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<p>(54) Title: PEPTIDES FOR USE IN VACCINATION AND INDUCTION OF NEUTRALIZING ANTIBODIES AGAINST HUMAN IMMUNODEFICIENCY VIRUS</p>		
<p>(57) Abstract</p> <p>Novel peptides are disclosed which correspond to epitopes of the HIV-1 gp120 protein. These antigenic peptides induce antibody-dependent cellular cytotoxicity against HIV, and thus are useful in immunization against HIV infection and induction of a heightened immune response to HIV.</p>		

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PEPTIDES FOR USE IN VACCINATION AND
INDUCTION OF NEUTRALIZING ANTIBODIES
AGAINST HUMAN IMMUNODEFICIENCY VIRUS

Background of the Invention

5 The present invention relates to peptides suitable for use in vaccination against AIDS.

 The human immunodeficiency virus (HIV) is responsible for the disease that has come to be known as acquired immune deficiency syndrome (AIDS). Although initially recognized in
10 1981, no cure has yet been found for this inevitably fatal disease. HIV is spread by a variety of means such as sexual contact, infected blood or blood products and perinatally. Due to the complexity of HIV infection and the paucity of effective therapies, eradication of AIDS will most likely
15 occur by preventing new infections rather than curing those persons already infected. To this end a great deal of effort has been expended in developing methods for detecting and preventing infection. Diagnostic procedures have been developed for identifying infected persons, blood and other
20 biological products.

 Like most viruses, HIV often elicits the production of neutralizing antibodies. Unlike many other viruses and other infectious agents for which infection leads to protective immunity, however, HIV specific antibodies are insufficient to
25 halt the progression of the disease. Therefore, in the case of HIV, a vaccine that elicits the immunity of natural infection could prove to be ineffective. In fact, vaccines prepared from the HIV protein gp160 appear to provide little immunity to HIV infection although they elicit neutralizing
30 antibodies. The failure to produce an effective anti-HIV vaccine has led to the prediction that an effective vaccine will not be available until the end of the 1990's.

 The HIV genome has been well characterized. Its approximately 10Kb encodes sequences that contain regulatory
35 segments for HIV replication as well as the gag, pol and env genes coding for the core proteins, the reverse transcriptase-

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protease-endonuclease, and the internal and external envelope glycoproteins respectively.

The HIV env gene encodes the intracellular glycoprotein, gp160, which is normally processed by proteolytic cleavage to form gp120, the external viral glycoprotein, and gp41, the viral transmembrane glycoprotein. The gp120 remains associated with HIV virions by virtue of noncovalent interactions with gp41. These noncovalent interactions are weak, consequently most of the gp120 is released from cells and virions in a soluble form.

Previous studies have shown that the proteins encoded by the gag and especially the env regions of the HIV-1 genome are immunogenic since antibodies to the products of the gag and env genes are found in the sera of HIV infected, AIDS and ARC ("AIDS Related Condition") patients.

It has previously been shown that some antibodies obtained from sera of AIDS and ARC patients, as well as asymptomatic individuals infected with the virus, are specific to gp120 and gp160. Occasionally these antibodies are neutralizing. The envelope glycoproteins are the HIV-1 antigen most consistently recognized by antibodies in AIDS and ARC patient sera. Allan et al., "Major Glycoprotein Antigens that Induce Antibodies in AIDS Patients are Encoded by HTLV-III," *Science*, 228:1091-1094 (1985); and Barin et al., "Virus Envelope Protein of HTLV-III Represents Major Target Antigen for Antibodies in AIDS Patients," *Science*, 228:1094-1096 (1985). In addition, antibodies in patient sera also recognize epitopes of the viral core proteins encoded by the gag gene.

Immunologically important HIV-1 antigens for use in diagnosis and as potential vaccine compositions have been prepared by cloning portions of the HIV-1 genome in various expression systems such as bacteria, yeast or vaccinia. Cabradilla et al., "Serodiagnosis of Antibodies to the Human AIDS Retrovirus With a Bacterially Synthesized env Polypeptide," *Biotechnology*, 4:128-133 (1986); and Chang et al., "Detection of Antibodies to Human T-Cell Lymphotropic

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Virus-III (HTLV-III) With an Immunoassay Employing a Recombinant Escherichia coli - Derived Viral Antigenic Peptide," Biotechnology, 3:905-909 (1985). HIV-1 antigens produced by recombinant DNA methods, however, must still be exhaustively purified to avoid adverse reactions upon vaccination and false positive reactions in ELISA assays due to any antibody reactivity to antigens of the expression system which may contaminate the HIV-1 antigen preparation. Also, denaturation of HIV-1 antigens during purification may destroy important antigen activity. Preparation of proteins from intact viruses can also result in contamination by intact virus.

Several publications have presented data showing immunologic reactivity of selected synthetic peptides corresponding to antigenic proteins of HIV-1. In one study, a peptide having the amino acid sequence Tyr-Asp-Arg-Pro-Glu-Gly-Ile-Glu-Glu-Gly-Gly-Glu-Arg-Asp-Arg-Asp-Arg-Ser-Gly-Cys which corresponds to amino acid residues 735-752 of HIV-1 was synthesized. Kennedy et al., "Antiserum to a Synthetic Peptide Recognizes the HTLV-III Envelope Glycoprotein," Science, 231:1556-1559 (1986). This peptide, derived from a portion of gp41, was used to immunize rabbits in an attempt to elicit a neutralizing antibody response to HIV-1. Furthermore, several sera from AIDS patients known to contain anti-gp41 antibodies were weakly reactive with this peptide, thus indicating that this peptide contains at least one epitope recognized, to some extent, by antibodies to native gp160/gp41. However, this peptide has not been shown to elicit neutralizing antibodies in mammals other than rabbits nor has it been suggested for use as a human vaccine.

In antigenic proteins of HIV-1 there are antigenic epitopes recognized by antibodies, cytotoxic T cells, helper T cells and also in antibody-dependent cellular cytotoxicity (ADCC). Traditionally, neutralizing antibodies are considered as essential in preventing viral infection. A neutralizing antibody binds to an infectious virus particle and in this process the infectivity of the virus particle is destroyed.

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Cellular mechanisms for elimination of virus infected cells involve cytotoxic T cells, T-helper cells and ADCC. The epitopes involved in neutralization and in the various cellular immune mechanisms need not necessarily be the same.

5 Previously it has been found that ADCC is an immunological defense mechanism that operates in viral infections. In this reaction, antigen-specific antibodies will bind to surface structures on the target cell and thus induce killing mediated by major histocompatibility complex
10 (MHC)-unrestricted CD16+, Fc receptor-bearing effector cells. HIV specific cytotoxicity in the peripheral blood of most seropositive individuals is also mediated by MHC-unrestricted ADCC effector cells which are armed with env-specific IgG antibodies, Tyler et al. J. Immunol., 142:1177 (1989); Tanneau
15 et al. J. Infect Dis., 162:837 (1990); Riviere et al. J. Virol., 63:2270 (1989). HIV-specific ADCC activity has been found in the majority of sera from HIV-1 infected individuals, Ljunggren et al. J. Immunol., 139:2263 (1987), Lyerly et al., AIDS Res. Hum. Retroviruses 3:409 (1987). Both type and
20 strain specific ADCC have been observed and antibodies in some sera mediated ADCC against all strains whereas other sera lacked ADCC activity completely, Ljunggren et al., 63:3376 (1989). In pediatric HIV-1 infection, presence of ADCC-mediating antibodies correlates significantly with a better
25 clinical stage, Ljunggren et al., 161:198 (1990). The ADCC reaction appears early after HIV-infection and broadly reacting ADCC against HIV-1_{HTLVIIIB} infected target cells appears between 2 and 12 months after seroconversion.

Activated cells expressing HIV antigens on their surface
30 are possible targets for ADCC. HIV-infected autologous CD4+ T-cell blasts have recently been shown to serve as targets for lysis by ADCC, Tanneau et al. J. Infect Dis., 162:837 (1990). The envelope glycoproteins of HIV have been suggested as target epitopes in a number of studies. Evans et al. AIDS,
35 3:273 (1989) used affinity purified human Ig or polyclonal rabbit sera against env proteins of HIV-1 and found antibodies mediating ADCC against gp120 and gp41. Koup et al. J Virol,

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63:584 (1989), have used vaccinia virus vectors expressing envelope glycoproteins (gp160, gp120 and gp41) or gag proteins (p55, p40, p24 and p17) in lymphoblastoid cell lines. Only the envelope glycoprotein complex gp120/gp41 was found to be the target antigen for HIV-specific ADCC which was also confirmed in another study using a similar system, Tanneau et al. J. Infect Dis., 162:837 (1990).

More defined regions have also been demonstrated in a number of studies. A murine monoclonal antibody directed to the V3 region (a.a. 309-318) of gp120 mediated both neutralization, titer 1:500, and ADCC, titer 1:800, against HTLVIIIIB. Brolden et al., J. Virol., 64:936 (1990). Also, a chimeric mouse-human antibody directed against the V3 region (a.a. 308-322) induced ADCC as well as neutralization and fusion inhibition, Liou et al. J. Immunol, 143:3967 (1989). Lysterly et al., AIDS Res Hum Retroviruses, 3:409 (1987), have localized an ADCC epitope in the C-terminal part of gp120 (a.a.467-511).

Summary of the Invention

In accordance with the present invention, novel peptides corresponding to epitopes of HIV-1 gp120 protein are disclosed and described. Each peptide comprises an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41, and wherein antisera raised in monkeys against the epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30.

In another embodiment of the present invention, each peptide has an epitopic sequence having an amino acid sequence that consists essentially of a sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41.

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In accordance with another aspect of the present invention, the novel peptides are used to formulate a vaccine composition. The vaccine composition comprises an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41, and wherein antisera raised in monkeys against the epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30, in an amount effective to induce an immune response in a mammal together with a pharmaceutically acceptable carrier. In a preferred embodiment, the vaccine composition further comprises an adjuvant such as Freund's complete adjuvant, Freund's incomplete adjuvant, muramyl dipeptide, levamisole, isoprinosine or tuftsin.

In accordance with yet another aspect of the present invention, at least two of the novel peptides are used in the vaccine composition. Each peptide comprises an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41, and wherein antisera raised in monkeys against the epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30. The peptides are present in an amount effective to induce an immune response in a mammal, and are combined with a pharmaceutically acceptable carrier.

In a preferred embodiment, this vaccine composition further comprises an adjuvant, such as Freund's complete adjuvant, Freund's incomplete adjuvant, muramyl dipeptide, levamisole, isoprinosine and tuftsin.

In accordance with yet another aspect of the present invention, there is disclosed a method of protecting a mammal from infection with human immunodeficiency virus, comprising

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administering to the mammal one of the vaccine compositions described herein. The administration can be by intravenous, intramuscular, subcutaneous, or intraperitoneal injection.

In accordance with still another aspect of the present invention, there is disclosed a method for inducing neutralizing anti-HIV antibodies in a mammal, comprising the step of administering an effective antibody-inducing amount of a composition comprising an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41, and wherein antisera raised in monkeys against the epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30, in an amount effective to induce an immune response in a mammal together with a pharmaceutically acceptable carrier.

In another embodiment of the invention, the vaccine composition comprises at least two of the novel peptides. Each peptide comprises an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41, and wherein antisera raised in monkeys against the epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30. The peptides are present in an amount effective to induce an immune response in a mammal, and are combined with a pharmaceutically acceptable carrier.

Detailed Description of the Invention

The present invention provides peptides which have been found to elicit production of HIV neutralizing antibodies by primate subjects. The peptides correspond to regions of the gp120 protein with coordinates as defined by Kennedy et al.

The peptides of the present invention are termed gp120-12 (amino acid coordinates 159-183), gp120-15 (amino acid coordinates 200-225), gp120-16 (amino acid coordinates 213-237) and gp120-19 (amino acid coordinates 255-276).
5 Although peptide gp120-19 is similar to a peptide that has been described (Ho et al., Science, 239:1021-1023 (1988)), it has now been found that gp120-19 elicits neutralizing antibodies in primates. The peptides of the present invention can be used as immunogens in vaccine compositions and to
10 elicit polyclonal or monoclonal antibody production; particularly important are HIV neutralizing antibodies.

Proteins contain a number of antigenic determinants or epitopes which are the regions of the proteins comprising the recognition and binding sites for specific antibodies. In
15 general, proteins contain between 5 to 10 epitopes, each of which contains a sequence of 6 to 8 amino acids. Epitopes can be either continuous, in which the 6 to 8 amino acids are present in linear sequence, or discontinuous, in which the amino acids that form the epitope are brought together by the
20 three dimensional folding of the protein. Even though an epitope constitutes only a relatively few amino acids, its reactivity with an antibody may be influenced by the amino acids in the protein which surround the epitope.

Studies aimed at mapping antigenic sites or epitopes of
25 proteins have been aided by the use of synthetic peptides corresponding to various regions of the proteins of interest. Lerner et al., in, The Biology of Immunological Disease: A Hospital Practice Book, (Dixon and Fisher, eds.) pp. 331-338 (1983); and Lerner, Adv. Immunol., 36:1 (1984). In addition.
30 to their usefulness in epitope mapping studies, synthetic peptides, if encompassing major antigenic determinants of a protein, have potential as vaccines and diagnostic reagents. Van Regenmortel, Ann. Inst. Pasteur/Virol 137E:497-528 (1986); and Van Regenmortel, Laboratory Techniques in Biochemistry and
35 Molecular Biology, Buroden and Van Knippenburg eds. Vol. 19, synthetic Peptides as Antigens, Elsevier ISBN 0-444-80974-0 (1988).

Synthetic peptides have several advantages with regard to specific antibody production and reactivity. The exact sequence of the synthesized peptide can be selected from the amino acid sequence of the protein as determined by amino acid sequencing of the protein or the predicted amino acid sequence determined from the DNA sequence encoding the protein. The use of specific synthetic peptides eliminates the need for the full-length protein in vaccination and the production of or assay for antibodies. Furthermore, the solid phase peptide synthetic techniques of Merrifield and coworkers allow for essentially unlimited quantities of the synthesized peptide of interest to be chemically produced. Erickson and Merrifield in *The Proteins*, 3rd Edit., Vol. 2, Academic Press, New York, Chapter 3 (1976). The availability of automated peptide synthesizers has further advanced such techniques.

Although a variety of criteria can be used to predict antigenic regions of proteins, peptides corresponding to such regions may not always be useful as vaccines. For example, antigenicity may be lost because the peptide is not in the proper spatial orientation to be recognized by antibodies which react with the protein. It has also been found that certain peptides derived from type C retroviruses and HIV act as immune-suppressive agents much as HIV itself. Cianciolo et al., *J. Immunol.*, 124:2900-2905 (1980); and Cianciolo et al., *Science*, 230:453-455 (1985). Peptides such as these, which have a deleterious effect on the patient, would not be suitable for use as vaccines.

Furthermore, as is particularly evident with HIV-1 and HIV-2, there is significant genetic variability within each of these two virus groups leading to many serotypes, or isolates, of the viruses. This has put a significant constraint on choosing a region of a protein from which to derive a peptide for use in formulating immunogens. However, certain immunodominant portions of HIV-1 and HIV-2 proteins have been found to be relatively invariant. Synthetic peptides may also be key to viral vaccines in that they may induce an immune response against type common sequences not normally

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immunogenic in the native molecule. These otherwise silent epitopes may be of broad protective specificity. Steward et al., *Immunol. Today*, 8:51-58 (1987). Several experimental vaccines have been formulated with the aim of preventing infection in those people who are likely to be exposed to the virus. Berman et al., "Protection of Chimpanzees from Infection by HIV-2 After Vaccination With Recombinant Glycoprotein gp120 but Not gp160," *Nature*, 345:622-625 (1990). Synthetic peptides corresponding to regions of immunologically important proteins of HIV have now found immediate use in diagnostic methods for detection of HIV, as potential vaccines for HIV and for the production of polyclonal and monoclonal antibodies.

A number of neutralization epitopes on gp120 have been found and defined by several investigators, for an overview see Bolognesi, *AIDS* (1989) 3(suppl 1):S111-s118. In this overview Bolognesi refers to four different virus neutralization epitopes with the following amino acid coordinates: 254-274, 303-337, 458-484 and 491-523. The peptide with amino acid coordinates 254-274 was used to immunize rabbits and the resulting antiserum was found to neutralize HIV-1 as described above. Ho et al.

The peptides encompassed by the invention comprise amino acid sequences each containing at least one continuous (linear) epitope that elicits production of HIV specific antibodies in the immunized host.

The invention thus encompasses immunogenic peptides corresponding to regions of HIV gp120 protein encoded by the envelope gene of HIV-1 HTLV III-B described by Muesing et al., "Nucleic Acid Structure and Expression of the Human AIDS/Lymphadenopathy retrovirus," *Nature*, 313:450-458 (1985). The nucleotide sequence is given in Genbank Release 63 under the name HIVPV22. The invention further encompasses functionally equivalent variants of the peptides which do not significantly affect the immunogenic properties of the peptides. For instance, conservative substitution of amino acid residues,

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one or a few amino acid residues by amino acid analogues are within the scope of the invention.

Homologs are peptides which have conservatively substituted amino acid residues. Amino acids which can be conservatively substituted for one another include but are not limited to: glycine/alanine; valine/isoleucine/leucine; asparagine/glutamine; aspartic acid/glutamic acid; serine/threonine; lysine/arginine; and phenylalanine/tyrosine. Homologous peptides are considered to be within the scope of the invention if they are recognized by antibodies which recognize the peptides designated gp120-12, gp120-15, gp120-16 and gp120-19, the sequences of which are shown below. Further, all homologous peptides corresponding to the peptides of the present invention but derived from different HIV isolates are also encompassed by the scope of this invention.

Analogues are defined as peptides which are functionally equivalent to the peptides of the present invention but which contain certain non-naturally occurring or modified amino acid residues. Additionally, polymers of one or more of the peptides, and peptide analogues or homologs are within the scope of the invention. Also within the scope of this invention are peptides of fewer amino acid residues than gp120-12, gp120-15, gp120-16 and gp120-19, respectively, but which encompass one or more immunogenic epitopes present in any one of the peptides and thus retain the immunogenic properties of the base peptide. Analytical techniques for determining the extent to which the peptides in question can be shortened at either end, while still retaining the immunogenic epitope of the longer sequence, are described below.

Addition of amino acids to either end of the peptides specifically disclosed herein is also considered within the scope of the present invention, so long as such addition does not significantly deleteriously affect the immunological properties of that peptide. Routine testing can determine whether the desired immunological properties are retained by such supplemented or truncated peptides. If amino acids are

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added, it is preferred that the resulting peptides are still relatively short, e.g., not more than about 50 amino acids long, preferably not more than about 40 or 45 amino acids long, and most preferably not more than about 25, 30, or 35 amino acids in length.

The peptides of the present invention were synthesized by known solid phase peptide synthesis techniques. Barany and Merrifield, *The Peptides: Analysis, synthesis, Biology*, Vol. 1, Gross and Meinenhofer, eds., Academic Press, New York, Chap. 1 (1980). The synthesis also allows for one or more amino acids not corresponding to the original protein sequence to be added to the amino or carboxyl terminus of the peptide. Such extra amino acids are useful for coupling the peptides to another peptide, to a large carrier protein or to a solid support. Amino acids that are useful for these purposes include but are not limited to tyrosine, lysine, glutamic acid, aspartic acid, cysteine and derivatives thereof. Additional protein modification techniques may be used, e.g., NH_2 -acetylation or COOH-terminal amidation, to provide additional means for coupling the peptides to another protein or peptide molecule or to a support. Procedures for coupling peptides to each other, carrier proteins and solid supports are well known in the art. Peptides containing the above-mentioned extra amino acid residues either carboxy or amino terminally, uncoupled or coupled to a carrier or solid support are consequently within the scope of the invention. Reference to the peptides of the present invention encompasses all of the embodiments discussed herein.

An alternative method of vaccine production is to use molecular biology techniques to produce a fusion protein containing one or more of the peptides of the present invention and a highly immunogenic protein. For instance, fusion proteins containing the antigen of interest and the B subunit of cholera toxin have been shown to induce an immune response to the antigen of interest. See Sanchez et al., "Recombinant System for Overexpression of Cholera Toxin B Submit In Vibrio cholerae as a Basis for Vaccine Development,"

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Proc. Natl. Acad. Sci. USA, 86:481-485 (1989). Such chimeric peptides may be orally administered.

The novel peptide sequences are set forth below. The amino acid residues are derived from the nucleotide sequence previously described by Muesing et al., "Nucleic Acid Structure and Express of the Human AIDS/Lymphadenopathy Retrovirus," Nature, 313:450-458 (1985). It is preferred that the peptides possess an amido group at their carboxy termini rather than a carboxyl group. The carboxy terminus can also be a carboxyl group as well as a moiety described below.

gp120-12

X-Gly-Glu-Ile-Lys-Asn-Cys-Ser-Phe-Asn-Ile-Ser-Thr-Ser-Ile-Arg-Gly-Lys-Val-Gln-Lys-Glu-Tyr-Ala-Phe-Phe-Y-Z

15 gp120-15

X-Leu-Thr-Ser-Cys-Asn-Thr-Ser-Val-Ile-Thr-Gln-Ala-Cys-Pro-Lys-Val-Ser-Phe-Glu-Pro-Ile-Pro-Ile-His-Tyr-Cys-Y-Z

gp120-16

20 X-Pro-Lys-Val-Ser-Phe-Glu-Pro-Ile-Pro-Ile-His-Tyr-Cys-Ala-Pro-Ala-Gly-Phe-Ala-Ile-Leu-Lys-Cys-Asn-Asn-Y-Z

gp120-19

25 X-Thr-His-Gly-Ile-Arg-Pro-Val-Val-Ser-Thr-gln-Leu-Leu-Leu-Asn-Gly-Ser-Leu-Ala-Glu-Glu-Y-Z

wherein X is either a hydrogen atom of the amino terminal NH₂ group of the peptide or an additional amino acid being selected to facilitate coupling of the peptide to a carrier; Y is absent or Cys; and Z is the carboxyl group of the carboxy terminal amino acid or an amido group. The amino acid abbreviations used are defined in Table 2.

The peptides are useful as vaccines to protect against future infection by HIV or to heighten the immune response to HIV in subjects already infected by HIV. Although any primate or preferably human subject could be vaccinated with the peptides, the most suitable subjects are people at risk for

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HIV infection. Such subjects include but are not limited to homosexuals, prostitutes, intravenous drug users and those in the medical professions who have contact with patients or biological samples. The invention also provides monoclonal and polyclonal antibodies which specifically recognize the peptides. The invention further provides antibodies which neutralize HIV.

In the preferred embodiment of the present invention, the peptides are formulated into compositions for use as immunogens. These immunogens can be used as vaccines in mammals including primates and humans or to elicit production of polyclonal and monoclonal antibodies in animals. For formulation of such compositions, an immunogenically effective amount of at least one of the peptides is admixed with a physiologically acceptable carrier suitable for administration to mammals including humans. The peptides may be covalently attached to each other, to other peptides, to a protein carrier or to other carriers, incorporated into liposomes or other such vesicles, and/or mixed with an adjuvant or adsorbent as is known in the vaccine art. For instance, the peptide or peptides can be mixed with immunostimulating complexes as described by Takahashi et al., "Induction of CD8+ Cytotoxic T Cells by Immunization With Purified HIV-1 Envelope Protein and ISCOMS," *Nature*, 344:873-875 (1990). Alternatively, the peptides are uncoupled and merely admixed with a physiologically acceptable carrier such as normal saline or a buffering compound suitable for administration to mammals including humans.

The immune response to the peptides of the present invention can be enhanced by a wide variety of agents. The list of available adjuvants is long and is rapidly growing. In a preferred embodiment, Freund's complete adjuvant is used to increase the immune response of the mammal receiving the peptide as a vaccine.

As with all immunogenic compositions for eliciting antibodies, the immunogenically effective amounts of the peptides of the invention must be determined empirically.

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Factors to be considered include the immunogenicity of the native peptide, whether or not the peptide will be complexed with or covalently attached to an adjuvant or carrier protein or other carrier and route of administration for the composition, i.e. intravenous, intramuscular, subcutaneous, etc., and the number of immunizing doses to be administered. Such factors are known in the vaccine art and it is well within the skill of immunologists to make such determinations without undue experimentation.

The invention is further illustrated by the following specific examples which are not intended in any way to limit the scope of the invention.

Example 1

Peptide Synthesis

An Applied Biosystems peptide-synthesizer Model 430 A, was utilized for the synthesis of the peptides of the present invention. Each synthesis used a p-methylbenzyl-hydrylamine solid phase support resin (Peptides International, Louisville, KY). The peptides were synthesized according to the Users Manual for Peptide Synthesizer Model 430A, Applied Biosystems, 1986.

All amino acids for use in synthesis contained t-butylcarbonyl groups (t-Boc) protecting the α -NH₂ group and were obtained from Novabiochem AG, Switzerland. Amino acids with reactive side chain groups contained additional protective groups to prevent unwanted and undesirable side chain reactions. The individual protected amino acids used in synthesizing all of the peptides are set forth in Table 1.

Table 1

Amino Acids Used in Peptides Synthesis

Boc-Ala-OH

Boc-Arg (Tos)-OH

Boc-Asn-OH

Boc-Asp (Obzl)-OH

Boc-Cys (Pmeobzl)-Oh

Boc-Glu (Obzl)-OH

Boc-Gln-OH

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- Boc-Gly-OH
Boc-His- (Tos) -OH
Boc-Ile-OH^{1/2} H₂O
Box-Leu-OH^{H₂O}
5 Box-Lys (2-CI-Z) -OH (cryst.)
Box-Met-OH
Boc-Phe-OH
Boc-Pro-OH
Boc-Ser (Bzl) -OH^{DCHA}
10 Boc-Thr (bzl) -OH
Boc-Trp (Formyl) -OH
Boc-Tyr (2-Br-Z) -OH
Boc-Val-OH
- 15 Tos: Tosyl or p-Toluene sulfonic acid
Obzl = Benzyloxy
Pmeobzl = p-Methylbenzyloxy
2-CL-Z = Carbobenzoxy chloride
2-Br-Z = Carbobenzoxy bromide
- 20 After completion of a particular synthesis, the protecting groups were removed from the synthesized peptide and the peptide was cleaved from the solid support resin by treatment with Trifluoromethane Sulfonic Acid (TFMSA) according to the method described by Bergot et al., "Utility
25 of Trifluoromethane Sulfonic Acid as a Cleavage Reagent in Solid Phase Peptide Synthesis," Applied Biosystems User Bulletin, Peptide Synthesizer, Issue No. 16, Sept. 2, 1986. The following is the detailed protocol used.
1. For 1 gram peptide-resin, 3 ml Thio-Anisol
30 1,2-Ethane-Dithiol (2:1) was added as scavenging agent and the mixture was incubated with continuous stirring for 10 min. at room temperature.
2. Trifluoroacetic Acid (TFA), 10 ml, was added and stirred continuously for 10 min. at room temperature.
- 35 3. TFMSA, 1 ml, was added dropwise with forceful stirring and reacted for 25 min. at room temperature.

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4. Following cleavage, the peptides were precipitated with and washed with anhydrous ether.

5. The precipitated and washed peptides were dissolved in a small volume of TFA (approximately 5 ml).

5 6. The dissolved peptides were again precipitated and washed as above in step 4 and the precipitate was dried under a stream of N₂.

Prior to use in specific assays, the peptides can be further purified, if desired, by reverse phase high performance liquid chromatography (HPLC). A particularly suitable column for such purification is the reverse-phase Vydak™ C-18 column using a water (TFA) - acetonitrile (TFA) gradient to elute the peptides. Forty peptides covering the entire sequence of HIV-1 gp120 were synthesized having the amino acid sequences shown in Table 2. A truncated peptide gp120-16/B with the amino acid coordinates 213-224 was also synthesized.

10

15

TABLE 2

Peptide	Amino Acid Coordinates*	Amino Acid Sequence**	SEQ. I.D. No.
gp120-1	1-28	MRVKEKYQHLWRWGWRWGTMLLGMLMIC	1
gp120-2	23-46	GMLMICSATEKLWVTVYYGVPVWK	2
gp120-3	41-64	GVPVWKEATTTLFCASDAKAYDTE	3
gp120-4	54-74	CASDAKAYDTEVHNVWATHAC	4
gp120-5	65-89	VHNVWATHACVPTDPNPQEVVLVNV	5
gp120-6	75-100	VPTDPNPQEVVLVNVTFNFMWKNDM	6
gp120-7	90-116	TENFMWKNDMVEQMHEDIISLWDQSL	7
gp120-8	101-126	VEQMHEDIISLWDQSLKPCVKLTPLC	8
gp120-9	117-141	KPCVKLTPLCVSLKCTDLKNDTNTN	9
gp120-10	127-151	VSLKCTDLKNDTNTNSSSGRMIMEK	10
gp120-11	142-164	SSSGRMIMEKGEIKNCSFNISTS	11
gp120-12	152-176	GEIKNCSFNISTSIRGKVQKEYAFF	12
gp120-13	165-192	IRGKVQKEYAFFYKLDIIPIDNDTTSYT	13
gp120-14	177-205	YKLDIIPIDNDTTSYTLTSCNTSVITQAC	14
gp120-15	193-218	LTSCNTSVITQACPKVSFEPIPIHYC	15
gp120-16	206-230	PKVSFEPIPIHYCAPAGFAILKCNN	16
gp120-16/B	213-224	IPIHYCAPAGFA	41
gp120-17	219-237	APAGHAILKCNNKTFNGTGPCTNVSTVQC	17
gp120-18	231-257	KTFNGTGPCTNVSTVQCTHGIRPVVST	18
gp120-19	248-269	THGIRPVVSTQLLLNGSLAEEE	19
gp120-20	258-282	QLLLNGSLAEEVVIRSANFTDNAK	20
gp120-21	270-295	VVIRSANFTDNAKTIIVQLNQSVEIN	21
gp120-22	283-306	TIIVQLNQSVEINCTRPNNNTRKS	22
gp120-23	296-320	CTRPNNNTRKSIRIQRGPGRAFVTI	23
gp120-24	307-330	IRIQRGPGRAFVTIGKIGNMRQAH	24
gp120-25	321-343	GKIGNMRQAHCNISRAKWNNTLK	25
gp120-26	331-353	CNISRAKWNNTLKQIDSKLREQF	26
gp120-27	344-366	QIDSKLREQFGNNKTIIFKQSSG	27

TABLE 2

Peptide	Amino Acid Coordinates*	Amino Acid Sequence**	SEQ. I.D. No.
gp120-28	354-377	GNNKTIIFKQSSGGDPEIVTHSFN	28
gp120-29	367-389	GDPEIVTHSFNCGGEFFYCNSTQ	29
gp120-30	378-400	CGGEFFYCNSTQLFNSTWFNSTW	30
gp120-31	390-409	LFNSTWFNSTWSTEGSNNTE	31
gp120-32	401-417	STEGSNNTEGSDTITLP	32
gp120-33	410-429	GSDTITLPCRKQFINMWQE	33
gp120-34	418-444	CRKQFINMWQEVGKAMYAPPISGQIR	34
gp120-35	430-453	VGKAMYAPPISGQIRCSSNITGLL	35
gp120-36	445-466	CSSNITGLLLTRDGGNNNESE	36
gp120-37	454-476	LTRDGGNNNESEIFRPGGGDMR	37
gp120-38	467-488	IFRPGGGDMRDNWRSELYKYKV	38
gp120-39	477-497	DNWRSELYKYKVVKIEPLGVA	39
gp120-40	489-511	VKIEPLGVAPTAKRRVVQREKR	40

**Amino acid abbreviations

Alanine	Ala	A	Leucine	Leu	L
Arginine	Arg	R	Lysine	Lys	K
Asparagine	Asn	N	Methionine	Met	M
Aspartic acid	Asp	D	Phenylalanine	Phe	F
Cysteine	Cys	C	Proline	Pro	P
Glutamine	Gln	Q	Serine	Ser	S
Glutamic acid	Glu	E	Threonine	Thr	T
Glycine	Gly	G	Tryptophan	Trp	W
Histidine	His	H	Tyrosine	Tyr	Y
Isoleucine	Ile	I	Valine	Val	V

* As previously described by Muesing et al.

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Example 2Cells and Virus Stocks

All neutralization tests were performed using H-9 cells and HTLV-111B virus (originating from R.C. Gallo and supplied by Dr. William Hall, North Shore Hospital, Manhasset, New York). H-9 cells (designated H9 NY) were maintained in RPMI Medium (Gibco) supplemented with 20% fetal calf serum (FCS), penicillin/streptomycin (PEN/STREP 50 μ g/ml each and without any fungicides). Cells were subcultured at a dilution of 1:3 every 4 days.

Cells were scraped from the plates and pelleted by centrifugation at 325 x g. Pelleted cells were resuspended in 1 ml of stock virus previously diluted 1/10 and allowed to adsorb for 60 min at 37°C with frequent stirring. After adsorption of the virus, the cells were recentrifuged and resuspended in 10 ml of RPMI with 20% FCS and polybrene (2 μ g/ml) (giving a final concentration of 5×10^5 cells/ml) and incubated at 37°C in 5% CO₂.

Infected cells were shown to be detectable at 4-5 days post-infection (p.i.) by monitoring syncytia formation, positive cells in immunofluorescence and p-24 production (assayed by the Abbott p-24 antigen test). The peak of HIV production was seen 10 - 15 days p.i. at which time virus was collected. After low speed centrifugation to remove debris, supernatants containing virus collected from infected cells were frozen in stocks at -90°C. One virus stock with endpoint titer of 40,000 50% tissue culture infective doses (TCID₅₀) was used throughout the studies (referred to as NT3-NT19).

Example 3Preparation of Peptides for Immunization

Peptides according to the present invention were covalently coupled to ovalbumin grade V (Sigma, St. Louis, MO, USA) at an approximate 10:1 (peptide:ovalbumin) molar ratio using N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP), (Pharmacia, Uppsala, Sweden) as bifunctional linker according

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to the manufacturer's instructions (Pharmacia) i.e., briefly as follows:

Ovalbumin was dissolved in coupling buffer (0.2M NaH₂PO₄, pH 8.5). The dissolved ovalbumin was then run through a
5 Sephadex G-25M column (Pharmacia, Sweden), using the same buffer. Protein concentration was measured at 280 nm and the recovery was determined. SPDP was dissolved in 99.5% ethanol to a final concentration of 40 mM. SPDP was then added dropwise to the ovalbumin solution under stirring. The SPDP-
10 ovalbumin mixture was then left at room temperature for approximately 30 minutes. The ovalbumin-SPDP conjugate was separated from unconjugated SPDP by running the mixture through a Sephadex G-25M column, using water as eluent. The degree of substitution for the ovalbumin-SPDP conjugate was
15 determined after diluting 50 µl conjugate in 2 ml of water, by measuring the diluted conjugate at 280 nm and the diluted conjugate plus 100 µl Dithiothreitol (DDT) (Sigma) at 343 nm, in order to determine the amount to be added to the peptide solution.

20 Finally, the synthetic peptide to be coupled to the ovalbumin-SPDP conjugate was dissolved in 10% acetic acid to a final concentration of 1 mg/ml and a suitable amount of ovalbumin-SPDP conjugate (as determined by the substitution degree above) was added and allowed to stand overnight at room
25 temperature.

Example 4

Immunization Protocols

Maccaca fascicularis were used to generate antibodies..
30 Prior to the initial peptide injection, a blood sample was drawn from the monkeys. This initial blood sample is termed "pre-immune" (Tables 3-6) and is used as an internal control and analyzed simultaneously with respective immunoserum.

The monkeys were injected with 100 µg peptide-SPDP-
35 ovalbumin suspended in 0.5 ml phosphate buffered saline (PBS). The monkeys were immunized intramuscularly three times, three weeks apart. As adjuvant, 0.5 ml of Freund's complete

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adjuvant was used for all immunizations. Two weeks after the final immunization, the monkeys were bled and pre-immune and hyperimmune sera were subject to neutralization assays as described in Example 5.

5

Example 5

HIV-1 Neutralization Assay

Sera containing antibodies that neutralize HTLV 11-B infectivity were detected by their ability to prevent HIV-1 syncytium formation, p-24 antigen production and decreased number of infected cells as determined by immuno-fluorescence markers, compared to control infections lacking peptide specific antisera. Stock virus, described in Example 2 was diluted to 100 TCID₅₀ and mixed with serial fourfold dilutions (1/5, 1/20, and 1/80) of complement-inactivated immunesera obtained from the monkeys immunized as described in Example 4. As a positive control, a guinea pig hyperimmune serum (referred to as MSV) with known HIV neutralizing titer of 1/40 - 1/160 was included in all experiments (kindly provided by Prof. B. Morein, Dept. Veterinary Virology, BMC, Uppsala, Sweden). After incubation for 60 min at 37°C or 16 hours at 4°C, the serum-virus mixture was added to 1x10⁶ H-9 cells and incubated for another 60 min at 37°C. Following incubation, the cells were washed once and placed in 24 well multidish plates with 2 ml of growth medium (RPMI, 10%, FCS, 2 µg polybrene/ml) per well.

Cells were examined under the microscope (magnification x200) for the presence of syncytia on days 5-12 p.i. Supernatants from infected cells were assayed for the presence of p-24 antigen according to the manufacturer's instructions (Abbott ag test HIVAG-1[®], Enzyme Immunoassay for the Detection of Human Immunodeficiency Virus Type I (HIV-1) Antigen(s) in Human Serum or Plasma) in tenfold serial dilutions (1/10 - 1/1,000) at 10 days p.i. The results are presented as absorbance values at 454 nm with higher absorbance values indicating higher protein concentration and hence HIV infection. Serial dilutions of the supernatants were made so

35

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as to detect p-24 concentrations in the most accurate range (< 2.0 absorbance units).

The number of infected cells were determined at the end of the experiment (usually on day 15 p.i.) by acetone-fixation of cells on slides adopted for immunofluorescence (IF). An indirect IF test was used according to standard procedures described in Jeansson et al., "Elimination of Mycoplasmas from Cell Cultures Utilizing Hyperimmune Sera", Ex. Cell Res., 161:181-188 (1985), with 1/400 dilution hyperimmune sera from HIV-infected individuals and a fluorescein isothiocyanate (FITC) labeled antihuman IgG antibody (Bio-Merieux France) diluted 1/100. Tables 3-6 show the results obtained from screening of hyperimmune sera from monkeys immunized with peptides 1-40.

In Tables 3(A-D)-6 the p24 antigen content of the supernatants was analyzed by ELISA as described above. The relative amount of antigen positive cells is depicted as AG POS cells wherein the percentages are represented by:

- = 0%, + = >0-2%, ++ = 3-10% and +++ = 11-20% where the percentage interval indicates the number of antigen positive cells.

Table 3A (HIVNT3P1.XLS) depicts the results obtained with sera derived from monkeys immunized with peptides gp120-1 - gp120-10. The cells used were H9 NY and the virus used was HTLV-IIIIB, Batch 18 described in Example 2. The incubation protocol was (virus plus serum) incubation at 37°C for one hour.

Table 3B (HIVNT4P1.XLS) depicts the results obtained with sera derived from monkeys immunized with peptides gp120-11 - gp120-20. The cells used were H9 NY and the virus used was HTLV-IIIIB, Batch 18 described in Example 2. The incubation protocol was (virus plus serum) incubation at 37°C for one hour.

Table 3C (HIVNT5P1.XLS) depicts the results obtained with sera derived from monkeys immunized with peptides gp120-21 - gp120-30. The cells used were H9 NY and the virus used was

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HTLV-IIIB, Batch 18 described in Example 2. The incubation protocol was virus plus serum incubated at 37°C for one hour.

5 Table 3D (HIVNT6P1.XLS) depicts the results obtained with sera derived from monkeys immunized with peptides gp120-31 - gp120-40. The cells used were H9 NY and the virus used was HTLV-IIIB, Batch 18 described in Example 2. The incubation protocol was (virus plus serum) incubation at 37°C for one hour.

10 Table 4 (HIVTAB4.XLS) shows the results of the first retest of putative neutralizing antibodies as determined by the first test (Tables 3A-D). In each test, the virus used was HTLV-IIIB, Batch 18 and the cells used were H9 NY. The First Retest results in rows 1-19 are the results of neutralization test number 5. The incubation protocol was
15 incubation at 37°C for one hour. The First Retest results in rows 20-32 are the results of neutralization test number 7. The incubation protocol was incubation of at 37°C for one hour.

20 Table 5 (HIVTAB5.XLS) shows second, third and fourth retest results of the positive peptides. In each test, the virus used was HTLV-IIIB, Batch 18 and the cells used were H9 NY. The Second Retest results in rows 1-4 are the results of neutralization test number 7. The incubation protocol was incubation at 37°C for one hour. The Second Retest results in
25 rows 5-13 are the results of neutralization test number 12. The Third Retest results shown in rows 14-16 are the results of neutralization test number 12. The incubation protocol was incubation of at 37°C for one hour. The Fourth Retest results shown in rows 17-39 are the results of neutralization test
30 number 16. The incubation protocol was incubation of at 4°C for 16 hours. The Second Retest results in rows 40-53 are the result of neutralization test 19. The incubation protocol was cells plus virus at 4°C for 16 hours.

35 Table 6 (HIVKOMBP.XLS) shows the neutralization assay results with combined hyperimmune sera. Note that the incubation of virus and cells was at 4°C for 16 hours.

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The results depicted in Tables 3 (A-D) -6 indicate that the peptides of the present invention elicit the production of HIV neutralizing antibodies in primate subjects. The use of the peptides in vaccination of human subjects is therefore applicable to prevent infection by HIV or to induce heightened immune response in subjects already infected by HIV.

TABLE 3A - ASSAYS OF ANTISERA TO PEPTIDES gp120-1 - gp120-10

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant DIL)			RELATIVE AMOUNT OF AG POS CELLS
			1/10	1/100	1/1000	
1.	Pos control		> 2.0	1.176	0.158	+++
2.	Pos control		> 2.0	1.194	0.177	+++
3.	Pos control		> 2.0	> 2.0	0.464	+++
4.	Neg control		0.056	-	-	-
5.	guinea pig	1/10	0.178	0.066	0.063	-
6.	Pos control	1/40	0.71	0.118	0.06	++
7.	Antiserum	1/160	> 2.0	0.742	0.11	++
8.		1/320	> 2.0	0.484	0.093	+++
9.	preimmune		ND	ND	ND	ND
10.	gp120-1	1/5	0.715	0.108	0.054	++
11.		1/20	> 2.0	0.36	0.073	++
12.		1/80	> 2.0	0.57	0.093	++
13.	preimmune		> 2.0	0.437	0.081	++
14.	gp120-2	1/5	> 2.0	0.86	0.138	++
15.		1/20	> 2.0	0.486	0.093	+++
16.		1/80	> 2.0	0.257	0.083	+++
17.	preimmune		> 2.0	0.466	0.09	++
18.	gp120-3	1/5	> 2.0	0.367	0.079	++

TABLE 3A - ASSAYS OF ANTISERA TO PEPTIDES gp120-1 - gp120-10

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant DIL)			RELATIVE AMOUNT OF AG POB CELLS
			1/10	1/100	1/1000	
19.		1/20	> 2.0	0.512	0.094	++
20.		1/80	> 2.0	0.724	0.113	++
21.	preimmune		> 2.0	0.536	0.094	++
22.	gp120-4	1/5	> 2.0	0.638	0.092	++
23.		1/20	> 2.0	0.448	0.082	++
24.		1/80	> 2.0	0.592	0.097	++
25.	preimmune		> 2.0	0.43	0.082	++
26.	gp120-5	1/5	> 2.0	0.638	0.098	++
27.		1/20	> 2.0	0.737	0.11	++
28.		1/80	> 2.0	0.786	0.119	+++
29.	preimmune		> 2.0	0.822	0.125	++
30.	gp120-6	1/5	> 2.0	0.716	0.131	+++
31.		1/20	> 2.0	0.977	0.119	++
32.		1/80	> 2.0	0.861	0.124	++
33.	preimmune		> 2.0	0.719	0.116	++
34.	gp120-7	1/5	> 2.0	0.587	0.106	++
35.		1/20	> 2.0	0.45	0.092	++
36.		1/80	> 2.0	0.756	0.117	++
37.	preimmune		> 2.0	0.507	0.096	+++

TABLE 3A - ASSAYS OF ANTISERA TO PEPTIDES gp120-1 - gp120-10

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant DIL)			RELATIVE AMOUNT OF AG POB CELLS
			1/10	1/100	1/1000	
38.	gp120-8	1/5	> 2.0	0.555	0.098	++
39.		1/20	> 2.0	0.59	0.103	++
40.		1/80	> 2.0	0.308	0.081	++
41.	preimmune		> 2.0	0.322	0.076	+++
42.	gp120-9	1/5	> 2.0	0.358	0.09	++
43.		1/20	> 2.0	0.403	0.082	+++
44.		1/80	> 2.0	0.612	0.102	+++
45.	preimmune		> 2.0	0.747	0.127	++
46.	gp120-10	1/5	> 2.0	0.3	0.074	++
47.		1/20	> 2.0	0.426	0.092	++
48.		1/80	> 2.0	0.442	0.083	++

TABLE 3B - ASSAYS OF ANTISERA TO PEPTIDES gp120-11 - gp120-20

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG FOB CELLS
			1/10	1/100	1/1000	
1.	preimmune	1/5	> 2.0	0.882	0.149	++
2.	gp120-11	1/5	> 2.0	0.73	0.135	++
3.		1/20	> 2.0	1.73	0.299	++
4.		1/80	> 2.0	0.700	0.148	++
5.	preimmune	1/5	> 2.0	1.07	0.151	++
6.	gp120-12	1/5	0.157	0.07	0.076	+
7.		1/20	> 2.0	1.45	0.22	++
8.		1/80	> 2.0	1.37	0.221	++
9.	preimmune	1/5	> 2.0	0.58	0.107	++
10.	gp120-13	1/5	> 2.0	1.16	0.194	++
11.		1/20	1.816	0.37	0.095	++
12.		1/80	> 2.0	1.16	0.187	++
13.	preimmune	1/5	> 2.0	> 2.0	0.281	++
14.	gp120-14	1/5	> 2.0	0.81	0.142	++
15.		1/20	> 2.0	1.39	0.219	++
16.		1/80	> 2.0	0.83	0.156	++
17.	preimmune	1/5	> 2.0	1.13	0.192	++
18.	gp120-15	1/5	> 2.0	1.43	0.243	+++
19.		1/20	0.069	0.05	0.05	-

TABLE 3B - ASSAYS OF ANTISERA TO PEPTIDES gp120-11 - gp120-20

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (supernatant DIL)			RELATIVE AMOUNT OF AG POB CELLS
			1/10	1/100	1/1000	
20.		1/80	> 2.0	0.57	0.104	++
21.	preimmune	1/5	> 2.0	1.78	0.303	++
22.	gp120-16	1/5	0.26	0.07	0.056	+
23.		1/20	0.067	0.06	0.054	-
24.		1/80	> 2.0	0.74	0.132	++
25.	preimmune	1/5	> 2.0	1.13	0.171	++
26.	gp120-17	1/5	> 2.0	0.76	0.161	++
27.		1/20	> 2.0	1.56	0.285	++
28.		1/80	> 2.0	0.7	0.129	++
29.	preimmune	1/5	> 2.0	1.41	0.177	++
30.	gp120-18	1/5	> 2.0	> 2.0	0.339	++
31.		1/20	> 2.0	1.36	0.218	++
32.		1/80	> 2.0	1.26	0.199	++
33.	preimmune	1/5	> 2.0	0.39	0.097	++
34.	gp120-19	1/5	0.476	0.1	0.061	+
35.		1/20	1.048	0.18	0.068	+
36.		1/80	> 2.0	1.62	0.303	++
37.	preimmune	1/5	> 2.0	1.11	0.189	++

TABLE 3B - ASSAYS OF ANTISERA TO PEPTIDES gp120-11 - gp120-20

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant DIL.)			RELATIVE AMOUNT OF AG POS CELLS
			1/10	1/100	1/1000	
38.	gp120-20	1/5	> 2.0	1.19	0.182	+++
39.		1/20	> 2.0	1.47	0.054	++
40.		1/80	> 2.0	1.42	0.264	++

TABLE 3C - ASSAY OF ANTISERA TO PEPTIDES 21-30

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL	
			1/10	1/100	1/1000		Day 5	Day 7
49.	pos control		> 2.0	0.65	0.09	++	12	72
50.	pos control		1.85	0.24	0.061	++	6	27
51.	neg control		0.4				0	0
52.	guinea pig	1/10	0.5	0.04	0.047	-	0	0
53.	pos control	1/40	0.05	0.04	0.04	-	1	0
54.	antisera	1/160	0.04	0.05	0.043	+	1	3
55.		1/640	1.07	0.14	0.056	+	2	19
56.	preimmune	1/5	> 2.0	1.57	0.275		12	85
57.	gp120-21	1/5	> 2.0	0.4	0.075	++	3	28
58.		1/20	1	0.17	0.059		5	21
59.		1/80	> 2.0	0.48	0.089		7	72
60.	preimmune	1/5	> 2.0	1.1	0.182		3	ND
61.	gp120-22	1/5	> 2.0	1.48	0.221	++	2	75
62.		1/20	> 2.0	1.07	0.16		0	80
63.		1/80	> 2.0	0.63	0.087		5	90
64.	preimmune	1/5	> 2.0	0.4	0.083		4	52
65.	gp120-23	1/5	1.97	0.26	0.067	ND	0	20
66.		1/20	> 2.0	1.63	0.236		5	98

TABLE 3C - ASSAY OF ANTISERA TO PEPTIDES 21-30

	PEPTIDE	Serum dilution	P-24 ANTIGEN (Supernatant DIL)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL	
			1/10	1/100	1/1000		Day 5	Day 7
67.		1/80	> 2.0	0.35	0.084		5	>150
68.	preimmune	1/5	> 2.0	> 2.0	0.355		2	49
69.	gp120-24	1/5	1.95	0.29	0.067	+	0	3
70.		1/20	> 2.0	0.37	0.081		5	34
71.		1/80	1.87	0.24	0.069		3	48
72.	preimmune	1/5	> 2.0	0.83	0.145		0	91
73.	gp120-25	1/5	> 2.0	0.73	0.11	++	1	25
74.		1/20	1.63	0.23	0.062		0	15
75.		1/80	1.88	0.22	0.064		0	38
76.	preimmune	1/5	> 2.0	0.48	0.089		0	79
77.	gp120-26	1/5	> 2.0	0.62	0.101	++	3	91
78.		1/20	> 2.0	0.34	0.063		3	35
79.	gp120-26	1/80	1.27	0.19	0.061		0	21
80.	preimmune	1/5	> 2.0	0.66	0.11		2	52
81.	gp120-27	1/5	> 2.0	0.58	0.098	++	1	26
82.		1/20	> 2.0	0.65	0.099		6	49
83.		1/80	> 2.0	0.3	0.062		2	35
84.	preimmune	1/5	> 2.0	> 2.0	0.317		7	31

TABLE 3C - ASSAY OF ANTISERA TO PEPTIDES 21-30

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL	
			1/10	1/100	1/1000		Day 5	Day 7
85.	gp120-28	1/5	> 2.0	0.39	0.078	++	2	22
86.		1/20	> 2.0	0.68	0.105		5	70
87.		1/80	0.99	0.15	0.05		3	>150
88.	preimmune	1/5	> 2.0	1.29	0.187		5	97
89.	gp120-29	1/5	> 2.0	0.55	0.096	++	3	112
90.		1/20	> 2.0	0.85	0.135		3	>150
91.		1/80	> 2.0	0.72	0.113		0	29
92.	preimmune	1/5	> 2.0	> 2.0	0.326		10	130
93.	gp120-30	1/5	> 2.0	0.27	0.073	+	3	38
94.		1/20	> 2.0	1.71	0.24		9	52
95.		1/80	> 2.0	0.44	0.082		6	ND

TABLE 3D - ASSAYS OF ANTISERA TO PEPTIDES 31-40

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL Day 6
			1/10	1/100	1/1000		
			96.	pos control	0.976		
97.	pos control	1.836	0.656	0.185		11	
98.	neg control						
99.	guinea pig	1/10	0.103	0.088	0.09		0
100.	pos control	1/40	0.104	0.087	0.093		0
101.	antisera	1/160	0.749	0.29	0.1		4
102.		1/640	1.066	0.238	0.237		7
103.	preimmune	1/5	0.824				
104.	gp120-31	1/5	1.769	0.675	0.186		47
105.		1/20	1.124	0.302	0.111		22
106.		1/80	0.978	0.258	ND		24
107.	preimmune	1/5	0.883				
108.	gp120-32	1/5	1.163	0.258	ND		7
109.		1/20	1.482	0.311	ND		8
110.		1/80	0.996	0.263	ND		0
111.	preimmune	1/5	1.76				
112.	gp120-33	1/5	0.84	0.239	0.156		20
113.		1/20	1.282	0.333	0.144		16

TABLE 3D - ASSAYS OF ANTISERA TO PEPTIDES 31-40

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL Day 6
			1/10	1/100	1/1000		
132.	gp120-38	1/5	1.386	0.59	0.114	11	
133.		1/20	0.576	0.214	0.106	17	
134.		1/80	1.23	0.329	ND		
135.	preimmune	1/5	1.854				
136.	gp120-39	1/5	1.376	0.495	0.182	28	
137.		1/20	0.711	0.296	0.118	17	
138.		1/80	0.929	0.237	ND		
139.	preimmune	1/5	ND				
140.	gp120-40	1/5	0.862	0.255	0.132	13	
141.		1/20	0.989	0.273	0.143	10	
142.		1/80	0.477	0.164	ND		

TABLE 4 - RETESTING OF HYPERIMMUNE SERA WITH THE CAPACITY TO NEUTRALIZE HIV									
PEPTIDE	Serum Dilution	P-24 ANTIGEN (DIL)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL			
		1/10	1/100	1/1000		Day 5	Day 7		
First Retest									
1.	pos control	> 2.0	0.646	0.09	++	12	72		
2.	pos control	1.853	0.244	0.061	++	6	27 ³		
3.	neg control	0.039				0	0		
4.	guinea pig 1/10	0.051	0.04	0.047	-	0	0		
5.	pos control 1/40	0.052	0.042	0.04	-	1	0		
6.	antiserum 1/160	0.042	0.046	0.043	+	1	3		
7.	1/640	1.067	0.144	0.056	+	2	19		
8.	preimmune 1/5	2	1.326	0.172		10	112		
9.	gp120-12 1/5	1.083	0.153	0.06	+	1	24		
10.	1/20	2	1.487	0.171		7	175		
11.	1/80	2	0.463	0.07		6	ND		
12.	preimmune 1/5	2	1.991	0.237		2	64		
13.	gp120-16 1/5	2	0.355	0.07	+	0	13		
14.	1/20	0.741	0.103	0.048	...	0	11		
15.	1/80	2	0.32	0.08		0	35		
16.	preimmune 1/5	> 2.0	0.547	0.082		3	42		
17.	gp120-19 1/5	0.141	0.062	0.053	+	0	6		

TABLE 4 - RETESTING OF HYPERIMMUNE SERA WITH THE CAPACITY TO NEUTRALIZE HIV									
	PEPTIDE	Serum Dilution	P-24 ANTIGEN (DIL)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL		
			1/10	1/100	1/1000		Day 5	Day 7	
18.		1/20	1.134	0.164	0.054		0	26	
19.		1/80	> 2.0	0.455	0.081		1	45	
	First Retest		1/5	1/50	1/500		Day 7	Day 10	
20.	pos control		1.175	0.426	0.201		9	46	
21.	pos control		1.529	0.401	0.161		32	167	
22.	neg control								
23.	guinea pig	1/10	0.139	0.165	0.145	-	0	0	
24.	pos control	1/40	0.211	0.159	0.168	-	1	0	
25.	antisera	1/160	0.961	0.299	0.163	++	9	26	
26.		1/640	0.989	0.26	0.159	++	5	20	
27.	gp120-24	1/5	1.067	0.245	0.166	++	4	34	
28.		1/20	0.795	0.204	0.167	++	5	41	
29.		1/80	0.433	0.167		-	15	80	
30.	gp120-25	1/5	1.237	0.282	0.155	++	19	144	
31.		1/20	1.312	0.373	0.187	++	42	116	
32.		1/80	ND	ND	ND	-	ND	ND	

TABLE 5 - RETESTING OF HYPERIMMUNE SERA WITH CAPACITY TO NEUTRALIZE HTLV-III

PEPTIDE	SERUM DILUTION	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL	
		1/5	1/50	1/500		Day 5	Day 7
Second Retest							
1.	gp120-16	1/5	ND	ND	ND	ND	ND
2.		1/5	1.924	1.062	0.282	++	
3.		1/20	0.365	0.172	0.145	-	2
4.		1/80	0.163	0.133		-	0
Second Retest							
			1/10	1/100	1/1,000		
5.	pos control		> 2.0	> 2.0	1.026	+++	320
6.	pos control		> 2.0	> 2.0	0.639	+++	220
7.	pos control		> 2.0	> 2.0	0.866	+++	290
8.	pos control		> 2.0	> 2.0	0.881	+++	
9.	neg control		0.223			-	
10.	neg control		0.16			-	
11.	gp120-24	1/5	> 2.0	> 2.0	0.545	+++	112
12.		1/20	> 2.0	> 2.0	0.819	+++	138
13.		1/80	> 2.0	> 2.0		+++	230
Third Retest							
14.	gp120-16	1/5	0.122	0.1	0.115	-	0

TABLE 5 - RETESTING OF HYPERIMMUNE SERA WITH CAPACITY TO NEUTRALIZE HTLV-III

	PEPTIDE	SERUM DILUTION	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF Ag POS CELLS	NO. OF SYNCYTIA/WELL	
			1/5	1/50	1/500		Day 5	Day 7
15.		1/20	> 2.0	1.14	0.352	++	0	
16.		1/80	> 2.0	> 2.0		+++	210	
Fourth Retest								
17.	pos control		1.425	0.732	0.154	++	16	
18.	pos control		1.346	0.672	0.152	+++	16	
19.	pos control		1.431	0.845	0.182	+++	17	
20.	pos control		1.414	0.931	0.251			
21.	neg control		0.067			-		
22.	neg control		0.045			-		
23.	neg control		0.042			-		
24.	guinea pig	1/10	0.044	0.037	0.029		0	
25.	pos control	1/40	0.063	0.039	0.029		0	
26.	antisera	1/160	0.036	0.035	0.055		0	
27.		1/640	0.556	0.072	0.034		1	
28.	gp120-12	1/8	0.072	0.043	0.046		0	
29.		1/32	0.169	0.054	0.047		0	
30.		1/128	> 2.0	1.124	0.241		19	
31.	gp120-16	1/8	0.043	0.045	0.049		0	

TABLE 5 - RETESTING OF HYPERIMMUNE SERA WITH CAPACITY TO NEUTRALIZE HTLV-III									
	PEPTIDE	SERUM DILUTION	P-24 ANTIGEN (Supernatant Dil.)			RELATIVE AMOUNT OF AG POS CELLS	NO. OF BINCYTIA/WELL		
			1/5	1/50	1/500		Day 5	Day 7	
32.		1/32	0.052	0.043	0.048		0		
33.		1/128	1.54	0.903	0.014		4		
34.	gp120-19	1/8	0.105	0.043	0.042		0		
35.		1/32	0.358	0.08	0.045		5		
36.		1/128	> 2.0	0.944	0.205		25		
37.	gp120-24	1/8	> 2.0	0.885	0.155		2		
38.		1/32	> 2.0	1.174	0.293		15		
39.		1/128	1.158	0.858	0.213		11		
Second Retest			1/5	1/50	1/500		Day 5	Day 7	
40.	pos control		0.916	0.166	0.099			74	
41.	pos control		1.607	0.469	0.151			130	
42.	pos control		> 2.0	0.943	0.203			123	
43.	pos control		1.445	0.319	0.082			195	
44.	neg control		0.145						
45.	neg control		0.328						
46.	guinea pig	1/10	0.09	0.111	0.075			0	
47.	pos control	1/140	0.096	0.082	0.078			0	
48.	antisera	1/160	0.094	0.109	0.091			0	

TABLE 5 - RETESTING OF HYPERIMMUNE SERA WITH CAPACITY TO NEUTRALIZE HTLV-III

	PEPTIDE	SERUM DILUTION	P-24 ANTIGEN (Supernatant Dil.)			*RELATIVE AMOUNT OF AG POB CELLS	NO. OF SYNCYTIA/WELL	
			1/5	1/50	1/500		Day 5	Day 7
49.		1/640	0.996	0.212	0.104			35
50.	preimmune	1/5	> 2.0	0.444	0.162			95
51.	gp120-15	1/5	0.155	0.094	0.111			ND
52.		1/20	0.152	0.109	0.158			4
53.		1/80	0.176	0.13	0.207			0

TABLE 6 - COMBINED NEUTRALIZATION EFFECTS OF SERA FROM MONKEYS

	PEPTIDE	Serum Dilution	P-24 ANTIGEN (supernatant dil.)			NT TITRE OF SERUM	RELATIVE AMOUNT OF AG FOB CELLS	NO. OF SYNCYTIA/WELL Day 6
			1/5	1/50	1/500			
1.	Pos control		1.4	0.7	0.154	++	16	
2.	Pos control		1.3	0.7	0.152	+++	16	
3.	Pos control		1.4	0.8	0.182		17	
4.	Pos control		1.4	0.9	0.251			
5.	neg control		0.1			-		
6.	neg control		0			-		
7.	neg control		0			-		
8.	guinea pig	1/10	0	0	0.029		0	
9.	pos control	1/40	0.1	0	0.029		0	
10.	antisera	1/160	0	0	0.055	160	0	
11.		1/640	0.6	0.1	0.034		1	
12.	Group I	1/8	0	0	0.038		1	
13.	gp120.mix	1/32	0	0	0.041		0	
14.	12+16+19+24	1/128	0.2	0.1	0.043	> 128	0	
15.	Group II	1/8	0.1	0	0.046		0	
16.	gp120.mix	1/32	0.1	0.1	0.046		0	
17.	16+19	1/128	0.1	0.2	0.043	> 128	0	

TABLE 6 - COMBINED NEUTRALIZATION EFFECTS OF SERA FROM MONKEYS

	PEPTIDE	Sera Dilution	P-24 ANTIGEN (Supernatant Dil)			NT TITRE OF SERUM	RELATIVE AMOUNT OF AG POS CELLS	NO. OF SYNCYTIA/WELL Day 6
			1/5	1/50	1/500			
18.	Group III	1/8	0	0	0.051		0	
19.	gp120.mix	1/32	0.1	0.1	0.043	-	0	
20.	16+24	1/128	1	0.3	0.065	++	1	
21.	Group IV	1/8	0.2	0	0.044		2	
22.	gp120.mix	1/32	0.1	0	0.045	-	1	
23.	16+12	1/128	0.2	0.1	0.048	> 128	0	
24.	gp120-12	1/8	0.1	0	0.046	-	0	
25.		1/32	0.2	0.1	0.047	+	0	
26.		1/128	> 3	1.1	0.241		19	
27.	gp120-16	1/8	0	0	0.049		0	
28.		1/32	0.1	0	0.048	-	0	
29.		1/128	1.5	0.9	0.138		4	
30.	gp120-19	1/8	0.1	0	0.042	-	0	
31.		1/32	0.4	0.1	0.045	-	5	
32.		1/128	> 3	0.9	0.205	++	25	
33.	gp120-24	1/8	> 3	0.9	0.155	neg	2	
34.		1/32	> 3	1.2	0.293		15	
35.		1/128	1.2	0.9	0.213		11	

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Example 6The ADCC Assay

The method used for determination of HIV specific ADCC has been described by Ljunggren et al. J. Immunol. Meth. 1987, 104:7; J. Immunol., 139:2263 (1987). Briefly, the cell line U937 clone 2, continuously infected with HIV-1_{HTLVIIIB} was used as target cells. Peripheral blood mononuclear cells (PBMC) obtained from HIV antibody negative blood donors were used as effector cells. The PBMC were collected by density centrifugation on Lymphoprep (Nycomed Pharma AS, Oslo, Norway) and adherent cells were removed by the scrubbed nylon wool technique, Merrill et al. Eur. J. Immunol., 11:536 (1981). ⁵¹Cr-labeled target cells, 1 x 10⁴, and lymphocytes as effector cells, 2 x 10⁵, were mixed with serum dilutions, six dilution steps in three-fold serial dilutions starting at 1:30. Supernatants were harvested after three hours and released radioactivity was calculated. The spontaneous release never exceeded 10%.

HIV specific ADCC was determined as follows: specific ⁵¹Cr-release with HIV positive sera minus specific ⁵¹Cr-release with HIV negative sera. Sera with a Specific ADCC Index (SAI) value > 0.5 at 1:30 were considered to be positive for HIV-specific ADCC, Ljunggren et al. J. Immunol. 1987, 139:2263. This value represents more than 3 SD above the specific ⁵¹Cr-release obtained by HIV-antibody negative sera. HIV antibody positive sera with known ADCC titer were included in each test. The reciprocal of the last dilution step with an SAI-value > 0.5 was taken as the ADCC titer. No ADCC activity could be detected in any sera against uninfected target cells or in any HIV antibody negative control sera.

The hyperimmune sera determined according to Example 5 above were tested in an ADCC assay as described above. The results for ADCC positive sera only are presented in Table 7 below. All other sera in the group were ADCC negative. All preimmune sera in monkeys 1-40 were negative against infected target cells except serum no. 36 that had a titer of 1:30.

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All preimmune and hyperimmune sera were ADCC negative against uninfected target cells.

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TABLE 7		
ADCC positive anti-sera raised in monkeys against peptides representing HIV-1 _{HTLVIII_B} gp120		
anti-sera against	amino acid #	ADCC titer
gp120-1	1-28	7290*
gp120-5	65-89	2430
gp120-6	75-100	2430
gp120-7	90-116	810
gp120-8	101-126	90
gp120-12	152-176	2430
gp120-14	177-205	90
gp120-16/B	213-224	2430
gp120-19	248-269	7290
gp120-20	258-282	2430
gp120-21	270-295	90
gp120-23	296-320	90
gp120-24	307-330	30
gp120-36	445-466	2430

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* This serum was negative in one out of three experiments; in two experiments the ADCC titer was 7290.

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The results depicted in Table 7 indicate that the peptides of the present invention include linear ADCC epitopes specific for HIV-1_{HTLVIII_B} gp120. Thus, the peptides of the present invention can be used to induce antibody-dependent cellular cytotoxicity to aid in the prevention of infection by HIV or to induce a heightened immune response in subjects already infected with HIV.

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To determine the precise amino acids necessary for the active epitope for each of the novel peptides of the present invention, deletion analysis can be performed as described in the following example.

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Example 7Deletion Analysis of the Peptides

The peptides of the present invention may be used in exactly the form described herein, or may be used in supplemented or truncated active form. In order to determine whether removal or addition of amino acids to the sequence affects the beneficial properties of that sequence as described above, routine experimentation may be conducted to identify that portion of the sequence containing the active epitope. For example, deletion analysis is performed on gp120-1 by synthesizing peptides lacking one, two, three, or more amino acids from the carboxy terminus, from the amino terminus, or both, and testing those peptides systematically in accordance with Examples 4-6. If the resulting truncated peptide is immunologically equivalent to the untruncated form in generating protective or neutralizing antibodies, then one can conclude that the epitope responsible for the properties in question is found within the truncated sequence. Similarly, the sequences can be tested after addition of one, two, three, or more amino acids (selected from any desired amino acid) to either end of the peptide. If the resulting peptide substantially retains the properties identified in Examples 4-6 for the unmodified peptide, the modified peptide is considered immunologically equivalent for purposes of the present invention.

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In addition to synthesizing the peptides to be tested *de novo*, amino acids can be chemically removed from the peptides of any of the SEQ ID NOs disclosed herein. For example, amino acids can be removed using the method disclosed in Morrison and Boyd, Organic Chemistry, 3d edition, pp. 1145-1146 (1976), the disclosure of which is hereby incorporated by reference. Briefly, phenyl isothiocyanate is used to form a substituted thiourea on the N-terminal residue of the peptide. Mild

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hydrolysis with hydrochloric acid selectively removes the N-terminal residue as the phenylthiohydantoin. The remaining peptide chain is left intact, and is assayed for immunologic activity according to the methods disclosed in Examples 4-6 described above. The procedure is then repeated, sequentially removing the N-terminal residue from the remaining peptide chain and testing the resulting peptide for its ability to induce HIV-specific ADCC, until this ability is lost. In this manner, the amino acid sequence of the active epitope is determined.

Alternatively, the C-terminal amino acid is removed selectively using the enzyme carboxypeptidase to cleave only the peptide linkages adjacent to the free alpha-carboxyl group. In addition, enzymes such as trypsin, chymotrypsin and pepsin may be used to reduce the peptides of the present invention into smaller fragments, which are then analyzed according to the methods described above in Examples 4-6.

While particular embodiments of the invention have been described in detail, it will be apparent to those skilled in the art that these embodiments are exemplary rather than limiting, and the true scope of the invention is that defined by the claims which follow.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Syntello Vaccine Development AB
- (ii) TITLE OF INVENTION: PEPTIDES FOR USE IN VACCINATION AND INDUCTION OF NEUTRALIZING ANTIBODIES AGAINST HUMAN IMMUNODEFICIENCY VIRUS
- (iii) NUMBER OF SEQUENCES: 41
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Syntello Vaccine Development AB
 - (B) STREET: Guldhedsgatan 10 B
 - (C) CITY: S-411 46 Göteborg
 - (D) STATE:
 - (E) COUNTRY: Sweden
 - (F) ZIP:
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: AWAPATENT AB, Stockholm
 - (B) REGISTRATION NUMBER:
 - (C) REFERENCE/DOCKET NUMBER: 2948411
- (ix) TELECOMMUNICATION INFORMATION:
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(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

WHAT IS CLAIMED IS:

1. A peptide comprising an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41 and wherein antisera raised in monkeys against said epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30.

2. A peptide according to Claim 1 wherein said epitopic sequence has an amino acid sequence that consists essentially of a sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41.

3. A vaccine composition comprising a peptide according to Claim 1, in an amount effective to induce an immune response in a mammal together with a pharmaceutically acceptable carrier.

4. The vaccine composition of Claim 3, further comprising an adjuvant.

5. The vaccine composition of Claim 4, wherein said adjuvant is selected from the group consisting of Freund's complete adjuvant, Freund's incomplete adjuvant, muramyl dipeptide, levamisole, isoprinosine and tuftsin.

6. A vaccine composition comprising at least two peptides, wherein each of said peptides comprises a peptide according to any one of Claim 1, said at least 2 peptides being in an amount effective to induce an immune response in a mammal together with a pharmaceutically acceptable carrier.

7. The vaccine composition of Claim 6, further comprising an adjuvant.

8. The vaccine composition of Claim 7, wherein said adjuvant is selected from the group consisting of Freund's complete adjuvant, Freund's incomplete adjuvant, muramyl dipeptide, levamisole, isoprinosine and tuftsin.

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9. A method of protecting a mammal from infection with human immunodeficiency virus, comprising administering to said mammal a composition according to Claim 3.

5 10. The method of Claim 9, wherein said composition is a composition according to Claim 6.

11. The method of Claim 9, wherein said administration step comprises intravenous, intramuscular, subcutaneous, or intraperitoneal injection.

10 12. A method for inducing neutralizing anti-HIV antibodies in a mammal, comprising the step of administering an effective antibody-inducing amount of a composition according to Claim 3.

13. The method of Claim 12, wherein said composition is a composition according to Claim 6.

15 14. Use of a peptide comprising an epitopic amino acid sequence from human immunodeficiency virus gp120 protein, wherein the epitope is located within SEQ ID NO:1, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:36 or SEQ ID NO:41 and wherein antisera raised in
20 monkeys against said epitopic sequence has a specific antibody-dependent cellular cytotoxicity index value greater than 0.5 at a dilution greater than 1:30 for the preparation of a pharmaceutical composition for immunizing a mammal
25 against infection with human immunodeficiency virus.

15. The use according to Claim 14, wherein said peptide is a peptide according to Claim 6.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/SE 94/00340

A. CLASSIFICATION OF SUBJECT MATTER
IPC 5 A61K39/21 C07K7/08 C07K7/10

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED
Minimum documentation searched (classification system followed by classification symbols)
IPC 5 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,92 21377 (SYNTELLO VACCINE DEVELOPMENT AB) 10 December 1992 see the whole document ---	1-15
X	WO,A,92 05800 (SYNTELLO VACCINE DEVELOPMENT AB) 16 April 1992 see the whole document ---	1-15
	-/--	

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search 22 August 1994	Date of mailing of the international search report 31. 08. 94
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Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax (+ 31-70) 340-3016	Authorized officer Cupido, M
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INTERNATIONAL SEARCH REPORT

International application No.

PCT/SE 94/00340

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 9-13 are directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged effects of the composition.
2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest.
- No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/SE 94/00340

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA. vol. 88, no. 23 , 1 December 1991 , WASHINGTON US pages 10744 - 10748 A.VAHLNE ET AL. 'Immunisation of monkeys with synthetic peptides disclose conserved areas on gp120 of human immunodeficiency virus type 1 associated with cross-neutralising antibodies and T-cell recognition'</p> <p style="text-align: center;">---</p>	1-15
X	<p>IMMUNOLOGY vol. 76, no. 4 , August 1992 , OXFORD, GB pages 515 - 534 D.F. NIXON ET AL. 'Cellular and humoral antigenic epitopes in HIV and SIV' see page 519 - page 522</p> <p style="text-align: center;">-----</p>	1-15

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/SE 94/00340

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9221377	10-12-92	AU-A- 1906592	08-01-93
		EP-A- 0594638	04-05-94

WO-A-9205800	16-04-92	AU-B- 650911	07-07-94
		AU-A- 8643591	28-04-92
		CA-A- 2091263	28-03-92
		EP-A- 0550599	14-07-93
		JP-T- 6501260	10-02-94
