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A vaccine effective against Marek's disease virus (MDV) comprises an MDV attenuated by virtue of being TK- or a host expressing an MDV antigen, namely the respective MDV homologues of the HSV gB, gC, gD or gH glycoproteins (or antigenic parts thereof) or the respective MDV homologues of the HSV-1 immediate early genes IE-68 or IE-175. The host may be a herpes virus of turkeys (HVT), more particularly HVT in which the MDV antigen is inserted in the HVT homologue of the HSV gC gene, the ribonucleotide reductase (large subunit) gene or the thyamidine kinase (TK) gene.



## **(54) Title: VIRAL VACCINES**

### **(57) Abstract**

A vaccine effective against Marek's disease virus (MDV) comprises (a) an MDV attenuated by virtue of being TK- or (b) a host expressing an MDV antigen, namely the respective MDV homologues of the HSV gB, gC, gD or gH glycoproteins (or antigenic parts thereof) or the respective MDV homologues of the HSV-1 immediate early genes IE-68 or IE-175. The host may be a herpes virus of turkeys (HVT), more particularly HVT in which the MDV antigen is inserted in the HVT homologue of the HSV **gC gene, the ribonucleotide reductase (large subunit) gene or the thymidine kinase (TK) gene.**

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## **VIRAL VACCINES**

The present invention relates to viral vaccines which may be used to provide immunity against disease and to nucleotide sequences for inclusion in the viruses of such vaccines.

## Background and Description of prior art

Herpesviruses are large double stranded DNA viruses consisting of an icosahedral capsid surrounded by an envelope. The group has been classified as alpha, beta and gammaherpesviruses on the basis of genome structure and biological properties [Roizman, B et al (1981) Inter-virology 16, 201-217], Avian herpes viruses include Marek's Disease Virus (MDV) (a gammaherpesvirus) which causes a lymphomatous disease of considerable economic importance in chickens [reviewed in Payne, L.N. (ed) Marek's Disease (1985), Martinus Nijhoff Publishing, Boston] and Infectious Laryngotracheitis Virus (ILTV) (an alphaherpesvirus) which causes an acute upper respiratory tract infection in chickens resulting in mortality and loss of egg production.

A recent unexpected finding in our laboratory is that there is sufficient amino acid homology between MDV, ILTV and mammalian herpesviruses, particularly varicella zoster (VZV) and Herpes Simplex Virus (HSV) to allow identification of numerous conserved genes. These include the MDV and Herpesvirus of

Turkeys (HVT) homologues of glycoproteins gB, gC and gH of HSV; the ILTV, MDV and HVT homologues of TK and ribonucleotide reductase genes and the ILTV homologue of gB and genes 34 and 35 of VZV fBuckmaster, A et al, (1988) J. gen. Virol, 69, 2033 2042).

Strains of MDV have been classified into three serotypes. Type <sup>1</sup> comprises pathogenic strains and their attenuated derivatives. Type <sup>2</sup> are a group of naturally-occurring nonpathogenic strains and type <sup>3</sup> is HVT. For more than a decade, vaccination with HVT has been remarkably effective in controlling Marek's disease. However, in recent years, new strains of MDV have been isolated which cause disease despite vaccination with HVT. Losses due to these 'very virulent' strains have occurred in parts of the U.S.A., Europe and the Middle East. Although the degree of protection can be improved by using a mixture of HVT, type <sup>2</sup> MDV and attenuated derivatives of very virulent strains for vaccination, the results have been erratic. These observations and the fact that there are MDV type-specific epitopes that are not shared by HVT or type <sup>2</sup> MDV have led us to the conclusion that improved vaccines might be constructed which are antigenically more related to MDV than existing vaccines. [Reviewed by Ross and Biggs in Goldman J.M. and Epstein M.A. (eds) Leukaemia and Lymphoma Research, Vaccine Intervention against Virus-Induced Tumour, p 13-31, Macmillan, 1986].

A number of herpesvirus antigens have been shown to confer protective immunity when expressed in a recombinant vaccinia virus. These include the gB gene of HSV [Cantin E.M. et al {1987) Proc, Natl. Acad. Sci, U.S.A. 84, 5908-5912], gD of HSV [Paoletti, E. et al (1984) Proc. Natl. Acad. Sci. U.S.A. 81, 193-197 ] and gp50 of pseudorabies virus (PRV), a homologue of HSV gD [Marchioli, C.C. et al (1987) J. Virol. 61, 3977-3981]. Because of the absolute requirement of gB for virus penetration and infectivity and because it is conserved among herpesviruses, gB and its homologues are important immunogens. Moreover, the presence of gB at the surface of infected cells has been shown to be an important target for humoral and cellmediated immune responses [Blacklaws, B.A. et al J.gen. Virol. 68, 1103-1114 (1987); McLaughlin-Taylor, E. et al (1988) J. gen. Virol. 69, 1731-1734]. The recently described glycoprotein gH of HSV is also essential for infectivity and may also be an important immunogen [Desai, P.J. et al (1988) J. gen. Virol. 69, 1147-1156]. It has also been shown that gIII of pseudorabies virus (PRV), a homologue of gC, is a major target for neutralizing antibody and for cytotoxic T cells although it is a non-essential protein. Also of interest is the unexpected participation of immediate early proteins in T cell mediated cytotoxic reactions in cells infected with cytomegalovirus (CMV) [Kozinowski U.H. et al (1987) J. Virol. 61, 2054-2058]. Similar antigens could play an important role in the rejection of latently infected and transformed lymphocytes in Marek's

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disease since immediate early RNA transcripts have been detected in lymphoblastoid cell lines established from Marek'<sup>s</sup> disease tumours.

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Although many recombinant vaccines have been constructed using' the poxvirus vaccinia as a vector, there are also reports of the use of herpesviruses as vectors for the expression of foreign genes. Thus hepatitis antigen has been expressed in HSV [Shih, M.F. et al (1984) Proc. Natl. Acad. Sci. U.S.A. 81, 5867-5870) and human tissue plasminogen activator has been expressed in PRV [Thomsen, D.R. et al (1987) Gene 57, 261-265. In both cases, foreign genes were inserted in cloned fragments of non-essential herpes genes which were then introduced into the virus vector by homologous recombination. The hepatitis virus gene was fused to a herpesvirus promoter and the recombinant DNA was inserted within the TK gene of HSV. Homologous recombination following co-transfection of the •recombinant DNA and wild-type HSV DNA resulted in TK- virus clones that expressed the hepatitis antigen.

In the case of PRV, the gX gene mapping in Us was used as the site for insertion of the foreign gene. The strategy used involved insertion of the TK gene of HSV in the gX gene of a PRV mutant that had a defect in its TK gene resulting in a TK positive virus. The human tissue plasminogen activator gene was then inserted within a cloned fragement of HSV TK and the

recombinant was introduced into the PRV mutant by homologous recombination. TK- virus was selected which expressed the human gene (Thomsen et al as above). Similarly, VZV has been used as a vector [Lowe et al (1987) Proc. Natl. Acad. Sci. U.S.A. 84, 3896-3900]. Several herpesvirus genes have also been shown to be associated with virulence and to be non-essential for growth in vitro. These include the TK genes of HSV [Jamieson, A.T. et al (1974) J. gen. Virol. 24, 466-480) Field, H. and Wildy, P., (1987) J. Hygiene (Cambridge) 81, 267-277] and of PRV. Indeed it has long been known that PRV is readily attenuated by deletion of TK activity [Tatarov, G. (1968) Zentralbl. Vet. Med 15B, 848-853]. Furthermore, attenuation of the Eartha strain of PRV has been attributed to a defect in gI, a non-essential structural glycoprotein mapping in U<sup>s</sup> [Mettenleiter, T. et al (1987) J. Virol. 61, 4030-4032].

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Genes of HSV mapping in the internal repeat region (TRS) flanking the long unique sequence have also been associated with pathogenicity [Rosen, A. et al (1986) Virus Research 5, 157-175) Thompson, R.L. et al (1983) Virology 131, 180-192]. Several additional genes of HSV have been shown to be nonessential for growth in vitro although it is not known whether they are associated with virulence. These include UL24 (Sanders, P.G., (1982), J. gen. Virol. 63, 277-295, large subunit of ribonucleotide reductase (Goldstein D.J. and Weller, S.K. (1988) J. Virol. 62, 196-205), gC (Draper K.G. et al

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(1984) J. Virol. 51, 578-585), dUTPase (Fisher, F.B. & Preston, V.G. (1986) Virology 148, 190-197), and U<sup>l</sup> 55 and U<sup>l</sup> 56 (MacLean, A.R. & Brown, S.M. (1987) J. gen. Virol. 68, 1339 — 1350). Moreover there is evidence that several genes of HSV mapping in Us are also non-essential for growth in vitro [Weber, P.C. et al (1987) Science 236, 576-579].

WO 88/07088 (published only on 22 September 1988) disclosed hybrid viral vectors based on HVT or MDV and including a gene of interest in a non-essential site, such as the TK region or the region encoding protein A. Protein A, in this context, appears to be the same as gC, disclosed by Velicer and Coussens.

### Summary of the invention

One aspect of the present invention provides a nucleotide sequence substantially free of the sequences which would adjoin it in the wild-type virus associated with the sequence, the sequence being selected from the group consisting of: (a) the MDV homologue of the HSV gB gene,

(b) the MDV homologue of the HSV gH gene,

(c) the TK gene of MDV,

(d) the MDV homologue of the immediate early gene IE-175 of HSV-1

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(e) the MDV homologue of the immediate early gene IE-68 **of HSV-I**

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(f) the MDV homologue of the HSV gD gene

and minor variations thereof.

In addition, the TK sequence of HVT, referred to hereinafter sometimes as sequence  $(x)$ , and the MDV analogue of HSV  $gC$ , referred to hereinafter sometimes as sequence (y), and minor variations of either may be used as insertion sites for certain heterologous sequences or as deletion sites to obtain less virulent viruses but are not novel per se.

Each of sequences (a) to  $(f)$ ,  $(x)$  and  $(y)$  may be associated with further elements such as suitable stop and start signals and other S' and 3' non-coding sequences, including promoters, enabling expression of the sequence. Such further elements may ibe those associated with the sequence in its naturallyoccurring state or may be heterologous to that sequence.

In particular the promoter may be one associated with one of the sequences (d) and (f) above.

The term "minor variations thereof" is intended to include .changes in the nucleotide sequence which do not affect its essential nature, for example minor substitutions of

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nucleotides for one another. In the case of sequences which are intended for insertion into a vector to encode an antigen, the "essential nature of the sequence refers to the protein or glycoprotein encoded. Conservative changes in the nucleotide •sequence which give rise to the same antigen will clearly be included, as will changes which cause conservative alterations in the amino acid sequence which do not affect adversely the antigenic nature of the antigen, in particular, antigenic portions of the antigen sequences may be used alone, for 'example, the regions corresponding to nucleotides 816-863, 1377-1S95, 1377-1630 or 1824-1985 of MDV gB, or nucleotides 483-633, 843-933 or 1203-1278 or MDV gC, and minor variations thereof. These sequences and the peptides encoded thereby form further aspects of the invention. In the case of a sequence 'which is an insertion site, it is necessary only that the sequence should be non-essential for the infectivity and replication of the virus and have sufficient homology with the defined sequence to enable recombination to occur. Thus an insertion of the nucleotide into the sequence could completely 'change the reading frame from then on in a downstream direction. In the case of an antigen-encoding sequence this would usually alter the amino acid sequence undesirably (depending on where the frameshift occurred) , but in the case of an insertion site, the degree of homology would be almost 'the same, thereby allowing recombination to take place with almost the same ease.

Generally· speaking, in an insertion site, if a nucleotide homology of at least 75% is present, the sequence is regarded as a "minor variation". Preferably, the sequence is at least 80, 85, 90, 95 or 99% homologous. It will be appreciated that such degrees of homology relate to substantially the entire portion of each sequence (a) to (f) and (x) defined above. Shorter sequences may be used as probes in the identification or isolation of such longer sequences, but in this case the degree of homology will in general need to be greater in order to ensure accurate hybridisation.

Thus, a further aspect of the invention provides sub-sequences of at least 13 nucleotides having at least 90% (preferably 95%, 99% or 100%) homology to at least one portion of any of the said sequences (a) to  $(f)$ ,  $(x)$  and  $(y)$  above.

In the above list, sequences (a), (b) and (d) to (f) are useful as antigen-expressing sequences and sequence (y) is useful as an insertion site for heterologous sequences. Sequence (c) is useful for deletion to provide TK- mutants.

The sequences may readily be isolated from naturally-occurring HVT and MDV viruses, using the sequence information given herein and standard techniques, for -example involving the preparation of oligonucleotide probes and use thereof to hybridise to the naturally-occurring DNA.

The isolated polypeptides encoded by sequences (a), (b) and (f) above are novel and form a further aspect of the invention, together with minor variations thereof and any glycosylated forms thereof which result from expression of the said sequences in MDV-susceptible cells.

A second aspect of the invention provides MDV mutants which are insertional or deletional mutants in the TK gene.

The mutation may be in the coding or non-coding sequences of the region identified.

An MDV antigen-expressing gene may be isolated from a virulent strain of MDV and inserted into the TK region of a less virulent strain of MDV; this insertion would result in a novel "virus" if it did not result in a naturally-occurring virus.

Other heterologous antigen-encoding sequences may be included, as well as an MDV antigen-encoding sequence, for example.

The heterologous sequence may alternatively be one coding for an antigen associated with any one of the following diseases: avian encephalomyelitis (epidemic tremor), avian influenza (fowl plague), avian leukosis, avian paramyxoviruses other than Newcastle disease (PMV2 to PMV7), avian reovirus diseases (enteric disease, tenosynovitis) , chicken anaemia (caused by

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chicken anaemia agent), coccidiosis, egg drop syndrome (EDS76), fowl pox, infectious bronchitis, infectious bursal disease (Gumboro), inclusion body hepatitis (adenovirus), lymphoproliferative disease of turkeys, Newcastle disease, reticuloendotheliosis in chickens, reticuloendotheliosis in turkeys, rotavirus enteritis, turkey haemorrhagic enteritis and turkey rhinotracheitis. The sequence may alternatively encode paramyosin (a muscle protein common to all invertebrate parasites) or an antigenic part thereof, somatostatin or a growth-promoting part thereof or an immune regulator.

The vectors in accordance with the invention will then provide multivalent vaccine protection.

The mutant viruses are potentially useful in vaccines as attenuated viruses, without necessarily having a heterologous sequence inserted.

A convenient process . for preparing the deletional or insertional mutants of the second aspect of the invention comprises simply introducing into a suitable cell, for example by co-transfection, a deletional or insertional mutant version of the TK region and either whole viral DNA or a whole virus (for example the wild-type virus). The naked DNA of such viruses has been found to be infectious, provided that it has not been sheared. A calcium phosphate precipitate of the DNA is generally advantageous. Suitable cells include chicken embryo

fibroblasts, chicken kidney cells and duck embryo fibroblasts, all preferably grown in sub-confluent monolayers in Petri dishes. The transfected DNA and the whole viral DNA will then recombine with one another in the infected cells by homologous recombination and the desired recombinants can be screened for, for example by the detection of hybridisation to suitable probes or by an immunoassay using suitable antibodies to the gene product of the region in question.

For homologous recombination to take place, the viral DNA must replicate. At present, no cell-free replication system for MDV is known. However, if such a system becomes available, then the process of the invention could be operated therein. The environment in which the replication and recombination occur is not critical.

Regions (a), (b) and (d) to (f), which were identified above as being responsible for encoding immunologically useful viral antigens, can be inserted into suitable vectors, for example into HVT or other vectors such as fowlpox-virus, bacteria or fungi. In the case of viral vectors, especially herpesvirus vectors and poxvirus vectors, such insertion can be achieved by recombination between the antigen-encoding sequence, flanked by suitable non-essential sequences, and the vector'<sup>s</sup> genome in a suitable host cell as described above. When HVT is the vector, the promoter will usually be an HVT or MDV vector. When fowlpox-virus or other virus is the vector, the promoter will

usually be a promoter which is endogenous to the vector. In the case of bacteria and fungi, the antigen-encoding sequence may be inserted using known or yet-to-be-discovered techniques of DNA manipulation. A non-pathogenic strain of Salmonella may be used as such a host. The heterologous sequence may be inserted into the host's genome or be carried on an independently-replicating plasmid. A promoter which is endogenous to the host will usually be used to control expression of the heterologous , (viral antigen-encoding) sequence .

The flanking sequences which are used may comprise all, virtually all or less of the region into which the heterologous sequence is to be inserted. If all the region is employed, then the sequence of that region will clearly still be present in •the resulting virus, but the function of that region will have been deleted. If less than the whole region is used as flanking sequences, then the result will be a structural as well as functional deletion. Either approach may be used.

Thus, three strategies can be envisaged for the construction of -improved Marek'<sup>s</sup> disease vaccines: (1) Construction of recombinant HVT that express selected MDV genes; (2) Construction of deletional or insertional mutants of highly virulent strains of MDV, which are attenuated and hence suitable for use in vaccines; (3) Construction of recombinant •viruses that express MDV proteins in other vectors such as fowl pox virus .

To prepare a vaccine in which HVT or MDV is the virus or vector, the virus is grown in suitable cells such as chick embryo fibroblasts in a standard culture medium which as 199 medium (Wellcome or Flow Laboratories) for <sup>3</sup> to <sup>4</sup> days at about 37oC. The cells are harvested by trypsinisation and suspended in medium containing 10% dimethyl sulphoxide and 4% calf serum before storage in liquid nitrogen in sealed ampoules.

For vaccination, typically, day-old chicks are injected intramuscularly with about 1,000 plaque-forming units. Immunity follows within a few days.

It should be noted that MDV and HVT are cell-associated viruses and are infectious only when present in cells. Thus, a vaccine based on such viruses will always include suitable infected cells.

The vaccines of the invention may be used to protect any fowl susceptible to MDV, including commercially-reared poultry such as chickens, turkeys, ducks and quail.

Preferred aspects of the invention will now be described by way of example and with reference to the accompanying drawings, in which :

Figure 1 is a map of the MDV genome showing in part the BamH1 site distribution and the location of the gB and TK genes;

Figure <sup>2</sup> (on 18 sheets) shows the nucleotide sequence of the gB gene of the RBIB strain of MDV, with the numbering referring to the MDV nucleotides, the sequence of part of the HVT gB gene shown under the line, homologies indicated by vertical bars, and amino acid differences between MDV gB and HVT gB shown above the line;

Figure <sup>3</sup> is a map of the HVT genome showing the positions of the gH (hatched), TK (solid black) and major capsid protein (MCP, dotted) genes, with Hindlll sites shown as "H";

Figure <sup>4</sup> (on <sup>8</sup> sheets) shows the nucleotide sequence of most of the HVT gH gene, with the corresponding amino acid sequence shown above the line;

Figure <sup>5</sup> (on 10 sheets) shows the nucleotide sequence of the HVT TK gene, with the numbering referring to the HVT nucleotides, the sequence of part of the MDV TK gene shown under the line, homologies indicated by vertical bars and amino acid differences between MDV TK and HVT TK shown above the line;

Figure <sup>6</sup> (on <sup>6</sup> sheets) shows the nucleotide sequence of the gC gene of the RBIB strain of MDV, with corresponding amino acids shown above the line;

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Figure 7 shows part of the nucleotide sequence of the HVT homoiogue of the VZV62/HSV-1 IE 175 gene with corresponding amino acids shown above the line;

Figure 8 shows part of the nucleotide sequence of the HVT ribonucleotide reductase (large subunit) gene with corresponding amino acids shown above the line;

Figure <sup>9</sup> (on <sup>2</sup> sheets) shows part of the nucleotide sequence of the MDV ribonucleotide reductase (large subunit) gene with corresponding amino acids shown above the line;

Figure 10 shows part of the nucleotide sequence of the MDV ribonucleotide reductase (small subunit) gene with corresponding amino acids shown above the line; .

Figure 11 shows part of the nucleotide sequence of the MDV homologue of the HSV-1 IE-175 gene with corresponding amino acids shown above the line;

Figure 12 shows part of the MDV homologue of the HSV-1 IE-68 gene with corresponding amino acids shown above the line;

Figure 13 is a schematic representation of homologous recombination at a non-essential region of a viral genome and a homologous region of DNA cloned within a plasmid vector;

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Figure 14 (on 27 sheets) supplements Figures <sup>4</sup> and 5, and shows the nucleotide and predicted amino acid sequences from the region containing the MDV and HVT TK and gH and flanking genes. The bracketed MDV amino acid sequences are those potentially encoded by this region of nucleotide sequence if the upstream ATG triplet were the true gene initiation site. Asterisks denote stop codons. Spaces have been inserted into the sequences in order to optimize alignments. Colons between the MDV and HVT DNA sequences indicate nucleotides conserved between the two viruses. MDV amino acids are only shown in positions where they differ from that in HVT; and

Figure 15 shows the partial nucleotide sequence of the MDV homologue of HSVgD, the predicted amino acids being shown above the MDV nucleotide sequence and residues in bold type being conserved between the MDV and HSV-1 gD regions.

## EXAMPLES: General Approaches

Selected short sequences of the avian herpesviruses cloned in the bacteriophage vector M13 were used as probes to identify longer fragments that might contain the entire genes of interest. This was achieved by Southern blot hybridization of restriction fragments. Full details are given below.

Virus Strains. The 'highly oncogenic' strain RB1B of MDV [Schat, K.A. et al (1982) Avian Pathol. 11, 593-605] was

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obtained from Professor B. Calnek, Cornell University, Ithaca, U.S.A. The virus received has been plaque purified in chicken kidney vells in tissue culture. It was passaged twice in SPF RIR chickens and <sup>4</sup> times in chick embryo fibroblasts (CEF). Its 'highly oncogenic' nature was demonstrated by a high incidence of gross tumours when inoculated in genetically resistant Nline chickens.

The FC126 strain of HVT [Witter, R.L. et al (1970) Am. J. Vet. Res. 31, 525-538], obtained from the Wellcome Research Laboratories, Beckenham, Kent, had been passaged 14 times in CEF. It was subsequently grown in duck embryo fibroblasts (DEF) and CEF in our laboratory. It was then plaque-purified and grown further in CEF. Viral DNA used for cloning in the present work was extracted from virus that had been passed 29 times since the original isolation.

Tissue culture. CEF were grown in roller bottles in 199 medium (Wellejme), supplemented with penicillin, streptomycin, Fungizone (Regd. T.M.) and calf serum as described previously [Ross, L.J.N. et al (1975) J. gen. Virol. 28, 37-47].

CKC were grown in 10 cm Petri dishes [Churchill, A.E. and Biggs P.M., (1967) Nature, 215, 528-530].

Isolation of MDV DNA. Cell associated RB1B was inoculated onto confluent monolayers of CEF in roller bottles at a multiplicity

of infection of approximately 0.001 plaque-forming units (pfu) per cell, and the cultures were incubated at 37°C. After <sup>3</sup> days, the medium was discarded and replaced with fresh 199 medium containing 2% calf serum. Cells were harvested for virus purification after <sup>2</sup> to <sup>3</sup> days when cytopathic effect was extensive. Virus was obtained by rate zonal centrifugation of the cytoplasmic fraction of infected cells [Lee, Y.S. et al. (1980) J. gen. Virol. 51, 245-253]. Viral DNA was extracted by treating purified virus with sarcosyl, proteinase K and Tris buffer pH <sup>9</sup> overnight at 37° C and purified by rate zonal centrifugation in glycerol gradients as described previously (Lee et al, 1980). High molecular weight viral DNA was precipitated with ethanol and resuspended in 10 mM Tris pH 7.5 im ImM EDTA (TE).

Cloning of MDV DNA. One  $\mu$ g of MDV DNA was cut with the restriction enzyme BamHl and ligated to BamHl-cut, dephosphorylated pUC13 DNA (Pharmacia). Competent E.coli strain TGI cells were transformed according to standard procedures [Hanahan, D. (1983) J. Mol. Biol. 16S, 557-580] and were grown in the presence of ampicillin and X-gal. White colonies were picked and tested for the presence or MDV inserts by hybridization to nick-translated MDV DNA [Grunstein M. and Hogness, D.S. (1975) Proc. Natl. Acad. Sci. U.S.A. 72, 3961 ], Positive colonies were cultured in small volume and plasmid DNA isolated by the procedure of Holmes, D.S. and Quigley, M.

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[(1981) Anal. Biochem. 114, 193-297]. The size of the inserts was determined by electrophoresis of BamH<sup>I</sup> digests of the recombinant DNA in agarose gels . Plasmids containing MDV inserts ranging from less than <sup>1</sup> to 18 Kbp were obtained.

Random sequencing of viral DNA. Sonicated fragments of viral DNA were cloned into Smal-cut, dephosphorylated M13.mp10 <sup>|</sup> (Amersham International PLC) and plaques containing MDV inserts were identified by hybridization to MDV DNA. The sequence was determined by the dideoxy method [Sanger, F. et al (1977) Proc. Natl. Acad. Sci. U.S.A. 74, 5463-5467] using 35S dATP).

The same procedure was used to sequence cloned fragments of MDV DNA except that plaques were identified by hybridization to labelled insert so as to avoid colonies containing pUC13 fragments.

### **<sup>i</sup>** EXAMPLE 1: gB gene of MDV

An M13 clone of HVT homologous to the gB gene of VZV and HSV hybridized to BamH<sup>I</sup> fragment 13 of MDV (see Figure 1). Sequencing of this fragment obtained from a BamHI library of the RB1B strain of MDV showed that two thirds of the gene, starting with the NH2 terminus, was contained within 13. The remainder of the gene was identified in the adjacent

restriction fragment K3. Figure <sup>1</sup> shows the map position of the gene which is 2.6Kbp long. Its mRNA has been estimated to be approximately 2.8 Kb. The translated protein is 865 amino acids long (Figure 2). This includes approximately 20 amino acids which may be part of a signal sequence domain. The primary translated sequence of MDV gB has a few features in common with gB of other herpes viruses such as the alignment of cysteine residues and the presence of hydrophobic sequences which are presumably capable of spanning a lipid bilayer [Pellet, P.E. et al (1985), J. Virol. 53, 243-253]. However, MDV gB has only 48% amino acid similarity with gB of HSV and has many unique features such as the insertion of 23 amino acids (residues 1851-1920, Figure 2) and the presence of extra sites with glycosylation potential. Comparison of the sequence of MDV gB with limited sequence data (702 bases) available for HVT gB (Figure 2) has shown 76.9% nucleic acid similarity and 87.1% amino acid similarity between these two glycoproteins. Amino acid substitutions in HVT gB compared to MDV gB were particularly marked in a region (residues 1323 - 1433) equivalent to a domain of HSV gB associated with virus neutralization [Pellet P.E. et al (1985) as above]. Amino acid substitutions between MDV and HVT gB were also noted in other regions of unknown function.

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## EXAMPLE 2: gH gene of HVT and gH gene of MDV

An M13 clone of HVT containing sequences homologous to HSV gH was isolated during our earlier work on gene identification and mapping (Buckmaster et al (1988) as above). This clone, when used as a probe, hybridized to a 6Kbp Hindlll fragment of HVT (Figure 3). Sequencing revealed that this fragment contained approximately one quarter of the gH gene including the carboxy terminus. The adjacent HindIII fragment (3.2 Kbp) containing the remainder of the gH gene was identified by hybridization using a cloned Hqa<sup>l</sup> fragment of HVT which overlapped the HindIII site. Figure 4 shows the sequence of the coding region of the gH gene of HVT (2.3 Kbp) and flanking sequences. The % amino acid identity between the gH gene of HVT and its homologue in HSV1, VZV and EBV was only 20, 24 and 20 respectively (estimated from maximised amino acid overlaps of 630, 644 and 153 respectively).

### EXAMPLE 3: TK gene of HVT and TK gene of MDV

The whole coding region of the TK gene of HVT (1053 bp) was contained within the 3.2 Kbp Hindlll fragment described above (Figure 3). The sequence of the entire gene and flanking regions is shown in Figure 5. Similarly the whole of the MDV TK gene is contained within the 3.6 Kbp BamHl K2 fragment of MDV (Figure 1). The complete sequence of MDV TK gene is shown in Figure 14. Comparison of the MDV and HVT TK sequences shows

that the two genes have 60% amino acid identity. By contrast, the % amino acid identities between the TK gene of HVT and the TK genes of HSV 1, VZV and EBV are only 30, 27 and 24 respectively (estimated from amino acid overlaps of 320, 332 and 133 respectively). The predicted amino acid sequences of HVT and MDV TK show characteristic ATP and/or CTP binding site motifs described for a number of virus and eukaryotic proteins that are associated with phosphorylation (Gentry, G.A. (198S) Proc. Natl. Acad. Sci. U.S.A. 82, 6815-6819). These conserved sequences are examples of useful sites for insertion and expression of foreign genes and for producing TK- deletion mutants .

## EXAMPLE 4: A antigen gene of MDV (gP57-65) (gC homologue)

The A antigen gene is of interest in vaccine development both as an immunogen (it encodes a major glycopolypeptide product) and also because we have identified it as the homologue of HSV gC, a potential non-essential region. The A antigen gene was mapped within the BamH1 B fragment of MDV (Isfort et al 1987), and the nucleotide sequence determined for the GA strain of MDV (Coussens and Velicer, Abstract OP18.51, VII International Congress of Virology, '9-14 August, (1987) Edmonton, Canada; J. Virol. 62, 2373-2379). During the random sequencing studies described earlier (Buckmaster et al 1988), we identified an M13 clone (No. 130) which came from the A antigen gene. This clone was then used to identify a 2.3 Kbp EcoR1/PvuII fragment from

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the RB1B strain of MDV containing the A antigen. This fragment was cloned into a Smal/EcoRI cleaved pUC13 vector by standard protocols. One plasmid (pMB419) was sequenced by the M13 dideoxynucleotide method. The sequence of the MDV RB1B A antigen and the predicted amino acid sequence of the protein are presented in Figure 6. The A antigen regions of MDV and HVT are non-essential genes and they can therefore be used as sites in MDV and HVT into which other genes can be inserted into the virus by homologous recombination. Several lines of evidence support this as outlined below.

l)During our study we isolated and sequenced another RB1B A antigen clone. This had one extra T residue in the string of T's <sup>45</sup> bases 3' to the <sup>A</sup> antigen ATG codon. This extra <sup>T</sup> would cause a frameshift which would make it impossible for the gene to encode functional A antigen. As it is probable that this gene was cloned from a replicating MDV, the results suggest that the A antigen is non-essential to the virus.

2) On conducting a similarity search it became clear that the MDV A antigen gene is the homologue of HSV gC and PRV gpIII glycoproteins. Both of these homologous genes are known to be non-essential [for the HSV homologue, see Rosenthal et al (1987) J. Virol. 61

3) Strains of MDV lacking A antigen as judged by agar gel diffusion tests [Churchill, A.E. et al (1969) J. gen. Virol. 4,

537-564] or producing low levels using the more sensitive 2D radio-immunoprecipitation (van Zaane, D. et al (1982) Virology 121, 133-146] have been reported.

Furthermore, in view of the fact that the A antigen is a major secreted glycoprotein, it may be a particularly suitable location for the presentation of foreign epitopes within the A antigen as soluble, secreted proteins. This may be achieved by cloning oligonucleotides encoding these epitopes in frame within the A antigen gene.

## STRATEGIES FOR INTRODUCING GENES INTO HVT VECTORS

Two possibilities can be envisaged: 1) insertion into nonessential genes of the vector or 2) substitution of foreign gene for corresponding gene of the vector. This would be possible only in regions which already have substantial homology as may be the case between some genes of MDV and HVT.

## EXAMPLE 5: Insertion into non-essential genes of HVT or MDV

(a)Insertion at, the TK locus of the vector.

1) HVT or MDV may be used as vectors for insertion and expression of avian herpesvirus genes. In particular gB, gH or gC of RB1B MDV may be inserted into HVT. One may use the promoter associated with the inserted gene or use heterologous

promoters, including those of a different class of genes (for example the immediate early promoter to optimise expression of  $gB$ ).

2) HVT or MDV may be used as general vectors for the insertion and expression of genes unrelated to avian herpes viruses and likely to require manipulation of promoters for optimal expression .

The procedure to be used for gene insertion is substantially as described previously for the insertion of hepatitis antigen in HSV [Shih et al, 1984 as above].

MDV and HVT DNA obtained as described above is infectious provided that precautions are taken not to shear the DNA during extraction. Calcium phosphate precipitates of viral DNA prepared as described by Stow and Wilkie [(1976) J. gen. Virol. 33, 477 ] were added to sub-confluent monolayers of CEF. After absorption for lh at 37°C, culture medium was added and cultures were incubated for <sup>1</sup> or <sup>2</sup> days until confluent. Monolayers were then trypsinised, replated (1:1 or 1:2) in 199 medium (Wellcome) containing <sup>2</sup> to 4% calf serum and incubated at 37° C until plaques developed, usually after <sup>4</sup> to <sup>5</sup> days. Approximately 200 plaques may be obtained per  $\mu$ g of HVT DNA and approximately 50 per  $\mu$ g of MDV DNA.

For homologous recombination and isolation of recombinant virus, genes of interest are inserted within non-essential genes such as TK or gC and co-transfected with wild-type viral DNA at molar ratios ranging from 10:1 to 2:1 as described above. Alternatively, intact wild-type virus may be used for co-infection .

Restriction enzyme sites that could be used for the insertion of foreign antigens into the TK of HVT strain Fc-126 include: BanII, Bsp1286, DraIII, EcoRI, HincII, Hpal, Nhel and NspbII.

RE sites that could be used to produce defined TK deletion mutants in MDV serotype I strain RB1B include; Ball, Haell, Nde<sup>l</sup> and Sph<sup>I</sup> as insertion sites for foreign DNA that would disrupt the TK gene, and double digests of combinations of these four restriction enzymes (EcoK could also be used) to remove a portion of the TK gene, thus inactivating it.

Some of these enzymes also have sites in the plasmid vector into which the virus DNA fragments are cloned. Thus in order to linearize the clone DNA without also cutting within the vector, partial, digests may be carried out.

None of the above enzymes should cause any disruption to flanking genes, HSV-1 homologues of which are known to play an important role in virus multiplication.

Virus recombination may be detected by 'plaque lifts' which involve transfer of infected cells and released virus which have adhered to the agar overlay to nitrocellulose and hybridization of the denatured DNA released from the cells and virus to suitable probes as described by Villareal, L. et al (1977) Science 196, 183-185. Virus which hybridizes to the probe may be recovered from the monolayer.

A similar procedure may be used to isolate recombinant virus which expressed epitopes of interest. In this instance the nitrocellulose "plaque lifts" are treated with antibody and the presence of bound antibody revealed using a suitable detection system such as labelled protein A or phosphatase conjugated anti-globulin antibody.

The gene of interest with appropriate promoters is first inserted within the cloned TK gene. The recombinant DNA is then co-transfected with infectious -DNA of the vector in chick embryo fibroblasts or chicken kidney cells and TK<sup>-</sup> virus may be selected by growth in medium containing acyclovir [Ross, N. (1985) as above] or FMAU [Schat, K.A. et al (1984) Antiviral Research 4, 159-270]. Alternatively, or in addition, plaques are screened for the presence of the gene of interest using 'plaque lifts' on nitrocellulose and hybridization to any relevant labelled probe. Plaques are also screened for expression of the epitopes of interest using monoclonal antibodies or antipeptide antibodies.

The main advantage of this strategy is that the selection procedure increases the chances of obtaining virus recombinants containing the gene of interest. It also offers the opportunity of using different promoters for optimum expression. Thus the use of an immediate early promoter may allow expression in latently infected cells.

(b)Insertion at other non-essential sites of the vector.

Since the A antigen (HVT and MDV homologues of HSV gC) is not essential for virus growth in vivo and in vitro (see section on gC above) it is a potentially useful site for the insertion and expression of foreign genes. Moreover, since it is one of the most abundant antigens and is excreted, it may be particularly useful for enhancing the immunogenic properties of foreign proteins. The isolation of virus recombinants at this locus may be achieved by first inserting at least part of the gene of interest in frame within the gC gene and then co-transfecting with infectious viral DNA. Screening of virus plaques with sequence specific probes or with specific antibody allows the isolation of recombinants.

An antigen-encoding sequence can also be inserted into the ribonucleotide reductase (large subunit) gene of HVT or of MDV - see Figures <sup>8</sup> and <sup>9</sup> .

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# EXAMPLE 6: Substitution of MDV genes for their homologues in HVT

Substitution may be achieved by co-transfection of cloned MDV sequences and infectious HVT DNA as described in Example 5. Substitution of the gB and gC genes derived from the RBIB strain of MDV for their counterparts in HVT may be effected as may substitution of the gH gene of MDV, other glycoproteins and immediate early genes.

Recombinants expressing MDV sequences and epitopes may be detected using MDV-specific monoclonal antibodies or antipeptide antibodies raised against unique MDV sequences as described above .

The advantage of this procedure is that it is relatively simple and does not require manipulation of promoters. However, it may be limited to genes which share substantial homology.

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## EXAMPLE 7: Strategies for obtaining TK- mutants of MDV

Deletion mutants. Deletions may be introduced within any suitable part of the gene, for example the domains of the gene that are required for nucleoside binding. This may be achieved by restriction enzyme double digestion, for example with Haell and any of the following enzymes: Ball, Ndel, SphI or EcoK. Appropriate fragments are then religated, followed by cotransfection with infectious viral DNA or transfection into virally-infected cells. Reference may be made to Figures <sup>7</sup> and 8, and to the section above relating to insertion of heterologous sequences, in choosing restriction enzymes and so on. TK- virus may be selected in the presence of acyclovir [Ross, N. (1985) as above] or FMAU [Schat, K.A. et al (1984) as above]. Plaque-purified clones may then be tested for the absence of the deleted portion of the TK gene by hybridization.

The deletion mutants of MDV may be used themselves as attenuated viruses for vaccine preparation, or may have sequences for heterologous antigens inserted.

Insertional mutants. A functional ß-galactosidase gene under the control of a herpesvirus promoter or any other suitable sequence or a single base is first introduced in a domain of the TK gene which is essential for TK activity. The recombinant DNA is then co-transfected with infectious viral DNA or transfected into virally-infected cells to allow homologous

recombination to occur. Selection in the presence of acylovir or FMAU will yield TK' insertional mutants. If a B-galactosidase gene is introduced, mutants can be detected by the production of blue plaques in the presence of X-gal.

The TK gene and surrounding sequences may be subcloned into another suitable vector if necessary.

### EXAMPLE 8: Insertion of MDV RB1B gB gene into HVT

The HVT TK gene is cloned in the plasmid vector pUC13 to generate a plasmid, which is termed pTKIB. This plasmid is linearised with, for example, the restriction endonuclease Rsr II which cleaves the plasmid only within the TK gene (nucleotide position 197 in Figure 5, enzyme recognition sequence CGGACCG). The "sticky" ends thus generated can be end repaired by standard techniques (see "Molecular Cloning: a Laboratory Manual", ed. Maniatis T., Fritsch E.F., and Sambrook J. Cold Spring Harbor Laboratory 1982).

The RB1B gB was originally cloned on two plasmids which may be termed RBIB-BamH1-I<sub>3</sub> and RB1B-BamH1-K<sub>3</sub>. (Note I<sub>3</sub> had lost one BamHI site during cloning.) To generate a complete gB copy on one plasmid, both plasmids were cleaved with BamHI and the fragments ligated. Recombinants containing the desired configuration were identified by restriction enzyme analysis of plasmid DNA's. However, as described above, the complete gB

sequence was subsequently obtained on an EcoRI/SalI fragment.

Further information regarding the sequence encoding MDV gB and its manipulation may be found in Ross et al J. gen. Virol ( 1989 ) 70. 1789-1804 .

The single recombinant plasmid of Ross et al is then cleaved with EcoRI and SalI, the ends are repaired and the plasmid is cloned into PTKIB prepared as above. Alternatively, the MDV gB open reading frame could be excised from plasmid MSB27 by digestion with HincII and Nael and the products ligated to HVT TK plasmid pTK1B, cleaved partially with HpaI. Recombinant plasmids containing both TK and gB sequences could be identified by hybridisation and further characterised by Southern blotting. The recombinant plasmids are then introduced into cells containing HVT virus (viral DNA) and homologous recombination will introduce the gB gene into the TK gene. HVT viral recombinants can be selected with acyclovir or FMAU or alternatively detected with labelled gB probes.

## EXAMPLE 9: RB1B gC (A antigen) gene into HVT

Blunt ended PTKIB is prepared as in Example 8. The RB1B gC is cleaved from the plasmid pMB419 (Example 4) with the restriction endonucleases EcoR<sup>I</sup> and Hindlll (site within the pUC13 polylinker). The sticky ends generated are again endrepaired by standard protocols. The end-repaired gC fragment is
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then cloned into the linearized end-repaired pTKIB as in Example 8. (The cloning can be verified by analysis of the resulting clones with restriction enzymes, probing with radioactively labelled fragments, or DNA sequencing , or any combination of these).

The resulting plasmid with the RB1B gC gene cloned into the HVT TK gene can then be introduced into the HVT genome by transfecting the plasmid into HVT-infected cells using calcium phosphate precipitation or electro-poration. Homologous recombination, ' involving cross-overs either side of the gC gene, between the HVT virus and the flanking sequences of the HVT TK plasmid will carry the RB1B gC gene into the HVT viral genome. Viral recombinants can be selected for (as they are TK-) or identified (eg by probing) as described above.

In analogous ways, the sequence information given above and in the Figures can be used to design cloning strategies for the insertion of these genes and others into the non-essential genes of the HVT described here or to generating combinations of antigen genes in HVT.

#### EXAMPLE 10: MDV gD gene

Figure 15 shows part of the sequence of the MDV gD gene. The sequence was obtained by sequencing random fragments of the U<sup>s</sup>

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region MDV DNA and comparing the sequence to the sequence of known herpesvirus genes (see Buckmaster et al, loc. cit.). The sequence gave homology scores of 189 and 216 respectively with HSV gD and PRV gp50. The sequence information assists in the preparation of suitable probes to isolate and characterise the gene .

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A nucleotide sequence substantially free of the sequences which would adjoin it in the wild-type virus associated with that sequence, the sequence being selected from the group consisting of :

(a) the MDV homologue of the HSV gB gene, or portions 816-863, 1377-1595, 1377-1630 or 1824-1985 of the said homologue,

(b) the MDV homologue of the HSV gH gene,

(c) the TK gene of MDV,

(d) the MDV homologue of the immediate early gene IE-175 of HSV-I,

 $(e)$  the MDV homologue of the immediate early gene IE-68 of HSV-I,

(f) the MDV homologue of the HSV gD gene, and portions 483—633, 843-933 or 1203—1278 of the MDV homologue of HSV gC

and minor variations thereof, as hereinbefore defined.

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2. A sequence according to Claim <sup>1</sup> comprising the coding portion of the said sequence and at least part of the 5' and/or 3' non-coding portions thereof. '

3. A plasmid vector comprising a sequence according to Claim <sup>1</sup> or <sup>2</sup> (except sequence c) which is suitable for transfection of an MDV- or HVT-susceptible cell.

A hybrid viral vector comprising, as a heterologous  $\Lambda$ . insert, sequence (a), (b), (d), (e) or (f) of Claim 1 and suitable for transfection of an MDV or HVT-susceptible cell.

5. A viral vector according to Claim 4 wherein the sequence is inserted into a non-essential site of HVT.

6. A viral vector according to Claim 5. wherein the nonessential site is in the.region homologous to the HSV gC gene or in the ribonucleotide reductase (large subunit) gene or the **TK gene.**

7. A peptide encoded by any one of sequences (a), (b) or (f) of Claim <sup>1</sup> or any of the said portions of sequence (a) or portions 483-633, 843-933 or 1203-1278 of the MDV homologue of HSV gC.

**<sup>q</sup> An MDV virus, mutated by virtue of being TK-.**

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**9. <sup>A</sup> vaccine comprising MDV-susceptible ceils and <sup>a</sup> viral vector according to any one of Claims <sup>4</sup> to <sup>g</sup> or a mutant MDV virus according to Claim <sup>8</sup> such that the virus is attenuated, at least partially as <sup>a</sup> result of such mutation.**

**<sup>J</sup> 1G. <sup>A</sup> method of vaccinating <sup>a</sup> fowl against a disease of that fowl comprising administering to the fowl a non-toxic immunityconferring amount of vaccine according to Claim 9.** disease of<br>-toxic imm<br>m 9.<br>ing to Clai

**II· <sup>A</sup> fowl when vaccinated by a method according to Claim 10.**

Dated this 7th day of May 199'2

RHONE-MERIEUX S.A.

By their Patent Attorneys GRIFFITH HACK f, CO



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FIG 2B ATATATATAACATATGAAACCGAATATCCACTTATAATGA 370 380 390 400 TTCTGGGGTCAGAATCAAGCACTTCAGAAACGCAAAATAT 410 420 430 440 GACTGCAATTATTGATACAGATGTTTTTTCGTTGCTTTAT 450 460 470 480 TCTATTTTGCAGTATATGGCCCCCGTTACGGCAGATCAGG 490 500 510 520 TGCGAGIAGAACAGATIACCAACAGCCACGCCCCCATCTG 530 540 · 550 560 ACCCGTCCAATATTCTTGTGTCCCTGCATTTTATCTCACA<br>570 580 590 600 570 580 590 600 $M$   $H$ 

CAATTTATGAACAGCATCATTAAGATCATCTCACTATGCA 610 620 630 640

YFRRNCI FFLIVI CTATTTTAGGCGGAATIGCATTTTTTTCCTTATAGTTATT<br>650 660 670 680 650 660 670 680

#### FIG 2C

LYGTNSSPSTQNVT CTATATGGTACGAACTCATCTCCGAGTACCCAAAATGTGA 690 700 710 720

SREVVSSVQLSEE CATCAAGAGAAGTTGTTTCGAGCGTCCAGTTGTCTGAGGA 730 740 750 760

ESTFYLCPPPVGS AGAGTCTACGTTTTATCTTTGTCCCCCACCAGTGGGTTCA 770 780 790 800

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TVIRLEPPRKCPEP ACCGTGATCCGTCTAGAACCGCCGCGAAAATGTCCCGAAC 810 820 830 840

RKATEWGEGIAIL CTAGAAAAGCCACCGAGTGGGGTGAAGGAATCGCGATATTA 850 860 870 880

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 $FIG 2H$ 

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#### FIG 21

RELSATFISNTTE CCGTGAACTTTCGGCAACGTTTATCAGTAATACGACTGAG 1610 1620 1630 1640

FDPNRI ILGQCIKR ITTGATCCAAATCGCATCATATTAGGAC^ATGTATTAAAC 1650 1660 1670 1680

EAEAAIEQIFRTK GCGAGGCAGAAGCAGCAATCGAGCAGATATTTAGGACAAA 1690 1700 1710 1720

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Y N D S H V K V G H V O Y ATATAATGACAGTCACGTCAAGGTTGGACATGTACAATA 1730 1740 1750 1760

FLA LGGFIVAYQPV TTTCTTGGCTCTCGGGGGATTTATTGTAGCATATCAGCCTG 1770 1780 1790 1800

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LSKSLAHMYLREL TTCTATCCAAATCCCTGGCTCATATGTACCTCAGAGAATT 1810 1820 1830 1840

MRDNRTDEMLDLV GATGAGAGACAACAGGACCGATGAGATGCTCGACCTGGTA 1850 1860 1870 1880

NNKHAI YKKNATSL AACAATAAGCATGCAATTTATAAGAAAAATGCTACCTCAT 1890 1900 1910 1920

SRLRRDIRNAPNR TGTCACGATTGCGGCGAGATATTCGAAATGCACCAAATAG 1930 1940 1950 1960 KITLDDTTAIKST AAAAATAACATTAGACGACACCA.CAGCTATTAAATCGACA 1970 1980 1990 2000

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SSVQFAMLQFLYDH TCGTCTGTTCAATTCGCCAIGCTCCAATTTCTTTATGATC 2010 2020 2030 2040

IQTHINDMFSRIA ATATACAAACCCATATTAATGATATGTTTAGTAGGATTCC<br>2050 2070 2080 2050 2060 2070 2080 **00**o

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TAWCELONRELVL CACAGCTTGGTGCGAATTGCAGAATAGAGAACTTGTTTTA 2090 2100 2120 2110  $\sim 10^{11}$  and  $\sim 10^{11}$ 

WHEGIKINPSATAS TGGCACGAAGGGATAAAGATTAATCCTAGCGCTACAGCGA 2130 2140 2150 2160



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#### FIG 2N

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LNLTLLEDREILP AGCTTAATCTAACCCTGCTAGAAGATCGGGAAATTTTGCC 2530 2540 ' 2550 2560

LSVYTKEELRDVG TITATCCGITTACACAAAAGAAGAGTTGCGTGATGTTGGT 2570 2580 2590 2600

VLDYAEVA RRNQLH GTATTGGATTATGCAGAAGTAGCTCGCCGCAATCAACTAC 2610 2620 2630 2640

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ELKFYDINKVIEV ATGAACTTAAATTTTATGACATAAACAAAGTAATAGAAGT 2650 2660 2670 2680

DTNYAFMNGLAEL GGATACAAATTACGCGTTTATGAACGGTTTGGCCGAATTG 2690 2700 2710 2720

FNG MGQVGQAIGKV TTTAACGGTATGGGTCAGGTAGGGCAAGCTATAGGCAAAG 2730 2740 2750 2760

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#### $FIG 2\phi$

VVGAAGAIVSTI S TTGTAGTAGGGGCTGCCGGTGCAATCGTATCTACCATATC 2770 2780 2790 2800

GVSAFMSIPLGLS TGGIGTCTCIGCTTTCATGTCAATCCCTTTGGGGCTTTCG 2810 2820 2830 2840

A I G L I I I A G L V A A F GCAATCGGTTTAATCATTATAGCAGGACTCGTGGCTGCAT 2850 2860 2870 2880

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<sup>L</sup> AY.RYVNKLKSNP TTTIAGCATATCGTTATGTAAACAAGCTTAAAAGCAATCC 2890 2900 2910 2920

MKALYPMTTEVLK AATGAAAGCCCTTTATCCTATGACAACAGAAGTGCTTAAG 2930 2940 2950 2960

AQATRELHGEESDD GCACAGGCAACGCGTGAGTTGCATGGCGAGGAATCAGATG 2970 2980 2990 3000

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### FIG 2P

LERTSIDERKLEE ATTTGGAACGAACATCTATTGATG/XAAGAAAATTAGAAGA 3010 ' 3020 3030 3040

AREMIKYMALVSA AGCTAGAGAAATGATAAAATATATGGCGTIAGTCTCCGCG 3050 3060 3070 3080

EERHEKKLRRKRRG GAAGAACGCCACGAGAAAAAACTGCGGAGAAAGAGGCGAG 3090 3100 3110 3120 TTAVLSDHLAKMR GCACTACCGCCGTTCTATCGGACCACCTGGCAAAAATGAG 3130 3140 3150 3160

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<sup>I</sup> KNSNPKYDKLPT GATTAAAAATAGTAACCCTAAATATGATAAGTTACCTACT 3170 3180 3190 3200

TYSDSEDDAV\* ACATATTCAGACTCAGAAGATGATGCTGTGTAAGTGGGCA 3210 3220 3230 3240

CTATTATATTTGAACTGAATAAAACGCATAGAGCATGATA 3250 3260 3270 3280

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FIG 2Q

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TGGAAGGCATAGGGCGTTCGACTCCCATGGGCCATGAAACTGTGGGATGT<br>3610 3620 3630 3640 3650 3630

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UL IRL IRS U TRL Bam HI B <sub>s</sub> IRs  $\Box$  $-++$ /  $\Delta H_{\rm H}$  $\overline{\phantom{a}}$  $\overline{\phantom{0}}$  $\overline{\phantom{a}}$  $\overline{\mathscr{C}}$  $\overline{\phantom{0}}$  $\overline{\phantom{a}}$  $\overline{\phantom{0}}$  $\overline{\phantom{0}}$ Z  $H<sub>1</sub>$ H  $\mathsf H$ —<br>———— ΕΞΙ-----------------  $\sigma$   $\rightarrow$   $\rightarrow$ **MCP** gH TK 6 Kbp  $*-32$ Kbp  $*$ -----------

Fig. 3

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H <sup>=</sup> Hind III sites ■> = Thymidine Kinase (TK) EZ3 = Glyco protein H(gH)

E3 = Major Capsid Protein (MCP)

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### $FIGA$

#### TATTATTGGTCCATGCTAGAATAGTCATACGCTACGATCT  $20$ 30  $10$  $40$

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GTTGCTATATATGACTATCGCCAAACTGTTAAACCCGCGA  $50$ 60 70  $80$ 

AGAATATATTTCATATAAACCTAAGGGCCCCTCAGTCTGA 90 100  $110$ 120

M  $\mathbf{K}$ Y T. F  $\mathcal{C}$ TTTTTTGTGAAAACGTGTATACCATGAAGTTTTACTGCCT  $1.30$ 140 150 160

 $R$  $F$ M  $\overline{A}$ N<sub>1</sub>  $T$  $\mathsf{T}$  $\mathsf{T}$  $\mathbf{T}$ . Y  $\overline{S}$  $S$ Y AATCCGTTTCATGATCATAGCGAATCTTTATTCATCTTAC 170 180 190  $200$ 

 $\overline{O}$  $\mathsf{T}$ <sub>S</sub>  $\mathbf{T}$ .  $\mathbf{P}$ G  $T$ Y  $\mathbf{P}$ S  $\overline{O}$  $T$ .  $\mathsf{T}$  $\mathsf{T}$ . CAAATATCGCTTCCAGGCACATATCCATCGCAAATATTGC 210 220  $230$  $2.40$ 

 $\mathbf{D}$ M  $\mathbf{K}$  $N$  $\mathcal{S}$  $\mathbf{T}$ P  $\mathbf{V}$  $\mathbf{R}$  $\mathbf F$  $\mathbf{N}$ S. T TTGACATGAAGAACTCGCCGCTCGTACGCTTTAATATATC  $250$ 260  $2.70$ 280

TRDYKDETLWIRK. GACGCGTGATTATAAAGACGAGACACTCTGGATACGGAAA 290 300 310 320 '

N S T F V Y I D T A V T T A<br>AFFICIACAFFECTERATIONALSCORETCACCACAC AATTCGACATTTGTTTATATCGATACGGCTGTGACGACAG 330 340 350 360 <sup>490</sup> <sup>500</sup> <sup>510</sup> <sup>520</sup> . **<sup>W</sup> <sup>O</sup> <sup>9</sup> <sup>0</sup> / <sup>0</sup> <sup>2</sup> <sup>8</sup> <sup>0</sup> <sup>3</sup> <sup>P</sup> <sup>C</sup> <sup>T</sup> / <sup>G</sup> <sup>B</sup> <sup>8</sup> <sup>9</sup> / <sup>0</sup> <sup>1</sup> <sup>0</sup> <sup>7</sup> 6**

N V I F Y L P I G Q V R Q<br>ACCEEARCEERARCE ACCESS ECCERCACCEACACA CGAACGTTATCTTTTATCTGCCGATCGGTCAGGTACGACA 370 380 390 400

M V F F K R P I S R L L T<br>M V F F K R P I S R L L T AATGGTTTTTTTCAAGCGTCCAATATCCAGGCTACTAACG 410 420 430 440

SNNLVKFINTGSYA<br>GGAATAAGGTGGTTAATTPATTAUL TCCAATAACCTGGTTAAATTTATT7ATACCGGTTCATACG 450 460 470 48Q

NHTFKTELSPYLS CCAATCATACATTCAAGACAGAACTTTCACCCTATTTGTC

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KTNTPLKKYEIVV. GAAAACCAATACACCGTTGAAGAAATATGAAATTGTTGTC 530 540 550 560

DQ PTGENPPAGFGS GATCAACCTACTGGAGAAAACCCTCCGGCAGGGTTCGGAA 570 580 590 600

L K P A D F L N P G Y K F GTTTAAAACCGGCAGACTTTCTCAACCCCGGATACAAGTT 610 620 630 640

VLTSELVGAYTKR CGTTCTCACAAGCGAGTTGGTAGGAGCCTACACAAAACGA 650 660 670 680 **<sup>e</sup>**

S C F V D P M D S L V P I D TCTTGTTTTGTCGATCCGATGGATTCTCTCGTCCCGATAG 690 700 710 720

YDHVRTIIFGSAG<br>ATTATGATCATGTACGAACCATTATATTCGGATCTGCTGC 730 740 750 760

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T T, M K M G T T L A S. M  $F_{\cdot}$ GATGGAGATTTTAATGAAGATGGGAATTACTTTGGCATCT 770 780 790 800

 $S$ T K Y N P P I M  $\mathbf{T}$  $\mathbf{T}$  $F$   $T$   $T$ ATGACCATTTCGACGAAATATAATCCTCCTATTGAACTGA 810  $820$ 830 840

SAKYRNLSLL  $T$  $W$  P TAATATCTGCAAAGTACCGAAATTTATCACTGTTGTGGCC 850 860 870 880

O O Y E P V N K G  $P$  R  $\mathbf{T}$ G ACCCCGACAACAATATGAACCTGTAAATAAAGGGACTGGA 890 900 910 920

R P H W I Y L L G V Y R N V CGCCCCCATTGGATCTACCTATTAGGTGTGTATAGAAACG 930 940 950 960

DSERDSYMNM  $S_{\cdot}$  $T$   $K$ TTTCGGACTCCGAGCGTGACTCATACATGAATATGATTAA  $9.70$ 980 990  $1000$  PCT/GB89/01076

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SLGDSMDY HFLIS. GAGTCTGGGCGATTCTATGGATTATCACTTCCTAATTAGC 1010 1020 1030 1040

RAHAQMLILAAEDR AGAGCGCATGCCCAGATGCTGATACTGGCAGCAGAGGACC 1050 1060 1070 1080

LVDEMHSFRNVIA GGCTCGTGGATGAAATGCATAGTTTCAGGAACGTTATTGC 1090 1100 1110 1120 1090 1100 1110 1120<br>RLFVSLFAFI RNA

GCGTTTATTTGTATCGTTGTTCGCATTCATACGTAACGCA 1130 1140 1150 1160 1130 1140 1150 1160<br>FOSGYTSLNDIIEI

TTTCAGTCTGGCTACACCTCTCTTAATGACATAATTGAAA  $1170$   $1180$   $1190$   $1200$ 1170 1180 1190 1200<br>EADLRLIVEGISS

TCGAAGCCGATTTGAGGTTAATTGTAGAAGGCATTTCTTC 1210 1220 1230 1240

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#### FIG 4F

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AAFRKDASTHFLI TGCTGCATTTCGTAAAGACGCTAGTACACACTTTCTTATA<br>1250 1270 1280  $1250$   $1260$   $1270$   $1280$ 1250 1260 1270 1280<br>SGTPIKDSKADLIK

T C G GGAAC G C C CATAAAAGATAG CAAAG C GGAT TTAATTA 1290. 1300 1310 1320

SLLSKVIRPISGH<br>SLLSKVIRPISGH AATCGTTGTTGTCTAAAGTCATTCGACCAATTTCCGGACA 1330 1340 1350 1360 1330 1340 1350 1360<br>TRPLSAIQHLFLL

TACACGTCCCTTATCTGCGATACAACATCTATTCCTTTTG 1370 1380 1390 1400

RSAYALDIPRQNGS AGATCCGCTTATGCATTGGATATACCCCGTCAAAACGGAT 1410 1420 1430 1440 1410 1420 1430 1440<br>LSEQVSTVAL SFI

CTTTGAGCGAACAGGTATCTACAGTGGCACTGTCGTTCAT 1450 1460 1470 1480

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#### FIG 4G

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**e**

ENIHSEAMRDILS. TGAAAATATTCACAGCGAGGCCATGAGGGACATTCTGTCA 1490 1500 1510 1520

WNTTTKHALY.YAFA TGGAACACTACAACAAAGCATGCGTTGTATTATGCATTCG 1530 1540 1550 1560

SILORPLTEWGAS CGAGTATTTTGCAACGGCCACTGACCGAATGGGGCGCCTC 1570 1580 1590 1600

<sup>R</sup> NARRAILLASSM AAGAAATGCACGGAGGGCAATACTATTAGCATCATCGATG 1610 1620 1630 1640 1610 1620 1630 1640<br>C T E E H V I A T E L A I O

TGTACAGAAGAGCATGTTATCGCAACTGAGTTGGCTATTC 1650 1660 1670 1680

ELYVKIRSNADPI AAGAACTGTATGTCAAAATCAGAAGTAATGCCGACCCAAT<br>1720 1720 1730 1700 1710 1720

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#### FIG 4H

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H L L D V Y  $\mathbf{P}$  $C = L$  $S-T$  $\mathbf{T}$  $\mathcal{S}$ ACACCTTCTAGACGTATATACACCATGTCTTTCTTCACTA 1730 1750 1760 1740

R L D L S  $E$  H H R I Y A M A CGATTGGACCTTTCCGAACACCATCGGATATACGCAATGG 1770 1780 1790 1800

 $D$   $V$   $V$   $F$ Y P D I  $\overline{O}$  $\overline{O}$ Y L K CAGATGTAGTTTTCTATCCAGACATTCAGCAGTATTTGAA 1820 1830 1810 1840

K K S H E G N M K E D D L AAAAAAATCCCATGAGGGTAATATGAAGGAAGATGATCTC 1850  $1860$  $1870$ 1880

E T K A E Y I  $\mathbf{L}$  $T$   $K$   $I$ GAAACAAAGGCGGAATACATCCTCACCAAGCTT 1890 1900  $-1910$ 

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## FIG 5A

AAGCTTTTTGTAAAAACGATTATGACCACGGACACCCGCT 10 20  $30<sup>°</sup>$  $40$ 

TTTAGCAATCCTGCCATAAGGTGGTTTCCCGCGTGCTTGC 60 50  $70$ 80

CTCGAAGACAATTGCCAGCTAATCCAGCATTACCATATTT 90 100 110 120

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FIG 58

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GAGAGTTTTCTTTATTTCAATCTAGCATGATTGTAACAGC 370 380 400 390  $-L$   $-S-K$ I Q A  $\mathbf{A}$  $\mathbf{P}$  $H$  $R$  $\mathbf{F}$ D  $\mathbf Y$ L. F TATACAAGCCAGATTTGCCGATCCATATTTGCTTTTTCAC TTTACAATCAAAGTTTGCAGATCCCTATCTT IGTATTTCAT 410 420 430 440

GAGCTTTATCACGATTCCAATCTGACATGATCATGGCATC

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 $FIG5F$ 



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GELLTQYSWIL WG ATGGGGAGCTGTTAACTCAGTATTCTTGGATATTGTGGGG 970 980 990 1000

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LLTKLHTINVELF ATTACTGACTAAACTACACACCATTAATGTCGAATTATTT 1010 1020 1030 1040



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# FIG 5J

# TATTATTGGTCCATGCTAGAATAGTCATACGCTACGATCT<br>1250 1270 1280 1260 1270

GTTGCTATATATGACTATCGCCAAACTGTTAAACCCGCGA<br>1290 1300 1310 1320 1290 1300 1310 1320

AGAATAIATTTCATATAAACCTAAGGGCCCCTCAGTCTGA, 1340

TTTTITGTGAAAACGTG'IATAC *C K 1370 1380*

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1 CAGCTGCCTATGTAGTGAAATCTATACTGGGATTT ATCATAACTAGTTTACTTGTTTGTATATTAGTAGCGCTATCT TGACCAAATCGTTGTTCACATCTTGGCCATATACGTATTGATC 121 GTTGTTTCGAACCGCGAATAAAACTTTCATACATAC TAAACGATGGAGTTGTGTTTTATGAGCGTTGAAAACAAAGGT ACCATCGGTTTAAAACTAAGTTGCATATCGTAATCCACAAAA 241 ATCATTTTATACATCATCCCGAAGAGACACCAAACG M<sub>L</sub>  $\mathbf{R}$  $\mathbf v$  $\mathbf{T}$  $\mathbf{p}$ TAACCCTCTACATATCTTCCCTCATGCTCACGCCGCGTGTGT L R A L G W  $\mathsf{T}_{\mathsf{L}}$ -S  $\mathbf{T}$  $\mathcal{G}$  $-F$  $\mathbf{F}$  $\mathbf{L} \cdot \mathbf{L}$  $\mathbf{L}$ TACGAGCTTIGGGGIGGACTGGACTCTTTTTTTTGCTTTTTAT  $\mathbf{P}$ S N V L G A S L  $\overline{R}$ S. 361 CTCCGAGCAACGTCCTAGGAGCCAGCCTTAGCCGG  $\mathsf{D}$  $\mathsf{T}$ .  $\mathbf{F}$ P  $\mathbf{p}$ E S S 

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NISING APLTEVPHAP ACATTTCAATTAACGGCGCGCCTTTAACTGAGGTACCTCATGCAC

STESVSTNSEST 481 CTTCCACAGAAAGTGTGTCAACAAATTCGGAAAGTACC

NEHTITETTGKNAY AATGAACATACCATAACAGAAACGACGGCCAAGAACGCATACA

I H N N A S T D K Q N A N D TCCACAACAATGCGTCTACGGACAAGCAAAATGCGAACG

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THK TPNIL CDTE 601 ACACTCATAAAACGCCCAATATACTCTGCGATACGGA

EVFVFLNETGRFVC 

T L K V D P P S D S E W S N ACTCTCAAAGTCGACCCCCCCCTCGGATAGTGAATGGTCCA

F V L D L I F N P I E Y 721 ACTTTGTTCTAGATCTGATCTTTAACCCAATTGAATA

HANEKNVEAARIAG CCACGCCAACGAAAAGAATGTGGAAGCGGCGCGTATCGCTGGT 40/80

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YKASCIVRHFYPPG TTACAAGGCATCTTGTATCGTTAGACACTTTTATCCCCCTGGA

DVLAPPVLSGEN 1081 TGGATGTACTGGCCCCTCCAGTCCTCAGCGGAGAAAA

TAGATAAACATATTTACATACGTGTGTGTCAACGACCTGCATCAG

MQVI MRDH FNRPL ATGCAAGTCATTATGCGTGACCATTTTAATCGGCCTTTAA IDKHIYIRVCQRPASV

IRNVDYADNGY 961 AAATTAGAAATGTAGATTATGCTGATAATGGCTAC

YFIWINK TTNTMGVE GTACTTCATATGGATAAACAAAACAACCAATACGATGGGCGTGG

QGTFWTSPSPHGNK AGGGCACATTTTGGACGAGCCCATCACCTCATGGAAACAA

SELISSIRRDP

841 AATCTGAATTAATTTCTTCGATTCGACGAGATCCCC

CTCTATGGAGTCCCCGGATCAGACTATGCATACCCACGTC

<sup>L</sup> YGVPGSDYAYPRQ

FIG 6C

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### FIG 6D

SVYVSWRQNGNIAT TCTGTCTATGTATCTTGGAGACAGAATGGAAACATTGCAA

PRKDRDGSFWWF 1201 CTCCTCGGAAAGATCGCGATGGAAGTTTTTGGTGGTT

ESGRGATLVSTITL CGAATCTGGTAGAGGAGCTACGTTGGTTTCTACAATAACATTG

GNSGID FP PKISCL GGAAATTCAGGAATTGATTTCCCCCCCAAAATATCTTGTC

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**SHEET**

<sup>V</sup> AWKQGDMI <sup>S</sup> <sup>T</sup> T 1321 TGGTTGCCTGGAAGCAGGGTGATATGATCAGCACGAC

ΝΑΤΑ <sup>I</sup> PTVYHHPRL GAATGCCACAGCTATCCCGACGGTATATCATCATCCCCGTTTA

SLAFKDGYAICTIE TCCCTCGCTTTTAAAGATGGGTATGCAATATGTACTATAG

CVPSEITVRWLV 1441 AATGTGTCCCCTCTGAGATTACTGTACGGTGGTTAGT

HDEAQPNTTYNTVV ACATGATGAAGCGCAGCCTAACACAACTTATAATACTGTGGTT  $42/8$ 

### FIG 6E

TGLCRTIDRHRNLL ACAGGTCTCTGCCGGACCATCGATCGCCATAGAAATCTCC

S R I P V W D N W T K T 1561 TCAGCCGCATTCCAGTATGGGACAATTGGACGAAAAC

K Y T C R L I G Y P F D E D AAAATATACGTGCAGACTCATAGGCTACCCCTTCGATGAAGAT

K F O D S E Y Y D A T P S A AAATTTCAAGATTCGGAATATTACGATGCAACTCCATCTG

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RGTPMVITVTAV 1681 CAAGAGGAACACCCATGGTTATTACGGTTACGGCAGT

L G L A V I L G M G I I M T TTTGGGATTGGCTGTAATTTTAGGGATGGGATAATCATGACT

ALCLYNSTRKNIRL GCCCTATGTTTATACAACTCCACACGAAAAAATATTCGAT

 $\star$ 1801 TATAATCTCATTGTTATGTAGTTGTGATTTATTAAAC

ATATTTTTTTATAACTCTAGTATTCTCCGAGTACTTATATATT

### FIG 6F

### TATTTGTCAGACAATAATGCAATAGIGGAGAAACGTGAGG

1921 GGAGTCTGTAAACAGAATACGTATAATCATCTATTTG AATAAAAGATTGIGGTATAAATGAAGATAGGGCAAGTCATTC

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PCT/GB89/01076 **P C T / G B 8 9 / 0 1 0 7 6**

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CAAGCTCTCCATTCTAITTAAACAATGTACAGTTIAAAGT

#### FIG 7

HVT HOMOLOGUES OF VZV62/ HSV-1 IE 175

SNVVRYMCGNTVL TCGAATGTGGTGCGATACATGTGCGGGAACACGGTACTCC 10 20 30 40

PPDPHGTPVVINVP CCCCGGATCCCCATGGCACCCCCGTGGTGATCAACGTTCC<br>50 60 70 80 50 60 . 70 80

EGT SETMAELTVA CGAGGGAACGTCCGAAACTATGGCGGAGCTTACTGTTGCT 90 100 110 120

HVG RKSIGWPT SE CACGTTGGGCGCAAGTCTATTGGGTGGCCGACCTCCGAGT 130 140 150 160

130 140 150 160<br>WHSATILQKD NDSRLVIIR GGCATTCCGCTACAATCCTGCAGAAAGATAATGATAGTCGGCTGGTAATTATACGCC 170 180 190 200 210

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## FIG 8

HVT HOMOLOGUE OF RIBONUCLEOTIDE REDUCTASE (LARGE SUBUNIT)

Q V T E V S E G F A P L F CAAGTGACCGAGGTTAGCGAAGGATTTGCCCCTTTGTTCA 10 20 30 40

SNMFSKVTSAGELL<br>CTAACATOTTGAGGAAGTGAGAAGTGGGGGGAAGTGGT GTAACATGTTCAGCAAGGTGACAAGTGCCGGGGAACTGCT 50 60 70 80

RPNSQLM RELRQI<br>GAGGGAAGAGTGAATTAATGGGGGAGGAGAGAAAT TAGACCCAACAGTCAATTAATGCGGGAGCTGAGACAAATA 90 100 110 120

Y P D N TATCCCGATAAT 130

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1 FIG 9A **W O 9 0 / 0 2 8 0 3** MDV HOMOLOGUE OF RIBONUCLEOTIDE REDUCTASE ( LARGE SUB-UNIT ) G I M E G S D V P T E K S<br>GGATAATGGAAGGAAGTGATGTAGGAGGAAAAATG GGGGATAATGGAAGGAAGTGATGTACCGACGGAAAAATCT  $10$  20 30 40 HSGRERNRSM GIG CATTCTGGCCGAGAACGTAACAGATCGATGGGCATCGGCG 50 60 70 80 VQG-FHTAFLSMGLD TGCAGGGCTTTCATACAGCTTTTCTATCTATGGGTCTTGA 90 100 110 120 90 100 110 120<br>LCDERARSLNKLI TTTATGCGATGAACGCGCTAGATCCCTCAACAAGCTAATT 130 140 150 160 FEFMLLEAMTVSC TTTGAATTCATGTTATTGGAGGCGATGACAGTTAGTTGCG 170 180 190 200  $\sqrt{6}$ PCT/GB89/01076 **P C T / G B 8 9 / 0 1 0 7 6**EFCERGLPPFADFS<br>AATTECTECAACCACCACCACTECACCACTER

AATTCTGCGAACGAGGCCTGCCGCCGTTTGCTGATTTCTC 210 220 230 240

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# FIG 9B

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NSYYARGRLHFDG TAACAGTTATTATGCACGAGGACGTCTGCATTTCGATGGG 250 260 270 280

WANVEL AAVE EWN TGGGCTAATGTAGAATTGGCTGCAGTGGAAGAGTGGAATA 290 300 310 320

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MDV HOMOLOGUE OF RIBONUCLEOTIDE REDUCTASE ( SMALL SUB-UNIT )

L D V E A I L C Y V R Y S<br>ECAECAECAACCAAEAEAEAECEACCEACCEEACA TATTGGATGTTGAAGCAATATTATGTTACGTACGTTACAG 10 20 30 40

FIG 10

RGQTTERIDMPPI CCGCGGACAGACTACTGAAAGAATAGATATGCCACCTATT 50 60 70 80

YNEPKPTADFPHAL TACAACGAACCTAAACCTACAGCTGATTTTCCGCATGCAC 90 100 110 120

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**SURGITETER** 

TASNNTNFFERRN<br>GACCERCAAARARACCAACRECERCACAACAAA TGACAGCTTCAAATAATACCAACTTCTTTGAGAGAAGAAA 130 140 150 160'

TAYSGSVSNDL\* TACTGCATACTCTGGAAGCGTGTCAAACGATCTTTAA 170 180 190

## FIG 11

### MDV HOMOLOGUE OF HSV-1 IE-175

PIPVYV EEMKDYA CCCATTCCCGTCTATGTAGAGGAAATGAAAGATTATGCCA 10 20 30 40

KOYDALVNSL F. HKS AACAATACGACGCTCTCGTAAACTCTTTGTTTCACAAAAG 50 60 70 80

MK VNPLNWMHHGK CATGAAAGTAAATCCTCTGAACTGGATGCACCACGGGAAG 90 100 110 120

LSTADAALNHIYV CTGTCTACCGCCGATGCTGCCCTAAACCACATATATGTTC<br>130 140 150 160 130 140 150 160

QKFQSSYDSPGAAV AGAAATTCCAGAGTTCATACGATTCGCCCGGAGCGCTGT<br>180 190 200 170 180 190 200

T G T V N AACTGGCACAGTTAACA 210

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## FIG 12

MDV HOMOLOGUE OF HSV-1 IE-68

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SD ODFELNNVG KF CGTCCGATCAAGACTTTGAACTTAATAATGTGGGCAAATT 10 20 30 40

CPLPWKPDVARLC TTGTCCTCTACCATGGAAACCCGATGTCGCTCGGTTATGT 50 60 70 80

ADTNKLFRCFIRCR<br>GCCAPACAAACAAACHAPPECCAPCEPRENAPELCCAPCEC GCGGATACAAACAAACTATTTCGATGTTTTATTCGATGTC 90 100 110 120

LN SGPFHDALRRA GACTAAATAGCGGTCCGTTCCACGATGCTCTTCGGAGAGC<br>160 160 160 130 140 150 160

L F D I H M I G R M G Y R L N<br>TATTCCATATTCATATCATTCCTCCAATCCCATATCCACTAAL ACTATTCGATATTCATATGATTGGTCGAATGGGATATCGACTAAA 170 180 190 200

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10 20 30 40 MDV AATGTCTTTTGAAGTCGAGCCCAATCGAAACCATATTTTTGTCTGCTA TKQLRAWDFGYKQRS

50 60 70 80 90 MDV TCAGAACTAGCAAGTCTCGTTGACAGATGCTCCAAATAAGTG<br>DSSALRTSLHELYT

100 110 120 130 MDV GGAACCGACTCAATCGCACTCATAAAGTTAGTGGGATGAGAAATATT P V S E I A S M F N T P H S I N

140 150 160 170 180 MDV AGTCCCAGTTTTTGCATAGAATGCATATAAACAAAGAATCGCA TGTKAYFA YLCL IA

 $190$  200 210 220 MDV CATTCTAGAGAGGAATAATAACGGGTGCCTACATATAAACGTCCGCA CELSSYYRTGVY LRGC

230 240 250 260 270 MDV TGATTGTAAAGATGTGATTGCCGTCACAATAAACGTTCGCGAC SUN 240<br>GATTGTAAAGATGTGATTGCCGTCACAATAAACGTTCGCG<br>SQLSTIATVIFTRS

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# FIG I4B



PCT/GB89/01076 **P C T / G B 8 9 / 0 I 0 7 6**

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### $FIG$  14C

550 560  $570$ 580 ATTGCAGCGTCGGCAAGTTCTGCTGCAGCCGCTGCATGTTCCAGATC **MDV** MAADALEAAAAAHELD

590 600 610  $620$ 630 CGCTAACGCTGTTGCGATATATTCAATTTTTTTCTTCTATTGGT **MDV** A L A T A I Y E  $\top$  $K$   $E$  $\mathbf{F}$  $\mathsf{T}$  $\mathbf{P}$ 

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640 650  $660$  $670$  $CGAAGTCTGCGGTCAATTTCTATTGGCAATAGAGTCGGTATGACCATC$ **MDV** R L R R D I E I A  $T-S$  $\mathbf{D}$  $\mathbf{T}$  $H$   $G$  $\Box$ 

680 690 700 710 720 CAAATTATTTAATGCTGCAGTGGCGGCATTGTTTCGTGCAGTA **MDV** L N N L A A T A A N N R A T

730 740 750 760 ATGATCGCAAGTTGTCGTTCCATATTGGCGCGGTTAGATGTAAATAC **MDV** I I A L Q R E M N A R N S  $\mathbf{T}$  $\mathbf{F}$  $\mathbf{V}$ 

770 780 790 800 810 CGGTTCCTTCCAGAACTCGATGGGCCATGGGGGAGCTATAAAG MDV PEKW FEI PW PPAIF

**Cn** 

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### FIG 14D

820 830 840 850 MDV TTCTTCACATCGGCAGGGAACATTTCCATTCCATCGCCTGTCAATAT<br>NKVDAPFMEMGDGTLI

860 870 880 890 900 MDV TCTCGCGTCCCAAATAAAGTTTGCCATGATGGTGCTACTCGAT<br>R A D W I F N A M

910 920 930 940 MDV ATAATCAGACAGAAGTTACAGGGAAACGCCACATGAGAAAATAATAC

950 960 970 980 990 MDV TACATTTAAACTACACAAGCTTATAAAAGTGTTACGGTCTCTG -  $\star$  P R Q

*1000 1010* 1020 1030 MDV AACAAGACGGGCGATAATATTAGCCATGTTTCGCATAGCCGTACCT<br>VLRAIINAMNRMATG

1040 1050 1060 1070 1080 MDV CCCGTTCTCTCCTGATTATTTGAAAATGATAAAGTAGCCGTTTT<br>
G T R E Q N N S F S L T A T K

1090 1100 1110 1120 MDV ATTACAAGCTATATGATTCCTCAAATCCGTTACGTTAGCAGACGCC<br>
NCAIHNRLDTVNASA **WO 90/02803** 90/02803

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FIG 14E

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MDV 1130 1140 1150 1160 1170 TTTCCACTGCGTCGTTGTATATGTATCGTGTTTGTATTATGACG <sup>K</sup> <sup>G</sup> <sup>S</sup> <sup>R</sup> <sup>R</sup> <sup>Q</sup> <sup>I</sup> <sup>I</sup> TNT <sup>N</sup> <sup>H</sup> <sup>R</sup>

**W O**

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MDV 1180 1190 1200 1210 TTTTAAAATTTTATGAGTGTCAGTTATCCGTGCTTTATAGTCAGAC<br>K L I K H T D T I R A K Y D S

MDV 1220 1230 1240 1250 1260 GCGGTCGCCAATATAGAGCATAGTCTATGAAAATCAGTCACTAT A <sup>T</sup> A <sup>L</sup> I <sup>S</sup> <sup>C</sup> <sup>L</sup> <sup>R</sup> <sup>H</sup> F <sup>D</sup> <sup>T</sup> <sup>V</sup> I

MDV 1270 1280 129 0 1300 GTGCCTTTTCTTTAGGCACATCACATGTAGAACAGACAGTTTTCGT<br>H R K K L C M V H L V S L K R

MDV 1310 1320 1330 1340 1350 CTTGCTACAAATACTAACATTGGACAAATAACGATACAATCTGA R A V F V L M P C I V I C D S

1360 1370 1380 1390 TCCTTGAGGCGCAATTTGCCCAATCAGAGATTTGGAATCCAATAAC<br>G Q P A I Q G I L S K S D L L MDV

### FIG 14F

1400 1410 1420 1430 1440 MDV TGCTTTATGCCGGTGAGTCTTTGTTCATGTTTACTGCGTGTCTT <sup>Q</sup> <sup>K</sup> I <sup>G</sup> T <sup>L</sup> <sup>R</sup> <sup>Q</sup> <sup>E</sup> <sup>K</sup> <sup>S</sup> R <sup>K</sup>

1450 1460 1470 1480 MDV CAGGTTACGAGAAAATTTGCAAGTTTTTAGTTCTAGAATGACGCAT<br>L N R S F K C T K L E L I V C

1490 1500 1510 1520 1530 MDV ' ACTCCATCACAGCCTACTTCCCACAAATCACGAGGCAACTTAAA V G D C G V E W L D R P L K F

1540 1550 1560 1570 MDV CATGCAAATACAATCCGGTCTACGTCGTTCTAGGTTTACTTCGAAG <sup>M</sup> <sup>C</sup> I <sup>C</sup> <sup>D</sup> <sup>P</sup> <sup>R</sup> <sup>R</sup> <sup>R</sup> <sup>E</sup> <sup>L</sup> <sup>N</sup> <sup>V</sup> <sup>E</sup> <sup>F</sup>

1580 1590 1600 1610 1620 MDV ACCAATCGAAAATCCGTCAACTGTTTAAATACATCTAATACCAT V L R F D T L Q F V D L V <sup>M</sup> MDV V Q S G L  $\mathcal{C}$ M A E A V D L V M<br>Y -----R--------HVT L.  $K$ K Y

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FIG 14G



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# FIG 14H







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FIG 141

. **I -------------------**

RPPTLTRVYLDGPFGI

— S — —<br>— —

 $M$ ------

 $T - S - A - Q - - - - -$ 

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FIG 14J





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FIG 14L





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MDV **HVT** T A D R N M L R T L N A V Y A S CGGCAGATAGAAATATGCTCAGAACACTCAATGCCGTATACGCATC HVT  $1:1:1$ ::::::::  $\mathbf{r}$  $1111$  $\cdots$  $\mathbf{r}$  $\pm$   $\pm$  $\mathbf{z}$   $\mathbf{z}$ **MDV** AAACAGATCTACTAATGCTCAGGGCACTTAATGCAGTGTATTCCTG 2440 2450 2460 2470





FIG 14M

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FIG 14N







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FIG 140

ATGACTA.....TCGCCAAACTGTTAAACCCGCGAAGAATATAT

 $\mathbf{r}$  :  $\mathbf{r}$ 

 $\ddot{z}$ 

 $\ddot{\phantom{1}}$ 

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**HVT** 

MDV

 $\mathbf{1}$ :

 $\mathbf{1}$ :





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 $MDV$ 

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 $---S---M---A---A---M---P------R---R---R---N---V---Q-$ 



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#### FIG 14 U

LN PGYKFVLTS.ELVG HVT CTCAACCCCGGATACAAGTTCGTTCTCACAAGCGAGTTGGTAGGAG 3620 3630 3640 3650

>

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**P C T / G B 8 9 / 0 1 0 7 6**

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A Y T K R S C F V D P M D S L HVT CCTACACAAAACGATCTTGTTTTGTCGATCCGATGGATTCTCTC 3660 3670 3680 3690 3700

VPIDYDHVRTIIFGS<br>TECCOLORACATION CARCATION CCAACOATION HVT GTCCCGATAGATTATGATCATGTACGAACCATTATATTCGGATCTG 3710 3720 3730 3740

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A G  $L_{\rm L}$ M<sub>K</sub> M G T L A S  $M$  E I I HVT CTGGGATGGAGATTTTAATGAAGATGGGAATTACTTTGGCATCT 3750 3760 3770 3780 3790

3/50 3760 3770 3780 3790<br>
MTISTKYNPPIELII<br>HVT ATGACCATTTCGACGAAATATAATCCTCCTATTGAACTGATAATAT 3800 3810 3820 3830

SAKYRNLSLLWPPRQ HVT CTGCAAAGTACCGAAATTTATCACTGTTGTGGCCACCCCGACAA 384.0 3850 3860 3870 3880

# FIG 14V

QYEPVNKGTGRPHWI<br>AATATGAAGGTGAAGGAAGGAAGGGGGAAFTGAA HVT CAATATGAACCTGTAAATAAAGGGACTGGACGCCCCCATTGGATCT 3890 3900 . 3910 3920

<sup>Y</sup> <sup>L</sup> <sup>L</sup> <sup>G</sup> <sup>V</sup> <sup>Y</sup> <sup>R</sup> <sup>N</sup> <sup>V</sup> SDS <sup>E</sup> <sup>R</sup> <sup>D</sup> HVT ACCTATTAGGTGTGTATAGAAACGTTTCGGACTCCGAGCGTGAC 3930 3940 3950 3960 3970 S Y M N M I K S L G D S M D Y HVT TCATACATGAATATGATTAAGAGTCTGGGCGATTCTATGGATTATC 3980 3990 4000 4010

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<sup>H</sup> <sup>F</sup> <sup>L</sup> <sup>I</sup> <sup>S</sup> <sup>R</sup> AHA <sup>Q</sup> <sup>M</sup> <sup>L</sup> ILA HVT ACTTCCTAATTAGCAGAGCGCATGCCCAGATGCTGATACTGGCA<br>4020 4030 4040 4050 4060 4020 4030 4040 4050 4060

A E D R L V D E H S F R N V HVT GCAGAGGACCGGCTCGTGGATGAAATGCATAGTTTCAGGAACGTTA 4070 4080 4090 4100

I A R L F V S L F A F I R N A HVT TTGCGCGTTTATTTGTATCGTTGTTCGCATTCATACGTAACGCA 4110 4120 4130 4140 4150

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### FIG 14W

HVT FOSGYTSLNDI.IEIE TTTCAGTCTGGCTACACCTCTCTTAATGACATAATTGAAATCGAAG 4160 4170 4180 4190

HVT ADLRL IVEGISSAAF CCGATTTGAGGTTAATTGTAGAAGGCATTTCTTCTGCTGCATTT 4200 4210 4220 4230 4240

HVT CGTAAAGACGCTAGTACACACTTTCTTATATCGGGAACGCCCATAA RKDAS THFLI SGTPI 4250 4260 4270 4280

HVT AAGATAGCAAAGCGGATTTAATTAAATCGTTGTTGTCTAAAGTC KDSKADLIKSLLSKV 4290 4300 4310 4320 4330

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HVT ATTCGACCAATTTCCGGACATACACGTCCCTTATCTGCGATACAAC IRPISGHTRPLSAIQ 4340 4350 4360 4370

HVT **c\_n** HLFLLRSAYALDIPR | ATCTATTCCTTTTGAGATCCGCTTATGCATTGGATATACCCCGT 4380 4390 4406 4410 4420

G8/<sub>5</sub>

## FIG 14X

QNGSLSEQVST.VALS HVT CAAAACGGATCTTTGAGCGAACAGGTATCTACAGTGGCACTGTCGT 4430 4440 4450 4460

FIENIHSEAMRDILS<br>TOATTOAAAATATTOAGAGOGAGOGAGAGOGAGAGTOTOTO HVT TCATTGAAAATATTCACAGCGAGGCCATGAGGGACATTCTGTCA 4470 4480 4490 4500 4510

WNTTTKHALYYAFAS HVT TGGAACACTACAACAAAGCATGCGTTGTATTATGCATTCGCGAGTA 4520 4530 4540 4550 4520 4530 4540 4550<br>ILQRPLTE WGAS RNA

HVT TTTTGCAACGGCCACTGACCGAATGGGCGCCTCAAGAAATGCA<br>4500 4500 4500 4560 4570 4580 . 4590 4600

RRAILLASSMCTEEH<br>GCAGGGGAADAGDADDAGGADGGADGGAAGAAGAAGG HVT CGGAGGGCAATACTATTAGCATCATCGATGTGTACAGAAGAGCATG 4610 4620 4630 4640

HVT VIATELAIQELYVKI<br>WIATELAIQELYVKI TTATCGCAACTGAGTTGGCTATTCAAGAACTGTATGTCAAAATC 4650 4660 4 6 4680 4690 WO 90/02803 90/02803

#### • FIG 14Y

RSNADPI HLLD.VYTP HVT AGAAGTAATGCCGACCCAATACACCTTCTAGACGTATATACACCAT 4700 4710 4720 4730

CLSSL RLDLSEHHRI<br>CLSSL RLDLSEHHRI  $\rm HVT = GTCTTTCTTCACTACGATTGGACCCTTTCCGAACACACACGATCGGATA$ 4740 4750 4760 4770 4780

Y A M A D V V F Y P D I Q Q Y HVT TACGCAATGGCAGATGTAGTTTTCTATCCAGACATTCAGCAGTATT 4790 4800 4810 4820

LKKKS HEGNMKEDDL HVT TGAAAAAAAAATCCCATGAGGGTAATATGAAGGAAGATGATCTC 4830 4840 4850 4860 4870

E T K A E Y I L T K L R S P L<br>AAACAAACCCCCAATACATCCTCACCAACCTTACCTCCCCCTT HVT GAAACAAAGGCGGAATACATCCTCACCAAGCTTAGGTCGCCGTTGA 4880 4890 4900 4910

IRTLSAYASEVLSCS HVT TCAGAACGCTGTCTGCCTATGCATCAGAAGTATTGTCCTGCTCC  $4920$   $4930$   $4940$   $4950$   $4960$ 

**W O**

**COO**

# FIG 14Z

DQDLL E I N A I L.I L P V<br>DQDLL E I N A I L.I L P V HVT GACCAGGATCTATTAGAAATAAATGCTATTTTAATTCTGCCCGTTT 4970 4980 4990 5000

S
G
I
G S Y v v 's R R A G M Q HVT CCGGTATTGGGAGCTATGTAGTCTCTCGAAGGGCAGGAATGCAA<br>5010 5020 5030 5040 5050 5010 5020 5030 5040 5050

<sup>G</sup> I <sup>V</sup> <sup>Y</sup> <sup>T</sup> <sup>V</sup> <sup>D</sup> <sup>G</sup> <sup>V</sup> <sup>D</sup> <sup>V</sup> <sup>N</sup> <sup>Q</sup>  $O$  L HVT GGCATTGTTTATACGGTAGACGGTGTTGATGTTAACAATCAGCTTT 5060 5070 5080 5090

 $T$ R  $\mathbf{M}$ T F  $\top$  $\mathbf{T}$ Y P C  $\mathbf T$  $\mathbf{T}$  $\mathsf{T}$ G N HVT TTATAACATATACCAGGATGCCGTGCACTACAACGATAGGTAAC 5100 5110 5120 5130 5140

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I V P T V L <sup>S</sup> R P S <sup>G</sup> <sup>K</sup> T <sup>C</sup>  $C \cdot P$ HVT ATTGTTCCAACAGTATTGTCAAGACCCTCGGGAAAAACGTGTCCGT 5150 5160 5170 5180

Y C G C V L L R Y S A D G N I HVT ATTGCGGCTGTGTTTTGCTGCGATATTCCGCCGATGGAAATATC<br>5230 5230 5230 5190 5200 5210 . 5220 5230 78/80

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FIG 14ZZ

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CGCTATTCTATTTACATTTCGTCCC<br>
5240 5250 HVT

# Figl5

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GRRKYDALVA-F GGGACGACGCAAATATGATGCTCTAGTAGCAT4GTTT

VL GRACGRPIYLR GTCTTGGGCAGAGCATGTGGGAGACCAATTTATTTACGT

E Y A N C S T N E P F G T GAATATGCCAACTGCTCTACTAATGAACCATTTGGAACT

CKLKSLGWWDRRY TGTAAATTAAAGTCCCTAGGATGGTGGGATAGAAGATAT

AMT SYIDRDELKL GCAATGACGAGTTATATCGATCGAGATGAATTGAAATTG

I I A A P S R E L S G L Y ATTATTGCAGCACCCAGTCGTGAGCTAAGTGGATTATAT

ATTATTGCAGCACCCAGTCGTGAGCTAAGTGGATTATAT<br>
TRLIINGEPISS<br>ACGCGTTTAATAATTATTAATGGAGAACCCATTTCGAGT

**CO** - **CO**

**PCT/GB89/01076 P C T / G B 8 9 / 0 1 0 7 6**

 $\sum_{i=1}^{n}$ 

<sup>D</sup> I <sup>L</sup> <sup>L</sup> <sup>V</sup> <sup>K</sup> GACATATTACTGACTGTTAAA