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DETERGENT COMPOSITIONS COMPRISING POLYPEPTIDES HAVING XANTHAN DEGRADING ACTIVITY

Reference to a Sequence Listing

This application contains a Sequence Listing in computer readable form, which is incorporated herein by reference.

Background of the invention

Field of the invention

The present invention relates to detergent compositions comprising polypeptides having xanthan degrading activity. In particular the invention relates to such detergent compositions comprising polypeptides within the glycosyl hydrolase family 5 (GH5) having xanthan degrading activity. The invention also relates to methods for producing said detergent compositions and to the use of said detergent compositions in cleaning applications.

Description of the related art

Xanthan gum is a polysaccharide secreted by the bacterium *Xanthomonas campestris*. It is produced by the fermentation of glucose, sucrose, or lactose in an aqueous growth medium by *X. campestris*. After a fermentation period, the polysaccharide is precipitated from the growth medium with isopropyl alcohol, dried, and ground into a fine powder. Later, the powder is added to a liquid medium to form the gum.

Xanthan is composed of pentasaccharide subunits, forming a cellulose backbone with trisaccharide side chains composed of mannose-(beta1,4)-glucuronic-acid-(beta1,2)-mannose attached to alternate glucose residues in the backbone by alpha1,3 linkages. This biopolymer is of great commercial significance because of its superior pseudoplasticity, thixotropy, and viscosity.

In recent years xanthan gum has been widely used as an ingredient in many consumer products including foods (e.g., as thickening agent in salat dressings and dairy products) and cosmetics (e.g., as stabilizer and thickener in toothpaste and make-up to prevent ingredients from separating) and cosmetics (e.g., sun creams).

In addition, xanthan gum has found use in the oil industry where xanthan gum is used in large quantities to thicken drilling mud. These fluids serve to carry the solids cut by the drilling bit back to the surface. When the circulation stops, the solids still remain suspended in the drilling fluid. The widespread use of horizontal drilling has led to its expanded use. Xanthan gum is also added to self-consolidating concrete, including concrete poured underwater, to increase its viscosity.

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The widespread use of xanthan gum has led to a desire to be able to degrade solutions or gels of xanthan gum. Complete enzymatic degradation of xanthan gum has till now required several enzymatic activities including xanthan lyase activity and endo-beta-1,4-glucanase activity. Xanthan lyases are enzymes that cleave the beta-D-mannosylalpha-beta-D-1,4-glucuronosyl bond of xanthan and have been described in the literature. Xanthan degrading enzymes are known in the art e.g., two xanthan lyases isolated from *Paenibacillus alginolyticus* XL-1.

Glycosyl hydrolases are enzymes that catalyze the hydrolysis of the glycosyl bond to release smaller sugars. There are over 100 classes of Glycosyl hydrolases which have been classified.. The glycosyl hydrolase family 5 (GH5) includes endo-glucanases (EC 3.2.1.4), endo-beta-1,4-xylanase (EC 3.2.1.8); beta-glucosidase (EC 3.2.1.21); beta-mannosidase (EC 3.2.1.25). However, until now identification of xanthan degrading enzymes have not been reported in glycosyl hydrolase family 5.

The mature peptide in SEQ ID NO: 2 is 45 % identical and the mature peptide in SEQ ID NO: 4 is 57 % identical to a predicted endoglucanase from the genome of *Echinicola vietnamensis* (UNIPROT: L0FVA9).

The mature peptide in SEQ ID NO: 6 is 47 % identical to an uncharacterized protein from the genome of *Barnesiella intestinihominis* (UNIPROT: K0WXE1).

The mature peptide in SEQ ID NO: 8 is 100 % identical to an uncharacterized protein from the genome of *Pseudomonas stutzeri* (UNIPROT: M2V1S3).

Summary of the invention

The invention provides new and improved detergent compositions comprising enzymes for the degradation of xanthan gum and methods for producing said detergent compositions and to the use of said detergent compositions in cleaning applications.

The present inventors have surprisingly discovered a new group of enzymes that have xanthan degrading activity – and which do not belong to any glycosyl hydrolase family previously known to comprise this enzymatic activity. The enzymes have no significant sequence similarity to any known enzyme having xanthan degrading activity.

The present invention provides detergent compositions comprising polypeptides having xanthan degrading activity, i.e., having activity on xanthan gum and/or having activity on xanthan gum pretreated with xanthan lyase.

Accordingly, the present invention provides a detergent composition comprising polypeptide of glycosyl hydrolase family 5 having xanthan degrading activity. More particularly, the present invention provides a detergent composition comprising polypeptide of glycosyl hydrolase family 5 having xanthan degrading activity, selected from the group consisting of:

(a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at

least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide of any of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 8;

- (b) a polypeptide encoded by a polynucleotide that hybridizes under medium stringency conditions with (i) the mature polypeptide coding sequence of any of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 7, (ii), or the full-length complement of (i);
- (c) a polypeptide encoded by a polynucleotide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide coding sequence of any of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5 or SEQ ID NO: 7;
- (d) a variant of the mature polypeptide of any of SEQ ID NO: 2 SEQ ID NO: 4, SEQ ID NO: 6 or SEQ ID NO: 8 comprising a substitution, deletion, and/or insertion at one or more positions;
- (e) a fragment of the polypeptide of (a), (b), (c), or (d) that has xanthan degrading activity; and
- (f) a polypeptide comprising the polypeptide of (a), (b), (c), (d), or (e) and a N-terminal and/or C-terminal His-tag.

The present invention also relates to methods of degrading xanthan gum using the detergent composition comprising the polypeptides.

Overview of Sequence Listing

SEQ ID NO: 1 is the DNA sequence of the EXa gene as isolated from an Opitutaceae sp.

SEQ ID NO: 2 is the amino acid sequence of the EXa GH5 polypeptide as deduced from SEQ ID NO: 1.

SEQ ID NO: 3 is the DNA sequence of the EXb gene as isolated from an environmental sample

SEQ ID NO: 4 is the amino acid sequence of the EXb GH5 polypeptide as deduced from SEQ ID NO: 3.

SEQ ID NO: 5 is the DNA sequence of the EXc gene as isolated from an environmental sample

SEQ ID NO: 6 is the amino acid sequence of the EXc GH5 polypeptide as deduced from SEQ ID NO: 5.

SEQ ID NO: 7 is the DNA sequence of the EXd gene as obtained from a public database (UNIPROT M2V1S3, originating from a strain of *Pseudomonas stutzeri* collected from a Galapagos Rift hydrothermal vent, Ecuador).

SEQ ID NO: 8 is the amino acid sequence of the EXd GH5 polypeptide as deduced from SEQ ID NO: 7.

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SEQ ID NO:9 is synth codon optimized DNA encoding the EXa GH5 polypeptide.

SEQ ID NO:10 is synth codon optimized DNA encoding the EXb GH5 polypeptide.

SEQ ID NO:11 is synth codon optimized DNA encoding the EXc GH5 polypeptide.

SEQ ID NO:12 is synth codon optimized DNA encoding the EXd GH5 polypeptide.

SEQ ID NO:13 is the EXa GH5 polypeptide + His affinity tag expressed in *E.coli*.

SEQ ID NO:14 is the EXb GH5 polypeptide + His affinity tag expressed in *E.coli*.

SEQ ID NO:15 the EXc GH5 polypeptide + His affinity tag expressed in E.coli.

SEQ ID NO:16 is the EXb GH5 polypeptide + His affinity tag expressed in B. subtilis.

SEQ ID NO:17 is the EXc GH5 polypeptide + His affinity tag expressed in B. subtilis.

SEQ ID NO:18 is the EXd GH5 polypeptide + His affinity tag expressed in B. subtilis.

SEQ ID NO:19 is the His affinity tag sequence.

SEQ ID NO:20 is the amino acid sequence of the Bacillus clausii secretion signal.

SEQ ID NO:21 is the amino acid sequence of a xanthan lyase XLa from a *Paenibacillus* sp (SEQ ID NO: 8 from WO2013167581).

SEQ ID NO:22 is the amino acid sequence of a xanthan lyase XLb from a *Paenibacillus* sp (SEQ ID NO: 66 from WO2013167581).

SEQ ID NO:23 is the amino acid sequence of a xanthan lyase XLc from a *Paenibacillus* sp (SEQ ID NO: 68 from WO2013167581).

SEQ ID NO:24 is the amino acid sequence of a xanthan lyase XLd from a *Paenibacillus* sp (SEQ ID NO: 120 from WO2013167581).

Identity Matrix for mature peptides							
	SEQ ID NO:2 EXa	SEQ ID NO:4 EXb	SEQ ID NO:6 EXc	SEQ ID NO:8 EXd			
SEQ ID NO:2 EXa		50	71	27			
SEQ ID NO:4 EXb			47	31			
SEQ ID NO:6 EXc				27			
SEQ ID NO:8 EXd							

Detailed Description of the Invention

The present invention provides detergent composition comprising GH5 polypeptides having xanthan degrading activity. The polypeptides do not belong to a GH family known to comprise enzymes, which degrade xanthan. In addition, the detergent composition comprising a combination of xanthan lyase and an enzyme of the invention having xanthan degrading activity shows a synergistic improved wash performance over using a detergent composition comprising either a xanthan lyase or a GH5 polypeptide alone having xanthan degrading activity.

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Definitions

Coding sequence: The term "coding sequence" means a polynucleotide, which directly specifies the amino acid sequence of a polypeptide. The boundaries of the coding sequence are generally determined by an open reading frame, which begins with a start codon such as ATG, GTG, or TTG and ends with a stop codon such as TAA, TAG, or TGA. The coding sequence may be a genomic DNA, cDNA, synthetic DNA, or a combination thereof.

Colour clarification: During washing and wearing loose or broken fibers can accumulate on the surface of the fabrics. One consequence can be that the colours of the fabric appear less bright or less intense because of the surface contaminations. Removal of the loose or broken fibers from the textile will partly restore the original colours and looks of the textile. By the term "colour clarification", as used herein, is meant the partial restoration of the initial colours of textile.

Detergent Composition:

The term "detergent composition", includes unless otherwise indicated, granular or powderform all-purpose or heavy-duty washing agents, especially cleaning detergents; liquid, gel or pasteform all-purpose washing agents, especially the so- called heavy-duty liquid (HDL) types; liquid finefabric detergents; hand dishwashing agents or light duty dishwashing agents, especially those of the high-foaming type; machine dishwashing agents, including the various tablet, granular, liquid and rinse-aid types for household and institutional use; liquid cleaning and disinfecting agents, including antibacterial hand-wash types, cleaning bars, soap bars, mouthwashes, denture cleaners, car or carpet shampoos, bathroom cleaners; hair shampoos and hair-rinses; shower gels, foam baths; metal cleaners; as well as cleaning auxiliaries such as bleach additives and "stain-stick" or pre-treat types. The terms "detergent composition" and "detergent formulation" are used in reference to mixtures which are intended for use in a wash medium for the cleaning of soiled objects. In some embodiments, the term is used in reference to laundering fabrics and/or garments (e.g., "laundry detergents"). In alternative embodiments, the term refers to other detergents, such as those used to clean dishes, cutlery, etc. (e.g., "dishwashing detergents"). It is not intended that the present invention be limited to any particular detergent formulation or composition. The term "detergent composition" is not intended to be limited to compositions that contain surfactants. It is intended that in addition to the variants according to the invention, the term encompasses detergents that may contain, e.g., surfactants, builders, chelators or chelating agents, bleach system or bleach components, polymers, fabric conditioners, foam boosters, suds suppressors, dyes, perfume, tannish inhibitors, optical brighteners, bactericides, fungicides, soil suspending agents, anticorrosion agents, enzyme inhibitors or stabilizers, enzyme activators, transferase(s), hydrolytic enzymes, oxido reductases, bluing agents and fluorescent dyes, antioxidants, and solubilizers.

Dish wash: The term "dish wash" refers to all forms of washing dishes, e.g., by hand or automatic dish wash. Washing dishes includes, but is not limited to, the cleaning of all forms of crockery such as plates, cups, glasses, bowls, all forms of cutlery such as spoons, knives, forks and serving utensils as well as ceramics, plastics, metals, china, glass and acrylics.

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Dish washing composition: The term "dish washing composition" refers to all forms of compositions for cleaning hard surfaces. The present invention is not restricted to any particular type of dish wash composition or any particular detergent.

Enzyme Detergency benefit: The term "enzyme detergency benefit" is defined herein as the advantageous effect an enzyme may add to a detergent compared to the same detergent without the enzyme. Important detergency benefits which can be provided by enzymes are stain removal with no or very little visible soils after washing and or cleaning, prevention or reduction of redeposition of soils released in the washing process an effect that also is termed anti-redeposition, restoring fully or partly the whiteness of textiles, which originally were white but after repeated use and wash have obtained a greyish or yellowish appearance an effect that also is termed whitening. Textile care benefits, which are not directly related to catalytic stain removal or prevention of redeposition of soils are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye transfer from one fabric to another fabric or another part of the same fabric an effect that is also termed dye transfer inhibition or anti-backstaining, removal of protruding or broken fibers from a fabric surface to decrease pilling tendencies or remove already existing pills or fuzz an effect that also is termed anti-pilling, improvement of the fabric-softness, colour clarification of the fabric and removal of particulate soils which are trapped in the fibers of the fabric or garment. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching component such as hydrogen peroxide or other peroxides.

Fragment: The term "fragment" means a polypeptide having one or more (*e.g.*, several) amino acids absent from the amino and/or carboxyl terminus of a mature polypeptide or domain; wherein the fragment has xanthan degrading activity.

Hard surface cleaning: The term "Hard surface cleaning" is defined herein as cleaning of hard surfaces wherein hard surfaces may include floors, tables, walls, roofs etc. as well as surfaces of hard objects such as cars (car wash) and dishes (dish wash). Dish washing includes but are not limited to cleaning of plates, cups, glasses, bowls, and cutlery such as spoons, knives, forks, serving utensils, ceramics, plastics, metals, china, glass and acrylics.

Improved wash performance: The term "improved wash performance" is defined herein as a (variant) enzyme (also a blend of enzymes, not necessarily only variants but also backbones, and in combination with certain cleaning composition etc.) displaying an alteration of the wash performance of a protease variant relative to the wash performance of the parent protease variant e.g. by increased stain removal. The term "wash performance" includes wash performance in laundry but also e.g. in dish wash.

Isolated: The term "isolated" means a substance in a form or environment that does not occur in nature. Non-limiting examples of isolated substances include (1) any non-naturally occurring substance, (2) any substance including, but not limited to, any enzyme, variant, nucleic acid, protein, peptide or cofactor, that is at least partially removed from one or more or all of the naturally occurring constituents with which it is associated in nature; (3) any substance modified by the hand of man

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relative to that substance found in nature; or (4) any substance modified by increasing the amount of the substance relative to other components with which it is naturally associated (e.g., recombinant production in a host cell; multiple copies of a gene encoding the substance; and use of a stronger promoter than the promoter naturally associated with the gene encoding the substance). An isolated substance may be present in a fermentation broth sample; e.g. a host cell may be genetically modified to express the polypeptide of the invention. The fermentation broth from that host cell will comprise the isolated polypeptide.

Mature polypeptide: The term "mature polypeptide" means a polypeptide in its final form following translation and any post-translational modifications, such as N-terminal processing, C-terminal truncation, glycosylation, phosphorylation, etc. In one aspect, the mature polypeptide is amino acids 1 to 802 of SEQ ID NO: 2. In a second aspect, the mature polypeptide is amino acids 1 to 808 of SEQ ID NO: 4. In a third aspect, the mature polypeptide is amino acids 1 to 800 of SEQ ID NO: 6. In a fourth aspect, the mature polypeptide is amino acids 1 to 657 of SEQ ID NO: 8. It is known in the art that a host cell may produce a mixture of two of more different mature polypeptides (*i.e.*, with a different C-terminal and/or N-terminal amino acid) expressed by the same polynucleotide. It is also known in the art that different host cells process polypeptides differently, and thus, one host cell expressing a polynucleotide may produce a different mature polypeptide (*e.g.*, having a different C-terminal and/or N-terminal amino acid) as compared to another host cell expressing the same polynucleotide.

Mature polypeptide coding sequence: The term "mature polypeptide coding sequence" means a polynucleotide that encodes a mature polypeptide having xanthan degrading activity. In one aspect, the mature polypeptide coding sequence is nucleotides 109 to 2514 of SEQ ID NO: 1. Nucleotides 1 to 108 of SEQ ID NO: 1 encode a signal peptide. In one aspect, the mature polypeptide coding sequence is nucleotides 112 to 2493 of SEQ ID NO: 3. Nucleotides 1 to 111 of SEQ ID NO: 3 encode a signal peptide. In one aspect, the mature polypeptide coding sequence is nucleotides 106 to 2505 of SEQ ID NO: 5. Nucleotides 1 to 105 of SEQ ID NO: 5 encode a signal peptide. In one aspect, the mature polypeptide coding sequence is nucleotides 109 to 2079 of SEQ ID NO: 7. Nucleotides 1 to 108 of SEQ ID NO: 7 encode a signal peptide.

Nucleic acid construct: The term "nucleic acid construct" means a nucleic acid molecule, either single- or double-stranded, which is isolated from a naturally occurring gene or is modified to contain segments of nucleic acids in a manner that would not otherwise exist in nature or which is synthetic, which comprises one or more control sequences.

Operably linked: The term "operably linked" means a configuration in which a control sequence is placed at an appropriate position relative to the coding sequence of a polynucleotide such that the control sequence directs expression of the coding sequence.

Sequence identity: The relatedness between two amino acid sequences or between two nucleotide sequences is described by the parameter "sequence identity".

For purposes of the present invention, the sequence identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970,

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J. Mol. Biol. 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice *et al.*, 2000, *Trends Genet.* 16: 276-277), preferably version 5.0.0 or later. The parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled "longest identity" (obtained using the –nobrief option) is used as the percent identity and is calculated as follows:

(Identical Residues x 100)/(Length of Alignment – Total Number of Gaps in Alignment)

For purposes of the present invention, the sequence identity between two deoxyribonucleotide sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, *supra*) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice *et al.*, 2000, *supra*), preferably version 5.0.0 or later. The parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EDNAFULL (EMBOSS version of NCBI NUC4.4) substitution matrix. The output of Needle labeled "longest identity" (obtained using the –nobrief option) is used as the percent identity and is calculated as follows:

(Identical Deoxyribonucleotides x 100)/(Length of Alignment – Total Number of Gaps in Alignment)

Textile: The term "textile" means any textile material including yarns, yarn intermediates, fibers, non-woven materials, natural materials, synthetic materials, and any other textile material, fabrics made of these materials and products made from fabrics (e.g., garments and other articles). The textile or fabric may be in the form of knits, wovens, denims, non-wovens, felts, yarns, and towelling. The textile may be cellulose based such as natural cellulosics, including cotton, flax/linen, jute, ramie, sisal or coir or manmade cellulosics (e.g. originating from wood pulp) including viscose/rayon, ramie, cellulose acetate fibers (tricell), lyocell or blends thereof. The textile or fabric may also be non-cellulose based such as natural polyamides including wool, camel, cashmere, mohair, rabit and silk or synthetic polymer such as nylon, aramid, polyester, acrylic, polypropylen and spandex/elastane, or blends thereof as well as blend of cellulose based and non-cellulose based fibers. Examples of blends are blends of cotton and/or rayon/viscose with one or more companion material such as wool, synthetic fibers (e.g. polyamide fibers, acrylic fibers, polyester fibers, polyvinyl alcohol fibers, polyvinyl chloride fibers, polyurethane fibers, polyurea fibers, aramid fibers), and cellulose-containing fibers (e.g. rayon/viscose, ramie, flax/linen, jute, cellulose acetate fibers, lyocell). Fabric may be conventional washable laundry, for example stained household laundry. When the term fabric or garment is used it is intended to include the broader term textiles as well.

Textile care benefit: "Textile care benefits", which are not directly related to catalytic stain removal or prevention of redeposition of soils, are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye transfer from one textile to another textile or another part of the same textile an effect that is also termed dye transfer inhibition or anti-backstaining, removal of protruding or broken fibers from a textile surface to decrease pilling tendencies or remove already existing pills or fuzz an effect that also is termed anti-pilling,

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improvement of the textile-softness, colour clarification of the textile and removal of particulate soils which are trapped in the fibers of the textile. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching component such as hydrogen peroxide or other peroxides or other bleaching species.

Wash performance: The term "wash performance" is used as an enzyme's ability to remove stains present on the object to be cleaned during e.g. wash or hard surface cleaning. The improvement in the wash performance may be quantified by calculating the so-called intensity value (Int) as defined in 'Automatic Mechanical Stress Assay (AMSA) for laundry' herein. See also the wash performance test in Example 18 herein.

Whiteness: The term "Whiteness" is defined herein as a broad term with different meanings in different regions and for different customers. Loss of whiteness can e.g. be due to greying, yellowing, or removal of optical brighteners/hueing agents. Greying and yellowing can be due to soil redeposition, body soils, colouring from e.g. iron and copper ions or dye transfer. Whiteness might include one or several issues from the list below: colorant or dye effects; incomplete stain removal (e.g. body soils, sebum ect.); re-deposition (greying, yellowing or other discolorations of the object) (removed soils re-associates with other part of textile, soiled or unsoiled); chemical changes in textile during application; and clarification or brightening of colours.

Xanthan Lyase: The term "xanthan lyase" is defined herein as an enzyme that cleaves the beta-D-mannosyl-beta-D-1,4-glucuronosyl bonds in xanthan gum (EC 4.2.2.12). For purposes of the present invention, xanthan lyase activity is determined according to the procedure described in the Examples in the 'Xanthan lyase activity assay.

Xanthan degrading activity: The term "xanthan degrading activity" is defined herein as ability to cause viscosity reduction of a xanthan solution. Xanthan solution is highly viscous even at low polymer concentrations, and this viscosity is associated with the polymer degree of xanthan. Therefore, viscosity reduction can be used to monitor xanthan degradation. The viscosity reduction may be detected using the viscosity pressure assay described in Example 6.

Xanthan degrading activity includes activity towards intact xanthan as well as activity towards xanthan pretreated with xanthan lyase (modified xanthan gum – see Example 8).

Activity on xanthan gum: The term "GH5 polypeptide having activity on xanthan gum" or a "polypeptide having activity on xanthan gum and belonging to the GH5 class of glycosyl hydrolases" is defined as a polypeptide comprising a domain belonging to the GH5 class of glycosyl hydrolases, and having significant activity on xanthan gum. In one aspect of the invention a GH5 polypeptide having activity on xanthan gum may be a polypeptide having a sequence selected among SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, and SEQ ID NO: 8.

Activity on xanthan gum pretreated with xanthan lyase: The term "GH5 polypeptide having activity on xanthan gum pretreated with xanthan lyase" or a "polypeptide having activity on xanthan gum pretreated with xanthan lyase and belonging to the GH5 class of glycosyl hydrolases" is defined as a polypeptide comprising a domain belonging to the GH5 class of glycosyl hydrolases, and having significant activity on xanthan gum pretreated with xanthan lyase (modified xanthan gum

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– see Example 8). In one aspect of the invention a GH5 polypeptide having activity on xanthan gum pretreated with xanthan lyase may be a polypeptide having a sequence selected among SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, and SEQ ID NO: 8.

Detergent compositions comprising Polypeptides having xanthan degrading activity

In an embodiment, the present invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, which have xanthan degrading activity. In one aspect, the polypeptides differ by up to 10 amino acids, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10, from the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 70% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 75% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 80% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 85% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least

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90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 90% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 95% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In a particular embodiment the invention relates to detergent compositions comprising polypeptides having a sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%, and wherein the polypeptide has at least at least 100% of the xanthan degrading activity of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8.

In an embodiment, the polypeptide comprised in the detergent composition of present invention has been isolated. A polypeptide preferably comprises or consists of the amino acid sequence of any of SEQ ID NO: 2, 4, 6 and 8 or an allelic variant thereof; or is a fragment thereof having xanthan degrading activity. In another aspect, the polypeptide comprises or consists of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8. In another aspect, the polypeptide comprises or consists of amino acids 1 to 802 of SEQ ID NO: 2, amino acids 1 to 808 of SEQ ID NO: 4, amino acids 1 to 800 of SEQ ID NO: 6, or amino acids 1 to 657 of SEQ ID NO: 8.

In another embodiment, the present invention relates to a detergent compositions comprising a polypeptide having xanthan degrading activity encoded by a polynucleotide that hybridizes under very low stringency conditions, low stringency conditions, medium stringency conditions, medium-high stringency conditions, high stringency conditions, or very high stringency conditions with (i) the mature polypeptide coding sequence of SEQ ID NO: 1, (ii), or (iii) the full-length complement of (i) or (ii). In an embodiment, the polypeptide comprised in the detergent composition has been isolated.

For purposes of the present invention, hybridization indicates that the polynucleotide hybridizes to a labeled nucleic acid probe corresponding to (i) any of SEQ ID NO: 1, 3, 5, or 7; (ii) the mature polypeptide coding sequence of any of SEQ ID NO: 1, 3, 5, or 7; (iii) the full-length complement thereof; or (iv) a subsequence thereof; under very low to very high stringency conditions. Molecules to which the nucleic acid probe hybridizes under these conditions can be detected using, for example, X-ray film or any other detection means known in the art.

In another embodiment, the present invention relates to a detergent compositions comprising a polypeptide having xanthan degrading activity encoded by a polynucleotide having a sequence identity to the mature polypeptide coding sequence of any of SEQ ID NO: 1, 3, 5, or 7 of at least 60%, e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least

91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100%. In a further embodiment, the polypeptide has been isolated.

In another embodiment, the present invention relates to detergent compositions comprising variants of the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 comprising a substitution, deletion, and/or insertion at one or more (e.g., several) positions. In an embodiment, the number of amino acid substitutions, deletions and/or insertions introduced into the mature polypeptide of any of SEQ ID NO: 2, 4, 6 and 8 is up to 10, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10. The amino acid changes may be of a minor nature, that is conservative amino acid substitutions or insertions that do not significantly affect the folding and/or activity of the protein; small deletions, typically of 1-30 amino acids; small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue; a small linker peptide of up to 20-25 residues; or a small extension that facilitates purification by changing net charge or another function, such as a poly-histidine tag, an antigenic epitope or a binding domain. SEQ ID NO: 13, 14 and 15 show the polypeptides of the invention (SEQ ID NO: 2, 4 and 6) with an N-terminal poly histidine tag (His-tag). SEQ ID NO: 16, 17 and 18 show the polypeptides of the invention (SEQ ID NO: 4, 6 and 8) with an N-terminal poly histidine tag.

Examples of conservative substitutions are within the groups of basic amino acids (arginine, lysine and histidine), acidic amino acids (glutamic acid and aspartic acid), polar amino acids (glutamine and asparagine), hydrophobic amino acids (leucine, isoleucine and valine), aromatic amino acids (phenylalanine, tryptophan and tyrosine), and small amino acids (glycine, alanine, serine, threonine and methionine). Amino acid substitutions that do not generally alter specific activity are known in the art and are described, for example, by H. Neurath and R.L. Hill, 1979, *In, The Proteins*, Academic Press, New York. Common substitutions are Ala/Ser, Val/Ile, Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Ser/Gly, Tyr/Phe, Ala/Pro, Lys/Arg, Asp/Asn, Leu/Ile, Leu/Val, Ala/Glu, and Asp/Gly.

Sources of polypeptides having xanthan degrading activity

A polypeptide having xanthan degrading activity as comprised in the detergent composition of the present invention may be obtained from microorganisms of any genus. For purposes of the present invention, the term "obtained from" as used herein in connection with a given source shall mean that the polypeptide encoded by a polynucleotide is produced by the source or by a strain in which the polynucleotide from the source has been inserted.

In an aspect, the polypeptide is a polypeptide obtained from an Opitutaceae species.

Polynucleotides

The present invention also relates to polynucleotides encoding a polypeptide, as described herein. In an embodiment, the polynucleotide encoding the polypeptide of the present invention has been isolated.

Detergent composition

In one embodiment of the present invention, the polypeptide of the present invention may be added to a detergent composition in an amount corresponding to 0.0001-200 mg of enzyme protein, such as 0.0005-100 mg of enzyme protein, preferably 0.001-30 mg of enzyme protein, more preferably 0.005-8 mg of enzyme protein, even more preferably 0.01-2 mg of enzyme protein per litre of wash liquor.

A composition for use in automatic dishwash (ADW), for example, may include 0.0001%-50%, such as 0.001%-20%, such as 0.01%-10%, such as 0.05-5% of enzyme protein by weight of the composition.

A composition for use in laundry powder, for example, may include 0.0001%-50%, such as 0.001%-20%, such as 0.01%-10%, such as 0.05%-5% of enzyme protein by weight of the composition.

A composition for use in laundry liquid, for example, may include 0.0001%-10%, such as 0.001-7%, such as 0.1%-5% of enzyme protein by weight of the composition.

The enzyme(s) of the detergent composition of the invention may be stabilized using conventional stabilizing agents, e.g., a polyol such as propylene glycol or glycerol, a sugar or sugar alcohol, lactic acid, boric acid, or a boric acid derivative, e.g., an aromatic borate ester, or a phenyl boronic acid derivative such as 4-formylphenyl boronic acid, and the composition may be formulated as described in, for example, WO92/19709 and WO92/19708.

In certain markets different wash conditions and, as such, different types of detergents are used. This is disclosed in e.g. EP 1 025 240. For example, In Asia (Japan) a low detergent concentration system is used, while the United States uses a medium detergent concentration system, and Europe uses a high detergent concentration system.

In one embodiment, the invention is directed to detergent compositions comprising an enzyme of the present invention in combination with one or more additional cleaning composition components. The choice of additional components is within the skill of the artisan and includes conventional ingredients, including the exemplary non-limiting components set forth below.

The choice of components may include, for textile care, the consideration of the type of textile to be cleaned, the type and/or degree of soiling, the temperature at which cleaning is to take place, and the formulation of the detergent product. Although components mentioned below are categorized by general header according to a particular functionality, this is not to be construed as a limitation, as a component may comprise additional functionalities as will be appreciated by the skilled artisan.

In one embodiment, the invention is directed to an ADW (Automatic Dish Wash) composition comprising an enzyme of the present invention in combination with one or more additional ADW composition components. The choice of additional components is within the skill of the artisan and includes conventional ingredients, including the exemplary non-limiting components set forth below.

In one embodiment the detergent composition of present invention comprises up to

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Surfactants

The detergent composition may comprise one or more surfactants, which may be anionic and/or cationic and/or non-ionic and/or semi-polar and/or zwitterionic, or a mixture thereof. In a particular embodiment, the detergent composition includes a mixture of one or more nonionic surfactants and one or more anionic surfactants. The surfactant(s) is typically present at a level of from about 0.1% to 60% by weight, such as about 1% to about 40%, or about 3% to about 20%, or about 3% to about 10%. The surfactant(s) is chosen based on the desired cleaning application, and may include any conventional surfactant(s) known in the art.

When included therein the detergent will usually contain from about 1% to about 40% by weight of an anionic surfactant, such as from about 5% to about 30%, including from about 5% to about 15%, or from about 15% to about 20%, or from about 20% to about 25% of an anionic surfactant. Non-limiting examples of anionic surfactants include sulfates and sulfonates, in particular, linear alkylbenzenesulfonates (LAS), isomers of LAS, branched alkylbenzenesulfonates (BABS), phenylalkanesulfonates, alpha-olefinsulfonates (AOS), olefin sulfonates, alkene sulfonates, alkane-2,3-diylbis(sulfates), hydroxyalkanesulfonates and disulfonates, alkyl sulfates (AS) such as sodium dodecyl sulfate (SDS), fatty alcohol sulfates (FAS), primary alcohol sulfates (PAS), alcohol ethersulfates (AES or AEOS or FES, also known as alcohol ethoxysulfates or fatty alcohol ether sulfates), secondary alkanesulfonates (SAS), paraffin sulfonates (PS), ester sulfonates, sulfonated fatty acid glycerol esters, alpha-sulfo fatty acid methyl esters (alpha-SFMe or SES) including methyl ester sulfonate (MES), alkylor alkenylsuccinic acid, dodecenyl/tetradecenyl succinic acid (DTSA), fatty acid derivatives of amino acids, diesters and monoesters of sulfo-succinic acid or salt of fatty acids (soap), and combinations thereof.

When included therein the detergent will usually contain from about 1% to about 40% by weigh of a cationic surfactant, for example from about 0.5% to about 30%, in particular from about 1% to about 20%, from about 3% to about 10%, such as from about 3% to about 5%, from about 8% to about 12% or from about 10% to about 12%. Non-limiting examples of cationic surfactants include alkyldimethylethanolamine quat (ADMEAQ), cetyltrimethylammonium bromide (CTAB), dimethyldistearylammonium chloride (DSDMAC), and alkylbenzyldimethylammonium, alkyl quaternary ammonium compounds, alkoxylated quaternary ammonium (AQA) compounds, ester quats, and combinations thereof.

When included therein the detergent will usually contain from about 0.2% to about 40% by weight of a nonionic surfactant, for example from about 0.5% to about 30%, in particular from about 1% to about 20%, from about 3% to about 10%, such as from about 3% to about 5%, from about 8% to about 12%, or from about 10% to about 12%. Non-limiting examples of nonionic surfactants include alcohol ethoxylates (AE or AEO), alcohol propoxylates, propoxylated fatty alcohols (PFA), alkoxylated fatty acid alkyl esters, such as ethoxylated and/or propoxylated fatty acid alkyl esters, alkylphenol ethoxylates (APE), nonylphenol ethoxylates (NPE), alkylpolyglycosides (APG), alkoxylated amines, fatty acid monoethanolamides (FAM), fatty acid diethanolamides (FADA), ethoxylated fatty acid monoethanolamides (EFAM), propoxylated fatty acid monoethanolamides (PFAM), polyhydroxyalkyl

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fatty acid amides, or *N*-acyl *N*-alkyl derivatives of glucosamine (glucamides, GA, or fatty acid glucamides, FAGA), as well as products available under the trade names SPAN and TWEEN, and combinations thereof.

When included therein the detergent will usually contain from about 0% to about 10% by weight of a semipolar surfactant. Non-limiting examples of semipolar surfactants include amine oxides (AO) such as alkyldimethylamineoxide, *N*-(coco alkyl)-*N*,*N*-dimethylamine oxide and *N*-(tallow-alkyl)-*N*,*N*-bis(2-hydroxyethyl)amine oxide, , and combinations thereof.

When included therein the detergent will usually contain from about 0% to about 10% by weight of a zwitterionic surfactant. Non-limiting examples of zwitterionic surfactants include betaines such as alkyldimethylbetaines, sulfobetaines, and combinations thereof.

Hydrotropes

A hydrotrope is a compound that solubilises hydrophobic compounds in aqueous solutions (or oppositely, polar substances in a non-polar environment). Typically, hydrotropes have both hydrophilic and a hydrophobic character (so-called amphiphilic properties as known from surfactants); however the molecular structure of hydrotropes generally do not favor spontaneous self-aggregation, see e.g. review by Hodgdon and Kaler (2007), Current Opinion in Colloid & Interface Science 12: 121-128. Hydrotropes do not display a critical concentration above which self-aggregation occurs as found for surfactants and lipids forming miceller, lamellar or other well defined meso-phases. Instead, many hydrotropes show a continuous-type aggregation process where the sizes of aggregates grow as concentration increases. However, many hydrotropes alter the phase behavior, stability, and colloidal properties of systems containing substances of polar and non-polar character, including mixtures of water, oil, surfactants, and polymers. Hydrotropes are classically used across industries from pharma, personal care, food, to technical applications. Use of hydrotropes in detergent compositions allow for example more concentrated formulations of surfactants (as in the process of compacting liquid detergents by removing water) without inducing undesired phenomena such as phase separation or high viscosity.

The detergent may contain 0-10% by weight, for example 0-5% by weight, such as about 0.5 to about 5%, or about 3% to about 5%, of a hydrotrope. Any hydrotrope known in the art for use in detergents may be utilized. Non-limiting examples of hydrotropes include sodium benzenesulfonate, sodium p-toluene sulfonate (STS), sodium xylene sulfonate (SXS), sodium cumene sulfonate (SCS), sodium cymene sulfonate, amine oxides, alcohols and polyglycolethers, sodium hydroxynaphthoate, sodium hydroxynaphthalene sulfonate, sodium ethylhexyl sulfate, and combinations thereof.

Builders and Co-Builders

The detergent composition may contain about 0-65% by weight, such as about 5% to about 50% of a detergent builder or co-builder, or a mixture thereof. In a dish wash detergent, the level of builder is typically 40-65%, particularly 50-65%. The builder and/or co-builder may particularly be a chelating agent that forms water-soluble complexes with Ca and Mg. Any builder and/or co-builder

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known in the art for use in detergents may be utilized. Non-limiting examples of builders include zeolites, diphosphates (pyrophosphates), triphosphates such as sodium triphosphate (STP or STPP), carbonates such as sodium carbonate, soluble silicates such as sodium metasilicate, layered silicates (*e.g.*, SKS-6 from Hoechst), ethanolamines such as 2-aminoethan-1-ol (MEA), diethanolamine (DEA, also known as 2,2'-iminodiethan-1-ol), triethanolamine (TEA, also known as 2,2',2"-nitrilotriethan-1-ol), and (carboxymethyl)inulin (CMI), and combinations thereof.

The detergent composition may also contain 0-50% by weight, such as about 5% to about 30%, of a detergent co-builder, . The detergent composition may include include a co-builder alone, or in combination with a builder, for example a zeolite builder. Non-limiting examples of co-builders include homopolymers of polyacrylates or copolymers thereof, such as poly(acrylic acid) (PAA) or copoly(acrylic acid/maleic acid) (PAA/PMA). Further non-limiting examples include citrate, chelators such as aminocarboxylates, aminopolycarboxylates and phosphonates, and alkyl- or alkenylsuccinic acid. Additional specific examples include 2,2',2"-nitrilotriacetic acid (NTA), ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTPA), iminodisuccinic acid (IDS), ethylenediamine-N,Ndisuccinic acid (EDDS), methylglycinediacetic acid (MGDA), glutamic acid-N,N-diacetic acid (GLDA), 1hydroxyethane-1,1-diphosphonic acid (HEDP), ethylenediaminetetra(methylenephosphonic acid) (EDTMPA), diethylenetriaminepentakis(methylenephosphonic acid) (DTMPA or DTPMPA), N-(2hydroxyethyl)iminodiacetic acid (EDG), aspartic acid-N-monoacetic acid (ASMA), aspartic acid-N,Ndiacetic acid (ASDA), aspartic acid-N-monopropionic acid (ASMP), iminodisuccinic acid (IDA), N-(2sulfomethyl)-aspartic acid (SMAS), N-(2-sulfoethyl)-aspartic acid (SEAS), N-(2-sulfomethyl)-glutamic acid (SMGL), N-(2-sulfoethyl)-glutamic acid (SEGL), N-methyliminodiacetic acid (MIDA), alpha-alanine-N,N-diacetic acid (α -ALDA), serine-N,N-diacetic acid (SEDA), isoserine-N,N-diacetic acid (ISDA), phenylalanine-N,N-diacetic acid (PHDA), anthranilic acid-N,N-diacetic acid (ANDA), sulfanilic acid-N,Ndiacetic acid (SLDA), taurine-N,N-diacetic acid (TUDA) and sulfomethyl-N,N-diacetic acid (SMDA), N-(2-hydroxyethyl)ethylenediamine-N,N',N"-triacetic acid (HEDTA), diethanolglycine (DEG). diethylenetriamine penta(methylenephosphonic acid) (DTPMP), aminotris(methylenephosphonic acid) (ATMP), and combinations and salts thereof. Further exemplary builders and/or co-builders are described in, e.g., WO 09/102854, US 5977053

Bleaching Systems

The detergent may contain 0-30% by weight, such as about 1% to about 20%, of a bleaching system. Any bleaching system known in the art for use in detergents may be utilized. Suitable bleaching system components include bleaching catalysts, photobleaches, bleach activators, sources of hydrogen peroxide such as sodium percarbonate, sodium perborates and hydrogen peroxide—urea (1:1), preformed peracids and mixtures thereof. Suitable preformed peracids include, but are not limited to, peroxycarboxylic acids and salts, diperoxydicarboxylic acids, perimidic acids and salts, peroxymonosulfuric acids and salts, for example, Oxone (R), and mixtures thereof. Non-limiting examples of bleaching systems include peroxide-based bleaching systems, which may comprise, for example, an inorganic salt, including alkali metal salts such as sodium salts of perborate (usually

mono- or tetra-hydrate), percarbonate, persulfate, perphosphate, persilicate salts, in combination with a peracid-forming bleach activator. The term bleach activator is meant herein as a compound which reacts with hydrogen peroxide to form a peracid via perhydrolysis. The peracid thus formed constitutes the activated bleach. Suitable bleach activators to be used herein include those belonging to the class of esters, amides, imides or anhydrides. Suitable examples are tetraacetylethylenediamine (TAED), sodium 4-[(3,5,5-trimethylhexanoyl)oxy]benzene-1-sulfonate (ISONOBS), 4-(dodecanoyloxy)benzene-1-sulfonate (LOBS), 4-(decanoyloxy)benzene-1-sulfonate, 4-(decanoyloxy)benzoate (DOBS or DOBA), 4-(nonanoyloxy)benzene-1-sulfonate (NOBS), and/or those disclosed in WO98/17767. A particular family of bleach activators of interest was disclosed in EP624154 and particulary preferred in that family is acetyl triethyl citrate (ATC). ATC or a short chain triglyceride like triacetin has the advantage that it is environmentally friendly Furthermore acetyl triethyl citrate and triacetin have good hydrolytical stability in the product upon storage and are efficient bleach activators. Finally ATC is multifunctional, as the citrate released in the perhydrolysis reaction may function as a builder. Alternatively, the bleaching system may comprise peroxyacids of, for example, the amide, imide, or sulfone type. The bleaching system may also comprise peracids such as 6-(phthalimido)peroxyhexanoic acid (PAP). The bleaching system may also include a bleach catalyst. In some embodiments the bleach component may be an organic catalyst selected from the group consisting of organic catalysts having the following formulae:

(i)
$$OSO_3^{\ominus}$$
 $O-R^1$
(ii) OSO_3^{\ominus}
 $O-R^1$

(iii) and mixtures thereof;

wherein each R¹ is independently a branched alkyl group containing from 9 to 24 carbons or linear alkyl group containing from 11 to 24 carbons, preferably each R¹ is independently a branched alkyl group containing from 9 to 18 carbons or linear alkyl group containing from 11 to 18 carbons, more preferably each R¹ is independently selected from the group consisting of 2-propylheptyl, 2-butyloctyl, 2-pentylnonyl, 2-hexyldecyl, dodecyl, tetradecyl, hexadecyl, octadecyl, isononyl, isodecyl, isotridecyl and isopentadecyl. Other exemplary bleaching systems are described, e.g. in WO2007/087258, WO2007/087244, WO2007/087259, EP1867708 (Vitamin K) and WO2007/087242. Suitable photobleaches may for example be sulfonated zinc or aluminium phthalocyanines.

Preferably the bleach component comprises a source of peracid in addition to bleach catalyst, particularly organic bleach catalyst. The source of peracid may be selected from (a) preformed peracid; (b) percarbonate, perborate or persulfate salt (hydrogen peroxide source) preferably in combination with a bleach activator; and (c) perhydrolase enzyme and an ester for forming peracid in situ in the presence of water in a textile or hard surface treatment step.

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Polymers

The detergent may contain 0-10% by weight, such as 0.5-5%, 2-5%, 0.5-2% or 0.2-1% of a polymer. Any polymer known in the art for use in detergents may be utilized. The polymer may function as a co-builder as mentioned above, or may provide antiredeposition, fiber protection, soil release, dye transfer inhibition, grease cleaning and/or anti-foaming properties. Some polymers may have more than one of the above-mentioned properties and/or more than one of the below-mentioned motifs. Exemplary polymers include (carboxymethyl)cellulose (CMC), poly(vinyl alcohol) (PVA), poly(vinylpyrrolidone) (PVP), poly(ethyleneglycol) or poly(ethylene oxide) (PEG), ethoxylated poly(ethyleneimine), carboxymethyl inulin (CMI), and polycarboxylates such as PAA, PAA/PMA, polyaspartic acid, and lauryl methacrylate/acrylic acid copolymers, hydrophobically modified CMC (HMCMC) and silicones, copolymers of terephthalic acid and oligomeric glycols, copolymers of poly(ethylene terephthalate) and poly(oxyethene terephthalate) (PET-POET), PVP, poly(vinylimidazole) (PVI), poly(vinylpyridine-N-oxide) (PVPO or PVPNO) and polyvinylpyrrolidone-vinylimidazole (PVPVI). Further exemplary polymers include sulfonated polycarboxylates, polyethylene oxide and polypropylene oxide (PEO-PPO) and diquaternium ethoxy sulfate. Other exemplary polymers are disclosed in, e.g., WO 2006/130575. Salts of the above-mentioned polymers are also contemplated.

Fabric hueing agents

The detergent compositions of the present invention may also include fabric hueing agents such as dyes or pigments, which when formulated in detergent compositions can deposit onto a fabric when said fabric is contacted with a wash liquor comprising said detergent compositions and thus altering the tint of said fabric through absorption/reflection of visible light. Fluorescent whitening agents emit at least some visible light. In contrast, fabric hueing agents alter the tint of a surface as they absorb at least a portion of the visible light spectrum. Suitable fabric hueing agents include dyes and dye-clay conjugates, and may also include pigments. Suitable dyes include small molecule dyes and polymeric dyes. Suitable small molecule dyes include small molecule dyes selected from the group consisting of dyes falling into the Colour Index (C.I.) classifications of Direct Blue, Direct Red, Direct Violet, Acid Blue, Acid Red, Acid Violet, Basic Blue, Basic Violet and Basic Red, or mixtures thereof, for example as described in WO2005/03274, WO2005/03275, WO2005/03276 and EP1876226 (hereby incorporated by reference). The detergent composition preferably comprises from about 0.00003 wt% to about 0.2 wt%, from about 0.00008 wt% to about 0.05 wt%, or even from about 0.0001 wt% to about 0.04 wt% fabric hueing agent. The composition may comprise from 0.0001 wt% to 0.2 wt% fabric hueing agent, this may be especially preferred when the composition is in the form of a unit dose pouch. Suitable hueing agents are also disclosed in, e.g. WO 2007/087257 and WO2007/087243.

Additional enzymes

The detergent additive as well as the detergent composition may comprise one or more additional enzymes such as a protease, a lipase, a cutinase, an amylase, a carbohydrase, a cellulase,

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a pectinase, a mannanase, an arabinase, a galactanase, a xylanase, an oxidase, *e.g.*, a laccase, and/or a peroxidase and/or a xanthan lyase.

In general the properties of the selected enzyme(s) should be compatible with the selected detergent, (i.e., pH-optimum, compatibility with other enzymatic and non-enzymatic ingredients, etc.), and the enzyme(s) should be present in effective amounts.

Cellulases

Suitable cellulases include those of bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Suitable cellulases include cellulases from the genera *Bacillus*, *Pseudomonas*, *Humicola*, *Fusarium*, *Thielavia*, *Acremonium*, e.g., the fungal cellulases produced from *Humicola insolens*, *Myceliophthora thermophila* and *Fusarium oxysporum* disclosed in US 4,435,307, US 5,648,263, US 5,691,178, US 5,776,757 and WO 89/09259.

Especially suitable cellulases are the alkaline or neutral cellulases having colour care benefits. Examples of such cellulases are cellulases described in EP 0 495 257, EP 0 531 372, WO 96/11262, WO 96/29397, WO 98/08940. Other examples are cellulase variants such as those described in WO 94/07998, EP 0 531 315, US 5,457,046, US 5,686,593, US 5,763,254, WO 95/24471, WO 98/12307 and WO99/001544.

Other cellulases are endo-beta-1,4-glucanase enzyme having a sequence of at least 97% identity to the amino acid sequence of position 1 to position 773 of SEQ ID NO:2 of WO 2002/099091 or a family 44 xyloglucanase, which a xyloglucanase enzyme having a sequence of at least 60% identity to positions 40-559 of SEQ ID NO: 2 of WO 2001/062903.

Commercially available cellulases include CelluzymeTM, and CarezymeTM (Novozymes A/S) Carezyme PremiumTM (Novozymes A/S), Celluclean TM (Novozymes A/S), Celluclean ClassicTM (Novozymes A/S), CellusoftTM (Novozymes A/S), WhitezymeTM (Novozymes A/S), ClazinaseTM, and Puradax HATM (Genencor International Inc.), and KAC-500(B)TM (Kao Corporation).

Mannanases

Suitable mannanases include those of bacterial or fungal origin. Chemically or genetically modified mutants are included. The mannanase may be an alkaline mannanase of Family 5 or 26. It may be a wild-type from *Bacillus* or *Humicola*, particularly *B. agaradhaerens*, *B. licheniformis*, *B. halodurans*, *B. clausii*, or *H. insolens*. Suitable mannanases are described in WO 1999/064619. A commercially available mannanase is Mannaway (Novozymes A/S).

Xanthan lyases

Suitable xanthan lyases include those of plant, bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful enzymes include the xanthan lyases disclosed in WO2013167581 and shown herein as SEQ ID NO:21, 22, 23 and 24.

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Proteases

Suitable proteases include those of bacterial, fungal, plant, viral or animal origin e.g. vegetable or microbial origin. Microbial origin is preferred. Chemically modified or protein engineered mutants are included. It may be an alkaline protease, such as a serine protease or a metalloprotease. A serine protease may for example be of the S1 family, such as trypsin, or the S8 family such as subtilisin. A metalloproteases protease may for example be a thermolysin from e.g. family M4 or other metalloprotease such as those from M5, M7 or M8 families.

The term "subtilases" refers to a sub-group of serine protease according to Siezen et al., Protein Engng. 4 (1991) 719-737 and Siezen et al. Protein Science 6 (1997) 501-523. Serine proteases are a subgroup of proteases characterized by having a serine in the active site, which forms a covalent adduct with the substrate. The subtilases may be divided into 6 sub-divisions, i.e. the Subtilisin family, the Thermitase family, the Proteinase K family, the Lantibiotic peptidase family, the Kexin family and the Pyrolysin family.

Examples of subtilases are those derived from *Bacillus* such as *Bacillus lentus*, *B. alkalophilus*, *B. subtilis*, *B. amyloliquefaciens*, *Bacillus pumilus* and *Bacillus gibsonii* described in; US7262042 and WO09/021867, and *subtilisin lentus*, *subtilisin Novo*, *subtilisin Carlsberg*, *Bacillus licheniformis*, *subtilisin BPN'*, *subtilisin 309*, *subtilisin 147* and *subtilisin 168* described in WO89/06279 and protease PD138 described in (WO93/18140). Other useful proteases may be those described in WO92/175177, WO01/016285, WO02/026024 and WO02/016547. Examples of trypsin-like proteases are trypsin (e.g. of porcine or bovine origin) and the *Fusarium* protease described in WO89/06270, WO94/25583 and WO05/040372, and the chymotrypsin proteases derived from *Cellumonas* described in WO05/052161 and WO05/052146.

A further preferred protease is the alkaline protease from *Bacillus lentus* DSM 5483, as described for example in WO95/23221, and variants thereof which are described in WO92/21760, WO95/23221, EP1921147 and EP1921148.

Examples of metalloproteases are the neutral metalloprotease as described in WO07/044993 (Genencor Int.) such as those derived from *Bacillus amyloliquefaciens*.

Examples of useful proteases are the variants described in: WO92/19729, WO96/034946, WO98/20115, WO98/20116, WO99/011768, WO01/44452, WO03/006602, WO04/03186, WO04/041979, WO07/006305, WO11/036263, WO11/036264, especially the variants with substitutions in one or more of the following positions: 3, 4, 9, 15, 27, 36, 57, 68, 76, 87, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 106, 118, 120, 123, 128, 129, 130, 160, 167, 170, 194, 195, 199, 205, 206, 217, 218, 222, 224, 232, 235, 236, 245, 248, 252 and 274 using the BPN' numbering. More preferred the subtilase variants may comprise the mutations: S3T, V4I, S9R, A15T, K27R, *36D, V68A, N76D, N87S,R, *97E, A98S, S99G,D,A, S99AD, S101G,M,R S103A, V104I,Y,N, S106A, G118V,R, H120D,N, N123S, S128L, P129Q, S130A, G160D, Y167A, R170S, A194P, G195E, V199M, V205I, L217D, N218D, M222S, A232V, K235L, Q236H, Q245R, N252K, T274A (using BPN' numbering).

Suitable commercially available protease enzymes include those sold under the trade names Alcalase®, DuralaseTm, DurazymTm, Relase®, Relase® Ultra, Savinase®, Savinase® Ultra, Primase®, Polarzyme®, Kannase®, Liquanase®, Liquanase® Ultra, Ovozyme®, Coronase®, Coronase® Ultra, Blaze®, Neutrase®, Everlase® and Esperase® (Novozymes A/S), those sold under the tradename Maxatase®, Maxacal®, Maxapem®, Purafect®, Purafect Prime®, , Purafect MA®, Purafect Ox®, Purafect OxP®, Puramax®, Properase®, , FN2®, FN3® , FN4®, Excellase®, Eraser®, Opticlean® and Optimase® (Danisco/DuPont), AxapemTM (Gist-Brocases N.V.), BLAP (sequence shown in Figure 29 of US5352604) and variants hereof (Henkel AG) and KAP (*Bacillus alkalophilus* subtilisin) from Kao.

Lipases and cutinases

Suitable lipases and cutinases include those of bacterial or fungal origin. Chemically modified or protein engineered mutant enzymes are included. Examples include lipase from *Thermomyces*, e.g. from *T. lanuginosus* (previously named *Humicola lanuginosa*) as described in EP258068 and EP305216, cutinase from *Humicola*, e.g. *H. insolens* (WO96/13580), lipase from strains of *Pseudomonas* (some of these now renamed to *Burkholderia*), e.g. *P. alcaligenes* or *P. pseudoalcaligenes* (EP218272), *P. cepacia* (EP331376), *P. sp.* strain SD705 (WO95/06720 & WO96/27002), *P. wisconsinensis* (WO96/12012), GDSL-type *Streptomyces* lipases (WO10/065455), cutinase from *Magnaporthe grisea* (WO10/107560), cutinase from *Pseudomonas mendocina* (US5,389,536), lipase from *Thermobifida fusca* (WO11/084412), *Geobacillus stearothermophilus* lipase (WO11/084417), lipase from *Bacillus subtilis* (WO11/084599), and lipase from *Streptomyces griseus* (WO11/150157) and *S. pristinaespiralis* (WO12/137147).

Other examples are lipase variants such as those described in EP407225, WO92/05249, WO94/01541, WO94/25578, WO95/14783, WO95/30744, WO95/35381, WO95/22615, WO96/00292, WO97/04079, WO97/07202, WO00/34450, WO00/60063, WO01/92502, WO07/87508 and WO09/109500.

Preferred commercial lipase products include include Lipolase[™], Lipex[™]; Lipolex[™] and Lipoclean[™] (Novozymes A/S), Lumafast (originally from Genencor) and Lipomax (originally from Gist-Brocades).

Still other examples are lipases sometimes referred to as acyltransferases or perhydrolases, e.g. acyltransferases with homology to *Candida antarctica* lipase A (WO10/111143), acyltransferase from *Mycobacterium smegmatis* (WO05/56782), perhydrolases from the CE 7 family (WO09/67279), and variants of the *M. smegmatis* perhydrolase in particular the S54V variant used in the commercial product Gentle Power Bleach from Huntsman Textile Effects Pte Ltd (WO10/100028).

Amylases

Suitable amylases which can be used together with the enzyme of the invention may be an alpha-amylase or a glucoamylase and may be of bacterial or fungal origin. Chemically modified or

protein engineered mutants are included. Amylases include, for example, alpha-amylases obtained from *Bacillus*, *e.g.*, a special strain of *Bacillus licheniformis*, described in more detail in GB 1,296,839.

Suitable amylases include amylases having SEQ ID NO: 2 in WO 95/10603 or variants having 90% sequence identity to SEQ ID NO: 3 thereof. Preferred variants are described in WO 94/02597, WO 94/18314, WO 97/43424 and SEQ ID NO: 4 of WO 99/019467, such as variants with substitutions in one or more of the following positions: 15, 23, 105, 106, 124, 128, 133, 154, 156, 178, 179, 181, 188, 190, 197, 201, 202, 207, 208, 209, 211, 243, 264, 304, 305, 391, 408, and 444.

Different suitable amylases include amylases having SEQ ID NO: 6 in WO 02/010355 or variants thereof having 90% sequence identity to SEQ ID NO: 6. Preferred variants of SEQ ID NO: 6 are those having a deletion in positions 181 and 182 and a substitution in position 193.

Other amylases which are suitable are hybrid alpha-amylase comprising residues 1-33 of the alpha-amylase derived from *B. amyloliquefaciens* shown in SEQ ID NO: 6 of WO 2006/066594 and residues 36-483 of the *B. licheniformis* alpha-amylase shown in SEQ ID NO: 4 of WO 2006/066594 or variants having 90% sequence identity thereof. Preferred variants of this hybrid alpha-amylase are those having a substitution, a deletion or an insertion in one of more of the following positions: G48, T49, G107, H156, A181, N190, M197, I201, A209 and Q264. Most preferred variants of the hybrid alpha-amylase comprising residues 1-33 of the alpha-amylase derived from *B. amyloliquefaciens* shown in SEQ ID NO: 6 of WO 2006/066594 and residues 36-483 of SEQ ID NO: 4 are those having the substitutions:

M197T;

H156Y+A181T+N190F+A209V+Q264S; or

G48A+T49I+G107A+H156Y+A181T+N190F+I201F+A209V+Q264S.

Further amylases which are suitable are amylases having SEQ ID NO: 6 in WO 99/019467 or variants thereof having 90% sequence identity to SEQ ID NO: 6. Preferred variants of SEQ ID NO: 6 are those having a substitution, a deletion or an insertion in one or more of the following positions: R181, G182, H183, G184, N195, I206, E212, E216 and K269. Particularly preferred amylases are those having deletion in positions R181 and G182, or positions H183 and G184.

Additional amylases which can be used are those having SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 2 or SEQ ID NO: 7 of WO 96/023873 or variants thereof having 90% sequence identity to SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3 or SEQ ID NO: 7. Preferred variants of SEQ ID NO: 1, SEQ ID NO: 3 or SEQ ID NO: 7 are those having a substitution, a deletion or an insertion in one or more of the following positions: 140, 181, 182, 183, 184, 195, 206, 212, 243, 260, 269, 304 and 476, using SEQ ID 2 of WO 96/023873 for numbering. More preferred variants are those having a deletion in two positions selected from 181, 182, 183 and 184, such as 181 and 182, 182 and 183, or positions 183 and 184. Most preferred amylase variants of SEQ ID NO: 1, SEQ ID NO: 2 or SEQ ID NO: 7 are those having a deletion in positions 183 and 184 and a substitution in one or more of positions 140, 195, 206, 243, 260, 304 and 476.

Other amylases which can be used are amylases having SEQ ID NO: 2 of WO 08/153815, SEQ ID NO: 10 in WO 01/66712 or variants thereof having 90% sequence identity to SEQ ID NO: 2

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of WO 08/153815 or 90% sequence identity to SEQ ID NO: 10 in WO 01/66712. Preferred variants of SEQ ID NO: 10 in WO 01/66712 are those having a substitution, a deletion or an insertion in one of more of the following positions: 176, 177, 178, 179, 190, 201, 207, 211 and 264.

Further suitable amylases are amylases having SEQ ID NO: 2 of WO 09/061380 or variants having 90% sequence identity to SEQ ID NO: 2 thereof. Preferred variants of SEQ ID NO: 2 are those having a truncation of the C-terminus and/or a substitution, a deletion or an insertion in one of more of the following positions: Q87, Q98, S125, N128, T131, T165, K178, R180, S181, T182, G183, M201, F202, N225, S243, N272, N282, Y305, R309, D319, Q320, Q359, K444 and G475. More preferred variants of SEQ ID NO: 2 are those having the substitution in one of more of the following positions: Q87E,R, Q98R, S125A, N128C, T131I, T165I, K178L, T182G, M201L, F202Y, N225E,R, N272E,R, S243Q,A,E,D, Y305R, R309A, Q320R, Q359E, K444E and G475K and/or deletion in position R180 and/or S181 or of T182 and/or G183. Most preferred amylase variants of SEQ ID NO: 2 are those having the substitutions:

N128C+K178L+T182G+Y305R+G475K;

N128C+K178L+T182G+F202Y+Y305R+D319T+G475K;

S125A+N128C+K178L+T182G+Y305R+G475K; or

S125A+N128C+T131I+T165I+K178L+T182G+Y305R+G475K wherein the variants are C-terminally truncated and optionally further comprises a substitution at position 243 and/or a deletion at position 180 and/or position 181.

Further suitable amylases are amylases having SEQ ID NO: 1 of WO13184577 or variants having 90% sequence identity to SEQ ID NO: 1 thereof. Preferred variants of SEQ ID NO: 1 are those having a substitution, a deletion or an insertion in one of more of the following positions: K176, R178, G179, T180, G181, E187, N192, M199, I203, S241, R458, T459, D460, G476 and G477. More preferred variants of SEQ ID NO: 1 are those having the substitution in one of more of the following positions: K176L, E187P, N192FYH, M199L, I203YF, S241QADN, R458N, T459S, D460T, G476K and G477K and/or deletion in position R178 and/or S179 or of T180 and/or G181. Most preferred amylase variants of SEQ ID NO: 1 are those having the substitutions:

E187P+I203Y+G476K

E187P+I203Y+R458N+T459S+D460T+G476K

wherein the variants optionally further comprises a substitution at position 241 and/or a deletion at position 178 and/or position 179.

Further suitable amylases are amylases having SEQ ID NO: 1 of WO10104675 or variants having 90% sequence identity to SEQ ID NO: 1 thereof. Preferred variants of SEQ ID NO: 1 are those having a substitution, a deletion or an insertion in one of more of the following positions: N21, D97, V128 K177, R179, S180, I181, G182, M200, L204, E242, G477 and G478. More preferred variants of SEQ ID NO: 1 are those having the substitution in one of more of the following positions: N21D, D97N, V128I K177L, M200L, L204YF, E242QA, G477K and G478K and/or deletion in position R179 and/or S180 or of I181 and/or G182. Most preferred amylase variants of SEQ ID NO: 1 are those having the substitutions:

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N21D+D97N+V128I

wherein the variants optionally further comprises a substitution at position 200 and/or a deletion at position 180 and/or position 181.

Other suitable amylases are the alpha-amylase having SEQ ID NO: 12 in WO01/66712 or a variant having at least 90% sequence identity to SEQ ID NO: 12. Preferred amylase variants are those having a substitution, a deletion or an insertion in one of more of the following positions of SEQ ID NO: 12 in WO01/66712: R28, R118, N174; R181, G182, D183, G184, G186, W189, N195, M202, Y298, N299, K302, S303, N306, R310, N314; R320, H324, E345, Y396, R400, W439, R444, N445, K446, Q449, R458, N471, N484. Particular preferred amylases include variants having a deletion of D183 and G184 and having the substitutions R118K, N195F, R320K and R458K, and a variant additionally having substitutions in one or more position selected from the group: M9, G149, G182, G186, M202, T257, Y295, N299, M323, E345 and A339, most preferred a variant that additionally has substitutions in all these positions.

Other examples are amylase variants such as those described in WO2011/098531, WO2013/001078 and WO2013/001087.

Commercially available amylases are DuramylTM, TermamylTM, FungamylTM, Stainzyme TM, Stainzyme PlusTM, NatalaseTM, Liquozyme X and BANTM (from Novozymes A/S), and RapidaseTM, PurastarTM/EffectenzTM, Powerase, Preferenz S1000, Preferenz S100 and Preferenz S110 (from Genencor International Inc./DuPont).

Peroxidases/Oxidases

A peroxidase according to the invention is a peroxidase enzyme comprised by the enzyme classification EC 1.11.1.7, as set out by the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology (IUBMB), or any fragment derived therefrom, exhibiting peroxidase activity.

Suitable peroxidases include those of plant, bacterial or fungal origin. Chemically modified or protein engineered mutants are included. Examples of useful peroxidases include peroxidases from *Coprinopsis*, *e.g.*, from *C. cinerea* (EP 179,486), and variants thereof as those described in WO 93/24618, WO 95/10602, and WO 98/15257.

A peroxidase according to the invention also include a haloperoxidase enzyme, such as chloroperoxidase, bromoperoxidase and compounds exhibiting chloroperoxidase or bromoperoxidase activity. Haloperoxidases are classified according to their specificity for halide ions. Chloroperoxidases (E.C. 1.11.1.10) catalyze formation of hypochlorite from chloride ions.

In an embodiment, the haloperoxidase of the invention is a chloroperoxidase. Preferably, the haloperoxidase is a vanadium haloperoxidase, *i.e.*, a vanadate-containing haloperoxidase. In a preferred method of the present invention the vanadate-containing haloperoxidase is combined with a source of chloride ion.

Haloperoxidases have been isolated from many different fungi, in particular from the fungus group dematiaceous hyphomycetes, such as *Caldariomyces*, e.g., *C. fumago*, *Alternaria*, *Curvularia*, e.g., *C. verruculosa* and *C. inaequalis*, *Drechslera*, *Ulocladium* and *Botrytis*.

Haloperoxidases have also been isolated from bacteria such as *Pseudomonas*, e.g., *P. pyrrocinia* and *Streptomyces*, e.g., *S. aureofaciens*.

In an preferred embodiment, the haloperoxidase is derivable from *Curvularia* sp., in particular *Curvularia verruculosa* or *Curvularia inaequalis*, such as *C. inaequalis* CBS 102.42 as described in WO 95/27046; or *C. verruculosa* CBS 147.63 or *C. verruculosa* CBS 444.70 as described in WO 97/04102; or from *Drechslera hartlebii* as described in WO 01/79459, *Dendryphiella salina* as described in WO 01/79458, *Phaeotrichoconis crotalarie* as described in WO 01/79461, or *Geniculosporium* sp. as described in WO 01/79460.

An oxidase according to the invention include, in particular, any laccase enzyme comprised by the enzyme classification EC 1.10.3.2, or any fragment derived therefrom exhibiting laccase activity, or a compound exhibiting a similar activity, such as a catechol oxidase (EC 1.10.3.1), an o-aminophenol oxidase (EC 1.10.3.4), or a bilirubin oxidase (EC 1.3.3.5).

Preferred laccase enzymes are enzymes of microbial origin. The enzymes may be derived from plants, bacteria or fungi (including filamentous fungi and yeasts).

Suitable examples from fungi include a laccase derivable from a strain of Aspergillus, Neurospora, e.g., N. crassa, Podospora, Botrytis, Collybia, Fomes, Lentinus, Pleurotus, Trametes, e.g., T. villosa and T. versicolor, Rhizoctonia, e.g., R. solani, Coprinopsis, e.g., C. cinerea, C. comatus, C. friesii, and C. plicatilis, Psathyrella, e.g., P. condelleana, Panaeolus, e.g., P. papilionaceus, Myceliophthora, e.g., M. thermophila, Schytalidium, e.g., S. thermophilum, Polyporus, e.g., P. pinsitus, Phlebia, e.g., P. radiata (WO 92/01046), or Coriolus, e.g., C. hirsutus (JP 2238885).

Suitable examples from bacteria include a laccase derivable from a strain of Bacillus.

A laccase derived from *Coprinopsis* or *Myceliophthora* is preferred; in particular a laccase derived from *Coprinopsis cinerea*, as disclosed in WO 97/08325; or from *Myceliophthora thermophila*, as disclosed in WO 95/33836.

The detergent enzyme(s) may be included in a detergent composition by adding separate additives containing one or more enzymes, or by adding a combined additive comprising all of these enzymes. A detergent additive of the invention, *i.e.*, a separate additive or a combined additive, can be formulated, for example, as a granulate, liquid, slurry, etc. Preferred detergent additive formulations are granulates, in particular non-dusting granulates, liquids, in particular stabilized liquids, or slurries.

Non-dusting granulates may be produced, e.g. as disclosed in US 4,106,991 and 4,661,452 and may optionally be coated by methods known in the art. Examples of waxy coating materials are polyethyleneglycol (PEG) with mean molar weights of 1000 to 20000; ethoxylated nonylphenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and monoand di- and triglycerides of fatty acids. Examples of film-forming coating materials suitable for application

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by fluid bed techniques are given in GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Protected enzymes may be prepared according to the method disclosed in EP 238,216.

Adjunct materials

Any detergent components known in the art for use in detergents may also be utilized. Other optional detergent components include anti-corrosion agents, anti-shrink agents, anti-soil redeposition agents, anti-wrinkling agents, bactericides, binders, corrosion inhibitors, disintegrants/disintegration agents, dyes, enzyme stabilizers (including boric acid, borates, CMC, and/or polyols such as propylene glycol), fabric conditioners including clays, fillers/processing aids, fluorescent whitening agents/optical brighteners, foam boosters, foam (suds) regulators, perfumes, soil-suspending agents, softeners, suds suppressors, tarnish inhibitors, and wicking agents, either alone or in combination. Any ingredient known in the art for use in detergents may be utilized. The choice of such ingredients is well within the skill of the artisan.

Dispersants

The detergent compositions of the present invention can also contain dispersants. In particular powdered detergents may comprise dispersants. Suitable water-soluble organic materials include the homo- or co-polymeric acids or their salts, in which the polycarboxylic acid comprises at least two carboxyl radicals separated from each other by not more than two carbon atoms. Suitable dispersants are for example described in Powdered Detergents, Surfactant science series volume 71, Marcel Dekker, Inc.

Dye transfer inhibiting agents

The detergent compositions of the present invention may also include one or more dye transfer inhibiting agents. Suitable polymeric dye transfer inhibiting agents include, but are not limited to, polyvinylpyrrolidone polymers, polyamine *N*-oxide polymers, copolymers of *N*-vinylpyrrolidone and *N*-vinylimidazole, polyvinyloxazolidones and polyvinylimidazoles or mixtures thereof. When present in a subject composition, the dye transfer inhibiting agents may be present at levels from about 0.0001 % to about 10%, from about 0.01% to about 5% or even from about 0.1% to about 3% by weight of the composition.

Fluorescent whitening agent

The detergent compositions of the present invention will preferably also contain additional components that may tint articles being cleaned, such as fluorescent whitening agent or optical brighteners. Where present the brightener is preferably at a level of about 0.01% to about 0.5%. Any fluorescent whitening agent suitable for use in a laundry detergent composition may be used in the composition of the present invention. The most commonly used fluorescent whitening agents are

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those belonging to the classes of diaminostilbene-sulfonic acid derivatives, diarylpyrazoline derivatives and bisphenyl-distyryl derivatives. Examples of the diaminostilbene-sulfonic acid derivative type of fluorescent whitening agents include the sodium salts of: 4,4'-bis-(2diethanolamino-4-anilino-s-triazin-6-ylamino) stilbene-2,2'-disulfonate, 4,4'-bis-(2,4-dianilino-striazin-6-ylamino) stilbene-2.2'-disulfonate, 4,4'-bis-(2-anilino-4-(N-methyl-N-2-hydroxy-ethylamino)stilbene-2,2'-disulfonate, s-triazin-6-ylamino) 4,4'-bis-(4-phenyl-1,2,3-triazol-2-yl)stilbene-2,2'disulfonate and sodium 5-(2H-naphtho[1,2-d][1,2,3]triazol-2-yl)-2-[(E)-2phenylvinyl]benzenesulfonate. Preferred fluorescent whitening agents are Tinopal DMS and Tinopal CBS available from Ciba-Geigy AG, Basel, Switzerland. Tinopal DMS is the disodium salt of 4,4'-bis-(2-morpholino-4-anilino-s-triazin-6-ylamino) stilbene-2,2'-disulfonate. Tinopal CBS is the disodium salt of 2,2'-bis-(phenyl-styryl)-disulfonate. Also preferred are fluorescent whitening agents is the commercially available Parawhite KX, supplied by Paramount Minerals and Chemicals, Mumbai, India. Other fluorescers suitable for use in the invention include the 1-3-diaryl pyrazolines and the 7alkylaminocoumarins.

Suitable fluorescent brightener levels include lower levels of from about 0.01, from 0.05, from about 0.1 or even from about 0.2 wt % to upper levels of 0.5 or even 0.75 wt%.

Soil release polymers

The detergent compositions of the present invention may also include one or more soil release polymers which aid the removal of soils from fabrics such as cotton and polyester based fabrics, in particular the removal of hydrophobic soils from polyester based fabrics. The soil release polymers may for example be nonionic or anionic terephthalte based polymers, polyvinyl caprolactam and related copolymers, vinyl graft copolymers, polyester polyamides see for example Chapter 7 in Powdered Detergents, Surfactant science series volume 71, Marcel Dekker, Inc. Another type of soil release polymers are amphiphilic alkoxylated grease cleaning polymers comprising a core structure and a plurality of alkoxylate groups attached to that core structure. The core structure may comprise a polyalkylenimine structure or a polyalkanolamine structure as described in detail in WO 2009/087523 (hereby incorporated by reference). Furthermore random graft co-polymers are suitable soil release polymers. Suitable graft co-polymers are described in more detail in WO 2007/138054, WO 2006/108856 and WO 2006/113314 (hereby incorporated by reference). Other soil release polymers are substituted polysaccharide structures especially substituted cellulosic structures such as modified cellulose deriviatives such as those described in EP 1867808 or WO 2003/040279 (both are hereby incorporated by reference). Suitable cellulosic polymers include cellulose, cellulose ethers, cellulose esters, cellulose amides and mixtures thereof. Suitable cellulosic polymers include anionically modified cellulose, nonionically modified cellulose, cationically modified cellulose, zwitterionically modified cellulose, and mixtures thereof. Suitable cellulosic polymers include methyl cellulose, carboxy methyl cellulose, ethyl cellulose, hydroxyl ethyl cellulose, hydroxyl propyl methyl cellulose, ester carboxy methyl cellulose, and mixtures thereof.

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Anti-redeposition agents

The detergent compositions of the present invention may also include one or more antiredeposition agents such as carboxymethylcellulose (CMC), polyvinyl alcohol (PVA), polyvinylpyrrolidone (PVP), polyoxyethylene and/or polyethyleneglycol (PEG), homopolymers of acrylic acid, copolymers of acrylic acid and maleic acid, and ethoxylated polyethyleneimines. The cellulose based polymers described under soil release polymers above may also function as antiredeposition agents.

Rheology Modifiers

The detergent compositions of the present invention may also include one or more rheology modifiers, structurants or thickeners, as distinct from viscosity reducing agents. The rheology modifiers are selected from the group consisting of non-polymeric crystalline, hydroxy-functional materials, polymeric rheology modifiers which impart shear thinning characteristics to the aqueous liquid matrix of a liquid detergent composition. The rheology and viscosity of the detergent can be modified and adjusted by methods known in the art, for example as shown in EP 2169040.

Formulation of detergent products

The detergent composition of the invention may be in any convenient form, e.g., a bar, a homogenous tablet, a tablet having two or more layers, a pouch having one or more compartments, a regular or compact powder, a granule, a paste, a gel, or a regular, compact or concentrated liquid.

Pouches can be configured as single or multicompartments. It can be of any form, shape and material which is suitable for hold the composition, e.g. without allowing the release of the composition to release of the composition from the pouch prior to water contact. The pouch is made from water soluble film which encloses an inner volume. Said inner volume can be divided into compartments of the pouch. Preferred films are polymeric materials preferably polymers which are formed into a film or sheet. Preferred polymers, copolymers or derivates thereof are selected polyacrylates, and water soluble acrylate copolymers, methyl cellulose, carboxy methyl cellulose, sodium dextrin, ethyl cellulose, hydroxyethyl cellulose, hydroxypropyl methyl cellulose, malto dextrin, poly methacrylates, most preferably polyvinyl alcohol copolymers and, hydroxypropyl methyl cellulose (HPMC). Preferably the level of polymer in the film for example PVA is at least about 60%. Preferred average molecular weight will typically be about 20,000 to about 150,000. Films can also be of blended compositions comprising hydrolytically degradable and water soluble polymer blends such as polylactide and polyvinyl alcohol (known under the Trade reference M8630 as sold by MonoSol LLC, Indiana, USA) plus plasticisers like glycerol, ethylene glycerol, propylene glycol, sorbitol and mixtures thereof. The pouches can comprise a solid laundry cleaning composition or part components and/or a liquid cleaning composition or part components separated by the water soluble film. The compartment for liquid components can be different in composition than compartments containing solids: US2009/0011970 A1.

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Detergent ingredients can be separated physically from each other by compartments in water dissolvable pouches or in different layers of tablets. Thereby negative storage interaction between components can be avoided. Different dissolution profiles of each of the compartments can also give rise to delayed dissolution of selected components in the wash solution.

A liquid or gel detergent, which is not unit dosed, may be aqueous, typically containing at least 20% by weight and up to 95% water, such as up to about 70% water, up to about 65% water, up to about 55% water, up to about 45% water, up to about 35% water. Other types of liquids, including without limitation, alkanols, amines, diols, ethers and polyols may be included in an aqueous liquid or gel. An aqueous liquid or gel detergent may contain from 0-30% organic solvent.

A liquid or gel detergent may be non-aqueous.

Laundry soap bars

The enzymes of the invention may be added to laundry soap bars and used for hand washing laundry, fabrics and/or textiles. The term laundry soap bar includes laundry bars, soap bars, combo bars, syndet bars and detergent bars. The types of bar usually differ in the type of surfactant they contain, and the term laundry soap bar includes those containing soaps from fatty acids and/or synthetic soaps. The laundry soap bar has a physical form which is solid and not a liquid, gel or a powder at room temperature. The term solid is defined as a physical form which does not significantly change over time, i.e. if a solid object (e.g. laundry soap bar) is placed inside a container, the solid object does not change to fill the container it is placed in. The bar is a solid typically in bar form but can be in other solid shapes such as round or oval.

The laundry soap bar may contain one or more additional enzymes, protease inhibitors such as peptide aldehydes (or hydrosulfite adduct or hemiacetal adduct), boric acid, borate, borax and/or phenylboronic acid derivatives such as 4-formylphenylboronic acid, one or more soaps or synthetic surfactants, polyols such as glycerine, pH controlling compounds such as fatty acids, citric acid, acetic acid and/or formic acid, and/or a salt of a monovalent cation and an organic anion wherein the monovalent cation may be for example Na⁺, K⁺ or NH₄⁺ and the organic anion may be for example formate, acetate, citrate or lactate such that the salt of a monovalent cation and an organic anion may be, for example, sodium formate.

The laundry soap bar may also contain complexing agents like EDTA and HEDP, perfumes and/or different type of fillers, surfactants e.g. anionic synthetic surfactants, builders, polymeric soil release agents, detergent chelators, stabilizing agents, fillers, dyes, colorants, dye transfer inhibitors, alkoxylated polycarbonates, suds suppressers, structurants, binders, leaching agents, bleaching activators, clay soil removal agents, anti-redeposition agents, polymeric dispersing agents, brighteners, fabric softeners, perfumes and/or other compounds known in the art.

The laundry soap bar may be processed in conventional laundry soap bar making equipment such as but not limited to: mixers, plodders, e.g a two stage vacuum plodder, extruders, cutters, logo-stampers, cooling tunnels and wrappers. The invention is not limited to preparing the laundry soap bars by any single method. The premix of the invention may be added to the soap at different stages of the

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process. For example, the premix containing a soap, the enzyme of the invention, optionally one or more additional enzymes, a protease inhibitor, and a salt of a monovalent cation and an organic anion may be prepared and and the mixture is then plodded. The enzyme of the invention and optional additional enzymes may be added at the same time as the protease inhibitor for example in liquid form. Besides the mixing step and the plodding step, the process may further comprise the steps of milling, extruding, cutting, stamping, cooling and/or wrapping.

Formulation of enzyme in co-granule

The enzyme comprised in the detergent compositions of the invention may be formulated as a granule for example as a co-granule that combines one or more enzymes. Each enzyme will then be present in more granules securing a more uniform distribution of enzymes in the detergent. This also reduces the physical segregation of different enzymes due to different particle sizes. Methods for producing multi-enzyme co-granulates for the detergent industry are disclosed in the IP.com disclosure IPCOM000200739D.

Another example of formulation of enzymes by the use of co-granulates are disclosed in WO 2013/188331, which relates to a detergent composition comprising (a) a multi-enzyme co-granule; (b) less than 10 wt zeolite (anhydrous basis); and (c) less than 10 wt phosphate salt (anhydrous basis), wherein said enzyme co-granule comprises from 10 to 98 wt% moisture sink component and the composition additionally comprises from 20 to 80 wt% detergent moisture sink component. WO 2013/188331 also relates to a method of treating and/or cleaning a surface, preferably a fabric surface comprising the steps of (i) contacting said surface with the detergent composition as claimed and described herein in an aqueous wash liquor, (ii) rinsing and/or drying the surface.

The multi-enzyme co-granule may comprise an enzyme of the invention and (a) one or more enzymes selected from the group consisting of first- wash lipases, cleaning cellulases, xyloglucanases, perhydrolases, peroxidases, lipoxygenases, laccases and mixtures thereof; and (b) one or more enzymes selected from the group consisting of hemicellulases, proteases, care cellulases, cellobiose dehydrogenases, xylanases, phospholipases, esterases, cutinases, pectinases, mannanases, pectate lyases, keratinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, tannases, pentosanases, lichenases glucanases, arabinosidases, hyaluronidase, chondroitinase, amylases, and mixtures thereof.

Use in degrading xanthan gum

Xanthan gum is used as an ingredient in many consumer products including foods and cosmetics as well as in the oil and drilling industry. Therefore, enzymes having xanthan degrading activity can be applied in improved cleaning processes, such as the easier removal of stains containing xanthan gum, as well as the degradation of xanthan gum, which is often used in the oil and drilling industry. Thus, the present invention is directed to the use of the detergent composition of the invention to degrade xanthan gum. The detergent composition of present invention can also comprise a combination of an enzyme as described herein and a xanthan lyase. The use of such a

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detergent composition to degrade xanthan gum is also envisaged.

Degradation of xanthan gum may be measured using the viscosity reduction assay as described herein on xanthan gum. Xanthan degrading activity may alternatively be measured as reducing ends on xanthan gum using the colorimetric assay developed by Lever (1972), *Anal. Biochem.* 47: 273-279, 1972.

Use in detergents

The present invention is directed to the use of the detergent compositions of the invention in cleaning processes such as the laundering of textiles and fabrics (e.g. household laundry washing and industrial laundry washing), as well as household and industrial hard surface cleaning, such as dish wash.

An embodiment is the use of a detergent composition comprising a combination of the enzymes as described herein together with xanthan lyases in cleaning processes such as the laundering of textiles and fabrics (e.g. household laundry washing and industrial laundry washing), as well as household and industrial hard surface cleaning, such as dish wash.

The invention also relates to methods for degrading xanthan gum on the surface of a textile or hard surface, such as dish wash, comprising applying a detergent composition comprising one or more enzymes as described herein to xanthan gum. The invention further relates to methods for degrading xanthan gum on the surface of a textile or hard surface, such as dish wash, comprising applying a detergent composition comprising one or more xanthan lyases to xanthan gum. An embodiment is a method for degrading xanthan gum on the surface of a textile or hard surface, such as dish wash, comprising applying a detergent composition comprising one or more enzymes as described herein together with one or more xanthan lyase to xanthan gum. An embodiment is the detergent composition comprising one or more detergent components as described above.

The present invention is further described by the following examples that should not be construed as limiting the scope of the invention.

Examples

Activity assays

Xanthan lyase activity assay

0.8 mL 100 mM HEPES buffer, pH 6.0 was mixed with 0.2 mL Xanthan gum (5 mg/mL) dissolved in water in a 1 mL 1 cm cuvette. The cuvette was inserted into a spectrophotometer (Agilent G1103A 8453A, CA, USA) with temperature control set at 40 °C. The solution was pre-incubated for 10 min and 0.1 mL sample was added and the solution was mixed by aspiring and dispensing the solution for at least 5 times using a pipette. Total reaction volume was 1.1 mL. Absorbance at 235

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nm was collected for 10 min using a 30 sec measuring interval. Initial activity was calculated by using the software (UV-Visible Chemstation Rev A.10.01 [81], Agilent).

Example 1: Strain and DNA

The DNA in SEQ ID NO: 1 encoding the GH5 polypeptide EXa of SEQ ID NO: 2 was obtained from an *Opitutaceae* species isolated from an environmental soil sample collected in Denmark.

The DNA SEQ ID NO: 3 encoding the GH5 polypeptide EXb of SEQ ID NO: 4 was isolated from an environmental sample collected in Denmark.

The DNA SEQ ID NO: 5 encoding the GH5 polypeptide EXc of SEQ ID NO: 5 was isolated from an environmental sample collected in Denmark.

The DNA SEQ ID NO: 7 encoding the GH5 polypeptide EXd of SEQ ID NO: 8 was obtained from the public database (UNIPROT M2V1S3) but originates from a strain of *Pseudomonas stutzeri* collected from a Galapagos Rift hydrothermal vent, Ecuador.

Codon optimized synthetic DNA encoding the mature peptide sequences of the four polypeptides were prepared (SEQ ID NO: 9; SEQ ID NO: 10, SEQ ID NO: 11; SEQ ID NO: 12).

Example 2: Cloning and expression of GH5 polypeptides

The GH5 encoding genes were either cloned by conventional techniques from the strains indicated above or from the synthetic DNA and inserted into a suitable plasmid as described below.

Example 2a: Cloning and expression of GH5 polypeptides in *E.coli*

The mature peptide encoding part of the GH5 endo-glucanase genes, SEQ ID NO: 1, 3, 5 and 7 was inserted with an N-terminal poly histidine tag with an extra proline and arginine (HHHHHHPR) (SEQ ID NO: 19) after the methionine in the E.coli pET-32a(+) vector from Novagen with standard recombinant techniques. The expression plasmid containing the insert was purified from an *E.coli* transformant harboring the plasmid and transformed into E.coli Xjb (DE3) host cells (from Zymo Research). A fresh clone of *E.coli* Xjb (DE3) containing the pET32-GH5 vector, was grown overnight in Terrific Broth containing 100 ug/ml ampicillin. Next day, a fresh 500 ml culture was inoculated with 1 ml overnight culture and cells were cultured (37 °C, 250 rpm) to an optical density (OD600) between 6-8. Protein expression was induced by 1 mM isopropylthio-D-galactosidase (IPTG) and 6 mM arabinose for 4.5 hours at 20 °C. After continued culture, cells were harvested by centrifugation and lysed by Bugbuster® (Novagen). The soluble fraction was used for polyhistidine tag purification of the GH5 polypeptides SEQ ID NO: 13, 14 and 15 as described in example 4.

Example 2b: Cloning and expression of GH5 polypeptides in Bacillus subtilis

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The synthetic codon optimized genes SEQ ID NO: 10, 11 and 12 were cloned into the *Bacillus* expression vector described in WO 2012/025577. The genes were expressed by replacing the native secretion signal sequence with the *Bacillus clausii* secretion signal MKKPLGKIVASTALLISVAFSSSIASA (SEQ ID NO: 20) with an extra affinity tag sequence (HHHHHHPR) (SEQ ID NO: 19) at the C-terminal of the signal peptide, to facilitate the purification process. This resulted in a recombinant mature polypeptide with a His tag at the front of the N-terminal of the mature wild type sequence (SEQ ID NO: 16, 17 and 18).

One clone with the correct recombinant gene sequence was selected and the corresponding plasmid was integrated by homologous recombination into the *Bacillus subtilis* host cell genome (pectate lyase locus) and the gene construct was expressed under the control of a triple promoter system as described in WO99/43835. The gene coding for chloramphenicol acetyltransferase was used as a marker (as described in Diderichsen *et al.*, 1993, Plasmid 30:312-315).

Chloramphenicol resistant transformants were analyzed by PCR to verify the correct size of the amplified fragment. A recombinant *B. subtilis* clone containing the integrated expression construct was selected and cultivated on a rotary shaking table in 500 mL baffled Erlenmeyer flasks each containing 100 ml yeast extract-based media. The clone was cultivated for 5 days at 30°C. The enzyme containing supernatants were harvested and the enzyme purified as described in Example 5.

Example 3: Purification of wild type GH5 polypeptide from the natural *Opitutaceae* strain

The *Opitutaceae* strain was cultivated on a rotary shaking table in 500 mL baffled Erlenmeyer flasks each containing 100 ml mineral solution with 0.5% xanthan gum. The strain was cultivated for 20 days at 30°C. A total of 2.0 L supernatant was harvested by centrifugation and was filtered using a 0.2 µm bottle top filter (Nalgene Nunc). The broth was concentrated to 300 mL using ultra-filtration (Sartorius) with 30 kDa cut-off. Equal volume of 3.2 M ammonium sulphate in 40 mM Tris-HCl, pH 7.9 was slowly added with continuous stirring. The sample was filtered using Whatman glass filters (1.7 µm – 0.7 µm) to remove larger particles. The sample was applied on a 20 mL Phenylsepharose high performance column (GE Healthcare) pre-equilibrated with 1.6 M ammonium sulphate in 20 mM Tris-HCl, pH 7.9 (equilibration buffer). Unbound protein was eluted by two column volumes of equilibration buffer. Elution was done by a 12 column volume linear gradient from 1.6 M to 0.0 M ammonium sulphate in 20 mM Tris-HCl, pH 7.9. A last elution step of 4 column volume with equilibration buffer was used to elute tightly bound protein. The absorbance at 280 nm was recorded during the entire purification. Protein containing fractions identified by the absorbance at 280 nm in the chromatogram were analyzed by SDS-PAGE (NuPAGE, Invitrogen). Fractions judged as pure were pooled. The sample was concentration from 30 to 4 mL using Macrosep ultra filtration device

with 3 kDa cut-off (Pall). The protein concentration was determined by measuring the absorbance at 280 nm using the calculated extinction coefficient where 1 mg/mL equaled 1.89 absorbance units.

Example 4: Purification of recombinant GH5 polypeptide produced in *E.coli*

200 mL lysed cells (grown as example 2a) were filtered through Fast PES 0.2 µm bottletop filters to remove debris and unbroken cells. 200 mL of equilibration buffer (20 mM Tris-HCl, pH 7.5 + 500 mM NaCl) was added to the crude protein solution. A 20 mL HisPrep column loaded with Ni²⁺ was equilibrated with equilibration buffer until a stable UV baseline was obtained. The absorbance at 280 nm was continuously monitored throughout the purification. Crude protein was loaded on the column using a flow rate of 4 mL/min. Unbound protein was removed by washing the column with equilibration buffer until a stable UV baseline was obtained. Elution was carried out by a two-step linear gradient using 20 mM Tris-HCl, pH 7.5 + 500 mM NaCl + 500 mM Imidazole (elution buffer). First elution gradient was 10 column volumes 0 to 40 % elution buffer followed by 4 column volumes from 40% to 100 %. Peaks absorbing at 280 nm were analyzed by SDS-PAGE (NuPAGE, Invitrogen). Fractions containing protein with the correct apparent molecular weight were pooled. The pool was desalted and buffer exchanged using a Sephadex G-25 super fine desalting column equilibrated with 20 mM Tris-HCl, pH 8.0. The pool was applied on a 20 mL Source15Q column preequillibrated with 20 mM Tris-HCl, pH 8.0. Unbound protein was washed out using 20 mM Tris-HCl, pH 8.0 until a stable UV baseline was obtained. Elution was done by a 10 column volume linear NaCl gradient from 0 to 500 mM NaCl in 20 mM Tris-HCl, pH 8.0. Protein containing fractions were analyzed by SDS-PAGE and fractions judged as pure were pooled. Protein concentration was measured using absorbance at 280 nm using a calculated extinction coefficient where 1 mg/mL corresponded to 1.86 absorbance units.

Example 5: Purification of recombinant GH5 polypeptide produced in B. subtilis

All His-tagged enzymes were purified by immobilized metal chromatography (IMAC) using Ni²⁺ as the metal ion on 5 mL HisTrap Excel columns (GE Healthcare Life Sciences). The purification was done at pH 8 and the bound proteins were eluted with imidazole. The purity of the purified enzymes was checked by SDS-PAGE and the concentration of each enzyme determined by Abs 280 nm after a buffer exchange.

Example 6: Xanthan degrading activity of GH5 polypeptide and xanthan lyase on xanthan gum by measurement of viscosity reduction

The viscosity reduction measurements were performed using the viscosity pressure assay described in WO2011/107472 and following the method described in WO2013167581. Results presented are the average of three measurements and are shown in table 1 and 2 below.

A sample size of was 400 μ L was used. The hydrolysis conditions were as follows: 30 °C, either 0.25% or 0.5% xanthan gum (XG) in 50 mM MES buffer + 0.01% triton x-100 pH 7.0 or 100mM CHES buffer + 0.01% triton x-100 pH10. Enzyme was added upon thermal equilibration. Prior to use all enzymes were buffer changed to the MES buffer using NAP 5 columns (GE Healthcare).

The purified enzyme preparations of Example 5 were used for the analysis at a concentration of 31.25 mg/L.

Table 1: Viscosity measurements (Pa) of EXa (SEQ ID NO:13) and/or Xanthan Lyase (SEQ ID NO: 21) on 0.5% xanthan gum at pH 7.						
	T= 0 minutes	T= 30 minutes	T= 1 hour	T= 2 hours	T= 3 hours	T= 4 hours
Water (control)	430±44	504±50	470±75	483±86	466±60	504±82
Xanthan gum (control)	1703±132	1738±26	1837±122	1803±64	1739±84	1757±21
Xanthan gum + EXa SEQ ID NO:13	1586±101	1154±38	1270±67	1230±36	1156±49	1184±44
Xanthan gum + XLa SEQ ID NO:21	1963±93	1884±67	1890±84	1840±131	1886±50	1950±25
Xanthan gum + EXa SEQ ID NO:13 + XLa SEQ ID NO:21	1370±197	861±23	973±59	840±62	916±47	904±79

The results presented above show that the GH5 polypeptide alone and in combination with xanthan lyase can degrade the xanthan gum present in the media at pH 7, thus leading to viscosity reduction. A synergistic effect is obtained with combination of GH5 and xanthan lyase.

Table 2: Viscosity measurements (Pa) of EXa (SEQ ID NO:13) and/or Xanthan Lyase (SEQ ID NO: 23) on 0.5% xanthan gum at pH10						
	T=0	T=0.5 hours	T=1 hours	T=2 hours	T=3,5 hours	
Water	370±10	454±15	519±60	411±29	554±180	
Xanthan gum (XG) control	1740±151	1734±21	1819±67	1795±29	1898±75	
XG + EXa SEQ ID NO:13	1676±50	1324±58	1223±12	1251±31	1318±62	
XG + XLc SEQ ID NO:23	2046±112	1811±82	1773±64	1781±92	1704±67	
XG + EXa SEQ ID NO:13 + XLc SEQ ID NO:23	1573±227	1057±21	1153±12	1161±40	1188±89	

The results presented above show that the GH5 polypeptide in alone or combination with xanthan lyase can degrade the xanthan gum present in the media at pH 10, thus leading to viscosity reduction.

Table 3: Viscosity measurements (Pa) of EXa (SEQ ID NO:13), EXd (SEQ ID NO:18) and/or Xanthan						
Lyase (XLa, SEQ ID NO: 21) on 0.5% xanthan gum at pH 7.						
T=C	T= 0	5 hours	T= 1 hours	T= 2 hours	T= 3 hours	

Water control	440	410	333	413	469
Xanthan gum (XG) control	1626	1590	1546	1566	1659
XG + EXa SEQ ID NO:13	1220	1080	1046	1040	1079
XG + EXa SEQ ID NO:13 + XLa					
SEQ ID NO:21	1263	850	786	793	815
XG + EXd SEQ ID NO:18	1476	1406	1313	1283	1245
XG + EXd SEQ ID NO:18 +					
XLa SEQ ID NO:21	1490	1056	1023	933	912

The results presented above show that the GH5 polypeptide alone and in combination with xanthan lyase can degrade the xanthan gum present in the media at pH 7, thus leading to viscosity reduction.

Table 4: Viscosity measurements (Pa) of EXa, EXb, EXc recombinantly expressed in E.coli (SEQ ID NO:13; SEQ ID NO:14, SEQ ID NO:15) and/or Xanthan Lyase (XLb, SEQ ID NO: 22) on 0.5% xanthan gum at pH7. T=00 is before addition of enzyme and T=0 is right after

gum at pH7. I=00 is before addition of enzyme and I=0 is right after.							
	T=00	T=0	T=30 min	T=1hr	T=2hrs	T=3hrs	T=4hrs
water	541±21	544±119	519±142	545±70	399±80	422±114	326±25
Xanthan gum control	1878±20	1444±15	1599±91	1571±64	1605±38	1586±40	1566±32
XG + XLb SEQ ID NO:22	1898±26	1511±12	1522±56	1505±20	1579±80	1516±21	1559±38
XG + EXb SEQ ID NO:14	1884±31	1281±55	1202±120	1145±52	1132±70	1096±60	1116±114
XG + EXc SEQ ID NO:15	1931±45	1444±80	1122±36	1108±42	1105±45	1019±10	1059±15
XG + EXa SEQ ID NO:13	1891±12	1441±38	1102±17	1051±25	1005±6	969±26	1036±25
XG + EXb SEQ ID NO:14 +XLb SEQ ID NO:22	1918±61	1121±6	862±17	731±31	689±25	652±40	576±40
XG + EXc SEQ ID NO:15 +XLb SEQ ID NO:22	1911±	1111±	935±	848±	832±	822±	789±
XG + EXa SEQ ID NO:13 +XLb SEQ ID NO:22	1934±31	1198±36	855±40	831±40	785±23	909±26	819±64

The results presented above show that the GH5 polypeptides EXa, EXb and EXc alone and in combination with xanthan lyase can degrade the xanthan gum present in the media at pH 7, thus leading to viscosity reduction. A synergistic effect is obtained with combination of GH5 polypeptide and xanthan lyase.

Table 5: Viscosity measurements (Pa) of EXa, recombinantly expressed in E. coli (SEQ ID NO:13) and EXb and EXc recombinantly expressed in B. subtilis (SEQ ID NO:16 and SEQ ID NO:17) and/or Xanthan Lyase (XLb, SEQ ID NO: 22) on 0.5% xanthan gum at pH 7. T=00 is before addition of enzyme and T=0 is right after.

addition of oneymound	. 0.0.19.	it aitor.					
	T=00	T=0	T=30	T=1 hour	T=2	T=3	T=4
			min		hours	hours	hours

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Water	441±25	421±40	646±44	535±59	599±74	492±15	494±32
Xanthan gum(XG)	2027±2 3	1707±3 5	1949±5 9	1785±11 6	1746±7 5	1726±1 0	1867±6
XG + EXa SEQ ID NO:13	2054±4 4	1514±1 7	1299±2 1	1112±57	1089±4 5	1046±0	1027±6
XG + EXb SEQ ID NO:16	2067±1 5	1527±8 1	1393±1 2	1229±53	1159±1 2	1136±0	1134±6
XG + EXc SEQ ID NO:17	2061±3 1	1501±5 5	1416±4 4	1175±6	1183±7 8	1169±4 0	1147±1 5
XG + EXa SEQ ID NO:13 + XLb SEQ ID NO:22	2061±6	1274±1 7	1063±4 7	812±59	769±46	729±15	671±26
XG + EXb SEQ ID NO:20 + XLb SEQ ID NO:22	2074±2 6	1411±6 5	1079±1 5	945±92	809±12	796±10	781±10
XG + EXc SEQ ID NO:17 + XLb SEQ ID NO:22	2094±3 0	1491±2 5	1166±0	959±46	889±40	846±0	847±57
XG + XLb SEQ ID NO:22	2097±4 9	1794±6 2	1863±2 3	1685±15	1653±1 0	1679±6	1667±2 9
XG + EXa SEQ ID NO:13 + XLa SEQ ID NO:21	2131±1 5	1227±8 1	1143±8 1	789±62	739±25	716±44	677±55
XG + EXb SEQ ID NO:16 +XLa SEQ ID NO:21	2104±7 9	1324±1 7	1096±4 4	795±31	803±26	792±21	767±12
XG + EXc SEQ ID NO:17 +XLa SEQ ID NO:21	2107±1 2	1241±5 0	1163±3 2	802±15	826±15	846±0	894±15
XG + XLa SEQ ID NO:21	2134±2 0	1741±5 7	1933±2 9	1639±30	1659±2 3	1666±1 7	1637±1 2

The results presented above show that the GH5 polypeptides EXa, EXb and EXc alone and in combination with xanthan lyase can degrade the xanthan gum present in the media at pH 7, thus leading to viscosity reduction. A synergistic effect is obtained with combination of GH5 polypeptide and xanthan lyase.

Table 6: Viscosity measurements (Pa) of EXa, EXb, EXc recombinantly expressed in E.coli (SEQ ID NO:13; SEQ ID NO: 14 or SEQ ID NO: 15) and/or Xanthan Lyase (XLc, SEQ ID NO: 23 or SEQ ID NO:24) on 0.5% xanthan gum at pH 10. T=00 is before addition of enzyme and T=0 is right after.

	T=00	T=0	T=30'	T=1hr	T=2hrs	T=3hrs
Water	429±66	502±110	504±50	434±29	478±42	479±26
Xanthan gum (XG)	1932±31	1485±81	1678±12	1641±70	1642±38	1592±92
XG + EXa SEQ ID NO:13	1992±13 8	1332±6	1254±21	1147±51	1192±35	1215±31
XG + EXb SEQ ID NO:14	1989±85	1415±50	1351±66	1321±17	1358±51	1252±21
XG + EXc SEQ ID NO:17	1892±45	1442±10 0	1408±21	1341±50	1332±31	1262±51
XG + EXa SEQ ID NO:13 +XLc SEQ ID NO:23	1899±69	1429±62	1084±76	1131±17	1092±25	1112±40

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XG + EXb SEQ ID NO: 14	2019±62	1465±13	1144±23	1121±53	1108±81	1012±59
+XLc SEQ ID NO: 23		2				
XG + EXc SEQ ID NO: 15	2085±80	1602±38	1344±15	1321±10	1262±55	1319±10
+XLc SEQ ID NO:23						
XG + XLc SEQ ID NO:23	2005±47	1702±75	1588±6	1524±67	1588±60	1569±36
XG + EXa SEQ ID NO:13	1959±72	1462±11	1158±38	1144±40	1148±72	1005±45
+XLd SEQ ID NO:24		0				
XG + EXb SEQ ID NO:14	1975±25	1442±35	1211±26	1177±15	1192±72	1182±67
+XLd SEQ ID NO:24						
XG + EXc SEQ ID NO: 15	1925±13	1422±95	1238±12	1274±58	1208±81	1215±67
+XLd SEQ ID NO:24	3					
XG + XLd SEQ ID NO:24	1839±40	1525±61	1488±21	1447±42	1432±15	1425±76

The results presented above show that the GH5 polypeptides GH5, EXb and EXc in combination with xanthan lyase can degrade the xanthan gum present in the media at pH 10, thus leading to viscosity reduction.

Table 7: Viscosity measuren	nents (Pa) d	of GH5 po	olypeptide	purified	from superr	atant of the
Opitutaceae sp strain and/or	Xanthan Lya	ase (XLa,	SEQ ID N	O: 21) c	on 0.25% xan	ithan gum at
pH7						
	T-0	T-0 5 b -	T_ 4		T- 0 h	T- 2 h

	T=0	T=0.5 hour	T= 1 hour	T= 2 hours	T= 3 hours
Water	471±99	390±46	423±61	433±64	438±36
Xanthan gum (XG)	898±12	880±40	900±17	820±40	908±50
XG + EXa SEQ ID NO:1	856±34	743±46	723±34	672±38	644±55
XG + XLa SEQ ID NO:21	908±29	865±22	860±35	857±32	856±61
XG + EXa SEQ ID NO:1 + XLa SEQ ID NO:21	800±28	597±30	612±31	577±45	648±89

Example 8: Xanthan degrading activity of GH5 polypeptide and xanthan lyase on xanthan gum by measurement of viscosity reduction

The viscosity measurements were performed using the viscosity pressure assay described in WO2011/107472. 150 μ L of each 1 mL hydrolysis or control was the sample size. Results presented are the average of four measurements and are shown in table 8 and 9 below.

Modified xanthan gum was prepared by an adaption of Nankai et al. 1999. "Microbial system for polysaccharide depolymerization: enzymatic route for xanthan depolymerization by *Bacillus sp* strain GL1." Applied and Environmental Microbiology 65(6): 2520-2526.

2.5 g of xanthan gum (CP Kelco) was wetted with 5 mL of 96 % ethanol in a 2 L beaker. 500 mL of 100 mM ACES buffer pH 7.00 was added and the solution stirred at ambient temperature for 2 h. 250 μL of xanthan lyase (*Bacillus* sp., Megazyme) was added and the solution incubated for 20 h at 50 °C. The sample was then cooled by placing the beaker on ice. After hydrolysis was 1400 mL of ice cold 96 % ethanol was added to the 500 mL sample, under stirring. Precipitation occurs, and after approximately 5 min the ethanol was decanted removing the pyruvated mannose residues. The sample was vacuum filtered and transferred to a glass plate. The glasses were dried at 50 °C for 20 h. The sample was collected, weighed, and grinded.

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The hydrolysis conditions were as follows: 40 °C, 0.35 % xanthan gum (XG) in 50 mM HEPES buffer + 0.01 % triton X-100 pH 7.0. The modified xanthan gum powder (mXG) was prepared as described above and a 0.7 % solution was prepared using the same procedure as outlined for XG. Enzyme was added upon thermal equilibration. The initial viscosity is measured prior to enzyme addition, after thermal equilibration. Controls are the same with buffer added instead of enzyme. Buffer was monitored to determine the ultimate end point of a total hydrolysis.

Table 8. Viscosity measurements (Pa). EXc SEQ ID NO:17 and XLb (SEQ ID NO:22). Each enzyme								
dosed in 1.5 ppm. pH 7.0								
Time (Minutes)	0	15	30	45	60	75	90	
Buffer 50 mM HEPES								
Control	645	610	521	502	620	632	600	
Xanthan Gum + Buffer								
Control	2140	2075	1948	2092	2033	2077	2005	
Xanthan Gum + EXc	2120	1295	991	957	935	1112	917	
Xanthan Gum + EXc +								
Xanthan Lyase	1977	808	811	837	773	807	777	
Xanthan Gum + Xanthan								
lyase	1972	1853	1838	1802	1750	1737	1677	
Modified Xanthan Gum +								
Buffer Control	2262	2100	2143	2134	2118	2150	2097	
Modified Xanthan Gum +								
EXc	2217	1225	1173	1157	1130	1155	1130	

Example 9: Wash performance of GH5 polypeptide and xanthan lyase

The wash performance of the GH5 enzyme was assessed in laundry wash experiments using a Mini wash assay, which is a test method where soiled textile is continuously lifted up and down into the test solution and subsequently rinsed. The wash experiment was conducted under the experimental conditions specified in Table 10.

The textiles were subsequently air-dried and the wash performance was measured as the brightness of the color of the textiles. Brightness can be expressed as the Remission (R), which is a measure for the light reflected or emitted from the test material when illuminated with white light. The Remission (R) of the textiles was measured at 460 nm using a Zeiss MCS 521 VIS spectrophotometer. The measurements were done according to the manufacturer's protocol.

The performance of the new enzyme (combination) was compared to the performance of detergent alone (blank). An enzyme (combination) is considered to exhibit improved wash performance, if it performs better than the detergent alone (i.e. Renzyme > RBLANK) (see Table 13 and 14).

Table 10: Experimental setup of Mini wash assay							
Detergent	Liquid Model detergent A or Model detergent T (see Table 11 and 12)						
Detergent dose	3.33 g/l						
pН	"as is" in the current detergent solution and was not adjusted						
Water hardness	16°dH, adjusted by adding CaCl₂*2H₂O, MgCl₂*6H₂O and NaHCO₃ (5:1:3) to milli-Q water.						
Enzymes	EXc (SEQ ID NO:17), xanthan lyase (XLb, SEQ ID NO:22 or XLc SEQ ID NO:23)						
Enzyme dosage	Dosage of GH5: 0.05 mg EP/L (enzyme protein), 0.10 mg EP/L, 0.2 mg EP/L, 0.5 mg EP/L, 1.0 mg EP/L; experiments with combinations of GH5 and XL were conducted with a fixed concentration of 1.0 mg EP/L XL						
Volume of test solution	50 ml						
Test material	Xanthan Gum with carbon black DN-31D textile swatches (23x3 cm). The test material was obtained from Center for Testmaterials BV, P.O. Box 120, 3133 KT Vlaardingen, the Netherlands, and WFK Testgewebe GmbH, Christenfeld 10, D-41379 Brüggen, Germany						
Temperature	40°C						
Wash time	30 min						
Rinse time	5 min						
Test system	Soiled textile continuously lifted up and down into the test solutions, 50 times per minute (up-time 0.4 sec, down-time 0.4 sec, lift time 0.4 sec). The test solutions are kept in 125 ml glass beakers. After wash of the textiles are continuously lifted up and down into tap water, 50 times per minute (up-time 0.4 sec, down-time 0.4 sec, lift time 0.4 sec).						

Table 11: Composition of Model Detergent A (Liquid) 1)				
Detergent ingredients	Wt %			
Linear alkylbenzenesulfonic acid (LAS) (Marlon AS3)	13			
Sodium alkyl(C12)ether sulfate (AEOS) (STEOL CS-370 E)	10			
Coco soap (Radiacid 631)	2.75			
Soy soap (Edenor SJ)	2.75			
Alcohol ethoxylate (AEO) (Bio-Soft N25-7)	11			
Sodium hydroxide	2			
Ethanol	3			
Propane-1,2-diol (MPG)	6			
Glycerol	2			
Triethanolamine (TEA)	3			
Sodium formate	1			
Sodium citrate	2			
Diethylenetriaminepentakis(methylenephosphonic acid) (DTMPA)	0.2			
Polycarboxylate polymer (PCA) (Sokalan CP-5)	0.2			
Water	Up to 100			
1)The pH of the detergent was adjusted to pH 8 with sodium hydroxide or citric acid.				

Table 12. Composition of Model detergent T (powder)					
Detergent ingredients	Wt %				
LAS, sodium salt	11.72				
AS, sodium salt	2.0				
Soap, sodium salt	2.15				
AEO	3.0				
Soda ash	14.98				
Hydrous sodium silicate	3.12				
Zeolite A	18.75				
HEDP-Na4	0.15				
Sodium citrate	2.0				
PCA, copoly(acrylic acid/maleic acid), sodium salt	1.65				
SRP	0.5				
Sodium sulfate	13.53				
Sodium percarbonate	22.20				
TAED	3.25				
Foam regulator	1.0				

Table 13: Remission (R) values obtained in Mini Wash using EXc with and without xanthan lyase (XLb) in liquid model A detergent

Enzyme dosage	No enzyme	EXc	EXc + xanthan lyase
0.05 mg EP/L	29.5	32.8	35.1
0.1 mg EP/L	29.5	33.6	35.4
0.2 mg EP/L	29.5	34.3	35.9
0.5 mg EP/L	29.5	35.1	36.7
1.0 mg EP/L	29.5	35.4	37.3

Table 14 Remission (R) values obtained in Mini Wash using EXc with and without Xanthan Lyase (XLc)in powder model T detergent

Enzyme dosage	No enzyme	EXc	EXc + xanthan lyase
0.05 mg EP/L	29.8	29.7	29.7
0.1 mg EP/L	29.8	29.8	29.8
0.2 mg EP/L	29.8	30.0	30.0
0.5 mg EP/L	29.8	30.6	30.9
1.0 mg EP/L	29.8	31.0	31.2

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Example 10: Wash performance of combinations of a GH5 polypeptide and xanthan lyase was tested on specific stains

The wash performance of variants in liquid and powder detergents was determined by using the following standardized stains, all obtainable from CFT (Center for Testmaterials) B.V., Vlaardingen, Netherlands:

A: Fluid make-up: product no. PCS17

B: Fluid make-up: product no. CS17

For the tests in liquid detergents, a liquid washing agent with the following composition was used as base formulation (all values in weight percent): 0 to 0.5% xanthan gum, 0.2 to 0.4% antifoaming agent, 6 to 7% glycerol, 0.3 to 0.5% ethanol, 0 to 7% FAEOS (fatty alcohol ether sulfate), 10 to 28% nonionic surfactants, 0.5-1% boric acid, 1 to 2% sodium citrate (dihydrate), 2 to 4% soda, 0 to 16% coconut fatty acid, 0.5% HEDP (1-hydroxyethane-(1,1-diphosphonic acid)), 0 to 0.4% PVP (polyvinylpyrrolidone), 0 to 0.05% optical brighteners, 0 to 0.001% dye, remainder deionized water.

Based on this base formulation, detergent was prepared by adding the respective enzyme combination as indicated in table 15. As a reference, the detergent composition without addition of the enzyme combinations was used.

The dosing ratio of the liquid washing agent was 4.7 grams per liter of washing liquor and the washing procedure was performed for 60 minutes at a temperature of 40°C, the water having a water hardness between 15.5 and 16.5° (German degrees of hardness).

For the tests in solid detergents, a European premium detergent was used as base formulation.

The whiteness, i.e. the brightening of the stains, was determined photometrically as an indication of wash performance. A Minolta CM508d spectrometer device was used, which was calibrated beforehand using a white standard provided with the unit.

The results obtained are the difference values between the remission units obtained with the detergents and the remission units obtained with the detergent containing the enzyme combinations. A positive value therefore indicates an improved wash performance due to the enzyme combinations present in the detergent. It is evident from table 15 that enzyme combinations according to the invention show improved wash performance.

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Table 15: Wash performance in liquid detergent

Enzyme combination		Α	В
XLb SEQ ID NO:22+ EXc SEQ ID NO:17	Diff	3.3	6.4
	HSD	2.4	1.2

Table 16: Wash performance in solid detergent

Enzyme combination		В
XLb SEQ ID NO:22 + EXc SEQ ID NO:17	Diff	1.9
	HSD	1.2

Example 11: Wash performance of GH5 polypeptides with and without Xanthan Lyase

In this example wash performance of GH5 polypeptides was evaluated in a liquid model detergent A washed in the Automatic Mechanical Stress Assay (AMSA) at 20°C or 40°C. The wash performance of the enzymes was evaluated either alone or in combination with a Xanthan Lyase. The wash conditions used are specified in Table 17 below.

Table 17. Wash conditions used in the example 11:

Detergent	Liquid model detergent A
Detergent conc.	3.3 g/L
рН	"as is" in the current detergent solution and was not adjusted
Temperature	20°C or 40°C
Dosages in AMSA-plate	140μL detergent per slot; 20μL enzyme per slot
Water hardness	16°dH, adjusted by adding CaCl ₂ *2H ₂ O, MgCl ₂ *6H ₂ O and NaHCO ₃ (5:1:3) to milli-Q water
Enzymes	EXb (SEQ ID NO:16); EXc (SEQ ID NO:17), xanthan lyase (XLb, SEQ ID NO:22)
Enzyme dosage	EXb and EXc concentrations: 0.7, 1.5, 20, 125 ppb XLb concentration: 400 ppb
Test solution volume	160 micro L
Wash time	20 minutes
Stain/ swatch	Mayonnaise with carbon black C-S-05 S from CFT, Center for Testmaterials BV.

The enzyme and wash liquid were dosed into the AMSA plate and washed according to conditions listed in Table 17. After wash the fabric was flushed in tap water and air-dried. The performance of the enzyme was subsequently measured as the brightness of the colour of the textile samples. Brightness was measured as the intensity of the light reflected from the textile sample

when illuminated with white light. Intensity was measured with a professional flatbed scanner EPSON EXPRESSION 10000XL with special designed software that extracted the intensity value from the scanned imagine through standard vector calculations.

The performance of the enzyme (or combination of enzymes) was compared to the performance of detergent alone (blank) or detergent with the Xanthan lyase (XL). An enzyme (or combination of enzymes) was considered to exhibit improved wash performance if it performed better than the detergent alone (i.e., $R_{\text{ENZYME}} > R_{\text{BLANK}}$) (see Tables 18, 19, 20 and 21).

Table 18. Intensity and delta intensity of GH5 polypeptides EXb (SEQ ID NO:16) and EXc (SEQ ID NO:17) tested in AMSA at 20°C in model detergent A.

		Inten	sity	Delta intensity					
Concentration [ppb]	0.7	1.5	20	125	0.7	1.5	20	125	
Blank	210.4	210.4	210.4	210.4					
EXb (SEQ ID									
NO:16)	210.8	212.8	217.2	217.8	0.4	2.4	6.8	7.5	
EXc (SEQ ID									
NO:17)	212.0	214.4	216.5	218.4	1.6	4.1	6.2	8.0	

Table 19. Intensity and delta intensity of GH5 polypeptides EXb (SEQ ID NO:16) and EXc (SEQ ID NO:17) tested in AMSA at 40°C in model detergent A.

		Inter	nsity		Delta intensity						
Concentration											
[ppb]	0.7	1.5	20	125	0.7	1.5	20	125			
Blank	220.0	220.0	220.0	220.0							
EXb (SEQ ID											
NO:16)	221.9	222.9	229.4	230.2	1.9	3.0	9.4	10.2			
EXc (SEQ ID											
NO:17)	223.2	225.4	228.3	229.0	3.3	5.4	8.3	9.0			

Table 20. Intensity and delta intensity of GH5 polypeptides EXb (SEQ ID NO:16) and EXc (SEQ ID NO:17) with Xanthan Iyase (XLb (SEQ ID NO:22) tested in AMSA at 20°C in model detergent A.

		Inter	nsity		Delta intensity						
Concentration											
[ppb]	0.7	1.5	20	125	0.7	1.5	20	125			
Blank with XLb											
(SEQ ID NO:22)	214.0	214.0	214.0	214.0							
EXb (SEQ ID											
NO:16 with XLb											
(SEQ ID NO:22)	213.0	215.3	220.4	223.7	-1.0	1.3	6.4	9.7			
EXc (SEQ ID											
NO:17) with XLb											
(SEQ ID NO:22)	212.4	215.1	220.2	221.4	-1.6	1.1	6.2	7.4			

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Table 21. Intensity and delta intensity of GH5 polypeptides EXb (SEQ ID NO:16) and EXc (SEQ ID NO:17) with Xanthan lyase (XLb (SEQ ID NO:22) tested in AMSA at 40°C in model detergent A.

		Inter	nsity		Delta intensity						
Concentration											
[ppb]	0.7	1.5	20	125	0.7	1.5	20	125			
Blank with XLb											
(SEQ ID NO:22)	220.6	220.6	220.6	220.6							
EXb (SEQ ID NO:16 with XLb (SEQ ID NO:22)	222.0	225.0	231.0	232.6	1.3	4.4	10.3	12.0			
EXc (SEQ ID NO:17) with XLb (SEQ ID NO:22)	222.3	223.9	230.1	231.5	1.7	3.2	9.5	10.9			

The results in above tables show that the GH5 polypeptides, e.g., EXb and EXc, have an improved wash performance both when evaluated alone or in combination with the Xanthan Lyase, e.g., XLb.

The invention described and claimed herein is not to be limited in scope by the specific aspects herein disclosed, since these aspects are intended as illustrations of several aspects of the invention. Any equivalent aspects are intended to be within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims. In the case of conflict, the present disclosure including definitions will control.

Claims

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- 1. A detergent composition comprising a polypeptide of glycosyl hydrolase family 5 having xanthan degrading activity.
- 2. The detergent composition of claim 1, wherein the polypeptide is selected from the group consisting of:
- (a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide of SEQ ID NO: 6;
- (b) a polypeptide encoded by a polynucleotide that hybridizes under medium stringency conditions with (i) the mature polypeptide coding sequence of any of SEQ ID NO: 5, (ii), or the full-length complement of (i);
- (c) a polypeptide encoded by a polynucleotide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 5;
- (d) a variant of the mature polypeptide of any of SEQ ID NO: 6 comprising a substitution, deletion, and/or insertion at one or more positions;
- (e) a fragment of the polypeptide of (a), (b), (c), or (d) that has xanthan degrading activity; and
- (f) a polypeptide comprising the polypeptide of (a), (b), (c), (d), or (e) and a N-terminal and/or C-terminal His-tag.
- 3. The detergent composition of claim 1, wherein the polypeptide is selected from the group consisting of:
- (a) a polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide of any of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 8;
- (b) a polypeptide encoded by a polynucleotide that hybridizes under medium stringency conditions with (i) the mature polypeptide coding sequence of any of SEQ ID NO: 1, SEQ ID NO: 3, or SEQ ID NO: 7 (ii), or the full-length complement of (i);
 - (c) a polypeptide encoded by a polynucleotide having at least 60%, at least 65%, at

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least 70%, at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% sequence identity to the mature polypeptide coding sequence of any of SEQ ID NO: 1, SEQ ID NO: 3, or SEQ ID NO: 7;

- (d) a variant of the mature polypeptide of any of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 8 comprising a substitution, deletion, and/or insertion at one or more positions;
- (e) a fragment of the polypeptide of (a), (b), (c), or (d) that has xanthan degrading activity; and
- (f) a polypeptide comprising the polypeptide of (a), (b), (c), (d), or (e) and a N-terminal and/or C-terminal His-tag.
- 4. The detergent composition of any one of claims 1 to 3, the polypeptide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide of any of SEQ ID NO: 2, 4, 6, or 8.
- 5. The detergent composition of any of claims 1 to 4, wherein the polypeptide is encoded by a polynucleotide that hybridizes under medium-high stringency conditions with (i) the mature polypeptide coding sequence of any of SEQ ID NO: 1, 3, 5, or 7, or (ii) the full-length complement of (i).
- 6. The detergent composition of any one of claims 1 to 5, wherein the polypeptide is encoded by a polynucleotide having at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99% or 100% sequence identity to the mature polypeptide coding sequence of any of SEQ ID NO: 1, 3, 5, or 7.
- 7. The detergent composition of any of claims 1 to 6, wherein the polypeptide is a variant of the mature polypeptide of any of SEQ ID NO: 2, 4, 6, or 8 comprising a substitution, deletion, and/or insertion at one or more positions, such as up to 10, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 positions.
- 8. The detergent composition of any one of claims 1 to 7, wherein the polypeptide is a fragment of any of SEQ ID NO: 2, 4, 6, or 8, wherein the fragment has xanthan degrading activity.
- 9. The detergent composition of any one of claims 1 to 8, further comprising a polypeptide having xanthan lyase activity.
- 10. The detergent composition of claim 9, wherein the polypeptide having xanthan lyase activity

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is a polypeptide having the amino acid sequence of any one of SEQ ID NO: 21, 22, 23 or 24.

- 11. The detergent composition according to any one of claims 1 10, wherein the composition is in form of a bar, a homogenous tablet, a tablet having two or more layers, a pouch having one or more compartments, a regular or compact powder, a granule, a paste, a gel, or a regular, compact or concentrated liquid.
- 12. The detergent composition of any one of claims 1-11, the composition further comprising one or more additional enzymes selected among protease, lipase, cutinase, amylase, carbohydrase, cellulase, pectinase, mannanase, arabinase, galactanase, xylanase, oxidase, xanthanase, laccase, and/or peroxidase.
- 13. The detergent composition of any one of claims 1-12, wherein the composition is a laundry detergent composition or a dishwashing composition, preferably a machine dishwashing composition.
- 14. Use of a detergent composition according to any one of claims 1-13 in a cleaning process.
- 15. The use according to claim 14, wherein the cleaning process is laundry.
- 16. The use according to claim 15, wherein the cleaning process is hard surface cleaning such as dish wash.
- 17. A method for removing a stain from a surface which comprises contacting the surface with a detergent composition according to any one of claims 1 13.
- 18. Use of a detergent composition according to any of claims 1 to 13 for degrading xanthan gum.
- 19. The use of claim 18, wherein the detergent composition has an enzyme detergency benefit.
- 20. A method for degrading xanthan gum comprising applying a detergent composition according to any of claims 1 to 13 to xanthan gum.
- 21. The method of claim 20, wherein the xanthan gum is on the surface of a textile or of a hard surface, such as in dish wash.

eolf-seql SEQUENCE LISTING

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ccg ggo Pro Gly	ggc Gly	gtg Val	ctc Leu 65	cgc Arg	ctt Leu	tcc Ser	ggc GI y	ggc GI y 70	gcc Al a	ggc GI y	gtc Val	atc IIe	ggc GI y 75	atg Met	336
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acc Thr	gac Asp	ggc GI y	gaa Gl u 160	cgc Arg	tac Tyr	ctc Leu	ggc GI y	gtc Val 165	gtc Val	cgc Arg	gtc Val	tcc Ser	tcc Ser 170	ggc GI y	gcg Al a	6	524
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caa GI n	agt Ser 350	ctc Leu	gcc Al a	ctc Leu	ggt GI y	gac Asp 355	tgg Trp	gag GI u	atc IIe	gcc Al a	tcc Ser 360	ggc GI y	atc IIe	acc Thr	aag Lys	12	200
ctc	tcc	gcc	ggc	tcc	ggt	cgc	agc	ctc		ttc ge 2	gac	atc	gac	tac	ctc	12	248

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agc Ser	ctc Leu	gac Asp	gac Asp 400	ggc GI y	tcc Ser	ggc GI y	tac Tyr	acc Thr 405	ggc GI y	acg Thr	ctc Leu	aac Asn	cac Hi s 410	gcg Al a	tcc Ser	1344
ggc GI y	gcg Al a	ctc Leu 415	cgc Arg	ttc Phe	gag GI u	tcc Ser	gtc Val 420	ttc Phe	tcc Ser	acc Thr	gag GI u	ggc GI y 425	gcg Al a	ctc Leu	acc Thr	1392
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Trp	Pro	Al a	Tyr 640	Al a	GI n	Al a	Al a		eol f Asn			Arg	GI u 650	Val	Asn	
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	aac Asn															2304
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ctc Leu	gtc Val 750	gtg Val	ctg Leu	gac Asp	aac Asn	ttc Phe 755	ctc Leu	gcc Al a	tac Tyr	ctc Leu	gcg Al a 760	gcc Al a	gag GI u	ggc GI y	gtg Val	2400
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Pro	Al a	Arg	Al a -1	Ala 1	Asp	Tyr	Tyr	Leu 5	Lys	Al a	Ser	GI n	GI y 10	Al a	Ser	
Asn	Hi s	Trp 15	Ser	Ser	Hi s	Leu	Thr 20	Asp	Trp	Thr	Al a	Asn 25	Al a	Asp	GI y	
Thr	GI y	Al a	Asn	Pro	Thr	Val	Пе	GI y		Ala ge 4	Asp	Thr	Phe	Asp	Thr	

35

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Ala Ser Thr Phe Gly Gly Gly Ala Leu Ala Leu Ala Ser Gly Ala Lys Page 5

Ala Ala Asn Gly Ser Gly Val Ala Pro Thr Ser Ile Asn Gly Tyr Asp

Phe Tyr IIe Asp Gln Val Ser Asn Arg Glu IIe Arg Thr Pro Ser Thr

300

290

Leu Thr Leu Lys Ser Ser Pro Gly Val Val Ser Thr IIe Pro Ala Phe Val Asn Thr Asn Ser Pro IIe IIe Val Asn Gly Gly Ser Phe Arg 335 340 345 Gln Ser Leu Ala Leu Gly Asp Trp Glu IIe Ala Ser Gly IIe Thr Lys 350 355 360 Leu Ser Ala Gly Ser Gly Arg Ser Leu Gly Phe Asp IIe Asp Tyr Leu 365 370 375 380 Gly Gly Ala Gly Gly Leu Val Thr Gln Asn Gly Gly Ser Tyr Phe Leu 385 390 395 Ser Leu Asp Asp Gly Ser Gly Tyr Thr Gly Thr Leu Asn His Ala Ser 400 410 Gly Ala Leu Arg Phe Glu Ser Val Phe Ser Thr Glu Gly Ala Leu Thr 415 420 425 lle Gly Ser Ser Ala Thr Val His Leu Asp Gln Gln Val Tyr Val Thr Ser Phe Ser Val Ala Gly Val Ala Lys Ala Ala Gly Ile His Thr Tyr Ala Ser Leu Asn Ala Ala His Pro Ala Gln Phe Thr Ala Gly Ala Ala Pro Gly Leu Val Ala Val Tyr Thr Pro Asp Thr Ala Gly Pro Val Arg Met Asn Gly Val Asn IIe Ser Gly Pro Glu Ser Asn Thr Ala Asn Leu 495 500 505 Pro Gly Thr Tyr Gly Tyr Asn Tyr Val Tyr Pro Thr Glu Ala Asp Phe 510 520 Asp Tyr Tyr Ala Ser Lys Gly Leu Asn Leu IIe Arg IIe Pro Phe Arg 525 535 540 Trp Glu Arg Met Gln His Gly Leu Asn Val Pro Leu Asn Thr Ala Gln 550 Leu Gly Tyr Met Asp Thr Ala Val Ala Arg Ala Ser Ala Arg Gly Met 560 565

Lys Val IIe Leu Asp Met His Asn Tyr Ala Arg Cys Lys Val Gly Gly

575 580 585

Val Thr Tyr Lys Phe Gly Asp Ala Gln Leu Pro Ala Ser Ala Tyr Ala 590 595 600

Asp Val Trp Arg Arg Leu Ala Asp His Tyr Lys Asn Glu Pro Ala IIe 605 610 615

Tyr Gly Phe Asp IIe Met Asn Glu Pro Asn Gly Leu Ser Gly Gly Val 625 630 635

Trp Pro Ala Tyr Ala Gln Ala Ala Val Asn Ala Ile Arg Glu Val Asn 640 650

Leu Ser Thr Trp Val IIe Val Glu Gly Glu Phe Trp Ala Asn Ala Trp 655 660 665

Gly Phe Glu Thr Lys Asn Pro Tyr Leu His Asn Val Arg Asp Pro Val 670 680

Gly Arg Leu Met Phe Ser Ala His Ser Tyr Trp Ser Asp Ala Gly Thr 685 695 700

Asp Val Tyr Lys Thr Tyr Asp Glu Glu Gly Ala Tyr Pro Glu Met Gly 705 710 715

Val Asn Asn Val Lys Pro Phe IIe Asp Trp Leu Lys Lys His Asp Ala 720 725 730

Lys Gly Phe Val Gly Glu Tyr Gly Val Pro Asn Asn Asp Pro Arg Trp 745

Leu Val Val Leu Asp Asn Phe Leu Ala Tyr Leu Ala Ala Glu Gly Val 750 755 760

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ccg t Pro F																96
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ccg a Pro T																624
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									P71	'F A						

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                                                                                              2400
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eal f-seal

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Gly Lys Leu Thr Gly Ser Gly Glu Thr Arg Leu His Gly Gly Gly Ala 145 150 155 160

Val Arg Leu Asp Val Thr Asp Gly Glu Arg Tyr Leu Gly Val Val Arg 165 170 175

Val Ser Ser Gly Ala Ala Asp Phe Asp Asn Asn Val Phe Val Ser Gly 180 185 190

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Ser Phe Ala Gly Leu Thr Val Ala Gly Thr Glu Tyr Ser Pro Gly Asn 210 215 220

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Asp Leu His Asn Tyr Tyr Arg Tyr Tyr Gly Lys Leu IIe Gly Ser Lys 445 Glu Val Pro IIe Ser Ser Phe Ala Ala Val Trp Lys Gln IIe Val Gln 450 460 Gln Val Val Asn His Pro Ala Val Glu Gly Tyr Gly Leu Met Asn Glu 465 470 475 480 Pro His Ser Thr Asn Gly Leu Trp Pro Gln Ala Ala Leu Ala Ala Ala Gln Ala IIe Arg Thr Val Asp Ser Lys Arg Trp IIe Tyr Val Ala Gly 500 505 510 Asp Arg Trp Ser Ser Ala Phe His Trp Pro His Tyr Asn Thr Gln Leu 515 Val Thr Asn Pro Trp Met Arg Asp Pro Lys Asn Asn Leu Val Tyr Glu Ala His Met Tyr Val Asp Lys Asp Phe Ser Gly Asn Tyr Phe Asp Lys 545 550 560 Ala Glu Lys Phe Asp Pro Met IIe Gly Val Asn Arg Val Lys Pro Phe 565 570 575 Val Asp Trp Leu Lys Gln His Lys Leu Arg Gly Tyr Ile Gly Glu His Gly Val Pro Asp Phe Ser Pro Ser Ala IIe Val Ala Thr Asp Asn Leu Leu Ala Tyr Leu Arg Gln Asn Cys IIe Pro Ser Thr Tyr Trp Ala Ala Gly Pro Trp Trp Gly Glu Tyr Ala Met Ser Leu Asp Val Ser Ser Gly Lys His Arg Pro Gln Leu Pro Val Leu Gln Lys His Ala Lys Thr Ala Asn Ser Cys Thr Ser IIe Gly Pro Leu <210> 19 <212> PRT <213> Arti fi ci al

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Ser Val Ala Phe Ser Ser Ser IIe Ala Ser Ala 20 25

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Trp Thr Ser Met Lys Lys Asp Ala Asn Arg Val Arg Leu Trp Asp Asn 55 60 65

Ala Pro Leu Gly Asn Asp Ser Ala Ser IIe Thr Thr Ser Tyr Arg Gln 70 80 85

Leu Ala Ala Met Ala Leu Ala Tyr Arg Thr Tyr Gly Ser Ser Leu Met 90 95 100

Gly Asp Pro Asp Leu Arg Asp Asp IIe IIe Asp Gly Leu Asp Trp IIe 105 110 115

Asn Thr Phe Gln His Gly Phe Cys Glu Gly Cys Ser Met Tyr Gln Asn 120 125 130 Trp Trp His Trp Gln IIe Gly Gly Pro IIe Ala Leu Asn Glu Val IIe 135 140 145 Ala Leu Met Tyr Asp Glu Leu Thr Gln Thr Gln Ile Asp Ser Tyr Ile Ala Ala IIe Asn Tyr Ala Gln Pro Ser Val Asn Met Thr Gly Ala Asn 170 175 180 Arg Leu Trp Glu Ser Gln Val IIe Ala Leu Ala Gly IIe Asn Gly Lys Asn Gly Asp Lys IIe Ala His Ala Arg Asp Gly Leu Ser Ala Leu Leu 200 205 210 Thr Tyr Val Val Gln Gly Asp Gly Phe Tyr Glu Asp Gly Ser Phe Val 215 220 225 Gln His Ser Tyr Tyr Ser Tyr Asn Gly Gly Tyr Gly Leu Asp Leu Leu 230 235 240 245 Lys Gly IIe Ala Asp Leu Thr Tyr Leu Leu His Asp Ser Asn Trp Glu 250 255 260 Val Val Asp Pro Asn Lys Gln Asn IIe Phe Asn Trp Val Tyr Asp Ser 265 270 275 Phe Glu Pro Phe IIe Tyr Asn Gly Asn Leu Met Asp Met Val Arg Gly 280 285 290 Arg Glu IIe Ser Arg His Ala Arg Gln Ser Asn Val Val Gly Val Glu 295 300 Ala Val Ala Ala IIe Leu Arg Leu Ser His Val Ala Pro Pro Ala Asp Ala Ala Ala Phe Lys Ser Met Val Lys His Trp Leu Gln Glu Gly Gly 330 335 340 Gly Ser Gln Phe Leu Gln Gln Ala Ser IIe Thr His IIe Leu Ser Ala 345 350 355 Gln Asp Val Leu Asn Asp Ser Gly IIe Val Pro Arg Gly Glu Leu Glu 360 365 370 Ala Tyr Arg Gln Phe Ala Gly Met Asp Arg Ala Leu Gln Leu Arg Gln 375 380 385

Gly Tyr Gly Phe Gly IIe Ser Met Phe Ser Ser Arg IIe Gly Gly His 395 Glu Ala IIe Asn Ala Glu Asn Asn Lys Gly Trp His Thr Gly Ala Gly 410 415 420 Met Thr Tyr Leu Tyr Asn Asn Asp Leu Ser Gln Phe Asn Asp His Phe Trp Pro Thr Val Asn Ser Tyr Arg Leu Pro Gly Thr Thr Val Leu Arg 440 445 450 Asp Thr Pro Gln Ala Ala Asn Thr Arg Gly Asp Arg Ser Trp Ala Gly 455 460 465 Gly Thr Asp Met Leu Gly Leu Tyr Gly IIe Thr Gly Met Glu Tyr His 470 480 485 Ala IIe Gly Lys Ser Leu Thr Ala Lys Lys Ser Trp Phe Met Phe Asp 490 495 500 Asp Glu IIe Val Ala Leu Gly Ala Asp IIe Thr Ser Gly Asp Gly Val 505 510 515 Ala Val Glu Thr IIe Val Glu Asn Arg Lys Leu Asn Gly Ala Gly Asp Asn Ser Leu Thr Val Asn Gly Thr Ala Lys Pro Ala Thr Leu Gly Trp 535 540 Ser Glu Thr Met Gly Thr Thr Ser Tyr Ala His Leu Gly Gly Ser Val 550 565 560 Ala Asp Ser Asp IIe Gly Tyr Tyr Phe Pro Asp Gly Gly Ala Thr Leu 570 His Ala Leu Arg Glu Ala Arg Thr Gly Asn Trp Arg Gln Ile Asn Ser Ala Gln Gly Ser Pro Asn Ala Pro His Thr Arg Asn Tyr Leu Thr Met 600 605 610 Trp Leu Glu His Gly Val Asn Pro Ser Asn Gly Ala Tyr Ser Tyr Val Leu Leu Pro Asn Lys Thr Ser Ala Ala Thr Ala Ser Tyr Ala Ala Ser Pro Asp IIe Thr IIe IIe Glu Asn Ser Ser Ser Ala Gln Ala Val Lys 655

Glu Asn Gly Leu Asn Met IIe Gly Val Asn Phe Trp Asn Asn Glu Arg 665 670 675

Lys Thr Ala Gly Gly IIe Thr Ser Asn Ala Lys Ala Ser Val Met Thr 680 685 690

Arg Glu Thr Ala Ser Glu Leu Asn Val Ser Val Ser Asp Pro Thr Gln 695 700 705

Ser Asn Val Gly Met lle Tyr lle Glu lle Asp Lys Ser Ala Thr Gly 710 715 720 725

Leu II e Ala Lys Asp Asp Ala Val Thr Val Leu Gln Tyr Ser Pro Thr 730 740

lle Lys Phe Lys Val Asp Val Asn Lys Ala Arg Gly Lys Ser Phe Lys 745 750 755

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Met Leu Asn Gly Gly Thr Thr Tyr Asn Leu Ser Asp Pro Asp IIe Ala $25 \hspace{1cm} 30 \hspace{1cm} 35$

Ala Arg Val Asn Ala IIe Thr Val Thr Ala Gln Gly Tyr Trp Asp Ser 40 45 50

Met Leu Lys Asp Pro Asn Arg Asn Arg Leu Trp Asn Asp Ala Pro Phe 55 60 65

Gly Ser Asp Ser Thr Ser IIe Thr Thr Thr Tyr Arg His Leu Tyr Asp 70 80 85

Met Ala Leu Ala Tyr Thr Thr Tyr Gly Ser Ser Leu Gln Gly Asn Ala 90 95 100 Ala Leu Lys Ala Asp IIe IIe Ser Gly Leu Asp Trp Met Asn Ala Asn 105 110 115 Gln Phe Tyr Asn Gly Cys Ser Gln Tyr Gln Asn Trp Trp His Trp Gln 120 125 130 lle Gly Gly Pro Met Ala Leu Asn Asp lle Val Ala Leu Met Tyr Thr 135 140 145 Glu Leu Thr Ala Thr Gln IIe Ser Asn Tyr Met Ala Ala IIe Tyr Tyr Thr Gln Ala Ser Val Thr Met Thr Gly Ala Asn Arg Leu Trp Glu Ser 170 175 180 Gln Val IIe Ala IIe Ser Gly IIe Leu Asn Lys Asp Ser Ala Arg Val 185 190 195 Ala Ala Gly Arg Asp Gly IIe Ser Ala Leu Leu Pro Tyr Val Ala Lys 200 205 210 Gly Asp Gly Phe Tyr Asn Asp Gly Ser Phe Val Gln His Thr Tyr Tyr 215 220 225 Ala Tyr Asn Gly Gly Tyr Gly Ser Glu Leu Leu Ser Gly Ile Ala Asp 230 235 240 245 Leu II e Phe II e Leu Asn Gly Ser Ser Trp Gln Val Thr Asp Pro Asn 250~ 255~ 260 260~Lys Asn Asn Val Tyr Arg Trp IIe Tyr Asp Ser Tyr Glu Pro Phe IIe 265 270 275 Tyr Lys Gly Asn Leu Met Asp Met Val Arg Gly Arg Glu IIe Ser Arg His Gly Leu Gln Asp Asp Lys Ala Ala Val Thr Val Met Ala Ser IIe 295 300 305 lle Arg Leu Ser Gln Thr Ala Ala Ser Ala Asp Ala Thr Ala Phe Lys 310 320 325 Arg Met Val Lys Tyr Trp Leu Leu Leu Asp Thr Asp Lys Thr Phe Leu 330 340 Lys Ala Val Ser IIe Asp Leu IIe IIe Ala Ala Asn Gln Leu Val Asn

350

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Thr Ser Ser Ala Val Ala Ser Tyr Ala Ala Thr Pro Gln Ile Ser Ile 630 635 Leu Glu Asn Ser Ser Ser Ala Gln Ala Val Lys Glu Thr Gln Leu Asn Val Thr Gly Ile Asn Phe Trp Asn Asp Glu Pro Thr Thr Val Gly Leu Val Thr Ser Asn Arg Lys Ala Ser Val Met Thr Lys Glu Thr Ala Ser Asp Phe Glu IIe Ser Val Ser Asp Pro Thr Gln Ser Asn Val Gly Thr Ile Tyr Ile Asp Val Asn Lys Ser Ala Thr Gly Leu Ile Ser Lys Asp Asn Glu IIe Thr Val IIe Gln Tyr Tyr Pro Thr Met Lys Phe Lys Val Asn Val Asn Asn Ser Gly Gly Lys Ser Tyr Lys Val Lys Phe Ser Leu 745 750 755 Thr Gly Thr Pro Gly Ser Asn Pro Ser Pro IIe Pro IIe Pro Asn Pro Tyr Glu Ala Glu Ala Leu Pro IIe Asn Ala Leu Thr Asp Thr Pro Val Val Tyr Asn Asp Ala Asn Ala Ser Gly Gly Lys Lys Leu Gly Phe Asn 790 800 805 Asn Asn Ala Val Asp Asp Tyr Val Glu Phe Ser Leu Asp Val Thr Gln 820 Pro Gly Thr Tyr Asp Val Lys Ser Arg IIe Met Lys Ser Thr Asn Ser Gly Ile Tyr Gln Leu Ser Ile Asn Gly Thr Asn Val Gly Ser Ala Gln 840 845 850 Asp Met Phe Trp Thr Thr Ser Glu Leu Ser Lys Glu Phe Thr Met Gly 860 Ser Tyr Ser Phe Ser Thr Pro Gly Ser Tyr Leu Phe Arg Leu Lys Thr Thr Gly Lys Asn Val Ser Ser Ser Gly Tyr Lys Leu Met Leu Asp Asn 890 895 900

Phe Ser Leu Val Ser Thr Gly II e Asp Thr Thr Val II e Val Asp Asn 905 915

Ala Asp Ala Ala Gly Val Thr Lys Val Gly Thr Trp Thr Gly Thr Asn 920 925 930

Thr Gln Thr Asp Arg Tyr Gly Ala Asp Tyr IIe His Asp Gly Asn Thr 935 940 945

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Tyr Ala Thr Met Leu Thr Gly Gly Thr Ala Tyr Ser Leu Ser Asp Pro 25 30 35

Asp IIe Ala Ala Arg Val Ala Ser IIe Thr Thr Asn Ala Gin Thr Leu Trp Thr Ser Met Lys Lys Asp Ala Asn Arg Val Arg Leu Trp Asp Asn 55 60 65 Ala Pro Leu Gly Asn Asp Ser Ala Ser IIe Thr Thr Ser Tyr Arg Gln 70 80 85 Leu Ala Met Ala Leu Ala Tyr Arg Thr Tyr Gly Ser Ser Leu Met 90 95 100 Gly Asp Pro Asp Leu Arg Asp Asp IIe IIe Asp Gly Leu Asp Trp IIe 105 110 115 Asn Thr Phe Gln His Gly Phe Cys Glu Gly Cys Ser Met Tyr Gln Asn 120 125 130 Trp Trp His Trp Gln IIe Gly Gly Pro IIe Ala Leu Asn Glu Val IIe 135 140 145 Ala Leu Met Tyr Asp Glu Leu Thr Gln Thr Gln Ile Asp Ser Tyr Ile 150 160 165 Ala Ala IIe Asn Tyr Ala Gln Pro Ser Val Asn Met Thr Gly Ala Asn 170 175 180 Arg Leu Trp Glu Ser Gln Val IIe Ala Leu Ala Gly IIe Asn Gly Lys 185 190 195 Asn Gly Asp Lys IIe Ala His Ala Arg Asp Gly Leu Ser Ala Leu Leu 200 205 210 Thr Tyr Val Val Gln Gly Asp Gly Phe Tyr Glu Asp Gly Ser Phe Val 215 220 225 Gln His Ser Tyr Tyr Ser Tyr Asn Gly Gly Tyr Gly Leu Asp Leu Leu 230 245 240 230 Lys Gly IIe Ala Asp Leu Thr Tyr Leu Leu His Asp Ser Asn Trp Glu 250 255 260 Val Val Asp Pro Asn Lys Gln Asn IIe Phe Asn Trp Val Tyr Asp Ser 265 270 275 Phe Glu Pro Phe IIe Tyr Asn Gly Asn Leu Met Asp Met Val Arg Gly 280 285 290 Arg Glu IIe Ser Arg His Ala Arg Gln Ser Asn Val Val Gly Val Glu 295 300 305

Ala Val Ala Ala IIe Leu Arg Leu Ser His Val Ala Pro Pro Ala Asp Ala Ala Ala Phe Lys Ser Met Val Lys His Trp Leu Gln Glu Gly Gly 330 335 340 Gly Ser Gln Phe Leu Gln Gln Ala Ser IIe Thr His IIe Leu Ser Ala 345 350 355 Gln Asp Val Leu Asn Asp Ser Gly IIe Val Pro Arg Gly Glu Leu Glu 360 365 370 Ala Tyr Arg Gln Phe Ala Gly Met Asp Arg Ala Leu Gln Leu Arg Gln 375 380 385 Gly Tyr Gly Phe Gly IIe Ser Met Phe Ser Ser Arg IIe Gly Gly His 390 400 405 Glu Ala IIe Asn Ala Glu Asn Asn Lys Gly Trp His Thr Gly Ala Gly 410 415 420 Met Thr Tyr Leu Tyr Asn Asn Asp Leu Ser Gln Phe Asn Asp His Phe Trp Pro Thr Val Asn Ser Tyr Arg Leu Pro Gly Thr Thr Val Leu Arg 440 445 450 Asp Thr Pro Gln Ala Ala Asn Thr Arg Gly Asp Arg Ser Trp Ala Gly 455 460 465 Gly Thr Asp Met Leu Gly Leu Tyr Gly IIe Thr Gly Met Glu Tyr His 470 475 480 $$ 485 Ala IIe Gly Lys Ser Leu Thr Ala Lys Lys Ser Trp Phe Met Phe Asp 490 495 500 Asp Glu IIe Val Ala Leu Gly Ala Asp IIe Thr Ser Gly Asp Gly Val 505 510 515 Ala Val Glu Thr IIe Val Glu Asn Arg Lys Leu Asn Gly Ala Gly Asp 520 525 530 Asn Ser Leu Thr Val Asn Gly Thr Ala Lys Pro Ala Thr Leu Gly Trp 535 540 545 Ser Glu Thr Met Gly Thr Thr Ser Tyr Ala His Leu Gly Gly Ser Val Ala Asp Ser Asp IIe Gly Tyr Tyr Phe Pro Asp Gly Gly Ala Thr Leu 570 575 580

His Ala Leu Arg Glu Ala Arg Thr Gly Asn Trp Arg Gln IIe Asn Ser 585 590 595 Ala Gln Gly Ser Pro Asn Ala Pro His Thr Arg Asn Tyr Leu Thr Met 600 605 610 Trp Leu Glu His Gly Val Asn Pro Ser Asn Gly Ala Tyr Ser Tyr Val Leu Leu Pro Asn Lys Thr Ser Ala Ala Thr Ala Ser Tyr Ala Ala Ser Pro Aspile Thrile IIe Glu Asn Ser Ser Ala Gln Ala Val Lys 655 Glu Asn Gly Leu Asn Met IIe Gly Val Asn Phe Trp Asn Asn Glu Arg Lys Thr Ala Gly Gly IIe Thr Ser Asn Ala Lys Ala Ser Val Met Thr Arg Glu Thr Ala Ser Glu Leu Asn Val Ser Val Ser Asp Pro Thr Gln Ser Asn Val Gly Met IIe Tyr IIe Glu IIe Asp Lys Ser Ala Thr Gly 710 715 720 725 Leu II e Ala Lys Asp Asp Ala Val Thr Val Leu Gln Tyr Ser Pro Thr 730 740 lle Lys Phe Lys Val Asp Val Asn Lys Ala Arg Gly Lys Ser Phe Lys
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Val Gly Asp Pro Val Asp Met Tyr Trp Asn Thr Ser Glu Ser Thr Lys 855 860 865

Ser Phe Ser Pro Gly Ser Tyr Thr Phe Ser Glu Pro Gly Ser Tyr Leu 870 885

Leu Arg Val Thr Val Thr Gly Lys His Pro Ser Ser Ser Gly Tyr Lys 890 895 900

Leu Met Leu Asp His Phe Thr Leu Glu Glu IIe Pro Val Ser Leu Pro 905 910 915

Asn Pro Tyr Glu Ala Glu Thr Leu Pro IIe His His Arg Thr Gln Thr 920 925 930

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Leu Asn His Lys Val Val Gly Asp Tyr Thr Glu Phe IIe Leu Asp Val 950 955 960 965

Pro Gln Ala Gly Thr Tyr Asp IIe Thr Ala Arg Val Leu Lys Phe Ser 970 975 980

Asp Asn Gly IIe Tyr Gln Phe Ser IIe Asp Gly Asn Pro Val Gly Ala 985 990 995

Pro IIe Asp Thr Tyr Trp Asn Thr Ala Gly Tyr IIe Arg Asp Phe 1000 1005 1010

Thr Pro Gly Ser Tyr Thr Phe Ser Glu Pro Gly Ser Tyr Leu Leu 1015 1020 1025

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Thr Ala Glu Ala Ala Glu Gly Glu Asn IIe Glu Asp Lys Met Val Ser $25 \hspace{1.5cm} 30 \hspace{1.5cm} 35$

Ala Tyr Asn Met Asp Ala Phe Asp IIe Met Arg Glu Val Arg Arg Thr 40 45 50

Met Leu Thr Gly Gly Ala Ala Leu Asn Pro Ala Asp Pro Asp Ala Ala 55 60 65

Ala Ala Val Ala Ala Leu Ala Ser Glu Ala Asn Gln Tyr Trp Gln Thr 70 75 80 85

Met Asp Asp Ser Pro Gly Arg Thr Ser Leu Trp Ser Asp Asn Pro Gly 90 95 100

Thr Gly Asn Ser IIe His IIe Arg IIe Thr Tyr Glu Arg Leu Lys Thr 105 110 115

Met Ala Leu Ala Tyr Ala Ala Ala Gly Ser Pro Leu His Ser Asn Ala 120 125 130

Ser Leu Glu Ala Asp IIe Val Asp Ala Leu Asp Tyr Met Tyr Ala Thr 135 140 145

Arg Tyr His Glu Asn Val Thr Thr Thr Pro Ser Gly Thr Ser Asn Trp 150 160 165

Trp Asp Trp Gln IIe Gly IIe Pro Met Gln Leu Asn Asp Thr Val Val 170 175 180

Leu Met Tyr Asp Ser Leu Thr Pro Ala Gln IIe Ala Asn Tyr Met Asn 185 190 195

Ala Val Glu Arg Phe Thr Pro Thr Val Asn Leu Thr Gly Ala Asn Arg 200 205 210

Ser Trp Lys Ala IIe Val Val Ala Val Arg Gly IIe Leu Val Lys Asp 215 220 225

Gly Ala Lys IIe Ala Ala Ala Arg Asp Gly Leu Ser Gln IIe Phe Asn 230 240 245

Tyr Ala Val Ser Gly Asp Gly Phe Tyr Arg Asp Gly Ser Phe Ile Gln 250 255 260

His Gly Asn IIe Pro Tyr Asn Gly Gly Tyr Gly Leu Asp Leu Leu Leu 265 270 275 Ser Asp Leu Met Thr Leu Leu His Gly Ser Ala Trp Gln Val Ala Val Thr Asp Pro Asn Gln Ala Asn Val Trp Glu Trp Val Tyr Arg Ala Tyr Gln Pro Leu IIe Tyr Lys Gly Ala Met Met Asp Met Val Arg Gly Arg 310 325 320 325 Glu lle Ser Arg Val Tyr Arg Gln Asp His Ala Ala Gly His lle Ala 335 Met Gln Gly IIe Leu Arg Leu Ser Ala Val Ala Pro Pro Ala Gln Ala Glu Asp Phe Lys Arg Met Val Lys Gly Trp Met Val Val Asp Gly Phe $360 \hspace{1cm} 365 \hspace{1cm} 370 \hspace{1cm}$ Met Arg Phe Tyr Glu Gln Ala Pro Leu Gly Leu IIe Pro Leu Ala Lys 375 380 385 Ala Val Glu Gly Asp Ala Ser IIe Ala Pro Ala Ser Glu Leu IIe Gln Tyr Arg Gln Tyr Ala Ala Met Asp Arg Ala Val Gln Leu Arg Pro Gly Tyr Gly Phe Gly Leu Ala Met Tyr Ser Ser Arg IIe Gly Ser Phe Glu 425 435 Ala IIe Asn Ser Glu Asn Leu Arg Gly Trp Tyr Thr Ser Ala Gly Met
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lle Val Ala Leu Gly Ala Gly IIe Ser Ser Ala Asp Gly IIe Pro Val 535 545 Glu Thr IIe IIe Glu Asn Arg Arg IIe Gly Gly Ala Gly Asp Asn Ala 550 565 Phe Leu Ala Asp Gly Ala Ala Met Pro Ala Glu Leu Gly Trp Ser Gly 570 575 580 Thr Leu Glu Gly Val Arg Trp Ala His Leu Thr Gly Thr Ala Ala Gly 585 590 595 Ala Asp Ile Gly Tyr Tyr Phe Pro Glu Pro Ala Ala Val His Ala Val 605 Arg Glu Ala Arg Thr Gly Asn Trp Arg Gln Ile Asn Asn Arg Pro Val Thr Pro Ala Ala Ser Val Thr Arg Asn Tyr Leu Thr Phe Trp Phe Asp His Gly Ala Asn Pro Thr Asn Ala Asp Tyr Gln Tyr Val Leu Leu Pro 650 660 Asn Lys Ser Gly Ala Gln Val Ala Gly Tyr Ala Ala Asn Pro Asp Val 665 670 675 Glu Val Leu Ala Asn Ser Pro Glu Val Gln Ala Val Lys Glu Ser Ser 680 685 690 Leu Gly IIe IIe Gly Ala Asn Phe Trp Ser Asp Gly Val Arg Thr Val 695 705 Asp Leu IIe Thr Val Asn Lys Lys Ala Ser Val Met Thr Arg Glu Thr Pro Gly Ala IIe Leu Asp Leu Ser Val Ser Asp Pro Thr Gln Val Asn Ala Gly Thr Ile Glu Ile Glu Leu Asn Arg Ala Ala Ser Gly Phe Thr 745 750 755 Ala Asp Pro Gly Val Thr Val Thr Arg Leu Ser Pro Thr Ile Lys Leu Thr Val Gln Val Ala Gly Ala Lys Gly Arg Ser Phe Lys Ala Ser Phe 775 780 785 Glu Leu Gly Glu Ala Ser Gly Pro Gly Pro Asp Pro Gly Pro 790 800 805

Ser Glu IIe IIe Val Asp Asn Gly Asp Ala Ala Gly Val Thr Lys IIe 810 815 820

Gly Ser Trp Lys Thr Gly Thr Val Gln Thr Asp Arg Tyr Gly Pro Asp 825 830 835

Tyr Leu His Asp Asp Asn Thr Gly Lys Gly Gly Lys Ser Val Arg Phe 840 845

Thr Pro Asp Leu Pro Thr Ala Gly Thr Tyr Asp Val Tyr Met Met Trp 855 860 865

Pro Gln His Phe Asn Arg Ala Thr Asn IIe Pro Val Thr IIe Ala His 870 880 885

Ala Gly Gly Thr Ala Thr Val Thr IIe Asp Gln Thr Val Ser Gly Gly 890 895 900

Val Trp Asn Tyr Leu Gly Ser Tyr Ser Phe Asp Thr Gly Ser Gly Gly 915

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Glu Ala Ser Asp Met Phe Asp Glu Leu Arg Glu Lys Tyr Ala Thr Met 35 40 45

Leu Thr Gly Gly Thr Ala Tyr Ser Leu Ser Asp Pro Asp Ile Ala Ala 50 60

Arg Val Ala Ser IIe Thr Thr Asn Ala Gln Thr Leu Trp Thr Ser Met 65 70 75 80

Lys Lys Asp Ala Asn Arg Val Arg Leu Trp Asp Asn Ala Pro Leu Gly Page 66

Asn Asp Ser Ala Ser IIe Thr Thr Ser Tyr Arg Gln Leu Ala Ala Met Ala Leu Ala Tyr Arg Thr Tyr Gly Ser Ser Leu Met Gly Asp Pro Asp 115 120 125 Leu Arg Asp Asp IIe IIe Asp Gly Leu Asp Trp IIe Asn Thr Phe Gln
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Lys Ser Met Val Lys His Trp Leu Gln Glu Gly Gly Ser Gln Phe

Page 67

355 360 365

Leu Gln Gln Ala Ser IIe Thr His IIe Leu Ser Ala Gln Asp Val Leu 370 375 380

Asn Asp Ser Gly IIe Val Pro Arg Gly Glu Leu Glu Ala Tyr Arg Gln 385 390 395 400

Phe Ala Gly Met Asp Arg Ala Leu Gln Leu Arg Gln Gly Tyr Gly Phe 405 410 415

Gly Ile Ser Met Phe Ser Ser Arg Ile Gly Gly His Glu Ala Ile Asn 420 425 430

Ala Glu Asn Asn Lys Gly Trp His Thr Gly Ala Gly Met Thr Tyr Leu 435 440 445

Tyr Asn Asn Asp Leu Ser Gln Phe Asn Asp His Phe Trp Pro Thr Val 450 455 460

Asn Ser Tyr Arg Leu Pro Gly Thr Thr Val Leu Arg Asp Thr Pro Gln 465 470 475 480

Ala Ala Asn Thr Arg Gly Asp Arg Ser Trp Ala Gly Gly Thr Asp Met 485 490 495

Leu Gly Leu Tyr Gly Ile Thr Gly Met Glu Tyr His Ala Ile Gly Lys 500 505 510

Ser Leu Thr Ala Lys Lys Ser Trp Phe Met Phe Asp Asp Glu IIe Val 515 520 525

Ala Leu Gly Ala Asp IIe Thr Ser Gly Asp Gly Val Ala Val Glu Thr 530 535 540

Val Asn Gly Thr Ala Lys Pro Ala Thr Leu Gly Trp Ser Glu Thr Met 565 570 575

Gly Thr Thr Ser Tyr Ala His Leu Gly Gly Ser Val Ala Asp Ser Asp 580 585 590

Ile Gly Tyr Tyr Phe Pro Asp Gly Gly Ala Thr Leu His Ala Leu Arg 595 600 605

Glu Ala Arg Thr Gly Asn Trp Arg Gln IIe Asn Ser Ala Gln Gly Ser 610 620

Pro Asn Ala Pro His Thr Arg Asn Tyr Leu Thr Met Trp Leu Glu His Page 68 Gly Val Asn Pro Ser Asn Gly Ala Tyr Ser Tyr Val Leu Leu Pro Asn 645 650 655

630

Lys Thr Ser Ala Ala Thr Ala Ser Tyr Ala Ala Ser Pro Asp IIe Thr 660 670

Ile Ile Glu Asn Ser Ser Ser Ala Gln Ala Val Lys Glu Asn Gly Leu 675 680 685

Asn Met IIe Gly Val Asn Phe Trp Asn Asn Glu Arg Lys Thr Ala Gly 690 695 700

Gly lle Thr Ser Asn Ala Lys Ala Ser Val Met Thr Arg Glu Thr Ala 705 710 715 720

Ser Glu Leu Asn Val Ser Val Ser Asp Pro Thr Gln Ser Asn Val Gly 725 730 735

Met IIe Tyr IIe Glu IIe Asp Lys Ser Ala Thr Gly Leu IIe Ala Lys 740 745 750

Asp Asp Ala Val Thr Val Leu Gln Tyr Ser Pro Thr IIe Lys Phe Lys 755 760 765

Val Asp Val Asn Lys Ala Arg Gly Lys Ser Phe Lys Ala Ala Phe Ser 770 780

Leu Thr Gly Ala Gln Gln Pro Asn Pro Ala Pro IIe Pro IIe Pro Asn 785 790 795 800

Pro Tyr Glu Ala Glu Leu Leu Pro IIe Ser Ala Thr Thr Lys Thr Pro 805 810 815

Thr Leu Ser Asn Asp Ser Asn Ala Ser Gly Gly Lys Lys Leu Gly Leu 820 825 830

Asn Ser Ser Val Val Gly Asp Tyr Thr Glu Phe Ser Leu Asp Val Thr 835 840 845

Gln Pro Gly Thr Tyr Asp IIe Ala Ala Lys IIe Met Lys Val Ser Asn 850 855 860

Asn Gly IIe Tyr Gln Phe Ser IIe Asn Gly Glu Pro Val Gly Asp Pro 865 870 875 880

Val Asp Met Tyr Trp Asn Thr Ser Glu Ser Thr Lys Ser Phe Ser Pro 885 890 895

Gly Ser Tyr Thr Phe Ser Glu Pro Gly Ser Tyr Leu Leu Arg Val Thr Page 69

900

910

Val Thr Gly Lys His Pro Ser Ser Ser Gly Tyr Lys Leu Met Leu Asp 915 920 925

His Phe Thr Leu Glu Glu IIe Pro Val Ser Leu Pro Asn Pro Tyr Glu 930 935 940

Ala Glu Thr Leu Pro IIe His His Arg Thr Gln Thr Val Thr IIe Tyr 945 950 955 960

Asn Asp Ser Asn Thr Ser Gly Gly Gln Arg Leu Gly Leu Asn His Lys $965 \hspace{1.5cm} 970 \hspace{1.5cm} 975$

Val Val Gly Asp Tyr Thr Glu Phe IIe Leu Asp Val Pro Gln Ala Gly 980 985 990

Thr Tyr Asp IIe Thr Ala Arg Val Leu Lys Phe Ser Asp Asn Gly IIe 995

Tyr Gln Phe Ser IIe Asp Gly Asn Pro Val Gly Ala Pro IIe Asp 1010 1015 1020

Thr Tyr Trp Asn Thr Ala Gly Tyr IIe Arg Asp Phe Thr Pro Gly 1025 1030 1035

Ser Tyr Thr Phe Ser Glu Pro Gly Ser Tyr Leu Leu Arg Leu Thr 1040 1045 1050

Ala Thr Gly Lys Asn Pro Ser Ala Ser Gly Leu Lys IIe Met Leu 1055 1060 1065

Asp Tyr IIe Trp Leu Asp 1070