

(12) PATENT
(19) AUSTRALIAN PATENT OFFICE

(11) Application No. AU 199741710 B2
(10) Patent No. 731162

(54) Title
Method, instruments and kit for autologous transplantation

(51)⁷ International Patent Classification(s)
A61F 002/30 A61B 017/326

(21) Application No: **199741710** (22) Application Date: **1997.08.29**

(87) WIPO No: **WO98/08469**

(30) Priority Data

(31) Number	(32) Date	(33) Country
08/704891	1996.08.30	US
08/857090	1997.05.15	US

(43) Publication Date : **1998.03.19**

(43) Publication Journal Date : **1998.05.14**

(44) Accepted Journal Date : **2001.03.22**

(71) Applicant(s)
VTS Holdings, Ltd.

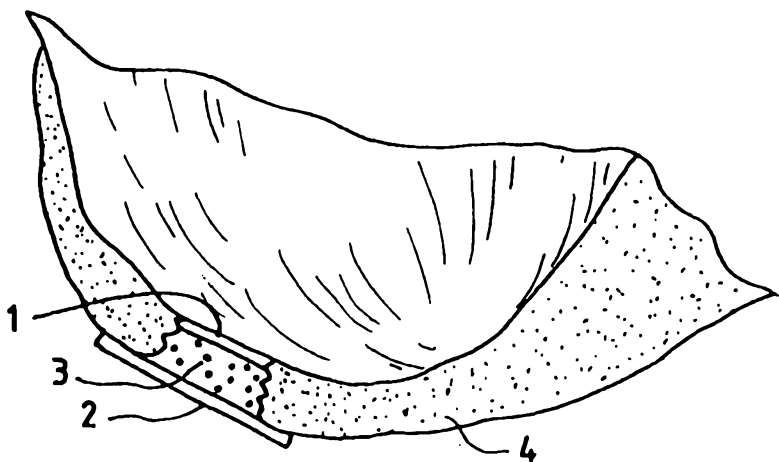
(72) Inventor(s)
Henrik Vibe-Hansen; Charlotte Lundsgaard; Ahmed Idouraine; Kurt B. Osther

(74) Agent/Attorney
**FREEHILLS CARTER SMITH BEADLE,Level 43,101 Collins Street,MELBOURNE
VIC 3000**

(56) Related Art
**EP 530804
WO 92/22312
US 5062845**



INTE

(51) International Patent Classification ⁶ : A61F 2/30, A61B 17/326		A2	(11) International Publication Number: WO 98/08469
			(43) International Publication Date: 5 March 1998 (05.03.98)
(21) International Application Number: PCT/US97/15258		(74) Agents: CHAO, Mark et al.; McDonnell Boehnen Hulbert & Berghoff, 300 S. Wacker Drive, Chicago, IL 60606 (US).	
(22) International Filing Date: 29 August 1997 (29.08.97)		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).	
(30) Priority Data: 08/704,891 30 August 1996 (30.08.96) US 08/857,090 15 May 1997 (15.05.97) US			
(60) Parent Application or Grant (63) Related by Continuation US 08/704,891 (CIP) Filed on 30 August 1996 (30.08.96)			
(71) Applicant (for all designated States except US): VTS HOLDINGS, LTD. [KN/KN]; Theodore L. Hobson, Main Street, P.O. Box 495, Charlestown, Nevis (KN).		Published <i>Without international search report and to be republished upon receipt of that report.</i>	
(72) Inventors; and (75) Inventors/Applicants (for US only): VIBE-HANSEN, Henrik [DK/DK]; Prinseevej 11, DK-2800 Lyngby (DK). LUNDS-GAARD, Charlotte [DK/DK]; Hvidorevej 36, DK-2930 Clampenborg (DK). IDOURAINE, Ahmed [DZ/US]; 2452 E. Springfield Place, Chandler, AZ 85249 (US). OSTHER, Kurt, B. [US/US]; Equestrian Manor, 6030 E. Laurel Lane, Scottsdale, AZ 85254 (US).			
(54) Title: METHOD, INSTRUMENTS AND KIT FOR AUTOLOGOUS TRANSPLANTATION			
(57) Abstract Method for the effective transplantation of chondrocytes/cartilage to an articular joint surface defect is taught, as well as a description of certain instruments and kit for practicing the invention.			
			

Method, Instruments and Kit for Autologous Transplantation

Field of the Invention

The instant invention concerns the field of chondrocyte transplantation, bone and cartilage grafting, healing, joint repair and the prevention of arthritic pathologies. In particular methods for the preparation of the graft site, instruments for such preparation and for the autologous transplantation of cells to the prepared graft site.

Background of the Invention

More than 500,000 arthroplastic procedures and total joint replacements are performed each year in the United States. Approximately the same number of similar procedures are performed in Europe. Included in these numbers are about 90,000 total-knee replacements and around 50,000 procedures to repair defects in the knee per year in Europe. The number of procedures are essentially the same in the U. S. (In: Praemer A., Furner S., Rice, D.P., Musculoskeletal conditions in the United States, American Academy of Orthopaedic Surgeons, Park Ridge, Ill., 1992, 125). A method for regeneration-treatment of cartilage would be most useful, and could be performed at an earlier stage of joint damage, thus reducing the number of patients needing artificial joint replacement surgery. With such preventative methods of treatment, the number of patients developing osteoarthritis would also decrease.

Techniques used for resurfacing the cartilage structure in joints have mainly attempted to induce the repair of cartilage using subchondral drilling, abrasion and other methods whereby there is excision of diseased cartilage and subchondral bone, leaving vascularized cancellous bone exposed (Insall, J., Clin. Orthop. 1974, 101, 61; Ficat R.P. et al, Clin Orthop. 1979, 144, 74; Johnson L.L., In: Operative Arthroscopy, McGinty J.B., Ed., Raven Press, New York, 1991, 341).

Coon and Cahn (Science 1966, 153, 1116) described a technique for the cultivation of cartilage synthesizing cells from chick embryo somites. Later Cahn and Lasher (PNAS USA 1967, 58, 1131) used the system for analysis of the involvement of DNA synthesis as a prerequisite for cartilage differentiation. Chondrocytes respond to both EFG and FGF by growth (Gospodarowicz and Mescher, J. Cell Physiology 1977, 93, 117), but ultimately

lose their differentiated function (Benya et al., Cell 1978, 15, 1313). Methods for growing chondrocytes were described and are principally being used with minor adjustments by Brittberg, M. et al. (New Engl. J. Med. 1994, 331, 889). Cells grown using these methods were used as autologous transplants into knee joints of patients. Additionally, Kolettas et al. (J. Cell Science 1995, 108, 1991) examined the expression of cartilage-specific molecules such as collagens and proteoglycans under prolonged cell culturing. They found that despite morphological changes during culturing in monolayer cultures (Aulthouse, A. et al., In Vitro Cell Dev. Biol., 1989,25,659; Archer, C. et al., J. Cell Sci. 1990,97,361; Hänselmann, H. et al., J. Cell Sci. 1994,107,17; Bonaventure, J. et al., Exp. Cell Res. 1994,212,97), when compared to suspension cultures grown over agarose gels, alginate beads or as spinner cultures (retaining a round cell morphology) tested by various scientists did not change the chondrocyte - expressed markers such as types II and IX collagens and the large aggregating proteoglycans, aggrecan, versican and link protein did not change (Kolettas, E. et al., J. Cell Science 1995,108,1991).

The articular chondrocytes are specialized mesenchymal derived cells found exclusively in cartilage. Cartilage is an avascular tissue whose physical properties depend on the extracellular matrix produced by the chondrocytes. During endochondral ossification chondrocytes undergo a maturation leading to cellular hypertrophy, characterized by the onset of expression of type X collagen (Upholt, W.B. and Olsen, R.R., In: Cartilage Molecular Aspects (Hall, B & Newman, S, Eds.) CRC Boca Raton 1991, 43; Reichenberger, E. et al., Dev. Biol. 1991, 148, 562; Kirsch, T. et al., Differentiation, 1992, 52, 89; Stephens, M. et al., J. Cell Sci. 1993, 103, 1111).

Excessive degradation of type II collagen in the outer layers or articular surfaces of joints is also caused by osteoarthritis. The collagen network is accordingly weakened and subsequently develops fibrillation whereby matrix substances such as proteoglycans are lost and eventually displaced entirely. Such fibrillation of weakened osteoarthritic cartilage can reach down to the calcified cartilage and into the subchondral bone (Kempson, G.E. et al., Biochim. Biophys. Acta 1976, 428, 741; Roth, V. and Mow, V.C., J. Bone Joint Surgery, 1980, 62A, 1102; Woo, S.L.-Y. et al., in Handbook of Bioengineering (R. Skalak and S. Chien eds.), McGraw-Hill, New York, 1987, pp. 4.1-4.44).

Descriptions of the basic development, histological and microscopic anatomy of bone, cartilage and other such connective tissues can be found for example in Wheater, Burkitt and Daniels, Functional Histology, 2nd Edition, (Churchill Livingstone, London, 1987, Chp. 4). Descriptions of the basic histological anatomy of defects in bone, cartilage and other connective tissue can be found for example in Wheater, Burkitt, Stevens and Lowe, Basic Histopathology, (Churchill Livingstone, London, 1985, Chp. 21).

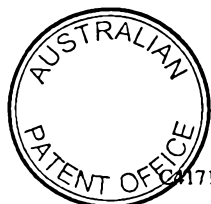
Despite the advances in cultivating chondrocytes, and manipulating bone and cartilage, there has not been great success with the attempts to transplant cartilage or chondrocytes for the repair of damaged articulating surfaces. The teachings of the instant invention provide for effective and efficient means of promoting the transplantation of cartilage and/or chondrocytes into a defect in an articulating joint or other cartilage covered bone surface, whereby cartilage is regenerated to fix the defect. The instant invention also provides for surgical instruments which are designed to prepare the graft site so as to facilitate the efficient integration of grafted material to the graft site.

It will be understood that the term "comprises" or its grammatical variants as used in this specification and claims is equivalent to the term "includes" and is not to be taken as excluding the presence of other elements or features.

Brief Summary of the Invention

The instant invention provides a method for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium, to a surface to be treated, with a covering patch. In particular, the instant method provides for a covering-patch that is a resorbable, semi-permeable collagen matrix. It may be cell-free, which is used herein as in the art, and means a material that is substantially free from intact cells which are capable of further cell division, promulgation or biological activity.

In one embodiment, the instant method encompasses the use of a cell-free covering patch which contains a semi-permeable collagen matrix. In another preferred embodiment of



the method, the porous surface of the cell-free covering-patch is directed towards the implant material.

Specifically, in a preferred embodiment, the invention provides a method for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium, to a surface to be treated, the method comprising the steps:

(a) placing chondrocytes in said culture medium upon the surface to be treated; and

(b) covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

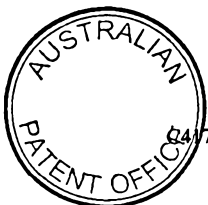
In another embodiment, the invention provides a method for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium to a surface to be treated, the method comprising the steps:

(a) sculpting the surface to be treated;

(b) placing chondrocytes in said culture medium upon the surface to be treated; and

(c) covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

Preferably the matrix is cell-free. It is also preferable that the porous surface of the covering-patch is directed toward the surface to be treated. Further, the covering-patch is preferably substantially free of intact cells and preferably resorbable. Further, it is also preferred that the covering-patch is partially attached to the surface to be treated prior to placing of the chondrocytes in said culture medium on the surface to be treated in step (b) above.



The invention also provides a kit for chondrocyte transplantation comprising a covering-patch which is a semi-permeable collagen matrix and an organic glue. In another embodiment, the invention provides a kit for chondrocyte transplantation for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium, the kit comprising a covering-patch which is a semi-permeable collagen matrix and which is adapted to cover the articulating joint surface cartilage, an organic glue, and a surgical instrument for sculpting the joint surface cartilage.

In another embodiment, the invention provides a cartilage repair structure for repair of a defect in articular cartilage to be treated comprising:

- (a) a cell-free membrane, said membrane having a porous surface and a dense surface; and
- (b) chondrocyte cells adjacent said porous surface of said membrane,

wherein the repair structure is adapted to be located adjacent a surface of the articular cartilage having the defect to be treated in order to repair the defect.

Preferably, the chondrocyte cells are adhered to said porous surface. Advantageously, the membrane is collagen, preferably Type I and Type III collagen. It is also desirable that the membrane is resorbable. It is also preferable that the chondrocyte cells are autologous. In two alternative embodiments, the chondrocyte cells are allogenic or xenogenic.

In another embodiment, the invention provides a method for producing a low density seeded chondrocyte culture *in vitro*, the chondrocytes being suitable for use for transplantation in a culture medium in a treatment of articulating joint surface cartilage, the method comprising the steps of:



(a) seeding chondrocyte cells in the culture medium at a density of 7.5×10^5 to 10×10^6 cells per millilitre; and

(b) allowing said chondrocyte cells to proliferate and differentiate for at least 48 hours.

5 In yet a further embodiment, the invention provides a method for repairing articular cartilage, the method comprising the steps of:

(a) removing a sample of healthy cartilage from a donor animal;

(b) isolating chondrocyte cells from said cartilage;

(c) seeding said chondrocyte cells in a culture medium at a density of between 7.5×10^5 to 10×10^6 cells per millilitre; and

(d) implanting said chondrocyte cells in the culture medium into a cartilage defect in a recipient animal and covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

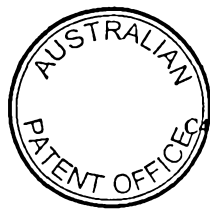
Brief Description of the Drawings

15 The present invention will be better understood by examining the following figures which illustrate certain properties of the instant invention wherein:

Figure 1A is a drawing showing a typical articulating end of a bone. Typically, the bone material is covered on the articulating surface with a cartilaginous.

20 Figure 1B shows an example of where a defect or injury to the cartilaginous cap occurs (gap in cartilage), and such a defect can be treated directly, enlarged slightly, or sculpted to accept the grafted material by surgical procedures prior to treatment.

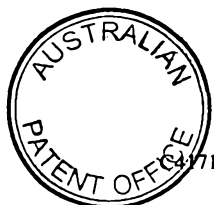
Figure 1C shows how the hemostatic barrier (numbered 1) is placed within the defect in the cartilage cap to inhibit or prevent vascularization into regenerating cartilage,



from the underlying bone. The chondrocytes to be implanted into the defect cavity are then layered on top of the hemostatic barrier.

Figure 2 is a drawing showing the treated defect (gap in cartilage) in the cartilaginous cap covered by a cell-free semi-permeable material (numbered 2) which is
5 used to form a cap/patch or bandage over the defect site. This cap is fixed in place, either

5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100



sutured to the edge of the cavity into healthy cartilage, or otherwise attached. This cap is covering the defective area of the joint into which the cultured chondrocytes/cartilage transplant has been placed, or will be placed under the partially attached cap.

Figure 3A is a diagram illustrating the differential response to compression and shearing forces by harder and softer cartilage with subsequent zone of demarcation.

Figure 3B illustrates the graft site, after the defect has been sculpted to have undulating walls.

Figure 3C illustrates the sculpted graft site with the hemostatic barrier (1), transplanted material (3), and cell-free covering-patch (2) in place within the articular surface cartilage (4).

Figure 4A illustrates one embodiment of the surgical device of the instant invention showing cutting teeth (5) and protruding placement pin (6). The cross-section illustrations to the right show two possible configurations of the cutting blades.

Figure 4B illustrates a second embodiment of the surgical device of the instant invention.

Figure 5 is a diagram illustrating the modified differential response to compression and shearing forces by harder cartilage and softer cartilage after sculpting the graft site.

Figure 6A is an MRI image of a pig knee showing cartilage defect in left (medial) condyle.

Figure 6B is an MRI image of the same pig knee three months after treatment.

Detailed Description of the Invention

This invention concerns the use of certain products that inhibit the formation of vascular tissue, for instance such as capillary loops projecting into the cartilage being established, during the process of autologous transplantation of chondrocytes into defects in the cartilage. The formation of vascular tissue from the underlying bone will tend to project into the new cartilage to be formed leading to appearance of cells other than the mesenchymal specialized chondrocytes desired.

The contaminating cells introduced by the vascularization may give rise to encroachment and over-growth into the cartilage to be formed by the implanted chondrocytes. One of the types of commercial products which can be used in this invention is Surgicel[®] (Ethicon Ltd., UK) which is absorbable after a period of 7 - 14 days. The use of this material in the method of the instant invention is contrary to the normal use of a

hemostatic device, such as Surgicel® as it is described in the package insert from Ethicon Ltd.

Surprisingly, we have found that in a situation where you wish to inhibit re-vascularization into cartilage, a hemostatic material will act like a gel-like artificial
5 coagulate. If red blood cells should be present within the full-thickness defect of articular cartilage that is capped by such a hemostatic barrier, these blood cells will be chemically changed to hematin, and thus rendered unable to induce vascular growth. Thus a hemostatic product used as a re-vascularization inhibitory barrier with or without fibrin adhesives, such as for example the Surgicel®, is effective for the envisioned method as taught by the instant
10 invention. Another part of this invention is the use of a cell-free component, that is used as a patch covering the defective area of the joint into which the cultured chondrocytes/cartilage are being transplanted, using autologous chondrocytes for the transplantation. The method of the invention also contemplates the use of suitable allogenic chondrocytes or xenogenic chondrocytes for the repair of a cartilage defect.

15 Thus the instant invention teaches methods for effective repair or treatment of cartilage defects in articular joint bone surfaces which comprises administering an agent or device to block vascular invasion into the cartilage site to be repaired, and also providing for a cell-free barrier which will isolate the repair site and keep transplanted cells in place. Thus the instant invention also provides for a kit comprising a hemostatic barrier
20 component for insertion into the site to be repaired, such that there is effective inhibition of vascularization into the site to be repaired; and once the chondrocytes to be transplanted are placed into the site to be repaired, a cell-free semi-permeable barrier is capped over the repair site such that the transplanted chondrocytes are held in place, but are still able to gain access to nutrients.

25 Certain aspects of the invention have been exemplified using an *in vitro* system to study the behavior of the chondrocytes when in contact with a certain product or a combination of certain products that inhibit the formation of vascular tissue. This *in vitro* testing predicts the ability of certain tested materials to inhibit vascularization, as will occur *in vivo* where capillary loops project into the cartilage being established during the process
30 of autologous transplantation of chondrocytes into defects in the cartilage.

Suitable hemostatic products will be characterized by having the ability to inhibit the growth, or invasion of vascular tissue, osteocytes, fibroblasts etc. into the developing cartilage. A suitable hemostatic material will achieve the goal of the method of the instant

invention in that vascular and cellular invasion into developing cartilage should be prevented in order to optimize the formation of cartilage and achieve repair of the full-thickness of any defects in the articular cartilage. Ideally, the hemostatic barrier will be stable for an extended period of time sufficient to allow for full cartilage repair, and then be able to be resorbed or otherwise broken down over time. One material identified as suitable is called Surgicel[®] W1912 (an absorbable hemostat containing oxidized regenerated sterile cellulose; Lot GG3DH, Ethicon Ltd. UK). Another example of a suitable material is BioGide[®] (a commercially available type I collagen matrix pad; Geistlich Söhne, Switzerland).

Suitable organic glue material can be found commercially, such as for example Tisseel[®] or Tissucol[®] (fibrin based adhesive; Immuno AG, Austria), Adhesive Protein (Cat. #A-2707, Sigma Chemical, USA), and Dow Corning Medical Adhesive B (Cat. #895-3, Dow Corning, USA).

The surgical instruments contemplated by the instant invention can be manufactured from metal and/or plastic suitable for making single-use disposable, or multi-use reusable surgical instruments. The cutting instrument may contain cutting teeth that are fully circular or flat, or anything in between. As cartilage is a relatively soft material it may be advantageous to manufacture hardened plastic cutting edges which will be able to sculpt cartilage without being able to damage bone. Such cutting instruments can be manufactured to incorporate openings for administration of fluid, suction removal of cutting debris and fluid, and fiber optic threads for illumination and visualization of the defect site.

Certain aspects of the instant invention may be better understood as illustrated by the following examples, which are meant by way of illustration and not limitation.

Example 1

In order for the Surgicel[®] to be used according to the invention for preventing development of blood vessels into autologous implanted cartilage or chondrocytes, Surgicel[®] was first treated with a fixative, such as glutaric aldehyde. Briefly, Surgicel[®] was treated with 0.6% glutaric aldehyde for 1 minute, followed by several washings to eliminate glutaric aldehyde residues that may otherwise be toxic to tissue. Alternatively, the Surgicel[®] was treated with the fibrin adhesive called Tisseel[®] prior to treatment with glutaric aldehyde as described in Example 2. It was found that the Surgicel[®] fixated for instance with a fixative such as glutaric aldehyde, washed with sterile physiological saline

(0.9%) and stored in refrigerator, does not dissolve for 1 to 2 months. Generally, Surgical is resorbed in a period between 7 and 14 days. This time would be too short, because a longer time is needed in preventing the development of blood vessels or vascularization as such from the bone structure into the implanted cartilage before the implanted chondrocytes have grown into a solid cartilage layer getting its nutrition requirements from the neighboring cartilage. In other words sufficient inhibition of the vascularization is needed for a longer time such as for instance one month. Therefore, the product should not be absorbed significantly prior to that time. On the other hand resorption is needed eventually. Hence, the organic material used as an inhibiting barrier shall have these capabilities, and it has been found that the Surgicel[®] treated in this manner provides that function.

Example 2

The Surgicel[®] was also coated with an organic glue, in this example the glue used was Tisseel[®] but others can also be used. This product, together with the Surgicel[®] produces a useable barrier for the particular purpose of the invention. Any other hemostat or vascular inhibiting barrier could be used. The Tisseel[®] was mixed as described below. The Surgicel[®] was then coated with Tisseel[®] by spraying the Surgicel[®] material on both sides until soaked. The Tisseel[®] (fibrin glue) was then allowed to solidify at room temperature. Immediately prior to completed solidification, the coated Surgicel[®] was then placed in 0.6% glutaric aldehyde for 1 minute and then washed with sterile physiological (0.9%) saline. The pH was then adjusted by PBS and/or with NaOH until pH was stable at 7.2 to 7.4. Afterwards the thus treated Surgicel[®] was then washed in tissue culture medium such as minimum essential medium/F12 with 15 mM Hepes buffer.

As mentioned in this example we have used Tisseel[®] as the fibrin adhesive to coat the Surgicel[®]. Furthermore the fibrin adhesive or glue may also be applied directly on the bottom of the lesion towards the bone, on which the Surgicel[®] is glued. The *in vitro* system used, in lieu of *in vivo* testing, consisted of a NUNCLON[™] Delta 6-well sterile disposable plate for cell research work (NUNC, InterMed, Roskilde, Denmark). Each well measures approximately 4 cm in diameter.

In the invention the fibrin adhesive can be any adhesive which together with the fibrin component will produce a glue that can be tolerated in humans (Ihara, N, et al., Burns Incl. Therm. Inj., 1984, 10, 396). The invention also anticipates any other glue component that can be used in lieu of the fibrin adhesive. In this invention we used Tisseel[®] or

Tissucol® (Immuno AG, Vienna, Austria). The Tisseel® kit consists of the following components:

Tisseel®, a lyophilized, virus-inactivated Sealer, containing clottable protein, thereof: fibrinogen, Plasma fibronectin (CIG) and Factor XIII, and Plasminogen.

5 Aprotinin Solution (bovine)

Thrombin 4 (bovine)

Thrombin 500 (bovine)

Calcium Chloride solution

The Tisseel® kit contains a DUPLOJECT® Application System. The fibrin adhesive
10 or the two-component sealant using Tisseel® Kit is combined in the following manner according to the Immuno AG product insert sheet:

Example 3

Chondrocytes were grown in minimal essential culture medium containing HAM
15 F12 and 15 mM Hepes buffer and 5 to 7.5% autologous serum in a CO₂ incubator at 37°C and handled in a Class 100 laboratory at Verigen Europe A/S, Symbion Science Park, Copenhagen, Denmark. Other compositions of culture medium may be used for culturing the chondrocytes. The cells were trypsinized using trypsin EDTA for 5 to 10 minutes and counted using Trypan Blue viability staining in a Bürker-Türk chamber. The cell count was
20 adjusted to 7.5×10^5 cells per ml. One NUNCLON™ plate was uncovered in the Class 100 laboratory.

The Surgicel® hemostatic barrier was cut to a suitable size fitting into the bottom of the well in the NUNCLON™ tissue culture tray. In this case a circle, of a size of approximately 4 cm (but could be of any possible size) and placed under aseptic conditions
25 on the bottom in well in a NUNCLON™ Delta 6-well sterile disposable plate for cell research work (NUNC, InterMed, Roskilde, Denmark). The hemostatic barrier to be placed on the bottom of the well was pre-treated as described in Example 1. This treatment delays the absorption of the Surgicel significantly. This hemostatic barrier was then washed several times in distilled water and subsequently several times until non-reacted
30 glutaraldehyde was washed out. A small amount of the cell culture medium containing serum was applied to be absorbed into the hemostatic barrier and at the same time keeping the hemostatic barrier wet at the bottom of the well.

Approximately 10^6 cells in 1 ml culture medium were placed directly on top of the

hemostatic barrier, dispersed over the surface of the hemostatic barrier pre-treated with 0.4% glutaraldehyde as described above. The plate was then incubated in a CO₂ incubator at 37°C for 60 minutes. An amount of 2 to 5 ml of tissue culture medium containing 5 to 7.5% serum was carefully added to the well containing the cells avoiding splashing the cells by holding the pipette tip tangential to the side of the well when expelling the medium. It appeared that the pH of the medium was too low (pH ~6.8). The pH was then adjusted to 7.4 to 7.5. The next day some chondrocytes had started to grow on the hemostatic barrier, arranged in clusters. Some of the cells had died due to the low pH exposure prior to the adjustment of the pH. The plate was incubated for 3 to 7 days with medium change at day 3.

At the end of the incubation period the medium was decanted and cold refrigerated 2.5% glutaraldehyde containing 0.1M sodium salt of dimethylarsinic acid, (also called sodium cacodylate, pH is adjusted with HCl to 7.4), was added as fixative for preparation of the cell and supporter (hemostatic barrier) for later preparation for electron microscopy.

Example 4

Chondrocytes were grown in minimal essential culture medium containing HAM F12 and 15 mM Hepes buffer and 5 to 7.5% autologous serum in a CO₂ incubator at 37°C and handled in a Class 100 laboratory at Verigen Europe A/S, Symbion Science Park, Copenhagen, Denmark. Other compositions of culture medium may be used for culturing the chondrocytes. The cells were trypsinized using trypsin EDTA for 5 to 10 minutes and counted using Trypan Blue viability staining in a Bürker-Türk chamber. The cell count was adjusted to 7.5×10^5 cells per ml. One NUNCLON™ plate was uncovered in the Class 100 laboratory.

The Surgicel® (for use as the hemostatic barrier) was treated with 0.6% glutaric aldehyde for one minute as described in Example 1, and washed with 0.9% sterile sodium chloride solution or, preferably, with a buffer such as a PBS buffer or the culture medium such as MEM/F12, because pH after the glutaric aldehyde treatment is 6.8 and should preferably be 7.0 to 7.5. The Tisseel® was applied on both side of the Surgicel® using the DUPLOJECT® system, thus coating both sides of the Surgicel®, the patch intended to be used, with fibrin adhesive. The glue is left to dry under aseptic condition for at least 3 to 5 minutes. The "coated" hemostatic barrier was placed on the bottom of the well in a NUNCLON™ Delta 6-well sterile disposable plate for cell research work. A small amount

of tissue culture medium containing serum was applied to be absorbed into the hemostatic barrier. Approximately 10^6 cells in 1 ml tissue culture medium containing serum was placed directly on top of the Hemostat, dispersed over the surface of the hemostatic barrier. The plate was then incubated in a CO_2 incubator at 37°C for 60 minutes. An amount of 2 to 5 ml of tissue culture medium containing 5 to 7.5 % serum was carefully added to the well containing the cells avoiding splashing the cells by holding the pipette tip tangential to the side of the well when expelling the medium. After 3 to 6 days, microscopic examination showed that the cells were adhering to and growing into the Surgicel[®] in a satisfactory way suggesting that Surgicel[®] did not show toxicity to the chondrocytes and that the chondrocytes grew in a satisfactory manner into the Surgicel[®].

The plate was incubated for 3 to 7 days with medium change at day 3. At the end of the incubation period the medium was decanted and cold refrigerated 2.5% glutaraldehyde containing 0.1M sodium salt of dimethylarsinic acid, also called sodium cacodylate, pH is adjusted with HCl to 7.4, was added as fixative for preparation of the cell and supporter (hemostatic barrier) for later preparation for electron microscopy.

Example 5

Chondrocytes were grown in minimal essential culture medium containing HAM F12 and 15 mM Hepes buffer and 5 to 7.5% autologous serum in a CO_2 incubator at 37°C and handled in a Class 100 laboratory at Verigen Europe A/S, Symbion Science Park, Copenhagen, Denmark. The cells were trypsinized using trypsin EDTA for 5 to 10 minutes and counted using Trypan Blue viability staining in a Bürker-Türk chamber. The cell count was adjusted to 7.5×10^5 to 2×10^6 cells per ml. One NUNCLON[™] plate was uncovered in the Class 100 laboratory.

It has been found that the Bio-Gide[®] can be used as a resorbable bilayer membrane which will be used as the patch or bandage covering the defective area of the joint into which the cultured chondrocytes are being transplanted as well as the hemostatic barrier. The Bio-Gide[®] is a pure collagen membrane obtained by standardized, controlled manufacturing processes (by E.D. Geistlich Söhne AG, CH-6110 Wolhusen). The collagen is extracted from veterinary certified pigs and is carefully purified to avoid antigenic reactions, and sterilized in double blisters by γ -irradiation. The bilayer membrane has a porous surface and a dense surface. The membrane is made of collagen type I and type III without further cross-linking or chemical treatment. The collagen is resorbed within 24

weeks. The membrane retains its structural integrity even when wet and it can be fixed by sutures or nails. The membrane may also be "glued" using fibrin adhesive such as Tisseel[®] to the neighboring cartilage or tissue either instead of sutures or together with sutures.

5 The Bio-Gide[®] was un-covered in a class 100 laboratory and placed under aseptic conditions on the bottom of the wells in a NUNCLON[™] Delta 6-well sterile disposable plate for cell research work, - either with the porous surface of the bilayer membrane facing up or with the dense surface facing up. Approximately 10⁶ cells in 1 ml tissue culture medium containing serum was placed directly on top of the Bio-Gide[®], dispersed either over the porous or the dense surface of the Bio-Gide[®]. The plate was then incubated in a
10 CO₂ incubator at 37°C for 60 minutes. An amount of 2 to 5 ml of tissue culture medium containing 5 to 7.5 % serum was carefully added to the well containing the cells avoiding splashing the cells by holding the pipette tip tangential to the side of the well when expelling the medium.

15 On day 2 after the chondrocytes were placed in the well containing the Bio-Gide[®] the cells were examined in a Nikon Inverted microscope. It was noticed that some chondrocytes had adhered to the edge of the Bio-Gide. It was of course not possible to be able to look through the Bio-Gide[®] itself using this microscope.

The plate was incubated for 3 to 7 days with medium change at day 3. At the end of the incubation period the medium was decanted and cold refrigerated 2.5% glutaraldehyde
20 containing 0.1M sodium salt of dimethylarsinic acid, also called sodium cacodylate, pH is adjusted with HCl to 7.4, was added as fixative for preparation of the cell and the Bio-Gide[®] supporter with the cells either cultured on the porous surface or the dense surface. The Bio-Gide[®] patches were then sent for electron microscopy at Department of Pathology, Herlev Hospital, Denmark.

25 The electron microscopy showed that the chondrocytes cultured on the dense surface of the Bio-Gide[®] did not grow into the collagen structure of the Bio-Gide[®], whereas the cells cultured on the porous surface did indeed grow into the collagen structure and furthermore, showed presence of proteoglycans and no signs of fibroblast structures. This result shows that when the collagen patch, as for instance a Bio-Gide[®] patch is sewn as a
30 patch covering a cartilage defect the porous surface shall be facing down towards the defect in which the cultured chondrocytes are to be injected. They will then be able to penetrate the collagen and produce a smooth cartilage surface in line with the intact surface, and in this area a smooth layer of proteoglycans will be built up. Whereas, if the dense surface of

the collagen is facing down into the defect the chondrocytes to be implanted will not integrate with the collagen, and the cells will not produce the same smooth surface as described above.

5 Example 6

Chondrocytes were grown in minimal essential culture medium containing HAM F12 and 15 mM Hepes buffer and 5 to 7.5% autologous serum in a CO₂ incubator at 37°C and handled in a Class 100 laboratory at Verigen Europe A/S, Symbion Science Park, Copenhagen, Denmark. The cells were trypsinized using trypsin EDTA for 5 to 10 minutes and counted using Trypan Blue viability staining in a Bürker-Türk chamber. The cell count was adjusted to 7.5 x 10⁵ to 2 x 10⁶ cells per ml. One NUNCLON™ plate was uncovered in the Class 100 laboratory.

The Bio-Gide® used as a resorbable bilayer membrane may also be used together with an organic glue such as Tisseel® with additional, significantly higher content of Aprotinin than normally found in Tisseel®, as described in the product insert. By increasing the content of Aprotinin to about 25,000 KIU/ml, the resorption of the material will be delayed by weeks instead of the normal span of days.

To test this feature *in vitro*, the Tisseel® is applied to the bottom of the well of the NUNCLON™ plate, and allowed to solidify incompletely. A collagen patch such as a Bio-Gide® is then applied over the Tisseel® and glued to the bottom of the well. This combination of Bio-Gide® and Tisseel® is designed to be a hemostatic barrier that will inhibit or prevent development or infiltration of blood vessels into the chondrocyte transplantation area. This hybrid collagen patch can now be used for both as a hemostatic barrier at the bottom of the lesion (most proximal to the surface to be repaired) but also as a support for cartilage formation because the distal surface can be the porous side of the collagen patch and thus encourage infiltration of chondrocytes and cartilage matrix. Thus this hybrid collagen patch can also be used to cover the top of the implant with the collagen porous surface directed down towards the implanted chondrocytes and the barrier forming the top. The hybrid collagen patch, with elevated Aprotinin component may also be used without any organic glue such as Tisseel® and placed within the defect directly, adhering by natural forces. Thus the collagen patch can be used both as the hemostatic barrier, and the cell-free covering of the repair/transplant site, with the porous surfaces of the patches oriented towards the transplanted chondrocytes/cartilage. Another variant would use a

collagen patch which consists of type II collagen (ie. from Geistlich Sohne AG, CH-6110 Wolhusen).

Thus the instant invention provides for a hybrid collagen patch where said patch is a collagen matrix with elevated levels of aprotinin component, preferably about 25,000
5 KIU/ml, in association with an organic matrix glue, where the collagen component is similar to the Bio-Gide[®] resorbable bilayer material or Type II collagen, and the organic glue is similar to the Tisseel[®] material. In another embodiment, the hybrid collagen patch does not use any organic glue to adhere to the site of the repair.

10 **Example 7**

Because of the weakened structure of osteoarthritic cartilage, adherence of cultured autologous chondrocytes transplanted to a graft site in defective cartilage may be inhibited, thus creating a marginal zone (zone of demarcation) between the newly implanted cartilage/chondrocytes and the surrounding established cartilage. This marginal zone will be
15 most pronounced if the graft site is prepared for the graft by creating straight, smooth walls cut in a linear fashion. The shearing and compression forces across such a marginal zone (as illustrated in Figure 3A) will exert great force to dislodge the graft when the graft site is cut in a linear fashion. This marginal zone, and differential movement of materials along this zone will inhibit confluent healing between the grafted material and the surrounding
20 material. This marginal zone shearing is exacerbated when the hardness of the abutting material is different. In many cases the graft material is softer than the surrounding material, however, in some instances of osteoarthritis disease, the surrounding cartilage may in fact be softer than the implanted chondrocytes/cartilage.

Therefore, in order to solve this problem, the method of the invention teaches the
25 use of surgical instruments to sculpt the walls of the graft site such that the walls are non-linear, and thus provide for undulated surfaces which will reduce marginal zone shearing and provide anchorage for grafted material. It is also possible to shape the graft site such that the diameter of the site proximal to the bone surface is of a greater dimension than the opening distal to the bone and at the surface of the cartilage to be repaired such that there is
30 a "reverse funnel" effect. A narrowed opening at the surface will aid in reducing marginal zone shearing and the expulsion of graft material from the graft site. A preferred embodiment describes the sculpting of the walls of the graft site in an fashion similar to a threaded opening for receiving a bolt or screw (as illustrated in Figure 3B), thus providing

mechanical resistance to the compression and or ejection of the grafted material from the graft site which can be described as "male" and "female" threading.

The surgical instruments contemplated by the instant invention can be manufactured from metal and/or plastic suitable for making single-use disposable, or multi-use reusable surgical instruments. As cartilage is a relatively soft material it may be advantageous to manufacture hardened plastic cutting edges which will be able to sculpt cartilage without being able to damage bone. Such cutting instruments can be manufactured to incorporate openings for administration of fluid, suction removal of cutting debris and fluid, and fiber optic threads for illumination and visualization of the defect site. In certain embodiments of the instrument, the base of the instrument may have protruding point or pin-like structure which will assist in guiding and placing the instrument in the graft site. Of course such a pin would be designed to minimized damage to the underlying bone.

While the cutting surface of the instrument may be single toothed, or multi-toothed, or describe a screw-like pattern such as that in a metal tap used to generate threaded holes in metal parts, the characteristic required of the cutting instrument is that the resulting sculpted sides of the graft site is undulated, and non-linear. For example, in certain embodiments, the cutting edge of the instrument can be shaped similar to that shown in Figure 4A, or as in Figure 4B. The cutting edge maybe flat, or circular in that it wraps around the diameter of the cutting instrument. Many other shapes can be designed to accomplish the purpose of the method of the invention to create an interface which provides for mechanical resistance to differential reaction to compression and shearing forces on the transplanted material and the surrounding material.

Example 8

A four month old mixed Yorkshire breed pig was subjected to general anesthesia and placed on its back. The pig was washed and draped in a surgical suite at Harrington Arthritis Research Center, Phoenix, Arizona. The entire surgical procedure was performed aseptically. The left hind-leg and adjacent abdomen and inguinal area was cleaned with iodine. The knee joint was localized, and the patella localized. A medial incision was performed approximately 3 cm from the posterior part of the patella and the several subcutis, muscle layers and ligaments was cut approximately in order to get access to the medial femoral condyle. Using a circular cutter a lesion was prepared in the white cartilage on the medial part of the medial condyle, leaving a 0.5 to 1 cm margin to the edge of the

cartilage covering the posterior-medial part of the condyle (left condyle, Figure 6A). The 0.5 to 1 cm defect was placed in a caudal weight bearing part of the medial condyle. The entire surgical procedure was done without tourniquet on the left femur. The different layers and skin was sutured appropriately.

5 On day 3 the animal was again brought to the surgical suite and positioned as above on the operating table and given general anesthesia. The left hind leg, abdomen and inguinal region was ionized as described above. Sutures were cut and the area opened. It was noticed that a moderate hematoma was present in the knee joint. The blood clot was removed and the defect inspected. There was a blood clot in the defect which was removed.

10 A sterile surgical instrument designed with a male thread cutting edge, with a size corresponding to, or slightly bigger than the circumference of the lesion was carefully screwed down into the defect. A BioGide[®] pad was cut to a size equal to the bottom of the defect. The first glue used, called Adhesive Protein (A-2707, Sigma Chemical, USA) was applied on the dense side of the trimmed hemostatic barrier pad, and the pad was placed

15 dense side down into the bottom of the lesion, using it as a barrier as described above. It was found that this glue did not seem to dry very fast. The slight bleeding from the bottom of the defect stopped immediately. A second BioGide[®] was cut somewhat bigger in circumference than the lesion and was placed with dense side up (thus the porous side down towards the graft) as described above.

20 This non-cellular covering-pad was then sutured over the cavity, leaving one edge open, where the chondrocyte to be explanted could be injected into the graft site. The surrounding part of the edge of the pad was covered with the second glue, Dow Corning Medical Adhesive B (Cat. #895-3, Dow Corning, USA). This second glue dried much faster and more efficiently than the first glue. It was found that during this particular procedure,

25 the first glue had not dried sufficiently to hold the hemostatic barrier in place when suturing of the cap was attempted. The main barrier formed on the proximal surface of the graft site was by the glue itself.

Using a 1 ml syringe and a 16 gauge needle, the chondrocyte cell suspension (about 0.6 ml) was drawn up into the barrel of the syringe. A 23 gauge short needle was switched

30 for the 16 gauge needle, and the cell suspension was injected under the sutured covering-patch into the graft site (about 10×10^6 cells). The open edge of the cap was then glued prior to removal of the needle, and the needle carefully withdrawn. No leakage of cells was seen. The wound was sutured and as above, no tourniquet was used, no bleeding was

observed. The final skin layers were sutured. No protrusion of the skin occurred after suturing, which indicates that there was no hematoma. Postoperative recovery was uneventful.

As expected, the grafted chondrocytes produced cartilage matrix sufficient to repair the defect made in the articular cartilage surface of the knee joint of the test pig. Figure 6A is an MRI image of a pig knee showing the cartilage defect created in the knee (left condyle, the medial condyle), and Figure 6B is an MRI image of the same pig knee three months after treatment showing repair of the defect.

Example 9

A kit comprising the components useful for practicing the method of the invention, will allow for the convenient practice of the method of the invention in a surgical setting. In a preferred embodiment, a kit of the invention will provide sterile components suitable for easy use in the surgical environment, and will provide a suitable hemostatic barrier, suitable covering patch, and if needed organic glue. A kit of the invention may also provide sterile, cell-free matrix material suitable for supporting autologous chondrocytes that are to be implanted into an articular joint surface defect. In one embodiment, a kit of the invention contains a Surgicel[®] hemostatic barrier and a Bio-Gide[®] covering patch with suitable coating of Tisseel[®] organic glue, where the Surgicel[®] and Bio-Gide[®] have been treated according to the teachings of the invention to increase the time till resorption. In instances where Tisseel[®] is pre-coated, in one embodiment the Tisseel[®] is supplemented with additional aprotinin to increase time till resorption.

In another preferred embodiment, the hemostatic barrier and covering-patch are both a semi-permeable collagen matrix which is treated to extend the time till resorption of the material. It is also possible to provide Tisseel[®] glue in enhanced form as a separate component to be applied as needed because of the inherent variability and unique circumstances every repair/transplantation procedure will encounter.

A further embodiment of a kit of the invention will include a surgical instrument as described in Example 7 above, or suitable variations thereof.

It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention shown in the specific embodiments without departing from the spirit and scope of the invention as described.

The invention is defined by the following claims:

1. A method for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium, to a surface to be treated, the method comprising the steps:

5 (a) placing chondrocytes in said culture medium upon the surface to be treated; and

(b) covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

2. The method according to claim 1, wherein the collagen matrix is cell-free.

10

3. The method according to claim 1 or 2, wherein the semi-permeable collagen matrix has a porous surface which is directed toward the surface to be treated.

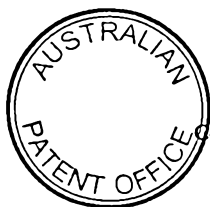
4. The method according to any of claims 1 to 3, wherein the covering-patch is substantially free of intact cells.

15

5. The method according to any of claims 1 to 4, wherein the covering-patch is resorbable.

6. The method according to any of claims 1 to 5, wherein the covering-patch is partially attached to the surface to be treated prior to placing of the chondrocytes in said culture medium on the surface to be treated in said step (b).

20 7. The use of chondrocytes in a culture medium in the manufacture of a pharmaceutical composition for the treatment of articulating joint surface cartilage in need of treatment wherein the chondrocytes in said culture medium are transplanted to a surface to be treated and the surface to be treated is covered with a covering-patch which is a semi-permeable collagen matrix.

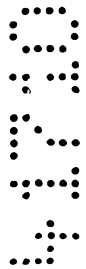


8. The use of chondrocytes according to claim 7, wherein the collagen matrix is cell-free.

9. The use of chondrocytes according to claim 7 or 8, wherein the semi-permeable collagen matrix has a porous surface which is directed toward the surface to be treated.

5 10. The use of chondrocytes according to any of claims 7 to 9, wherein the covering-patch is substantially free of intact cells.

11. The use of chondrocytes according to any of claims 7 to 10, wherein the covering-patch is resorbable.



10 12. The use of chondrocytes according to any of claims 7 to 11, wherein the covering-patch is partially attached to the surface to be treated prior to placing of the chondrocytes in said culture medium on the surface to be treated.

13. A kit for chondrocyte transplantation comprising a covering-patch which is a semi-permeable collagen matrix and an organic glue.



15 14. The kit according to claim 13 wherein the semi-permeable collagen matrix has a porous surface suitable for the placement thereon of chondrocytes.

15. The kit according to claim 13 or 14 wherein the collagen matrix is cell-free.

16. A method for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium to a surface to be treated, the method comprising the steps:

- 20
- (a) sculpting the surface to be treated;
 - (b) placing chondrocytes in said culture medium upon the surface to be treated; and



(c) covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

17. The method according to claim 16, wherein the collagen matrix is cell-free.

18. The method according to any of claims 16 to 17, wherein the semi-permeable
5 collagen matrix has a porous surface.

19. The method according to claim 18, wherein the porous surface of the covering-patch is directed toward the surface to be treated.

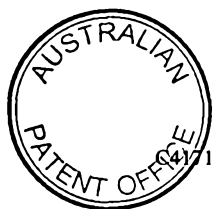
20. The method according to any of claims 16 to 19, wherein the semi-permeable collagen matrix is substantially free of intact cells.

10 21. The method according to any of claims 16 to 20, wherein the covering-patch is resorbable.

22. The method according to any of claims 16 to 21, wherein the covering-patch is partially attached to the surface to be treated prior to placing of the chondrocytes in said culture medium on the surface to be treated in said step (b).

15 23. The use of chondrocytes in a culture medium in the manufacture of a pharmaceutical composition for the effective treatment of articulating joint surface cartilage in need of treatment by the transplantation of the chondrocytes in a culture medium to the surface, wherein the surface to be treated is sculpted, the chondrocytes in said culture medium are then placed upon the surface to be treated and then the surface to be treated is covered
20 with a covering-patch which is a semi-permeable collagen matrix.

24. The use of chondrocytes according to claim 23, wherein the collagen matrix is cell-free.

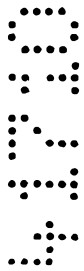


25. The use of chondrocytes according to claim 23 or 24, wherein the semi-permeable collagen matrix has a porous surface.

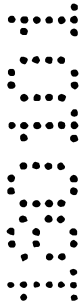
26. The use of chondrocytes according to claim 25, wherein the porous surface of the covering patch is directed toward the surface to be treated.

5 27. The use of chondrocytes according to any of claims 23 to 26, wherein the semi-permeable collagen matrix is substantially free of intact cells.

28. The use of chondrocytes according to any of claims 23 to 27, wherein the covering-patch is resorbable.



10 29. The use of chondrocytes according to any of claims 23 to 28, wherein the covering-patch is partially attached to the surface to be treated prior to placing of the chondrocytes in said culture medium on the surface to be treated.



15 30. A kit for chondrocyte transplantation for the effective treatment of articulating joint surface cartilage by the transplantation of chondrocytes in a culture medium, the kit comprising a covering-patch which is a semi-permeable collagen matrix and which is adapted to cover the articulating joint surface cartilage, an organic glue, and a surgical instrument for sculpting the joint surface cartilage.

31. The kit according to claim 30 wherein the semi-permeable collagen matrix has a porous surface suitable for the placement thereon of chondrocytes.

32. The kit according to claim 30 or 31 wherein the collagen matrix is cell-free.

20 33. A cartilage repair structure for repair of a defect in articular cartilage to be treated comprising:

- (a) a cell-free membrane, said membrane having a porous surface and a dense surface; and



(b) chondrocyte cells adjacent said porous surface of said membrane,

wherein the repair structure is adapted to be located adjacent a surface of the articular cartilage having the defect to be treated in order to repair the defect.

34. A cartilage repair structure according to claim 33, wherein said chondrocyte cells are
5 adhered to said porous surface.

35. A cartilage repair structure according to claim 33 or 34, wherein said membrane is collagen.

36. A cartilage repair structure according to claim 35, wherein said membrane is Type I and Type III collagen.

10 37. A cartilage repair structure according to any of claims 33 to 36, wherein said membrane is resorbable.

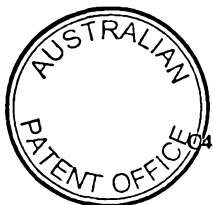
38. A cartilage repair structure according to any of claims 33 to 37, wherein said chondrocyte cells are autologous.

15 39. A cartilage repair structure according to any of claims 33 to 37, wherein said chondrocyte cells are allogenic.

40. A cartilage repair structure according to any of claims 33 to 37, wherein said chondrocytes are xenogenic.

41. A cartilage repair structure according to any of claims 33 to 40, wherein said structure is adapted to be disposed over the articular cartilage defect.

20 42. A cartilage repair structure according to any of claims 33 to 40, wherein said structure is adapted to be disposed in the articular cartilage defect.



43. A cartilage repair structure according to any of claims 33 to 40, wherein said structure is disposed over the articular cartilage defect.

44. A cartilage repair structure according to any of claims 33 to 40, wherein said structure is disposed in the articular cartilage defect.

5 45. A method for the effective treatment of a defect in articular cartilage in need of treatment by the transplantation of chondrocytes to the defect to be treated, the method comprising the steps:

(a) obtaining a cartilage repair structure according to any one of claims 33 to 44;

(b) placing the structure adjacent the articular cartilage to be treated such that the chondrocytes are between the membrane and the articular cartilage.

46. A method according to claim 45 further comprising the step of applying a biocompatible adhesive to the articular cartilage prior to step (b).

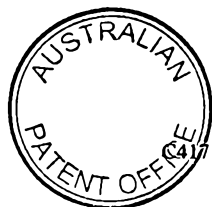
47. A method for producing a low density seeded chondrocyte culture *in vitro*, the chondrocytes being suitable for use for transplantation in a culture medium in a treatment of articulating joint surface cartilage, the method comprising the steps of:

(a) seeding chondrocyte cells in the culture medium at a density of 7.5×10^5 to 10×10^6 cells per millilitre; and

(b) allowing said chondrocyte cells to proliferate and differentiate for at least 48 hours.

20 48. A method for repairing articular cartilage, the method comprising the steps of:

(a) removing a sample of healthy cartilage from a donor animal;



(b) isolating chondrocyte cells from said cartilage;

(c) seeding said chondrocyte cells in a culture medium at a density of between 7.5×10^5 to 10×10^6 cells per millilitre; and

5 (d) implanting said chondrocyte cells in the culture medium into a cartilage defect in a recipient animal and covering the surface to be treated with a covering-patch which is a semi-permeable collagen matrix.

49. A method according to claim 47, wherein in said step (a) said chondrocyte cells are seeded in the culture medium at a density of 7.5×10^5 to 2×10^6 cells per millilitre.

10 50. A method according to claim 48, wherein in said step (c) said chondrocyte cells are seeded in the culture medium at a density of 7.5×10^5 to 2×10^6 cells per millilitre.

15 51. Use of chondrocyte cells in the manufacture of a pharmaceutical composition for the repair of the articular cartilage in need of repair wherein the cells are isolated from a removed sample of healthy cartilage from a donor animal and seeded in the culture medium at a density of between 7.5×10^5 to 10×10^6 cells per millilitre, and then the chondrocyte cells are implanted into a cartilage defect in a recipient animal.

52. The use according to claim 51 in which the cells are seeded in the culture medium at a density of 7.5×10^5 to 2×10^6 cells per millilitre.

53. A method for the effective treatment of articulating joint surface cartilage substantially as described herein with reference to the examples.

20 54. A kit for chondrocyte transplantation substantially as described herein with reference to the examples.

55. A cartilage repair structure for repair of a defect in articular cartilage substantially as described herein with reference to the examples.



56. A method for producing a low density seeded chondrocyte culture substantially as described herein with reference to the examples.

57. A method for repairing articular cartilage substantially as herein described with reference to the examples.

5

Verigen Transplantation Service International
By its registered patent attorneys
Freehills Carter Smith Beadle

15 January 2001

2001
15
JAN
2001

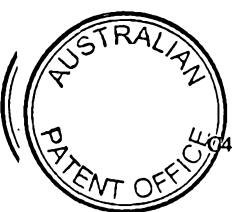


FIG. 1A

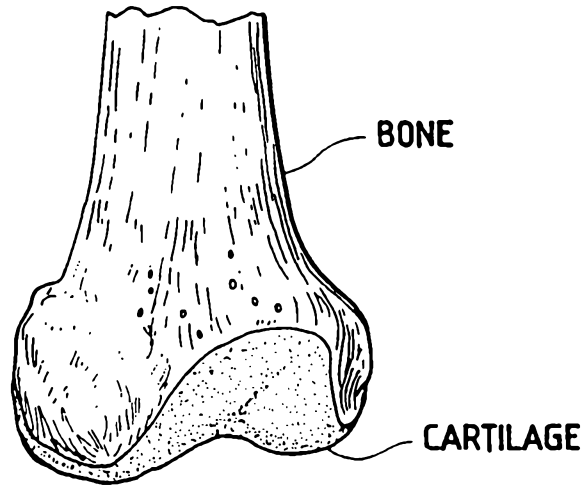


FIG. 1B

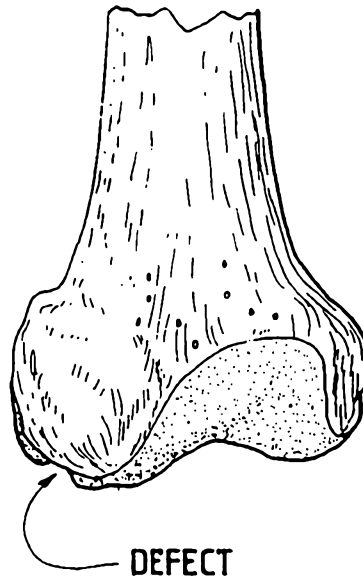


FIG. 1C

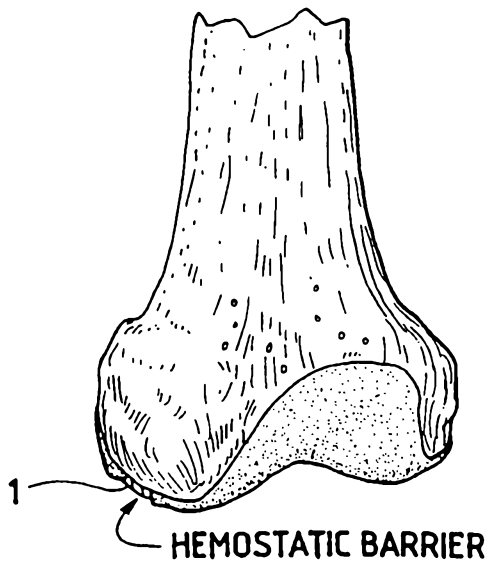


FIG. 4A

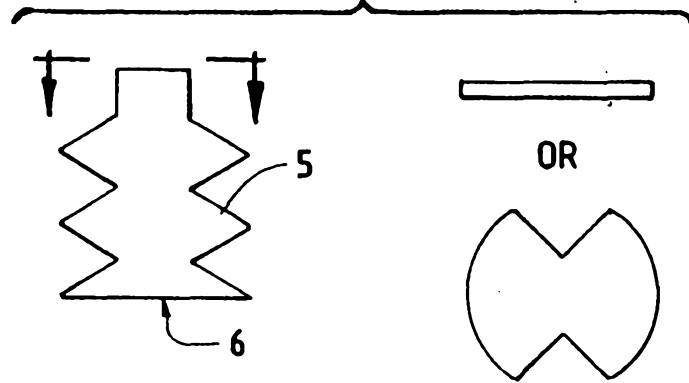


FIG. 4B

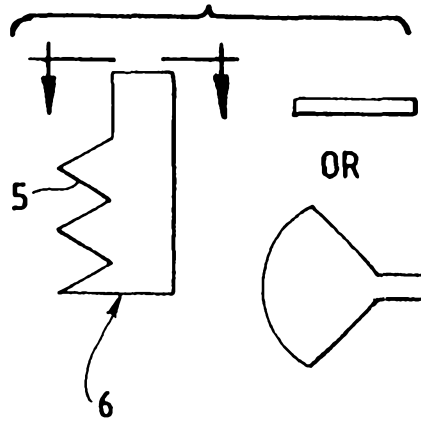
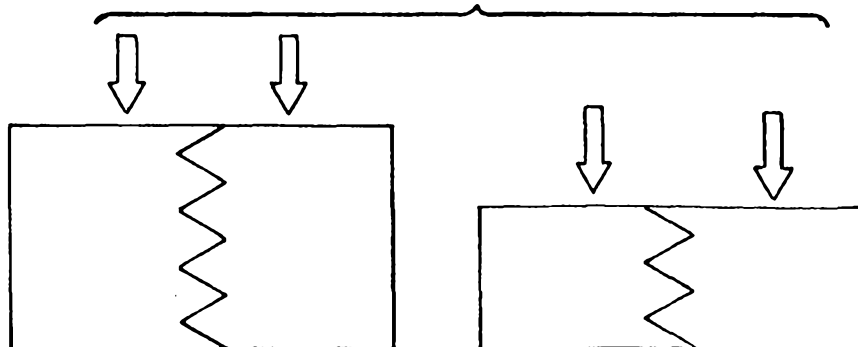


FIG. 5



4 / 5



FIG. 6A

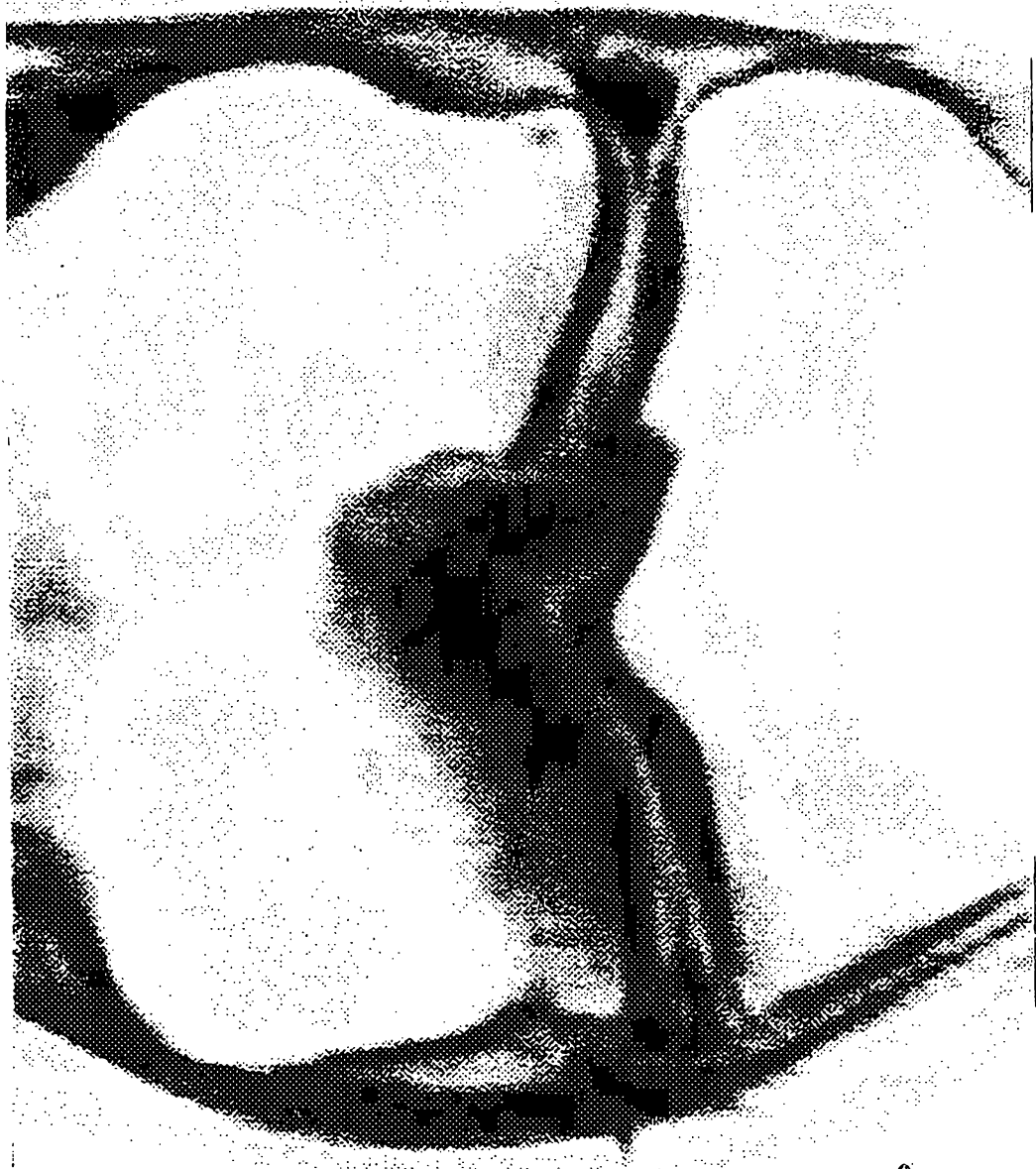


FIG. 6B