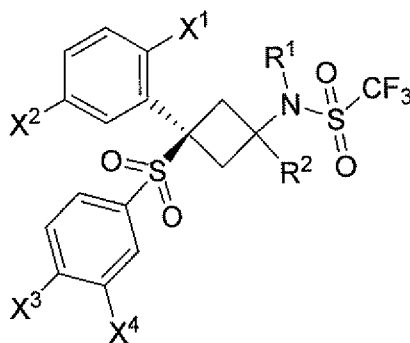
In order to examine the effects on the initial cleavage of other γ -secretase substrates, MRK-560 and Example 2 were tested in a transactivation assay described above. MRK-560 inhibited ICD release of all examined substrates, whereas Example 2 retained initial ϵ /S3-cleavage, as evidenced by ICD release and subsequent translocation of the ICD-GVP construct to allow for reporter activation. See Fig. 2.

Cell-based multi-substrate assay confirmed full inhibition of ϵ /S3- and γ -cleavages by MRK-560 resulting in SCN2b-, ECAD- and CD43-CTF accumulation. In contrast, Example 2 treatment showed no or less CTF accumulation indicating that NS-GSIs spare initial cleavage of γ -secretase substrates. See Fig. 3.

5 MRK-560 and Example 2 were also tested in the compound binding assay described above. Tritiated GSI tracers L-458 (transition state, red) or L-881 (non transition state, blue) were incubated with semi-purified γ -secretase complex and increasing concentrations of the respective compounds. MRK-560 showed full and partial displacement of at L-881 and L-458 sites, respectively. Example 2 was able to fully displace L-881 but not L-458. The results
10 demonstrate that notch sparing compounds such as Example 2 have a shifted binding site as compared to traditional inhibitors such as MRK-560. This deregulates enzymatic cleavage in a manner that spares ϵ /S3 (AICD/NICD release) while potently inhibiting all γ -cleavage sites. See Fig. 4.

WHAT IS CLAIMED IS:

1. A compound according to Formula I



5

I

or pharmaceutically acceptable salt thereof, wherein:

X¹ is selected from the group consisting of: F and CN;

X² is selected from the group consisting of: F, Cl and CN;

10 X³ is selected from the group consisting of: F, Br, Cl, CN, CF₃, OCF₃, C(O)-OCH₃ and S-CH₃;

X⁴ is selected from the group consisting of: H, F and Cl;

R¹ is selected from the group consisting of:

(a) H,

(b) CH₃,

15 (c) -(CH₂)_n-OR³;

(d) -(CH₂)_n-C(O)-OR⁴ and

(e) -SO₂-CF₃;

R² is H or CH₃ when the compound of formula I is in the *cis* configuration, otherwise R² is H;

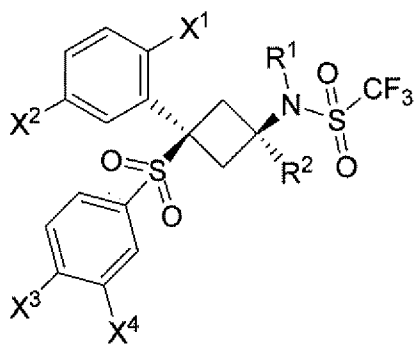
R³ is a five- or six-membered non-aromatic heterocycle having one oxygen heteroatom;

20 R⁴ is H or CH₃; and

n is 1 to 4.

2. The compound according to Claim 1 of Formula Ia

25



Ia

or a pharmaceutically acceptable salt thereof.

5

3. The compound according to Claim 2 wherein:

X¹ is F and

X⁴ is H.

10

4. The compound according to Claim 3 wherein:

X² is F and

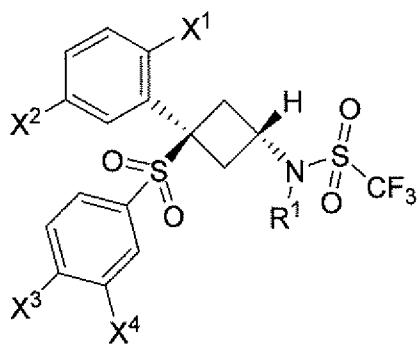
15 X³ is Cl.

5. The compound according to Claim 4 wherein R² is H.

6. The compound according to Claim 5 wherein R¹ is -(CH₂)_n-C(O)-OR⁴.

20

7. The compound according to Claim 1 of Formula Ib



Ib

or a pharmaceutically acceptable salt thereof.

5

8. The compound according to Claim 7 wherein:

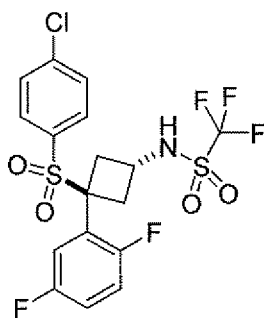
X¹ and X² are F;

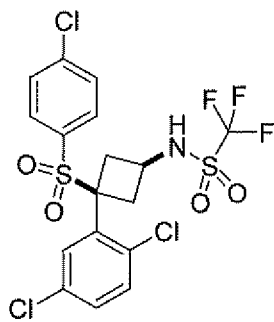
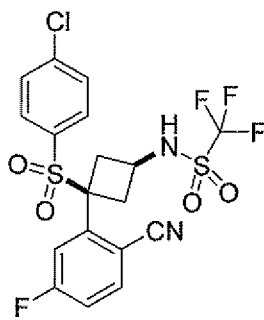
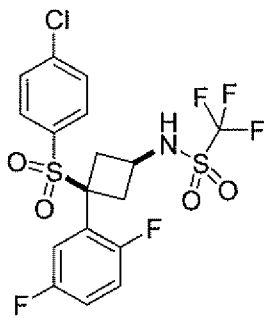
10 X³ is Cl; and

X⁴ is H.

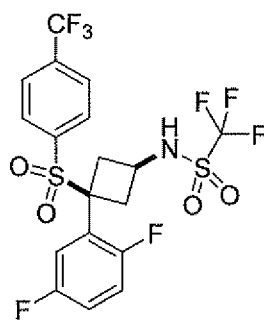
9. A compound according to Claim 1 selected from the following group:

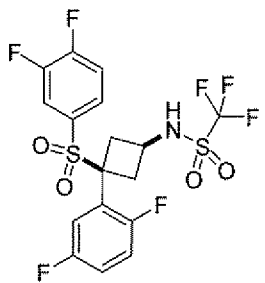
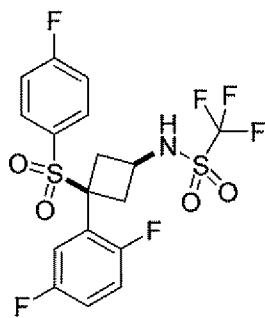
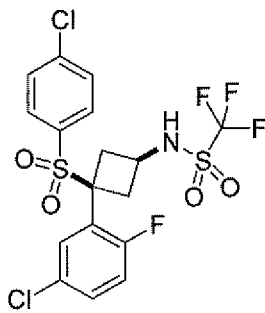
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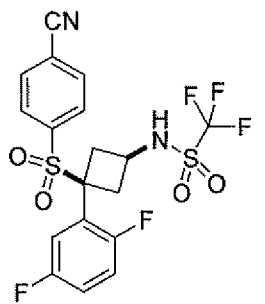


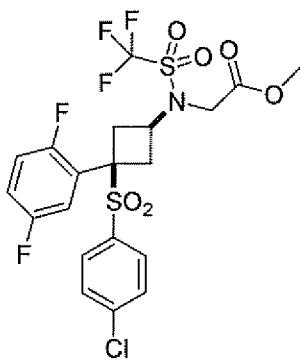
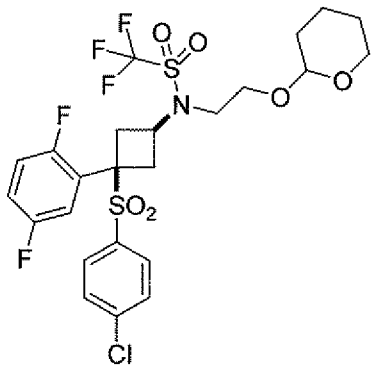
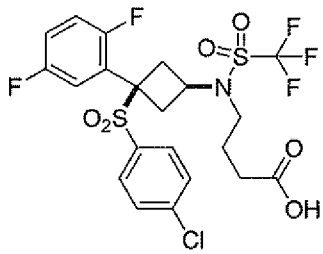
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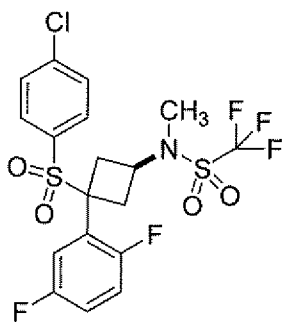


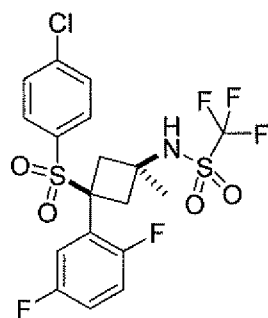
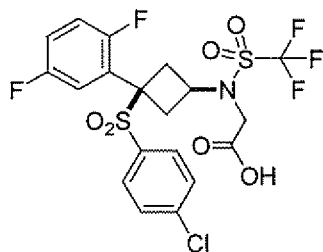
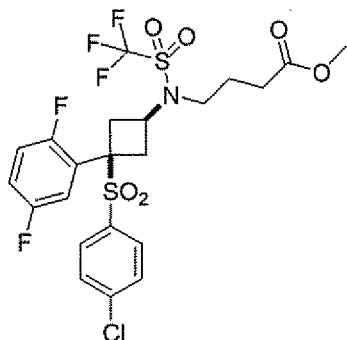
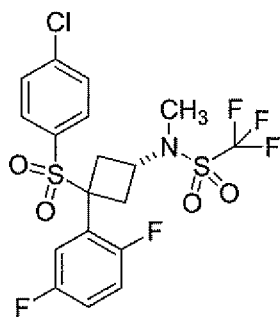
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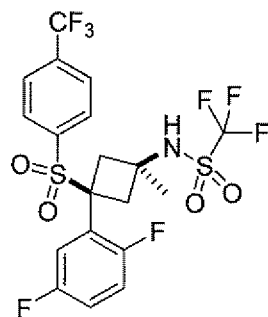


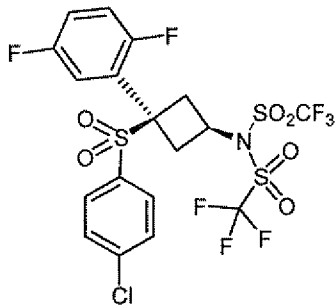
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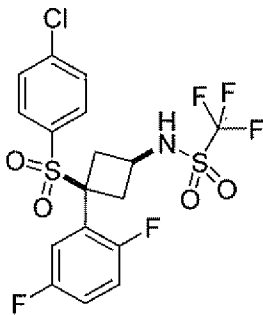
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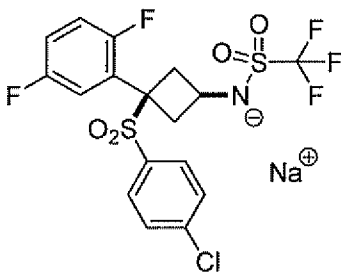
or a pharmaceutically acceptable salt of any of the foregoing compounds.

5 10. A compound according to Claim 9 which is

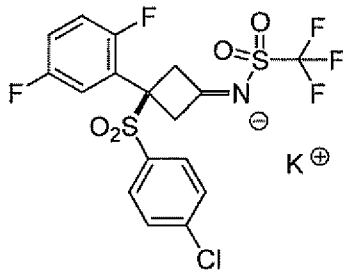


or a pharmaceutically acceptable salt thereof.

10 11. A compound according to Claim 10 which is



15 12. A compound according to Claim 10 which is



13. A pharmaceutical composition comprising a compound according to Claim 1 or a pharmaceutically acceptable salt thereof and a pharmaceutically acceptable carrier.

5

14. A method of treatment of a subject suffering or prone to a condition associated with the deposition of β -amyloid which comprises administering to that subject an effective amount of a compound according to Claim 1 or a pharmaceutically acceptable salt thereof.

10

15. The use of a compound according to Claim 1 or a pharmaceutically acceptable salt thereof for the manufacture of a medicament for treating or preventing Alzheimer's disease.

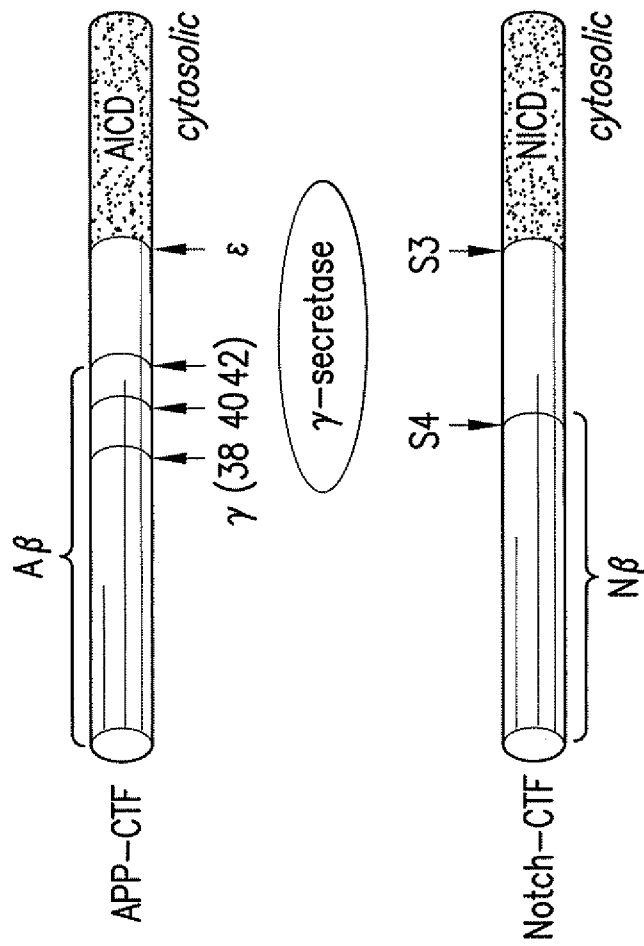


FIG. 1

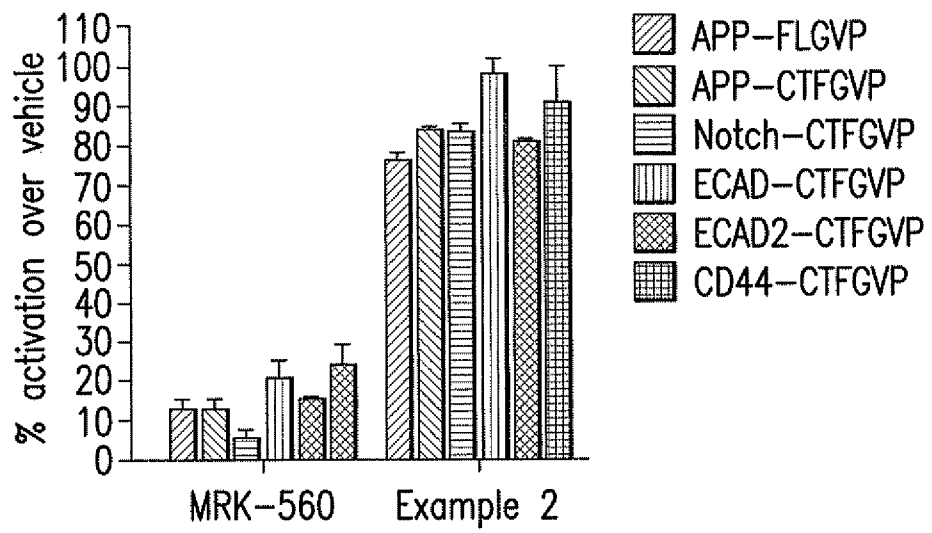


FIG. 2

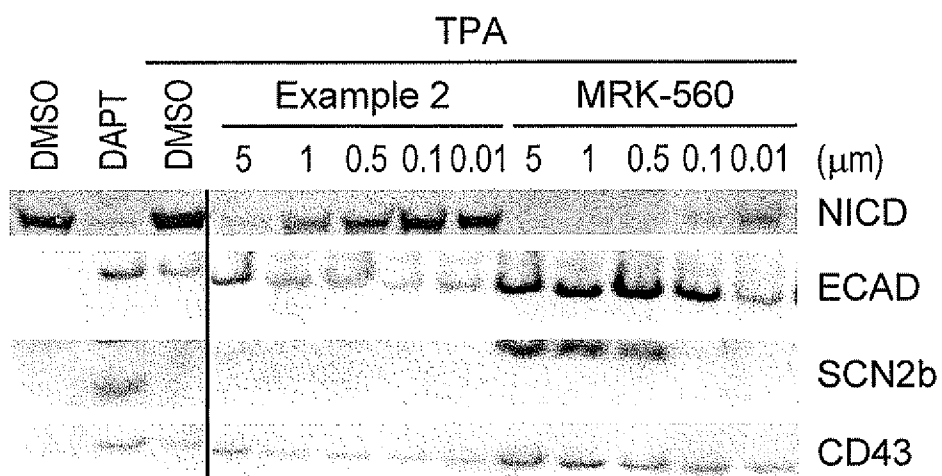
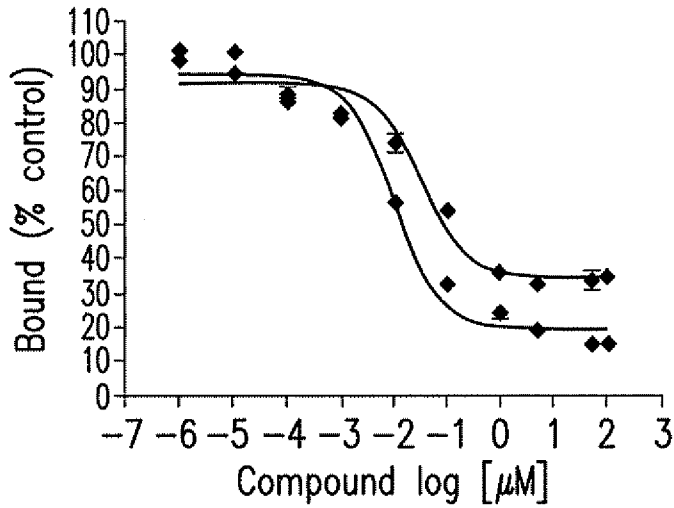


FIG.3

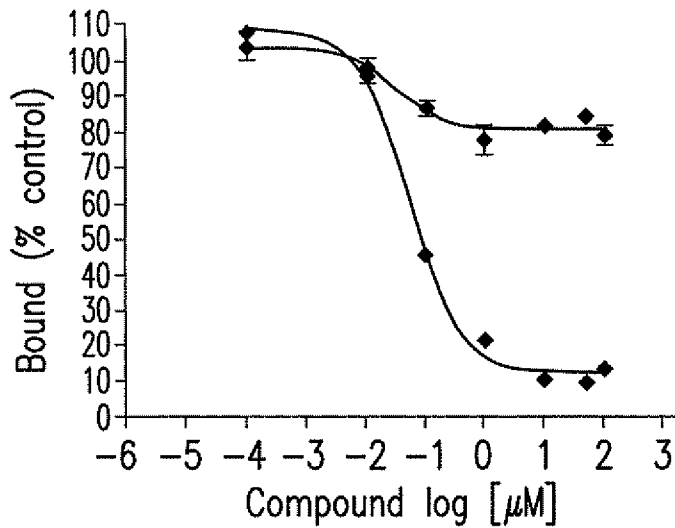
MRK-560:



	L-458	L-881
Log IC ₅₀	-1.498	-1.986
IC ₅₀	31.75nM	10.34nM
Top	91.99%	93.32%
Bottom	34.34%	19.40%
K _I	30.72nM	8.20nM

FIG.4A

Example 2:



	L-458	L-881
Log IC ₅₀	N/A	1.24
IC ₅₀	N/A	57.03nM
Top	103.50%	108.50%
Bottom	80.36%	11.95%
K _I	N/A	45.26nM

FIG.4B

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 09/40913

A. CLASSIFICATION OF SUBJECT MATTER IPC(8) - A01N 41/10; A61K 31/10 (2009.01) USPC - 514/709 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) IPC(8): A01N 41/10; A61K 31/10 (2009.01) USPC- 514/709 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched USPC- 514/570, 520, 485, 532; 568/34, 28; 562/429; Patents and NPL Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) USPTO PubWest (US Patent, PgPub: classification best fit), DialogClassic (Derwent, EPO, JPO, USPTO, WIPO: keyword), GoogleScholar; search terms: cyclobutylsulfon?, notch?, secretase?, alzheimer?, trifluoromethanesulfonamide?, cyclobutanol sulfonyl, tricyl?		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	BEST, et al. 'In Vivo Characterization of ABeta(40) Changes in Brain and Cerebrospinal Fluid Using the Novel gamma-Secretase Inhibitor N-[cis-4-[(4-Chlorophenyl)sulfonyl]-4-(2,5-difluorophenyl)cyclohexyl]-1,1,1-trifluoromethanesulfonamide (MRK-560) in the Rat.' The Journal of Pharmacology and Experimental Therapeutics [online], 2006 [Retrieved on 2009-07-10], Vol. 317, No. 2, pg 786-790. Retrieved from the Internet: <URL: http://jpet.aspetjournals.org/cgi/content/abstract/317/2/786 >.	1-15
Y	US 2007/0203120 A1 (MCDEVITT, et al.) 30 August 2007 (30.08.2007), para [0016], [0262], [0492], [0562]	1-15
Y	WO 2005/030709 A1 (OH, et al.) 07 April 2005 (07.04.2005), pg 4, ln 19 to pg 6, ln 14; pg 9, ln 15 to pg 10, ln 4; pg 12, ln 5 to pg 13, ln 4	6
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/>		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier application or patent but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search 10 July 2009 (10.07.2009)		Date of mailing of the international search report 27 JUL 2009
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450 Facsimile No. 571-273-3201		Authorized officer: Lee W. Young PCT Helpdesk: 571-272-4300 PCT OSP: 571-272-7774