



(11) **EP 3 330 351 A1**

(12) **EUROPEAN PATENT APPLICATION**

(43) Date of publication:  
**06.06.2018 Bulletin 2018/23**

(51) Int Cl.:  
**C11D 3/386 (2006.01)**

(21) Application number: **17204763.1**

(22) Date of filing: **30.11.2017**

(84) Designated Contracting States:  
**AL AT BE BG CH CY CZ DE DK EE ES FI FR GB  
GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO  
PL PT RO RS SE SI SK SM TR**  
Designated Extension States:  
**BA ME**  
Designated Validation States:  
**MA MD**

(72) Inventors:  
• **LANT, Neil Joseph**  
**Newcastle upon Tyne, NE12 9TS (GB)**  
• **FERNANDEZ PRIETO, Susana**  
**1853 Strombeek-Bever (BE)**

(30) Priority: **02.12.2016 EP 16202072**

(71) Applicant: **The Procter & Gamble Company**  
**Cincinnati, OH 45202 (US)**

(74) Representative: **Peet, Jillian Wendy**  
**Procter & Gamble Technical Centres Limited**  
**Whitley Road**  
**Longbenton**  
**Newcastle upon Tyne**  
**NE12 9TS (GB)**

(54) **CLEANING COMPOSITIONS INCLUDING ENZYME AND PLANT FIBER**

(57) Cleaning compositions that include a galactanase enzyme and water-insoluble plant fibers. Methods of making and using such cleaning compositions. Use of water-insoluble plant fibers.

**EP 3 330 351 A1**

**Description**

REFERENCE TO A SEQUENCE LISTING

5 [0001] This application contains a Sequence Listing in computer readable form, which is incorporated herein by reference.

FIELD OF THE INVENTION

10 [0002] The present invention relates to cleaning compositions that include a galactanase enzyme and water-insoluble plant fibers. The present invention also relates to methods of making and using such cleaning compositions. The present invention also relates to the use of galactanase enzymes and water-insoluble plant fibers.

BACKGROUND OF THE INVENTION

15 [0003] The laundry detergent formulator is constantly aiming to improve the performance of detergent compositions. Enzymes may be added to liquid detergent formulations in order to improve cleaning performance, but soils may remain on the targeted surface.

20 [0004] There is a need for improved cleaning compositions that provide improved soil removal benefits.

SUMMARY OF THE INVENTION

25 [0005] The present invention provides a cleaning composition comprising an endo-beta-1,6-galactanase enzyme and water-insoluble plant fibers.

[0006] The present invention also relates to a method of cleaning a surface, preferably a textile, where the method comprises mixing the cleaning composition described herein with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step.

30 [0007] The present invention also relates to a use of an endo-beta-1,6-galactanase enzyme and water-insoluble plant fibers in a cleaning composition to enhance stain-removal and/or malodor-reducing benefits.

DETAILED DESCRIPTION OF THE INVENTION

35 [0008] The present invention relates to cleaning compositions, for example liquid cleaning compositions, comprising a specific galactanase enzyme and water-insoluble plant fiber. Without wishing to be bound by theory, it is believed that the water-insoluble plant fibers result in microabrasion of a target surface, such as a soiled fabric, thereby complementing the cleaning mechanism of the galactanase enzyme and enhancing the removal of the soil matrix. This effect may be particularly strong in instances of direct application of a neat liquid detergent onto the fabric surface, such as in a pretreatment process.

[0009] The components of the compositions and processes of the present invention are described in more detail below.

40 [0010] As used herein, the articles "a" and "an" when used in a claim, are understood to mean one or more of what is claimed or described. As used herein, the terms "include," "includes," and "including" are meant to be non-limiting. The compositions of the present invention can comprise, consist essentially of, or consist of, the components of the present disclosure.

45 [0011] The terms "substantially free of" or "substantially free from" may be used herein. This means that the indicated material is at the very minimum not deliberately added to the composition to form part of it, or, preferably, is not present at analytically detectable levels. It is meant to include compositions whereby the indicated material is present only as an impurity in one of the other materials deliberately included. The indicated material may be present, if at all, at a level of less than 1%, or less than 0.1%, or less than 0.01%, or even 0%, by weight of the composition.

50 [0012] As used herein, "insoluble" means having a water solubility of less than 10% when 1g of the dry material is stirred in 100g of deionized water in a 250ml beaker for 15 minutes at 20°C using a magnetic stirrer set at 100rpm. The degree of solubility is calculated by comparing the mass of dry fiber before ( $m_i = 1g$ ) and after ( $m_f$ ) the solubility test as follows:

55 
$$\% \text{ Solubility} = 100[1 - (m_f / m_i)]$$

[0013] Unless otherwise noted, all component or composition levels are in reference to the active portion of that component or composition, and are exclusive of impurities, for example, residual solvents or by-products, which may

be present in commercially available sources of such components or compositions.

**[0014]** All temperatures herein are in degrees Celsius (°C) unless otherwise indicated. Unless otherwise specified, all measurements herein are conducted at 20°C and under the atmospheric pressure.

**[0015]** In all embodiments of the present disclosure, all percentages are by weight of the total composition, unless specifically stated otherwise. All ratios are weight ratios, unless specifically stated otherwise.

**[0016]** It should be understood that every maximum numerical limitation given throughout this specification includes every lower numerical limitation, as if such lower numerical limitations were expressly written herein. Every minimum numerical limitation given throughout this specification will include every higher numerical limitation, as if such higher numerical limitations were expressly written herein. Every numerical range given throughout this specification will include every narrower numerical range that falls within such broader numerical range, as if such narrower numerical ranges were all expressly written herein.

**[0017]** As used herein, the term "alkoxy" is intended to include C1-C8 alkoxy and C1-C8 alkoxy derivatives of polyols having repeating units such as butylene oxide, glycidol oxide, ethylene oxide or propylene oxide.

**[0018]** As used herein, unless otherwise specified, the terms "alkyl" and "alkyl capped" are intended to include C1-C18 alkyl groups, or even C1-C6 alkyl groups.

**[0019]** As used herein, unless otherwise specified, the term "aryl" is intended to include C3-12 aryl groups.

**[0020]** As used herein, unless otherwise specified, the term "arylalkyl" and "alkaryl" are equivalent and are each intended to include groups comprising an alkyl moiety bound to an aromatic moiety, typically having C1-C18 alkyl groups and, in one aspect, C1-C6 alkyl groups.

**[0021]** The terms "ethylene oxide," "propylene oxide" and "butylene oxide" may be shown herein by their typical designation of "EO," "PO" and "BO," respectively.

**[0022]** As used herein, the term "cleaning and/or treatment composition" includes, unless otherwise indicated, granular, powder, liquid, gel, paste, unit dose, bar form and/or flake type washing agents and/or fabric treatment compositions.

**[0023]** As used herein, "cellulosic substrates" are intended to include any substrate which comprises cellulose, either 100% by weight cellulose or at least 20% by weight, or at least 30 % by weight or at least 40 or at least 50 % by weight or even at least 60 % by weight cellulose. Cellulose may be found in wood, cotton, linen, jute, and hemp. Cellulosic substrates may be in the form of powders, fibers, pulp and articles formed from powders, fibers and pulp. Cellulosic fibers, include, without limitation, cotton, rayon (regenerated cellulose), acetate (cellulose acetate), triacetate (cellulose triacetate), and mixtures thereof. Typically cellulosic substrates comprise cotton. Articles formed from cellulosic fibers include textile articles such as fabrics. Articles formed from pulp include paper.

**[0024]** As used herein, the term "maximum extinction coefficient" is intended to describe the molar extinction coefficient at the wavelength of maximum absorption (also referred to herein as the maximum wavelength), in the range of 400 nanometers to 750 nanometers.

**[0025]** As used herein "average molecular weight" is reported as a weight average molecular weight, as determined by its molecular weight distribution; as a consequence of their manufacturing process, polymers disclosed herein may contain a distribution of repeating units in their polymeric moiety.

**[0026]** As used herein the term "variant" refers to a polypeptide that contains an amino acid sequence that differs from a wild type or reference sequence. A variant polypeptide can differ from the wild type or reference sequence due to a deletion, insertion, or substitution of a nucleotide(s) relative to said reference or wild type nucleotide sequence. The reference or wild type sequence can be a full-length native polypeptide sequence or any other fragment of a full-length polypeptide sequence. A polypeptide variant generally has at least about 70% amino acid sequence identity with the reference sequence, but may include 75% amino acid sequence identity within the reference sequence, 80% amino acid sequence identity within the reference sequence, 85% amino acid sequence identity with the reference sequence, 86% amino acid sequence identity with the reference sequence, 87% amino acid sequence identity with the reference sequence, 88% amino acid sequence identity with the reference sequence, 89% amino acid sequence identity with the reference sequence, 90% amino acid sequence identity with the reference sequence, 91% amino acid sequence identity with the reference sequence, 92% amino acid sequence identity with the reference sequence, 93% amino acid sequence identity with the reference sequence, 94% amino acid sequence identity with the reference sequence, 95% amino acid sequence identity with the reference sequence, 96% amino acid sequence identity with the reference sequence, 97% amino acid sequence identity with the reference sequence, 98% amino acid sequence identity with the reference sequence, 98.5% amino acid sequence identity with the reference sequence or 99% amino acid sequence identity with the reference sequence.

**[0027]** As used herein, the term "solid" includes granular, powder, bar and tablet product forms.

**[0028]** As used herein, the term "fluid" includes liquid, gel, paste, and gas product forms.

#### Cleaning Composition

**[0029]** The present disclosure relates to cleaning and/or treatment compositions. The cleaning composition may be

selected from the group of light duty liquid detergents compositions, heavy duty liquid detergent compositions, solid, for example particulate/powder or "dry" cleaning compositions, hard surface cleaning compositions, detergent gels commonly used for laundry, bleaching compositions, laundry additives, fabric enhancer compositions, shampoos, body washes, other personal care compositions, and mixtures thereof. The cleaning composition may be a hard surface cleaning composition (such as a dishwashing composition) or a laundry composition (such as a heavy duty liquid or solid detergent composition).

**[0030]** The cleaning compositions may be in any suitable form. The composition can be selected from a liquid, solid, or combination thereof. As used herein, "liquid" includes free-flowing liquids, as well as pastes, gels, foams and mousses. Non-limiting examples of liquids include light duty and heavy duty liquid detergent compositions, fabric enhancers, detergent gels commonly used for laundry, bleach and laundry additives. Gases, e.g., suspended bubbles, or solids, e.g. particles, may be included within the liquids. A "solid" as used herein includes, but is not limited to, powders, agglomerates, and mixtures thereof. Non-limiting examples of solids include: granules, micro-capsules, beads, noodles, and pearlised balls. Solid compositions may provide a technical benefit including, but not limited to, through-the-wash benefits, pre-treatment benefits, and/or aesthetic effects.

**[0031]** The cleaning composition may be in the form of a unitized dose article, such as a tablet or in the form of a pouch. Such pouches typically include a water-soluble film, such as a polyvinyl alcohol water-soluble film, that at least partially encapsulates a composition. Suitable films are available from MonoSol, LLC (Indiana, USA). The composition can be encapsulated in a single or multi-compartment pouch. A multi-compartment pouch may have at least two, at least three, or at least four compartments. A multi-compartmented pouch may include compartments that are side-by-side and/or superposed. The composition contained in the pouch may be liquid, solid (such as powders), or combinations thereof.

#### Galactanase Enzyme

**[0032]** The endo-beta-1,6-galactanase enzyme is an extracellular polymer-degrading enzyme. The term "endo-beta-1,6-galactanase" or "a polypeptide having endo-beta-1,6-galactanase activity" means an endo-beta-1,6-galactanase activity (EC 3.2.1.164) that catalyzes the hydrolytic cleavage of 1,6-3-D-galactooligosaccharides with a degree of polymerization (DP) higher than 3, and their acidic derivatives with 4-O-methylglucosyluronate or glucosyluronate groups at the non-reducing terminals.

**[0033]** For purposes of the present disclosure, endo-beta-1,6-galactanase activity is determined according to the procedure described in WO 2015185689 in Assay I. Suitable examples from class EC 3.2.1.164 are described in WO 2015185689, such as the mature polypeptide SEQ ID NO: 2 described therein. Preferably the galactanase enzyme is selected from Glycoside Hydrolase (GH) Family 30.

**[0034]** Preferably, the endo-beta-1,6-galactanase comprises a microbial enzyme. The endo-beta-1,6-galactanase may be fungal or bacterial in origin. Bacterial endo-beta-1,6-galactanase may be most preferred. Fungal endo-beta-1,6-galactanase may be most preferred.

**[0035]** A bacterial endo-beta-1,6-galactanase is obtainable from *Streptomyces*, for example *Streptomyces davawensis*. A preferred endo-beta-1,6-galactanase is obtainable from *Streptomyces davawensis* JCM 4913 defined in SEQ ID NO: 1 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identity thereto.

**[0036]** Other bacterial endo-beta-1,6-galactanase include those encoded by the DNA sequences of *Streptomyces avermitilis* MA-4680 with amino acid sequence defined in SEQ ID NO: 2 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identity thereto.

**[0037]** A fungal endo-beta-1,6-galactanase is obtainable from *Trichoderma*, for example *Trichoderma harzianum*. A preferred endo-beta-1,6-galactanase is obtainable from *Trichoderma harzianum* defined in SEQ ID NO: 3 herein, or a variant thereof, for example having at least 40% or 50% or 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

**[0038]** Other fungal endo-beta-1,6-galactanases include those encoded by the DNA sequences of *Ceratocystis fimbriata* f. sp. Platani, *Muscodor strobilii* WG-2009a, *Oculimacula yallundae*, *Trichoderma viride* GD36A, *Thermomyces stellatus*, *Myceliophthora thermophila*.

**[0039]** Preferably the galactanase has an amino acid sequence having at least 60%, or at least 80%, or at least 90% or at least 95% identity with the amino acid sequence shown in SEQ ID NO:1, SEQ ID NO:2 or SEQ ID NO:3. Preferably the galactanase is an isolated galactanase.

**[0040]** Preferably the galactanase enzyme is present in a laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the galactanase enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm.

**[0041]** The galactanase or composition comprising galactanase may also give rise to biofilm-disrupting effects.

Water-Insoluble Plant Fiber

5 [0042] The liquid cleaning compositions of the present invention include water-soluble plant fiber. Without wishing to be bound by theory, it is believed that the insoluble fibers may microabrade a target surface, thereby synergistically facilitating the cleaning benefits provided by other components of the composition. Additionally, such fibers may be particularly useful in liquid compositions, as they can provide structuring benefits.

[0043] The liquid cleaning compositions of the present invention may include from about 0.01% to about 5%, by weight of the cleaning composition, of water-insoluble fiber.

10 [0044] The water-insoluble plant fiber may be derived from any member of the plant kingdom, including trees, herbaceous plants and the fruits of these plants. Example sources of such fibers are wood, chicory root, sugar beet and citrus or other fruits such as apple. The fibers may be produced as a side-stream from the processing of such crops for other purposes, for example in sugar refining, inulin production and fruit juice production.

[0045] Examples of suitable materials include parenchymal cellulose compositions and activated citrus fibers.

15 [0046] Suitable parenchymal cellulose compositions include particulate cellulose material containing, by dry weight of the particulate cellulose material, at least 70% cellulose, less than 10% pectin and at least 3% hemicellulose, wherein the particulate material has a volume-weighted median major particle dimension within the range of 25-75  $\mu\text{m}$ , preferably within the range of 35-65  $\mu\text{m}$ , as measured by laser light diffractometry in accordance with the established protocol ISO 13320 (2009). The particulate cellulose material of this invention may contain particles of specific structure, shape and size. Typically the material contains particles having the form of platelets comprising parenchymal cellulose structures or networks. It is preferred that the size distribution of the particulate material falls within certain limits. When the distribution is measured with a laser light scattering particle size analyzer, such as the Malvern Mastersizer or another instrument of equal or better sensitivity, the diameter data is preferably reported as a volume distribution. Thus the reported median for a population of particles will be volume-weighted, with about one-half of the particles, on a volume basis, having diameters less than the median diameter for the population. Typically, the median major dimension of the particles of the parenchymal cellulose composition is within the range of 25-75  $\mu\text{m}$ . More preferably the median major dimension of the particles of the parenchymal cellulose composition is within the range of 35-65  $\mu\text{m}$ . Typically at least about 90%, on a volume basis, of the particles has a diameter less than about 120  $\mu\text{m}$ , more preferably less than 110  $\mu\text{m}$ , more preferably less than 100  $\mu\text{m}$ . Preferably, the particulate cellulose material has a volume-weighted median minor dimension larger than 0.5  $\mu\text{m}$ , preferably larger than 1  $\mu\text{m}$ .

20 [0047] The parenchymal cellulose is characterized by the fact that the majority of the cellulose material is present in the form of particles that are distinct from the nanofibrillated cellulose described in the prior art in that the cellulose nanofibrils are not substantially unraveled. Preferably, less than 10%, or more preferably less than 1% or less than 0.1% by dry weight of the cellulose within the composition is in the form of nanofibrillated cellulose. This is advantageous as nanofibrillated cellulose negatively affects the redispersability of the material, as indicated herein before. By 'nanofibrils' we refer to the fibrils making up the cellulose fibers, typically having a width in the nanometer range (e.g., less than 1  $\mu\text{m}$ ) and a length of up to 20  $\mu\text{m}$ . The nomenclature used in the field over the past decades has been somewhat inconsistent in that the terms 'microfibril' and 'nanofibril' have been used to denote the same material. In the context of this invention, the two terms are deemed to be fully interchangeable.

25 [0048] In accordance with the invention, the plant parenchymal cellulose material has been treated, modified and/or some components may have been removed but the cellulose at no time has been broken down to individual microfibrils, thereby losing the structure of plant cell wall sections. As mentioned before, the cellulose material of this invention has a reduced pectin content, as compared to the parenchymal cell wall material from which it is derived. Removal of some of the pectin is believed to result in enhanced thermal stability. The term "pectin" as used herein refers to a class of plant cell-wall heterogeneous polysaccharides that can be extracted by treatment with acids and chelating agents. Typically, 70-80% of pectin is found as a linear chain of alpha-(1,4)-linked D-galacturonic acid monomers. The smaller RG-I fraction of pectin is comprised of alternating (1-4)-linked galacturonic acid and (1,2)-linked L-rhamnose, with substantial arabinogalactan branching emanating from the L-rhamnose residue. Other monosaccharides, such as D-fucose, D-xylose, apiose, aceric acid, Kdo, Dha, 2-O-methyl-D-fucose, and 2-O-methyl-D-xylose, are found either in the RG-II pectin fraction (<2%), or as minor constituents in the RG-I fraction. Proportions of each of the monosaccharides in relation to D-galacturonic acid vary depending on the individual plant and its micro-environment, the species, and time during the growth cycle. For the same reasons, the homogalacturonan and RG-I fractions can differ widely in their content of methyl esters on GalA residues, and the content of acetyl residue esters on the C-2 and C-3 positions of GalA and neutral sugars. It is preferred that the particulate cellulose material of the invention comprises less than 5 wt.% of pectin, by dry weight of the particulate cellulose material, more preferably less than 2.5 wt.%. The presence of at least some pectin in the cellulose material is nevertheless desired. Without wishing to be bound by any theory it is assumed that pectin plays a role in the electrostatic interactions between particles contained in the material and/or in supporting the network/structure of the cellulose. Hence, it is preferred that the particulate cellulose material contains at least 0.5 wt% of pectin by dry weight of the particulate cellulose material, more preferably at least 1 wt.%.

**[0049]** The composition of the present invention comprises, based on the total composition weight, from 0.01 to 5 %, preferably 0.05 to 1 %, more preferably from 0.1 to 0.75 % of water insoluble plant derived fibers.

**[0050]** By water insoluble plant derived fibers it is meant herein cellulose micro or nano fibrils and micro or nano crystals. The plant fibers can be extracted from plants, fruits or wood. Water insoluble plant derived fibers sources may be selected from the group consisting of citrus peels, such as lemons, oranges and/or grapefruit; fruits, such as apples, bananas and/or pear; vegetables such as carrots, peas, potatoes and/or chicory; plants such as bamboo, jute, abaca, flax, cotton and/or sisal, cereals, and different wood sources such as spruces, eucalyptus and/or oak. Preferably, the cellulose fibers source may be selected from the group consisting of wood or plants, in particular, spruce, eucalyptus, jute and sisal. After water-insoluble fibers have been activated by high pressure homogenizer (from 80 to 350 bars), the mean hydrodynamic diameter of such fibers is preferably from 3 microns to 130 microns (as measured using the hydrodynamic method), more preferably from 5 microns to 110 microns, even more preferably from 10 to 100microns and the average diameter is from 1nm to 1 micron, preferably from 10 nm to 850 nm, even more preferably from 15 to 350 nm. Without wishing to be bound by any theory, acid hydrolysis of water-insoluble plant derived fibers, would lead to micro or nanocrystals having an average diameter from 1 nm to 100 nm and a length from 200 nm to 3 microns (as measured using the AFM method). Such materials are commercially available from American process under the tradename of Bioplus.

**[0051]** The content of cellulose will vary depending on the source and treatment applied for the extraction of the fibers, and will range from 15 to 100%, preferably above 30%, more preferably above 50%, and even more preferably above 80%.

**[0052]** Such cellulose fibers may comprise pectin, hemicellulose, proteins, lignin and other impurities inherent to the cellulose based material source such as ash, metals, salts and combinations thereof. The cellulose fibers are preferably non-ionic.

**[0053]** Suitable activated citrus fruits may be produced from lemons and limes. These fruits may be de-juiced to leave an insoluble plant cell wall material with some internally contained sugars and pectin. The 'spongy microstructure', known as albedo, may be used to make acidic, powdered citrus fiber. The structure is dried, sieved and then washed to increase the fiber content. Dried materials are typically large (with cell fragments greater than 100 microns), consisting of tightly bound/bonded fibrils). After milling a powdered citrus fiber material is obtained. This procedure leaves much of the natural cell wall intact whilst sugars are removed. The resultant swellable citrus fiber materials are typically used as food additives and are often employed for example in low fat mayonnaise.

**[0054]** A preferred type of powdered citrus fiber for detergent formulations and used in accordance with the present invention is available from Herbafood Ingredients GmbH under the tradename, Herbacel™ AQ+ type N citrus fiber and Citri-Fi 100FG from Fiberstar. This citrus fiber has a total (soluble and insoluble) fiber content of greater than 80% by weight and soluble fiber content of greater than 20% by weight. It is supplied as a fine dried powder with low colour and has a water binding capacity of about 20 kg water per kg of powder.

**[0055]** The citrus fiber of the present disclosure may be activated citrus fiber. To activate the citrus fibers, powdered citrus fiber may be activated (hydrated and opened up structurally) by using a high shear dispersion process at low concentration, in water. It is also advantageous to include a preservative into the premix as the dispersed activated citrus fiber is biodegradable.

**[0056]** A particularly preferred plant fiber may be provided by is Exilva® (from Borregaard).

#### Adjuncts

**[0057]** The cleaning compositions described herein may optionally include other adjunct components, for example selected from surfactants, fabric shading dyes, fabric care benefit agent; additional enzyme; deposition aid; rheology modifier; builder; chelant; bleach; bleaching agent; bleach precursor; bleach booster; bleach activator, bleach catalyst; perfume and/or perfume microcapsules; perfume loaded zeolite; starch encapsulated accord; polyglycerol esters; whitening agent; pearlescent agent; enzyme stabilizing systems; scavenging agents including fixing agents for anionic dyes, complexing agents for anionic surfactants, and mixtures thereof; optical brighteners or fluorescers; polymer including but not limited to soil release polymer and/or soil suspension polymer; dispersants; antifoam agents; non-aqueous solvent; fatty acid; suds suppressors, e.g., silicone suds suppressors; cationic starches; scum dispersants; substantive dyes; colorants; opacifier; antioxidant; hydrotropes such as toluenesulfonates, cumenesulfonates and naphthalenesulfonates; color speckles; colored beads, spheres or extrudates; clay softening agents; anti-bacterial agents. Additionally or alternatively, the compositions may comprise surfactants, and/or solvent systems. Quaternary ammonium compounds may be present, particularly in fabric enhancer compositions, such as fabric softeners, and comprise quaternary ammonium cations that are positively charged polyatomic ions of the structure  $NR_4^+$ , where R is an alkyl group or an aryl group.

Additional Enzymes

**[0058]** Preferably the composition of the invention comprises additional enzymes, for example selected from lipases, amylases, proteases, nucleases, pectate lyases, cellulases, cutinases, and mixtures thereof. The cleaning compositions preferably comprise one or more additional enzymes from the group selected from nucleases. The cleaning compositions preferably comprises one or more additional enzymes selected from the group amylases, lipases, proteases, pectate lyases, cellulases, cutinases, and mixtures thereof. Preferably, the cleaning compositions comprises one or more additional enzymes selected from amylases and proteases and mixtures thereof. Preferably the cleaning compositions comprise one or more additional enzymes selected from lipases. The compositions may also comprise hemicellulases, peroxidases, xylanases, pectinases, keratinases, reductases, oxidases, phenoloxidases, lipoxygenases, ligninases, pullulanases, tannases, pentosanases, malanases,  $\beta$ -glucanases, arabinosidases, hyaluronidase, chondroitinase, lacase and mixtures thereof. When present in the composition, the aforementioned additional enzymes may be present at levels from about 0.00001% to about 2%, from about 0.0001 % to about 1% or even from about 0.001 % to about 0.5% enzyme protein by weight of the composition. Preferably the or each additional enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the active enzyme protein, or from 0.05 or from 0.1ppm to 750 or 500ppm.

Nucleases

**[0059]** Preferably the composition additionally comprises a nuclease enzyme. The nuclease enzyme is an enzyme capable of cleaving the phosphodiester bonds between the nucleotide subunits of nucleic acids. Suitable nuclease enzymes may be deoxyribonuclease or ribonuclease enzyme or a functional fragment thereof. By functional fragment or part is meant the portion of the nuclease enzyme that catalyzes the cleavage of phosphodiester linkages in the DNA backbone and so is a region of said nuclease protein that retains catalytic activity. Thus it includes truncated, but functional versions, of the enzyme and/or variants and/or derivatives and/or homologues whose functionality is maintained.

**[0060]** Preferably the nuclease enzyme is a deoxyribonuclease, preferably selected from any of the classes E.C. 3.1.21.x, where x=1, 2, 3, 4, 5, 6, 7, 8 or 9, E.C. 3.1.22.y where y=1, 2, 4 or 5, E.C. 3.1.30.z where z= 1 or 2, E.C. 3.1.31.1 and mixtures thereof. Nuclease enzymes from class E.C. 3.1.21.x and especially where x=1 are particularly preferred. Nucleases in class E.C. 3.1.22.y cleave at the 5' hydroxyl to liberate 3' phosphomonoesters. Enzymes in class E.C. 3.1.30.z may be preferred as they act on both DNA and RNA and liberate 5'-phosphomonoesters. Suitable examples from class E.C. 3.1.31.2 are described in US2012/0135498A, such as SEQ ID NO:3 therein. Such enzymes are commercially available as DENARASE® enzyme from c-LECTA. Nuclease enzymes from class E.C. 3.1.31.1 produce 3' phosphomonoesters.

**[0061]** Preferably, the nuclease enzyme comprises a microbial enzyme. The nuclease enzyme may be fungal or bacterial in origin. Bacterial nucleases may be most preferred. Fungal nucleases may be most preferred.

**[0062]** The microbial nuclease is obtainable from *Bacillus*, such as a *Bacillus licheniformis* or *Bacillus subtilis* bacterial nucleases. A preferred nuclease is obtainable from *Bacillus licheniformis*, preferably from strain EI-34-6. A preferred deoxyribonuclease is a variant of *Bacillus licheniformis*, from strain EI-34-6 nucB deoxyribonuclease defined in SEQ ID NO:4 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto. Other suitable nucleases are defined in SEQ ID NO: 5 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto. Other suitable nucleases are defined in SEQ ID NO: 6 herein, or variant thereof, for example having at least 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

**[0063]** A fungal nuclease is obtainable from *Aspergillus*, for example *Aspergillus oryzae*. A preferred nuclease is obtainable from *Aspergillus oryzae* defined in SEQ ID NO: 7 herein, or variant thereof, for example having at least 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

**[0064]** Another suitable fungal nuclease is obtainable from *Trichoderma*, for example *Trichoderma harzianum*. A preferred nuclease is obtainable from *Trichoderma harzianum* defined in SEQ ID NO: 8 herein, or variant thereof, for example having at least 60% or 70% or 75% or 80% or 85% or 90% or 95%, 96%, 97%, 98%, 99% or 100% identical thereto.

**[0065]** Other fungal nucleases include those encoded by the DNA sequences of *Aspergillus oryzae* RIB40, *Aspergillus oryzae* 3.042, *Aspergillus flavus* NRRL3357, *Aspergillus parasiticus* SU-1, *Aspergillus nomius* NRRL13137, *Trichoderma reesei* QM6a, *Trichoderma virens* Gv29-8, *Oidiodendron maius* Zn, *Metarhizium guizhouense* ARSEF 977, *Metarhizium majus* ARSEF 297, *Metarhizium robertsii* ARSEF 23, *Metarhizium acridum* CQMa 102, *Metarhizium brunneum* ARSEF 3297, *Metarhizium anisopliae*, *Colletotrichum fiorinae* PJ7, *Colletotrichum sublineola*, *Trichoderma atroviride* IMI 206040, *Tolyposcladium ophioglossoides* CBS 100239, *Beauveria bassiana* ARSEF 2860, *Colletotrichum higginsianum*, *Hirsutella minnesotensis* 3608, *Scedosporium apiospermum*, *Phaeomonilla chlamydospora*, *Fusarium verticillioides* 7600, *Fusarium oxysporum* f. sp. cubense race 4, *Colletotrichum graminicola* M1.001, *Fusarium oxysporum* FOSC 3-a, *Fusarium avenaceum*, *Fusarium langsethiae*, *Grosmannia clavigera* kw 1407, *Claviceps purpurea* 20.1, *Verticillium*

## EP 3 330 351 A1

*longisporum*, *Fusarium oxysporum* f. sp. cubense race 1, *Magnaporthe oryzae* 70-15, *Beauveria bassiana* D1-5, *Fusarium pseudograminearum* CS3096, *Neonectria ditissima*, *Magnaportheopsis poae* ATCC 64411, *Cordyceps militaris* CM01, *Marssonina brunnea* f. sp. 'multigermtubi' MB\_m1, *Diaporthe ampelina*, *Metarhizium album* ARSEF 1941, *Colletotrichum gloeosporioides* Nara gc5, *Madurella mycetomatis*, *Metarhizium brunneum* ARSEF 3297, *Verticillium alfalfae* VaMs.102, *Gaeumannomyces graminis* var. tritici R3-111a-1, *Nectria haematococca* mpVI 77-13-4, *Verticillium longisporum*, *Verticillium dahliae* VdLs.17, *Torubiella hemipterigena*, *Verticillium longisporum*, *Verticillium dahliae* VdLs.17, *Botrytis cinerea* B05.10, *Chaetomium globosum* CBS 148.51, *Metarhizium anisopliae*, *Stemphylium lycopersici*, *Sclerotinia borealis* F-4157, *Metarhizium robertsii* ARSEF 23, *Myceliophthora thermophila* ATCC 42464, *Phaeosphaeria nodorum* SN15, *Phialophora attae*, *Ustilagoidea virens*, *Diplodia seriata*, *Ophiostoma piceae* UAMH 11346, *Pseudogymnoascus pannorum* VKM F-4515 (FW-2607), *Bipolaris oryzae* ATCC 44560, *Metarhizium guizhouense* ARSEF 977, *Chaetomium thermophilum* var. thermophilum DSM 1495, *Pestalotiopsis fici* W106-1, *Bipolaris zeicola* 26-R-13, *Setosphaeria turcica* Et28A, *Arthroderma otae* CBS 113480 and *Pyrenophora tritici-repentis* Pt-1C-BFP.

**[0066]** Preferably the nuclease is an isolated nuclease.

**[0067]** Preferably the nuclease enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the nuclease enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm.

### Acetylglucosaminidases.

**[0068]** Preferably the composition comprises an acetylglucosaminidase enzyme, preferably a  $\beta$ -N-acetylglucosaminidase enzyme from E.C. 3.2.1.52, preferably an enzyme having at least 70%, or at least 75% or at least 80% or at least 85% or at least 90% or at least 95% or at least 96% or at least 97% or at least 98% or at least 99% or at least or 100% identity to SEQ ID NO: 9.

### Mannanases

**[0069]** Preferably the composition comprises a mannanase enzyme. The term "mannanase" means a polypeptide having mannan endo-1,4- beta-mannosidase activity (EC 3.2.1.78) from the glycoside hydrolase family 26 that catalyzes the hydrolysis of 1,4-3-D-mannosidic linkages in mannans, galactomannans and glucomannans. Alternative names of mannan endo-1,4-beta-mannosidase are 1,4-3-D-mannan mannanohydrolase; endo-1,4-3-mannanase; endo-  $\beta$ -1,4-mannase;  $\beta$ -mannanase B; 3-1,4-mannan 4-mannanohydrolase; endo-3-mannanase; and  $\beta$ -D-mannanase. Preferred mannanases are members of the glycoside hydrolase family 26.

**[0070]** For purposes of the present disclosure, mannanase activity may be determined using the Reducing End Assay as described in the experimental section of WO 2015040159. Suitable examples from class EC 3.2.1.78 are described in WO 2015040159, such as the mature polypeptide SEQ ID NO: 2 described therein.

**[0071]** Preferred mannanases are variants having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81 %, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 10 from *Ascobolus stictoides*;

**[0072]** Preferred mannanases are variants having at least 81 %, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 11 from *Chaetomium virens*.

**[0073]** Preferred mannanases are variants having at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 12 from *Preussia aemulans*.

**[0074]** Preferred mannanases are variants having at least at least 65%, at least 66%, at least 67%, at least 68%, at least 69%, at least 70%, at least 71%, at least 72%, at least 73%, at least 74%, at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 13 from *Yunnania penicillata*.

**[0075]** Preferred mannanases are variants having at least at least 75%, at least 76%, at least 77%, at least 78%, at least 79%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91 %, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide SEQ ID NO: 14 from *Myrothecium roridum*. Preferably the mannanase is an isolated mannanase.



**[0076]** Preferably the mannanase enzyme is present in the cleaning compositions in an amount from 0.001 to 1 wt% based on active protein in the composition, or from 0.005 to 0.5 wt% or from 0.01 to 0.25 wt%. Preferably the mannanase enzyme is present in the laundering aqueous liquor in an amount of from 0.01ppm to 1000 ppm of the mannanase enzyme, or from 0.05 or from 0.1ppm to 750 or 500ppm. The compositions of the invention comprising both galactanase and mannanase may be particularly effective against sticky soils and for improved cleaning. It is believed the two enzymes function together in a complementary way.

#### Further Glycosyl Hydrolases

**[0077]** The composition may comprise a glycosyl hydrolase selected from GH family 39 and GH family 114 and mixtures thereof, for example as described in co-pending applications having applicants reference numbers CM4645FM and CM4646 FM, respectively.

#### Proteases.

**[0078]** Preferably the composition comprises one or more proteases. Suitable proteases include metalloproteases and serine proteases, including neutral or alkaline microbial serine proteases, such as subtilisins (EC 3.4.21.62). Suitable proteases include those of animal, vegetable or microbial origin. In one aspect, such suitable protease may be of microbial origin. The suitable proteases include chemically or genetically modified mutants of the aforementioned suitable proteases. In one aspect, the suitable protease may be a serine protease, such as an alkaline microbial protease or/and a trypsin-type protease. Examples of suitable neutral or alkaline proteases include:

(a) subtilisins (EC 3.4.21.62), preferably those derived from *Bacillus sp.*, such as *B. lentus*, *B. alkalophilus*, *B. subtilis*, *B. amyloliquefaciens*, *B. pumilus* and *B. gibsonii* and *B. akibaii* described in WO2004067737, WO2015091989, WO2015091990, WO2015024739, WO2015143360, US 6,312,936 B1, US 5,679,630, US 4,760,025, US7,262,042 and WO09/021867, DE102006022216A1, DE102006022224A1, WO2015089447, WO2015089441, WO2016066756, WO2016066757, WO2016069557, WO2016069563, WO2016069569.

(b) trypsin-type or chymotrypsin-type proteases, such as trypsin (e.g., of porcine or bovine origin), including the *Fusarium* protease described in WO 89/06270 and the chymotrypsin proteases derived from *Cellulomonas* described in WO 05/052161 and WO 05/052146.

(c) metalloproteases, preferably those derived from *Bacillus amyloliquefaciens* described in WO 07/044993A2; from *Bacillus*, *Brevibacillus*, *Thermoactinomyces*, *Geobacillus*, *Paenibacillus*, *Lysinibacillus* or *Streptomyces spp.* Described in WO2014194032, WO2014194054 and WO2014194117; from *Kribella alluminosa* described in WO2015193488; and from *Streptomyces* and *Lysobacter* described in WO2016075078.

(d) protease having at least 90% identity to the subtilase from *Bacillus sp.* TY145, NCIMB 40339, described in WO92/17577 (Novozymes A/S), including the variants of this *Bacillus sp* TY145 subtilase described in WO2015024739, and WO2016066757.

**[0079]** Preferred proteases include those derived from *Bacillus gibsonii* or *Bacillus Lentus*.

**[0080]** Suitable commercially available protease enzymes include those sold under the trade names Alcalase®, Savinase®, Primase®, Durazym®, Polarzyme®, Kannase®, Liquanase®, Liquanase Ultra®, Savinase Ultra®, Ovozyme®, Neutrase®, Everlase® and Esperase® by Novozymes A/S (Denmark), those sold under the tradename Maxatase®, Maxacal®, Maxapem®, Properase®, Purafect®, Purafect Prime®, Purafect Ox®, FN3®, FN4®, Excellase® and Purafect OXP® by Genencor International, those sold under the tradename Opticlean® and Optimase® by Solvay Enzymes, those available from Henkel/ Kemira, namely BLAP (sequence shown in Figure 29 of US 5,352,604 with the following mutations S99D + S101 R + S103A + V104I + G159S, hereinafter referred to as BLAP), BLAP R (BLAP with S3T + V4I + V199M + V205I + L217D), BLAP X (BLAP with S3T + V4I + V205I) and BLAP F49 (BLAP with S3T + V4I + A194P + V199M + V205I + L217D) - all from Henkel/Kemira; and KAP (*Bacillus alkalophilus* subtilisin with mutations A230V + S256G + S259N) from Kao, or as disclosed in WO2009/149144, WO2009/149145, WO2010/56653, WO2010/56640, WO2011/072117, US2011/0237487, WO2011/140316, WO2012/151480, EP2510092, EP2566960 OR EP2705145.

#### Amylases

**[0081]** Preferably the composition may comprise an amylase. Suitable alpha-amylases include those of bacterial or fungal origin. Chemically or genetically modified mutants (variants) are included. A preferred alkaline alpha-amylase is derived from a strain of *Bacillus*, such as *Bacillus licheniformis*, *Bacillus amyloliquefaciens*, *Bacillus stearothermophilus*, *Bacillus subtilis*, or other *Bacillus sp.*, such as *Bacillus sp.* NCIB 12289, NCIB 12512, NCIB 12513, DSM 9375 (USP 7,153,818) DSM 12368, DSMZ no. 12649, KSM AP1378 (WO 97/00324), KSM K36 or KSM K38 (EP 1,022,334).

## EP 3 330 351 A1

Preferred amylases include:

(a) the variants described in WO 94/02597, WO 94/18314, WO96/23874 and WO 97/43424, especially the variants with substitutions in one or more of the following positions versus the enzyme listed as SEQ ID No. 2 in WO 96/23874: 15, 23, 105, 106, 124, 128, 133, 154, 156, 181, 188, 190, 197, 202, 208, 209, 243, 264, 304, 305, 391, 408, and 444.  
(b) the variants described in USP 5,856,164 and WO99/23211, WO 96/23873, WO00/60060 and WO 06/002643, especially the variants with one or more substitutions in the following positions versus the AA560 enzyme listed as SEQ ID No. 12 in WO 06/002643:

26, 30, 33, 82, 37, 106, 118, 128, 133, 149, 150, 160, 178, 182, 186, 193, 203, 214, 231, 256, 257, 258, 269, 270, 272, 283, 295, 296, 298, 299, 303, 304, 305, 311, 314, 315, 318, 319, 339, 345, 361, 378, 383, 419, 421, 437, 441, 444, 445, 446, 447, 450, 461, 471, 482, 484, preferably that also contain the deletions of D183\* and G184\*.

(c) variants exhibiting at least 90% identity with SEQ ID No. 4 in WO06/002643, the wild-type enzyme from Bacillus SP722, especially variants with deletions in the 183 and 184 positions and variants described in WO 00/60060, which is incorporated herein by reference.

(d) variants exhibiting at least 95% identity with the wild-type enzyme from Bacillus sp.707 (SEQ ID NO:7 in US 6,093, 562), especially those comprising one or more of the following mutations M202, M208, S255, R172, and/or M261. Preferably said amylase comprises one or more of M202L, M202V, M202S, M202T, M202I, M202Q, M202W, S255N and/or R172Q. Particularly preferred are those comprising the M202L or M202T mutations.

(e) variants described in WO 09/149130, preferably those exhibiting at least 90% identity with SEQ ID NO: 1 or SEQ ID NO:2 in WO 09/149130, the wild-type enzyme from Geobacillus Stearophermophilus or a truncated version thereof;

(f) variants as described in EP2540825 and EP2357220, EP2534233; (g) variants as described in WO2009100102 and WO2010115028.

**[0082]** Suitable commercially available alpha-amylases include DURAMYL®, LIQUEZYME®, TERMAMYL®, TERMAMYL ULTRA®, NATALASE®, SUPRAMYL®, STAINZYME®, STAINZYME PLUS®, FUNGAMYL® and BAN® (Novozymes A/S, Bagsvaerd, Denmark), KEMZYM® AT 9000 Biozym Biotech Trading GmbH Wehlstrasse 27b A-1200 Wien Austria, RAPIDASE®, PURASTAR®, ENZYSE®, OPTISIZE HT PLUS®, POWERASE® and PURASTAR OX-AM® (Genencor International Inc., Palo Alto, California) and KAM® (Kao, 14-10 Nihonbashi Kayabacho, 1-chome, Chuo-ku Tokyo 103-8210, Japan). In one aspect, suitable amylases include NATALASE®, STAINZYME® and STAINZYME PLUS® and mixtures thereof.

### Lipases

**[0083]** Preferably the composition comprises one or more lipases, including "first cycle lipases" such as those described in U.S. Patent 6,939,702 B1 and US PA 2009/0217464. Preferred lipases are first-wash lipases. In one embodiment of the invention the composition comprises a first wash lipase. First wash lipases includes a lipase which is a polypeptide having an amino acid sequence which: (a) has at least 90% identity with the wild-type lipase derived from *Humicola lanuginosa* strain DSM 4109; (b) compared to said wild-type lipase, comprises a substitution of an electrically neutral or negatively charged amino acid at the surface of the three-dimensional structure within 15Å of E1 or Q249 with a positively charged amino acid; and (c) comprises a peptide addition at the C-terminal; and/or (d) comprises a peptide addition at the N-terminal and/or (e) meets the following limitations: i) comprises a negative amino acid in position E210 of said wild-type lipase; ii) comprises a negatively charged amino acid in the region corresponding to positions 90-101 of said wild-type lipase; and iii) comprises a neutral or negative amino acid at a position corresponding to N94 or said wild-type lipase and/or has a negative or neutral net electric charge in the region corresponding to positions 90-101 of said wild-type lipase. Preferred are variants of the wild-type lipase from *Thermomyces lanuginosus* comprising one or more of the T231R and N233R mutations. The wild-type sequence is the 269 amino acids (amino acids 23 - 291) of the Swissprot accession number Swiss-Prot O59952 (derived from *Thermomyces lanuginosus* (*Humicola lanuginosa*)). Preferred lipases include those sold under the tradenames Lipex® and Lipolex® and Lipoclean®. Other suitable lipases include those described in European Patent Application No. 12001034.3 or EP2623586.

### Endoglucanases

**[0084]** Other preferred enzymes include microbial-derived endoglucanases exhibiting endo-beta-1,4-glucanase activity (E.C. 3.2.1.4), including a bacterial polypeptide endogenous to a member of the genus Bacillus which has a

sequence of at least 90%, 94%, 97% and even 99% identity to the amino acid sequence SEQ ID NO:2 in US7,141,403B2) and mixtures thereof. Suitable endoglucanases are sold under the tradenames Celluclean® and Whitezyme® (Novozymes A/S, Bagsvaerd, Denmark).

#### 5 Pectate Lyases

[0085] Other preferred enzymes include pectate lyases sold under the tradenames Pectawash®, Pectaway®, Xpect® and mannanases sold under the tradenames Mannaway® (all from Novozymes A/S, Bagsvaerd, Denmark), and Purabrite® (Genencor International Inc., Palo Alto, California).

10

#### Cleaning Cellulase

[0086] The cleaning composition described herein may additionally comprise a cleaning cellulase. The cellulase may be an endoglucanase. The cellulase may have endo beta 1,4-glucanase activity and a structure which does not comprise a class A Carbohydrate Binding Module (CBM). A class A CBM is defined according to A. B. Boraston et al. Biochemical Journal 2004, Volume 382 (part 3) pages 769-781. In particular, the cellulase does not comprise a class A CBM from families 1, 2a, 3, 5 and 10.

15

[0087] The cellulase may be a glycosyl hydrolase having enzymatic activity towards amorphous cellulose substrates, wherein the glycosyl hydrolase is selected from GH families 5, 7, 12, 16, 44 or 74. Preferably, the cellulase is a glycosyl hydrolase selected from GH family 5. A preferred cellulase is Celluclean, supplied by Novozymes. This preferred cellulase is described in more detail in WO2002/099091. The glycosyl hydrolase (GH) family definition is described in more detail in Biochem J. 1991, v280, 309-316. Another preferred cellulase is a glycosyl hydrolase having enzymatic activity towards both xyloglucan and amorphous cellulose substrates, wherein the glycosyl hydrolase is selected from GH families 5, 12, 44 or 74. Preferably, the glycosyl hydrolase selected from GH family 44.

20

[0088] For purposes of the present invention, the degree of identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, J. Mol. Biol. 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, Trends in Genetics 16: 276-277), preferably version 3.0.0 or later. The optional parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled "longest identity" (obtained using the -nobrief option) is used as the percent identity and is calculated as follows: (Identical Residues x 100)/(Length of Alignment - Total Number of Gaps in Alignment).

25

[0089] Suitable cleaning cellulase glycosyl hydrolases are selected from the group consisting of: GH family 44 glycosyl hydrolases from *Paenibacillus polyxyrna* (wild-type) such as XYG1006 described in WO 01/062903 or are variants thereof; GH family 12 glycosyl hydrolases from *Bacillus licheniformis* (wild-type) such as Seq. No. ID: 1 described in WO 99/02663 or are variants thereof; GH family 5 glycosyl hydrolases from *Bacillus agaradhaerens* (wild type) or variants thereof; GH family 5 glycosyl hydrolases from *Paenibacillus* (wild type) such as XYG1034 and XYG 1022 described in WO 01/064853 or variants thereof; GH family 74 glycosyl hydrolases from *Jonesia sp.* (wild type) such as XYG1020 described in WO 2002/077242 or variants thereof; and GH family 74 glycosyl hydrolases from *Trichoderma Reesei* (wild type), such as the enzyme described in more detail in Sequence ID no. 2 of WO03/089598, or variants thereof.

35

[0090] Preferred glycosyl hydrolases are selected from the group consisting of: GH family 44 glycosyl hydrolases from *Paenibacillus polyxyrna* (wild-type) such as XYG1006 or are variants thereof.

40

[0091] Typically, the cellulase modifies the fabric surface during the laundering process so as to improve the removal of soils adhered to the fabric after the laundering process during wearing and usage of the fabric, in subsequent wash cycles. Preferably, the cellulase modifies the fabric surface during the laundering process so as to improve the removal of soils adhered to the fabric after the laundering process during wearing and usage of the fabric, in the subsequent two, or even three wash cycles.

45

[0092] Typically, the cellulase is used at a concentration of 0.005ppm to 1.0ppm in the aqueous liquor during the first laundering process. Preferably, the cellulase is used at a concentration of 0.02ppm to 0.5ppm in the aqueous liquor during the first laundering process.

50

#### Surfactant system

[0093] The cleaning composition may comprise a surfactant system. The cleaning composition may comprise from about 1% to about 80%, or from 1% to about 60%, preferably from about 5% to about 50% more preferably from about 8% to about 40%, by weight of the cleaning composition, of a surfactant system.

55

[0094] Surfactants suitable for use in the surfactant system may be derived from natural and/or renewable sources.

[0095] The surfactant system may comprise an anionic surfactant, more preferably an anionic surfactant selected from the group consisting of, alkyl benzene sulfonate, alkyl sulfate, alkyl alkoxy sulfate, especially alkyl ethoxy sulfate,

paraffin sulfonate and mixtures thereof, alkyl benzene sulfonates are particularly preferred. The surfactant system may further comprise a surfactant selected from the group consisting of nonionic surfactant, cationic surfactant, amphoteric surfactant, zwitterionic surfactant, and mixtures thereof. The surfactant system preferably comprises a nonionic surfactant, for example an ethoxylated nonionic surfactant. The surfactant system may comprise an amphoteric surfactant, for example an amine oxide surfactant, such as an alkyl dimethyl amine oxide. The surfactant system may comprise a zwitterionic surfactant, such as a betaine.

**[0096]** The most preferred surfactant system for the detergent composition of the present invention comprises from 1% to 40%, preferably 6% to 35%, more preferably 8% to 30% weight of the total composition of an anionic surfactant, preferably comprising an alkyl benzene sulphonate. The preferred surfactant system may optionally in addition comprise an alkyl alkoxy sulfate surfactant, more preferably an alkyl ethoxy sulfate, optionally combined with 0.5% to 15%, preferably from 1% to 12%, more preferably from 2% to 10% by weight of the composition of amphoteric and/or zwitterionic surfactant, more preferably an amphoteric and even more preferably an amine oxide surfactant, especially an alkyl dimethyl amine oxide.

**[0097]** Preferably the composition further comprises a nonionic surfactant, especially an alcohol alkoxyate in particular an alcohol ethoxyate nonionic surfactant. Most preferably the surfactant system comprises an anionic and a nonionic surfactant, preferably the weight ratio of the anionic to nonionic surfactant is from 25:1 to 1:2.

#### Anionic surfactant

**[0098]** Anionic surfactants may be in salt form or acid form, typically in the form of a water-soluble sodium, potassium, ammonium, magnesium or mono-, di- or tri- C2-C3 alkanolammonium salt, with the sodium cation being the usual one chosen.

#### Sulfonate Surfactant

**[0099]** Suitable anionic sulfonate surfactants for use herein include water-soluble salts of C8-C18 alkyl or hydroxyalkyl sulfonates; C11-C18 alkyl benzene sulfonates (LAS), modified alkylbenzene sulfonate (MLAS) as discussed in WO 99/05243, WO 99/05242, WO 99/05244, WO 99/05082, WO 99/05084, WO 99/05241, WO 99/07656, WO 00/23549, and WO 00/23548; methyl ester sulfonate (MES); and alpha-olefin sulfonate (AOS). Those also include the paraffin sulfonates may be monosulfonates and/or disulfonates, obtained by sulfonating paraffins of 10 to 20 carbon atoms. The sulfonate surfactant may also include the alkyl glyceryl sulfonate surfactants.

#### Sulfated anionic surfactant

**[0100]** Preferably the sulfated anionic surfactant is alkoxyated, more preferably, an alkoxyated branched sulfated anionic surfactant having an alkoxylation degree of from about 0.2 to about 4, even more preferably from about 0.3 to about 3, even more preferably from about 0.4 to about 1.5 and especially from about 0.4 to about 1. Preferably, the alkoxy group is ethoxy. When the sulfated anionic surfactant is a mixture of sulfated anionic surfactants, the alkoxylation degree is the weight average alkoxylation degree of all the components of the mixture (weight average alkoxylation degree). In the weight average alkoxylation degree calculation the weight of sulfated anionic surfactant components not having alkoxyated groups should also be included.

$$\text{Weight average alkoxylation degree} = (x_1 * \text{alkoxylation degree of surfactant 1} + x_2 * \text{alkoxylation degree of surfactant 2} + \dots) / (x_1 + x_2 + \dots)$$

wherein  $x_1$ ,  $x_2$ , ... are the weights in grams of each sulfated anionic surfactant of the mixture and alkoxylation degree is the number of alkoxy groups in each sulfated anionic surfactant.

**[0101]** Preferably, the branching group is an alkyl. Typically, the alkyl is selected from methyl, ethyl, propyl, butyl, pentyl, cyclic alkyl groups and mixtures thereof. Single or multiple alkyl branches could be present on the main hydrocarbyl chain of the starting alcohol(s) used to produce the sulfated anionic surfactant used in the detergent of the invention. Most preferably the branched sulfated anionic surfactant is selected from alkyl sulfates, alkyl ethoxy sulfates, and mixtures thereof.

**[0102]** The branched sulfated anionic surfactant can be a single anionic surfactant or a mixture of anionic surfactants. In the case of a single surfactant the percentage of branching refers to the weight percentage of the hydrocarbyl chains that are branched in the original alcohol from which the surfactant is derived.

**[0103]** In the case of a surfactant mixture the percentage of branching is the weight average and it is defined according

to the following formula:

$$\text{Weight average of branching (\%)} = [(x_1 * \text{wt\% branched alcohol 1 in alcohol 1} + x_2 * \text{wt\% branched alcohol 2 in alcohol 2} + \dots) / (x_1 + x_2 + \dots)] * 100$$

wherein  $x_1, x_2, \dots$  are the weight in grams of each alcohol in the total alcohol mixture of the alcohols which were used as starting material for the anionic surfactant for the detergent of the invention. In the weight average branching degree calculation the weight of anionic surfactant components not having branched groups should also be included.

**[0104]** Suitable sulfate surfactants for use herein include water-soluble salts of C8-C18 alkyl or hydroxyalkyl, sulfate and/or ether sulfate. Suitable counterions include alkali metal cation or ammonium or substituted ammonium, but preferably sodium.

**[0105]** The sulfate surfactants may be selected from C8-C18 primary, branched chain and random alkyl sulfates (AS); C8-C18 secondary (2,3) alkyl sulfates; C8-C18 alkyl alkoxy sulfates (AExS) wherein preferably  $x$  is from 1-30 in which the alkoxy group could be selected from ethoxy, propoxy, butoxy or even higher alkoxy groups and mixtures thereof.

**[0106]** Alkyl sulfates and alkyl alkoxy sulfates are commercially available with a variety of chain lengths, ethoxylation and branching degrees. Commercially available sulfates include, those based on Neodol alcohols ex the Shell company, Lial - Isalchem and Safol ex the Sasol company, natural alcohols ex The Procter & Gamble Chemicals company.

**[0107]** Preferred alkyl sulfates are those in which the anionic surfactant is an alkyl ethoxy sulfate with a degree of ethoxylation of from about 0.2 to about 3, more preferably from about 0.3 to about 2, even more preferably from about 0.4 to about 1.5, and especially from about 0.4 to about 1. They are also preferred anionic surfactant having a level of branching of from about 5% to about 40%, even more preferably from about 10% to 35% and especially from about 20% to 30%.

#### Nonionic surfactant

**[0108]** Preferably the surfactant system comprises a nonionic surfactant, in an amount of from 0.1% to 40%, preferably 0.2% to 20%, most preferably 0.5% to 10% by weight of the composition. Suitable nonionic surfactants include the condensation products of aliphatic alcohols with from 1 to 25 moles of ethylene oxide. The alkyl chain of the aliphatic alcohol can either be straight or branched, primary or secondary, and generally contains from 8 to 22 carbon atoms. Particularly preferred are the condensation products of alcohols having an alkyl group containing from 10 to 18 carbon atoms, preferably from 10 to 15 carbon atoms with from 2 to 18 moles, preferably 2 to 15, more preferably 5-12 of ethylene oxide per mole of alcohol. Highly preferred nonionic surfactants are the condensation products of guerbet alcohols with from 2 to 18 moles, preferably 2 to 15, more preferably 5-12 of ethylene oxide per mole of alcohol.

**[0109]** Other suitable non-ionic surfactants for use herein include fatty alcohol polyglycol ethers, alkylpolyglucosides and fatty acid glucamides.

#### Amphoteric surfactant

**[0110]** The surfactant system may include amphoteric surfactant, such as amine oxide. Preferred amine oxides are alkyl dimethyl amine oxide or alkyl amido propyl dimethyl amine oxide, more preferably alkyl dimethyl amine oxide and especially coco dimethyl amino oxide. Amine oxide may have a linear or mid-branched alkyl moiety. Typical linear amine oxides include water-soluble amine oxides containing one R1 C8-18 alkyl moiety and 2 R2 and R3 moieties selected from the group consisting of C1-3 alkyl groups and C1-3 hydroxyalkyl groups. Preferably amine oxide is characterized by the formula  $R_1 - N(R_2)(R_3) O$  wherein R1 is a C8-18 alkyl and R2 and R3 are selected from the group consisting of methyl, ethyl, propyl, isopropyl, 2-hydroxyethyl, 2-hydroxypropyl and 3-hydroxypropyl. The linear amine oxide surfactants in particular may include linear C10-C18 alkyl dimethyl amine oxides and linear C8-C12 alkoxy ethyl dihydroxy ethyl amine oxides. Preferred amine oxides include linear C10, linear C10-C12, and linear C12-C14 alkyl dimethyl amine oxides. As used herein "mid-branched" means that the amine oxide has one alkyl moiety having  $n_1$  carbon atoms with one alkyl branch on the alkyl moiety having  $n_2$  carbon atoms. The alkyl branch is located on the  $\alpha$  carbon from the nitrogen on the alkyl moiety. This type of branching for the amine oxide is also known in the art as an internal amine oxide. The total sum of  $n_1$  and  $n_2$  is from 10 to 24 carbon atoms, preferably from 12 to 20, and more preferably from 10 to 16. The number of carbon atoms for the one alkyl moiety ( $n_1$ ) should be approximately the same number of carbon atoms as the one alkyl branch ( $n_2$ ) such that the one alkyl moiety and the one alkyl branch are symmetric. As used herein "symmetric" means that  $|n_1 - n_2|$  is less than or equal to 5, preferably 4, most preferably from 0 to 4 carbon atoms in at least 50 wt%, more preferably at least 75 wt% to 100 wt% of the mid-branched amine oxides for use herein.

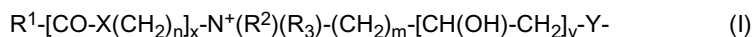
**[0111]** The amine oxide may further comprise two moieties, independently selected from a C1-3 alkyl, a C1-3 hydroxy-

## EP 3 330 351 A1

alkyl group, or a polyethylene oxide group containing an average of from about 1 to about 3 ethylene oxide groups. Preferably the two moieties are selected from a C1-3 alkyl, more preferably both are selected as a C1 alkyl.

### Zwitterionic surfactant

**[0112]** Other suitable surfactants include betaines, such as alkyl betaines, alkylamidobetaine, amidazoliniumbetaine, sulfobetaine (INCI Sultaines) as well as the Phosphobetaine and preferably meets formula (I):



wherein

R<sup>1</sup> is a saturated or unsaturated C6-22 alkyl residue, preferably C8-18 alkyl residue, in particular a saturated C10-16 alkyl residue, for example a saturated C12-14 alkyl residue;

X is NH, NR<sup>4</sup> with C1-4 Alkyl residue R<sup>4</sup>, O or S,

n a number from 1 to 10, preferably 2 to 5, in particular 3,

x 0 or 1, preferably 1,

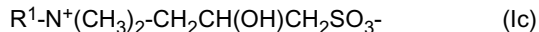
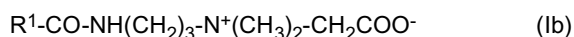
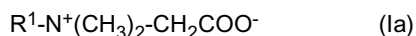
R<sup>2</sup>, R<sup>3</sup> are independently a C1-4 alkyl residue, potentially hydroxy substituted such as a hydroxyethyl, preferably a methyl.

m a number from 1 to 4, in particular 1, 2 or 3,

y 0 or 1 and

Y is COO, SO<sub>3</sub>, OPO(OR<sup>5</sup>)O or P(O)(OR<sup>5</sup>)O, whereby R<sup>5</sup> is a hydrogen atom H or a C1-4 alkyl residue.

**[0113]** Preferred betaines are the alkyl betaines of the formula (Ia), the alkyl amido propyl betaine of the formula (Ib), the Sulfo betaines of the formula (Ic) and the Amido sulfobetaine of the formula (Id);



**[0114]** R<sup>1</sup>-CO-NH-(CH<sub>2</sub>)<sub>3</sub>-N<sup>+</sup>(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>CH(OH)CH<sub>2</sub>SO<sub>3</sub><sup>-</sup> (Id) in which R<sup>1</sup> as the same meaning as in formula I. Particularly preferred betaines are the Carbobetaine [wherein Y<sup>-</sup>=COO<sup>-</sup>], in particular the Carbobetaine of the formula (Ia) and (Ib), more preferred are the Alkylamidobetaine of the formula (Ib).

**[0115]** Examples of suitable betaines and sulfobetaine are the following [designated in accordance with INCI]: Almondamidopropyl of betaines, Apricotamidopropyl betaines, Avocamidopropyl of betaines, Babassamidopropyl of betaines, Behenam idopropyl betaines, Behenyl of betaines, betaines, Canolamidopropyl betaines, Capryl/Capram idopropyl betaines, Carnitine, Cetyl of betaines, Cocamidoethyl of betaines, Cocamidopropyl betaines, Cocamidopropyl Hydroxysultaine, Coco betaines, Coco Hydroxysultaine, Coco/Oleamidopropyl betaines, Coco Sultaine, Decyl of betaines, Dihydroxyethyl Oleyl Glycinate, Dihydroxyethyl Soy Glycinate, Dihydroxyethyl Stearyl Glycinate, Dihydroxyethyl Tallow Glycinate, Dimethicone Propyl of PG-betaines, Erucamidopropyl Hydroxysultaine, Hydrogenated Tallow of betaines, Isostearamidopropyl betaines, Lauramidopropyl betaines, Lauryl of betaines, Lauryl Hydroxysultaine, Lauryl Sultaine, Milkamidopropyl betaines, Minkamidopropyl of betaines, Myristamidopropyl betaines, Myristyl of betaines, Oleamidopropyl betaines, Oleamidopropyl Hydroxysultaine, Oleyl of betaines, Olivamidopropyl of betaines, Palmamidopropyl betaines, Palm itamidopropyl betaines, Palmitoyl Carnitine, Palm Kernelamidopropyl betaines, Polytetrafluoroethylene Acetoxypropyl of betaines, Ricinoleamidopropyl betaines, Sesamidopropyl betaines, Soyamidopropyl betaines, Stearamidopropyl betaines, Stearyl of betaines, Tallowamidopropyl betaines, Tallowamidopropyl Hydroxysultaine, Tallow of betaines, Tallow Dihydroxyethyl of betaines, Undecylenamidopropyl betaines and Wheat Germamidopropyl betaines. A preferred betaine is, for example, Cocoamidopropylbetaine.

### Fatty Acid

**[0116]** Especially when in liquid form, preferably, the detergent composition comprises between 1.5% and 20%, more preferably between 2% and 15%, even more preferably between 3% and 10%, most preferably between 4% and 8% by weight of the liquid detergent composition of soap, preferably a fatty acid salt, more preferably an amine neutralized fatty acid salt, wherein preferably the amine is an alkanolamine more preferably selected from monoethanolamine, diethanolamine, triethanolamine or a mixture thereof, more preferably monoethanolamine.

Perfume

**[0117]** Preferred compositions of the invention comprise perfume. Typically the composition comprises a perfume that comprises one or more perfume raw materials, selected from the group as described in WO08/87497. However, any perfume useful in a detergent may be used. A preferred method of incorporating perfume into the compositions of the invention is via an encapsulated perfume particle comprising either a water-soluble hydroxylic compound or melamine-formaldehyde or modified polyvinyl alcohol. In one aspect the encapsulate comprises (a) an at least partially water-soluble solid matrix comprising one or more water-soluble hydroxylic compounds, preferably starch; and (b) a perfume oil encapsulated by the solid matrix. In a further aspect the perfume may be pre-complexed with a polyamine, preferably a polyethylenimine so as to form a Schiff base.

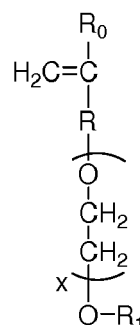
Polymers

**[0118]** The detergent composition may comprise one or more polymers for example for cleaning and/or care. Examples are optionally modified carboxymethylcellulose, poly (ethylene glycol), poly(vinyl alcohol), polycarboxylates such as polyacrylates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid co-polymers and carboxylate polymers.

**[0119]** Suitable carboxylate polymers include maleate/acrylate random copolymer or polyacrylate homopolymer. The carboxylate polymer may be a polyacrylate homopolymer having a molecular weight of from 4,000 Da to 9,000 Da, or from 6,000 Da to 9,000 Da. Other suitable carboxylate polymers are co-polymers of maleic acid and acrylic acid, and may have a molecular weight in the range of from 4,000 Da to 90,000 Da.

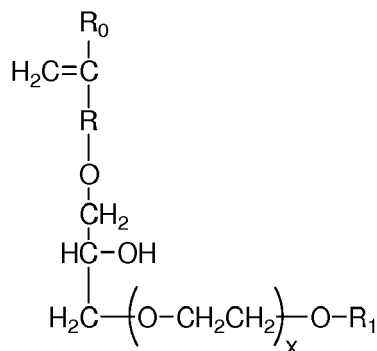
**[0120]** Other suitable carboxylate polymers are co-polymers comprising: (i) from 50 to less than 98 wt% structural units derived from one or more monomers comprising carboxyl groups; (ii) from 1 to less than 49 wt% structural units derived from one or more monomers comprising sulfonate moieties; and (iii) from 1 to 49 wt% structural units derived from one or more types of monomers selected from ether bond-containing monomers represented by formulas (I) and (II):

formula (I):



wherein in formula (I),  $R_0$  represents a hydrogen atom or  $CH_3$  group, R represents a  $CH_2$  group,  $CH_2CH_2$  group or single bond, X represents a number 0-5 provided X represents a number 1-5 when R is a single bond, and  $R_1$  is a hydrogen atom or C1 to C20 organic group;

formula (II)



in formula (II),  $R_0$  represents a hydrogen atom or  $\text{CH}_3$  group, R represents a  $\text{CH}_2$  group,  $\text{CH}_2\text{CH}_2$  group or single bond, X represents a number 0-5, and  $R_1$  is a hydrogen atom or C1 to C20 organic group.

**[0121]** The composition may comprise one or more amphiphilic cleaning polymers such as the compound having the following general structure:  $\text{bis}((\text{C}_2\text{H}_5\text{O})(\text{C}_2\text{H}_4\text{O})_n)(\text{CH}_3)\text{-N}^+\text{-C}_x\text{H}_{2x}\text{-N}^+\text{-}(\text{CH}_3)\text{-bis}((\text{C}_2\text{H}_5\text{O})(\text{C}_2\text{H}_4\text{O})_n)$ , wherein  $n =$  from 20 to 30, and  $x =$  from 3 to 8, or sulphated or sulphonated variants thereof. In one aspect, this polymer is sulphated or sulphonated to provide a zwitterionic soil suspension polymer.

**[0122]** The composition preferably comprises amphiphilic alkoxyated grease cleaning polymers which have balanced hydrophilic and properties such that they remove grease particles from fabrics and surfaces. Preferred amphiphilic alkoxyated grease cleaning polymers comprise a core structure and a plurality of alkoxyate groups attached to that core structure. These may comprise alkoxyated polyalkylenimines, preferably having an inner polyethylene oxide block and an outer polypropylene oxide block. Typically these may be incorporated into the compositions of the invention in amounts of from 0.005 to 10 wt%, generally from 0.5 to 8 wt%.

**[0123]** Alkoxyated polycarboxylates such as those prepared from polyacrylates are useful herein to provide additional grease removal performance. Such materials are described in WO 91/08281 and PCT 90/01815. Chemically, these materials comprise polyacrylates having one ethoxy side-chain per every 7-8 acrylate units. The side-chains are of the formula  $\text{-(CH}_2\text{CH}_2\text{O)}_m(\text{CH}_2)_n\text{CH}_3$  wherein  $m$  is 2-3 and  $n$  is 6-12. The side-chains are ester-linked to the polyacrylate "backbone" to provide a "comb" polymer type structure. The molecular weight can vary, but is typically in the range of about 2000 to about 50,000. Such alkoxyated polycarboxylates can comprise from about 0.05% to about 10%, by weight, of the compositions herein.

**[0124]** The composition may comprise polyethylene glycol polymers and these may be particularly preferred in compositions comprising mixed surfactant systems. Suitable polyethylene glycol polymers include random graft co-polymers comprising: (i) hydrophilic backbone comprising polyethylene glycol; and (ii) side chain(s) selected from the group consisting of: C4-C25 alkyl group, polypropylene, polybutylene, vinyl ester of a saturated C1-C6 mono-carboxylic acid, C1-C6 alkyl ester of acrylic or methacrylic acid, and mixtures thereof. Suitable polyethylene glycol polymers have a polyethylene glycol backbone with random grafted polyvinyl acetate side chains. The average molecular weight of the polyethylene glycol backbone can be in the range of from 2,000 Da to 20,000 Da, or from 4,000 Da to 8,000 Da. The molecular weight ratio of the polyethylene glycol backbone to the polyvinyl acetate side chains can be in the range of from 1:1 to 1:5, or from 1:1.2 to 1:2. The average number of graft sites per ethylene oxide units can be less than 1, or less than 0.8, the average number of graft sites per ethylene oxide units can be in the range of from 0.5 to 0.9, or the average number of graft sites per ethylene oxide units can be in the range of from 0.1 to 0.5, or from 0.2 to 0.4. A suitable polyethylene glycol polymer is Sokalan HP22.

**[0125]** Typically these polymers when present are each incorporated into the compositions of the invention in amounts from 0.005 to 10 wt%, more usually from 0.05 to 8 wt%.

**[0126]** Preferably the composition comprises one or more carboxylate polymer, such as a maleate/acrylate random copolymer or polyacrylate homopolymer. In one aspect, the carboxylate polymer is a polyacrylate homopolymer having a molecular weight of from 4,000 Da to 9,000 Da, or from 6,000 Da to 9,000 Da. Typically these are incorporated into the compositions of the invention in amounts from 0.005 to 10 wt%, or from 0.05 to 8 wt%.

**[0127]** Preferably the composition comprises one or more soil release polymers.

**[0128]** Suitable soil release polymers are polyester soil release polymers such as Repel-o-tex polymers, including Repel-o-tex SF, SF-2 and SRP6 supplied by Rhodia. Other suitable soil release polymers include Texcare polymers, including Texcare SRA100, SRA300, SRN100, SRN170, SRN240, SRN260, SRN300 and SRN325 supplied by Clariant. Other suitable soil release polymers are Marloquest polymers, such as Marloquest SL supplied by Sasol.

**[0129]** Preferably the composition comprises one or more cellulosic polymer, including those selected from alkyl cellulose, alkyl alkoxyalkyl cellulose, carboxyalkyl cellulose, alkyl carboxyalkyl cellulose. Preferred cellulosic polymers are selected from the group comprising carboxymethyl cellulose, methyl cellulose, methyl hydroxyethyl cellulose, methyl carboxymethyl cellulose, and mixtures thereof. In one aspect, the carboxymethyl cellulose has a degree of carboxymethyl substitution from 0.5 to 0.9 and a molecular weight from 100,000 Da to 300,000 Da.

**[0130]** The composition preferably comprises a cationically-modified polysaccharide polymer. Preferably, the cationic polysaccharide polymer is selected from cationically modified hydroxyethyl cellulose, cationically modified hydroxypropyl cellulose, cationically and hydrophobically modified hydroxyethyl cellulose, cationically and hydrophobically modified hydroxypropyl cellulose, or a mixture thereof, more preferably cationically modified hydroxyethyl cellulose, cationically and hydrophobically modified hydroxyethyl cellulose, or a mixture thereof.

#### Amines

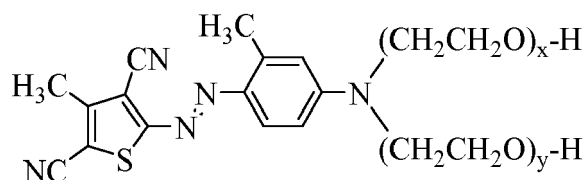
**[0131]** The cleaning compositions described herein may contain an amine. The cleaning compositions may include from about 0.1% to about 10%, or from about 0.2% to about 5%, or from about 0.5% to about 4%, or from about 0.1% to about 4%, or from about 0.1% to about 2%, by weight of the composition, of an amine. The amine can be subjected



to protonation depending on the pH of the cleaning medium in which it is used. Non-limiting examples of amines include, but are not limited to, etheramines, cyclic amines, polyamines, oligoamines (e.g., triamines, diamines, pentamines, tetraamines), or combinations thereof. The compositions described herein may comprise an amine selected from the group consisting of oligoamines, etheramines, cyclic amines, and combinations thereof. In some aspects, the amine is not an alkanolamine. In some aspects, the amine is not a polyalkyleneimine. Examples of suitable oligoamines include tetraethylenepentamine, triethylenetetraamine, diethylenetriamine, and mixtures thereof. Etheramines and cyclic amines may be particularly preferred.

#### Fabric Shading Dye

**[0132]** The composition may comprise a fabric shading agent. Suitable fabric shading agents include dyes, dye-clay conjugates, and pigments. Suitable dyes include small molecule dyes and polymeric dyes. Suitable small molecule dyes include small molecule dyes selected from the group consisting of dyes falling into the Colour Index (C.I.) classifications of Direct Blue, Direct Red, Direct Violet, Acid Blue, Acid Red, Acid Violet, Basic Blue, Basic Violet and Basic Red, or mixtures thereof. Preferred dyes include alkoxyated azothiophenes, Solvent Violet 13, Acid Violet 50 and Direct Violet 9. Particularly preferred dyes are polymeric dyes, particularly comprising polyalkoxy, most preferably polyethoxy groups, for example:



wherein the index values x and y are independently selected from 1 to 10.

#### Dye Transfer Inhibitors

**[0133]** Suitable dye transfer inhibitors include polyamine N-oxide polymers, copolymers of N-vinylpyrrolidone and N-vinylimidazole, polyvinylpyrrolidone, polyvinylloxazolidone, polyvinylimidazole and mixtures thereof. Preferred are poly(vinyl pyrrolidone), poly(vinylpyridine betaine), poly(vinylpyridine N-oxide), poly(vinyl pyrrolidone-vinyl imidazole) and mixtures thereof. Suitable commercially available dye transfer inhibitors include PVP-K15 and K30 (Ashland), Sokalan® HP165, HP50, HP53, HP59, HP56K, HP56, HP66 (BASF), Chromabond® S-400, S403E and S-100 (Ashland).

#### Chelant

**[0134]** The composition may comprise chelant for example selected from phosphonic, sulphonic, succinic and acetic chelants or mixtures thereof. Suitable examples include HEDP, DTPA, EDTA, MGDA, GLDA, EDDS and 4,5-dihydroxy-1,3-benzenedisulfonic acids and salts thereof.

#### Encapsulated Benefit Agent

**[0135]** The composition may further comprise an encapsulated benefit agent. The encapsulated benefit may comprise a shell surrounding a core. The core may comprise a benefit agent. The benefit agent may comprise perfume raw materials.

**[0136]** The shell may comprise a material selected from the group consisting of aminoplast copolymer, an acrylic, an acrylate, and mixtures thereof. The aminoplast copolymer may be melamine-formaldehyde, urea-formaldehyde, cross-linked melamine formaldehyde, or mixtures thereof.

**[0137]** The shell may be coated with one or more materials, such as a polymer, that aids in the deposition and/or retention of the perfume microcapsule on the site that is treated with the composition disclosed herein. The polymer may be a cationic polymer selected from the group consisting of polysaccharides, cationically modified starch, cationically modified guar, polysiloxanes, poly diallyl dimethyl ammonium halides, copolymers of poly diallyl dimethyl ammonium chloride and vinyl pyrrolidone, acrylamides, imidazoles, imidazolium halides, imidazolium halides, poly vinyl amine, copolymers of poly vinyl amine and N-vinyl formamide, and mixtures thereof.

**[0138]** The core may comprise a benefit agent. Suitable benefit agents include a material selected from the group consisting of perfume raw materials, silicone oils, waxes, hydrocarbons, higher fatty acids, essential oils, lipids, skin coolants, vitamins, sunscreens, antioxidants, glycerine, catalysts, bleach particles, silicon dioxide particles, malodor

reducing agents, odor-controlling materials, chelating agents, antistatic agents, softening agents, insect and moth repellent agents, colorants, antioxidants, chelants, bodying agents, drape and form control agents, smoothness agents, wrinkle control agents, sanitization agents, disinfecting agents, germ control agents, mold control agents, mildew control agents, antiviral agents, drying agents, stain resistance agents, soil release agents, fabric refreshing agents and freshness extending agents, chlorine bleach odor control agents, dye fixatives, dye transfer inhibitors, color maintenance agents, optical brighteners, color restoration/rejuvenation agents, anti-fading agents, whiteness enhancers, anti-abrasion agents, wear resistance agents, fabric integrity agents, anti-wear agents, anti-pilling agents, defoamers, anti-foaming agents, UV protection agents, sun fade inhibitors, anti-allergenic agents, enzymes, water proofing agents, fabric comfort agents, shrinkage resistance agents, stretch resistance agents, stretch recovery agents, skin care agents, glycerin, and natural actives, antibacterial actives, antiperspirant actives, cationic polymers, dyes and mixtures thereof. The benefit agent may comprise perfume raw materials.

**[0139]** The composition may comprise, based on total composition weight, from about 0.01% to about 10%, or from about 0.1% to about 5%, or from about 0.2% to about 1%, of encapsulated benefit agent. The encapsulated benefit agent may be friable and/or have a mean particle size of from about 10 microns to about 500 microns or from about 20 microns to about 200 microns.

**[0140]** Suitable encapsulated benefit agents may be obtained from Encapsys, LLC, of Appleton, Wisconsin USA.

**[0141]** Formaldehyde scavengers may also be used in or with such encapsulated benefit agents.

**[0142]** In a further preferred aspect of the invention, the composition is preferably liquid and comprises particulate benefit agents such as the encapsulated benefit agents mentioned above. The combination of the galactanase enzyme in addition to the plant fiber and particulate benefit agent has been found to provide the additional benefit of enhanced deposition of the particulate benefit agent. Thus, the present invention also provides a method of enhancing deposition of a particulate benefit agent comprising contacting a textile with an aqueous liquor comprising a composition defined herein, comprising a galactanase enzyme and a plant fiber and in addition a particulate benefit agent in a textile treatment step, preferably a laundering step, and optionally rinsing and drying the textile. In a preferred method, the aqueous liquor is an aqueous wash liquor. In a preferred method, the particulate benefit agent comprises an encapsulated perfume particle, most preferably comprising a shell which comprises a material selected from the group consisting of polymers or copolymers comprising acrylic acid and/or acrylates, and mixtures thereof.

#### Methods of Making the Composition

#### Methods of Making the Composition

**[0143]** The present invention relates to methods of making the compositions described herein. The compositions of the invention may be solid (for example granules or tablets) or liquid form. Preferably the compositions are in liquid form. They may be made by any process chosen by the formulator, including by a batch process, a continuous loop process, or combinations thereof.

**[0144]** When in the form of a liquid, the compositions of the invention may be aqueous (typically above 2 wt% or even above 5 or 10 wt% total water, up to 90 or up to 80wt% or 70 wt% total water) or non-aqueous (typically below 2 wt% total water content). Typically the compositions of the invention will be in the form of an aqueous solution or uniform dispersion or suspension of optical brightener, DTI and optional additional adjunct materials, some of which may normally be in solid form, that have been combined with the normally liquid components of the composition, such as the liquid alcohol ethoxylate nonionic, the aqueous liquid carrier, and any other normally liquid optional ingredients. Such a solution, dispersion or suspension will be acceptably phase stable. When in the form of a liquid, the detergents of the invention preferably have viscosity from 1 to 1500 centipoises (1-1500 mPa\*s), more preferably from 100 to 1000 centipoises (100-1000 mPa\*s), and most preferably from 200 to 500 centipoises (200-500 mPa\*s) at 20s<sup>-1</sup> and 21°C. Viscosity can be determined by conventional methods. Viscosity may be measured using an AR 550 rheometer from TA instruments using a plate steel spindle at 40 mm diameter and a gap size of 500 μm. The high shear viscosity at 20s<sup>-1</sup> and low shear viscosity at 0.05-1 can be obtained from a logarithmic shear rate sweep from 0.1-1 to 25-1 in 3 minutes time at 21°C. The preferred rheology described therein may be achieved using internal existing structuring with detergent ingredients or by employing an external rheology modifier. More preferably the detergents, such as detergent liquid compositions have a high shear rate viscosity of from about 100 centipoise to 1500 centipoise, more preferably from 100 to 1000 cps. Unit Dose detergents, such as detergent liquid compositions have high shear rate viscosity of from 400 to 1000cps. Detergents such as laundry softening compositions typically have high shear rate viscosity of from 10 to 1000, more preferably from 10 to 800 cps, most preferably from 10 to 500 cps. Hand dishwashing compositions have high shear rate viscosity of from 300 to 4000 cps, more preferably 300 to 1000 cps.

**[0145]** The cleaning and/or treatment compositions in the form of a liquid herein can be prepared by combining the components thereof in any convenient order and by mixing, e.g., agitating, the resulting component combination to form a phase stable liquid detergent composition. In a process for preparing such compositions, a liquid matrix is formed

containing at least a major proportion, or even substantially all, of the liquid components, e.g., nonionic surfactant, the non-surface active liquid carriers and other optional liquid components, with the liquid components being thoroughly admixed by imparting shear agitation to this liquid combination. For example, rapid stirring with a mechanical stirrer may usefully be employed. While shear agitation is maintained, substantially all of any anionic surfactants and the solid form ingredients can be added. Agitation of the mixture is continued, and if necessary, can be increased at this point to form a solution or a uniform dispersion of insoluble solid phase particulates within the liquid phase. After some or all of the solid-form materials have been added to this agitated mixture, particles of any enzyme material to be included, e.g., enzyme granulates, are incorporated. As a variation of the composition preparation procedure hereinbefore described, one or more of the solid components may be added to the agitated mixture as a solution or slurry of particles premixed with a minor portion of one or more of the liquid components. After addition of all of the composition components, agitation of the mixture is continued for a period of time sufficient to form compositions having the requisite viscosity and phase stability characteristics. Frequently this will involve agitation for a period of from about 30 to 60 minutes.

**[0146]** The adjunct ingredients in the compositions of this invention may be incorporated into the composition as the product of the synthesis generating such components, either with or without an intermediate purification step. Where there is no purification step, commonly the mixture used will comprise the desired component or mixtures thereof (and percentages given herein relate to the weight percent of the component itself unless otherwise specified) and in addition unreacted starting materials and impurities formed from side reactions and/or incomplete reaction. For example, for an ethoxylated or substituted component, the mixture will likely comprise different degrees of ethoxylation/substitution.

#### Method of Use

**[0147]** The present invention relates to methods of using the cleaning compositions of the present invention to clean a surface, such as a textile. In general, the method includes mixing the cleaning composition as described herein with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step. The target surface may include a greasy soil such as a body soil. The compositions herein, typically prepared as hereinbefore described, can be used to form aqueous washing/treatment solutions for use in the laundering/treatment of fabrics and/or hard surfaces. Generally, an effective amount of such a composition is added to water, for example in a conventional fabric automatic washing machine, to form such aqueous liquor laundering solutions. The aqueous liquor so formed is then contacted, typically under agitation, with the fabrics to be laundered/treated therewith. An effective amount of the cleaning composition herein added to water to form aqueous liquors for washing can comprise amounts sufficient to form from about 500 to 25,000 ppm, or from 500 to 15,000 ppm of composition in aqueous liquor, or from about 1,000 to 3,000 ppm of the cleaning compositions herein will be provided in aqueous liquor.

**[0148]** Typically, the aqueous liquor is formed by contacting the detergent with wash water in such an amount so that the concentration of the cleaning composition in the aqueous liquor is from above 0.1 g/l to 5g/l, or from 1g/l, and to 4.5g/l, or to 4.0g/l, or to 3.5g/l, or to 3.0g/l, or to 2.5g/l, or even to 2.0g/l, or even to 1.5g/l. The method of laundering fabric or textile may be carried out in a top-loading or front-loading automatic washing machine, or can be used in a hand-wash laundry application. In these applications, the aqueous liquor formed and concentration of laundry detergent composition in the aqueous liquor is that of the main wash cycle. Any input of water during any optional rinsing step(s) is not included when determining the volume of the aqueous liquor.

**[0149]** The aqueous liquor may comprise 40 litres or less of water, or 30 litres or less, or 20 litres or less, or 10 litres or less, or 8 litres or less, or even 6 litres or less of water. The wash liquor may comprise from above 0 to 15 litres, or from 2 litres, and to 12 litres, or even to 8 litres of water. Typically from 0.01kg to 2kg of fabric per litre of aqueous liquor is dosed into said aqueous liquor. Typically from 0.01kg, or from 0.05kg, or from 0.07kg, or from 0.10kg, or from 0.15kg, or from 0.20kg, or from 0.25kg fabric per litre of aqueous liquor is dosed into said aqueous liquor. Optionally, 50g or less, or 45g or less, or 40g or less, or 35g or less, or 30g or less, or 25g or less, or 20g or less, or even 15g or less, or even 10g or less of the composition is contacted to water to form the aqueous liquor. Such compositions are typically employed at concentrations of from about 500 ppm to about 15,000 ppm in solution. When the wash solvent is water, the water temperature typically ranges from about 5 °C to about 90 °C and, when the situs comprises a fabric, the water to fabric ratio is typically from about 1:1 to about 30:1. Typically the aqueous liquor comprising the detergent of the invention has a pH of from 3 to 11.5.

**[0150]** In one aspect, such method comprises the steps of optionally washing and/or rinsing said surface or fabric, contacting said surface or fabric with any composition disclosed in this specification then optionally washing and/or rinsing said surface or fabric is disclosed, with an optional drying step.

**[0151]** Drying of such surfaces or fabrics may be accomplished by any one of the common means employed either in domestic or industrial settings: machine drying or open-air drying. The fabric may comprise any fabric capable of being laundered in normal consumer or institutional use conditions, and the invention is particularly suitable for synthetic textiles such as polyester and nylon and especially for treatment of mixed fabrics and/or fibres comprising synthetic and cellulosic fabrics and/or fibres. As examples of synthetic fabrics are polyester, nylon, these may be present in mixtures

with cellulosic fibres, for example, polycotton fabrics. The solution typically has a pH of from 7 to 11, more usually 8 to 10.5. The compositions are typically employed at concentrations from 500 ppm to 5,000 ppm in solution. The water temperatures typically range from about 5 °C to about 90 °C. The water to fabric ratio is typically from about 1:1 to about 30:1.

5  
Use of Water-Insoluble Plant Fiber

10  
[0152] The present invention further relates to a use of water-insoluble plant fiber in a cleaning composition to enhance the stain-removal and/or malodor-reducing benefits of a galactanase enzyme or composition comprising a galactanase enzyme.

Measurement Methods for Fibers

15  
Sample preparation:

[0153]

20  
A) Cellulose fibers raw material: A cellulose fibers sample is prepared by adding 1% dry matter of cellulose fibers to water and activating it with a high pressure homogenizer (PANDA from GEA, 350 bars, 10 passes). Obtained sample is analyzed.

B) Composition comprising cellulose fibers:

25  
The composition sample is centrifuged at 4,000 rpm for 10 minutes using a 5804 centrifuge from Eppendorf, in order to remove potential particles to avoid interference in the measurement of the fiber size. The clarified composition is then decanted as the supernatant. The water insoluble plant derived fibers present in the composition (supernatant) are redispersed in ethanol using an Ultra Turrax device from IKA, T25 S 25 N - 25 G - ST, at a speed of 21,000rpm for 10 minutes. Then, sample is centrifuged at 4,000 rpm for 10 minutes using a 5804 centrifuge from Eppendorf and supernatant is removed. Remaining cellulose fibers at the bottom are analyzed. Repeat the process as many times as needed to have enough amount for the analysis.

30  
Measuring hydrodynamic diameter:

35  
[0154] The instrument cell is cleaned and then filled with demineralised water. If the background has a laser intensity above 79%, the system is considered clean and the sample can be added to the vessel until the desired obscuration is achieved. Then ultrasounds are switched on for 30 seconds and once the sample is well dispersed, the measurement can start.

[0155] Then, the hydrodynamic diameter (volume weight mean [4,3]) is measured. The hydrodynamic diameter is the diameter of the equivalent sphere that has the same translational diffusion coefficient as the fiber being measured assuming a hydration layer surrounding the fiber.

40  
Sampler selection: Hydro 2000MU

Sampler settings: Pump/stir speed: 2500rpm; Ultrasonics: 30 seconds

Material: Refractive Index of the material: 1.53; Dispersant used: demineralised water in an amount as needed; Particle shape: Irregular.

45  
Measurement: Measurement cycles: 3 measurements per aliquot with a delay of 10 seconds, Measurement time: 10 seconds; Measurement snaps: 10,000; Background time: 10 seconds; o Background snaps: 10,000; Lower obscuration limit: 5; Upper obscuration limit: 15

50  
Method for determining average cellulose fiber diameter:

[0156] The average water-insoluble plant derived fiber diameter can be determined directly from the cellulose fiber raw material or from the composition comprising cellulose fibers following sample preparation described above.

55  
[0157] Water-insoluble plant derived fiber diameter is analysed using Atomic force microscopy (AFM). A 0,02% w/w cellulose fiber dispersion in demineralized water is prepared, and a drop of this dispersion is deposited onto freshly cleaved mica (highest grade V1 Mica, 15x15mm - TED PELLA, INC., or equivalent). The sample is then allowed to dry in an oven at 40°C.

[0158] The mica sheet is mounted in an AFM (Nanosurf Flex AFM, ST Instruments or equivalent) and imaged in air under ambient conditions using a Si cantilever in dynamic mode with dynamic mode tip (ACTA -50 - APPNANO or

**EP 3 330 351 A1**

equivalent). The image dimensions are 20 micron by 20 micron, and 256 points per line are captured.

**[0159]** The AFM image is opened using suitable AFM data analysis software (such as Mountainsmap SPM 7.3, ST Instruments, or equivalent). Each image is leveled line by line. One or more profiles are extracted crossing perpendicularly one or multiple fibers avoiding bundles of fibers, and from each profile, a distance measurement is performed to obtain the diameter of the fibers. Ten diameter measurements are performed per picture counting each fiber only once.

**[0160]** Three sets of measurements (sample preparation, AFM measurement and image analysis) are made. The arithmetic mean of all fibers measured in all images is the Average water-insoluble plant derived fiber Diameter.

**EXAMPLES**

**[0161]** The following are illustrative examples of cleaning compositions according to the present invention and are not intended to be limiting.

Examples 1-7: Heavy Duty Liquid laundry detergent compositions.

**[0162]**

<u>Ingredients</u>	1	2	3	4	5	6	7
	% weight						
AE <sub>1,8</sub> S	6.77	5.16	1.36	1.30	-	-	-
AE <sub>3</sub> S	-	-	-	-	0.45	-	-
LAS	0.86	2.06	2.72	0.68	0.95	1.56	3.55
HSAS	1.85	2.63	1.02	-	-	-	-
AE9	6.32	9.85	10.20	7.92			
AE8							35.45
AE7					8.40	12.44	
C <sub>12-14</sub> dimethyl Amine Oxide	0.30	0.73	0.23	0.37	-	-	-
C <sub>12-18</sub> Fatty Acid	0.80	1.90	0.60	0.99	1.20	-	15.00
Citric Acid	2.50	3.96	1.88	1.98	0.90	2.50	0.60
Optical Brightener 1	1.00	0.80	0.10	0.30	0.05	0.50	0.001
Optical Brightener 3	0.001	0.05	0.01	0.20	0.50	-	1.00
Sodium formate	1.60	0.09	1.20	0.04	1.60	1.20	0.20
DTI 1	0.32	0.05	-	0.60	0.10	0.60	0.01
DTI 2	0.32	0.10	0.60	0.60	0.05	0.40	0.20
Sodium hydroxide	2.30	3.80	1.70	1.90	1.70	2.50	2.30
Monoethanolamine	1.40	1.49	1.00	0.70	-	-	-
Diethylene glycol	5.50	-	4.10	-	-	-	-
Chelant 1	0.15	0.15	0.11	0.07	0.50	0.11	0.80
4-formyl-phenylboronic acid	-	-	-	-	0.05	0.02	0.01
Sodium tetraborate	1.43	1.50	1.10	0.75	-	1.07	-
Ethanol	1.54	1.77	1.15	0.89	-	3.00	7.00
Polymer 1	0.10	-	-	-	-	-	2.00
Polymer 2	0.30	0.33	0.23	0.17	-	-	-
Polymer 3	-	-	-	-	-	-	0.80
Polymer 4	0.80	0.81	0.60	0.40	1.00	1.00	-

**EP 3 330 351 A1**

(continued)

	<b>Ingredients</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>
		<b>% weight</b>						
5	1,2-Propanediol	-	6.60	-	3.30	0.50	2.00	8.00
	Structurant	0.10	-	-	-	-	-	0.10
	Perfume	1.60	1.10	1.00	0.80	0.90	1.50	1.60
10	Perfume encapsulate	0.10	0.05	0.01	0.02	0.10	0.05	0.10
	Protease	0.80	0.60	0.70	0.90	0.70	0.60	1.50
	Mannanase	0.07	-	-	0.06	0.04	0.045	0.10
15	Amylase 1	0.30	-	0.30	0.10	-	0.40	0.10
	Amylase 2	-	0.20	0.10	0.15	0.07	-	0.10
	Xyloglucannase	0.20	0.10	-	-	0.05	0.05	0.20
	Lipase	0.40	0.20	0.30	0.10	0.20	-	-
20	Polishing enzyme	-	0.04	-	-	-	0.004	-
	Galactanase	0.05	0.03	0.01	0.03	0.03	0.003	0.003
	Dispersin B	-	-	-	0.05	0.03	0.001	0.001
25	Acid Violet 50	0.05	-	-	-	-	-	0.005
	Direct Violet 9	-	-	-	-	-	0.05	-
	Violet DD	-	0.035	0.02	0.037	0.04	-	-
	Water insoluble plant fiber	0.2	0.6	0.2	0.03	1.2	0.3	0.3
30	Water, dyes & minors	<b>Balance</b>						
	pH	<b>8.2</b>						

**[0163]** Based on total cleaning and/or treatment composition weight. Enzyme levels are reported as raw material.

Examples 8 to 18: Unit Dose Compositions.

**[0164]** These examples provide various formulations for unit dose laundry detergents. Compositions 8 to 12 comprise a single unit dose compartment. The film used to encapsulate the compositions is polyvinyl alcohol-based film.

	<b>Ingredients</b>	<b>8</b>	<b>9</b>	<b>10</b>	<b>11</b>	<b>12</b>
		<b>% weight</b>				
45	LAS	19.09	16.76	8.59	6.56	3.44
	AE3S	1.91	0.74	0.18	0.46	0.07
	AE7	14.00	17.50	26.33	28.08	31.59
	Citric Acid	0.6	0.6	0.6	0.6	0.6
50	C12-15 Fatty Acid	14.8	14.8	14.8	14.8	14.8
	Polymer 3	4.0	4.0	4.0	4.0	4.0
	Chelant 2	1.2	1.2	1.2	1.2	1.2
55	Optical Brightener 1	0.20	0.25	0.01	0.01	0.50
	Optical Brightener 2	0.20	-	0.25	0.03	0.01
	Optical Brightener 3	0.18	0.09	0.30	0.01	-

EP 3 330 351 A1

(continued)

5  
10  
15  
20  
25  
30  
35

Ingredients	8	9	10	11	12
	% weight				
DTI 1	0.10	-	0.20	0.01	0.05
DTI 2	-	0.10	0.20	0.25	0.05
Glycerol	6.1	6.1	6.1	6.1	6.1
Monoethanol amine	8.0	8.0	8.0	8.0	8.0
Tri-isopropanol amine	-	-	2.0	-	-
Tri-ethanol amine	-	2.0	-	-	-
Cumene sulfonate	-	-	-	-	2.0
Protease	0.80	0.60	0.07	1.00	1.50
Mannanase	0.07	-	0.05	-	0.01
Amylase 1	0.20	0.11	0.30	0.50	0.05
Amylase 2	0.11	0.20	0.10	-	0.50
Polishing enzyme	0.005	0.05	-	-	-
Galactanase	0.005	0.05	0.005	0.010	0.005
Dispersin B	0.010	0.05	0.005	0.005	-
Cyclohexyl dimethanol	-	-	-	2.0	-
Acid violet 50	0.03	0.02			
Violet DD			0.01	0.05	0.02
Structurant	0.14	0.14	0.14	0.14	0.14
Perfume	1.9	1.9	1.9	1.9	1.9
Water insoluble plant fiber	0.02	0.3	0.02	0.03	0.3
Water and miscellaneous	To 100%				
pH	7.5-8.2				

[0165] Based on total cleaning and/or treatment composition weight. Enzyme levels are reported as raw material.

[0166] In the following examples the unit dose has three compartments, but similar compositions can be made with two, four or five compartments. The film used to encapsulate the compartments is polyvinyl alcohol.

45  
50  
55

Base compositions Ingredients	13	14	15	16
	% weight			
HLAS	26.82	16.35	7.50	3.34
AE7	17.88	16.35	22.50	30.06
Citric Acid	0.5	0.7	0.6	0.5
C12-15 Fatty acid	16.4	6.0	11.0	13.0
Polymer 1	2.9	0.1	-	-
Polymer 3	1.1	5.1	2.5	4.2
Cationic cellulose polymer	-	-	0.3	0.5
Polymer 6	-	1.5	0.3	0.2
Chelant 2	1.1	2.0	0.6	1.5

EP 3 330 351 A1

(continued)

5

10

15

20

25

30

<u>Base compositions Ingredients</u>	13	14	15	16
	% weight			
Optical Brightener 1	0.20	0.25	0.01	0.005
Optical Brightener 3	0.18	0.09	0.30	0.005
DTI 1	0.1	-	0.2	-
DTI 2	-	0.1	0.2	-
Glycerol	5.3	5.0	5.0	4.2
Monoethanolamine	10.0	8.1	8.4	7.6
Polyethylene glycol	-	-	2.5	3.0
Potassium sulfite	0.2	0.3	0.5	0.7
Protease	0.80	0.60	0.40	0.80
Amylase 1	0.20	0.20	0.200	0.30
Polishing enzyme	-	-	0.005	0.005
Galactanase	0.05	0.010	0.005	0.005
Dispersin B	-	0.010	0.010	0.010
MgCl <sub>2</sub>	0.2	0.2	0.1	0.3
Structurant	0.2	0.1	0.2	0.2
Acid Violet 50	0.04	0.03	0.05	0.03
Perfume / encapsulates	0.10	0.30	0.01	0.05
Water-insoluble plant fiber	0.2	0.03	0.4	2.0
Solvents and misc.	To 100%			
pH	7.0-8.2			

35

40

45

50

<u>Finishing compositions</u>	17			18		
Compartment	A	B	c	A	B	c
Volume of each compartment	40 ml	5 ml	5 ml	40 ml	5 ml	5 ml
<u>Ingredients</u>	Active material in Wt. %					
Perfume	1.6	1.6	1.6	1.6	1.6	1.6
Violet DD	0	0.006	0	0	0.004	-
TiO <sub>2</sub>	-	-	0.1	-		0.1
Sodium Sulfite	0.4	0.4	0.4	0.3	0.3	0.3
Polymer 5	-			2	-	-
Hydrogenated castor oil	0.14	0.14	0.14	0.14	0.14	0.14
Base Composition 13, 14, 15 or 16	Add to 100%					

[0167] Based on total cleaning and/or treatment composition weight, enzyme levels are reported as raw material.

55

Examples 19 to 24

[0168] Granular laundry detergent compositions for hand washing or washing machines, typically top-loading washing



EP 3 330 351 A1

machines.

5

10

15

20

25

30

35

40

45

50

Ingredient	19	20	21	22	23	24
	% weight					
LAS	11.33	10.81	7.04	4.20	3.92	2.29
Quaternary ammonium	0.70	0.20	1.00	0.60	-	-
AE3S	0.51	0.49	0.32	-	0.08	0.10
AE7	8.36	11.50	12.54	11.20	16.00	21.51
Sodium Tripolyphosphate	5.0	-	4.0	9.0	2.0	-
Zeolite A	-	1.0	-	1.0	4.0	1.0
Sodium silicate 1.6R	7.0	5.0	2.0	3.0	3.0	5.0
Sodium carbonate	20.0	17.0	23.0	14.0	14.0	16.0
Polyacrylate MW 4500	1.0	0.6	1.0	1.0	1.5	1.0
Polymer 6	0.1	0.2	-	-	0.1	-
Carboxymethyl cellulose	1.0	0.3	1.0	1.0	1.0	1.0
Acid Violet 50	0.05	-	0.02	-	0.04	-
Violet DD	-	0.03	-	0.03	-	0.03
Protease 2	0.10	0.10	0.10	0.10	-	0.10
Amylase	0.03	-	0.03	0.03	0.03	0.03
Lipase	0.03	0.07	0.30	0.10	0.07	0.40
Polishing enzyme	0.002	-	0.05	-	0.02	-
Galactanase	0.001	0.001	0.01	0.05	0.002	0.02
Dispersin B	0.001	0.001	0.05	-	0.001	-
Optical Brightener 1	0.200	0.001	0.300	0.650	0.050	0.001
Optical Brightener 2	0.060	-	0.650	0.180	0.200	0.060
Optical Brightener 3	0.100	0.060	0.050	-	0.030	0.300
Chelant 1	0.60	0.80	0.60	0.25	0.60	0.60
DTI 1	0.32	0.15	0.15	-	0.10	0.10
DTI 2	0.32	0.15	0.30	0.30	0.10	0.20
Sodium Percarbonate	-	5.2	0.1	-	-	-
Sodium Perborate	4.4	-	3.85	2.09	0.78	3.63
Nonanoyloxybenzenesulfonate	1.9	0.0	1.66	0.0	0.33	0.75
Tetraacetylenediamine	0.58	1.2	0.51	0.0	0.015	0.28
Photobleach	0.0030	0.0	0.0012	0.0030	0.0021	-
S-ACMC	0.1	0.0	0.0	0.0	0.06	0.0
Water-insoluble plant fiber	1.5	3.2	2.4	2.2	2.8	0.9
Sulfate/Moisture	Balance					

55 Examples 25-30

**[0169]** Granular laundry detergent compositions typically for front-loading automatic washing machines.

EP 3 330 351 A1

	Ingredient	25	26	27	28	29	30
		% weight					
5	LAS	6.08	5.05	4.27	3.24	2.30	1.09
	AE3S	-	0.90	0.21	0.18	-	0.06
	AS	0.34	-	-	-	-	-
10	AE7	4.28	5.95	6.72	7.98	9.20	10.35
	Quaternary ammonium	0.5	-	-	0.3	-	-
	Crystalline layered silicate	4.1	-	4.8	-	-	-
	Zeolite A	5.0	-	2.0	-	2.0	2.0
15	Citric acid	3.0	4.0	3.0	4.0	2.5	3.0
	Sodium carbonate	11.0	17.0	12.0	15.0	18.0	18.0
	Sodium silicate 2R	0.08	-	0.11	-	-	-
20	Optical Brightener 1	-	0.25	0.05	0.01	0.10	0.02
	Optical Brightener 2	-	-	0.25	0.20	0.01	0.08
	Optical Brightener 3	-	0.06	0.04	0.15	-	0.05
	DTI 1	0.08	-	0.04	-	0.10	0.01
25	DTI 2	0.08	-	0.04	0.10	0.10	0.02
	Soil release agent	0.75	0.72	0.71	0.72	-	-
	Acrylic /maleic acid copolymer	1.1	3.7	1.0	3.7	2.6	3.8
30	Carboxymethyl cellulose	0.2	1.4	0.2	1.4	1.0	0.5
	Protease 3	0.20	0.20	0.30	0.15	0.12	0.13
	Amylase 3	0.20	0.15	0.20	0.30	0.15	0.15
	Lipase	0.05	0.15	0.10	-	-	-
35	Amylase 2	0.03	0.07	-	-	0.05	0.05
	Cellulase 2	-	-	-	-	0.10	0.10
	Polishing enzyme	0.003	0.005	0.020	-	-	-
40	Galactanase	0.002	0.010	0.020	0.020	0.010	0.003
	Dispersin B	0.002	0.010	0.020	0.020	0.010	0.002
	Tetraacetylenehtylenediamine	3.6	4.0	3.6	4.0	2.2	1.4
45	Sodium percarbonate	13.0	13.2	13.0	13.2	16.0	14.0
	Chelant 3	-	0.2	-	0.2	-	0.2
	Chelant 2	0.2	-	0.2	-	0.2	0.2
	MgSO <sub>4</sub>	-	0.42	-	0.42	-	0.4
50	Perfume	0.5	0.6	0.5	0.6	0.6	0.6
	Suds suppressor agglomerate	0.05	0.10	0.05	0.10	0.06	0.05
	Soap	0.45	0.45	0.45	0.45	-	-
55	Acid Violet 50	0.04	-	0.05	-	0.04	-
	Violet DD	-	0.04	-	0.05	-	0.04
	S-ACMC	0.01	0.01	-	0.01	-	-

EP 3 330 351 A1

(continued)

Ingredient	25	26	27	28	29	30
	% weight					
Direct Violet 9 (active)	-	-	0.0001	0.0001	-	-
Water-insoluble plant fiber	1.23	2.2	0.87	4.4	2.6	2.8
Sulfate/ Water & Miscellaneous	Balance					

Examples 31-37: Heavy Duty Liquid laundry detergent compositions.

[0170]

Ingredients	31	32	33	34	35	36	37
	% weight						
AE <sub>1.8</sub> S	6.77	5.16	1.36	1.30	-	-	-
AE <sub>3</sub> S	-	-	-	-	0.45	-	-
LAS	0.86	2.06	2.72	0.68	0.95	1.56	3.55
HSAS	1.85	2.63	1.02	-	-	-	-
AE9	6.32	9.85	10.20	7.92			
AE8							35.45
AE7					8.40	12.44	
C <sub>12-14</sub> dimethyl Amine Oxide	0.30	0.73	0.23	0.37	-	-	-
C <sub>12-18</sub> Fatty Acid	0.80	1.90	0.60	0.99	1.20	-	15.00
Citric Acid	2.50	3.96	1.88	1.98	0.90	2.50	0.60
Optical Brightener 1	1.00	0.80	0.10	0.30	0.05	0.50	0.001
Optical Brightener 3	0.001	0.05	0.01	0.20	0.50	-	1.00
Sodium formate	1.60	0.09	1.20	0.04	1.60	1.20	0.20
DTI 1	0.32	0.05	-	0.60	0.10	0.60	0.01
DTI 2	0.32	0.10	0.60	0.60	0.05	0.40	0.20
Sodium hydroxide	2.30	3.80	1.70	1.90	1.70	2.50	2.30
Monoethanolamine	1.40	1.49	1.00	0.70	-	-	-
Diethylene glycol	5.50	-	4.10	-	-	-	-
Chelant 1	0.15	0.15	0.11	0.07	0.50	0.11	0.80
4-formyl-phenylboronic acid	-	-	-	-	0.05	0.02	0.01
Sodium tetraborate	1.43	1.50	1.10	0.75	-	1.07	-
Ethanol	1.54	1.77	1.15	0.89	-	3.00	7.00
Polymer 1	0.10	-	-	-	-	-	2.00
Polymer 2	0.30	0.33	0.23	0.17	-	-	-
Polymer 3	-	-	-	-	-	-	0.80
Polymer 4	0.80	0.81	0.60	0.40	1.00	1.00	-
1,2-Propanediol	-	6.60	-	3.30	0.50	2.00	8.00
Structurant	0.10	-	-	-	-	-	0.10

EP 3 330 351 A1

(continued)

Ingredients	31	32	33	34	35	36	37
	% weight						
Perfume	1.60	1.10	1.00	0.80	0.90	1.50	1.60
Perfume encapsulate	0.10	0.05	0.01	0.02	0.10	0.05	0.10
Protease	0.80	0.60	0.70	0.90	0.70	0.60	1.50
Galactanase of any of SEQ ID Nos: 1-3	0.07	0.05	0.045	0.06	0.04	0.045	0.10
Amylase 1	0.30	-	0.30	0.10	-	0.40	0.10
Amylase 2	-	0.20	0.10	0.15	0.07	-	0.10
Xyloglucanase	0.20	0.10	-	-	0.05	0.05	0.20
Lipase	0.40	0.20	0.30	0.10	0.20	-	-
Polishing enzyme	-	0.04	-	-	-	0.004	-
Nuclease	0.05	0.03	0.01	0.03	0.03	0.003	0.003
Dispersin B	-	-	-	0.05	0.03	0.001	0.001
Acid Violet 50	0.05	-	-	-	-	-	0.005
Direct Violet 9	-	-	-	-	-	0.05	-
Violet DD	-	0.035	0.02	0.037	0.04	-	-
Water insoluble plant fiber	0.2	0.1	0.3	0.25	1.2	1.5	0.25
Dye control agent	-	0.3	-	0.5	-	0.3	-
Alkoxylated polyaryl/ polyalkyl phenol	-	-	1.2	-	-	-	3.1
Water, dyes & minors	Balance						
pH	8.2						

Based on total cleaning and/or treatment composition weight. Unless indicated otherwise, enzyme levels are reported as raw material.

AE1.8S is C<sub>12-15</sub> alkyl ethoxy sulfate with an average degree of ethoxylation of 1.8  
 AE3S is C<sub>12-15</sub> alkyl ethoxy sulfate with an av degree of ethoxylation of 3  
 AE7 is C<sub>12-13</sub> alcohol ethoxylate, with an average degree of ethoxylation of 7  
 AE8 is C<sub>12-13</sub> alcohol ethoxylate, with an average degree of ethoxylation of 8  
 AE9 is C<sub>12-13</sub> alcohol ethoxylate, with an average degree of ethoxylation of 9  
 Alkoxylated polyaryl / polyalkyl phenol is alkoxylated polyaryl/polyalkyl phenol for example Emulsogen® TS160, Hostapal® BV conc., Sapogenat® T110 or Sapogenat® T139, all from Clariant

Amylase 1 is Stainzyme®, 15 mg active/g  
 Amylase 2 is Natalase®, 29 mg active/g  
 Amylase 3 is Stainzyme Plus®, 20 mg active/g,  
 AS is C<sub>12-14</sub> alkylsulfate  
 Cellulase 2 is Celluclean™, 15.6 mg active/g  
 Xyloglucanase is Whitezyme®, 20mg active/g  
 Chelant 1 is diethylene triamine pentaacetic acid  
 Chelant 2 is 1-hydroxyethane 1,1-diphosphonic acid  
 Chelant 3 is sodium salt of ethylenediamine-N,N'-disuccinic acid, (S,S) isomer (EDDS)  
 Dispersin B is a glycoside hydrolase, reported as 1000mg active/g  
 DTI 1 is poly(4-vinylpyridine-1-oxide) (such as Chromabond S-403E®),  
 DTI 2 is poly(1-vinylpyrrolidone-co-1-vinylimidazole) (such as Sokalan HP56®).  
 Dye Control Agent is for example Suparex® O.IN (M1), Nylofixan® P (M2), Nylofixan® PM (M3), or Nylofixan®

EP 3 330 351 A1

	HF (M4)
	is mid-branched alkyl sulfate as disclosed in US 6,020,303 and US6,060,443
	is linear alkylbenzenesulfonate having an average aliphatic carbon chain length C <sub>9</sub> -C <sub>15</sub> (HLAS is acid form).
5	Galactanase is SEQ ID NO: 1, 2 or 3, as active protein.
	Lipase is Lipex®, 18 mg active/g
	Mannanase is Mannaway®, 25 mg active/g
	Optical Brightener 1 is disodium 4,4'-bis[[4-anilino-6-morpholino-s-triazin-2-yl]-amino]-2,2'-stilbenedisulfonate
10	Optical Brightener 2 is disodium 4,4'-bis-(2-sulfostyryl)biphenyl (sodium salt)
	Optical Brightener 3 is Optiblanc SPL10® from 3V Sigma
	Perfume encapsulate is a core-shell melamine formaldehyde perfume microcapsules.
	Photobleach is a sulfonated zinc phthalocyanine
	Polishing enzyme is Para-nitrobenzyl esterase, reported as 1000mg active/g as described in present disclosure.
15	Polyetheramine is bis((C <sub>2</sub> H <sub>5</sub> O)(C <sub>2</sub> H <sub>4</sub> O) <sub>n</sub> )(CH <sub>3</sub> )-N <sup>+</sup> -C <sub>x</sub> H <sub>2x</sub> -N <sup>+</sup> -(CH <sub>3</sub> )-bis((C <sub>2</sub> H <sub>5</sub> O)(C <sub>2</sub> H <sub>4</sub> O) <sub>n</sub> ), wherein n = 20-30, x = 3 to 8 or sulphated or sulfonated variants thereof
	Polymer 2 is ethoxylated (EO <sub>15</sub> ) tetraethylene pentamine
	Polymer 3 is ethoxylated polyethylenimine
20	Polymer 4 is ethoxylated hexamethylene diamine
	Polymer 5 is Acusol 305, provided by Rohm&Haas
	Polymer 6 is a polyethylene glycol polymer grafted with vinyl acetate side chains, provided by BASF.
	Protease is Purafect Prime®, 40.6 mg active/g
	Protease 2 is Savinase®, 32.89 mg active/g
25	Protease 3 is Purafect®, 84 mg active/g
	Quaternary ammonium is C <sub>12-14</sub> Dimethylhydroxyethyl ammonium chloride
	S-ACMC is Reactive Blue 19 Azo-CM-Cellulose provided by Megazyme
	Soil release agent is Repel-o-tex® SF2
	Structurant is Hydrogenated Castor Oil
30	Violet DD is a thiophene azo dye provided by Milliken
	Water insoluble plant fiber Water insoluble plant fiber in accordance with the present disclosure, for example Herbacel AQ+ Type N, supplied by Herbafood Ingredients GmbH, Werder, Germany and activated as a 2% aqueous slurry using a high pressure homogenizer PandaPlus from GEA (350 bars, 10 passes), then this slurry is added in the last step by using a Ultra-turrax with S 25 N - 18 G - ST Dispersing element from IKA.
35	

**[0171]** The dimensions and values disclosed herein are not to be understood as being strictly limited to the exact numerical values recited. Instead, unless otherwise specified, each such dimension is intended to mean both the recited value and a functionally equivalent range surrounding that value. For example, a dimension disclosed as "40 mm" is intended to mean "about 40 mm."

45

50

55

EP 3 330 351 A1

SEQUENCE LISTING

<110> P&G

5 <120> CLEANING COMPOSITIONS INCLUDING ENZYME AND BLEACH

<130> CM04648FM

<160> 14

10 <170> PatentIn version 3.5

<210> 1

<211> 463

<212> PRT

15 <213> Streptomyces davawensis

<400> 1

20 Asp Ala Thr Ile Val Ile Asn Pro Gly Thr Arg Tyr Gly Thr Trp Glu  
1 5 10 15

Gly Trp Gly Thr Ser Leu Ala Trp Trp Gly Asn Val Phe Gly Thr Arg  
20 25 30

25 Asp Asp Phe Ala Asp Leu Phe Phe Thr Thr Lys Ser Val Thr Tyr Asn  
35 40 45

30 Gly Thr Ser Leu Pro Gly Leu Gly Leu Asn Ile Ala Arg Tyr Asn Leu  
50 55 60

35 Gly Ala Cys Ser Trp Asn Ala Val Asn Gly Glu Thr Met Val Lys Ser  
65 70 75 80

Pro Asn Ile Pro Ala Phe Lys Gln Ile Glu Gly Phe Trp Gln Asp Trp  
85 90 95

40 Asn Asn Glu Asp Pro Thr Ser Ser Ala Trp Asp Trp Thr Ala Asp Ala  
100 105 110

45 Thr Gln Arg Ala Met Leu Val Lys Ala Thr Gln Arg Gly Ala Val Thr  
115 120 125

Glu Leu Phe Ala Asn Ser Pro Met Trp Trp Met Cys Tyr Asn His Asn  
130 135 140

50 Pro Ser Gly Ala Ala Asp Gly Gly Asn Asn Leu Gln Thr Trp Asn Tyr  
145 150 155 160

55 Arg Gln His Ala Ser His Leu Ala Ala Val Ala Leu Tyr Ala Arg Thr  
165 170 175

EP 3 330 351 A1

Asn Trp Gly Val Asn Phe Ala Thr Val Asp Pro Phe Asn Glu Pro Ala  
 180 185 190

5 Ser Ser Trp Trp Thr Ala Ser Gly Thr Gln Glu Gly Cys His Leu Asp  
 195 200 205

10 Pro Ala Val Gln Ala Ala Val Leu Pro Tyr Met Arg Ser Glu Leu Asp  
 210 215 220

Lys Arg Gly Leu Thr Gly Val Arg Ile Ser Ala Ser Asp Glu Thr Asn  
 225 230 235 240

15 Tyr Asp Thr Ala Arg Ser Thr Trp Ser Ser Phe Gly Ser Ala Thr Lys  
 245 250 255

20 Ala Leu Val Ser Gln Val Asn Val His Gly Tyr Gln Gly Thr Gly Gly  
 260 265 270

Arg Arg Asp Leu Leu Tyr Thr Asp Val Val Thr Thr Ser Gly Lys Lys  
 275 280 285

25 Leu Trp Asn Ser Glu Thr Gly Asp Ser Asp Gly Thr Gly Leu Ser Met  
 290 295 300

30 Ala Arg Asn Leu Cys Tyr Asp Phe Arg Trp Leu His Pro Thr Ala Trp  
 305 310 315 320

Cys Tyr Trp Gln Val Met Asp Pro Ser Thr Gly Trp Ala Met Ile Ala  
 325 330 335

35 Tyr Asp Ala Asn Thr Leu Gln Pro Thr Thr Val Gln Pro Lys Tyr Tyr  
 340 345 350

40 Val Met Ala Gln Phe Ser Arg His Ile Arg Pro Gly Met Thr Ile Leu  
 355 360 365

45 Asp Thr Gly Val Ser Phe Ala Ala Ala Tyr Asp Ala Ser Ala Arg  
 370 375 380

Arg Leu Val Leu Val Ala Val Asn Thr Ser Thr Ser Pro Gln Thr Phe  
 385 390 395 400

50 Thr Phe Asp Leu Ser Arg Phe Thr Thr Val Thr Gly Gly Ser Gly Gly  
 405 410 415

55 Leu Val Pro Arg Trp Asn Thr Val Thr Gly Gly Gly Asp Met Tyr Arg  
 420 425 430

EP 3 330 351 A1

Ala Tyr Thr Asn Thr Tyr Val Thr Gly Lys Ser Val Ser Ala Thr Phe  
435 440 445

5 Ala Ala Gly Ser Val Gln Thr Leu Gln Val Asp Gly Val Thr Thr  
450 455 460

<210> 2  
<211> 464  
10 <212> PRT  
<213> Streptomyces avermitilis

<400> 2

15 Asp Ala Thr Ile Ala Val Asn Pro Ser Thr Thr Tyr Gly Lys Trp Glu  
1 5 10 15

Gly Trp Gly Thr Ser Leu Ala Trp Trp Ala Asn Val Phe Gly Ala Arg  
20 20 25 30

Asp Asp Phe Ala Asp Leu Phe Phe Thr Thr Lys Ser Val Thr Tyr Asn  
25 35 40 45

Gly Arg Thr Leu Pro Gly Leu Gly Leu Asn Ile Ala Arg Tyr Asn Leu  
30 50 55 60

Gly Ala Cys Ser Trp Asn Ser Val Ser Gly Glu Ser Met Val Ala Ser  
35 65 70 75 80

Ala Asn Ile Pro Ala Phe Lys Gln Ile Glu Gly Tyr Trp Gln Asp Trp  
40 85 90 95

Asn Asn Glu Asp Pro Thr Ser Ser Ala Trp Lys Trp Thr Ala Asp Ala  
45 100 105 110

Ala Gln Arg Thr Met Leu Val Lys Ala Thr Ala Arg Gly Ala Thr Thr  
50 115 120 125

Glu Leu Phe Ala Asn Ser Pro Met Trp Trp Met Cys Leu Asn His Asn  
55 130 135 140

Pro Ser Gly Ala Ser Gly Gly Gly Asn Asn Leu Gln Ser Trp Asn Tyr  
60 145 150 155 160

Arg Gln His Ala Ser His Leu Ala Ala Val Ala Leu Tyr Ala Lys Ser  
65 165 170 175

Asn Trp Gly Val Asn Phe Ala Thr Val Asp Pro Phe Asn Glu Pro Ser  
70 180 185 190



EP 3 330 351 A1

Ser Ser Trp Trp Thr Ala Thr Gly Thr Gln Glu Gly Cys His Met Asp  
 195 200 205

5  
 Ala Ser Val Gln Ala Ala Val Leu Pro Tyr Leu Arg Ser Glu Leu Asp  
 210 215 220

10  
 Arg Arg Gly Leu Thr Gly Thr Lys Ile Ser Ala Ser Asp Glu Thr Ser  
 225 230 235 240

15  
 Tyr Asp Leu Ala Arg Thr Thr Trp Gly Ser Phe Gly Ser Ser Thr Lys  
 245 250 255

20  
 Ala Leu Val Asn Arg Val Asn Val His Gly Tyr Gln Gly Ser Gly Gly  
 260 265 270

25  
 Arg Arg Asp Leu Leu Tyr Thr Asp Val Val Thr Thr Ala Gly Lys Ala  
 275 280 285

30  
 Leu Trp Asn Ser Glu Thr Gly Asp Ser Asp Gly Thr Gly Leu Thr Leu  
 290 295 300

35  
 Ala Ser Asn Leu Cys Leu Asp Phe Arg Trp Leu His Pro Thr Ala Trp  
 305 310 315 320

40  
 Val Tyr Trp Gln Val Met Asp Pro Ser Ser Gly Trp Ala Met Ile Ala  
 325 330 335

45  
 Tyr Asp Ala Ser Thr Leu Gln Pro Gly Ala Val Gln Thr Lys Tyr Tyr  
 340 345 350

50  
 Val Met Ala Gln Phe Ser Arg His Ile Arg Ala Gly Met Thr Ile Val  
 355 360 365

55  
 Asp Thr Gly Val Gly Tyr Ala Ala Ala Ala Tyr Asp Ala Thr Ala Arg  
 370 375 380

60  
 Arg Leu Val Ile Val Ala Val Asn Thr Ser Thr Ser Ala Gln Thr Leu  
 385 390 395 400

65  
 Thr Phe Asp Leu Ser Arg Phe Ser Thr Val Thr Gly Gly Thr Gly Gly  
 405 410 415

70  
 Leu Val Arg Arg Trp Asn Thr Val Thr Gly Gly Gly Gly Asp Leu Tyr  
 420 425 430

75  
 Ala Ala His Ser Asp Thr Tyr Leu Ser Gly Lys Ser Leu Ser Val Pro  
 435 440 445

EP 3 330 351 A1

Phe Ala Ala Gly Ala Val Gln Thr Leu Glu Val Asp Gly Val Thr Val  
 450 455 460

5 <210> 3  
 <211> 458  
 <212> PRT  
 <213> Trichoderma harzianum

10 <400> 3

Asp Thr Thr Leu Ser Ile Asp Pro Thr Ser Asn Trp Gly Thr Trp Glu  
 1 5 10 15

15 Gly Trp Gly Val Ser Leu Ala Trp Trp Ala Lys Ala Phe Gly Asn Arg  
 20 25 30

20 Asp Asp Leu Ala Asn Val Phe Phe Thr Arg Asn Asn Gln Val Ile Asn  
 35 40 45

Gly Gln Asn Leu Pro Gly Leu Gly Phe Asn Ile Ala Arg Tyr Asn Ala  
 50 55 60

25 Gly Ala Cys Ser Thr Asn Thr Tyr Asn Gly Ser Ser Met Val Val Ser  
 65 70 75 80

30 Ser Ser Ile Lys Pro Ser Arg Gln Val Asp Gly Tyr Trp Leu Asp Trp  
 85 90 95

35 Ala Ser Thr Asp Pro Ala Ser Ser Ser Trp Asn Trp Asn Val Asp Ala  
 100 105 110

Asn Gln Arg Ala Met Leu Gln Lys Ala Lys Ala Asn Gly Ala Asn Ile  
 115 120 125

40 Phe Glu Leu Phe Ser Asn Ser Pro Met Trp Trp Met Cys Leu Asn His  
 130 135 140

45 Asn Pro Ser Gly Ser Gly Ser Ser Asp Asn Leu Gln Ser Trp Asn Tyr  
 145 150 155 160

Gln Asn His Ala Val Tyr Leu Ala Asn Ile Ala Gln His Ala Gln Gln  
 165 170 175

50 Asn Trp Gly Ile Gln Phe Gln Ser Val Glu Ala Phe Asn Glu Pro Ser  
 180 185 190

55 Ser Gly Trp Gly Pro Thr Gly Thr Gln Glu Gly Cys His Phe Ala Val  
 195 200 205

EP 3 330 351 A1

Ser Thr Met Ala Thr Val Ile Gly Tyr Leu Asn Thr Glu Leu Ala Gln  
 210 215 220  
 5 Arg Gly Leu Ser Ser Phe Ile Ser Ala Ser Asp Glu Thr Ser Tyr Asp  
 225 230 235 240  
 10 Leu Ala Ile Ser Thr Trp Gln Gly Leu Gly Ser Ser Ala Gln Asn Ala  
 245 250 255  
 15 Val Lys Arg Val Asn Val His Gly Tyr Gln Gly Gly Gly Gly Arg Arg  
 260 265 270  
 20 Asp Thr Leu Tyr Ser Leu Val Ser Gln Ala Gly Lys Arg Leu Trp Asn  
 275 280 285  
 25 Ser Glu Tyr Gly Asp Ala Asp Ala Ser Gly Lys Ser Met Tyr Thr Asn  
 290 295 300  
 30 Leu Leu Leu Asp Phe Thr Trp Leu His Pro Thr Ala Trp Val Tyr Trp  
 305 310 315 320  
 35 Gln Ala Ile Asp Gly Ser Gly Trp Gly Leu Ile Val Gly Asp Asn Asp  
 325 330 335  
 40 Gln Leu Thr Leu Ser Ser Ala Ser Thr Lys Tyr Phe Val Leu Ala Gln  
 340 345 350  
 45 Leu Thr Arg His Ile Arg Pro Gly Met Gln Ile Leu Thr Thr Pro Asp  
 355 360 365  
 50 Gly Asn Thr Val Ala Ala Tyr Asp Ser Gly Ser Gln Lys Leu Val Ile  
 370 375 380  
 55 Val Ala Ala Asn Trp Gly Ser Ala Gln Thr Ile Thr Phe Asp Leu Thr  
 385 390 395 400  
 Arg Ala Lys Thr Ala Gly Ser Asn Gly Ala Thr Val Pro Arg Trp Ser  
 405 410 415  
 50 Thr Gln Thr Ser Gly Gly Asp Gln Tyr Lys Ser Tyr Ser Asp Thr Lys  
 420 425 430  
 55 Ile Asn Asn Gly Lys Phe Ser Val Ser Phe Ser Thr Gly Gln Val Gln  
 435 440 445  
 Thr Phe Glu Ile Ser Gly Val Val Leu Lys

EP 3 330 351 A1

450

455

5

<210> 4  
 <211> 109  
 <212> PRT  
 <213> Bacillus licheniformis

<400> 4

10

Ala Arg Tyr Asp Asp Val Leu Tyr Phe Pro Ala Ser Arg Tyr Pro Glu  
 1 5 10 15

15

Thr Gly Ala His Ile Ser Asp Ala Ile Lys Ala Gly His Ala Asp Val  
 20 25 30

20

Cys Thr Ile Glu Arg Ser Gly Ala Asp Lys Arg Arg Gln Glu Ser Leu  
 35 40 45

25

Lys Gly Ile Pro Thr Lys Pro Gly Phe Asp Arg Asp Glu Trp Pro Met  
 50 55 60

30

Ala Met Cys Glu Glu Gly Gly Lys Gly Ala Ser Val Arg Tyr Val Ser  
 65 70 75 80

35

Ser Ser Asp Asn Arg Gly Ala Gly Ser Trp Val Gly Asn Arg Leu Asn  
 85 90 95

40

Gly Tyr Ala Asp Gly Thr Arg Ile Leu Phe Ile Val Gln  
 100 105

45

<210> 5  
 <211> 109  
 <212> PRT  
 <213> Bacillus subtilis

<400> 5

Ala Ser Ser Tyr Asp Lys Val Leu Tyr Phe Pro Leu Ser Arg Tyr Pro  
 1 5 10 15

50

Glu Thr Gly Ser His Ile Arg Asp Ala Ile Ala Glu Gly His Pro Asp  
 20 25 30

55

Ile Cys Thr Ile Asp Asp Gly Ala Asp Lys Arg Arg Glu Glu Ser Leu  
 35 40 45

Lys Gly Ile Pro Thr Lys Pro Gly Tyr Asp Arg Asp Glu Trp Pro Met  
 50 55 60

60

Ala Val Cys Glu Glu Gly Gly Ala Gly Ala Asp Val Arg Tyr Val Thr  
 65 70 75 80

EP 3 330 351 A1

Pro Ser Asp Asn Arg Gly Ala Gly Ser Trp Val Gly Asn Gln Met Ser  
 85 90 95

5 Ser Tyr Pro Asp Gly Thr Arg Val Leu Phe Ile Val Gln  
 100 105

10 <210> 6  
 <211> 109  
 <212> PRT  
 <213> Bacillus licheniformis  
 <400> 6

15 Ala Arg Tyr Asp Asp Ile Leu Tyr Phe Pro Ala Ser Arg Tyr Pro Glu  
 1 5 10 15

20 Thr Gly Ala His Ile Ser Asp Ala Ile Lys Ala Gly His Ser Asp Val  
 20 25 30

25 Cys Thr Ile Glu Arg Ser Gly Ala Asp Lys Arg Arg Gln Glu Ser Leu  
 35 40 45

Lys Gly Ile Pro Thr Lys Pro Gly Phe Asp Arg Asp Glu Trp Pro Met  
 50 55 60

30 Ala Met Cys Glu Glu Gly Gly Lys Gly Ala Ser Val Arg Tyr Val Ser  
 65 70 75 80

35 Ser Ser Asp Asn Arg Gly Ala Gly Ser Trp Val Gly Asn Arg Leu Ser  
 85 90 95

40 Gly Phe Ala Asp Gly Thr Arg Ile Leu Phe Ile Val Gln  
 100 105

45 <210> 7  
 <211> 204  
 <212> PRT  
 <213> Aspergillus oryzae  
 <400> 7

50 Lys Thr Gly Ser Gly Asp Ser Gln Ser Asp Pro Ile Lys Ala Asp Leu  
 1 5 10 15

Glu Val Lys Gly Gln Ser Ala Leu Pro Phe Asp Val Asp Cys Trp Ala  
 20 25 30

55 Ile Leu Cys Lys Gly Ala Pro Asn Val Leu Gln Arg Val Asn Glu Lys  
 35 40 45

EP 3 330 351 A1

Thr Lys Asn Ser Asn Arg Asp Arg Ser Gly Ala Asn Lys Gly Pro Phe  
 50 55 60  
 5 Lys Asp Pro Gln Lys Trp Gly Ile Lys Ala Leu Pro Pro Lys Asn Pro  
 65 70 75 80  
 10 Ser Trp Ser Ala Gln Asp Phe Lys Ser Pro Glu Glu Tyr Ala Phe Ala  
 85 90 95  
 15 Ser Ser Leu Gln Gly Gly Thr Asn Ala Ile Leu Ala Pro Val Asn Leu  
 100 105 110  
 20 Ala Ser Gln Asn Ser Gln Gly Gly Val Leu Asn Gly Phe Tyr Ser Ala  
 115 120 125  
 25 Asn Lys Val Ala Gln Phe Asp Pro Ser Lys Pro Gln Gln Thr Lys Gly  
 130 135 140  
 Thr Trp Phe Gln Ile Thr Lys Phe Thr Gly Ala Ala Gly Pro Tyr Cys  
 145 150 155 160  
 30 Lys Ala Leu Gly Ser Asn Asp Lys Ser Val Cys Asp Lys Asn Lys Asn  
 165 170 175  
 Ile Ala Gly Asp Trp Gly Phe Asp Pro Ala Lys Trp Ala Tyr Gln Tyr  
 180 185 190  
 35 Asp Glu Lys Asn Asn Lys Phe Asn Tyr Val Gly Lys  
 195 200  
 40 <210> 8  
 <211> 188  
 <212> PRT  
 <213> Trichoderma harzianum  
 <400> 8  
 45 Ala Pro Ala Pro Met Pro Thr Pro Pro Gly Ile Pro Thr Glu Ser Ser  
 1 5 10 15  
 50 Ala Arg Thr Gln Leu Ala Gly Leu Thr Val Ala Val Ala Gly Ser Gly  
 20 25 30  
 Thr Gly Tyr Ser Arg Asp Leu Phe Pro Thr Trp Asp Ala Ile Ser Gly  
 35 40 45  
 55 Asn Cys Asn Ala Arg Glu Tyr Val Leu Lys Arg Asp Gly Glu Gly Val  
 50 55 60

EP 3 330 351 A1

Gln Val Asn Asn Ala Cys Glu Ser Gln Ser Gly Thr Trp Ile Ser Pro  
 65 70 75 80  
 5 Tyr Asp Asn Ala Ser Phe Thr Asn Ala Ser Ser Leu Asp Ile Asp His  
 85 90 95  
 10 Met Val Pro Leu Lys Asn Ala Trp Ile Ser Gly Ala Ser Ser Trp Thr  
 100 105 110  
 15 Thr Ala Gln Arg Glu Ala Leu Ala Asn Asp Val Ser Arg Pro Gln Leu  
 115 120 125  
 20 Trp Ala Val Ser Ala Ser Ala Asn Arg Ser Lys Gly Asp Arg Ser Pro  
 130 135 140  
 25 Asp Gln Trp Lys Pro Pro Leu Thr Ser Phe Tyr Cys Thr Tyr Ala Lys  
 145 150 155 160  
 30 Ser Trp Ile Asp Val Lys Ser Phe Tyr Lys Leu Thr Ile Thr Ser Ala  
 165 170 175  
 35 Glu Lys Thr Ala Leu Ser Ser Met Leu Asp Thr Cys  
 180 185  
 40 <210> 9  
 <211> 361  
 <212> PRT  
 <213> *Aggregatibacter actinomycetemcomitans*  
 45 <400> 9  
 50 Asn Cys Cys Val Lys Gly Asn Ser Ile Tyr Pro Gln Lys Thr Ser Thr  
 1 5 10 15  
 55 Lys Gln Thr Gly Leu Met Leu Asp Ile Ala Arg His Phe Tyr Ser Pro  
 20 25 30  
 60 Glu Val Ile Lys Ser Phe Ile Asp Thr Ile Ser Leu Ser Gly Gly Asn  
 35 40 45  
 65 Phe Leu His Leu His Phe Ser Asp His Glu Asn Tyr Ala Ile Glu Ser  
 50 55 60  
 70 His Leu Leu Asn Gln Arg Ala Glu Asn Ala Val Gln Gly Lys Asp Gly  
 65 70 75 80  
 75 Ile Tyr Ile Asn Pro Tyr Thr Gly Lys Pro Phe Leu Ser Tyr Arg Gln  
 85 90 95

EP 3 330 351 A1

Leu Asp Asp Ile Lys Ala Tyr Ala Lys Ala Lys Gly Ile Glu Leu Ile  
 100 105 110  
 5 Pro Glu Leu Asp Ser Pro Asn His Met Thr Ala Ile Phe Lys Leu Val  
 115 120 125  
 10 Gln Lys Asp Arg Gly Val Lys Tyr Leu Gln Gly Leu Lys Ser Arg Gln  
 130 135 140  
 Val Asp Asp Glu Ile Asp Ile Thr Asn Ala Asp Ser Ile Thr Phe Met  
 145 150 155 160  
 15 Gln Ser Leu Met Ser Glu Val Ile Asp Ile Phe Gly Asp Thr Ser Gln  
 165 170 175  
 20 His Phe His Ile Gly Gly Asp Glu Phe Gly Tyr Ser Val Glu Ser Asn  
 180 185 190  
 His Glu Phe Ile Thr Tyr Ala Asn Lys Leu Ser Tyr Phe Leu Glu Lys  
 195 200 205  
 25 Lys Gly Leu Lys Thr Arg Met Trp Asn Asp Gly Leu Ile Lys Asn Thr  
 210 215 220  
 30 Phe Glu Gln Ile Asn Pro Asn Ile Glu Ile Thr Tyr Trp Ser Tyr Asp  
 225 230 235 240  
 Gly Asp Thr Gln Asp Lys Asn Glu Ala Ala Glu Arg Arg Asp Met Arg  
 245 250 255  
 35 Val Ser Leu Pro Glu Leu Leu Ala Lys Gly Phe Thr Val Leu Asn Tyr  
 260 265 270  
 40 Asn Ser Tyr Tyr Leu Tyr Ile Val Pro Lys Ala Ser Pro Thr Phe Ser  
 275 280 285  
 45 Gln Asp Ala Ala Phe Ala Ala Lys Asp Val Ile Lys Asn Trp Asp Leu  
 290 295 300  
 Gly Val Trp Asp Gly Arg Asn Thr Lys Asn Arg Val Gln Asn Thr His  
 305 310 315 320  
 50 Glu Ile Ala Gly Ala Ala Leu Ser Ile Trp Gly Glu Asp Ala Lys Ala  
 325 330 335  
 55 Leu Lys Asp Glu Thr Ile Gln Lys Asn Thr Lys Ser Leu Leu Glu Ala  
 340 345 350



EP 3 330 351 A1

Val Ile His Lys Thr Asn Gly Asp Glu  
 355 360

5 <210> 10  
 <211> 541  
 <212> PRT  
 <213> *Ascobolus stictoides*

10 <400> 10

Gln Thr Tyr Thr Leu Glu Ala Glu Ala Gly Thr Leu Thr Gly Val Thr  
 1 5 10 15

15 Val Met Asn Glu Ile Ala Gly Phe Ser Gly Thr Gly Tyr Val Gly Gly  
 20 25 30

20 Trp Asp Glu Asp Ala Asp Thr Val Ser Leu Thr Phe Thr Ser Asp Ala  
 35 40 45

25 Thr Lys Leu Tyr Asp Val Lys Ile Arg Tyr Ser Gly Pro Tyr Gly Ser  
 50 55 60

Lys Tyr Thr Arg Ile Ser Tyr Asn Gly Ala Thr Gly Gly Asp Ile Ser  
 65 70 75 80

30 Leu Pro Glu Thr Thr Glu Trp Ala Thr Val Asn Ala Gly Gln Ala Leu  
 85 90 95

35 Leu Asn Ala Gly Ser Asn Thr Ile Lys Leu His Asn Asn Trp Gly Trp  
 100 105 110

Tyr Leu Ile Asp Ala Val Ile Leu Thr Pro Ser Val Pro Arg Pro Pro  
 115 120 125

40 His Gln Val Thr Asp Ala Leu Val Asn Thr Asn Ser Asn Ala Val Thr  
 130 135 140

45 Lys Gln Leu Met Lys Phe Leu Val Ser Lys Tyr His Lys Ala Tyr Ile  
 145 150 155 160

50 Thr Gly Gln Gln Glu Leu His Ala His Gln Trp Val Glu Lys Asn Val  
 165 170 175

Gly Lys Ser Pro Ala Ile Leu Gly Leu Asp Phe Met Asp Tyr Ser Pro  
 180 185 190

55 Ser Arg Val Glu Phe Gly Thr Thr Ser Gln Ala Val Glu Gln Ala Ile  
 195 200 205

EP 3 330 351 A1

Asp Phe Asp Lys Arg Gly Gly Ile Val Thr Phe Ala Trp His Trp Asn  
 210 215 220  
 5  
 Ala Pro Ser Gly Leu Ile Asn Thr Pro Gly Ser Glu Trp Trp Arg Gly  
 225 230 235 240  
 10  
 Phe Tyr Thr Glu His Thr Thr Phe Asp Val Ala Ala Ala Leu Gln Asn  
 245 250 255  
 15  
 Thr Thr Asn Ala Asn Tyr Asn Leu Leu Ile Arg Asp Ile Asp Ala Ile  
 260 265 270  
 20  
 Ala Val Gln Leu Lys Arg Leu Gln Thr Ala Gly Val Pro Val Leu Trp  
 275 280 285  
 25  
 Arg Pro Leu His Glu Ala Glu Gly Gly Trp Phe Trp Trp Gly Ala Lys  
 290 295 300  
 30  
 Gly Pro Glu Pro Ala Lys Lys Leu Tyr Lys Ile Leu Tyr Asp Arg Leu  
 305 310 315 320  
 35  
 Thr Asn Tyr His Lys Leu Asn Asn Leu Ile Trp Val Trp Asn Ser Val  
 325 330 335  
 40  
 Ala Lys Asp Trp Tyr Pro Gly Asp Glu Ile Val Asp Val Leu Ser Phe  
 340 345 350  
 45  
 Asp Ser Tyr Pro Ala Gln Pro Gly Asp His Gly Pro Val Ser Ala Gln  
 355 360 365  
 50  
 Tyr Asn Ala Leu Val Glu Leu Gly Lys Asp Lys Lys Leu Ile Ala Ala  
 370 375 380  
 55  
 Thr Glu Val Gly Thr Ile Pro Asp Pro Asp Leu Met Gln Leu Tyr Glu  
 385 390 395 400  
 Ser Tyr Trp Ser Phe Phe Val Thr Trp Glu Gly Glu Phe Ile Glu Asn  
 405 410 415  
 Gly Val His Asn Ser Leu Glu Phe Leu Lys Lys Leu Tyr Asn Asn Ser  
 420 425 430  
 Phe Val Leu Asn Leu Asp Thr Ile Gln Gly Trp Lys Asn Gly Ala Gly  
 435 440 445  
 Ser Ser Thr Thr Thr Val Lys Ser Thr Thr Thr Thr Pro Thr Thr Thr

EP 3 330 351 A1

450 455 460

5 Ile Lys Ser Thr Thr Thr Thr Pro Val Thr Thr Pro Thr Thr Val Lys  
465 470 475 480

10 Thr Thr Thr Thr Pro Thr Thr Thr Ala Thr Thr Val Lys Ser Thr Thr  
485 490 495

15 Thr Thr Ala Gly Pro Thr Pro Thr Ala Val Ala Gly Arg Trp Gln Gln  
500 505 510

20 Cys Gly Gly Ile Gly Phe Thr Gly Pro Thr Thr Cys Glu Ala Gly Thr  
515 520 525

25 Thr Cys Asn Val Leu Asn Pro Tyr Tyr Ser Gln Cys Leu  
530 535 540

<210> 11  
<211> 526  
<212> PRT  
<213> Chaetomium virescens

30 Pro Arg Asp Pro Gly Ala Thr Ala Arg Thr Phe Glu Ala Glu Asp Ala  
1 5 10 15

35 Thr Leu Ala Gly Thr Asn Val Asp Thr Ala Leu Ser Gly Phe Thr Gly  
20 25 30

40 Thr Gly Tyr Val Thr Gly Phe Asp Gln Ala Ala Asp Lys Val Thr Phe  
35 40 45

45 Thr Val Asp Ser Ala Ser Thr Glu Leu Tyr Asp Leu Ser Ile Arg Val  
50 55 60

50 Ala Ala Ile Tyr Gly Asp Lys Arg Thr Ser Val Val Leu Asn Gly Gly  
65 70 75 80

55 Ala Ser Ser Glu Val Tyr Phe Pro Ala Gly Glu Thr Trp Thr Asn Val  
85 90 95

Ala Ala Gly Gln Leu Leu Leu Asn Gln Gly Ser Asn Thr Ile Asp Ile  
100 105 110

Val Ser Asn Trp Gly Trp Tyr Leu Ile Asp Ser Ile Thr Leu Thr Pro  
115 120 125

55 Ser Thr Pro Arg Pro Ala His Gln Ile Asn Glu Ala Pro Val Asn Ala

EP 3 330 351 A1

	130		135		140												
5	Ala 145	Ala	Asp	Lys	Asn	Ala 150	Lys	Ala	Leu	Tyr	Ser 155	Tyr	Leu	Arg	Ser	Ile 160	
10	Tyr	Gly	Lys	Lys	Ile 165	Leu	Ser	Gly	Gln	Gln 170	Glu	Leu	Ser	Leu	Ser	Asn 175	
15	Trp	Ile	Ala	Gln 180	Gln	Thr	Gly	Lys	Thr 185	Pro	Ala	Leu	Val	Ser 190	Val	Asp	
20	Leu	Met	Asp 195	Tyr	Ser	Pro	Ser	Arg 200	Val	Glu	Arg	Gly	Thr 205	Val	Gly	Thr	
25	Ala	Val 210	Glu	Glu	Ala	Ile	Gln 215	His	His	Asn	Arg	Gly 220	Gly	Ile	Val	Ser	
30	Val 225	Leu	Trp	His	Trp	Asn 230	Ala	Pro	Thr	Gly	Leu 235	Tyr	Asp	Thr	Glu	Glu 240	
35	His	Arg	Trp	Trp	Ser 245	Gly	Phe	Tyr	Thr	Ser 250	Ala	Thr	Asp	Phe	Asp 255	Val	
40	Ala	Ala	Ala	Leu 260	Ser	Ser	Thr	Thr	Asn 265	Ala	Asn	Tyr	Thr	Leu 270	Leu	Ile	
45	Arg	Asp	Ile 275	Asp	Ala	Ile	Ala	Val 280	Gln	Leu	Lys	Arg	Leu 285	Gln	Ser	Ala	
50	Gly	Val 290	Pro	Val	Leu	Phe	Arg 295	Pro	Leu	His	Glu	Ala 300	Glu	Gly	Gly	Trp	
55	Phe 305	Trp	Trp	Gly	Ala	Lys 310	Gly	Pro	Glu	Pro	Ala 315	Lys	Lys	Leu	Trp	Gly 320	
60	Ile	Leu	Tyr	Asp	Arg 325	Val	Thr	Asn	His 330	His	Gln	Ile	Asn	Asn	Leu 335	Leu	
65	Trp	Val	Trp	Asn 340	Ser	Ile	Leu	Pro	Glu 345	Trp	Tyr	Pro	Gly	Asp 350	Ala	Thr	
70	Val	Asp	Ile 355	Leu	Ser	Ala	Asp 360	Val	Tyr	Ala	Gln	Gly	Asn 365	Gly	Pro	Met	
75	Ser	Thr 370	Gln	Tyr	Asn	Gln	Leu 375	Ile	Glu	Leu	Gly	Lys 380	Asp	Lys	Lys	Met	

EP 3 330 351 A1

Ile Ala Ala Ala Glu Val Gly Ala Ala Pro Leu Pro Asp Leu Leu Gln  
385 390 395 400

5 Ala Tyr Glu Ala His Trp Leu Trp Phe Thr Val Trp Gly Asp Ser Phe  
405 410 415

10 Ile Asn Asn Ala Asp Trp Asn Ser Leu Asp Thr Leu Lys Lys Val Tyr  
420 425 430

15 Thr Ser Asp Tyr Val Leu Thr Leu Asp Glu Ile Gln Gly Trp Gln Gly  
435 440 445

20 Ser Thr Pro Ser Ala Thr Thr Thr Ser Ser Thr Thr Thr Pro Ser Ala  
450 455 460

25 Thr Thr Thr Thr Thr Thr Pro Ser Thr Thr Ala Thr Thr Ala Thr Pro  
465 470 475 480

30 Ser Ala Thr Thr Thr Ala Ser Pro Val Thr Tyr Ala Glu His Trp Gly  
485 490 495

35 Gln Cys Ala Gly Lys Gly Trp Thr Gly Pro Thr Thr Cys Arg Pro Pro  
500 505 510

40 Tyr Thr Cys Lys Tyr Gln Asn Asp Trp Tyr Ser Gln Cys Leu  
515 520 525

45 <210> 12  
<211> 452  
<212> PRT  
<213> *Preussia aemulans*

50 <400> 12

55 Gln Thr Val Ile Tyr Gln Ala Glu Gln Ala Lys Leu Ser Gly Val Thr  
1 5 10 15

60 Val Glu Phe Ser Ile Ile Lys Gln Val Val Gly Thr Gly Tyr Val Glu  
20 25 30

65 Gly Phe Asp Glu Ser Thr Asp Ser Ile Thr Phe Thr Val Glu Ser Thr  
35 40 45

70 Thr Ala Ala Leu Tyr Asp Leu Ala Leu Thr Tyr Asn Gly Pro Tyr Gly  
50 55 60

75 Asp Lys Tyr Thr Asn Val Val Leu Asn Asn Ala Ala Gly Ser Gln Val  
65 70 75 80

EP 3 330 351 A1

Ser Leu Pro Ala Thr Thr Ala Trp Thr Thr Val Pro Ala Gly Gln Val  
 85 90 95  
 5 Leu Leu Asn Ala Gly Ala Asn Thr Ile Gln Ile Gln Asn Asn Trp Gly  
 100 105 110  
 Trp Tyr Leu Val Asp Ser Ile Ser Leu Lys Pro Ala Ala Thr Arg Gly  
 115 120 125  
 10 Ala His Gln Ile Thr Thr Lys Pro Val Asn Lys Asn Ala Asn Ser Asp  
 130 135 140  
 15 Ala Lys Ala Leu Leu Lys Tyr Leu Gly Ser Ile Tyr Gly Lys Lys Ile  
 145 150 155 160  
 20 Leu Ser Gly Gln Gln Asp Leu Ser Ser Leu Asp Trp Val Thr Lys Asn  
 165 170 175  
 Val Gly Lys Thr Pro Ala Val Leu Gly Leu Asp Thr Met Asp Tyr Ser  
 180 185 190  
 25 Glu Ser Arg Lys Ser Arg Gly Ala Val Ser Thr Asp Val Asp Lys Ala  
 195 200 205  
 30 Ile Ala Phe Ala Lys Lys Gly Gly Ile Val Thr Phe Cys Trp His Trp  
 210 215 220  
 Gly Ala Pro Thr Gly Leu Phe Asp Ser Ala Ala Gln Pro Trp Tyr Arg  
 225 230 235 240  
 35 Gly Phe Tyr Thr Asp Ala Thr Asp Phe Asn Ile Glu Thr Ala Leu Lys  
 245 250 255  
 40 Asp Thr Thr Asn Ala Asn Tyr Thr Leu Leu Met Lys Asp Ile Asp Thr  
 260 265 270  
 45 Ile Ala Val Gln Leu Lys Lys Leu Gln Asp Ala Gly Val Pro Val Ile  
 275 280 285  
 Trp Arg Pro Leu His Glu Ala Glu Gly Gly Trp Phe Trp Trp Gly Ala  
 290 295 300  
 50 Lys Gly Pro Glu Pro Ala Lys Lys Leu Trp Lys Ile Met Tyr Asp Arg  
 305 310 315 320  
 55 Leu Thr Asn Gln His Gly Leu Asn Asn Leu Val Trp Thr Trp Asn Ser  
 325 330 335

EP 3 330 351 A1

Val Ala Pro Asn Trp Tyr Pro Gly Asp Asp Thr Val Asp Ile Val Ser  
 340 345 350

5 Ala Asp Thr Tyr Ser Gln Gly Asp His Gly Pro Ile Ser Ala Thr Tyr  
 355 360 365

10 Asn Asn Leu Leu Ala Leu Thr Asn Asp Thr Lys Ile Ile Ala Ala Ala  
 370 375 380

15 Glu Ile Gly Ser Val Met Glu Pro Ala Gln Leu Gln Ala Tyr Gln Ala  
 385 390 395 400

20 Asp Trp Val Tyr Phe Cys Val Trp Ser Gly Glu Phe Ile Asp Gly Gly  
 405 410 415

25 Val Trp Asn Ser Leu Asp Phe Leu Lys Lys Val Tyr Asn Asp Pro Tyr  
 420 425 430

30 Val Leu Thr Leu Asp Glu Ile Gln Gly Trp Lys Thr Ala Arg Gly Lys  
 435 440 445

35 Pro Arg Val Ser  
 450

<210> 13  
 <211> 312  
 <212> PRT  
 <213> Yunnania penicillata

40 Ala Pro Ser Thr Thr Pro Val Asn Glu Lys Ala Thr Asp Ala Ala Lys  
 1 5 10 15

45 Asn Leu Leu Ser Tyr Leu Val Glu Gln Ala Ala Asn Gly Val Thr Leu  
 20 25 30

50 Ser Gly Gln Gln Asp Leu Glu Ser Ala Gln Trp Val Ser Asp Asn Val  
 35 40 45

55 Gly Lys Trp Pro Ala Ile Leu Gly Ile Asp Phe Met Asp Tyr Ser Pro  
 50 55 60

Ser Arg Val Glu Tyr Gly Ala Val Gly Ser Thr Val Pro Asp Ala Ile  
 65 70 75 80

Ser Tyr Asp Ser Asp Gly Gly Ile Val Thr Phe Cys Trp His Trp Gly  
 85 90 95

EP 3 330 351 A1

Ser Pro Ser Gly Thr Tyr Asn Thr Thr Asp Gln Pro Trp Trp Ser Asn  
 100 105 110  
 5 Phe Tyr Thr Glu Ala Thr Ala Phe Asp Ile Ala Ala Ala Met Asp Asp  
 115 120 125  
 10 Pro Asp Ser Ala Asp Tyr Asn Leu Leu Val Arg Asp Ile Asp Ala Ile  
 130 135 140  
 15 Ser Glu Leu Leu Leu Gln Leu Gln Asp Leu Asp Ile Pro Ile Leu Trp  
 145 150 155 160  
 20 Arg Pro Leu His Glu Ala Glu Gly Gly Trp Phe Trp Trp Gly Ala Lys  
 165 170 175  
 25 Gly Pro Glu Ala Cys Ile Ala Leu Tyr Arg Leu Met Phe Asp Arg Met  
 180 185 190  
 30 Thr Asn His His Gly Leu Asn Asn Leu Leu Trp Val Trp Asn Ser Val  
 195 200 205  
 35 Asp Pro Ser Trp Tyr Pro Gly Asn Asp Val Val Asp Ile Val Ser Ala  
 210 215 220  
 40 Asp Ile Tyr Ala Asp Ala Gly Asp His Ser Pro Gln Glu Glu Thr Phe  
 225 230 235 240  
 45 Ala Ser Leu Gln Ser Leu Thr Gly Asp Thr Lys Leu Val Ala Leu Gly  
 245 250 255  
 50 Glu Val Gly Asn Ile Pro Asp Pro Ala Ser Thr Gly Gly Val Ala Asp  
 260 265 270  
 55 Trp Ala Tyr Trp Val Thr Trp Asn Gly Asp Phe Ile Lys Gly Glu Asp  
 275 280 285  
 60 Tyr Asn Pro Leu Glu Tyr Lys Lys Glu Val Phe Ser Ala Glu Asn Ile  
 290 295 300  
 65 Ile Thr Arg Asp Glu Val Asp Val  
 305 310  
 <210> 14  
 <211> 327  
 <212> PRT  
 <213> Myrothecium roridum  
 <400> 14



EP 3 330 351 A1

Gly Thr Ile Glu Asn Arg Gln Trp Leu Thr Tyr Asn Pro Val Asp Ser  
 1 5 10 15  
 Ala Ala Thr Thr Glu Ala Arg Ala Leu Leu Arg Tyr Ile Gln Ser Gln  
 5 20 25 30  
 Tyr Gly Trp Arg Tyr Leu Ser Gly Gln Gln Glu Arg Ala Glu Val Gln  
 10 35 40 45  
 Trp Leu Lys Ser Asn Ile Gly Lys Thr Pro Ala Ile Gln Gly Ser Asp  
 50 55 60  
 Leu Ile Asp Tyr Ser Pro Ser Arg Val Ser Tyr Gly Ala Thr Ser Thr  
 65 70 75 80  
 Ala Val Glu Asp Ala Ile Ala Phe Asp Arg Gln Gly Gly Ile Val Thr  
 20 85 90 95  
 Phe Thr Trp His Trp Asn Ala Pro Asn Cys Leu Tyr Asn Ser Ala Asp  
 25 100 105 110  
 Gln Pro Trp Tyr Phe Gly Phe Tyr Thr Lys Ala Thr Cys Phe Asn Ile  
 115 120 125  
 Gln Ala Ala Leu Ala Gln Gly Ser Asn Gly Ala Asp Tyr Lys Leu Leu  
 30 130 135 140  
 Ile Arg Asp Ile Asp Ala Ile Ala Val Gln Leu Lys Arg Leu Arg Asp  
 35 145 150 155 160  
 Ala Lys Val Pro Ile Leu Phe Arg Pro Leu His Glu Pro Asp Gly Ala  
 165 170 175  
 Trp Phe Trp Trp Gly Ala Lys Gly Ser Gly Pro Phe Lys Gln Leu Trp  
 180 185 190  
 Asp Ile Leu Tyr Asp Arg Leu Thr Lys Tyr His Gly Leu His Asn Met  
 45 195 200 205  
 Leu Trp Val Cys Asn Thr Glu Lys Ser Asp Trp Tyr Pro Gly Asn Asn  
 210 215 220  
 Lys Cys Asp Ile Ala Thr Thr Asp Val Tyr Val Asn Ala Gly Asp His  
 225 230 235 240  
 Ser Val Gln Lys Ser His Trp Asp Ala Leu Tyr Gly Val Ser Gly Gly  
 245 250 255

# EP 3 330 351 A1

Gln Arg Ile Leu Ala Leu Gly Glu Val Gly Val Ile Pro Asp Pro Glu  
260 265 270

5 Arg Gln Ala Ser Glu Asn Val Pro Trp Ala Tyr Trp Met Thr Trp Asn  
275 280 285

10 Gly Tyr Phe Ile Arg Asp Gly Asn Tyr Asn Ser Arg Asn Phe Leu Gln  
290 295 300

15 Ser Thr Phe Ser Asn Ala Arg Val Val Thr Leu Asp Gly Thr Ser Pro  
305 310 315 320

Leu Gly Asn Trp Lys Ser Ser  
325

20

## Claims

1. A cleaning composition comprising:
  - 25 a) an endo-beta-1,6-galactanase enzyme; and
  - b) from about 0.01% to about 5%, by weight of the cleaning composition, of water-insoluble plant fiber.
- 30 2. A cleaning composition according to claim 1, wherein the enzyme has an amino acid sequence having at least 60%, or at least 80%, or at least 90% or at least 95% identity with the amino acid sequence shown in SEQ ID NO:1, SEQ ID NO:2 or SEQ ID NO:3.
3. A cleaning composition according claims 1 and 2, wherein the galactanase enzyme is selected from Glycoside Hydrolase Family 30.
- 35 4. A cleaning composition according to any preceding claim, wherein the galactanase enzyme is obtainable from *Streptomyces davawensis*, *Trichoderma harzianum*, *Streptomyces avermitilis*, or a mixture thereof.
- 40 5. A cleaning composition according to any preceding claim, wherein the composition further comprises a  $\beta$ -N-acetylglucosaminidase enzyme from E.C. 3.2.1.52, preferably an enzyme having at least 70% identity to SEQ ID NO:9.
6. A cleaning composition according to any preceding claim, wherein the water-insoluble plant fiber is selected from the group consisting of particulate cellulose material, activated citrus fiber, and mixtures thereof.
- 45 7. A cleaning composition according to claim 6, wherein the water-insoluble plant fiber comprises particulate cellulose material containing, by dry weight of the particulate cellulose material, at least 70% cellulose.
8. A cleaning composition according to claim 6 or claim 7, wherein the particulate cellulose material further comprises less than about 10% pectin, and at least about 3% hemicellulose.
- 50 9. A cleaning composition according to any of claims 6 to 8, wherein the water-insoluble plant fiber comprises particulate cellulose material having a volume-weighted median major particle dimension of from about 25 $\mu$ m to about 75 $\mu$ m, preferably from about 35 $\mu$ m to about 65 $\mu$ m, as measured by laser light diffractometry.
- 55 10. A cleaning composition according to any of claims 6 to 9, wherein the water-insoluble plant fiber comprises particulate cellulose material, wherein less than about 10% by dry weight of the cellulose material is in the form of nanofibrillated cellulose.
11. A cleaning composition according to any preceding claim, wherein the composition further comprises fabric shading

## EP 3 330 351 A1

agent and/or an additional enzyme selected from lipases, amylases, proteases, mannanases, pectate lyases, cellulases, cutinases, and mixtures thereof, and/or an encapsulated benefit agent, wherein the encapsulated benefit agent comprises a shell surrounding a core, the core comprising a benefit agent, preferably the benefit agent comprising perfume raw materials.

5

**12.** A method of cleaning a surface, preferably a textile, comprising mixing the cleaning composition according to any preceding claim with water to form an aqueous liquor and contacting a surface, preferably a textile, with the aqueous liquor in a laundering step.

10

**13.** A method of enhancing deposition of a particulate benefit agent comprising contacting a surface, preferably a textile with an aqueous liquor comprising a composition comprising an endo-beta-1,6-galactanase enzyme, a water-insoluble plant fiber and a particulate benefit agent in a textile treatment step, preferably a laundering step, and optionally rinsing and drying the textile.

15

**14.** The use of an endo-beta-1,6-galactanase enzyme and a water-insoluble plant fiber in a cleaning composition to enhance the stain-removal and/or malodor-reducing benefits.

20

25

30

35

40

45

50

55



EUROPEAN SEARCH REPORT

Application Number  
EP 17 20 4763

5

10

15

20

25

30

35

40

45

50

55

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (IPC)
X,D	WO 2015/185689 A1 (NOVOZYMES AS [DK]) 10 December 2015 (2015-12-10) * page 38, line 31 - page 54, line 24; claims; examples; sequence 2 * -----	1-14	INV. C11D3/386
			TECHNICAL FIELDS SEARCHED (IPC)
			C11D
The present search report has been drawn up for all claims			
Place of search <b>Munich</b>		Date of completion of the search <b>11 January 2018</b>	Examiner <b>Vernier, Frédéric</b>
<b>CATEGORY OF CITED DOCUMENTS</b> X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons ..... & : member of the same patent family, corresponding document	

EPO FORM 1503 03/02 (P04C01)

**ANNEX TO THE EUROPEAN SEARCH REPORT  
ON EUROPEAN PATENT APPLICATION NO.**

EP 17 20 4763

5 This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report.  
The members are as contained in the European Patent Office EDP file on  
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

11-01-2018

10	Patent document cited in search report	Publication date	Patent family member(s)	Publication date
	WO 2015185689 A1	10-12-2015	CN 106414698 A	15-02-2017
			EP 3152290 A1	12-04-2017
15			WO 2015185689 A1	10-12-2015
	-----			
20				
25				
30				
35				
40				
45				
50				
55				

EPO FORM P0459

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82

## REFERENCES CITED IN THE DESCRIPTION

*This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.*

## Patent documents cited in the description

- WO 2015185689 A [0033]
- US 20120135498 A [0060]
- WO 2015040159 A [0070]
- WO 2004067737 A [0078]
- WO 2015091989 A [0078]
- WO 2015091990 A [0078]
- WO 2015024739 A [0078]
- WO 2015143360 A [0078]
- US 6312936 B1 [0078]
- US 5679630 A [0078]
- US 4760025 A [0078]
- US 7262042 B [0078]
- WO 09021867 A [0078]
- DE 102006022216 A1 [0078]
- DE 102006022224 A1 [0078]
- WO 2015089447 A [0078]
- WO 2015089441 A [0078]
- WO 2016066756 A [0078]
- WO 2016066757 A [0078]
- WO 2016069557 A [0078]
- WO 2016069563 A [0078]
- WO 2016069569 A [0078]
- WO 8906270 A [0078]
- WO 05052161 A [0078]
- WO 05052146 A [0078]
- WO 07044993 A2 [0078]
- WO 2014194032 A [0078]
- WO 2014194054 A [0078]
- WO 2014194117 A [0078]
- WO 2015193488 A [0078]
- WO 2016075078 A [0078]
- WO 9217577 A [0078]
- US 5352604 A [0080]
- WO 2009149144 A [0080]
- WO 2009149145 A [0080]
- WO 201056653 A [0080]
- WO 201056640 A [0080]
- WO 2011072117 A [0080]
- US 20110237487 A [0080]
- WO 2011140316 A [0080]
- WO 2012151480 A [0080]
- EP 2510092 A [0080]
- EP 2566960 A [0080]
- EP 2705145 A [0080]
- US 7153818 B [0081]
- WO 9700324 A [0081]
- EP 1022334 A [0081]
- WO 9402597 A [0081]
- WO 9418314 A [0081]
- WO 9623874 A [0081]
- WO 9743424 A [0081]
- US 5856164 A [0081]
- WO 9923211 A [0081]
- WO 9623873 A [0081]
- WO 0060060 A [0081]
- WO 06002643 A [0081]
- US 6093 A [0081]
- US 562 A [0081]
- WO 09149130 A [0081]
- EP 2540825 A [0081]
- EP 2357220 A [0081]
- EP 2534233 A [0081]
- WO 2009100102 A [0081]
- WO 2010115028 A [0081]
- US 6939702 B1 [0083]
- US PA20090217464 A [0083]
- EP 12001034 A [0083]
- EP 2623586 A [0083]
- US 7141403 B2 [0084]
- WO 2002099091 A [0087]
- WO 01062903 A [0089]
- WO 9902663 A [0089]
- WO 01064853 A [0089]
- WO 2002077242 A [0089]
- WO 03089598 A [0089]
- WO 9905243 A [0099]
- WO 9905242 A [0099]
- WO 9905244 A [0099]
- WO 9905082 A [0099]
- WO 9905084 A [0099]
- WO 9905241 A [0099]
- WO 9907656 A [0099]
- WO 0023549 A [0099]
- WO 0023548 A [0099]
- WO 0887497 A [0117]
- WO 9108281 A [0123]
- WO 9001815 A [0123]
- US 6020303 A [0170]
- US 6060443 A [0170]

**Non-patent literature cited in the description**

- **A. B. BORASTON et al.** *Biochemical Journal*, 2004, vol. 382, 769-781 [0086]
- *Biochem J.*, 1991, vol. 280, 309-316 [0087]
- **NEEDLEMAN ; WUNSCH.** *J. Mol. Biol.*, 1970, vol. 48, 443-453 [0088]
- **RICE et al.** EMBOSS: The European Molecular Biology Open Software Suite. *Trends in Genetics*, 2000, vol. 16, 276-277 [0088]