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(54) **CYCLIC GMP-BINDING, CYCLIC GMP-SPECIFIC PHOSPHODIESTERASE MATERIALS AND METHODS**

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Publication Classification

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(57) **ABSTRACT**

The present invention provides novel purified and isolated nucleotide sequences encoding the cGMP-binding, cGMP-specific phosphodiesterase designated cGB-PDE. Also provided by the invention are methods and materials for the recombinant production of cGB-PDE polypeptide products and methods for identifying compounds which modulate the enzymatic activity of cGB-PDE polypeptides.

CGB-PDE	FQMKHEVLCK	WILSVKKNYR	K.NVAYHNWR	HAFNTAOCMF	AALKAGKIQK	626
ROS- α	FHIPOEALVR	FMYSLSKGYR	R..ITYHNWR	HGFNVGQTMF	SLLVTGKLRK	582
ROS- β	FQIPOEVLVR	FLFSVSKGYR	R..ITYHNWR	HGFNVAQTMF	TLLMTGKLS	580
CONE- α '	FKVPVEVLTR	WNTYVRKGYR	A..VTYHNWR	HGFNVGQTMF	TLLMTGRLKK	580
CGS	YKIDCPTLAR	FCLMVKKGYR	D.P.PYHNWM	HAFSVSHFCY	LLYKNLELTN	639
61 KCAM	FKIPVSCLIA	FAEALVGYG	KYKNPYHNLI	HAADVTQTVH	YIMLHTGIMH	242
63 KCAM	FKIPTVFLMT	FLDALETGYG	KYKNPYHNQI	HAADVTQTVH	CFLLRTGMVH	244
RATDUNCE	FOIPADTLLR	YLLTLEGHYH	S.NVAYHNSI	HAADVVSASH	VLLGTPALEA	125
DROSDUNCE	.MIPPKTFLN	FMSTLEDHYV	K.DNPFHNSL	HAADVTOSTN	VLLNTPALEG	48
CONSERVED	-----*	-----Y-	-----HN--*	H-----**	-----*	

CGB-PDE	RLTDLEILAL	LIAALSHDLD	HRGVNNSYIQ	RSEHPLAQLY	CH..SIMEHH	674
ROS- α	YFTDLEALAM	VTAAFCHDID	HRGTNNLYQM	KSQNPLAKLH	GS..SILERH	630
ROS- β	YYTDLEAFAM	VTAGLCHDID	HRGTNNLYQM	KSQNPLAKLH	GS..SILERH	628
CONE- α '	YYTDLEAFAM	LAAAFCHDID	HRGTNNLYQM	KSTSPLARLH	GS..SILERH	628
CGS	YLEDMEIFAL	FISCMCHDLD	HRGTNNSFQV	ASKSVLAALY	SSEGSVMERH	709
61 KCAM	WLTELEILAM	VFAAAIHDYE	HIGTTNFI	QTRSDVAILY	.NDRSVLENH	291
63 KCAM	CLSEIEVLAI	IFAAAIHDYE	HIGTTNSFHI	QTKSEQAILY	.NDRSVLENH	293
RATDUNCE	VFTDLEVLA	IFACAIHDVD	HPGVSNQFLI	NTNSELALMY	.NDSSVLENH	174
DROSDUNCE	VFTPLEVGA	LFAACIHDVD	HPGLTNQFLV	NSSSELALMY	.NDESVLENH	97
CONSERVED	-----E	-----HD--	H-G--N--*	-----*-A	-----S--E-H	

FIGURE 1A

CGB-PDE
 ROS- α
 ROS- β
 CONE- α
 CGS
 61 KCAM
 63 KCAN
 RATDUNCE
 DROSDUNCE
 CONSERVED

HFDXICLHILN SP6NOILSGL
 HLEFGKTLRL DESLNIFONL
 HLEFGKFLLS EETLNLYONL
 HLEYSKTLLO DESLHIFONL
 HFAQATAILN THGCHIFDHF
 HVSAAYRLMQ EEEMVLLNL
 HISSVFRHMQ DDEHNIIFINL
 HLAVGFKLLQ GENCDIFONL
 HLAVAFKLLQ HOGCDIFCNH
 H-----

SIEEYKTTLK
 HRRQHEHAIH
 HRRQHEIVIH
 NKROYETVHH
 SRKDYORMLD
 SKDDWRDLRH
 TKDEFVELRA
 STKOKLSLRR
 QKKOROTLRK

IIKOAILATD
 MNDIAIATD
 LMDIAIATD
 LFEVAIATD
 LMRDIIATD
 LVIEKVLSTD
 LVIEKVLATD
 HVIDNVLATD
 HVIDIVLSTD
 -----T)

LALYIKRRGE
 LALYCKKRTM
 LALYFKKRTK
 LALYFKKRTM
 LAHWLRIFKD
 HSGHFOQIKH
 HSCHFQOVKS
 HSKHMSLLAD
 HSKHMSLLAD
 ---***--*--

724
680
678
678
748
326
328
220
143

CGB-PDE
 ROS- β
 ROS- β
 CONE- α
 CGS
 61 KCAM
 63 KCAN
 RATDUNCE
 DROSDUNCE
 CONSERVED

FFELIMKH..
 FOKIVDLSKT
 FOKIVDESKH
 FOKIVDACEK
 LOKMAE.....
 IRRHSLOOPEG
 HKTALQQLER
 LKTHVETKKV
 LKTHVETKKV

HLEDPIKEL
 HMLDQTRKEI
 LSLETRKEI
 VTIDPTKKEI
 DRTHKQHSLS
DKAK
DKSK
 LLDNYSDRID
 LLDNYDRID

FLAMLMTACD
 VMAMMTACD
 VMAMMTACD
 IMAMMTACD
 LLCLLMTSCD
 THSLILHAAD
 ALSLLHAAD
 VLOSLYHCAD
 VLENLVHCAD

LSAITKPWPI
 LSAITKPWEV
 LSAITKPWEV
 LSAITKPWEV
 LSDQTKGWKT
 ISHPAKSWKL
 ISHPAKSWSV
 LSNPAKPLPL
 LSNPTKPLPL
 -S*-K-

.....OF
 YETQDQNTQY
 YEDRKSWEY
 METEEAICY
VGY
 L.....
 I.....
 T.....SLGVL
 A.....GSGVL

.....D
 ---***--D

764
730
728
728
798
376
378
270
193

FIGURE 1B

CGB - PDE	QQRIAELVAT	EFFDQGDRLER	KELNIEPADL	MNREKKNKIP	SMQVGFID...	812
ROS - α	QSKVALLVAA	EFWEOGDLER	TVLQONPIPM	MDRNKADEL P	KLOVGFID..	778
ROS - β	QSKVALLVAA	EFWEOGDLER	TVLDOQPIPM	MDRNKAAEL P	KLOVGFID..	776
CONE - α	QSQVALLVAN	EFWEOGDLER	TVLQOQPIPM	MDRNKKOEL P	KLOVGFID..	776
C6S	TRKIAELIYK	EFFSQGDLEK	A.MGNRPHEM	MDREKAY.IP	ELQISFME..	844
61 KCAM	HHRWTMALME	EFFLOGDKEA	EL..GLPFSP	LCDRKSTMVA	QSQIGFID..	422
63 KCAM	HSRNTKALME	EFFRQGDKEA	EL..GLPFSP	LCDRTSTLVA	QSQIGFID..	424
RATDUNCE	YRQWTERIMA	EFFQOQDRER	ES..GLDISP	MCDKHTASVE	KSQVGFID..	316
DROSDUNCE	YKRWVALLME	EFFLOGDKER	ES..GMDISP	MCDRHNATIE	KSQVGFID..	239
CONSERVED	*-----*-	EF--QGD-E-	-----	-----	--Q--F----	

FIGURE 1C

CGB-PDE	LLELVKDISS	HLDVTALCHK	I FLHIGHLIS	ADRYSLFLVC	EDSSNDKFLI	188
CGS	ILQLCGELYD	.LDASSLQLK	VLOYLQGETQ	ASRCCLLLV	EDN..LQ.LS	245
CONE- α '	LLEVL..LEE	AGSVELAAHR	ALORLAQLLQ	ADRCMFLCR	ARNGTPE.VA	106
ROS- β	LFELVQDMQE	NVNMERVVFK	ILRRLCSILH	ADRCSLFMYR	QRNGVAE.LA	107
ROS- α	...LLRDFOD	NLQAEKCVFN	VMKKLCFLLO	ADRMSLFMYR	ARNGIAE.LA	109
CONSERVED	-----	-----	-----	A-R-----	-----	
CGB-PDE	SRLFDAEGS	TLEE...ASN	NCIRLEMKG	IVGHVAAFGE	PLNIKDAYED	237
CGS	CKVIG...DK	VLEE.....	.EISFPLTTG	RLGQVVEDKK	SIQLKDLTSE	292
CONE- α '	SKLLDVTPTS	KFEDNLVVPD	REAVFPLDVG	IVGWVAHTKK	TFNVDPDVKKN	154
ROS- β	TRLFSVOPDS	VLEDCLVPPD	SEIVFPLDIG	VVGHVAQTKK	MVNVQDVMK	155
ROS- α	TRLFNVHKDA	VLEECLVAPD	SEIVFPLDMG	VVGHVALSKK	IYNVPTTEED	157
CONSERVED	-----	E-----	-----	--G-V-----	-----	
CGB-PDE	PRFNAEVDQI	TGYKTSILC	MPIKNHR.EE	VVGVAQAINK	KSGNGGTFTTE	287
CGS	DM..OQLQSM	LGCEVOAMLC	VPVISRATDQ	VVALACAFNK	..LGGDLFTD	342
CONE- α '	SHFSDFMDKQ	TGYVTRNLLA	TPIV..MGKE	VLAVFMVAVNK	..VDASEFSK	204
ROS- β	PHFSSFADDEL	TDYVTRNILA	TPIM..NGKD	VVAVIMAVNK	..LDGPCFCTS	205
ROS- α	EHFCDFVDTL	TEYQTKNILA	SPIH..NGKD	VVAIIMAVNK	..VDGPHFTE	207
CONSERVED	-----	-----	-P-----	V-----	A-NK-----	F--

FIGURE 2A

CGB-PDE	KDEKFAAYL	AFCGIVLHMA	QLYETSLEN	KRNQVLLDLA	SLIFEEQOQL	337
CGS	ODEHVIOHCF	HYTSTVLTST	LAFQKEOKLK	CECOALLOVA	KHLFTHLDDV	390
CONE- α '	ODEEVFSKYL	SFYSIILKLH	HTNYLYMIES	RRSQILMWSA	NKVFEELTDV	252
ROS- β	EDEDVFLKYL	NFGTLNLKIY	HYSYLHNCET	RRGOVLLWSA	NKVFEELTDI	253
ROS- α	NDEEILLKYL	HFANLIMKVF	HLSYLHNCET	RRGQILLMSG	SKVFEELTOI	255
CONSERVED	-DE-----	-----	-----	---Q-L---	---F----	
CGB-PDE	EVILKKIAAT	IISFMQVQKC	TIFIVD.EDC	SDFSSEVFMH	ECEELEKSSD	361
CGS	SVLLDEIITE	ARNLSNAEIC	SVFLID...Q	NELVAKVFDG	GVLEDESY..	409
CONE- α '	ERQFHKALYT	VRTYLNLCERY	SIGLLDNTKE	KEYF.DEWVPV	KPGEVEPEYKG	301
ROS- β	ERQFHKAFYT	VRAYLNCDRY	SVGLDNTKE	KEFF.DYWPV	LMGEAQAYS	302
ROS- α	ERQFHKALYT	VRAFLNCDRY	SVGLDNTKQ	KEFF.DVWPV	LMGEAPPYAG	304
CONSERVED	-----	-----	-----D-----	-----	---E----	
CGB-PDE	TLTRE.....RDANRINY	MYAQYVKNTM	411
CGSEIRI...	.PADQ.....	GIAGHVATTG	459
CONE- α '	PKTPDGREVI	FYKIIDYILH	GKEEIKVIPT	PPMDHMTLIS	GLPTYVAENG	351
ROS- β	PRTPDGREIL	FYKVIDYILH	GKEDIKVIPI	PPADHWALAS	GLPTYVAESG	352
ROS- α	PRTPDGREIN	FYKVIDYILH	GKEDIKVIPI	PPPDHWALYS	GLPTYVAOENG	354
CONSERVED	-----	-----	-----	---D-----	---Y----	

FIGURE 2B

CGB-PDE	EPLNIPDVSK	DKRFPWTNEN	MGNINQQCIR	SLLCTPIKNG	KKNKVIGVCCQ	459
CGS	QILNIPDAYA	HPLFY..RGV	DDSTGRF.TR	NILCFPIKN.	ENQEVIGVAE	499
CONE- α '	FICNMLNAPA	DEYFTFOKGP	VDETGWV.IK	NVLSLPIVN.	KKEDIVGVAT	399
ROS- β	FICNIMNAPA	DEMFNFOEGP	LDDSGWI.VK	NVLSMPIVN.	KKEEIVGVAT	400
ROS- α	LICNIMNAPS	EDFFAFOKEP	LDES6WM.IK	NVLSMPIVN.	KKEEIVGVAT	402
CONSERVED	---N-----	---F-----	-----	--L--PI-N-	-----GV--	
CGB-PDE	LVNKMEETTG	KVKAFHRNDE	QFLEAFVIFC	GLGIONTOMY	EAVRAMAKQ	506
CGS	LVNKING...	..PWFSKFDE	DLATAFSIYC	GISIAHSLLY	KKVNEAQYRS	541
CONE- α '	FYNRKDG...	..KPFDEYDE	HIAETLTQFL	GWSLLNTDITY	EKNKLENRKR	441
ROS- β	FYNRKDG...	..KPFDEQDE	VLMESLTQFL	GWSVLNTDITY	DKKNKLENRKR	442
ROS- α	FYNRKDG...	..KPFDEMD	TLMESLAQFL	GWSVLNPDITY	ELMNKLENRKR	444
CONSERVED	--N-----	----F----DE	-----	G-----Y	-----	
CGB-PDE	MVTLEVLSYH	ASAAEEE				526
CGS	HLANEMMYH	MKVSDDE				561
CONE- α '	DIAQEMLMNH	TKATPDE				461
ROS- β	DIAQDMVLYH	VRCDRER				462
ROS- α	DIFQDMVKYH	VKCDNEE				464
CONSERVED	-----H	-----E				

FIGURE 2C

CGB-PDE
 ROS- α
 ROS- β
 CONE- α '
 CGS
 CGB-PDE
 ROS- α
 ROS- β
 CONE- α '
 CGS
 CONSERVED

A
 A
 A
 A
 A
 B
 B
 B
 B
 B

EPLNIKDAYEDPRF...NAEVDQITGYKTSILCMPKMH.REEVVGVAQAIN.KKSGN
 KIVHVPNTEDEHF...CDFVDTLTEYQTKNILASPIHNG.K.DVVAIIMAVN.KVDGP
 KMVNVDVMECPHF...SSFADDELTDYVTRNIIATPIHNG.K.DVVAIIMAVN.KLDGP
 XTFNVDPVKKNSHF...SDFMOKDTGYVTRNIIATPIVNG.K.EVLAVFMVAVN.KVDAS
 KSIQLKDLTSEDM...QQLQSHLGCVEOAMLCVVISRATDQVVALACAFN.KLGGD
 EPLNIPDVSQDKRFPWTNENMGNIQCIRSLCTPIKNGKKNKVIQVQQLVN.KMHEET
 LICNIMNAPSEDFFAFOKEPDE.SGMIKNVLSPIVNK.KEEIVGVATFYNRKD&KP
 FICNIMNAPADEMFNFOEGPLDD.SGWIVKNVLSMPIVNK.KEEIVGVATFYNRKD&KP
 FICNMLNAPADEYFTFOKGPVDE.TGWIVKNVLSLPIVNK.KEDIYGVATFYNRKD&KP
 QILNIPDAYAHPLF...YRGVDDSTGFRTRNIIICFPPIKNE.HQEVIGVAELVN.KINGP
 ---*---L---P*---*****N-K---

CGB-PDE
 ROS- α
 ROS- β
 CONE- α '
 CGS
 CGB-PDE
 ROS- α
 ROS- β
 CONE- α '
 CGS
 CONSERVED

A
 A
 A
 A
 A
 B
 B
 B
 B
 B

GG...TFTEKDEKFAAYLAFCGIVLHMAQL.YE
 ...HFTENDEEILLKYLNFANLIMKVFHLSY.
 ...CFTSEDEDFLKYLNFGTLNLKIYHLSY.
 ...EFSKQDEEVFSKYLSFVSIILKLHHTNY.
 ...LFTDODEHVIOHCCHYTSTVL.TSTLAFQ
 TGKYKAFNRNDEQFLFAFYFCGLGIONTQM.YE
 ...FDEMDELMEADFLGWSV.LNPDITYE
 ...FVEODEVLMESLTOFLGWSV.LNTDTYD
 ...FDEYDEHIAETLQFLGWSL.LNTDTYE
 ...WFSKFDLATAFSIYCGISI.AHSLLYK
 ---F---DE---*---

FIGURE 3

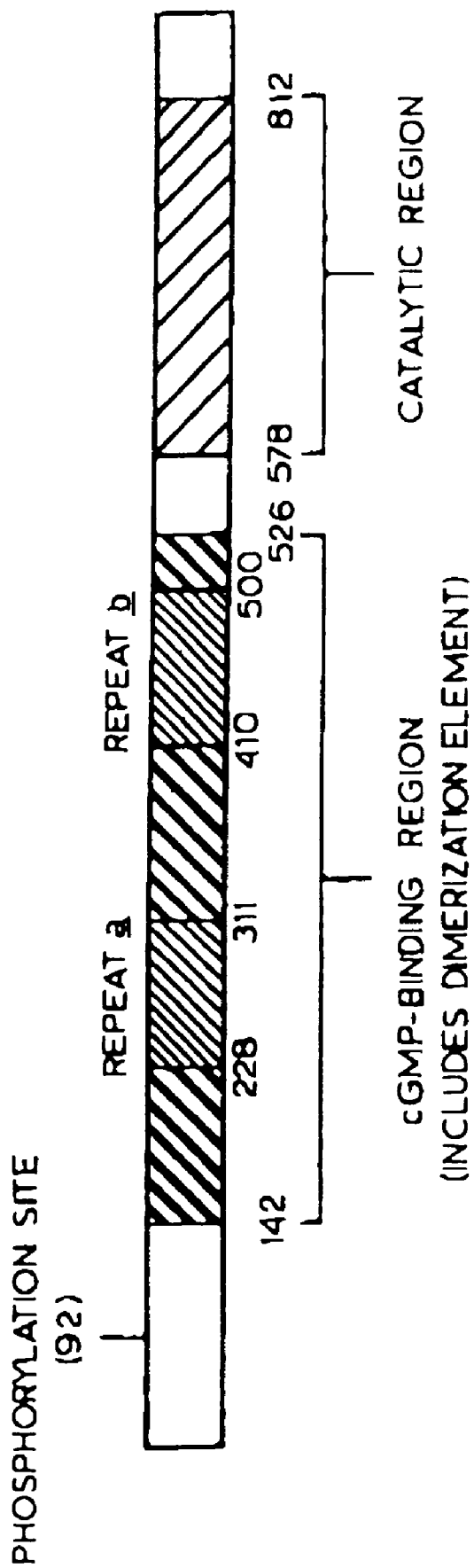


FIGURE 4

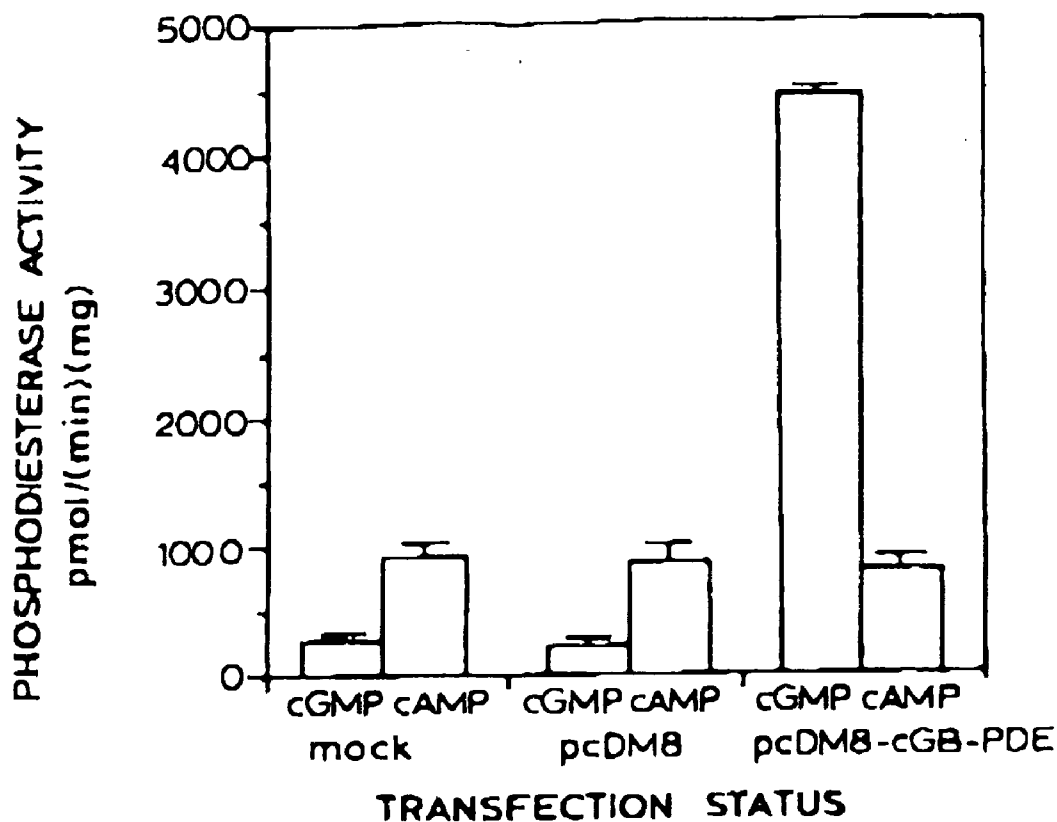


FIGURE 5

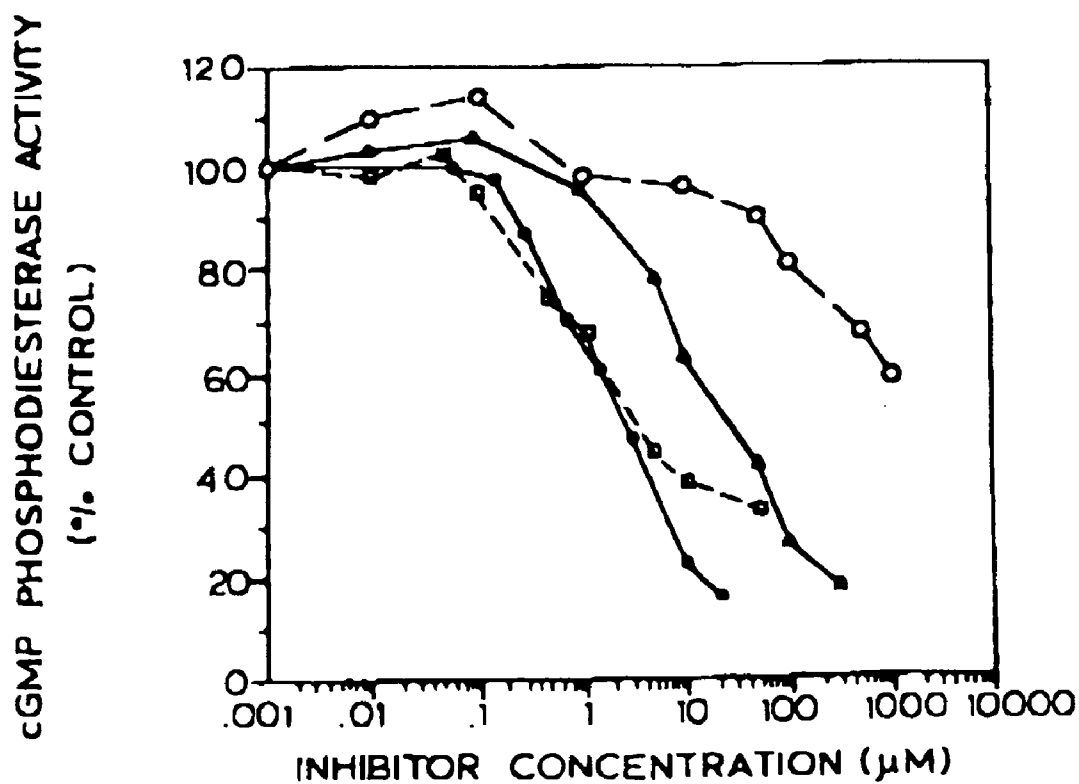


FIGURE 6

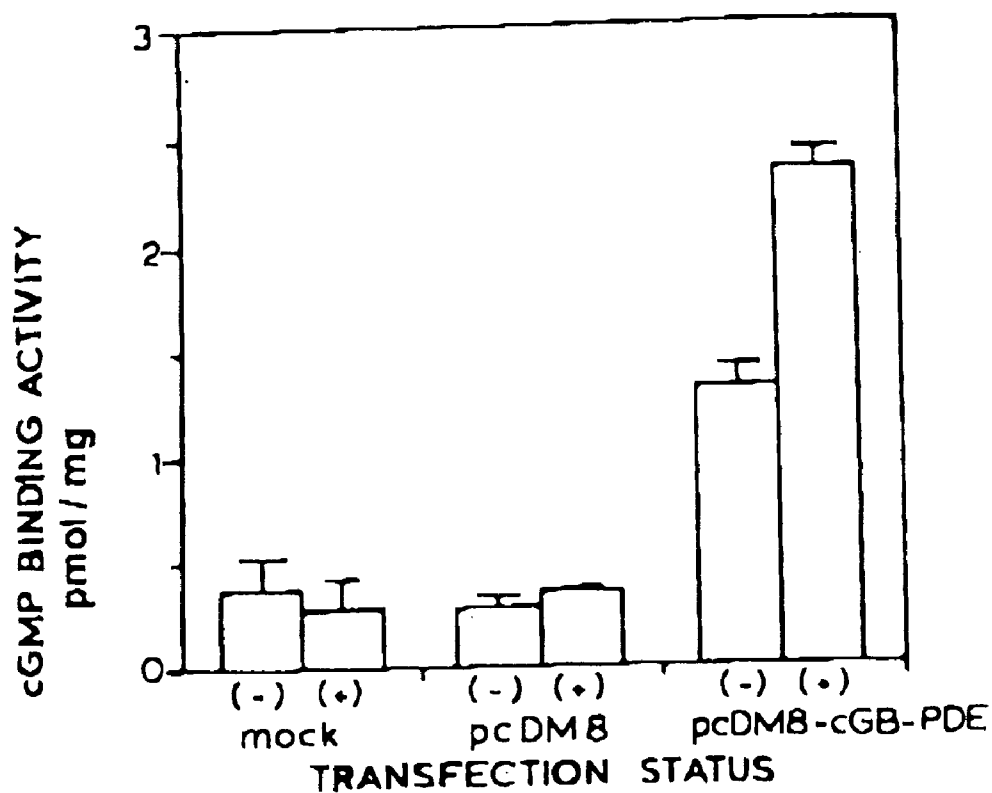


FIGURE 7

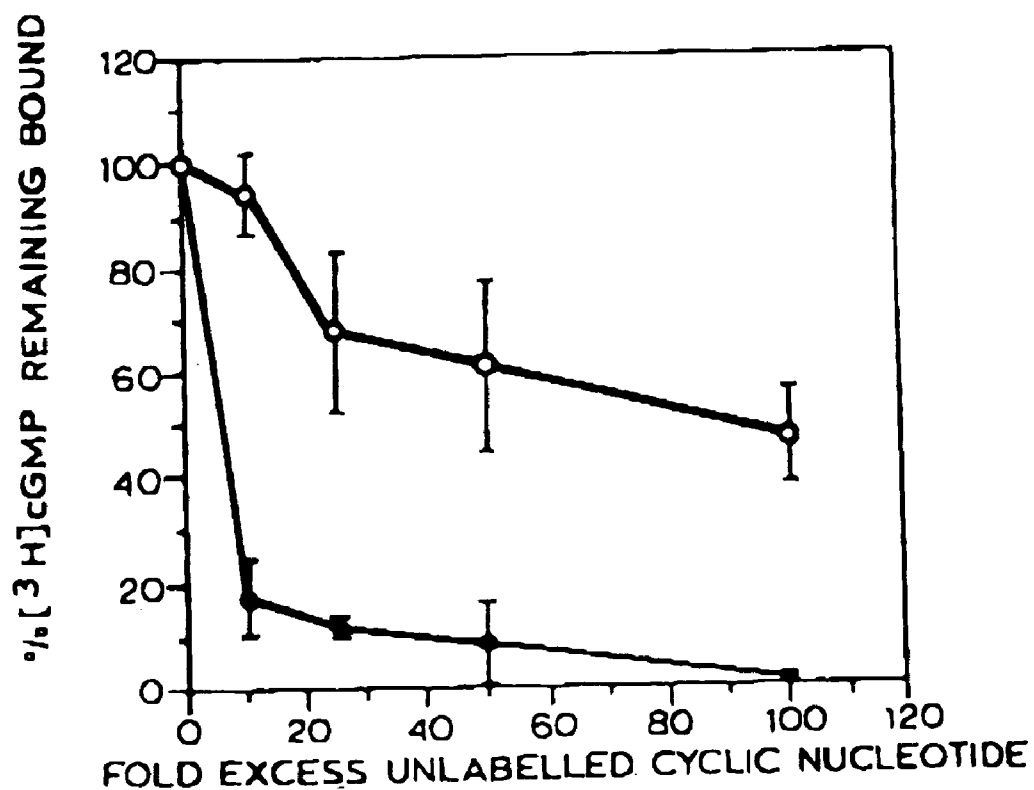


FIGURE 8

CYCLIC GMP-BINDING, CYCLIC GMP-SPECIFIC PHOSPHODIESTERASE MATERIALS AND METHODS

[0001] This application is a continuation-in-part of co-pending U.S. patent application Ser. No. 08/068,051 filed May 27, 1993.

[0002] Experimental work described herein was supported in part by Research Grants GM15731, DK21723, DK40029 and GM41269 and the Medical Scientist Training Program Grant GM07347 awarded by the National Institutes of Health. The United States government has certain rights in the invention.

FIELD OF THE INVENTION

[0003] The present invention relates generally to a cyclic guanosine monophosphate-binding, cyclic guanosine monophosphate-specific phosphodiester designated cGB-PDE and more particularly to novel purified and isolated polynucleotides encoding cGB-PDE polypeptides, to methods and materials for recombinant production of cGB-PDE polypeptide, and to methods for identifying modulators of cGB-PDE activity.

BACKGROUND

[0004] Cyclic nucleotide phosphodiesterases (PDEs) that catalyze the hydrolysis of 3'5' cyclic nucleotides such as cyclic guanosine monophosphate (cGMP) and cyclic adenosine monophosphate (cAMP) to the corresponding nucleoside 5' monophosphates constitute a complex family of enzymes. By mediating the intracellular concentration of the cyclic nucleotides, the PDE isoenzymes function in signal transduction pathways involving cyclic nucleotide second messenger.

[0005] A variety of PDEs have been isolated from different tissue sources and many of the PDEs characterized to date exhibit differences in biological properties including physicochemical properties, substrate specificity, sensitivity to inhibitor, immunological reactivity and mode of regulation. [See Beavo et al., *Cyclic Nucleotide Phosphodiesterases: Structure, Regulation and Drug Action*, John Wiley & Sons, Chichester, U.K. (1990)]. Comparison of the known amino acid sequences of various PDEs indicates that most PDEs are chimeric multidomain proteins that have distinct catalytic and regulatory domains. [See Charbonneau, pp. 267 in Beavo et al., supra] All mammalian PDEs characterized to date share a sequence of approximately 250 amino acid residues in length that appear to comprise the catalytic site and is located in the carboxyl terminal region of the enzyme. PDE domains that interact with allosteric or regulatory molecules are thought to be located within the amino-terminal regions of the isoenzymes. Based on their biological properties, the PDEs may be classified into six general families: the Ca²⁺/calmodulin-stimulated PDEs (Type I), the cGMP-stimulated PDEs (Type II), the cGMP-inhibited PDEs (Type III), the cAMP-specific PDEs (Type IV), the cGMP-specific phosphodiesterase COB-PDE (Type V) which is the subject of the present invention and the cGMP-specific photoreceptor PDEs (Type VI).

[0006] The cGMP-binding PDEs (Type II, Type V and Type VI PDEs), in addition to having a homologous catalytic domain near their carboxyl terminus, have a second

conserved sequence which is located closer to their amino terminus and which may comprise an allosteric cGMP-binding domain. See Charbonneau et al., *Proc. Natl. Acad. Sci. USA*, 87: 288-292 (1990).

[0007] The Type II cGMP-stimulated PDEs (cGs-PDEs) are widely distributed in different tissue types and are thought to exist as homodimers of 100-105 kDa subunits. The cGs-PDEs respond under physiological conditions to evaluated cGMP concentrations by increasing the rate of cAMP hydrolysis. The amino acid sequence of a bovine heart cGs-PDE and a partial cDNA sequence of a bovine adrenal cortex cGs-PDE are reported in LeTrong et al., *Biochemistry*, 29: 10280-10288 (1990) and full length bovine adrenal and human fetal brain cGB-PDE cDNA sequences are described in Patent Cooperation Treaty International Publication No. WO 92/18541 published on Oct. 29, 1992. The full length bovine adrenal cDNA sequence is also described in Sonnenburg et al., *J. Biol. Chem.*, 266: 17655-17661 (1991).

[0008] The photoreceptor PDEs and the cGB-PDE have been described as cGMP-specific PDEs because they exhibit a 50-fold or greater selectivity for hydrolyzing cGMP over cAMP.

[0009] The photoreceptor PDEs are the rod outer segment PDE (ROS-PDE) and the cone PDE (COS-PDE). The holoenzyme structure of the ROS-PDE consists of two large subunits α (88 kDa) and β (84 kDa) which are both catalytically active and two smaller γ regulatory subunits (both 11 kDa). A soluble form of the ROS-PDE has also been identified which includes α , β , and γ subunits and a δ subunit (15 kDa) that appears to be identical to the COS-PDE 15 kDa subunit. A full-length cDNA corresponding to the bovine membrane-associated ROS-PDE α subunit is described in Ovchinnikov et al., *FEBS Lett.*, 223: 169-173 (1987) and a full length cDNA corresponding to the bovine rod outer segment PDE β subunit is described in Lipkin et al., *J. Biol. Chem.*, 265: 12955-12959 (1990). Ovchinnikov et al., *FEBS Lett.*, 204: 169-173 (1986) presents a full-length cDNA corresponding to the bovine ROS-PDE γ subunit and the amino acid sequence of the δ subunit. Expression of the ROS-PDE has also been reported in brain in Collins et al., *Genomics*, 13: 698-704 (1992). The COS-PDE is composed of two identical α' (94 kDa) subunits and three smaller subunits of 11 kDa, 13 kDa and 15 kDa. A full-length cDNA corresponding to the bovine COS-PDE α' subunit is reported in Li et al., *Proc. Natl. Acad. Sci. USA*, 87: 293-297 (1990).

[0010] cGB-PDE has been purified to homogeneity from rat [Francis et al., *Methods Enzymol.*, 159: 722-729 (1988)] and bovine lung tissue [Thomas et al., *J. Biol. Chem.*, 265: 14964-14970 (1990), hereinafter "Thomas I"]. The presence of this or similar enzymes has been reported in a variety of tissues and species including rat and human platelets [Hamet et al., *Adv. Cyclic Nucleotide Protein Phosphorylation Res.*, 16: 119-136 (1984)], rat spleen [Coquil et al., *Biochem. Biophys. Res. Commun.*, 127: 226-231 (1985)], guinea pig lung [Davis et al., *J. Biol. Chem.*, 252: 4078-4084 (1977)], vascular smooth muscle [Coquil et al., *Biochim. Biophys. Acta*, 631: 148-165 (1980)], and sea urchin sperm [Francis et al., *J. Biol. Chem.*, 255: 620-626 (1979)]. cGB-PDE may be a homodimer comprised of two 93 kDa subunits. [See Thomas I, supra] cGB-PDE has been shown to contain a single site not found in other known cGMP-binding PDEs

which is phosphorylated by cGMP-dependent protein kinase (cGK) and, with a lower affinity, by cAMP-dependent protein kinase (cAK). [See Thomas et al., *J. Biol. Chem.*, 265: 14971-14978 (1990), hereinafter "Thomas II"]The primary amino acid sequence of the phosphorylation site and of the amino-terminal end of a fragment generated by chymotryptic digestion of cGB-PDE are described in Thomas II, supra, and Thomas I; supra, respectively. However, the majority of the amino acid sequence of cGB-PDE has not previously been described.

[0011] Various inhibitors of different types of PDEs have been described in the literature. Two inhibitors that exhibit some specificity for Type V PDEs are zaprinast and dipyridamole. See Francis et al., pp. 117-140 in Beavo et al., supra.

[0012] Elucidation of the DNA and amino acid sequences encoding the cGB-PDE and production of cGB-PDE polypeptide by recombinant methods would provide information and material to allow the identification of novel agent that selectively modulate the activity of the cGB-PDEs. The recognition that there are distinct types or families of PDE isoenzymes and that different tissues express different complements of PDEs has led to an interest in the development of PDE modulation which may have therapeutic indications for disease states that involve signal transduction pathways utilizing cyclic nucleotides as second messenger. Various selective and non-selective inhibitors of PDE activity are discussed in Murray et al., *Biochem. Soc. Trans.*, 20(2): 460-464 (1992). Development of PDE modulators without the ability to produce a specific PDE by recombinant DNA techniques is difficult because all PDEs catalyze the same basic reaction, have overlapping substrate specificities and occur only in trace amounts. As a result, purification to homogeneity of many PDEs is a tedious and difficult process.

[0013] There thus continues to exist a need in the art for DNA and amino acid sequence information for the cGB-PDE, for methods and means for the recombinant production of cGB-PDE polypeptides and for methods for identifying specific modulators of cGB-PDE activity.

SUMMARY OF THE INVENTION

[0014] The present invention provides novel purified and isolate polynucleotides (e.g., DNA sequences and RNA transcripts, both sense and antisense strands, including splice variants thereof) encoding the cGMP-binding, cGMP-specific PDE designated cGB-PDE. Preferred DNA sequences of the invention include genomic and cDNA sequences as well as wholly or partially chemically synthesized DNA sequences. DNA sequences encoding cGB-PDE that are set out in SEQ ID NO: 9 or 20 and DNA sequences which hybridize thereto under stringent conditions or DNA sequences which would hybridize thereto but for the redundancy of the genetic code are contemplated by the invention. Also contemplated by the invention are biological replicas (i.e., copies of isolated DNA sequences made in vivo or in vitro) of DNA sequences of the invention. Autonomously replicating recombinant constructions such as plasmid and viral DNA vectors incorporating cGB-PDE sequences and especially vectors wherein DNA encoding cGB-PDE is operatively linked to an endogenous or exogenous expression control DNA sequence and a transcrip-

tional terminator are also provided. Specifically illustrating expression plasmids of the invention is the plasmid hcgmet156-2 6n in *E. coli* strain JM109 which was deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Md. 20852, on May 4, 1993 as Accession No. 69296.

[0015] According to another aspect of the invention, host cells including procaryotic and eucaryotic cells, are stably transformed with DNA sequences of the invention in a manner allowing the desired polypeptides to be expressed therein. Host cells expressing cGB-PDE products can serve a variety of useful purposes. Such cells constitute a valuable source of immunogen for the development of antibody substances specifically immunoreactive with cGB-PDE. Host cells of the invention are conspicuously useful in methods for the large scale production of cGB-PDE polypeptides wherein the cells are grown in a suitable culture medium and the desired polypeptide products are isolated from the cells or from the medium in which the cells are grown by, for example, immunoaffinity purification.

[0016] cGB-PDE products may be obtained as isolates from natural cell sources or may be chemically synthesized, but are preferably produced by recombinant procedures involving host cells of the invention. Use of mammalian host cells is expected to provide for such post-translational modifications (e.g., glycosylation, truncation, lipidation and tyrosine, serine or threonine phosphorylation) as may be needed to confer optimal biological activity on recombinant expression products of the invention. cGB-PDE products of the invention may be full length polypeptides, fragments or variants. Variants may comprise cGB-PDE polypeptide analogs wherein one or more of the specified (i.e., naturally encoded) amino acids is deleted or replaced or wherein one or more nonspecified amino acids is added: (1) without loss of one or more of the biological activities or immunological characteristics specific for cGB-PDE; or (2) with specific disablement of a particular biological activity of cGB-PDE.

[0017] Also comprehended by the present invention are antibody substances (e.g., monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies, CDR-grafted antibodies and the like) and other binding proteins specific for cGB-PDE. Specific binding proteins can be developed using isolated or recombinant cGB-PDE or cGB-PDE variants or cells expressing such products. Binding proteins are useful, in turn, in compositions for immunization as well as for purifying cGB-PDE polypeptides and detection or quantification of cGB-PDE polypeptides in fluid and tissue samples by known immunological procedure. They are also manifestly useful in modulating (i.e., blocking, inhibiting or stimulating) biochemical activities of cGB-PDE, especially those activities involved in signal transduction. Anti-idiotypic antibodies specific for anti-cGB-PDE antibody substances are also contemplated.

[0018] The scientific value of the information contributed through the disclosures of DNA and amino acid sequence of the present invention is manifest. As one series of examples, knowledge of the sequence of a cDNA for cGB-PDE makes possible the isolation by DNA/DNA hybridization of genomic DNA sequences encoding cGB-PDE and specifying cGB-PDE expression control regulatory sequences such as promoters, operators and the like. DNA/DNA hybridization procedures carried out with DNA sequences of the

invention under stringent conditions are likewise expected to allow the isolation of DNAs encoding allelic variants of cGB-PDE, other structurally related proteins sharing one or more of the biochemical and/or immunological properties specific to cGB-PDE, and non-human species proteins homologous to cGB-PDE. Polynucleotides of the invention when suitably labelled are useful in hybridization assays to detect the capacity of cells to synthesize cGB-PDE. Polynucleotides of the invention may also be the basis for diagnostic methods useful for identifying a genetic alteration(s) in the cGB-PDE locus that underlies a disease state or states. Also made available by the invention are anti-sense polynucleotides relevant to regulating expression of cGB-PDE by those cells which ordinarily express the same.

[0019] The DNA and amino acid sequence information provided by the present invention also makes possible the systematic analysis of the structure and function of cGB-PDE and definition of those molecules with which it will interact. Agents that modulate cGB-PDE activity may be identified by incubating a putative modulator with lysate from eucaryotic cells expressing recombinant cGB-PDE and determining the effect of the putative modulator on cGB-PDE phosphodiesterase activity. In a preferred embodiment the eucaryotic cell lacks endogenous cyclic nucleotide phosphodiesterase activity. Specifically illustrating such a eucaryotic cell is the yeast strain YKS45 which was deposited with the ATCC on May 19, 1993 as Accession No. 74225. The selectivity of a compound that modulates the activity of the cGB-PDE can be evaluated by comparing its activity on the cGB-PDE to its activity on other PDE isozymes. The combination of the recombinant cGB-PDE products of the invention with other recombinant PDE products in a series of independent assays provides a system for developing selective modulators of cGB-PDE.

[0020] Selective modulators may include, for example, antibodies and other proteins or peptides which specifically bind to the cGB-PDE or cGB-PDE nucleic acid, oligonucleotides which specifically bind to the cGB-PDE or cGB-PDE nucleic acid and other non-peptide compounds (e.g., isolated or synthetic organic molecules) which specifically react with cGB-PDE or cGB-PDE nucleic acid. Mutant forms of cGB-PDE which affect the enzymatic activity or cellular location of the wild-type cGB-PDE are also contemplated by the invention. Presently preferred targets for the development of selective modulators include, for example: (1) the regions of the cGB-PDE which contact other proteins and/or localize the cGB-PDE within a cell, (2) the regions of the cGB-PDE which bind substrate, (3) the allosteric cGMP-binding site(s) of cGB-PDE, (4) the phosphorylation site(s) of cGB-PDE and (5) the regions of the cGB-PDE which are involved in dimerization of cGB-PDE subunits. Modulators of cGB-PDE activity may be therapeutically useful in treatment of a wide range of diseases and physiological conditions.

BRIEF DESCRIPTION OF THE DRAWINGS

[0021] Numerous other aspects and advantages of the present invention will be apparent upon consideration of the following detailed description thereof, reference being made to the drawing wherein:

[0022] FIG. 1A to 1C is an alignment of the conserved catalytic domains of several PDE isoenzymes wherein residues which are identical in all PDEs listed are indicated by

their one letter amino acid abbreviation in the "conserved" line, residues which are identical in the cGB-PDE and photoreceptor PDEs only are indicated by a star in the "conserved" line and gaps introduced for optimum alignment are indicated by periods;

[0023] FIG. 2A to 2C is an alignment of the cGMP-binding domains of several PDE isoenzymes wherein residues which are identical in all PDEs listed are indicated by their one letter amino acid abbreviation in the "conserved" line and gaps introduced for optimum alignment are indicated by periods;

[0024] FIG. 3 is an alignment of internally homologous repeats from several PDE isoenzymes wherein residues identical in each repeat A and B from all cGMP-binding PDEs listed are indicated by their one letter amino acid abbreviation in the "conserved" line and stars in the "conserved" line represent positions in which all residues are chemically conserved;

[0025] FIG. 4 schematically depicts the domain organization of cGB-PDE,

[0026] FIG. 5 is a bar graph representing the results of experiments in which extracts of COS cells transfected with bovine cGB-PDE sequences or extracts of untransfected COS cells were assayed for phosphodiesterase activity using either 20 μ M cGMP or 20 μ M cAMP as the substrate;

[0027] FIG. 6 is a graph depicting results of assays of extracts from cells transfected with bovine cGB-PDE sequences for cGMP phosphodiesterase activity in the presence of a series of concentrations of phosphodiesterase inhibitors including dipyrindamole (closed squares), zaprinast (closed circles), methoxymethylxanthine (closed triangles) and rolipram (open circles);

[0028] FIG. 7 is a bar graph presenting results of experiments in which cell extracts from COS cells transfected with bovine cGB-PDE sequences or control untransfected COS cells were assayed for [³H]cGMP-binding activity in the absence (-) or presence (+) of 0.2 mM IBMX; and

[0029] FIG. 8 is a graph of the results of assays in which extracts from cells transfected with bovine cGB-PDE sequences were assayed for [³H]cGMP-binding activity in the presence of excess unlabelled cAMP (open circles) or cGMP (closed circles) at the concentrations indicated.

DETAILED DESCRIPTION

[0030] The following examples illustrate the invention. Example 1 describes the isolation of a bovine cGB-PDE cDNA fragment by PCR and subsequent isolation of a full length cGB-PDE cDNA using the PCR fragment as a probe. Example 2 presents an analysis of the relationship of the bovine cGB-PDE amino acid sequence to sequences reported for various other PDEs. Northern blot analysis of cGB-PDE mRNA in various bovine tissues is presented in Example 3. Expression of the bovine cGB-PDE cDNA in COS cells is described in Example 4. Example 5 presents results of assays of the cGB-PDE COS cell expression product for phosphodiesterase activity, cGMP-binding activity and Zn²⁺ hydrolase activity. Example 6 describes the isolation of human cDNAs homologous to the bovine cGB-PDE cDNA. The expression of a human cGB-PDE cDNA in yeast cells is presented in Example 7. RNase protection

assays to detect cGB-PDE in human tissues are described in Example 8. Example 9 describes the bacterial expression of human cGB-PDE cDNA and the development of antibodies reactive with the bacterial cGB-PDE expression product. Example 10 describes cGB-PDE analogs and fragments. The generation of monoclonal antibodies that recognize cGB-PDE is described in Example 11. Example 12 relates to utilizing recombinant cGB-PDE products of the invention to develop agents that selectively modulate the biological activities of cGB-PDE.

EXAMPLE 1

[0031] The polymerase chain reaction (PCR) was utilized to isolate a cDNA fragment encoding a portion of cGB-PDE from bovine lung first strand cDNA. Fully degenerate sense and antisense PCR primers were designed based on the partial cGB-PDE amino acid sequence described in Thomas I, supra, and novel partial amino-acid sequence information.

[0032] A. Purification of cGB-PDE Protein

[0033] cGB-PDE was purified as described in Thomas I, supra, or by a modification of that method as described below.

[0034] Fresh bovine lungs (5-10 kg) were obtained from a slaughterhouse and immediately placed on ice. The tissue was ground and combined with cold PEM buffer (20 mM sodium phosphate, pH 6.8, containing 2 mM EDTA and 25 mM β -mercaptoethanol). After homogenization and centrifugation, the resulting supernatant was incubated with 4-7 liters of DEAE-cellulose (Whatman, UK) for 3-4 hours. The DEAE slurry was then filtered under vacuum and rinsed with multiple volumes of cold PEM. The resin was poured into a glass column and washed with three to four volumes of PEM. The protein was eluted with 100 mM NaCl in PEM and twelve 1-liter fractions were collected. Fractions were assayed for IBMX-stimulated cGMP binding and cGMP phosphodiesterase activities by standard procedures described in Thomas et al., supra. Appropriate fractions were pooled, diluted 2-fold with cold, deionized water and subjected to Blue Sepharose® CL-6B (Pharmacia LKB Biotechnology Inc., Piscataway, N.J.) chromatography. Zinc chelate affinity adsorbent chromatography was then performed using either an agarose or Sepharose-based gel matrix. The resulting protein pool from the zinc chelation step treated as described in the Thomas I, supra, or was subjected to a modified purification procedure.

[0035] As described in Thomas I, supra, the protein pool was applied in multiple loads to an HPLC Bio-Sil TSK-545 DEAE column (150x21.5 mm) (BioRad Laboratories, Hercules, Calif.) equilibrated in PEM at 4° C. After in equilibration period, a 120-ml wash of 50 mM NaCl in PEM was followed by a 120-ml linear gradient (50-200 mM NaCl in PEM) elution at a flow rate of 2 ml/minute. Appropriate fractions were pooled and concentrated in dialysis tubing against Sephadex G-200 (Boehringer Mannheim Biochemicals, UK) to a final volume of 1.5 ml. The concentrated cGB-PDE pool was applied to an HPLC gel filtration column (Bio-Sil TSK-250, 500x215 mm) equilibrated in 100 mM sodium phosphate, pH 6.8, 2 mM EDTA, 25 mM β -mercaptoethanol and eluted with a flow rate of 2 ml/minute at 4° C.

[0036] If the modified, less cumbersome procedure was performed, the protein pool was dialyzed against PEM for 2

hours and loaded onto a 10 ml preparative DEAE Sephacel column (Pharmacia) equilibrated in PEM buffer. The protein was eluted batchwise with 0.5M NaCl in PEM, resulting in an approximately 10-15 fold concentration of protein. The concentrated protein sample was loaded onto an 800 ml (2.5 cmx154 cm) Sephacryl S400 gel filtration column (Boehringer) equilibrated in 0.1M NaCl in PEM, and eluted at a flow rate: of 1.7 ml/minute.

[0037] The purity of the protein was assessed by Coomassie staining after sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). Approximately 0.5-3.0 mg of pure cGB-PDE were obtained per 10 kg bovine lung.

[0038] Rabbit polyclonal antibodies specific for the purified bovine cGB-PDE were generated by standard procedures.

[0039] B. Amino Acid Sequencing of cGB-PDE

[0040] cGB-PDE phosphorylated with [³²P]ATP and was then digested with protease yield ³²P-labelled phosphopeptides. Approximately 100 μ g of purified cGB-PDE was phosphorylated in a reaction mixture containing 9 mM MgCl₂, 9 μ M [³²P]ATP, 10 μ M cGMP, and 4.2 μ g purified bovine catalytic subunit of cAMP-dependent protein kinase (cAK) in a final volume of 900 μ l. Catalytic subunit of cAK was prepared according to the method of Flockhart et. al., pp. 209-215 in Marangos et al., *Brain Receptor Methodologies, Part A*, Academic Press, Orlando, Fla. (1984). The reaction was incubated for 30 minutes at 30° C., and stopped by addition of 60 μ l of 200 mM EDTA.

[0041] To obtain a first peptide sequence from cGB-PDE, 3.7 μ l of a 1 mg/ml solution of a α -chymotrypsin in KPE buffer (10 mM potassium phosphate, pH 6.8, with 2 mM EDTA) was added to 100 μ g purified, phosphorylated cGB-PDE and the mixture was incubated for 30 minutes at 30° C. Proteolysis was stopped by addition of 50 μ l of 10% SDS and 25 μ l of β -mercaptoethanol. The sample was boiled until the volume was reduced to less than 400 μ l, and was loaded onto an 8% preparative SDS-polyacrylamide gel and subjected to electrophoresis at 50 mAmps. The separated digestion products were electroblotted onto Immobilon polyvinylidene difluoride (Millipore, Bedford, Mass.), according to the method of Matsudaira, *J. Biol. Chem*, 262: 10035-10038 (1987). Transferred protein was identified by Coomassie Blue staining, and a 50 kDa band was excised from the membrane for automated gas-phase amino acid sequencing. The sequence of the peptide obtained by the α -chymotryptic digestion procedure is set out below as SEQ ID NO: 1.

REXDANRINMYAQVVKNTM SEQ ID NO: 1

[0042] A second sequence was obtained from a cGB-PDE peptide fragment generated by V8 proteolysis. Approximately 200 μ g of purified cGB-PDE was added to 10 mM MgCl₂, 10 μ M [³²P]ATP, 100 μ M cGMP, and 1 μ g/ml purified catalytic subunit of cAK in a final volume of 1.4 ml. The reaction was incubated for 30 minutes at 30° C., and was terminated by the addition of 160 μ l of 0.2M EDTA. Next, 9 μ l of 1 mg/ml *Staphylococcal aureus* V8 protease (International Chemical Nuclear Biomedicals, Costa Mesa, Calif.) diluted in KPE was added, followed by a 15 minute

incubation at 30° C. Proteolysis was stopped by addition of 88 μ l of 20% SDS and 45 μ l β -mercaptoethanol. The digestion products were separated by electrophoresis on a preparative 10% SDS-polyacrylamide gel run at 25 mAmps for 4.5 hours. Proteins were electroblotted and stained as described above. A 28 kDa protein band was excised from the membrane and subjected to automated gas-phase amino acid sequencing. The sequence obtained is set out below as SEQ ID NO: 2.

QSLAAAVVP SEQ ID NO: 2

[0043] C. PCR Amplification of Bovine cDNA

[0044] The partial amino acid sequences utilized to design primers (SEQ ID NO: 3, below, and amino acids 9-20 of SEQ ID NO: 1) and the sequences of the corresponding PCR primers (in IUPAC nomenclature) are set below wherein SEQ ID NO: 3 is the sequence reported in Thomas I, supra.

SEQ ID NO: 3

F D N D E G E Q
5' TTY GAY AAY GAY GAR GGN GAR CA 3'

3' AAR CTR TTR CTR CTY CCN CTY GT 5'

SEQ ID NO: 1, Amino acids 9-20

N Y M Y A Q Y V K N T M
5' AAY TAY ATG TAY GCN CAR TAY GT 3'

3' TTR ATR TAC ATR CGN GTY ATR CA 5'

3' TTR ATR TAC ATR CGN GTY ATR CAN TTY TTR TGN TAC 5'

not homologous to any known PDE sequence or to the known partial cGB-PDE sequences.

[0047] PCR was performed again on bovine lung first strand cDNA using the primers set out in SEQ ID NOs; 4 and 7. A clone containing a 0.8 Kb insert with a single large open reading frame was identified. The open reading frame method a polypeptide that included the amino acids KNTM (amino acids 17-20 of SEQ ID NO: 1 which were not utilized to design the primer sequence which is set out in SEQ ID NO: 7) and that possessed a high degree of homology to the deduced amino acid sequences of the cGs-, ROS- and COS-PDEs. The clone identified corresponds to nucleotides 489-1312 of SEQ ID NO: 9.

[0048] D. Construction and Hybridization Screening of a Bovine cDNA Library

[0049] In order to obtain a cDNA encoding a full-length cGB-PDE, a bovine lung cDNA library was screened using the ³²P-labelled PCR-generated cDNA insert as a probe.

(SEQ ID NO: 4)

(SEQ ID NO: 5)

(SEQ ID NO: 6)

(SEQ ID NO: 7)

(SEQ ID NO: 8)

[0045] The sense and antisense primers, synthesized using an Applied Biosystems Model 380A DNA Synthesizer (Foster City, Calif.), were used in all possible combinations to amplify cGB-PDE-specific sequences from bovine lung first strand cDNA as described below.

[0046] After ethanol precipitation, pairs of oligonucleotides were combined (SEQ ID NO: 4 or 5 combined with SEQ ID NOs: 6, 7 or 8) at 400 nM each in a PCR reaction. The reaction was run using 50 ng first strand bovine lung cDNA (generated using AMV reverse transcriptase and random primers on oligo dT selected bovine lung mRNA), 200 μ M dNTPs, and 2 units of Taq polymerase. The initial denaturation step was carried out at 94° C. for 5 minutes, followed by 30 cycles of a 1 minute denaturation step at 94° C., a two minute annealing step at 50° C., and a 2 minute extension step at 72° C. PCR was performed using a Hybrid Thermal Reactor (ENK Scientific Products, Saratoga, Calif.) and products were treated by gel electrophoresis on a 1% low melting point agarose gel run in 40 mM Tris-acetate, 2 mM EDTA. A weak band of about 800-840 bp was seen with the primers set out in SEQ ID NOs: 4 and 7 and with primers set out in SEQ ID NOs: 4 and 8. None of the other primer pairs yielded visible bands. The PCR product generated by amplification with the primers set out in SEQ ID NOs: 4 and 7 was isolated using the Gene Clean® (Bio101, La Jolla, Calif.) DNA purification kit according to the manufacturer's protocol. The PCR product (20 ng) was ligated into 200 ng of linearized pBluescript KS(+) (Stratagene, La Jolla, Calif.), and the resulting plasmid construct was used to transform *E. coli* XL1 Blue cells (Stratagene Cloning Systems, La Jolla, Calif.). Putative transformation positives were screened by sequencing. The sequences obtained were

[0050] Polyadenylated RNA was prepared from bovine lung as described Sonnenburg et al., *J. Biol. Chem.*, 266: 17655-17661 (1991). First strand cDNA was synthesized using AMV reverse transcriptase (Life Sciences, St. Petersburg, Fla.) with random hexanucleotides primers as described in Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley & Sons, New York (1987). Second strand cDNA was synthesized using *E. coli* DNA polymerase in the presence of *E. coli* DNA ligase and *E. coli* RNase H. Selection of cDNAs larger than 500 bp was performed by Sepharose® CL-4B (Millipore) chromatography. EcoRI adaptors (Promega, Madison, Wis.) were ligated to the cDNA using T4 DNA ligase. Following heat inactivation of the ligase, the cDNA was phosphorylated using T4 polynucleotide kinase. Unligated adaptors were removed by Sepharose® CL-4B chromatography (Pharmacia, Piscataway, N.J.). The cDNA was ligated into EcoRI-digested, dephosphorylated lambda Zap®II arms (Stratagene) and packaged with Gigapack® Gold (Stratagene) extracts according to the manufacturer's protocol. The titer of the unamplified library was 9.9×10^5 with 18% nonrecombinants. The library was amplified by plating 50,000 plaque forming units (pfu) on to twenty 150 mm plates, resulting in a final titer of 5.95×10^6 pfu/ml with 21% nonrecombinants.

[0051] The library was plated on twenty-four 150 mm plates at 50,000 pfu/plate, and screened with the ³²P-labelled cDNA clone. The probe was prepared using the method of Feinberg et al., *Anal Biochem*, 137: 266-267 (1984), and the ³²P-labelled DNA was purified using Elutip-D® columns (Schleicher and Schuell Inc., Keene, N.H.) using the manu-

facturer's protocol. Plaque-lifts were performed using 15 cm nitrocellulose filters. Following denaturation and neutralization, DNA was fixed onto the filter by baking at 80° C. for 2 hours. Hybridization was carried out at 42° C. overnight in a solution containing 50% formamide, 5×SSC (0.75M NaCl, 0.75M sodium citrate, pH 7), 25 mM sodium phosphate (pH 7.0), 2× Denhardt's solution, 10% dextran sulfate, 90 µg/ml yeast tRNA, and approximately 10⁶ cpm/ml ³²P-labelled probe (5×10⁶[text missing or illegible when filed]cpm/µg). The filters were washed twice in 0.1×SSC, 0.1% SDS at room temperature for 15 minutes per wash, followed by a single 20 minute wash in 0.1×SSC, 1% SDS at 45° C. The filters were then exposed to X-ray film at -70° C. for several days.

[0052] Plaques that hybridized with the labelled probe were purified by several rounds of replating and rescreening. Insert cDNAs were subcloned into the pBluescript SK(-) vector (Stratagene) by the in vivo excision method described by the manufacturer's protocol. Southern blots were performed in order to verify that the rescued cDNA hybridized to the PCR probe. Putative cGB-PDE cDNAs were sequenced using Sequenase® Version 2.0 (United States Biochemical Corporation, Cleveland, Ohio) or TaqTrack® kits (Promega).

[0053] distinct cDNA clones designated cGB-2, cGB-8 and cGB-10 were isolated. The DNA and deduced amino acid sequences of clone cGB-8 are set out in SEQ ID NOs: 9 and 10. The DNA sequence downstream of nucleotide 2686 may represent a cloning artifact. The DNA sequence of cGB-10 is identical to the sequence of cGB-8 with the exception of one nucleotide. The DNA sequence of clone cGB-2 diverges from that of clone cGB-8 5' to nucleotide 219 of clone cgb-8 (see SEQ ID NO: 9) and could encode a protein with a different amino terminus.

[0054] The cGB-8 cDNA clone is 4474 bp in length and contains a large open reading frame of 2625 bp. The triplet ATG at position 99-101 in the nucleotide sequence is predicted to be the translation initiation site of the cGB-PDE gene because it is preceded by an in-frame stop codon and the surrounding bases are compatible with the Kozak consensus initiation site for eucaryotic mRNAs. The stop codon TAG is located at positions 2724-2726, and is followed by 1748 bp of 3' untranslated sequence. The sequence of cGB-8 does not contain a transcription termination consensus sequence, therefore the clone may not represent the entire 3' untranslated region of the corresponding rRNA.

[0055] The open reading frame of the cGB-8 cDNA encodes an 875 amino acid polypeptide with a calculated molecular mass of 99.5 kD. This calculated molecular mass is only slightly larger than the reported molecular mass of purified cGB-PDE, estimated by SDS-PAGE analysis to be approximately 93 kDa. The deduced amino acid sequence of cGB-8 corresponded exactly to all peptide sequences obtained from purified bovine lung cGB-PDE providing strong evidence that cGB-8 encodes cGB-PDE.

EXAMPLE 2

[0056] A search of the SWISS-PROT and GEnEmbl data banks (Release of February, 1992) conducted using the FASTA program supplied with the Genetics Computer Group (GCG) Software Package (Madison, Wis.) revealed

that only DNA and amino acid sequences reported for other PDEs displayed significant similarity to the DNA and deduced amino acid of clone cGB-8.

[0057] Pairwise comparisons of the cGB-PDE deduced amino acid sequence with the sequences of eight other PDEs were conducted using the ALIGN [Dayhoff et al., *Methods Enzymol.*, 92; 524-545 (1983)] and BESTFIT [Wilbur et al., *Proc. Natl. Acad. Sci. USA*, 80: 726-730 (1983)] programs. Like all mammalian phosphodiesterases sequenced to date, cGB-PDE contains a conserved catalytic domain sequence of approximately 250 amino acids in the carboxyl-terminal half of the protein that is thought to be essential for catalytic activity. This segment comprises amino acids 578-812 of SEQ ID NO: 9 and exhibits sequence conservation with the corresponding regions of other PDEs. Table 1 below sets out the specific identity values obtained in pairwise comparisons of other PDEs with amino acids 578-812 of cGB-PDE, wherein "ratdunce" is the rat cAMP-specific PDE; "61 kCaM" is the bovine 61 kDa calcium/calmodulin-dependent PDE; "63 kCaM" is the bovine 63 kDa calcium/calmodulin-dependent PDE; "drosdunce" is the *drosophila* cAMP-specific PDE; "ROS-α" is the bovine ROS-PDE α-subunit; "ROS-β" is the bovine ROS-PDE β-subunit; "COS-α" is the bovine COS-PDE α subunit; and "cGs" is the bovine cGs-PDE (612-844).

TABLE 1

Phosphodiesterase	Catalytic Domain Residues	% Identity
Ratdunce	77-316	31
61 kCaM	193-422	29
63 kcam	195-424	29
drosdunce	1-239	28
ROS-α	535-778	45
ROS-β	533-776	46
COS-α'	533-776	48
cGs	612-844	40

[0058] Multiple sequence alignments were performed using the Progressive Alignment Algorithm [Feng et al., *Methods Enzymol.*, 183: 375-387 (1990)] implemented in the PILEUP program (GCG Software). FIG. 1A to 1C shows a multiple sequence alignment of the proposed catalytic domain of cGB-PDE with the all the corresponding regions of the PDEs of Table 1. Twenty-eight residues (see residues indicated by one letter amino acid abbreviations in the "conserved" line on FIG. 1A to 1C) are invariant among the isoenzymes including several conserved histidine residues predicted to play a functional role in catalysis. See Charbonneau et al., *Proc. Natl. Acad. Sci. USA*, supra. The catalytic domain of cGB-PDE more closely resembles the catalytic domains of the ROS-PDEs and COS-PDEs than the corresponding regions of other PDE isoenzymes. That are several conserved regions among the photoreceptor PDEs and cGB-PDE that are not shared by other PDEs. Amino acid positions in these regions that are invariant in the photoreceptor PDE and cGB-PDE sequences are indicated by stars in the "conserved" line of FIG. 1A to 1C. Regions of homology among cGB-PDE and the ROS- and COS-PDEs may serve important roles in conferring specificity for cGMP hydrolysis relative to cAMP hydrolysis or for sensitivity to specific pharmacological agents.

[0059] Sequence similarity between cGB-PDE, cGs-PDE and the photoreceptor PDEs, is not limited to the conserved

catalytic domain but also includes the noncatalytic cGMP binding domain in the amino-terminal half of the protein. Optimization of the alignment between cGB-PDE, cGs-PDE and the photoreceptor PDEs indicates that an amino-terminal conserved segment may exist including amino acids 142-526 of SEQ ID NO: 9. Pairwise analysis of the sequence of the proposed cGMP-binding domain of cGB-PDE with the corresponding regions of the photoreceptor PDEs and cGs-PDE revealed 26-28% sequence identity. Multiple sequence alignment of the proposed cGMP-binding domains with the cGMP-binding PDEs is shown in FIG. 2A to 2C wherein abbreviations are the same as indicated for Table 1. Thirty-eight positions in this non-catalytic domain appear to be invariant among all cGMP-binding PDEs (see positions indicated by one letter amino acid abbreviations in the "conserved" line of FIG. 2A to 2C).

[0060] The cGMP-binding domain of the cGMP-binding PDEs contains internally homologous repeats which may form two similar but distinct inter- or intra-subunit cGMP-binding sites. FIG. 3 shows a multiple sequence alignment of the repeats a (corresponding to amino acids 228-311 of cGB-PDE) and b (corresponding to amino acids 410-500 of cGB-PDE) of the cGMP-binding PDEs. Seven residues are invariant in each A and B regions (see residue indicated by one letter amino acid abbreviations in the "conserved" line of FIG. 3). Residue that are chemically conserved in the A and B regions are indicated by stars in the "conserved" line of FIG. 3. cGMP analog studies of cGB-PDE support the existence of a hydrogen bond between the cyclic nucleotide binding site on cGB-PDE and the 2' OH of cGMP.

[0061] Three regions of cGB-PDE have no significant sequence similarity to other PDE isoenzymes. These regions include the sequence flanking the carboxyl-terminal end of the catalytic domain (amino acids 812-875), the sequence separating the cGMP-binding and catalytic domains (amino acids 527-577) and the amino-terminal sequence spanning amino acids 1-141. The site (the serine at position 92 of SEQ ID NO: 10) of phosphorylation of cGB-PDE by cGK is located in this amino-terminal region of sequence. Binding of cGMP to the allosteric site on cGB-PDE is required for its phosphorylation.

[0062] A proposed domain structure of cGB-PDE based on the foregoing comparisons with other PDE isoenzymes is presented in FIG. 4. This domain structure is supported by the biochemical studies of cGB-PDE purified from bovine lung.

EXAMPLE 3

[0063] The presence of cGB-PDE mRNA in various bovine tissues was examined by Northern blot hybridization.

[0064] Polyadenylated RNA was purified from total RNA preparations using the Poly(A) Quick® mRNA purification kit (Stratagene) according to the manufacturer's protocol. RNA sample (5 µg) were loaded onto a 1.2% agarose, 67% formaldehyde gel. Electrophoresis and RNA transfer were performed as previously described in Sonnenburg et al., supra. Prehybridization of the RNA blot was carried out for 4 hours at 45° C. in a solution containing 50% formamide, 5×SSC, 25 mM sodium phosphate, pH 7, 2× Denhardt's solution, 10% dextran sulfate, and 0.1 mg/ml yeast tRNA. A random hexanucleotide-primer-labelled probe (5×10⁶ [text missing or illegible when filed], cpm/µg) was prepared

as described in Feinberg et al., supra, using the 4.7 kb cGB-8 cDNA clone of Example 2 exercised by digestion with AccI and SacII. The probe was heat denatured and injected into a blotting bag (6×10⁵ cpm/ml) following prehybridization. The Northern blot was hybridized overnight at 45° C., followed by one 15 minute wash with 2×SSC, 0.1% SDS at room temperature, and three 20 minute washes with 0.1×SSC, 0.1% SDS at 45° C. The blot was exposed to X-ray film for 24 hours at -70° C. The size of the RNA that hybridized with the cGB-PDE probe was estimated using a 0.24-9.5 kb RNA ladder that was stained with ethidium bromide and visualized with UV light.

[0065] The ³²P-labeled cGB-PDE cDNA hybridized to a single 6.8 kb bovine lung RNA species. A mRNA band of the identical size was also detected in polyadenylated RNA isolated from bovine trachea, aorta, kidney and spleen.

EXAMPLE 4

[0066] The cGB-PDE cDNA in clone cGB-8 of Example 2 was expressed in COS-7 cells (ATCC CRL1651).

[0067] A portion of the cGB-8 cDNA was isolated following digestion with the restriction enzyme XbaI. XbaI cut at a position in the pBluescript polylinker sequence located 30 bp upstream of the 5' end of the cGB-8 insert and at position 3359 within the cGB-8 insert. The resulting 3399 bp fragment, which contains the entire coding region of cGB-8, was then ligated into the unique XbaI cloning site of the expression vector pCDM8 (Invitrogen, San Diego, Calif.). The pCDM8 plasmid is a 4.5 kb eucaryotic expression vector containing a cytomegalovirus promoter and enhancer, an SV40-derived origin of replication, a polyadenylation signal, a procaryotic origin of replication (derived from pBR322) and a procaryotic genetic marker (supF). *E. coli* MC1061/P3 cells (Invitrogen) were transformed with the resulting ligation products, and transformation positive colonies were screened for proper orientation of the cGB-8 insert using PCR and restriction enzyme analysis. The resulting expression construct containing the cGB-8 insert in the proper orientation is referred to as pCDM8-cGB-PDE.

[0068] The pCDM8-cGB-PDE DNA was purified from large-scale plasmid preparations using Qiagen pack-500 columns (Chatsworth, Calif.) according to the manufacturer's protocol. COS-7 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum, 50 µg/ml penicillin and 50 µg/ml streptomycin at 37° C. in a humidified 5% CO₂ atmosphere. Approximately 24 hours prior to transfection, confluent 100 mm dishes of cells were replated at one-fourth or one-fifth the original density. In a typical transfection experiment, cells were washed with buffer containing 137 mM NaCl, 2.7 mM KCl, 1.1 mM potassium phosphate, and 8.1 mM sodium phosphate, pH 7.2 (PBS). Then 4-5 ml of DMEM containing 10% NuSerum (Collaborative Biomedical Products, Bedford, Mass.) was added to each plate. Transfection with 10 µg pCDM8-cGB-PDE DNA or pCDM8 vector DNA mixed with 400 µg DEAE-dextran (Pharmacia) in 60 µl TBS [Tris-buffered saline: 25 mM Tris-HCl (pH 7.4), 137 mM NaCl, 5 mM KCl, 0.6 mM Na₂HPO₄, 0.7 mM CaCl₂, and 0.5 mM MgCl₂] was carried out by dropwise addition of the mixture to each plate. The cells were incubated at 37° C., 5% CO₂ for 4 hours, and then treated with 10% dimethyl sulfoxide in PBS for 1 minute. After 2 minutes, the dimethyl sulfoxide was removed, the cells were washed with PBS and

incubated in complete medium. After 48 hours, cells were suspended in 0.5-1 ml of cold homogenization buffer [40 mM Tris-HCl (pH 7.5), 15 mM benzamidine, 15 mM β -mercaptoethanol, 0.7 μ g/ml pepstatin A, 0.5 μ g/ml leupeptin, and 5 μ M EDTA] per plate of cells, and disrupted using a Dounce homogenizer. The resulting whole extracts were assayed for phosphodiesterase activity, cGMP-binding activity, and total protein concentration as described below in Example 5.

EXAMPLE 5

[0069] Phosphodiesterase activity in extracts of the transfected COS cells of Example 4 or in extracts of mock transfected COS cells was measured using a modification of the assay procedure described for the cGs-PDE in Martins et al., *J. Biol. Chem.* 257: 1973-1979 (1982). Cells were harvested and extracts prepared 48 hours after transfection. Incubation mixtures contained 40 mM MOPS buffer (pH 7), 0.8 mM EDTA, 15 mM magnesium acetate, 2 mg/ml bovine serum albumin, 20 μ M [3 H]cGMP or [3 H]cAMP (100,000-200,000 cpm/assay) and COS-7 cell extract in a total volume of 250 μ l. The reaction mixture was incubated for 10 minutes at 30° C., and then stopped by boiling. Next, 10 μ l of 10 mg/ml *Crotalus atrax* venom (Sigma) was added followed by a 10 minute incubation at 30° C. Nucleoside products were separated from unreacted nucleotides as described in Martins et al., supra. In all studies, less than 15% of the total [3 H]cyclic nucleoside was hydrolyzed during the reaction.

[0070] The results of the assays are presented in FIG. 5 wherein the results shown are averages of three separate transfections. Transfection of COS-7 cells with pCDM8-PDE DNA resulted in the expression of approximately 15-fold higher levels of cGMP phosphodiesterase activity than in mock-transfected cells or in cells transfected with pCDM8 vector alone. No increase in cAMP phosphodiesterase activity over mock or vector-only transfected cells was detected in extracts from cells transfected with pCDM8-cGB-PDE DNA. These results confirm that the cGB-PDE bovine cDNA encodes a cGMP-specific phosphodiesterase.

[0071] Extracts from the transfected COS cells of Example 4 were also assayed for cGMP PDE activity in the presence of a series of concentrations of the PDE inhibitors zaprinast, dipyrindamole (Sigma), isobutyl-1-methyl-8-methoxymethylxanthine (MeOxMeMIX) and rolipram.

[0072] The results of the assays are presented in FIG. 6 wherein PDE activity in the absence of inhibitor is taken as 100% and each data point represents the average of two separate determinations. The relative potencies of PDE inhibitors for inhibition of cGMP hydrolysis by the expressed cGB-PDE cDNA protein product were identical to those relative potencies reported for native cGB-PDE purified from bovine lung (Thomas I, supra). IC₅₀ values calculated from the curves in FIG. 6 are as follows: zaprinast (closed circles), 2 μ M; dipyrindamole (closed squares), 3.5 μ M; MeOxMeMIX (closed triangles), 30 μ M; and rolipram (open circles), >300 μ M. The IC₅₀ value of zaprinast, a relatively specific inhibitor of cGMP-specific phosphodiesterases, was at least two orders of magnitude lower than that reported for inhibition of phosphodiesterase activity of the cGs-PDE or of the cGMP-inhibited phosphodiesterase (cGi-PDEs) (Reeves et al., pp. 300-316 in Beavo et al., supra). Dipyrindamole, an effective inhibitor of selected cAMP- and cGMP-specific phosphodiesterases, was also a potent inhibitor of the expressed cGB-PDE. The relatively selective inhibitor of calcium/calmodulin-stimulated phospho-

phodiesterase (CaM-PDEs), MeOxMeMIX, was approximately 10-fold less potent than zaprinast and dipyrindamole, in agreement with results using cGB-PDE activity purified from bovine lung. Rolipram, a potent inhibitor of low K_m cAMP phosphodiesterases, was a poor inhibitor of expressed cGB-PDE cDNA protein product. These results show that the cGB-PDE cDNA encodes a phosphodiesterase that possesses catalytic activity characteristic of cGB-PDE isolated from bovine tissue, thus verifying the identity of the cGB-8 cDNA clone as a cGB-PDE.

[0073] It is of interest to note that although the relative potencies of the PDE inhibitors for inhibition of cGMP hydrolysis were identical for the recombinant and bovine isolate cGB-PDE, the absolute IC₅₀ values for all inhibitors tested were 2-7 fold higher for the recombinant cGB-PDE. This difference could not be attributed to the effects of any factors present in COS-7 cell extracts on cGMP hydrolytic activity, since cGB-PDE isolated from bovine tissue exhibited identical kinetics of inhibition as a pure enzyme, or when added back to extracts of mock-transfected COS-7 cells. This apparent difference in pharmacological sensitivity may be due to a subtle difference in the structure of the recombinant cGB-PDE cDNA protein product and bovine lung cGB-PDE, such as a difference in post-translational modification at or near the catalytic-site. Alternatively, this difference may be due to an alteration of the catalytic activity of bovine lung cGB-PDE over several purification steps.

[0074] Cell extracts were assayed for [3 H]cGMP-binding activity in the absence or presence of 0.2 mM 3-isobutyl-1-methylxanthine (IBMX) (Sigma), a competitive inhibitor of cGMP hydrolysis. The cGMP binding assay, modified from the assay described in Thomas I, supra, was conducted in a total volume of 80 μ l. Sixty μ l of cell extract was combined with 20 μ l of a binding cocktail such that the final concentration of components of the mixture were 1 μ M [3 H]cGMP, 5 μ M cAMP, and 10 μ M 8-bromo-cGMP. The cAMP and 8-bromo-cGMP were added to block [3 H]cGMP binding to cAK and cGK, respectively. Assays were carried out in the absence and presence of 0.2 mM IBMX. The reaction was initiated by the addition of the cell extract, and was incubated for 60 minutes at 0° C. Filtration of the reaction mixtures was carried out as described in Thomas I, supra. Blanks were determined by parallel incubations with homogenization buffer replacing cell extracts, or with a 100-fold excess of unlabelled cGMP. Similar results were obtained with both methods. Total protein concentration of the cell extracts was determined by the method of Bradford, *Anal. Biochem.*, 72:248-254 (1976) using bovine serum albumin as the standard.

[0075] Results of the assay are set out in FIG. 7. When measured at 1 μ M [3 H]cGMP in the presence of 0.2 mM IBMX, extracts from COS-7 cells transfected with pCDM8-cGB-PDE exhibited 8-fold higher cGMP-binding activity than extracts from mock-transfected cells. No IBMX stimulation of background cGMP binding was observed suggesting that little or no endogenous cGB-PDE was present in the COS-7 cell extracts. In extracts of pCDM8-cGB-PDE transfected cells cGMP-specific activity was stimulated approximately 1.8-fold by the addition of 0.2 mM IBMX. The ability of IBMX to stimulate cGMP binding 2-5 fold is a distinctive property of the cGMP-binding phosphodiesterases.

[0076] Cell extracts were assayed as described above for [3 H]cGMP-binding activity (wherein concentration of [3 H]

cGMP was 2.5 μM) in the presence of excess unlabelled cAMP or cGMP. Results are presented in FIG. 8 wherein cGMP binding in the absence of unlabelled computer was taken 100% and each data point represents the average of three separate determinations. The binding activity of the problem product encoded by the cGB-PDE cDNA was specific for cGMP relative to cAMP. Less than 10-fold higher concentrations of unlabelled cGMP were required to inhibit [^3H]cGMP binding activity by 50% whereas approximately 100-fold higher concentrations of cAMP were required for the same degree of inhibition.

[0077] The results presented in this example show that the cGB-PDE cDNA encodes a phosphodiesterase which possesses biochemical activities characteristic of native cGB-PDE.

[0078] The catalytic domains of mammalian PDEs and a *Drosophila* PDE contain two tandem conserved sequences (HX₃HX₂₄₋₂₆E) that are typical Zn²⁺-binding motifs in Zn²⁺hydrolases such as thermolysin [Vallee and Auld, *Biochem.*, 29: 5647-5659 (1990)]. cGB-PDE binds Zn²⁺ in the presence of large excesses of Me²⁺, Mn²⁺, Fe²⁺, Fe²⁺, Ca²⁺ or Cd²⁺. In the absence of added, metal, cGB-PDE has a PDE activity that is approximately 20% of the maximum activity that occurs in the presence of 40 mM Mg²⁺, and this basal activity is inhibited by 1,10-phenanthroline or EDTA. This suggests that a trace metal(s) accounts for the basal PDE activity despite exhaustive treatments to remove metal(s), PDE activity is stimulated by addition of Zn²⁺ (0.02-1 μM) or Co²⁺ (1-20 μM), but not by Fe²⁺, Fe²⁺, Ca²⁺, Cd²⁺, or Cu²⁺. Zn²⁺ increases the basal PDE activity up to 70% of the maximum stimulation produced by 40 mM Me²⁺. The stimulatory effect of Zn²⁺ in these assays may be compromised by an inhibitory effect that is caused by Zn²⁺ concentrations >1 μM . The Zn²⁺-supported PDE activity and Zn²⁺ binding by cGB-PDE occur at similar concentrations of Zn²⁺. cGB-PDE thus appears to be a Zn²⁺hydrolase and Zn²⁺ appears to play a critical role in the activity of the enzyme. See, Colbran et al., *The FASEB J.*, 8: Abstract 2148 (Mar. 15, 1994).

EXAMPLES 6

[0079] Several human cDNA clones, homologous to the bovine cDNA clone encoding cGB-PDE, were isolated by hybridization under stringent conditions using a nucleic acid probe corresponding to a portion of the bovine cGB-8 clone (nucleotides 489-1312 of SEQ ID NO: 9).

[0080] Isolation of cDNA Fragments Encoding Human cGB-PDE

[0081] Three human cDNA libraries (two glioblastoma and one lung) in the vector lambda Zap were probed with the bovine cGB-PDE sequence. The PCR-generated clone corresponding to nucleotide 484-1312 of SEQ ID NO: 9 which is described in Example 1 was digested with EcoRI and SalI and the resulting 0.8 kb cDNA insert was isolated and purified by agarose gel electrophoresis. The fragment was labelled with radioactive nucleotides using a random primed DNA labelling kit (Boehringer).

[0082] The cDNA libraries were plated on 150 mm petri plates at a density of approximately 50,000 plaques per plate. Duplicate nitrocellulose filter replicas were prepared. The prehybridization buffer was 3 \times SSC, 0.1% sarkosyl, 10 \times

Denhardt's, 20 mM sodium phosphate (pH 6.8) and 50 $\mu\text{g/ml}$ salmon testis DNA. Prehybridization was carried out at 65 $^{\circ}$ C. for a minimum of 30 minutes. Hybridization was carried out at 65 $^{\circ}$ C. overnight in buffer of the same composition with the addition of 1 \times 10⁵ cpm/ml of probe. The filter were washed at 65 $^{\circ}$ C. in 2 \times SSC, 0.1% SDS. Hybridizing plaques were detected by autoradiography. The number of cDNAs that hybridized to the bovine probe and the number of cDNAs screened are indicated in Table 2 below.

TABLE 2

cDNA Library	Type	Positive Plaques	Plaques Screened
Human SW 1088 glioblastoma	dT-primed	1	1.5 \times 10 ⁴
Human lung	dT-primed	2	1.5 \times 10 ⁶
Human SW 1088 glioblastoma	dT-primed	4	1.5 \times 10 ⁶

[0083] Plasmids designated cgbS2.1, cgbS3.1, cgbL23.1, cgbL27.1 and cgbS27.1 were excised in vivo from the lambda Zap clones and sequenced.

[0084] Clone cgbS3.1 contains 2060 bp of a PDE open reading frame followed by a putative intron. Analysis of clone cgbS2.1 reveals that it corresponds to clone cgbS3.1 positions 664 to 2060 and extends the PDE open reading frame an additional 585 bp before reading into a putative intron. The sequence of the putative 5' untranslated region and the protein encoding portions of the cgbS2.1 and cgbS3.1 clones are set out in SEQ ID NOS: 11 and 12, respectively. Combining the two cDNAs yields a sequence containing approximately 2.7 kb of an open reading encoding a PDE. The three other cDNAs did not extend any further 5' or 3' than cDNA cgbS3.1 or cDNA cgbS2.1.

[0085] To isolate additional cDNAs, probes specific for the 5' end of clone cgbS3.1 and the 3' end of clone cgbS2.1 were prepared and used to screen a SW1088 glioblastoma cDNA library and a human aorta cDNA library. A 5' probe was derived from clone cgbS3.1 by PCR using the primers cgbS3.1S311 and cgbL23.1A1286 whose sequences are set out in SEQ ID NOS: 8 and 9, respectively, and below,

Primer cgb53.15311 (SEQ ID NO: 13)
5' GCCACCAGAGAAATGGTC 3'

Primer cgbL23.1A1286 (SEQ ID) NO: 14)
5' ACAATGGGTCTAAGAGGC 3'

[0086] The PCR reaction was carried out in a 50 μl reaction volume containing 50 pg cgbS3.1 cDNA, 0.2 mM dNTP, 10 $\mu\text{g/ml}$ each primer, 50 mM KCl, 10 mM Tris-HCl pH 8.2, 1.5 mM MgCl₂ and Taq polymerase. After an initial four minute denaturation at 94 $^{\circ}$ C., 30 cycles of one minute at 94 $^{\circ}$ C., two minutes at 50 $^{\circ}$ C. and four minutes at 72 $^{\circ}$ C. were carried out. An approximately 0.2 kb fragment was generated by the PCR reaction which corresponded to nucleotides 300-496 of clone cgbS3.1.

[0087] A 3' probe was derived from cDNA cgbS2.1 by PCR using the oligos cgbL3.1 S1190 and cgbS2.1A231 whose sequences are set out below.

Primer cgbL23.151190 (SEQ ID NO: 15)
5' TCAGTGCATGTTTCTGC 3'

Primer cgbS2.1A231 (SEQ ID NO: 16)
5' TACAACATGTTTCATCAG 3'

[0088] The PCR reaction as carried out similarly to that described above for generating the 5' probe, and yielded a fragment of approximately 0.8 kb corresponding to nucleotides 1358-2139 of cDNA cgbS2.1. The 3' 157 nucleotides of the PCR fragment (not shown in SEQ ID NO: 12) are within the presumptive intron.

[0089] The two PCR fragments were purified and isolated by agarose gel electrophoresis, and were labelled with radioactive nucleotides by random priming. A random-printed SW1088 glioblastoma cDNA library (1.5×10^6 plaques) was screened with the labelled fragments as described above, and 19 hybridizing plaques were isolated. An additional 50 hybridizing plaques were isolated from a human aorta cDNA library (dT and random primed, Clontech, Palo Alto, Calif.).

[0090] Plasmids were excised *in vivo* from some of the positive lambda Zap clones and sequenced. A clone designated cgbS53.2, the sequence of which is set out in SEQ ID NO: 17, contains an approximately 1.1 kb insert whose sequence overlaps the last 61 bp of cgbS3.1 and extends the open reading frame an additional 135 bp beyond that found in cgbS2.1. The clone contains a termination codon and approximately 0.3 kb of putative 3' untranslated sequence.

[0091] Generation of a Composite cDNA Encoding Human cGB-PDE

[0092] Clones cgbS3.1, cgbS2.1 and cgbS53.2 were used as described in the following paragraphs to build a composite cDNA that contained a complete human cGB-PDE opening reading frame. The composite cDNA is designated cgbmet156-2 and was inserted in the yeast ADH1 expression vector pBNI6N.

[0093] First, a plasmid designated cgb stop-2 was generated that contained the 3' end of the cGB-PDE open reading frame. A portion of the insert of the plasmid was generated by PCR using clone cgbS53.2 as a template. The PCR primers utilized were cgbS2.1S1700 and cgbstop-2.

Primer cgbS2.1S1700 (SEQ ID NO: 18)
5' TTGGAAGATCCTCATCA 3'

Primer cgbstop-2 (SEQ ID NO: 19)
5' ATGTCTCGAGTCAGTCCGCTTGGCCCTG 3'

[0094] The PCR reaction was carried out in 50 ul containing 50 pg template DNA, 0.2 mM dNTPs, 20 mM Tris-HCl pH 8.2, 10 mM KCl, 6 mM $(\text{NH}_4)_2\text{SO}_4$, 1.5 mM MgCl_2 , 0.1% Triton-X-100, 500 ng each primer and 0.5 units of Pfu polymerase (Stratagene). The reaction was heated to 94° C. for 4 minutes and then 30 cycles of 1 minute at 94° C., 2 minutes at 50° C. and four minutes at 72° C. were performed. The polymerase was added during the first cycle at 50° C. The resulting PCR product was phenol/chloroform extracted, chloroform extracted, ethanol precipi-

tated and cut with the restriction enzymes BclI and XhoI. The restriction fragment was purified on in agarose gel and eluted.

[0095] This fragment was ligated to the cDNA cgbS2.1 that had been grown in darn *E. coli*, cut with the restriction enzymes BclI and XhoI, and gel-purified using the Promega magic PCR kit. The resulting plasmid was sequenced to verify that cgbstop-2 contains the 3' portion of the cGB-PDE open reading frame.

[0096] Second, a plasmid carrying the 5' end of the human cGB-PDE open reading frame was generated. Its insert was generated by PCR using clone cgbS3.1 as a template. PCR was performed as described above using primers cgbmet156 and cgbS2.1A2150.

Primer cgbmet156 (SEQ ID NO: 20)
5' TACAGAATICTGACCATGGAGCGGCCCGC 3'

Primer cgbS2.1A2150 (SEQ ID NO: 21)
5' CATTCTAAGCGGATACAG 3'

[0097] The resulting PCR fragment was phenol/chloroform extracted, chloroform extracted, ethanol precipitated and purified on a Sepharose CL-6B column. The fragment was cut with the restriction enzymes EcoRV and EoRI, run on an agarose gel and purified by spinning through glass wool. Following phenol/chloroform extraction, chloroform extraction and ethanol precipitation, the fragment was ligated into EcoRI/EcoRV digested BluescriptII SK(+) to generate plasmid cgbmet156. The DNA sequence of the insert and junctions was determined. The insert contains a new EcoRI site and an additional 5 nucleotides that together replace the original 155 nucleotides 5' of the initiation codon. The insert extends to an EcoRV site beginning 531 nucleotides from the initiation codon.

[0098] The 5' and 3' portions of the cGB-PDE open reading frame were then assembled in vector pBNI6a. The vector pBNI6a was cut with EcoRI and XhoI, isolated from a gel and combined with the agarose gel purified EcoRI/EcoRV fragment from cgbmet156 and the agarose gel purified EcoRV/XhoI fragment from cgbstop-2. The junctions of the insert were sequenced and the construct was named hcbmet15-2 6a.

[0099] The cGB-PDE insert from hcbmet15-2 6a was then moved into the expression vector pBNI6n. Expression of DNA inserted in this vector is directed from the yeast ADH1 promoter and terminator. The vector contains the yeast 2 micron origin of replication, the pUC19 origin of replication and an ampicillin resistance gene. Vector pBNI6n was cut with EcoRI and XhoI and gel-purified. The EcoRI/XhoI insert from hcbmet156-2 6a was gel purified using Promega magic PCR construct, and ligated into the cut pBNI6n. All new junctions in the resulting construct, hcbmet156-2 6n, were sequenced. The DNA and deduced amino acid sequences of the insert of hcbmet156-2 6n which encodes a composite human cGB-PDE is set out in SEQ ID NOs: 22 and 23. The insert extends from the first methionine in clone cgbS3.1 (nucleotide 156) to the stop codon (nucleotide 2781) in the composite cDNA. Because the methionine is the most 5' methionine in clone cgbS3.1

and because there are no stop codons in frame with the methionine and upstream of it, the insert in pBNY6n may represent a truncated form of the open reading frame.

[0100] Variant cDNAs

[0101] Four human cGB-PDE cDNAs that are different from the hcgmet156-2 6n composite cDNA have been isolated. One cDNA, cgbL23.1, is missing an internal region of hcgmet156-2 6n (nucleotides 997-1000 to 1444-1447). The exact end points of the deletion cannot be determined from the cDNA sequence at the positions. Three of the four variant cDNAs have 5' end sequences that diverge from the hcgmet156-2 6n sequence upstream of nucleotide 151 (cDNAs cgbA7f, cgbA5C, cgb12). These cDNAs presumably represent alternatively spliced or unspliced mRNAs.

EXAMPLE 7

[0102] The composite human cGB-PDE cDNA construct, hcgmet156-2 6n, was transformed into the yeast strain YKS45 (ATCC 74225) (MAT α his3 trp1 ura3 leu3 pde::HIS3 pde2::TRP1) in which two endogenous PDE genes are deleted. Transformants complementing the leu deficiency of the YKS45 strain were selected and assayed for cGB-PDE activity. Extracts from cells bearing the plasmid hcgmet156-2 6n were determined to display cyclic GMP-specific phosphodiesterase activity by the assay described below.

[0103] One liter of YKS45 cells transformed with the plasmid cgbmet156-2 6n and grown in SC-leu medium to a density of $1-2 \times 10^7$ cells/ml was harvested by centrifugation, washed once with deionized water, frozen in dry ice/ethanol and stored at -70° C. Cell pellets (1-1.5 ml) were thawed on ice in the presence of an equal volume of 25 mM Tris-Cl (pH 8.0)/5 mM EDTA/5 mM EGTA/1 mM o-phenanthroline/0.5 mM AEBSE (Calbiochem)/0.1% β -mercaptoethanol and 10 μ g/ml each of aprotinin, leupeptin, and pepstatin A. The thawed cells were added to 2 ml of acid-washed glass beads (425-600 μ m, Sigma) in 15 ml Corex tube. Cells were broken with 4 cycle consisting of a 30 second vortexing on setting 1 followed by a 60 second incubation on ice. The cell lysate was centrifuged at 12,000 \times g for 10 minutes and the supernatant was passed through a 0.8 μ filter. The supernatant was assayed for cGMP PDE activity as follows. Samples were incubated for 20 minutes at 30° C. in the presence of 45 mM Tris-Cl (pH 8.0), 2 mM EGTA, 1 mM EDTA, 0.2 mg/ml BSA, 5 mM MgCl₂, 0.2 mM o-phenanthroline, 2 μ g/ml each of pepstatin A, leupeptin, and aprotinin, 0.1 mM AEBSE, 0.02% β -mercaptoethanol and 0.1 mM [³H]cGMP as substrate. [¹⁴C]-AMP (0.5 nCi/assay) was added as a recovery standard. The reaction was terminated with stop buffer (0.1M ethanolamine pH 9.0, 0.5M ammonium sulfate, 10 nM EDTA, 0.05% SDS final concentration). The product was separated from the cyclic nucleotide substrate by chromatography on BioRad Affi-Gel 601. The sample was applied to a column containing approximately 0.25 ml of Affi-Gel 601 equilibrated in column buffer (0.1M ethanolamine pH 9.0 containing 0.5M ammonium sulfate). The column was washed five times with 0.5 ml of column buffer. The product was eluted with four 0.5 ml aliquots of 0.25 acetic acid and mixed with 5 ml Ecolume (ICN Biochemicals). The radioactive product was measured by scintillation counting.

EXAMPLE 8

[0104] Analysis of expression of cGB-PDE mRNA in human tissues was carried out by RNase protection assay.

[0105] A probe corresponding to a portion of the putative cGMP binding domain of cGB-PDE (402 bp corresponding to nucleotides 1450 through 1851 of SEQ ID NO: 13) was generated by PCR. The PCR fragment was inserted into the EcoRI site of the plasmid pBSII SK(-) to generate the plasmid RP3. RP3 plasmid DNA was linearized with XbaI and antisense probes were generated by a modification of the Stratagene T7 RNA polymerase kit. Twenty-five ng of linearized plasmid was combined with 20 microcuries of alpha ³²rUTP (800 Ci/mmol, 10 mCi/ml), 1 \times transcription buffer (40 mM Tris Cl, pH 8, 8 mM MgCl₂, 2 mM spermidine, 50 mM NaCl), 0.25 mM each rATP, rGTP and rCTP, 0.1 units of RNase Block II, 5 mM DTT, 8 μ M rUTP and 5 units of T7 RNA Polymerase in a total volume of 5 μ l. The reaction was allowed to proceed 1 hour at room temperature and then the DNA template was removed by digestion with RNase free DNase. The reaction was diluted into 100 μ l of 40 mM TrisCl, pH 8, 6 mM MgCl₂ and 10 mM NaCl. Five units of RNase-free DNase were added and the reaction was allowed to continue another 15 minutes at 37° C. The reaction was stopped by a phenol extraction followed by a phenol chloroform extraction. One half volume of 7.5M NH₄OAc was added and the probe was ethanol precipitated.

[0106] The RNase protection assays were carried out using the Ambion RNase Protection kit (Austin, Tex.) and 10 μ g RNA isolated from human tissues by an acid guanidinium extraction method. Expression of cGB-PDE mRNA was easily detected in RNA extracted from skeletal muscle, uterus, bronchus, skin, right saphenous vein, aorta and SW1018 glioblastoma cells. Barely detectable expression was found in RNA extracted from right atrium, right ventricle, kidney cortex, and kidney medulla. Only complete protection of the RP3 probe was seen. The lack of particle protection argues against the cDNA cgbL23.1 (a variant cDNA described in Example 7) representing a major transcript, at least in these RNA samples.

EXAMPLE 9

[0107] Polyclonal antisera was raised to *E. coli*-produced fragments of the human cGB-PDE.

[0108] A portion of the human cGB-PDE cDNA (nucleotides 1668-2612 of SEQ ID NO: 22, amino acids 515-819 of SEQ ID NO: 23) was amplified by PCR and inserted into the *E. coli* expression vector pGEX2T (Pharmacia) as a BamHI/EcoRI fragment. The pGEX2T plasmid carries an ampicillin resistance gene, an *E. coli* laq I^a gene and a portion of the *Schistosoma japonicum* glutathione-S-transferase (GST) gene. DNA inserted in the plasmid can be expressed as a fusion protein with GST and can then be cleaved from the GST portion of the protein with thrombin. The resulting plasmid, designated cgbPE3, was transformed into *E. coli* strain LE392 (Stratagene). Transformed cells were grown at 37° C. to an OD₆₀₀ of 0.6. IPTG (isopropylthioalactopyranoside) was added to 0.1 mM and the cells were grown at 37° C. for an additional 2 hours. The cells were collected by centrifugation and lysed by sonication. Cell debris was removed by centrifugation and the supernatant was fractionated by SDS-PAGE. The gel was stained with cold 0.4M KCl and the GST-cgb fusion protein band

was excised and electrocuted. The PDE portion of the protein was separated from the GST portion by digestion with thrombin. The digest was fractionated by SDS-PAGE, the PDE protein was electroeluted and injected subcutaneously into a rabbit. The resultant antisera recognizes both the bovine cGB-PDE fragment that was utilized as antigen and the full length human cGB-PDE protein expressed in yeast (see Example 8).

EXAMPLE 10

[0109] Polynucleotides encoding various cGB-PDE analogs and cGB-PDE fragments were generated by standard methods.

[0110] A. cGB-PDE Analogs

[0111] All known cGMP-binding PDEs contain two internally homologous tandem repeats within their putative cGMP-binding domain. In the bovine cGB-PDE of the invention, the repeats span at least residues 228-311 (repeat A) and 410-500 (repeat B) of SEQ ID NO: 10. Site-directed mutagenesis of an aspartic acid that is conserved in repeats A and B of all known cGMP-binding PDEs was used to create analogs of cGB-PDE having either Asp-289 replaced with Ala (D289A) or Asp-478 replaced with Ala (D478A). Recombinant wild type (WT) bovine and mutant bovine cGB-PDEs were expressed in COS-7 cells cGB-PDE purified from bovine lung (native cGB-PDE) and WT cGB-PDE displayed identical cGMP-binding kinetics with a K_d of approximately 2 μ M and a curvilinear dissociation profile ($t=1.3$ hours at 4° C.). This curvilinearity may have been due to the presence of distinct high affinity (slow) and low affinity (fast) sites of cGMP binding. The D289A mutant had significantly decreased affinity for cGMP ($K_d > 20 \mu$ M) and a single rate of cGMP-association ($t_{\text{[text missing or illegible when filed]}}=0.5$ hours), that was similar to that calculated for the fast site of WT and native cGB-PDE. This suggested the loss of a slow cGMP-binding site in repeat A of this mutant. Conversely, the D478A mutant showed higher affinity for cGMP (K_d of approximately 0.5 μ M) and a single cGMP-dissociation rate ($t_{\text{[text missing or illegible when filed]}}=2.8$ hours) that was similar to the calculated rate of the slow site of WT and native cGB-PDE. This suggested the loss of a fast site when repeat was modified. These results indicate that dimeric cGB-PDE possesses two homologous but kinetically distinct cGMP-binding sites, with the conserved aspartic acid being critical for interaction with cGMP at each site. See, Colbran et al., *FASEB J.*, 8: Abstract 2149 (May 15, 1994).

[0112] B. Amino-Terminal Truncated cGB-PDE Polypeptides

[0113] A truncated human cGB-PDE polypeptide including amino acids 516-875 of SEQ ID NO: 23 was expressed in yeast. A cDNA insert extending from the NcoI site at nucleotide 1555 of SEQ ID NO: 22 through the XhoI site at the 3' end of SEQ ID NO: 22 was inserted into the ADH2 yeast expression vector YEpC-PADH2d [Price et al., *Meth. Enzymol.*, 118: 308-318 (1990)] that had been digested with NcoI and Sall to generate plasmid YEpC-PADH2d HcGB. The plasmid was transformed into spheroplasts of the yeast strain yBJ2-54 (prc1-407 prb1-1122 pep4-3 leu2 trp1 ura3-52 Δ pde1::URA3, HIS3 Δ pdc::TRP1 cir). The endogenous PDE genes are deleted in this strain. Cells were grown in SC-leu media with 2% glucose to 10^7 cells/ml, collected by filtration and grown 24 hours in YEP media containing 3% glycerol. Cells were pelleted by centrifugation and stored frozen. Cells were disrupted with glass beads and the cell

homogenate was assayed for phosphodiesterase activity essentially as described in Prpic et al., *Anal. Biochem.*, 208: 155-160 (1993). The truncated human cGB-PDE polypeptide exhibited phosphodiesterase activity.

[0114] C. Carboxy-Terminal Truncated cGB-PDE Polypeptides

[0115] Two different plasmids encoding carboxy-terminal truncated human cGB-PDE polypeptides were constructed.

[0116] Plasmid pBJ6-84Hin contains a cDNA encoding amino acids 1-494 of SEQ ID NO: 23 inserted into the NcoI and Sall sites of vector YEpC-PADH2d. The cDNA insert extends from the NcoI site at nucleotide position 10 of SEQ ID NO: 22 through the HindIII site at nucleotide position 1494 of SEQ ID NO: 22 followed by a linker and the Sall site of YEpC-PADH2d.

[0117] Plasmid pBJ6-84Ban contains a cDNA encoding amino acids 1-549 of SEQ ID NO: 23 inserted into the NcoI and Sall sites of vector YEpC-PADH2d. The cDNA insert extends from the NcoI site at nucleotide position 10 of SEQ ID NO: 22 through the BanI site at nucleotide position 1657 of SEQ ID NO: 22 followed by a linker and the Sall site of YEpC-PADH2d.

[0118] The truncated cGB-PDE polypeptides are useful for screening for modulators of cGB-PDE activity.

EXAMPLE 11

[0119] Monoclonal antibodies reactive with human cGB-PDE were generated.

[0120] Yeast yB12-54 containing the plasmid YEpADH2 HcGB (Example 10B) were fermented in a New Brunswick Scientific 10 liter Microferm. The cGB-PDE cDNA insert in plasmid YEpADH2 HcGB extends from the NcoI site at nucleotide 12 of SEQ ID NO: 22 to the XhoI site at the 3' end of SEQ ID NO: 22. An inoculum of 4×10^8 $t_{\text{[text missing or illegible when filed]}}$ cells was added to 8 liters of media containing SC-leu, 5% glucose, trace metals, and trace vitamins. Fermentation was maintained at 26° C., agitated at 600 rpm with the standard microbial impeller, and aerated with compressed air at 10 volumes per minute. When glucose done to 0.3% at 24 hours post-inoculation the culture was infused with 2 liters of 5xYEP media containing 15% glycerol. At 66 hours post-inoculation the yeast from the ferment was harvested by centrifugation at 4,000xg for 30 minutes at 4° C. Total yield of biomass from this fermentation approached 350 g wet weight.

[0121] Human cGB-PDE enzyme was purified from the yeast cell pellet. Assays for PDE activity using 1 mM cGMP as substituted was employed to follow the chromatography of the enzyme. All chromatographic manipulations were performed at 4° C.

[0122] Yeast (29 g wet weight) were resuspended in 70 ml of buffer A (25 mM Tris pH 8.0, 0.25 mM DTT, 5 mM $MgCl_2$, 10 μ M $ZnSO_4$, 1 mM benzamide) and lysed by passing through a microfluidizer at 22-24,000 psi. The lysate was centrifuged at 10,000xg for 30 minutes and the supernatant was applied to a 2.6x28 cm column containing Pharmacia Fast Flow Q anion exchange resin equilibrated with buffer B containing 20 mM BisTris-propane pH 6.8, 0.25 mM DTT, 1 mM $MgCl_2$, and 10 μ M $ZnSO_4$. The column was washed with 5 column volumes of buffer B containing 0.125M NaCl and then developed with a linear gradient from 0.125 to 1.0M NaCl. Fractions containing the enzyme were pooled and applied directly to a 5x20 cm column of ceramic hydroxyapatite (BioRad) equilibrated in buffer C containing 20 mM BisTris-propane pH 6.8, 0.25 mM DTT, 0.25 MKCl, 1 mM $MgCl_2$, and 10 μ M $ZnSO_4$.

The column was washed with 5 column volumes of buffer C and eluted with a linear gradient from 0 to 250 mM potassium phosphate in buffer C. The pooled enzyme was concentrated 8-fold by ultrafiltration (YM30 membrane, Amicon). The concentrated enzyme was chromatographed on a 2.6×90 cm column of Pharmacia Sephacryl S300 (Piscataway, N.J.) equilibrated in 25 mM BisTris-propane pH 6.8, 0.25 mM DTT, 0.25M NaCl, 1 mM MgCl₂, and 20 μM ZnSO₄. Approximately 4 mg of protein was obtained. The recombinant human cGB-PDE enzyme accounted for approximately 90% of protein obtained as judged by SDS polyacrylamide gel electrophoresis followed by Coomassie blue staining.

[0123] The purified protein was used as an antigen to raise monoclonal antibodies. Each of 19 week old Balb/c mice (Charles River Biotechnical Services, Inc., Wilmington, Mass.) was immunized sub-cutaneously with 50 ug purified human cGB-PDE enzyme in a 200 ul emulsion consisting of 50% Freund's complete adjuvant (Sigma Chemical Co.). Subsequent boosts on day 20 and day 43 were administered in incomplete Freund's adjuvant. A pre-fusion boost was done on day 86 using 50 ug enzyme in FBS. The fusion was performed on day 90.

[0124] The spleen from mouse #1817 was removed sterilely and placed in 10 ml serum free RPMI 1640. A single-cell suspension was formed and filtered through sterile 70 mesh Nitex cell strainer (Becton Dickinson, Parsippany, N.J.), and washed twice by centrifuging at 200 g for 5 minutes and resuspending the pellet in 20 ml serum free RPMI. Thymocytes taken from 3 naive Balb/c mice were prepared in a similar manner.

[0125] NS-1 myeloma cells, kept in log phase in RPMI with 11% Fetalclone (FBS) Hyclone Laboratories, Inc., Logan, Utah) for three days prior to fusion, were centrifuged 200 g for 5 minutes, and the pellet was washed twice as described in the foregoing paragraph. After washing each cell suspension was brought to a final volume of 10 ml in serum free RPMI, and 20 μl was diluted 1:50 in 1 ml serum free RPMI: 20 μl of each dilution was removed, mixed with 20 μl 0.4% trypan blue sun in 0.85% saline (Gibco), loaded onto a hemocytometer (Baxter Healthcare Corp., Deerfield, Ill.) and counted.

[0126] Two×10⁷[text missing or illegible when filed]spleen cells were combined with 4.0×10⁷ NS-1 cells, centrifuged and the supernatant was aspirated. The cell pellet was dislodged by tapping the tube and 2 ml of 37° C. PEG 1500 (50% in 75 mM Hepes, pH 8.0) (Boehringer Mannheim) was added with stirring over the course of 1 minute, followed by adding 14 ml of serum free RPMI over 7 minutes. An additional 16 ml RPMI was added and the cells were centrifuged at 200 g for 10 minutes. After discarding the supernatant, the pellet was resuspended in 200 ml RPMI containing 15% FBS, 100 μM sodium hypoxanthine, 0.4 μM aminopterin, 16 μM thymidine (HAT) (Gibco), 25 units/ml IL-6 (Boehringer Mannheim) and 1.5×10⁷[text missing or illegible when filed]thymocytes/ml. The suspension was first placed in a T225 flask (Corning, United Kingdom) at 37° C. for two hours before being dispensed into ten 96 well flat bottom tissue culture plates (Corning, United Kingdom) at 200 μl/well. Cells in plates were fed on days 3, 4, 5 post fusion day by aspirating approximately 100 μl from each well with a 20 G needle (Becton Dickinson), and adding 100 μl/well plating medium described above except containing 10 units/ml IL-6 and lacking thymocytes.

[0127] The fusion was screened initially by ELISA. Immulon 4 plates (Dynatech) were coated at 4° C. overnight

with purified recombinant human cGB-PDE enzyme (100 ng/well in 50 mM carbonate buffer pH9.6). The plates were washed 3× with PBS containing 0.05% Tween 20 (PBST). The supernatants from the individual hybridoma wells were added to the enzyme coated wells (50 μl/well). After incubation at 37° C. for 30 minutes, and washing as above, 50 μl of horseradish peroxidase conjugated goat anti-mouse IgG(fc) (Jackson ImmunoResearch, West Grove, Pa.) diluted 1:3500 in PBST was added. Plates were incubated as above, washed 4× with PBST and 100 μl substrate consisting of 1 mg/ml o-phenylene diamine (Sigma) and 0.1 μl/ml 30% H₂O₂ in 100 mM citrate, pH 4.5, was added. The color reaction was stopped in 5 minutes with the addition of 50 μl of 15% H₂SO₄. A₄₉₀ was read on a plate reader (Dynatech).

[0128] Wells C5G, [text missing or illegible when filed]4D, F1G, F9H, F11G, J4A, and J5D were picked and renamed 102A, 102B, 102C, 102D, 102E, 102F, and 102G respectively, cloned two or three times, successively, by doubling dilution in RPMI, 15% FBS, 100 μM sodium hypoxanthine, 16 μM thymidine, and 10 units/ml IL-6. Wells of clone plates were scored visually after 4 days and the number of colonies in the least dense wells were recorded. Selected wells of the each cloning were tested by ELISA.

[0129] The monoclonal antibodies produced by above hybridomas were isotyped in an ELISA assay. Results showed that monoclonal antibodies 102A to 102E were IgG1, 102F was IgG2b and 102G was IgG2a.

[0130] All seven monoclonal antibodies reacted with human cGS-PDE as determined by Western analysis.

EXAMPLE 12

[0131] Developing modulators of the biological activities of specific PDEs requires differentiating PDE isozymes present in a particular assay preparation. The classical enzymological approach of isolating PDEs from natural tissue sources and studying each new isozyme is hampered by the limits of purification techniques and the inability to definitively assess whether complete resolution of a isozyme has been achieved. Another approach has been to identify assay conditions which might favor the contribution of one isozyme and minimize the contribution of others in a preparation. Still another approach has been the separation of PDEs by immunological means. Each of the foregoing approaches for differentiating PDE isozymes is time consuming and technically difficult. As a result many attempts to develop selective PDE modulators have been performed with preparations containing more than one isozyme. Moreover, PDE preparations from natural tissue sources are susceptible to limited proteolysis and may contain mixtures of active proteolytic products that have different kinetic, regulatory and physiological properties than the full length PDEs.

[0132] Recombinant cGB-PDE polypeptide products of the invention greatly facilitate the development of new and specific cGB-PDE modulators. The use of human recombinant enzymes for screening for modulators has many inherent advantages. The need for purification of an isozyme can be avoided by expressing it recombinantly in a host cell that lacks endogenous phosphodiesterase activity (e.g. yeast swain YKS45 deposited as ATCC 74225). Screening compounds against human protein avoids complications that often arise from screening against non-human protein where a compound optimized on a non-human protein may fail to be specific for or react with the human protein. For example, a single amino acid difference between the human and rodent 5HT_{1B} serotonin receptors accounts for the difference in binding of a compound to the receptors [See Oskenberg

et al., *Nature*, 360: 161-163 (1992)]. Once a compound that modulates the activity of the cGB-PDE is discovered, its selectivity can be evaluated by comparing its activity on the cGB-PDE to its activity on other PDE isozymes. Thus, the combination of the recombinant cGB-PDE products of the invention with other recombinant PDE products in a series of independent assays provides a system for developing selective modulators of cGB-PDE. Selective modulators many include, for example, antibodies and other proteins or peptides which specifically bind to the cGB-PDE or cGB-PDE nucleic acid, oligonucleotides which specifically bind to the cGB-PDE (so Patent Cooperation Treaty International Publication No. WO93/05182 published Mar. 18, 1993 which describes methods for selecting oligonucleotides which selectively bind to target biomolecules) or cGB-PDE nucleic acid (e.g., antisense oligonucleotides) and other non-peptide natural or synthetic compounds which specifically bind to the cGB-PDE or cGB-PDE nucleic acid. Mutant forms of the cGB-PDE which alter the enzymatic activity of the cGB-PDE or its localization in a cell are also contemplated. Crystallization of recombinant cGB-PDE

alone and bound to a modulator, analysis of atomic structure by X-ray crystallography, and computer modulating of those structures are methods useful for designing and optimizing non-peptide selective modulators. See, for example, Erickson et al., *Ann. Rep. Med. Chem.* 27: 271-289 (1992) for a general review of structure-based drug design.

[0133] Targets for the development of selective modulators include, for example: (1) the regions of the cGB-PDE which contact other proteins and/or localize the cGB-PDE within a cell, (2) the regions of the cGB-PDE which bind substrate, (3) the allosteric cGMP-binding site(s) of cGB-PDE, (4) the metal-binding regions of the cGB-PDE, (5) the phosphorylation site(s) of cGB-PDE and (6) the regions of the cGB-PDE which are involved in dimerization of cGB-PDE subunits.

[0134] While the present invention has been described in terms of specific embodiments, it is understood that variations and modifications will occur to those skilled in the art. Accordingly, only such limitations as appear in the appended claims should be placed on the invention.

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gaa ctt ata atg aaa aat caa ttc aat ttg gaa gat cct cat caa aag Glu Leu Ile Met Lys Asn Gln Phe Asn Leu Glu Asp Pro His Gln Lys 730 735 740			2324
gag ttg ttt tta gcg atg ctg atg aca gct tgt gat ctt tct gca att Glu Leu Phe Leu Ala Met Leu Met Thr Ala Cys Asp Leu Ser Ala Ile 745 750 755			2372
aca aaa ccc tgg cct att caa caa cgg ata gca gaa ctt gtt gcc act Thr Lys Pro Trp Pro Ile Gln Gln Arg Ile Ala Glu Leu Val Ala Thr 760 765 770			2420
gaa ttt ttt gac caa gga gat aga gag agg aaa gaa ctc aac ata gag Glu Phe Phe Asp Gln Gly Asp Arg Glu Arg Lys Glu Leu Asn Ile Glu 775 780 785 790			2468
ccc gct gat cta atg aac cgg gag aag aaa aac aaa atc cca agt atg Pro Ala Asp Leu Met Asn Arg Glu Lys Lys Asn Lys Ile Pro Ser Met 795 800 805			2516
caa gtt gga ttc ata gat gcc atc tgc ttg caa ctg tat gag gcc ttg Gln Val Gly Phe Ile Asp Ala Ile Cys Leu Gln Leu Tyr Glu Ala Leu			2564

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810	815	820	
acc cat gtg tcg gag gac tgt ttc cct ttg ctg gac ggc tgc aga aag			2612
Thr His Val Ser Glu Asp Cys Phe Pro Leu Leu Asp Gly Cys Arg Lys			
825	830	835	
aac agg cag aaa tgg cag gct ctt gca gaa cag cag gag aag aca ctg			2660
Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu Gln Gln Glu Lys Thr Leu			
840	845	850	
atc aat ggt gaa agc agc cag acc aac cga cag caa cgg aat tcc gtt			2708
Ile Asn Gly Glu Ser Ser Gln Thr Asn Arg Gln Gln Arg Asn Ser Val			
855	860	865	870
gct gtc ggg aca gtg tagccagggtg tatcagatga gtgagtgtgt gctcagctca			2763
Ala Val Gly Thr Val			
875			
gtcctctgca acaccatgaa gctaggcatt ccagcttaat tcctgcagtt gactttaaaa			2823
aactggcata aagcactagt cagcatctag ttctagcttg accagtgaag agtagaacac			2883
caccacagtc aggggtcaga gcagttggca gtctcctttc gaaccagac tgggaattt			2943
aaagaagagc agtcgtcgtt tatactctctg tcttttccta agcggggtgt ggaatctcta			3003
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cactgtgctg tgacctctca atctgagaaa cgtgtaagga aggtttcagc gaattccctt			3123
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aattttccaa gatgatacac tcctccctag tctaggggtc agaccctgta tggtggtgt			3423
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attatcttgg cagtttttct aatgacttg cacagacttc tcctgtactt catggctgtg			3843
cagtgttcca tgctgtgagg gcaccatcgt gtattaaatc agttccctgg tcacacatag			3903
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aaaaaatcaa actatagttt atttacaatg ttgtgttaat ttcgggtgta cagcaaagt			4263
actcagtggt caagtacatt taaaacactg ggcatactct ctccctctcc ttgtgtacct			4323
ggttggtatt tccagaaacc atgctcttgt ctgtcctgta gttttggaag cgctttctct			4383
ttgaagactg ccttctctcc tctgtctgcc ctacatggac tagttcgttt attgtcctac			4443
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<210> SEQ ID NO 10
<211> LENGTH: 875
<212> TYPE: PRT
<213> ORGANISM: bovine

<400> SEQUENCE: 10

Met Glu Arg Ala Gly Pro Gly Cys Arg Ala Ala Ala Thr Ala Met Gly
1          5          10          15
Pro Gly Leu Gly Arg Ser Val Ala Gly Arg Ser Leu Gly Leu Tyr Leu
20          25          30
Leu Tyr Phe Val Arg Lys Gly Thr Arg Glu Met Val Asn Ala Trp Phe
35          40          45
Ala Glu Arg Val His Thr Ile Pro Val Cys Lys Glu Gly Ile Lys Gly
50          55          60
His Thr Glu Ser Cys Ser Cys Pro Leu Gln Pro Ser Pro Arg Ala Glu
65          70          75          80
Ser Ser Val Pro Gly Thr Pro Thr Arg Lys Ile Ser Ala Ser Glu Phe
85          90          95
Asp Arg Pro Leu Arg Pro Ile Val Ile Lys Asp Ser Glu Gly Thr Val
100         105         110
Ser Phe Leu Ser Asp Ser Asp Lys Lys Glu Gln Met Pro Leu Thr Ser
115         120         125
Pro Arg Phe Asp Asn Asp Glu Gly Asp Gln Cys Ser Arg Leu Leu Glu
130         135         140
Leu Val Lys Asp Ile Ser Ser His Leu Asp Val Thr Ala Leu Cys His
145         150         155         160
Lys Ile Phe Leu His Ile His Gly Leu Ile Ser Ala Asp Arg Tyr Ser
165         170         175
Leu Phe Leu Val Cys Glu Asp Ser Ser Asn Asp Lys Phe Leu Ile Ser
180         185         190
Arg Leu Phe Asp Val Ala Glu Gly Ser Thr Leu Glu Glu Ala Ser Asn
195         200         205
Asn Cys Ile Arg Leu Glu Trp Asn Lys Gly Ile Val Gly His Val Ala
210         215         220
Ala Phe Gly Glu Pro Leu Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg
225         230         235         240
Phe Asn Ala Glu Val Asp Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile
245         250         255
Leu Cys Met Pro Ile Lys Asn His Arg Glu Glu Val Val Gly Val Ala
260         265         270
Gln Ala Ile Asn Lys Lys Ser Gly Asn Gly Gly Thr Phe Thr Glu Lys
275         280         285
Asp Glu Lys Asp Phe Ala Ala Tyr Leu Ala Phe Cys Gly Ile Val Leu
290         295         300
His Asn Ala Gln Leu Tyr Glu Thr Ser Leu Leu Glu Asn Lys Arg Asn
305         310         315         320
Gln Val Leu Leu Asp Leu Ala Ser Leu Ile Phe Glu Glu Gln Gln Ser
325         330         335
Leu Glu Val Ile Leu Lys Lys Ile Ala Ala Thr Ile Ile Ser Phe Met
340         345         350
Gln Val Gln Lys Cys Thr Ile Phe Ile Val Asp Glu Asp Cys Ser Asp
355         360         365

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Ser Phe Ser Ser Val Phe His Met Glu Cys Glu Glu Leu Glu Lys Ser
 370 375 380

Ser Asp Thr Leu Thr Arg Glu Arg Asp Ala Asn Arg Ile Asn Tyr Met
 385 390 395 400

Tyr Ala Gln Tyr Val Lys Asn Thr Met Glu Pro Leu Asn Ile Pro Asp
 405 410 415

Val Ser Lys Asp Lys Arg Phe Pro Trp Thr Asn Glu Asn Met Gly Asn
 420 425 430

Ile Asn Gln Gln Cys Ile Arg Ser Leu Leu Cys Thr Pro Ile Lys Asn
 435 440 445

Gly Lys Lys Asn Lys Val Ile Gly Val Cys Gln Leu Val Asn Lys Met
 450 455 460

Glu Glu Thr Thr Gly Lys Val Lys Ala Phe Asn Arg Asn Asp Glu Gln
 465 470 475 480

Phe Leu Glu Ala Phe Val Ile Phe Cys Gly Leu Gly Ile Gln Asn Thr
 485 490 495

Gln Met Tyr Glu Ala Val Glu Arg Ala Met Ala Lys Gln Met Val Thr
 500 505 510

Leu Glu Val Leu Ser Tyr His Ala Ser Ala Ala Glu Glu Glu Thr Arg
 515 520 525

Glu Leu Gln Ser Leu Ala Ala Val Val Pro Ser Ala Gln Thr Leu
 530 535 540

Lys Ile Thr Asp Phe Ser Phe Ser Asp Phe Glu Leu Ser Asp Leu Glu
 545 550 555 560

Thr Ala Leu Cys Thr Ile Arg Met Phe Thr Asp Leu Asn Leu Val Gln
 565 570 575

Asn Phe Gln Met Lys His Glu Val Leu Cys Lys Trp Ile Leu Ser Val
 580 585 590

Lys Lys Asn Tyr Arg Lys Asn Val Ala Tyr His Asn Trp Arg His Ala
 595 600 605

Phe Asn Thr Ala Gln Cys Met Phe Ala Ala Leu Lys Ala Gly Lys Ile
 610 615 620

Gln Lys Arg Leu Thr Asp Leu Glu Ile Leu Ala Leu Leu Ile Ala Ala
 625 630 635 640

Leu Ser His Asp Leu Asp His Arg Gly Val Asn Asn Ser Tyr Ile Gln
 645 650 655

Arg Ser Glu His Pro Leu Ala Gln Leu Tyr Cys His Ser Ile Met Glu
 660 665 670

His His His Phe Asp Gln Cys Leu Met Ile Leu Asn Ser Pro Gly Asn
 675 680 685

Gln Ile Leu Ser Gly Leu Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys
 690 695 700

Ile Ile Lys Gln Ala Ile Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys
 705 710 715 720

Arg Arg Gly Glu Phe Phe Glu Leu Ile Met Lys Asn Gln Phe Asn Leu
 725 730 735

Glu Asp Pro His Gln Lys Glu Leu Phe Leu Ala Met Leu Met Thr Ala
 740 745 750

Cys Asp Leu Ser Ala Ile Thr Lys Pro Trp Pro Ile Gln Gln Arg Ile
 755 760 765

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Ala Glu Leu Val Ala Thr Glu Phe Phe Asp Gln Gly Asp Arg Glu Arg
 770 775 780

Lys Glu Leu Asn Ile Glu Pro Ala Asp Leu Met Asn Arg Glu Lys Lys
 785 790 795 800

Asn Lys Ile Pro Ser Met Gln Val Gly Phe Ile Asp Ala Ile Cys Leu
 805 810 815

Gln Leu Tyr Glu Ala Leu Thr His Val Ser Glu Asp Cys Phe Pro Leu
 820 825 830

Leu Asp Gly Cys Arg Lys Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu
 835 840 845

Gln Gln Glu Lys Thr Leu Ile Asn Gly Glu Ser Ser Gln Thr Asn Arg
 850 855 860

Gln Gln Arg Asn Ser Val Ala Val Gly Thr Val
 865 870 875

<210> SEQ ID NO 11

<211> LENGTH: 2060

<212> TYPE: DNA

<213> ORGANISM: human cDNA

<400> SEQUENCE: 11

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gcggccgcgc tccggccgct ttgtcgaag cgggcccgac tggagcagga cgaaggggga    60
gggtctcgag gccgagtcct gttcttctga gggacggacc ccagctgggg tggaaaagca    120
gtaccagaga gcctccgagg cgcgcggtgc caaccatgga gcgggcccgc cccagcttcg    180
ggcagcagcg acagcagcag cagccccagc agcagaagca gcagcagagg gatcaggact    240
cggtcgaagc atggctggac gatcactggg actttacctt ctcatacttt gttagaaaag    300
ccaccagaga aatggtcaat gcatggtttg ctgagagagt tcacaccatc cctgtgtgca    360
aggaaggtat cagaggccac accgaatctt gctcttgtcc cttgcagcag agtcctctgtg    420
cagataacag tgtccctgga acaccaacca ggaaaatctc tgcctctgaa tttgaccggc    480
ctcttagacc cattgttgtc aaggattctg agggactgtg gagcttcctc tctgactcag    540
aaaagaagga acagatgcct ctaaccctc caaggtttga tcatgatgaa ggggaccagt    600
gtcacaagact cttggaatta gtgaaggata tttctagtca tttggatgac acagccttat    660
gtcacaataa tttcttgcat atccatggac tgatatctgc tgaccgctat tccctgttcc    720
ttgtctgtga agacagctcc aatgacaagt ttcttatcag ccgcctcttt gatgttgctg    780
aaggttcaac actggaagaa gtttcaaata actgtatccg cttagaatgg aacaaaaggca    840
ttgtgggaca tgtggcagcg cttggtgagc cottgaacat caaagatgca tatgaggatc    900
ctcgggtcaa tgcagaagtt gaccaaata caggctacaa gacacaaagc attcctttgta    960
tgccaattaa gaatcatagg gaagaggttg ttggtgtagc ccaggccatc aacaagaaat    1020
caggaaacgg tgggacattt actgaaaaag atgaaaagga ctttgctgct tatttgcat    1080
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gaaatcaggt gctgcttgac cttgctagtt taatttttga agaacaacaa tcattagaag    1200
taattttgaa gaaaatagct gccactatta tctctttcat gcaagtgcag aaatgcacca    1260
ttttcatagt ggatgaagat tgctccgatt ctttttctag tgtgtttcac atggagtgtg    1320
aggaattaga aaaatcatct gatacattaa caagggaaca tgatgcaaac aaaatcaatt    1380
acatgtatgc tcagtatgct aaaaatacta tggaaacctt ttatatccca gatgtcagta    1440

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aggataaaag atttccctgg acaactgaaa atacaggaaa tgtaaaccag cagtgcatta 1500
gaagtttgct ttgtacacct ataaaaaatg gaaagaagaa taaagttata ggggtttgcc 1560
aacttgtaa taagatggag gagaatactg gcaaggtaa gcctttcaac cgaatgacg 1620
aacagtttct ggaagctttt gtcactcttt gtggcttggg gatccagaac acgcagatgt 1680
atgaagcagt ggagagagcc atggccaagc aaatggtcac attggagggt ctgtcgtatc 1740
atgcttcagc agcagaggaa gaaacaagag agctacagtc gttagcggct gctgtgggtc 1800
catctgcccc gacccttaaa attactgact ttactctcag tgactttgag ctgtctgatc 1860
tggaacagc actgtgtaca attcggatgt ttactgacct caacctgtg cagaacttcc 1920
agatgaaaca tgaggttctt tgcagatgga ttttaagtgt taagaagaat ttcggaaga 1980
atgttgccca tcataattgg agacatgcct ttaatacagc tcagtgcag tttgctgctc 2040
taaaagcag caaaattcag 2060

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<210> SEQ ID NO 12
<211> LENGTH: 1982
<212> TYPE: DNA
<213> ORGANISM: human cDNA

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<400> SEQUENCE: 12

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acaaaatctt ctgcatatc catggactga tatctgctga ccgctattcc ctgttccttg 60
tctgtgaaga cagctccaat gacaagtctt ttatcagccg cctctttgat gttgctgaag 120
gttcaacact ggaagaagt tcaaataact gtatccgctt agaatggaac aaaggcattg 180
tgggacatgt ggcagcgctt ggtgagccct tgaacatcaa agatgcatat gaggatcctc 240
ggttcaatgc agaagtgtac caaattacag gctacaagac acaaagcatt ctttgtatgc 300
caattaagaa tcatagggaa gaggttgttg gtgtagccca ggccatcaac aagaaatcag 360
gaaacggtgg gacatttact gaaaagatg aaaaggactt tgctgcttat ttggcatttt 420
gtggatttgt tcttcataat gctcagctct atgagacttc actgctggag aacaagagaa 480
atcaggtgct gcttgacctt gctagttaa tttttgaaga acaacaatca ttagaagtaa 540
ttttgaagaa aatagctgcc actattatct ctttcatgca agtgcagaaa tgcaccattt 600
tcatagtgga tgaagattgc tccgattctt tttctagtgt gtttcacatg gagtgtgagg 660
aattagaaaa atcatctgat acattaacaa gggaacatga tgcaaacaaa atcaattaca 720
tgtatgctca gtatgtcaaa aatactatgg aaccacttaa tatcccagat gtcagtaagg 780
ataaaagatt tccttgaca actgaaaata caggaaatgt aaaccagcag tgcattagaa 840
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ttgttaataa gatggaggag aatactggca aggttaagcc tttcaaccga aatgacgaac 960
agtttctgga agcttttctc atcttttctg gottggggat ccagaacacg cagatgtatg 1020
aagcagtgga gagagccatg gccaaagcaa tggtcacatt ggaggttctg tcgtatcatg 1080
cttcagcagc agaggaagaa acaagagagc tacagtcgtt agcggctgct gtggtgccat 1140
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aaacagcact gtgtacaatt cggatgttta ctgacctcaa ccttgtgcag aacttccaga 1260
tgaaacatga gtttctttgc agatggattt taagtgttaa gaagaattat cggagaagt 1320
ttgctatca taattggaga catgccttta atacagctca gtgcatgttt gctgctctaa 1380

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aagcaggcaa aattcagaac aagctgactg acctggagat acttgacattg ctgattgctg 1440
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 atccacttgc ccagctttac tgccattcaa tcatggaaca ccatcatttt gaccagtgcc 1560
 tgatgattct taatagtcca ggcaatcaga ttctcagtgg cctctccatt gaagaatata 1620
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 aaccctggcc tattcaacaa cggatagcag aacttgtagc aactgaattt ttgatcaag 1860
 gagacagaga gagaaaagaa ctcaacatag aaccactga tctaatgaac agggagaaga 1920
 aaaaaaaat cccaagtatg caagttgggt tcatagatgc catctgcttg caactgtatg 1980
 ag 1982

<210> SEQ ID NO 13
 <211> LENGTH: 18
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 13

gccaccagag aaatggtc 18

<210> SEQ ID NO 14
 <211> LENGTH: 18
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 14

acaatgggtc taagaggc 18

<210> SEQ ID NO 15
 <211> LENGTH: 18
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 15

tcagtgcctg tttgctgc 18

<210> SEQ ID NO 16
 <211> LENGTH: 18
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer

<400> SEQUENCE: 16

tacaaacatg ttcacatg 18

<210> SEQ ID NO 17
 <211> LENGTH: 1107
 <212> TYPE: DNA
 <213> ORGANISM: human cDNA

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<400> SEQUENCE: 17

gagacatgcc ttaatacag ctcaatgcat gtttgctgct ctaaaagcag gcaaaattca 60
gaacaagctg actgacctgg agatacttgc attgctgatt gctgactaa gccacgattt 120
ggatcaccgt ggtgtgaata actcttacat acagcgaagt gaacatccac ttgccagct 180
ttactgccat tcaatcatgg aacaccatca ttttgaccag tgcctgatga ttcttaatag 240
tccaggcaat cagattctca gtggcctctc cattgaagaa tataagacca cgttgaaaat 300
aatcaagcaa gctatttttag ctacagacct agcactgtac attaagaggc gaggagaatt 360
tttgaactt ataagaaaa atcaattcaa tttggaagat cctcatcaaa aggagtgtt 420
tttgcaatg ctgatgacag cttgtgatct ttctgcaatt acaaaacctt ggctattca 480
acaacggata gcagaacttg tagcaactga attttttgat caaggagaca gagagagaaa 540
agaactcaac atagaacca ctgatctaata gaacagggag aagaaaaaca aaatcccaag 600
tatgcaagtt gggttcatag atgcatctg cttgcaactg tatgaggccc tgaccacgt 660
gtcagaggac tgtttccctt tgctagatgg ctgcagaaag aacaggcaga aatggcaggc 720
ccttgacaga cagcaggaga agatgctgat taatggggaa agcggccagg ccaagcggaa 780
ctgagtggcc tatttcatgc agagttgaag tttacagaga tgggtgtgtc tgcaaatatgc 840
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tatttttatt tttgcacaac ttttgagagt atagcatgaa tgtttttaga ggactattac 960
atatttttg tatatttgtt ttatgctact gaactgaaag gatcaacaac atccactgtt 1020
agcacattga taaaagcatt gtttgatgata tttcgtgtac tgcaaagtgt atgcagtatt 1080
cttgactga ggttttttg cttgggg 1107

<210> SEQ ID NO 18

<211> LENGTH: 18

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 18

tttgaagat cctcatca 18

<210> SEQ ID NO 19

<211> LENGTH: 28

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 19

atgtctcgag tcagttccgc ttggcctg 28

<210> SEQ ID NO 20

<211> LENGTH: 30

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 20

tacagaattc tgaccatgga gcgggcccgc 30

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<210> SEQ ID NO 21
<211> LENGTH: 18
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 21

cattctaagc ggatacag                                     18

<210> SEQ ID NO 22
<211> LENGTH: 2645
<212> TYPE: DNA
<213> ORGANISM: human cDNA
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (12)..(2636)

<400> SEQUENCE: 22

gaattctgac c atg gag cgg gcc ggc ccc agc ttc ggg cag cag cga cag   50
          Met Glu Arg Ala Gly Pro Ser Phe Gly Gln Gln Arg Gln
            1             5             10

cag cag cag ccc cag cag cag aag cag cag cag agg gat cag gac tcg   98
Gln Gln Gln Pro Gln Gln Lys Gln Gln Gln Arg Asp Gln Asp Ser
  15             20             25

gtc gaa gca tgg ctg gac gat cac tgg gac ttt acc ttc tca tac ttt   146
Val Glu Ala Trp Leu Asp Asp His Trp Asp Phe Thr Phe Ser Tyr Phe
  30             35             40             45

gtt aga aaa gcc acc aga gaa atg gtc aat gca tgg ttt gct gag aga   194
Val Arg Lys Ala Thr Arg Glu Met Val Asn Ala Trp Phe Ala Glu Arg
  50             55             60

gtt cac acc atc cct gtg tgc aag gaa ggt atc aga ggc cac acc gaa   242
Val His Thr Ile Pro Val Cys Lys Glu Gly Ile Arg Gly His Thr Glu
  65             70             75

tct tgc tct tgt ccc ttg cag cag agt cct cgt gca gat aac agt gtc   290
Ser Cys Ser Cys Pro Leu Gln Gln Ser Pro Arg Ala Asp Asn Ser Val
  80             85             90

cct gga aca cca acc agg aaa atc tct gcc tct gaa ttt gac cgg cct   338
Pro Gly Thr Pro Thr Arg Lys Ile Ser Ala Ser Glu Phe Asp Arg Pro
  95             100            105

ctt aga ccc att gtt gtc aag gat tct gag gga act gtg agc ttc ctc   386
Leu Arg Pro Ile Val Val Lys Asp Ser Glu Gly Thr Val Ser Phe Leu
  110            115            120            125

tct gac tca gaa aag aag gaa cag atg cct cta acc cct cca agg ttt   434
Ser Asp Ser Glu Lys Lys Glu Gln Met Pro Leu Thr Pro Pro Arg Phe
  130            135            140

gat cat gat gaa ggg gac cag tgc tca aga ctc ttg gaa tta gtg aag   482
Asp His Asp Glu Gly Asp Gln Cys Ser Arg Leu Leu Glu Leu Val Lys
  145            150            155

gat att tct agt cat ttg gat gtc aca gcc tta tgt cac aaa att ttc   530
Asp Ile Ser Ser His Leu Asp Val Thr Ala Leu Cys His Lys Ile Phe
  160            165            170

ttg cat atc cat gga ctg ata tct gct gac cgc tat tcc ctg ttc ctt   578
Leu His Ile His Gly Leu Ile Ser Ala Asp Arg Tyr Ser Leu Phe Leu
  175            180            185

gtc tgt gaa gac agc tcc aat gac aag ttt ctt atc agc cgc ctc ttt   626
Val Cys Glu Asp Ser Ser Asn Asp Lys Phe Leu Ile Ser Arg Leu Phe
  190            195            200            205

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gat gtt gct gaa ggt tca aca ctg gaa gaa gtt tca aat aac tgt atc	674
Asp Val Ala Glu Gly Ser Thr Leu Glu Glu Val Ser Asn Asn Cys Ile	
210 215 220	
cgc tta gaa tgg aac aaa ggc att gtg gga cat gtg gca gcg ctt ggt	722
Arg Leu Glu Trp Asn Lys Gly Ile Val Gly His Val Ala Ala Leu Gly	
225 230 235	
gag ccc ttg aac atc aaa gat gca tat gag gat cct cgg ttc aat gca	770
Glu Pro Leu Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg Phe Asn Ala	
240 245 250	
gaa gtt gac caa att aca ggc tac aag aca caa agc att ctt tgt atg	818
Glu Val Asp Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile Leu Cys Met	
255 260 265	
cca att aag aat cat agg gaa gag gtt gtt ggt gta gcc cag gcc atc	866
Pro Ile Lys Asn His Arg Glu Glu Val Val Gly Val Ala Gln Ala Ile	
270 275 280 285	
aac aag aaa tca gga aac ggt ggg aca ttt act gaa aaa gat gaa aag	914
Asn Lys Lys Ser Gly Asn Gly Gly Thr Phe Thr Glu Lys Asp Glu Lys	
290 295 300	
gac ttt gct gct tat ttg gca ttt tgt ggt att gtt ctt cat aat gct	962
Asp Phe Ala Ala Tyr Leu Ala Phe Cys Gly Ile Val Leu His Asn Ala	
305 310 315	
cag ctc tat gag act tca ctg ctg gag aac aag aga aat cag gtg ctg	1010
Gln Leu Tyr Glu Thr Ser Leu Leu Glu Asn Lys Arg Asn Gln Val Leu	
320 325 330	
ctt gac ctt gct agt tta att ttt gaa gaa caa caa tca tta gaa gta	1058
Leu Asp Leu Ala Ser Leu Ile Phe Glu Glu Gln Gln Ser Leu Glu Val	
335 340 345	
att ttg aag aaa ata gct gcc act att atc tct ttc atg caa gtg cag	1106
Ile Leu Lys Lys Ile Ala Ala Thr Ile Ile Ser Phe Met Gln Val Gln	
350 355 360 365	
aaa tgc acc att ttc ata gtg gat gaa gat tgc tcc gat tct ttt tct	1154
Lys Cys Thr Ile Phe Ile Val Asp Glu Asp Cys Ser Asp Ser Phe Ser	
370 375 380	
agt gtg ttt cac atg gag tgt gag gaa tta gaa aaa tca tct gat aca	1202
Ser Val Phe His Met Glu Cys Glu Glu Leu Glu Lys Ser Ser Asp Thr	
385 390 395	
tta aca agg gaa cat gat gca aac aaa atc aat tac atg tat gct cag	1250
Leu Thr Arg Glu His Asp Ala Asn Lys Ile Asn Tyr Met Tyr Ala Gln	
400 405 410	
tat gtc aaa aat act atg gaa cca ctt aat atc cca gat gtc agt aag	1298
Tyr Val Lys Asn Thr Met Glu Pro Leu Asn Ile Pro Asp Val Ser Lys	
415 420 425	
gat aaa aga ttt ccc tgg aca act gaa aat aca gga aat gta aac cag	1346
Asp Lys Arg Phe Pro Trp Thr Thr Glu Asn Thr Gly Asn Val Asn Gln	
430 435 440 445	
cag tgc att aga agt ttg ctt tgt aca cct ata aaa aat gga aag aag	1394
Gln Cys Ile Arg Ser Leu Leu Cys Thr Pro Ile Lys Asn Gly Lys Lys	
450 455 460	
aat aaa gtt ata ggg gtt tgc caa ctt gtt aat aag atg gag gag aat	1442
Asn Lys Val Ile Gly Val Cys Gln Leu Val Asn Lys Met Glu Glu Asn	
465 470 475	
act ggc aag gtt aag cct ttc aac cga aat gac gaa cag ttt ctg gaa	1490
Thr Gly Lys Val Lys Pro Phe Asn Arg Asn Asp Glu Gln Phe Leu Glu	
480 485 490	
gct ttt gtc atc ttt tgt ggc ttg ggg atc cag aac acg cag atg tat	1538
Ala Phe Val Ile Phe Cys Gly Leu Gly Ile Gln Asn Thr Gln Met Tyr	
495 500 505	

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gaa gca gtg gag aga gcc atg gcc aag caa atg gtc aca ttg gag gtt Glu Ala Val Glu Arg Ala Met Ala Lys Gln Met Val Thr Leu Glu Val 510 515 520 525	1586
ctg tcg tat cat gct tca gca gca gag gaa gaa aca aga gag cta cag Leu Ser Tyr His Ala Ser Ala Ala Glu Glu Thr Arg Glu Leu Gln 530 535 540	1634
tcg tta gcg gct gct gtg gtg cca tct gcc cag acc ctt aaa att act Ser Leu Ala Ala Val Val Pro Ser Ala Gln Thr Leu Lys Ile Thr 545 550 555	1682
gac ttt agc ttc agt gac ttt gag ctg tct gat ctg gaa aca gca ctg Asp Phe Ser Phe Ser Asp Phe Glu Leu Ser Asp Leu Glu Thr Ala Leu 560 565 570	1730
tgt aca att cgg atg ttt act gac ttc aac ctt gtg cag aac ttc cag Cys Thr Ile Arg Met Phe Thr Asp Leu Asn Leu Val Gln Asn Phe Gln 575 580 585	1778
atg aaa cat gag gtt ctt tgc aga tgg att tta agt gtt aag aag aat Met Lys His Glu Val Leu Cys Arg Trp Ile Leu Ser Val Lys Lys Asn 590 595 600 605	1826
tat cgg aag aat gtt gcc tat cat aat tgg aga cat gcc ttt aat aca Tyr Arg Lys Asn Val Ala Tyr His Asn Trp Arg His Ala Phe Asn Thr 610 615 620	1874
gct cag tgc atg ttt gct gct cta aaa gca ggc aaa att cag aac aag Ala Gln Cys Met Phe Ala Ala Leu Lys Ala Gly Lys Ile Gln Asn Lys 625 630 635	1922
ctg act gac ctg gag ata ctt gca ttg ctg att gct gca cta agc cac Leu Thr Asp Leu Glu Ile Leu Ala Leu Leu Ile Ala Ala Leu Ser His 640 645 650	1970
gat ttg gat cac cgt ggt gtg aat aac tct tac ata cag cga agt gaa Asp Leu Asp His Arg Gly Val Asn Asn Ser Tyr Ile Gln Arg Ser Glu 655 660 665	2018
cat cca ctt gcc cag ctt tac tgc cat tca atc atg gaa cac cat cat His Pro Leu Ala Gln Leu Tyr Cys His Ser Ile Met Glu His His His 670 675 680 685	2066
ttt gac cag tgc ctg atg att ctt aat agt cca ggc aat cag att ctc Phe Asp Gln Cys Leu Met Ile Leu Asn Ser Pro Gly Asn Gln Ile Leu 690 695 700	2114
agt ggc ttc tcc att gaa gaa tat aag acc acg ttg aaa ata atc aag Ser Gly Leu Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys Ile Ile Lys 705 710 715	2162
caa gct att tta gct aca gac cta gca ctg tac att aag agg cga gga Gln Ala Ile Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys Arg Arg Gly 720 725 730	2210
gaa ttt ttt gaa ctt ata aga aaa aat caa ttc aat ttg gaa gat cct Glu Phe Phe Glu Leu Ile Arg Lys Asn Gln Phe Asn Leu Glu Asp Pro 735 740 745	2258
cat caa aag gag ttg ttt ttg gca atg ctg atg aca gct tgt gat ctt His Gln Lys Glu Leu Phe Leu Ala Met Leu Met Thr Ala Cys Asp Leu 750 755 760 765	2306
tct gca att aca aaa ccc tgg cct att caa caa cgg ata gca gaa ctt Ser Ala Ile Thr Lys Pro Trp Pro Ile Gln Gln Arg Ile Ala Glu Leu 770 775 780	2354
gta gca act gaa ttt ttt gat caa gga gac aga gag aga aaa gaa ctc Val Ala Thr Glu Phe Phe Asp Gln Gly Asp Arg Glu Arg Lys Glu Leu 785 790 795	2402
aac ata gaa ccc act gat cta atg aac agg gag aag aaa aac aaa atc Asn Ile Glu Pro Thr Asp Leu Met Asn Arg Glu Lys Lys Asn Lys Ile 800 805 810	2450

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cca agt atg caa gtt ggg ttc ata gat gcc atc tgc ttg caa ctg tat    2498
Pro Ser Met Gln Val Gly Phe Ile Asp Ala Ile Cys Leu Gln Leu Tyr
      815                               820                               825

gag gcc ctg acc cac gtg tca gag gac tgt ttc cct ttg cta gat ggc    2546
Glu Ala Leu Thr His Val Ser Glu Asp Cys Phe Pro Leu Leu Asp Gly
830                               835                               840                               845

tgc aga aag aac agg cag aaa tgg cag gcc ctt gca gaa cag cag gag    2594
Cys Arg Lys Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu Gln Gln Glu
                               850                               855                               860

aag atg ctg att aat ggg gaa agc ggc cag gcc aag cgg aac tgactcgag    2645
Lys Met Leu Ile Asn Gly Glu Ser Gly Gln Ala Lys Arg Asn
      865                               870                               875

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<210> SEQ ID NO 23

<211> LENGTH: 875

<212> TYPE: PRT

<213> ORGANISM: human cDNA

<400> SEQUENCE: 23

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Met Glu Arg Ala Gly Pro Ser Phe Gly Gln Gln Arg Gln Gln Gln Gln
 1                               5                               10                               15

Pro Gln Gln Gln Lys Gln Gln Gln Arg Asp Gln Asp Ser Val Glu Ala
 20                               25                               30

Trp Leu Asp Asp His Trp Asp Phe Thr Phe Ser Tyr Phe Val Arg Lys
 35                               40                               45

Ala Thr Arg Glu Met Val Asn Ala Trp Phe Ala Glu Arg Val His Thr
 50                               55                               60

Ile Pro Val Cys Lys Glu Gly Ile Arg Gly His Thr Glu Ser Cys Ser
 65                               70                               75                               80

Cys Pro Leu Gln Gln Ser Pro Arg Ala Asp Asn Ser Val Pro Gly Thr
 85                               90                               95

Pro Thr Arg Lys Ile Ser Ala Ser Glu Phe Asp Arg Pro Leu Arg Pro
 100                              105                              110

Ile Val Val Lys Asp Ser Glu Gly Thr Val Ser Phe Leu Ser Asp Ser
 115                              120                              125

Glu Lys Lys Glu Gln Met Pro Leu Thr Pro Pro Arg Phe Asp His Asp
 130                              135                              140

Glu Gly Asp Gln Cys Ser Arg Leu Leu Glu Leu Val Lys Asp Ile Ser
 145                              150                              155                              160

Ser His Leu Asp Val Thr Ala Leu Cys His Lys Ile Phe Leu His Ile
 165                              170                              175

His Gly Leu Ile Ser Ala Asp Arg Tyr Ser Leu Phe Leu Val Cys Glu
 180                              185                              190

Asp Ser Ser Asn Asp Lys Phe Leu Ile Ser Arg Leu Phe Asp Val Ala
 195                              200                              205

Glu Gly Ser Thr Leu Glu Glu Val Ser Asn Asn Cys Ile Arg Leu Glu
 210                              215                              220

Trp Asn Lys Gly Ile Val Gly His Val Ala Ala Leu Gly Glu Pro Leu
 225                              230                              235                              240

Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg Phe Asn Ala Glu Val Asp
 245                              250                              255

Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile Leu Cys Met Pro Ile Lys
 260                              265                              270

Asn His Arg Glu Glu Val Val Gly Val Ala Gln Ala Ile Asn Lys Lys

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275					280					285					
Ser	Gly	Asn	Gly	Gly	Thr	Phe	Thr	Glu	Lys	Asp	Glu	Lys	Asp	Phe	Ala
290						295					300				
Ala	Tyr	Leu	Ala	Phe	Cys	Gly	Ile	Val	Leu	His	Asn	Ala	Gln	Leu	Tyr
305					310					315					320
Glu	Thr	Ser	Leu	Leu	Glu	Asn	Lys	Arg	Asn	Gln	Val	Leu	Leu	Asp	Leu
				325					330					335	
Ala	Ser	Leu	Ile	Phe	Glu	Glu	Gln	Gln	Ser	Leu	Glu	Val	Ile	Leu	Lys
			340					345					350		
Lys	Ile	Ala	Ala	Thr	Ile	Ile	Ser	Phe	Met	Gln	Val	Gln	Lys	Cys	Thr
		355					360					365			
Ile	Phe	Ile	Val	Asp	Glu	Asp	Cys	Ser	Asp	Ser	Phe	Ser	Ser	Val	Phe
	370					375					380				
His	Met	Glu	Cys	Glu	Glu	Leu	Glu	Lys	Ser	Ser	Asp	Thr	Leu	Thr	Arg
385					390					395					400
Glu	His	Asp	Ala	Asn	Lys	Ile	Asn	Tyr	Met	Tyr	Ala	Gln	Tyr	Val	Lys
				405					410					415	
Asn	Thr	Met	Glu	Pro	Leu	Asn	Ile	Pro	Asp	Val	Ser	Lys	Asp	Lys	Arg
			420					425					430		
Phe	Pro	Trp	Thr	Thr	Glu	Asn	Thr	Gly	Asn	Val	Asn	Gln	Gln	Cys	Ile
		435					440					445			
Arg	Ser	Leu	Leu	Cys	Thr	Pro	Ile	Lys	Asn	Gly	Lys	Lys	Asn	Lys	Val
	450					455					460				
Ile	Gly	Val	Cys	Gln	Leu	Val	Asn	Lys	Met	Glu	Glu	Asn	Thr	Gly	Lys
465					470					475					480
Val	Lys	Pro	Phe	Asn	Arg	Asn	Asp	Glu	Gln	Phe	Leu	Glu	Ala	Phe	Val
				485					490					495	
Ile	Phe	Cys	Gly	Leu	Gly	Ile	Gln	Asn	Thr	Gln	Met	Tyr	Glu	Ala	Val
			500				505						510		
Glu	Arg	Ala	Met	Ala	Lys	Gln	Met	Val	Thr	Leu	Glu	Val	Leu	Ser	Tyr
		515				520						525			
His	Ala	Ser	Ala	Ala	Glu	Glu	Glu	Thr	Arg	Glu	Leu	Gln	Ser	Leu	Ala
	530					535					540				
Ala	Ala	Val	Val	Pro	Ser	Ala	Gln	Thr	Leu	Lys	Ile	Thr	Asp	Phe	Ser
545					550					555					560
Phe	Ser	Asp	Phe	Glu	Leu	Ser	Asp	Leu	Glu	Thr	Ala	Leu	Cys	Thr	Ile
				565					570					575	
Arg	Met	Phe	Thr	Asp	Leu	Asn	Leu	Val	Gln	Asn	Phe	Gln	Met	Lys	His
			580					585					590		
Glu	Val	Leu	Cys	Arg	Trp	Ile	Leu	Ser	Val	Lys	Lys	Asn	Tyr	Arg	Lys
		595					600					605			
Asn	Val	Ala	Tyr	His	Asn	Trp	Arg	His	Ala	Phe	Asn	Thr	Ala	Gln	Cys
	610					615						620			
Met	Phe	Ala	Ala	Leu	Lys	Ala	Gly	Lys	Ile	Gln	Asn	Lys	Leu	Thr	Asp
625					630					635					640
Leu	Glu	Ile	Leu	Ala	Leu	Leu	Ile	Ala	Ala	Leu	Ser	His	Asp	Leu	Asp
				645					650					655	
His	Arg	Gly	Val	Asn	Asn	Ser	Tyr	Ile	Gln	Arg	Ser	Glu	His	Pro	Leu
			660					665					670		
Ala	Gln	Leu	Tyr	Cys	His	Ser	Ile	Met	Glu	His	His	His	Phe	Asp	Gln
		675					680						685		

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Cys Leu Met Ile Leu Asn Ser Pro Gly Asn Gln Ile Leu Ser Gly Leu
 690 695 700
 Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys Ile Ile Lys Gln Ala Ile
 705 710 715 720
 Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys Arg Arg Gly Glu Phe Phe
 725 730 735
 Glu Leu Ile Arg Lys Asn Gln Phe Asn Leu Glu Asp Pro His Gln Lys
 740 745 750
 Glu Leu Phe Leu Ala Met Leu Met Thr Ala Cys Asp Leu Ser Ala Ile
 755 760 765
 Thr Lys Pro Trp Pro Ile Gln Gln Arg Ile Ala Glu Leu Val Ala Thr
 770 775 780
 Glu Phe Phe Asp Gln Gly Asp Arg Glu Arg Lys Glu Leu Asn Ile Glu
 785 790 795 800
 Pro Thr Asp Leu Met Asn Arg Glu Lys Lys Asn Lys Ile Pro Ser Met
 805 810 815
 Gln Val Gly Phe Ile Asp Ala Ile Cys Leu Gln Leu Tyr Glu Ala Leu
 820 825 830
 Thr His Val Ser Glu Asp Cys Phe Pro Leu Leu Asp Gly Cys Arg Lys
 835 840 845
 Asn Arg Gln Lys Trp Gln Ala Leu Ala Glu Gln Gln Glu Lys Met Leu
 850 855 860
 Ile Asn Gly Glu Ser Gly Gln Ala Lys Arg Asn
 865 870 875

<210> SEQ ID NO 24
 <211> LENGTH: 235
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 24

Phe Gln Met Lys His Glu Val Leu Cys Lys Trp Ile Leu Ser Val Lys
 1 5 10 15
 Lys Asn Tyr Arg Lys Asn Val Ala Tyr His Asn Trp Arg His Ala Phe
 20 25 30
 Asn Thr Ala Gln Cys Met Phe Ala Ala Leu Lys Ala Gly Lys Ile Gln
 35 40 45
 Lys Arg Leu Thr Asp Leu Glu Ile Leu Ala Leu Leu Ile Ala Ala Leu
 50 55 60
 Ser His Asp Leu Asp His Arg Gly Val Asn Asn Ser Tyr Ile Gln Arg
 65 70 75 80
 Ser Glu His Pro Leu Ala Gln Leu Tyr Cys His Ser Ile Met Glu His
 85 90 95
 His His Phe Asp Gln Cys Leu Met Ile Leu Asn Ser Pro Gly Asn Gln
 100 105 110
 Ile Leu Ser Gly Leu Ser Ile Glu Glu Tyr Lys Thr Thr Leu Lys Ile
 115 120 125
 Ile Lys Gln Ala Ile Leu Ala Thr Asp Leu Ala Leu Tyr Ile Lys Arg
 130 135 140
 Arg Gly Glu Phe Phe Glu Leu Ile Met Lys Asn Gln Phe Asn Leu Glu
 145 150 155 160
 Asp Pro His Gln Lys Glu Leu Phe Leu Ala Met Leu Met Thr Ala Cys

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165	170	175
Asp Leu Ser Ala Ile Thr Lys Pro Trp Pro Ile Gln Gln Arg Ile Ala 180 185 190		
Glu Leu Val Ala Thr Glu Phe Phe Asp Gln Gly Asp Arg Glu Arg Lys 195 200 205		
Glu Leu Asn Ile Glu Pro Ala Asp Leu Met Asn Arg Glu Lys Lys Asn 210 215 220		
Lys Ile Pro Ser Met Glu Val Gly Phe Ile Asp 225 230 235		

<210> SEQ ID NO 25
 <211> LENGTH: 244
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 25

Phe His Ile Pro Gln Glu Ala Leu Val Arg Phe Met Tyr Ser Leu Ser 1 5 10 15		
Lys Gly Tyr Arg Arg Ile Thr Tyr His Asn Trp Arg His Gly Phe Asn 20 25 30		
Val Gly Gln Thr Met Phe Ser Leu Val Thr Gly Lys Leu Lys Arg 35 40 45		
Tyr Phe Thr Asp Leu Glu Ala Leu Ala Met Val Thr Ala Ala Phe Cys 50 55 60		
His Asp Ile Asp His Arg Gly Thr Asn Asn Leu Tyr Gln Met Lys Ser 65 70 75 80		
Gln Asn Pro Leu Ala Lys Leu His Gly Ser Ile Leu Glu Arg His 85 90 95		
His Leu Glu Phe Gly Lys Thr Leu Leu Arg Asp Glu Ser Leu Asn Ile 100 105 110		
Phe Gln Asn Leu Asn Arg Arg Gln His Glu His Ala Ile His Met Met 115 120 125		
Asp Ile Ala Ile Ile Ala Thr Asp Leu Ala Leu Tyr Cys Lys Lys Arg 130 135 140		
Thr Met Phe Gln Lys Ile Val Asp Gln Ser Lys Thr Tyr Glu Thr Gln 145 150 155 160		
Gln Glu Trp Thr Gln Tyr Met Met Leu Asp Gln Thr Arg Lys Glu Ile 165 170 175		
Val Met Ala Met Met Met Thr Ala Cys Asp Leu Ser Ala Ile Thr Lys 180 185 190		
Pro Trp Glu Val Gln Ser Lys Val Ala Leu Leu Val Ala Ala Glu Phe 195 200 205		
Trp Glu Gln Gly Asp Leu Glu Arg Thr Val Leu Gln Gln Asn Pro Ile 210 215 220		
Pro Met Met Asp Arg Asn Lys Ala Asp Glu Leu Pro Lys Leu Gln Val 225 230 235 240		

Gly Phe Ile Asp

<210> SEQ ID NO 26
 <211> LENGTH: 244
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 26

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Phe Gln Ile Pro Gln Glu Val Leu Val Arg Phe Leu Phe Ser Val Ser
1           5           10           15
Lys Gly Tyr Arg Arg Ile Thr Tyr His Asn Trp Arg His Gly Phe Asn
20          25          30
Val Ala Gln Thr Met Phe Thr Leu Leu Met Thr Gly Lys Leu Lys Ser
35          40          45
Tyr Tyr Thr Asp Leu Glu Ala Phe Ala Met Val Thr Ala Gly Leu Cys
50          55          60
His Asp Ile Asp His Arg Gly Thr Asn Asn Leu Tyr Gln Met Lys Ser
65          70          75          80
Gln Asn Pro Leu Ala Lys Leu His Gly Ser Ser Ile Leu Glu Arg His
85          90          95
His Leu Glu Phe Gly Lys Phe Leu Leu Ser Glu Glu Thr Leu Asn Ile
100         105         110
Tyr Gln Asn Leu Asn Arg Arg Gln His Glu His Val Ile His Leu Met
115        120        125
Asp Ile Ala Ile Ile Ala Thr Asp Leu Ala Leu Tyr Phe Lys Lys Arg
130        135        140
Thr Met Phe Gln Lys Ile Val Asp Glu Ser Lys Asn Tyr Glu Asp Arg
145        150        155        160
Lys Ser Trp Val Glu Tyr Leu Ser Leu Glu Thr Thr Arg Lys Glu Ile
165        170        175
Val Met Ala Met Met Met Thr Ala Cys Asp Leu Ser Ala Ile Thr Lys
180        185        190
Pro Trp Glu Val Gln Ser Lys Val Ala Leu Leu Val Ala Ala Glu Phe
195        200        205
Trp Glu Gln Gly Asp Leu Glu Arg Thr Val Leu Asp Gln Gln Pro Ile
210        215        220
Pro Met Met Asp Arg Asn Lys Ala Ala Glu Leu Pro Lys Leu Gln Val
225        230        235        240
Gly Phe Ile Asp

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<210> SEQ ID NO 27
<211> LENGTH: 244
<212> TYPE: PRT
<213> ORGANISM: bovine

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<400> SEQUENCE: 27

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Phe Lys Val Pro Val Glu Val Leu Thr Arg Trp Met Thr Tyr Val Arg
1           5           10           15
Lys Gly Tyr Arg Ala Val Thr Tyr His Asn Trp Arg His Gly Phe Asn
20          25          30
Val Gly Gln Thr Met Phe Thr Leu Leu Met Thr Gly Arg Leu Lys Lys
35          40          45
Tyr Tyr Thr Asp Leu Glu Ala Phe Ala Met Leu Ala Ala Ala Phe Cys
50          55          60
His Asp Ile Asp His Arg Gly Thr Asn Asn Leu Tyr Gln Met Lys Ser
65          70          75          80
Thr Ser Pro Leu Ala Arg Leu His Gly Ser Ser Ile Leu Glu Arg His
85          90          95
His Leu Glu Tyr Ser Lys Thr Leu Leu Gln Asp Glu Ser Leu Asn Ile
100         105         110

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Pro Glu Leu Gln Ile Ser Phe Met Glu
225 230

<210> SEQ ID NO 29
<211> LENGTH: 230
<212> TYPE: PRT
<213> ORGANISM: bovine

<400> SEQUENCE: 29

Phe Lys Ile Pro Val Ser Cys Leu Ile Ala Phe Ala Glu Ala Leu Glu
1 5 10 15
Val Gly Tyr Ser Lys Tyr Lys Asn Pro Tyr His Asn Leu Ile His Ala
20 25 30
Ala Asp Val Thr Gln Thr Val His Tyr Ile Met Leu His Thr Gly Ile
35 40 45
Met His Trp Leu Thr Glu Leu Glu Ile Leu Ala Met Val Phe Ala Ala
50 55 60
Ala Ile His Asp Tyr Glu His Ile Gly Thr Thr Asn Asn Phe His Ile
65 70 75 80
Gln Thr Arg Ser Asp Val Ala Ile Leu Tyr Asn Asp Arg Ser Val Leu
85 90 95
Glu Asn His His Val Ser Ala Ala Tyr Arg Leu Met Gln Glu Glu Glu
100 105 110
Met Asn Val Leu Ile Asn Leu Ser Lys Asp Asp Trp Arg Asp Leu Arg
115 120 125
Asn Leu Val Ile Glu Met Val Leu Ser Thr Asp Met Ser Gly His Phe
130 135 140
Gln Gln Ile Lys Asn Ile Arg Asn Ser Leu Gln Gln Pro Glu Gly Leu
145 150 155 160
Asp Lys Ala Lys Thr Met Ser Leu Ile Leu His Ala Ala Asp Ile Ser
165 170 175
His Pro Ala Lys Ser Trp Lys Leu His His Arg Trp Thr Met Ala Leu
180 185 190
Met Glu Glu Phe Phe Leu Gln Gly Asp Lys Glu Ala Glu Leu Gly Leu
195 200 205
Pro Phe Ser Pro Leu Cys Asp Arg Lys Ser Thr Met Val Ala Gln Ser
210 215 220
Gln Ile Gly Phe Ile Asp
225 230

<210> SEQ ID NO 30
<211> LENGTH: 230
<212> TYPE: PRT
<213> ORGANISM: bovine

<400> SEQUENCE: 30

Phe Lys Ile Pro Thr Val Phe Leu Met Thr Phe Leu Asp Ala Leu Glu
1 5 10 15
Thr Gly Tyr Gly Lys Tyr Lys Asn Pro Tyr His Asn Gln Ile His Ala
20 25 30
Ala Asp Val Thr Gln Thr Val His Cys Phe Leu Leu Arg Thr Gly Met
35 40 45
Val His Cys Leu Ser Glu Ile Glu Val Leu Ala Ile Ile Phe Ala Ala
50 55 60

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Ala Ile His Asp Tyr Glu His Ile Gly Thr Thr Asn Ser Phe His Ile
65 70 75 80

Gln Thr Lys Ser Glu Gln Ala Ile Leu Tyr Asn Asp Arg Ser Val Leu
85 90 95

Glu Asn His His Ile Ser Ser Val Phe Arg Met Met Gln Asp Asp Glu
100 105 110

Met Asn Ile Phe Ile Asn Leu Thr Lys Asp Glu Phe Val Glu Leu Arg
115 120 125

Ala Leu Val Ile Glu Met Val Leu Ala Thr Asp Met Ser Cys His Phe
130 135 140

Gln Gln Val Lys Ser Met Lys Thr Ala Leu Gln Gln Leu Glu Arg Ile
145 150 155 160

Asp Lys Ser Lys Ala Leu Ser Leu Leu Leu His Ala Ala Asp Ile Ser
165 170 175

His Pro Thr Lys Gln Trp Ser Val His Ser Arg Trp Thr Lys Ala Leu
180 185 190

Met Glu Glu Phe Phe Arg Gln Gly Asp Lys Glu Ala Glu Leu Gly Leu
195 200 205

Pro Phe Ser Pro Leu Cys Asp Arg Thr Ser Thr Leu Val Ala Gln Ser
210 215 220

Gln Ile Gly Phe Ile Asp
225 230

<210> SEQ ID NO 31
 <211> LENGTH: 240
 <212> TYPE: PRT
 <213> ORGANISM: rat

<400> SEQUENCE: 31

Phe Gln Ile Pro Ala Asp Thr Leu Leu Arg Tyr Leu Leu Thr Leu Glu
1 5 10 15

Gly His Tyr His Ser Asn Val Ala Tyr His Asn Ser Ile His Ala Ala
20 25 30

Asp Val Val Gln Ser Ala His Val Leu Leu Gly Thr Pro Ala Leu Glu
35 40 45

Ala Val Phe Thr Asp Leu Glu Val Leu Ala Ala Ile Phe Ala Cys Ala
50 55 60

Ile His Asp Val Asp His Pro Gly Val Ser Asn Gln Phe Leu Ile Asn
65 70 75 80

Thr Asn Ser Glu Leu Ala Leu Met Tyr Asn Asp Ser Ser Val Leu Glu
85 90 95

Asn His His Leu Ala Val Gly Phe Lys Leu Leu Gln Gly Glu Asn Cys
100 105 110

Asp Ile Phe Gln Asn Leu Ser Thr Lys Gln Lys Leu Ser Leu Arg Arg
115 120 125

Met Val Ile Asp Met Val Leu Ala Thr Asp Met Ser Lys His Met Ser
130 135 140

Leu Leu Ala Asp Leu Lys Thr Met Val Glu Thr Lys Lys Val Thr Ser
145 150 155 160

Leu Gly Val Leu Leu Leu Asp Asn Tyr Ser Asp Arg Ile Gln Val Leu
165 170 175

Gln Ser Leu Val His Cys Ala Asp Leu Ser Asn Pro Ala Lys Pro Leu

-continued

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          180              185              190
Pro Leu Tyr Arg Gln Trp Thr Glu Arg Ile Met Ala Glu Phe Phe Gln
   195                          200                205

Gln Gly Asp Arg Glu Arg Glu Ser Gly Leu Asp Ile Ser Pro Met Cys
   210                          215                220

Asp Lys His Thr Ala Ser Val Glu Lys Ser Gln Val Gly Phe Ile Asp
   225                          230                235                240

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<210> SEQ ID NO 32
<211> LENGTH: 239
<212> TYPE: PRT
<213> ORGANISM: drosophila

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<400> SEQUENCE: 32

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Met Ile Pro Pro Lys Thr Phe Leu Asn Phe Met Ser Thr Leu Glu Asp
 1          5          10          15

His Tyr Val Lys Asp Asn Pro Phe His Asn Ser Leu His Ala Ala Asp
 20         25         30

Val Thr Gln Ser Thr Asn Val Leu Leu Asn Thr Pro Ala Leu Glu Gly
 35         40         45

Val Phe Thr Pro Leu Glu Val Gly Gly Ala Leu Phe Ala Ala Cys Ile
 50         55         60

His Asp Val Asp His Pro Gly Leu Thr Asn Gln Phe Leu Val Asn Ser
 65         70         75         80

Ser Ser Glu Leu Ala Leu Met Tyr Asn Asp Glu Ser Val Leu Glu Asn
 85         90         95

His His Leu Ala Val Ala Phe Lys Leu Leu Gln Asn Gln Gly Cys Asp
100        105        110

Ile Phe Cys Asn Met Gln Lys Lys Gln Arg Gln Thr Leu Arg Lys Met
115        120        125

Val Ile Asp Ile Val Leu Ser Thr Asp Met Ser Lys His Met Ser Leu
130        135        140

Leu Ala Asp Leu Lys Thr Met Val Glu Thr Lys Lys Val Ala Gly Ser
145        150        155        160

Gly Val Leu Leu Leu Asp Asn Tyr Thr Asp Arg Ile Gln Val Leu Glu
165        170        175

Asn Leu Val His Cys Ala Asp Leu Ser Asn Pro Thr Lys Pro Leu Pro
180        185        190

Leu Tyr Lys Arg Trp Val Ala Leu Leu Met Glu Glu Phe Phe Leu Gln
195        200        205

Gly Asp Lys Glu Arg Glu Ser Gly Met Asp Ile Ser Pro Met Cys Asp
210        215        220

Arg His Asn Ala Thr Ile Glu Lys Ser Gln Val Gly Phe Ile Asp
225        230        235

```

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<210> SEQ ID NO 33
<211> LENGTH: 385
<212> TYPE: PRT
<213> ORGANISM: bovine

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<400> SEQUENCE: 33

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```

Leu Leu Glu Leu Val Lys Asp Ile Ser Ser His Leu Asp Val Thr Ala
 1          5          10          15

Leu Cys His Lys Ile Phe Leu His Ile His Gly Leu Ile Ser Ala Asp

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-continued

20		25		30											
Arg	Tyr	Ser	Leu	Phe	Leu	Val	Cys	Glu	Asp	Ser	Ser	Asn	Asp	Lys	Phe
	35						40					45			
Leu	Ile	Ser	Arg	Leu	Phe	Asp	Val	Ala	Glu	Gly	Ser	Thr	Leu	Glu	Glu
	50					55					60				
Ala	Ser	Asn	Asn	Cys	Ile	Arg	Leu	Glu	Trp	Asn	Lys	Gly	Ile	Val	Gly
	65				70					75					80
His	Val	Ala	Ala	Phe	Gly	Glu	Pro	Leu	Asn	Ile	Lys	Asp	Ala	Tyr	Glu
				85					90					95	
Asp	Pro	Arg	Phe	Asn	Ala	Glu	Val	Asp	Gln	Ile	Thr	Gly	Tyr	Lys	Thr
			100					105					110		
Gln	Ser	Ile	Leu	Cys	Met	Pro	Ile	Lys	Asn	His	Arg	Glu	Glu	Val	Val
		115					120					125			
Gly	Val	Ala	Gln	Ala	Ile	Asn	Lys	Lys	Ser	Gly	Asn	Gly	Gly	Thr	Phe
	130					135					140				
Thr	Glu	Lys	Asp	Glu	Lys	Asp	Phe	Ala	Ala	Tyr	Leu	Ala	Phe	Cys	Gly
	145				150					155					160
Ile	Val	Leu	His	Asn	Ala	Gln	Leu	Tyr	Glu	Thr	Ser	Leu	Leu	Glu	Asn
				165					170					175	
Lys	Arg	Asn	Gln	Val	Leu	Leu	Asp	Leu	Ala	Ser	Leu	Ile	Phe	Glu	Glu
			180					185					190		
Gln	Gln	Ser	Leu	Glu	Val	Ile	Leu	Lys	Lys	Ile	Ala	Ala	Thr	Ile	Ile
		195					200					205			
Ser	Phe	Met	Gln	Val	Gln	Lys	Cys	Thr	Ile	Phe	Ile	Val	Asp	Glu	Asp
	210					215					220				
Cys	Ser	Asp	Ser	Phe	Ser	Ser	Val	Phe	His	Met	Glu	Cys	Glu	Glu	Leu
	225				230					235					240
Glu	Lys	Ser	Ser	Asp	Thr	Leu	Thr	Arg	Glu	Arg	Asp	Ala	Asn	Arg	Ile
				245					250					255	
Asn	Tyr	Met	Tyr	Ala	Gln	Tyr	Val	Lys	Asn	Thr	Met	Glu	Pro	Leu	Asn
			260					265					270		
Ile	Pro	Asp	Val	Ser	Lys	Asp	Lys	Arg	Phe	Pro	Trp	Thr	Asn	Glu	Asn
		275					280					285			
Met	Gly	Asn	Ile	Asn	Gln	Gln	Cys	Ile	Arg	Ser	Leu	Leu	Cys	Thr	Pro
	290					295					300				
Ile	Lys	Asn	Gly	Lys	Lys	Asn	Lys	Val	Ile	Gly	Val	Cys	Gln	Leu	Val
	305				310					315					320
Asn	Lys	Met	Glu	Glu	Thr	Thr	Gly	Lys	Val	Lys	Ala	Phe	Asn	Arg	Asn
				325					330					335	
Asp	Glu	Gln	Phe	Leu	Glu	Ala	Phe	Val	Ile	Phe	Cys	Gly	Leu	Gly	Ile
			340					345					350		
Gln	Asn	Thr	Gln	Met	Tyr	Glu	Ala	Val	Glu	Arg	Ala	Met	Ala	Lys	Gln
		355					360					365			
Met	Val	Thr	Leu	Glu	Val	Leu	Ser	Tyr	His	Ala	Ser	Ala	Ala	Glu	Glu
	370					375					380				
Glu															
	385														

<210> SEQ ID NO 34
 <211> LENGTH: 353
 <212> TYPE: PRT
 <213> ORGANISM: bovine

-continued

<400> SEQUENCE: 34

```

Ile Leu Gln Leu Cys Gly Glu Leu Tyr Asp Leu Asp Ala Ser Ser Leu
1           5           10           15
Gln Leu Lys Val Leu Gln Tyr Leu Gln Gln Glu Thr Gln Ala Ser Arg
20           25           30
Cys Cys Leu Leu Leu Val Ser Glu Asp Asn Leu Gln Leu Ser Cys Lys
35           40           45
Val Ile Gly Asp Lys Val Leu Glu Glu Glu Ile Ser Phe Pro Leu Thr
50           55           60
Thr Gly Arg Leu Gly Gln Val Val Glu Asp Lys Lys Ser Ile Gln Leu
65           70           75           80
Lys Asp Leu Thr Ser Glu Asp Met Gln Gln Leu Gln Ser Met Leu Gly
85           90           95
Cys Glu Val Gln Ala Met Leu Cys Val Pro Val Ile Ser Arg Ala Thr
100          105          110
Asp Gln Val Val Ala Leu Ala Cys Ala Phe Asn Lys Leu Gly Gly Asp
115          120          125
Leu Phe Thr Asp Gln Asp Glu His Val Ile Gln His Cys Phe His Tyr
130          135          140
Thr Ser Thr Val Leu Thr Ser Thr Leu Ala Phe Gln Lys Glu Gln Lys
145          150          155          160
Leu Lys Cys Glu Cys Gln Ala Leu Leu Gln Val Ala Lys Asn Leu Phe
165          170          175
Thr His Leu Asp Asp Val Ser Val Leu Leu Gln Glu Ile Ile Thr Glu
180          185          190
Ala Arg Asn Leu Ser Asn Ala Glu Ile Cys Ser Val Phe Leu Ile Asp
195          200          205
Gln Asn Glu Leu Val Ala Lys Val Phe Asp Gly Gly Val Leu Glu Asp
210          215          220
Glu Ser Tyr Glu Ile Arg Ile Pro Ala Asp Gln Gly Ile Ala Gly His
225          230          235          240
Val Ala Thr Thr Gly Gln Ile Leu Asn Ile Pro Asp Ala Tyr Ala His
245          250          255
Pro Leu Phe Tyr Arg Gly Val Asp Asp Ser Thr Gly Arg Phe Thr Arg
260          265          270
Asn Ile Leu Cys Phe Pro Ile Lys Asn Glu Asn Gln Glu Val Ile Gly
275          280          285
Val Ala Glu Leu Val Asn Lys Ile Asn Gly Pro Trp Phe Ser Lys Phe
290          295          300
Asp Glu Asp Leu Ala Thr Ala Phe Ser Ile Tyr Cys Gly Ile Ser Ile
305          310          315          320
Ala His Ser Leu Leu Tyr Lys Lys Val Asn Glu Ala Gln Tyr Arg Ser
325          330          335
His Leu Ala Asn Glu Met Met Met Tyr His Met Lys Val Ser Asp Asp
340          345          350

```

Glu

<210> SEQ ID NO 35

<211> LENGTH: 402

<212> TYPE: PRT

<213> ORGANISM: bovine

-continued

<400> SEQUENCE: 35

Leu Leu Glu Val Leu Leu Glu Glu Ala Gly Ser Val Glu Leu Ala Ala
 1 5 10 15
 His Arg Ala Leu Gln Arg Leu Ala Gln Leu Leu Gln Ala Asp Arg Cys
 20 25 30
 Ser Met Phe Leu Cys Arg Ala Arg Asn Gly Thr Pro Glu Val Ala Ser
 35 40 45
 Lys Leu Leu Asp Val Thr Pro Thr Ser Lys Phe Glu Asp Asn Leu Val
 50 55 60
 Val Pro Asp Arg Glu Ala Val Phe Pro Leu Asp Val Gly Ile Val Gly
 65 70 75 80
 Trp Val Ala His Thr Lys Lys Thr Phe Asn Val Pro Asp Val Lys Lys
 85 90 95
 Asn Ser His Phe Ser Asp Phe Met Asp Lys Gln Thr Gly Tyr Val Thr
 100 105 110
 Arg Asn Leu Leu Ala Thr Pro Ile Val Met Gly Lys Glu Val Leu Ala
 115 120 125
 Val Phe Met Ala Val Asn Lys Val Asp Ala Ser Glu Phe Ser Lys Gln
 130 135 140
 Asp Glu Glu Val Phe Ser Lys Tyr Leu Ser Phe Val Ser Ile Ile Leu
 145 150 155 160
 Lys Leu His His Thr Asn Tyr Leu Tyr Asn Ile Glu Ser Arg Arg Ser
 165 170 175
 Gln Ile Leu Met Trp Ser Ala Asn Lys Val Phe Glu Glu Leu Thr Asp
 180 185 190
 Val Glu Arg Gln Phe His Lys Ala Leu Tyr Thr Val Arg Thr Tyr Leu
 195 200 205
 Asn Cys Glu Arg Tyr Ser Ile Gly Leu Leu Asp Met Thr Lys Glu Lys
 210 215 220
 Glu Phe Tyr Asp Glu Trp Pro Val Lys Pro Gly Glu Val Glu Pro Tyr
 225 230 235 240
 Lys Gly Pro Lys Thr Pro Asp Gly Arg Glu Val Ile Phe Tyr Lys Ile
 245 250 255
 Ile Asp Tyr Ile Leu His Gly Lys Glu Glu Ile Lys Val Ile Pro Thr
 260 265 270
 Pro Pro Met Asp His Trp Thr Leu Ile Ser Gly Leu Pro Thr Tyr Val
 275 280 285
 Ala Glu Asn Gly Phe Ile Cys Asn Met Leu Asn Ala Pro Ala Asp Glu
 290 295 300
 Tyr Phe Thr Phe Gln Lys Gly Pro Val Asp Glu Thr Gly Trp Val Ile
 305 310 315 320
 Lys Asn Val Leu Ser Leu Pro Ile Val Asn Lys Lys Glu Asp Ile Val
 325 330 335
 Gly Val Ala Thr Phe Tyr Asn Arg Lys Asp Gly Lys Pro Phe Asp Glu
 340 345 350
 Tyr Asp Glu His Ile Ala Glu Thr Leu Thr Gln Phe Leu Gly Trp Ser
 355 360 365
 Leu Leu Asn Thr Asp Thr Tyr Glu Lys Met Asn Lys Leu Glu Asn Arg
 370 375 380
 Lys Asp Ile Ala Gln Glu Met Leu Met Asn His Thr Lys Ala Thr Pro

-continued

385 390 395 400

Asp Glu

<210> SEQ ID NO 36
 <211> LENGTH: 404
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 36

Leu Phe Glu Leu Val Gln Asp Met Gln Glu Asn Val Asn Met Glu Arg
 1 5 10 15

Val Val Phe Lys Ile Leu Arg Arg Leu Cys Ser Ile Leu His Ala Asp
 20 25 30

Arg Cys Ser Leu Phe Met Tyr Arg Gln Arg Asn Gly Val Ala Glu Leu
 35 40 45

Ala Thr Arg Leu Phe Ser Val Gln Pro Asp Ser Val Leu Glu Asp Cys
 50 55 60

Leu Val Pro Pro Asp Ser Glu Ile Val Phe Pro Leu Asp Ile Gly Val
 65 70 75 80

Val Gly His Val Ala Gln Thr Lys Lys Met Val Asn Val Gln Asp Val
 85 90 95

Met Glu Cys Pro His Phe Ser Ser Phe Ala Asp Glu Leu Thr Asp Tyr
 100 105 110

Val Thr Arg Asn Ile Leu Ala Thr Pro Ile Met Asn Gly Lys Asp Val
 115 120 125

Val Ala Val Ile Met Ala Val Asn Lys Leu Asp Gly Pro Cys Phe Thr
 130 135 140

Ser Glu Asp Glu Asp Val Phe Leu Lys Tyr Leu Asn Phe Gly Thr Leu
 145 150 155 160

Asn Leu Lys Ile Tyr His Tyr Ser Tyr Leu His Asn Cys Glu Thr Arg
 165 170 175

Arg Gly Gln Val Leu Leu Trp Ser Ala Asn Lys Val Phe Glu Glu Leu
 180 185 190

Thr Asp Ile Glu Arg Gln Phe His Lys Ala Phe Tyr Thr Val Arg Ala
 195 200 205

Tyr Leu Asn Cys Asp Arg Tyr Ser Val Gly Leu Leu Asp Met Thr Lys
 210 215 220

Glu Lys Glu Phe Phe Asp Val Trp Pro Val Leu Met Gly Glu Ala Gln
 225 230 235 240

Ala Tyr Ser Gly Pro Arg Thr Pro Asp Gly Arg Glu Ile Leu Phe Tyr
 245 250 255

Lys Val Ile Asp Tyr Ile Leu His Gly Lys Glu Asp Ile Lys Val Ile
 260 265 270

Pro Ser Pro Pro Ala Asp His Trp Ala Leu Ala Ser Gly Leu Pro Thr
 275 280 285

Tyr Val Ala Glu Ser Gly Phe Ile Cys Asn Ile Met Asn Ala Pro Ala
 290 295 300

Asp Glu Met Phe Asn Phe Gln Glu Gly Pro Leu Asp Asp Ser Gly Trp
 305 310 315 320

Ile Val Lys Asn Val Leu Ser Met Pro Ile Val Asn Lys Lys Glu Glu
 325 330 335

Ile Val Gly Val Ala Thr Phe Tyr Asn Arg Lys Asp Gly Lys Pro Phe

-continued

340	345	350
Asp Glu Gln Asp Glu Val Leu Met Glu Ser Leu Thr Gln Phe Leu Gly		
355	360	365
Trp Ser Val Leu Asn Thr Asp Thr Tyr Asp Lys Met Asn Lys Leu Glu		
370	375	380
Asn Arg Lys Asp Ile Ala Gln Asp Met Val Leu Tyr His Val Arg Cys		
385	390	395
Asp Arg Glu Glu		

<210> SEQ ID NO 37
 <211> LENGTH: 401
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 37

Leu Leu Arg Asp Phe Gln Asp Asn Leu Gln Ala Glu Lys Cys Val Phe		
1	5	10
Asn Val Met Lys Lys Leu Cys Phe Leu Leu Gln Ala Asp Arg Met Ser		
20	25	30
Leu Phe Met Tyr Arg Ala Arg Asn Gly Ile Ala Glu Leu Ala Thr Arg		
35	40	45
Leu Phe Asn Val His Lys Asp Ala Val Leu Glu Glu Cys Leu Val Ala		
50	55	60
Pro Asp Ser Glu Ile Val Phe Pro Leu Asp Met Gly Val Val Gly His		
65	70	75
Val Ala Leu Ser Lys Lys Ile Val Asn Val Pro Asn Thr Glu Glu Asp		
85	90	95
Glu His Phe Cys Asp Phe Val Asp Thr Leu Thr Glu Tyr Gln Thr Lys		
100	105	110
Asn Ile Leu Ala Ser Pro Ile Met Asn Gly Lys Asp Val Val Ala Ile		
115	120	125
Ile Met Ala Val Asn Lys Val Asp Gly Pro His Phe Thr Glu Asn Asp		
130	135	140
Glu Glu Ile Leu Leu Lys Tyr Leu Asn Phe Ala Asn Leu Ile Met Lys		
145	150	155
Val Phe His Leu Ser Tyr Leu His Asn Cys Glu Thr Arg Arg Gly Gln		
165	170	175
Ile Leu Leu Trp Ser Gly Ser Lys Val Phe Glu Glu Leu Thr Asp Ile		
180	185	190
Glu Arg Gln Phe His Lys Ala Leu Tyr Thr Val Arg Ala Phe Leu Asn		
195	200	205
Cys Asp Arg Tyr Ser Val Gly Leu Leu Asp Met Thr Lys Gln Lys Glu		
210	215	220
Phe Phe Asp Val Trp Pro Val Leu Met Gly Glu Ala Pro Pro Tyr Ala		
225	230	235
Gly Pro Arg Thr Pro Asp Gly Arg Glu Ile Asn Phe Tyr Lys Val Ile		
245	250	255
Asp Tyr Ile Leu His Gly Lys Glu Asp Ile Lys Val Ile Pro Asn Pro		
260	265	270
Pro Pro Asp His Trp Ala Leu Val Ser Gly Leu Pro Thr Tyr Val Ala		
275	280	285
Gln Asn Gly Leu Ile Cys Asn Ile Met Asn Ala Pro Ser Glu Asp Phe		

-continued

290	295	300
Phe Ala Phe Gln Lys	Glu Pro Leu Asp Glu Ser Gly Trp Met Ile Lys	
305	310	315 320
Asn Val Leu Ser Met	Pro Ile Val Asn Lys Lys Glu Glu Ile Val Gly	
	325	330 335
Val Ala Thr Phe Tyr	Asn Arg Lys Asp Gly Lys Pro Phe Asp Glu Met	
	340	345 350
Asp Glu Thr Leu Met	Glu Ser Leu Ala Gln Phe Leu Gly Trp Ser Val	
	355	360 365
Leu Asn Pro Asp Thr Tyr	Glu Leu Met Asn Lys Leu Glu Asn Arg Lys	
	370	375 380
Asp Ile Phe Gln Asp Met	Val Lys Tyr His Val Lys Cys Asp Asn Glu	
385	390	395 400

Glu

<210> SEQ ID NO 38
 <211> LENGTH: 84
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 38

Glu Pro Leu Asn Ile Lys Asp Ala Tyr Glu Asp Pro Arg Phe Asn Ala																			
1			5					10											15
Glu Val Asp Gln Ile Thr Gly Tyr Lys Thr Gln Ser Ile Leu Cys Met																			
			20					25											30
Pro Ile Lys Met His Arg Glu Glu Val Val Gly Val Ala Gln Ala Ile																			
			35					40											45
Asn Lys Lys Ser Gly Asn Gly Gly Thr Phe Thr Glu Lys Asp Glu Lys																			
			50					55											60
Asp Phe Ala Ala Tyr Leu Ala Phe Cys Gly Ile Val Leu His Met Ala																			
65			70																80

Gln Leu Tyr Glu

<210> SEQ ID NO 39
 <211> LENGTH: 81
 <212> TYPE: PRT
 <213> ORGANISM: bovine

<400> SEQUENCE: 39

Lys Ile Val Asn Val Pro Asn Thr Glu Glu Asp Glu His Phe Cys Asp																			
1			5						10										15
Phe Val Asp Thr Leu Thr Glu Tyr Gln Thr Lys Asn Ile Leu Ala Ser																			
			20						25										30
Pro Ile Met Asn Gly Lys Asp Val Val Ala Ile Ile Met Ala Val Asn																			
			35						40										45
Lys Val Asp Gly Pro His Phe Thr Glu Asn Asp Glu Glu Ile Leu Leu																			
			50						55										60
Lys Tyr Leu Asn Phe Ala Asn Leu Ile Met Lys Val Phe His Leu Ser																			
65			70																80

Tyr

<210> SEQ ID NO 40
 <211> LENGTH: 81
 <212> TYPE: PRT

-continued

<213> ORGANISM: bovine

<400> SEQUENCE: 40

Lys Met Val Asn Val Gln Asp Val Met Glu Cys Pro His Phe Ser Ser
 1 5 10 15
 Phe Ala Asp Glu Leu Thr Asp Tyr Val Thr Arg Asn Ile Leu Ala Thr
 20 25 30
 Pro Ile Met Asn Gly Lys Asp Val Val Ala Val Ile Met Ala Val Asn
 35 40 45
 Lys Leu Asp Gly Pro Cys Phe Thr Ser Glu Asp Glu Asp Val Phe Leu
 50 55 60
 Lys Tyr Leu Asn Phe Gly Thr Leu Asn Leu Lys Ile Tyr His Leu Ser
 65 70 75 80

Tyr

<210> SEQ ID NO 41

<211> LENGTH: 81

<212> TYPE: PRT

<213> ORGANISM: bovine

<400> SEQUENCE: 41

Lys Thr Phe Asn Val Pro Asp Val Lys Lys Asn Ser His Phe Ser Asp
 1 5 10 15
 Phe Met Asp Lys Gln Thr Gly Tyr Val Thr Arg Asn Ile Leu Ala Thr
 20 25 30
 Pro Ile Val Met Gly Lys Glu Val Leu Ala Val Phe Met Ala Val Asn
 35 40 45
 Lys Val Asp Ala Ser Glu Phe Ser Lys Gln Asp Glu Glu Val Phe Ser
 50 55 60
 Lys Tyr Leu Ser Phe Val Ser Ile Ile Leu Lys Leu His His Thr Asn
 65 70 75 80

Tyr

<210> SEQ ID NO 42

<211> LENGTH: 81

<212> TYPE: PRT

<213> ORGANISM: bovine

<400> SEQUENCE: 42

Lys Ser Ile Gln Leu Lys Asp Leu Thr Ser Glu Asp Met Gln Gln Leu
 1 5 10 15
 Gln Ser Met Leu Gly Cys Glu Val Gln Ala Met Leu Cys Val Pro Val
 20 25 30
 Ile Ser Arg Ala Thr Asp Gln Val Val Ala Leu Ala Cys Ala Phe Asn
 35 40 45
 Lys Leu Gly Gly Asp Leu Phe Thr Asp Gln Asp Glu His Val Ile Gln
 50 55 60
 His Cys Phe His Tyr Thr Ser Thr Val Leu Thr Ser Thr Leu Ala Phe
 65 70 75 80

Gln

<210> SEQ ID NO 43

<211> LENGTH: 91

<212> TYPE: PRT

<213> ORGANISM: bovine

-continued

<400> SEQUENCE: 43

```

Glu Pro Leu Asn Ile Pro Asp Val Ser Lys Asp Lys Arg Phe Pro Trp
1           5           10           15
Thr Asn Glu Asn Met Gly Asn Ile Asn Gln Gln Cys Ile Arg Ser Leu
          20           25           30
Leu Cys Thr Pro Ile Lys Asn Gly Lys Lys Asn Lys Val Ile Gly Val
          35           40           45
Cys Gln Leu Val Asn Lys Met Glu Glu Thr Thr Gly Lys Val Lys Ala
          50           55           60
Phe Asn Arg Asn Asp Glu Gln Phe Leu Glu Ala Phe Val Ile Phe Cys
65           70           75           80
Gly Leu Gly Ile Gln Asn Thr Gln Met Tyr Glu
          85           90

```

<210> SEQ ID NO 44

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: bovine

<400> SEQUENCE: 44

```

Leu Ile Cys Asn Ile Met Asn Ala Pro Ser Glu Asp Phe Phe Ala Phe
1           5           10           15
Gln Lys Glu Pro Leu Asp Glu Ser Gly Trp Met Ile Lys Asn Val Leu
          20           25           30
Ser Met Pro Ile Val Asn Lys Lys Glu Glu Ile Val Gly Val Ala Thr
          35           40           45
Phe Tyr Asn Arg Lys Asp Gly Lys Pro Phe Asp Glu Met Asp Glu Thr
          50           55           60
Leu Met Glu Ser Leu Ala Gln Phe Leu Gly Trp Ser Val Leu Asn Pro
65           70           75           80
Asp Thr Tyr Glu

```

<210> SEQ ID NO 45

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: bovine

<400> SEQUENCE: 45

```

Phe Ile Cys Asn Ile Met Asn Ala Pro Ala Asp Glu Met Phe Asn Phe
1           5           10           15
Gln Glu Gly Pro Leu Asp Asp Ser Gly Trp Ile Val Lys Asn Val Leu
          20           25           30
Ser Met Pro Ile Val Asn Lys Lys Glu Glu Ile Val Gly Val Ala Thr
          35           40           45
Phe Tyr Asn Arg Lys Asp Gly Lys Pro Phe Val Glu Gln Asp Glu Val
          50           55           60
Leu Met Glu Ser Leu Thr Gln Phe Leu Gly Trp Ser Val Leu Asn Thr
65           70           75           80
Asp Thr Tyr Asp

```

<210> SEQ ID NO 46

<211> LENGTH: 84

<212> TYPE: PRT

<213> ORGANISM: bovine

-continued

<400> SEQUENCE: 46

```

Phe Ile Cys Asn Met Leu Asn Ala Pro Ala Asp Glu Tyr Phe Thr Phe
1           5           10           15
Gln Lys Gly Pro Val Asp Glu Thr Gly Trp Val Ile Lys Asn Val Leu
                20           25           30
Ser Leu Pro Ile Val Asn Lys Lys Glu Asp Ile Val Gly Val Ala Thr
                35           40           45
Phe Tyr Asn Arg Lys Asp Gly Lys Pro Phe Asp Glu Tyr Asp Glu His
                50           55           60
Ile Ala Glu Thr Leu Thr Gln Phe Leu Gly Trp Ser Leu Leu Asn Thr
65           70           75           80
Asp Thr Tyr Glu

```

```

<210> SEQ ID NO 47
<211> LENGTH: 82
<212> TYPE: PRT
<213> ORGANISM: bovine

```

<400> SEQUENCE: 47

```

Gln Ile Leu Asn Ile Pro Asp Ala Tyr Ala His Pro Leu Phe Tyr Arg
1           5           10           15
Gly Val Asp Asp Ser Thr Gly Phe Arg Thr Arg Asn Ile Leu Cys Phe
                20           25           30
Pro Ile Lys Asn Glu Asn Gln Glu Val Ile Gly Val Ala Glu Leu Val
                35           40           45
Asn Lys Ile Asn Gly Pro Trp Phe Ser Lys Phe Asp Glu Asp Leu Ala
                50           55           60
Thr Ala Phe Ser Ile Tyr Cys Gly Ile Ser Ile Ala His Ser Leu Leu
65           70           75           80
Tyr Lys

```

1. A purified and isolated polynucleotide encoding cGB-PDE.

2. The polynucleotide of claim 1 which is a DNA sequence.

3. The DNA sequence of claim 2 which is a cDNA sequence or a biological replica thereof.

4. The DNA sequence of claim 2 which is a genomic DNA sequence or a biological replica thereof.

5. An RNA transcript of the genomic DNA sequence of claim 4.

6. The DNA sequence of claim 2 which is a wholly or partially chemically synthesized DNA sequence or a biological replica thereof.

7. The DNA sequence of claim 4 further comprising an endogenous expression control DNA sequence.

8. A DNA vector comprising a DNA sequence according to claim 2.

9. The vector of claim 8 wherein said DNA sequence is operatively linked to an expression control DNA sequence.

10. A host cell stably transformed or transfected with a DNA sequence according to claim 7 in a manner allowing the expression in said host cell of cGB-PDE polypeptide

possessing a ligand/receptor binding biological activity or immunological property specific to cGB-PDE.

11. A method for producing cGB-PDE polypeptide, said method comprising growing a host cell according to claim 10 in a suitable nutrient medium and isolating cGB-PDE polypeptide from said cell or the medium of its growth.

12. A polypeptide or peptide capable of specifically binding to cGB-PDE.

13. An antibody substance according to claim 12.

14. A monoclonal antibody according to claim 13.

15. A hybridoma cell line producing a monoclonal antibody according to claim 14.

16. A humanized antibody substance according to claim 13.

17. An antisense polynucleotide specific for a polynucleotide encoding cGB-PDE.

18. A DNA sequence encoding cGB-PDE and selected from the group consisting of:

(a) the DNA sequence set out in SEQ ID NO: 9 or 22;

(b) a DNA which hybridizes under stringent conditions to the DNA of (a); and

(c) a DNA sequence which, but for the redundancy of the generic code, would hybridize under stringent conditions to a DNA sequence of (a) or (b).

19. A method for modulating the enzymatic activity of cGB-PDE, comprising contacting cGB-PDE with an effective amount of an agent that binds CGB-PDE and activates or inhibits cGB-PDE.

20. The method of claim 19 wherein the agent that binds cGB-PDE is selected from the group consisting of antibodies, peptides, proteins, oligonucleotides, antisense molecules, non-peptide compounds, and small molecules.

21. The method of claim 20, wherein the agent that binds cGB-PDE is an anti-cGB-PDE antibody.

22. A method for identifying an agent that specifically binds to cGB-PDE comprising:

(a) contacting cGB-PDE with an effective amount of a test agent; and

(b) determining if the test agent specifically binds cGB-PDE.

23. A method for identifying an agent that specifically binds to cGB-PDE so as to modulate the enzymatic activity of cGB-PDE comprising:

(a) contacting cGB-PDE with an effective amount of a test agent;

(b) determining if the test agent specifically binds cGB-PDE; and

(c) testing for modulation of cGB-PDE activity.

24. The method of claim 23, wherein the cGB-PDE is recombinant cGB-PDE.

25. The method of claim 23, wherein the agent is selected from the group consisting of antibodies, peptides, proteins, oligonucleotides, antisense molecules, non-peptide compounds, and small molecules.

26. A method of using an agent that modulates the enzymatic activity of cGB-PDE for treating a condition that involves signal transduction pathways utilizing cyclic nucleotides as second messengers, comprising administering to a subject an agent that modulates the activity of cGB-PDE.

27. An agent identified by the method of claim 24.

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