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(54) Title: METHODS FOR BONE REGENERATION USING ENDOTHELIAL PROGENITOR CELL PREPARATIONS

(57) Abstract: The present invention relates in general to orthopedics and to a method for promoting repair of large bone defects, in particular non-union or delayed union fractures. Specifically the invention concerns the use of endothelial progenitor cell preparations for bone repair.

METHODS FOR BONE REGENERATION USING ENDOTHELIAL PROGENITOR CELL PREPARATIONS

FIELD OF THE INVENTION

5 The present invention relates to a method for treating bone defects, in particular for promoting repair of large bone defects, including non-union and delayed union fractures. Specifically the invention concerns compositions comprising endothelial progenitor cells and methods of use thereof for bone repair.

10 BACKGROUND

Bone is a unique type of tissue composed of an inorganic mineral phase and cellular and extracellular matrix phases. Bone is a vital organ that undergoes modeling and remodeling wherein old bone is lost (resorption) and new bone is formed (replacement).

15 In children and young adults, bone remodeling results in the growth and increase in density of the skeleton. In adults, remodeling normally results in no net change in skeletal size since bone replacement matches bone resorption. Osteoporosis and related diseases ensue when bone resorption exceeds bone replacement. Bone restoration or replacement is a viable consideration in indications including osteopenia, osteoporosis,
20 bone tumors, spinal fusion and fractures.

Each year, more than 6.3 million people in the U.S. experience bone fractures, of which almost 1 million require hospitalization. Natural healing of bone with mechanical fixation can, in most cases, adequately mend minor fractures over time. However, in approximately 10% of all fractures, the defect is too large for the body's natural healing
25 response, and delayed unions or non-unions develop at the fracture site (Bancroft and Mikos, 2001; Rozen et al., 2007). In these cases, supplementary bone material may be required to fill in the defect and restore structure and function.

Current therapies for treatment of severe bone defects

30 The standard treatment for healing severe bone defects is transplantation of autologous bone tissue (autografts) (reviewed in Mistry and Mikos, 2005). However, this process has several drawbacks including: limited body sites from which bone may

be harvested without loss of function; autografts are less effective in irregularly shaped defects; and the procedure may be associated with complications such as infection, pain and nerve injury. Allografts derived from cadavers are another commonly used bone graft material. However, un-processed grafts carry a risk of disease transmission and
5 immune rejection, while demineralized bone matrix is poor in bone growth inducing factors. Xenografts are also a poor option due to the danger of disease transmission or rejection.

Metal implants may be permanently placed in bone to fill a defect however corrosion, infection and poor implant-tissue interface create many problems.

10 Ceramics may also be used in the treatment of bone injuries. While offering excellent biocompatibility, they are often brittle and degrade too slowly thereby inhibiting natural bone re-growth.

Another potential treatment is distraction osteogenesis, which entails the lengthening of limbs across a defect through temporary external fixation devices.

15 As outlined above, the most advanced treatments are limited in effectiveness and are often associated with complications. Thus, there is a significant need for an alternative strategy for the treatment of severe bone loss or fracture.

Cell-based therapies

20 An emerging approach to damage repair is tissue engineering, which involves treatment with one or more of the following elements: cells, signaling molecules and scaffolds. Thus, by employing the body's natural healing response, a bone defect may be replaced by natural bone tissue in the absence of an exogenous permanent implant (Bancroft and Mikos, 2001; Rozen et al., 2007).

25 Cell-based strategies for bone tissue engineering involve, *inter alia*, the transplantation of osteogenic cells. Cell-based therapies may include fresh bone marrow, as well as mesenchymal stem cells (MSCs) expanded in culture, or differentiated osteoblasts. Autologous bone-marrow injected into patients' un-connected tibial fractures with fixation demonstrated efficacy equal to that of autografts (Connolly et al., 1991).

A key factor for the success of bone marrow in healing non-union fractures is the presence of MSCs. The limited quantity of MSCs in marrow has led to the development of methods to isolate progenitor cells from bone marrow and expand them *in-vitro*, prior to transplantation.

5 Richards et al., (1999) demonstrated the osteogenic capabilities of cultured MSCs in a collagen gel carrier injected into distraction gaps of rats. Kadiyala et al., (1997) loaded MSCs onto ceramic cylinders and implanted them into critical-sized defects in rat femora. In US 6,541,024 and 6,863,900 to Kadiyala et al. regeneration and augmentation of bone repair following administration of MSCs was disclosed.

10 Isolated MSCs in culture can be selectively differentiated into osteoblasts with media supplements (Bruder et al., 1997; Pittenger et al., 1999). These osteogenic cells, when combined with a biomaterial carrier, can begin bone reconstruction immediately upon delivery to the injury site. For example, when rat marrow stromal cells cultured on porous hydroxyapatite scaffolds with osteogenic supplements were implanted
15 subcutaneously in rats, rapid bone formation was observed (Yoshikawa et al., 1996).

US 5,811,094 (Caplan et al.) provides a method for isolating, purifying, culturing and expanding human mesenchymal stem cells (MSCs) for the purpose of repairing connective tissue defects (including bone and cartilage repair). US 6,355,239 (Bruder et al.) provides methods and preparations for promoting connective tissue
20 growth, including bone, by transplanting allogeneic, mesenchymal stem cells.

US 7,029,666 (Bruder et al.) demonstrates use of non-autologous MSCs for treatment and regeneration of connective tissue and enhancement of bone marrow engraftment.

US 5,972,703 (Long et al.) discloses a process for preparing an enriched
25 population of bone precursor cells (expressing osteocalcin or osteonectin) obtained from bone marrow for promoting bone fracture repair.

Endothelial progenitor cells

Endothelial progenitor cells (EPCs) have been identified in adult bone marrow as well as in peripheral blood and human umbilical cord blood, and have been shown to maintain their
30 potency to proliferate and to differentiate into mature endothelial cells (Ashara et al., 1997; Murohara et al., 2000). Vasculogenesis, the development of new blood vessels during

embryogenesis begins with the formation of blood islands comprising endothelial progenitor cells (EPCs) and hematopoietic stem cells (Risau, 1997; Risau, 1995; Risau et al., 1988; Flamme et al., 1992; Hatzopoulos et al., 1998; Doyle et al., 2006; Ribatti, 2007).

EPCs have been shown to participate in postnatal neovascularization (Takahashi et al., 1999; Isner and Asahara, 1999). Furthermore, EPCs were found to participate in angiogenesis, vascular repair and vasculoprotection (Humpert et al., 2005; Doyle et al., 2006).

Recent studies have shown that EPCs significantly participate in constructing endothelium of new vessels in situations of tissue regeneration such as burns, bypass coronary artery grafting, and acute myocardial infarction. In these instances, bone marrow-derived EPCs are recruited to the blood circulation and home to injured and regenerating tissues for their participation in the buildup of new blood vessels. For example, addition of a purified and *ex-vivo* expanded population of these cells to nude mice with hind limb ischemia improved blood flow recovery and reduced limb loss (Kalka et al., 2000). Moreover, growth factors and ischemic conditions augment the number of circulating EPC (Takahashi et al., 1999).

EPCs can be identified by tube formation in Matrigel™ (Bellik et al., 2005), acetylated low-density lipoprotein (Ac-LDL) incorporation or expression of characteristic cell markers including Tie-2, CD34+ and von Willebrand factor (vWf) (Neumuller et al., 2006).

Early studies in a distraction osteogenesis model in sheep described the appearance of cellular colonies of vascular nature (immunopositive for Tie-2 and factor VIII-related antigen), the origin of which was not clear (Rachmiel et al., 2002; Lewinson et al., 2001(a, b)).

Cetrulo et al., (2005) demonstrated that distraction osteogenesis in a rat mandible model results in the generation of an ischemic region, to which concomitantly injected human EPCs were shown to home.

US 6,878,371 to Ueno et al. provides methods of forming new blood vessels in diseased or damaged tissue, specifically cardiac muscle comprising transplanting locally autologous bone marrow mononuclear cells.

Vascular injury was shown to promote an increase in circulating endothelial cells and EPCs (Hunting et al., 2005). In another study, bone marrow derived circulating EPCs were used to enhance angiogenesis following tissue ischemia (Werner et al. 2006).

US 6,720,340 (Cooke et al.) discloses recruitment of bone marrow derived EPCs and hematopoietic stem cells to the site of deficiency or injury, by administration of nicotine or nicotine receptor agonists.

5 Nowhere in the background art is it taught or suggested that local transplantation of autologous endothelial progenitor cells augments bone regeneration and may therefore be used as a therapeutic strategy for critical-gap bone fracture repair. There remains an unmet need in the medical community for an effective treatment for non-union and delayed union fractures.

10 SUMMARY OF THE INVENTION

The present invention provides methods for treating bone defects comprising administering to the site of bone defect a composition comprising endothelial progenitor cells (EPCs). In contrast to other prevailing techniques that employ scaffolds into the area of bone defect, the current invention entails implantation of endothelial progenitor
15 cells. Unlike MSCs, which are known to differentiate into bone cells, the current invention reveals the surprising finding that endothelial progenitor cells, which are known to promote vasculogenesis and angiogenesis, can also successfully promote bone regeneration. The present invention thus describes for the first time a therapeutic use of endothelial progenitor cells for repair of large bone defects.

20 In one of its aspects the present invention is directed to a method of repairing bone defects in a subject in need thereof by administering to the subject an effective amount of a cell preparation comprising endothelial progenitor cells (EPCs). EPCs can be identified by tube formation in Matrigel™, acetylated low-density lipoprotein (Ac-LDL) incorporation or expression of characteristic cell markers including Tie-2, CD34+
25 and von Willebrand factor (vWf).

In one embodiment the EPCs are injected systemically into the subject's blood circulation and home towards the bone defect area.

In a preferred embodiment the step of administering the cell preparation comprises applying said cell preparation locally to the site of bone defect. Local
30 administration assures that the administered EPCs remain in the desired area, thereby allowing a direct and rapid effect of the cells on the injured tissue.

In one embodiment the administered EPCs are allogeneic, i.e. the cells originate from a member of the same species but not from the individual being treated.

In a preferred embodiment the administered EPCs are autologous, i.e. the cells are recovered from the individual being treated for the bone defect. The cells are
5 isolated from the bone marrow or the peripheral blood and expanded ex-vivo to reach the desired cell number.

According to various embodiments the method of the invention is suitable for the repair of large bone defects, for example, non-union or delayed fractures, and critical sized bone defects caused by trauma, bone resection or radiotherapy.

10 In a certain embodiment of the present invention, the subject is a mammal. In a preferred embodiment the subject is human.

EPCs may be obtained from various sources including but not limited to bone marrow, peripheral blood and umbilical cord blood. Preferably, the EPCs are obtained by isolating mononuclear cells (MNC) from either the bone marrow, or from peripheral
15 blood and expanding these cells *in-vitro* under conditions favorable for the expansion of endothelial cells. The cells may be derived from human CD34⁺ mononuclear cells (EPC-enriched fraction).

In a particular embodiment a suspension of 10^6 - 10^{11} EPCs per ml in a pharmaceutically acceptable carrier is applied to the site of bone defect. In a preferred
20 embodiment the EPCs are applied in a concentration of 10^7 - 10^{10} cells/ml. In a more preferred embodiment the EPCs are applied in a concentration of 10^8 - 10^9 cells/ml.

The administration of EPCs may be performed at any time after the injury. According to some embodiments the EPCs are administered at least 10 days after the injury. According to another embodiment the EPCs are administered at least two weeks
25 after the injury.

In a particular embodiment the EPCs may be administered in combination with other cell types, e.g. mesenchymal stem cells.

The EPCs may be administered alone or in combination with a conductive i.e. support material. Non-limiting examples of such conductive materials include paste
30 (e.g. amorphous calcium phosphate paste, hydroxy apatite, calcium sulfate paste and demineralized bone), a natural or synthetic suitable scaffold (e.g. a fibrin matrix), a

viscous milieu based on a biopolymer such as hyaluronic acid or a combination of these materials. Examples for suitable fibrin matrices may be found for example in patents US 7,009,039, WO 2004/067704 and WO 2006/008748 assigned in part to the assignee of the present invention.

5 The EPCs may be administered in combination with an inductive material that would enhance their expansion *in-vivo*. Non limiting examples of inductive agents include growth factors. Examples for growth factors include: vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF), epidermal growth factor (EGF), insulin-like growth factor 1 (IGF1), bone morphogenetic proteins (BMP), and
10 transforming growth factor (TGF). In one embodiment EPCs are expanded *ex-vivo* with FGF, preferably with FGF2.

In another aspect, the present invention uses EPCs for the preparation of a pharmaceutical composition for repairing bone defects in a subject in need thereof. In yet another aspect, the present invention provides a pharmaceutical composition
15 comprising EPCs and a pharmaceutically acceptable carrier for the treatment of bone defects.

These and further aspects of the current invention will be better understood in conjunction with the figures, detailed description, examples and claims that follow.

20 **BRIEF DESCRIPTION OF THE FIGURES**

Fig. 1 is a microphotograph of human EPCs 3 days in culture (A, B) and after trypsinization and reseeded about 2 weeks afterwards (C, D). The cells are shown at a magnification of x 4 (A, C) and x 40 (B, D).

25 Fig. 2 is a microphotograph of capillary-like structures formed by human umbilical vein endothelial cells (HUVEC) (A) and sheep EPC (B) plated in MatrigelTM. The cells are shown at a magnification of x 10.

Fig. 3 is a microphotograph of sheep EPC positively stained for cytoplasmic Von Willebrand factor (vWf) (left panel). The right panel shows cells stained without the primary anti-vWf antibody (as negative control). The cells are shown at a
30 magnification of x 10.

Fig. 4 is a microphotograph of sheep EPC (A) and HUVEC (B-D) stained with Dil-Ac LDL (A, B) and lectin (C). D shows double staining of HUVEC with Dil-Ac LDL and lectin. The cells are shown at a magnification of x 10.

Fig. 5 is an X-ray radiograph of sheep tibia with a critical gap of 3.2 cm at the day of bone removal (A). (B) is an X-ray radiograph of the critical gap taken two weeks after bone removal prior to EPC or paste implantation.

Fig. 6 is an X-ray radiograph of the critical gap taken 2 weeks post treatment with EPC and/or paste versus control. (A) control; (B) treated with EPC; (C) treated with paste; (D) treated with both paste and EPC.

Fig. 7 is an X-ray radiograph of the critical gap taken 4 weeks post treatment with EPC and/or paste versus control. (A) control; (B) treated with EPC; (C) treated with paste; (D) treated with both paste and EPC.

Fig. 8 is an X-ray radiograph of the critical gap taken 8 weeks post treatment with EPC and/or paste versus control. (A) control; (B) treated with EPC; (C) treated with paste; (D) treated with both paste and EPC.

Fig. 9 is an X-ray radiograph of the critical gap taken 12 weeks post treatment with EPC and/or paste versus control. (A) control; (B) treated with EPC; (C) treated with paste; (D) treated with both paste and EPC.

Fig. 10 is a Micro Computed Tomography (μ CT) 3D visualization of the critical gap 12 weeks post implantation versus control. (A) control; (B) treated with EPC; (C) treated with paste; (D) treated with both paste and EPC.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed to treatment of skeletal defects, in particular large bone gaps and non-union fragments, by implantation of endothelial progenitor cells (EPCs). This therapeutic method takes advantage of the natural healing process, enhanced by the addition of these growth (regeneration) promoting cells.

Several methods have been reported in the art to induce bone repair or enhance bone regeneration by implanting bone marrow or bone marrow derived mesenchymal stem cells (MSCs). MSCs are multipotent stem cells that can differentiate into a variety

of cell types including osteoblasts, chondrocytes, myocytes, adipocytes, neuronal cells, and, as described lately, into pancreatic beta-islets cells. Since these cells can differentiate into bone cells, it is not surprising that they would enhance bone regeneration. Without wishing to be bound by any theory, the current invention
5 discloses for the first time the finding that endothelial progenitor cells, which are known to promote vasculogenesis and angiogenesis, can also successfully promote bone regeneration.

The extraction of MSCs from peripheral blood is extremely difficult, and extraction from bone marrow yields only a limited quantity. In contrast, EPCs may be
10 easily extracted from peripheral blood and expanded *ex-vivo* to obtain a sufficient quantity for implantation.

The advantages of this method include the following:

- i) Regenerative capacity of EPCs, which are easily isolated and expanded *ex-vivo*.
- 15 ii) The surrounding healing tissue serves as a natural scaffold.
- iii) Usage of autologous cells eliminates disease transmission and immunologic rejection and also minimizes the chances of infection.
- iv) Implantation of endothelial progenitor cells directly to the damaged tissue reduces homing to other tissues thereby allowing efficient beneficial effect of the EPCs
20 and also reducing the risk of promoting vascularization.
- v) Obviates requirement for carrier, though the EPCs may be used in conjunction with methods employing scaffolds or carriers.

Bone is a vital organ that undergoes modeling and remodeling wherein old bone is lost (resorption) and new bone is formed (replacement). Although bone has an
25 inherent capacity for repair and regeneration when damaged by disease or trauma, the renewed bone is often fragile and not weight bearing. Bone restoration or replacement is a viable consideration in indications including osteopenia, osteoporosis, bone tumors, spinal fusion, fractures and non-union fractures.

The process of bone formation is initiated by endochondral ossification and
30 intramembranous ossification. Endochondral ossification is the fundamental mechanism

for longitudinal bone formation whereby cartilage is replaced by bone. It requires the sequential formation and degradation of cartilaginous structures in the growth plates that serve as templates for the developing bones. During intramembranous ossification bone is formed directly in the connective tissues. Both processes require the infiltration
5 of osteoblasts and subsequent matrix deposition. Bone formation may be enhanced either by recruiting osteoblasts, the bone forming cells, or by inhibiting recruitment or activity of osteoclasts, the bone resorbing cells. Osteoblasts and osteoclasts work together in a coordinated fashion to form and remodel bone tissue.

It is within the scope of the present invention to utilize compositions comprising
10 EPCs, i.e. the compositions of the present invention, together with any such matrices or scaffolds as are known in the art.

Many materials have been suggested for bone repair, specifically materials that avoid the harvesting problems associated with autologous matter and the health risks associated with allogeneic material. Inorganic materials such as calcium phosphate have
15 been utilized as bone and dental fillers (reviewed in LeGeros, Clin Orthop 395:81-98, 2002). Apatite, a particulate calcium phosphate, is particularly appealing by virtue of the fact that it is the naturally occurring mineral component in bone and teeth. Bone apatite exhibits low crystallinity due to the presence of magnesium and carbonate (CO_3) ions. Lack of crystallinity in apatites is associated with increased solubility in vivo.
20 Hydroxyapatite, in contrast, exhibits high crystallinity and represents a small component of natural bone. Synthetic bone substitute materials comprising calcium phosphate or hydroxyapatite have been disclosed for use as bone grafts implants and cements. Highly crystalline form of hydroxyapatite can be produced by solution precipitation followed by sintering at high temperatures.

25 Various types of inorganic fillers or matrices that may be used in bone repair are well known. For example, US 4,880,610 to Constantz teaches a method for producing an injectable calcium phosphate mineral bone-like material using highly concentrated phosphoric acid, a calcium source and a neutralizing source, to which various additives may be incorporated, including sugars or proteins such as collagen, fibrinogen or
30 elastin. US 5,650,176; 5,676,976 and 5,683,461 to Lee et al. teach the synthesis of reactive amorphous calcium phosphates (ACP) and their use for promoting bone growth.

It is further to be explicitly understood that the compositions and methods of the present may be used alone or in conjunction with other compositions and methods for bone repair as are known in the art. Thus, the EPCs are beneficial alone, but may be used together with other known cell types including but not limited to MSCs.

5 Furthermore the EPCs may be administered in a composition comprising inert ingredients or within a composition comprising additional active ingredients such as known growth factors. The compositions comprising EPCs may be administered to the individual systemically or locally. According to currently preferred embodiments the administration is designed for local administration. For local administration it may be

10 advantageous to add a supportive matrix or scaffold to assist in retaining the cells at the site of injury or fracture.

Definitions

For convenience and clarity certain terms employed in the specification, examples and claims are described herein.

15 As herein, the terms "bone defect" or "bone disorder" is meant an imbalance in the ratio of bone formation to bone resorption, such that, if unmodified, the subject will exhibit less bone than desirable, or the subject's bones will be less intact than desired. Bone deficit may also result from mutation, fracture, surgical intervention or from dental or periodontal disease.

20 Bones have a tremendous capacity for self-healing, and with proper treatment, most fractures will heal without complication. However, some fractured bones have difficulty healing. A "non-union fracture" occurs when a broken bone does not heal. When a bone is slow to heal, it is termed "delayed union fracture".

The term "critical bone gap" or "large bone gap" refer to bone gaps which are

25 too large for spontaneous bone regeneration and require external interference.

Non limiting examples for bone defect are delayed or non-union fractures. Additional non-limiting examples are large bone defects caused by trauma, bone resection or radiotherapy.

Pharmacology

The term "therapeutic" refers to any pharmaceutical, drug or prophylactic agent which may be used in the treatment (including the prevention, diagnosis, alleviation, or cure) of a malady, affliction, disease or injury in a patient.

Pharmaceutical compositions suitable for use in context of the present invention include compositions wherein the active ingredients are contained in an amount effective to achieve the intended purpose. More specifically, a "therapeutically effective amount" relates an amount of a compound sufficient to promote bone regeneration. Determination of a therapeutically effective amount is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein. The present invention contemplates pharmaceutical compositions, both for veterinary and for human medical use, which comprise as the active agent EPCs, as well as the use of EPCs in the manufacture of a medicament for the treatment of bone defects.

By the term "carrier" is meant any physiologically acceptable vehicle such as phosphate-buffered saline (PBS) or an analogous physiological solution including but not limited to biopolymers such as hyaluronic acid, viscous or non viscous vehicles, natural or synthetic vehicles, etc.

The dose of the pharmaceutical composition of the present invention may vary with the type of bone defect, the age of the patient, body weight, the route of administration, etc.; typically, it can be administered locally to the site of bone defect in a suspension of 10^6 - 10^{11} EPCs per ml of the pharmaceutically acceptable carrier.

Endothelial progenitor cells (EPCs)

The term "stem cell" refers to an undifferentiated cell that is capable of proliferation and further differentiation. Stem cells are capable of producing either new stem cells or cells called "progenitor cells" that differentiate to produce the specialized cells found in mammalian tissue and organs.

"Endothelial progenitor cells" (EPCs) refer to precursor cells produced in the bone marrow that are able of entering the bloodstream, reaching areas of blood vessel injury and participating in vascular repair. Studies have established that bone marrow-derived endothelial progenitor cells (EPCs) are present in the systemic circulation. They

were also found to contribute to targeted tumor therapy. The current study demonstrates that these cells are also capable of participating in bone repair.

EPCs may be isolated from any one of bone marrow, peripheral blood or umbilical-cord blood. Isolation from peripheral blood, which is performed by density gradient centrifugation, provides sufficient quantities of EPCs to permit their harvest and administration following *ex-vivo* expansion. After culturing for 7 days, EPCs can be identified by demonstrating positive fluorescence staining for 1,1'-dioctadecyl-3,3,3',3'-tetramethylindocarbocyanine (DiI)-labeled acetylated low density lipoprotein (Ac-LDL) (Kalka et al., 2000). In addition, EPCs are characterized by homogenous staining for von Willebrand factor (vWF). EPCs also demonstrated spindle-like structures during maturation (Neumuller et al., 2006)

Cultured EPCs have a few advantages over freshly isolated CD34 antigen-positive (CD34+) EPCs. First of all, the number of EPCs obtained by *ex-vivo* expansion exceed the number of CD34+ cells that can be freshly isolated. Secondly, the purity and quality of EPCs in a cultured population are superior to that of freshly isolated CD34+ cells; as CD34+ was originally described as the prototypical antigen expressed by both HSCs and endothelial lineage cells, hematopoietic cells may contaminate freshly isolated CD34+ cells. Indeed, pilot studies demonstrated that the extent of neovascularization achieved after transplantation of freshly isolated CD34+ cells was inferior to culture-expanded EPCs. Thirdly, for therapeutic strategies designed to employ transplanted cells that constitutively express pro- or anti-angiogenic factors, gene transfer of EPCs is facilitated by the use of culture-committed vs. less-differentiated CD34+ EPCs (Kalka et al., 2000).

According to one aspect the present invention relates to a method of repairing bone defects in a subject in need thereof by administering to said subject an effective amount of a cell preparation comprising endothelial progenitor cells (EPC).

In a preferred embodiment the cell preparation is administered locally to the site of bone defect. Local administration ensures a direct and rapid effect of the cells on the injured tissue, and reduces homing to other tissues. In addition, local administration may have another advantage compared to injection of cells into the blood circulation, since bone marrow-derived EPCs along with hematopoietic progenitor cells were shown to contribute to the vascularization of specific primary tumors (Lyden et al., 2001;

Kaplan et al., 2005). It may therefore be advisable to use local administration of the EPCs directly at the bone defect area.

The present invention is not limited by a method of preparing the implant. EPCs may be isolated from various sources including but not limited to bone marrow, peripheral blood and umbilical cord blood derived mononuclear cells (MNC). The cells
5 may be freshly isolated from a mammalian subject (CD34+ mononuclear cells, EPC-enriched fraction).

The term "autologous" as used herein, denotes that the cells are recovered from the individual being treated. The cells are isolated from the bone marrow or the
10 peripheral blood and expanded in culture to reach the desired cell number.

The term "allogeneic" as used herein refers to being genetically different although belonging to or obtained from the same species.

The term "mammal" as used herein indicates human treatment as well as veterinary use.

In a preferred embodiment of the current invention, EPCs are obtained by
15 isolating mononuclear cells (MNC) from either the bone marrow, or from peripheral blood of a mammal, expanding these cells *in vitro* under conditions favorable for the expansion of endothelial cells, and implanting the cells into the damaged tissue. In a preferred embodiment isolated EPCs are autologous to an individual in need of such
20 treatment.

The term "conductive material" as used herein refers to a material which helps convey the EPC to the site of the bone defect. Use of a conductive material aims to enhance the therapeutic effect of the EPCs, by providing a milieu conducive to their survival or by aiding in retention of the cells at the site in need of repair.

The EPCs may be administered alone or in combination with a conductive support material. Non-limiting examples of such conductive materials include paste (e.g. amorphous calcium phosphate paste, hydroxy apatite, calcium sulfate paste and demineralized bone), a suitable scaffold (e.g. a fibrin matrix), a viscous milieu based on
25 a biopolymer such as hyaluronic acid or a combination of these materials. Examples for
30 suitable fibrin matrices may be found for example in patents US 7,009,039; WO

2004/067704 and WO 2006/008748 assigned in part to the assignee of the present invention.

Amorphous calcium phosphate paste may be prepared according to the protocol exemplified hereinbelow, or by using any one of the other methods known in the art.
5 Highly crystalline form of hydroxyapatite can be produced by solution precipitation followed by sintering at high temperatures.

The term "inductive material" as used herein refers to a substance which enhances the therapeutic (regenerative) effect of the EPCs. The inductive material may act directly to promote bone regeneration or it may act by promoting proliferation of the
10 EPCs or both.

The EPCs may also be administered in combination with an inductive material such as a growth factor that would enhance their expansion in vivo. Non limiting examples of such growth factors are: vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF) and their variants, including FGF2, FGF4, FGF9 and
15 FGF18 which promote bone and cartilage healing, epidermal growth factor (EGF), insulin-like growth factor 1 (IGF1), bone morphogenetic proteins (BMP), and transforming growth factor (TGF).

FGFs comprise a large family of evolutionarily conserved polypeptides involved in a variety of biological processes including morphogenesis, angiogenesis, and tissue
20 remodeling as well as in the pathogenesis of numerous diseases. The various members of this family stimulate the proliferation of a wide spectrum of cells, ranging from mesenchymal to epithelial and neuroectodermal origin in vitro and in vivo. FGFs are expressed in a strict temporal and spatial pattern during development and have important roles in patterning and limb formation. FGFs are powerful mitogens and are
25 critical in the regulation of many biological processes including angiogenesis, vasculogenesis, wound healing, limb formation, tumorigenesis and cell survival.

FGFs and their analogs have been shown useful for treating indications including wounds (US 4,950,483; 5,859,208 and 6,294,359), myocardial infarction (US
4,296,100 and 4,378,347), skeletal disorders (US 5,614,496 and 5,656,598) and
30 remodeling cardiac tissue (US 6,352,971).

WO 02/36732 and WO 03/094835 assigned in part to the assignee of the present invention disclose FGF variants, having at least one amino acid substitution in the beta 8-beta 9 loop, useful for the preparation of medicaments aimed at bone and cartilage formation and regeneration, wound healing, neovascularization and treating FGFR related skeletal and proliferative disorders. By way of non-limiting examples, the FGF2-N111X variants, including FGF2(3,5Q)-N111X, are more potent mitogens than the native FGF2.

Growth factors may be administered at a wide range of concentrations, depending on the type of bone defect, the age of the patient, body weight, the route of administration, etc.

"Mesenchymal stem cells" or MSCs as referred to herein are multipotent stem cells which can differentiate into a variety of cell types. EPCs may also be administered in combination with other cell types, e.g. mesenchymal stem cells, which would enhance their regenerative effect.

The term "implantation" as used herein refers to the insertion of the composition of the invention into damaged tissue, whereby the implant serves to promote the regeneration of the tissue that has been damaged or removed.

EPCs can be injected systemically into the subject's blood circulation or implanted directly onto the area of bone defect. In a preferred embodiment the cell preparation is administered locally to the area of bone defect. Local administration ensures that most of the implanted EPCs reach the defected tissue and reduces homing to other tissues, thereby allowing a direct and rapid effect of the cells on the injured tissue. A further advantage of direct implantation over injection of cells into the blood circulation, is reducing the risk of contributing to the vascularization of specific primary tumors. Local administration may be performed via several procedures, including but not limited to open surgery and direct injection to the site of bone defect guided by either x-ray or an endoscope which is inserted through a small incision in the skin.

Another aspect of the invention is a pharmaceutical composition comprising EPC for repairing bone defects.

The following examples are intended to be merely illustrative in nature and to be construed in a non-limitative fashion.

EXAMPLESExample 1: *Ex-vivo* expansion and characterization of EPCIsolation and culture of EPC from bone marrow derived and peripheral blood derived
5 mononuclear cells (MNC)

EPC were prepared from sheep mononuclear cells (MNC). Mononuclear cells (MNC) were separated from the bone-marrow of a removed tibia segment (3.2 cm) or from samples of 10 to 40 ml of peripheral venous blood taken from the jugular vein, using Uni-Sep^{MAXI}® U-16 Ficoll™ (Novamed, Jerusalem, Israel) by density gradient
10 centrifugation at 400 x g for 30 min at room temperature (RT). The MNC fraction was collected, washed twice, by centrifugation at 200 x g for 15 min, with 45 ml Dulbecco's phosphate-buffered saline (DPBS, without calcium and magnesium; Biological Industries Ltd., Beit Haemek, Israel) containing 3% fetal calf serum (FCS; Biological Industries Ltd., Beit Haemek, Israel), and plated on fibronectin-coated six-well plates
15 (Sigma Chemical Co., St. Louis, MO). Cells were cultured in endothelial basal medium-2 (EBM-2; Clonetics, Walkersville, MD, USA), supplemented with EGM-2MV SingleQuot® containing 5% fetal bovine serum, vascular endothelial growth factor (VEGF), fibroblast growth factor-2, epidermal growth factor, insulin-like growth factor-1 and ascorbic acid. Cells were grown at 37°C with humidified 95% air/5% CO₂. The
20 initially seeded cells were round. After 3-5 days, attached cells appeared elongated and spindle shaped (Fig. 1).

After 6 days of culture, non-adherent cells were discarded by gentle washing with PBS, and fresh medium was applied. The attached cells were continually cultured with complete endothelial growth medium-2 (EGM-2 medium). Cells were fed 3 times
25 per week and split when confluent by brief trypsinization using 0.5% trypsin/0.2% EDTA (Biological Industries Ltd., Beit Haemek, Israel) and dispersed. The re-plated cells rapidly replicated from several cells to colonies and formed a monolayer with full confluence (Fig. 1). These cells were expanded and underwent serial passages while maintaining a homogenous appearance, and were characterized thereafter.

Characterization of mononuclear cell derived EPC

The incubated cells were identified as EPC using the following criteria: tube formation in Matrigel™, Ac-LDL incorporation and expression of von Willebrand factor (vWf).

5 Tube formation in Matrigel™: For demonstration of capillary tube formation, 250 µl of growth factor-reduced Matrigel™ (BD Biosciences Discovery Labware, Bedford, MA) was added per well of a 24-well plate and allowed to polymerize at 37°C for at least 30 min. Adherent cells (suspected to be EPC) or human umbilical vein endothelial cells (HUVEC, which serve as an endothelial cell positive control) were
10 trypsinized and 5×10^4 cells were re-suspended in 300 µl EGM2-MV medium and seeded onto Matrigel™ (Ingram et al. 2004). The cells were incubated at 37°C with humidified 95% air/5% CO₂ for 5-24 h. The tube networks were observed with an Olympus inverted microscope (Olympus, CKX41-RFA). When placed in Matrigel™-based media the cells rapidly formed capillary-like structures similar to those formed by
15 HUVEC in Matrigel™ (Fig. 2).

Acetylated LDL (Ac-LDL) incorporation: To confirm the EPC phenotype, adherent cells were incubated with 10 mg/ml Ac-LDL coupled with fluorescent 1, 1'-dioctadecyl-3, 3, 3', 3'-tetra-methyl-indocarbocyanide perchlorate (Dil-Ac-LDL) (Molecular Probes Inc., Eugene, Oregon, USA) for 3-24 hours (Ingram et al. 2004).
20 Cells were visualized with an Olympus CXX41 inverted fluorescent microscope. These cells demonstrated the ability to rapidly incorporate Ac-LDL (Fig. 4) indicating EPC characteristics. Also, Ac-LDL staining of sheep EPC was similar to the staining of HUVEC.

Von Willebrand factor (vWf): On day 7 of culture, adherent cells were
25 trypsinized, seeded and grown on chamber-slides for 2-3 days. Slides were subjected to immunocytochemistry to detect the expression of vWf. In brief, following fixation with 4% paraformaldehyde for 10 min at 4°C and endogenous peroxidase inactivation immunostaining was performed. After 3 washes with PBS, slides were incubated with rabbit anti human vWf diluted 1:50 (DAKO, Glostrup, Denmark) for 1 hour, RT.
30 Negative control slides were incubated without the antibody. After 3 washes with PBS, slides were incubated for 10 min with HRP polymer conjugate (SuperPicTure™

Polymer Detection Kit-Zymed Laboratories, San Francisco, CA, USA). To visualize the final immunoreaction products, AEC (RED) substrate (SuperPicTure™ Polymer Detection Kit-Zymed Laboratories, San Francisco, CA, USA) was incubated with the slides for 5 min. Immunohistochemical staining of these cells demonstrated the presence of vWf (Fig. 3)

Example 2: A critical gap model in sheep tibiae

A critical bone gap model was generated by removing a 3.2 cm bone segment from sheep tibia (Fig. 5). Anesthesia was performed in 13 sheep (aged about 2 years, 40-70 Kilograms) by 0.4 xylasin and 600 mg ketamin, induced by 200 mg propapoi and maintained by an intubation of 2% isofloran and oxygen ventilation. One gm cephalosporin was given during surgery and then amoxicillin 6 ml twice a day for the next 10 days. A longitudinal incision was performed along the skin of the anterior aspect of the right lower leg. The periosteum was similarly cut and elevated. Following the adjustment of a metal plate by 8 screws to the posterior aspect of the tibia, a segment of 3.2 cm was removed from the mid-diaphysis. This segment was preserved on ice for future isolation and culture of autologous EPC (as described above). Periosteum, fascia and skin were replaced and closed by sutures and pins and sprayed with antibiotics. A plaster cast was placed over the tibia and was removed one week later. All surgical procedures were approved by the Institutional Animals Health and Care Committee.

Example 3: Implantation of EPC or paste into a critical bone gap

Preparation of amorphous calcium phosphate paste

1.3 gr of Hydroxyapatite (ProChon Biotech) was dissolved in 10ml of phosphate buffered saline (PBS). Following filtration, 2.5gr of Hyaluronic acid (MW 3×10^6 in 1% concentration, ProChon Biotech) were added.

Amorphous calcium phosphate paste containing FGF

1.3 gr of Hydroxyapatite (ProChon Biotech) was dissolved in 10ml of phosphate buffered saline (PBS). 25 μ gr/ml FGF2(3,5Q)-N111G (an FGF2 variant disclosed in WO 03/094835 assigned in part to the assignee of the present invention) was precipitated on the Hydroxyapatite for 1hr at 37°C. Following filtration, 2.5gr of Hyaluronic acid (MW 3×10^6 in 1% concentration, ProChon Biotech) were added.

Two weeks after the removal of the bone segment, the sheep underwent a second operation using the same procedures. A nicely enveloped healing tissue was observed filling the whole segmental gap. A longitudinal wedged-shaped incision was made within the gap. Each sheep received a different treatment:

5 In 3 sheep the gap was filled with 200 microliter of phosphate buffered saline (PBS) and covered by the wedged-shaped excised tissue. These sheep served as segmental defect controls.

In 5 sheep the gap was filled by a suspension of about 2×10^7 EPC in 200 microliter of PBS (10^8 EPCs per ml) and covered by the wedged-shaped excised tissue

10 In one sheep the gap was filled by a suspension of about 2×10^7 EPC in 200 microliter of PBS (10^8 EPCs per ml) that was layered upon a layer of an amorphous calcium phosphate paste and then covered by another layer of the paste.

In one sheep the gap was filled only by amorphous calcium phosphate paste and in 3 additional sheep the gap was filled by amorphous calcium phosphate paste containing $25 \mu\text{g}/\text{ml}$ FGF2(3,5Q)-N111G (an FGF2 variant disclosed in WO 03/094835 assigned in part to the assignee of the present invention).

Example 4: Radiological follow-up:

In each sheep, the right lower leg was X-rayed every 2 weeks until the animal was sacrificed after 3 months of healing. Upon sacrifice, the healed gap with the limiting bone edges were excised and immersed in neutral-buffered formalin for 4 days which was then replaced by 70% alcohol. The specimens were then scanned by μCT (Micro Computed Tomography). This method is used for taking qualitative and quantitative total bone volume, mineralized bone volume and bone mineral density measurements.

25 Compared to control gaps, bone regeneration bridging the whole length of the defect was observed in all five experimental sheep. Figures 6-9 demonstrate time course of bone regeneration in one representative sheep from each treatment group. In sheep where the gap was implanted with EPCs, a weak cloudy opaqueness along the gap was observed already 2 weeks following the implantation of the cells (Fig. 6). It gradually increased in volume and density until complete bridging was observed by the time of sacrifice, 12 weeks after the implantation. In sheep where the gap was filled either with

paste, paste containing FGF or paste plus cells, the regeneration process was somewhat slower. However, in these sheep a marked bone formation bridged the gap by 12 weeks of healing (Figures 7-9). The control gap showed no bridging, except for a small radio-opaque focus after 4 weeks in the middle of the gap or minor mineralization along the plate, that did not develop any further. Table 1 shows quantitative evaluation of x-ray radiographs for gap mineralization at sacrifice for each sheep, where prox denotes an area 5 mm proximal to the gap center, and dist denotes an area 5 mm distal to the gap center. Bone formation was scored on a scale from 0 to +++, where 0 denotes negative results and +++ denotes complete bridging.

10

Table 1- Quantitative evaluation of gap mineralization at sacrifice

	center	prox	dist
Control	+	0	0
	+	+	+
	0	0	0
EPC	+++	++++	++++
	++++	++++	++++
	++++	++++	++++
	++++	++++	++++
	++++	++++	++++
Paste	+	++	++
Paste + FGF	+++	+++	+++
	++	++	+
	+++	+++	+
Paste + EPC	0	+	+

μCT 3D visualization (presented in figure 10) confirmed the x-ray results demonstrating a very dense trabecular bone formation in gaps filled with EPC, minor mineralization in control sheep and partial bridging/mineralization in gaps filled with either paste+FGF or paste+EPC.

Representative quantitative analyses of bone formation (μCT imaging system, Scanco Medical, Basserdorf, Switzerland, at a resolution of 36 micrometer), is presented in Table 2.

20

Table 2 - μCT quantitative analysis for representative sheep

Sheep Identification	Total Tissue Volume (mm³)	Bone Volume (mm³)	Material Density (mg HA/cm³)
Control	2240.2	1008.3	329.75
EPC	11002.8	7854.3	705.16
Paste	4444.6	2789.0	615.59
Paste + EPC	2729.2	2034.6	440.58

While certain embodiments of the invention have been illustrated and described, it will be clear that the invention is not limited to the embodiments described herein. Numerous modifications, changes, variations, substitutions and equivalents will be apparent to those skilled in the art without departing from the spirit and scope of the present invention as described by the claims, which follow.

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CLAIMS

1. A method of repairing a bone defect in a subject in need thereof the method comprising the step of administering to the subject a therapeutically effective amount of a cell preparation comprising endothelial progenitor cells.
- 5 2. The method according to claim 1 wherein the step of administering the cell preparation comprises applying said cell preparation locally to an area of bone defect.
3. The method according to claim 1 wherein the endothelial progenitor cells are allogeneic.
4. The method according to claim 1 wherein the endothelial progenitor cells are
10 recovered from the individual being treated.
5. The method according to claim 1 wherein the bone defect is a delayed or non-union fracture.
6. The method according to claim 1 wherein the bone defect is a large bone defect caused by trauma, bone resection or radiotherapy.
- 15 7. The method according to claim 1 wherein the subject is a mammal.
8. The method according to claim 7 wherein the subject is human.
9. The method according to claim 1 wherein the endothelial progenitor cells are obtained from bone marrow-derived mononuclear cells.
10. The method according to claim 1 wherein the endothelial progenitor cells are
20 obtained from peripheral blood- derived mononuclear cells.
11. The method according to any one of claims 9 or 10 wherein the endothelial progenitor cells are derived from human CD34⁺ mononuclear cells.
12. The method according to claim 2 wherein the endothelial progenitor cells are administered in a suspension of 10^6 - 10^{11} cells per ml of a pharmaceutically acceptable
25 carrier.
13. The method according to claim 12 wherein the concentration of endothelial progenitor cells ranges between 10^7 - 10^{10} cells per ml of the pharmaceutically acceptable carrier.

14. The method according to claim 12 wherein the concentration of endothelial progenitor cells ranges between 10^8 - 10^9 cells per ml of the pharmaceutically acceptable carrier.

5 15. The method according to claim 1 wherein the endothelial progenitor cells are administered at least 10 days after the injury.

16. The method according to claim 15 wherein the endothelial progenitor cells are administered at least two weeks after the injury.

17. The method according to claim 1 wherein said cell preparation further comprises mesenchymal stem cells.

10 18. The method according to claim 2 which further comprises administering to said subject a conductive material.

19. The method according to claim 18 wherein the conductive material is selected from a group consisting of a paste, a scaffold and a viscous biopolymer.

15 20. The method according to claim 19 wherein the paste is selected from the group consisting of an amorphous calcium phosphate paste, hydroxy apatite, calcium sulfate paste and demineralized bone.

21. The method according to claim 1 which further comprises administering to said subject an inductive material.

20 22. The method according to claim 21 in which the inductive material is a growth factor.

23. The method according to claim 22 wherein the growth factor is selected from the group consisting of vascular endothelial growth factor, fibroblast growth factor, epidermal growth factor, insulin-like growth factor 1, bone morphogenetic proteins, and transforming growth factor.

25 24. Use of endothelial progenitor cells for the preparation of a pharmaceutical composition for repairing bone defects in a subject in need thereof.

25. A pharmaceutical composition for repairing bone defects in a subject in need thereof comprising a cell preparation containing endothelial progenitor cells and a pharmaceutically acceptable carrier.

26. The pharmaceutical composition according to claim 25 wherein the cell preparation is suitable for local application to an area of bone defect.

27. The pharmaceutical composition according to claim 25 wherein the endothelial progenitor cells are allogeneic.

5 28. The pharmaceutical composition according to claim 25 wherein the endothelial progenitor cells are recovered from the individual being treated.

29. The pharmaceutical composition according to claim 25 wherein the endothelial progenitor cells are obtained from bone marrow-derived mononuclear cells.

10 30. The pharmaceutical composition according to claim 29 wherein the peripheral blood- derived mononuclear cells incorporate Ac-LDL following culture.

31. The pharmaceutical composition according to claim 29 wherein the peripheral blood- derived mononuclear cells express Von Willebrand factor (vWf) following culture.

15 32. The pharmaceutical composition according to claim 29 wherein the endothelial progenitor cells are derived from human CD34⁺ mononuclear cells.

33. The pharmaceutical composition according to claim 25 wherein the endothelial progenitor cells are obtained from peripheral blood derived mononuclear cells.

20 34. The pharmaceutical composition according to claim 33 wherein the peripheral blood- derived mononuclear cells incorporate Ac-LDL following culture.

35. The pharmaceutical composition according to claim 33 wherein the peripheral blood- derived mononuclear cells express Von Willebrand factor (vWf) following culture.

25 36. The pharmaceutical composition according to claim 33 wherein the peripheral blood- derived mononuclear cells form tubes in MatrigelTM following culture.

37. The pharmaceutical composition according to claim 33 wherein the endothelial progenitor cells are derived from human CD34⁺ mononuclear cells.

30 38. The pharmaceutical composition according to claim 26 wherein the endothelial progenitor cells are present in a suspension of 10⁶-10¹¹ cells per ml of the pharmaceutically acceptable carrier.

39. The pharmaceutical composition according to claim 38 wherein the concentration of endothelial progenitor cells ranges between 10^7 - 10^{10} cells per ml of the pharmaceutically acceptable carrier.

5 40. The pharmaceutical composition according to claim 39 wherein the concentration of endothelial progenitor cells ranges between 10^8 - 10^9 cells per ml of the pharmaceutically acceptable carrier.

41. The pharmaceutical composition according to claim 25 wherein said cell preparation further comprises mesenchymal stem cells.

10 42. The pharmaceutical composition according to claim 26 which further comprises a conductive material.

43. The pharmaceutical composition according to claim 42 wherein the conductive material is selected from a group consisting of a paste, a scaffold and a viscous biopolymer.

15 44. The pharmaceutical composition according to claim 43 wherein the paste is selected from the group consisting of an amorphous calcium phosphate paste, hydroxy apatite, calcium sulfate paste and demineralized bone.

45. The pharmaceutical composition according to claim 25 which further comprises an inductive material.

20 46. The pharmaceutical composition according to claim 45 in which the inductive material is a growth factor.

47. The pharmaceutical composition according to claim 46 wherein the growth factor is selected from the group consisting of vascular endothelial growth factor, fibroblast growth factor, epidermal growth factor, insulin-like growth factor 1, bone morphogenetic proteins, and transforming growth factor.

25 48. Use of a pharmaceutical composition according to any of claims 25-47 for the preparation of a medicament for treating bone defects in a subject in need thereof.

Figure 1A

Figure 1B

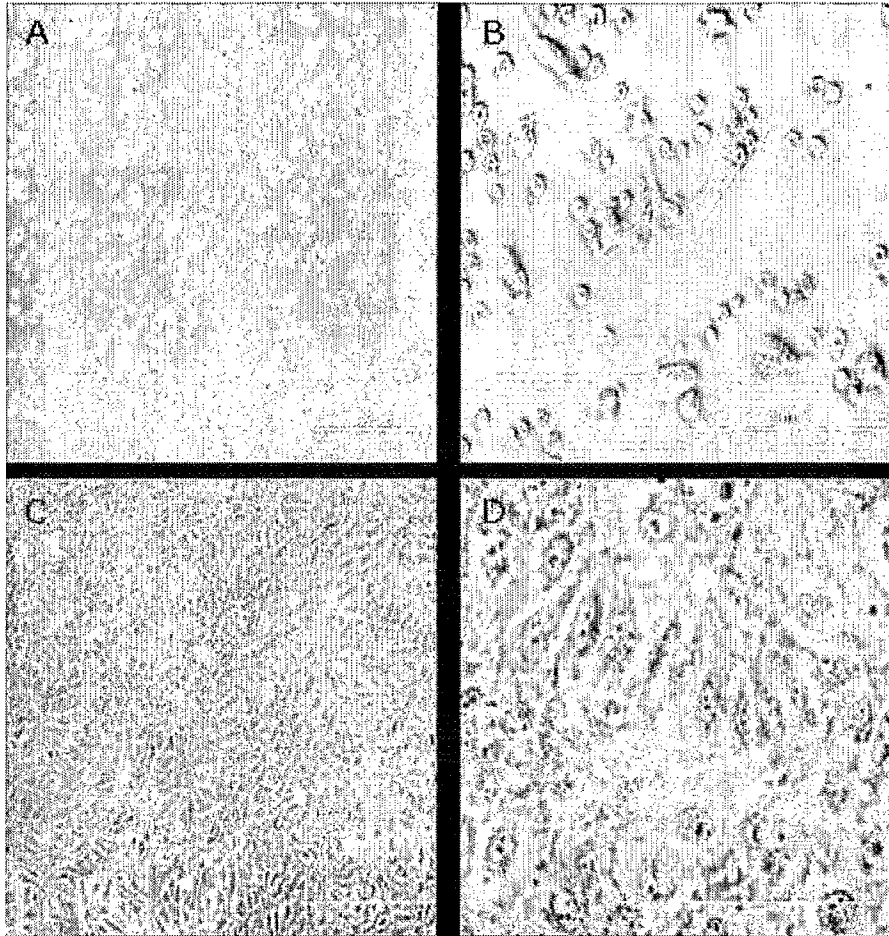


Figure 1C

Figure 1D

Figure 2A

Figure 2B

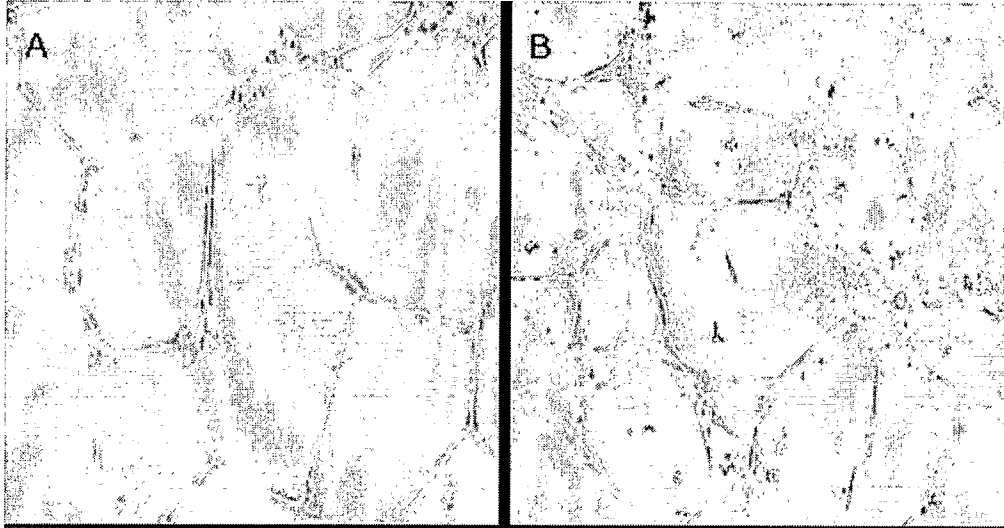


Figure 3A



Figure 3B

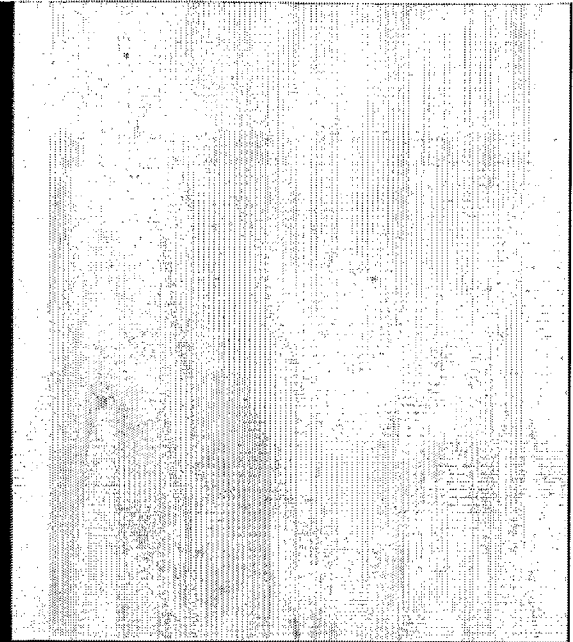


Figure 4A

Figure 4B

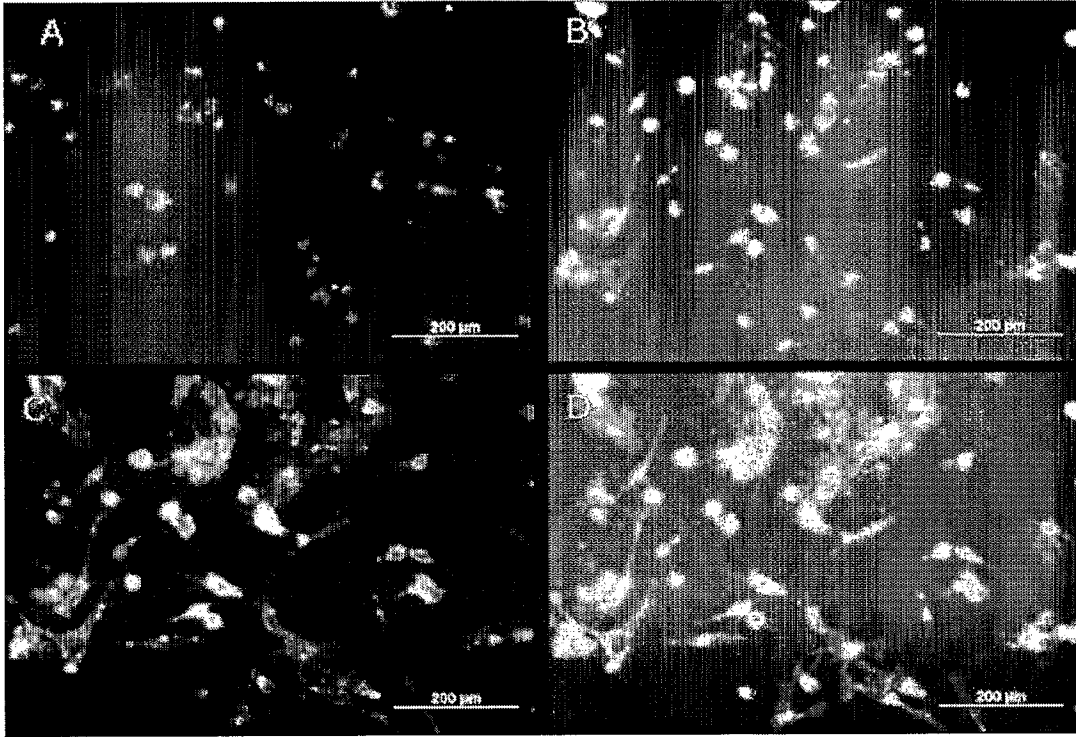


Figure 4C

Figure 4D

Figure 5A

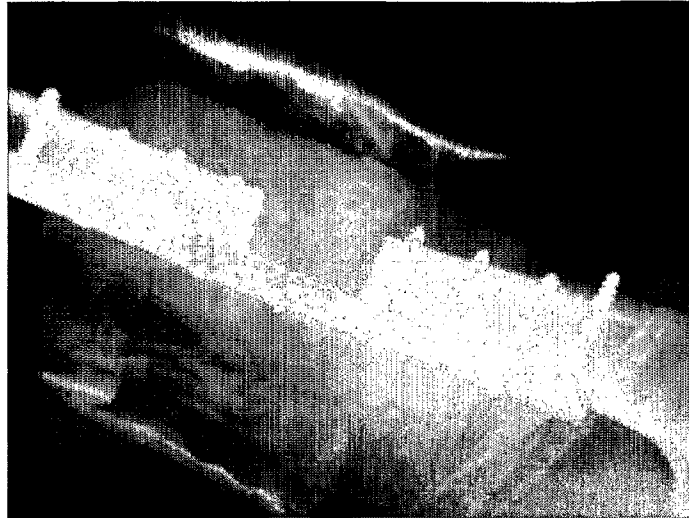


Figure 5B

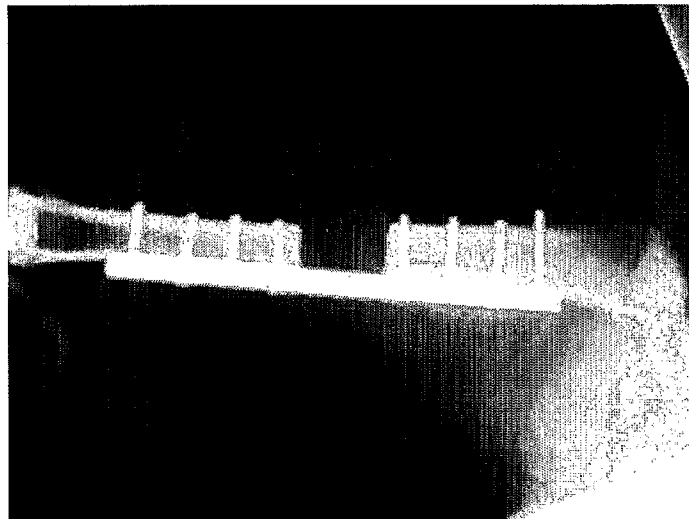


Figure 6A

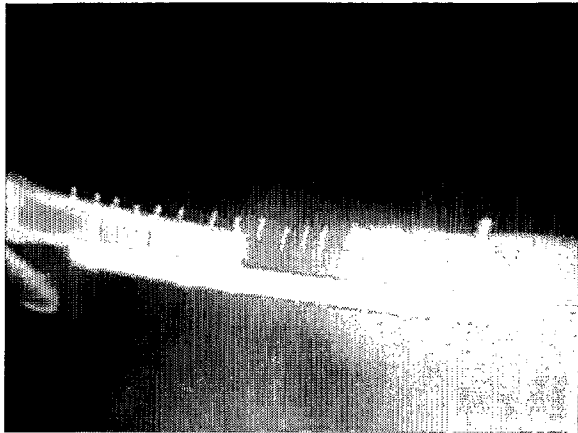


Figure 6B

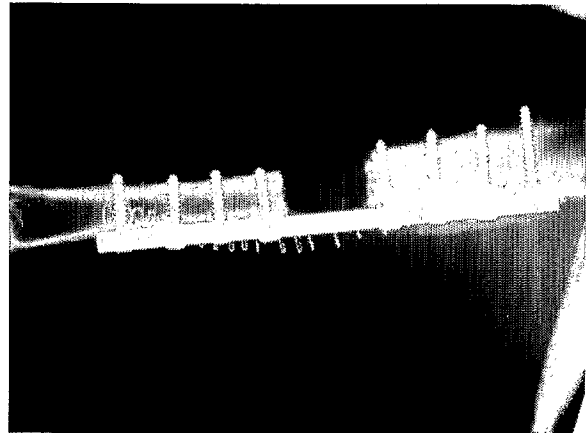


Figure 6C

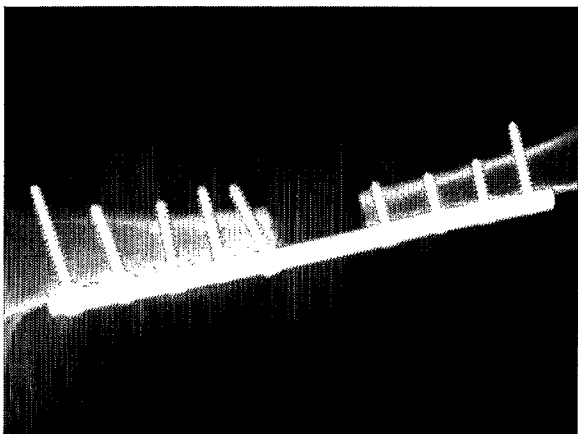


Figure 6D

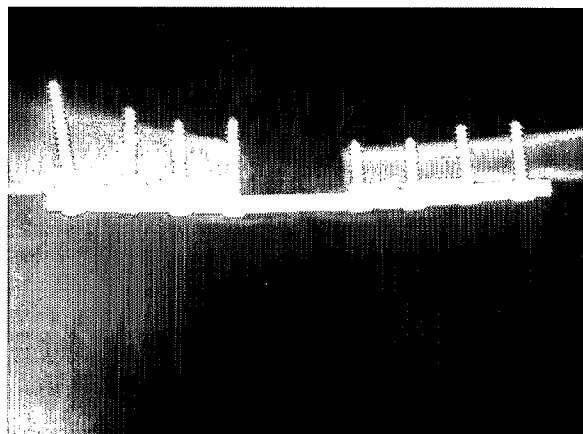


Figure 7A



Figure 7B

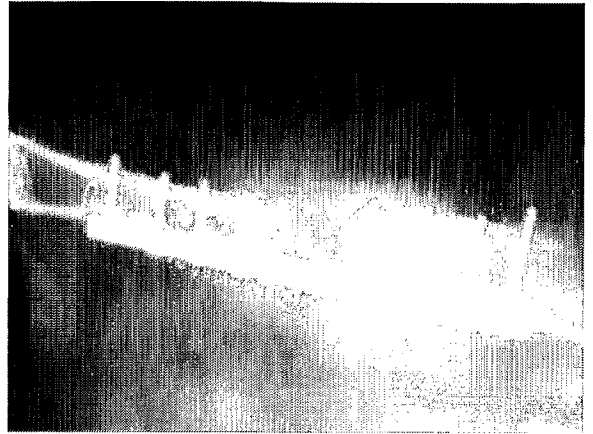


Figure 7C

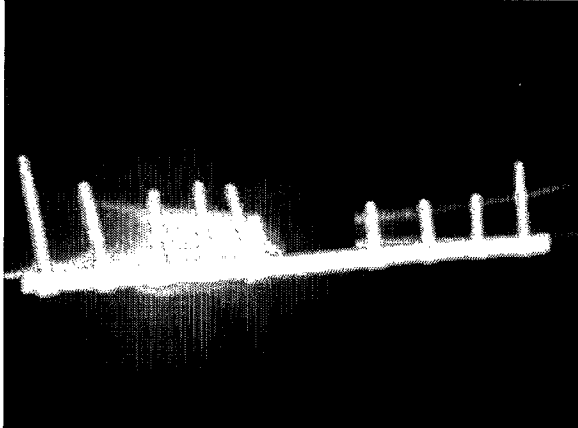


Figure 7D

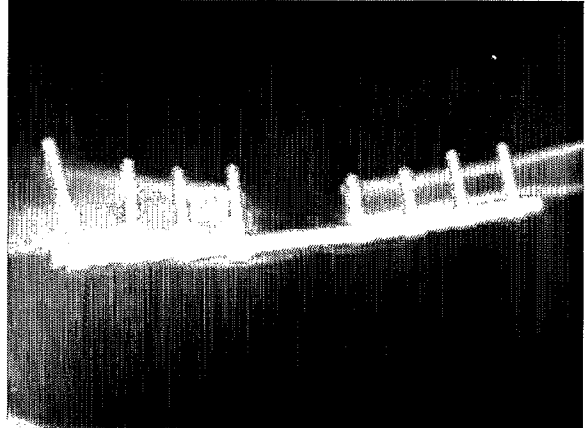


Figure 8A

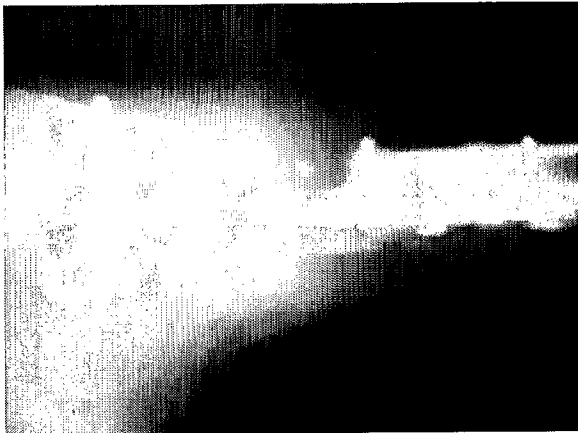


Figure 8B

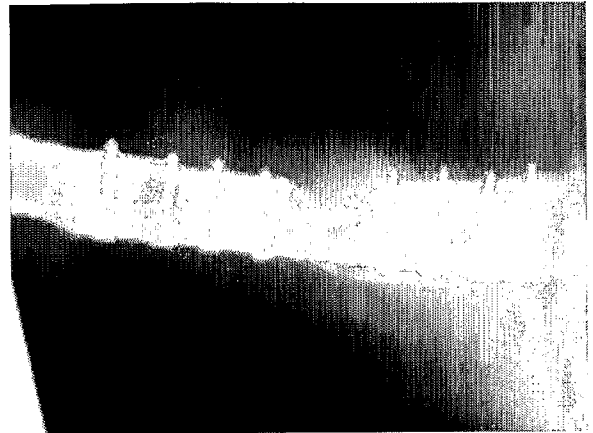


Figure 8C

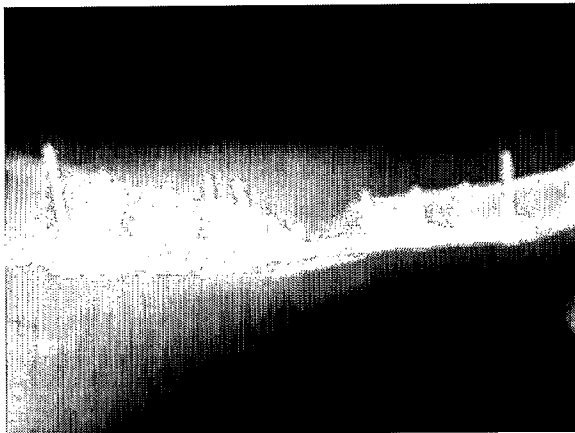


Figure 8D

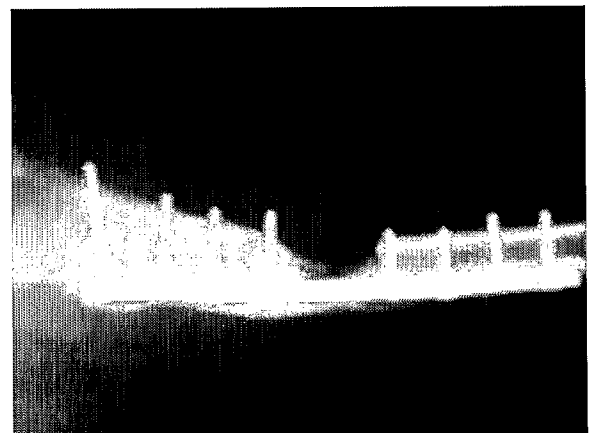


Figure 9

Figure 9A

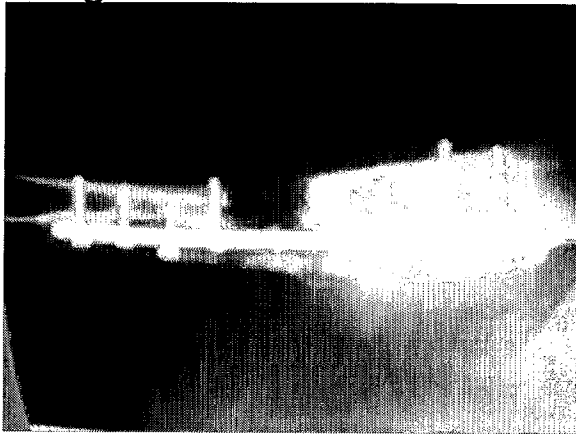


Figure 9B

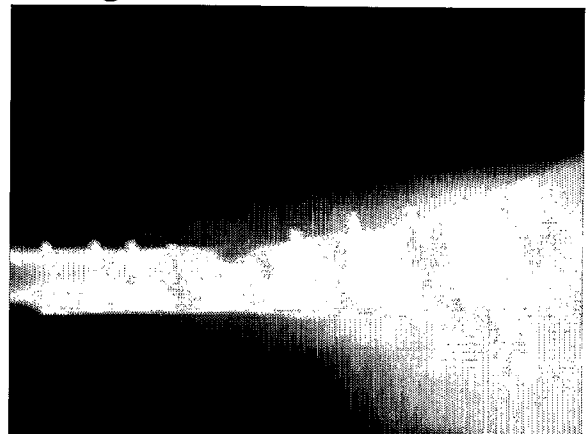


Figure 9C

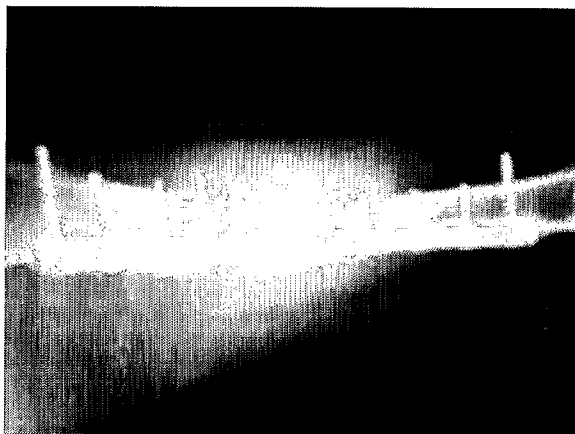


Figure 9D





Figure 10A

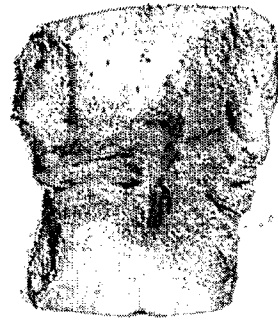


Figure 10B



Figure 10C



Figure 10D