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(54) Title: PRODUCTION OF STEVIOL GLYCOSIDES IN RECOMBINANT HOSTS

(57) Abstract: The invention relates to recombinant microorganisms and methods for producing steviol glycosides and steviol glycoside precursors.

PRODUCTION OF STEVIOL GLYCOSIDES IN RECOMBINANT HOSTS

BACKGROUND OF THE INVENTION

Field of the Invention

[0001] This disclosure relates to recombinant production of steviol glycosides and steviol glycoside precursors in recombinant hosts. In particular, this disclosure relates to production of steviol glycosides comprising steviol-13-O-glucoside (13-SMG), rubusoside, rebaudioside B (RebB), rebaudioside A (RebA), rebaudioside D (RebD), and rebaudioside M (RebM) in recombinant hosts comprising genes involved in uridine diphosphate (UDP)-glucose formation.

Description of Related Art

[0002] Sweeteners are well known as ingredients used most commonly in the food, beverage, or confectionary industries. The sweetener can either be incorporated into a final food product during production or for stand-alone use, when appropriately diluted, as a tabletop sweetener or an at-home replacement for sugars in baking. Sweeteners include natural sweeteners such as sucrose, high fructose corn syrup, molasses, maple syrup, and honey and artificial sweeteners such as aspartame, saccharine, and sucralose. Stevia extract is a natural sweetener that can be isolated and extracted from a perennial shrub, *Stevia rebaudiana*. Stevia is commonly grown in South America and Asia for commercial production of stevia extract. Stevia extract, purified to various degrees, is used commercially as a high intensity sweetener in foods and in blends or alone as a tabletop sweetener. Extracts of the Stevia plant generally comprise steviol glycosides that contribute to the sweet flavor, although the amount of each steviol glycoside often varies, *inter alia*, among different production batches.

[0003] Chemical structures for several steviol glycosides are shown in Figure 2, including the diterpene steviol and various steviol glycosides. Extracts of the Stevia plant generally comprise steviol glycosides that contribute to the sweet flavor, although the amount of each steviol glycoside often varies, *inter alia*, among different production batches.

[0004] As recovery and purification of steviol glycosides from the Stevia plant have proven to be labor intensive and inefficient, there remains a need for a recombinant production system that can accumulate high yields of desired steviol glycosides, such as RebM. There also remains a need for improved production of steviol glycosides in recombinant hosts for

commercial uses. As well, there remains a need for increasing UDP-glucose formation in recombinant hosts in order to produce higher yields of steviol glycosides, including RebM.

SUMMARY OF THE INVENTION

[0005] It is against the above background that the present invention provides certain advantages and advancements over the prior art.

[0006] Although this invention as disclosed herein is not limited to specific advantages or functionalities, the invention provides a recombinant host cell capable of producing one or more steviol glycosides or a steviol glycoside composition in a cell culture, comprising:

- (a) a recombinant gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP);
- (b) a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate; and/or
- (c) a recombinant gene encoding a polypeptide capable of synthesizing uridine diphosphate glucose (UDP-glucose) from UTP and glucose-1-phosphate.

[0007] In one aspect of the recombinant host cell disclosed herein:

- (a) the polypeptide capable of synthesizing UTP from UDP comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
- (b) the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:143 or a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:141, SEQ ID NO:145, or SEQ ID NO:147; and/or
- (c) the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:127, a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID

NO:137, or SEQ ID NO:139 or a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131.

[0008] In one aspect, the recombinant host cell disclosed herein further comprises:

- (a) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof;
- (b) a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;
- (c) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof; and/or
- (d) a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside.

[0009] In one aspect, the recombinant host cell disclosed herein further comprises:

- (e) a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP);
- (f) a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP;
- (g) a gene encoding an a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate;
- (h) a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid from *ent*-kaurene;
- (i) a gene encoding a polypeptide capable of reducing cytochrome P450 complex; and/or
- (j) a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid.

[0010] In one aspect of the recombinant host cell disclosed herein:

- (a) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;

- (b) the polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (c) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
- (d) the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16;
- (e) the polypeptide capable of synthesizing GGPP comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, or SEQ ID NO:116;
- (f) the polypeptide capable of synthesizing ent-copalyl diphosphate comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, or SEQ ID NO:120;
- (g) the polypeptide capable of synthesizing ent-kaurene comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:52;
- (h) the polypeptide capable of synthesizing ent-kaurenoic acid comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:117, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, or SEQ ID NO:76;
- (i) the polypeptide capable of reducing cytochrome P450 complex comprises a polypeptide having at least 70% sequence identity to the amino acid sequence

set forth in SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92; and/or

- (k) the polypeptide capable of synthesizing steviol comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:94, SEQ ID NO:97, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO:110, SEQ ID NO:112, or SEQ ID NO:114.

[0011] In one aspect, the recombinant host cell disclosed herein comprises:

- (a) a gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP) having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
- (b) one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, each having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2 and/or SEQ ID NO:119; and
- (c) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121.

[0012] In one aspect, the recombinant host cell disclosed herein comprises:

- (a) a gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP);
- (b) a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate;
- (c) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121;
- (d) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139; at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:127; or at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131; and

one or more of:

- (e) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;
- (b) a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (c) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
- (d) a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16.

[0013] In one aspect, the recombinant host cell disclosed herein comprises:

- (a) a recombinant gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP) having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
- (b) one or more recombinant genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, each having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2 and/or SEQ ID NO:119; and/or
- (c) a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121;

wherein the gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP), the one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or the gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and

glucose-1-phosphate are overexpressed relative to a corresponding host cell lacking the one or more recombinant genes.

[0014] In one aspect of the recombinant host cell disclosed herein, the gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP), the one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or the gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate are overexpressed by at least 10%, or at least 15%, or at least 20%, or at least 30%, or at least 40%, or at least 50%, or at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 100%, or at least 125%, or at least 150%, or at least 175%, or at least 200% relative to a corresponding host cell lacking the one or more recombinant genes.

[0015] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increase the amount of UDP-glucose accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.

[0016] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases the amount of UDP-glucose accumulated by the cell by at least about 10%, at least about 25%, or at least about 50%, at least about 100%, at least about 150%, at least about 200%, or at least about 250% relative to a corresponding host lacking the one or more recombinant genes.

[0017] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases an amount of the one or more steviol glycosides or the steviol glycoside composition produced by the cell relative to a corresponding host lacking the one or more recombinant genes.

[0018] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases the amount of the one or more steviol glycosides produced by the cell by at least about 5%, at least about 10%, at least about 25%, at least about 50%, at least about 75%, or at least about 100% relative to a corresponding host lacking the one or more recombinant genes.

[0019] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases the amount of RebA, RebB, Reb D, and/or RebM produced by the cell relative to a corresponding host lacking the one or more recombinant genes.

[0020] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes decreases the one of one or more steviol glycosides or the steviol glycoside composition accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.

[0021] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes decreases the amount of the one or more steviol glycosides accumulated by the cell by at least about 5%, at least about 10%, at least about 25%, or at least about 50% relative to a corresponding host lacking the one or more recombinant genes.

[0022] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes decreases the amount of RebB, RebD, and/or 13-SMG accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.

[0023] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases or decreases the amount of total steviol glycosides produced by the cell by less than 5%, less than 2.5%, or less than 1% relative to a corresponding host lacking the one or more recombinant genes.

[0024] In one aspect of the recombinant host cell disclosed herein, expression of the one or more recombinant genes increases the amount of total steviol glycosides produced by the cell by at least about 5%, at least about 10%, or at least about 25% relative to a corresponding host lacking the one or more recombinant genes.

[0025] In one aspect of the recombinant host cell disclosed herein, the one or more steviol glycosides is, or the steviol glycoside composition comprises, steviol-13-O-glucoside (13-SMG), steviol-1,2-Bioside, steviol-1,3-Bioside, steviol-19-O-glucoside (19-SMG), 1,2-Stevioside, 1,3-stevioside (RebG), rubusoside, rebaudioside A (RebA), rebaudioside B (RebB), rebaudioside C (RebC), rebaudioside D (RebD), rebaudioside E (RebE), rebaudioside F (RebF), rebaudioside M (RebM), rebaudioside Q (RebQ), rebaudioside I (RebI), dulcoside A, and/or an isomer thereof.

[0026] In one aspect of the recombinant host cell disclosed herein, the recombinant host cell is a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell or a bacterial cell.

[0027] The invention also provides method of producing one or more steviol glycosides or a steviol glycoside composition in a cell culture, comprising culturing the recombinant host cell disclosed herein, under conditions in which the genes are expressed, and wherein the one or

more steviol glycosides or the steviol glycoside composition is produced by the recombinant host cell.

[0028] In one aspect of the methods disclosed herein, the genes are constitutively expressed and/or expression of the genes is induced.

[0029] In one aspect of the methods disclosed herein, the amount of UDP-glucose accumulated by the cell is increased by at least by at least about 10% relative to a corresponding host lacking the one or more recombinant genes.

[0030] In one aspect of the methods disclosed herein, the amount of RebA, RebB, RebD, and/or RebM produced by the cell is increased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.

[0031] In one aspect of the methods disclosed herein, the amount of RebB, RebD, and/or 13-SMG accumulated by the cell is decreased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.

[0032] In one aspect of the methods disclosed herein, the amount of total steviol glycosides produced by the cell is increased or decreased by less than about 5% relative to a corresponding host lacking the one or more recombinant genes.

[0033] In one aspect of the methods disclosed herein, the amount of total steviol glycosides produced by the cell is increased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.

[0034] In one aspect of the methods disclosed herein, the recombinant host cell is grown in a fermentor at a temperature for a period of time, wherein the temperature and period of time facilitate the production of the one or more steviol glycosides or the steviol glycoside composition.

[0035] In one aspect of the methods disclosed herein, the amount of UDP-glucose present in the cell culture is increased by at least about 10%, at least about 25%, or at least about 50%, at least about 100%, at least about 150%, at least about 200%, or at least about 250% at any point throughout the period of time.

[0036] In one aspect, the methods disclosed herein further comprise isolating the produced one or more steviol glycosides or the steviol glycoside composition from the cell culture.

[0037] In one aspect of the methods disclosed herein, the isolating step comprises:

- (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;
 - (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
 - (c) providing one or more adsorbent resins, comprising providing the adsorbent resins in a packed column; and
 - (d) contacting the supernatant of step (b) with the one or more adsorbent resins in order to obtain at least a portion of the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition;
- or
- (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;
 - (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
 - (c) providing one or more ion exchange or ion exchange or reversed-phase chromatography columns; and
 - (d) contacting the supernatant of step (b) with the one or more ion exchange or ion exchange or reversed-phase chromatography columns in order to obtain at least a portion of the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition;
- or
- (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;
 - (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
 - (c) crystallizing or extracting the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition.

[0038] In one aspect, the methods disclosed herein further comprise recovering the one or more steviol glycosides or the steviol glycoside composition from the cell culture.

[0039] In one aspect of the methods disclosed herein, the recovered one or more steviol glycosides or the steviol glycoside composition has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

[0040] The invention also provides a method for producing one or more steviol glycosides or a steviol glycoside composition, comprising whole-cell bioconversion of plant-derived or synthetic steviol and/or steviol glycosides in a cell culture medium of a recombinant host cell using:

- (a) a polypeptide capable of synthesizing UTP from UDP having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
- (b) a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, or SEQ ID NO:143; at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:141, SEQ ID NO:145, or SEQ ID NO:147; and/or
- (c) a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:127; at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139; or at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131; and

one or more of:

- (d) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof;
- (e) a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;
- (f) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof; and/or
- (g) a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;

wherein at least one of the polypeptides is a recombinant polypeptide expressed in the recombinant host cell; and producing the one or more steviol glycosides or the steviol glycoside composition thereby.

[0041] In one aspect of the methods disclosed herein:

- (d) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;
- (e) the polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (f) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
- (g) the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16.

[0042] In one aspect of the methods disclosed herein, the recombinant host cell is a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell or a bacterial cell.

[0043] In one aspect of the methods disclosed herein, the one or more steviol glycosides is, or the steviol glycoside composition comprises, steviol-13-O-glucoside (13-SMG), steviol-1,2-Bioside, steviol-1,3-Bioside, steviol-19-O-glucoside (19-SMG), 1,2-stevioside, 1,3-stevioside (RebG), rubusoside, rebaudioside A (RebA), rebaudioside B (RebB), rebaudioside C (RebC), rebaudioside D (RebD), rebaudioside E (RebE), rebaudioside F (RebF), rebaudioside M (RebM), rebaudioside Q (RebQ), rebaudioside I (RebI), dulcoside A, and/or an isomer thereof.

[0044] The invention also provides a cell culture, comprising the recombinant host cell disclosed herein, the cell culture further comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids;

wherein the one or more steviol glycosides or the steviol glycoside composition is present at a concentration of at least 1 mg/liter of the cell culture;

wherein the cell culture is enriched for the one or more steviol glycosides or the steviol glycoside composition relative to a steviol glycoside composition from a Stevia plant and has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

[0045] The invention also provides a cell culture, comprising the recombinant host cell disclosed herein, the cell culture further comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids;

wherein UDP-glucose is present in the cell culture at a concentration of at least 100 μ M;

wherein the cell culture is enriched for UGP-glucose relative to a steviol glycoside composition from a Stevia plant and has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

[0046] The invention also provides cell lysate from the recombinant host cell disclosed herein grown in the cell culture, comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or

(c) supplemental nutrients comprising trace metals, vitamins, salts, yeast nitrogen base, YNB, and/or amino acids;

wherein the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell is present at a concentration of at least 1 mg/liter of the cell culture.

[0047] The invention also provides one or more steviol glycosides produced by the recombinant host cell disclosed herein;

wherein the one or more steviol glycosides produced by the recombinant host cell are present in relative amounts that are different from a steviol glycoside composition from a Stevia plant and have a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

[0048] The invention also provides one or more steviol glycosides produced by the method disclosed herein;

wherein the one or more steviol glycosides produced by the recombinant host cell are present in relative amounts that are different from a steviol glycoside composition from a Stevia plant and have a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

[0049] The invention also provides a sweetener composition, comprising the one or more steviol glycosides disclosed herein.

[0050] The invention also provides a food product comprising, the sweetener composition disclosed herein.

[0051] The invention also provides a beverage or a beverage concentrate, comprising the sweetener composition disclosed herein.

[0052] These and other features and advantages of the present invention will be more fully understood from the following detailed description taken together with the accompanying claims. It is noted that the scope of the claims is defined by the recitations therein and not by the specific discussion of features and advantages set forth in the present description.

BRIEF DESCRIPTION OF THE DRAWINGS

[0053] The following detailed description of the embodiments of the present invention can be best understood when read in conjunction with the following drawings, where like structure is indicated with like reference numerals and in which:

[0054] Figure 1 shows the biochemical pathway for producing steviol from geranylgeranyl diphosphate using geranylgeranyl diphosphate synthase (GGPPS), ent-copalyl diphosphate synthase (CDPS), ent-kaurene synthase (KS), ent-kaurene oxidase (KO), and ent-kaurenoic acid hydroxylase (KAH) polypeptides.

[0055] Figure 2 shows representative primary steviol glycoside glycosylation reactions catalyzed by suitable UGT enzymes and chemical structures for several of the compounds found in *Stevia* extracts.

[0056] Figure 3 shows representative reactions catalyzed by enzymes involved in the UDP-glucose biosynthetic pathway, including uracil permease (FUR4), uracil phosphoribosyltransferase (FUR1), orotate phosphoribosyltransferase 1 (URA5), orotate phosphoribosyltransferase 2 (URA10), orotidine 5'-phosphate decarboxylase (URA3), uridylylate kinase (URA6), nucleoside diphosphate kinase (YNK1), phosphoglucomutase-1 (PGM1), phosphoglucomutase-2 (PGM2), and UTP-glucose-1-phosphate uridylyltransferase (UGP1). See, e.g., Daran *et al.*, 1995, *Eur J Biochem.* 233(2):520-30.

[0057] Skilled artisans will appreciate that elements in the Figures are illustrated for simplicity and clarity and have not necessarily been drawn to scale. For example, the dimensions of some of the elements in the Figures can be exaggerated relative to other elements to help improve understanding of the embodiment(s) of the present invention.

DETAILED DESCRIPTION OF THE INVENTION

[0058] All publications, patents and patent applications cited herein are hereby expressly incorporated by reference for all purposes.

[0059] Before describing the present invention in detail, a number of terms will be defined. As used herein, the singular forms "a," "an," and "the" include plural referents unless the context

clearly dictates otherwise. For example, reference to a “nucleic acid” means one or more nucleic acids.

[0060] It is noted that terms like “preferably,” “commonly,” and “typically” are not utilized herein to limit the scope of the claimed invention or to imply that certain features are critical, essential, or even important to the structure or function of the claimed invention. Rather, these terms are merely intended to highlight alternative or additional features that can or cannot be utilized in a particular embodiment of the present invention.

[0061] For the purposes of describing and defining the present invention it is noted that the term “substantially” is utilized herein to represent the inherent degree of uncertainty that can be attributed to any quantitative comparison, value, measurement, or other representation. The term “substantially” is also utilized herein to represent the degree by which a quantitative representation can vary from a stated reference without resulting in a change in the basic function of the subject matter at issue.

[0062] Methods well known to those skilled in the art can be used to construct genetic expression constructs and recombinant cells according to this invention. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, *in vivo* recombination techniques, and polymerase chain reaction (PCR) techniques. See, for example, techniques as described in Green & Sambrook, 2012, MOLECULAR CLONING: A LABORATORY MANUAL, Fourth Edition, Cold Spring Harbor Laboratory, New York; Ausubel *et al.*, 1989, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Greene Publishing Associates and Wiley Interscience, New York, and PCR Protocols: A Guide to Methods and Applications (Innis *et al.*, 1990, Academic Press, San Diego, CA).

[0063] As used herein, the terms “polynucleotide,” “nucleotide,” “oligonucleotide,” and “nucleic acid” can be used interchangeably to refer to nucleic acid comprising DNA, RNA, derivatives thereof, or combinations thereof, in either single-stranded or double-stranded embodiments depending on context as understood by the skilled worker.

[0064] As used herein, the terms “microorganism,” “microorganism host,” “microorganism host cell,” “recombinant host,” and “recombinant host cell” can be used interchangeably. As used herein, the term “recombinant host” is intended to refer to a host, the genome of which has been augmented by at least one DNA sequence. Such DNA sequences include but are not limited to genes that are not naturally present, DNA sequences that are not normally transcribed into RNA or translated into a protein (“expressed”), and other genes or DNA sequences which

one desires to introduce into a host. It will be appreciated that typically the genome of a recombinant host described herein is augmented through stable introduction of one or more recombinant genes. Generally, introduced DNA is not originally resident in the host that is the recipient of the DNA, but it is within the scope of this disclosure to isolate a DNA segment from a given host, and to subsequently introduce one or more additional copies of that DNA into the same host, e.g., to enhance production of the product of a gene or alter the expression pattern of a gene. In some instances, the introduced DNA will modify or even replace an endogenous gene or DNA sequence by, e.g., homologous recombination or site-directed mutagenesis. Suitable recombinant hosts include microorganisms.

[0065] As used herein, the term “recombinant gene” refers to a gene or DNA sequence that is introduced into a recipient host, regardless of whether the same or a similar gene or DNA sequence may already be present in such a host. “Introduced,” or “augmented” in this context, is known in the art to mean introduced or augmented by the hand of man. Thus, a recombinant gene can be a DNA sequence from another species or can be a DNA sequence that originated from or is present in the same species but has been incorporated into a host by recombinant methods to form a recombinant host. It will be appreciated that a recombinant gene that is introduced into a host can be identical to a DNA sequence that is normally present in the host being transformed, and is introduced to provide one or more additional copies of the DNA to thereby permit overexpression or modified expression of the gene product of that DNA. In some aspects, said recombinant genes are encoded by cDNA. In other embodiments, recombinant genes are synthetic and/or codon-optimized for expression in *S. cerevisiae*.

[0066] As used herein, the term “engineered biosynthetic pathway” refers to a biosynthetic pathway that occurs in a recombinant host, as described herein. In some aspects, one or more steps of the biosynthetic pathway do not naturally occur in an unmodified host. In some embodiments, a heterologous version of a gene is introduced into a host that comprises an endogenous version of the gene.

[0067] As used herein, the term “endogenous” gene refers to a gene that originates from and is produced or synthesized within a particular organism, tissue, or cell. In some embodiments, the endogenous gene is a yeast gene. In some embodiments, the gene is endogenous to *S. cerevisiae*, including, but not limited to *S. cerevisiae* strain S288C. In some embodiments, an endogenous yeast gene is overexpressed. As used herein, the term “overexpress” is used to refer to the expression of a gene in an organism at levels higher than

the level of gene expression in a wild type organism. See, e.g., Prelich, 2012, *Genetics* 190:841-54. See, e.g., Giaever & Nislow, 2014, *Genetics* 197(2):451-65. In some aspects, overexpression can be performed by integration using the USER cloning system; see, e.g., Nour-Eldin *et al.*, 2010, *Methods Mol Biol.* 643:185-200. As used herein, the terms “deletion,” “deleted,” “knockout,” and “knocked out” can be used interchangeably to refer to an endogenous gene that has been manipulated to no longer be expressed in an organism, including, but not limited to, *S. cerevisiae*.

[0068] As used herein, the terms “heterologous sequence” and “heterologous coding sequence” are used to describe a sequence derived from a species other than the recombinant host. In some embodiments, the recombinant host is an *S. cerevisiae* cell, and a heterologous sequence is derived from an organism other than *S. cerevisiae*. A heterologous coding sequence, for example, can be from a prokaryotic microorganism, a eukaryotic microorganism, a plant, an animal, an insect, or a fungus different than the recombinant host expressing the heterologous sequence. In some embodiments, a coding sequence is a sequence that is native to the host.

[0069] A “selectable marker” can be one of any number of genes that complement host cell auxotrophy, provide antibiotic resistance, or result in a color change. Linearized DNA fragments of the gene replacement vector then are introduced into the cells using methods well known in the art (see below). Integration of the linear fragments into the genome and the disruption of the gene can be determined based on the selection marker and can be verified by, for example, PCR or Southern blot analysis. Subsequent to its use in selection, a selectable marker can be removed from the genome of the host cell by, e.g., Cre-LoxP systems (see, e.g., Gossen *et al.*, 2002, *Ann. Rev. Genetics* 36:153-173 and U.S. 2006/0014264). Alternatively, a gene replacement vector can be constructed in such a way as to include a portion of the gene to be disrupted, where the portion is devoid of any endogenous gene promoter sequence and encodes none, or an inactive fragment of, the coding sequence of the gene.

[0070] As used herein, the terms “variant” and “mutant” are used to describe a protein sequence that has been modified at one or more amino acids, compared to the wild-type sequence of a particular protein.

[0071] As used herein, the term “inactive fragment” is a fragment of the gene that encodes a protein having, e.g., less than about 10% (e.g., less than about 9%, less than about 8%, less than about 7%, less than about 6%, less than about 5%, less than about 4%, less than about

3%, less than about 2%, less than about 1%, or 0%) of the activity of the protein produced from the full-length coding sequence of the gene. Such a portion of a gene is inserted in a vector in such a way that no known promoter sequence is operably linked to the gene sequence, but that a stop codon and a transcription termination sequence are operably linked to the portion of the gene sequence. This vector can be subsequently linearized in the portion of the gene sequence and transformed into a cell. By way of single homologous recombination, this linearized vector is then integrated in the endogenous counterpart of the gene with inactivation thereof.

[0072] As used herein, the term “steviol glycoside” refers to rebaudioside A (RebA) (CAS # 58543-16-1), rebaudioside B (RebB) (CAS # 58543-17-2), rebaudioside C (RebC) (CAS # 63550-99-2), rebaudioside D (RebD) (CAS # 63279-13-0), rebaudioside E (RebE) (CAS # 63279-14-1), rebaudioside F (RebF) (CAS # 438045-89-7), rebaudioside M (RebM) (CAS # 1220616-44-3), Rubusoside (CAS # 63849-39-4), Dulcoside A (CAS # 64432-06-0), rebaudioside I (RebI) (MassBank Record: FU000332), rebaudioside Q (RebQ), 1,2-Stevioside (CAS # 57817-89-7), 1,3-Stevioside (RebG), Steviol-1,2-Bioside (MassBank Record: FU000299), Steviol-1,3-Bioside, Steviol-13-O-glucoside (13-SMG), Steviol-19-O-glucoside (19-SMG), a tri-glycosylated steviol glycoside, a tetra-glycosylated steviol glycoside, a penta-glycosylated steviol glycoside, a hexa-glycosylated steviol glycoside, a hepta-glycosylated steviol glycoside, and isomers thereof. See Figure 2; see *also*, Steviol Glycosides Chemical and Technical Assessment 69th JECFA, 2007, prepared by Harriet Wallin, Food Agric. Org.

[0073] As used herein, the terms “steviol glycoside precursor” and “steviol glycoside precursor compound” are used to refer to intermediate compounds in the steviol glycoside biosynthetic pathway. Steviol glycoside precursors include, but are not limited to, geranylgeranyl diphosphate (GGPP), *ent*-copalyl-diphosphate, *ent*-kaurene, *ent*-kaurenol, *ent*-kaurenal, *ent*-kaurenoic acid, and steviol. See Figure 1. In some embodiments, steviol glycoside precursors are themselves steviol glycoside compounds. For example, 19-SMG, rubusoside, 1,2-stevioside, and RebE are steviol glycoside precursors of RebM. See Figure 2.

[0074] Also as used herein, the terms “steviol precursor” and “steviol precursor compound” are used to refer to intermediate compounds in the steviol biosynthetic pathway. Steviol precursors may also be steviol glycoside precursors, and include, but are not limited to, geranylgeranyl diphosphate (GGPP), *ent*-copalyl-diphosphate, *ent*-kaurene, *ent*-kaurenol, *ent*-kaurenal, and *ent*-kaurenoic acid. Steviol glycosides and/or steviol glycoside precursors can be produced *in vivo* (*i.e.*, in a recombinant host), *in vitro* (*i.e.*, enzymatically), or by whole cell

bioconversion. As used herein, the terms “produce” and “accumulate” can be used interchangeably to describe synthesis of steviol glycosides and steviol glycoside precursors *in vivo*, *in vitro*, or by whole cell bioconversion.

[0075] As used herein, the terms “culture broth,” “culture medium,” and “growth medium” can be used interchangeably to refer to a liquid or solid that supports growth of a cell. A culture broth can comprise glucose, fructose, sucrose, trace metals, vitamins, salts, yeast nitrogen base (YNB), and/or amino acids. The trace metals can be divalent cations, including, but not limited to, Mn^{2+} and/or Mg^{2+} . In some embodiments, Mn^{2+} can be in the form of $MnCl_2$ dihydrate and range from approximately 0.01 g/L to 100 g/L. In some embodiments, Mg^{2+} can be in the form of $MgSO_4$ heptahydrate and range from approximately 0.01 g/L to 100 g/L. For example, a culture broth can comprise i) approximately 0.02-0.03 g/L $MnCl_2$ dihydrate and approximately 0.5-3.8 g/L $MgSO_4$ heptahydrate, ii) approximately 0.03-0.06 g/L $MnCl_2$ dihydrate and approximately 0.5-3.8 g/L $MgSO_4$ heptahydrate, and/or iii) approximately 0.03-0.17 g/L $MnCl_2$ dihydrate and approximately 0.5-7.3 g/L $MgSO_4$ heptahydrate. Additionally, a culture broth can comprise one or more steviol glycosides produced by a recombinant host, as described herein.

[0076] Recombinant steviol glycoside-producing *Saccharomyces cerevisiae* (*S. cerevisiae*) strains are described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328, each of which is incorporated by reference in their entirety. Methods of producing steviol glycosides in recombinant hosts, by whole cell bio-conversion, and *in vitro* are also described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328.

[0077] In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP) (e.g., geranylgeranyl diphosphate synthase (GGPPS)); a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP (e.g., *ent*-copalyl diphosphate synthase (CDPS)); a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate (e.g., kaurene synthase (KS)); a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene (e.g., kaurene oxidase (KO)); a gene encoding a polypeptide capable of reducing cytochrome P450 complex (e.g., cytochrome P450 reductase (CPR) or P450 oxidoreductase (POR); for example, but not limited to a polypeptide capable of electron transfer from NADPH to cytochrome P450 complex during conversion of NADPH to $NADP^+$, which is utilized as a cofactor for terpenoid biosynthesis); a gene encoding a

polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid (e.g., steviol synthase (KAH)); and/or a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate (e.g., an *ent*-copalyl diphosphate synthase (CDPS) – *ent*-kaurene synthase (KS) polypeptide) can produce steviol *in vivo*. See, e.g., Figure 1. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

[0078] In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group (e.g., UGT85C2 polypeptide); a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., UGT76G1 polypeptide); a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group (e.g., UGT74G1 polypeptide); and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., UGT91D2 and EUGT11 polypeptide) can produce a steviol glycoside *in vivo*. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

[0079] In some embodiments, steviol glycosides and/or steviol glycoside precursors are produced *in vivo* through expression of one or more enzymes involved in the steviol glycoside biosynthetic pathway in a recombinant host. For example, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP); a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP; a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate; a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenol from *ent*-kaurene; a gene encoding a polypeptide capable of reducing cytochrome P450 complex; a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate; a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group (e.g., UGT85C2 polypeptide); a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-

glucose of a steviol glycoside (e.g., UGT76G1 polypeptide); a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group (e.g., UGT74G1 polypeptide); and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., UGT91D2 and EUGT11 polypeptide) can produce a steviol glycoside and/or steviol glycoside precursors *in vivo*. See, e.g., Figures 1 and 2. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

[0080] In some embodiments, a steviol-producing recombinant microorganism comprises heterologous nucleic acids encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group; a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group; and a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside.

[0081] In some embodiments, a steviol-producing recombinant microorganism comprises heterologous nucleic acids encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group, a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside, and a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside polypeptides.

[0082] In some aspects, a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group, a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside, a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group, and/or a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside, transfers a glucose molecule from uridine diphosphate glucose (UDP-glucose) to steviol and/or a steviol glycoside.

[0083] In some aspects, UDP-glucose is produced *in vivo* through expression of one or more enzymes involved in the UDP-glucose biosynthetic pathway in a recombinant host. For example, a recombinant host comprising a gene encoding a polypeptide capable of transporting

uracil into the host cell (e.g., uracil permease (FUR4)); a gene encoding a polypeptide capable of synthesizing uridine monophosphate (UMP) from uracil (e.g., uracil phosphoribosyltransferase (FUR1)); a gene encoding a polypeptide capable of synthesizing orotidine monophosphate (OMP) from orotate or orotic acid (e.g., orotate phosphoribosyltransferase 1 (URA5) and orotate phosphoribosyltransferase 2 (URA10)); a gene encoding a polypeptide capable of synthesizing UMP from OMP (e.g., orotidine 5'-phosphate decarboxylase (URA3)); a gene encoding a polypeptide capable of synthesizing uridine diphosphate (UDP) from UMP (e.g., uridylate kinase (URA6)); a gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from UDP (i.e., a polypeptide capable of catalyzing the transfer of gamma phosphates from nucleoside triphosphates, e.g., nucleoside diphosphate kinase (YNK1)); a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., phosphoglucomutase-1 (PGM1) and phosphoglucomutase-2 (PGM2)); and/or a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., UTP-glucose-1-phosphate uridylyltransferase (UGP1) can produce UDP-glucose *in vivo*. See, e.g., Figure 3. The skilled worker will appreciate that one or more of these genes may be endogenous to the host.

[0084] In some embodiments, a recombinant host comprises a gene encoding a polypeptide capable of synthesizing UTP from UDP. In some aspects, the gene encoding a polypeptide capable of synthesizing UTP from UDP is a recombinant gene. In some aspects, the recombinant gene comprises a nucleotide sequence native to the host. In other aspects, the recombinant gene comprises a heterologous nucleotide sequence. In some aspects, the recombinant gene is operably linked to a promoter. In some aspects, the recombinant gene is operably linked to a terminator, for example but not limited to, tCYC1 (SEQ ID NO:154) or tADH1 (SEQ ID NO:155). In some aspects, the promoter and terminator drive high expression of the recombinant gene. In some aspects, the recombinant gene is operably linked to a strong promoter, for example but not limited to, pTEF1 (SEQ ID NO:148), pPGK1 (SEQ ID NO:149), pTDH3 (SEQ ID NO:150), pTEF2 (SEQ ID NO:151), pTPI1 (SEQ ID NO:152), or pPDC1 (SEQ ID NO:153). In some aspects, the recombinant gene comprises a nucleotide sequence that originated from or is present in the same species as the recombinant host. In some aspects, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP results in a total expression level of genes encoding a polypeptide capable of synthesizing UTP from UDP that is higher than the expression level of endogenous genes encoding a

polypeptide capable of synthesizing UTP from UDP, *i.e.*, an overexpression of a polypeptide capable of synthesizing UTP from UDP.

[0085] In some aspects, the gene encoding the polypeptide capable of synthesizing UTP from UDP is a gene present in the same species as the recombinant host, *i.e.*, an endogenous gene. In some embodiments, the wild-type promoter of an endogenous gene encoding the polypeptide capable of synthesizing UTP from UDP can be exchanged for a strong promoter. In some aspects, the strong promoter drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In other embodiments, the wild-type enhancer of an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP can be exchanged for a strong enhancer. In some embodiments, the strong enhancer drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In some embodiments, both the wild-type enhancer (*i.e.*, operably linked to the promoter) and the wild-type promoter (*i.e.*, operably linked to the endogenous gene) of the endogenous gene can be exchanged for a strong enhancer and strong promoter, respectively, resulting in overexpression of a polypeptide capable of synthesizing UTP from UDP (*i.e.*, relative to the expression level of endogenous genes operably linked to wild-type enhancers and/or promoters). The endogenous gene operably linked to the strong enhancer and/or promoter may be located at the native loci, and/or may be located elsewhere in the genome.

[0086] For example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, comprising a nucleotide sequence native to the host, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In another example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, comprising a heterologous nucleotide sequence, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In yet another example, in some embodiments, a recombinant host comprises an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP, operably linked to, *e.g.*, a strong promoter native to the host, or a heterologous promoter.

[0087] The person of ordinary skill in the art will appreciate that, e.g., expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP; expression of a recombinant gene and an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP, and expression of an endogenous gene encoding a polypeptide capable of synthesizing UTP from UDP, wherein the wild-type promoter and/or enhancer of the endogenous gene are exchanged for a strong promoter and/or enhancer, each result in overexpression of a polypeptide capable of synthesizing UTP from UDP relative to a corresponding host not expressing a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP and/or a corresponding host expressing only a native gene encoding a polypeptide capable of synthesizing UTP from UDP, operably linked to the wild-type promoter and enhancer—*i.e.*, as used herein, the term “expression” may include “overexpression.”

[0088] In some embodiments, a polypeptide capable of synthesizing UTP from UDP is overexpressed such that the total expression level of genes encoding the polypeptide capable of synthesizing UTP from UDP is at least 5% higher than the expression level of endogenous genes encoding a polypeptide capable of synthesizing UTP from UDP. In some embodiments, the total expression level of genes encoding a polypeptide capable of synthesizing UTP from UDP is at least 10%, or at least 15%, or at least 20%, or at least 30%, or at least 40%, or at least 50%, or at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 100%, or at least 125%, or at least 150%, or at least 175%, or at least 200% higher than the expression level of endogenous genes encoding a polypeptide capable of synthesizing UTP from UDP.

[0089] In some embodiments, a recombinant host comprises a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate. In some aspects, the gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate is a recombinant gene. In some aspects, the recombinant gene comprises a nucleotide sequence native to the host. In other aspects, the recombinant gene comprises a heterologous nucleotide sequence. In some aspects, the recombinant gene is operably linked to a promoter. In some aspects, the recombinant gene is operably linked to a terminator, for example but not limited to, tCYC1 (SEQ ID NO:154) or tADH1 (SEQ ID NO:155). In some aspects, the promoter and terminator drive high expression of the recombinant gene. In some aspects, the recombinant gene is operably linked to a strong promoter, for example but not limited to, pTEF1 (SEQ ID NO:148), pPGK1 (SEQ ID NO:149), pTDH3 (SEQ ID NO:150), pTEF2 (SEQ ID NO:151), pTPI1 (SEQ ID NO:152), or pPDC1 (SEQ ID NO:153). In some aspects, the recombinant gene

comprises a nucleotide sequence that originated from or is present in the same species as the recombinant host. In some aspects, expression of a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate results in a total expression level of genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate that is higher than the expression level of endogenous genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, *i.e.*, an overexpression of a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate.

[0090] In some aspects, the gene encoding the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate is a gene present in the same species as the recombinant host, *i.e.*, an endogenous gene. In some embodiments, the wild-type promoter of an endogenous gene encoding the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate can be exchanged for a strong promoter. In some aspects, the strong promoter drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In other embodiments, the wild-type enhancer of an endogenous gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate can be exchanged for a strong enhancer. In some embodiments, the strong enhancer drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In some embodiments, both the wild-type enhancer (*i.e.*, operably linked to the promoter) and the wild-type promoter (*i.e.*, operably linked to the endogenous gene) of the endogenous gene can be exchanged for a strong enhancer and strong promoter, respectively, resulting in overexpression of a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate (*i.e.*, relative to the expression level of endogenous genes operably linked to wild-type enhancers and/or promoters). The endogenous gene operably linked to the strong enhancer and/or promoter may be located at the native loci, and/or may be located elsewhere in the genome.

[0091] For example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, comprising a nucleotide sequence native to the host, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In another example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable

of converting glucose-6-phosphate to glucose-1-phosphate, comprising a heterologous nucleotide sequence, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In yet another example, in some embodiments, a recombinant host comprises an endogenous gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, operably linked to, *e.g.*, a strong promoter native to the host, or a heterologous promoter.

[0092] In some embodiments, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate is overexpressed such that the total expression level of genes encoding the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate is at least 5% higher than the expression level of endogenous genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate. In some embodiments, the total expression level of genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate is at least 10%, or at least 15%, or at least 20%, or at least 30%, or at least 40%, or at least 50%, or at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 100%, or at least 125%, or at least 150%, or at least 175%, or at least 200% higher than the expression level of endogenous genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate.

[0093] In some embodiments, a recombinant host comprises a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate. In some aspects, the gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate is a recombinant gene. In some aspects, the recombinant gene comprises a nucleotide sequence native to the host. In other aspects, the recombinant gene comprises a heterologous nucleotide sequence. In some aspects, the recombinant gene is operably linked to a promoter. In some aspects, the recombinant gene is operably linked to a terminator, for example but not limited to, tCYC1 (SEQ ID NO:154) or tADH1 (SEQ ID NO:155). In some aspects, the promoter and terminator drive high expression of the recombinant gene. In some aspects, the recombinant gene is operably linked to a strong promoter, for example but not limited to, pTEF1 (SEQ ID NO:148), pPGK1 (SEQ ID NO:149), pTDH3 (SEQ ID NO:150), pTEF2 (SEQ ID NO:151), pTPI1 (SEQ ID NO:152), or pPDC1 (SEQ ID NO:153). In some aspects, the recombinant gene comprises a nucleotide sequence that originated from or is present in the same species as the recombinant host. In some aspects, expression of a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate results in a total expression level of genes encoding a polypeptide capable

of synthesizing UDP-glucose from UTP and glucose-1-phosphate that is higher than the expression level of endogenous genes encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, *i.e.*, an overexpression of a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate.

[0094] In some aspects, the gene encoding the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate is a gene present in the same species as the recombinant host, *i.e.*, an endogenous gene. In some embodiments, the wild-type promoter of an endogenous gene encoding the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate can be exchanged for a strong promoter. In some aspects, the strong promoter drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In other embodiments, the wild-type enhancer of an endogenous gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate can be exchanged for a strong enhancer. In some embodiments, the strong enhancer drives high expression of the endogenous gene (*i.e.*, overexpression of the gene). In some embodiments, both the wild-type enhancer (*i.e.*, operably linked to the promoter) and the wild-type promoter (*i.e.*, operably linked to the endogenous gene) of the endogenous gene can be exchanged for a strong enhancer and strong promoter, respectively, resulting in overexpression of a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*i.e.*, relative to the expression level of endogenous genes operably linked to wild-type enhancers and/or promoters). The endogenous gene operably linked to the strong enhancer and/or promoter may be located at the native loci, and/or may be located elsewhere in the genome.

[0095] For example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, comprising a nucleotide sequence native to the host, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In another example, in some embodiments, a recombinant host comprising an endogenous gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, operably linked to a wild-type promoter, further comprises a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, comprising a heterologous nucleotide sequence, operably linked to, *e.g.*, a wild-type promoter, a promoter native to the host, or a heterologous promoter. In yet another example, in some embodiments,

a recombinant host comprises an endogenous gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, operably linked to, e.g., a strong promoter native to the host, or a heterologous promoter.

[0096] In some embodiments, a recombinant host comprising a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate is overexpressed such that the total expression level of genes encoding the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate is at least 5% higher than the expression level of endogenous genes encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate. In some embodiments, the total expression level of genes encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate is at least 10%, or at least 15%, or at least 20%, or at least 30%, or at least 40%, or at least 50%, or at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 100%, or at least 125%, or at least 150%, or at least 175%, or at least 200% higher than the expression level of endogenous genes encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate.

[0097] In some aspects, a recombinant host comprising one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP, one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate may further comprise a recombinant gene encoding a polypeptide capable of transporting uracil into the host cell; a recombinant gene encoding a polypeptide capable of synthesizing uridine monophosphate (UMP) from uracil; a recombinant gene encoding a polypeptide capable of synthesizing orotidine monophosphate (OMP) from orotate or orotic acid; a recombinant gene encoding a polypeptide capable of synthesizing UMP from OMP; and/or a recombinant gene encoding a polypeptide capable of synthesizing uridine diphosphate (UDP) from UMP. In some embodiments, a recombinant host comprising one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP, one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate may overexpress a gene encoding a polypeptide capable of transporting uracil into the host cell; a gene encoding a polypeptide capable of synthesizing uridine monophosphate (UMP) from uracil; a gene encoding a polypeptide capable of synthesizing orotidine monophosphate (OMP) from orotate

or orotic acid; a gene encoding a polypeptide capable of synthesizing UMP from OMP; and/or a gene encoding a polypeptide capable of synthesizing uridine diphosphate (UDP) from UMP.

[0098] In some aspects, the polypeptide capable of synthesizing UTP from UDP comprises a polypeptide having the amino acid sequence set forth in SEQ ID NO:123 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:122).

[0099] In some aspects, the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate comprises a polypeptide having the amino acid sequence set forth in SEQ ID NO:2 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:1), SEQ ID NO:119 (encoded by the nucleotide sequence set forth in SEQ ID NO:118), SEQ ID NO:141 (encoded by the nucleotide sequence set forth in SEQ ID NO:140), SEQ ID NO:143 (encoded by the nucleotide sequence set forth in SEQ ID NO:142), SEQ ID NO:145 (encoded by the nucleotide sequence set forth in SEQ ID NO:144), or SEQ ID NO:147 (encoded by the nucleotide sequence set forth in SEQ ID NO:146).

[00100] In some aspects, the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate comprises a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:120), SEQ ID NO:125 (encoded by the nucleotide sequence set forth in SEQ ID NO:124), SEQ ID NO:127 (encoded by the nucleotide sequence set forth in SEQ ID NO:126), SEQ ID NO:129 (encoded by the nucleotide sequence set forth in SEQ ID NO:128), SEQ ID NO:131 (encoded by the nucleotide sequence set forth in SEQ ID NO:130), SEQ ID NO:133 (encoded by the nucleotide sequence set forth in SEQ ID NO:132), SEQ ID NO:135 (encoded by the nucleotide sequence set forth in SEQ ID NO:134), SEQ ID NO:137 (encoded by the nucleotide sequence set forth in SEQ ID NO:136), or SEQ ID NO:139 (encoded by the nucleotide sequence set forth in SEQ ID NO:138).

[00101] In some embodiments, a recombinant host comprises a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP and a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate. In some embodiments, a recombinant host comprises a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate. In some embodiments, a recombinant host comprises a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate and a recombinant gene

encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate. In some embodiments, a recombinant host comprises a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate.

[00102] In some embodiments, a recombinant host comprises two or more recombinant genes encoding a polypeptide involved in the UDP-glucose biosynthetic pathway, e.g., a gene encoding a polypeptide capable of converting glucose-6-phosphate having a first amino acid sequence and a gene encoding a polypeptide capable of converting glucose-6-phosphate having a second amino acid sequence distinct from the first amino acid sequence. For example, in some embodiments, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence of PGM1 (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2) and a gene encoding a polypeptide having the amino acid sequence of PGM2 (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, or SEQ ID NO:147). In certain such embodiments, the two or more genes encoding a polypeptide involved in the UDP-glucose biosynthetic pathway comprise nucleotide sequences native to the recombinant host cell (e.g., a recombinant *S. cerevisiae* host cell comprising a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:2 and a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:119). In other such embodiments, one of the two or more genes encoding a polypeptide involved in the UDP-glucose biosynthetic pathway comprises a nucleotide sequence native to the recombinant host cell, while one or more of the two or more genes encoding a polypeptide involved in the UDP-glucose biosynthetic pathway comprises a heterologous nucleotide sequence. For example, in some embodiments, a recombinant *S. cerevisiae* host cell expressing a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121 (i.e., a recombinant host overexpressing the polypeptide) further expresses a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in, e.g., SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139. In another example, in some embodiments, a recombinant *S. cerevisiae* host cell expressing a recombinant gene encoding a

polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:119 (*i.e.*, a recombinant host overexpressing the polypeptide) further expresses a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in, *e.g.*, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, or SEQ ID NO:147. Accordingly, as used herein, the term “a recombinant gene” may include “one or more recombinant genes.”

[00103] In some embodiments, a recombinant host comprises two or more copies of a recombinant gene encoding a polypeptide involved in the UDP-glucose biosynthetic pathway or the steviol glycoside biosynthetic pathway. In some embodiments, a recombinant host is preferably transformed with, *e.g.*, two copies, three copies, four copies, or five copies of a recombinant gene encoding a polypeptide involved in the UDP-glucose biosynthetic pathway or the steviol glycoside biosynthetic pathway. For example, in some embodiments, a recombinant host is transformed with two copies of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123). The person of ordinary skill in the art will appreciate that, in some embodiments, recombinant genes may be replicated in a host cell independently of cell replication; accordingly, a recombinant host cell may comprise, *e.g.*, more copies of a recombinant gene than the number of copies the cell was transformed with. Accordingly, as used herein, the term “a recombinant gene” may include “one or more copies of a recombinant gene.”

[00104] In some aspects, expression of a polypeptide capable of synthesizing UTP from UDP, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a recombinant host cell increases the amount of UDP-glucose produced by the cell. In some aspects, expression of a polypeptide capable of synthesizing UTP from UDP, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a recombinant host cell maintains, or even increases, the pool of UDP-glucose available for, *e.g.*, glycosylation of steviol or a steviol glycoside. In some aspects, expression of a polypeptide capable of synthesizing UTP from UDP, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a polypeptide capable synthesizing UDP-glucose from UTP and glucose-1-phosphate in a recombinant host cell increases the speed which which UDP-glucose

is regenerated, thus maintaining, or even increasing, the UDP-glucose pool, which can be used to synthesize one or more steviol glycosides.

[00105] In some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.* a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, or SEQ ID NO:147), and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139) in a recombinant host cell increases the amount of UDP-glucose produced by the cell by at least about 10%, *e.g.*, at least about 25%, or at least about 50%, or at least about 75%, or at least about 100%, or at least about 125%, or at least about 150%, or at least about 175%, or at least about 200%, or at least about 225%, or at least about 250%, or at least about 275%, or at least about 300%, calculated as an increase in intracellular UDP-glucose concentration relative to a corresponding host lacking the recombinant genes.

[00106] In certain such embodiments, one or more of the recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, the recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and the recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate comprise a nucleotide sequence native to the host cell. For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP having the amino acid sequence set forth in SEQ ID NO:123, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:2 and/or SEQ ID NO:119, and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121 in a steviol glycoside-producing *S. cerevisiae* host cell (*i.e.*, providing a recombinant host overexpressing the polypeptides) increases the amount of UDP-glucose produced by the cell by at least about 10%, *e.g.*, at least about 25%, or at least about 50%, or at least about 75%, or at least about 100%, or at least about 125%, or at least about 150%, or at least about 175%, or at least about 200%, or at least about 225%, or at least about 250%, or at least about

275%, or at least about 300%, calculated as an increase in intracellular UDP-glucose concentration relative to a corresponding host lacking the recombinant genes.

[00107] In some aspects, expression of a polypeptide capable of synthesizing UTP from UDP, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a steviol-glycoside producing recombinant host cell further expressing a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group; a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group; and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside, increases the amount of one or more steviol glycosides produced by the cell, and/or decreases the amount of one or more steviol glycosides produced by the cell. In some embodiments, the steviol glycoside-producing host further expresses a gene encoding a polypeptide capable of synthesizing GGPP from FPP and IPP; a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP; a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate; a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene; a gene encoding a polypeptide capable of reducing cytochrome P450 complex; and a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid; and/or a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate.

[00108] In some aspects, the polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP) comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:20 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:19), SEQ ID NO:22 (encoded by the nucleotide sequence set forth in SEQ ID NO:21), SEQ ID NO:24 (encoded by the nucleotide sequence set forth in SEQ ID NO:23), SEQ ID NO:26 (encoded by the nucleotide sequence set forth in SEQ ID NO:25), SEQ ID NO:28 (encoded by the nucleotide sequence set forth in SEQ ID NO:27), SEQ ID NO:30 (encoded by the nucleotide sequence set forth in SEQ ID NO:29), SEQ ID NO:32 (encoded by the nucleotide sequence set forth in SEQ ID NO:31), or SEQ ID NO:116 (encoded by the nucleotide sequence set forth in SEQ ID NO:115). In some

embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP) further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00109] In some aspects, the polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:34 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:33), SEQ ID NO:36 (encoded by the nucleotide sequence set forth in SEQ ID NO:35), SEQ ID NO:38 (encoded by the nucleotide sequence set forth in SEQ ID NO:37), SEQ ID NO:40 (encoded by the nucleotide sequence set forth in SEQ ID NO:39), or SEQ ID NO:42 (encoded by the nucleotide sequence set forth in SEQ ID NO:41). In some embodiments, the polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP lacks a chloroplast transit peptide. In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135,

SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00110] In some aspects, the polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:44 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:43), SEQ ID NO:46 (encoded by the nucleotide sequence set forth in SEQ ID NO:45), SEQ ID NO:48 (encoded by the nucleotide sequence set forth in SEQ ID NO:47), SEQ ID NO:50 (encoded by the nucleotide sequence set forth in SEQ ID NO:49), or SEQ ID NO:52 (encoded by the nucleotide sequence set forth in SEQ ID NO:51). In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00111] In some embodiments, a recombinant host comprises a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate. In some aspects, the bifunctional polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:54 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:53), SEQ ID NO:56 (encoded by the nucleotide sequence set forth in SEQ ID NO:55), or SEQ ID NO:58 (encoded by the nucleotide sequence set forth in SEQ ID NO:57). In some embodiments, a recombinant host comprising a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl

diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00112] In some aspects, the polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:60 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:59), SEQ ID NO:62 (encoded by the nucleotide sequence set forth in SEQ ID NO:61), SEQ ID NO:117 (encoded by the nucleotide sequence set forth in SEQ ID NO:63 or SEQ ID NO:64), SEQ ID NO:66 (encoded by the nucleotide sequence set forth in SEQ ID NO:65), SEQ ID NO:68 (encoded by the nucleotide sequence set forth in SEQ ID NO:67), SEQ ID NO:70 (encoded by the nucleotide sequence set forth in SEQ ID NO:69), SEQ ID NO:72 (encoded by the nucleotide sequence set forth in SEQ ID NO:71), SEQ ID NO:74 (encoded by the nucleotide sequence set forth in SEQ ID NO:73), or SEQ ID NO:76 (encoded by the nucleotide sequence set forth in SEQ ID NO:75). In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and

glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00113] In some aspects, the polypeptide capable of reducing cytochrome P450 complex comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:78 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:77), SEQ ID NO:80 (encoded by the nucleotide sequence set forth in SEQ ID NO:79), SEQ ID NO:82 (encoded by the nucleotide sequence set forth in SEQ ID NO:81), SEQ ID NO:84 (encoded by the nucleotide sequence set forth in SEQ ID NO:83), SEQ ID NO:86 (encoded by the nucleotide sequence set forth in SEQ ID NO:85), SEQ ID NO:88 (encoded by the nucleotide sequence set forth in SEQ ID NO:87), SEQ ID NO:90 (encoded by the nucleotide sequence set forth in SEQ ID NO:89), or SEQ ID NO:92 (encoded by the nucleotide sequence set forth in SEQ ID NO:91). In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of reducing cytochrome P450 complex further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00114] In some aspects, the polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:94 (which

can be encoded by the nucleotide sequence set forth in SEQ ID NO:93), SEQ ID NO:97 (encoded by the nucleotide sequence set forth in SEQ ID NO:95 or SEQ ID NO:96), SEQ ID NO:100 (encoded by the nucleotide sequence set forth in SEQ ID NO:98 or SEQ ID NO:99), SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106 (encoded by the nucleotide sequence set forth in SEQ ID NO:105), SEQ ID NO:108 (encoded by the nucleotide sequence set forth in SEQ ID NO:107), SEQ ID NO:110 (encoded by the nucleotide sequence set forth in SEQ ID NO:109), SEQ ID NO:112 (encoded by the nucleotide sequence set forth in SEQ ID NO:111), or SEQ ID NO:114 (encoded by the nucleotide sequence set forth in SEQ ID NO:113). In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00115] In some embodiments, a recombinant host comprises a nucleic acid encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group (*e.g.*, UGT85C2 polypeptide) (SEQ ID NO:7), a nucleic acid encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (*e.g.*, UGT76G1 polypeptide) (SEQ ID NO:9), a nucleic acid encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group (*e.g.*, UGT74G1 polypeptide) (SEQ ID NO:4), a nucleic acid encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (*e.g.*, EUGT11 polypeptide) (SEQ ID NO:16). In some aspects, the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-

O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., UGT91D2 polypeptide) can be a UGT91D2e polypeptide (SEQ ID NO:11) or a UGT91D2e-b polypeptide (SEQ ID NO:13). In some embodiments, a recombinant host comprising a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside further comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139). In some embodiments, the recombinant host is an *S. cerevisiae* host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00116] In some aspects, the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group is encoded by the nucleotide sequence set forth in SEQ ID NO:5 or SEQ ID NO:6, the polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside is encoded by the nucleotide sequence set forth in SEQ ID NO:8, the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group is encoded by the nucleotide sequence set forth in SEQ ID NO:3, the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside is encoded by the nucleotide sequence set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, or SEQ ID NO:15. The skilled worker will appreciate that expression of these genes may be necessary to produce a particular steviol glycoside but that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

[00117] In some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and a recombinant gene encoding a

polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a steviol glycoside-producing recombinant host increases the amount of one or more steviol glycosides, *e.g.*, rubusoside, RebB, RebA, RebD, and RebM, produced by the cell by at least about 5%, *e.g.*, at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about 30%, or at least about 35%, or at least about 40%, or at least about 45%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 100%, calculated as an increase in intracellular steviol glycoside concentration relative to a corresponding steviol glycoside-producing host lacking the recombinant genes.

[00118] For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.* a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, or SEQ ID NO:147), and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139) in a steviol glycoside-producing host increases the amount of one or more steviol glycosides, *e.g.*, rubusoside, RebB, RebA, RebD, and RebM, produced by the cell by at least about 5%, *e.g.*, at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about 30%, or at least about 35%, or at least about 40%, or at least about 45%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 100%, calculated as an increase in intracellular glycoside concentration relative to a corresponding steviol glycoside-producing host lacking the recombinant genes.

[00119] In some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a steviol glycoside-producing recombinant host decreases the amount of one or more steviol glycosides, *e.g.*, 13-SMG and RebD, produced by the cell by at least about 5%, *e.g.*, at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about

30%, or at least about 35%, or at least about 40%, or at least about 45%, or at least about 50%, calculated as a decrease in intracellular steviol glycoside concentration relative to a corresponding steviol glycoside-producing host lacking the recombinant genes.

[00120] For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP having the amino acid sequence set forth in SEQ ID NO:123, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:2, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:119, a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121, and further expression of a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in, e.g., SEQ ID NO:127, SEQ ID NO:133, SEQ ID NO:129, SEQ ID NO:125, SEQ ID NO:139, or SEQ ID NO:135, in a steviol glycoside-producing recombinant host decreases the amount of 13-SMG produced by the cell by at least about 5%, e.g., at least about 7.5%, or at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about 30%, or at least about 35%.

[00121] In some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate in a steviol glycoside-producing recombinant host increases the total amount of steviol glycosides (*i.e.*, the total amount of mono-, di-, tri-, tetra- penta-, hexa-, and hepta-glycosylated steviol compounds) by at least about 5%, e.g., at least about 7.5%, or at least about 10%, or at least about 12.5%, or at least about 15%, or at least about 17.5%, or at least about 20%, or at least about 25%, or at least about 27.5%, or at least about 30%, or at least about 35%, calculated as an increase in intracellular steviol glycoside concentration relative to a corresponding steviol glycoside-producing host lacking the recombinant genes.

[00122] For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP having the amino acid sequence set forth in SEQ ID NO:123, a recombinant gene encoding a polypeptide capable of converting glucose-6-

phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:2, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:119, a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121, and further expression of a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in, e.g., SEQ ID NO:133, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:125, SEQ ID NO:139, or SEQ ID NO:135, in a steviol glycoside-producing recombinant host increases the total amount of steviol glycosides (*i.e.*, the total amount of mono-, di-, tri-, tetra- penta-, hexa-, and hepta-glycosylated steviol compounds) by at least about 5%, e.g., at least about 7.5%, or at least about 10%, or at least about 12.5%, or at least about 15%, or at least about 17.5%, or at least about 20%, or at least about 25%, or at least about 27.5%, or at least about 30%, or at least about 35%, calculated as an increase in intracellular steviol glycoside concentration relative to a corresponding steviol glycoside-producing host lacking the recombinant genes.

[00123] In some other embodiments, the total amount of steviol glycosides produced by a steviol glycoside-producing recombinant host cell is unchanged (*i.e.*, increased or decreased by less than about 5%, or less than about 4%, or less than about 3%, or less than about 2%, or less than about 1%) by expression in the host of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate. For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP having the amino acid sequence set forth in SEQ ID NO:123, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:2, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:119, a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121 in a steviol glycoside-producing recombinant host increases the total amount of steviol glycosides produced by the host by less than about 5%, e.g., less than about 4%, or less than about 3%, or less than about 2%.

[00124] The person of ordinary skill in the art will appreciate that, in such embodiments, expression of one or more genes encoding a polypeptide involved in the involved in the UDP-glucose biosynthetic pathway may affect the relative levels of steviol glycosides produced by the recombinant host, *e.g.*, by increasing the level of UDP-glucose available as a substrate for a polypeptide capable of glycosylating steviol or a steviol glycoside. For example, in some embodiments, expression of a recombinant gene encoding a polypeptide capable of synthesizing UTP from UDP having the amino acid sequence set forth in SEQ ID NO:123, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:2, a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:119, a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having the amino acid sequence set forth in SEQ ID NO:121 in a steviol glycoside-producing recombinant host increases the total amount of steviol glycosides produced by the host by less than about 5%, *e.g.*, less than about 4%, or less than about 3%, or less than about 2%, increases the amount of RebM produced by the host by at least about 50%, *e.g.*, at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, and decreases the amount of RebD produced by the host by at least about 10%, *e.g.*, at least about 20%, or at least about 30%, or at least about 40%.

[00125] In some embodiments, a recombinant host cell comprises one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139).

[00126] In certain embodiments, a recombinant host comprises one or more recombinant genes having a nucleotide sequence native to the host that encode one or more polypeptides capable of synthesizing UTP from UDP, one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or one or more polypeptides capable of

synthesizing UDP-glucose from UTP and glucose-1-phosphate, *i.e.*, a recombinant host overexpresses one or more polypeptides capable of synthesizing UTP from UDP, one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate.

[00127] In certain such embodiments, a recombinant host cell overexpresses one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, an *S. cerevisiae* host cell expressing a recombinant gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, an *S. cerevisiae* host cell expressing a recombinant gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, and/or SEQ ID NO:119), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, an *S. cerevisiae* host cell expressing a recombinant gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121). In one example, a recombinant *S. cerevisiae* host cell overexpresses a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:123, a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:119, and a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121.

[00128] In certain embodiments, a recombinant host cell comprising one or more genes encoding one or more polypeptides capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139), further comprises a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:7); a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (*e.g.*, a polypeptide

having the amino acid sequence set forth in SEQ ID NO:9); a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:4); and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:11, SEQ ID NO:13, or SEQ ID NO:16). In certain such embodiments, the recombinant host cell further comprises a gene encoding a polypeptide capable of synthesizing GGPP from FPP and IPP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:20); a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:40); a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:52); a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:60 or SEQ ID NO:117); a gene encoding a polypeptide capable of reducing cytochrome P450 complex (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:78, SEQ ID NO:86, or SEQ ID NO:92); and/or a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:94).

[00129] In some embodiments, a recombinant host comprises two or more genes encoding two or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or two or more genes encoding two or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139).

[00130] In certain such embodiments, a recombinant host comprises two or more genes encoding two or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate, e.g., two or more genes encoding two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147. In one example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:2 and a

polypeptide having the amino acid sequence set forth in SEQ ID NO:119. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:2, a polypeptide having the amino acid sequence set forth in SEQ ID NO:119, and a polypeptide having the amino acid sequence set forth in SEQ ID NO:145. In some embodiments, the recombinant host further comprises a gene encoding a polypeptide capable of synthesizing UTP from UDP (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:123) and/or one or more genes encoding one or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (*e.g.*, a polypeptide having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139).

[00131] In certain such embodiments, a recombinant host comprises two or more genes encoding two or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, *e.g.*, two or more genes encoding two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139. In one example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a polypeptide having the amino acid sequence set forth in SEQ ID NO:125. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a polypeptide having the amino acid sequence set forth in SEQ ID NO:127. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a polypeptide having the amino acid sequence set forth in SEQ ID NO:129. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a polypeptide having the amino acid sequence set forth in SEQ ID NO:131. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:133. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:135. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a gene encoding a polypeptide having the

amino acid sequence set forth in SEQ ID NO:137. In another example, a recombinant host comprises a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:121 and a gene encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO:139. In some embodiments, the recombinant host further comprises a gene encoding a polypeptide capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123) and/or one or more genes encoding one or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., one or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147).

[00132] In certain such embodiments, a recombinant host comprising two or more genes encoding two or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or two or more genes encoding two or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139) is a host cell overexpressing one or more genes encoding one or more polypeptides involved in the UDP-glucose biosynthetic pathway (e.g., an *S. cerevisiae* host cell expressing one or more genes encoding one or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:121, and/or SEQ ID NO:123).

[00133] In certain embodiments, a recombinant host cell comprising two or more genes encoding two or more polypeptides capable of converting glucose-6-phosphate to glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:141, SEQ ID NO:143, SEQ ID NO:145, and/or SEQ ID NO:147), and/or two or more genes encoding two or more polypeptides capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate (e.g., two or more polypeptides having the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:125, SEQ ID NO:127, SEQ ID NO:129, SEQ ID NO:131, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, and/or SEQ ID NO:139), further comprises a gene encoding polypeptide capable of synthesizing UTP from UDP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:123), a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:7);

a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:9); a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:4); and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:11, SEQ ID NO:13, or SEQ ID NO:16). In certain such embodiments, the recombinant host cell further comprises a gene encoding a polypeptide capable of synthesizing GGPP from FPP and IPP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:20); a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:40); a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:52); a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:60 or SEQ ID NO:117); a gene encoding a polypeptide capable of reducing cytochrome P450 complex (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:78, SEQ ID NO:86, or SEQ ID NO:92); and/or a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid (e.g., a polypeptide having the amino acid sequence set forth in SEQ ID NO:94).

[00134] In some embodiments, a steviol glycoside or steviol glycoside precursor is produced by whole cell bioconversion. For whole cell bioconversion to occur, a host cell expressing one or more enzymes involved in the steviol glycoside pathway takes up and modifies a steviol glycoside precursor in the cell; following modification *in vivo*, a steviol glycoside remains in the cell and/or is excreted into the culture medium. For example, a host cell expressing a gene encoding a polypeptide capable of synthesizing UTP from UDP, a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate; and further expressing a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group; a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; a gene encoding a polypeptide capable of glycosylating steviol or

a steviol glycoside at its C-19 carboxyl group; and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside can take up steviol and glycosylate steviol in the cell; following glycosylation *in vivo*, a steviol glycoside can be excreted into the culture medium. In certain such embodiments, the host cell may further express a gene encoding a polypeptide capable of synthesizing GGPP from FPP and IPP; a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP; a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate; a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene; a gene encoding a polypeptide capable of reducing cytochrome P450 complex; a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid; and/or a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate.

[00135] In some embodiments, the method for producing one or more steviol glycosides or a steviol glycoside composition disclosed herein comprises whole-cell bioconversion of plant-derived or synthetic steviol and/or steviol glycosides in a cell culture medium of a recombinant host cell using: (a) a polypeptide capable of synthesizing UTP from UDP; (b) a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate; and/or (c) a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate, and one or more of: (d) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof; (e) a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; (f) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof; and/or (g) a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; wherein at least one of the polypeptides is a recombinant polypeptide expressed in the recombinant host cell; and producing the one or more steviol glycosides or the steviol glycoside composition thereby.

[00136] In some embodiments of the methods for producing one or more steviol glycosides or a steviol glycoside composition disclosed herein comprises whole-cell bioconversion of plant-derived or synthetic steviol and/or steviol glycosides in a cell culture medium of a recombinant host cell disclosed herein, the polypeptide capable of synthesizing UTP from UDP comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ

ID NO:123; the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, or SEQ ID NO:143; or at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:141, SEQ ID NO:145, or SEQ ID NO:147; and/or the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:127; at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139; or at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131.

[00137] In some embodiments, a polypeptide capable of synthesizing UTP from UDP, a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate can be displayed on the surface of the recombinant host cells disclosed herein by fusing it with the anchoring motifs.

[00138] In some embodiments, the cell is permeabilized to take up a substrate to be modified or to excrete a modified product. In some embodiments, a permeabilizing agent can be added to aid the feedstock entering into the host and product getting out. In some embodiments, the cells are permeabilized with a solvent such as toluene, or with a detergent such as Triton-X or Tween. In some embodiments, the cells are permeabilized with a surfactant, for example a cationic surfactant such as cetyltrimethylammonium bromide (CTAB). In some embodiments, the cells are permeabilized with periodic mechanical shock such as electroporation or a slight osmotic shock. For example, a crude lysate of the cultured microorganism can be centrifuged to obtain a supernatant. The resulting supernatant can then be applied to a chromatography column, *e.g.*, a C18 column, and washed with water to remove hydrophilic compounds, followed by elution of the compound(s) of interest with a solvent such as methanol. The compound(s) can then be further purified by preparative HPLC. See *also*, WO 2009/140394.

[00139] In some embodiments, steviol, one or more steviol glycoside precursors, and/or one or more steviol glycosides are produced by co-culturing of two or more hosts. In some embodiments, one or more hosts, each expressing one or more enzymes involved in the steviol glycoside pathway, produce steviol, one or more steviol glycoside precursors, and/or one or more steviol glycosides. For example, a host expressing a gene encoding a polypeptide

capable of synthesizing GGPP from FPP and IPP; a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP; a gene encoding a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate; a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid, *ent*-kaurenol, and/or *ent*-kaurenal from *ent*-kaurene; a gene encoding a polypeptide capable of reducing cytochrome P450 complex; a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid; and/or a gene encoding a bifunctional polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP and synthesizing *ent*-kaurene from *ent*-copalyl diphosphate and a host expressing a gene encoding a polypeptide capable of synthesizing UTP from UDP, a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate; and further expressing a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group; a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside; a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group; and/or a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside, produce one or more steviol glycosides.

[00140] In some embodiments, the steviol glycoside comprises, for example, but not limited to, 13-SMG, steviol-1,2-bioside, steviol-1,3-bioside, 19-SMG, 1,2-stevioside, 1,3-stevioside (RebG), rubusoside, RebA, RebB, RebC, RebD, RebE, RebF, RebM, RebQ, Rebl, dulcoside A, di-glycosylated steviol, tri-glycosylated steviol, tetra-glycosylated steviol, penta-glycosylated steviol, hexa-glycosylated steviol, hepta-glycosylated steviol, or isomers thereof.

[00141] In some embodiments, a steviol glycoside or steviol glycoside precursor composition produced *in vivo*, *in vitro*, or by whole cell bioconversion does not comprise or comprises a reduced amount or reduced level of plant-derived components than a *Stevia* extract from, *inter alia*, a *Stevia* plant. Plant-derived components can contribute to off-flavors and include pigments, lipids, proteins, phenolics, saccharides, spathulenol and other sesquiterpenes, labdane diterpenes, monoterpenes, decanoic acid, 8,11,14-eicosatrienoic acid, 2-methyloctadecane, pentacosane, octacosane, tetracosane, octadecanol, stigmasterol, β -sitosterol, α - and β -amyrin, lupeol, β -amryin acetate, pentacyclic triterpenes, centauredin, quercitin, epi- α -cadinol, carophyllenes and derivatives, beta-pinene, beta-sitosterol, and

gibberellin. In some embodiments, the plant-derived components referred to herein are non-glycoside compounds.

[00142] As used herein, the terms “detectable amount,” “detectable concentration,” “measurable amount,” and “measurable concentration” refer to a level of steviol glycosides measured in AUC, $\mu\text{M}/\text{OD}_{600}$, mg/L, μM , or mM. Steviol glycoside production (*i.e.*, total, supernatant, and/or intracellular steviol glycoside levels) can be detected and/or analyzed by techniques generally available to one skilled in the art, for example, but not limited to, liquid chromatography-mass spectrometry (LC-MS), thin layer chromatography (TLC), high-performance liquid chromatography (HPLC), ultraviolet-visible spectroscopy/ spectrophotometry (UV-Vis), mass spectrometry (MS), and nuclear magnetic resonance spectroscopy (NMR).

[00143] As used herein, the term “undetectable concentration” refers to a level of a compound that is too low to be measured and/or analyzed by techniques such as TLC, HPLC, UV-Vis, MS, or NMR. In some embodiments, a compound of an “undetectable concentration” is not present in a steviol glycoside or steviol glycoside precursor composition.

[00144] After the recombinant microorganism has been grown in culture for the period of time, wherein the temperature and period of time facilitate the production of a steviol glycoside, steviol and/or one or more steviol glycosides can then be recovered from the culture using various techniques known in the art. Steviol glycosides can be isolated using a method described herein. For example, following fermentation, a culture broth can be centrifuged for 30 min at 7000 rpm at 4°C to remove cells, or cells can be removed by filtration. The cell-free lysate can be obtained, for example, by mechanical disruption or enzymatic disruption of the host cells and additional centrifugation to remove cell debris. Mechanical disruption of the dried broth materials can also be performed, such as by sonication. The dissolved or suspended broth materials can be filtered using a micron or sub-micron prior to further purification, such as by preparative chromatography. The fermentation media or cell-free lysate can optionally be treated to remove low molecular weight compounds such as salt; and can optionally be dried prior to purification and re-dissolved in a mixture of water and solvent.

[00145] The supernatant or cell-free lysate can be purified as follows: a column can be filled with, for example, HP20 Diaion resin (aromatic type Synthetic Adsorbent; Supelco) or other suitable non-polar adsorbent or reversed-phase chromatography resin, and an aliquot of supernatant or cell-free lysate can be loaded on to the column and washed with water to remove the hydrophilic components. The steviol glycoside product can be eluted by stepwise

incremental increases in the solvent concentration in water or a gradient from, e. g., 0% → 100% methanol). The levels of steviol glycosides, glycosylated *ent*-kaurenol, and/or glycosylated *ent*-kaurenoic acid in each fraction, including the flow-through, can then be analyzed by LC-MS. Fractions can then be combined and reduced in volume using a vacuum evaporator. Additional purification steps can be utilized, if desired, such as additional chromatography steps and crystallization. For example, steviol glycosides can be isolated by methods not limited to ion exchange chromatography, reversed-phase chromatography (*i.e.*, using a C18 column), extraction, crystallization, and carbon columns and/or decoloring steps.

[00146] As used herein, the terms “or” and “and/or” is utilized to describe multiple components in combination or exclusive of one another. For example, “x, y, and/or z” can refer to “x” alone, “y” alone, “z” alone, “x, y, and z,” “(x and y) or z,” “x or (y and z),” or “x or y or z.” In some embodiments, “and/or” is used to refer to the exogenous nucleic acids that a recombinant cell comprises, wherein a recombinant cell comprises one or more exogenous nucleic acids selected from a group. In some embodiments, “and/or” is used to refer to production of steviol glycosides and/or steviol glycoside precursors. In some embodiments, “and/or” is used to refer to production of steviol glycosides, wherein one or more steviol glycosides are produced. In some embodiments, “and/or” is used to refer to production of steviol glycosides, wherein one or more steviol glycosides are produced through one or more of the following steps: culturing a recombinant microorganism, synthesizing one or more steviol glycosides in a recombinant microorganism, and/or isolating one or more steviol glycosides.

Functional Homologs

[00147] Functional homologs of the polypeptides described above are also suitable for use in producing steviol glycosides in a recombinant host. A functional homolog is a polypeptide that has sequence similarity to a reference polypeptide, and that carries out one or more of the biochemical or physiological function(s) of the reference polypeptide. A functional homolog and the reference polypeptide can be a natural occurring polypeptide, and the sequence similarity can be due to convergent or divergent evolutionary events. As such, functional homologs are sometimes designated in the literature as homologs, or orthologs, or paralogs. Variants of a naturally occurring functional homolog, such as polypeptides encoded by mutants of a wild type coding sequence, can themselves be functional homologs. Functional homologs can also be

created via site-directed mutagenesis of the coding sequence for a polypeptide, or by combining domains from the coding sequences for different naturally-occurring polypeptides (“domain swapping”). Techniques for modifying genes encoding functional polypeptides described herein are known and include, *inter alia*, directed evolution techniques, site-directed mutagenesis techniques and random mutagenesis techniques, and can be useful to increase specific activity of a polypeptide, alter substrate specificity, alter expression levels, alter subcellular location, or modify polypeptide-polypeptide interactions in a desired manner. Such modified polypeptides are considered functional homologs. The term “functional homolog” is sometimes applied to the nucleic acid that encodes a functionally homologous polypeptide.

[00148] Functional homologs can be identified by analysis of nucleotide and polypeptide sequence alignments. For example, performing a query on a database of nucleotide or polypeptide sequences can identify homologs of steviol glycoside biosynthesis polypeptides. Sequence analysis can involve BLAST, Reciprocal BLAST, or PSI-BLAST analysis of non-redundant databases using a UGT amino acid sequence as the reference sequence. Amino acid sequence is, in some instances, deduced from the nucleotide sequence. Those polypeptides in the database that have greater than 40% sequence identity are candidates for further evaluation for suitability as a steviol glycoside biosynthesis polypeptide. Amino acid sequence similarity allows for conservative amino acid substitutions, such as substitution of one hydrophobic residue for another or substitution of one polar residue for another. If desired, manual inspection of such candidates can be carried out in order to narrow the number of candidates to be further evaluated. Manual inspection can be performed by selecting those candidates that appear to have domains present in steviol glycoside biosynthesis polypeptides, *e.g.*, conserved functional domains. In some embodiments, nucleic acids and polypeptides are identified from transcriptome data based on expression levels rather than by using BLAST analysis.

[00149] Conserved regions can be identified by locating a region within the primary amino acid sequence of a steviol glycoside biosynthesis polypeptide that is a repeated sequence, forms some secondary structure (*e.g.*, helices and beta sheets), establishes positively or negatively charged domains, or represents a protein motif or domain. See, *e.g.*, the Pfam web site describing consensus sequences for a variety of protein motifs and domains on the World Wide Web at sanger.ac.uk/Software/Pfam/ and pfam.janelia.org/. The information included at the Pfam database is described in Sonnhammer *et al.*, *Nucl. Acids Res.*, 26:320-322 (1998); Sonnhammer *et al.*, *Proteins*, 28:405-420 (1997); and Bateman *et al.*, *Nucl. Acids Res.*, 27:260-

262 (1999). Conserved regions also can be determined by aligning sequences of the same or related polypeptides from closely related species. Closely related species preferably are from the same family. In some embodiments, alignment of sequences from two different species is adequate to identify such homologs.

[00150] Typically, polypeptides that exhibit at least about 40% amino acid sequence identity are useful to identify conserved regions. Conserved regions of related polypeptides exhibit at least 45% amino acid sequence identity (e.g., at least 50%, at least 60%, at least 70%, at least 80%, or at least 90% amino acid sequence identity). In some embodiments, a conserved region exhibits at least 92%, 94%, 96%, 98%, or 99% amino acid sequence identity.

[00151] For example, polypeptides suitable for producing steviol in a recombinant host include functional homologs of UGTs.

[00152] Methods to modify the substrate specificity of, for example, a UGT, are known to those skilled in the art, and include without limitation site-directed/rational mutagenesis approaches, random directed evolution approaches and combinations in which random mutagenesis/saturation techniques are performed near the active site of the enzyme. For example see Osmani *et al.*, 2009, *Phytochemistry* 70: 325–347.

[00153] A candidate sequence typically has a length that is from 80% to 200% of the length of the reference sequence, e.g., 82, 85, 87, 89, 90, 93, 95, 97, 99, 100, 105, 110, 115, 120, 130, 140, 150, 160, 170, 180, 190, or 200% of the length of the reference sequence. A functional homolog polypeptide typically has a length that is from 95% to 105% of the length of the reference sequence, e.g., 90, 93, 95, 97, 99, 100, 105, 110, 115, or 120% of the length of the reference sequence, or any range between. A % identity for any candidate nucleic acid or polypeptide relative to a reference nucleic acid or polypeptide can be determined as follows. A reference sequence (e.g., a nucleic acid sequence or an amino acid sequence described herein) is aligned to one or more candidate sequences using the computer program Clustal Omega (version 1.2.1, default parameters), which allows alignments of nucleic acid or polypeptide sequences to be carried out across their entire length (global alignment). Chenna *et al.*, 2003, *Nucleic Acids Res.* 31(13):3497-500.

[00154] ClustalW calculates the best match between a reference and one or more candidate sequences, and aligns them so that identities, similarities and differences can be determined. Gaps of one or more residues can be inserted into a reference sequence, a candidate sequence, or both, to maximize sequence alignments. For fast pairwise alignment of nucleic

acid sequences, the following default parameters are used: word size: 2; window size: 4; scoring method: % age; number of top diagonals: 4; and gap penalty: 5. For multiple alignment of nucleic acid sequences, the following parameters are used: gap opening penalty: 10.0; gap extension penalty: 5.0; and weight transitions: yes. For fast pairwise alignment of protein sequences, the following parameters are used: word size: 1; window size: 5; scoring method: % age; number of top diagonals: 5; gap penalty: 3. For multiple alignment of protein sequences, the following parameters are used: weight matrix: blosum; gap opening penalty: 10.0; gap extension penalty: 0.05; hydrophilic gaps: on; hydrophilic residues: Gly, Pro, Ser, Asn, Asp, Gln, Glu, Arg, and Lys; residue-specific gap penalties: on. The ClustalW output is a sequence alignment that reflects the relationship between sequences. ClustalW can be run, for example, at the Baylor College of Medicine Search Launcher site on the World Wide Web (searchlauncher.bcm.tmc.edu/multi-align/multi-align.html) and at the European Bioinformatics Institute site on the World Wide Web (ebi.ac.uk/clustalw).

[00155] To determine a % identity of a candidate nucleic acid or amino acid sequence to a reference sequence, the sequences are aligned using Clustal Omega, the number of identical matches in the alignment is divided by the length of the reference sequence, and the result is multiplied by 100. It is noted that the % identity value can be rounded to the nearest tenth. For example, 78.11, 78.12, 78.13, and 78.14 are rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 are rounded up to 78.2.

[00156] It will be appreciated that functional UGT proteins (e.g., a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group) can include additional amino acids that are not involved in the enzymatic activities carried out by the enzymes. In some embodiments, UGT proteins are fusion proteins. The terms "chimera," "fusion polypeptide," "fusion protein," "fusion enzyme," "fusion construct," "chimeric protein," "chimeric polypeptide," "chimeric construct," and "chimeric enzyme" can be used interchangeably herein to refer to proteins engineered through the joining of two or more genes that code for different proteins. In some embodiments, a nucleic acid sequence encoding a UGT polypeptide (e.g., a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group) can include a tag sequence that encodes a "tag" designed to facilitate subsequent manipulation (e.g., to facilitate purification or detection), secretion, or localization of the encoded polypeptide. Tag sequences can be inserted in the nucleic acid sequence encoding the polypeptide such that the encoded tag is located at either the carboxyl or amino terminus of the polypeptide. Non-limiting examples of encoded tags include green fluorescent protein (GFP), human influenza

hemagglutinin (HA), glutathione S transferase (GST), polyhistidine-tag (HIS tag), and Flag™ tag (Kodak, New Haven, CT). Other examples of tags include a chloroplast transit peptide, a mitochondrial transit peptide, an amyloplast peptide, signal peptide, or a secretion tag.

[00157] In some embodiments, a fusion protein is a protein altered by domain swapping. As used herein, the term “domain swapping” is used to describe the process of replacing a domain of a first protein with a domain of a second protein. In some embodiments, the domain of the first protein and the domain of the second protein are functionally identical or functionally similar. In some embodiments, the structure and/or sequence of the domain of the second protein differs from the structure and/or sequence of the domain of the first protein. In some embodiments, a UGT polypeptide (*e.g.*, a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group) is altered by domain swapping.

[00158] In some embodiments, a fusion protein is a protein altered by circular permutation, which consists in the covalent attachment of the ends of a protein that would be opened elsewhere afterwards. Thus, the order of the sequence is altered without causing changes in the amino acids of the protein. In some embodiments, a targeted circular permutation can be produced, for example but not limited to, by designing a spacer to join the ends of the original protein. Once the spacer has been defined, there are several possibilities to generate permutations through generally accepted molecular biology techniques, for example but not limited to, by producing concatemers by means of PCR and subsequent amplification of specific permutations inside the concatemer or by amplifying discrete fragments of the protein to exchange to join them in a different order. The step of generating permutations can be followed by creating a circular gene by binding the fragment ends and cutting back at random, thus forming collections of permutations from a unique construct. In some embodiments, DAP1 polypeptide is altered by circular permutation.

Steviol and Steviol Glycoside Biosynthesis Nucleic Acids

[00159] A recombinant gene encoding a polypeptide described herein comprises the coding sequence for that polypeptide, operably linked in sense orientation to one or more regulatory regions suitable for expressing the polypeptide. Because many microorganisms are capable of expressing multiple gene products from a polycistronic mRNA, multiple polypeptides can be expressed under the control of a single regulatory region for those microorganisms, if desired.

A coding sequence and a regulatory region are considered to be operably linked when the regulatory region and coding sequence are positioned so that the regulatory region is effective for regulating transcription or translation of the sequence. Typically, the translation initiation site of the translational reading frame of the coding sequence is positioned between one and about fifty nucleotides downstream of the regulatory region for a monocistronic gene.

[00160] In many cases, the coding sequence for a polypeptide described herein is identified in a species other than the recombinant host, *i.e.*, is a heterologous nucleic acid. Thus, if the recombinant host is a microorganism, the coding sequence can be from other prokaryotic or eukaryotic microorganisms, from plants or from animals. In some case, however, the coding sequence is a sequence that is native to the host and is being reintroduced into that organism. A native sequence can often be distinguished from the naturally occurring sequence by the presence of non-natural sequences linked to the exogenous nucleic acid, *e.g.*, non-native regulatory sequences flanking a native sequence in a recombinant nucleic acid construct. In addition, stably transformed exogenous nucleic acids typically are integrated at positions other than the position where the native sequence is found. "Regulatory region" refers to a nucleic acid having nucleotide sequences that influence transcription or translation initiation and rate, and stability and/or mobility of a transcription or translation product. Regulatory regions include, without limitation, promoter sequences, enhancer sequences, response elements, protein recognition sites, inducible elements, protein binding sequences, 5' and 3' untranslated regions (UTRs), transcriptional start sites, termination sequences, polyadenylation sequences, introns, and combinations thereof. A regulatory region typically comprises at least a core (basal) promoter. A regulatory region also may include at least one control element, such as an enhancer sequence, an upstream element or an upstream activation region (UAR). A regulatory region is operably linked to a coding sequence by positioning the regulatory region and the coding sequence so that the regulatory region is effective for regulating transcription or translation of the sequence. For example, to operably link a coding sequence and a promoter sequence, the translation initiation site of the translational reading frame of the coding sequence is typically positioned between one and about fifty nucleotides downstream of the promoter. A regulatory region can, however, be positioned as much as about 5,000 nucleotides upstream of the translation initiation site, or about 2,000 nucleotides upstream of the transcription start site.

[00161] The choice of regulatory regions to be included depends upon several factors, including, but not limited to, efficiency, selectability, inducibility, desired expression level, and preferential expression during certain culture stages. It is a routine matter for one of skill in the

art to modulate the expression of a coding sequence by appropriately selecting and positioning regulatory regions relative to the coding sequence. It will be understood that more than one regulatory region may be present, *e.g.*, introns, enhancers, upstream activation regions, transcription terminators, and inducible elements.

[00162] One or more genes can be combined in a recombinant nucleic acid construct in “modules” useful for a discrete aspect of steviol and/or steviol glycoside production. Combining a plurality of genes in a module, particularly a polycistronic module, facilitates the use of the module in a variety of species. For example, a steviol biosynthesis gene cluster, or a UGT gene cluster, can be combined in a polycistronic module such that, after insertion of a suitable regulatory region, the module can be introduced into a wide variety of species. As another example, a UGT gene cluster can be combined such that each UGT coding sequence is operably linked to a separate regulatory region, to form a UGT module. Such a module can be used in those species for which monocistronic expression is necessary or desirable. In addition to genes useful for steviol or steviol glycoside production, a recombinant construct typically also contains an origin of replication, and one or more selectable markers for maintenance of the construct in appropriate species.

[00163] It will be appreciated that because of the degeneracy of the genetic code, a number of nucleic acids can encode a particular polypeptide; *i.e.*, for many amino acids, there is more than one nucleotide triplet that serves as the codon for the amino acid. Thus, codons in the coding sequence for a given polypeptide can be modified such that optimal expression in a particular host is obtained, using appropriate codon bias tables for that host (*e.g.*, microorganism). As isolated nucleic acids, these modified sequences can exist as purified molecules and can be incorporated into a vector or a virus for use in constructing modules for recombinant nucleic acid constructs.

[00164] In some cases, it is desirable to inhibit one or more functions of an endogenous polypeptide in order to divert metabolic intermediates towards steviol or steviol glycoside biosynthesis. For example, it may be desirable to downregulate synthesis of sterols in a yeast strain in order to further increase steviol or steviol glycoside production, *e.g.*, by downregulating squalene epoxidase. As another example, it may be desirable to inhibit degradative functions of certain endogenous gene products, *e.g.*, glycohydrolases that remove glucose moieties from secondary metabolites or phosphatases as discussed herein. In such cases, a nucleic acid that overexpresses the polypeptide or gene product may be included in a recombinant construct that

is transformed into the strain. Alternatively, mutagenesis can be used to generate mutants in genes for which it is desired to increase or enhance function.

Host Microorganisms

[00165] Recombinant hosts can be used to express polypeptides for the producing steviol glycosides, including mammalian, insect, plant, and algal cells. A number of prokaryotes and eukaryotes are also suitable for use in constructing the recombinant microorganisms described herein, e.g., gram-negative bacteria, yeast, and fungi. A species and strain selected for use as a steviol glycoside production strain is first analyzed to determine which production genes are endogenous to the strain and which genes are not present. Genes for which an endogenous counterpart is not present in the strain are advantageously assembled in one or more recombinant constructs, which are then transformed into the strain in order to supply the missing function(s).

[00166] Typically, the recombinant microorganism is grown in a fermenter at a temperature(s) for a period of time, wherein the temperature and period of time facilitate the production of a steviol glycoside. The constructed and genetically engineered microorganisms provided by the invention can be cultivated using conventional fermentation processes, including, *inter alia*, chemostat, batch, fed-batch cultivations, semi-continuous fermentations such as draw and fill, continuous perfusion fermentation, and continuous perfusion cell culture. Depending on the particular microorganism used in the method, other recombinant genes such as isopentenyl biosynthesis genes and terpene synthase and cyclase genes may also be present and expressed. Levels of substrates and intermediates, e.g., isopentenyl diphosphate, dimethylallyl diphosphate, GGPP, *ent*-kaurene and *ent*-kaurenoic acid, can be determined by extracting samples from culture media for analysis according to published methods.

[00167] Carbon sources of use in the instant method include any molecule that can be metabolized by the recombinant host cell to facilitate growth and/or production of the steviol glycosides. Examples of suitable carbon sources include, but are not limited to, sucrose (e.g., as found in molasses), fructose, xylose, ethanol, glycerol, glucose, cellulose, starch, cellobiose or other glucose-comprising polymer. In embodiments employing yeast as a host, for example, carbon sources such as sucrose, fructose, xylose, ethanol, glycerol, and glucose are suitable. The carbon source can be provided to the host organism throughout the cultivation period or

alternatively, the organism can be grown for a period of time in the presence of another energy source, e.g., protein, and then provided with a source of carbon only during the fed-batch phase.

[00168] It will be appreciated that the various genes and modules discussed herein can be present in two or more recombinant hosts rather than a single host. When a plurality of recombinant hosts is used, they can be grown in a mixed culture to accumulate steviol and/or steviol glycosides.

[00169] Alternatively, the two or more hosts each can be grown in a separate culture medium and the product of the first culture medium, e.g., steviol, can be introduced into second culture medium to be converted into a subsequent intermediate, or into an end product such as, for example, RebA. The product produced by the second, or final host is then recovered. It will also be appreciated that in some embodiments, a recombinant host is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

[00170] Exemplary prokaryotic and eukaryotic species are described in more detail below. However, it will be appreciated that other species can be suitable. For example, suitable species can be in a genus such as *Agaricus*, *Aspergillus*, *Bacillus*, *Candida*, *Corynebacterium*, *Eremothecium*, *Escherichia*, *Fusarium/Gibberella*, *Kluyveromyces*, *Laetiporus*, *Lentinus*, *Phaffia*, *Phanerochaete*, *Pichia*, *Physcomitrella*, *Rhodoturula*, *Saccharomyces*, *Schizosaccharomyces*, *Sphaceloma*, *Xanthophyllomyces* or *Yarrowia*. Exemplary species from such genera include *Lentinus tigrinus*, *Laetiporus sulphureus*, *Phanerochaete chrysosporium*, *Pichia pastoris*, *Cyberlindnera jadinii*, *Physcomitrella patens*, *Rhodoturula glutinis*, *Rhodoturula mucilaginoso*, *Phaffia rhodozyma*, *Xanthophyllomyces dendrorhous*, *Fusarium fujikuroi/Gibberella fujikuroi*, *Candida utilis*, *Candida glabrata*, *Candida albicans*, and *Yarrowia lipolytica*.

[00171] In some embodiments, a microorganism can be a prokaryote such as *Escherichia* bacteria cells, for example, *Escherichia coli* cells; *Lactobacillus* bacteria cells; *Lactococcus* bacteria cells; *Cornibacterium* bacteria cells; *Acetobacter* bacteria cells; *Acinetobacter* bacteria cells; or *Pseudomonas* bacterial cells.

[00172] In some embodiments, a microorganism can be an Ascomycete such as *Gibberella fujikuroi*, *Kluyveromyces lactis*, *Schizosaccharomyces pombe*, *Aspergillus niger*, *Yarrowia lipolytica*, *Ashbya gossypii*, or *S. cerevisiae*.

[00173] In some embodiments, a microorganism can be an algal cell such as *Blakeslea trispora*, *Dunaliella salina*, *Haematococcus pluvialis*, *Chlorella sp.*, *Undaria pinnatifida*, *Sargassum*, *Laminaria japonica*, *Scenedesmus almeriensis* species.

[00174] In some embodiments, a microorganism can be a cyanobacterial cell such as *Blakeslea trispora*, *Dunaliella salina*, *Haematococcus pluvialis*, *Chlorella sp.*, *Undaria pinnatifida*, *Sargassum*, *Laminaria japonica*, *Scenedesmus almeriensis*.

Saccharomyces spp.

[00175] *Saccharomyces* is a widely used chassis organism in synthetic biology, and can be used as the recombinant microorganism platform. For example, there are libraries of mutants, plasmids, detailed computer models of metabolism and other information available for *S. cerevisiae*, allowing for rational design of various modules to enhance product yield. Methods are known for making recombinant microorganisms.

Aspergillus spp.

[00176] *Aspergillus* species such as *A. oryzae*, *A. niger* and *A. sojae* are widely used microorganisms in food production and can also be used as the recombinant microorganism platform. Nucleotide sequences are available for genomes of *A. nidulans*, *A. fumigatus*, *A. oryzae*, *A. clavatus*, *A. flavus*, *A. niger*, and *A. terreus*, allowing rational design and modification of endogenous pathways to enhance flux and increase product yield. Metabolic models have been developed for *Aspergillus*, as well as transcriptomic studies and proteomics studies. *A. niger* is cultured for the industrial production of a number of food ingredients such as citric acid and gluconic acid, and thus species such as *A. niger* are generally suitable for producing steviol glycosides.

E. coli

[00177] *E. coli*, another widely used platform organism in synthetic biology, can also be used as the recombinant microorganism platform. Similar to *Saccharomyces*, there are libraries of mutants, plasmids, detailed computer models of metabolism and other information available for *E. coli*, allowing for rational design of various modules to enhance product yield. Methods similar to those described above for *Saccharomyces* can be used to make recombinant *E. coli* microorganisms.

Agaricus, Gibberella, and Phanerochaete spp.

[00178] *Agaricus*, *Gibberella*, and *Phanerochaete* spp. can be useful because they are known to produce large amounts of isoprenoids in culture. Thus, the terpene precursors for producing large amounts of steviol glycosides are already produced by endogenous genes. Thus, modules comprising recombinant genes for steviol glycoside biosynthesis polypeptides can be introduced into species from such genera without the necessity of introducing mevalonate or MEP pathway genes.

Arxula adenivorans (Blastobotrys adenivorans)

[00179] *Arxula adenivorans* is dimorphic yeast (it grows as budding yeast like the baker's yeast up to a temperature of 42°C, above this threshold it grows in a filamentous form) with unusual biochemical characteristics. It can grow on a wide range of substrates and can assimilate nitrate. It has successfully been applied to the generation of strains that can produce natural plastics or the development of a biosensor for estrogens in environmental samples.

Yarrowia lipolytica

[00180] *Yarrowia lipolytica* is dimorphic yeast (see *Arxula adenivorans*) and belongs to the family Hemiascomycetes. The entire genome of *Yarrowia lipolytica* is known. *Yarrowia* species is aerobic and considered to be non-pathogenic. *Yarrowia* is efficient in using hydrophobic substrates (e.g., alkanes, fatty acids, oils) and can grow on sugars. It has a high potential for industrial applications and is an oleaginous microorganism. *Yarrowia lipolytica* can accumulate lipid content to approximately 40% of its dry cell weight and is a model organism for lipid accumulation and remobilization. See e.g., Nicaud, 2012, *Yeast* 29(10):409-18; Beopoulos et al., 2009, *Biochimie* 91(6):692-6; Bankar et al., 2009, *Appl Microbiol Biotechnol.* 84(5):847-65.

Rhodotorula sp.

[00181] *Rhodotorula* is unicellular, pigmented yeast. The oleaginous red yeast, *Rhodotorula glutinis*, has been shown to produce lipids and carotenoids from crude glycerol (Saenge et al., 2011, *Process Biochemistry* 46(1):210-8). *Rhodotorula toruloides* strains have been shown to be an efficient fed-batch fermentation system for improved biomass and lipid productivity (Li et al., 2007, *Enzyme and Microbial Technology* 41:312-7).

Rhodospiridium toruloides

[00182] *Rhodospiridium toruloides* is oleaginous yeast and useful for engineering lipid-production pathways (See e.g. Zhu *et al.*, 2013, *Nature Commun.* 3:1112; Ageitos *et al.*, 2011, *Applied Microbiology and Biotechnology* 90(4):1219-27).

Candida boidinii

[00183] *Candida boidinii* is methylotrophic yeast (it can grow on methanol). Like other methylotrophic species such as *Hansenula polymorpha* and *Pichia pastoris*, it provides an excellent platform for producing heterologous proteins. Yields in a multigram range of a secreted foreign protein have been reported. A computational method, IPRO, recently predicted mutations that experimentally switched the cofactor specificity of *Candida boidinii* xylose reductase from NADPH to NADH. See, e.g., Mattanovich *et al.*, 2012, *Methods Mol Biol.* 824:329-58; Khoury *et al.*, 2009, *Protein Sci.* 18(10):2125-38.

Hansenula polymorpha (*Pichia angusta*)

[00184] *Hansenula polymorpha* is methylotrophic yeast (see *Candida boidinii*). It can furthermore grow on a wide range of other substrates; it is thermo-tolerant and can assimilate nitrate (see also *Kluyveromyces lactis*). It has been applied to producing hepatitis B vaccines, insulin and interferon alpha-2a for the treatment of hepatitis C, furthermore to a range of technical enzymes. See, e.g., Xu *et al.*, 2014, *Virologica Sinica* 29(6):403-9.

Kluyveromyces lactis

[00185] *Kluyveromyces lactis* is yeast regularly applied to the production of kefir. It can grow on several sugars, most importantly on lactose which is present in milk and whey. It has successfully been applied among others for producing chymosin (an enzyme that is usually present in the stomach of calves) for producing cheese. Production takes place in fermenters on a 40,000 L scale. See, e.g., van Ooyen *et al.*, 2006, *FEMS Yeast Res.* 6(3):381-92.

Pichia pastoris

[00186] *Pichia pastoris* is methylotrophic yeast (see *Candida boidinii* and *Hansenula polymorpha*). It provides an efficient platform for producing foreign proteins. Platform elements are available as a kit and it is worldwide used in academia for producing proteins. Strains have been engineered that can produce complex human N-glycan (yeast glycans are similar but not identical to those found in humans). See, e.g., Piirainen *et al.*, 2014, *N Biotechnol.* 31(6):532-7.

Physcomitrella spp.

[00187] *Physcomitrella mosses*, when grown in suspension culture, have characteristics similar to yeast or other fungal cultures. This genera can be used for producing plant secondary metabolites, which can be difficult to produce in other types of cells.

[00188] It can be appreciated that the recombinant host cell disclosed herein can comprise a plant cell, comprising a plant cell that is grown in a plant, a mammalian cell, an insect cell, a fungal cell, comprising a yeast cell, wherein the yeast cell is a cell from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Yarrowia lipolytica*, *Candida glabrata*, *Ashbya gossypii*, *Cyberlindnera jadinii*, *Pichia pastoris*, *Kluyveromyces lactis*, *Hansenula polymorpha*, *Candida boidinii*, *Arxula adenivorans*, *Xanthophyllomyces dendrorhous*, or *Candida albicans species* or is a Saccharomycete or is a *Saccharomyces cerevisiae* cell, an algal cell or a bacterial cell, comprising *Escherichia* cells, *Lactobacillus* cells, *Lactococcus* cells, *Cornebacterium* cells, *Acetobacter* cells, *Acinetobacter* cells, or *Pseudomonas* cells.

Steviol Glycoside Compositions

[00189] Steviol glycosides do not necessarily have equivalent performance in different food systems. It is therefore desirable to have the ability to direct the synthesis to steviol glycoside compositions of choice. Recombinant hosts described herein can produce compositions that are selectively enriched for specific steviol glycosides (e.g., RebD or RebM) and have a consistent taste profile. As used herein, the term “enriched” is used to describe a steviol glycoside composition with an increased proportion of a particular steviol glycoside, compared to a steviol glycoside composition (extract) from a stevia plant. Thus, the recombinant hosts described herein can facilitate the production of compositions that are tailored to meet the sweetening profile desired for a given food product and that have a proportion of each steviol glycoside that is consistent from batch to batch. In some embodiments, hosts described herein do not produce or produce a reduced amount of undesired plant by-products found in Stevia extracts. Thus, steviol glycoside compositions produced by the recombinant hosts described herein are distinguishable from compositions derived from *Stevia* plants.

[00190] The amount of an individual steviol glycoside (e.g., RebA, RebB, RebD, or RebM) accumulated can be from about 1 to about 7,000 mg/L, e.g., about 1 to about 10 mg/L, about 3 to about 10 mg/L, about 5 to about 20 mg/L, about 10 to about 50 mg/L, about 10 to about 100 mg/L, about 25 to about 500 mg/L, about 100 to about 1,500 mg/L, or about 200 to about 1,000

mg/L, at least about 1,000 mg/L, at least about 1,200 mg/L, at least about at least 1,400 mg/L, at least about 1,600 mg/L, at least about 1,800 mg/L, at least about 2,800 mg/L, or at least about 7,000 mg/L. In some aspects, the amount of an individual steviol glycoside can exceed 7,000 mg/L. The amount of a combination of steviol glycosides (e.g., RebA, RebB, RebD, or RebM) accumulated can be from about 1 mg/L to about 7,000 mg/L, e.g., about 200 to about 1,500, at least about 2,000 mg/L, at least about 3,000 mg/L, at least about 4,000 mg/L, at least about 5,000 mg/L, at least about 6,000 mg/L, or at least about 7,000 mg/L. In some aspects, the amount of a combination of steviol glycosides can exceed 7,000 mg/L. In general, longer culture times will lead to greater amounts of product. Thus, the recombinant microorganism can be cultured for from 1 day to 7 days, from 1 day to 5 days, from 3 days to 5 days, about 3 days, about 4 days, or about 5 days.

[00191] It will be appreciated that the various genes and modules discussed herein can be present in two or more recombinant microorganisms rather than a single microorganism. When a plurality of recombinant microorganisms is used, they can be grown in a mixed culture to produce steviol and/or steviol glycosides. For example, a first microorganism can comprise one or more biosynthesis genes for producing a steviol glycoside precursor, while a second microorganism comprises steviol glycoside biosynthesis genes. The product produced by the second, or final microorganism is then recovered. It will also be appreciated that in some embodiments, a recombinant microorganism is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

[00192] Alternatively, the two or more microorganisms each can be grown in a separate culture medium and the product of the first culture medium, e.g., steviol, can be introduced into second culture medium to be converted into a subsequent intermediate, or into an end product such as RebA. The product produced by the second, or final microorganism is then recovered. It will also be appreciated that in some embodiments, a recombinant microorganism is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

[00193] Steviol glycosides and compositions obtained by the methods disclosed herein can be used to make food products, dietary supplements and sweetener compositions. See, e.g., WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328.

[00194] For example, substantially pure steviol or steviol glycoside such as RebM or RebD can be included in food products such as ice cream, carbonated 2s, fruit juices, yogurts, baked

goods, chewing gums, hard and soft candies, and sauces. Substantially pure steviol or steviol glycoside can also be included in non-food products such as pharmaceutical products, medicinal products, dietary supplements and nutritional supplements. Substantially pure steviol or steviol glycosides may also be included in animal feed products for both the agriculture industry and the companion animal industry. Alternatively, a mixture of steviol and/or steviol glycosides can be made by culturing recombinant microorganisms separately, each producing a specific steviol or steviol glycoside, recovering the steviol or steviol glycoside in substantially pure form from each microorganism and then combining the compounds to obtain a mixture comprising each compound in the desired proportion. The recombinant microorganisms described herein permit more precise and consistent mixtures to be obtained compared to current Stevia products.

[00195] In another alternative, a substantially pure steviol or steviol glycoside can be incorporated into a food product along with other sweeteners, *e.g.*, saccharin, dextrose, sucrose, fructose, erythritol, aspartame, sucralose, monatin, or acesulfame potassium. The weight ratio of steviol or steviol glycoside relative to other sweeteners can be varied as desired to achieve a satisfactory taste in the final food product. See, *e.g.*, U.S. 2007/0128311. In some embodiments, the steviol or steviol glycoside may be provided with a flavor (*e.g.*, citrus) as a flavor modulator.

[00196] Compositions produced by a recombinant microorganism described herein can be incorporated into food products. For example, a steviol glycoside composition produced by a recombinant microorganism can be incorporated into a food product in an amount ranging from about 20 mg steviol glycoside/kg food product to about 1800 mg steviol glycoside/kg food product on a dry weight basis, depending on the type of steviol glycoside and food product. For example, a steviol glycoside composition produced by a recombinant microorganism can be incorporated into a dessert, cold confectionary (*e.g.*, ice cream), dairy product (*e.g.*, yogurt), or beverage (*e.g.*, a carbonated beverage) such that the food product has a maximum of 500 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into a baked good (*e.g.*, a biscuit) such that the food product has a maximum of 300 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into a sauce (*e.g.*, chocolate syrup) or vegetable product (*e.g.*, pickles) such that the food product has a maximum of 1000 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into

bread such that the food product has a maximum of 160 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism, plant, or plant cell can be incorporated into a hard or soft candy such that the food product has a maximum of 1600 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism, plant, or plant cell can be incorporated into a processed fruit product (e.g., fruit juices, fruit filling, jams, and jellies) such that the food product has a maximum of 1000 mg steviol glycoside/kg food on a dry weight basis. In some embodiments, a steviol glycoside composition produced herein is a component of a pharmaceutical composition. See, e.g., Steviol Glycosides Chemical and Technical Assessment 69th JECFA, 2007, prepared by Harriet Wallin, Food Agric. Org.; EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS), "Scientific Opinion on the safety of steviol glycosides for the proposed uses as a food additive," 2010, *EFSA Journal* 8(4):1537; U.S. Food and Drug Administration GRAS Notice 323; U.S Food and Drug Administration GRAS Notice 329; WO 2011/037959; WO 2010/146463; WO 2011/046423; and WO 2011/056834.

[00197] For example, such a steviol glycoside composition can have from 90-99 weight % RebA and an undetectable amount of stevia plant-derived contaminants, and be incorporated into a food product at from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis.

[00198] Such a steviol glycoside composition can be a RebB-enriched composition having greater than 3 weight % RebB and be incorporated into the food product such that the amount of RebB in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebB-enriched composition has an undetectable amount of stevia plant-derived contaminants.

[00199] Such a steviol glycoside composition can be a RebD-enriched composition having greater than 3 weight % RebD and be incorporated into the food product such that the amount of RebD in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebD-enriched composition has an undetectable amount of stevia plant-derived contaminants.

[00200] Such a steviol glycoside composition can be a RebE-enriched composition having greater than 3 weight % RebE and be incorporated into the food product such that the amount of RebE in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000

mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebE-enriched composition has an undetectable amount of stevia plant-derived contaminants.

[00201] Such a steviol glycoside composition can be a RebM-enriched composition having greater than 3 weight % RebM and be incorporated into the food product such that the amount of RebM in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebM-enriched composition has an undetectable amount of stevia plant-derived contaminants.

[00202] In some embodiments, a substantially pure steviol or steviol glycoside is incorporated into a tabletop sweetener or “cup-for-cup” product. Such products typically are diluted to the appropriate sweetness level with one or more bulking agents, e.g., maltodextrins, known to those skilled in the art. Steviol glycoside compositions enriched for RebA, RebB, RebD, RebE, or RebM, can be package in a sachet, for example, at from 10,000 to 30,000 mg steviol glycoside/kg product on a dry weight basis, for tabletop use. In some embodiments, a steviol glycoside produced *in vitro*, *in vivo*, or by whole cell bioconversion

[00203] The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

EXAMPLES

[00204] The Examples that follow are illustrative of specific embodiments of the invention, and various uses thereof. They are set forth for explanatory purposes only, and are not to be taken as limiting the invention.

Example 1: Strain Engineering

[00205] Steviol glycoside-producing *S. cerevisiae* strains were constructed as described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328, each of which is incorporated by reference in its entirety. For example, yeast strains comprising and expressing a native gene encoding a YNK1 polypeptide (SEQ ID NO:122, SEQ ID NO:123), a native gene encoding a PGM1 polypeptide (SEQ ID NO:1, SEQ ID NO:2), a native gene encoding a PGM2 polypeptide (SEQ ID NO:118, SEQ ID NO:119), a native gene encoding a UGP1 polypeptide (SEQ ID NO:120, SEQ ID NO:121), a recombinant gene encoding a GGPPS polypeptide (SEQ ID NO:19, SEQ ID NO:20), a recombinant gene encoding a truncated CDPS

polypeptide (SEQ ID NO:39, SEQ ID NO:40), a recombinant gene encoding a KS polypeptide (SEQ ID NO:51, SEQ ID NO:52), a recombinant gene encoding a KO polypeptide (SEQ ID NO:59, SEQ ID NO:60), a recombinant gene encoding a KO polypeptide (SEQ ID NO:63, SEQ ID NO:64), a recombinant gene encoding an ATR2 polypeptide (SEQ ID NO:91, SEQ ID NO:92), a recombinant gene encoding a KAHe1 polypeptide (SEQ ID NO:93, SEQ ID NO:94), a recombinant gene encoding a CPR8 polypeptide (SEQ ID NO:85, SEQ ID NO:86), a recombinant gene encoding a CPR1 polypeptide (SEQ ID NO:77, SEQ ID NO:78), a recombinant gene encoding a UGT76G1 polypeptide (SEQ ID NO:8, SEQ ID NO:9), a recombinant gene encoding a UGT85C2 polypeptide (SEQ ID NO:5/SEQ ID NO:6, SEQ ID NO:7), a recombinant gene encoding a UGT74G1 polypeptide (SEQ ID NO:3, SEQ ID NO:4), a recombinant gene encoding a UGT91d2e-b polypeptide (SEQ ID NO:12, SEQ ID NO:13) and a recombinant gene encoding an EUGT11 polypeptide (SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16) were engineered to accumulate steviol glycosides.

Example 2: Overexpression of PGM1, PGM2, UGP1, and YNK1

[00206] A steviol glycoside-producing *S. cerevisiae* strain as described in Example 1, further engineered to comprise and express a recombinant gene encoding a KAH polypeptide (SEQ ID NO:96, SEQ ID NO:97) and a recombinant gene encoding a KO polypeptide (SEQ ID NO:117, SEQ ID NO:64), was transformed with vectors comprising an additional copy of the gene encoding a YNK1 polypeptide (SEQ ID NO:122, SEQ ID NO:123), operably linked to a pTEF1 promoter (SEQ ID NO:148) and a CYC1 terminator (SEQ ID NO:154), an additional copy of the gene encoding a PGM1 polypeptide (SEQ ID NO:1, SEQ ID NO:2), operably linked to a pTEF1 promoter (SEQ ID NO:148) and a CYC1 terminator (SEQ ID NO:154), an additional copy of the gene encoding a PGM2 polypeptide (SEQ ID NO:118, SEQ ID NO:119), operably linked to a pPGK1 promoter (SEQ ID NO:149) and a tADH1 terminator (SEQ ID NO:155), and an additional copy of the gene encoding a UGP1 polypeptide (SEQ ID NO:120, SEQ ID NO:121), operably linked to a pPGK1 promoter (SEQ ID NO:149) and a tADH1 terminator (SEQ ID NO:155).

[00207] Fed-batch fermentation with cultures of the transformed *S. cerevisiae* strain and a control *S. cerevisiae* strain (a steviol glycoside-producing *S. cerevisiae* strain as described in Example 2, further engineered to comprise and express a recombinant gene encoding a KAH

polypeptide and a recombinant gene encoding a KO polypeptide) was carried out aerobically in 2L fermenters at 30°C with an approximate 16 h growth phase in minimal medium comprising glucose, ammonium sulfate, trace metals, vitamins, salts, and buffer followed by an approximate 100 h feeding phase with a glucose-comprising defined feed medium. A pH near 6.0 and glucose-limiting conditions were maintained. Extractions of whole culture samples (without cell removal) were performed and extracts were analyzed by LC-UV to determine levels of steviol glycosides.

[00208] LC-UV was conducted with an Agilent 1290 instrument comprising a variable wavelength detector (VWD), a thermostatted column compartment (TCC), an autosampler, an autosampler cooling unit, and a binary pump, using SB-C18 rapid resolution high definition (RRHD) 2.1 mm x 300 mm, 1.8 µm analytical columns (two 150 mm columns in series; column temperature of 65°C). Steviol glycosides were separated by a reversed-phase C18 column followed by detection by UV absorbance at 210 nm. Quantification of steviol glycosides was done by comparing the peak area of each analyte to standards of RebA and applying a correction factor for species with differing molar absorptivities. For LC-UV, 0.5 mL cultures were spun down, the supernatant was removed, and the wet weight of the pellets was calculated. The LC-UV results were normalized by pellet wet weight. Total steviol glycoside values of the fed-batch fermentation were calculated based upon the measured levels of steviol glycosides calculated as a sum (in g/L RebD equivalents) of measured RebA, RebB, RebD, RebE, RebM, 13-SMG, rubusoside, steviol-1,2-bioside, di-glycosylated steviol, tri-glycosylated steviol, tetra-glycosylated steviol, penta-glycosylated steviol, hexa-glycosylated steviol, and hepta-glycosylated steviol. Results are shown in Table 1.

Table 1: Steviol Glycoside accumulation by transformed *S. cerevisiae* strain and *S. cerevisiae* control strain.

	Transformed Strain		Control Strain	
	Accumulation (g/L RebD Equiv.)	Std. Error (g/L RebD Equiv.)	Accumulation (g/L RebD Equiv.)	Std. Error (g/L RebD Equiv.)
13-SMG	2.40	0.14	4.2	0.02
RebA	0.59	0.007	0.45	0.07
RebD	1.21	0.16	2.16	0.12
RebM	6.31	0.22	3.22	0.06
Total SG	11.90	0.33	11.76	0.34

[00209] A decrease in 13-SMG and RebD accumulation, and an increase in RebA and RebM accumulation were observed for the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, relative to the control strain. Furthermore, RebD + RebM accumulation levels increased upon overexpression of UGP1, YNK1, PGM1, and PGM2, while the total steviol glycosides produced by the experimental strain increased negligibly. In addition, RebD / RebM ratios of 0.2 and below were observed for the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, relative to the control strain.

Example 3: UGP1, PGM2 Activity Assay

[00210] Fed-batch fermentation with cultures of a *S. cerevisiae* strain overexpressing PGM1, PGM2, UGP1, and YNK1, as described in Example 2, and a control *S. cerevisiae* strain (a steviol glycoside-producing *S. cerevisiae* strain as described in Example 1) was carried out aerobically in 2L fermenters at 30°C with an approximate 16 h growth phase in minimal medium comprising glucose, ammonium sulfate, trace metals, vitamins, salts, and buffer followed by an approximate 100 h feeding phase with a glucose-comprising defined feed medium. A pH near 6.0 and glucose-limiting conditions were maintained. Whole culture samples (without cell removal) were analyzed to determine the activity levels of PGM and UGP.

[00211] For both assays, frozen fermentation cell pellets were resuspended in CellLytic™ Y Cell Lysis Reagent (Sigma) to an OD₆₀₀ of 44. Samples were shaken 30 min at 25°C and then centrifuged at 13,000 rpm for 10 min. The supernatant was recovered and stored on ice.

[00212] The PGM enzyme assay relies on a coupled activity assay wherein supplied glucose-1-phosphate is first converted to glucose-6-phosphate by a PGM polypeptide/PGM polypeptide containing cell lysate, followed by glucose-6-phosphate conversion by a glucose-6-phosphate dehydrogenase (added to the assay as a purified enzyme in excess) to phosphogluconolactone under β -NADP⁺ consumption. The kinetics of the concomitant β -NAPDH released are recorded by monitoring the absorbance at 340 nm.

[00213] 180 mM glycylglycine, pH 7.4 (adjusted with NaOH/HCl); 5.0 mM glucose-1-phosphate; 3.00 mM β -NADP⁺; 0.4 mM G1,6-bisphosphate; 30 mM MgCl₂, 43 mM L-cysteine; 0.65 U/ml G6P-DH, and previously stored cell lysate were mixed together at 30°C at different cell-lysate/buffer concentrations (0.5% (v/v), 1%(v/v), 2%(v/v), and 3%(v/v)). The kinetics for the release of β -NAPDH were followed over a maximum of 1000 sec. for each concentration of

supernatant added. PGM activity for each cell-lysate/buffer concentration was defined by the maximum slope of the curve of OD₃₄₀ versus time. Cell-lysate/buffer concentration corrected PGM activity was defined as the slope of the curve of OD₃₄₀/sec as a function of Cell-lysate/buffer concentrations. The value obtained in this way for a certain strain can be compared to the values from other strains and differences in PGM activity can be pointed out. The increase in activity of the cell-lysate of the *S. cerevisiae* strain overexpressing PGM1, PGM2, UGP1, and YNK1 is shown in Table 3, below, relative to that of the control strain.

[00214] The UGP assay relies on a coupled activity assay of the yeast UDP-glucose pyrophosphorylase wherein supplied glucose-1-phosphate is first converted to UDP-glucose by a UGP polypeptide/UGP polypeptide-containing cell-lysate under UTP consumption, followed by UDP-glucose conversion to UDP-Glucuronate and β -NADH by UDP-glucose dehydrogenase (added to the assay as a purified enzyme in excess) under β -NAD⁺ consumption. The kinetics for the release of β -NADH are followed by monitoring the change in absorbance at 340 nm. Alternative UGP assays using, for example but not limited to, hydrophilic interaction liquid chromatography coupled with tandem mass spectrometry for the quantification of UDP-glucose (see Warth *et al.*, Journal of Chromatography A, 1423, pp. 183–189 (2016)) may be used as well.

[00215] 100 mM Tris/HCl, pH 8.5; 10 mM MgCl₂; 100 mM NaCl; 5.0 mM β -NAD⁺; 2 mM UTP; 2 mM ATP; 0.12 mg/ml UDPG-DH; 5 mM; and previously stored cell lysate were mixed together at 30°C at different supernatant/buffer concentrations (0.5% (v/v), 1%(v/v), 1.5%(v/v), and 2%(v/v)). The kinetics for the release of β -NADH were followed over a maximum of 1000 sec. for each supernatant/buffer concentration. UGP activity for each cell-lysate/buffer concentration was defined by the maximum slope of the curve of OD₃₄₀ versus time. Cell-lysate/buffer concentration corrected UGP activity was defined as the slope of the curve of OD₃₄₀/sec as a function of Cell-lysate/buffer concentrations. The value obtained in this way for a certain strain can be compared to the values from other strains and differences in UGP activity can be pointed out. The increase in activity of the lysate of the *S. cerevisiae* strain overexpressing PGM1, PGM2, UGP1, and YNK1 is shown in Table 2, below, relative to that of the control strain.

Table 2. Relative UGP and PGM activity

	Transformed Strain	Control Strain
UGP Activity relative to control strain	250%	100%

PGM Activity relative to control stain	160%	100%
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[00216] Individual and total steviol glycoside values of the fed-batch fermentation were calculated according to Example 2. Results are shown in Table 3.

Table 3: Steviol Glycoside accumulation by transformed *S. cerevisiae* strain and *S. cerevisiae* control strain.

	Transformed Strain	Control Strain
	Accumulation (g/L RebD Equiv.)	Accumulation (g/L RebD Equiv.)
RebD	2.19	1.21
RebM	5.71	5.12
Total SG	12.10	9.43

[00217] An increase in both UGP and PGM activity was observed for the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, relative to the control strain. As shown in Table 3, RebD and total steviol glycoside accumulation increased upon overexpression of UGP1, YNK1, PGM1, and PGM2. Without being bound to a particular theory, the results suggest that increasing UGP and PGM activity (*i.e.*, by expressing genes encoding polypeptides involved in the UDP-glucose biosynthetic pathway) allows for conversion of partially glycosylated steviol glycosides to higher molecular weight steviol glycosides, including, *e.g.*, RebD.

Example 4: LC-MS Analytical Procedures (UDP-glucose Analysis)

[00218] LC-MS analyses were performed on a Thermo Scientific Accela UPLC (Ultra Performance Liquid Chromatography system; Thermo Scientific) with a Thermo Scientific PAL autosampler system (Thermo Scientific) SeQuant ZIC-cHILIC column (2.1 mm x 150 mm, 3.0 μ m analytical column, 100 Å pore size) coupled to a Thermo Scientific Exactive Orbitrap mass spectrometer with electrospray ionization (ESI) operated in negative ionization mode. Compound separation was achieved using a gradient of the two mobile phases: A (water with 0.1% ammonium acetate) and B (MeCN). Separation was achieved by using a gradient from time 0 min with 15% A holding until 0.5 min and increasing to 50% A at time 15.50 min, holding until time 17.50 min, and reducing to 15% A at time 17.60 min, equilibrating at 15% A until 25.50

min. The flow rate was 0.3 mL/min, and the column was maintained at room temperature. UDP-glucose was monitored by full-scan analysis in the mass range 130-1400 m/z. EIC (Extracted ion chromatogram) of 565.04492-565.05058 corresponding to UDP-glucose was extracted and quantified by comparing against authentic standards. See Table 4 for m/z trace and retention time values of UDP-glucose.

Table 4: LC-MS Analytical Data for UDP-glucose

Compound	MS Trace	RT (mins)
UDP-glucose	565.04775	8.4

[00219] To determine the intracellular concentration of UDP-Glucose, full fermentation broth was sampled (via syringe) at desired time points during different stages of fermentation. Biomass (cells) was quickly separated by centrifugation and supernatant was removed. Cell pellets were quenched and extracted using a mixture of methanol, chloroform and an aqueous buffer solution. The final intracellular extracts were stored at -80°C prior to LC-MS analysis.

Example 5: UDP-glucose Accumulation Quantification

[00220] Fed-batch fermentation with cultures of a *S. cerevisiae* strain overexpressing PGM1, PGM2, UGP1, and YNK1, as described in Example 2, and a control *S. cerevisiae* strain (a *S. cerevisiae* strain comprising and expressing a native gene encoding a YNK1 polypeptide (SEQ ID NO:122, SEQ ID NO:123), a native gene encoding a PGM1 polypeptide (SEQ ID NO:1, SEQ ID NO:2), a native gene encoding a PGM2 polypeptide (SEQ ID NO:118, SEQ ID NO:119), a native gene encoding a UGP1 polypeptide (SEQ ID NO:120, SEQ ID NO:121), a recombinant gene encoding a GGPPS polypeptide (SEQ ID NO:19, SEQ ID NO:20), a recombinant gene encoding a truncated CDPS polypeptide (SEQ ID NO:39, SEQ ID NO:40), a recombinant gene encoding a KS polypeptide (SEQ ID NO:51, SEQ ID NO:52), a recombinant gene encoding a KO polypeptide (SEQ ID NO:59, SEQ ID NO:60), a recombinant gene encoding a KAHe1 polypeptide (SEQ ID NO:93, SEQ ID NO:94), a recombinant gene encoding a CPR8 polypeptide (SEQ ID NO:85, SEQ ID NO:86), a recombinant gene encoding a CPR1 polypeptide (SEQ ID NO:77, SEQ ID NO:78), a recombinant gene encoding an ATR2 polypeptide (SEQ ID NO:91, SEQ ID NO:92), a recombinant gene encoding a UGT85C2

polypeptide (SEQ ID NO:5/SEQ ID NO:6, SEQ ID NO:7), and a recombinant gene encoding a UGT74G1 polypeptide (SEQ ID NO:3, SEQ ID NO:4)) was carried out aerobically in 2L fermenters at 30°C with an approximate 16 h growth phase in minimal medium comprising glucose, ammonium sulfate, trace metals, vitamins, salts, and buffer followed by an approximate 100 h feeding phase with a glucose-comprising defined feed medium. A pH near 6.0 and glucose-limiting conditions were maintained. Whole culture samples (without cell removal) were analyzed by LC-UV to determine the levels of steviol glycosides, according to Example 2, and by LC-MS to analyze the intracellular level of UDP-glucose, according to Example 4. Results are shown in Tables 5-6.

Table 5: Steviol Glycoside accumulation by transformed *S. cerevisiae* strain and *S. cerevisiae* control strain.

	Transformed Strain	Control Strain
	Accumulation (g/L RebD Equiv.)	Accumulation (g/L RebD Equiv.)
RebD	1.05	1.92
RebM	5.75	2.23
Total SG	10.18	7.40

Table 6: UDP-glucose accumulation by transformed *S. cerevisiae* strain and *S. cerevisiae* control strain.

Time (h)	Transformed Strain		Control Strain	
	UDP-glucose Accumulation (μM)	Std. Deviation (μM)	UDP-glucose Accumulation (μM)	Std. Deviation (μM)
22	450.52	54.96	306.50	51.75
30	495.66	10.83	198.88	36.95
46	518.26	26.13	241.30	45.69
55	425.39	70.01	221.35	64.36
72	398.08	41.85	206.26	19.54
76	299.16	33.57	159.96	5.06
96	270.53	82.67	160.74	9.19
104	310.97	24.57	132.08	21.17
120	359.92	24.30	119.32	37.39

[00221] An increase in UDP-glucose accumulation, by up to 300%, was observed for the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, relative to the control strain. RebD + RebM accumulation levels increased upon overexpression of UGP1, YNK1, PGM1, and PGM2; this result further demonstrates a beneficial effect of expression of UDP-glucose biosynthetic pathway genes on the production of higher molecular weight steviol glycosides such as RebD or RebM.

[00222] One of skill in the art would appreciate a distinction between improving the total amount of UDP-glucose as compared to the recycling of UDP-glucose. As shown in Table 6 above, taking the highest and lowest number over fermentation time, the worst decrease in parental strain is 2.5 while the worst decrease in UDP-glucose boosted strain (*i.e.*, the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2) is 1.9 times. This demonstrates that overexpressing UGP1, YNK1, PGM1, and PGM2 increases the UDP-glucose pool and UDP-glucose. In fact, the net increase (consumption/formation) is higher in the UDP-glucose boosted strain.

[00223] Without being bound to a particular theory, the results observed in Examples 2-5 suggest that increasing UDP-glucose levels (*i.e.*, by expressing genes encoding polypeptides involved in the UDP-glucose biosynthetic pathway) allows for conversion of 13-SMG and other partially glycosylated steviol glycosides to higher molecular weight steviol glycosides, including, *e.g.*, RebM. Furthermore, the difference between the magnitude of the increase in accumulation levels of, *e.g.*, RebM and/or RebD and that of the increase in accumulation levels of the total steviol glycosides suggests that maintaining and/or increasing UDP-glucose levels allows for more efficient production of higher molecular weight steviol glycosides, including, *e.g.*, RebM (*i.e.*, by shifting the profile of produced steviol glycosides away from lower molecular weight steviol glycosides).

Example 6: Expression of Heterologous UGP1 and PGM2

[00224] A steviol glycoside-producing *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, as described in Example 2, was transformed with vectors comprising a gene encoding a UGP1 polypeptide (SEQ ID NO:132, SEQ ID NO:133) operably linked to a pPDC1 promoter (SEQ ID NO:153) and a tCYC1 terminator (SEQ ID NO:154) and a gene encoding a

PGM2 polypeptide (SEQ ID NO:144, SEQ ID NO:145), operably linked to a pTPI1 promoter (SEQ ID NO:152) and an tADH1 terminator (SEQ ID NO:155).

[00225] Fed-batch fermentation with cultures of the transformed *S. cerevisiae* strain and a control *S. cerevisiae* strain (a steviol glycoside-producing *S. cerevisiae* strain as described in Example 2, further engineered to comprise and express a recombinant gene encoding a ~~Stevia KAH polypeptide~~, KAH polypeptide and a recombinant gene encoding a KO polypeptide) was carried out aerobically in 2L fermenters at 30°C with an approximate 16 h growth phase in minimal medium comprising glucose, ammonium sulfate, trace metals, vitamins, salts, and buffer followed by an approximate 100 h feeding phase with a glucose-comprising defined feed medium. A pH near 6.0 and glucose-limiting conditions were maintained. Whole culture samples (without cell removal) were analyzed by LC-UV to determine levels of steviol glycosides, as described in Example 2. Results are shown in Table 7.

Table 7: Steviol Glycoside accumulation by transformed *S. cerevisiae* strain and *S. cerevisiae* control strain.

	Transformed Strain	Control Strain
	Accumulation (g/L RebD Equiv.)	Accumulation (g/L RebD Equiv.)
RebD	2.27	1.80
RebM	5.33	4.50
Total SG	14.27	12.39

[00226] An increase in RebD and RebM accumulation were observed for the *S. cerevisiae* strain expressing PGM2 and UGP1, relative to the control strain. Furthermore, total steviol glycosides produced by the experimental strain also increased. Without being bound to a particular theory, the results observed in Table 7 suggest that increasing UDP-glucose levels (*i.e.*, by expressing genes encoding polypeptides involved in the UDP-glucose biosynthetic pathway) allows for conversion of 13-SMG and other partially glycosylated steviol glycosides to higher molecular weight steviol glycosides, including, *e.g.*, RebM.

Example 7: LC-MS Analytical Procedures (Steviol Glycoside Analysis)

[00227] LC-MS analyses were performed on a Waters ACQUITY UPLC (Ultra Performance Liquid Chromatography system; Waters Corporation) with a Waters ACQUITY UPLC (Ultra Performance Liquid Chromatography system; Waters Corporation) BEH C18 column (2.1 x 50 mm, 1.7 μ m particles, 130 Å pore size) equipped with a pre-column (2.1 x 5 mm, 1.7 μ m particles, 130 Å pore size) coupled to a Waters ACQUITY TQD triple quadrupole mass spectrometer with electrospray ionization (ESI) operated in negative ionization mode. Compound separation was achieved using a gradient of the two mobile phases, A (water with 0.1% formic acid) and B (MeCN with 0.1% formic acid), by increasing from 20% to 50 % B between 0.3 to 2.0 min, increasing to 100% B at 2.01 min and holding 100% B for 0.6 min, and re-equilibrating for 0.6 min. The flow rate was 0.6 mL/min, and the column temperature was set at 55°C. Steviol glycosides were monitored using SIM (Single Ion Monitoring) and quantified by comparing against authentic standards. See Table 1 for m/z trace and retention time values of steviol glycosides and glycosides of steviol precursors detected.

Table 8: LC-MS Analytical Data for Steviol and Glycosides of Steviol and Steviol Precursors

Compound	MS Trace	RT (mins)
steviol+5Glc (#22) [also referred to as compound 5.22]	1127.48	0.85
steviol+6Glc (isomer 1) [also referred to as compound 6.1]	1289.53	0.87
steviol+7Glc (isomer 2) [also referred to as compound 7.2]	1451.581	0.94
steviol+6Glc (#23) [also referred to as compound 6.23]	1289.53	0.97

Compound	MS Trace	RT (mins)
RebE	965.42	1.06
RebD	1127.48	1.08
RebM	1289.53	1.15
steviol+7Glc (isomer 5) [also referred to as compound 7.5]	1451.581	1.09
steviol+4Glc (#26) [also referred to as compound 4.26]	965.42	1.21
steviol+5Glc (#24) [also referred to as compound 5.24]	1127.48	1.18
steviol+4Glc (#25) [also referred to as compound 5.25]	1127.48	1.40
RebA	965.42	1.43
1,2-Stevioside	803.37	1.43
steviol+4Glc (#33) [also referred to as compound 4.33]	965.42	1.49
steviol+3Glc (#1) [also referred to as compound 3.1]	803.37	1.52
steviol+2Glc (#57) [also referred to as compound 2.57]	641.32	1.57
RebQ	965.42	1.59
1,3-Stevioside (RebG)	803.37	1.60
Rubusoside	641.32	1.67
RebB	803.37	1.76
Steviol-1,2-Bioside	641.32	1.80
Steviol-1,3-Bioside	641.32	1.95
19-SMG	525.27	1.98
13-SMG	479.26	2.04
<i>ent</i> -kaurenoic acid+3Glc (isomer 1) [also referred to as compound KA3.1]	787.37	2.16

Compound	MS Trace	RT (mins)
<i>ent</i> -kaurenoic acid+3Glc (isomer 2) [also referred to as compound KA3.2]	787.37	2.28
<i>ent</i> -kaurenol+3Glc (isomer 1) co-eluted with <i>ent</i> -kaurenol+3Glc (#6) [also referred to as compounds KL3.1 and KL3.6]	773.4	2.36
<i>ent</i> -kaurenoic acid+2Glc (#7) [also referred to as compound KA2.7]	625.32	2.35
<i>ent</i> -kaurenol+2Glc (#8) [also referred to as compound KL2.8]	611.34	2.38
Steviol	317.21	2.39

[00228] Steviol glycosides can be isolated using a method described herein. For example, following fermentation, a culture broth can be centrifuged for 30 min at 7000 rpm at 4°C to remove cells, or cells can be removed by filtration. The cell-free lysate can be obtained, for example, by mechanical disruption or enzymatic disruption of the host cells and additional centrifugation to remove cell debris. Mechanical disruption of the dried broth materials can also be performed, such as by sonication. The dissolved or suspended broth materials can be filtered using a micron or sub-micron filter prior to further purification, such as by preparative chromatography. The fermentation media or cell-free lysate can optionally be treated to remove low molecular weight compounds such as salt, and can optionally be dried prior to purification and re-dissolved in a mixture of water and solvent. The supernatant or cell-free lysate can be purified as follows: a column can be filled with, for example, HP20 Diaion resin (aromatic-type Synthetic Adsorbent; Supelco) or another suitable non-polar adsorbent or reverse phase chromatography resin, and an aliquot of supernatant or cell-free lysate can be loaded on to the column and washed with water to remove the hydrophilic components. The steviol glycoside product can be eluted by stepwise incremental increases in the solvent concentration in water or a gradient from, e.g., 0% → 100% methanol. The levels of steviol glycosides, glycosylated *ent*-kaurenol, and/or glycosylated *ent*-kaurenoic acid in each fraction, including the flow-through, can then be analyzed by LC-MS. Fractions can then be combined and reduced in volume using

a vacuum evaporator. Additional purification steps can be utilized, if desired, such as additional chromatography steps and crystallization.

Example 8: Expression of Heterologous UGP1

[00229] A steviol glycoside-producing *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, as described in Example 2, was transformed with a vector comprising a codon-optimized nucleotide sequence encoding a UGP1 polypeptide (SEQ ID NO:132, SEQ ID NO:133) operably linked to a pTDH3 promoter (SEQ ID NO:150) and a tCYC1 terminator (SEQ ID NO:154), as summarized in Table 9, below.

Table 9. UGP1 Polypeptides Expressed

Strain	SEQ ID
1	126, 127
2	132, 133
3	128, 129
4	130, 131
5	124, 125
6	138, 139
7	136, 137
8	134, 135

[00230] Single colonies of the transformed strains provided in Table 9, and a control strain, transformed with a blank vector, were grown in 500 μ L of Delft medium in a 96-well plate for 2 days at 30°C, shaking at 280 rpm. 50 μ L of the cell culture of each strain was then transferred to a second 96-well plate and grown in 450 μ L Feed-in-Time medium (m2p-labs GmbH, Baesweiler, Germany) for 4 days at 30°C, shaking at 280 rpm. Samples for LC-MS analysis were prepared by extracting 100 μ L of cell solution with 100 μ L of DMSO, vortexing until mixed, and incubating at 80°C for 10 minutes. The resultant extract was clarified by centrifugation at 15,000 *g* for 10 min. 20 μ L of the supernatant was diluted with 140 μ L of 50% (v/v) DMSO for LC-MS injection. LC-MS data was normalized to the OD₆₀₀ of a mixture of 100 μ L of the cell solution and 100 μ L of water, measured on an ENVISION® Multilabel Reader (PerkinElmer, Waltham, MA).

[00231] LC-MS analysis was performed according to Example 7. Whole culture accumulation of compounds in $\mu\text{M}/\text{OD}_{600}$ was quantified by LC-MS against a known standard. Results are shown in Table 10, below. Each value is an average of 6 independent clones.

Table 10. Concentration of Steviol Glycosides

Strain	Accumulated Concentration ($\mu\text{M}/\text{OD}_{600}$)						
	13-SMG	Rubu.	RebB	RebA	RebD	RebM	Total
Control	9.96 \pm 2.19	0.05 \pm 0.08	0.67 \pm 0.14	1.95 \pm 0.79	3.89 \pm 0.60	20.73 \pm 4.48	37.38 \pm 6.71
1	6.15 \pm 1.83	0.26 \pm 0.04	0.59 \pm 0.09	2.37 \pm 0.65	1.49 \pm 0.36	25.91 \pm 1.35	37.38 \pm 3.03
2	7.06 \pm 2.48	0.23 \pm 0.12	0.76 \pm 0.30	2.03 \pm 0.37	1.34 \pm 0.24	27.99 \pm 3.17	39.43 \pm 5.88
3	8.73 \pm 3.20	0.25 \pm 0.08	0.69 \pm 0.24	2.50 \pm 0.81	1.69 \pm 0.43	29.41 \pm 6.19	43.34 \pm 9.22
4	13.02 \pm 2.39	0.14 \pm 0.08	0.99 \pm 0.23	2.88 \pm 0.51	4.89 \pm 0.75	30.41 \pm 5.90	52.50 \pm 9.51
5	7.91 \pm 2.30	0.28 \pm 0.08	0.62 \pm 0.14	2.55 \pm 0.96	1.42 \pm 0.33	29.54 \pm 4.23	42.37 \pm 5.98
6	8.89 \pm 2.94	0.28 \pm 0.04	0.68 \pm 0.18	2.36 \pm 0.66	1.43 \pm 0.49	27.64 \pm 3.49	41.32 \pm 5.08
7	5.68 \pm 2.05	0.23 \pm 0.09	0.51 \pm 0.19	2.04 \pm 0.50	1.26 \pm 0.28	23.63 \pm 2.27	33.38 \pm 4.98
8	6.59 \pm 2.65	0.22 \pm 0.12	0.63 \pm 0.17	2.28 \pm 1.03	1.49 \pm 0.59	26.64 \pm 6.51	37.90 \pm 10.21

[00232] Increases in steviol glycoside accumulation, by up to about 600%, was observed for the *S. cerevisiae* strain overexpressing UGP1, YNK1, PGM1, and PGM2, and further expressing heterologous UGP1, relative to the control strain. RebD + RebM accumulation levels increased upon expression of heterologous UGP1, further demonstrating a beneficial effect of expression of heterologous UDP-glucose biosynthetic pathway genes on the production of higher molecular weight steviol glycosides such as RebD or RebM.

[00233] Having described the invention in detail and by reference to specific embodiments thereof, it will be apparent that modifications and variations are possible without departing from the scope of the invention defined in the appended claims. More specifically, although some aspects of the present invention are identified herein as particularly advantageous, it is

contemplated that the present invention is not necessarily limited to these particular aspects of the invention.

Table 11. Sequences disclosed herein.

SEQ ID NO:1

S. cerevisiae

atgtcacttc	taatagattc	tgtaccaaca	gttgcttata	aggacaaaa	accgggtact	60
tcaggtttac	gtaagaagac	caagggtttc	atggatgagc	ctcattatac	tgagaacttc	120
attcaagcaa	caatgcaatc	tatcccataa	ggctcagagg	gaaccacttt	agttgttgga	180
ggagatggtc	gtttctacaa	cgatgttatc	atgaacaaga	ttgccgcagt	aggtgctgca	240
aacgggtgca	gaaagttagt	cattgggtcaa	ggcgggtttac	tttcaacacc	agctgcttct	300
catataatta	gaacatacga	ggaaaagtgt	accggtggtg	gtatcatatt	aactgcctca	360
cacaaccag	gcggtccaga	gaatgattta	ggatcaagt	ataatttacc	taatggtggg	420
ccagctccag	agagtgtcac	taacgctatc	tgggaagcgt	ctaaaaaatt	aactcactat	480
aaaattataa	agaacttccc	caagttgaat	ttgaacaagc	ttggtaaaaa	ccaaaaatat	540
ggccccattgt	tagtggacat	aattgatcct	gccaaagcat	acgttcaatt	tctgaaggaa	600
atTTTTgatt	ttgacttaat	taaaagcttc	ttagcgaaac	agcgcaaaga	caaaggggtg	660
aagttgttgt	ttgactcctt	aaatgggtatt	acaggaccat	atggtaaggc	tatatttgtt	720
gatgaatttg	gtttaccggc	agaggaagtt	cttcaaaatt	ggcacccttt	acctgatttc	780
ggcggtttac	atcccgatcc	gaatctaacc	tatgcacgaa	ctcttggtga	cagggttgac	840
cgcgaaaaaa	ttgcctttgg	agcagcctcc	gatgggtgat	gtgataggaa	tatgatttac	900
ggttatggcc	ctgctttcgt	ttcgccagg	gattctgttg	ccattattgc	cgaatatgca	960
cccgaatttc	catacttcgc	caaacaaggt	atTTatggct	tggcacgttc	atTtctaca	1020
tctcagcca	ttgatcgtgt	tgcagcaaaa	aagggattaa	gatgttacga	agttccaacc	1080
ggctggaaat	tcttctgtgc	cttatttgat	gctaaaaagc	tatcaatctg	tggtgaagaa	1140
tccttcggta	caggttccaa	tcatatcaga	gaaaaggacg	gtctatgggc	cattattgct	1200
tggtaaata	tcttggtctat	ctaccatagg	cgtaaccctg	aaaaggaagc	ttcgatcaaa	1260
actattcagg	acgaattttg	gaacgagtat	ggcogtactt	tcttcacaag	atacgattac	1320
gaacatatcg	aatgcgagca	ggccgaaaaa	gtttagcttc	ttttgagtga	atTtTtatca	1380
aggccaaacg	tttTgtgctc	ccacttccca	gctgatgagt	ctTtaaccgt	tatcgattgt	1440
ggtgattttt	cgtatagaga	tctagatggc	tccatctctg	aaaatcaagg	cctTtTcgta	1500
aagttttcga	atgggactaa	atTtTgtTtTg	aggttatccg	gcacaggcag	tTctggtgca	1560
acaataagat	tatacgtaga	aaagtatact	gataaaaagg	agaactatgg	ccaaacagct	1620
gacgtcttct	tgaaacccgt	catcaactcc	attgtaaaat	tcttaagatt	taaagaaatt	1680
ttaggaacag	acgaaccaac	agtccgcaca	tag			1713

SEQ ID NO:2

S. cerevisiae

MSLLIDSVPT	VAYKDQKPGT	SGLRKKTKVF	MDEPHYTENF	IQATMQSIPN	GSEGTTLVVG	60
GDGRFYNDVI	MNKIAAVGAA	NGVRKLVIGQ	GGLLSTPAAS	HIIRTYEEKC	TGGGIILTAS	120
HNPGGPENDL	GIKYNLPNGG	PAPESVTNAI	WEASKKLTHY	KIIKNFPKLN	LNKLGNQKY	180
GPLLVDIIDP	AKAYVQFLKE	IFDFDLIKSF	LAKQRKDKGW	KLLFDSLNGI	TGPYKAI FV	240
DEFGLPAEEV	LQNWHPPLDF	GGLHPDPNLT	YARTLVDRVD	REKIAFGAAS	DGDGDRNMIY	300
GYGPAFVSPG	DSVAIIAEYA	PEI PYFAKQG	IYGLARSFPT	SSAIDRVAAK	KGLRCYEVPT	360
GWKFFCALFD	AKKLSICGEE	SFGTGSNHIR	EKDGLWAI IA	WLNILAIYHR	RNPEKEASIK	420
TIQDEFWNEY	GRTFFTRYDY	EHIECEQAEK	VVALLSEFVS	RPNVCGSHFP	ADESLTVIDC	480
GDFSyrDLdG	SISENqGLFV	KFSNGTKFVL	RLSGTGSSGA	TIRLYVEKYT	DKKENYgQTA	540
DVFLKPVINS	IVKFLRFKEI	LGTDEPTVRT				570

SEQ ID NO:3

S. rebaudiana

atggcagagc	aacaaaaagat	caaaaagtca	cctcacgtct	tacttattcc	atTtctctctg	60
caaggacata	tcaaccatt	catacaattt	gggaaaaagat	tgattagtaa	gggtgtaaag	120
acaacactgg	taaccactat	ccacactttg	aattctactc	tgaaccactc	aaatactact	180
actacaagta	tagaaattca	agctatatca	gacggatgcg	atgaggggtg	ctTtatgtct	240

gccggtgaat	cttacttggga	aacattcaag	caagtgggat	ccaagtctct	ggccgatcta	300
atcaaaaagt	tacagagtga	aggcaccaca	attgacgcca	taatctacga	ttctatgaca	360
gagtggttt	tagacgttgc	tatcgaat	ggtattgatg	gaggttcctt	tttcacacaa	420
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ggtgaaactg	tttcagttcc	aggttttcca	gtgttacaac	ggtgggaaac	cccatgac	540
ttacaaaatc	atgaacaaat	acaatcacct	tggctccaga	tgttggttg	tcaattcgct	600
aacatcgatc	aagcaagatg	ggtctttact	aattcattct	ataagttaga	ggaagaggta	660
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aatggtatcg	tcagaagagg	gaacctagct	tcatgtatca	aaatgatcat	ggaagaggaa	1260
agaggagtta	tcataaggaa	aaacgcagtt	aagtgggaag	atcttgcaaa	ggttgccgtc	1320
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taa						1383

SEQ ID NO:4

S. rebaudiana

MAEQQKIKKS	PHVLLIPFPL	QGHINPFIQF	GKRLISKGVK	TTLVTTIHTL	NSTLNHSNTT	60
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EWVLDVAIEF	GIDGGSFFTQ	ACVVNSLYYH	VHKGLISLPL	GETVSVPGFP	VLQRWETPLI	180
LQNHEDIQSP	WSQMLFGQFA	NIDQARWVFT	NSFYKLEEEV	IEWTRKIWNL	KVIGPTLPSM	240
YLDKRLDDDK	DNGFNLYKAN	HHECMNWLDD	KPKESVYVVA	FGSLVKHGPE	QVEEITRALI	300
DSDVNFLWVI	KHKEEGKLEPE	NLSEVIKTGK	GLIVAWCKQL	DVLAHESVGC	FVTHCGFNST	360
LEAISLGPV	VAMPQFSDQT	TNAKLLDEIL	GVGVRVKADE	NGIVRRGNLA	SCIKMIMEEE	420
RGVIRKNAV	KWKDLAKVAV	HEGSSDNDI	VEFVSELIKA			460

SEQ ID NO:5

S. rebaudiana

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ccggaagcga	gcatcccaat	cagagaatca	ctcttgagat	ccattgaaac	caacttcttg	300
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gttttgcccc	ctgaacttga	ggaacatata	aagaaaagag	gctttattgc	tagctggtgt	1080
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gaccagctga	ccaactgtag	gtatatatgc	aaagaatggg	aggttgggct	cgagatggga	1260
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aacggttcat	cttctttgaa	catagacaaa	atggtaagg	aaatcaccgt	gctagcaaga	1440
aactagttac	aaagttgttt	cacattgtgc	tttctattta	agatgtaact	ttgttctaata	1500
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SEQ ID NO:6

S. rebaudiana

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aactaa						1446

SEQ ID NO:7

S. rebaudiana

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKQOTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:8

S. rebaudiana

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ttttctatta	caatctttca	caccaatttc	aacaaacca	aaacatcaa	ttaccacat	180

ttcacattca	gattcatact	tgataatgat	ccacaagatg	aacgtatttc	aaacttacct	240
accacaggtc	ctttagctgg	aatgagaatt	ccaatcatca	atgaacatgg	tgccgatgag	300
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gatagtaagc	agtcaatcct	ttgggtcgtg	cgtccagggt	tcgtgaaagg	ctcaacatgg	960
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caacaggaag	ttttagctca	tggcgctatt	ggggcattct	ggactcattc	cggatggaat	1080
tcaactttag	aatcagtatg	cgaaggggta	cctatgatct	tttcagattt	tggctcttgat	1140
caaccactga	acgcaagata	catgtctgat	gttttgaaag	tgggtgtata	tctagaaaaat	1200
ggctgggaaa	gggtgaaat	agctaatgca	ataagacgtg	ttatggttga	tgaagagggg	1260
gagtatatca	gacaaaacgc	aagagtgctg	aagcaaaagg	ccgacgtttc	tctaatagaag	1320
ggaggctctt	catacgaatc	cttagaatct	cttgtttcct	acatttcac	actgtaa	1377

SEQ ID NO:9

S. rebaudiana

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGFLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWVW	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNRVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:10

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ggacataaag	tgtcattcct	ttcaacaact	agaaacattc	aaagattatc	ttccacata	180
tcaccattga	ttaacgtcgt	tcaattgaca	cttccaagag	tacaggaatt	accagaagat	240
gctgaagcta	caacagatgt	gcatcctgaa	gatataccctt	acttgaaaaa	ggcatccgat	300
ggattacagc	ctgaggtcac	tagattcctt	gagcaacaca	gtccagattg	gatcatatac	360
gactacactc	actatgtggt	gccttcaatt	gcagcatcac	taggcatttc	tagggcacat	420
ttcagtgtaa	ccacaccttg	ggccattgct	tacatgggtc	catccgctga	tgctatgatt	480
aacggcagtg	atggtagaac	taccgttgaa	gatttgacaa	ccccaccaa	gtggtttcca	540
tttccaacta	aagtctgttg	gagaaaaac	gacttagcaa	gactggttcc	atacaaggca	600
ccaggaatct	cagacggcta	tagaatgggt	ttagtcctta	aagggtctga	ctgcctattg	660
tctaagtgtt	accatgagtt	tgggacacaa	tggctaccac	ttttgaaac	attacaccaa	720
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gactgggtt	ccgaagtgtt	agtatctcaa	acagaagtgt	tggaacttgc	cttaggtttg	900
gaactatctg	gattgccatt	tgtctgggcc	tacagaaaac	caaaaggccc	tgcaaagtcc	960
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acttcatggg	ctccacaatt	gagaatcctg	agtcacgaat	ctgtgtgcgg	tttcctaaca	1080
cattgtgggt	ctggttctat	agttgaagga	ctgatgtttg	gtcatccact	tatcatgttg	1140
ccaatctttg	gtgaccaagc	tttgaatgca	cgtctgttag	aagataaaca	agttggaatt	1200
gaaatcccac	gtaatgagga	agatggatgt	ttaaccaagg	agtctgtggc	cagatcatta	1260

cgttccggtg	tcggtgaaaa	ggaaggcgaa	atctacaagg	ccaatgcccc	tgaactttca	1320
aagatctaca	atgacacaaa	agtagagaag	gaatatgttt	ctcaatttgt	agattaccta	1380
gagaaaaacg	ctagagccgt	agctattgat	catgaatcct	aa		1422

SEQ ID NO:11

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDRKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:12

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ggtcacaagg	tttcattctt	gtctaccacc	agaaacatcc	aaagattgtc	ctctcataatc	180
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tttccaacaa	aagtctgttg	gagaaaacac	gatttggtta	gattggttcc	atacaaagct	600
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cattgtggtt	ctggttctat	cggtgaaggt	ttgatgtttg	gtcaccatt	gattatgttg	1140
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gaaatcccaa	gaaatgaaga	agatggttgc	ttgaccaaag	aatctgttgc	tagatctttg	1260
agatccgttg	tcggtgaaaa	agaaggtgaa	atctacaagg	ctaacgctag	agaattgtcc	1320
aagatctaca	acgataccaa	ggtcgaaaaa	gaatacgttt	cccaattcgt	tgactacttg	1380
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SEQ ID NO:13

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DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEALVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDRKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:14

O. sativa

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ctccaccgga	ggccttcga	cgggctcgcc	gogcccttct	cggagttctt	gggcaccgcg	360
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gacagacggc	tcgagcgcgc	ggagacagag	tcgcctgctg	ctgccgggca	gggacgcca	540
gcggcgggcg	caacgttcga	ggtggcgagg	atgaagttga	tacgaaccaa	aggctcatcg	600
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gtggaggaag	aaagcagcaa	agtgtttcaa	gccaaagcca	agaagctgca	ggagatcgtc	1320
gcggacatgg	cctgccatga	gaggtacatc	gacggattca	ttcagcaatt	gagatcttac	1380
aaggattga						1389

SEQ ID NO:15

O. sativa

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tgtgcagact	gggttatagt	cgatgtatct	catcactggg	ctgctgcagc	cgattggaa	420
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aactcaacaa	tagaaggact	gatgtttggt	catccactta	ttatgttacc	aatctttggc	1140
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aatgatggtg	atggttcctt	tgatagagaa	ggcgttgcag	ctgccatcag	agcagtcgcc	1260
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SEQ ID NO:16

O. sativa

MDSGYSSSYA	AAAGMHVIC	PWLAFGHLLP	CLDLAQRLLAS	RGHRVSFVST	PRNISRLPPV	60
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RPALAPLVAF	VALPLPRVEG	LPDGAESTND	VPHDRPDMVE	LHRRAFDGLA	APFSEFLGTA	120
CADWVIVDVF	HHWAAAAALE	HKVPCAMMLL	GSAHMIASIA	DRRLERAETE	SPAAAGQGRP	180
AAAPTFEVAR	MKLIRTKGSS	GMSLAERFSL	TLRSSSLVVG	RSCVEFEPET	VPLLSTLRGK	240
PITFLGLMPP	LHEGRREDGE	DATVRWLDAQ	PAKSVVYVAL	GSEVPLGVEK	VHELALGLEL	300
AGTRFLWALR	KPTGVSDADL	LPAGFEERTR	GRGVVATRWV	PQMSILAHAA	VGAFSLTHCGW	360
NSTIEGLMFG	HPLIMLPIFG	DQGNARLIE	AKNAGLQVAR	NDGDGSFDRE	GVAAAIRAVA	420
VEEESKVFQ	AKAKKLQIV	ADMACHERYI	DGFIQQLRSY	KD		462

SEQ ID NO:17

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RPALAPLVAF	VALPLPRVEG	LPDGAESTND	VPHDRPDMVE	LHRRAFDGLA	APFSEFLGTA	120
CADWVIVDVF	HHWAAAAALE	HKVPCAMMLL	GSAHMIASIA	DRRLERAETE	SPAAAGQGRP	180
AAAPTFEVAR	MKLIRTKGSS	GMSLAERFSL	TLRSSSLVVG	RSCVEFEPET	VPLLSTLRGK	240
PITFLGLLPP	EIPGDEKDET	WVSIKKWLDG	KQKGSVVYVA	LGSEALVSQT	EVVELALGLE	300
LSGLPFVWAY	RKPKGPAKSD	SVELPDGFVE	RTRDRGLVWT	SWAPQLRILS	HESVCGFLTH	360
CGSGSIVEGL	MFHGHLIMLP	IFGDQPLNAR	LLEDKQVGIE	IARNDDGDSF	DREGVAAAIR	420
AVAVEEESK	VFQAKAKLQ	EIVADMACHE	RYIDGFIQQL	RSYKD		465

SEQ ID NO:18

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SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DI PYLKKASD	GLQPEVTRFL	EQHSPDWI IY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDDL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLMP	PLHEGRREDG	EDATVRWLDA	QPAKSVVYVA	LGSEVPLGVE	KVELALGLE	300
LAGTRFLWAL	RKPTGVSDAD	LLPAGFEERT	RGRGVVATRW	VPQMSILAHA	AVGAFSLTHCG	360
WNSTIEGLMF	GHPLIMLPIF	GDQGNARLI	EAKNAGLQVP	RNEEDGCLTK	ESVARSLRSV	420
VVEKEGEIYK	ANARELSKIY	NDTKVEKEYV	SQFVDYLEKN	ARAVAI DHE S		470

SEQ ID NO:19

Synechococcus sp.

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aattga						1086

SEQ ID NO:20

Synechococcus sp.

MALVNPTALF	YGTSIRTRPT	NLLNPTQKLR	PVSSSSLPSP	SSVSAILTEK	HQSNPSENNN	60
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ACEIVGGNIL	NAMPACAVE	MIHTMSLVHD	DLPCMDNDDF	RRGKPISHKV	YGEEMAVLTG	180
DALLSLSFEH	IATATKGVSK	DRIVRAIGEL	ARSVGSEGLV	AGQVVDILSE	GADVGLDHLE	240
YIHIHKTAML	LESSVIGAI	MGGGSDQQIE	KLRKFARSIG	LLFQVDDIL	DVTKSTEELE	300
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N						361

SEQ ID NO:21

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tcctcatctt	ctgaaggcgg	ttcattgtct	agatacgacg	agagaagagt	ctctttgcct	180
ctcagtcata	atgctgcctc	tccagatatt	gtatcacaac	atggtttttc	cactgcaatg	240
tcttcagagt	tgaatcacag	atggaaatct	caaagattaa	aggtggccga	ttctccttac	300
aactatatcc	taacattacc	atcaaaagga	attagaggtg	cctttatcga	ttccctgaac	360
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cacaactctt	cattaatcat	tgatgacttc	caagataatt	ctccacttag	aagaggaaag	480
ccatctacc	atacagtctt	cggccctgcc	caggctatca	atactgctac	ttacgttata	540
gttaaagcaa	tcgaaaagat	acaagacata	gtgggacacg	atgcattggc	agatgttacg	600
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tggaaatga						1029

SEQ ID NO:22

MAEQQISNLL	SMFDASHASQ	KLEITVQMMD	TYHYRETPPD	SSSSEGGSL	RYDERRVSLP	60
LSHNAASPDI	VSQLCFSTAM	SSELNHRWKS	QRLKVADSPY	NYILTLPSKG	IRGAFIDSLN	120
VWLEVPEDET	SVIKEVIGML	HNSSLIIDDF	QDNSPLRRGK	PSTHTVFGPA	QAINATATYVI	180
VKAIEKIQDI	VGHDALADVT	GTITTFQGG	AMDLLWWTANA	IVPSIQEYLL	MVNDKTGALF	240
RLSLELLALN	SEASISDSAL	ESLSSAVSLL	GQYFQIRDDY	MNLIDNKYTD	QKGFCELDLE	300
GKYSLTLIHA	LQTDSSDLLT	NILSMRRVQG	KLTAQKRCWF	WK		342

SEQ ID NO:23

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tccatatacg	gggtaccaag	tgtaatcaac	tcagctaatt	acgtctactt	cttgggattg	300
gaaaaagtat	tgacattaga	tcatccagac	gctgtaaagc	tattcaccag	acaacttctt	360
gaattgcatc	aaggccaagg	tttgatatac	tattggagag	acacttatac	ttgccaaca	420
gaagaggagt	acaaagcaat	ggttctacaa	aagactggcg	gtttgttcgg	acttgccgtt	480
ggtctgatgc	aacttttctc	tgattacaag	gaggacttaa	agcctctggt	ggataccttg	540
ggcttgtttt	tccagattag	agatgactac	gctaacttac	attcaaagga	atattcagaa	600
aacaaatcat	tctgtgaaga	tttgactgaa	gggaagttta	gttttccaac	aatccacgcc	660
atttggtaa	gaccagaatc	tactcaagtg	caaaacattc	tgcgctcagag	aacagagaat	720
attgacatca	aaaagtattg	tgttcagtac	ttggaagatg	ttggttcttt	tgcttacaca	780
agacatacac	ttagagaatt	agaggcaaaa	gcatacaagc	aaatagaagc	ctgtggaggc	840
aatccttctc	tagtggcatt	ggttaaacat	ttgtccaaaa	tgttcaccga	ggaaaacaag	900
taa						903

SEQ ID NO:24

MEKTKEKAER	ILLEPYRYLL	QLPGKQVRSK	LSQAFNHWLK	VPEDKLQIII	EVTEMLHNAS	60
LLIDDIEDSS	KLRRGFVAH	SIYGVPSVIN	SANYVYFLGL	EKVLTLDHPD	AVKLFTRQLL	120
ELHQGQGLDI	YWRDITYCPT	EEEYKAMVLQ	KTGGLFGLAV	GLMQLFSDYK	EDLKPLDRTL	180
GLFFQIRDDY	ANLHSKEYSE	NKSFCEDLTE	GKFSFPTIHA	IWSRPESTQV	QNILRQRTE	240
IDIKKYCVQY	LEDVGSFAYT	RHTLRELEAK	AYKQIEACGG	NPSLVALVKH	LSKMFTEENK	300

SEQ ID NO:25

atggcaagat	tctatcttct	taacgcacta	ttgatgggta	tctcattaca	atcaactaca	60
gccttcactc	cagctaaact	tgcttatcca	acaacaacaa	cagctctaaa	tgctgcctcc	120
gccgaaactt	ctttcagtct	agatgaatac	ttggcctcta	agataggacc	tatagagtct	180
gccttgggaag	catcagtcaa	atccagaatt	ccacagaccg	ataagatctg	cgaatctatg	240
gcctactctt	tgatggcagg	aggcaagaga	attagaccag	tggtgtgat	cgctgcatgt	300
gagatgttcg	gtggatcca	agatgtcgt	atgcctactg	ctgtggcatt	agaaatgata	360
cacacaatgt	cttgattca	tgatgatttg	ccatccatgg	ataacgatga	cttgagaaga	420
ggtaaaccua	caaaccatgt	cgttttcggc	gaagatgtag	ctattcttgc	aggtgactct	480
ttattgtcaa	cttccttcga	gcacgtcgt	agagaaacaa	aaggagtgtc	agcagaaaag	540
atcgtggatg	ttatcgctag	attaggcaaa	tctgttggtg	ccgagggcct	tgctggcggt	600
caagttatgg	acttagaatg	tgaagctaaa	ccaggtacca	cattagacga	cttgaaatgg	660
attcatatcc	ataaaaccgc	tacattgtta	caagttgctg	tagcttctgg	tgcatgtcta	720
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actgcaggca	aagatgaagc	tactgataag	acaacttacc	caaagttatt	aggattagaa	900
gagagtaagg	catacgcaag	acaactaatc	gatgaagcca	aggaaagttt	ggctcctttt	960
ggagatagag	ctgccccttt	attggccatt	gcagatttca	ttattgatag	aaagaattga	1020

SEQ ID NO:26

MARFYFLNAL	LMVISLQSTT	AFTPAKLAYP	TTTTALNVAS	AETSFLSLEY	LASKIGPIES	60
ALEASVKSRI	PQTDKICESM	AYSIMAGGKR	IRPVLICIAAC	EMFGGSQDVA	MPTAVALEMI	120
HTMSLIHDDL	PSMDNDDLRR	GKPTNHVVFV	EDVAI LAGDS	LLSTSFEHVA	RETKGVSAEK	180
IVDVIARL GK	SVGAEGLAGG	QVMDLECEAK	PGTTLDDLKW	IHIHKATALL	QVAVASGAVL	240
GGATPEEVAA	CELFAMNIGL	AFQVADDILD	VTASSEDLGK	TAGKDEATDK	TTYPKLLGLE	300
ESKAYARQLI	DEAKESLAPF	GDRAAPLLAI	ADFIIDRKN			339

SEQ ID NO:27

atgcacttag	caccacgtag	agtccctaga	ggtagaagat	caccacctga	cagagttcct	60
gaaagacaag	gtgccttggg	tagaagacgt	ggagctggct	ctactggctg	tgcccgtgct	120
gctgctggtg	ttcaccgtag	aagaggagga	ggcgaggctg	atccatcagc	tgctgtgcat	180
agaggctggc	aagccgggtg	tgccaccggt	ttgcctgatg	aggtggtgtc	taccgcagcc	240
gccttagaaa	tgtttcatgc	ttttgcttta	atccatgatg	atatcatgga	tgatagtga	300
actagaagag	gctcccaac	tgttcacaga	gccctagctg	atcgtttagg	cgctgctctg	360
gaccagatc	aggccggtca	actaggagtt	tctactgcta	tcttggttgg	agatctggct	420
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gatgacctgc	taggcgtctt	cggtgatcca	agacgtacag	ggaaacctga	cctagatgat	780
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cagagacaca	cattggatac	attattgggt	acaccaggtc	ttgatagaca	aggcgcttca	900
agactaagat	gcgtattggt	agcaactggt	gcaagagccg	aagccgaaag	acttattaca	960
gagagaagag	atcaagcatt	aactgcattg	aacgcattaa	cactgccacc	tccttagctt	1020
gaggcattag	caagattgac	attagggtct	acagctcatc	ctgcctaa		1068

SEQ ID NO:28

MHLAPRRVPR	GRRSPPDRVP	ERQGALGRRR	GAGSTGCARA	AAGVHRRRGG	GEADPSAAVH	60
RGWQAGGGTG	LPDEVVSTAA	ALEMFHAFAL	IHDDIMDDSA	TRRGSPTVHR	ALADRLGAAL	120
DPDQAGQLGV	STAILVGDIA	LTWSDELLYA	PLTPHRLAAV	LPLVTAMRAE	TVHGQYLDIT	180
SARRPGTDT	LALRIARYKT	AAYTMERPLH	IGAALAGARP	ELLAGLSAYA	LPAGEAFQLA	240
DDLLGVFGDP	RRTGKPDLLD	LRGKHTVLV	ALAREHATPE	QRHTLDTLGG	TPGLDRQGAS	300
RLRCVLVATG	ARAEAEERLIT	ERRDQALTAL	NALTLPPPLA	EALARLTLGS	TAHPA	358

SEQ ID NO:29

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tcttacatct	ctggcgacgt	accaaacta	tacgaagcct	cctaccattt	gtttacatca	120
ggaggaaaga	gactaagacc	attgacccct	acaatttctt	ctgatctttt	cgggtggacag	180
agagaaagag	catactatgc	tggcgacgca	atcgaagttt	tgcacacatt	cactttgggt	240
cacgatgata	tcatggatca	agataacatt	cgtagaggct	ttcctactgt	acatgtcaag	300
tatggcctac	ctttggccat	tttagctggt	gacttattgc	atgcaaaagc	ctttcaattg	360
ttgactcagg	cattgagagg	tctaccatct	gaaactatca	tcaaggcgtt	tgatatcttt	420
acaagatcta	tcattatcat	atcagaaggt	caagctgtcg	atatggaatt	cgaagataga	480
attgatatca	aggaacaaga	gtatttggat	atgatatctc	gtaaaaccgc	tgcttattc	540
tcagcttctt	cttccattgg	ggcgttgata	gctggagcta	atgataacga	tgtgagatta	600
atgtccgatt	tcggtacaaa	tcttgggatc	gcatttcaaa	ttgtagatga	tatacttgggt	660
ttaacagctg	atgaaaaaga	gctaggaaaa	cctgttttca	gtgatatcag	agaaggtaaa	720
aagaccatat	tagtcattaa	gactttagaa	ttgtgtaagg	aagacgagaa	aaagattgtg	780
ttaaaagcgc	taggcaacaa	gtcagcatca	aaggaagagt	tgatgagttc	tgctgacata	840
atcaaaaagt	actcattgga	ttacgcctac	aacttagctg	agaaatacta	caaaaacgcc	900
atcgattctc	taaatcaagt	ttcaagtaaa	agtgatattc	caggggaaggc	attgaaatat	960
cttgctgaat	tcaccatcag	aagacgtaag	taa			993

SEQ ID NO:30

MSYFDNYFNE	IVNSVNDIIK	SYISGDVPKL	YEASYHLFTS	GGKRLRPLIL	TISSDLFGGQ	60
RERAYYAGAA	IEVLHTFTLV	HDDIMDQDNI	RRGLPTVHVK	YGLPLAILAG	DLLHAKAFQL	120
LTQALRGLPS	ETIIKAFDIF	TRSI III ISEG	QAVDMEFEDR	IDIKEQEYLD	MISRKTAALF	180
SASSSIGALI	AGANDNDVRL	MSDFGTNLGI	AFQIVDDILG	LTADEKELGK	PVFS DIREGK	240
KTILVIKTL	LCKEDEKKIV	LKALGNKSAS	KEELMSSADI	IKKYSLDYAY	NLAEKYYKNA	300
IDSLNQVSSK	SDIPGKALKY	LAEFTIRRRK				330

SEQ ID NO:31

atggtcgcac	aaactttcaa	cctggataacc	tacttatccc	aaagacaaca	acaagttgaa	60
gaggccctaa	gtgctgctct	tgtgccagct	tatcctgaga	gaatatacga	agctatgaga	120
tactccctcc	tggcaggtgg	caaaagatta	agacctatct	tatgttttagc	tgcttgcgaa	180
ttggcaggtg	gttctgttga	acaagccatg	ccaactgcgt	gtgcacttga	aatgatccat	240
acaatgtcac	taattcatga	tgacctgcca	gccatggata	acgatgattt	cagaagagga	300
aagccaacta	atcacaaggt	gttcggggaa	gatatagcca	tcttagcggg	tgatgcgctt	360
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tcacataaga	ctggagcctt	gctggaagca	tcagttgtct	caggcgggat	tctcgcaggg	600
gcagatgaag	agcttttggc	cagattgtct	cattacgcta	gagatatagg	cttggctttt	660
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ggtaaagacc	aggcagccgc	aaaggcaact	tatccaagtc	tattgggttt	agaagcctct	780
agacagaaag	cggaagagtt	gattcaatct	gctaaggaag	ccttaagacc	ttacggttca	840
caagcagagc	cactcctagc	gctggcagac	ttcatcacac	gctcgtcagca	taa	894

SEQ ID NO:32

MVAQTFNLDT	YLSQRQQVE	EALSAALVPA	YPERIYEAMR	YSLLAGGKRL	RPILCLAACE	60
LAGGSVEQAM	PTACALEMIH	TMSLIHDDL	AMDNDFFRRG	KPTNHKVFGE	DIAILAGDAL	120
LAYAFEHIAS	QTRGVPPQLV	LQVIARIGHA	VAATGLVGGQ	VVDLESEGKA	ISLETLEYIH	180
SHKTGALLEA	SVVSGGILAG	ADEELLARLS	HYARDIGLAF	QIVDDILDVT	ATSEQLGKTA	240
GKDQAAAKAT	YPSLLGLEAS	RQKAEELIQS	AKEALRPYGS	QAEPLLALAD	FITRRQH	297

SEQ ID NO:33

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aagttctctt	acaattactt	aaaggaaaag	caaagtacca	acgaattgct	ggataaatgg	1380
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gcaaagctgg	attacaataa	ctatgttgca	gtccttcaat	tagaatggta	cacaatacaa	1620
caatggtacg	tcgatattgg	tatagagaag	ttcgaatctg	acaacatcaa	gtcagtcctg	1680

SEQ ID NO:34

MKTGFISPAT	VFHHRISPAT	TFRHHLSPAT	TNSTGIVALR	DINFRCKAVS	KEYSDLLQKD	60
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QDVDGSGSPQ	FPSSLEWIAN	NQLSDGSGWD	HLLFSAHDRI	INTLACVIAL	TSWNVHPSKC	180
EKGLNFLREN	ICKLEDENAE	HMPIGFVTF	PSLIDIAKKL	NIEVPEDTPA	LKEIYARRDI	240
KLTKIPMEVL	HKVPTLLHS	LEGMPDLEWE	KLLKLQCKDG	SFLFSPSSTA	FALMQTKDEK	300
CLQYLTVNVT	KFNGGVPNVY	PVDLFEHIWV	VDRLQRLGIA	RYFKSEIKDC	VEYINKYWTK	360
NGICWARNTN	VQDIDDTAMG	FRVLAHGYD	VPDVFVRQFE	KDGKFCVCFAG	QSTQAVTGMF	420
NVYRASQMLF	PGERILEDAL	KFSYNYLKEK	QSTNELLDKW	IIAKDLPGEV	GYALDIPWYA	480
SLPRLETRY	LEQYGGEDDV	WIGKTLYRMG	YVSNNTYLEM	AKLDYNNYVA	VLQLEWYTIQ	540
QWYVDIGIEK	FESDNIKSVL	VSYYLAAASI	FEPERSKERI	AWAKTTILVD	KITSIFDSSQ	600
SSKEDITAFI	DKFRNKSSSK	KHSINGEPWH	EVMVALKCTL	HGFALDALMT	HSQDIHPQLH	660
QAWEMWLTKL	QDGVDTVTAEL	MVQMINMTAG	RWVSKELLTH	PQYQRLSTVT	NSVCHDITKL	720
HNFKENSTTV	DSKVQELVQL	VFSDTPDDL	QDMKQTFELTV	MKTFYYKAWC	DPNTINDHIS	780
KVFEIVI						787

SEQ ID NO:35

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agaagattgg	ggacatctga	ctccccacct	gatactatag	cagttgagct	ggttatcca	420
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gcattggcaa	cacatgggag	aggtagaaga	ccagaagtag	tgatggatta	caggactgac	1020
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gccatggata	ccgcatcagc	ttggctgctg	gcagctcaaa	agcaagatgg	ctcttggtta	1200
gataaatggc	atgcctcacc	atactacgct	actgtttgtt	gcacacaagc	cctagccgct	1260
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gatctattgt	taccaccatt	gtaa				1584

SEQ ID NO:36

MPDAHDAAPP	QIRQRTLVD	ATQLLITESAE	DAWGEVSVSE	YETARLVAHA	TWLGGHATRV	60
AFLLERQHED	GSWGPPGGYR	LVPTLSAVHA	LLTCLASPAQ	DHGVPHDRLL	RAVDAGLTAL	120
RRLGTSDSPP	DTIAVELVIP	SLLEGIQHLL	DPAHPHSRPA	FSQHRGSLVC	PGGLDGRITLG	180
ALRSHAAAGT	PVPGKVVHAS	ETLGLSTEEA	SHLQPAQGI	GGSAATATW	LTRVAPSQQS	240
DSARRYLEEL	QHRYSGPVPS	ITPITYFERA	WLLNNFAAAG	VPCEAPAALL	DSLEAALTPQ	300
GAPAGAGLPP	DADDTAAVLL	ALATHGRGRR	PEVLMDYRTD	GYFQCFIGER	TPSISTNAHV	360
LETGLHHVAQ	HPQDRARYGS	AMDTASAWLL	AAQKQDGSWL	DKWHASPYYA	TVCCTQALAA	420
HASPATAPAR	QRAVRWVLAT	QRSDGGWGLW	HSTVEETAYA	LQILAPPSGG	GNI PVQQALT	480
RGRARLCGAL	PLTPLWHDKD	LYTPVRVVRA	ARAAALYTTR	DLLLPL		527

SEQ ID NO:37

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tacgtcgaag	caaatagaaa	tccacatggt	ttgtgggaca	acgaaaaatg	gcacgtttca	1140
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gaattgtact	gtcctactag	agtcgtaaga	gtagctgagc	tagctggcct	gtgggtagca	1500
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SEQ ID NO:38

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AELILPQFCG	EAAWLLGGVA	FPRHPALLPL	RQACLVKLGA	VAMLPSGHPL	LHSWEAWGTS	180
PTTACPDDDG	SIGISPAATA	AWRAQAVTRG	STPQVGRADA	YLQMASRATR	SGIEGVFPNV	240
WPINVFPCW	SLYTLHLAGL	FAHPALAEAV	RVIVAQLEAR	LGVHGLGPAL	HFAADADDTA	300
VALCVLHLAG	RDPAVDALRH	FEIGELFVTF	PGERNASVST	NIHALHALRL	LGKPAAGASA	360
YVEANRNPHG	LWDNEKWHVS	WLYPTAHAVA	ALAQGKPQWR	DERALAALLQ	AQRDDGGWGA	420
GRGSTFEETA	YALFALHVMD	GSEEATGRRR	IAQVVARALE	WMLARHAAHG	LPQTPWLWIGK	480
ELYCPTRVVR	VAELAGLWLA	LRWGRRVLAE	GAGAAP			516

SEQ ID NO:39

Z. mays

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gcaaagggca	gcagtttgac	ccctatagtg	agaactgacg	ctgagtcaag	gagaacaaga	240
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SEQ ID NO:40

Z. mays

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GRGLSFLGRN	MWKLATEDEE	SMPIGFELAF	PSLIELAKSL	GVHDFPYDHQ	ALQGIYSSRE	240
IKMKRIPKEV	MHTVPTSILH	SLEGMPGLDW	AKLLKLQSSD	GSFLFSPAAT	AYALMNTGDD	300
RCFSYIDRTV	KKFNGGVPNV	YPVDLFEHIW	AVDRLERLGI	SRYFQKEIEQ	CMDYVNRHWT	360
EDGICWARNS	DVKEVDDTAM	AFRLRLRHGY	SVSPDVFKNF	EKDGEFFAFV	GQSNQAVTGM	420
YNLNRASQIS	FPGEDVLHRA	GAFSYEFLRR	KEAEGALRDK	WIISKDLPGE	VVYTLDFPWY	480
GNLPRVEARD	YLEQYGGDD	VWIGKTLYRM	PLVNNDVYLE	LARMDFNHCQ	ALHQLEWQGL	540
KRWYTENRLM	DFGVAQEDAL	RAYFLAAASV	YEP CRAAERL	AWARAAI LAN	AVSTHLRNSP	600
SFRERLEHSL	RCRPSEETDG	SWFNSSSGSD	AVLVKAVLRL	TDSLAREAQP	IHGDPEDII	660
HKLLRSAAWE	WVREKADAAD	SVCNGSSAVE	QEGSRMVHDK	QTCLLLLAMI	EISAGRAAGE	720
AASEDGDRRI	IQLTGSICDS	LKQKMLVSD	PEKNEEMMSH	VDDELKLRIR	EFVQYLLRLG	780
EKKTGSSETR	QTFLSIVKSC	YYAAHCPPHV	VDRHISRIVF	EPVSAAK		827

SEQ ID NO:41

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SEQ ID NO:42

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AWVALIDAGD	KTPAFPSAVK	WIAENQLSDG	SWGDAYLFSY	HDRLINTLAC	VVALRSWNLF	180
PHQCNGKITF	FRENIGKLED	ENDEHMPIGF	EVAFPSLLEI	ARGINIDVPY	DSPVLKDIYA	240
KKELKLTRIP	KEIMHKIPTT	LLHSLEGMRD	LDWEKLLKLO	SQDGSFLFSP	SSTAFAFMQT	300
RDSNCLEYLR	NAVKRFNGGV	PNVFPVDLFE	HIWIVDRLQR	LGISRYFEEE	IKECLDYVHR	360
YWTDNGICWA	RCSHVQDIDD	TAMAFRLLRQ	HGYQVSADVF	KNFEKEGEFF	CFVQSQSNQAV	420
TGMFNLYRAS	QLAFPREEIL	KNAKEFSYNY	LLEKREREEL	IDKWIIMKDL	PGEIGFALEI	480
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DIFQKWYEEN	RLSEWVRRS	ELLECYLAA	ATIFESERSH	ERMVWAKSSV	LVKAISSSFG	600
ESSDSRRSFS	DQFHEYIANA	RRSDHHFNDR	NMRLDRPGSV	QASRLAGVLI	GTLNQMSFDL	660
FMSHGRDVNN	LLYLSWGDWM	EKWKLYGDEG	EGELMVKMI I	LMKNNDLTNF	FTHTHFVRLA	720
EI INRICLPR	QYLKARRNDE	KEKTIKSM EK	EMGKMVELAL	SESDFTRDVS	ITFLDVAKAF	780
YYFALCGDHL	QTHISKVLFQ	KV				802

SEQ ID NO:43

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SEQ ID NO:44

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LSKQTDfSLM	LHKRELEQKR	CHSNEMDGYL	AYISEGLGNL	YDWNMVKKYQ	MKNGSVFNSP	240
SATAAAFINH	QNPgCLNYLN	SLLDKFGNAV	PTVYPHDLFI	RLSMVDTIER	LGISHHFRVE	300
IKNVLDETYR	CWVERDEQIF	MDVVTCALAF	RLLRINGYEV	SPDPLAEITN	ELALKDEYAA	360
LETYHASHIL	YQEDLSSGKQ	ILKSADFLKE	IISTDSNRLS	KLIHKEVENA	LKFPINTGLE	420
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MNSMLREAIW	TRDAYVPTLN	EYMENAYVSF	ALGPVVKPAI	YFVGPKLSEE	IVESSEYHNL	660
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EEQR						784

SEQ ID NO:45

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ttagaaacat	accatgcatc	ccaaatactt	taccaggaag	acctaagttc	aggaaaacaa	1140
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agaatcaata	ctaggagaaa	cattcagctg	tacaacgtag	ataatacaag	gattcttaag	1320
accacctacc	atagttcaaa	catttccaac	acctattact	taagattagc	tgtcgaagac	1380
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gaggaacaaa	gataa					2355

SEQ ID NO:46

MNLSLCIASP	LLTKSSRPTA	LSAIHTASTS	HGGQTNPTNL	IIDTTKERIQ	KLFKNVEISV	60
SSYDTAWVAM	VSPNSPKSP	CFPECLNWL	NNQLNDGSWG	LVNHTNHNH	PLLKDSLST	120
LACIVALKRW	NVGEDQINKG	LSFIESNLAS	ATDKSQPSPI	GFDIIFPGLL	EYAKNLDINL	180
LSKQTDFSLM	LHKRELEQKR	CHSNEIDGYL	AYISEGLGNL	YDWNMVKKYQ	MKNGSVFNSP	240
SATAAAFINH	QNPGLNYLN	SLLDKFGNAV	PTVYPLDLYI	RLSMVDTIER	LGISHHFRVE	300
IKNVLDETYR	CWVERDEQIF	MDVVTALAF	RLLRIHGYKV	SPDQLAEITN	ELAFKDEYAA	360
LETYHASQIL	YQEDLSSGKQ	ILKSADFLKG	ILSTDSNRLS	KLIHKEVENA	LKFPINTGLE	420
RINTRRNIQL	YNVDNTRILK	TTYHSSNISN	TYYLRLAVED	FYTCQSIYRE	ELKGLERWV	480
QNKLDQLKFA	RQKTAYCYFS	VAATLSSPEL	SDARISWAKN	GILTTVVDDF	FDIGGTIDEL	540
TNLIQCVEKW	NVDVDKCCS	EHVRILFLAL	KDAICWIGDE	AFKWQARDVT	SHVIQTWLEL	600
MNSMLREAIW	TRDAYVPTLN	EYMENAYVSF	ALGPVVKPAI	YFVGPKLSEE	IVESSEYHNL	660
FKLMSTQGRL	LNDIHSFKRE	FKEGKLNVA	LHLSNGESGK	VEEEVVEEMM	MMIKNKRKEL	720
MKLIFEENG	IVPRACKDAF	WNMCHVLNFF	YANDDGFTGN	TILDTVKDII	YNPLVLNEN	780
EEQR						784

SEQ ID NO:47

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ctagacagaa	cttacagatc	ttggttacaa	agacacgagg	aaatcatgct	ggacactatg	360
acatgtgcta	tggcttttag	aatcctaaga	ttgaacggat	acaacgtttc	atcagatgaa	420
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accagaacac	tacttgaatt	acacaaggct	tcaacagtta	gtatctctga	ggatgaatct	540
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ggcgactga	gaaagccttc	tttattcaaa	gaggttgaac	atgcactgga	tggacctttt	660
tacaccacac	ttgatagact	tcatcatagg	tggaatattg	aaaacttcaa	cattattgag	720
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caaggttctc	atacactggt	atctgatggt	taa			1773

SEQ ID NO:48

MAMPVKLTPA	SLSLKAVCCR	FSSGGHALRF	GSSLPCWRRT	PTQRSTSSST	TRPAAEVSSG	60
KSKQHDQEAS	EATIRQQLQL	VDVLENMGIS	RHFAAEIKCI	LDRTYRSWLQ	RHEEIMLDTM	120
TCAMAFRI LR	LNGYNVSSDE	LYHVVEASGL	HNSLGGYLND	TRTLLELHKA	STVSISEDES	180
ILDSIGSRSR	TLLREQLESG	GALRKPSL FK	EVEHALDGPF	YTTLDR LHHR	WNIENFNIE	240
QHMLETPYLS	NQHTSRDILA	LSIRDFSSSQ	FTYQQELQHL	ESWVKECR LD	QLQFARQKLA	300
YFYLSAAGTM	FSPELSDART	LWAKNGVLT T	IVDDFFDVAG	SKEELENL VM	LVEMWDEHHK	360
VEFYSEQVEI	IFSSYDSVN	QLGEKASLVQ	DRSITKHLVE	IWLDLLKSMM	TEVEWRLSKY	420
VPTEKEYMIN	ASLIFGLGPI	VLPALYFVGP	KISESIVKDP	EYDELFLKMS	TCGRLLNDVQ	480
TFEREYNEGK	LNSVSLLV LH	GGPMSISDAK	RKLQKPIDTC	RRDLLSLV LR	EESVVP RPCK	540
ELFWKMCKVC	YFFYSTTDGF	SSQVERAKEV	DAVINEPLKL	QGSHTLVSDV		590

SEQ ID NO:49

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SEQ ID NO:50

MSCIRPWFCP	SSISATLTDP	ASKLVTGEFK	TTSLNPHGTK	ERIKKMFDKI	ELSVSSYDTA	60
WVAMVPSDC	PETPCFPECT	KWILENQLGD	GSWSLPHGNP	LLVKDALSSST	LACILALKRW	120
GIGEEQINKG	LRFIELNSAS	VDNEQHKPI	GFDIIFPGMI	EYAKDLDLNL	PLKPTDINSM	180
LHRRALELTS	GGGNLEGR	AYLAYVSEGI	GKLQDWEMAM	KYQRKNGSLF	NSPSTTAAAF	240
IHIQDAECLH	YIRSLQKFG	NAVPTIYPLD	IYARLSMVDA	LERLGIDRHF	RKERKFLVDE	300
TYRFWLQGEE	EIFSDNATCA	LAFRILRLNG	YDVSLEDHFS	NSLGGYLKDS	GAALELYRAL	360
QLSYPDESLL	EKQNSRTSYF	LKQGLSNVSL	CGDRLRKNII	GEVHDALNFP	DHANLQRLAI	420
RRRIKHYATD	DTRILKTSYR	CSTIGNQDFL	KLAVEDFNIC	QSIQREEFKH	IERWVVERRL	480
DKLKFARQKE	AYCYFSAAAT	LFAPELSDAR	MSWAKNGVLT	TVVDDFFDVG	GSEELVNLI	540
ELIERWDVNG	SADFCSEEVE	I IYSAIHSTI	SEIGDKSFGW	QGRDVKSHVI	KIWLDLLKSM	600
LTEAQWSSNK	SVPTLDEYMT	TAHVSFALGP	IVLPALYFVG	PKLSEEVAGH	PELLNLYKVM	660
STCGRLNDW	RSFKRESEEG	KLNAISLYMI	HSGGASTEEE	TIEHFKGLID	SQRRQLQLV	720
LQEKDSIIPR	PCKDLFWNMI	KLLHTFYMKD	DGFTSNEMRN	VVKAIINEPI	SLDEL	775

SEQ ID NO:51

A. thaliana

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aaagaatctt	taacttga					2358

SEQ ID NO:52

A. thaliana

MSINLRSSGC	SSPISATLER	GLDSEVQTRA	NNVSFEQTK	KIRKMLEKVE	LSVSAYDTSW	60
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GIGERQINKG	LQFIELNSAL	VTDETIQKPT	GFDIIFPGMI	KYARDLNLTI	PLGSEVVDDM	180
IRKRDLKLC	DSEKFSKGRE	AYLAYVLEGT	RNLKDWDLIV	KYQRKNGSLF	DSPATTAAAF	240
TQFGNDGCLR	YLCSLLQKFE	AAVPSVYVYD	QYARLSIIVT	LESLGIDRDF	KTEIKSILDE	300
TYRYWLRGDE	EICLDLATCA	LAFRLLLAHG	YDVSYPDKP	FAEESGFSDT	LEGYVKNFVS	360
VLELFKAAQS	YPHESALKKQ	CCWTKQYLEM	ELSSVWVTSV	RDKYLKKEVE	DALAFPSYAS	420
LERSDHRRKI	LNGSAVENTR	VTKTSYRLHN	ICTSDILKLA	VDDFNFCQSI	HREEMERLDR	480
WIVENRLQEL	KFARQKLAYC	YFSGAATLFS	PELSDARISW	AKGGVLTTVV	DDFFDVGGSK	540
EELNLIHLV	EKWDLNGVPE	YSSEHVEIIF	SVLRDITILET	GDKAFTYQGR	NVTHHIVKIW	600
LDLLKSMRE	AEWSSDKSTP	SLEDYMENAY	ISFALGPVIVL	PATYLIGPPL	PEKTVDSHQY	660
NQLYKLVSTM	GRLNLDIQGF	KRESAEGKLN	AVSLHMKHER	DNRSKEVIIE	SMKGLAERKR	720
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KESLT						785

SEQ ID NO:53

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SEQ ID NO:54

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AASLRAQLAA	LDVSTTEHVG	FEIIVPAMLD	PLEAEDPSLV	FDFPARKPLM	KIHDAKMSRF	180
RPEYLYGKQP	MTALHSLEAF	IGKIDFDKVR	HHRTHGSMMG	SPSSTAAYLM	HASQWDGSE	240
AYLRHVIKHA	AGQGTGAVPS	AFPSTHFESS	WILTTLFRAG	FSASHLACDE	LNKLVEILEG	300
SFEKEGGAIG	YAPGFQADVD	DTAKTISTLA	VLGRDATPRQ	MIKVFEANTH	FRTYPGERDP	360
SLTANCNALS	ALLHQPDAAAM	YGSQIQKITK	FVCDYWWKSD	GKIKDKWNTC	YLYPSVLLVE	420
VLVDLVSLLE	QKGLPDVLDQ	ELQYRVAITL	FQACLRPLLD	QDAEGSWNKS	IEATAYGILI	480
LTEARRVCFE	DRLSEPLNEA	IRRGIAFADS	MSGTEAQLNY	IWIEKVSYP	ALLTKSYLLA	540
ARWAAKSPLG	ASVGSLSLWTP	PREGLDKHVR	LFHQAELEFRS	LPEWELRASM	IEAALFTPLL	600
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ATAGILFRDH	MDDLRLIHD	LLAEKTSFKS	SGRSSQGTKD	ADSGIEEDVS	MSDSASDSQD	720
RSPEYDLVFS	ALSTFTKHVL	QHPSIQSASV	WDRKLLAREM	KAYLLAHIQQ	AEDSTPLSEL	780
KDVPQKTDVT	RVSTSTTTF	NWVRTTSADH	ISCPYSFHFV	ACHLGAALSP	KGSNGDCYPS	840
AGEKFLAAAV	CRHLATMCRM	YNDLGSAERD	SDEGNLNSLD	FPEFADSAGN	GGIEIQKAAL	900
LRLAEFERDS	YLEAFRRLQD	ESNRVHGPAG	GDEARLSRRR	MAILEFFAQQ	VDLYGQVYVI	960
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SEQ ID NO:55

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SEQ ID NO:56

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LHSLEGLHRE	VDWNKLLQLQ	SENGSFLYSP	ASTACALMYT	KDVKCFDYLN	QLLIKFDHAC	360
PNVYPVDLFE	RLWMVDRLQR	LGISRYFERE	IRDCLQYVYR	YWKDCGIGWA	SNSSVQDVDD	420
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KKARTFSRNF	LRTKHENNEC	FDKWIITKDL	AGEVEYNLTF	PWYASLPRLE	HRTYLDQYGI	540
DDIWIGKSLY	KMPAVTNEVF	LKLAKADFNM	CQALHKKELE	QVIKWNASCQ	FRDLEFARQK	600
SVECYFAGAA	TMFPEPMVQA	RLVWARCCVL	TTVLDDYFDH	GTPVEELRVF	VQAVRTWNPE	660
LINGLPEQAK	ILFMGLYKTV	NTIAEEAFMA	QKRDRVHHLK	HYWDKLITSA	LKEAEWAESG	720
YVPTFDEYME	VAEISVALEP	IVCSTLFFAG	HRLDEDVLDS	YDYHLMHLV	NRVGRI LNDI	780
QGMKREASQG	KISSVQIYME	EHPSVPSEAM	AIAHLQELVD	NSMQQLTYEV	LRFTAVPKSC	840
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SEQ ID NO:57

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SEQ ID NO:58

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LERMHGEKLG	HFDLEQVYVK	PSSLLHSLEA	FLGKLD FDR L	SHHLYHGSMM	ASPSSTAAYL	240
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FTTFGSRDP	SLTSLNLHVL L	SLLKQSNLSQ	YHPQILKTTL	FTCRWWWGSD	HCVKDKWNLS	420
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IESSFFVPLL	QQRVEIYPR	DNIKVDEDKY	LSIIPFTWVG	CNNRSRTFAS	NRWLYDMMYL	660
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SEQ ID NO:59

S. rebaudiana

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SEQ ID NO:60

S. rebaudiana

MDAVTGLLTV	PATAITIGGT	AVALAVALIF	WYLKSYTSAR	RSQSNHLPRV	PEVPGVPLLG	60
NLLQLKEKKP	YMTFTRWAAT	YGPIYSIKTG	ATSMVVVSSN	EIAKEALVTR	FQSISTRNLS	120
KALKVLTADK	TMVAMSDYDD	YHKTVKRHIL	TAVLGPNAQK	KHRIHRDIMM	DNISTQLHEF	180
VKNNPEQEEV	DLRKIFQSEL	FGLAMRQALG	KDVESLYVED	LKITMNRDEI	FQVLVDPMM	240
GAIDVDWRDF	FPYLKWVPNK	KFENTIQQMY	IRREAVMKSL	IKEHKKRIAS	GEKLNSYIDY	300
LLSEAQTLTD	QQLLMSLWEP	IESSDTTMV	TTEWAMYELA	KNPKLQDRLY	RDIKSVCGSE	360
KITEEHLSQL	PYITAI FHET	LRRHSPVPII	PLRHVHEDTV	LGGYHVPAGT	ELAVNIYGCN	420
MDKNVWENPE	EWNPERFMKE	NETIDFQKTM	AFGGGKRVCA	GSLQALLTAS	IGIGRMVQEF	480
EWKLDKDMTQE	EVNTIGLTTQ	MLRPLRAIIK	PRI			513

SEQ ID NO:61

aagcttacta	gtaaaatgga	cggtgtcatc	gatatgcaaa	ccattccatt	gagaaccgct	60
attgctattg	gtggtactgc	tgttgctttg	gttggttgc	tatacttttg	gttcttgaga	120
tctacagctt	ccccatctca	tcattcta	caattgcccac	agttccagct	agttccagct	180
gttccagctt	tgggtaattt	gttgcaattg	aaagaaaaaa	agccttacat	gaccttcacc	240
aagtgggctg	aaatgtatgg	tccaatctac	tctattagaa	ctgggtgctac	ttccatgggt	300
gttgtctctt	ctaacgaaat	cgccaaagaa	gttggttgtta	ccagattccc	atctatctct	360
accagaaaat	tgtcttacgc	cttgaagggt	ttgaccgaag	ataagtctat	ggttgccatg	420
tctgattatc	acgattacca	taagaccgtc	aagagacata	ttttgactgc	tgttttgggt	480
ccaaacgccc	aaaaaaagtt	tagagcacat	agagacacca	tgatggaaaa	cgtttccaat	540
gaattgcatg	ccttcttcga	aaagaacca	aatcaagaag	tcaacttgag	aaagatcttc	600
caatcccaat	tattcggttt	ggctatgaag	caagccttg	gtaaagatgt	tgaatccatc	660
tacgttaagg	atttggaaac	caccatgaag	agagaagaaa	tcttcgaagt	tttggttgtc	720
gatccaatga	tgggtgctat	tgaagttgat	tggagagact	ttttcccata	cttgaatgg	780
gttccaaaca	agtccttcga	aaacatcatc	catagaatgt	acactagaag	agaagctggt	840
atgaaggcct	tgatccaaga	acacaagaaa	agaattgctt	ccggtgaaaa	cttgaactcc	900
tacattgatt	acttgttgtc	tgaagcccaa	accctgaccg	ataagcaatt	attgatgtct	960
ttgtgggaac	ctattatcga	atcttctgat	accactatgg	ttactactga	atgggctatg	1020
tacgaattgg	ctaagaatcc	aaacatgcaa	gacagattat	acgaagaaat	ccaatccggt	1080
tgcggttccg	aaaagattac	tgaagaaaac	ttgtcccaat	tgccatactt	gtacgctggt	1140
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gaaaacaccg	ttttgggtgg	ttatcatggt	ccagctggta	ctgaagttgc	tattaacatc	1260
tacggttgca	acatggataa	gaaggtctgg	gaaaatccag	aagaatggaa	tccagaaaga	1320
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ttgactacc	aaaagttgca	tccattattg	gccttgatta	acccaagaaa	gtaactcgag	1560
ccgcg						1566

SEQ ID NO:62

MDGVIDMQTI	PLRTAIAIGG	TAVALVVALY	FWFLRSYASP	SHHSNHLPPV	PEVPGVPVLG	60
NLLQLKEKKP	YMTFTKWAEM	YGPIYSIRTG	ATSMVVVSSN	EIAKEVVVTR	FPSISTRKLS	120
YALKVLTEDK	SMVAMSDYHD	YHKTVKRHIL	TAVLGPNAQK	KFRAHRDTMM	ENVSNELHAF	180
FEKNPNQEVN	LRKIFQSQLF	GLAMKQALGK	DVESIYVKDL	ETTMKREEIF	EVLVDPMMG	240
AIEVDWRDF	PYLKWPVNS	FENI IHRMYT	RREAVMKALI	QEHKKRIASG	ENLNSYIDYL	300
LSEAQTLTDK	QLLMSLWEP	IESSDTTMVT	TEWAMYELAK	NPNMQDRLYE	EIQSVCGSEK	360

ITEENLSQLP	YLYAVFQETL	RKHCPVPIMP	LRYVHENTVL	GGYHVPAGTE	VAINIYGCNM	420
DKKVWENPEE	WNPFRFLSEK	ESMDLYKTMA	FGGGRVCAG	SLQAMVISCI	GIGRLVQDFE	480
WKLKDDAEED	VNTLGLTTQK	LHPLLLALINP	RK			512

SEQ ID NO:63

R. suavis

atggccacc	tccttgagca	tttccaagct	atgccctttg	ccatccctat	tgcactggct	60
gctctgtctt	ggctgttctt	cttttacatc	aaagtttcat	tcttttccaa	caagagtgtc	120
caggctaagc	tccctcctgt	gccagtggtt	cctgggctgc	cggtgattgg	gaatttactg	180
caactcaagg	agaagaaacc	ctaccagact	tttacaaggt	gggctgagga	gtatggacca	240
atctattcta	tcaggactgg	tgcttccacc	atggteggtc	tcaataccac	ccaagttgca	300
aaagaggcca	tggtgaccag	atatttatcc	atctcaacca	gaaagctatc	aaacgcacta	360
aagattccta	ctgctgataa	atgtatgggtt	gcaataagtg	actacaacga	ttttcacaag	420
atgataaagc	gatacatact	ctcaaagtgt	cttggaccta	gtgctcagaa	gcgtcaccgg	480
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tctcctcgag	aagctgtgaa	tttcagaaga	gtttttgagt	gggaactcct	tggaattgca	600
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gaggttgatt	ggagagattt	cttcccttac	ctgagatgga	ttccgaatac	gcgcatggaa	780
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gaagggaaga	cactgacaat	ggaccaaaata	agtatgttgc	tttgggagac	ggttattgaa	960
acagcagata	ctacaatggt	aacgcagaaa	tgggctatgt	atgaagttgc	taaagactca	1020
aagcgtcagg	atcgtctcta	tcaggaaatc	caaaagggtt	gtggatcggg	gatggttaca	1080
gaggaatact	tgtcccaact	gccgtacctg	aatgcagttt	tccatgaaac	gctaaggaag	1140
cacagtcagg	ctgctgtagt	tcctttaaga	tatgcacatg	aagataacca	actaggaggt	1200
tactacattc	cagctggaac	tgagattgct	ataaacatat	acgggtgtaa	catggacaag	1260
catcaatggg	aaagccctga	ggaatggaaa	ccggagagat	ttttggacct	gaaatttgat	1320
cctatggatt	tgtacaagac	catggctttt	ggggctggaa	agagggtatg	tgctggttct	1380
cttcaggcaa	tgttaatagc	gtgcccagcg	attggtaggc	tgggtcagga	gtttgagtgg	1440
aagctgagag	atggagaaga	agaaaatgta	gatactgttg	ggctcaccac	tcacaaacgc	1500
tatccaatgc	atgcaatcct	gaagccaaga	agtta			1535

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R. suavis

atggctacct	tgttggaaaca	ttttcaagct	atgccattcg	ctattccaat	tgctttggct	60
gctttgtctt	ggttgttttt	gttctacatc	aaggtttctt	tcttctccaa	caaatccgct	120
caagctaaat	tgccaccagt	tccagttggt	ccaggtttgc	cagttattgg	taatttgttg	180
caattgaaag	aaaagaagcc	ataccaaacc	ttcactagat	gggctgaaga	atatggtcca	240
atctactcta	ttagaactgg	tgcttctact	atggttgtct	tgaacactac	tcaagttgcc	300
aaagaagcta	tggttaccag	atacttgtct	atctctacca	gaaagttgtc	caacgccttg	360
aaaattttga	ccgctgataa	gtgcatgggt	gccatttctg	attacaacga	tttccacaag	420
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gaagttgatt	ggagagattt	tttcccatac	ttgcgttgga	ttccaaacac	cagaatggaa	780
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caaaagaaaa	gaattgcctc	cggtgaagaa	atcaactgct	acatcgattt	cttgttgaaa	900
gaaggttaaga	ccttgaccat	ggaccaaatc	tctatgttgt	tgtgggaaac	cgttattgaa	960
actgctgata	ccacaatggt	tactactgaa	tgggctatgt	acgaagttgc	taaggattct	1020
aaaagacaag	acagattata	ccaagaaatc	caaaaggctc	gcggttctga	aatggttaca	1080
gaagaatact	tgtcccaatt	gccatacttg	aatgctgttt	tccacgaaac	tttgagaaaa	1140

cattctccag	ctgctttggt	tccattgaga	tatgctcatg	aagatactca	attgggtggt	1200
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caccaatggg	aatctccaga	agaatggaag	ccagaaaagat	ttttggatcc	taagtttgac	1320
ccaatggact	tgtacaaaac	tatggcTTTT	ggtgctggta	aaagagtttg	cgctggttct	1380
ttacaagcta	tgTTgattgc	ttgtccaacc	atcggtagat	tggttcaaga	atttgaatgg	1440
aagttgagag	atggtgaaga	agaaaacggt	gatactgTTg	gTTtgaccac	ccataagaga	1500
tatccaatgc	atgctatTTT	gaagccaaga	tcttaa			1536

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aagcttacta	gtaaaaatggc	ctccatcacc	cattttcttac	aagattttca	agctactcca	60
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ggtttccact	ctactaagaa	aaacgaatat	tacaagTTgc	caccagttcc	agttgTTcca	180
ggtttgccag	ttgTTgTaa	tttggTgcaa	ttgaaagaaa	agaagccata	caagactttc	240
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gTTgTTgTTa	actctactca	tgTTgcccAAA	gaagctatgg	ttaccagatt	ctcttcaatc	360
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ttcgaatctg	aattattcgg	tttggctatg	aagcaagcct	tggttatga	tgTTgattcc	660
ttgTTcgTTg	aagaattggg	tactaccttg	tccagagaag	aaatctacaa	cgTTttggTc	720
agtgacatgt	tgaagggtgc	tattgaagtt	gattggagag	actTTttccc	atacttGaaa	780
tggtcccaa	acaagtctt	cgaaatgaag	attcaaagat	tggtctctag	aagacaagcc	840
gTtatgaact	ctattgtcaa	agaacaaaag	aagtccattg	cctctgTgTaa	gggtgaaaac	900
gttacttga	attacttgtt	gtccgaagct	aagactTTga	ccgaaaagca	aatttccatt	960
ttggcctggg	aaaccattat	tgaaactgct	gatacaactg	ttgttaccac	tgaatgggct	1020
atgtacgaat	tggtcaaaaa	cccaaagcaa	caagacagat	tatacaacga	aatccaaaac	1080
gtctgcggtg	ctgataagat	taccgaagaa	catttgtcca	agttgcctta	cttgtctgct	1140
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catgaagata	ctcaattggg	tggttattat	gttccagccg	gtactgaaat	tgctgttaat	1260
atctacgggt	gcaacatgga	caagaatcaa	tgggaaactc	cagaagaatg	gaagccagaa	1320
agatttttgg	acgaaaagta	cgatccaatg	gacatgtaca	agactatgTc	ttttggTtcc	1380
ggtaaaaagag	tttgcgctgg	ttctttacaa	gctagTTtga	ttgcttgtac	ctccatcggt	1440
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ctcgagccgc	gg					1572

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MASITHFLQD	FQATPFATAF	AVGGVSLLI	FFFIRGFHST	KKNEYKLP	VPVPLPV	60
GNULLQLKEK	PYKTFWRWAE	IHGPIYSIRT	GASTMVVNS	THVAKEAMVT	RFSSISTRKL	120
SKALELLTSN	KSMVATSDYN	EFHKMVKKYI	LAELLGANAQ	KRHRHRDTL	IENVLNKLHA	180
HTKNSPLQAV	NFRKIFESEL	FGLAMKQALG	YDVDSLVEE	LGTTLSREEI	YNVLVSDMLK	240
GAIEVDWRDF	FPYLKWIIPNK	SFEMKIQLA	SRRQAVMNSI	VKEQKKSIA	GKGENCYLNY	300
LLSEAKTLTE	KQISILAWET	IIETADTTVV	TTEWAMYELA	KNPKQDRLY	NEIQNVCGTD	360
KITEEHLKSL	PYLSAVFHET	LRKYSPLV	PLRYAHEDTQ	LGGYVPAGT	EIAVNIYGCN	420
MDKNQWETPE	EWKPERFLDE	KYDPMYKT	MSFGSGKRV	AGSLQASLIA	CTSIGRLVQE	480
FEWRLKDG	ENVDTLGLTT	HKLYPMQAIL	QPRN			514

SEQ ID NO:67

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ccagttccag	gTTttccatt	gattggTaac	ttgTTgcaat	tGaaagaaaa	gaagccacac	180
aagactttca	ccaagtggTc	tgaattatat	ggTccaatct	actctatcaa	gatgggtTcc	240

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tcttcaatct	ctaccagaaa	gttgtcctaac	gcttttgactg	ttttgacctg	caacaaatct	360
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agattggctg	ttatgaacgc	cttgatccaa	gacagattga	atcaaaacga	ttccgaatcc	840
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attgctatct	tggtttggga	aaccattatc	gaaactgctg	ataccacttt	ggttactact	960
gaatgggcta	tgtacgaatt	ggccaaacat	caatctgttc	aagatagatt	attcaaagaa	1020
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tgccagataa	gatttttggg	agatagatac	gaatcctccg	acttgcataa	gactatggct	1320
tttggtgctg	gtaaaaagag	ttgtgctggt	gctttacaag	ctagtttgat	ggctggtatt	1380
gctatcggtg	gattggttca	agaattcgaa	tggaaagttg	gagatggtga	agaagaaaac	1440
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agaagatctt	aa					1512

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MASMSLLLG	FVVSFLFIF	FLKLLFFFS	RHKMSEVSR	PSVPVPGFPL	IGNLLQLKEK	60
KPHKTFKWS	ELYGPIYSIK	MGSSSLIVLN	SIETAKEAMV	SRFSSISTRK	LSNALTVLTC	120
NKSMVATSDY	DDFHKFKVRC	LLNGLLGANA	QERKRHYRDA	LIENVTSKLH	AHTRNHPQEP	180
VNFRAIFEHE	LFGVALKQAF	GKDVESIYVK	ELGVTLSRDE	IFKVLVHDM	EGAIDVDWRD	240
FFPYLKWIPN	NSFEARIQQK	HKRRLAVMNA	LIQDRLNQND	SESDDDCYLN	FLMSEAKTLT	300
MEQIAILVWE	TIIETADTTL	VTEWAMYEL	AKHQSVQDRL	FKEIQSVC	EKIKEEQLPR	360
LPYVNGVFHE	TLRKYSAPPL	VPIRYAHEDT	QIGGYHIPAG	SEIAINIYGC	NMDKKRWERP	420
EEWWPERFLE	DRYESSDLHK	TMAFGAGKRV	CAGALQASLM	AGIAIGRLVQ	EFEWKLDRGE	480
EENVDTYGLT	SQKLYPLMAI	INPRRS				506

SEQ ID NO:69

aagcttacta	gtaaaatgga	catgatgggt	attgaagctg	ttccatttgc	tactgctggt	60
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aacttgttgc	aattgaaaga	aaagaagcca	cataagacct	ttgctagatg	ggctgaaact	240
tacggtccaa	tttctctat	tagaactggt	gcttctacca	tgatcgtctt	gaattcttct	300
gaagttgcca	agaagctat	ggctactaga	ttctcttcaa	tctctaccag	aaagttgtcc	360
aacgccttga	agattttgac	cttcgataag	tgtatggttg	ccacctctga	ttacaacgat	420
tttcacaaaa	tggtcaaggg	tttcatcttg	agaaacgttt	taggtgctcc	agcccaaaaa	480
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gtaagactt	ctccattgga	accagttgct	ttgaagaaga	ttttcgaatc	cgaatttttc	600
ggtttggcct	tgaacaagc	cttgggtaag	gatatcgaat	ccatctatgt	tgaagaattg	660
ggtactacct	tgtccagaga	agaaattttt	gccgttttgg	ttgttgatcc	aatggctggt	720
gctattgaag	ttgattggag	agattttttc	ccatacttgt	cctggattcc	aaacaagtct	780
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gacccaata	gacaagaaat	cttgtagaga	gaaatccaca	aggtttgagg	ttctaacaag	1080
ttgactgaag	aaaacttgtc	caagttgcca	tacttgaact	ctgttttcca	cgaaaccttg	1140

agaaagtatt	ctccagctcc	aatggttcca	gttagatatg	ctcatgaaga	tactcaattg	1200
ggtggttacc	atattccagc	tggttctcaa	attgccatta	acatctacgg	ttgcaacatg	1260
aacaaaaagc	aatgggaaaa	tcctgaagaa	tggaagccag	aaagattcct	ggacgaaaag	1320
tatgacttga	tggaacttgc	taagactatg	gcttttgggtg	gtggtaaaag	agtttgtgct	1380
ggtgctttac	aagcaatggt	gattgcttgc	acttccatcg	gtagattcgt	tcaagaattt	1440
gaatggaagt	tgatgggtgg	tgaagaagaa	aacggtgata	ctggttgctt	gacctccaa	1500
aaattgcatc	caatgcaagc	cattattaag	gccagagaat	gactcgagcc	gcgg	1554

SEQ ID NO:70

MDMMGIEAVP	FATAVVLGGI	SLVVLIFIRR	FVSNRKRSVE	GLPPVPDIPG	LPLIGNLLQL	60
KEKKPHKTF	RWAETYGPIF	SIRTGASTMI	VLNSSEVAKE	AMVTRFSSIS	TRKLSNALKI	120
LTFDKCMVAT	SDYNDFHKMV	KGFILRNVLG	APAQKRHRCH	RDTLIENISK	YLHAHVKTSP	180
LEPVVLKKIF	ESEIFGLALK	QALGKDIESI	YVEELGTTLS	REEIFAVLVV	DPMAGAIEVD	240
WRDFFPYLSW	IPNKSMEMKI	QRMDFRRGAL	MKALIGEQQK	RIGSGEEKNS	YIDFLLSEAT	300
TLTEKQIAML	IWETIEISD	TTLVTSEWAM	YELAKDPNRQ	EILYREIHKV	CGSNKLTEEN	360
LSKLPYLNVS	FHETLRKYSP	APMVPVRYAH	EDTQLGGYHI	PAGSQIAINI	YGCNMNKKQW	420
ENPEEWKPER	FLDEKYDLMD	LHKTMAFGGG	KRVCAGALQA	MLIACTSIGR	FVQEFEWKLM	480
GGEEENVDTV	ALTSQKLHPM	QAI IKARE				508

SEQ ID NO:71

aagcttaaaa	tgagtaagtc	taatagtatg	aattctacat	cacacgaaac	cctttttcaa	60
caattggtct	tggtttgga	ccgtatgcca	ttgatggatg	ttcactggtt	gatctacggt	120
gctttcggcg	catggttatg	ttcttatgtg	atacatgttt	tatcatcttc	ctctacagta	180
aaagtgccag	ttgttgata	caggtctgta	ttcgaacct	catggttgct	tagacttaga	240
ttcgtctggg	aagtggtctc	tatcataggt	caaggttaca	ataagtttaa	agactctatt	300
ttccaagtta	ggaaattggg	aactgatatt	gtcattatac	cacctaaact	tattgatgaa	360
gtgagaaaat	tgtcacagga	caagactaga	tcagttgaac	ctttcattaa	tgattttgca	420
ggtcaataca	caagagcat	ggttttcttg	caatctgact	tacaaaaccg	tgttatacaa	480
caaagactaa	ctccaaaatt	ggtttccttg	accaaggtca	tgaaggaaga	gttggattat	540
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agtataatgg	tgagattgat	ttccaggatc	tccgccagag	tctttctagg	gcctgaacac	660
tgctgtaacc	aggaatggtt	gactactaca	gcagaatatt	cagaatcact	tttcattaca	720
gggtttatct	taagagttgt	acctcatatc	ttaagaccat	tcatcgcccc	tctattacct	780
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ggagaggaaa	agcaaatcga	taacattgct	cagagaaatg	taattctttc	tttagcatca	960
atccacacta	ctgcatgac	catgacacat	gccatgtacg	atctatgtgc	ttgccctgag	1020
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acagcgtaa	acagatttca	taagttggac	tccttcctaa	aagagtcaca	aagattcaac	1140
ccagtattct	tattgacatt	caatagaatc	taccatcaat	ctatgacctt	atcagatggc	1200
actaacattc	catctggaac	acgtattgct	gttccatcac	acgcaatggt	gcaagattct	1260
gcacatgtcc	caggtccaac	cccacctact	gaatttgatg	gattcagata	tagtaagata	1320
cgttctgata	gtaactacgc	acaaaagtac	ctattctcca	tgaccgattc	ttcaaacatg	1380
gctttcggat	acggcaagta	tgcttgctca	ggtagatttt	acgcgtctaa	tgagatgaaa	1440
ctaacattag	ccattttggt	gctacaattt	gagttcaaac	taccagatgg	taaaggtcgt	1500
cctagaaata	tcactatcga	ttctgatatg	attccagacc	caagagctag	actttgcgtc	1560
agaaaaagat	cacttagaga	tgaatgaccg	cgg			1593

SEQ ID NO:72

MSKSNMNST	SHETLFQQLV	LGLDRMPLMD	VHWLIYVAFG	AWLCSYVIHV	LSSSSTVKVP	60
VVGYRSVFEP	TWLLRLRFVW	EGGSIIIGQY	NKFKDSIFQV	RKLGTDIVII	PPNYIDEVRK	120
LSQDKTRSVE	PFINDFAGQY	TRGMVFLQSD	LQNRVIQQL	TPKLVSLTKV	MKEELDYALT	180
KEMPMKND	WVEVDISSIM	VRLISRISAR	VFLGPEHCRN	QEWLTTTAEY	SESLFITGFI	240

LRVVPHILRP	FIAPLLPSYR	TLLRNVSSGR	RVIGDIIRSQ	QGDGNEDILS	WMRDAATGEE	300
KQIDNIAQRM	LILSLASIHT	TAMTMTHAMY	DLCACPEYIE	PLRDEVKSVV	GASGWDKTAL	360
NRFHKLDSFL	KESQRFNPVF	LLTFNRIYHQ	SMTLSDGTNI	PSGTRIAVPS	HAMLQDSAHV	420
PGPTPTEFD	GFRYSKIRSD	SNYAQKYLFS	MTDSSNMAFG	YGKYACPGRF	YASNEMKLTL	480
AILLLQFEFK	LPDGKGRPRN	ITIDSMDIPD	PRARLCVRKR	SLRDE		525

SEQ ID NO:73

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gccgtgcttc	ctgatgtcat	tgaagagttg	acacttgctg	ttagacagta	cattccaaca	480
gaaggtgatg	aatgggtgtc	cgtaaactgt	tcaaaggccg	caagagatat	tggttctaga	540
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gttccttttg	ttgctccatt	ggtggaggaa	agacgtagac	ttatggaaga	gtacggtgaa	780
gactggtctg	aaaaacctaa	tgatatgtta	cagtgataaa	tgatgaagc	tgcatccaga	840
gatagttcag	tgaaggcaat	cgcagagaga	ttgttaatgg	tgaacttcgc	ggctattcat	900
acctcatcaa	acactatcac	tcatgctttg	taccaccttg	ccgaaatgcc	tgaaactttg	960
caaccactta	gagaagagat	cgaaccatta	gtcaaagagg	agggctggac	caaggctgct	1020
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aacatcgtat	ctttaactag	aatggctgac	aaagatatta	cattgagtga	tggcacattt	1140
ttgccaaaag	gtactctagt	ggccggtcca	gcgtattcta	ctcatagaga	tgatgctgtc	1200
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gaaggtacaa	agcaccagtt	cgtaataact	tcagtcgagt	acgttccatt	tggtcacgga	1320
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agtctataac	cgcg					1515

SEQ ID NO:74

MEDPTVLYAC	LAIAVATFVY	RWYRDPLRSI	PTVGGSDLPI	LSYIGALRWT	RRGREILQEG	60
YDGYRGSTFK	IAMLDRWIVI	ANGPKLADEV	RRRPDEELNF	MDGLGAFVQT	KYTLGEAIHN	120
DPYHVDIIRE	KLTRGLPAVL	PDVIEELTLA	VRQYIPTEGD	EWVSVNCSKA	ARDIVARASN	180
RVFVGLPACR	NQGYLDLAI	FTLSVVKDRA	IINMFPELLK	PIVGRVVGNA	TRNVRRVAVPF	240
VAPLVEERRR	LMEYGEDWS	EKPNMLQWI	MDEAASRDSS	VKAIAERLLM	VNFAAIHTSS	300
NTITHALYHL	AEMPETLQPL	REEIEPLVKE	EGWTKAAMGK	MWWLDSFLRE	SQRYNGINIV	360
SLTRMADKDI	TLSDGTFLPK	GTLVAVPAYS	THRDDAVYAD	ALVFDPPFRFS	RMRAREGEGT	420
KHQFVNTSVE	YVPFGHGKHA	CPGRFFAANE	LKAMLAYIVL	NYDVKLPGDG	KRPLNMYWGP	480
TVLPAPAGQV	LFRRKQVSL					499

SEQ ID NO:75

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ataaagatgg	gttcttcatc	tcttattgta	ttgaacagta	cagaaactgc	taaggaagca	300
atggctcacta	gattttcatc	aatatctacc	agaaaattgt	caaacgccct	aacagttcta	360
acctgcgata	agtctatggt	cgccacttct	gattatgatg	acttccacaa	attagtttaag	420

agatgtttgc	taaatggact	tcttgggtgct	aatgctcaaa	agagaaaaag	acactacaga	480
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gagccagtta	actttagagc	aatttttcgaa	cacgaattgt	ttggtgtagc	attaaagcaa	600
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ttgactaagg	aacagatcgc	aatccttgtc	tgggaaacaa	tcattgaaac	agcagatact	960
accttagtca	caactgaatg	ggccatatac	gagctagcca	aacatccatc	tgtgcaagat	1020
aggttgtgta	aggagatcca	gaacgtgtgt	ggtaggagaga	aattcaagga	agagcagttg	1080
tcacaagttc	cttaccttaa	cggcgttttc	catgaaacct	tgagaaaata	ctcacctgca	1140
ccattagttc	ctattagata	cgccccacgaa	gatacacaaa	tcggtggcta	ccatgttcca	1200
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ttgcataaaa	caatggcttt	cggagctggc	aaaagagtgt	gtgccgggtg	tctacaagcc	1380
tcctaatagg	ctggtatcgc	tattggtaga	ttggtccaag	agttcgaatg	gaaacttaga	1440
gatggtgaag	aggaaaatgt	cgatacttat	gggttaacat	ctcaaaagtt	ataccacta	1500
atggcaatca	tcaatcctag	aagatcctaa				1530

SEQ ID NO:76

MAFFSMISIL	LGFVISSFIF	IFFFKLLSF	SRKNMSEVST	LPSVPVPGF	PVIGNLLQLK	60
EKKPHKTFTR	WSEIYGPIYS	IKMGSSSLIV	LNSTETAKEA	MVTRFSSIST	RKLSNALTVL	120
TCDKSMVATS	DYDDFHKLVK	RCLLNGLLGA	NAQKRKRHYR	DALIENVSSK	LHAHARDHPQ	180
EPVNFRAIFE	HELFGVALKQ	AFGKDVEISY	VKELGVTLK	DEIFKVLVHD	MMEGAIDVDW	240
RDFFPYLKWI	PNKSFEARIQ	QKHKRRLAVM	NALIQDRLKQ	NGSESDDDCY	LNFLMSEAKT	300
LTKEQIAILV	WETIETADT	TLVTTEWAIY	ELAKHPSVQD	RLCKEIQNVC	GGEKFKEEQL	360
SQVPYLNQVF	HETLRKYSPA	PLVPIRYAHE	DTQIGGYHVP	AGSEIAINIY	GCMNDKKRWE	420
RPEDWWPERF	LDDGKYETSD	LHKTMAFGAG	KRVCAGALQA	SLMAGIAIGR	LVQEFWKLK	480
DGEEENVDTY	GLTSQKLYPL	MAIINPRRS				509

SEQ ID NO:77

S. rebaudiana

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aagatgctag	ttgaaaatag	agaattgttg	acactgttca	caacttcctt	cgcagttctt	180
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ggtcatggtg	ttcatgatgc	acagcatcct	tcaagatcta	atgtggcttt	caaaaaggaa	960
ctacacacct	ctcaatcaga	taggtcttgt	actcacttag	aattcgatat	ttctcacaca	1020
ggactgtctt	acgaaactgg	cgatcacggt	ggcgtttatt	ccgagaactt	gtccgaagtt	1080
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cgtaatagaa	aagttgactt	tatctacgag	gacgagctta	acaattttgt	tgagacagga	1860
gcattgtcag	aattgatcgt	cgcatcttca	agagaagggga	ctgccaaga	gtacgttcag	1920
cacaagatga	gtcaaaaagc	ctccgatata	tggaaacttc	taagtgaagg	tgccatctt	1980
tatgtctgtg	gcgatgcaaa	gggcatggcc	aaggatgtcc	atagaactct	gcatacaatt	2040
gttcaggaac	aaggagtgct	ggattcttcc	aaggctgaat	tgtacgtcaa	aaacttacag	2100
atgtctggaa	gatacttaag	agatgtttgg	taa			2133

SEQ ID NO:78

S. rebaudiana

MQSDSVKVSP	FDLVSAMNG	KAMEKLNASE	SEDPTTLPAL	KMLVENRELL	TLFTTSFAVL	60
IGCLVFLMWR	RSSSKLKVQD	PVPQVIVVKK	KEKESEVDDG	KKKVSIFYGT	QTGTAEGFAK	120
ALVEEAKVRY	EKTSFKVIDL	DDYAADDDEY	EEKLKESLA	FFFLATYGDG	EPTDNAANFY	180
KWFTEGDDKG	EWLKKLQYGV	FGLGNRQYEH	FNKIAIVVDD	KLTEMGAKRL	VPVGLGDDQ	240
CIEDDFTAWK	ELVWPELDQL	LRDEDDTSVT	TPYTAAVLEY	RVVYHDKPAD	SYAEDQHTN	300
GHVVHDAQHP	SRSNVAFKKE	LHTSQSDRSC	THLEFDISHT	GLSYETGDHV	GVYSENLSEV	360
VDEALKLLGL	SPDTYFSVHA	DKEDGTFPIGG	ASLPPFPFPC	TLRDALTRYA	DVLSSPKKVA	420
LLALAAHASD	PSEADRLKFL	ASPAGKDEYA	QWIVANQRSL	LEVMSQFPSA	KPPLGVFFAA	480
VAPRLQPRYY	SISSPKMSP	NRIHVTCALV	YETTPAGRIH	RGLCSTWMKN	AVPLTESPDC	540
SQASIFVRTS	NFRLPVPDKV	PVIMIGPGTG	LAPFRGFLQE	RLALKESGTE	LGSSIFFFGC	600
RNRKVDFIYE	DELNNFVETG	ALSELIVAFS	REGTAKEYVQ	HKMSQKASDI	WKLLSEGAYL	660
YVCGDAKGMA	KDVHRTLHTI	VQEQGSLDSS	KAELYVKNLQ	MSGRYLRDVW		710

SEQ ID NO:79

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gcccaatctg	ttatcggttc	ccaaaagtct	ttggttgaag	ttatggctga	attcccatct	1380
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tctagagaag	gtcctaccaa	agaatacgtc	caacataaga	tggtgaaaa	ggcttctgat	1920
atctggaact	tgatttctga	aggtgcttac	ttgtacgttt	gtggtgatgc	taaaggtatg	1980
gctaaggatg	ttcatagaac	cttgcatacc	atcatgcaag	aacaagggtc	tttgatttct	2040
tccaagctg	aatccatggt	caagaacttg	caaatgaatg	gtagatactt	aagagatggt	2100
tggtaa						2106

SEQ ID NO:80

MKVSPFEFMS	AIKGRMDPS	NSSFESTGEV	ASVIFENREL	VAILTTSIAV	MIGCFVVLW	60
RRAGSRKVK	VELPKPLIVH	EPEPEVEDGK	KKVSIFFGTQ	TGTAEGFAKA	LADEAKARYE	120
KATFRVVDLD	DYAADDQYE	EKLKNESFAV	FLLATYGDGE	PTDNAARFYK	WFAEGKERGE	180
WLQNLHYAVF	GLGNRQYEHF	NKIAKVADEL	LEAQGGNRLV	KVGLGDDQD	IEDDFSARE	240
SLWPELDMLL	RDEDDATTVT	TPYTAAVLEY	RVVFHDSADV	AAEDKSWINA	NGHAVHDAQH	300
PFRSNVVRK	ELHTSASDRS	CSHLEFNISG	SALNYETGDH	VGVCENLTE	TVDEALNLLG	360
LSPETYFSIY	TDNEDGTPLG	GSSLPPFFPS	CTLRTALTRY	ADLLNSPKKS	ALLALAAHAS	420
NPVEADRLRY	LASPAKDEY	AQSVIGSQKS	LLEVMAEFPS	AKPPLGVFFA	AVAPRLQPRF	480
YSISSPRMA	PSRIHVTCAL	VYDKMPTGRI	HKGVCSTWMK	NSVPMESHE	CSWAPIFVRQ	540
SNFKLPAESK	VPIIMVPGGT	GLAPFRGFLQ	ERLALKESGV	ELGPSILFFG	CRNRRMDYIY	600
EDELNNFVET	GALSELVIAF	SREGPTKEYV	QHKMAEKASD	IWNLISEGAY	LYVCGDAKGM	660
AKDVHRTLHT	IMQEQGSILDS	SKAESMVKNL	QMNGRYLRDV	W		701

SEQ ID NO:81

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gctgcaggtg	gtgcttccaa	gcctggcaga	actagaaca	tcgtcgaagc	tatggaggaa	180
tcaggtaaaa	actgtgttgt	tttctacggc	agtcaaacag	gtacagcgga	ggattacgca	240
tcaagacttg	caaaggaagg	aaagtccaga	ttcggtttga	acactatgat	cgccgatcta	300
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agatcagcaa	atcaatacca	agtgtgttct	gatttcgtaa	ctttacactg	taaagagaca	2100
acatacgcga	attcagaatt	gcaagaggat	gtctggagtt	aa		2142

SEQ ID NO:82

MAELDTLDIV	VLGVIFLGTV	AYFTKGLWGW	VTKDPYANGF	AAGGASKPGR	TRNIVEAMEE	60
SGKNCVVFYG	SQTGTAEDYA	SRLAKEGKSR	FGLNTMIADL	EDYDFDNLDT	VPSDNIVMFV	120
LATYGEGETP	DNAVDFYEFI	TGEDASFNEG	NDPPLGNLNY	VAFGLGNNTY	EHYNSMVRNV	180
NKALEKLGAAH	RIGEAGEGDD	GAGTMEEDFL	AWKDPMWEAL	AKKMGLEERE	AVYEPIFAIN	240
ERDDLTPKAN	EVYLGEPNKL	HLEGTAKGPF	NSHNPYIAPI	AESYELFSAK	DRNCLHMEID	300
ISGSNLKYET	GDHIAIWPTN	PGEEVNKFLD	ILDLSGKQHS	VVTVKALEPT	AKVPFPNPTT	360
YDAILRYHLE	ICAPVSRQFV	STLAAAFAPND	DIKAEMNRLG	SDKDYFHEKT	GPHYNYNIARF	420
LASVSKGEKW	TKIPFSAFIE	GLTKLQPRYY	SISSSSLVQP	KKISITAVVE	SQQIPGRDDP	480
FRGVATNYLF	ALKQKQNGDP	NPAPFGQSYE	LTGPRNKYDG	IHVPVHVRHS	NFKLPSDPGK	540
PIIMIGPGTG	VAPFRGFVQE	RAKQARDGVE	VGKTLFFFGC	RKSTEDFMYQ	KEWQEYKEAL	600
GDKFEMITAF	SREGSKKVYV	QHRLKERSKE	VSDLLSQKAY	FYVCGDAAHM	AREVNTVLAQ	660
IIAEGRGVSE	AKGEEIVKNM	RSANQYQVCS	DFVTLHCKET	TYANSELQED	VWS	713

SEQ ID NO:83

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gaagtcatgg	aagctttccc	gtcagctaga	ccgccacttg	gtgttttctt	tgacgcggtt	1440
gcaccgcggt	tacagcctcg	ttactactct	atcttcttct	ccccaaagat	ggaaccaaac	1500
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gtatgtggtg	atgctaaagg	catggctaaa	gatgtacacc	gtacacttca	caccattgtg	2040
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SEQ ID NO:84

MQSESVEAST	IDLMTAVLKD	TVIDTANASD	NGDSKMPPAL	AMMFEIRDLL	LILTTSVAVL	60
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ALFEEAKARY	EKAAPKVIDL	DDYAADLDEY	AEKLLKETYA	FFFLATYGDG	EPTDNAAKFY	180
KWFTEGDEKG	VWLQKLQYGV	FGLGNRQYEH	FNKIGIVVDD	GLTEQGAKRI	VPVGLGDDDDQ	240
SIEDDFSQWK	ELVWPELDDL	LRDEDDKAAA	TPYTAAIPEY	RVVFHDKPDA	FSDDHTQTNG	300
HAVHDAQHPC	RSNVAVKKEK	HTPESDRSCT	HLEFDISHTG	LSYETGDHVG	VYCENLIEVV	360
EEAGKLLGLS	TDTYFSLHID	NEDGSPGGP	SLQPPFPFCT	LRKALTNYAD	LLSSPKKSTL	420
LALAAHASDP	TEADRLRFLA	SREGKDEYAE	WVVANQRSLL	EVMEAFPSAR	PPLGVFFAAV	480
APRLQPRYYS	ISSSPKMEPN	RIHVTCALVY	EKTPAGRIHK	GICSTWMKNA	VPLTESQDCS	540
WAPIFVRTSN	FRLPIDPKVP	VIMIGPGTGL	APFRGFLQER	LALKESGTEL	GSSILFFGCR	600
NRKVDYIYEN	ELNNEFVENGA	LSELDVAFSR	DGPTKEYVQH	KMTQKASEIW	NMLSEGAYLY	660
VCGDAKGMMAK	DVHRTLHTIV	QEQGLDSSK	AELYVKNLQM	SGRYLRDWW		709

SEQ ID NO:85

S. rebaudiana

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SEQ ID NO:86

S. rebaudiana

MQSNVSVKISP	LDLVTALFSG	KVLDTSNASE	SGESAMLPTI	AMIMENRELL	MILTTSVAVL	60
IGCVVVLVWR	RSSTKKSAL	PPVIVVVKRV	QEEEVDDGKK	KVTVFFGTQT	GTAEGFAKAL	120
VEEAKARYEK	AVFKVIDLDD	YAADDDEYEE	KLKESLAFF	FLATYGDGEP	TDNAARFYKW	180
FTEGDAKGEW	LNKLQYGVFG	LGNRQYEHFN	KIAKVVDDGL	VEQGAKRLVP	VGLGDDQCI	240
EDDFTAWKEL	VWPELDQLLR	DEDDTTVATP	YTAAVAEYRV	VFHEKPDALS	EDYSYTNNGHA	300
VHDAQHPCRS	NVAVKELHS	PESDRSCTHL	EFDISNTGLS	YETGDHVGVI	CENLSEVND	360
AERLVGLPPD	TYSSIHTDSE	DGSPGGASL	PPFPPTLR	KALTCYADVL	SSPKKSALLA	420
LAAHATDPSE	ADRLKFLASP	AGKDEYSQWI	VASQRSLLLEV	MEAFPSAKPS	LGVFFASVAP	480
RLQPRYYSIS	SSPKMAPDRI	HVTCALVYEK	TPAGRIHKGV	CSTWMKNAV	MTESQDCSWA	540
PIYVRTSNFR	LPSDPKVPVI	MIGPGTGLAP	FRGFLQERLA	LKEAGTDLGL	SILFFGCRNR	600
KVDFIYENEL	NNFVETGALS	ELIVAFSREG	PTKEYVQHKM	SEKASDIWNL	LSEGAYLYVC	660
GDAKGMADV	HRTLHTIVQE	QGSLDSSKAE	LYVKNLQMSG	RYLRDVW		707

SEQ ID NO:87

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SEQ ID NO:88

MSSNSDLVRR	LESVLGVSFG	GSVTDSVVVI	ATTSIALVIG	VLVLLWRRSS	DRSREVKQLA	60
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DYTAEDDKYG	EKLKKETMAF	FMLATYGDGE	PTDNAARFYK	WFTEGTRGV	WLEHLRYGVF	180
GLGNRQYEHF	NKI AKVVDDL	LVEQGA KRLV	TVGLGDDDQC	IEDDFSAWKE	ALWPELDQLL	240
QDDTNTVSTP	YTAVIPEYRV	VIHDP SVTSY	EDPYSNMANG	NASYDIHHP	RANVAVQKEL	300
HKPESDRSCI	HLEFDI FATG	LT YETGDHVG	VYADNCDDTV	E EAAKLLGQP	LDLLFSIHTD	360
NNDGTS LGSS	LPPFP GPCT	LRTALARYAD	LLNPPKKAAL	I ALAAHADEP	SEAERLKFLS	420
SPQGKDEYSK	WV VGSQRSLV	EVMAEFPSAK	PPLGVFFAAV	V PRLQPRYS	I SSSPRFAPH	480
RVHVT CALVY	GPTPTGRIHR	GVCSFWMKNV	VPLEKSQNC	WAPI FIRQSN	FKLPADHSVP	540
IVMVGPGTGL	APFRGFLQER	LALKEEGAQV	GPALLFFGCR	NRQMDFIYEV	ELNNFVEQGA	600
LSELIVAFSR	EGPSKEYVQH	KMVEKAA YMW	NLISQGGYFY	VCGDAKGMAR	DVHRTLHTIV	660
QQEEKVDSTK	AESIVKKLQM	DGRYLRD VW				689

SEQ ID NO:89

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ggtttcgttg	tcttattgtg	gaaaaagacc	acggcagatc	gttccggcga	gctaaagcca	180
cta atgatcc	cta agtctct	gatggcgaaa	gatgaggatg	atgacttaga	tctaggttct	240
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aaagcacttt	cagaagagat	caaagcaaga	tacgaaaagg	cggtctgtaa	agtaatcgat	360
ttggatgatt	acgctgccga	tgatgaccaa	tatgaggaaa	agttgaaaa	ggaaacattg	420
gctttctttt	gtgtagccac	gtatggtgat	ggtgaaccaa	ccgataacgc	cgcaagattc	480
tacaagtggt	ttactgaaga	gaacgaaaga	gatatcaagt	tcgagcaact	tgcttacggc	540
gtttttgcct	taggtaacag	acaatacagag	cactttaaca	agataggtat	tgtcttagat	600
gaagagttat	gcaaaaaggg	tgcgaaagaga	ttgattgaag	tcggttttagg	agatgatgat	660
caatctatcg	aggatgactt	taatgcatgg	aaggaatctt	tgtggtctga	attagataag	720
ttacttaagg	acgaagatga	taaatccggt	gccactccat	acacagccgt	cattccagaa	780
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gcta atggta	atactaccat	cgatattcat	catccatgta	gagtagacgt	tcgagttcaa	900
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catgccgata	aagaggatgg	ctcaccacta	gaaagtgcag	tgctccacc	atttccagga	1140
ccatgcacc	taggtaccgg	tttagctcgt	tacgcggatc	tgttaaatcc	tccacgtaaa	1200
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tctttactag	aagttatggc	tgctttccca	tccgctaaac	ctcctttggg	tgttttcttc	1380
gccgcaatag	cgctagact	gcaaccaaga	tactattcaa	tttcatcctc	acctagactg	1440
gcaccatcaa	gagttcatgt	cacatccgct	ttagtgtacg	gtccaactcc	tactggtaga	1500
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caaggagtta	tttcagagtt	gataatggct	ttttctagag	aaggtgctca	gaaggagtc	1860

gtccaacaca	aatgatgga	aaaggccgca	caagtttggg	acttaatcaa	agaggaaggc	1920
tatctatatg	tctgtggtga	tgcaaagggg	atggcaagag	atgttcacag	aacacttcat	1980
actatagtc	aggaacagga	aggcgtagt	tcttctgaag	cggaagcaat	tgtgaaaaag	2040
ttacaaacag	aggaagata	cttgagagat	gtgtggttaa			2079

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MTSALYASDL	FKQLKSI	MGTDLSLSD	DVVIVL	IATTS	LALVA	GFVLL	LWKK	TADR	SGEL	KP	60	
LMI	PKSL	MAK	DEDD	LDL	GS	GKTR	VSI	FFG	TQT	GTA	E	120
LDDYA	ADDD	Q	YEEK	LK	KETL	AFFC	VAT	YGD	GEPT	DNA	ARF	180
VFAL	GNR	QYE	HFNK	I	GIVLD	EELCK	K	GAKR	LIEV	GL	GDDD	240
LLK	DEDD	KSV	ATPY	T	AVIPE	YRVV	T	HDP	RF	TTQ	KSM	300
KELH	THE	SDR	SCIH	L	FDIS	RTG	I	YET	GD	HVG	VYA	360
HADK	ED	GSPL	ESAV	P	PPFP	PCT	L	G	TGLAR	YAD	LLN	420
HLT	SPD	GKDE	YSQ	W	IVASQR	S	L	L	VMAAFP	SAK	P	480
APSR	VH	V	TSA	LV	GPT	TGR	I	H	K	V	C	540
STPI	V	M	V	G	P	G	T	L	A	P	R	600
QGV	I	S	E	L	I	M	A					660
TIV	Q	E	Q	E	G	V	S	S	E	A	E	692

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A. thaliana

atgtc	tctc	ctt	c	t	c	t	c	t	c	t	c	c	a	g	t	a	c	c	t	c	t	a	t	t	a	t	a	a	a	60
ggt	g	a	a	c	c	a	c	a	g	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	120
g	a	a	t	t	g	t	c	t	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	180
g	c	t	g	t	t	t	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	240
aaa	a	g	a	g	a	g	a	t	c	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	300
g	g	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	360
aa	a	g	c	c	t	t	a	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	420
t	t	g	a	t	g	a	t	g	a	c	c	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	480
g	c	a	t	t	t	t	t	t	t	c	a	a	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	540
t	a	c	a	a	a	t	t	a	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	600
g	t	t	t	c	g	g	t	a	a	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	660
g	a	a	t	t	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	720
ca	a	t	g	t	a	t	a	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	780
a	t	c	t	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	840
t	a	c	a	g	a	g	t	t	t	c	a	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	900
g	g	t	a	a	c	g	g	t	t	a	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	960
a	g	a	a	a	t	a	c	a	c	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1020
g	g	t	c	c	g	g	t	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1080
g	a	a	a	c	t	g	t	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1140
c	a	c	g	c	t	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1200
t	g	t	a	a	c	t	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1260
g	c	c	t	t	g	g	t	t	g	c	c	g	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1320
t	t	a	g	c	a	t	c	t	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1380
t	t	g	t	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1440
g	a	a	g	t	a	g	a	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1500
g	a	a	a	c	t	a	g	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1560
c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1620
t	t	g	t	t	c	t	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1680
a	a	a	g	t	t	c	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1740
c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1800
g	g	t	t	g	t	a	g	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1860
t	c	t	g	g	t	g	c	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1920
g	t	t	c	a	a	c	a	t	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	a	1980

tatttgtacg	tttgcggtga	cgcaaagggg	atggccagag	atgtccatag	atctttgcac	2040
acaattgctc	aagaacaagg	ttccatggat	agtaccaaag	ctgaaggttt	cgtaaagaac	2100
ttacaaactt	ccggtagata	cttgagagat	gtctggtga			2139

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A. thaliana

MSSSSSSSTS	MIDLMAAIK	GEPVIVSDPA	NASAYESVAA	ELSSMLIENR	QFAMIVTTSI	60
AVLIGCIVML	VWRRSGSGNS	KRVEPLKPLV	IKPREEEIDD	GRKKVTIFFG	TQTGTAEGFA	120
KALGEEAKAR	YEKTRFKIVD	LDDYAADDDE	YEEKLKKEDV	AFFFLATYGD	GEPTDNAARF	180
YKWFTEGNDR	GEWLKLNLYG	VFGLGNRQYE	HFNKVAKVVD	DILVEQGAQR	LVQVGLGDDD	240
QCIEDDFTAW	REALWPELDT	ILREEGDTAV	ATPYTAAVLE	YRVSIHDSER	AKFNDDITLAN	300
GNGYTVFDAQ	HPYKANVAVK	RELHTPESDR	SCIHLEFDIA	GSGLTMKLG	HVGVLCNLS	360
ETVDEALRLL	DMSPDYFSL	HAEKEDGTPI	SSSLPPFPFP	CNLRALTRY	ACLLSSPKKS	420
ALVALAAHAS	DPTAERLKH	LASPAGKDEY	SKWVVSQRS	LLEVMAEFPS	AKPPLGVFFA	480
GVAPRLQPRF	YSISSPKIA	ETRIHVTCAL	VYEKMPTGRI	HKGVCSTWMK	NAVPEKSEK	540
LFLGRPIFVR	QSNFKLPSDS	KVPIIMIGPG	TGLAPFRGFL	QERLALVESG	VELGPSVLFF	600
GCRNRRMDFI	YEEELQRFVE	SGALAELSVA	FSREGPTKEY	VQHKMMDKAS	DIWNMISQGA	660
YLYVCGDAKG	MARDVHRS LH	TIAQEQGSMD	STKAEGFVKN	LQTSGRYLRD	VW	712

SEQ ID NO:93

S. rebaudiana

atggaagcct	cttacctata	catttctatt	ttgcttttac	tggcatcata	cctgttcacc	60
actcaactta	gaaggaagag	cgctaactta	ccaccaaccg	tgtttccatc	aataccaatc	120
attggacact	tatacttact	caaaaagcct	ctttatagaa	ctttagcaaa	aattgccgct	180
aagtacggac	caatactgca	attacaactc	ggctacagac	gtgttctggt	gatttctca	240
ccatcagcag	cagaagagtg	ctttaccaat	aacgatgtaa	tcttcgcaa	tagacctag	300
acattgtttg	gcaaaaatag	gggtggaaca	tcccttggca	gtttatccta	cggcgatcaa	360
tggcgtaatc	taaggagagt	agcttctatc	gaaatcctat	cagttcatag	gttgaacgaa	420
tttcatgata	tcagagtgga	tgagaacaga	ttgttaatta	gaaaacttag	aagttcatct	480
tctcctgtta	ctcttataac	agtcttttat	gctctaacat	tgaacgtcat	tatgagaatg	540
atctctggca	aaagatattt	cgacagtggg	gatagagaat	tggaggagga	aggtaagaga	600
tttcgagaaa	tcttagacga	aacgttgctt	ctagccgggtg	cttctaattg	tggcgactac	660
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aaagttagca	aagtagaaa	aacgatgac	gaactcttat	tatctttgca	agagtcagaa	840
cctgagtact	atacagatgc	tatgataaga	tcttttgtcc	taggtctgct	ggctgcaggt	900
agtgatactt	cagcgggcac	tatggaatgg	gccatgagct	tactggtcaa	tcaccacat	960
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gagtcagaca	ttggaaatat	cccttacatc	gggtgtatta	tcaatgaaac	tctaagactc	1080
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agagtaggag	atgagatggt	tgacatgaca	gaaggtttgg	gtgtcacact	tcctaaggcc	1440
gttccattag	ttgccaatg	taagccacgt	tccgaaatga	ctaattctct	atccgaactt	1500
taa						1503

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S. rebaudiana

MEASYLYISI	LLLLASYLFT	TQLRRKSANL	PPTVFPSIPI	IGHLYLLKPP	LYRTLAKIAA	60
KYGPILQLQL	GYRRVLVISS	PSAAEECFN	NDVIFANRPK	TLFGKIVGGT	SLGSLSYGDQ	120
WRNLRRVASI	EILSVHRLNE	FHDIRVDENR	LLIRKLRSSS	SPVTLITVFY	ALTLNVIMRM	180

ISGKRYFDSG	DRELEEEGKR	FREILDETL	LAGASNVGDY	LPILNWLGVK	SLEKKLIALQ	240
KKRDDFFQGL	IEQVRKSRGA	KVGKGRKMTI	ELLLSLQESE	PEYYTDAMIR	SFVLGLLAAG	300
SDTSAGTMEW	AMSLLVNHPH	VLKKAQAEID	RVIGNNRLID	ESDIGNIPYI	GCIINETLRL	360
YPAGPLLFPH	ESSADCVISG	YNI PRGTMLI	VNQWAIHHD	KVWDDPETFK	PERFQGLEGT	420
RDGFKLMPFG	SGRRGCPGEG	LAIRLLGMTL	GSVIQCFDWE	RVGDEMVDMT	EGLGVTLPKA	480
VPLVAKCKPR	SEMTNLLSEL					500

SEQ ID NO:95

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ttgagggagc	aaggccttaa	aggcaattcc	tacaggtttt	tatatggaga	catgaaggag	180
aactctatcc	tgctcaaaaca	agcaagatcc	aaacctatga	acctctccac	ctcccatgac	240
atagcacctc	aagtcacccc	ttttgtcgac	caaaccgtga	aagcttacgg	taagaactct	300
tttaattggg	ttggcccat	accaaggggtg	aacataatga	atccagaaga	tttgaaggac	360
gtcttaacaa	aaaatggtga	ctttgttaag	ccaatatcaa	accacttat	caagttgcta	420
gctacaggta	ttgcaatcta	tgaagggtgag	aaatggacta	aacacagaag	gattatcaac	480
ccaacattcc	attcggagag	gctaaagcgt	atgttacctt	catttcacca	aagttgtaat	540
gagatggcca	aggaatggga	gagcttgggtg	tcaaaagagg	gttcatcatg	tgagttggat	600
gtctggcctt	ttcttgaaaa	tatgtcggca	gatgtgatct	cgagaacagc	atttggaaact	660
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aaaggctttc	aaagttttta	cattccagga	tggagggtttc	tcccaactaa	gatgaacaag	780
aggatgaatg	agattaacga	agaaataaaa	ggattaatca	ggggattat	aattgacaga	840
gagcaaatca	ttaaggcagg	tgaagaaacc	aacgatgact	tattaggtgc	acttatggag	900
tcaaacttga	aggacattcg	ggaacatggg	aaaaacaaca	aaaatggttg	gatgagtatt	960
gaagatgtaa	ttcaggagtg	taagctggtt	tactttgctg	ggcaagaaac	cacttcagtg	1020
ttgctggcct	ggacaatggt	tttacttggg	caaaatcaga	actggcaaga	tcgagcaaga	1080
caagagggtt	tgcaagtctt	tggaaagcagc	aagccagatt	ttgatggtct	agctcacctt	1140
aaagtctgta	ccatgatctt	gcttgaagtt	cttcgattat	accaccagt	cattgaactt	1200
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cagttcaatc	cagagaggtt	ttcggaaagga	gtttccaaag	caacaaagaa	ccgactctca	1380
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gcaaagttgg	ccttagcatt	gatcttgcaa	cacttcacct	ttgagctttc	tccatctcat	1500
gcacatgctc	cttcccacg	tataaccctt	caaccacagt	atggtgttcg	tatcatttta	1560
catcgacgtt	ag					1572

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R. suavis

atggaagtca	ctgtcgcctc	ttctgtcgtc	ttatccttag	tcttcatttc	cattgtcgtc	60
agatgggctt	ggccggtgt	caactgggtt	tggttcaaac	caaagaagtt	ggaaagattc	120
ttgagagagc	aaggtttgaa	gggtaattct	tatagattct	tgtacggtga	catgaaggaa	180
aattctatct	tgttgaagca	agccagatcc	aaaccaatga	acttgtctac	ctctcatgat	240
attgctccac	aagttaactcc	attcgtcgat	caaactgtta	aagcctacgg	taagaactct	300
ttcaattggg	ttggtccaat	tcctagagtt	aacatcatga	accagaaga	tttgaaggat	360
gtcttgacca	agaacgttga	cttcgttaag	ccaatttcca	accattgat	taaattgttg	420
gctactggta	ttgccaattta	cgaaggtgaa	aagtggacta	agcatagaag	aatcatcaac	480
cctacccttc	actctgaaaag	attgaagaga	atgttaccat	ctttccatca	atcctgtaat	540
gaaatgggta	aggaatggga	atccttgggt	tctaaagaag	gttcttcttg	cgaattggat	600
gtttgcccat	tcttgaaaaa	tatgtctgct	gatgtcattt	ccagaaccgc	tttcggtacc	660
tcctacaaga	agggtaaaaa	gattttcgaa	ttggtgagag	agcaagttat	ttacgttacc	720
aagggtttcc	aatccttcta	catcccaggt	tggagattct	tgccaactaa	aatgaacaag	780
cgtatgaacg	agatcaacga	agaaattaa	ggtttgatca	gaggtattat	tatcgacaga	840
gaacaaatta	ttaaagctgg	tgaagaaacc	aacgatgatt	tgttgggtgc	tttgatggag	900
tccaacttga	aggaatttag	agaacatggt	aagaacaaca	agaatggttg	tatgtctatt	960

gaagatgtta	ttcaagaatg	taagttatct	tacttcgctg	gtcaagagac	cacttctggt	1020
ttgtagcct	ggactatggt	cttgtagggt	caaaaccaa	attggcaaga	tagagctaga	1080
caagaagttt	tgcaagtctt	cggttcttcc	aagccagact	ttgatgggtt	ggcccacttg	1140
aaggttgta	ctatgatgtt	gtagaaggt	ttgagattgt	accaccagt	cattgagtta	1200
atcagaacca	ttcataaaaa	gactcaattg	ggtaaattat	ctttgccaga	aggtggtgaa	1260
gtcagattac	caaccttggt	gattcaccac	gataaggaat	tatgggggtga	cgacgcta	1320
caatttaatc	cagaaagatt	ttccgaaggt	gtttccaagg	ctaccaaaaa	ccgtttgtcc	1380
ttcttcccat	ttggtgctgg	tccacgtatt	tgtatcggtc	aaaacttttc	catgatggaa	1440
gccaagttgg	ctttggcttt	aatcttgcaa	cacttcactt	tccaattgtc	tccatcccat	1500
gccacgctc	cttctcatag	aatcacttta	caaccacaat	acgggtgtcag	aatcatctta	1560
cacagaagat	aa					1572

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R. suavis

MEVTVASSVA	LSLVFISIVV	RWAWSVVNWV	WFKPKLERF	LREQGLKGN	YRFLYGDMKE	60
NSILLKQARS	KPMNLSTSHD	IAPQVTFPVD	QTVKAYGKNS	FNWVGPIPRV	NIMNPEDLKD	120
VLTKNVDFVK	PISNPLIKLL	ATGIAIYEGE	KWTKHRRIN	PTFHSERLKR	MLPSFHQSCN	180
EMVKEWESLV	SKEGSSCELD	VWPFLENMSA	DVISRTAFGT	SYKKGQKIFE	LLREQVIYVT	240
KGFQSFYIPG	WRFLPTKMNK	RMNEINEEIK	GLIRGIIDR	EQIIKAGEET	NDDLGLALME	300
SNLKDIREHG	KNNKNVMSI	EDVIOECKLF	YFAGQETTSV	LLAWTMVLLG	QNQNWQDRAR	360
QEVLQVFGSS	KPDFDGLAHL	KVVTMILLEV	LRLYPPVIEL	IRTIHKKTQL	GKLSLPEGVE	420
VRLPTLLIHH	DKELWGDDAN	QFNPERFSEG	VSKATKNRLS	FFPFGAGPRI	CIGQNFMSME	480
AKLALALILQ	HFTFELSPSH	AHAPSHRITL	QPQYGVRIIL	HRR		523

SEQ ID NO:98

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ttgagggagc	aaggccttac	aggcaattct	tacaggcttt	tgtttgaga	caccaaggat	180
ctctcgaaga	tgctggaaca	aacacaatcc	aaaccatca	aactctccac	ctcccatgat	240
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gccttcaaca	gacatgatga	ttttcataag	acagtaaaaa	atcctatcat	gaagtctcca	420
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gagatgatta	acaaatggga	gagcttggtg	tccaaagaga	gttcatgtga	gttggatgtg	600
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tatgaagagg	gaaggaaaat	atttcaacta	ctaagagagg	aagcaaaagt	ttattcggta	720
gctctacgaa	gtgtttacat	tccaggatgg	aggtttctac	caaccaagca	gaacaagaag	780
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accactcaca	agaaaacaca	gcttggaaaa	ttatcattac	cagctggagt	ggaagtctcc	1260
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aagccagaga	ggttttcaga	gggagtttca	aaggcaacaa	agaacaaatt	tacatactta	1380
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ttggccttgg	ccctgatgtt	acaacacttt	gcctttgagc	tttctccatc	ctatgctcat	1500
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cggtga						1566

SEQ ID NO:99

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gctttcaaca	gacatgatga	ttccataag	accgtcaaga	accaattat	gaagtctcca	420
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gaaatgatta	acaagtggga	atccttgggt	tccaaagaat	cttctgtga	attggatgct	600
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tacgaagaag	gtagaaagat	cttccaatta	ttgagagaag	aagccaaggt	ttactccggt	720
gctttgagat	ctgtttacat	tccaggttgg	agattcttgc	caactaagca	aaacaaaaag	780
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gaagctatga	aggctggtga	agctacaaaa	gatgatttgt	tggttatctt	gatggaatcc	900
aacttcagag	aatccaaga	acacggtaac	aacaagaatg	ccggtatgct	tattgaagat	960
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agataac						1567

SEQ ID NO:100

MEASRASCVA	LCVVVWSIVI	TLAWRVLNWV	WLRPKKLERC	LREQGLTGNS	YRLLFGDTKD	60
LSKMLEQTQS	KPIKLSHSD	IAPRVTPFFH	RTVNSNGKNS	FVWVGPIPRV	HIMNPEDLKD	120
AFNRHDDFHK	TVKNPIMKSP	PPGIVGIEGE	QWAKHRKIIN	PAFHLEKLGK	MVPIFYQSCS	180
EMINKWESLV	SKESSCELDV	WPYLENFTSD	VISRAAFGSS	YEEGRKIFQL	LREEAKVYSV	240
ALRSVYIPGW	RFLPTKQNKK	TKEIHNEIKG	LLKGIINKRE	EAMKAGEATK	DDLLGILMES	300
NFREIQEHGN	NKNAGMSIED	VIGECKLFYF	AGQETTSVLL	VWTMILLSQN	QDWQARAREE	360
VLKVFSGSNIP	TYEELSHLKV	VTMILLEVLR	LYPSVVALPR	TTHKKTQLGK	LSLPAGVEVS	420
LPILLVHHDK	ELWGEDANEF	KPERFSEGV	KATKNKFTYL	PFGGGPRICI	QONFAMVEAK	480
LALALILQHF	AFELSPSYAH	APSAVITLQP	QFGAHIILHK	R		521

SEQ ID NO:101

ASWVAVLSV	WVSMVIAWAW	RVLNWWLWRP	KKLEKCLREQ	GLAGNSYRLL	FGDTKDLSKM	60
LEQTQSKPIK	LSTSHDIAPH	VTPFFHQTVN	SYGKNSFVWM	GPIPRVHIMN	PEDLKDTFNR	120
HDDDFHKVKN	PIMKSLPQGI	VGIEGEQWAK	HRKIINPAFH	LEKLGKMPVI	FYRSCSEMIN	180
KWESLVSKES	SCELDVWPYL	ENFTSDVISR	AAFSSYEEG	RKIFQLLREE	AKIYTVAMRS	240
VYIPGWRFLP	TKQNKKAKEI	HNEIKGLLKG	IINKREEAMK	AGEATKDDLL	GILMESNFRE	300
IQEHGNNKNA	GMSIEDVIGE	CKLFYFAGQE	TTSVLLVWTM	VLLSQNDWQ	ARAREEVLQV	360
FGSNIPTYEE	LSQLKVVTMI	LLEVLRLYPS	VVALPRTHK	KTQLGKLSLP	AGVEVSLPIL	420
LVHHDKELWG	EDANEFKPER	FSEGVSKATK	NQFTYFPFGG	GPRICIGQNF	AMMEAKLALS	480
LILRHFALEL	SPLYAHAPSV	TITLQPQYGA	HIILHKR			517

SEQ ID NO:102

MEASRPSCVA	LSVVLVSIVI	AWAWRVLNWV	WLRPNKLERC	LREQGLTGNS	YRLLFGDTKE	60
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ISMVVEQAQS	KPIKLSSTHD	IAPRVIPFSH	QIVYTYGRNS	FVWMGPTPRV	TIMNPEDLKD	120
AFNKSDEFQR	AISNPVKS	SQGLSSLEGE	KWAKHRKIIN	PAFHLEKLGK	MLPTFYQSCS	180
EMINKWESLV	FKEGSREMDV	WPYLENLTS	VISRAAFGSS	YEEGRKIFQL	LREEAKFYTI	240
AARSVYIPGW	RFLPTKQNK	MKEIHKEVRG	LLKGIINKRE	DAIKAGEAAK	GNULLGILMES	300
NFREIQEHGN	NKNAGMSIED	VIGECKLFYF	AGQETTSVLL	VWTLVLLSQN	QDWQARAREE	360
VLQVFGTNI	TYDQLSHLKV	VTMILLEVLR	LYPAVVELPR	TTYKKTQLGK	FLLPAGVEVS	420
LHIMLAHHDK	ELWGEDAKEF	KPERFSEGV	KATKNQFTYF	PFAGPRICI	GQNFAMLEAK	480
LALSILQHF	TFELSPSYAH	APSVTITLHP	QFGAHFILHK	R		521

SEQ ID NO:103

CVALSVLV	IVIAWAWRVL	NVWVLRPNKL	ERCLREQGLT	GNSYRLLFGD	TKEISMVVEQ	60
AQSKPIKLST	THDIAPRVIP	FSHQIVYTYG	RNSFVWMGPT	PRVTIMNPED	LKDAFNKSDE	120
FQRAISNPIV	KSISQGLSSL	EGEKWAKHRK	IINPAFHLEK	LKGMMLPTFYQ	SCSEMINKWE	180
SLVFKEGSRE	MDVWPYLENL	TSDVISRAAF	GSSYEGRKI	FQLLREEAKF	YTIAARSVYI	240
PGWRFLPTKQ	NKRMKEIHKE	VRGLLKGIIN	KREDAIKAGE	AAKGNLLGIL	MESNFREIQE	300
HGNNKNAGMS	IEDVIGECKL	FYFAGQETTS	VLLVWTLVLL	SONQDWQARA	REEVLQVFGT	360
NIPTYDQLSH	LKVVTMILLE	VLRLYPVAVE	LPRTTYKKTQ	LGKFLLPAGV	EVSLHIMLAH	420
HDKELWGEDA	KEFKPERFSE	GVSKATKNQF	TYFPFAGGPR	ICIGQNFAML	EAKLALSIL	480
QHFTFELSPS	YAHAPSVTIT	LHPQFGAHFI	LHKR			514

SEQ ID NO:104

MGPIPRVHIM	NPEDLKDTFN	RHDDFHKVVK	NPIMKSLPQG	IVGIEGDQWA	KHRKIINPAF	60
HLEKLGKGMVP	IFYQSCSEMI	NIWKSLSKE	SSCELDVWPY	LENFTSDVIS	RAAFGSSYEE	120
GRKIFQLLRE	EAKVYTVAVR	SVYIPGWRFL	PTKQNKKTKE	IHNEIKGLLK	GIINKKREEM	180
KAGEATKDDL	LGI LMESNFR	EIQEHGNKN	AGMSIEDVIG	ECKLFYFAGQ	ETTSVLLVWT	240
MVLLSQNDW	QARAREEVQ	VFGSNIPTYE	ELSHLKVVTM	ILLEVLRLYP	SVVALPRTH	300
KKTQLGKLSL	PAGVEVSLPI	LLVHHDKELW	GEDANFKPE	RFSEGVSKAT	KNQFTYFPFG	360
GGPRICIGQN	FAMMEAKLAL	SLILQHFTE	LSPQYSHAPS	VTITLQPQYG	AHLILHKR	418

SEQ ID NO:105

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caagctaacg	atgacaaaaa	ggaggatatg	gatttcatgg	atatcatgat	ctccatgaca	900
gaagcaaatt	caccacttga	aggatacggc	actgatacta	ttatcaagac	cacatgtatg	960
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cgtgttaaat	ggtcctaa					1578

SEQ ID NO:106

MGLFPLEDSY	ALVFEGLAIT	LALYYLLSFI	YKTSKKTCTP	PKASGEHPIT	GHLNLLSGSS	60
GLPHLALASL	ADRCGPIFTI	RLGIRRVLVV	SNWEIAKEIF	TTHDLIVSNR	PKYLAAKILG	120
FNYVSFSFAP	YGPYWVGIRK	I IATKLMSSS	RLQKLQFVRV	FELENSMKSI	RESWKEKKDE	180
EGKVLVEMKK	WFWELNMNIV	LRTVAGKQYT	GTVDDADAKR	ISELFREWFH	YTRGFVVGDA	240
FPFLGWLDLG	GYKKTMEELVA	SRLDSMVSKW	LDEHRKKQAN	DDKKEDMDFM	DIMISMTEAN	300
SPLEGYGTDI	I IKTTCMTLI	VSGVDTTSIV	LTWALSLLL	NRDTLKKAQE	ELDMCVGKGR	360
QVNESDLVNL	IYLEAVLKEA	LRLYPAAFLG	GPRAFLEDCT	VAGYRIPKGT	CLLINMWKLNH	420
RDPNIWSDPC	EFKPERFLT	NQKDVVDVIGM	DFELIPFGAG	RRYCPGTRLA	LQMLHIVLAT	480
LLQNFEMSTP	NDAPVDMTAS	VGMTNAKASP	LEVLLSPRVK	WS		522

SEQ ID NO:107

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gcgttggttg	atatcgacta	tgctggttac	actatcccaa	aaggatggaa	gttgcattgg	1140
tcagctgttt	ctactcaaag	agacgaagcc	aatttcgaag	atgtaactag	attcgatcca	1200
tccagatttg	aaggggcagg	ccctactcca	ttcacatttg	tgcttttcgg	tggaggctct	1260
agaatgtgtt	taggcaaaga	gtttgccagg	ttagaagtgt	tagcatttct	ccacaacatt	1320
gttaccact	ttaagtggga	tcttctaate	cctgatgaga	agatcgaata	tgatccaatg	1380
gctactccag	ctaagggtct	gccaaattaga	cttcatccac	accaagtcta	a	1431

SEQ ID NO:108

MIQVLTPILL	FLIFFVFWKV	YKHQTKINL	PPGSFGWPF	GETLALLRAG	WDSEPERFVR	60
ERIKKHGSPL	VFKTSLFGDR	FAVLCGPAGN	KFLFCNENKL	VASWWPVPVR	KLFGKSLITI	120
RGDEAKWMRK	MLLSYLGPD	FATHYAVTMD	VVTRRHIDVH	WRGKEEVNMF	QTVKLYAFEL	180
ACRLFMNLDD	PNHIAKLGSL	FNIFLKGIEE	LPIDVPGTRF	YSSKAAAAI	RIELKLIKA	240
RKLELKEGKA	SSSQDLLSHL	LTSPDENGFM	LTEEEIVDNI	LLLLFAGHDT	SALSITLLMK	300
TLGEHSDVYD	KVLKEQLEIS	KTKEAWESLK	WEDIQMKYS	WSVICEVMRL	NPPVIGTYRE	360
ALVDIDYAGY	TIPKGWKLHW	SAVSTQRDEA	NFEDVTRFDP	SRFEGAGPTP	FTFVFPFGGGP	420
RMCLGKEFAR	LEVLAFLHNI	VTNFKWDLII	PDEKIEYDPM	ATPAKGLPIR	LHPHQV	476

SEQ ID NO:109

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gagctatctc	agactaacac	attgaacttg	ggtagaatca	cccatataac	caaaagattg	420
aatcctatct	taggtaacgg	aatcataacc	tctaattggtc	ctcattgggc	ccatcagcgt	480
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tatcctcaaa	gttacattcc	atgttggtctg	ggtcctagaa	catgcgttgg	taaaaacttt	1440
ggcatgatgg	aagtaaaggt	tcttgtttcc	ctgattgtct	ccaagttctc	tttactctca	1500
tctcctacct	accaacatag	tcctagtcac	aaacttttag	tagaaccaca	acatggggtg	1560
gtaattagag	tggtttaa					1578

SEQ ID NO:110

MESLVVHTVN	AIWCIVIVGI	FSVGYHVIYGR	AVVEQWRMRR	SLKLQGVKGP	PPSIFNGNVS	60
EMQRIQSEAK	HCSGDNIIISH	DYSSSLFPHF	DHWRKQYGRI	YTYSTGLKQH	LYINHPPEMVK	120
ELSQTNTLNL	GRITHITKRL	NPILGNGIIT	SNGPHWAHQ	RIIAYEFTHD	KIKGMVGLMV	180
ESAMPMLNKW	EEMVKRGGEM	GCDIRVDEDL	KDVSADVIAK	ACFGSSFSKG	KAIFSMIRDL	240
LTAITKRSVL	FRFNGFTDMV	FGSKKHGDVD	IDALEMELES	SIWETVKERE	IECKDTHKKD	300
LMQLILEGAM	RSCDGNLWDK	SAYRRFVVDN	CKSIYFAGHD	STAVSVSWCL	MLLALNPSWQ	360
VKIRDEILSS	CKNGIPDAES	IPNLKTVTMV	IQETMRLYPP	APIVGREASK	DIRLGDLVVP	420
KGVCIWTLIP	ALHRDPEIWG	PDANDFKPER	FSEGISKACK	YPQSYIPFGL	GPRTCVGKNF	480
GMMEVKVLVS	LIVSKFSEFL	SPTYQHSPSH	KLLVEPQHG	VIRVV		525

SEQ ID NO:111

atgtacttcc	tactacaata	cctcaacatc	acaaccgttg	gtgtctttgc	cacattgttt	60
ctctcttatt	gtttacttct	ctggagaagt	agagcgggta	acaaaaagat	tgccccagaa	120
gctgccgctg	catggcctat	tatcgccac	ctccacttac	ttgcaggtgg	atccccatcaa	180
ctaccacata	ttacattggg	taacatggca	gataagtacg	gtcctgtatt	cacaatcaga	240
ataggcttgc	atagagctgt	agttgtctca	tcttgggaaa	tgccaaagga	atggtcaaca	300
gctaatgatac	aagtgtcttc	ttcaagacct	gaactattag	cttctaagtt	gttgggttat	360
aactacgcca	tgtttggttt	ttcacatac	ggttcatact	ggagagaaat	gagaaagatc	420
atctctctcg	aattactatc	taattccaga	ttggaactat	tgaaagatgt	tagagcctca	480
gaagttgtca	catctattaa	ggaactatac	aaattgtggg	cggaaaagaa	gaatgagtca	540
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agaatgggtg	ctggtaaaag	atacttctcc	gcgagtgcag	cttcagaaaa	caaacaggcc	660
cagcgttgta	gaagagtctt	cagagaaatc	ttccatctct	ccggcttggt	tgtgggtgct	720
gatgctatac	cttttcttgg	atggctcgat	tggggaagac	acgagaagac	cttgaaaaag	780
accgccatag	aatggattc	catcgccag	gagtggttgg	aggaacatag	acgtagaaaa	840

gattctggag	atgataattc	tacccaagat	ttcatggacg	ttatgcaatc	tgtgctagat	900
ggcaaaaatc	taggcggata	cgatgctgat	acgattaaca	aggctacatg	cttaactctt	960
atatcagggtg	gcagtgatac	tactgtagtt	tctttgacat	gggctccttag	tcttgtgtta	1020
aacaatagag	atactttgaa	aaaggcacag	gaagagttag	acatccaagt	cggtaaggaa	1080
agattgggta	acgagcaaga	catcagtaag	ttagtttact	tgcaagcaat	agtaaaagag	1140
acactcagac	tttatccacc	aggtcctttg	ggtgggttga	gacaattcac	tgaagattgt	1200
acactaggtg	gctatcacgt	ttcaaaagga	actagattaa	tcatgaactt	atccaagatt	1260
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actcataaag	atgtcgatcc	acgtggtaaa	cactttgaat	tattccatt	cgggtgcagga	1380
agacgtgcat	gtcctggtat	cacattcggg	ttacaagtac	tacatctaac	attggcatct	1440
ttcttgcag	cgtttgaatt	ttcaacacca	tcaaatgagc	aggttaacat	gagagaatca	1500
ttaggtctta	cgaatatgaa	atctacccca	ttagaagttt	tgatttctcc	aagactatcc	1560
cttaattgct	tcaaccttat	gaaaatttga				1590

SEQ ID NO:112

MYFLLQYLNI	TTVGVFATLF	LSYCLLLWRS	RAGNKKIAPE	AAAAPWIIGH	LHLLAGGSHQ	60
LPHITLGNMA	DKYGPVFTIR	IGLHRAVVVS	SWEMAKECST	ANDQVSSSRP	ELLASKLLGY	120
NYAMFGFSPY	GSYWREMRKI	ISLELLSNSR	LELLKDVRAS	EVVTSIKELY	KLWAEKKNES	180
GLVSVEMKQW	FGDLTLNVIL	RMVAGKRYFS	ASDASENKQA	QRCRRVFREF	FHLSGLFVVA	240
DAIPFLGWLD	WGRHEKTLKK	TAIEMDSIAQ	EWLEEHRRRK	DSGDDNSTQD	FMDVMQSVLD	300
GKNLGGYDAD	TINKATCLTL	ISGGSDTTVV	SLTWALSLVL	NNRDTLKKAQ	EELDIQVGKE	360
RLVNEQDISK	LVYLQAIVKE	TLRLYPPGPL	GGLRQFTEDC	TLGGYHVSKG	TRLIMNLSKI	420
QKDPRIWSDP	TEFQPERFLT	THKDVDPRGK	HFEFIPFGAG	RRACPGITFG	LQVLHLTLAS	480
FLHAFEFSTP	SNEQVNMRES	LGLTNMKSTP	LEVLISPRLS	SCSLYN		526

SEQ ID NO:113

atggaacct	acttttactt	gtcattacta	ttgttgctcg	tgaccttcat	ttctttaagt	60
ctgtttttca	tcttttacia	acaaaagtcc	ccattgaatt	tgccaccagg	gaaaatgggt	120
taccctatca	taggtgaaag	tttagaattc	ctatccacag	gctggaagg	acatcctgaa	180
aagttcatat	ttgatagaat	gcgtaagtac	agtagtgagt	tattcaagac	ttctattgta	240
ggcgaatcca	cagttgtttg	ctgtggggca	gctagtaaca	aattcctatt	ctctaacgaa	300
aacaaactgg	taactgcctg	gtggccagat	tctgttaaca	aaatcttccc	aacacttca	360
ctggattcta	attgaaagga	ggaatctata	aagatgagaa	agttgctgcc	acagttcttc	420
aaaccagaag	cacttcaaaag	atacgtcggc	gttatggatg	taatcgca	aagacatttt	480
gtcactcact	gggacaacaa	aaatgagatc	acagtttata	cacttgctaa	aagatacact	540
ttcttgcttg	cggtgtagact	gttcatgtct	gttgaggatg	aaaatcatgt	ggcgaaattc	600
tcagacccat	tccaactaat	cgctgcaggc	atcatttcac	ttcctatcga	tcttctgggt	660
actccattca	acaaggccat	aaaggcttca	aatttcatta	gaaaagagct	gataaagatt	720
atcaaacaaa	gacgtgttga	tctggcagag	ggtacagcat	ctccaacca	ggatatcttg	780
tcacatatgc	tattaacatc	tgatgaaaac	ggtaaactca	tgaacgagtt	gaacattgcc	840
gacaagattc	ttggactatt	gataggaggc	cacgatacag	cttcagtagc	ttgcacattt	900
ctagtgaagt	acttaggaga	attaccacat	atctacgata	aagtctacca	agagcaaatg	960
gaaattgcc	agtccaaacc	tgctggggaa	ttgttgaatt	gggatgactt	gaaaaagatg	1020
aagtattcat	ggaatgtggc	atgtgaggta	atgagattgt	caccaccttt	acaagggtgg	1080
tttagagagg	ctataactga	ctttatgttt	aacggtttct	ctattccaaa	agggtggaag	1140
ttatactggt	ccgccaactc	tacacacaaa	aatgcagaat	gtttcccaat	gcctgagaaa	1200
ttcgatccta	ccagatttga	aggtaatggg	ccagcgcctt	atacatttgt	accattcggt	1260
ggaggcccta	gaatgtgtcc	tggaaaggaa	tacgctagat	tagaaatctt	ggttttcatg	1320
cataatctgg	tcaaactgtt	taagtgggaa	aaggttattc	cagacgaaaa	gattattgtc	1380
gatccattcc	caatcccagc	taaagatctt	ccaatccggt	tgtatcctca	caaagcttaa	1440

SEQ ID NO:114

MEPNFYLSLL	LLFVTFISLS	LFFIFYKQKS	PLNLPPGKMG	YPIIGESLEF	LSTGWKGHPE	60
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KFIFDRMRKY	SSELFKTSIV	GESTVVCCGA	ASNKFLFSNE	NKLVTAWWPD	SVNKIFPTTS	120
LDSNLKEESI	KMRKLLPQFF	KPEALQRYVG	VMDVIAQRHF	VTHWDNKNEI	TVYPLAKRYT	180
FLLACRLFMS	VEDENHVAKF	SDPFQLIAAG	IISLPIDLPG	TPFNKAIKAS	NFIRKELIKI	240
IKQRRVDLAE	GTASPTQDIL	SHMLLTSDEN	GKSMNELNIA	DKILGLLIGG	HDTASVACTF	300
LVKYLGELPH	IYDKVYQEQM	EIAKSKPAGE	LLNWDDLKKM	KYSWNVACEV	MRLSPPLQGG	360
FREAITDFMF	NGFSIPKGWK	LYWSANSTHK	NAECFPMPEK	FDPTRFEGNG	PAPYTFVFPFG	420
GGPRMC PGKE	YARLEILVFM	HNLVKRFKWE	KVIPDEKIV	DPFPIPAKDL	PIRLYPHKA	479

SEQ ID NO:115

atggcctctg	ttactttggg	ttcctggatc	gtcgtccacc	accataacca	tcaccatcca	60
tcactctatcc	taactaaatc	tcgttcaaga	tcctgtccta	ttacactaac	caaaccaatc	120
tcttttcggt	caaagagaac	agtttcctct	agtagttcta	tcgtgtcctc	tagtgcgctc	180
actaaggaag	acaatctgag	acagtctgaa	ccttcttctc	ttgatttcat	gtcatatatac	240
attactaagg	cagaactagt	gaataaggct	cttgattcag	cagttccatt	aagagagcca	300
ttgaaaatcc	atgaagcaat	gagatactct	cttctagctg	gcgggaagag	agtcagacct	360
gtactctgca	tagcagcgtg	cgaattagtt	ggtagcagag	aatcaaccgc	tatgcctgcc	420
gcttgctgctg	tagaaaatgat	tcatacaatg	tcactgatac	acgatgattt	gccatgtatg	480
gataacgatg	atctgagaag	gggtaagcca	actaaccata	agggttttcgg	cgaagatggt	540
gccgtcttag	ctggtgatgc	tttgttatct	ttcgcgcttcg	aacatttggc	atccgcaaca	600
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attggaactg	agggtttagt	tgcaggctcaa	gtcgtcgata	tctcttccga	aggctttgat	720
ttgaatgatg	taggtcttga	acatctcgaa	ttcatccatc	ttcacaagac	agctgcactt	780
ttagaagcca	gtgcggttct	cggcgcaatt	gtagcgggag	ggagtgatga	cgaaattgag	840
agattgagga	agtttgctag	atgtatagga	ttactgttcc	aagtagtaga	cgatatacta	900
gatgtgacaa	agtcttccaa	agagttggga	aaaacagctg	gtaaagattt	gattgccgac	960
aaattgacct	accctaagat	tatggggcta	gaaaaatcaa	gagaatttgc	cgagaaactc	1020
aatagagagg	cgctgatca	actggtgggt	ttcgattctg	ataaagttgc	accactctta	1080
gccttagcca	actacatcgc	ttacagacaa	aactaa			1116

SEQ ID NO:116

MASVTLGSKI	VVHHHNNHHH	SSILTKSRSR	SCPITLTKPI	SFRSKRTVSS	SSSIVSSSVV	60
TKEDNLRQSE	PSSFDFMSYI	ITKAEVLNKA	LDSAVPLREP	LKIHEAMRYS	LLAGGKRVRP	120
VLCIAACELV	GGEESTAMPA	ACAVEMIHTM	SLIHDDLPCM	DNDDLRRGKP	TNHKVFGEDEV	180
AVLAGDALLS	FAFEHLASAT	SSDVVSPVRV	VRAVDELAKA	IGTEGLVAGQ	VVDISSEGLD	240
LNDVGLHLE	FIHLHKTAAL	LEASAVLGAI	VGGGSDDEIE	RLRKFARCIG	LLFQVDDIL	300
DVTKSSKELG	KTAGKDLIAD	KLTYPKIMGL	EKSREFAEKL	NREARDQLLG	FDSKVAPLL	360
ALANYIAYRQ	N					371

SEQ ID NO:117

R. suavis

MATLLEHFQA	MPFAIPIALA	ALSWLFLFYI	KVSFFSNKSA	QAKLPPVPV	PGLPVIIGNLL	60
QLKEKKPYQT	FTRWAEYGP	IYSIRTGAST	MVVLNTTQVA	KEAMVTRYLS	ISTRKLSNAL	120
KILTADKCMV	AISDYDNFHK	MIKRYILSNV	LGPSAQKRHR	SNRDTLRANV	CSRLHSQVKN	180
SPREAVNFR	VFEWELFGIA	LKQAFGKDIE	KPIYVEELGT	TLRDEIFKV	LVLDMEGAI	240
EVDWRDFFPY	LRWIPNTRME	TKIQRLYFR	KAVMTALINE	QKKRIASGEE	INCYIDFLLK	300
EGKTLTMDQI	SMLLWETVIE	TADTTMVTTE	WAMYEVAKDS	KRQDRLYQEI	QKVCGSEMVT	360
EEYLSQLPYL	NAVFHETLRK	HSPAALVPLR	YAHEDTQLGG	YYIPAGTEIA	INIYGCNMDK	420
HQWESPEEWK	PERFLDPKFD	PMDLYKTMAF	GAGKRVCAGS	LQAMLIACPT	IGRLVQEFEW	480
KLRDGEENV	DTVGLTTHKR	YPMHAILKPR	S			511

SEQ ID NO:118

S. cerevisiae

atgtcatttc	aaattgaaac	ggttcccacc	aaaccatatag	aagacaaaa	gcctggtacc	60
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tctggtttgc	gtaagaagac	aaaggtgttt	aaagacgaac	ctaactacac	agaaaatttc	120
attcaatcga	tcatggaagc	tattccagag	ggttctaaag	gtgccactct	tgttgctcgg	180
ggtgatgggc	gttactacaa	tgatgtcatt	cttcataaga	ttgccgctat	cggtgctgcc	240
aacggtatta	aaaagttagt	tattggccag	catggtcttc	tgtctacgcc	agccgcttct	300
cacatcatga	gaacctacga	ggaaaaatgt	actggtggta	ttatcttaac	cgccctacat	360
aatccaggtg	gtccagaaaa	tgacatgggt	attaagtata	acttatccaa	tgggggtcct	420
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gaatttggtt	taccggcgga	tgaggtttta	caaaactggc	atccttctcc	ggattttggt	780
ggtatgcatc	cagatccaaa	cttaacttat	gccagttcgt	tagtgaaaag	agtagatcgt	840
gaaaagattg	agtttgggtg	tgcatccgat	ggtgatgggtg	atagaaatat	gatttacgg	900
tacggcccat	ctttcgtttc	tccaggtgac	tccgtcgcaa	ttattgccga	atatgcagct	960
gaaatcccat	atttcgccaa	gcaaggtata	tatggtctgg	cccgttcatt	ccctacctca	1020
ggagccatag	accgtgttgc	caaggcccat	ggtctaaact	gttatgaggt	cccaactggc	1080
tggaaatttt	tctgtgcttt	gttcgacgct	aaaaaattat	ctatttgggg	tgaagaatcg	1140
tttgactactg	tctccaacca	cgtaagggaa	aaggacggtg	tttgggccat	tatggcgtgg	1200
ttgaacatct	tggccattta	caacaagcat	catccggaga	acgaagcttc	tattaagacg	1260
atacagaatg	aattctgggc	aaagtacggc	cgtactttct	tcaactcgta	tgattttgaa	1320
aaagttgaaa	cagaaaaagc	taacaagatt	gtcgatcaat	tgagagcata	tgttaccaa	1380
tcgggtgttg	ttaattccgc	cttcccagcc	gatgagtctc	ttaaggtcac	cgattgtggt	1440
gatttttcat	acacagattt	ggacggttct	gtttctgacc	atcaagggtt	atatgtcaag	1500
ctttccaatg	gtgcaagatt	cgttctaaga	ttgtcaggta	caggttcttc	aggtgctacc	1560
attagattgt	acattgaaaa	atactgcat	gataaatcac	aatacaaaaa	gacagctgaa	1620
gaataactga	agccaattat	taactcggtc	atcaagttct	tgaactttaa	acaagtttta	1680
ggaactgaag	aaccaacggt	tcgtacttaa				1710

SEQ ID NO:119

S. cerevisiae

MSFQIETVPT	KPYEDQKPGT	SGLRKKTKVF	KDEPNYTENF	IQSIMEAIP	GSKGATLVVG	60
GDGRYNDVI	LHKIAAIGAA	NGIKKLVIGQ	HGLLSTPAAS	HIMRTYEKC	TGGIILTASH	120
NPGGPENDMG	IKYNLSNGGP	APESVTNAIW	EISKKLTSYK	IIKDFPELDL	GTIGKNKKYG	180
PLLVDIIDIT	KDYVNFLEKI	FDLFIKKFI	DNQRSTKNWK	LLFDSMNGVT	GPYKAI FVD	240
EFGLPADEVL	QNWHPSPDFG	GMHPDPNLTY	ASSLVKRVDR	EKIEFGAASD	GDGDRNMIY	300
YGPSFVSPGD	SVAIIAEYAA	EIPYFAKQGI	YGLARSFPTS	GAIDRVAKAH	GLNCYEVPTG	360
WKFFCALFDA	KKLSICGEES	FGTGSNHVRE	KDGVWAIMAW	LNILAIYNKH	HPENEASIKT	420
IQNEFWAKYK	RTFFTRYDFE	KVETEKANKI	VDQLRAYVTK	SGVVNSAFPA	DESLKVTDCG	480
DFSYTDLDGS	VSDHQGLYVK	LSNGARFVLR	LSGTGSSGAT	IRLYIEKYCD	DKSQYQKTAE	540
EYLPKIINSV	IKFLNFKQVL	GTEEPTVRT				560

SEQ ID NO:120

S. cerevisiae

atgtccacta	agaagcacac	caaaacacat	tccacttatg	cattcgagag	caacacaaac	60
agcgttgctg	cctcacaaa	gagaaacgcc	ttaaacaagt	tggcggactc	tagtaaactt	120
gacgatgctg	ctcgcgctaa	gtttgagaac	gaactggatt	cgtttttcac	gcttttcagg	180
agatatttgg	tagagaagtc	ttctagaacc	accttggaat	gggacaagat	caagtctccc	240
aaccggtatg	aagtgggtta	gtatgaaatt	atctctcagc	agcccagaaa	tgtctcaaac	300
ctttccaaat	tggtgtttt	gaagttgaac	ggtgggctgg	gtacctccat	gggctgcgtt	360
ggccctaaat	ctgttattga	agtgagagag	ggaaacacct	ttttggattt	gtctgttcgt	420
caaattgaat	acttgaacag	acagtacgat	agcgcagctg	cattgttatt	gatgaattct	480
ttcaacactg	acaaggatac	ggaacacttg	attaagaagt	attccgctaa	cagaatcaga	540
atcagatctt	tcaatcaatc	caggttccca	agagtctaca	aggattcttt	attgcctgtc	600

cccaccgaat	acgattctcc	actggatgct	tggtatccac	caggtcacgg	tgatttgttt	660
gaatctttac	acgtatctgg	tgaactggat	gccttaattg	cccaaggaag	agaaatatta	720
tttgtttcta	acggtgacaa	cttgggtgct	accgtcgact	taaaaatfff	aaaccacatg	780
atcgagactg	gtgccgaata	tataatggaa	ttgactgata	agaccagagc	cgatgttaaa	840
ggtggactct	tgatttctta	cgatgggtcaa	gtccggtttat	tggaagtcgc	ccaagttcca	900
aaagaacaca	ttgacgaatt	caaaaatatac	agaaaagtta	ccaacttcaa	cacgaataac	960
ttatggatca	atctgaaagc	agtaaagagg	ttgatcgaat	cgagcaatff	ggagatggaa	1020
atcattccaa	accaaaaaac	tataacaaga	gacggatcatg	aaattaatgt	cttacaatta	1080
gaaaccgctt	gtgggtgctg	tatcaggcat	tttgatgggtg	ctcacgggtg	tgctgtttcca	1140
agatcaagat	tcttgctctg	caagacctgt	tccgatttgg	tgctgggttaa	atcagatcta	1200
ttccgtctgg	aacacggttc	tttgaagtta	gacccatccc	gttttggtcc	aaaccatta	1260
atcaagttgg	gctcgcattt	caaaaagggt	tctgggtttta	acgcaagaat	ccctcacatc	1320
ccaaaaatcg	tcgagctaga	tcatcttgacc	atcactggta	acgtcttttt	aggtaaagat	1380
gtcactttga	gggtactgtg	catcatcgtt	tgctccgacg	gtcataaaat	cgatattcca	1440
aacggctcca	tattggaaaa	tgttgtcgtt	actggtaatt	tgcaaatctt	ggaacattga	1500

SEQ ID NO:121

S. cerevisiae

MSTKKHTKTH	STYAFESNTN	SVAASQMRNA	LNKLADSSKL	DDAARAKFEN	ELDSFFTLFR	60
RYLVEKSSRT	TLEWDKIKSP	NPDEVVKYEI	ISQQPENVSN	LSKLAVLKLN	GGLGTSMGCV	120
GPKSVIEVRE	GNTFLDLVSR	QIEYLNRYD	SDVPLLLMNS	FNTDKDTEHL	IKKYSANRIR	180
IRSFNQSRFP	RVKDSLLEPV	PTEYDSPLDA	WYPPGHGDLF	ESLHVSGLD	ALIAQGREIL	240
FVSNGDNLGA	TVDLKILNHM	IETGAEYIME	LTDKTRADV	GGTLISYDGO	VRLEVAQVP	300
KEHIDEFKNI	RKFTNFNTNN	LWINLKAVKR	LISSNLEME	IIPNQKTITR	DGHEINVLQL	360
ETACGAAIRH	FDGAHVVP	RSRFLPVKTC	SDLLLKSD	FRLEHGSLKL	DPSRFGPNPL	420
IKLGSHFKKV	SGFNARIPHI	PKIVELDHLT	ITGNVFLGKD	VTLRGTVIIV	CSDGHKIDIP	480
NGSILENVVV	TGNLQILEH					499

SEQ ID NO:122

S. cerevisiae

atgtctagtc	aaacagaaaag	aacttttatt	gcggtaaaac	cagatgggtg	ccagagggggc	60
ttagtatctc	aaattctatc	tcgttttgaa	aaaaaaggtt	acaaactagt	tgctattaaa	120
ttagttaaac	cgatgataa	attactagag	caacattacg	cagagcatgt	tggtaaacca	180
tttttcccaa	agatggtatc	ctttatgaag	tctgggtccca	ttttggccac	ggtctgggag	240
ggaaaagatg	tggttagaca	aggaagaact	attcttggtg	ctactaatcc	tttgggcagt	300
gcaccaggta	ccattagagg	tgatttcggt	attgacctag	gcagaaacgt	ctgtcacggc	360
agtgattctg	ttgatagcgc	tgaacgtgaa	atcaatttgg	ggtttaagaa	ggaagagtta	420
gttgattggg	aatctaataca	agctaagtgg	atztatgaat	ga		462

SEQ ID NO:123

S. cerevisiae

MSSQTERTFI	AVKPDGVQRG	LVSQILSRFE	KKGYKLVAIK	LVKADDKLE	QHYAEHVGKP	60
FFPKMVSFMK	SGPILATVWE	GKDVRQGR	ILGATNPLGS	APGTIRGDFG	IDLGRNVCHG	120
SDSVDSAERE	INLWFKKEEL	VDWESNQAKW	IYE			153

SEQ ID NO:124

S. rebaudiana

atggctgctg	ctgatactga	aaagttgaac	aatttgagat	ccgccgtttc	tggtttgacc	60
caaatttctg	ataacgaaaa	gtccggtttc	atcaacttgg	tcagtagata	tttgtctggt	120
gaagctcaac	acgttgaatg	gtctaaaatt	caaacctcaa	ccgataagat	cgttgtttcca	180
tagcatactt	tgctgctgt	tccagaagat	gctgctcaaa	caaaatcttt	ggtgataaag	240
ttggtcgtct	tgaagttgaa	cggtggtttg	ggtactacta	tgggtttgtac	tggtccaaag	300
tctgttatcg	aagttagaaa	cggtttgacc	ttcttgatt	tgatcgtcat	ccaaatcgaa	360

tccttgaaca	agaagtacgg	ttgttctggt	cctttgttgt	tgatgaactc	tttcaacacc	420
catgaagata	cccaaaagat	cgtcgaaaag	tactccggtt	ctaacattga	agttcacacc	480
ttcaatcaat	cccaataccc	aagattgggt	gtcgatgaat	ttttgccatt	gccatctaaa	540
ggtgaaactg	gtaaagatgg	ttggtatcca	ccaggatcatg	gtgatgtttt	tccatccttg	600
atgaattccg	gtaagttgga	tgctttggtg	tcccaaggtg	aagaatacgt	tttcggtgcc	660
aactctgata	acttgggtgc	agttggtgat	ttgaagatct	tgaaccactt	gatccaaaac	720
aagaacgaat	actgcatgga	agttactcca	aagactttgg	ctgatgttaa	gggtgggtact	780
ttgatttctt	acgatggtaa	ggttcaatta	ttggaaatcg	cccaagttcc	agatgaacac	840
gttaatgaat	tcaagtccat	cgaaaagttt	aagatcttta	acactaacia	cttgtgggtc	900
aacttgaacg	ccattaagag	attggttcaa	gctgatgctt	tgaagatgga	aattattcca	960
aatccaaaag	aagcaacgg	tgtcaaggta	ttgcaattgg	aaactgctgc	tggtgctgct	1020
attaagtttt	tcgataatgc	catcggatc	aacgtcccaa	gatctagatt	tttgcctggt	1080
aaggcttctt	ctgacttggt	gtagttcaa	tcagacttgt	acaccgaaaa	ggatgggtac	1140
gttattagaa	accagctag	aaaggatcca	gctaaccat	ctattgaatt	gggtccagaa	1200
ttcaaaaagg	tcggtgattt	cttgaagaga	ttcaagtcta	tcccatccat	catcgaattg	1260
gactcattga	aagtttctgg	tgatgtctgg	tttggttcca	acgttgtttt	gaaaggtgaa	1320
gttggtggtg	ctgccaatc	cggtgaaaa	ttggaaatc	cagatgggtgc	cttgattgaa	1380
aacaaagaag	ttcatggtgc	ctccgacatt	tga			1413

SEQ ID NO:125

S. rebaudiana

MAAADTEKLN	NLRSAVSLT	QISDNEKSGF	INLVSRYLSG	EAQHVEWSKI	QTPTDKIVVP	60
YDTLSAVPED	AAQTKSLLDK	LVVLKLNGLL	GTTMGCTGPK	SVIEVRNGLT	FLDLIVIQIE	120
SLNKYGCYSV	PLLLMNSFNT	HEDTQKIVEK	YSGSNIEVHT	FNQSQYPRLV	VDEFLPLPSK	180
GETGKDGWYP	PGHGDVFPST	MNSGKLDALL	SQGKEYVFVA	NSDNLGAVVD	LKILNHLIQN	240
KNEYCMEVTP	KTLDVKGGT	LISYDGVQVQ	LEIAQVPDEH	VNEFKSIEKF	KIFNTNLLWV	300
NLNAIKRLVQ	ADALKMEIIP	NPKEVNGVKV	LQLETAAGAA	IKFFDNAIGI	NVPRSRFLPV	360
KASSDLLLVQ	SDLYTEKDG	VIRNPARKDP	ANPSIELGPE	FKKVGDFLKR	FKSIPSIIEEL	420
DSLKVSQDVW	FGSNVVLKKG	VVVAKSGEK	LEIPDGALIE	NKEVHGASDI		470

SEQ ID NO:126

A. pullulans

atgtcctctg	aaatggctac	tcatttgaaa	cctaattggtg	gtgccgaatt	cgaaaaaaga	60
catcatggta	agacccaatc	ccatggtgct	tttgaaaaca	cttctacatc	tgttgctgcc	120
tcccaaatga	gaaatgcttt	gaatactttg	tgcgattccg	ttactgatcc	agctgaaaag	180
caaagattcg	aaaccgaaat	ggataacttc	ttcgcttctg	ttagaagata	cttgaacgat	240
aaggctaagg	gtaacgaaat	cgaaatggtct	agaattgctc	cacccaaaacc	agaacaagtt	300
gttgcttatc	aagacttgcc	tgaacaagaa	tcogttgaat	tcttgaacia	attggccgctc	360
ttgaagttga	atggtgggtt	gggtacttct	atgggttctg	ttggtccaaa	gtctgttatc	420
gaagttagag	atggtatgct	cttcttggat	ttgtccgcta	gacaaatcga	atacttgaat	480
agaacctacg	gtgttaacgt	tccattcgtc	ttgatgaatt	ctttcaacac	tgatgctgat	540
accgccaaca	ttatcaaaaa	gtacgaagg	cacaacatcg	acatcatgac	cttcaatcaa	600
tctagatacc	caagaatctt	gaaggattct	ttggtgccag	ctccaaaatc	tgccaactct	660
caaatttctg	attggtatcc	accaggtcat	ggtgacggtt	ttgaatcctt	gtacaactct	720
ggtatcttgg	ataagttggt	ggaaagagg	gtcgaatcg	ttttcttctc	caatgctgat	780
aatttgggtg	ccgttgttga	tttgaagatc	ttgcaacata	tggttgatac	caaggccgaa	840
tatatcatgg	aattgactga	taagactaag	gccgatgcta	aggggtgtac	tattattgac	900
tatgaaggct	aagccagatt	attggaat	gcccagttc	caaaagaaca	cgtcaacgaa	960
ttcaagtcca	tcaagaagtt	taagtacttc	aacaccaaca	acatctggat	gaacttgaga	1020
gctgttaaga	gaatcgctga	aaacaacgaa	ttggccatgg	aaattatccc	aaacggtaaa	1080
tctattccag	ccgacaaaaa	aggtgaagcc	gatgtttcta	tagttcaatt	ggaaactgct	1140
gttggtgctg	ccattagaca	ttttaacaat	gctcatggtg	tcaacgtccc	aagaagaaga	1200
tttttgccag	ttaagacctg	ctccgatttg	atggttggtta	agtctgactt	gtacactttg	1260
aagcacggct	aattgattat	ggacccaaat	agatttggtc	cagccccatt	gattaagttg	1320

ggtggtgatt	ttaagaaggt	ttcctcattc	caatccagaa	tcccatccat	tcctaaaatc	1380
ttggaattgg	atcatttgac	cattaccggg	ccagttaact	tgggtagagg	tgttactttt	1440
aagggtagctg	ttattatcgt	tgctccgaa	gggtcaaacca	ttgatattcc	acctggttcc	1500
atthttgaaa	acgtttgtgt	tcaaggttcc	ttgagattat	tagaacatta	a	1551

SEQ ID NO:127

A. pullulans

MSSEMATHLK	PNGGAEFEKR	HHGKTQSHVA	FENTSTSVAA	SQMRNALNTL	CDSVTDPAEK	60
QRFETEMDNF	FALFRYLND	KAKGNEIEWS	RIAPPKPEQV	VAYQDLPEQE	SVEFLNKLAV	120
LKLNGLGTS	MGCVGPKSVI	EVRDGMFLD	LSVRQIEYLN	RTYGVNVPFV	LMNSFNTDAD	180
TANIIKKEYG	HNIDIMTFNQ	SRYPRIKDS	LLPAPKSANS	QISDWYPPGH	GDVFESLYNS	240
GILDKLLERG	VEIVFLSNAD	NLGAVVDLKI	LQHMVDTKAE	YIMELTDKTK	ADVKGGTIID	300
YEQARLLEI	AQVPKEHVNE	FKSIKKFKYF	NTNNIWMNLR	AVKRIVENNE	LAMEIIPNGK	360
SIPADKKGEA	DVSIVQLETA	VGAAIRHFNN	AHGVNVPRRR	FLPVKTCSDL	MLVKSPLYTL	420
KHGQLIMDPN	RFGPAPLIK	GGDFKKVSSF	QSRIPIPKI	LELDHLTITG	PVNLGRGVTF	480
KGTVIIIVASE	GQTIDIPPGS	I LENVVQGS	LRLLEH			516

SEQ ID NO:128

A. thaliana

atggctgcta	ctactgaaaa	cttgccacaa	ttgaaatctg	ccggtgatgg	tttgactgaa	60
atgtccgaat	ctgaaaagtc	cggtttcatc	tctttgggtca	gtagatattt	gtctggtgaa	120
gccaacata	tcgaatggct	taaaattcaa	actccaaccg	acgaaatcgt	tgtcccatac	180
gaaaaaatga	ctccagtttc	tcaagatgtc	gccgaaacta	agaatttggt	ggataagttg	240
gtcgtcttga	agttgaatgg	tggtttgggt	actactatgg	gttgtactgg	tccaaagtct	300
gttatcgaag	ttagagatgg	tttaaccttc	ttggacttga	tcgtcatcca	aatcgaaaac	360
ttgaacaaca	agtacggttg	caaggttcca	ttggtcttga	tgaattcttt	caacacccat	420
gatgataccc	acaagatcgt	tgaaaagtac	accaactcca	acggtgatat	ccacaccttc	480
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aactccggta	agttggatac	tttcttgtec	caaggtaaag	aatacgtttt	cgttgccaac	660
tctgataact	tggtgctat	agttgatttg	accatcttga	agcacttgat	ccaaaacaag	720
aacgaatact	gcatggaagt	tactccaaag	actttggctg	atgttaaggg	tggtactttg	780
atthctttag	aaggtaaagt	tcaattattg	gaaatcgccc	aagttccaga	tgaacacggt	840
aatgaattca	agtccatcga	aaagttcaag	atcttcaaca	ccaacaactt	gtggggtaac	900
ttgaaggcca	tcaagaaatt	ggttgaagct	gatgctttga	agatggaaat	tatcccaaac	960
ccaaaagaag	ttgacggtgt	taaggtattg	caattggaaa	ctgctgctgg	tgctgctatt	1020
agatthttcg	ataatgccat	cggtgttaac	gtcccaagat	ctagatthtt	gccagtttaag	1080
gcttctctcg	atthgttgtt	ggttcaatct	gacttgta	ccttggttga	cggttttgtt	1140
acaagaaaca	aggctagaac	taaccatctc	aaccatctta	ttgaattggg	tccagaattc	1200
aaaaaggttg	ccacattctt	gtccagattc	aagtctattc	catccatcgt	cgaattggac	1260
tcattgaaag	thtctggtga	tgtctgggtt	ggttctctta	tagthttgaa	gggtaaggtt	1320
actgthgctg	ctaaatctgg	tgthtaagttg	gaaattccag	atagagccgt	tgthgaaaa	1380
aaaaacatta	acgthctctga	agatthgtga				1410

SEQ ID NO:129

A. thaliana

MAATTENLPQ	LKSAVDGLTE	MSESEKSGFI	SLVSRYLSE	AQHIEWSKIQ	TPTDEIVVPY	60
EKMTPVSDV	AETKNLLDKL	VVLKLNGLG	TTMGCTGPKS	VIEVRDGLTF	LDLIVIQIEN	120
LNNKYGCKVP	LVLNNSFNTH	DDTHKIVEKY	TNSNVDIHTF	NQSKYPRVVA	DEFVPWPSKG	180
KTDKEGWYPP	GHGDVFPALM	NSGKLDTFLS	QGKEYVVFAN	SDNLGAIVDL	TILKHLIQNK	240
NEYCMEVTPK	TLADVKGGLT	ISYEGKVQLL	EIAQVPDEHV	NEFKSIEKFK	IFNTNLLWVN	300
LKAIKKLVEA	DALKMEIIPN	PKEVDGVKVL	QLETAAGAAI	RFFDNAIGVN	VPRSRFLPVK	360
ASSDLLLVQS	DLYTLVDGFV	TRNKARTNPS	NPSIELGPEF	KKVATFLSRF	KSIPSIVELD	420

SLKVSQDVWF GSSIVLKGKV TVAAKSGVKL EIPDRAVVEN KNINGPEDL 469

SEQ ID NO:130

E. coli

atggctgcta	taaacaccaa	ggtaagaag	gctgttattc	cagttgctgg	tttgggtact	60
agaatgttgc	cagctacaaa	agccattcca	aaagaaatgt	taccattggg	cgataagcca	120
ttgatccaat	acgttgtaaa	cgaatgtatt	gctgctggta	ttaccgaaat	cgttttgggt	180
actcactcct	ccaagaactc	cattgaaaat	catttcgaca	cctcattcga	attggaagcc	240
atggttgaaa	agagagtcaa	gagacaatta	ttggacgaag	tccaatctat	ttgccacca	300
catgttacta	tcatgcaagt	tagacaagg	ttggctaaag	gtttgggtca	tgctgttttg	360
tgtgctcatc	cagttgttgg	tgatgaacca	gttgcagtta	ttttgccaga	tgttatcttg	420
gacgaatacg	aatccgattt	gtctcaagat	aacttggctg	aaatgatcag	aagattcgac	480
gaaactggtc	actcccaa	tatggttgaa	cctgttggctg	atgttactgc	ttatgggtgt	540
gttgattgca	agggtgttga	attggctcca	ggtgaatctg	ttccaatggg	tggtgttga	600
gaaaagccaa	aagctgatgt	tgctccatct	aatttggcta	tcgttggtag	atatgttttg	660
tccgctgata	tttggccttt	gttggctaaa	actccaccag	gtgctgggta	cgaaattcaa	720
ttgactgatg	ctatcgacat	gttgatcgaa	aaagaaaccg	ttgaagccta	ccacatgaag	780
ggtaaactctc	atgattgtgg	taacaagttg	ggttacatgc	aagcttttgt	tgaatacggg	840
atcagacata	acaccttagg	tactgaattc	aaggcttggg	tggaagaaga	aatgggtatc	900
aagaagtaa						909

SEQ ID NO:131

E. coli

MAAINTKVKK	AVI PVAGLGT	RMLPATKAIP	KEMLPLVDKP	LIQYVVNECI	AAGITEIVLV	60
THSSKNSIEN	HFDTSFELEA	MLEKRVKRQL	LDEVQSI CPP	HVTIMQVRQG	LAKGLGHAVL	120
CAHPVVGDEP	VAVILPDVIL	DEYESDLSQD	NLAEMIRRFD	ETGHSQIMVE	PVADVTAYGV	180
VDCKGVELAP	GESVPMVGVV	EKPKADVAPS	NLAIVGRYVL	SADIWPLLAK	TPPGAGDEIQ	240
LTD AIDMLIE	KETVEAYHMK	GKSHDCGNKL	GYMQAFVEYG	IRHNTLGTEF	KAWLEEMGI	300
KK						302

SEQ ID NO:132

R. suavis

atggctgctg	ttgctactga	taagatctct	aagttgaagt	ctgaagttgc	tgcttgtcc	60
caaatttctg	aaaacgaaaa	gtccggtttc	atcaacttgg	tcagtagata	tttgtctggg	120
actgaagcta	ctcacgttga	atggtctaaa	attcaaactc	caaccgatga	agttgttgg	180
ccatgatgata	ctttggctcc	aactccagaa	gatccagctg	aaactaagaa	gttgttagat	240
aagttggctg	tcttgaagtt	gaacgggtgg	ttgggtacta	ctatgggttg	tactgggtcca	300
aagtctgtta	tcgaagttag	aaacggtttg	accttcttgg	atgtgatcgt	cattcaaactc	360
gaaaccttga	acaacaagta	cggttgtaac	gttcctttgt	tgttgatgaa	ctctttcaac	420
acccatgatg	acaccttcaa	gatcgttgaa	agatacacca	agtccaacgt	tcaaatccat	480
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aagggctcaa	ctggtaaaaga	tggttgggat	ccaccaggtc	atgggtgatgt	ttttccatct	600
ttgagaact	ccggttaagtt	ggatttggtt	ttatcccaag	gtaaagaata	cgttttcatc	660
tccaactctg	ataacttggg	tgcagttggt	gatttgaaga	tcttgtccca	tttgggtcca	720
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cacgttaacg	aattcaagtc	catcgaaaag	ttcaagatct	ttaacaccaa	caatttgggg	900
gtcaacttga	acgccattaa	gagattagtt	gaagctgatg	ccttgaaaat	ggaaatcatc	960
ccaaatccaa	aagaagtcga	cggtattaag	gtccttgcaat	tggaactgc	tgctgggtgct	1020
gctattagat	ttttcaatca	tgccatcggg	atcaacgtcc	caagatctag	atttttgcca	1080
gttaaggcta	cctccgattt	gttattgggt	caatctgact	tgtacaccgt	cgaagatggg	1140
ttcgttatta	gaaacactgc	tagaaagaat	ccagccaacc	catctgttga	attgggtcca	1200
gaattcaaaa	aggttgccaa	cttcttgtcc	agattcaagt	ctattccatc	catcatcgaa	1260

ttggactcat	tgaaggttgt	tggatgatgta	tggtttgggtg	ctgggtggtt	tttgaaaggt	1320
aaggttacta	ttactgctaa	gccaggtggt	aagttggaaa	ttccagataa	ggctgtcttg	1380
gaaaacaagg	atattaacgg	tcctgaagat	ttgtga			1416

SEQ ID NO:133

R. suavis

MAAVATDKIS	KLKSEVAALS	QISENEKSGF	INLVSRYLSG	TEATHVEWSK	IQTPTDEVVV	60
PYDTLAPTPE	DPAETKLLD	KLVLKLNNG	LGTTMGCTGP	KSVIEVRNGL	TFLDLIVIQI	120
ETLNKYGCN	VPLLLMNSFN	THDDTFKIVE	RYTKSNVQIH	TFNQSQYPR	VVEDNSPLPS	180
KGQTGKDGWY	PPGHGDVFP	LRNSGKLDLL	LSQGKEYVFI	SNSDNLGAVV	DLKILSHLVQ	240
KKNEYCMEVT	PKTLADVKGG	TLISYEGRTQ	LLEIAQVPDQ	HVNEFKSIEK	FKIFNTNNLW	300
VNLNAIKRLV	EADALKMEII	PNPKEVDGIK	VLQLETAAGA	AIRFFNHAIG	INVPRSRFLP	360
VKATSDLLLV	QSDLYTVEDG	FVIRNTARKN	PANPSVELGP	EFKKVANFLS	RFKSIPSIIE	420
LDSLKVVDV	WFGAGVVLKG	KVTITAKPGV	KLEIPDKAVL	ENKNDINGPED	L	471

SEQ ID NO:134

H. vulgare

atggctgctg	ctgcagttgc	tgctgattct	aaaattgatg	gtttgagaga	tgctggtgcc	60
aagttgggtg	aaatttctga	aaacgaaaag	gccggtttca	tctccttgg	ttctagatat	120
ttgtctggtg	aagccgaaca	aatcgaatgg	tctaaaattc	aaactccaac	cgatgaagtt	180
gttggtccat	atgatacttt	ggctccacca	cctgaagatt	tggatgctat	gaaggctttg	240
ttggataagt	tggttgtctt	gaagttgaat	gggtggttgg	gtactactat	gggttgtact	300
ggtcacaagt	ctgttatcga	agttagaaac	ggtttcacct	tcttggattt	gatcgttatc	360
caaattgaat	ccttgaacaa	gaagtacggt	tgctctgttc	ctttggtggt	gatgaactct	420
ttcaacacc	atgatgacac	ccaaaagatc	gttgaaaagt	actccaactc	caacatcgaa	480
atccacacct	tcaatcaatc	tcaataccca	agaatcgtca	ccgaagattt	tttgccattg	540
ccatctaaag	gtcaaaactg	taaagatgg	tggatccac	caggatcatg	tgatgttttt	600
ccatctttga	acaactccg	taagttggat	accttgttgt	ctcaaggtaa	agaatacgtt	660
ttcgttgcca	actctgataa	cttgggtgct	atcgttgata	ttaagatctt	gaaccacttg	720
atccacaatc	aaaacgaata	ctgcatggaa	gttactccaa	agactttggc	tgatgttaag	780
ggtggtactt	tgatttctta	cgaaggtaga	gttcaattat	tggaaatcgc	ccaagttcca	840
gatgaacacg	ttgatgaatt	caagtccatc	gaaaagttca	aaatcttcaa	caccaacaac	900
ttgtgggtta	acttgaaggc	cattaagaga	ttggttgatg	ctgaagcttt	gaaaatggaa	960
atcatcccaa	accctaaaga	agttgacggt	gttaaggtat	tgcaattgga	aactgctgct	1020
ggtgctgcta	ttagattctt	tgaaaaagcc	atcggtatca	acgtcccaag	atctagattt	1080
ttgctcagta	aggctacctc	tgacttgttg	ttggttcaat	cagacttgta	caccttggtt	1140
gacggttacg	ttattagaaa	tccagctaga	gttaagccat	ccaaccatc	tattgaattg	1200
ggtccagaat	tcaagaaggt	cgctaatttc	ttggctagat	tcaagtctat	cccatccatc	1260
gttgaattgg	actcattgaa	agtttctggt	gatgtctctt	ttggttccgg	tgttgttttg	1320
aagggtaatg	ttactattgc	tgctaaggct	ggtgttaagt	tggaaattcc	agatggtgct	1380
gttttgaaa	acaaggatat	taacggtcca	gaagatattt	ga		1422

SEQ ID NO:135

H. vulgare

MAAAVAADS	KIDGLRDAVA	KLGEISENEK	AGFISLVSRY	LSGEAEQIEW	SKIQTPTDEV	60
VVPYDTLAPP	PEDLDAMKAL	LDKLVVLKLN	GGLGTTMGCT	GPKSVIEVRN	GFTFLDLIVI	120
QIESLNKKYG	CSVPLLLMNS	FNTHDDTQKI	VEKYSNSNIE	IHTFNQSQYP	RIVTEDFLPL	180
PSKGQTGKDG	WYPPGHGDVF	PSLNNSGKLD	TLLSQGKEYV	FVANSNDLGA	IVDIKILNHL	240
IHNQNEYCME	VTPKTLADVK	GGTLISYEGR	VQLLEIAQVP	DEHVDEFKSI	EKFKIFNTNN	300
LWVNLKAIKR	LVDAEALKME	IIPNPKEVDG	VKVLQLETAA	GAAIRFFEKA	IGINVPRSRF	360
LPVKATSDLL	LVQSDLYTLV	DGYVIRNPAR	VKPSNPSIEL	GPEFKKVANF	LARFKSIPSI	420
VELDSLKVS	DVSFGSGVVL	KGNVTIAAKA	GVKLEIPDGA	VLENKNDINGP	EDI	473

SEQ ID NO:136

O. sativa

atggctgacg	aaaaattggc	caaattgaga	gaagctgttg	ctggtttgtc	tcaaatctct	60
gataacgaaa	agtcgggttt	catttccttg	gttgctagat	atgtgtccgg	tgaagaagaa	120
catggtgaat	gggtctaaaat	tcatacccca	accgatgaag	ttgttgttcc	atatgatact	180
ttggaagctc	caccagaaga	tttggaga	acaaaaaagt	tgttgaacaa	gttggccgctc	240
ttgaagttga	atggtgggtt	gggtactact	atgggttgta	ctggtcctaaa	gtctgttatc	300
gaagttagaa	acggtttcac	cttcttggt	ttgatcgtca	tccaaatcga	atccttgaac	360
aaaaagtacg	gttccaacgt	tcctttggtg	ttgatgaact	ctttcaacac	ccatgaagat	420
accttgaaga	tcgttgaaaa	gtacaccaac	tccaacatcg	aagttcacac	cttcaatcaa	480
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ggtaagttgg	acttgttgtt	gtcccaaggt	aaagaatacg	ttttcattgc	caactccgat	660
aacttgggtg	ctatagttga	tatgaagatt	ttgaaccact	tgatccacaa	gcaaaacgaa	720
tactgtatgg	aagtactcc	aaagactttg	gctgatgtta	aggggtgtac	tttgatctct	780
tacgaagata	aggttcaatt	attggaaatc	gcccaagttc	cagatgctca	tgtaaagaa	840
ttcaagtcca	tcgaaaagtt	caagatcttt	aacaccaaca	acttgtgggt	taacttgaag	900
gccattaaga	gattagttga	agctgacgct	ttgaagatgg	aaattatccc	aaacccaaaa	960
gaagttgacg	gtgttaaggt	attgcaattg	gaaactgctg	ctgggtgctgc	tattagattt	1020
ttcgatcatg	ctatcggtat	caacgtccca	agatctagat	ttttaccagt	taaggctacc	1080
tccgacttgc	aactagtcca	atctgacttg	tacaccttgg	ttgatgggtt	cgttactaga	1140
aatccagcta	gaactaatcc	atccaaccca	tctattgaat	tgggtccaga	attcaagaag	1200
gttggttgtt	ttttgggtag	attcaagtct	atcccatcca	tcgttgaatt	ggacactttg	1260
aaagtttctg	gtgatgtttg	gttcggttcc	tccattacat	tgaaaggtaa	ggttactatt	1320
accgctcaac	caggtgttaa	gttggaaatt	ccagatgggtg	ctgtcatcga	aaacaaggat	1380
attaacggtc	ctgaagattt	gtga				1404

SEQ ID NO:137

O. sativa

MADEKLAKLR	EAVAGLSQIS	DNEKSGFISL	VARYLSGEEE	HVEWAKIHTP	TDEVVVPYDT	60
LEAPPEDLEE	TKKLLNKLAV	LKLNGLGTT	MGCTGPKSVI	EVRNGFTFLD	LIVIQIESLN	120
KKYGSNVPLL	LMNSFNTHED	TLKIVEKYTN	SNIEVHTFNQ	SQYPRVVADE	FLPWPSKGKT	180
CKDGWYPPGH	GDIFFPSLMNS	GKLDLLLSQG	KEYVFIANS	NLGAIVDMKI	LNHLIHKQNE	240
YCMVTPKTL	ADVKGGLTIS	YEDKVQLLEI	AQVPDAHVNE	FKSIEKFKIF	NTNNLWVNLK	300
AIKRLVEADA	LKMEIIPNPK	EVDGVKVLQL	ETAAGAAIRF	FDHAIGINVP	RSRFLPVKAT	360
SDLQLVQSDL	YTLVDGFVTR	NPARTNPSNP	SIELGPEFKK	VGCFLGRFKS	IPSIVELDTL	420
KVSGDVWFGS	SITLKGKVTI	TAQPGVKLEI	PDGAVIENKD	INGPEDL		467

SEQ ID NO:138

S. tuberosum

atggctactg	ctactacttt	gtctccagct	gatgctgaaa	agttgaacaa	tttgaatct	60
gctgtcgccg	gtttgaatca	aatctctgaa	aacgaaaagt	ccggtttcat	caacttgggt	120
ggtagatatt	tgctctggtg	agcccaacat	attgactggt	ctaaaattca	aactccaacc	180
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ggttgactg	gtccaaagtc	tgttatcgaa	gttagaacag	gtttgacctt	cttggatttg	360
atcgtcaagc	aaattgaagc	tttgaacgct	aagttcgggt	gttctgttcc	tttgttgttg	420
atgaactctt	tcaacaccca	tgatgacacc	ttgaagatcg	ttgaaaagta	cgccaactcc	480
aacattgata	tccacacctt	caatcaatcc	caatacccaa	gattggttac	cgaagatttt	540
gctccattgc	catgtaaagg	taactctggt	aaagatgggt	ggtatccacc	aggtcatggt	600
gatgtttttc	catccttgat	gaattccggt	aagttggatg	ctttgttggc	taagggtaaa	660
gaatacgttt	tcgttgccaa	ctctgataac	ttgggtgcta	tcgttgattt	gaaaatcttg	720
aaccacttga	tcttgaacaa	gaacgaatac	tgcatggaag	ttactccaaa	gactttggct	780
gatgttaagg	gtggtacttt	gatttcttac	gaaggtaagg	ttcaattatt	ggaaatcgcc	840

caagttccag	atgaacacgt	taatgaatc	aagtccatcg	aaaagttaa	gatcttcaac	900
actaacaact	tgtgggtcaa	cttgtctgcc	attaagagat	tgggtgaagc	tgatgccttg	960
aaaatggaaa	ttattccaaa	cccaaaagaa	gtcgatggtg	tcaaagtatt	gcaattggaa	1020
actgctgctg	gtgctgctat	taagtttttc	gatagagcta	ttgggtgcaa	cgttccaaga	1080
tctagatttt	tgccagttaa	ggctacctct	gacttgttgt	tggttcaatc	agacttgta	1140
actttgactg	atgaaggtta	cgttattaga	aaccagcta	gatccaatcc	atccaacca	1200
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atcccatcca	tcatcgattt	ggattctttg	aaagttactg	gtgatgtctg	gtttggttct	1320
ggtgttactt	tgaaggttaa	agttaccggt	gctgctaagt	caggtgttaa	gttggaaatt	1380
ccagatggtg	ctgttattgc	caacaaggat	attaacggtc	cagaagatat	ctaa	1434

SEQ ID NO:139

S. tuberosum

MATATTLSPA	DAEKLNNLKS	AVAGLNQISE	NEKSGFINLV	GRYLSGEAQH	IDWSKIQTPT	60
DEVVVPYDKL	APLSEDPAET	KKLLDKLVVL	KLNGGLGTTM	GCTGPKSVIE	VRNGLTFLDL	120
IVKQIEALNA	KFGCSVPLLL	MNSFNTHDDT	LKIVEKYANS	NIDIHTFNQS	QYPRLVTEDE	180
APLPCKGNNG	KDGYWPPGHG	DVFPMSLMSG	KLDALLAKGK	EYVVFVANSN	LGAIVDLKIL	240
NHLILNKNEY	CMEVTPKTLA	DVKGGTLISY	EGKVQLLEIA	QVPDEHVNEF	KSIEKFKIFN	300
TNNLWVNLSA	IKRLVEADAL	KMEIIPNPKE	VDGVKVLQLE	TAAGAAIKFF	DRAIGANVPR	360
SRFLPVKATS	DLLLVQSDLY	TLTDEGYVIR	NPARSNPSNP	SIELGPEFKK	VANFLGRFKS	420
IPSIIDLDSL	KVTGDVWFGS	GVTLKGVKTV	AAKSGVKLEI	PDGAVIANKD	INGPEDI	477

SEQ ID NO:140

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ttgttcatct	tcggtgtctc	attggtttct	acctctccaa	ttgatgggtca	aaaaccaggt	120
acttctgggt	tgagaaagaa	ggtcaagggt	ttcaagcaac	ctaactactt	ggaaaacttc	180
gttcaagcta	ctttcaacgc	tttgactacc	gaaaaagtta	aggggtgctac	tttggttgtt	240
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gctaacggtg	ttagaagagt	ttgggttggt	caaaactctt	tgttgtctac	tccagctggt	360
tccgccatta	ttagagaaaag	agttgggtgct	gatggttcta	aagctactgg	tgctttcatt	420
ttgactgctt	ctcataatcc	aggtgggtcca	actgaagatt	tcggtattaa	gtacaacatg	480
gaaaatgggt	gtccagcccc	agaatctatt	actgataaga	tatacgaaaa	caccaagacc	540
atcaagaagt	acccaattgc	agaagatttg	ccaagagttg	atatctctac	tatcggtatc	600
acttctttcg	aaggtcctga	aggtaaatc	gacggtgaag	tttttgattc	cgctgatgat	660
tacgtcaagt	tgatgaagtc	catcttcgac	ttcgaatcca	tcaagaagtt	gttgtcttac	720
ccaaagttca	ccttttggtta	cgatgcattg	catggtggtg	ctggtgctta	tgctcataga	780
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aaaggtggtg	ctagatctat	gccaaacttct	gctgcttttg	atggtgttgc	taagaatttg	1140
ggtttgaagt	tcttcgaagt	tccaactggt	tggaaattct	tcggtaat	gatggatgca	1200
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aaggatggta	tttgggctgt	tttggcttgg	ttgtctat	tggtcacia	gaacaaagaa	1320
accttgatg	gtaatgcaa	ggtggttact	ggtgaagata	tcggttagaca	acattgggct	1380
acttacggta	gacattacta	cactagatac	gactacgaaa	acggtgatgc	tacagctgct	1440
aaagaattga	tggttttatt	ggtcaagttg	caatcctcat	tgccagaagt	taacaagatc	1500
atcaagggta	tccatcctga	agttgctaat	ggtgcttctg	ctgatgaatt	cgaatacaag	1560
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ccattggttg	atgttgcttt	gaagttgtcc	aagatgcaag	aattcactgg	tagatcttct	1800
ccaaccgtta	ttacctga					1818

SEQ ID NO:141

MFLLVTS CFL	PDSGSSSVKVS	LFIFGVSLVS	TSPIDGQKPG	TSGLRKKVKV	FKQPNYLENF	60
VQATFNALTT	EKVKGATLVV	SGDGRYYSEQ	AIQIIVKMAA	ANGVRRVWVG	QNSLLSTPAV	120
SAIIRERVGA	DGSKATGAFI	LTASHNPGGP	TEDFGIKYNM	ENGGPAPESI	TDKIYENTKT	180
IKEYPIAEDL	PRVDISTIGI	TSFEGPEGKF	DVEVFDSADD	YVKLMKSIFD	FESIKKLLSY	240
PKFTFCYDAL	HGVAGAYAHR	IFVEELGAPE	SSLLNCVPKE	DFGGGHPDPN	LYAKELVAR	300
MGLSKTDDAG	GEPPEFGAAA	DGDADRNMIL	GKRFFVTPSD	SVAIIAANAV	GAIPIYFSSGL	360
KGVARSMPTS	AALDVVAKNL	GLKFFEVP TG	WKFFGNLMDA	GMCSVCGEES	FGTGS DHIRE	420
KDGIWAVLAW	LSILAHKNKE	TLDGNAKLVT	VEDIVRQHWA	TYGRHYTRY	DYENV DATAA	480
KELMGLLVKL	QSSLPEVNKI	IKGIHPEVAN	VASADEFEYK	DPVDGVS SKH	QGIRYLFEDG	540
SRLVFRLSGT	GSEGATIRLY	IEQYEKDASK	IGRDSQDALG	PLVDVALKLS	KMQEFTGRSS	600
PTVIT						605

SEQ ID NO:142

atggccattc	ataatagagc	tggtcaacca	gcacaacaat	ccgatttgat	taacgttgct	60
caattgaccg	cccaatatta	cgttttgaaa	cctgaagctg	gtaacgctga	acatgctggt	120
aagtttggtg	cttctgggtc	tagaggttct	gctgctagac	attcctttta	cgaaccacat	180
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gttttggctg	ctaacggtgt	tgatggtatc	gttcaagaaa	acaacggttt	cactccaact	360
ccagctgttt	ctaactgctat	tttggttcac	aacaaaaagg	gtggtccatt	ggctgatggt	420
atagttatta	ctccatctca	taaccacact	gaagatggtg	gtattaagta	caatccacca	480
aatggtggtc	cagctgatac	aaatggtact	aagggtggtg	aagatagagc	caacgctttg	540
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catgtcaaag	aacaagatth	ggttcaacca	ttcggtgaag	gtttggctga	tatagttgat	660
atggctgcta	ttcaaaaggc	tggtttgact	ttgggtggtg	atccattggg	tggttctggt	720
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aaggatggtg	ctgttggtaa	aactttggtt	tctctcgcta	tgatcgatag	agttgttaac	1080
gatttggtg	gaaagtgggt	tgaagttcca	gttggtttca	agtggtttgt	tgacggtttg	1140
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tccttcttgg	gtgaagaaca	tagaaagcaa	attgaaaaag	aagccgtcga	aatcgtcagt	1620
gaagttttga	agaatgccta	a				1641

SEQ ID NO:143

MAIHN RAGQP	AQQSDLINVA	QLTAQYYVLK	PEAGNAEHAV	KFGTSGHRGS	AARHSFNEPH	60
ILAI AQIAIE	ERAKNGITGP	CYVGK DTHAL	SEPAFISVLE	VLAANGVDVI	VQENNGFTPT	120
PAVSNAILVH	NKKGGLADG	IVITPSHNPP	EDGGIKYNPP	NGGPADTNVT	KVEDRANAL	180
LADGLKGVKR	ISLDEAMASG	HVKEQDLVQP	FVEGLADIVD	MAAIQKAGLT	LGVDPLGSG	240
IEYWKRI GEY	YNLNLTI VND	QVDQTFRFMH	LDKDG AIRMD	CSSECAMAGL	LALRDKFDLA	300
FANDPDYDRH	GIVTPAGLMN	PNHYLAVAIN	YLFQHRPQWG	KDVAVGKTLV	SSAMIDRVVN	360
DLGRKLV EVP	VGFKWFVDGL	FDGSFGFGGE	ESAGASFLRF	DGTPWSTDKD	GIIMCLLAAE	420
ITAVTGKNPQ	EHYNELAKRF	GAPSYNRLQA	AATSAQKAAL	SKLSPEMVSA	STLAGDPITA	480

RLTAAPGNGA SIGGLKVMTD NGWFAARPSG TEDAYKIYCE SFLGEEHRKQ IEKEAVEIVS 540
 EVLKNA 546

SEQ ID NO:144

R. suavis

atgtcctccg gtaagattaa gagagttcaa actactccat tcgacgggtca aaaaccagggt 60
 acttctgggt tgagaaagaa ggtaaggtt ttcacccaac ctaactactt gcaaaacttc 120
 gttcaatcta cttcaacgc tttgccatct gataaggtaa aagggtgctag attggttggt 180
 tctgggtgat gtagatactt ctccaaagaa gccattcaaa tcatcattaa gatggctgct 240
 ggtaacggtg ttaagtctgt ttgggttggt caaaatgggt tggtgtctac tccagctggt 300
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 gaaaatgggt gtccagctcc agaatctatt accaacaataa tctacgaaaa caccaccaa 480
 atcaagaat acttgaccgt tgatttgcca gaagttgata ttactaagcc aggtgttact 540
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 gtcaagttga tgaagctcat tttcgacttc gaatccatca gaaagttggt gtcctctcca 660
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 gatgctgata gaaatatggt tttgggtaag agattcttcg ttaccccatc tgattccggt 960
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 atggcatcat tggtaactt gcaatcatct ttgcctgaag ttaacaagat cgtaagggt 1440
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 gttttcagat tgcctggtac aggttctgaa ggtgctacta ttagattgta catcgaaca 1620
 tacgaaaatg acccatcaa gatctccaga gaatcttctg aagctttggc tccattggtt 1680
 gaagttgctt tgaattgtc caagatgcaa gaattcactg gtagatcagc tccaactggt 1740
 attacctga 1749

SEQ ID NO:145

R. suavis

MSSGKIKRVQ TTPFDGQKPG TSGLRKKVKV FTQPNYLQNF VQSTFNLPS DKVKGARLVV 60
 SGDGRYFSKE AIQIIKMAA GNGVKSVMVG QNGLLSTPAV SAVRERVGA DGCKASGAFI 120
 LTASHNPGGP NEDFGIKYNM ENGGPAPESI TNKIYENTTQ IKEYLTVDLP EVDITKPGVT 180
 TFEVEGGTFT VDVFSASDY VKLMKSI FDF ESIRKLLSSP KFTFCFDALH GVGAYAKRI 240
 FVEELGAKES SLLNCVPKED FGGGHPDNL TYAKELVARM GLSKSNTQNE PPEFGAAADG 300
 DADRNMLGK RFFVTPSDSV AI IANA VEA I PYFSTGLKG VARSMPTSAA LDVVAKHLNL 360
 KFFEVP TGWK FFGNLM DAGL CSVCGEESFG TGS DHI REKD GIWAVLAWLS IIAIKNKDNI 420
 GGDKLTVED IVRKH WATY G RHYYTRYDYE NVDAGKAKDL MASLVNLQSS LPEVNKIVKG 480
 ICSDVANVVG ADEFYKDSV DGSISKHOGI RYLFEDGSRL VFRLSGTGSE GATIRLYIEQ 540
 YENDPSKISR ESSEALAPLV EVALKLSKMQ EFTGRSAPT V IT 582

SEQ ID NO:146

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 gttcaatcta ctttcaacgc tttgtctgcc gaaaaagtta agggttctac tttggttggt 180

tccggtgatg	gtagatatta	ctccaaggat	gccattcaaa	tcatcattaa	gatggctgct	240
gctaacggtg	ttagaagagt	ttggggttgg	caaaatgggt	tggtgtctac	tccagctggt	300
tctgctggtg	ttagagaaaag	agttgggtgct	gatggttcta	aatctaacgg	tgctttcatt	360
ttgactgcct	ctcataatcc	aggtgggtcca	aatgaagatt	tcggtatcaa	gtacaacatg	420
gaaaaatggtg	gtccagctcc	agaaggtatt	actgataaga	tttttgaaaa	caccaagacc	480
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aagttcttcg	aagtccaac	tggttgaag	tttttcggta	atltgatgga	tgctgggttg	1140
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agacattact	acactagata	cgactacgaa	aacggtgatg	ctggtgctgc	aaaagaattg	1380
atggctcatt	tggttaagtt	gcaatcctcc	atctctgatg	ttaacacctt	cattaagggg	1440
atcagatccg	atgttgctaa	tggtgcatct	gctgatgaat	tcgaatacaa	ggatccagtt	1500
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attacctga						1749

SEQ ID NO:147

MASFKVNRVE	SSPIEGQKPG	TSGLRKKVKV	FTQPHYLHNF	VQSTFNALSA	EKVKGSTLVV	60
SGDGRYYSKD	AIQIIKMAA	ANGVRRVWVG	QNGLLSTPAV	SAVVRERVGA	DGSKSNGAFI	120
LTASHNPGGP	NEDFGIKYNM	ENGGPAPEGI	TDKIFENTKT	IKEYFIAEGL	PDVDISAIGI	180
SSFSGPDGQF	DVDVFDSSSD	YVKLMKSIFD	FQSIKKLITS	PQFSFCYDAL	HGVGGAYAKP	240
IFVDELGAKE	SSLLNCVPKE	DFGGGHPDPN	LTYAKELVSR	MGLGKNPDSN	PPEFGAAADG	300
DADRNMILGK	RFVTPSDSV	AIIAANAVQS	IPYFSSGLKG	VARSMPTSAA	LDVVAKSLNL	360
KFFEVP TGWK	FFGNLMDAGL	CSVCGEESFG	TGSDHIREKD	GIWAVLAWLS	ILAHKNKDNL	420
NGGNLVTVED	IVKQHWATYG	RHYTRYDYE	NVDAGAAKEL	MAHLVKLQSS	ISDVNTFIKG	480
IRSDVANVAS	ADEFYKDPV	DGSISKHQGI	RYLFEDGSRL	VFRLSGTGSE	GATIRLYIEQ	540
YEKSSKTGR	DSQEALAPLV	EVALKLSKMQ	EFTGRSAPTV	IT		582

SEQ ID NO:148

gcacacacca	tagcttcaaa	atgtttctac	tcctttttta	ctcttccaga	ttttctcgga	60
ctccgcgcat	cgccgtacca	cttcaaaaaca	cccaagcaca	gcatactaaa	tttcccctct	120
ttcttctct	agggtgtcgt	taattaccgg	tactaaaggt	ttggaaaaga	aaaaagagac	180
cgctcgttt	cttttcttc	gtcgaaaag	gcaataaaaa	tttttatcac	gtttctttt	240
cttgaaaatt	ttttttttg	atltttttct	ctttcgatga	cctcccattg	atatttaagt	300
taataaacgg	tcttcaattt	ctcaagtttc	agtttcattt	ttcttgttct	attacaactt	360
tttttacttc	ttgctcatta	gaaagaaagc	atagcaatct	aatctaagtt	ttaattacaa	420
ggatcc						426

SEQ ID NO:149

ggaagtacct	tcaaagaatg	gggtcttata	ttgttttgca	agtaccactg	agcaggataa	60
taatagaaat	gataatatac	tatagtagag	ataacgtcga	tgacttccca	tactgtaatt	120

gcttttagtt	gtgtattttt	agtgtgcaag	tttctgtaaa	tcgattaatt	tttttttctt	180
tcctcttttt	attaacctta	atttttatatt	tagattcctg	acttcaactc	aagacgcaca	240
gatattataa	catctgcata	ataggcattt	gcaagaatta	ctcgtgagta	aggaaagagt	300
gaggaactat	cgcatacctg	catttaaaga	tgccgatttg	ggcgcgaatc	ctttattttg	360
gcttcaccct	catactatta	tcagggccag	aaaaaggaag	tgtttccctc	cttcttgaat	420
tgatgttacc	ctcataaagc	acgtggcctc	ttatcgagaa	agaaattacc	gtcgcctcgtg	480
atgtgtttgc	aaaaagaaca	aaactgaaaa	aaccagaca	cgctcgactt	cctgtcttcc	540
tattgattgc	agcttccaat	ttcgtcacac	aacaaggtcc	tagcgacggc	tcacaggttt	600
tgtaacaagc	aatcgaaggt	tctggaatgg	cgggaaaggg	tttagtacca	catgctatga	660
tgcccactgt	gatctccaga	gcaaagtctg	ttcgcgta	ctgttactct	ctctctttca	720
aacagaattg	tccgaatcgt	gtgacaacaa	cagcctgttc	tcacacactc	ttttcttcta	780
accaaggggg	tggttagtt	tagtagaacc	tcgtgaaact	tacatttaca	tatatataaa	840
cttgcataaa	ttgtcaatg	caagaaatac	atatttggtc	ttttctaatt	cgtagttttt	900
caagttctta	gatgctttct	ttttctcttt	tttacagatc	atcaaggaag	taattatcta	960
ctttttacaa	caaatataaa	acaa				984

SEQ ID NO:150

cattatcaat	actgccattt	caaagaatac	gtaaataatt	aatagtagtg	attttcctaa	60
ctttatttag	tcaaaaaatt	agccttttaa	ttctgctgta	acccgtaac	gcccaaaata	120
gggggctggg	tacacagaat	atataacatc	gtaggtgtct	gggtgaacag	tttattcctg	180
gcatccacta	aatataatgg	agcccgtctt	ttaagctggc	atccagaaaa	aaaaagaatc	240
ccagcaccaa	aatattgttt	tcttcaccaa	ccatcagttc	ataggtccat	tctcttagcg	300
caactacaga	gaacaggggc	acaaacaggg	aaaaaacggg	cacaacctca	atggagtgat	360
gcaacctgcc	tgagtaaat	gatgacacaa	ggcaattgac	ccacgcatgt	atctatctca	420
ttttcttaca	ccttctatta	ccttctgctc	tctctgattt	ggaaaaagct	gaaaaaaaag	480
ggtgaaacca	gttccctgaa	attattcccc	tacttgacta	ataagtatat	aaagacggta	540
ggtattgatt	gtaattctgt	aaatctatct	cttaaacttc	ttaaattcta	cttttatagt	600
tagtcttttt	tttagtttta	aaacaccaag	aacttagttt	cgaataaaca	cacataaaca	660
aacaaa						666

SEQ ID NO:151

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gtgagggcgc	cataaccaag	gtatctatag	accgccaatc	agcaaactac	ctccgtacat	120
tcatgttgca	cccacacatt	tatacaccca	gaccgcgaca	aattacccat	aaggttgttt	180
gtgacggcgt	cgtacaagag	aacgtgggaa	cttttttaggc	tcaccaaaaa	agaaagaaaa	240
aatacgagtt	gctgacagaa	gcctcaagaa	aaaaaaaaatt	cttcttcgac	tatgctggag	300
gcagagatga	tcgagccggt	agttaactat	atatagctaa	attggttcca	tcaccttctt	360
ttctgggtgc	gctccttcta	gtgctatctc	tggtctttcc	tatttttttt	tttccatttt	420
tctttctctc	tttctaatat	ataaattctc	ttgcattttc	tatttttctc	tctatctatt	480
ctacttggtt	attcccttca	aggttttttt	ttaaggagta	cttgttttta	gaatatacgg	540
tcaacgaact	ataattaact	aaaca				565

SEQ ID NO:152

agttataata	atcctacgtt	agtgtgagcg	ggatttaaac	tgtgaggacc	ttaatacatt	60
cagacacttc	tgcggtatca	ccctacttat	tccttctgag	attatatcta	ggaaccctac	120
agggttggtg	aagattacc	gttctaagac	ttttcagctt	cctctattga	tgttacacct	180
ggacaccctt	ttctgycat	ccagttttta	atcttcagtg	gcatgtgaga	ttctccgaaa	240
ttaattaaag	caatcacaca	attctctcgg	ataccacctc	ggttgaaact	gacaggtggt	300
ttgttacgca	tgctaatgca	aaggagccta	tatacctttg	gctcggctgc	tgtaacaggg	360
aatataaagg	gcagcataat	ttaggagttt	agtgaacttg	caacatttac	tattttccct	420
tcttacgtaa	atatttttct	ttttaattct	aaatcaatct	ttttcaattt	tttgtttgta	480
ttcttttctt	gcttaaatct	ataactacaa	aaaacacata	cataaactaa	aa	532

SEQ ID NO:153

gatctatgcg	actgggtgag	catatgttcc	gctgatgtga	tgtgcaagat	aaacaagcaa	60
ggcagaaact	aacttcttct	tcatgtaata	aacacacccc	gcgtttat	acctatctct	120
aaacttcaac	accttatatc	ataactaata	tttcttgaga	taagcacact	gcaccatac	180
cttccttaaa	aacgtagctt	ccagtttttg	gtggttccgg	cttccttccc	gattccgccc	240
gctaaacgca	tatTTTTgtt	gcctgggtggc	atttgcaaaa	tgcataacct	atgcatttaa	300
aagattatgt	atgctcttct	gacttttctg	gtgatgaggc	tcgtggaaaa	aatgaataat	360
ttatgaattt	gagaacaatt	ttgtgttggt	acggtat	actatggaat	aatcaatcaa	420
ttgaggattt	tatgcaaata	tcgtttggaat	atTTTTccga	ccctttgagt	acttttcttc	480
ataattgcat	aatattgtcc	gctgcccctt	tttctgttag	acgggtgtctt	gatctacttg	540
ctatcgttca	acaccacctt	atTTTtctaac	tatTTTTttt	ttagctcatt	tgaatcagct	600
tatgggtgatg	gcacattttt	gcataaacct	agctgtcctc	gttgaacata	ggaaaaaaaa	660
atatataaac	aaggctcttt	cactctcctt	gcaatcagat	ttgggtttgt	tccctttatt	720
ttcatat	ttgtcatatt	cctttctcaa	ttattat	ctactcataa	cctcacgcaa	780
aataacacag	tcaaatctat	caaaa				805

SEQ ID NO:154

atccgctcta	accgaaaagg	aaggagttag	acaacctgaa	gtctaggtcc	ctatttattt	60
tttttaatag	ttatgttagt	attaagaacg	ttatttatat	ttcaaatttt	tctttttttt	120
ctgtacaaac	gcgtgtacgc	atgtaacatt	atactgaaaa	ccttgcttga	gaaggttttg	180
ggacgctcga	ag					192

SEQ ID NO:155

gtagatacgt	tgttgacact	tctaaataag	cgaatttctt	atgatttatg	atTTTTatta	60
ttaaataagt	tataaaaaaa	ataagtgtat	acaaatttta	aagtgactct	taggttttaa	120
aacgaaaatt	cttattcttg	agtaactctt	tcctgtaggt	caggttgctt	tctcaggtat	180
agcatgaggt	cgctc					195

WHAT IS CLAIMED IS:

1. A recombinant host cell capable of producing one or more steviol glycosides or a steviol glycoside composition in a cell culture, comprising:
 - (a) a recombinant gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP);
 - (b) a recombinant gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate; and/or
 - (c) a recombinant gene encoding a polypeptide capable of synthesizing uridine diphosphate glucose (UDP-glucose) from UTP and glucose-1-phosphate.

2. The recombinant host cell of claim 1, wherein:
 - (a) the polypeptide capable of synthesizing UTP from UDP comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
 - (b) the polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, SEQ ID NO:143 or a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:141, SEQ ID NO:145, or SEQ ID NO:147; and/or
 - (c) the polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate comprises a polypeptide having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:127, a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139 or a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131.

3. The recombinant host cell of claim 1 or 2, further comprising:

- (a) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof;
 - (b) a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;
 - (c) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof; and/or
 - (d) a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside.
4. The recombinant host cell of claim 3, further comprising:
- (e) a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP);
 - (f) a gene encoding a polypeptide capable of synthesizing *ent*-copalyl diphosphate from GGPP;
 - (g) a gene encoding an a polypeptide capable of synthesizing *ent*-kaurene from *ent*-copalyl diphosphate;
 - (h) a gene encoding a polypeptide capable of synthesizing *ent*-kaurenoic acid from *ent*-kaurene;
 - (i) a gene encoding a polypeptide capable of reducing cytochrome P450 complex; and/or
 - (j) a gene encoding a polypeptide capable of synthesizing steviol from *ent*-kaurenoic acid.
5. The recombinant host cell of claim 3 or 4, wherein:
- (a) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;
 - (b) the polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a

- steviol glycoside comprises a polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (c) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
 - (d) the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16;
 - (e) the polypeptide capable of synthesizing GGPP comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, or SEQ ID NO:116;
 - (f) the polypeptide capable of synthesizing ent-copalyl diphosphate comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, SEQ ID NO:42, or SEQ ID NO:120;
 - (g) the polypeptide capable of synthesizing ent-kaurene comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:52;
 - (h) the polypeptide capable of synthesizing ent-kaurenoic acid comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:117, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, or SEQ ID NO:76;
 - (i) the polypeptide capable of reducing cytochrome P450 complex comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82,

SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92; and/or

- (k) the polypeptide capable of synthesizing steviol comprises a polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:94, SEQ ID NO:97, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO:110, SEQ ID NO:112, or SEQ ID NO:114.
6. The recombinant host cell of any one of claims 1-5, wherein the recombinant host cell comprises:
- (a) a gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP) having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
 - (b) one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, each having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2 and/or SEQ ID NO:119; and
 - (c) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121;
7. The recombinant host cell of any one of claims 1-6, wherein the recombinant host cell comprises:
- (a) a gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP);
 - (b) a gene encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate;
 - (c) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121;
 - (d) a gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 55% sequence

identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129; SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139; or at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:127; or at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131; and

one or more of:

- (e) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;
- (b) a gene encoding a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (c) a gene encoding a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
- (d) a gene encoding a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16.

8. The recombinant host cell of any one of claims 1-7, wherein the recombinant host cell comprises:

- (a) a recombinant gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP) having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
- (b) one or more recombinant genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, each having at

least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2 and/or SEQ ID NO:119; and/or

- (c) a recombinant gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121;

wherein the gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP), the one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or the gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate are overexpressed relative to a corresponding host cell lacking the one or more recombinant genes.

9. The recombinant host cell of claim 8, wherein the gene encoding a polypeptide capable of synthesizing uridine 5'-triphosphate (UTP) from uridine diphosphate (UDP), the one or more genes encoding a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate, and/or the gene encoding a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate are overexpressed by at least 10%, or at least 15%, or at least 20%, or at least 30%, or at least 40%, or at least 50%, or at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 100%, or at least 125%, or at least 150%, or at least 175%, or at least 200% relative to a corresponding host cell lacking the one or more recombinant genes.
10. The recombinant host cell of any one of claims 1-9, wherein expression of the one or more recombinant genes increase the amount of UDP-glucose accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.
11. The recombinant host cell of claim 10, wherein expression of the one or more recombinant genes increases the amount of UDP-glucose accumulated by the cell by at least about 10%, at least about 25%, or at least about 50%, at least about 100%, at least about 150%, at least about 200%, or at least about 250% relative to a corresponding host lacking the one or more recombinant genes.

12. The recombinant host cell of any one of claims 1-11, wherein expression of the one or more recombinant genes increases an amount of the one or more steviol glycosides or the steviol glycoside composition produced by the cell relative to a corresponding host lacking the one or more recombinant genes.
13. The recombinant host cell of claim 12, wherein expression of the one or more recombinant genes increases the amount of the one or more steviol glycosides produced by the cell by at least about 5%, at least about 10%, at least about 25%, at least about 50%, at least about 75%, or at least about 100% relative to a corresponding host lacking the one or more recombinant genes.
14. The recombinant host cell of claim 12 or 13, wherein expression of the one or more recombinant genes increases the amount of RebA, RebB, Reb D, and/or RebM produced by the cell relative to a corresponding host lacking the one or more recombinant genes.
15. The recombinant host cell of any one of claims 1-14, wherein expression of the one or more recombinant genes decreases the one of one or more steviol glycosides or the steviol glycoside composition accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.
16. The recombinant host cell of claim 15, wherein expression of the one or more recombinant genes decreases the amount of the one or more steviol glycosides accumulated by the cell by at least about 5%, at least about 10%, at least about 25%, or at least about 50% relative to a corresponding host lacking the one or more recombinant genes.
17. The recombinant host cell of claim 15 or 16, wherein expression of the one or more recombinant genes decreases the amount of RebB, RebD, and/or 13-SMG accumulated by the cell relative to a corresponding host lacking the one or more recombinant genes.
18. The recombinant host cell of any one of claims 1-17, wherein expression of the one or more recombinant genes increases or decreases the amount of total steviol glycosides

- produced by the cell by less than 5%, less than 2.5%, or less than 1% relative to a corresponding host lacking the one or more recombinant genes.
19. The recombinant host cell of any one of claims 1-17, wherein expression of the one or more recombinant genes increases the amount of total steviol glycosides produced by the cell by at least about 5%, at least about 10%, or at least about 25% relative to a corresponding host lacking the one or more recombinant genes.
 20. The recombinant host cell of any one of claims 1-18, wherein the one or more steviol glycosides is, or the steviol glycoside composition comprises, steviol-13-O-glucoside (13-SMG), steviol-1,2-Bioside, steviol-1,3-Bioside, steviol-19-O-glucoside (19-SMG), 1,2-Stevioside, 1,3-stevioside (RebG), rubusoside, rebaudioside A (RebA), rebaudioside B (RebB), rebaudioside C (RebC), rebaudioside D (RebD), rebaudioside E (RebE), rebaudioside F (RebF), rebaudioside M (RebM), rebaudioside Q (RebQ), rebaudioside I (RebI), dulcoside A, and/or an isomer thereof.
 21. The recombinant host cell of any one of claims 1-20, wherein the recombinant host cell is a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell or a bacterial cell.
 22. A method of producing one or more steviol glycosides or a steviol glycoside composition in a cell culture, comprising culturing the recombinant host cell of any one of claims 1-21 in the cell culture, under conditions in which the genes are expressed, and wherein the one or more steviol glycosides or the steviol glycoside composition is produced by the recombinant host cell.
 23. The method of claim 22, wherein the genes are constitutively expressed and/or expression of the genes is induced.
 24. The method of claim 22 or 23, wherein the amount of UDP-glucose accumulated by the cell is increased by at least about 10% relative to a corresponding host lacking the one or more recombinant genes.

25. The method of any one of claims 22-24, wherein the amount of RebA, RebB, RebD, and/or RebM produced by the cell is increased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.
26. The method of any one of claims 22-25, wherein the amount of RebB, RebD, and/or 13-SMG accumulated by the cell is decreased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.
27. The method of any one of claims 22-26, wherein the amount of total steviol glycosides produced by the cell is increased or decreased by less than about 5% relative to a corresponding host lacking the one or more recombinant genes.
28. The method of any one of claims 22-26, wherein the amount of total steviol glycosides produced by the cell is increased by at least about 5% relative to a corresponding host lacking the one or more recombinant genes.
29. The method of any one of claims 22-28, wherein the recombinant host cell is grown in a fermentor at a temperature for a period of time, wherein the temperature and period of time facilitate the production of the one or more steviol glycosides or the steviol glycoside composition.
30. The method of claim 29, wherein the amount of UDP-glucose present in the cell culture is increased by at least about 10%, at least about 25%, or at least about 50%, at least about 100%, at least about 150%, at least about 200%, or at least about 250% at any point throughout the period of time.
31. The method of any one of claims 22-30, further comprising isolating the produced one or more steviol glycosides or the steviol glycoside composition from the cell culture.
32. The method of claim 31, wherein the isolating step comprises:
 - (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;

- (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
- (c) providing one or more adsorbent resins, comprising providing the adsorbent resins in a packed column; and
- (d) contacting the supernatant of step (b) with the one or more adsorbent resins in order to obtain at least a portion of the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition;

or

- (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;
- (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
- (c) providing one or more ion exchange or ion exchange or reversed-phase chromatography columns; and
- (d) contacting the supernatant of step (b) with the one or more ion exchange or ion exchange or reversed-phase chromatography columns in order to obtain at least a portion of the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition;

or

- (a) providing the cell culture comprising the one or more steviol glycosides or the steviol glycoside composition;
- (b) separating a liquid phase of the cell culture from a solid phase of the cell culture to obtain a supernatant comprising the produced one or more steviol glycosides or the steviol glycoside composition;
- (c) crystallizing or extracting the produced one or more steviol glycosides or the steviol glycoside composition, thereby isolating the produced one or more steviol glycosides or the steviol glycoside composition.

33. The method of any one of claims 22-30, further comprising recovering the one or more steviol glycosides or the steviol glycoside composition from the cell culture.
34. The method of claim 33, wherein the recovered one or more steviol glycosides or the steviol glycoside composition has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.
35. A method for producing one or more steviol glycosides or a steviol glycoside composition, comprising whole-cell bioconversion of plant-derived or synthetic steviol and/or steviol glycosides in a cell culture medium of a recombinant host cell using:
- (a) a polypeptide capable of synthesizing UTP from UDP having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:123;
 - (b) a polypeptide capable of converting glucose-6-phosphate to glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:2, SEQ ID NO:119, or SEQ ID NO:143; or at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:141, SEQ ID NO:145, or SEQ ID NO:147; and/or
 - (c) a polypeptide capable of synthesizing UDP-glucose from UTP and glucose-1-phosphate having at least 60% sequence identity to the amino acid sequence set forth in SEQ ID NO:121, SEQ ID NO:127; at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:125, SEQ ID NO:129, SEQ ID NO:133, SEQ ID NO:135, SEQ ID NO:137, or SEQ ID NO:139; or at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:131, and
- one or more of:
- (d) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof;
 - (e) a polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;
 - (f) a polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof; and/or

- (g) a polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside;

wherein at least one of the polypeptides is a recombinant polypeptide expressed in the recombinant host cell; and producing the one or more steviol glycosides or the steviol glycoside composition thereby.

36. The method of claim 35, wherein:

- (d) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-13 hydroxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7;
- (e) the polypeptide capable of beta 1,3 glycosylation of the C3' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- (f) the polypeptide capable of glycosylating steviol or a steviol glycoside at its C-19 carboxyl group thereof comprises a polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:4;
- (g) the polypeptide capable of beta 1,2 glycosylation of the C2' of the 13-O-glucose, 19-O-glucose, or both 13-O-glucose and 19-O-glucose of a steviol glycoside comprises a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:11; a polypeptide having 80% or greater identity to the amino acid sequence set forth in SEQ ID NO:13; or a polypeptide having at least 65% sequence identity to the amino acid sequence set forth in SEQ ID NO:16.

37. The method of any one of claims 22-36, wherein the recombinant host cell is a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell or a bacterial cell.

38. The method of any one of claims 22-37, wherein the one or more steviol glycosides is, or the steviol glycoside composition comprises, steviol-13-O-glucoside (13-SMG), steviol-

1,2-Bioside, steviol-1,3-Bioside, steviol-19-O-glucoside (19-SMG), 1,2-stevioside, 1,3-stevioside (RebG), rubusoside, rebaudioside A (RebA), rebaudioside B (RebB), rebaudioside C (RebC), rebaudioside D (RebD), rebaudioside E (RebE), rebaudioside F (RebF), rebaudioside M (RebM), rebaudioside Q (RebQ), rebaudioside I (RebI), dulcoside A, and/or an isomer thereof.

39. A cell culture, comprising the recombinant host cell of any one of claims 1-21, the cell culture further comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids;

wherein the one or more steviol glycosides or the steviol glycoside composition is present at a concentration of at least 1 mg/liter of the cell culture;

wherein the cell culture is enriched for the one or more steviol glycosides or the steviol glycoside composition relative to a steviol glycoside composition from a Stevia plant and has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

40. A cell culture, comprising the recombinant host cell of any one of claims 1-21, the cell culture further comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids;

wherein UDP-glucose is present in the cell culture at a concentration of at least 100 μ M;

wherein the cell culture is enriched for UGP-glucose relative to a steviol glycoside composition from a Stevia plant and has a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

41. A cell lysate from the recombinant host cell of any one of claims 1-21 grown in the cell culture, comprising:

- (a) the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell;
- (b) glucose, fructose, sucrose, xylose, rhamnose, UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, yeast nitrogen base, YNB, and/or amino acids;

wherein the one or more steviol glycosides or the steviol glycoside composition produced by the recombinant host cell is present at a concentration of at least 1 mg/liter of the cell culture.

42. One or more steviol glycosides produced by the recombinant host cell of any one of claims 1-21;

wherein the one or more steviol glycosides produced by the recombinant host cell are present in relative amounts that are different from a steviol glycoside composition from a Stevia plant and have a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

43. One or more steviol glycosides produced by the method of any one of claims 22-38;

wherein the one or more steviol glycosides produced by the recombinant host cell are present in relative amounts that are different from a steviol glycoside composition from a Stevia plant and have a reduced level of Stevia plant-derived components relative to a plant-derived Stevia extract.

44. A sweetener composition, comprising the one or more steviol glycosides of claim 42 or 43.

45. A food product comprising, the sweetener composition of claim 44.

46. A beverage or a beverage concentrate, comprising the sweetener composition of claim 44.

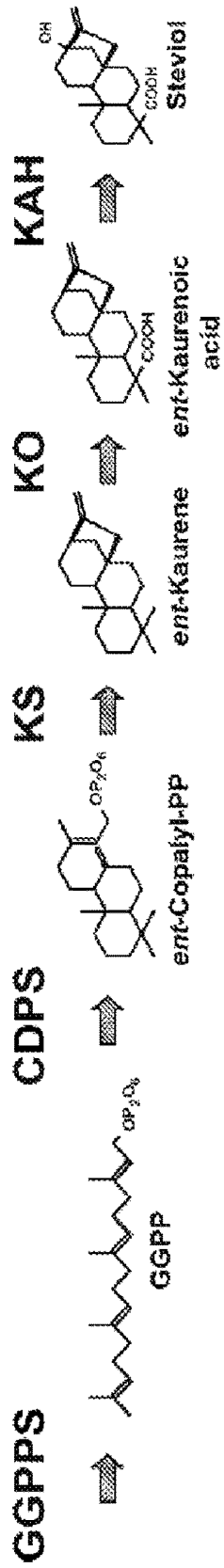


Figure 1

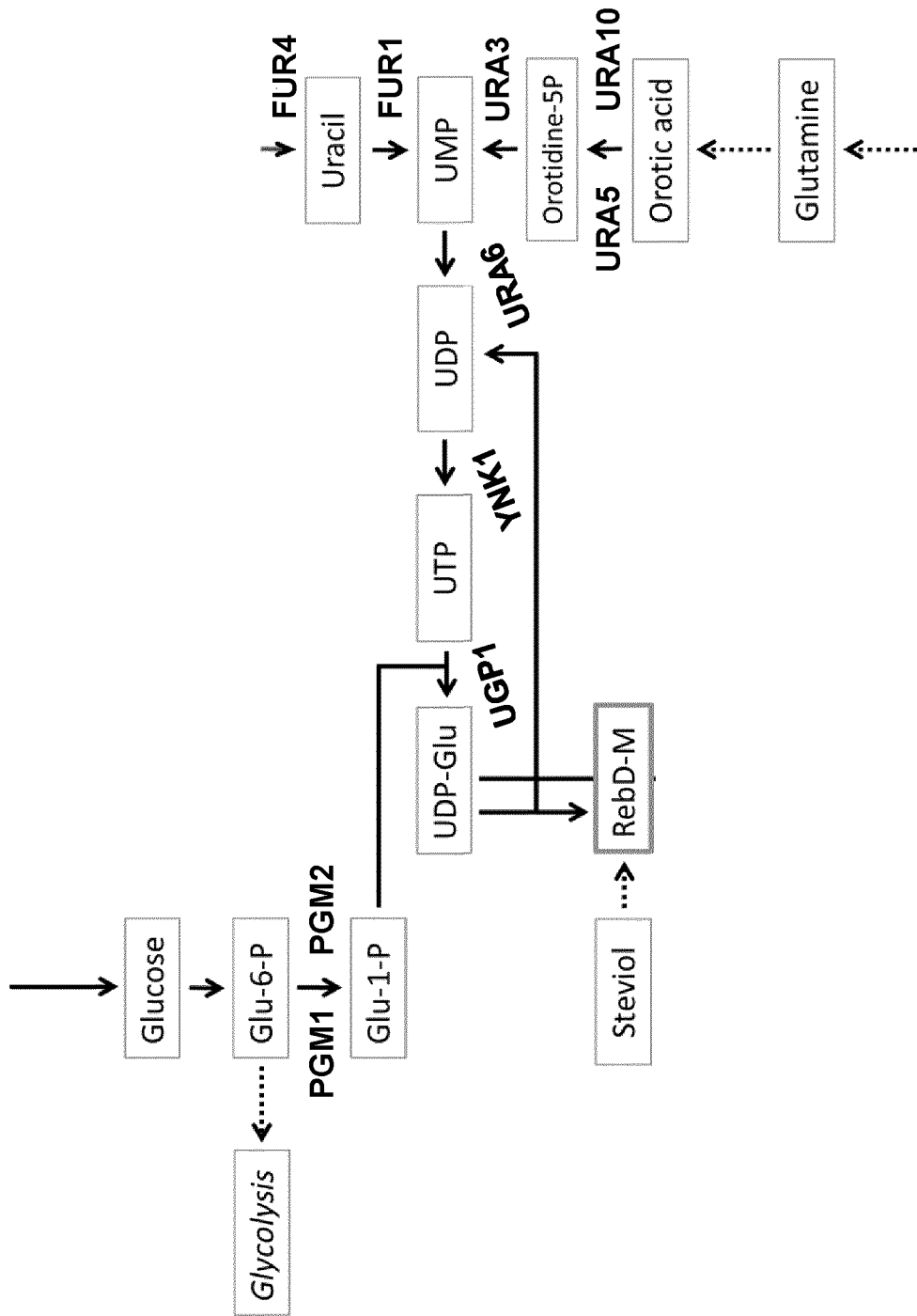


Figure 3

INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2017/059028

A. CLASSIFICATION OF SUBJECT MATTER
INV. A23L27/30 A23L2/60 C12N15/63
ADD.
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED
Minimum documentation searched (classification system followed by classification symbols)
A23L C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal, BIOSIS, COMPENDEX, EMBASE, FSTA, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	KR 2015 0000258 A (KOREA RES INST OF BIOSCIENCE [KR]) 2 January 2015 (2015-01-02)	42-46
Y	the whole document, in particular Figure 1 and claims	1-46
X	WO 2015/014969 A1 (DSM IP ASSETS BV [NL]) 5 February 2015 (2015-02-05)	42-46
Y	the whole document, in particular the claims	1-46
X	WO 2016/038095 A2 (EVOLVA SA [CH]; ROBERTSEN HELENE LUNDE [DK]; ANDERSEN IBEN NORDMARK [D]) 17 March 2016 (2016-03-17)	42-46
Y	the whole document, in particular the claims	1-46
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Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier application or patent but published on or after the international filing date
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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search 2 June 2017	Date of mailing of the international search report 27/06/2017
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Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer Bassias, Ioannis
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INTERNATIONAL SEARCH REPORT

International application No
PCT/EP2017/059028

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2014/122328 A1 (EVOLVA SA [CH]) 14 August 2014 (2014-08-14)	42-46
Y	the whole document, in particular the claims	1-46

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