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(54) **NOVEL METALLOPROTEASES**

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(2013.01); **C11D 3/386** (2013.01); **C12Y 304/24**  
(2013.01)

(57) **ABSTRACT**

Aspects of the present compositions and methods relate to novel metalloproteases, polynucleotides encoding the novel metalloproteases, and compositions and methods for use thereof.

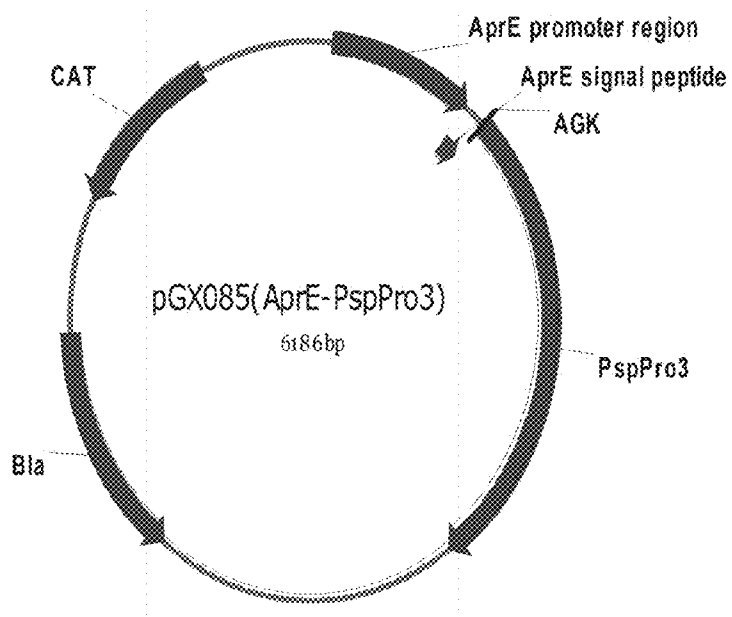


Figure 1.1. Plasmid map of pGX085(AprE-PspPro3).

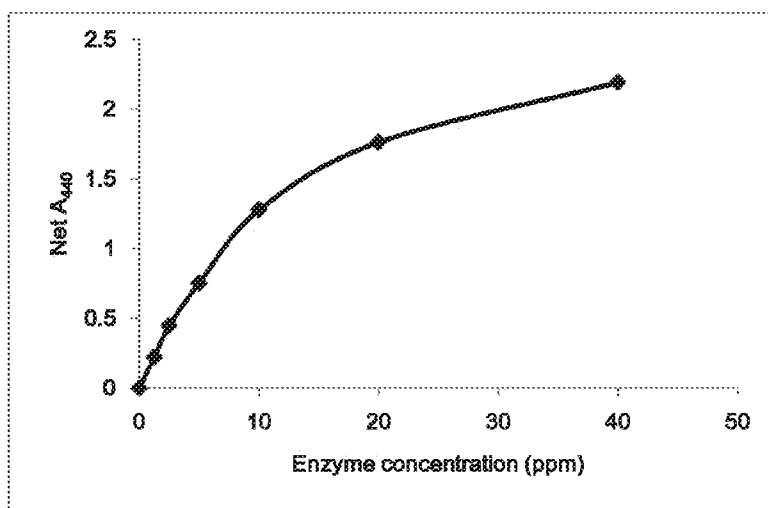


Figure 1.2. Dose response of PspPro3 in azo-casein assay at pH 7.

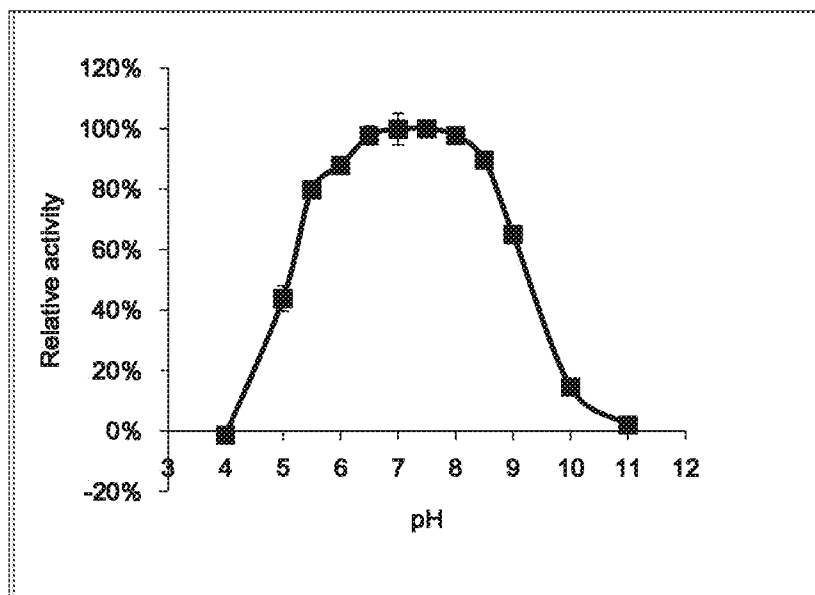


Figure 1.3. pH profile of PspPro3.

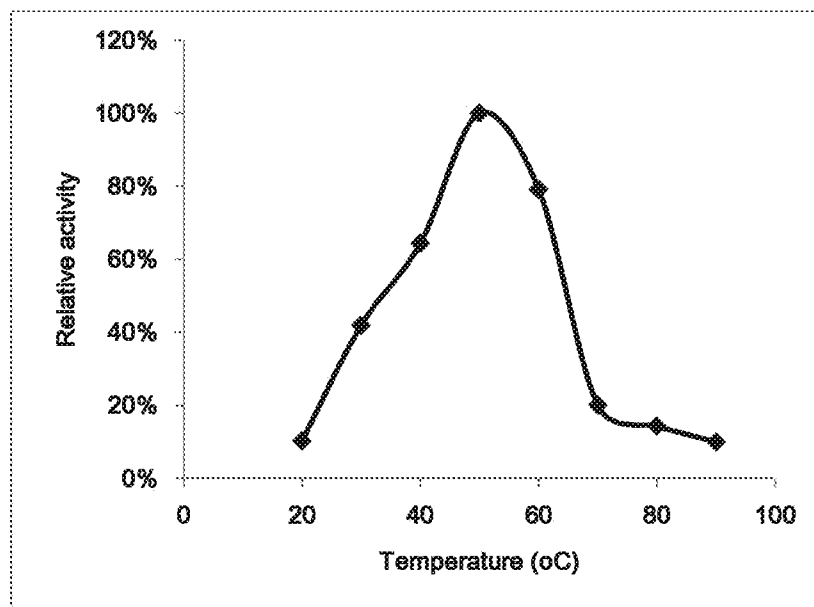


Figure 1.4. Temperature profile of PspPro3.

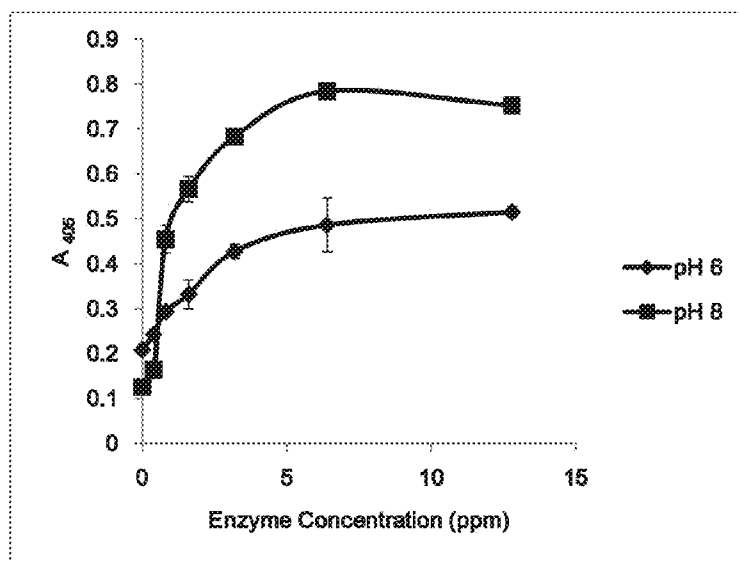


Figure 1.5A. Cleaning performance of PspPro3 at pH 6 and 8 in AT dish detergent.

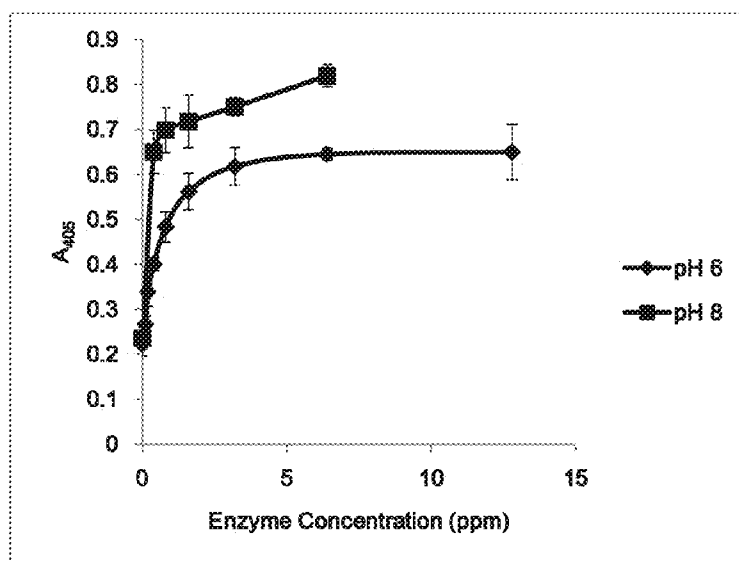


Figure 1.5B. Cleaning performance of PspPro3 at pH 6 and 8 in AT dish detergent with bleach

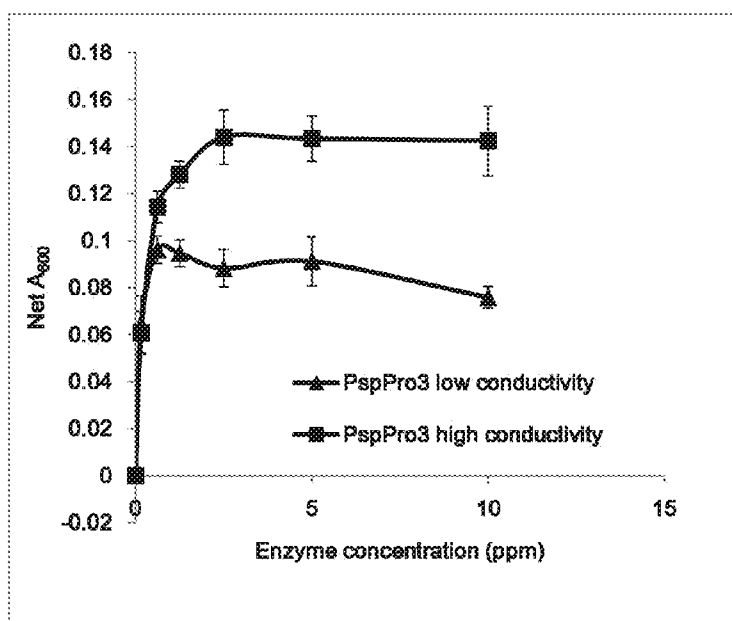


Figure 1.6. Cleaning performance of PspPro3 in liquid laundry detergent at pH 8

CLUSTAL W (1.83) multiple sequence alignment

```

PspPro3          -----ATGTCCKGVLGDTKTFNNTASGSSVQLRDTTRGNGIVTYTASNRRQS
Paenibacillus_sp_Aloe-11  ---NRATGTGKGVLDGDKTFNNTASGSSVQLRDTTRGNGIVTYTASNRRQS
B_thermoproteolyticus_P00800  ITGTSTVGVGRGVLGDQKMINITYG-TYYYLQDNTRGGIGTYDANKRYRT
      :*:*:*:*:*:* *:*:*:* * : * *:*:*:*:*:*:* * * :
PspPro3          IPGTILTDADENVWN----OPAGVDAHAYAANTIDYINEKFPNRNSIDGRGLQ
Paenibacillus_sp_Aloe-11  IPGTILTDADENVWN----DPAGVDAHAYAANTYGYEYKFPNRNSIDGRGLQ
B_thermoproteolyticus_P00800  LFGSLWADADNQFFASVDAPAVDAHYAGVTYDYKKNVHNRLSYDGNNA
      :*:*: :*:*: : *..*:* *:.*****: .* * *..
PspPro3          LRSTVHYGNRYNNAFWNGSQMTYGGDGTFFIAFSGDPPDVVGHETHGVT
Paenibacillus_sp_Aloe-11  LRSTVHYGNRYNNAFWNGSQMTYGGDGTFFIAFSGDPPDVVGHETHGVT
B_thermoproteolyticus_P00800  IRSVYHYGQYNNNAFWNGSQMVYGDGQGTFFIFLSGGIDVVAHELTHAVT
      :*:*:*:*: *****:***** *:*:*:*:*:*:*:*:*:*:*
PspPro3          EYTSNLEYYGESGALNEAFSDIIGNDIQ-----RKNWLVGDDIYTPRIAG
Paenibacillus_sp_Aloe-11  EYTSNLEYYGESGALNEAFSDIIGNDIQ-----RKNWLVGDDIYTPRIAG
B_thermoproteolyticus_P00800  DYTAGLTYQNESGALNEAISDIFPGTLVEFYANKRNPDWEIGEDVYTPGLSG
      :*:*:* * * .*****:*:*:*:*:*: . : . :^ :^:*:* *:*
PspPro3          DALRSMSENPTLYDQPDHYSNLYRGSDDNGGVHTNSGIINKAYYLLAQQGT
Paenibacillus_sp_Aloe-11  DALRSMSENPTLYDQPDHYSNLYRGSDDNGGVHTNSGIINKAYYLLAQQGT
B_thermoproteolyticus_P00800  DSLRSMSSDPAKYGFDPHYSNRYTGTQDNGGVHTNSGIINKAYYLLISQGT
      *:*:*:*:*:*: *:*:*:*:*: * *:*:*:*:* *:*:*:*:*:*:*:*:*:*
PspPro3          FHGVTVNGIGRDAAVQIYYSAFNTNYLTSDDFSNARDAVVQAAKDLYGAS
Paenibacillus_sp_Aloe-11  FHGVTVNGIGRDAAVQIYYSAFNTNYLTSDDFSNARDAVVQAAKDLYGAS
B_thermoproteolyticus_P00800  HYGVSVVVIGRDKLGIKIFRALTQYLTPTSNFSQLRAAAVQSATDLYGST
      :*:*:* * * * * * :*:* *:*:*:*:*:*:*: * *:*:*:*:*:*:*
PspPro3          SAQATAAAKSFDAVGVN (SEQ ID NO: 3)
Paenibacillus_sp_Aloe-11  SAQATAAAKSFDAVGVN (SEQ ID NO: 44)
B_thermoproteolyticus_P00800  SQEVASVRQAPDAVGVK (SEQ ID NO: 45)
      * :*:*: .:*****:

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Figure 1.7. Alignment of PspPro3 with protease homologs

Phylogenetic tree

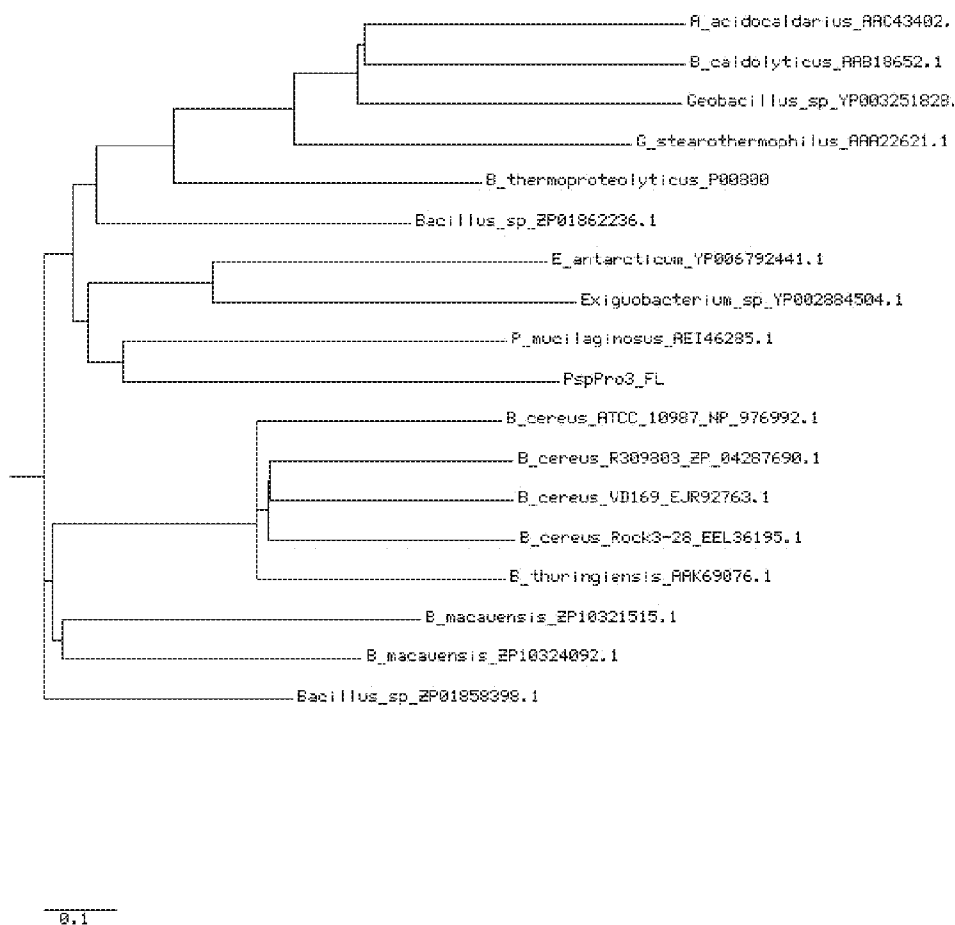


Figure 1.8. Phylogenetic tree of PspPro3 and homologs.

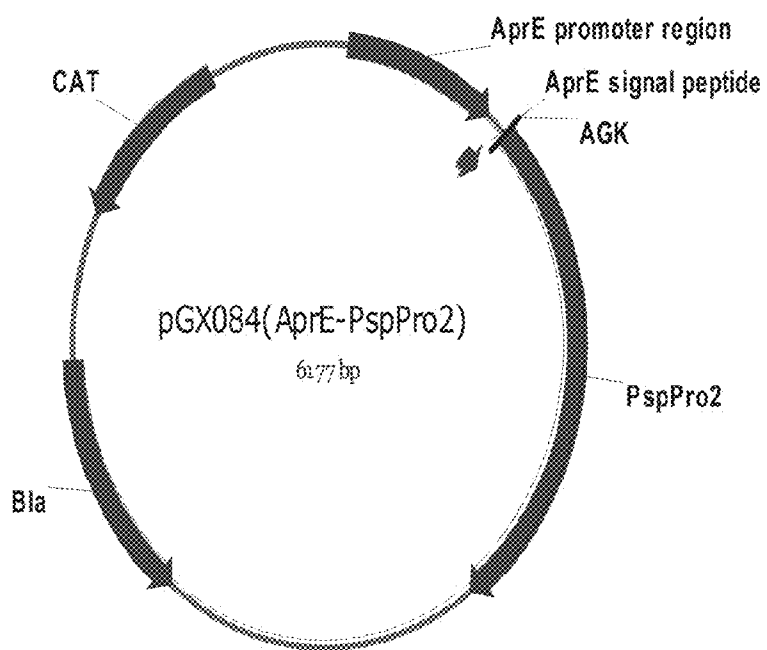


Figure 2.1. The map of plasmid pGX084(AprE-PspPro2).



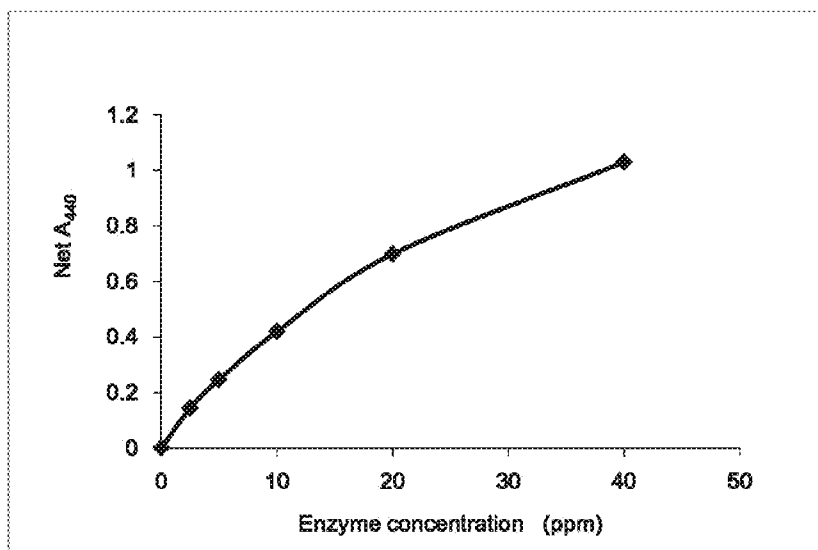


Figure 2.2. Dose response curve of PspPro2 in azo-casein assay at pH 7.

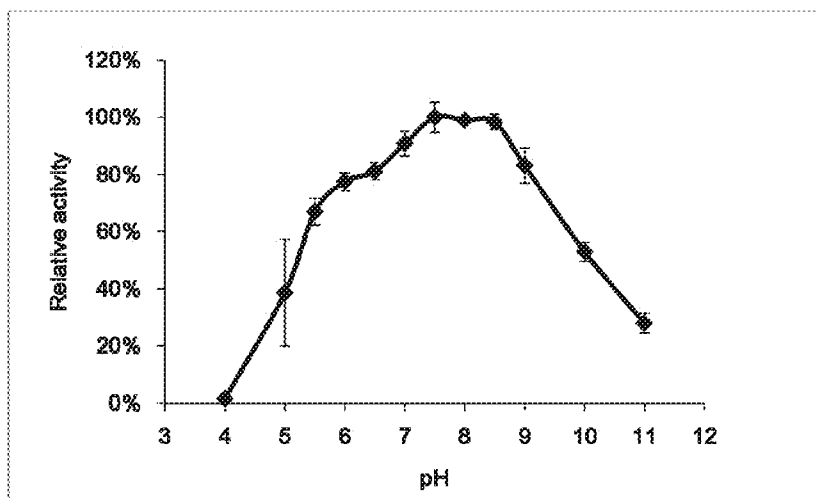


Figure 2.3. pH profile of purified PspPro2.

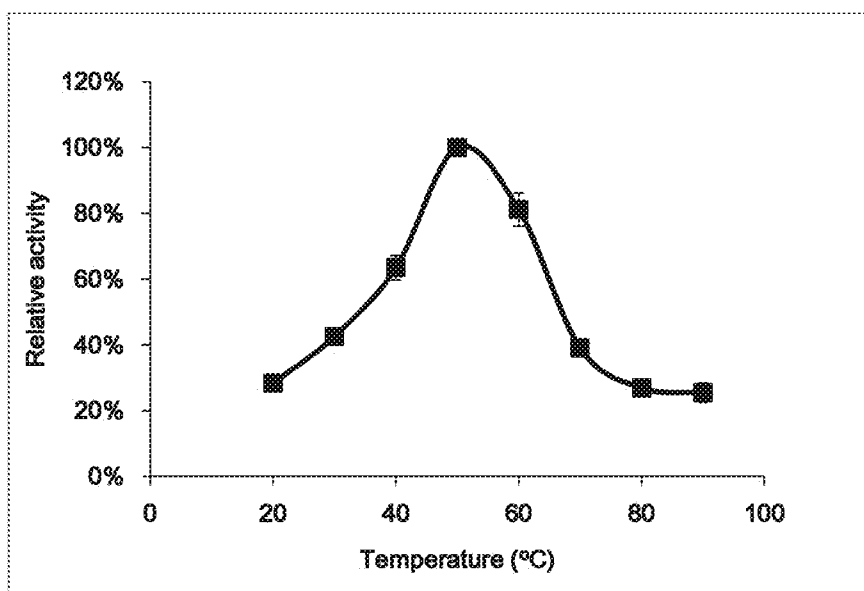


Figure 2.4. Temperature profile of purified PspPro2.

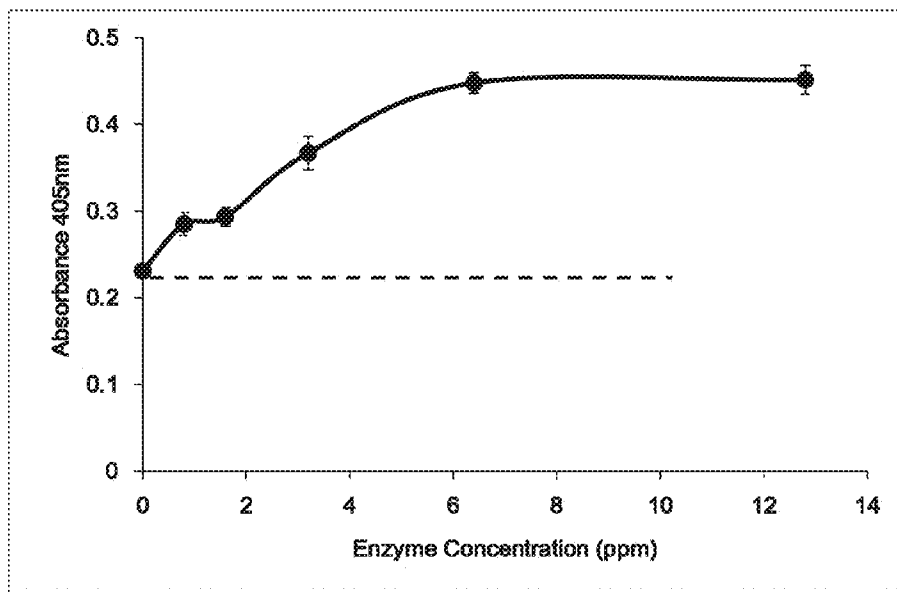


Figure 2.5A. Cleaning performance of PspPro2 protein at pH 6 in AT detergent with bleach.

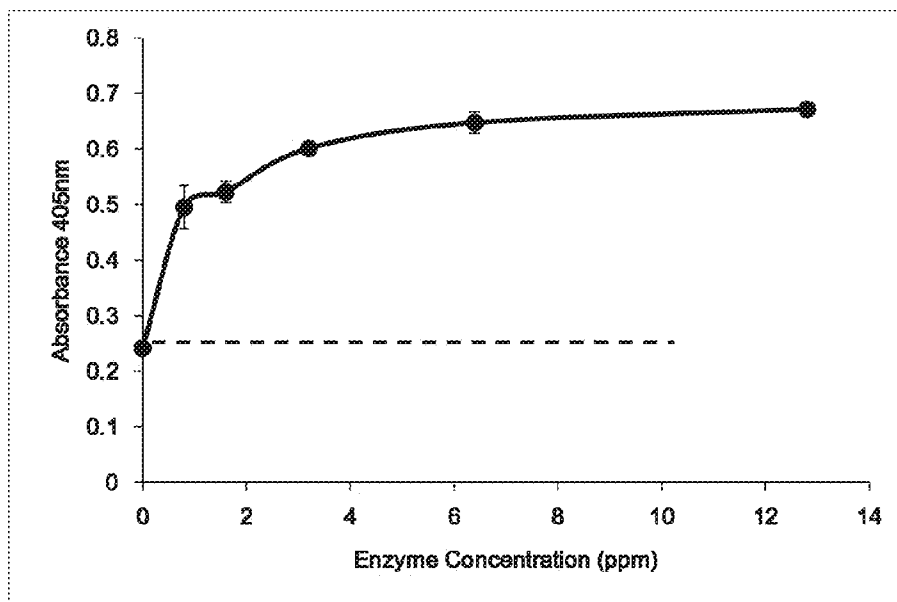


Figure 2.5B. Cleaning performance of PspPro2 protein at pH 8 in AT detergent with bleach.

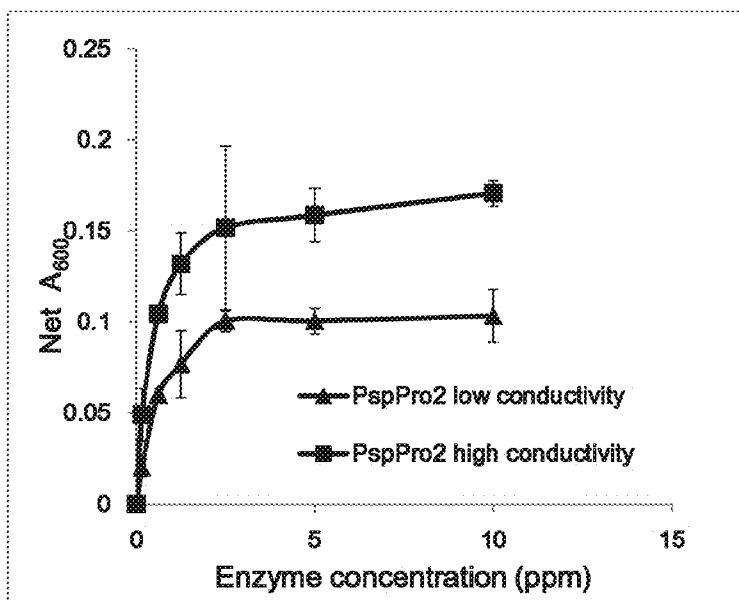


Figure 2.6A. Cleaning performance of PspPro2 protein in liquid laundry detergent.

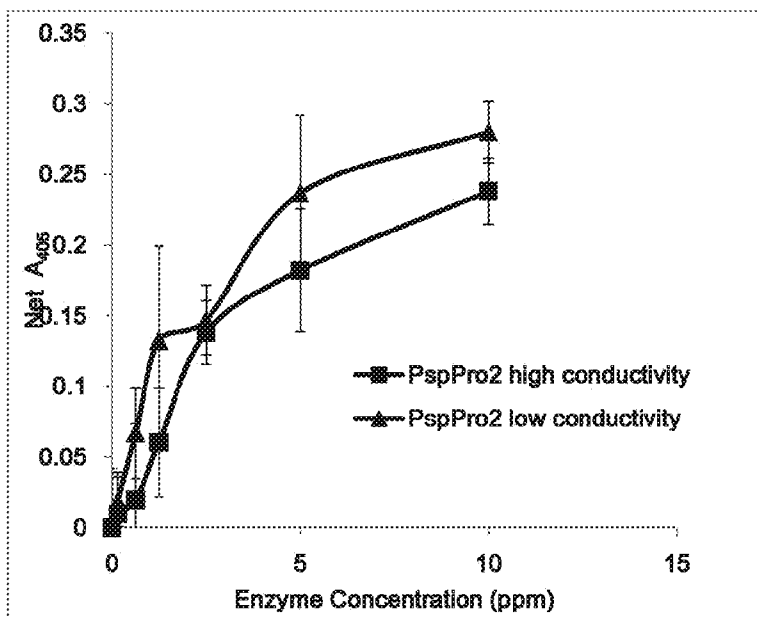


Figure 2.6B. Cleaning performance of PspPro2 protein in powder laundry detergent.

CLUSTAL W (1.83) multiple sequence alignment

```

PspPro2          -----A TGTGRGVDGKTKSFTTTASGNRYQLKDTTRSNGLVITYTAGNROF
ZP_09775365.1_P_sp_Aloe-11  -----ATGTGRGVDGKTKSFTTTASGNRYQLKDTTRSNGLVITYTAGNROF
E_thermoproteolyticus_F00800  ITGTSTVWVGRGVLGDQNNINITYS--TYYYLQDNIRGMGIPTYDAKYRTT
      :*,*****.*,*.,**.*. * *,*,**,* ** * * *

PspPro2          TPGTILTDIDNVW---EDPAAVDAHAYAIKTYDYKKNKFGKRSIDGRGMQ
ZP_09775365.1_P_sp_Aloe-11  TPGTILTDIDNVW---EDPAAVDAHAYAIKTYDYKKNKFGKRSIDGRGMQ
E_thermoproteolyticus_F00800  LPCS LWADADNQPFASYDAPAVDAHYYACVTYDYKKNVHNRLSYDGNNA
      **::*:** : *..***** ** ***** ..* * **..

PspPro2          IRSTVHYGKYNNAFWNGSQMTYGDGDSFTFFSGDQDVVGHLELTHGVT
ZP_09775365.1_P_sp_Aloe-11  IRSTVHYGKYNNAFWNGSQMTYGDGDSFTFFSGDQDVVGHLELTHGVT
E_thermoproteolyticus_F00800  IRSSVHYSGYNNAFWNGSQMVYGDGDSFTFFSGGIDVVAHELTHAVT
      ***:**.: *****,** ** :*, **,* ** **

PspPro2          EFTSNLEYEGESSALNEAFSDIIGNDID-----GTSWLLGDGIYTPNIPG
ZP_09775365.1_P_sp_Aloe-11  EFTSNLEYEGESSALNEAFSDIIGNDID-----GTSWLLGDGIYTPNIPG
E_thermoproteolyticus_F00800  DYTAGL IYQNESSALNEAFSDIFGTLVEFYAKKNPDWEIGEDVYTPGISG
      :*,*.* ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** **
      :*,*.* ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** **

PspPro2          DALRSLSDPTRFGQPDHYSNFPDPMNDDECGVHINSGIINKAYYLLAQC
ZP_09775365.1_P_sp_Aloe-11  DALRSLSDPTRFGQPDHYSNFPDPMNDDECGVHINSGIINKAYYLLAQC
E_thermoproteolyticus_F00800  DSLRSMSDPARYGLPDHYSRKYT--GTQDNGGVHINSGIINKAAYLISQC
      *:**:**:*,** ** : * ..*:** ** ** ** ** ** ** ** ** ** ** **

PspPro2          GTSHGVIVTGTGREAAVFIYNAFTNYLITSTSNF SNARA AVIQAAKDFYQ
ZP_09775365.1_P_sp_Aloe-11  GTSHGVIVTGTGREAAVFIYNAFTNYLITSTSNF SNARA AVIQAAKDFYQ
E_thermoproteolyticus_F00800  GTHYGVSVVGTGRDKLGMIFYRALTQYLTPTS KPSQLRAAAVQSACDLYG
      ** :*,*.* ** ** : *,*.* ** ** ** ** ** ** ** ** ** ** ** ** ** ** ** **

PspPro2          ADSEAVTSAIQSFDVAVGIK SEQ ID NO: 8
ZP_09775365.1_P_sp_Aloe-11  ADSEAVTSAIQSFDVAVGIK SEQ ID NO: 46
E_thermoproteolyticus_F00800  STSQEVASVKQAPDVAVGK SEQ ID NO: 45
      : * *.* * ** ** **
    
```

Figure 2.7: Alignment of PspPro2 protein with homologous protease sequences.

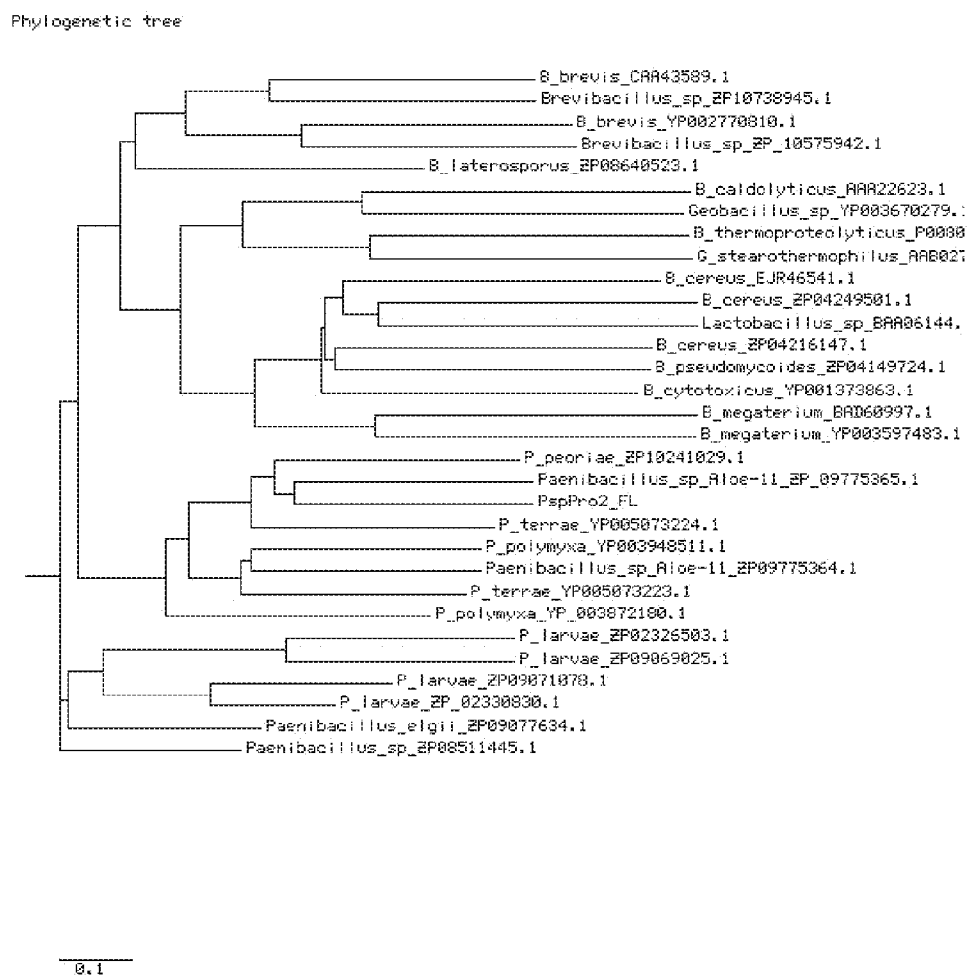


Figure 2.8: Phylogenetic tree for PspPro2 and its homologs

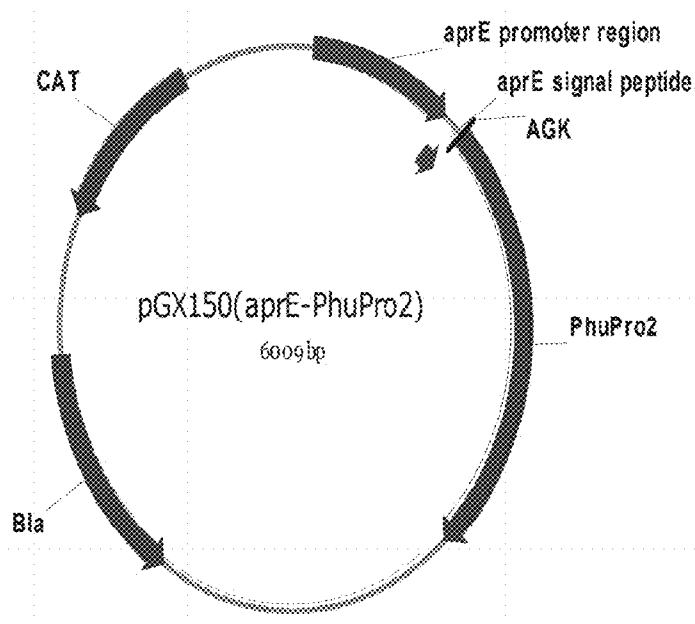


Figure 3.1. The plasmid map of pGX150 (AprE-PhuPro2).

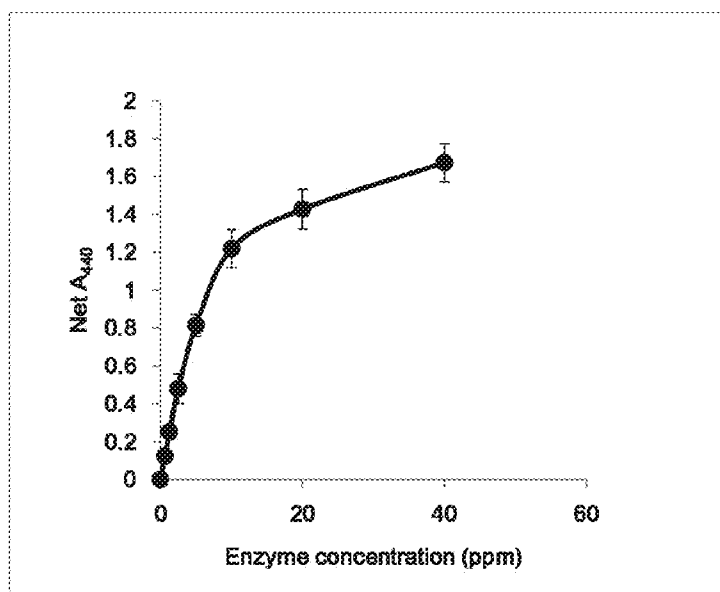


Figure 3.2. Dose response curve of PhuPro2 in azo-casein assay at pH 7.

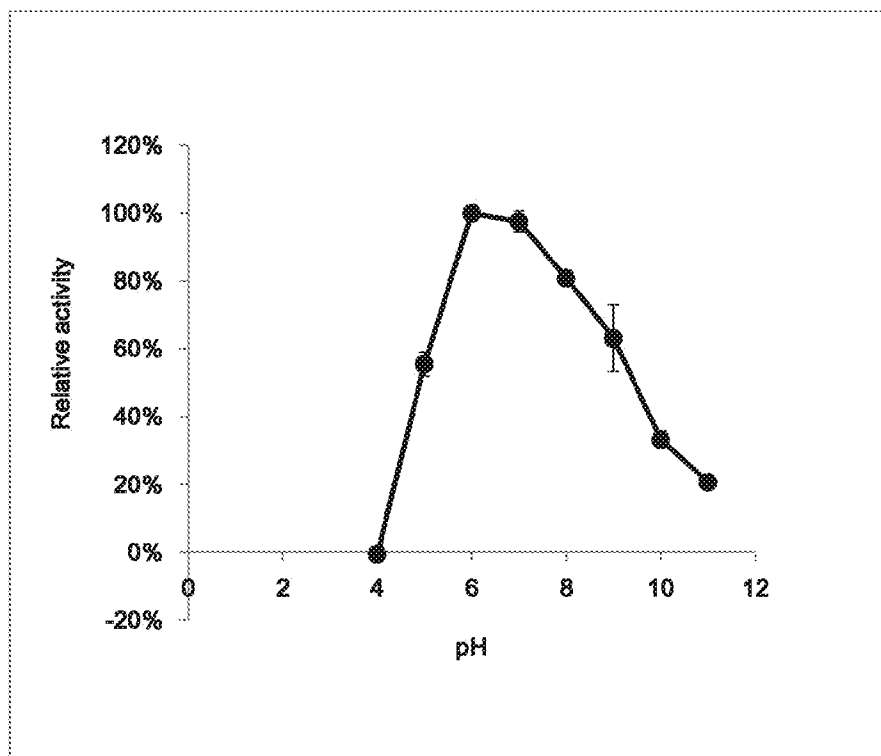


Figure 3.3. pH profile of PhuPro2.

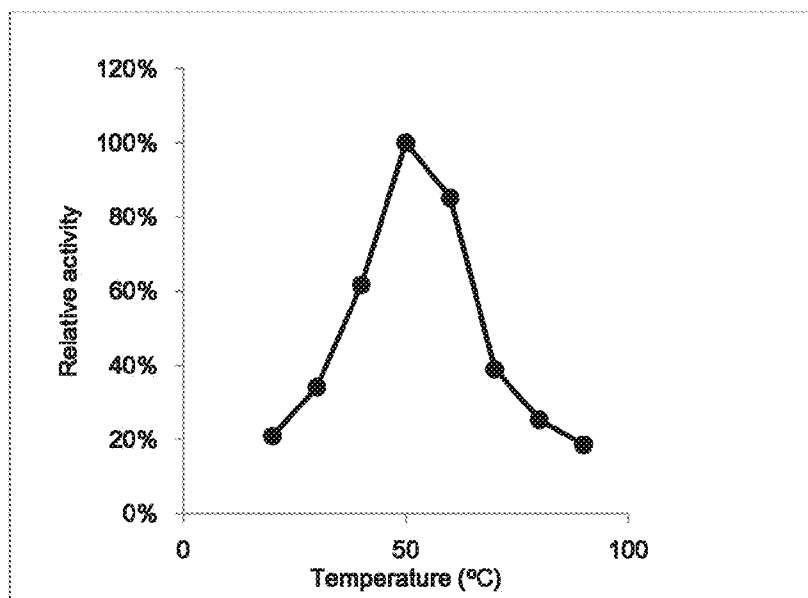


Figure 3.4. Temperature profile of PhuPro2.



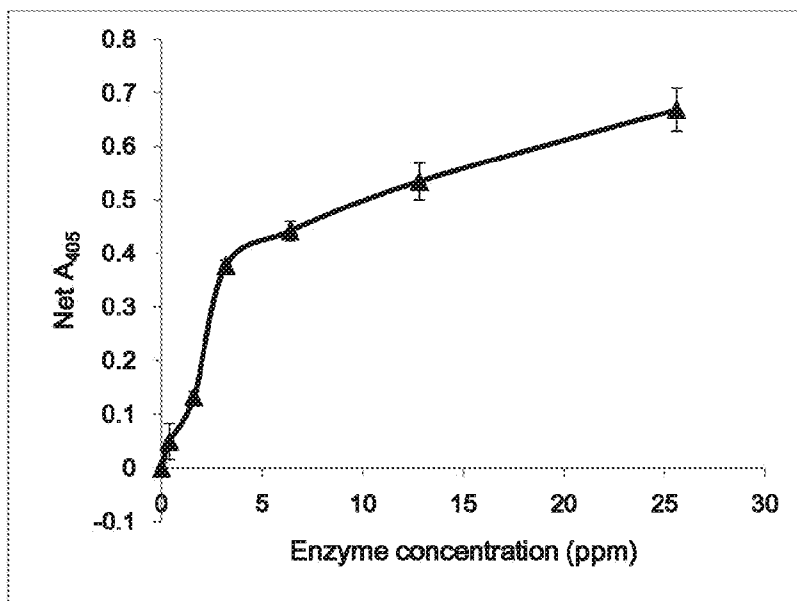


Figure 3.5A. Cleaning performance of PhuPro2 in AT dish detergent at pH 6.

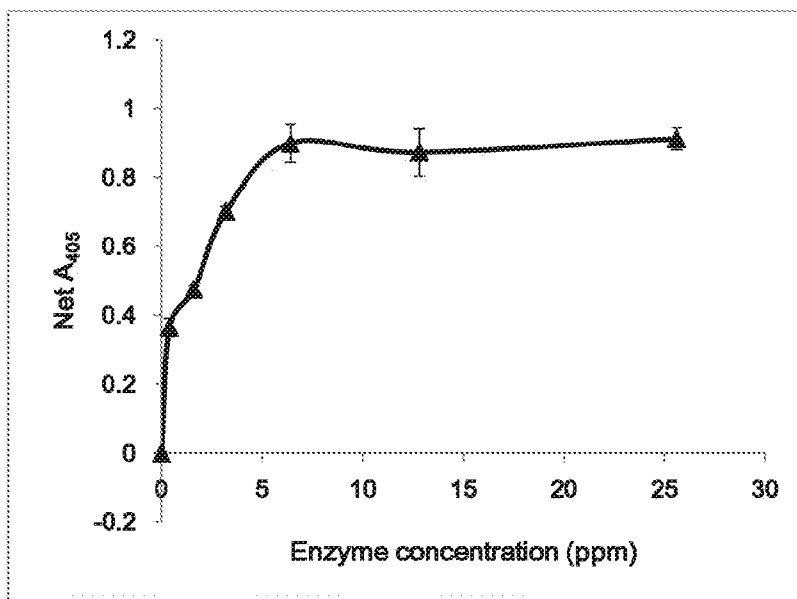


Figure 3.5B. Cleaning performance of PhuPro2 in AT dish detergent at pH 8.

CLUSTAL W (1.83) multiple sequence alignment

```

PhuPro2          -----ATGSGTGVVLGDNKTFQTTLSCSTYQLKDTTRGNGIYTYTASNRFT
P__terrae__HPL-003_YP_005073223.  -----ATGTGKGVLDGDKSFNTTQSGS SYQLKDTTRGNGIYTYTASNRQT
E__thermoproteolyticus_P00800      ITSTSIYGVGGRVLDQQRNINTYVS-TYYLQDNTRGNGIFTYDAKYRRT
                                   :.* * ***** *.:*** * : * *;*.***** * * . * *

PhuPro2          IPGTLLTDADNVWT---DGRAVDAHYAGKVYDFYKTKFGRN SLDGNGLL
P__terrae__HPL-003_YP_005073223.  IPGTLLTDADNVWN----DFAGVDANAYAAKTYDYKDFGRNSIDGRGLQ
E__thermoproteolyticus_P00800      LFGSLWADADNQFFASYDAPAVDAHYYAGVYDYDKVNHRLSYDCNNAA
                                   :**:* :***** : * .:***** ** .:**;* ..* * *..

PhuPro2          IRSSVHYSSRYNNAFWNGTQIVFGDGDGSTEPLPSGLDLDVVSHEL SSGVI
P__terrae__HPL-003_YP_005073223.  IRSTVHYGSRYNNAFWNGSQMTYGDGDGTTFTAFSGDPDVVGHLLTHGVI
E__thermoproteolyticus_P00800      IRSSVHYSQYNNAFWNGSQMVGCDGCGQTFIPLSGGIDVVAHELLHAVI
                                   :**:* * . *****;*.:***** **;**. **.* **.*

PhuPro2          EYTSNLQYLNESGALNESYADV LGNISIQ-----AKNWLIGDDVYTPGISG
P__terrae__HPL-003_YP_005073223.  EYTSNLQYKQNESGALNESYSDI KGNISIQ-----RNWLVGDDVYTPGISG
E__thermoproteolyticus_P00800      DYTAGLIYQNESGALNESAI SDI FGLVDFYANENPCWEIGEDVYTPGISG
                                   :**:* * .*****:**: *:* .: : * :*:***.**:

PhuPro2          DALRSMGNPTLYGQPDNYANRYIGSSDNGGVHINSGI INKAFYLLAQGGT
P__terrae__HPL-003_YP_005073223.  DALRSMGNPTLYDQPDHYSNLYKGS SDNGGVHINSGI INKAFYLLAQGGT
E__thermoproteolyticus_P00800      DSI RSMSPANYPDPKYSKRYTGTQDNGGVHINSGI INKAFYLLAQGGT
                                   *:*****:*.:***:*.: *:* .***** * ** * ** **;***

PhuPro2          QNGVTVAGIGRDAAVNI FYNTVAYILYLTSTSNFAAAKNASIQAAKDL YGTC
P__terrae__HPL-003_YP_005073223.  PHNVTVSGIGRDAAVQIYYSAFINYLSTSNFSNTRAAVVQAAKDL YGAN
E__thermoproteolyticus_P00800      HYGVSVVGI GRDKLGI FYRALTYLTPFSNFSQLRAAVQSATDLKGST
                                   .*: * ***** :*:* .: : ***,*****: : * :*:***.

PhuPro2          SSYVTSVTNAPRAVGL- SEQ ID NO: 13
P__terrae__HPL-003_YP_005073223.  SAQATAAAKSFDAVGVN SEQ ID NO: 47
E__thermoproteolyticus_P00800      SQEVASVQAQFQAVGVK SEQ ID NO: 45
                                   * .: .: .: * **;

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Figure 3.6: Alignment of PhuPro2 with homologous protease sequences.

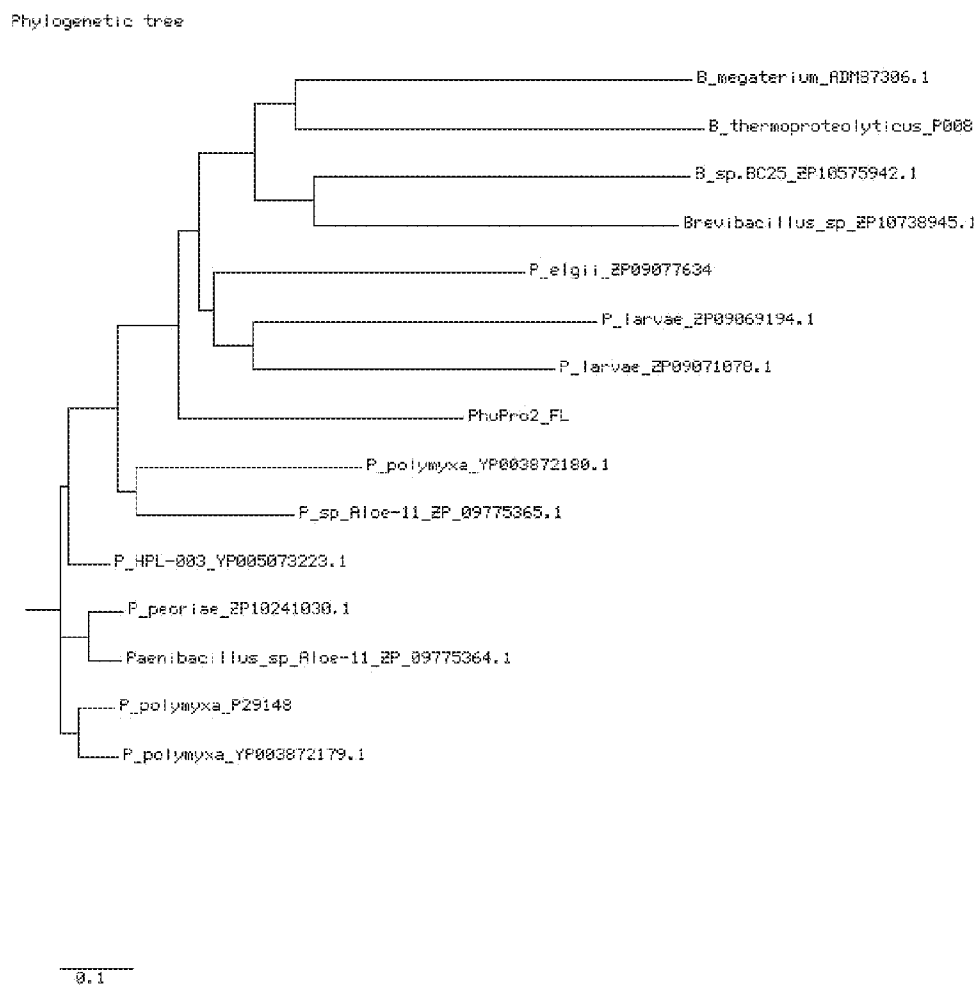


Figure 3.7: Phylogenetic tree for PhuPro2 and homologs.

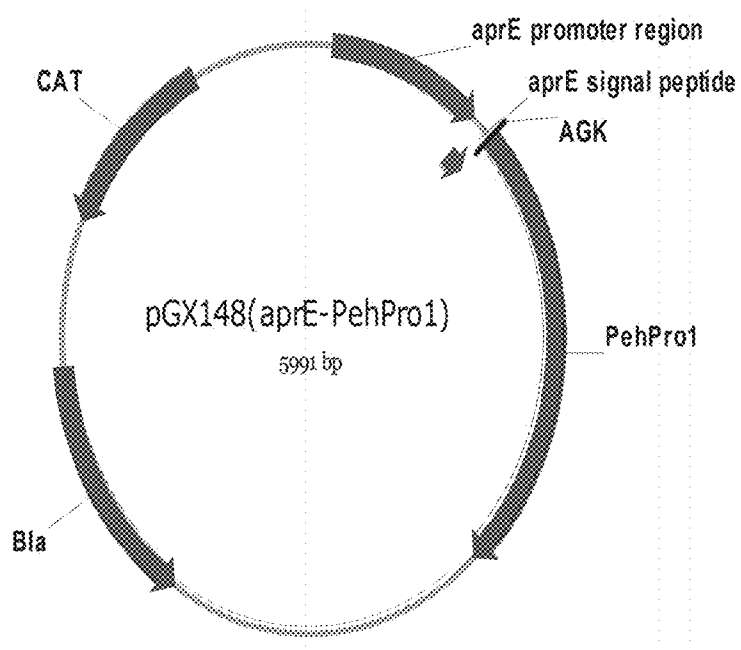


Figure 4.1. The plasmid map of pGX148 (AprE-PehPro1).

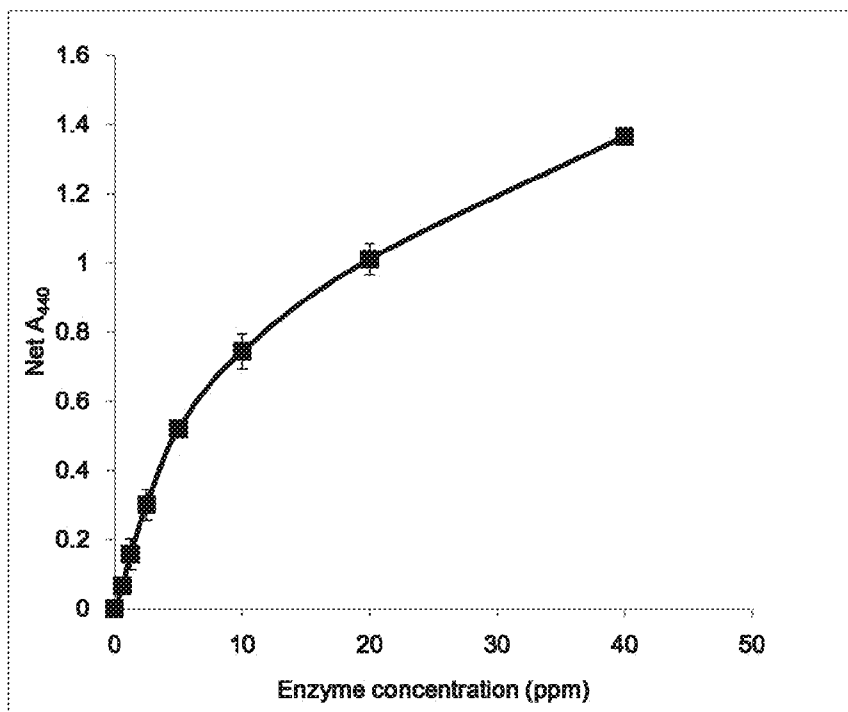


Figure 4.2. Dose response curve of PehPro1 in azo-casein assay at pH 7.

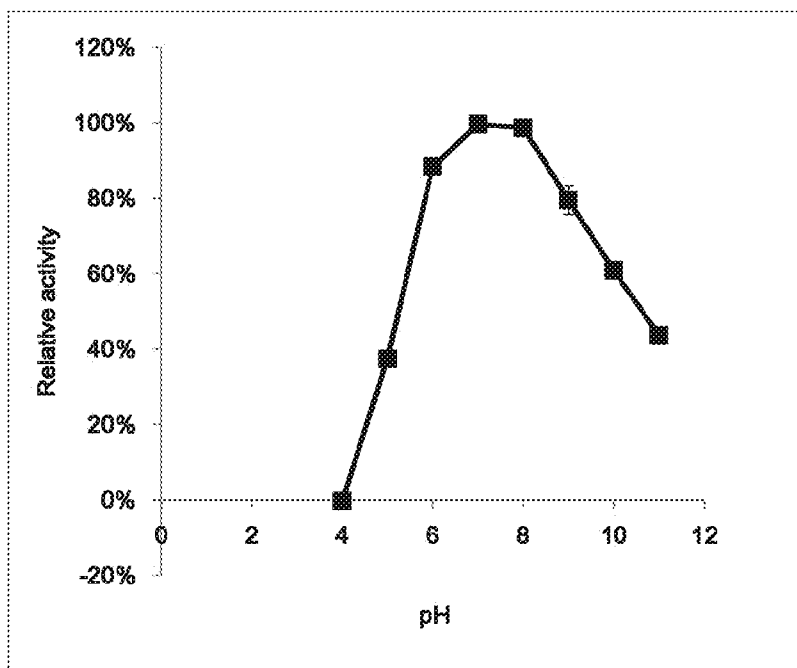


Figure 4.3. pH profile of PehPro1.

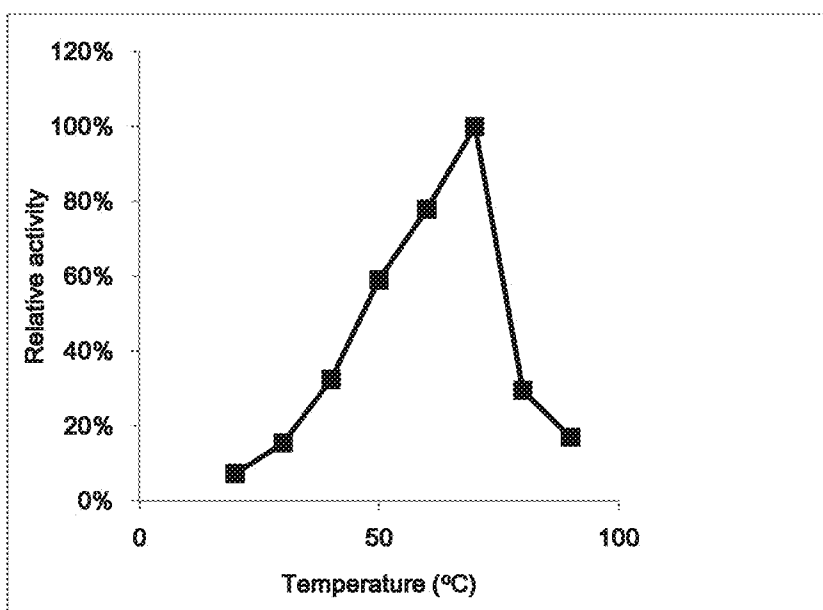


Figure 4.4. Temperature profile of PehPro1.

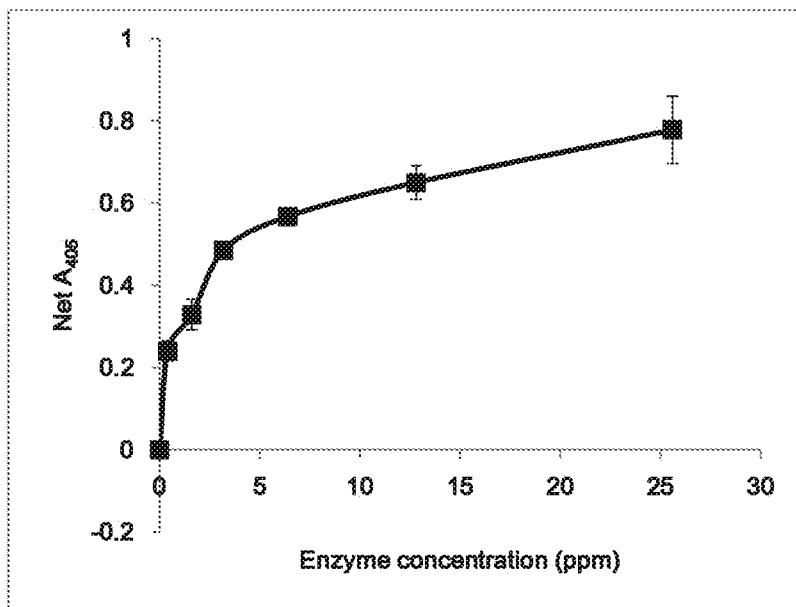


Figure 4.5A: Cleaning performance of PehPro1 in AT detergent at pH 6.

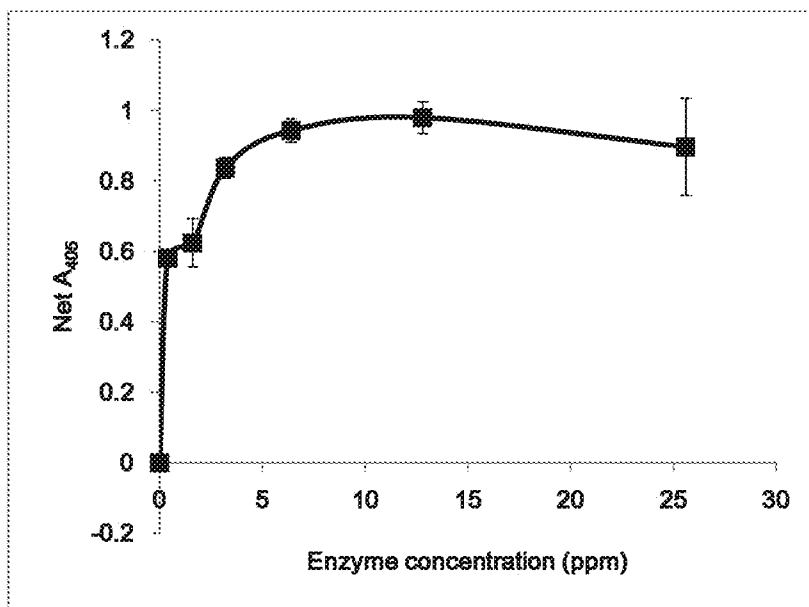


Figure 4.5B: Cleaning performance of PehPro1 in AT detergent at pH 8.

CLUSTAL W (1.83) multiple sequence alignment

```

PehProl_mature          -----ATGTGKGVLDGDKSFTI TQS GS TQQLKDTT RQGGI VI YSAGNP SS
Paenibacillus_elgii_B69_ZP_090  -----ATGTGKGVLDGDKSFTI TQS GS SYQLNDTTRGQGI VI YSAGNR TS
E_thermoproteolyticus_P00800  ITGTI SVVGVGPGVLDGDKMINTI TYS -I XXY LQDNTRNGI FT YDAKRY TT
      !..*!***** *..!* * : * *!*..***!..*..* * !!

PehProl_mature          LPGLLLTSSSNIWN----DGAAVDAHAY TAKVYDYYKNKFGKNSIDGNGPQ
Paenibacillus_elgii_B69_ZP_090  LPGLLLTSTINIW----DSSAVDAHAY TGRVYDYYKNKFGKNSIDGNGLQ
E_thermoproteolyticus_P00800  LPGLLWADADNQFFASYDAPAVDAHYYAGVYDYYKNVHNRLSYDGNNA
***:*:!!:* : *..***** *!..***** !* * **

PehProl_mature          LKSTVHYSSRYNNAFWNGVQVYGDGDGVTFI PFSADPDVIGHELTHGVT
Paenibacillus_elgii_B69_ZP_090  LKSTVHYSTRYNNAFWNGVQVYGDGDGVTFRSPAPADPDVIGHELTHGVT
E_thermoproteolyticus_P00800  IRRSVHYSDGYNNAFWNGSQVYGDGDGVTFIRLSGGIDVVAHELTHAVT
:!*:**** * * * * * * * * * * * * * * * * * * * * * * * * * *

PehProl_mature          EHTAGLEYYGESGALNEISDIIGNATD-----GKNWLEGDLIYTPNTPG
Paenibacillus_elgii_B69_ZP_090  ESTAGLEYYGESGALNEISDIIGNAIE----GKNWLEGDLI---TLNA
E_thermoproteolyticus_P00800  DYTAGLIYQNESGALNEAISDI PGTIVREYANKNPDWREIGEDVYTPCI SG
: **** * ..*****!*:******:!. : * * * :

PehProl_mature          DALRSMENPKLYNQDRYQDFYTCP SDNGGVHINSGINMKAFYLIAGGST
Paenibacillus_elgii_B69_ZP_090  DALRSMENPKLYRQDPDRYQDFYTCP SDNGGVHTNSGINNKAFHLIAGGST
E_thermoproteolyticus_P00800  DGLRSMSEDPANYGDPDHYSKRYTGTQDNGGVHINSGINKAAVLIAGGST
: *****: * : **:*..*****!****** **** * * * :***

PehProl_mature          HYGVTVNGIGPDAAVQI FYCALINYLTPTS NF SAMPAAAIQAATDLYGAN
Paenibacillus_elgii_B69_ZP_090  HYGVTVNGIGRSAAEQI FYDALTHYLTPTS NF SAI RAAAIQAATD SFGAN
E_thermoproteolyticus_P00800  HYGVSVVGI GHDKLGKI FYRALTQYLTPTS NF SQLRAAAVQSATDLYGST
***:* **** * :*** ** :***** :****:*:*** :!..

PehProl_mature          SSQVNVKKAYTAVGVN SEQ ID NO: 16
Paenibacillus_elgii_B69_ZP_090  SSQVDVKKAYNAVGVN SEQ ID NO: 48
E_thermoproteolyticus_P00800  SQEVASVKQAEAVGVK SEQ ID NO: 45
*..* :**:* :****:

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Figure 4.6: Alignment of PehProl protein with homologous protease sequences.



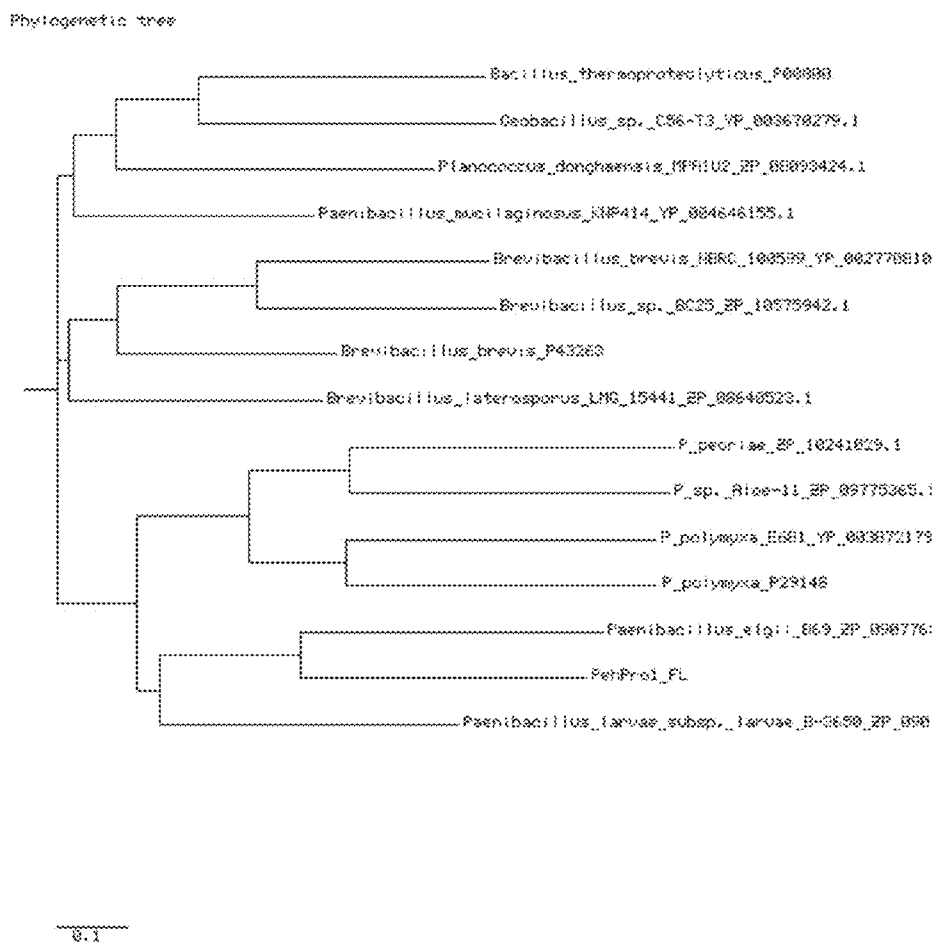


Figure 4.7: Phylogenetic tree for PehPro1 and its homologs.

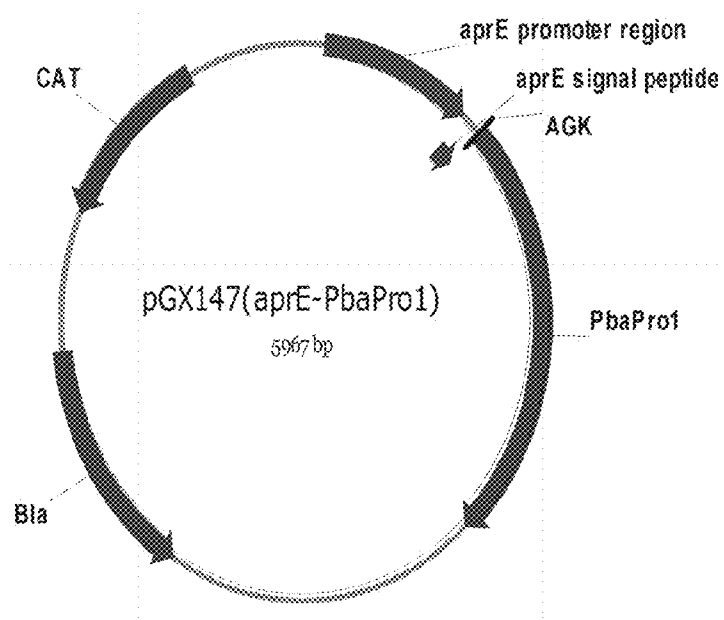


Figure 5.1. The plasmid map of pGX147(AprE-PbaPro1).

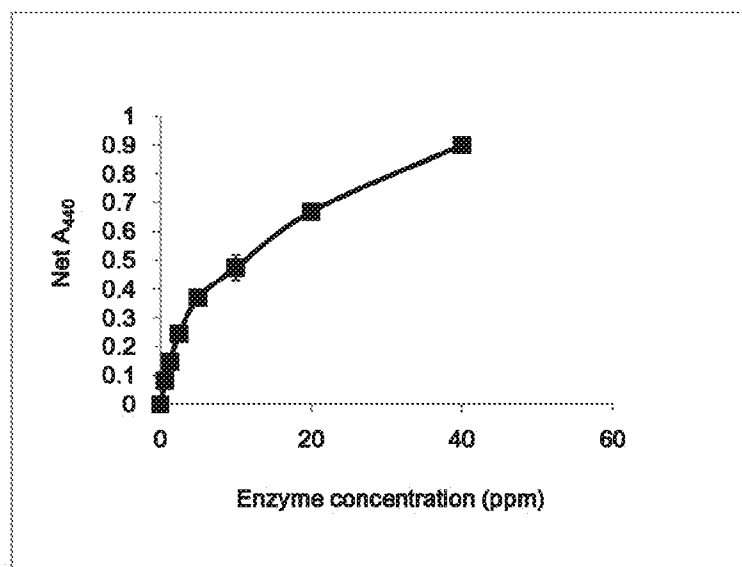


Figure 5.2. Dose response curve of PbaPro1 in azo-casein assay at pH 7.

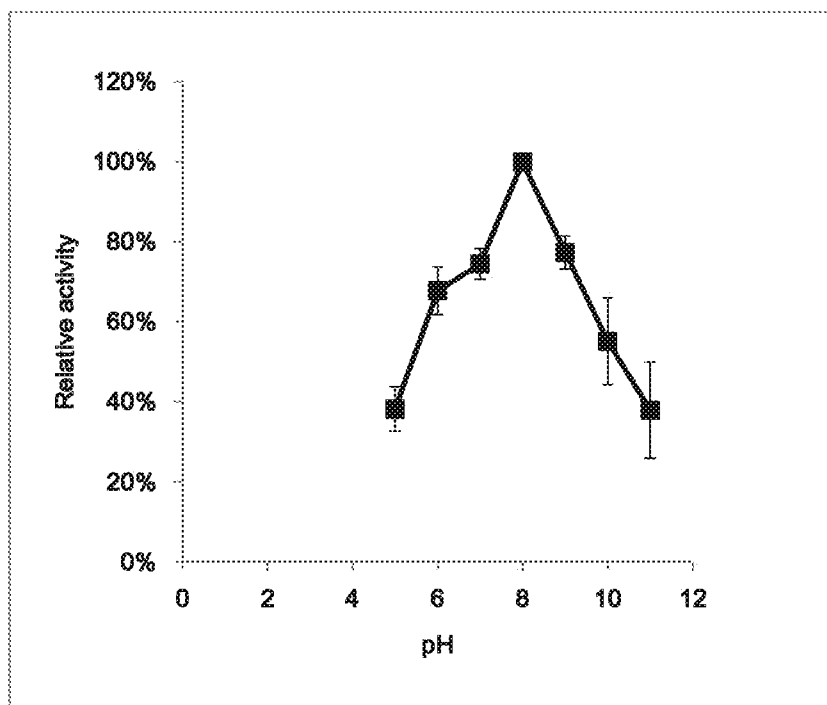


Figure 5.3. pH profile of PbaPro1.

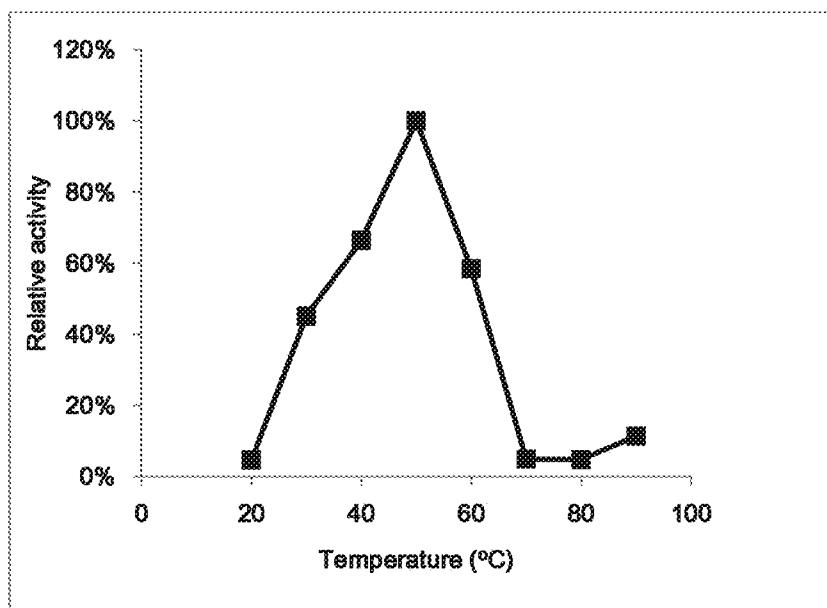


Figure 5.4. Temperature profile of PbaPro1.

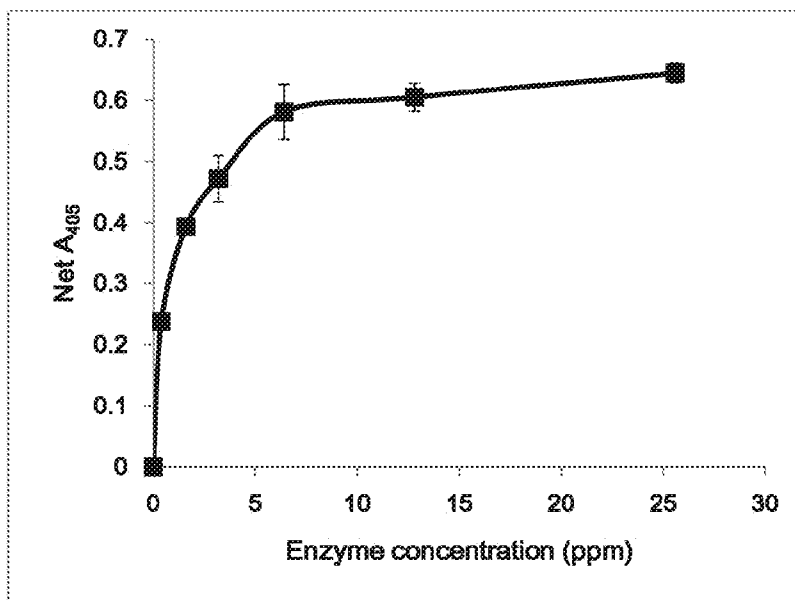


Figure 5.5A: Cleaning performance of PbaPro1 in AT dish detergent at pH 6.

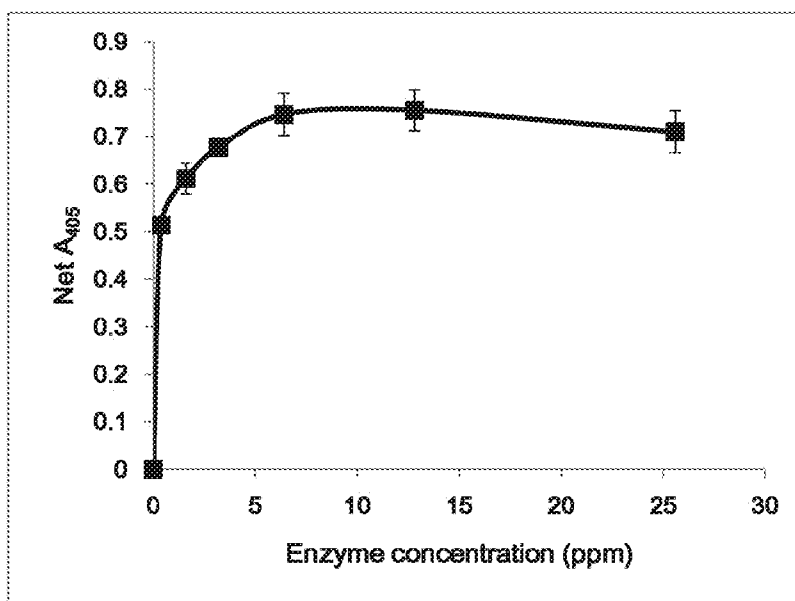


Figure 5.5B: Cleaning performance of PbaPro1 in AT dish detergent at pH 8.

CLUSTAL W (1.83) multiple sequence alignment

```

PbaProl          -----ATGTGTGVHGDTKLLITQSGSTYQLKDTTRGRNGIQTYIAMNRSS
P_polymyxa_SC2   -----NEATGTGKGVLDGSKSFTTASGSSYQLKDTTRGNGIVTYTASNRGS
B_thermoproteolyticus_P00800  ITGTSTVGVGRGVLGDQKNINITYS-ITYYLLQDNRGNGIETTDKAYRIT
                :.* ** ** * :.*** * : * *;*.***;** ** * . * :

PbaProl          LPGSLSTSSNNVWT----DRAAVDAHAYAAATYDFYKMKFNRNGIDGNGLL
P_polymyxa_SC2   LPGTLITLDADNVWN--DPAGVDAHAYAAKTYDYKAKFGRNSIDGRGLQ
B_thermoproteolyticus_P00800  LPGLWADAQNGQFFASYGAPAVDAHYAGVTYDYKKNVNRSLSYDGNNA
                :***: :.:** : * .,***** **; **;* ** . **..

PbaProl          IRSTVHYGSSNYKNAFWNGAQIVYGGDGIIEFGPFSGDLDDVVGHELTGVI
P_polymyxa_SC2   LRSTVHYGSRYNNAFWNGSQMTYGDGGSTFIAFSGDPDVVGHLELTGVT
B_thermoproteolyticus_P00800  IRSSVHYGQGYNNAFWNGSQMIVYGDGGQTFIPLSGGIDVVABELTHAVT
                :**;***. *;*****;.;***** * .;*. **;*****.*

PbaProl          EYTNLEYYRNEFGALNEAFADIMGNTIE-----SKWLLSDGIYTPNIHG
P_polymyxa_SC2   EYTSNLEYYCESCALNEAFSDVIGNDIQ-----RKNWLVGDDIYTPNIAG
B_thermoproteolyticus_P00800  DYTAGLIYQNESGAINELSDIFGTLVEFYANKNPOWEIGEDVYTPGLSG
                :**;. * * .;***;***;:**;*. : : : * :.***.***.

PbaProl          DALRSLSDPTLYNOPDKYSDRYTGSQDNGGVHNSGIINKAYYLLAQQGST
P_polymyxa_SC2   DALRSMNPTLYDQPDHYSNLYRGSQDNGGVHNSGIINKAYYLLAQQGM
B_thermoproteolyticus_P00800  DSLKSMSPAKYQDPDHYSKRYTGTQDNGGVHNSGIINKAAYLISQGGT
                *;***;*. *; .;***;**. * *;.***** ***** ** ;***.

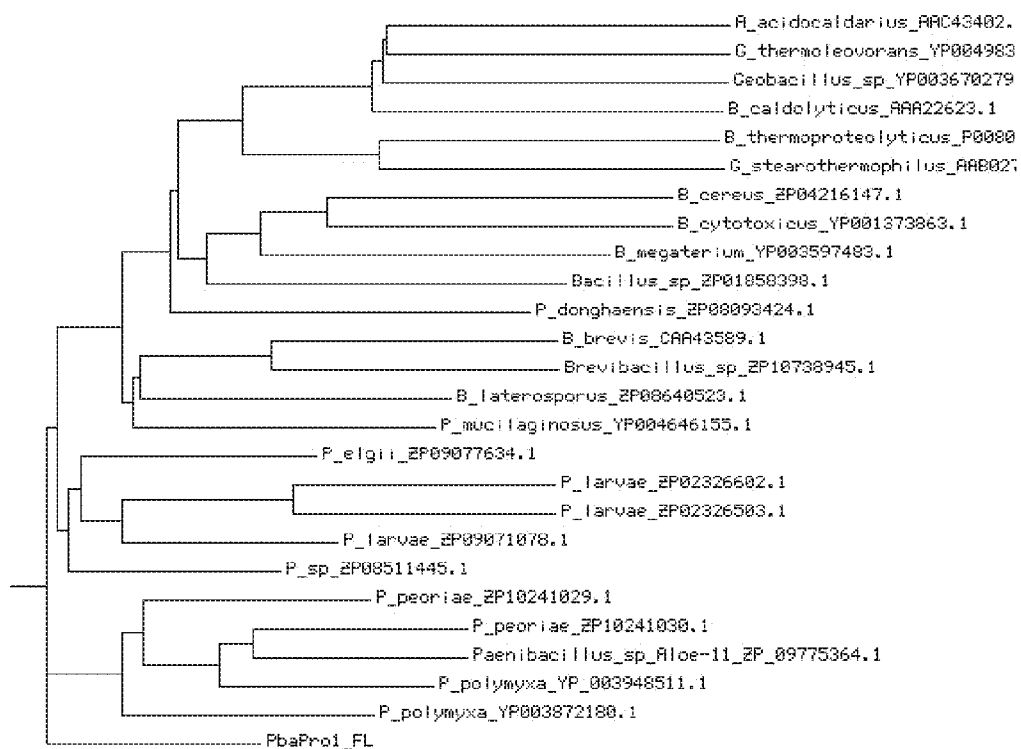
PbaProl          HNGVTVSGTGRDKAVRTFYSTLVNLYLPTSKFAAAKTATIQAAKDLYGAN
P_polymyxa_SC2   FHGVTVNGIGRDAVQIYYSATNYLTSSSGFNRRAAVIQARDLYGAN
B_thermoproteolyticus_P00800  HYGVSVVGIGRDLGKIFRYALTQYLPTGNFGQLRAAAVQSATDLYGST
                . **;* ***** :;* :.***;*. * : :.***.***.

PbaProl          SAEATAITKAYQAVGL-- SEQ ID NO: 23
P_polymyxa_SC2   SAEATAAARKSPDAVGVN SEQ ID NO: 49
B_thermoproteolyticus_P00800  SQEVASVRQAFDAVGVK SEQ ID NO: 45
                * *;. :.***;

```

Figure 5.6: Alignment of PbaProl protein with homologous protease sequences.

Phylogenetic tree



0.1

Figure 5.7: Phylogenetic tree for PbaPro1 and homologs.

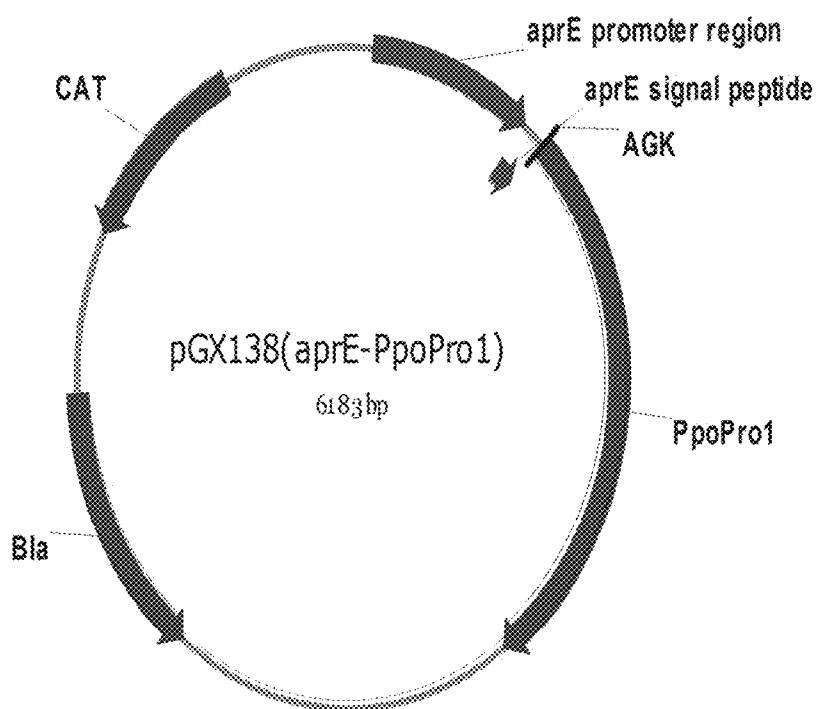


Figure 6.1. The plasmid map of pGX138 (AprE-PpoPro1).

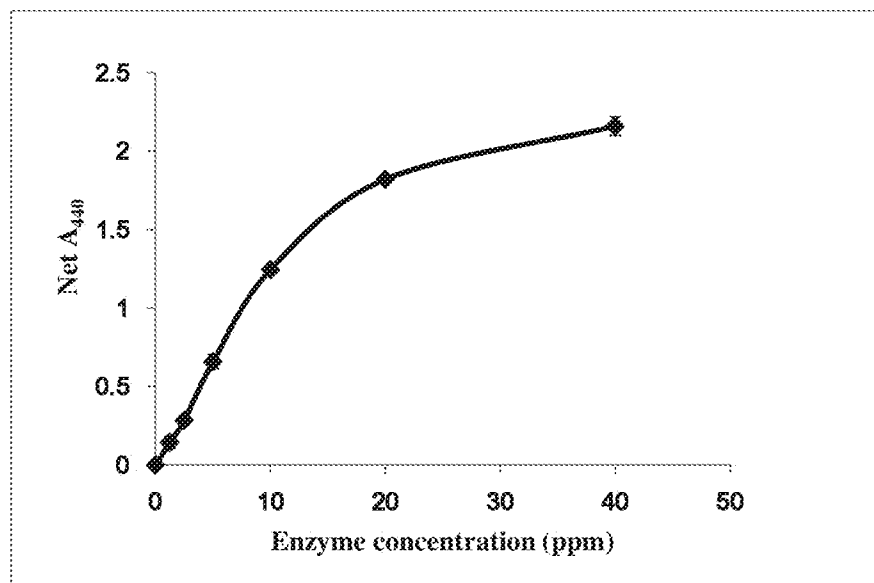


Figure 6.2. Dose response of PpoPro1 in azo-casein assay at pH 7.

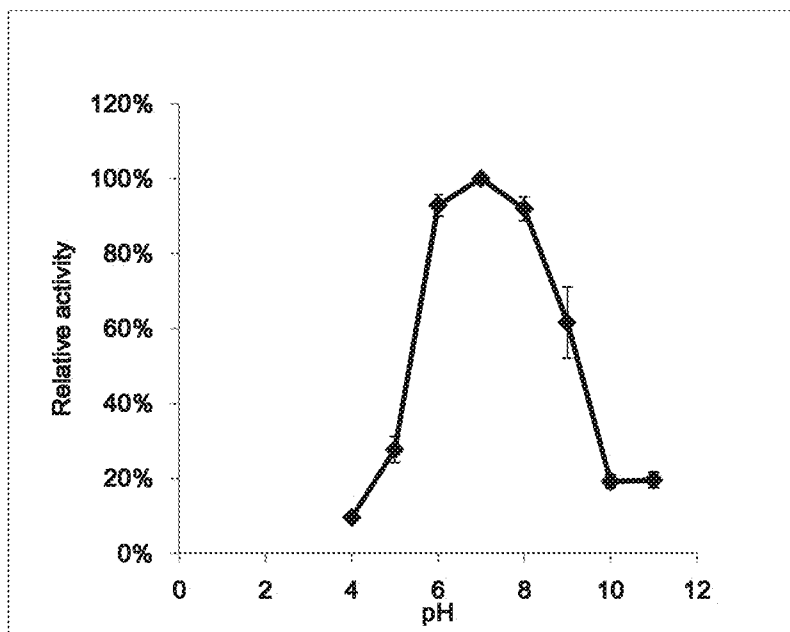


Figure 6.3. pH profile of purified PpoProI.

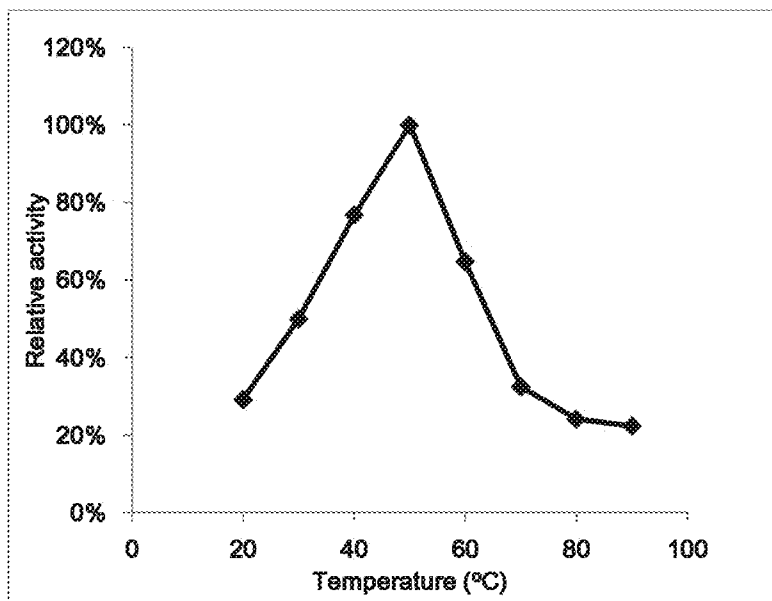


Figure 6.4. Temperature profile of purified PpoProI.



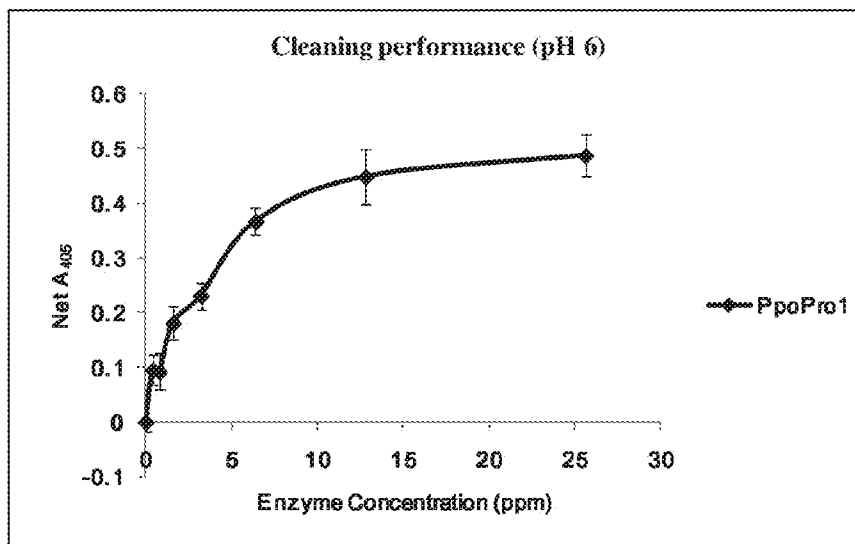


Figure 6.5A: Cleaning performance of PpoPro1 at pH 6 in AT detergent with PAP.

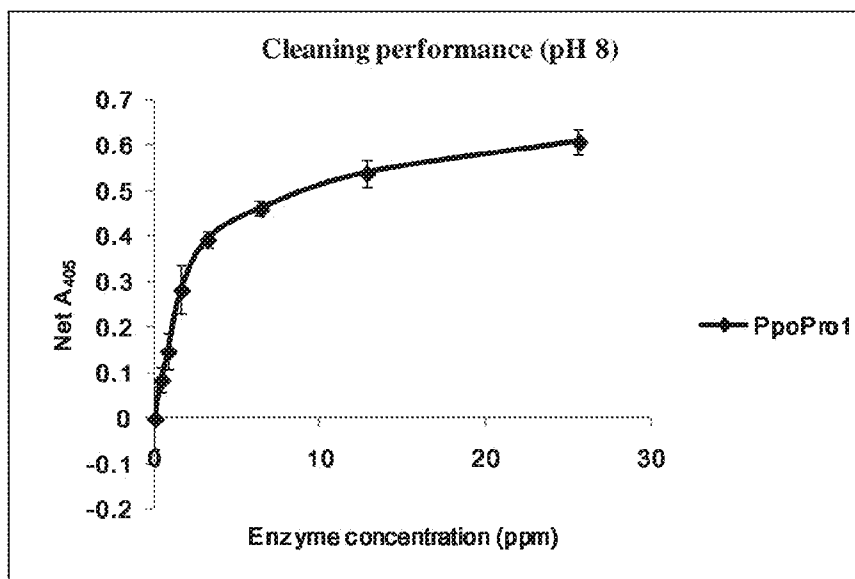


Figure 6.5B: Cleaning performance of PpoPro1 at pH 8 in AT detergent with PAP.

```

CLUSTAL W (1.83) multiple sequence alignment

EpoProl      -----ATGTGKGVLSGDSKSF TTTASGSSYQLKDDTTRNGIIVYTTASNRQS
E__polymyxa__SC2__YP__003948511.1  ---NEATGTGKGVLSGDSKSF TTTASGSSYQLKDDTTRNGIIVYTTASNRQS
E__thermoproteolyticus__P00800    ITGTSTVGVGRGVLSGDKNINITYS--TYYYLQDNTRNGIIFTYDAKYRIT
                                   :.*.*:*****.*.:** * : * *:*.******.*** * * :

EpoProl      IFGTILTDADNVWN---DFAGVDAHAYAARTDYKAKKFRNSIDGRGLQ
E__polymyxa__SC2__YP__003948511.1  IPCTILTDADNVWN---DPAGVDAHAYAARTDYKAKKFRNSIDGRGLQ
E__thermoproteolyticus__P00800    LPSLWADADNQFFASYDAPAVDAHYYAGVTYDYKKNVHNRLSYDGNNA
                                   :*:* : :***** : *...***** **.****** ..* * **..

EpoProl      LRSTVHYGSRYNNAFWNGSQMTYGDGDSGTFIAFSGDPDVPVGHETHGVT
E__polymyxa__SC2__YP__003948511.1  LRSTVHYGSRYNNAFWNGSQMTYGDGDSGTFIAFSGDPDVPVGHETHGVT
E__thermoproteolyticus__P00800    IRSVHYGSGYNNAFWNGSQMVYGDGDSGTFIFLPSGGIDVVAHELTHAVT
                                   :*:*:*.. *****.*****.**.:**.* **.******.***

EpoProl      EYTSNLEYEYGESGALNEAFSDVIGNDIQ----RKNWLVGDDIYTPNIAG
E__polymyxa__SC2__YP__003948511.1  EYTSNLEYEYGESGALNEAFSDVIGNDIQ----RKNWLVGDDIYTPNIAG
E__thermoproteolyticus__P00800    DYTASLEIYQNESGALNEAIGDIEFGTLVEFYANKNPDWEIGEDVYTPGISG
                                   :*:*.* * .*****:*:*:*:*.:* : : . : * :*:*:*:*:*:*

EpoProl      DALRSMNSPTLYDQPDHYSNLYRGS SDNGGVHTNSGI INKAYYLLAQGGN
E__polymyxa__SC2__YP__003948511.1  DALRSMNSPTLYDQPDHYSNLYRGS SDNGGVHTNSGI INKAYYLLAQGGN
E__thermoproteolyticus__P00800    DSLRSMNSDPARYGDPDHYSKRYTGTQDNGGVHINSGI INKAAAYLISQSGT
                                   *:******:* :*:******: * *:*.****** ***** **:***.

EpoProl      FHGVTVNGIGRDAAVQIYSAFTNYLTSSSDFSNARA AVIQA AKDLYGAN
E__polymyxa__SC2__YP__003948511.1  FHGVTVNGIGRDAAVQIYSAFTNYLTSSSDFSNARA AVIQA AKDLYGAN
E__thermoproteolyticus__P00800    HYGVSVVVIGIKDKLGI FYRALTQYLIFTSNFSQLRAAAVQSATDLYGST
                                   :.*:* * ***** :*:* *:*:*:*:*:*:* *:*:*:*:*.******.:

EpoProl      SAEATAA AKSFD AVGVN SEQ ID NO: 28
E__polymyxa__SC2__YP__003948511.1  SAEATAA AKSFD AVGVN SEQ ID NO: 50
E__thermoproteolyticus__P00800    SQEVASVRQA FDAVGVK SEQ ID NO: 45
                                   * *.:.. :*****:

```

Figure 6.6. Alignment of PpoProl with protease homologs.

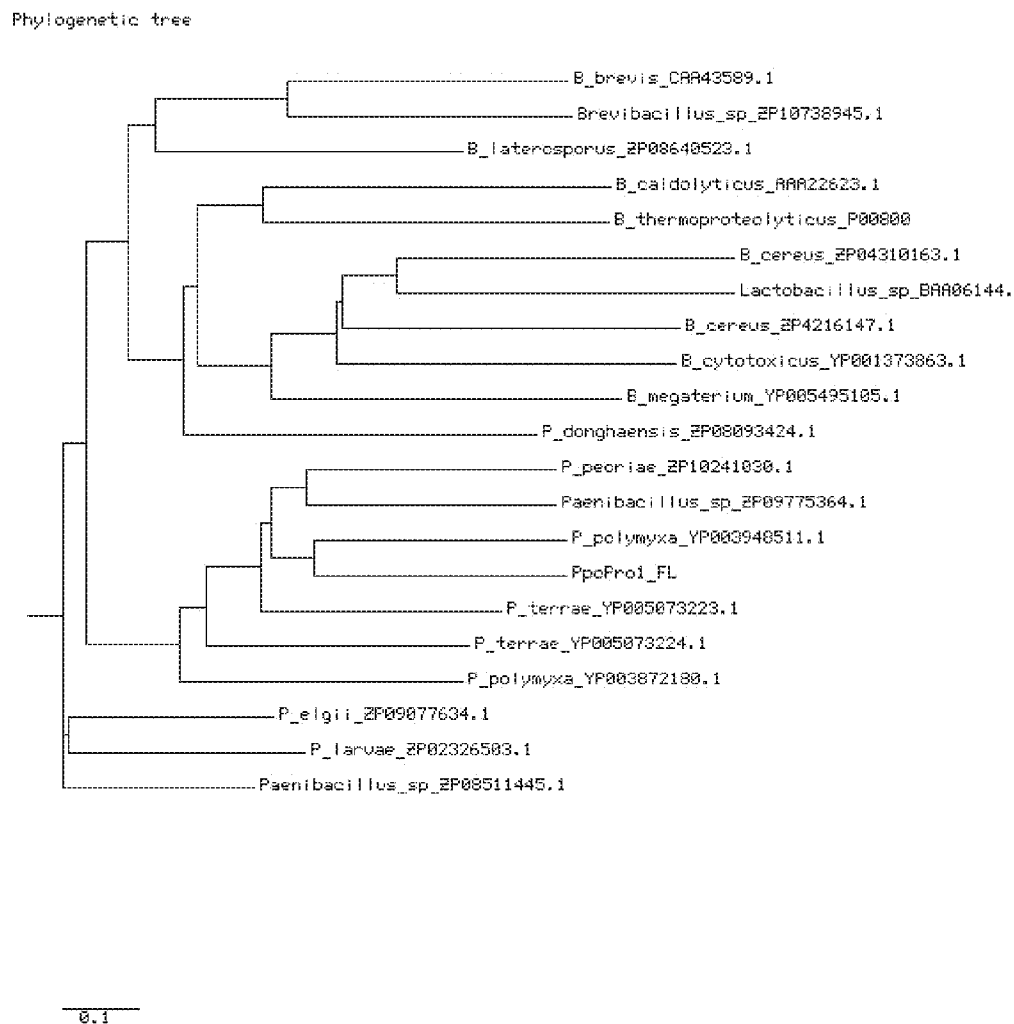


Figure 6.7. Phylogenetic tree of PpoPro1 and homologs.

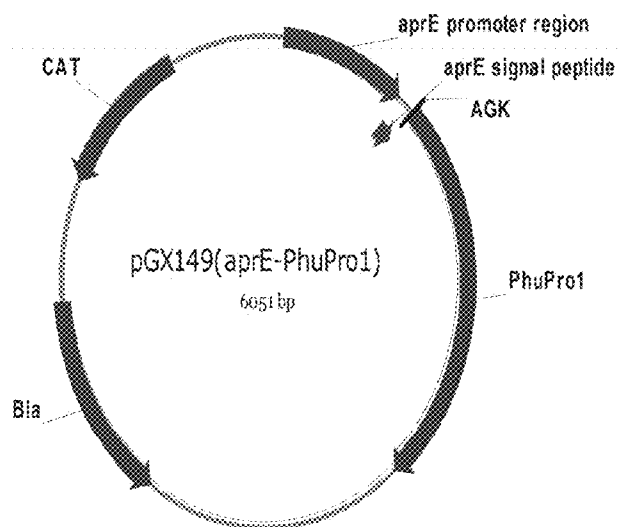


Figure 7.1. The plasmid map of pGX149(AprE-PhuPro1).

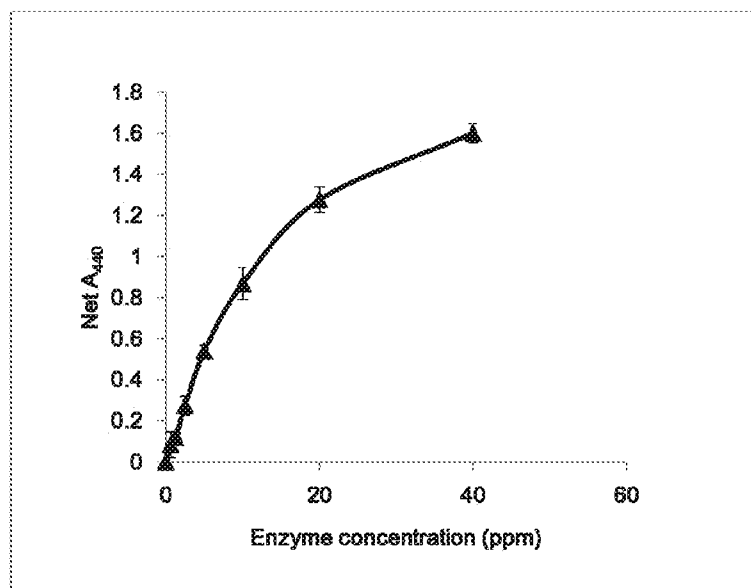


Figure 7.2. Dose response curve of PhuPro1 in azo-casein assay at pH 7.

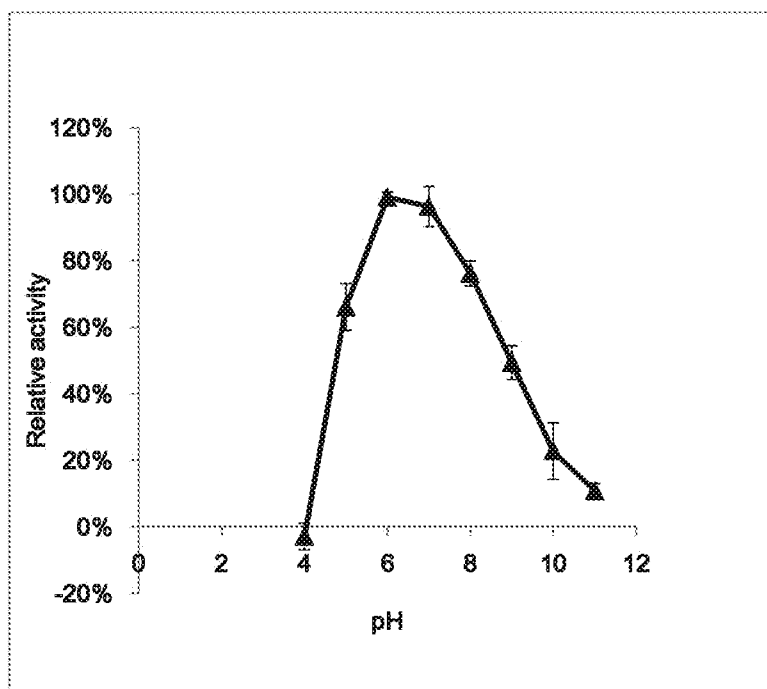


Figure 7.3. pH profile of PhuPro1.

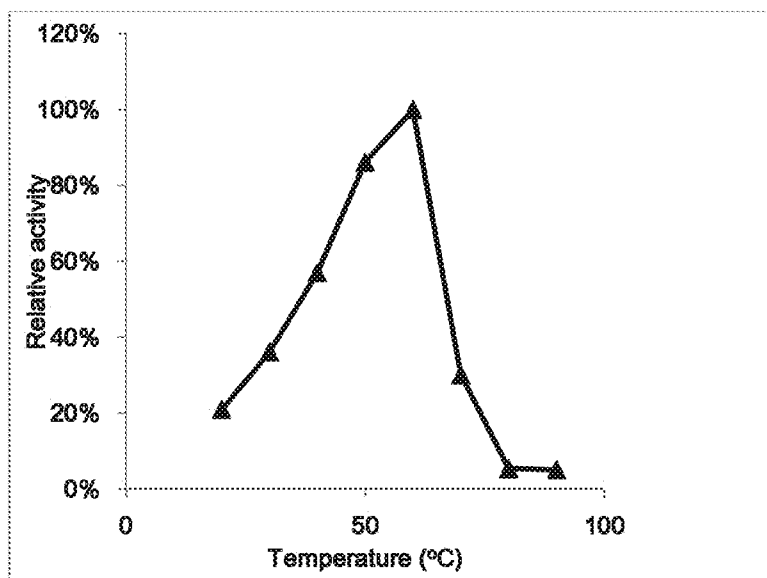


Figure 7.4. Temperature profile of PhuPro1.

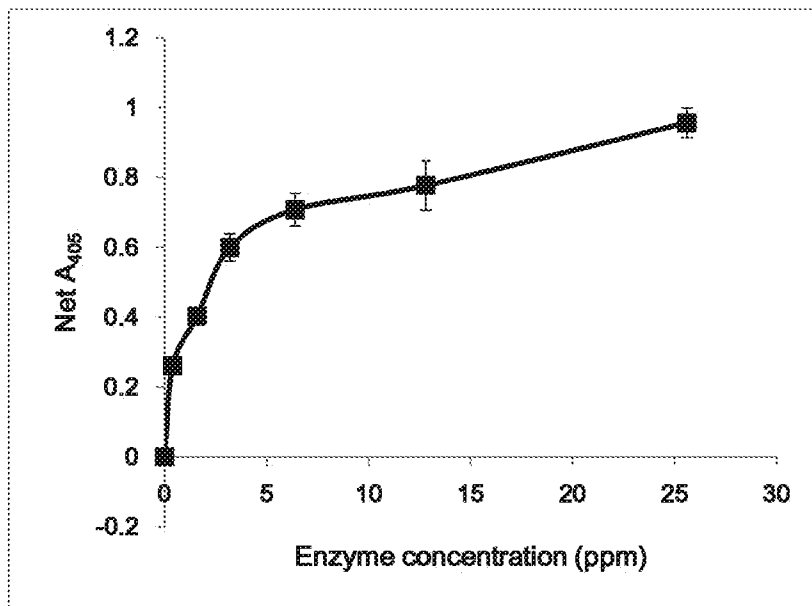


Figure 7.5A. Cleaning performance of PhuPro1 in dish detergent at pH 6.

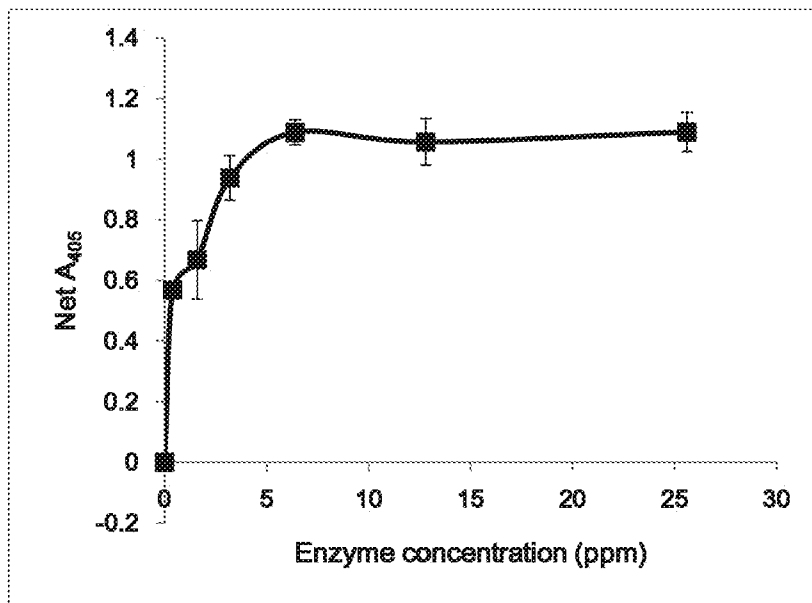


Figure 7.5B. Cleaning performance of PhuPro1 in dish detergent at pH 8.

CIGSTAT W (1.83) multiple sequence alignment

```

PhuFrol          -----ATGTGKGVLDGDKSPFVGTSGSSYVMI DSTRGKGIQTITASNRFS
P_terrae_HPL-003_YP_005073223. -----ATGTGKGVLDGDKSPFVGTSGSSYVMI DSTRGKGIQTITASNRFS
B_thermoproteolyticus_P00800 ITGTSTVGVGRGVLGDQHNINITYS-TYYYLQDNTRGNGIPTYDAKYRIT
      :.*:*:***** *.:. . * : * : *:*:*:* * * * . * :

PhuFrol          LPGSEVTSSESTFN-----DPASVDAHAYAQKVYDFYKSNFNRRNSIDGNGLA
P_terrae_HPL-003_YP_005073223. LPGSEVTSSESTFN-----DPAGVDAHAYAAKTYDYYKDFPGRNSIDGRGLQ
B_thermoproteolyticus_P00800 LPGSLMADADNQFFASVDAPAVDAAHYACVITYDYYKNVNRSLSYDGNWAA
      :** : : : : : * : : * * : * : * : * : * : * :

PhuFrol          IRSTTHYSTRYNNAFWNGSQMVYGDGDGSOFIAPSGDLVVGHETHGVT
P_terrae_HPL-003_YP_005073223. LRSTVHYGSKYNNAFWNGSQMTYGDGDGTFIAPSGDPDVVGHETHGVT
B_thermoproteolyticus_P00800 IRSSVHYSGYNNAFWNGSQMVYGDGDGQTFIPLSGGLDVVAHELTHAVT
      :**:* * : * : * : * : * : * : * : * : * : * : * : * :

PhuFrol          EYTSNLEYYGESGALNESISDIIGNDIQ-----RKNWLVGDDIYTPSGS
P_terrae_HPL-003_YP_005073223. EYTSNLEYYGESGALNESISDIIGNDIQ-----RKNWLVGDDIYTPSGS
B_thermoproteolyticus_P00800 DYTAGLIYQNESGALNEATSDIFGTLVFEFYANKNFDWEIGEDVYTPGISG
      :**:* * : * : * : * : * : * : * : * : * : * : * : * :

PhuFrol          QALRYNDDEPNEGQPPARMADEVNTEADNGGVHTNSGIPNNAYYLLAQGGT
P_terrae_HPL-003_YP_005073223. DALRSMSENPILYDQPDHYSNLYKGSNDGGVHTNSGIPNNAYYLLAQGGT
B_thermoproteolyticus_P00800 ESLRSMSEDPAKYGDPDHYSKRYTGTQDNGGVHTNSGIPNNAYYLLAQGGT
      :** * : * : * : * : * : * : * : * : * : * : * : * :

PhuFrol          FGGVWVTGIGHSQAIQIVYPALTYLLTSSNFSNYRSAMVQASTDLKGAN
P_terrae_HPL-003_YP_005073223. FHNVEVSGIGRDAAVQIYYSAFTNYLTSSNFSNTRAAVVQAAKDLKGAN
B_thermoproteolyticus_P00800 HYGVSVVGIQRDLKGI FYRALIQYLTPTSNFSQLRAAVQASATDLYGST
      :.*:* * * * * : * * * : * * * : * * * : * * * : * * * :

PhuFrol          STQTAVKNSLSAVGIN SEQ ID NO: 33
P_terrae_HPL-003_YP_005073223. SAQATAAAKSFDAVGNN SEQ ID NO: 51
B_thermoproteolyticus_P00800 SQEVASVKQAFDAVGVK SEQ ID NO: 45
      * : : : : : * : : : : * : : : : * : : : :
    
```

Figure 7.6: Alignment of PhuFrol with homologous protease sequences.

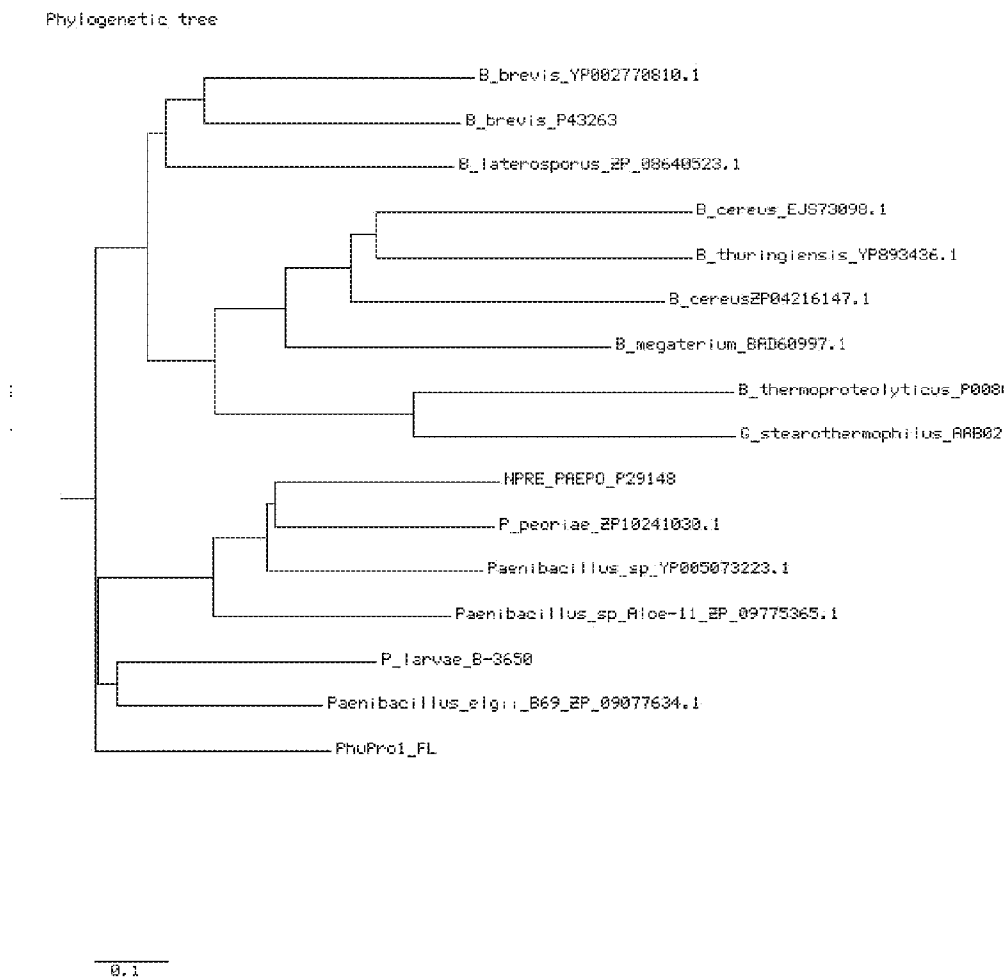


Figure 7.7: Phylogenetic tree for PhuPro1 and homologs.



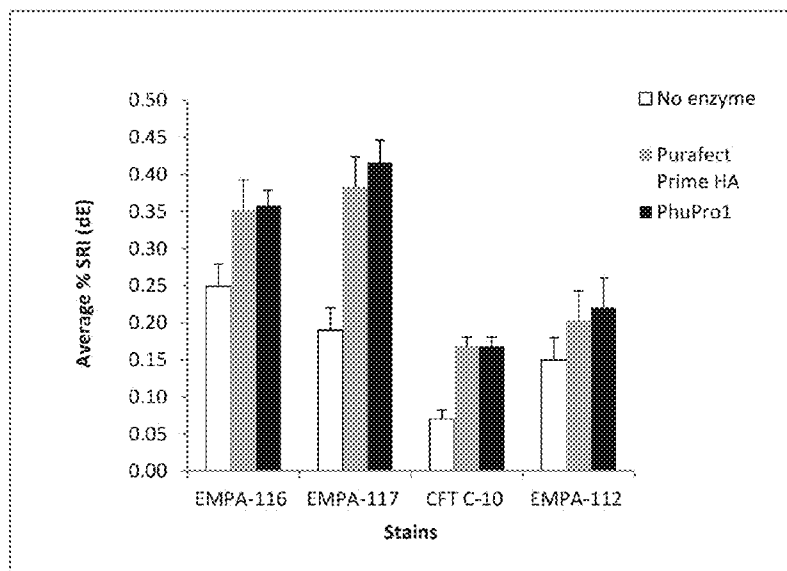


Figure 7.8A: Cleaning performance in Terg-o-Tometer assay at 32°C, on four technical stains.

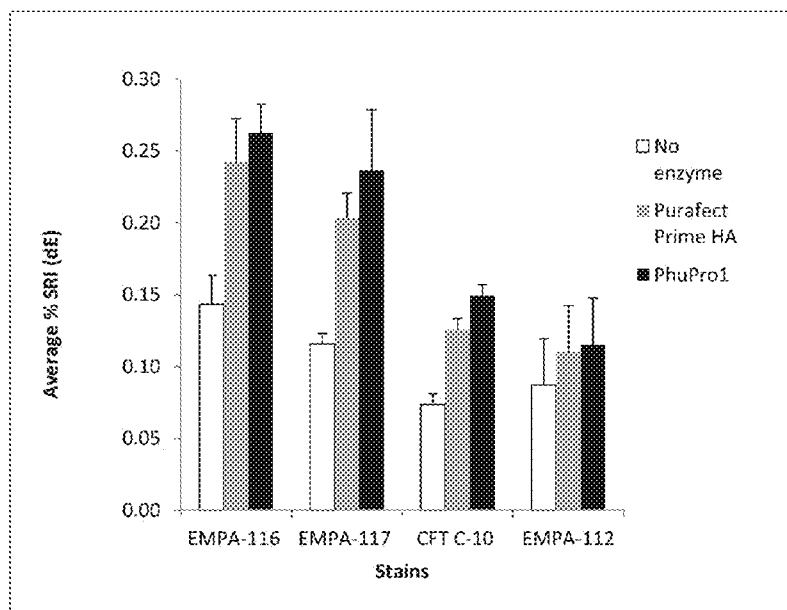


Figure 7.8B: Cleaning performance in Terg-o-Tometer assay at 16°C, on four technical stains.

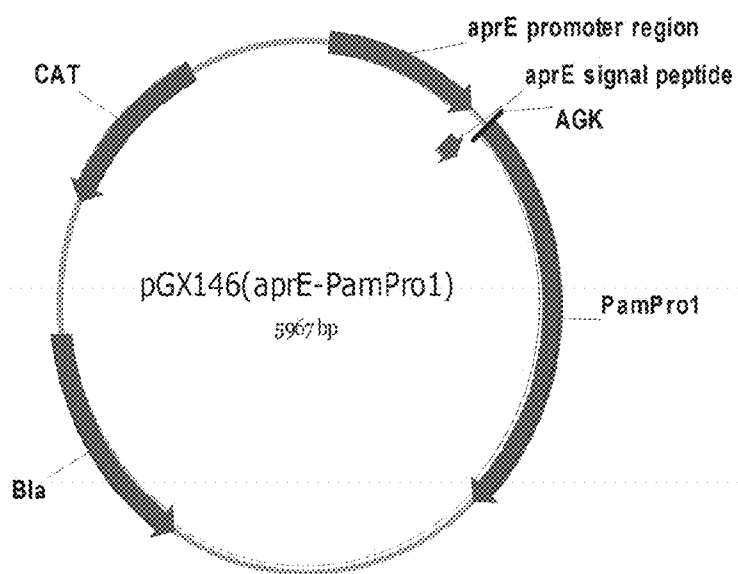


Figure 8.1. The plasmid map of pGX146(AprE-PamPro1).

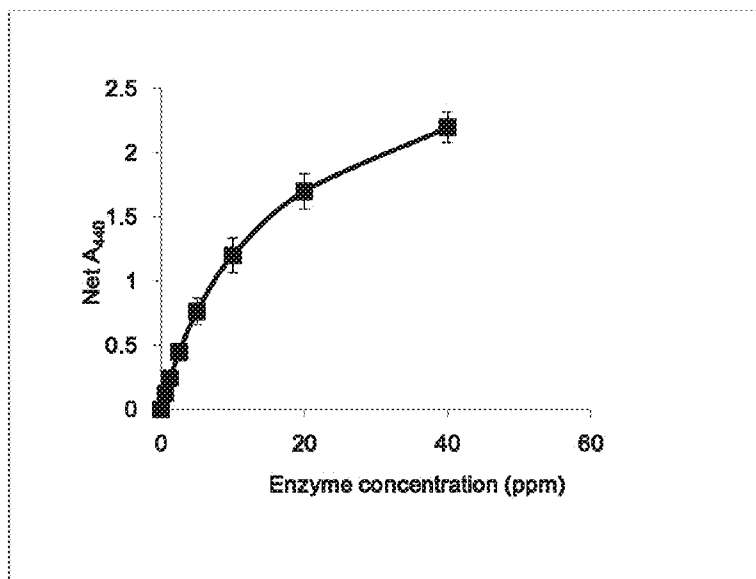


Figure 8.2. Dose response curve of PamPro1 the azo-casein assay at pH 7.

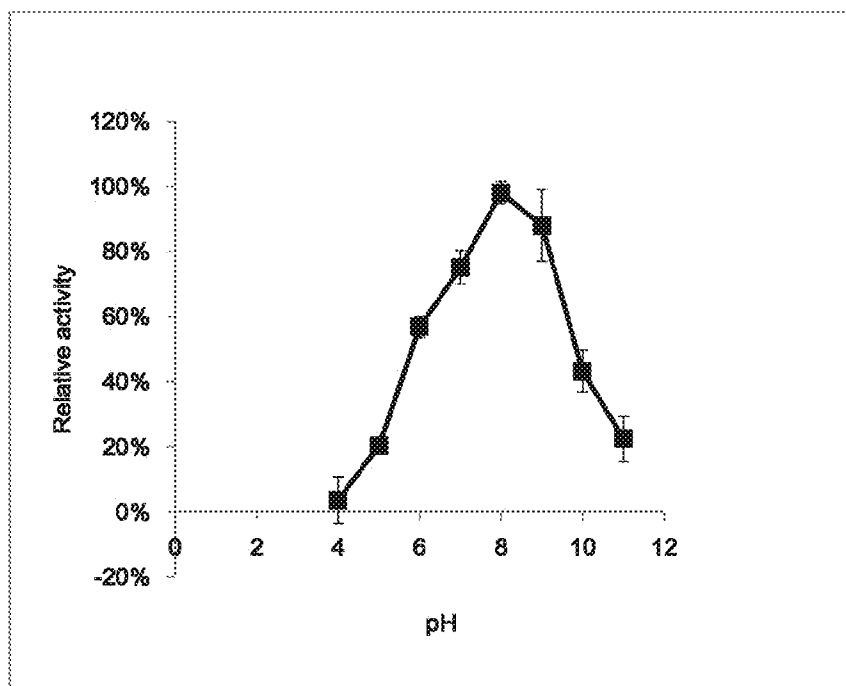


Figure 8.3. pH profile of PamPro1.

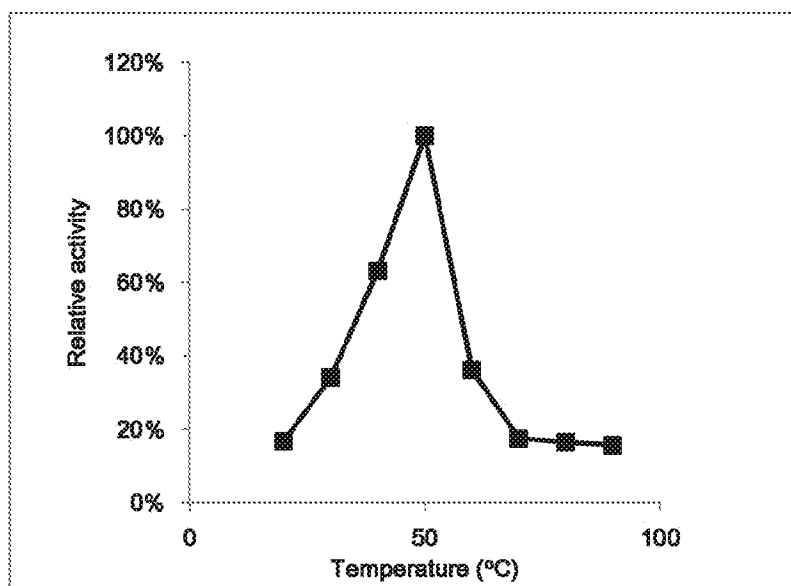


Figure 8.4. Temperature profile of PamPro1.

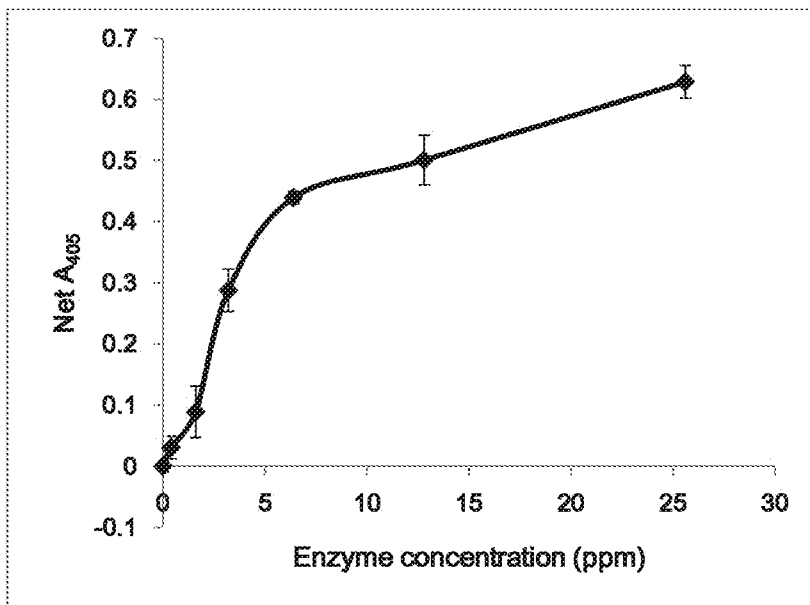


Figure 8.5A: Cleaning performance of PamPro1 in AT dish detergent at pH 6.

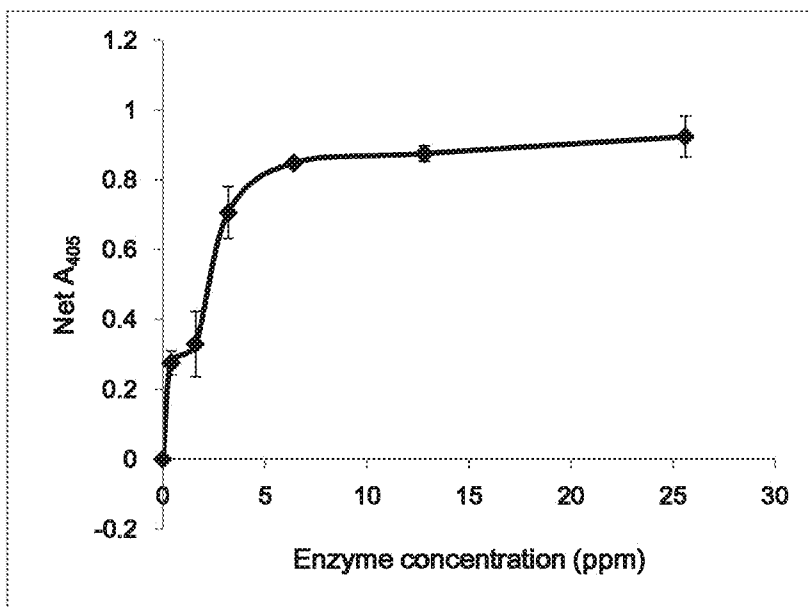


Figure 8.5B: Cleaning performance of PamPro1 in AT dish detergent at pH 8.

CLUSTAL W (1.83) multiple sequence alignment

```

PamPro1          -----ATGTGTGVLGDTKTLITTTQSGSTPQLRDTTRCNGIQTYTAMNGSS
P_peoriae_KCTC  DIINEAFGTGKGVLDTKSFTTASGSSYQLRDTTRCNGIVTYTASNROE
E_thermoproteolyticus_P00800  ITGISTVGVGRGVLGDQKKNINTYS-TYYLQDNTRCNGIFTDAKYRRT
      :.* ***** *.:** * : : *:****** ** * . :

PamPro1          LPGSLLTDSQNVVT---DRAGVDAHAAHAATYDFYKKNFNRCNGINGNGLL
P_peoriae_KCTC  IEGTLLTGADNVWN----DPAGVDAAHAYAAKTYDYKKEKFNNSIDGRGLQ
E_thermoproteolyticus_P00800  LPGSLWADADNQFEASVDAPAVDAHYAGVTYDYKKNVHNRLSYDGNNAE
      :**.: :*:** : * ..**** :*, **:***.: ** . :*..

PamPro1          IRSTVHYGSNYNNAFWNGAQIVFGGSDGTMFRSLSGGLEVVGHETHGVY
P_peoriae_KCTC  LRSVHYGNRYNNAFWNGSQMTYGGDGGTTFIAESGDFDVVGHETHGVY
E_thermoproteolyticus_P00800  IRSVHYGQYNNAFWNGSQMVMYGGDGGQTFIEPLSGGLEVVVAHELTHAVT
      :**:***.. *****:*.:***** * :**.***.***.*

PamPro1          EYTANLEYRNEPGALNEAFADIFGNTIQ-----SRNWLLCDDIYTPNTPG
P_peoriae_KCTC  EYTSNLEYYSSESGALNESESDIIGNCIQ-----RKNWLVGGDIYTPKIAG
E_thermoproteolyticus_P00800  DYTAGLIYQNESGAINAISEDFGTLVEFYANKNPDRBIEGEDVYTPGISS
      :**:* * .*.**:***.:**:* . : : * :*:**.* **

PamPro1          DALRSLSNPTLYGQFDNYSDRYTGSDQDNGGVHNSGIINKAYFLAQGGT
P_peoriae_KCTC  DALRSMNSNPTLYDQPDHYSNLYRGSQDNGGVHNSGIINKAYYLLAQGGT
E_thermoproteolyticus_P00800  DSLRSMSEFAKYGDFDHYSKRYTGTQDNGGVHNSGIINKAAYLISQGGT
      :**:***:* *.:**:***.* * :***** ***** * :****

PamPro1          HNGVIVTGIQRDKAIQIFYSTLVNILEPTSKFAAAKTATIQAAARDLYGAT
P_peoriae_KCTC  FHGVTVNGIGKDAAVQIYSAFTNYLTSSEDFSNAPDAVVQBAKDLYGAS
E_thermoproteolyticus_P00800  HYGVSVVBIGRDKLGGKIFYRALTQYLTPTSNFSQLRAAAVQSAATDLYGST
      . **:* ***** :*:* :.:**:***.* * : : *.:**.***.*:

PamPro1          SAEATAITKAYQAVGL- SEQ ID NO: 38
P_peoriae_KCTC  SAQATAAARAFDAVGVN SEQ ID NO: 52
E_thermoproteolyticus_P00800  SQEVASVKQAFDAVGVK SEQ ID NO: 45
      * :.: :*:**:*
    
```

Figure 8.6: Alignment of PamPro1 with homologous protease sequences

Phylogenetic tree

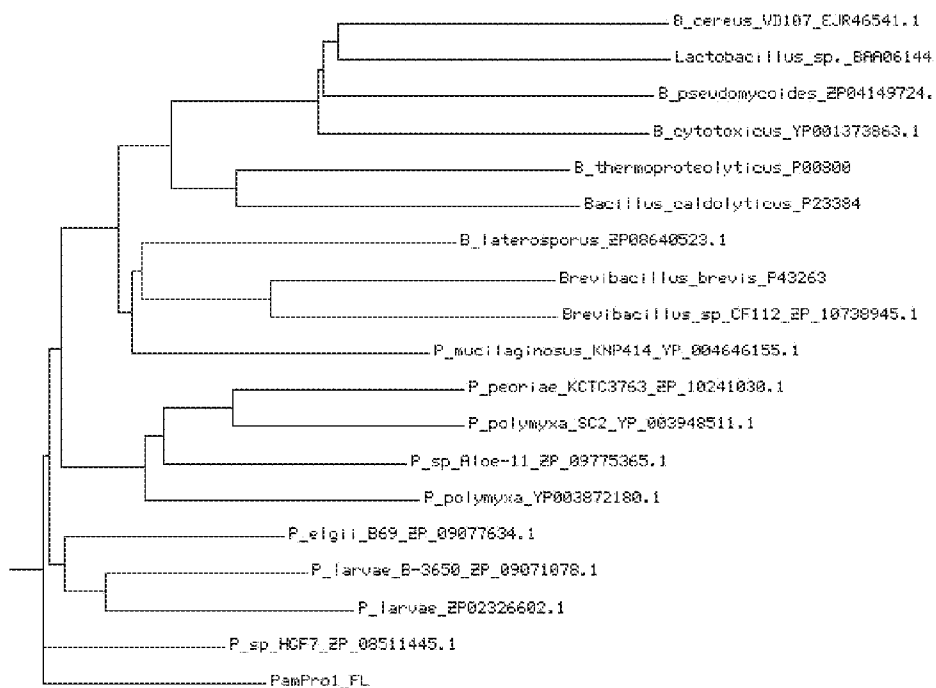


Figure 8.7: Phylogenetic tree for PamPro1 and homologs

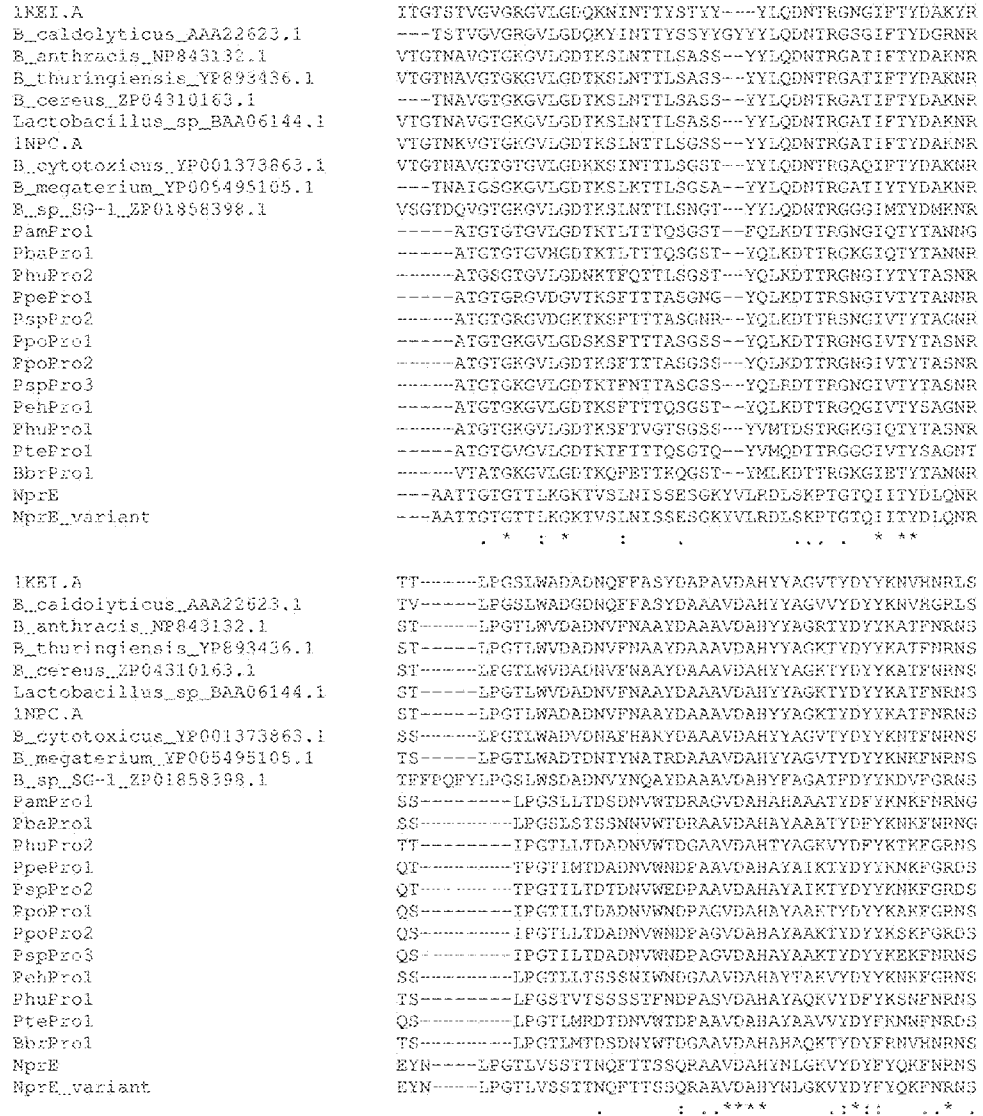


Figure 9.1A CLUSTAL 2.0.10 multiple sequence alignment of the various *Paenibacillus* metalloproteases with other bacterial metalloprotease homologs.

```
1KEI.A            YDGNNAAIRSSVHYSGYNNNAFWNGSQMVYGDGGDTTFIPLSGGIDVVVAH
E_caldolyticus_AAA22623.1   YDGSNAAIRSTVHYGKRYNNAFWNGSQMVYGDGGDTFLFPSSGGIDVVVGH
E_anthraxis_NP843132.1     INDAGAPLKRSTVHYGSRYNNAFWNGSQMVYGDGGDTFTSLSGGIDVIGH
B_thuringiensis_NP893436.1  INDAGAPLKRSTVHYGSRYNNAFWNGSQMVYGDGGDTFTSLSGGIDVIGH
E_cereus_ZP04310163.1     INDAGAPLKRSTVHYGSRYNNAFWNGSQMVYGDGGDTFTSLSGGIDVIGH
Lactobacillus_sp_BAA06144.1 INDAGAPLKRSTVHYGSRYNNAFWNGSQMVYGDGGDTFTSLSGGIDVIGH
1NPC.A            INDAGAAELKRSTVHYGSRYNNAFWNGSQMVYGDGGDTFTSLSGGIDVIGH
E_cytotoxica_XP001373863.1  YDNACAPLKRSTVHYSSGYNNNAFWNGSQMVYGDGGDTFTVPLSGGLDVIKH
B_megaterium_YP005495105.1  YDNKGTTIQSSVHYSKNYNNAFWNGSQMVYGDGGDTFTIPLSGGLDVVVAH
E_sp_SG-1_ZP01858398.1     INGNGLLIRSTVHYGSRYNNAFWNGAQIVFGDDGGDTMFRLSLGGDLVVVGH
FamPro1           EDGNGLLIRSTVHYGSRYNNAFWNGAQIVFGDDGGDTIEFGPFGDLDVVVGH
PhuPro2           LDGNGLLIRSSVHYSSRYNNAFWNGTQIVFGDDGGDTFTIPLSGGLDVVVGH
PpePro1           LDGRGMQIRSTVHYGKRYNNAFWNGSQMITYGDGGDTFTFPFGDFFVVVGH
PspPro2           IDGRGMQIRSTVHYGKRYNNAFWNGSQMITYGDGGDTFTFPFGDFFVVVGH
EpoPro1           IDGRGLQIRSTVHYGSRYNNAFWNGSQMITYGDGGDTFTIAFSGGPDVVVGH
PpePro2           VDGRGLQIRSTVHYGSRYNNAFWNGSQMITYGDGGDTFTIAFSGDFFVVVGH
PspPro3           IDGRGLQIRSTVHYGNRYNNAFWNGSQMITYGDGGDTFTIAFSGGPDVVVGH
PehPro1           IDGNQFLKIRSTVHYSSRYNNAFWNGVQMVYGDGGDTFTIPLSADFFVIGH
PhuPro1           IDGNGLAIRSTVHYSTRYNNNAFWNGSQMVYGDGGDTFTIAFSGGLDVVVGH
FtePro1           LDGRGMAIKIRSTVHYGSRYNNAFWNGTQIAYVGDGGDTFTFRAFSGGLDVIGH
BbrPro1           YDGNCAVIRSTVHYSTRYNNNAFWNGSQMVYGDGGDTFTIPLSGGIDVVVAH
NprE              YDNKGGKILVSSVHYGSRYNNAAWIGDQMIYGDGGDTFTSPSLSGSMDVTAH
NprE_variant      YDNKGGKRVSSVHYGSRYNNAAWIGDQMIYGDGGDTFTSPSLSGSMDVTAH
. . . : : * : * * * * * : : * * * * * : : * : * * : :

1KEI.A            ELTHAVTDYTAGLTYQNESGALNEAISDFGTLEVFYANRNPDWIEIGEDV
E_caldolyticus_AAA22623.1   ELTHAVTDYTAGLVYQNESGALNEAMSDIFGTLEVFYANRNPDWIEIGEDI
E_anthraxis_NP843132.1     ELTHAVTEYSSDLTYQNESGALNEAISDFVFGTEVEYDNRNPDWEIIGEDI
B_thuringiensis_NP893436.1  ELTHAVTEYSSDLTYQNESGALNEAISDFVFGTEVEYDNRNPDWEIIGEDI
E_cereus_ZP04310163.1     ELTHAVTEYSSDLTYQNESGALNEAISDFVFGTEVEYDNRNPDWEIIGEDI
Lactobacillus_sp_BAA06144.1 ELTHAVTEYSSDLTYQNESGALNEAISDFVFGTEVEYDNRNPDWEIIGEDI
1NPC.A            ELTHAVTEYSSDLTYQNESGALNEAISDFGTLEVFYDNRNPDWEIIGEDI
E_cytotoxica_XP001373863.1  ELTHAVTEYSSDLTYQNESGALNEAISDFGTLEVFYDNRNPDWEIIGEDI
B_megaterium_YP005495105.1  ELTHAVTEYSSDLTYQNESGALNEAISDFGTLEVFYDNRNPDWEIIGEDI
E_sp_SG-1_ZP01858398.1     ELTHAVTEYSSDLTYQNESGALNEAISDFGTLEVFYHENHNPDPWEIIGEDI
FamPro1           ELTHGVTEYTSANLEYRNEPGALNEAFADIFGNTIQ-----SKNWLIGDDI
PbaPro1           ELTHGVTEYTSANLEYRNEPGALNEAFADIFGNTIQ-----SKNWLIGDDI
PhuPro2           ELTHGVTEYTSNLEYNQYVNESGALNEAFADIFGNTIQ-----AKNWLIGDDI
EpoPro1           ELTHGVTEYTSNLEYYGESGALNEAFSDIFGNDIQ-----KANWLLGDCI
PspPro2           ELTHGVTEYTSNLEYYGESGALNEAFSDIFGNDIQ-----GTWLLGDCI
PpePro1           ELTHGVTEYTSNLEYYGESGALNEAFSDIFGNDIQ-----RKNWLIGDDI
PpePro2           ELTHGVTEYTSNLEYYGESGALNEAFSDIFGNDIQ-----RKNWLIGDDI
PspPro3           ELTHGVTEYTSNLEYYGESGALNEAFSDIFGNDIQ-----RKNWLIGDDI
PehPro1           ELTHGVTEYTAGLEYYGESGALNESISDFIGNALID-----GKNWLLIGDII
PhuPro1           ELTHGVTEYTSANLEYYGQSGALNESISDFIGNALIE-----GKNWLVGDAI
FtePro1           ELTHGVTEYTAGLEYYGESGALNESISDFVFNITIQ-----GKNWLLIGDDI
BbrPro1           ELTHAVTEYTAGLVYQNESGALNESISDFVFNITIQ-----NDDWLLIGDDI
NprE              EMTHGVTEYTSANLEYRNEPGALNESISDFVFNITIQ-----TEDWLLIGDDI
NprE_variant      EMTHGVTEYTSANLEYRNEPGALNESISDFVFNITIQ-----TEDWLLIGDDI
*:*:* : : : * * : : * * * * * : * * * * * : : * : * * : :


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Figure 9.1B CLUSTAL 2.0.10 multiple sequence alignment of the various *Paenibacillus* metalloproteases with other bacterial metalloprotease homologs.



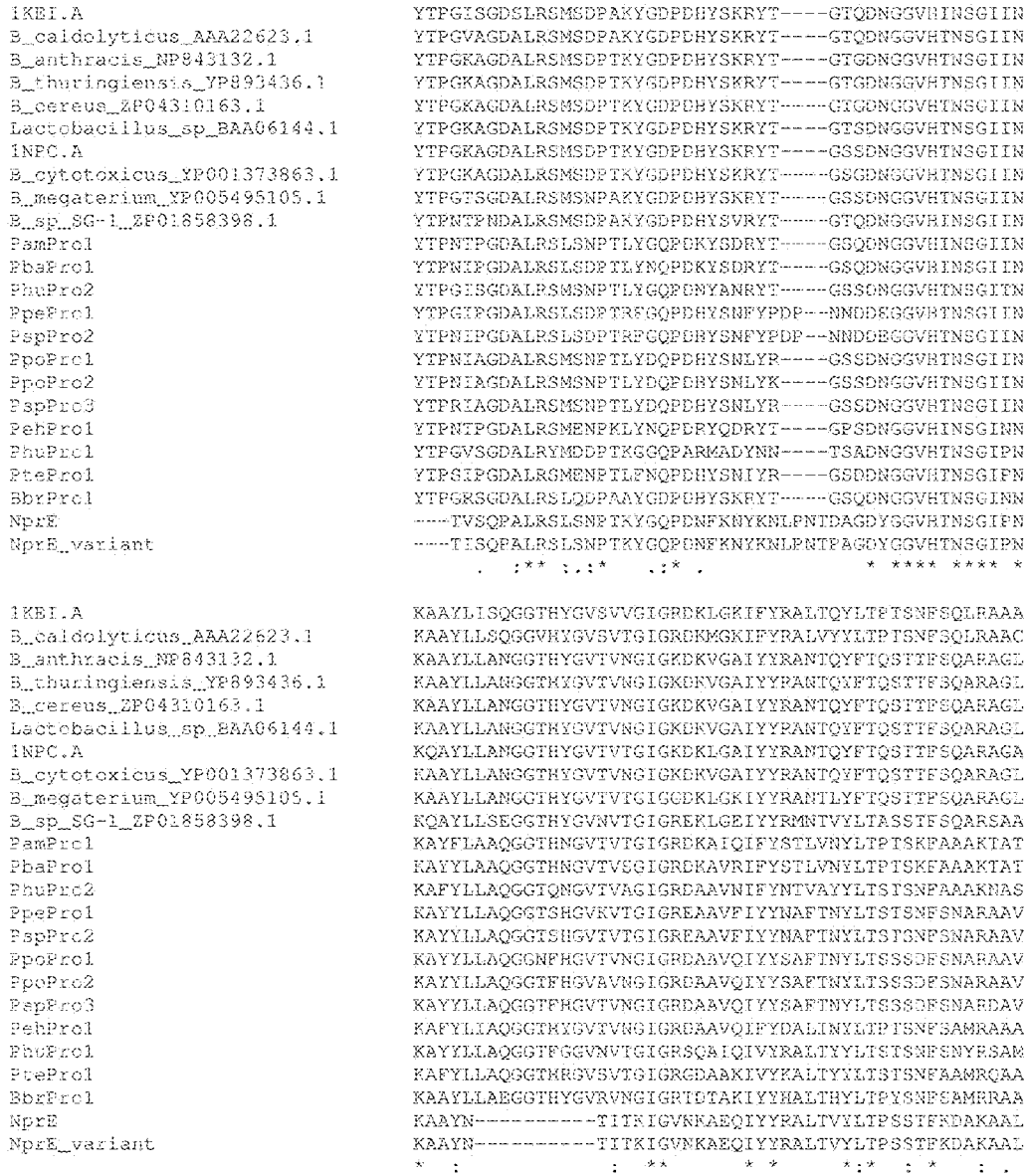


Figure 9.1C CLUSTAL 2.0.10 multiple sequence alignment of the various Paenibacillus metalloproteases with other bacterial metalloprotease homologs.

```

1KEI.A          VQSATDLYGSTSQEVASVKQAFDAVGVK  SEQ ID NO: 53
E_caldolyticus_AAA22623.1  VQAAADLYGSTSQEVNSVKQAFNAVGVY  SEQ ID NO: 54
E_anthraxis_NP843132.1    VQA-----  SEQ ID NO: 55
E_thuringiensis_YP893436.1 VQAAADLYGASSAEVAAVKQSYSAVGVN  SEQ ID NO: 56
E_cereus_ZP04310163.1    VQAAADLYGASSAEVAAVKQSYSAVGVN  SEQ ID NO: 57
Lactobacillus_sp_BAA06144.1 VQAAADLYGASSAEVAAVKQSYSAVGVN  SEQ ID NO: 58
INPC.A          VQAAADLYGANSAEVAAVKQSFSAVGVN  SEQ ID NO: 59
E_cytotoxicus_YP001373863.1 VQAAADLYGANSAEVTAVKQSYDAVGVK  SEQ ID NO: 60
E_megaterium_YF005495105.1 VQAAADLYSGSGSQEIVSVGKSFDAVGVQ  SEQ ID NO: 61
B_sp_SG-1_ZPG1858398.1    VQAAADLYGANSPEVQSVNQSFDAVGIN  SEQ ID NO: 62
PamPro1         IQAAKDLYGATSAEATAITKAYQAVGL-  SEQ ID NO: 38
PbaPro1         IQAAKDLYGANSAEATAITKAYQAVGL-  SEQ ID NO: 23
PhuPro2         IQAAKDLYGTGSSYVTSVTNAFRAVGL-  SEQ ID NO: 13
PpePro1         IQAAKDFYGADSLAVTSAIKSFDAVGIK  SEQ ID NO: 63
PspPro2         IQAAKDFYGADSLAVTSAIQSFDAVGIK  SEQ ID NO: 8
PpoPro1         IQAAKDLYGANSAEATAAAKSFDAVGVN  SEQ ID NO: 28
PpoPro2         IQAAKDLYGANSAEATAAAKSFDAVGVN  SEQ ID NO: 64
PspPro3         VQAAKDLYGASSAQATAAAKSFDAVGVN  SEQ ID NO: 3
FehPro1         IQAAADLYGANSQVNAVKKAYTAVGVN  SEQ ID NO: 18
PhuPro1         VQASTDLYGANSTQTTAVKNSLSAVGIN  SEQ ID NO: 33
PtePro1         ISSATDLEFGANSAQVNSVKAAYAVGI-  SEQ ID NO: 65
EbrPro1         VLSATDLEFGANSRQVQAVNAAVDVAVGVK  SEQ ID NO: 66
NprE           IQSARDLYGSSQDAASVEAAWNAVGL-  SEQ ID NO: 67
NprE_variant    IQSARDLYGSSQDAASVEAAWNAVGL-  SEQ ID NO: 68
:
:

```

Figure 9.1D CLUSTAL 2.0.10 multiple sequence alignment of the various Paenibacillus metalloproteases with other bacterial metalloprotease homologs.

Phylogenetic tree

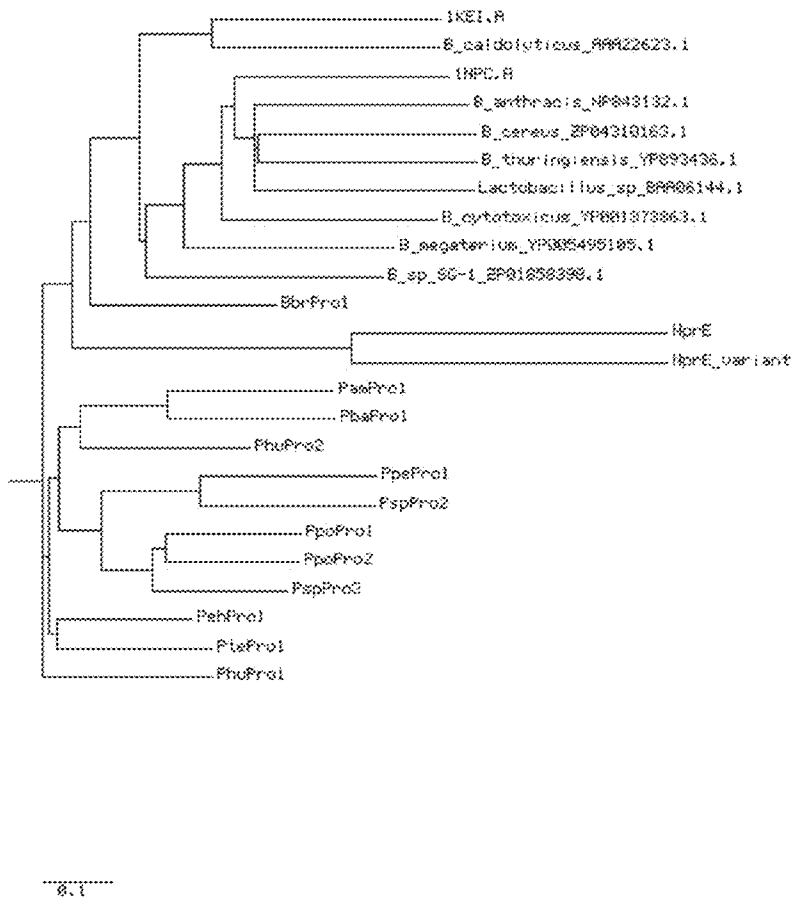


Figure 9.2 The phylogenetic tree of the various Paenibacillus metalloproteases with other bacterial metalloprotease homologs.

## NOVEL METALLOPROTEASES

### CROSS REFERENCE TO RELATED APPLICATIONS

[0001] This application claims benefit of priority from International patent applications Serial No. PCT/CN2013/076419; Serial No. PCT/CN2013/076387; Serial No. PCT/CN2013/076401; Serial No. PCT/CN2013/076406; Serial No. PCT/CN2013/076414; Serial No. PCT/CN2013/076384; Serial No. PCT/CN2013/076398; and Serial No. PCT/CN2013/076415; all filed on 29 May 2013, the contents of which are incorporated herein by reference in their entirety.

### FIELD OF THE INVENTION

[0002] The present disclosure relates to proteases and variants thereof. Compositions containing the proteases are suitable for use in cleaning, food and feed as well as in a variety of other industrial applications.

### BACKGROUND

[0003] Metalloproteases (MPs) are among the hydrolases that mediate nucleophilic attack on peptide bonds using a water molecule coordinated in the active site. In their case, a divalent ion, such as zinc, activates the water molecule. This metal ion is held in place by amino acid ligands, usually 3 in number. The clan MA consists of zinc-dependent MPs in which two of the zinc ligands are the histidines in the motif: HisGluXXHis (SEQ ID NO: 41). This Glu is the catalytic residue. These are two domain proteases with the active site between the domains. In subclass MA(E), also known as Glu-zincins, the 3<sup>rd</sup> ligand is a Glu located C-terminal to the HDXXH (SEQ ID NO: 42) motif. Members of the families: M1, 3, 4, 13, 27 and 34 are all secreted proteases, almost exclusively from bacteria (Rawlings and Salvesen (2013) Handbook of Proteolytic Enzymes, Elsevier Press). They are generally active at elevated temperatures and this stability is attributed to calcium binding. Thermolysin-like proteases are found in the M4 family as defined by MEROPS (Rawlings et al., (2012) Nucleic Acids Res 40:D343-D350). Although proteases have long been known in the art of industrial enzymes, there remains a need for novel proteases that are suitable for particular conditions and uses.

### SUMMARY

[0004] The present disclosure provides novel metalloprotease enzymes, nucleic acids encoding the same, and compositions and methods related to the production and use thereof.

[0005] In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 3. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, Alicyclobacillaceae, Lactobacillaceae, or a *Bacillus*, *Alicyclobacillus*, *Geobacillus*, *Exiguobacterium*, *Lactobacillus*, or *Paenibacillus* spp., such as *Paenibacillus polymyxa*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the Pseudococcidae, or a *Planococcus* spp., such as *Planococcus donghaensis*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50%

of its maximal activity between pH 5 and 9.5. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 30° C. and 70° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

[0006] In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 8. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, or Brevibacillaceae, or a *Bacillus*, *Brevibacillus*, or *Paenibacillus* spp., such as *Paenibacillus* sp. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from *Brevibacillus* sp. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5 and 10. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 35° C. and 70° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

[0007] In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 13. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, or Brevibacillaceae, or a *Bacillus*, *Geobacillus*, *Brevibacillus*, or *Paenibacillus* spp., such as *Paenibacillus humicus*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from *Bacillus polymyxa*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5 and 9.5. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 35° C. and 70° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

[0008] In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 18. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, or Brevibacillaceae, or a *Bacillus*, *Geobacillus*, *Brevibacillus*, or *Paenibacillus* spp., such as *Paenibacillus ehimensis*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from *Brevibacillus* sp. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5 and 10.5. In various

embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 45° C. and 75° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

**[0009]** In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 23. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, Alicyclobacillaceae, Lactobacillaceae, or a *Bacillus*, *Geobacillus*, *Alicyclobacillus*, *Brevibacillus*, *Paenibacillus*, or *Lactobacillus* spp., such as *Paenibacillus barcinonensis*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the family Pseudococcidae, or a *Planococcus* spp., such as *Planococcus donghaensis*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5 and 10. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 35° C. and 65° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

**[0010]** In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 28. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, or a *Bacillus*, *Brevibacillus*, *Paenibacillus*, or *Lactobacillus* spp., such as *Paenibacillus polymyxa*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the family Pseudococcidae, or a *Planococcus* spp., such as *Planococcus donghaensis*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5 and 9.5. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 30° C. and 65° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

**[0011]** In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 33. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, or a *Bacillus*, *Geobacillus*, *Brevibacillus*, or *Paenibacillus* spp., such as *Paenibacillus humanensis*. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from *Bacillus polymyxa*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein

hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 4.5 and 9.0. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 35° C. and 70° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

**[0012]** In some embodiments, the invention is a polypeptide comprising an amino acid sequence having at least 60%, at least 80%, or at least 95% sequence identity to the amino acid sequence of SEQ ID NO: 38. In some embodiments, the invention is any of the above, wherein said polypeptide is derived from a member of the order Bacillales; family Bacillaceae, Paenibacillaceae, Lactobacillaceae, or a *Bacillus*, *Brevibacillus*, *Lactobacillus*, *Paenibacillus*, or *Geobacillus* spp., such as *Paenibacillus amylolyticus*. In various embodiments of the invention, any of the above polypeptides has protease activity, such as azo-casein hydrolysis. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between pH 5.5 and 10. In various embodiments of the invention, any of the above polypeptides retains at least 50% of its maximal activity between 35° C. and 65° C. In various embodiments of the invention, any of the above polypeptides has cleaning activity in a detergent composition, such as an ADW, laundry, liquid laundry, or powder laundry detergent composition.

**[0013]** In some embodiments, the invention is a composition comprising any of the above, such as a cleaning or detergent composition. In some embodiments, the composition further comprises a surfactant, at least one calcium ion and/or zinc ion, at least one stabilizer, at least one bleaching agent, and can contain phosphate, or be phosphate-free. In some embodiments, the composition further comprises one or more additional enzymes or enzyme derivatives selected from the group consisting of acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageninases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1,4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxygenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoloxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases, pullulanases, reductases, rhamnogalacturonases, beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases, and combinations thereof. In some embodiments, the composition is formulated at a pH of from about 5.5 to about 8.5. In some embodiments, the invention is a method of cleaning using any of the above polypeptides or compositions. In some embodiments, the invention is a textile processing composition, animal feed composition, leather processing composition, or feather processing composition.

#### BRIEF DESCRIPTION OF THE DRAWINGS

**[0014]** FIG. 1.1 provides a plasmid map of pGX085 (aprE-PspPro3), described in Example 1.2.

**[0015]** FIG. 1.2 provides a dose response curve of PspPro3 in the azo-casein assay.

**[0016]** FIG. 1.3 provides the pH profile of PspPro3.

- [0017] FIG. 1.4 provides the temperature profile of PspPro3.
- [0018] FIG. 1.5A shows dose response for cleaning of PAS-38 microswatches by PspPro3 protein in ADW detergent at pH 6 and 8.
- [0019] FIG. 1.5B shows dose response for cleaning of PAS-38 microswatches shows by PspPro3 protein in ADW detergent at pH 6 and 8 in the presence of bleach.
- [0020] FIG. 1.6 shows cleaning performance of PspPro3 protein in liquid laundry detergent.
- [0021] FIG. 1.7 (SEQ ID NOS: 3, 44, and 45, respectively) shows alignment of PspPro3 with other protein homologs.
- [0022] FIG. 1.8 provides the phylogenetic tree for PspPro3 and its homologs.
- [0023] FIG. 2.1 provides a plasmid map of pGX084 (aprE-PspPro2), described in Example 2.2.
- [0024] FIG. 2.2 provides a dose response curve of PspPro2 in the azo-casein assay.
- [0025] FIG. 2.3 provides the pH profile of purified PspPro2.
- [0026] FIG. 2.4 provides the temperature profile of purified PspPro2.
- [0027] FIG. 2.5A shows dose response for cleaning performance of PspPro2 at pH 6 in AT dish detergent with bleach.
- [0028] FIG. 2.5B shows dose response for cleaning performance of purified PspPro2 at pH 8 in AT detergent with bleach.
- [0029] FIG. 2.6A shows cleaning performance of PspPro2 protein in liquid laundry detergent.
- [0030] FIG. 2.6B shows cleaning performance of PspPro2 protein in powder laundry detergent.
- [0031] FIG. 2.7 (SEQ ID NOS: 8, 46, and 45, respectively) shows alignment of PspPro2 with other protein homologs.
- [0032] FIG. 2.8 provides the phylogenetic tree for PspPro2 and its homologs.
- [0033] FIG. 3.1 provides a plasmid map of pGX150 (aprE-PhuPro2), described in Example 3.2.
- [0034] FIG. 3.2 provides a dose response curve of PhuPro2 in the azo-casein assay.
- [0035] FIG. 3.3 provides the pH profile of purified PhuPro2.
- [0036] FIG. 3.4 provides the temperature profile of purified PhuPro2.
- [0037] FIG. 3.5A shows dose response for cleaning performance of PhuPro2 in AT dish detergent at pH 6.
- [0038] FIG. 3.5B shows dose response for cleaning performance of PhuPro2 in AT dish detergent at pH 8.
- [0039] FIG. 3.6 (SEQ ID NOS: 13, 47 and 45, respectively) shows alignment of PhuPro2 with other protein homologs.
- [0040] FIG. 3.7 provides the phylogenetic tree for PhuPro2 and its homologs.
- [0041] FIG. 4.1 provides a plasmid map of pGX148 (aprE-PehPro1), described in Example 4.2.
- [0042] FIG. 4.2 provides a dose response curve of PehPro1 in the azo-casein assay.
- [0043] FIG. 4.3 provides the pH profile of purified PehPro1.
- [0044] FIG. 4.4 provides the temperature profile of purified PehPro1.
- [0045] FIG. 4.5A shows dose response for cleaning performance of PehPro1 at pH 6 in AT dish detergent with bleach.
- [0046] FIG. 4.5B shows dose response for cleaning performance of purified PehPro1 at pH 8 in AT detergent with bleach.
- [0047] FIG. 4.6 (SEQ ID NOS: 18, 48, and 45, respectively) shows alignment of PehPro1 with other protein homologs.
- [0048] FIG. 4.7 provides the phylogenetic tree for PehPro1 and its homologs.
- [0049] FIG. 5.1 provides a plasmid map of pGX147 (aprE-PbaPro1), described in Example 5.2.
- [0050] FIG. 5.2 provides a dose response curve of PbaPro1 in the azo-casein assay.
- [0051] FIG. 5.3 provides the pH profile of purified PbaPro1.
- [0052] FIG. 5.4 provides the temperature profile of purified PbaPro1.
- [0053] FIG. 5.5A shows dose response for cleaning of PAS-38 microswatches by PbaPro1 protein in ADW detergent at pH 6.
- [0054] FIG. 5.5B shows dose response for cleaning of PAS-38 microswatches shows by PbaPro1 protein in ADW detergent at pH 8.
- [0055] FIG. 5.6 (SEQ ID NOS: 23, 49, and 45, respectively) shows the alignment of PbaPro1 with protease homologs.
- [0056] FIG. 5.7 provides the phylogenetic tree for PbaPro1 and its homologs.
- [0057] FIG. 6.1 provides a plasmid map of pGX138 (aprE-PpoPro1), described in Example 6.2.
- [0058] FIG. 6.2 provides a dose response curve of PpoPro1 in the azo-casein assay.
- [0059] FIG. 6.3 provides the pH profile of purified PpoPro1.
- [0060] FIG. 6.4 provides the temperature profile of purified PpoPro1.
- [0061] FIG. 6.5A shows dose response for cleaning of PAS-38 microswatches by PpoPro1 protein in ADW detergent at pH 6 in the presence of bleach.
- [0062] FIG. 6.5B shows dose response for cleaning of PAS-38 microswatches shows by PpoPro1 protein in ADW detergent at pH 8 in the presence of bleach.
- [0063] FIG. 6.6 (SEQ ID NOS: 28, 50, and 45, respectively) shows the alignment of PpoPro1 with protease homologs.
- [0064] FIG. 6.7 provides the phylogenetic tree for PpoPro1 and its homologs.
- [0065] FIG. 7.1 provides a plasmid map of pGX149 (aprE-PhuPro1), described in Example 7.2.
- [0066] FIG. 7.2 provides a dose response curve of PhuPro1 in the azo-casein assay.
- [0067] FIG. 7.3 provides the pH profile of purified PhuPro1.
- [0068] FIG. 7.4 provides the temperature profile of purified PhuPro1.
- [0069] FIG. 7.5A shows dose response for cleaning of PAS-38 microswatches by PhuPro1 protein in ADW detergent at pH 6.
- [0070] FIG. 7.5B shows dose response for cleaning of PAS-38 microswatches shows by Phu Pro1 protein in ADW detergent at pH 8.
- [0071] FIG. 7.6 (SEQ ID NOS: 33, 51, and 45, respectively) shows alignment of PhuPro1 with other protein homologs.
- [0072] FIG. 7.7 provides the phylogenetic tree for PhuPro1 and its homologs.
- [0073] FIGS. 7.8A and 7.8B show cleaning performances of PhuPro1 and Purafect® Prime HA proteases.
- [0074] FIG. 8.1 provides a plasmid map of pGX146 (aprE-PamPro1), described in Example 8.2.
- [0075] FIG. 8.2 provides a dose response curve of PamPro1 in the azo-casein assay.

**[0076]** FIG. 8.3 provides the pH profile of purified PamPro1.

**[0077]** FIG. 8.4 provides the temperature profile of purified PamPro1.

**[0078]** FIG. 8.5A shows dose response for cleaning of PA-S-38 microswatches by PamPro1 protein in ADW detergent at pH 6.

**[0079]** FIG. 8.5B shows dose response for cleaning of PA-S-38 microswatches shows by PamPro1 protein in ADW detergent at pH 8.

**[0080]** FIG. 8.6 (SEQ ID NOS: 38, 52, and 45, respectively) shows the alignment of PamPro1 with protease homologs.

**[0081]** FIG. 8.7 provides the phylogenetic tree for PamPro1 and its homologs.

**[0082]** FIGS. 9.1A thru 9.1D (SEQ ID NOS: 53-62, 38, 23, 13, 63, 8, 28, 64, 3, 18, 33, 65-68, respectively) show the alignment of the various *Paenibacillus* metalloproteases with other bacterial metalloprotease homologs.

**[0083]** FIG. 9.2 provides the phylogenetic tree of the various *Paenibacillus* metalloproteases with other bacterial metalloprotease homologs.

#### DETAILED DESCRIPTION

**[0084]** The present invention provides novel metalloprotease enzymes, especially enzymes useful for detergent compositions cloned from various *Paenibacillus* sp. The compositions and methods are based, in part, on the observation that the novel metalloproteases of the present invention have proteolytic activity in the presence of detergent compositions. This feature makes metalloproteases of the present invention particularly well suited to and useful in a variety of cleaning applications where the enzyme can hydrolyze polypeptides in the presence of surfactants and other components found in detergent compositions. The invention includes compositions comprising at least one of the novel metalloprotease enzymes set forth herein. Some such compositions comprise detergent compositions. The metalloprotease enzymes of the present invention can be combined with other enzymes useful in detergent compositions. The invention also provides methods of cleaning using metalloprotease enzymes of the present invention.

#### DEFINITIONS AND ABBREVIATIONS

**[0085]** Unless otherwise indicated, the practice of the present invention involves conventional techniques commonly used in molecular biology, protein engineering, microbiology, and recombinant DNA technology, which are within the skill of the art. Such techniques are known to those of skill in the art and are described in numerous texts and reference works well known to those of skill in the art. All patents, patent applications, articles and publications mentioned herein, both supra and infra, are hereby expressly incorporated herein by reference.

**[0086]** Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Many technical dictionaries are known to those of skill in the art. Although any methods and materials similar or equivalent to those described herein find use in the practice of the present invention, some suitable methods and materials are described herein. Accordingly, the terms defined immediately below are more fully described by reference to the Specification as a whole. Also, as used herein,

the singular “a”, “an” and “the” includes the plural reference unless the context clearly indicates otherwise. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. It is to be understood that this invention is not limited to the particular methodology, protocols, and reagents described, as these may vary, depending upon the context they are used by those of skill in the art.

**[0087]** Furthermore, the headings provided herein are not limitations of the various aspects or embodiments of the invention.

**[0088]** It is intended that every maximum numerical limitation given throughout this specification includes every lower numerical limitation, as if such lower numerical limitations were expressly written herein. Every minimum numerical limitation given throughout this specification will include every higher numerical limitation, as if such higher numerical limitations were expressly written herein. Every numerical range given throughout this specification will include every narrower numerical range that falls within such broader numerical range, as if such narrower numerical ranges were all expressly written herein.

**[0089]** As used herein, the terms “protease” and “proteinase” refer to an enzyme that has the ability to break down proteins and peptides. A protease has the ability to conduct “proteolysis,” by hydrolysis of peptide bonds that link amino acids together in a peptide or polypeptide chain forming the protein. This activity of a protease as a protein-digesting enzyme is referred to as “proteolytic activity.” Many well known procedures exist for measuring proteolytic activity (See e.g., Kalisz, “Microbial Proteinases,” In: Fiechter (ed.), *Advances in Biochemical Engineering/Biotechnology*, (1988)). For example, proteolytic activity may be ascertained by comparative assays which analyze the respective protease’s ability to hydrolyze a suitable substrate. Exemplary substrates useful in the analysis of protease or proteolytic activity, include, but are not limited to, di-methyl casein (Sigma C-9801), bovine collagen (Sigma C-9879), bovine elastin (Sigma E-1625), and bovine keratin (ICN Biomedical 902111). Colorimetric assays utilizing these substrates are well known in the art (See e.g., WO 99/34011 and U.S. Pat. No. 6,376,450, both of which are incorporated herein by reference). The pNA peptidyl assay (See e.g., Del Mar et al., *Anal. Biochem.* 99:316-320 [1979]) also finds use in determining the active enzyme concentration. This assay measures the rate at which p-nitroaniline is released as the enzyme hydrolyzes a soluble synthetic substrate, such as succinyl-alanine-alanine-proline-phenylalanine-p-nitroanilide (suc-AAPF-pNA) (SEQ ID NO: 43). The rate of production of yellow color from the hydrolysis reaction is measured at 410 nm on a spectrophotometer and is proportional to the active enzyme concentration. In addition, absorbance measurements at 280 nanometers (nm) can be used to determine the total protein concentration in a sample of purified protein. The activity on substrate/protein concentration gives the enzyme specific activity.

**[0090]** As used herein, the term “variant polypeptide” refers to a polypeptide comprising an amino acid sequence that differs in at least one amino acid residue from the amino acid sequence of a parent or reference polypeptide (including but not limited to wild-type polypeptides).

**[0091]** As used herein, “the genus *Bacillus*” includes all species within the genus “*Bacillus*,” as known to those of skill

in the art, including but not limited to *B. subtilis*, *B. licheniformis*, *B. lentus*, *B. brevis*, *B. stearothermophilus*, *B. alkalophilus*, *B. amyloliquefaciens*, *B. clausii*, *B. halodurans*, *B. megaterium*, *B. coagulans*, *B. circulans*, and *B. thuringiensis*. It is recognized that the genus *Bacillus* continues to undergo taxonomical reorganization. Thus, it is intended that the genus include species that have been reclassified, including but not limited to such organisms as *B. stearothermophilus*, which is now named “*Geobacillus stearothermophilus*.” The production of resistant endospores under stressful environmental conditions is considered the defining feature of the genus *Bacillus*, although this characteristic also applies to the recently named *Alicyclobacillus*, *Amphibacillus*, *Aneurinibacillus*, *Paenibacillus*, *Brevibacillus*, *Filobacillus*, *Gracilibacillus*, *Halobacillus*, *Paenibacillus*, *Salibacillus*, *Thermobacillus*, *Ureibacillus*, and *Virgibacillus*.

**[0092]** The terms “polynucleotide” and “nucleic acid,” which are used interchangeably herein, refer to a polymer of any length of nucleotide monomers covalently bonded in a chain. DNA (deoxyribonucleic acid), a polynucleotide comprising deoxyribonucleotides, and RNA (ribonucleic acid), a polymer of ribonucleotides, are examples of polynucleotides or nucleic acids having distinct biological function. Polynucleotides or nucleic acids include, but are not limited to, a single-, double- or triple-stranded DNA, genomic DNA, cDNA, RNA, DNA-RNA hybrid, or a polymer comprising purine and pyrimidine bases, or other natural, chemically, biochemically modified, non-natural or derivatized nucleotide bases. The following are non-limiting examples of polynucleotides: genes, gene fragments, chromosomal fragments, expressed sequence tag(s) (EST(s)), exons, introns, messenger RNA (mRNA), transfer RNA (tRNA), ribosomal RNA (rRNA), ribozymes, complementary DNA (cDNA), recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers.

**[0093]** As used herein, the term “mutation” refers to changes made to a reference amino acid or nucleic acid sequence. It is intended that the term encompass substitutions, insertions and deletions.

**[0094]** As used herein, the term “vector” refers to a nucleic acid construct used to introduce or transfer nucleic acid(s) into a target cell or tissue. A vector is typically used to introduce foreign DNA into a cell or tissue. Vectors include plasmids, cloning vectors, bacteriophages, viruses (e.g., viral vector), cosmids, expression vectors, shuttle vectors, and the like. A vector typically includes an origin of replication, a multicloning site, and a selectable marker. The process of inserting a vector into a target cell is typically referred to as transformation. The present invention includes, in some embodiments, a vector that comprises a DNA sequence encoding a metalloprotease polypeptide (e.g., precursor or mature metalloprotease polypeptide) that is operably linked to a suitable prosequence (e.g., secretory, signal peptide sequence, etc.) capable of effecting the expression of the DNA sequence in a suitable host, and the folding and translocation of the recombinant polypeptide chain.

**[0095]** As used herein, the term “expression cassette,” “expression plasmid” or “expression vector” refers to a nucleic acid construct or vector generated recombinantly or synthetically for the expression of a nucleic acid of interest in a target cell. An expression vector or expression cassette typically comprises a promoter nucleotide sequence that drives expression of the foreign nucleic acid. The expression

vector or cassette also typically includes any other specified nucleic acid elements that permit transcription of a particular nucleic acid in a target cell. A recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Many prokaryotic and eukaryotic expression vectors are commercially available.

**[0096]** In some embodiments, the ends of the sequence are closed such that the DNA construct forms a closed circle. The nucleic acid sequence of interest, which is incorporated into the DNA construct, using techniques well known in the art, may be a wild-type, mutant, or modified nucleic acid. In some embodiments, the DNA construct comprises one or more nucleic acid sequences homologous to the host cell chromosome. In other embodiments, the DNA construct comprises one or more non-homologous nucleotide sequences. Once the DNA construct is assembled in vitro, it may be used, for example, to: 1) insert heterologous sequences into a desired target sequence of a host cell; and/or 2) mutagenize a region of the host cell chromosome (i.e., replace an endogenous sequence with a heterologous sequence); 3) delete target genes; and/or 4) introduce a replicating plasmid into the host. “DNA construct” is used interchangeably herein with “expression cassette.”

**[0097]** As used herein, a “plasmid” refers to an extrachromosomal DNA molecule which is capable of replicating independently from the chromosomal DNA. A plasmid is double stranded (ds) and may be circular and is typically used as a cloning vector.

**[0098]** As used herein in the context of introducing a nucleic acid sequence into a cell, the term “introduced” refers to any method suitable for transferring the nucleic acid sequence into the cell. Such methods for introduction include but are not limited to protoplast fusion, transfection, transformation, electroporation, conjugation, and transduction (See e.g., Ferrari et al., “Genetics,” in Hardwood et al. (eds.), *Bacillus*, Plenum Publishing Corp., pp. 57-72 [1989]).

**[0099]** Transformation refers to the genetic alteration of a cell which results from the uptake, optional genomic incorporation, and expression of genetic material (e.g., DNA).

**[0100]** As used herein, a nucleic acid is “operably linked” with another nucleic acid sequence when it is placed into a functional relationship with another nucleic acid sequence. For example, a promoter or enhancer is operably linked to a nucleotide coding sequence if the promoter affects the transcription of the coding sequence. A ribosome binding site may be operably linked to a coding sequence if it is positioned so as to facilitate translation of the coding sequence. Typically, “operably linked” DNA sequences are contiguous. However, enhancers do not have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, synthetic oligonucleotide adaptors or linkers may be used in accordance with conventional practice.

**[0101]** As used herein the term “gene” refers to a polynucleotide (e.g., a DNA segment), that encodes a polypeptide and includes regions preceding and following the coding regions as well as intervening sequences (introns) between individual coding segments (exons).

**[0102]** As used herein, “recombinant” when used with reference to a cell typically indicates that the cell has been modified by the introduction of a foreign nucleic acid sequence or that the cell is derived from a cell so modified. For example, a recombinant cell may comprise a gene not found in identical form within the native (non-recombinant) form of



the cell, or a recombinant cell may comprise a native gene (found in the native form of the cell) but which has been modified and re-introduced into the cell. A recombinant cell may comprise a nucleic acid endogenous to the cell that has been modified without removing the nucleic acid from the cell; such modifications include those obtained by gene replacement, site-specific mutation, and related techniques known to those of ordinary skill in the art. Recombinant DNA technology includes techniques for the production of recombinant DNA in vitro and transfer of the recombinant DNA into cells where it may be expressed or propagated, thereby producing a recombinant polypeptide. "Recombination," "recombining," and "recombined" of polynucleotides or nucleic acids refer generally to the assembly or combining of two or more nucleic acid or polynucleotide strands or fragments to generate a new polynucleotide or nucleic acid. The recombinant polynucleotide or nucleic acid is sometimes referred to as a chimera. A nucleic acid or polypeptide is "recombinant" when it is artificial or engineered.

**[0103]** A nucleic acid or polynucleotide is said to "encode" a polypeptide if, in its native state or when manipulated by methods known to those of skill in the art, it can be transcribed and/or translated to produce the polypeptide or a fragment thereof. The anti-sense strand of such a nucleic acid is also said to encode the sequence.

**[0104]** "Host strain" or "host cell" refers to a suitable host for an expression vector comprising a DNA sequence of interest.

**[0105]** A "protein" or "polypeptide" comprises a polymeric sequence of amino acid residues. The terms "protein" and "polypeptide" are used interchangeably herein. The single and 3-letter code for amino acids as defined in conformity with the IUPAC-IUB Joint Commission on Biochemical Nomenclature (JCBN) is used through out this disclosure. The single letter X refers to any of the twenty amino acids. It is also understood that a polypeptide may be coded for by more than one nucleotide sequence due to the degeneracy of the genetic code. Mutations can be named by the one letter code for the parent amino acid, followed by a position number and then the one letter code for the variant amino acid. For example, mutating glycine (G) at position 87 to serine (S) is represented as "G087S" or "G87S". Mutations can also be named by using the three letter code for an amino acid followed by its position in the polypeptide chain as counted from the N-terminus; for example, Ala10 for alanine at position 10. Multiple mutations are indicated by inserting a "-" between the mutations. Mutations at positions 87 and 90 are represented as either "G087S-A090Y" or "G87S-A90Y" or "G87S+A90Y" or "G087S+A090Y". For deletions, the one letter code "Z" is used. For an insertion relative to the parent sequence, the one letter code "Z" is on the left side of the position number. For a deletion, the one letter code "Z" is on the right side of the position number. For insertions, the position number is the position number before the inserted amino acid(s), plus 0.01 for each amino acid. For example, an insertion of three amino acids alanine (A), serine (S) and tyrosine (Y) between position 87 and 88 is shown as "Z087.01A-Z087.02S-Z087.03Y." Thus, combining all the mutations above plus a deletion at position 100 is: "G087S-Z087.01A-Z087.02S-Z087.03Y-A090Y-A100Z." When describing modifications, a position followed by amino acids listed in parentheses indicates a list of substitutions at that

position by any of the listed amino acids. For example, 6(L,I) means position 6 can be substituted with a leucine or isoleucine.

**[0106]** A "prosequence" or "propeptide sequence" refers to an amino acid sequence between the signal peptide sequence and mature protease sequence that is necessary for the proper folding and secretion of the protease; they are sometimes referred to as intramolecular chaperones. Cleavage of the prosequence or propeptide sequence results in a mature active protease. Bacterial metalloproteases are often expressed as pro-enzymes.

**[0107]** The term "signal sequence" or "signal peptide" refers to a sequence of amino acid residues that may participate in the secretion or direct transport of the mature or precursor form of a protein. The signal sequence is typically located N-terminal to the precursor or mature protein sequence. The signal sequence may be endogenous or exogenous. A signal sequence is normally absent from the mature protein. A signal sequence is typically cleaved from the protein by a signal peptidase after the protein is transported.

**[0108]** The term "mature" form of a protein, polypeptide, or peptide refers to the functional form of the protein, polypeptide, or peptide without the signal peptide sequence and propeptide sequence.

**[0109]** The term "precursor" form of a protein or peptide refers to a mature form of the protein having a prosequence operably linked to the amino or carbonyl terminus of the protein. The precursor may also have a "signal" sequence operably linked to the amino terminus of the prosequence. The precursor may also have additional polypeptides that are involved in post-translational activity (e.g., polypeptides cleaved therefrom to leave the mature form of a protein or peptide).

**[0110]** The term "wild-type" in reference to an amino acid sequence or nucleic acid sequence indicates that the amino acid sequence or nucleic acid sequence is native or naturally occurring sequence. As used herein, the term "naturally-occurring" refers to anything (e.g., proteins, amino acids, or nucleic acid sequences) that are found in nature.

**[0111]** As used herein, the term "non-naturally occurring" refers to anything that is not found in nature (e.g., recombinant nucleic acids and protein sequences produced in the laboratory), as modification of the wild-type sequence.

**[0112]** As used herein with regard to amino acid residue positions, "corresponding to" or "corresponds to" or "corresponds" refers to an amino acid residue at the enumerated position in a protein or peptide, or an amino acid residue that is analogous, homologous, or equivalent to an enumerated residue in a protein or peptide. As used herein, "corresponding region" generally refers to an analogous position in a related proteins or a reference protein.

**[0113]** The terms "derived from" and "obtained from" refer to not only a protein produced or producible by a strain of the organism in question, but also a protein encoded by a DNA sequence isolated from such strain and produced in a host organism containing such DNA sequence. Additionally, the term refers to a protein which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the protein in question. To exemplify, "proteases derived from *Bacillus*" refers to those enzymes having proteolytic activity which are naturally produced by *Bacillus*, as well as to serine proteases like those produced by *Bacillus* sources but which through the use of genetic engi-

neering techniques are produced by non-*Bacillus* organisms transformed with a nucleic acid encoding the serine proteases.

**[0114]** The term “identical” in the context of two nucleic acids or polypeptide sequences refers to the residues in the two sequences that are the same when aligned for maximum correspondence, as measured using one of the following sequence comparison or analysis algorithms.

**[0115]** As used herein, “homologous genes” refers to a pair of genes from different, but usually related species, which correspond to each other and which are identical or very similar to each other. The term encompasses genes that are separated by speciation (i.e., the development of new species) (e.g., orthologous genes), as well as genes that have been separated by genetic duplication (e.g., paralogous genes).

**[0116]** As used herein, “% identity or percent identity” refers to sequence similarity. Percent identity may be determined using standard techniques known in the art (See e.g., Smith and Waterman, *Adv. Appl. Math.* 2:482 [1981]; Needleman and Wunsch, *J. Mol. Biol.* 48:443 [1970]; Pearson and Lipman, *Proc. Natl. Acad. Sci. USA* 85:2444 [1988]; software programs such as GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package (Genetics Computer Group, Madison, Wis.); and Devereux et al., *Nucl. Acid Res.* 12:387-395 [1984]). One example of a useful algorithm is PILEUP. PILEUP creates a multiple sequence alignment from a group of related sequences using progressive, pair-wise alignments. It can also plot a tree showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng and Doolittle (See, Feng and Doolittle, *J. Mol. Evol.* 35:351-360 [1987]). The method is similar to that described by Higgins and Sharp (See, Higgins and Sharp, *CABIOS* 5:151-153 [1989]). Useful PILEUP parameters include a default gap weight of 3.00, a default gap length weight of 0.10, and weighted end gaps. Other useful algorithm is the BLAST algorithms described by Altschul et al., (See, Altschul et al., *J. Mol. Biol.* 215:403-410 [1990]; and Karlin and Altschul, *Proc. Natl. Acad. Sci. USA* 90:5873-5787 [1993]). The BLAST program uses several search parameters, most of which are set to the default values.

**[0117]** The NCBI BLAST algorithm finds the most relevant sequences in terms of biological similarity but is not recommended for query sequences of less than 20 residues (Altschul, S F et al. (1997) *Nucleic Acids Res.* 25:3389-3402 and Schaffer, A A et al. (2001) *Nucleic Acids Res.* 29:2994-3005). Example default BLAST parameters for a nucleic acid sequence searches are:

- [0118]** Neighboring words threshold: 11
- [0119]** E-value cutoff: 10
- [0120]** Scoring Matrix: NUC.3.1 (match=1, mismatch=-3)
- [0121]** Gap Opening: 5
- [0122]** Gap Extension: 2

and the following parameters for amino acid sequence searches:

- [0123]** Word size: 3
- [0124]** E-value cutoff: 10
- [0125]** Scoring Matrix: BLOSUM62
- [0126]** Gap Opening: 11
- [0127]** Gap extension: 1

**[0128]** A percent (%) amino acid sequence identity value is determined by the number of matching identical residues divided by the total number of residues of the “reference”

sequence including any gaps created by the program for optimal/maximum alignment. If a sequence is 90% identical to SEQ ID NO: A, SEQ ID NO: A is the “reference” sequence. BLAST algorithms refer the “reference” sequence as “query” sequence.

**[0129]** The CLUSTAL W algorithm is another example of a sequence alignment algorithm. See Thompson et al. (1994) *Nucleic Acids Res.* 22:4673-4680. Default parameters for the CLUSTAL W algorithm are:

- [0130]** Gap opening penalty: 10.0
- [0131]** Gap extension penalty: 0.05
- [0132]** Protein weight matrix: BLOSUM series
- [0133]** DNA weight matrix: IUB
- [0134]** Delay divergent sequences %: 40
- [0135]** Gap separation distance: 8
- [0136]** DNA transitions weight: 0.50
- [0137]** List hydrophilic residues: GPSNDQEKR
- [0138]** Use negative matrix: OFF
- [0139]** Toggle Residue specific penalties: ON
- [0140]** Toggle hydrophilic penalties: ON
- [0141]** Toggle end gap separation penalty OFF.

**[0142]** In CLUSTAL algorithms, deletions occurring at either terminus are included. For example, a variant with five amino acid deletion at either terminus (or within the polypeptide) of a polypeptide of 500 amino acids would have a percent sequence identity of 99% (495/500 identical residues $\times$ 100) relative to the “reference” polypeptide. Such a variant would be encompassed by a variant having “at least 99% sequence identity” to the polypeptide.

**[0143]** A polypeptide of interest may be said to be “substantially identical” to a reference polypeptide if the polypeptide of interest comprises an amino acid sequence having at least about 60%, least about 65%, least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or at least about 99.5% sequence identity to the amino acid sequence of the reference polypeptide. The percent identity between two such polypeptides can be determined manually by inspection of the two optimally aligned polypeptide sequences or by using software programs or algorithms (e.g., BLAST, ALIGN, CLUSTAL) using standard parameters. One indication that two polypeptides are substantially identical is that the first polypeptide is immunologically cross-reactive with the second polypeptide. Typically, polypeptides that differ by conservative amino acid substitutions are immunologically cross-reactive. Thus, a polypeptide is substantially identical to a second polypeptide, for example, where the two peptides differ only by a conservative amino acid substitution or one or more conservative amino acid substitutions.

**[0144]** A nucleic acid of interest may be said to be “substantially identical” to a reference nucleic acid if the nucleic acid of interest comprises a nucleotide sequence having least about 60%, least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 91%, at least about 92%, at least about 93%, at least about 94%, at least about 95%, at least about 96%, at least about 97%, at least about 98%, at least about 99%, or at least about 99.5% sequence identity to the nucleotide sequence of the reference nucleic acid. The percent identity between two such nucleic acids can be determined manually by inspection of the two optimally aligned nucleic

acid sequences or by using software programs or algorithms (e.g., BLAST, ALIGN, CLUSTAL) using standard parameters. One indication that two nucleic acid sequences are substantially identical is that the two nucleic acid molecules hybridize to each other under stringent conditions (e.g., within a range of medium to high stringency).

**[0145]** A nucleic acid or polynucleotide is “isolated” when it is at least partially or completely separated from other components, including but not limited to for example, other proteins, nucleic acids, cells, etc. Similarly, a polypeptide, protein or peptide is “isolated” when it is at least partially or completely separated from other components, including but not limited to for example, other proteins, nucleic acids, cells, etc. On a molar basis, an isolated species is more abundant than are other species in a composition. For example, an isolated species may comprise at least about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% (on a molar basis) of all macromolecular species present. Preferably, the species of interest is purified to essential homogeneity (i.e., contaminant species cannot be detected in the composition by conventional detection methods). Purity and homogeneity can be determined using a number of techniques well known in the art, such as agarose or polyacrylamide gel electrophoresis of a nucleic acid or a protein sample, respectively, followed by visualization upon staining. If desired, a high-resolution technique, such as high performance liquid chromatography (HPLC) or a similar means can be utilized for purification of the material.

**[0146]** “Hybridization” refers to the process by which one strand of nucleic acid forms a duplex with, i.e., base pairs with, a complementary strand. A nucleic acid sequence is considered to be “selectively hybridizable” to a reference nucleic acid sequence if the two sequences specifically hybridize to one another under moderate to high stringency hybridization and wash conditions. Hybridization conditions are based on the melting temperature ( $T_m$ ) of the nucleic acid binding complex or probe. For example, “maximum stringency” typically occurs at about  $T_m - 5^\circ \text{C}$ . ( $5^\circ$  below the  $T_m$  of the probe); “high stringency” at about  $5 - 10^\circ \text{C}$ . below the  $T_m$ ; “intermediate stringency” at about  $10 - 20^\circ \text{C}$ . below the  $T_m$  of the probe; and “low stringency” at about  $20 - 25^\circ \text{C}$ . below the  $T_m$ . Functionally, maximum stringency conditions can be used to identify sequences having strict identity or near-strict identity with the hybridization probe; while intermediate or low stringency hybridization can be used to identify or detect polynucleotide sequence homologs.

**[0147]** Moderate and high stringency hybridization conditions are well known in the art. Stringent hybridization conditions are exemplified by hybridization under the following conditions:  $65^\circ \text{C}$ . and  $0.1 \times \text{SSC}$  (where  $1 \times \text{SSC} = 0.15 \text{ M NaCl}$ ,  $0.015 \text{ M Na}_3\text{ citrate}$ ,  $\text{pH } 7.0$ ). Hybridized, duplex nucleic acids are characterized by a melting temperature ( $T_m$ ), where one half of the hybridized nucleic acids are unpaired with the complementary strand. Mismatched nucleic acids within the duplex lower the  $T_m$ . Very stringent hybridization conditions involve  $68^\circ \text{C}$ . and  $0.1 \times \text{SSC}$ . A nucleic acid encoding a variant metalloprotease can have a  $T_m$  reduced by  $1^\circ \text{C}$ .- $3^\circ \text{C}$ . or more compared to a duplex formed between the nucleic acid and its identical complement.

**[0148]** Another example of high stringency conditions includes hybridization at about  $42^\circ \text{C}$ . in 50% formamide,  $5 \times \text{SSC}$ ,  $5 \times \text{Denhardt's solution}$ , 0.5% SDS and  $100 \mu\text{g/ml}$

denatured carrier DNA followed by washing two times in  $2 \times \text{SSC}$  and 0.5% SDS at room temperature and two additional times in  $0.1 \times \text{SSC}$  and 0.5% SDS at  $42^\circ \text{C}$ . An example of moderate stringent conditions include an overnight incubation at  $37^\circ \text{C}$ . in a solution comprising 20% formamide,  $5 \times \text{SSC}$  (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6),  $5 \times \text{Denhardt's solution}$ , 10% dextran sulfate and 20 mg/ml denatured sheared salmon sperm DNA, followed by washing the filters in  $1 \times \text{SSC}$  at about  $37 - 50^\circ \text{C}$ . Those of skill in the art know how to adjust the temperature, ionic strength, etc. to accommodate factors such as probe length and the like.

**[0149]** The term “purified” as applied to nucleic acids or polypeptides generally denotes a nucleic acid or polypeptide that is essentially free from other components as determined by analytical techniques well known in the art (e.g., a purified polypeptide or polynucleotide forms a discrete band in an electrophoretic gel, chromatographic eluate, and/or a media subjected to density gradient centrifugation). For example, a nucleic acid or polypeptide that gives rise to essentially one band in an electrophoretic gel is “purified.” A purified nucleic acid or polypeptide is at least about 50% pure, usually at least about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, about 99.5%, about 99.6%, about 99.7%, about 99.8% or more pure (e.g., percent by weight on a molar basis). In a related sense, the invention provides methods of enriching compositions for one or more molecules of the invention, such as one or more polypeptides or polynucleotides of the invention. A composition is enriched for a molecule when there is a substantial increase in the concentration of the molecule after application of a purification or enrichment technique. A substantially pure polypeptide or polynucleotide of the invention (e.g., substantially pure metalloprotease polypeptide or polynucleotide encoding a metalloprotease polypeptide of the invention, respectively) will typically comprise at least about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98, about 99%, about 99.5% or more by weight (on a molar basis) of all macromolecular species in a particular composition.

**[0150]** The term “enriched” refers to a compound, polypeptide, cell, nucleic acid, amino acid, or other specified material or component that is present in a composition at a relative or absolute concentration that is higher than a starting composition.

**[0151]** In a related sense, the invention provides methods of enriching compositions for one or more molecules of the invention, such as one or more polypeptides of the invention (e.g., one or more metalloprotease polypeptides of the invention) or one or more nucleic acids of the invention (e.g., one or more nucleic acids encoding one or more metalloprotease polypeptides of the invention). A composition is enriched for a molecule when there is a substantial increase in the concentration of the molecule after application of a purification or enrichment technique. A substantially pure polypeptide or polynucleotide will typically comprise at least about 55%, about 60%, about 65%, about 70%, about 75%, about 80%, about 85%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98, about 99%, about 99.5% or more by weight (on a molar basis) of all macromolecular species in a particular composition.

**[0152]** As used herein, the term “functional assay” refers to an assay that provides an indication of a protein’s activity. In some embodiments, the term refers to assay systems in which a protein is analyzed for its ability to function in its usual capacity. For example, in the case of a protease, a functional assay involves determining the effectiveness of the protease to hydrolyze a proteinaceous substrate.

**[0153]** The terms “modified nucleic acid sequence” and “modified gene” are used interchangeably herein to refer to a nucleic acid sequence that includes a deletion, insertion or interruption of naturally occurring (i.e., wild-type) nucleic acid sequence. In some embodiments, the expression product of the modified nucleic acid sequence is a truncated protein (e.g., if the modification is a deletion or interruption of the sequence). In some embodiments, the truncated protein retains biological activity. In alternative embodiments, the expression product of the modified nucleic acid sequence is an elongated protein (e.g., modifications comprising an insertion into the nucleic acid sequence). In some embodiments, a nucleotide insertion in the nucleic acid sequence leads to a truncated protein (e.g., when the insertion results in the formation of a stop codon). Thus, an insertion may result in either a truncated protein or an elongated protein as an expression product.

**[0154]** A “mutant” nucleic acid sequence typically refers to a nucleic acid sequence that has an alteration in at least one codon occurring in a host cell’s wild-type sequence such that the expression product of the mutant nucleic acid sequence is a protein with an altered amino acid sequence relative to the wild-type protein. The expression product may have an altered functional capacity (e.g., enhanced enzymatic activity).

**[0155]** As used herein, the phrase “alteration in substrate specificity” refers to changes in the substrate specificity of an enzyme. In some embodiments, a change in substrate specificity is defined as a change in  $k_{cat}$  and/or  $K_m$  for a particular substrate, resulting from mutations of the enzyme or alteration of reaction conditions. The substrate specificity of an enzyme is determined by comparing the catalytic efficiencies it exhibits with different substrates. These determinations find particular use in assessing the efficiency of mutant enzymes, as it is generally desired to produce variant enzymes that exhibit greater ratios of  $k_{cat}/K_m$  for substrates of interest. However, it is not intended that the present invention be limited to any particular substrate composition or substrate specificity.

**[0156]** As used herein, “surface property” is used in reference to electrostatic charge, as well as properties such as the hydrophobicity and hydrophilicity exhibited by the surface of a protein. As used herein, the term “net charge” is defined as the sum of all charges present in a molecule. “Net charge changes” are made to a parent protein molecule to provide a variant that has a net charge that differs from that of the parent molecule (i.e., the variant has a net charge that is not the same as that of the parent molecule). For example, substitution of a neutral amino acid with a negatively charged amino acid or a positively charged amino acid with a neutral amino acid results in net charge of  $-1$  with respect to the parent molecule. Substitution of a positively charged amino acid with a negatively charged amino acid results in a net charge of  $-2$  with respect to the parent. Substitution of a neutral amino acid with a positively charged amino acid or a negatively charged amino acid with a neutral amino acid results in net charge of  $+1$  with respect to the parent. Substitution of a negatively

charged amino acid with a positively charged amino acid results in a net charge of  $+2$  with respect to the parent. The net charge of a parent protein can also be altered by deletion and/or insertion of charged amino acids. A net charge change applies to changes in charge of a variant versus a parent when measured at the same pH conditions.

**[0157]** The terms “thermally stable” and “thermostable” and “thermostability” refer to proteases that retain a specified amount of enzymatic activity after exposure to identified temperatures over a given period of time under conditions prevailing during the proteolytic, hydrolyzing, cleaning or other process of the invention, while being exposed to altered temperatures. “Altered temperatures” encompass increased or decreased temperatures. In some embodiments, the proteases retain at least about 50%, about 60%, about 70%, about 75%, about 80%, about 85%, about 90%, about 92%, about 95%, about 96%, about 97%, about 98%, or about 99% proteolytic activity after exposure to altered temperatures over a given time period, for example, at least about 60 minutes, about 120 minutes, about 180 minutes, about 240 minutes, about 300 minutes, etc.

**[0158]** The term “enhanced stability” in the context of an oxidation, chelator, thermal, chemical, autolytic and/or pH stable protease refers to a higher retained proteolytic activity over time as compared to other proteases (e.g., thermolysin proteases) and/or wild-type enzymes.

**[0159]** The term “diminished stability” in the context of an oxidation, chelator, thermal and/or pH stable protease refers to a lower retained proteolytic activity over time as compared to other proteases (e.g., thermolysin proteases) and/or wild-type enzymes.

**[0160]** The term “cleaning activity” refers to a cleaning performance achieved by a metalloprotease polypeptide or reference protease under conditions prevailing during the proteolytic, hydrolyzing, cleaning, or other process of the invention. In some embodiments, cleaning performance of a metalloprotease polypeptide or reference protease may be determined by using various assays for cleaning one or more various enzyme sensitive stains on an item or surface (e.g., a stain resulting from food, grass, blood, ink, milk, oil, and/or egg protein). Cleaning performance of a variant or reference protease can be determined by subjecting the stain on the item or surface to standard wash condition(s) and assessing the degree to which the stain is removed by using various chromatographic, spectrophotometric, or other quantitative methodologies. Exemplary cleaning assays and methods are known in the art and include, but are not limited to those described in WO 99/34011 and U.S. Pat. No. 6,605,458, both of which are herein incorporated by reference, as well as those cleaning assays and methods included in the Examples provided below.

**[0161]** The term “cleaning effective amount” of a metalloprotease polypeptide or reference protease refers to the amount of protease that achieves a desired level of enzymatic activity in a specific cleaning composition. Such effective amounts are readily ascertained by one of ordinary skill in the art and are based on many factors, such as the particular protease used, the cleaning application, the specific composition of the cleaning composition, and whether a liquid or dry (e.g., granular, tablet, bar) composition is required, etc.

**[0162]** The term “cleaning adjunct material” refers to any liquid, solid, or gaseous material included in cleaning composition other than a metalloprotease polypeptide of the invention. In some embodiments, the cleaning compositions

of the present invention include one or more cleaning adjunct materials. Each cleaning adjunct material is typically selected depending on the particular type and form of cleaning composition (e.g., liquid, granule, powder, bar, paste, spray, tablet, gel, foam, or other composition). Preferably, each cleaning adjunct material is compatible with the protease enzyme used in the composition.

**[0163]** The term “enhanced performance” in the context of cleaning activity refers to an increased or greater cleaning activity by an enzyme with respect to a parent or reference protein as measured on certain enzyme sensitive stains such as egg, milk, grass, ink, oil, and/or blood, as determined by usual evaluation after a standard wash cycle and/or multiple wash cycles.

**[0164]** The term “diminished performance” in the context of cleaning activity refers to a decreased or lesser cleaning activity by an enzyme on certain enzyme sensitive stains such as egg, milk, grass or blood, as determined by usual evaluation after a standard wash cycle and/or multiple wash cycles.

**[0165]** Cleaning compositions and cleaning formulations include any composition that is suited for cleaning, bleaching, disinfecting, and/or sterilizing any object, item, and/or surface. Such compositions and formulations include, but are not limited to for example, liquid and/or solid compositions, including cleaning or detergent compositions (e.g., liquid, tablet, gel, bar, granule, and/or solid laundry cleaning or detergent compositions and fine fabric detergent compositions; hard surface cleaning compositions and formulations, such as for glass, wood, ceramic and metal counter tops and windows; carpet cleaners; oven cleaners; fabric fresheners; fabric softeners; and textile, laundry booster cleaning or detergent compositions, laundry additive cleaning compositions, and laundry pre-spotter cleaning compositions; dishwashing compositions, including hand or manual dishwash compositions (e.g., “hand” or “manual” dishwashing detergents) and automatic dishwashing compositions (e.g., “automatic dishwashing detergents”).

**[0166]** Cleaning composition or cleaning formulations, as used herein, include, unless otherwise indicated, granular or powder-form all-purpose or heavy-duty washing agents, especially cleaning detergents; liquid, granular, gel, solid, tablet, or paste-form all-purpose washing agents, especially the so-called heavy-duty liquid (HDL) detergent or heavy-duty powder detergent (HDD) types; liquid fine-fabric detergents; hand or manual dishwashing agents, including those of the high-foaming type; hand or manual dishwashing, automatic dishwashing, or dishware or tableware washing agents, including the various tablet, powder, solid, granular, liquid, gel, and rinse-aid types for household and institutional use; liquid cleaning and disinfecting agents, including antibacterial hand-wash types, cleaning bars, mouthwashes, denture cleaners, car shampoos, carpet shampoos, bathroom cleaners; hair shampoos and/or hair-rinses for humans and other animals; shower gels and foam baths and metal cleaners; as well as cleaning auxiliaries, such as bleach additives and “stain-stick” or pre-treat types. In some embodiments, granular compositions are in “compact” form; in some embodiments, liquid compositions are in a “concentrated” form.

**[0167]** As used herein, “fabric cleaning compositions” include hand and machine laundry detergent compositions including laundry additive compositions and compositions suitable for use in the soaking and/or pretreatment of stained fabrics (e.g., clothes, linens, and other textile materials).

**[0168]** As used herein, “non-fabric cleaning compositions” include non-textile (i.e., non-fabric) surface cleaning compositions, including, but not limited to for example, hand or manual or automatic dishwashing detergent compositions, oral cleaning compositions, denture cleaning compositions, and personal cleansing compositions.

**[0169]** As used herein, the term “fabric and/or hard surface cleaning and/or treatment composition” is a subset of cleaning and treatment compositions that includes, unless otherwise indicated, granular or powder-form all-purpose or “heavy-duty” washing agents, especially cleaning detergents; liquid, gel or paste-form all-purpose washing agents, especially the so-called heavy-duty liquid types; liquid fine-fabric detergents; hand dishwashing agents or light duty dishwashing agents, especially those of the high-foaming type; machine dishwashing agents, including the various tablet, granular, liquid and rinse-aid types for household and institutional use; liquid cleaning and disinfecting agents, car or carpet shampoos, bathroom cleaners including toilet bowl cleaners; fabric conditioning products including softening and/or freshening that may be in liquid, solid and/or dryer sheet form; as well as cleaning auxiliaries such as bleach additives and “stain-stick” or pre-treat types, substrate-laden products such as dryer added sheets. All of such products which are applicable may be in standard, concentrated or even highly concentrated form even to the extent that such products may in certain aspect be non-aqueous.

**[0170]** As used herein, the term “detergent composition” or “detergent formulation” is used in reference to a composition intended for use in a wash medium for the cleaning of soiled or dirty objects, including particular fabric and/or non-fabric objects or items. Such compositions of the present invention are not limited to any particular detergent composition or formulation. Indeed, in some embodiments, the detergents of the invention comprise at least one metalloprotease polypeptide of the invention and, in addition, one or more surfactants, transferase(s), hydrolytic enzymes, oxido reductases, builders (e.g., a builder salt), bleaching agents, bleach activators, bluing agents, fluorescent dyes, caking inhibitors, masking agents, enzyme activators, antioxidants, and/or solubilizers. In some instances, a builder salt is a mixture of a silicate salt and a phosphate salt, preferably with more silicate (e.g., sodium metasilicate) than phosphate (e.g., sodium tripolyphosphate). Some compositions of the invention, such as, but not limited to, cleaning compositions or detergent compositions, do not contain any phosphate (e.g., phosphate salt or phosphate builder).

**[0171]** As used herein, the term “bleaching” refers to the treatment of a material (e.g., fabric, laundry, pulp, etc.) or surface for a sufficient length of time and/or under appropriate pH and/or temperature conditions to effect a brightening (i.e., whitening) and/or cleaning of the material. Examples of chemicals suitable for bleaching include, but are not limited to, for example,  $\text{ClO}_2$ ,  $\text{H}_2\text{O}_2$ , peracids,  $\text{NO}_2$ , etc.

**[0172]** As used herein, “wash performance” of a protease (e.g., a metalloprotease polypeptide of the invention) refers to the contribution of a metalloprotease polypeptide to washing that provides additional cleaning performance to the detergent as compared to the detergent without the addition of the metalloprotease polypeptide to the composition. Wash performance is compared under relevant washing conditions. In some test systems, other relevant factors, such as detergent composition, sud concentration, water hardness, washing mechanics, time, pH, and/or temperature, can be controlled in

such a way that condition(s) typical for household application in a certain market segment (e.g., hand or manual dishwashing, automatic dishwashing, dishware cleaning, tableware cleaning, fabric cleaning, etc.) are imitated.

**[0173]** The term “relevant washing conditions” is used herein to indicate the conditions, particularly washing temperature, time, washing mechanics, sud concentration, type of detergent and water hardness, actually used in households in a hand dishwashing, automatic dishwashing, or laundry detergent market segment.

**[0174]** The term “improved wash performance” is used to indicate that a better end result is obtained in stain removal under relevant washing conditions, or that less metalloprotease polypeptide, on weight basis, is needed to obtain the same end result relative to the corresponding wild-type or starting parent protease.

**[0175]** As used herein, the term “disinfecting” refers to the removal of contaminants from the surfaces, as well as the inhibition or killing of microbes on the surfaces of items. It is not intended that the present invention be limited to any particular surface, item, or contaminant(s) or microbes to be removed.

**[0176]** The “compact” form of the cleaning compositions herein is best reflected by density and, in terms of composition, by the amount of inorganic filler salt. Inorganic filler salts are conventional ingredients of detergent compositions in powder form. In conventional detergent compositions, the filler salts are present in substantial amounts, typically about 17 to about 35% by weight of the total composition. In contrast, in compact compositions, the filler salt is present in amounts not exceeding about 15% of the total composition. In some embodiments, the filler salt is present in amounts that do not exceed about 10%, or more preferably, about 5%, by weight of the composition. In some embodiments, the inorganic filler salts are selected from the alkali and alkaline-earth-metal salts of sulfates and chlorides. In some embodiments, the filler salt is sodium sulfate.

**[0177]** As used herein in connection with a numerical value, the term “about” refers to a range of  $\pm 0.5$  of the numerical value, unless the term is otherwise specifically defined in context. For instance, the phrase a “pH value of about 6” refers to pH values of from 5.5 to 6.5, unless the pH value is specifically defined otherwise.

**[0178]** The position of an amino acid residue in a given amino acid sequence is typically numbered herein using the numbering of the position of the corresponding amino acid residue of the wild type *Paenibacillus* metalloprotease amino acid sequences shown in SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38. The *Paenibacillus* sp. metalloprotease amino acid sequences, thus serves as a reference parent sequence. A given amino acid sequence, such as a metalloprotease enzyme amino acid sequence and variants thereof described herein, can be aligned with the wild type metalloprotease sequence (e.g., SEQ ID NO: 3) using an alignment algorithm as described herein, and an amino acid residue in the given amino acid sequence that aligns (preferably optimally aligns) with an amino acid residue in the wild type sequence can be conveniently numbered by reference to the corresponding amino acid residue in the metalloprotease sequence.

**[0179]** Oligonucleotide synthesis and purification steps are typically performed according to specifications. Techniques and procedures are generally performed according to conventional methods well known in the art and various general references that are provided throughout this document. Pro-

cedures therein are believed to be well known to those of ordinary skill in the art and are provided for the convenience of the reader.

#### Metalloprotease Polypeptides of the Present Invention

**[0180]** The present invention provides novel metalloprotease enzyme polypeptides, which may be collectively referred to as “enzymes of the invention” or “polypeptides of the invention.” Polypeptides of the invention include isolated, recombinant, substantially pure, or non-naturally occurring polypeptides. In some embodiments, polypeptides of the invention are useful in cleaning applications and can be incorporated into cleaning compositions that are useful in methods of cleaning an item or a surface in need of cleaning.

**[0181]** In some embodiments, the enzyme of the present invention has 50, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 or 100% identity to SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38. In various embodiments, the enzyme of the present invention has 50, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 or 100% identity to a metalloprotease enzyme from any genus in Tables 1.2, 2.2, 3.2, 4.2, 5.2, 6.2, 7.2 or 8.2.

**[0182]** In some embodiments, the enzyme of the present invention, including all embodiments supra, can be derived from a member of the order Bacillales or family Bacillaceae, Paenibacillaceae, Alicyclobacillaceae, or Lactobacillaceae. In some embodiments, the enzyme of the present invention, including all embodiments supra, can be derived from a *Bacillus*, *Alicyclobacillus*, *Geobacillus*, *Exiguobacterium*, *Lactobacillus*, or *Paenibacillus* species. In some embodiments, the enzyme of the present invention, including all embodiments supra, can be derived from a member of the Pseudococcidae family. In some embodiments, the enzyme of the present invention, including all embodiments supra, can be derived from a *Planococcus* species. Various enzyme metalloproteases have been found that have a high identity to each other and to the *Paenibacillus* enzymes as shown in SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38.

**[0183]** In a particular embodiment, the invention is an enzyme derived from the genus *Paenibacillus*. In a particular embodiment, the invention is an enzyme derived from the genus *Paenibacillus* and from the species *Paenibacillus* sp., *Paenibacillus ehimensis*, *Paenibacillus humanensis*, *Paenibacillus barcinonensis*, *Paenibacillus amylolyticus*, *Paenibacillus humicus* and *Paenibacillus polymyxa*.

**[0184]** Described are compositions and methods relating to enzymes cloned from *Paenibacillus*. The compositions and methods are based, in part, on the observation that cloned and expressed enzymes of the present invention have proteolytic activity in the presence of a detergent composition. Enzymes of the present invention also demonstrate excellent stability in detergent compositions. These features makes enzymes of the present invention well suited for use in a variety of cleaning applications, where the enzyme can hydrolyze proteins in the presence of surfactants and other components found in detergent compositions.

**[0185]** In some embodiments, the invention includes an isolated, recombinant, substantially pure, or non-naturally occurring enzyme having protease activity, which polypeptide comprises a polypeptide sequence having at least about 60%, 65%, 70%, 75%, 80%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% sequence identity to a parent enzyme as provided herein.

**[0186]** In some embodiments, the polypeptide of the present invention, is a polypeptide having a specified degree of amino acid sequence homology to the exemplified polypeptides, e.g., at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or even at least 99% sequence homology to the amino acid sequences of SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38. Homology can be determined by amino acid sequence alignment, e.g., using a program such as BLAST, ALIGN, or CLUSTAL, as described herein.

**[0187]** Also provided are polypeptide enzymes of the present invention, having protease activity, said enzymes comprising an amino acid sequence which differs from the amino acid sequence of SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38 by no more than 50, no more than 40, no more than 30, no more than 35, no more than 25, no more than 20, no more than 19, no more than 18, no more than 17, no more than 16, no more than 15, no more than 14, no more than 13, no more than 12, no more than 11, no more than 10, no more than 9, no more than 8, no more than 7, no more than 6, no more than 5, no more than 4, no more than 3, no more than 2, or no more than 1 amino acid residue(s), when aligned using any of the previously described alignment methods.

**[0188]** As noted above, the variant enzyme polypeptides of the invention have enzymatic activities (e.g., protease activities) and thus are useful in cleaning applications, including but not limited to, methods for cleaning dishware items, tableware items, fabrics, and items having hard surfaces (e.g., the hard surface of a table, table top, wall, furniture item, floor, ceiling, etc.). Exemplary cleaning compositions comprising one or more variant metalloprotease enzyme polypeptides of the invention are described infra. The enzymatic activity (e.g., protease enzyme activity) of an enzyme polypeptide of the invention can be determined readily using procedures well known to those of ordinary skill in the art. The Examples presented infra describe methods for evaluating the enzymatic activity and cleaning performance. The performance of polypeptide enzymes of the invention in removing stains (e.g., a protein stain such as blood/milk/ink or egg yolk), cleaning hard surfaces, or cleaning laundry, dishware or tableware item(s) can be readily determined using procedures well known in the art and/or by using procedures set forth in the Examples.

**[0189]** The metalloprotease polypeptides of the invention have protease activity such that they are useful in casein hydrolysis, collagen hydrolysis, elastin hydrolysis, keratin hydrolysis, soy protein hydrolysis or corn meal protein hydrolysis. Thus, the polypeptides of the invention find use in other applications such as pretreatments for food, feed, or protein degradation.

**[0190]** The polypeptides of the invention are also useful in pretreatment of animal feed products, such as soy protein, corn meal, and other protein rich components. Pretreatment of these animal feed products with a polypeptide of the invention may help in the breakdown of complex proteins into their hydrolysates which are easily digestible by animals.

**[0191]** In yet other embodiments, the disclosed metalloprotease polypeptides find use in hydrolysis of corn soy protein. The disclosed metalloprotease polypeptides may be used alone or in combination with other proteases, amylases or lipases to produce peptides and free amino acids from the corn or soy protein. In some embodiments, the recovered

proteins, peptides or amino acids can be subsequently used in animal feed or human food products.

**[0192]** The polypeptides of the invention are also useful in treatment of wounds, particularly in wound debridement. Wound debridement is the removal of dead, damaged or infected tissue to improve the healing potential of the remaining healthy tissue. Debridement is an important part of the healing process for burns and other serious wounds. The wounds or burns may be treated with a composition comprising a polypeptide of the invention which would result in removal of unwanted damaged tissue and improving the healthy tissue.

The metalloprotease polypeptides of the present invention can have protease activity over a broad range of pH conditions. In some embodiments, the metalloprotease polypeptides have protease activity on azo-casein as a substrate, as demonstrated in Examples 3.1 to 3.8. In some embodiments, the metalloprotease polypeptides have protease activity at a pH of from about 3.0 to about 12.0. In some embodiments, the metalloprotease polypeptides have protease activity at a pH of from about 4.0 to about 10.5. In some embodiments, the metalloprotease polypeptides have at least 70% of maximal protease activity at a pH of from about 5.5 to about 9.0. In some embodiments, the metalloprotease polypeptides have at least 80% of maximal protease activity at a pH of from about 6.0 to about 8.5. In some embodiments, the metalloprotease polypeptides have maximal protease activity at a pH of about 7.5.

**[0193]** In some embodiments, the metalloprotease polypeptides of the present invention have protease activity at a temperature range of from about 10° C. to about 100° C. In some embodiments, the metalloprotease polypeptides of the present invention have protease activity at a temperature range of from about 20° C. to about 90° C. In some embodiments, the metalloprotease polypeptides have at least 70% of maximal protease activity at a temperature of from about 45° C. to about 60° C. In some embodiments, the metalloprotease polypeptides have maximal protease activity at a temperature of 50° C.

**[0194]** In some embodiments, the metalloprotease polypeptides of the present invention demonstrate cleaning performance in a cleaning composition. Cleaning compositions often include ingredients harmful to the stability and performance of enzymes, making cleaning compositions a harsh environment for enzymes, e.g. metalloproteases, to retain function. Thus, it is not trivial for an enzyme to be put in a cleaning composition and expect enzymatic function (e.g. metalloprotease activity, such as demonstrated by cleaning performance). In some embodiments, the metalloprotease polypeptides of the present invention demonstrate cleaning performance in automatic dishwashing (ADW) detergent compositions. In some embodiments, the cleaning performance in automatic dishwashing (ADW) detergent compositions includes cleaning of egg yolk stains. In some embodiments, the metalloprotease polypeptides of the present invention demonstrate cleaning performance in laundry detergent compositions. In some embodiments, the cleaning performance in laundry detergent compositions includes cleaning of blood/milk/ink stains. In each of the cleaning compositions, the metalloprotease polypeptides of the present invention demonstrate cleaning performance with or without a bleach component.

**[0195]** The metalloprotease polypeptides of the invention have protease activity such that they are useful in casein

hydrolysis, collagen hydrolysis, elastin hydrolysis, keratin hydrolysis, soy protein hydrolysis or corn meal protein hydrolysis. Thus, the polypeptides of the invention find use in other applications such as pretreatments for food, feed, or protein degradation.

**[0196]** A polypeptide of the invention can be subject to various changes, such as one or more amino acid insertions, deletions, and/or substitutions, either conservative or non-conservative, including where such changes do not substantially alter the enzymatic activity of the polypeptide. Similarly, a nucleic acid of the invention can also be subject to various changes, such as one or more substitutions of one or more nucleotides in one or more codons such that a particular codon encodes the same or a different amino acid, resulting in either a silent variation (e.g., when the encoded amino acid is not altered by the nucleotide mutation) or non-silent variation, one or more deletions of one or more nucleic acids (or codons) in the sequence, one or more additions or insertions of one or more nucleic acids (or codons) in the sequence, and/or cleavage of or one or more truncations of one or more nucleic acids (or codons) in the sequence. Many such changes in the nucleic acid sequence may not substantially alter the enzymatic activity of the resulting encoded polypeptide enzyme compared to the polypeptide enzyme encoded by the original nucleic acid sequence. A nucleic acid sequence of the invention can also be modified to include one or more codons that provide for optimum expression in an expression system (e.g., bacterial expression system), while, if desired, said one or more codons still encode the same amino acid(s).

**[0197]** In some embodiments, the present invention provides a genus of enzyme polypeptides having the desired enzymatic activity (e.g., protease enzyme activity or cleaning performance activity) which comprise sequences having the amino acid substitutions described herein and also which comprise one or more additional amino acid substitutions, such as conservative and non-conservative substitutions, wherein the polypeptide exhibits, maintains, or approximately maintains the desired enzymatic activity (e.g., proteolytic activity, as reflected in the cleaning activity or performance of the polypeptide enzymes of SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 and 38). Amino acid substitutions in accordance with the invention may include, but are not limited to, one or more non-conservative substitutions and/or one or more conservative amino acid substitutions. A conservative amino acid residue substitution typically involves exchanging a member within one functional class of amino acid residues for a residue that belongs to the same functional class (conservative amino acid residues are considered functionally homologous or conserved in calculating percent functional homology). A conservative amino acid substitution typically involves the substitution of an amino acid in an amino acid sequence with a functionally similar amino acid. For example, alanine, glycine, serine, and threonine are functionally similar and thus may serve as conservative amino acid substitutions for one another. Aspartic acid and glutamic acid may serve as conservative substitutions for one another. Asparagine and glutamine may serve as conservative substitutions for one another. Arginine, lysine, and histidine may serve as conservative substitutions for one another. Isoleucine, leucine, methionine, and valine may serve as conservative substitutions for one another. Phenylalanine, tyrosine, and tryptophan may serve as conservative substitutions for one another.

**[0198]** Other conservative amino acid substitution groups can be envisioned. For example, amino acids can be grouped by similar function or chemical structure or composition (e.g., acidic, basic, aliphatic, aromatic, sulfur-containing). For instance, an aliphatic grouping may comprise: Glycine (G), Alanine (A), Valine (V), Leucine (L), Isoleucine (I). Other groups containing amino acids that are considered conservative substitutions for one another include: aromatic: Phenylalanine (F), Tyrosine (Y), Tryptophan (W); sulfur-containing: Methionine (M), Cysteine (C); Basic: Arginine (R), Lysine (K), Histidine (H); Acidic: Aspartic acid (D), Glutamic acid (E); non-polar uncharged residues, Cysteine (C), Methionine (M), and Proline (P); hydrophilic uncharged residues: Serine (S), Threonine (T), Asparagine (N), and Glutamine (Q). Additional groupings of amino acids are well-known to those of skill in the art and described in various standard textbooks. Listing of a polypeptide sequence herein, in conjunction with the above substitution groups, provides an express listing of all conservatively substituted polypeptide sequences.

**[0199]** More conservative substitutions exist within the amino acid residue classes described above, which also or alternatively can be suitable. Conservation groups for substitutions that are more conservative include: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, and asparagine-glutamine.

**[0200]** Conservatively substituted variations of a polypeptide sequence of the invention (e.g., variant metalloproteases of the invention) include substitutions of a small percentage, sometimes less than 25%, 20%, 15%, 14%, 13%, 12%, 11%, 10%, 9%, 8%, 7%, or 6% of the amino acids of the polypeptide sequence, or less than 5%, 4%, 3%, 2%, or 1%, or less than 10, 9, 8, 7, 6, 5, 4, 3, 2, or 1 amino acid substitution of the amino acids of the polypeptide sequence, with a conservatively selected amino acid of the same conservative substitution group.

**[0201]** As described elsewhere herein in greater detail and in the Examples provided herein, polypeptides of the invention may have cleaning abilities that may be compared to known proteases, including known metalloproteases.

#### Nucleic Acids of the Invention

**[0202]** The invention provides isolated, non-naturally occurring, or recombinant nucleic acids which may be collectively referred to as "nucleic acids of the invention" or "polynucleotides of the invention", which encode polypeptides of the invention. Nucleic acids of the invention, including all described below, are useful in recombinant production (e.g., expression) of polypeptides of the invention, typically through expression of a plasmid expression vector comprising a sequence encoding the polypeptide of interest or fragment thereof. As discussed above, polypeptides include metalloprotease polypeptides having enzymatic activity (e.g., proteolytic activity) which are useful in cleaning applications and cleaning compositions for cleaning an item or a surface (e.g., surface of an item) in need of cleaning.

**[0203]** In some embodiments, the invention provides an isolated, recombinant, substantially pure, or non-naturally occurring nucleic acid comprising a nucleotide sequence encoding any polypeptide (including any fusion protein, etc.) of the invention described above in the section entitled "Polypeptides of the Invention" and elsewhere herein. The invention also provides an isolated, recombinant, substantially pure, or non-naturally-occurring nucleic acid compris-



ing a nucleotide sequence encoding a combination of two or more of any polypeptides of the invention described above and elsewhere herein. In some embodiments, the nucleic acids of the present invention has 50, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99 or 100% identity to SEQ ID NO: 4, 9, 14, 19, 24, 29, 34 and 39.

**[0204]** The present invention provides nucleic acids encoding a metalloprotease polypeptide of the present invention, wherein the metalloprotease polypeptide is a mature form having proteolytic activity, wherein the amino acid positions of the variant are numbered by correspondence with the amino acid sequence of *Paenibacillus* metalloprotease polypeptides set forth as SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 or 38.

**[0205]** Nucleic acids of the invention can be generated by using any suitable synthesis, manipulation, and/or isolation techniques, or combinations thereof. For example, a polynucleotide of the invention may be produced using standard nucleic acid synthesis techniques, such as solid-phase synthesis techniques that are well-known to those skilled in the art. In such techniques, fragments of up to 50 or more nucleotide bases are typically synthesized, then joined (e.g., by enzymatic or chemical ligation methods) to form essentially any desired continuous nucleic acid sequence. The synthesis of the nucleic acids of the invention can be also facilitated by any suitable method known in the art, including but not limited to chemical synthesis using the classical phosphoramidite method (See e.g., Beaucage et al. *Tetrahedron Letters* 22:1859-69 [1981]); or the method described by Matthes et al. (See, Matthes et al., *EMBO J.* 3:801-805 [1984], as is typically practiced in automated synthetic methods. Nucleic acids of the invention also can be produced by using an automatic DNA synthesizer. Customized nucleic acids can be ordered from a variety of commercial sources (e.g., The Midland Certified Reagent Company, the Great American Gene Company, Operon Technologies Inc., and DNA2.0). Other techniques for synthesizing nucleic acids and related principles are known in the art (See e.g., Itakura et al., *Ann Rev. Biochem.* 53:323 [1984]; and Itakura et al., *Science* 198:1056 [1984]).

**[0206]** As indicated above, recombinant DNA techniques useful in modification of nucleic acids are well known in the art. For example, techniques such as restriction endonuclease digestion, ligation, reverse transcription and cDNA production, and polymerase chain reaction (e.g., PCR) are known and readily employed by those of skill in the art. Nucleotides of the invention may also be obtained by screening cDNA libraries using one or more oligonucleotide probes that can hybridize to or PCR-amplify polynucleotides which encode a metalloprotease polypeptide(s) of the invention. Procedures for screening and isolating cDNA clones and PCR amplification procedures are well known to those of skill in the art and described in standard references known to those skilled in the art. Some nucleic acids of the invention can be obtained by altering a naturally occurring polynucleotide backbone (e.g., that encodes an enzyme or parent protease) by, for example, a known mutagenesis procedure (e.g., site-directed mutagenesis, site saturation mutagenesis, and in vitro recombination).

Methods for Making Modified Metalloprotease polypeptides of the Invention

**[0207]** A variety of methods are known in the art that are suitable for generating modified polynucleotides of the invention that encode metalloprotease polypeptides of the inven-

tion, including, but not limited to, for example, site-saturation mutagenesis, scanning mutagenesis, insertional mutagenesis, deletion mutagenesis, random mutagenesis, site-directed mutagenesis, and directed-evolution, as well as various other recombinatorial approaches. Methods for making modified polynucleotides and proteins (e.g., metalloprotease polypeptides) include DNA shuffling methodologies, methods based on non-homologous recombination of genes, such as ITCHY (See, Ostermeier et al., 7:2139-44 [1999]), SCRACHY (See, Lutz et al. 98:11248-53 [2001]), SHIPREC (See, Sieber et al., 19:456-60 [2001]), and NRR (See, Bittker et al., 20:1024-9 [2001]; Bittker et al., 101:7011-6 [2004]), and methods that rely on the use of oligonucleotides to insert random and targeted mutations, deletions and/or insertions (See, Ness et al., 20:1251-5 [2002]; Coco et al., 20:1246-50 [2002]; Zha et al., 4:34-9 [2003]; Glaser et al., 149:3903-13 [1992]).

Vectors, Cells, and Methods for Producing Metalloprotease Polypeptides of the Invention

**[0208]** The present invention provides vectors comprising at least one metalloprotease polynucleotide of the invention described herein (e.g., a polynucleotide encoding a metalloprotease polypeptide of the invention described herein), expression vectors or expression cassettes comprising at least one nucleic acid or polynucleotide of the invention, isolated, substantially pure, or recombinant DNA constructs comprising at least one nucleic acid or polynucleotide of the invention, isolated or recombinant cells comprising at least one polynucleotide of the invention, and compositions comprising one or more such vectors, nucleic acids, expression vectors, expression cassettes, DNA constructs, cells, cell cultures, or any combination or mixtures thereof.

**[0209]** In some embodiments, the invention provides recombinant cells comprising at least one vector (e.g., expression vector or DNA construct) of the invention which comprises at least one nucleic acid or polynucleotide of the invention. Some such recombinant cells are transformed or transfected with such at least one vector. Such cells are typically referred to as host cells. Some such cells comprise bacterial cells, including, but are not limited to *Bacillus* sp. cells, such as *B. subtilis* cells. The invention also provides recombinant cells (e.g., recombinant host cells) comprising at least one metalloprotease polypeptide of the invention.

**[0210]** In some embodiments, the invention provides a vector comprising a nucleic acid or polynucleotide of the invention. In some embodiments, the vector is an expression vector or expression cassette in which a polynucleotide sequence of the invention which encodes a metalloprotease polypeptide of the invention is operably linked to one or additional nucleic acid segments required for efficient gene expression (e.g., a promoter operably linked to the polynucleotide of the invention which encodes a metalloprotease polypeptide of the invention). A vector may include a transcription terminator and/or a selection gene, such as an antibiotic resistance gene, that enables continuous cultural maintenance of plasmid-infected host cells by growth in antimicrobial-containing media.

**[0211]** An expression vector may be derived from plasmid or viral DNA, or in alternative embodiments, contains elements of both. Exemplary vectors include, but are not limited to pC194, pJH101, pE194, pHP13 (See, Harwood and Cutting [eds.], Chapter 3, *Molecular Biological Methods for Bacillus*, John Wiley & Sons [1990]; suitable replicating plasmids for *B. subtilis* include those listed on p. 92) See also,

Perego, Integrational Vectors for Genetic Manipulations in *Bacillus subtilis*, in Sonenshein et al., [eds.] *Bacillus subtilis* and Other Gram-Positive Bacteria: Biochemistry, Physiology and Molecular Genetics, American Society for Microbiology, Washington, D.C. [1993], pp. 615-624), and p2JM103BBI.

**[0212]** For expression and production of a protein of interest (e.g., metalloprotease polypeptide) in a cell, at least one expression vector comprising at least one copy of a polynucleotide encoding the metalloprotease polypeptide, and in some instances comprising multiple copies, is transformed into the cell under conditions suitable for expression of the metalloprotease. In some embodiments of the present invention, a polynucleotide sequence encoding the metalloprotease polypeptide (as well as other sequences included in the vector) is integrated into the genome of the host cell, while in other embodiments, a plasmid vector comprising a polynucleotide sequence encoding the metalloprotease polypeptide remains as autonomous extra-chromosomal element within the cell. The invention provides both extrachromosomal nucleic acid elements as well as incoming nucleotide sequences that are integrated into the host cell genome. The vectors described herein are useful for production of the metalloprotease polypeptides of the invention. In some embodiments, a polynucleotide construct encoding the metalloprotease polypeptide is present on an integrating vector that enables the integration and optionally the amplification of the polynucleotide encoding the metalloprotease polypeptide into the host chromosome. Examples of sites for integration are well known to those skilled in the art. In some embodiments, transcription of a polynucleotide encoding a metalloprotease polypeptide of the invention is effectuated by a promoter that is the wild-type promoter for the selected precursor protease. In some other embodiments, the promoter is heterologous to the precursor protease, but is functional in the host cell. Specifically, examples of suitable promoters for use in bacterial host cells include, but are not limited to, for example, the amyE, amyQ, amyL, pstS, sacB, pSPAC, pAprE, pVeg, pHpaII promoters, the promoter of the *B. stearothermophilus* maltogenic amylase gene, the *B. amyloliquefaciens* (BAN) amylase gene, the *B. subtilis* alkaline protease gene, the *B. clausii* alkaline protease gene the *B. pumilis* xylosidase gene, the *B. thuringiensis* cryIIIA, and the *B. licheniformis* alpha-amylase gene. Additional promoters include, but are not limited to the A4 promoter, as well as phage Lambda PR or PL promoters, and the *E. coli* lac, trp or tac promoters.

**[0213]** Metalloprotease polypeptides of the present invention can be produced in host cells of any suitable microorganism, including bacteria and fungi. In some embodiments, metalloprotease polypeptides of the present invention can be produced in Gram-positive bacteria. In some embodiments, the host cells are *Bacillus* spp., *Streptomyces* spp., *Escherichia* spp., *Aspergillus* spp., *Trichoderma* spp., *Pseudomonas* spp., *Corynebacterium* spp., *Saccharomyces* spp., or *Pichia* spp. In some embodiments, the metalloprotease polypeptides are produced by *Bacillus* sp. host cells. Examples of *Bacillus* sp. host cells that find use in the production of the metalloprotease polypeptides of the invention include, but are not limited to *B. licheniformis*, *B. lentus*, *B. subtilis*, *B. amyloliquefaciens*, *B. lentus*, *B. brevis*, *B. stearothermophilus*, *B. alkalophilus*, *B. coagulans*, *B. circulans*, *B. pumilis*, *B. thuringiensis*, *B. clausii*, and *B. megaterium*, as well as other organisms within the genus *Bacillus*. In some embodiments, *B. subtilis* host cells are used for production of metallopro-

tease polypeptides. U.S. Pat. Nos. 5,264,366 and 4,760,025 (RE 34,606) describe various *Bacillus* host strains that can be used for producing metalloprotease polypeptide of the invention, although other suitable strains can be used.

**[0214]** Several bacterial strains that can be used to produce metalloprotease polypeptides of the invention include non-recombinant (i.e., wild-type) *Bacillus* sp. strains, as well as variants of naturally-occurring strains and/or recombinant strains. In some embodiments, the host strain is a recombinant strain, wherein a polynucleotide encoding a polypeptide of interest has been introduced into the host. In some embodiments, the host strain is a *B. subtilis* host strain and particularly a recombinant *Bacillus subtilis* host strain. Numerous *B. subtilis* strains are known, including, but not limited to for example, 1A6 (ATCC 39085), 168 (1A01), SB19, W23, Ts85, B637, PB1753 through PB1758, PB3360, JH642, 1A243 (ATCC 39,087), ATCC 21332, ATCC 6051, MI113, DE100 (ATCC 39,094), GX4931, PBT 110, and PEP 211 strain (See e.g., Hoch et al., Genetics 73:215-228 [1973]; See also, U.S. Pat. Nos. 4,450,235 and 4,302,544, and EP 0134048, each of which is incorporated by reference in its entirety). The use of *B. subtilis* as an expression host cells is well known in the art (See e.g., Palva et al., Gene 19:81-87 [1982]; Fahnestock and Fischer, J. Bacteriol., 165:796-804 [1986]; and Wang et al., Gene 69:39-47 [1988]).

**[0215]** In some embodiments, the *Bacillus* host cell is a *Bacillus* sp. that includes a mutation or deletion in at least one of the following genes, degU, degS, degR and degQ. In some embodiments, the mutation is in a degU gene, and in some embodiments the mutation is degU(Hy)32 (See e.g., Msadek et al., J. Bacteriol. 172:824-834 [1990]; and Olmos et al., Mol. Gen. Genet. 253:562-567 [1997]). In some embodiments, the *Bacillus* host comprises a mutation or deletion in scoC4 (See e.g., Caldwell et al., J. Bacteriol. 183:7329-7340 [2001]); spoIIIE (See e.g., Arigoni et al., Mol. Microbiol. 31:1407-1415 [1999]); and/or oppA or other genes of the opp operon (See e.g., Perego et al., Mol. Microbiol. 5:173-185 [1991]). Indeed, it is contemplated that any mutation in the opp operon that causes the same phenotype as a mutation in the oppA gene will find use in some embodiments of the altered *Bacillus* strain of the invention. In some embodiments, these mutations occur alone, while in other embodiments, combinations of mutations are present. In some embodiments, an altered *Bacillus* host cell strain that can be used to produce a metalloprotease polypeptide of the invention is a *Bacillus* host strain that already includes a mutation in one or more of the above-mentioned genes. In addition, *Bacillus* sp. host cells that comprise mutation(s) and/or deletions of endogenous protease genes find use. In some embodiments, the *Bacillus* host cell comprises a deletion of the aprE and the nprE genes. In other embodiments, the *Bacillus* sp. host cell comprises a deletion of 5 protease genes, while in other embodiments, the *Bacillus* sp. host cell comprises a deletion of 9 protease genes (See e.g., U.S. Pat. Appln. Pub. No. 2005/0202535, incorporated herein by reference).

**[0216]** Host cells are transformed with at least one nucleic acid encoding at least one metalloprotease polypeptide of the invention using any suitable method known in the art. Methods for introducing a nucleic acid (e.g., DNA) into *Bacillus* cells or *E. coli* cells utilizing plasmid DNA constructs or vectors and transforming such plasmid DNA constructs or vectors into such cells are well known. In some embodiments, the plasmids are subsequently isolated from *E. coli* cells and transformed into *Bacillus* cells. However, it is not essential to

use intervening microorganisms such as *E. coli*, and in some embodiments, a DNA construct or vector is directly introduced into a *Bacillus* host.

**[0217]** Those of skill in the art are well aware of suitable methods for introducing nucleic acid sequences of the invention into *Bacillus* cells (See e.g., Ferrari et al., "Genetics," in Harwood et al. [eds.], *Bacillus*, Plenum Publishing Corp. [1989], pp. 57-72; Saunders et al., *J. Bacteriol.* 157:718-726 [1984]; Hoch et al., *J. Bacteriol.* 93:1925-1937 [1967]; Mann et al., *Current Microbiol.* 13:131-135 [1986]; Holubova, *Folia Microbiol.* 30:97 [1985]; Chang et al., *Mol. Gen. Genet.* 168:11-115 [1979]; Vorobjeva et al., *FEMS Microbiol. Lett.* 7:261-263 [1980]; Smith et al., *Appl. Env. Microbiol.* 51:634 [1986]; Fisher et al., *Arch. Microbiol.* 139:213-217 [1981]; and McDonald, *J. Gen. Microbiol.* 130:203 [1984]). Indeed, such methods as transformation, including protoplast transformation and transfection, transduction, and protoplast fusion are well known and suited for use in the present invention. Methods known in the art to transform *Bacillus* cells include such methods as plasmid marker rescue transformation, which involves the uptake of a donor plasmid by competent cells carrying a partially homologous resident plasmid (See, Contente et al., *Plasmid* 2:555-571 [1979]; Haima et al., *Mol. Gen. Genet.* 223:185-191 [1990]; Weinrauch et al., *J. Bacteriol.* 154:1077-1087 [1983]; and Weinrauch et al., *J. Bacteriol.* 169:1205-1211 [1987]). In this method, the incoming donor plasmid recombines with the homologous region of the resident "helper" plasmid in a process that mimics chromosomal transformation.

**[0218]** In addition to commonly used methods, in some embodiments, host cells are directly transformed with a DNA construct or vector comprising a nucleic acid encoding a metalloprotease polypeptide of the invention (i.e., an intermediate cell is not used to amplify, or otherwise process, the DNA construct or vector prior to introduction into the host cell). Introduction of the DNA construct or vector of the invention into the host cell includes those physical and chemical methods known in the art to introduce a nucleic acid sequence (e.g., DNA sequence) into a host cell without insertion into the host genome. Such methods include, but are not limited to calcium chloride precipitation, electroporation, naked DNA, liposomes and the like. In additional embodiments, DNA constructs or vector are co-transformed with a plasmid, without being inserted into the plasmid. In further embodiments, a selective marker is deleted from the altered *Bacillus* strain by methods known in the art (See, Stahl et al., *J. Bacteriol.* 158:411-418 [1984]; and Palmeros et al., *Gene* 247:255-264 [2000]).

**[0219]** In some embodiments, the transformed cells of the present invention are cultured in conventional nutrient media. The suitable specific culture conditions, such as temperature, pH and the like are known to those skilled in the art and are well described in the scientific literature. In some embodiments, the invention provides a culture (e.g., cell culture) comprising at least one metalloprotease polypeptide or at least one nucleic acid of the invention.

**[0220]** In some embodiments, host cells transformed with at least one polynucleotide sequence encoding at least one metalloprotease polypeptide of the invention are cultured in a suitable nutrient medium under conditions permitting the expression of the present protease, after which the resulting protease is recovered from the culture. In some embodiments, the protease produced by the cells is recovered from the culture medium by conventional procedures, including, but

not limited to for example, separating the host cells from the medium by centrifugation or filtration, precipitating the proteinaceous components of the supernatant or filtrate by means of a salt (e.g., ammonium sulfate), chromatographic purification (e.g., ion exchange, gel filtration, affinity, etc.).

**[0221]** In some embodiments, a metalloprotease polypeptide produced by a recombinant host cell is secreted into the culture medium. A nucleic acid sequence that encodes a purification facilitating domain may be used to facilitate purification of proteins. A vector or DNA construct comprising a polynucleotide sequence encoding a metalloprotease polypeptide may further comprise a nucleic acid sequence encoding a purification facilitating domain to facilitate purification of the metalloprotease polypeptide (See e.g., Kroll et al., *DNA Cell Biol.* 12:441-53 [1993]). Such purification facilitating domains include, but are not limited to, for example, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals (See, Porath, *Protein Expr. Purif.* 3:263-281 [1992]), protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system. The inclusion of a cleavable linker sequence such as Factor XA or enterokinase (e.g., sequences available from Invitrogen, San Diego, Calif.) between the purification domain and the heterologous protein also find use to facilitate purification.

**[0222]** Assays for detecting and measuring the enzymatic activity of an enzyme, such as a metalloprotease polypeptide of the invention, are well known. Various assays for detecting and measuring activity of proteases (e.g., metalloprotease polypeptides of the invention), are also known to those of ordinary skill in the art. In particular, assays are available for measuring protease activity that are based on the release of acid-soluble peptides from casein or hemoglobin, measured as absorbance at 280 nm or colorimetrically using the Folin method. Other exemplary assays involve the solubilization of chromogenic substrates (See e.g., Ward, "Proteinases," in Fogarty (ed.), *Microbial Enzymes and Biotechnology*, Applied Science, London, [1983], pp. 251-317). Other exemplary assays include, but are not limited to succinyl-Ala-Ala-Pro-Phe-para nitroanilide assay (suc-AAPF-pNA)(SEQ ID NO: 43) and the 2,4,6-trinitrobenzene sulfonate sodium salt assay (TNBS assay). Numerous additional references known to those in the art provide suitable methods (See e.g., Wells et al., *Nucleic Acids Res.* 11:7911-7925 [1983]; Christianson et al., *Anal. Biochem.* 223:119-129 [1994]; and Hsia et al., *Anal. Biochem.* 242:221-227 [1999]).

**[0223]** A variety of methods can be used to determine the level of production of a mature protease (e.g., mature metalloprotease polypeptides of the present invention) in a host cell. Such methods include, but are not limited to, for example, methods that utilize either polyclonal or monoclonal antibodies specific for the protease. Exemplary methods include, but are not limited to enzyme-linked immunosorbent assays (ELISA), radioimmunoassays (RIA), fluorescent immunoassays (FIA), and fluorescent activated cell sorting (FACS). These and other assays are well known in the art (See e.g., Maddox et al., *J. Exp. Med.* 158:1211 [1983]).

**[0224]** In some other embodiments, the invention provides methods for making or producing a mature metalloprotease polypeptide of the invention. A mature metalloprotease polypeptide does not include a signal peptide or a propeptide sequence. Some methods comprise making or producing a metalloprotease polypeptide of the invention in a recombi-

nant bacterial host cell, such as for example, a *Bacillus* sp. cell (e.g., a *B. subtilis* cell). In some embodiments, the invention provides a method of producing a metalloprotease polypeptide of the invention, the method comprising cultivating a recombinant host cell comprising a recombinant expression vector comprising a nucleic acid encoding a metalloprotease polypeptide of the invention under conditions conducive to the production of the metalloprotease polypeptide. Some such methods further comprise recovering the metalloprotease polypeptide from the culture.

[0225] In some embodiments the invention provides methods of producing a metalloprotease polypeptide of the invention, the methods comprising: (a) introducing a recombinant expression vector comprising a nucleic acid encoding a metalloprotease polypeptide of the invention into a population of cells (e.g., bacterial cells, such as *B. subtilis* cells); and (b) culturing the cells in a culture medium under conditions conducive to produce the metalloprotease polypeptide encoded by the expression vector. Some such methods further comprise: (c) isolating the metalloprotease polypeptide from the cells or from the culture medium.

#### Fabric and Home Care Products

[0226] In some embodiments, the metalloprotease polypeptides of the present invention can be used in compositions comprising an adjunct material and a metalloprotease polypeptide, wherein the composition is a fabric and home care product.

[0227] In some embodiments, the fabric and home care product compositions comprising at least one metalloprotease polypeptide comprise one or more of the following ingredients (based on total composition weight): from about 0.0005 wt % to about 0.1 wt %, from about 0.001 wt % to about 0.05 wt %, or even from about 0.002 wt % to about 0.03 wt % of said metalloprotease polypeptide; and one or more of the following: from about 0.00003 wt % to about 0.1 wt % fabric hueing agent; from about 0.001 wt % to about 5 wt %, perfume capsules; from about 0.001 wt % to about 1 wt %, cold-water soluble brighteners; from about 0.00003 wt % to about 0.1 wt % bleach catalysts; from about 0.00003 wt % to about 0.1 wt % first wash lipases; from about 0.00003 wt % to about 0.1 wt % bacterial cleaning cellulases; and/or from about 0.05 wt % to about 20 wt % Guerbet nonionic surfactants.

[0228] In some embodiments, the fabric and home care product composition is a liquid laundry detergent or a dishwashing detergent, such as an automatic dishwashing (ADW) detergent or hand dishwashing detergent.

[0229] It is intended that the fabric and home care product is provided in any suitable form, including a fluid or solid, or granular, powder, solid, bar, liquid, tablet, gel, or paste form. The fabric and home care product may be in the form of a unit dose pouch, especially when in the form of a liquid, and typically the fabric and home care product is at least partially, or even completely, enclosed by a water-soluble pouch. In addition, in some embodiments of the fabric and home care products comprising at least one metalloprotease polypeptide, the fabric and home care product may have any combination of parameters and/or characteristics detailed above.

#### Compositions Having the Metalloprotease Polypeptide of the Present Invention

[0230] Unless otherwise noted, all component or composition levels provided herein are made in reference to the active

level of that component or composition, and are exclusive of impurities, for example, residual solvents or by-products, which may be present in commercially available sources. Enzyme components weights are based on total active protein. All percentages and ratios are calculated by weight unless otherwise indicated. All percentages and ratios are calculated based on the total composition unless otherwise indicated. Compositions of the invention include cleaning compositions, such as detergent compositions. In the exemplified detergent compositions, the enzymes levels are expressed by pure enzyme by weight of the total composition and unless otherwise specified, the detergent ingredients are expressed by weight of the total compositions.

[0231] As indicated herein, in some embodiments, the cleaning compositions of the present invention further comprise adjunct materials including, but not limited to, surfactants, builders, bleaches, bleach activators, bleach catalysts, other enzymes, enzyme stabilizing systems, chelants, optical brighteners, soil release polymers, dye transfer agents, dispersants, suds suppressors, dyes, perfumes, colorants, filler salts, hydrotropes, photoactivators, fluorescers, fabric conditioners, hydrolyzable surfactants, preservatives, anti-oxidants, anti-shrinkage agents, anti-wrinkle agents, germicides, fungicides, color speckles, silvercare, anti-tarnish and/or anti-corrosion agents, alkalinity sources, solubilizing agents, carriers, processing aids, pigments, and pH control agents (See e.g., U.S. Pat. Nos. 6,610,642, 6,605,458, 5,705,464, 5,710,115, 5,698,504, 5,695,679, 5,686,014 and 5,646,101, all of which are incorporated herein by reference). Embodiments of specific cleaning composition materials are exemplified in detail below. In embodiments in which the cleaning adjunct materials are not compatible with the metalloprotease polypeptides of the present invention in the cleaning compositions, then suitable methods of keeping the cleaning adjunct materials and the protease(s) separated (i.e., not in contact with each other) until combination of the two components is appropriate are used. Such separation methods include any suitable method known in the art (e.g., gencaps, encapsulation, tablets, physical separation, etc.).

[0232] The cleaning compositions of the present invention are advantageously employed for example, in laundry applications, hard surface cleaning, dishwashing applications, including automatic dishwashing and hand dishwashing, as well as cosmetic applications such as dentures, teeth, hair and skin. In addition, due to the unique advantages of increased effectiveness in lower temperature solutions, the enzymes of the present invention are ideally suited for laundry applications. Furthermore, the enzymes of the present invention find use in granular and liquid compositions.

[0233] The metalloprotease polypeptides of the present invention also find use in cleaning additive products. In some embodiments, low temperature solution cleaning applications find use. In some embodiments, the present invention provides cleaning additive products including at least one enzyme of the present invention is ideally suited for inclusion in a wash process when additional bleaching effectiveness is desired. Such instances include, but are not limited to low temperature solution cleaning applications. In some embodiments, the additive product is in its simplest form, one or more proteases. In some embodiments, the additive is packaged in dosage form for addition to a cleaning process. In some embodiments, the additive is packaged in dosage form for addition to a cleaning process where a source of peroxygen is employed and increased bleaching effectiveness is desired.

Any suitable single dosage unit form finds use with the present invention, including but not limited to pills, tablets, gencaps, or other single dosage units such as pre-measured powders or liquids. In some embodiments, filler(s) or carrier material(s) are included to increase the volume of such compositions. Suitable filler or carrier materials include, but are not limited to, various salts of sulfate, carbonate and silicate as well as talc, clay and the like. Suitable filler or carrier materials for liquid compositions include, but are not limited to water or low molecular weight primary and secondary alcohols including polyols and diols. Examples of such alcohols include, but are not limited to, methanol, ethanol, propanol and isopropanol. In some embodiments, the compositions contain from about 5% to about 90% of such materials. Acidic fillers find use to reduce pH. Alternatively, in some embodiments, the cleaning additive includes adjunct ingredients, as more fully described below.

**[0234]** The present cleaning compositions and cleaning additives require an effective amount of at least one of the metalloprotease polypeptides provided herein, alone or in combination with other proteases and/or additional enzymes. The required level of enzyme is achieved by the addition of one or more metalloprotease polypeptides of the present invention. Typically the present cleaning compositions comprise at least about 0.0001 weight percent, from about 0.0001 to about 10, from about 0.001 to about 1, or from about 0.01 to about 0.1 weight percent of at least one of the metalloprotease polypeptides of the present invention.

**[0235]** The cleaning compositions herein are typically formulated such that, during use in aqueous cleaning operations, the wash water will have a pH of from about 4.0 to about 11.5, or even from about 5.0 to about 11.5, or even from about 5.0 to about 8.0, or even from about 7.5 to about 10.5. Liquid product formulations are typically formulated to have a pH from about 3.0 to about 9.0 or even from about 3 to about 5. Granular laundry products are typically formulated to have a pH from about 9 to about 11. Techniques for controlling pH at recommended usage levels include the use of buffers, alkalis, acids, etc., and are well known to those skilled in the art.

**[0236]** Suitable "low pH cleaning compositions" typically have a pH of from about 3 to about 5, and are typically free of surfactants that hydrolyze in such a pH environment. Such surfactants include sodium alkyl sulfate surfactants that comprise at least one ethylene oxide moiety or even from about 1 to about 16 moles of ethylene oxide. Such cleaning compositions typically comprise a sufficient amount of a pH modifier, such as sodium hydroxide, monoethanolamine or hydrochloric acid, to provide such cleaning composition with a pH of from about 3 to about 5. Such compositions typically comprise at least one acid stable enzyme. In some embodiments, the compositions are liquids, while in other embodiments, they are solids. The pH of such liquid compositions is typically measured as a neat pH. The pH of such solid compositions is measured as a 10% solids solution of said composition wherein the solvent is distilled water. In these embodiments, all pH measurements are taken at 20° C., unless otherwise indicated.

**[0237]** In some embodiments, when the metalloprotease polypeptide (s) is/are employed in a granular composition or liquid, it is desirable for the metalloprotease polypeptide to be in the form of an encapsulated particle to protect the metalloprotease polypeptide from other components of the granular composition during storage. In addition, encapsulation is also a means of controlling the availability of the metallopro-

tease polypeptide during the cleaning process. In some embodiments, encapsulation enhances the performance of the metalloprotease polypeptide (s) and/or additional enzymes. In this regard, the metalloprotease polypeptides of the present invention are encapsulated with any suitable encapsulating material known in the art. In some embodiments, the encapsulating material typically encapsulates at least part of the metalloprotease polypeptide (s) of the present invention. Typically, the encapsulating material is water-soluble and/or water-dispersible. In some embodiments, the encapsulating material has a glass transition temperature ( $T_g$ ) of 0° C. or higher. Glass transition temperature is described in more detail in WO 97/11151. The encapsulating material is typically selected from consisting of carbohydrates, natural or synthetic gums, chitin, chitosan, cellulose and cellulose derivatives, silicates, phosphates, borates, polyvinyl alcohol, polyethylene glycol, paraffin waxes, and combinations thereof. When the encapsulating material is a carbohydrate, it is typically selected from monosaccharides, oligosaccharides, polysaccharides, and combinations thereof. In some typical embodiments, the encapsulating material is a starch (See e.g., EP 0 922 499; U.S. Pat. No. 4,977,252; U.S. Pat. No. 5,354,559, and U.S. Pat. No. 5,935,826). In some embodiments, the encapsulating material is a microsphere made from plastic such as thermoplastics, acrylonitrile, methacrylonitrile, polyacrylonitrile, polymethacrylonitrile and mixtures thereof; commercially available microspheres that find use include, but are not limited to those supplied by EXPANCEL® (Stockviksverken, Sweden), and PM 6545, PM 6550, PM 7220, PM 7228, EXTENDOSPHERES®, LUXSIL®, Q-CEL®, and SPHERICEL® (PQ Corp., Valley Forge, Pa.).

**[0238]** As described herein, the metalloprotease polypeptides of the present invention find particular use in the cleaning industry, including, but not limited to laundry and dish detergents. These applications place enzymes under various environmental stresses. The metalloprotease polypeptides of the present invention provide advantages over many currently used enzymes, due to their stability under various conditions.

**[0239]** Indeed, there are a variety of wash conditions including varying detergent formulations, wash water volumes, wash water temperatures, and lengths of wash time, to which proteases involved in washing are exposed. In addition, detergent formulations used in different geographical areas have different concentrations of their relevant components present in the wash water. For example, European detergents typically have about 4500-5000 ppm of detergent components in the wash water, while Japanese detergents typically have approximately 667 ppm of detergent components in the wash water. In North America, particularly the United States, detergents typically have about 975 ppm of detergent components present in the wash water.

**[0240]** A low detergent concentration system includes detergents where less than about 800 ppm of the detergent components are present in the wash water. Japanese detergents are typically considered low detergent concentration system as they have approximately 667 ppm of detergent components present in the wash water.

**[0241]** A medium detergent concentration includes detergents where between about 800 ppm and about 2000 ppm of the detergent components are present in the wash water. North American detergents are generally considered to be medium detergent concentration systems as they have approximately 975 ppm of detergent components present in the wash water.

Brazil typically has approximately 1500 ppm of detergent components present in the wash water.

**[0242]** A high detergent concentration system includes detergents where greater than about 2000 ppm of the detergent components are present in the wash water. European detergents are generally considered to be high detergent concentration systems as they have approximately 4500-5000 ppm of detergent components in the wash water.

**[0243]** Latin American detergents are generally high suds phosphate builder detergents and the range of detergents used in Latin America can fall in both the medium and high detergent concentrations as they range from 1500 ppm to 6000 ppm of detergent components in the wash water. As mentioned above, Brazil typically has approximately 1500 ppm of detergent components present in the wash water. However, other high suds phosphate builder detergent geographies, not limited to other Latin American countries, may have high detergent concentration systems up to about 6000 ppm of detergent components present in the wash water.

**[0244]** In light of the foregoing, it is evident that concentrations of detergent compositions in typical wash solutions throughout the world varies from less than about 800 ppm of detergent to about 6000 ppm in high suds phosphate builder geographies.

**[0245]** The concentrations of the typical wash solutions are determined empirically. For example, in the U.S., a typical washing machine holds a volume of about 64.4 L of wash solution. Accordingly, in order to obtain a concentration of about 975 ppm of detergent within the wash solution about 62.79 g of detergent composition must be added to the 64.4 L of wash solution. This amount is the typical amount measured into the wash water by the consumer using the measuring cup provided with the detergent.

**[0246]** As a further example, different geographies use different wash temperatures. The temperature of the wash water in Japan is typically less than that used in Europe. For example, the temperature of the wash water in North America and Japan is typically between about 10 and about 40° C. (e.g., about 20° C.), whereas the temperature of wash water in Europe is typically between about 30 and about 60° C. (e.g., about 40° C.). However, in the interest of saving energy, many consumers are switching to using cold water washing. In addition, in some further regions, cold water is typically used for laundry, as well as dish washing applications. In some embodiments, the “cold water washing” of the present invention utilizes “cold water detergent” suitable for washing at temperatures from about 10° C. to about 40° C., or from about 20° C. to about 30° C., or from about 15° C. to about 25° C., as well as all other combinations within the range of about 15° C. to about 35° C., and all ranges within 10° C. to 40° C.

**[0247]** As a further example, different geographies typically have different water hardness. Water hardness is usually described in terms of the grains per gallon mixed  $\text{Ca}^{2+}/\text{Mg}^{2+}$ . Hardness is a measure of the amount of calcium ( $\text{Ca}^{2+}$ ) and magnesium ( $\text{Mg}^{2+}$ ) in the water. Most water in the United States is hard, but the degree of hardness varies. Moderately hard (60-120 ppm) to hard (121-181 ppm) water has 60 to 181 parts per million (parts per million converted to grains per U.S. gallon is ppm # divided by 17.1 equals grains per gallon) of hardness minerals.

| Water           | Grains per gallon | Parts per million |
|-----------------|-------------------|-------------------|
| Soft            | less than 1.0     | less than 17      |
| Slightly hard   | 1.0 to 3.5        | 17 to 60          |
| Moderately hard | 3.5 to 7.0        | 60 to 120         |
| Hard            | 7.0 to 10.5       | 120 to 180        |
| Very hard       | greater than 10.5 | greater than 180  |

**[0248]** European water hardness is typically greater than about 10.5 (for example about 10.5 to about 20.0) grains per gallon mixed  $\text{Ca}^{2+}/\text{Mg}^{2+}$  (e.g., about 15 grains per gallon mixed  $\text{Ca}^{2+}/\text{Mg}^{2+}$ ). North American water hardness is typically greater than Japanese water hardness, but less than European water hardness. For example, North American water hardness can be between about 3 to about 10 grains, about 3 to about 8 grains or about 6 grains. Japanese water hardness is typically lower than North American water hardness, usually less than about 4, for example about 3 grains per gallon mixed  $\text{Ca}^{2+}/\text{Mg}^{2+}$ .

**[0249]** Accordingly, in some embodiments, the present invention provides metalloprotease polypeptides that show surprising wash performance in at least one set of wash conditions (e.g., water temperature, water hardness, and/or detergent concentration). In some embodiments, the metalloprotease polypeptides of the present invention are comparable in wash performance to other metalloprotease polypeptide proteases. In some embodiments of the present invention, the metalloprotease polypeptides provided herein exhibit enhanced oxidative stability, enhanced thermal stability, enhanced cleaning capabilities under various conditions, and/or enhanced chelator stability. In addition, the metalloprotease polypeptides of the present invention find use in cleaning compositions that do not include detergents, again either alone or in combination with builders and stabilizers.

**[0250]** In some embodiments of the present invention, the cleaning compositions comprise at least one metalloprotease polypeptide of the present invention at a level from about 0.0001% to about 10% by weight of the composition and the balance (e.g., about 99.999% to about 90.0%) comprising cleaning adjunct materials by weight of composition. In some other embodiments of the present invention, the cleaning compositions of the present invention comprises at least one metalloprotease polypeptide at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% by weight of the composition and the balance of the cleaning composition (e.g., about 99.9999% to about 90.0%, about 99.999% to about 98%, about 99.995% to about 99.5% by weight) comprising cleaning adjunct materials.

**[0251]** In some embodiments, the cleaning compositions of the present invention comprise one or more additional detergent enzymes, which provide cleaning performance and/or fabric care and/or dishwashing benefits. Examples of suitable enzymes include, but are not limited to, acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageenases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1, 4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxigenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases,

pullulanases, reductases, rhamnogalacturonases, beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases, or any combinations or mixtures thereof. In some embodiments, a combination of enzymes is used (i.e., a “cocktail”) comprising conventional applicable enzymes like protease, lipase, cutinase and/or cellulase in conjunction with amylase is used.

**[0252]** In addition to the metalloprotease polypeptides provided herein, any other suitable protease finds use in the compositions of the present invention. Suitable proteases include those of animal, vegetable or microbial origin. In some embodiments, microbial proteases are used. In some embodiments, chemically or genetically modified mutants are included. In some embodiments, the protease is a serine protease, preferably an alkaline microbial protease or a trypsin-like protease. Examples of alkaline proteases include subtilisins, especially those derived from *Bacillus* (e.g., subtilisin, *lentus*, *amyloliquefaciens*, subtilisin Carlsberg, subtilisin 309, subtilisin 147 and subtilisin 168). Additional examples include those mutant proteases described in U.S. Pat. Nos. RE 34,606, 5,955,340, 5,700,676, 6,312,936, and 6,482,628, all of which are incorporated herein by reference. Additional protease examples include, but are not limited to trypsin (e.g., of porcine or bovine origin), and the *Fusarium* protease described in WO 89/06270. In some embodiments, commercially available protease enzymes that find use in the present invention include, but are not limited to MAX-ATASE®, MAXACAL™, MAXAPEM™, OPTICLEAN®, OPTIMASE®, PROPERASE®, PURAFECT®, PURAFECT® OXP, PURAMAX™, EXCELLASE™, and PURAFAST™ (Genencor); ALCALASE®, SAVINASE®, PRIMASE®, DURAZYM™, POLARZYME®, OVOZYME®, KANNASE®, LIQUANASE®, NEUTRASE®, RELEASE® and ESPERASE® (Novozymes); BLAP™ and BLAP™ variants (Henkel Kommanditgesellschaft auf Aktien, Duesseldorf, Germany), and KAP (*B. alkalophilus* subtilisin; Kao Corp., Tokyo, Japan). Various proteases are described in WO95/23221, WO 92/21760, WO 09/149200, WO 09/149144, WO 09/149145, WO 11/072099, WO 10/056640, WO 10/056653, WO 11/140364, WO 12/151534, U.S. Pat. Publ. No. 2008/0090747, and U.S. Pat. Nos. 5,801,039, 5,340,735, 5,500,364, 5,855,625, US RE 34,606, 5,955,340, 5,700,676, 6,312,936, and 6,482,628, and various other patents. In some further embodiments, metalloproteases find use in the present invention, including but not limited to the neutral metalloprotease described in WO 07/044993.

**[0253]** In addition, any suitable lipase finds use in the present invention. Suitable lipases include, but are not limited to those of bacterial or fungal origin. Chemically or genetically modified mutants are encompassed by the present invention. Examples of useful lipases include *Humicola lanuginosa* lipase (See e.g., EP 258 068, and EP 305 216), *Rhizomucor miehei* lipase (See e.g., EP 238 023), *Candida* lipase, such as *C. antarctica* lipase (e.g., the *C. antarctica* lipase A or B; See e.g., EP 214 761), *Pseudomonas* lipases such as *P. alcaligenes* lipase and *P. pseudoalcaligenes* lipase (See e.g., EP 218 272), *P. cepacia* lipase (See e.g., EP 331 376), *P. stutzeri* lipase (See e.g., GB 1,372,034), *P. fluorescens* lipase, *Bacillus* lipase (e.g., *B. subtilis* lipase [Dartois et al., Biochem. Biophys. Acta 1131:253-260 [1993]]; *B. stearothermophilus* lipase [See e.g., JP 64/744992]; and *B. pumilus* lipase [See e.g., WO 91/16422]).

**[0254]** Furthermore, a number of cloned lipases find use in some embodiments of the present invention, including but not limited to *Penicillium camembertii* lipase (See, Yamaguchi et al., Gene 103:61-67 [1991]), *Geotricum candidum* lipase (See, Shimada et al., J. Biochem., 106:383-388 [1989]), and various *Rhizopus* lipases such as *R. delemar* lipase (See, Hass et al., Gene 109:117-113 [1991]), a *R. niveus* lipase (Kugimiya et al., Biosci. Biotech. Biochem. 56:716-719 [1992]) and *R. oryzae* lipase.

**[0255]** Other types of lipase polypeptide enzymes such as cutinases also find use in some embodiments of the present invention, including but not limited to the cutinase derived from *Pseudomonas mendocina* (See, WO 88/09367), and the cutinase derived from *Fusarium solani pisi* (See, WO 90/09446).

**[0256]** Additional suitable lipases include commercially available lipases such as M1 LIPASE™, LUMA FAST™, and LIPOMAX™ (Genencor); LIPEX®, LIPOLASE® and LIPOLASE® ULTRA (Novozymes); and LIPASE P™ “Amano” (Amano Pharmaceutical Co. Ltd., Japan).

**[0257]** In some embodiments of the present invention, the cleaning compositions of the present invention further comprise lipases at a level from about 0.00001% to about 10% of additional lipase by weight of the composition and the balance of cleaning adjunct materials by weight of composition. In some other embodiments of the present invention, the cleaning compositions of the present invention also comprise lipases at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% lipase by weight of the composition.

**[0258]** In some embodiments of the present invention, any suitable amylase finds use in the present invention. In some embodiments, any amylase (e.g., alpha and/or beta) suitable for use in alkaline solutions also find use. Suitable amylases include, but are not limited to those of bacterial or fungal origin. Chemically or genetically modified mutants are included in some embodiments. Amylases that find use in the present invention, include, but are not limited to  $\alpha$ -amylases obtained from *B. licheniformis* (See e.g., GB 1,296,839). Additional suitable amylases include those found in WO9510603, WO9526397, WO9623874, WO9623873, WO9741213, WO9919467, WO0060060, WO0029560, WO9923211, WO9946399, WO0060058, WO0060059, WO9942567, WO0114532, WO02092797, WO0166712, WO0188107, WO0196537, WO0210355, WO9402597, WO0231124, WO9943793, WO9943794, WO2004113551, WO2005001064, WO2005003311, WO0164852, WO2006066594, WO2006066594, WO2006066596, WO2006012899, WO2008092919, WO2008000825, WO2005018336, WO2005066338, WO2009140504, WO2005019443, WO2010091221, WO2010088447, WO0134784, WO2006012902, WO2006031554, WO2006136161, WO2008101894, WO2010059413, WO2011098531, WO2011080352, WO2011080353, WO2011080354, WO2011082425, WO2011082429, WO2011076123, WO2011087836, WO2011076897, WO94183314, WO9535382, WO9909183, WO9826078, WO9902702, WO9743424, WO9929876, WO9100353, WO9605295, WO9630481, WO9710342, WO2008088493, WO2009149419, WO2009061381, WO2009100102, WO2010104675, WO2010117511, and WO2010115021. Commercially available amylases that find use in the present invention include, but are not limited to DURAMYL®, TERMAMYL®, FUNGAMYL®, STAINZYME®, STAIN-

ZYME PLUS®, STAINZYME ULTRA®, and BAN™ (Novozymes), as well as POWERASE™, RAPIDASE® and MAXAMYL® P (Genencor).

[0259] In some embodiments of the present invention, the cleaning compositions of the present invention further comprise amylases at a level from about 0.00001% to about 10% of additional amylase by weight of the composition and the balance of cleaning adjunct materials by weight of composition. In some other embodiments of the present invention, the cleaning compositions of the present invention also comprise amylases at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% amylase by weight of the composition.

[0260] In some further embodiments, any suitable cellulase finds used in the cleaning compositions of the present invention. Suitable cellulases include, but are not limited to those of bacterial or fungal origin. Chemically or genetically modified mutants are included in some embodiments. Suitable cellulases include, but are not limited to *Humicola insolens* cellulases (See e.g., U.S. Pat. No. 4,435,307). Especially suitable cellulases are the cellulases having color care benefits (See e.g., EP 0 495 257). Commercially available cellulases that find use in the present include, but are not limited to CELLUZYME, CELLUCLEAN, CAREZYME (Novozymes), PURADEX AND REVITALENZ (Danisco US Inc.), and KAC-500(B) (Kao Corporation). In some embodiments, cellulases are incorporated as portions or fragments of mature wild-type or variant cellulases, wherein a portion of the N-terminus is deleted (See e.g., U.S. Pat. No. 5,874,276). Additional suitable cellulases include those found in WO2005054475, WO2005056787, U.S. Pat. Nos. 7,449,318, and 7,833,773. In some embodiments, the cleaning compositions of the present invention further comprise cellulases at a level from about 0.00001% to about 10% of additional cellulase by weight of the composition and the balance of cleaning adjunct materials by weight of composition. In some other embodiments of the present invention, the cleaning compositions of the present invention also comprise cellulases at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% cellulase by weight of the composition.

[0261] Any mannanase suitable for use in detergent compositions also finds use in the present invention. Suitable mannanases include, but are not limited to those of bacterial or fungal origin. Chemically or genetically modified mutants are included in some embodiments. Various mannanases are known which find use in the present invention (See e.g., U.S. Pat. Nos. 6,566,114; 6,602,842; 5,476,775 and 6,440,991, and U.S. Prov. App. Ser. No. 61/739,267; all of which are incorporated herein by reference). Commercially available mannanases that find use in the present invention include, but are not limited to MANNASTAR, PURABRITE, and MANNAWAY. In some embodiments, the cleaning compositions of the present invention further comprise mannanases at a level from about 0.00001% to about 10% of additional mannanase by weight of the composition and the balance of cleaning adjunct materials by weight of composition. In some embodiments of the present invention, the cleaning compositions of the present invention also comprise mannanases at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% mannanase by weight of the composition.

[0262] In some embodiments, peroxidases are used in combination with hydrogen peroxide or a source thereof (e.g., a

percarbonate, perborate or persulfate) in the compositions of the present invention. In some alternative embodiments, oxidases are used in combination with oxygen. Both types of enzymes are used for "solution bleaching" (i.e., to prevent transfer of a textile dye from a dyed fabric to another fabric when the fabrics are washed together in a wash liquor), preferably together with an enhancing agent (See e.g., WO 94/12621 and WO 95/01426). Suitable peroxidases/oxidases include, but are not limited to those of plant, bacterial or fungal origin. Chemically or genetically modified mutants are included in some embodiments. In some embodiments, the cleaning compositions of the present invention further comprise peroxidase and/or oxidase enzymes at a level from about 0.00001% to about 10% of additional peroxidase and/or oxidase by weight of the composition and the balance of cleaning adjunct materials by weight of composition. In some other embodiments of the present invention, the cleaning compositions of the present invention also comprise, peroxidase and/or oxidase enzymes at a level of about 0.0001% to about 10%, about 0.001% to about 5%, about 0.001% to about 2%, about 0.005% to about 0.5% peroxidase and/or oxidase enzymes by weight of the composition.

[0263] In some embodiments, additional enzymes find use, including but not limited to perhydrolases (See e.g., WO 05/056782). In addition, in some embodiments, mixtures of the above mentioned enzymes are encompassed herein, in particular one or more additional protease, amylase, lipase, mannanase, and/or at least one cellulase. Indeed, it is contemplated that various mixtures of these enzymes will find use in the present invention. It is also contemplated that the varying levels of the metalloprotease polypeptide (s) and one or more additional enzymes may both independently range to about 10%, the balance of the cleaning composition being cleaning adjunct materials. The specific selection of cleaning adjunct materials are readily made by considering the surface, item, or fabric to be cleaned, and the desired form of the composition for the cleaning conditions during use (e.g., through the wash detergent use).

[0264] Examples of suitable cleaning adjunct materials include, but are not limited to, surfactants, builders, bleaches, bleach activators, bleach catalysts, other enzymes, enzyme stabilizing systems, chelants, optical brighteners, soil release polymers, dye transfer agents, dye transfer inhibiting agents, catalytic materials, hydrogen peroxide, sources of hydrogen peroxide, preformed peracids, polymeric dispersing agents, clay soil removal agents, structure elasticizing agents, dispersants, suds suppressors, dyes, perfumes, colorants, filler salts, hydrotropes, photoactivators, fluorescers, fabric conditioners, fabric softeners, carriers, hydrotropes, processing aids, solvents, pigments, hydrolyzable surfactants, preservatives, anti-oxidants, anti-shrinkage agents, anti-wrinkle agents, germicides, fungicides, color speckles, silvercare, anti-tarnish and/or anti-corrosion agents, alkalinity sources, solubilizing agents, carriers, processing aids, pigments, and pH control agents (See e.g., U.S. Pat. Nos. 6,610,642; 6,605,458; 5,705,464; 5,710,115; 5,698,504; 5,695,679; 5,686,014 and 5,646,101, all of which are incorporated herein by reference). Embodiments of specific cleaning composition materials are exemplified in detail below. In embodiments in which the cleaning adjunct materials are not compatible with the metalloprotease polypeptides of the present invention in the cleaning compositions, then suitable methods of keeping the cleaning adjunct materials and the protease(s) separated (i.e., not in contact with each other) until combination of the



two components is appropriate are used. Such separation methods include any suitable method known in the art (e.g., gelpcaps, encapsulation, tablets, physical separation, etc.).

**[0265]** In some embodiments, an effective amount of one or more metalloprotease polypeptide (s) provided herein is included in compositions useful for cleaning a variety of surfaces in need of proteinaceous stain removal. Such cleaning compositions include cleaning compositions for such applications as cleaning hard surfaces, fabrics, and dishes. Indeed, in some embodiments, the present invention provides fabric cleaning compositions, while in other embodiments, the present invention provides non-fabric cleaning compositions. Notably, the present invention also provides cleaning compositions suitable for personal care, including oral care (including dentrifices, toothpastes, mouthwashes, etc., as well as denture cleaning compositions), skin, and hair cleaning compositions. It is intended that the present invention encompass detergent compositions in any form (i.e., liquid, granular, bar, semi-solid, gels, emulsions, tablets, capsules, etc.).

**[0266]** By way of example, several cleaning compositions wherein the metalloprotease polypeptides of the present invention find use are described in greater detail below. In some embodiments in which the cleaning compositions of the present invention are formulated as compositions suitable for use in laundry machine washing method(s), the compositions of the present invention preferably contain at least one surfactant and at least one builder compound, as well as one or more cleaning adjunct materials preferably selected from organic polymeric compounds, bleaching agents, additional enzymes, suds suppressors, dispersants, lime-soap dispersants, soil suspension and anti-redeposition agents and corrosion inhibitors. In some embodiments, laundry compositions also contain softening agents (i.e., as additional cleaning adjunct materials). The compositions of the present invention also find use in detergent additive products in solid or liquid form. Such additive products are intended to supplement and/or boost the performance of conventional detergent compositions and can be added at any stage of the cleaning process. In some embodiments, the density of the laundry detergent compositions herein ranges from about 400 to about 1200 g/liter, while in other embodiments, it ranges from about 500 to about 950 g/liter of composition measured at 20° C.

**[0267]** In embodiments formulated as compositions for use in manual dishwashing methods, the compositions of the invention preferably contain at least one surfactant and preferably at least one additional cleaning adjunct material selected from organic polymeric compounds, suds enhancing agents, group II metal ions, solvents, hydrotropes and additional enzymes.

**[0268]** In some embodiments, various cleaning compositions such as those provided in U.S. Pat. No. 6,605,458, find use with the metalloprotease polypeptides of the present invention. Thus, in some embodiments, the compositions comprising at least one metalloprotease polypeptide of the present invention is a compact granular fabric cleaning composition, while in other embodiments, the composition is a granular fabric cleaning composition useful in the laundering of colored fabrics, in further embodiments, the composition is a granular fabric cleaning composition which provides softening through the wash capacity, in additional embodiments, the composition is a heavy duty liquid fabric cleaning composition. In some embodiments, the compositions comprising at least one metalloprotease polypeptide of the present

invention are fabric cleaning compositions such as those described in U.S. Pat. Nos. 6,610,642 and 6,376,450. In addition, the metalloprotease polypeptides of the present invention find use in granular laundry detergent compositions of particular utility under European or Japanese washing conditions (See e.g., U.S. Pat. No. 6,610,642).

**[0269]** In some alternative embodiments, the present invention provides hard surface cleaning compositions comprising at least one metalloprotease polypeptide provided herein. Thus, in some embodiments, the compositions comprising at least one metalloprotease polypeptide of the present invention is a hard surface cleaning composition such as those described in U.S. Pat. Nos. 6,610,642; 6,376,450, and 6,376,450.

**[0270]** In yet further embodiments, the present invention provides dishwashing compositions comprising at least one metalloprotease polypeptide provided herein. Thus, in some embodiments, the compositions comprising at least one metalloprotease polypeptide of the present invention is a hard surface cleaning composition such as those in U.S. Pat. Nos. 6,610,642 and 6,376,450. In some still further embodiments, the present invention provides dishwashing compositions comprising at least one metalloprotease polypeptide provided herein. In some further embodiments, the compositions comprising at least one metalloprotease polypeptide of the present invention comprise oral care compositions such as those in U.S. Pat. Nos. 6,376,450, and 6,376,450. The formulations and descriptions of the compounds and cleaning adjunct materials contained in the aforementioned U.S. Pat. Nos. 6,376,450; 6,605,458; 6,605,458, and 6,610,642, find use with the metalloprotease polypeptides provided herein.

**[0271]** The cleaning compositions of the present invention are formulated into any suitable form and prepared by any process chosen by the formulator, non-limiting examples of which are described in U.S. Pat. Nos. 5,879,584; 5,691,297; 5,574,005; 5,569,645; 5,565,422; 5,516,448; 5,489,392, and 5,486,303, all of which are incorporated herein by reference. When a low pH cleaning composition is desired, the pH of such composition is adjusted via the addition of a material such as monoethanolamine or an acidic material such as HCl.

**[0272]** In some embodiments, the cleaning compositions of the present invention can be formulated to have an alkaline pH under wash conditions, such as a pH of from about 8.0 to about 12.0, or from about 8.5 to about 11.0, or from about 9.0 to about 11.0. In some embodiments, the cleaning compositions of the present invention can be formulated to have a neutral pH under wash conditions, such as a pH of from about 5.0 to about 8.0, or from about 5.5 to about 8.0, or from about 6.0 to about 8.0, or from about 6.0 to about 7.5. In some embodiments, the neutral pH conditions can be measured when the cleaning composition is dissolved 1:100 (wt:wt) in de-ionised water at 20° C., measured using a conventional pH meter.

**[0273]** While not essential for the purposes of the present invention, the non-limiting list of adjuncts illustrated hereinafter are suitable for use in the instant cleaning compositions. In some embodiments, these adjuncts are incorporated for example, to assist or enhance cleaning performance, for treatment of the substrate to be cleaned, or to modify the aesthetics of the cleaning composition as is the case with perfumes, colorants, dyes or the like. It is understood that such adjuncts are in addition to the metalloprotease polypeptides of the present invention. The precise nature of these additional components, and levels of incorporation thereof, will depend on

the physical form of the composition and the nature of the cleaning operation for which it is to be used. Suitable adjunct materials include, but are not limited to, surfactants, builders, chelating agents, dye transfer inhibiting agents, deposition aids, dispersants, additional enzymes, and enzyme stabilizers, catalytic materials, bleach activators, bleach boosters, hydrogen peroxide, sources of hydrogen peroxide, preformed peracids, polymeric dispersing agents, clay soil removal/anti-redeposition agents, brighteners, suds suppressors, dyes, perfumes, structure elasticizing agents, fabric softeners, carriers, hydrotropes, processing aids and/or pigments. In addition to the disclosure below, suitable examples of such other adjuncts and levels of use are found in U.S. Pat. Nos. 5,576,282, 6,306,812, and 6,326,348, incorporated by reference. The aforementioned adjunct ingredients may constitute the balance of the cleaning compositions of the present invention.

**[0274]** In some embodiments, the cleaning compositions according to the present invention comprise an acidifying particle or an amino carboxylic builder. Examples of an amino carboxylic builder include aminocarboxylic acids, salts and derivatives thereof. In some embodiment, the amino carboxylic builder is an aminopolycarboxylic builder, such as glycine-N,N-diacetic acid or derivative of general formula  $\text{MOOC}-\text{CHR}-\text{N}(\text{CH}_2\text{COOM})_2$  where R is  $\text{C}_{1-12}$  alkyl and M is alkali metal. In some embodiments, the amino carboxylic builder can be methylglycine diacetic acid (MGDA), GLDA (glutamic-N,N-diacetic acid), iminodisuccinic acid (IDS), carboxymethyl inulin and salts and derivatives thereof, aspartic acid-N-monoacetic acid (ASMA), aspartic acid-N,N-diacetic acid (ASDA), aspartic acid-N-monopropionic acid (ASMP), iminodisuccinic acid (IDA), N-(2-sulfomethyl)aspartic acid (SMAS), N-(2-sulfoethyl)aspartic acid (SEAS), N-(2-sulfomethyl)glutamic acid (SMGL), N-(2-sulfoethyl)glutamic acid (SEGL), IDS (iminodiacetic acid) and salts and derivatives thereof such as N-methyliminodiacetic acid (MIDA), alpha-alanine-N,N-diacetic acid (alpha-ALDA), serine-N,N-diacetic acid (SEDA), isoserine-N,N-diacetic acid (ISDA), phenylalanine-N,N-diacetic acid (PHDA), anthranilic acid-N,N-diacetic acid (ANDA), sulfanilic acid-N,N-diacetic acid (SLDA), taurine-N,N-diacetic acid (TUDA) and sulfomethyl-N,N-diacetic acid (SMDA) and alkali metal salts and derivative thereof. In some embodiments, the acidifying particle has a weight geometric mean particle size of from about  $400\mu$  to about  $1200\mu$  and a bulk density of at least  $550\text{ g/L}$ . In some embodiments, the acidifying particle comprises at least about 5% of the builder.

**[0275]** In some embodiments, the acidifying particle can comprise any acid, including organic acids and mineral acids. Organic acids can have one or two carboxyls and in some instances up to 15 carbons, especially up to 10 carbons, such as formic, acetic, propionic, capric, oxalic, succinic, adipic, maleic, fumaric, sebacic, malic, lactic, glycolic, tartaric and glyoxylic acids. In some embodiments, the acid is citric acid. Mineral acids include hydrochloric and sulphuric acid. In some instances, the acidifying particle of the invention is a highly active particle comprising a high level of amino carboxylic builder. Sulphuric acid has been found to further contribute to the stability of the final particle.

**[0276]** In some embodiments, the cleaning compositions according to the present invention comprise at least one surfactant and/or a surfactant system wherein the surfactant is selected from nonionic surfactants, anionic surfactants, cationic surfactants, ampholytic surfactants, zwitterionic surfactants, semi-polar nonionic surfactants and mixtures thereof.

In some low pH cleaning composition embodiments (e.g., compositions having a neat pH of from about 3 to about 5), the composition typically does not contain alkyl ethoxylated sulfate, as it is believed that such surfactant may be hydrolyzed by such compositions the acidic contents. In some embodiments, the surfactant is present at a level of from about 0.1% to about 60%, while in alternative embodiments the level is from about 1% to about 50%, while in still further embodiments the level is from about 5% to about 40%, by weight of the cleaning composition.

**[0277]** In some embodiments, the cleaning compositions of the present invention comprise one or more detergent builders or builder systems. In some embodiments incorporating at least one builder, the cleaning compositions comprise at least about 1%, from about 3% to about 60% or even from about 5% to about 40% builder by weight of the cleaning composition. Builders include, but are not limited to, the alkali metal, ammonium and alkanolammonium salts of polyphosphates, alkali metal silicates, alkaline earth and alkali metal carbonates, aluminosilicates, polycarboxylate compounds, ether hydroxypolycarboxylates, copolymers of maleic anhydride with ethylene or vinyl methyl ether, 1,3,5-trihydroxy benzene-2,4,6-trisulphonic acid, and carboxymethyloxysuccinic acid, the various alkali metal, ammonium and substituted ammonium salts of polyacetic acids such as ethylenediamine tetraacetic acid and nitrilotriacetic acid, as well as polycarboxylates such as mellitic acid, succinic acid, citric acid, oxydisuccinic acid, polymaleic acid, benzene 1,3,5-tricarboxylic acid, carboxymethyloxysuccinic acid, and soluble salts thereof. Indeed, it is contemplated that any suitable builder will find use in various embodiments of the present invention.

**[0278]** In some embodiments, the builders form water-soluble hardness ion complexes (e.g., sequestering builders), such as citrates and polyphosphates (e.g., sodium tripolyphosphate and sodium tripolyphosphate hexahydrate, potassium tripolyphosphate, and mixed sodium and potassium tripolyphosphate, etc.). It is contemplated that any suitable builder will find use in the present invention, including those known in the art (See e.g., EP 2 100 949).

**[0279]** In some embodiments, builders for use herein include phosphate builders and non-phosphate builders. In some embodiments, the builder is a phosphate builder. In some embodiments, the builder is a non-phosphate builder. If present, builders are used in a level of from 0.1% to 80%, or from 5 to 60%, or from 10 to 50% by weight of the composition. In some embodiments the product comprises a mixture of phosphate and non-phosphate builders. Suitable phosphate builders include mono-phosphates, di-phosphates, tri-polyphosphates or oligomeric-polyphosphates, including the alkali metal salts of these compounds, including the sodium salts. In some embodiments, a builder can be sodium tripolyphosphate (STPP). Additionally, the composition can comprise carbonate and/or citrate, preferably citrate that helps to achieve a neutral pH composition of the invention. Other suitable non-phosphate builders include homopolymers and copolymers of polycarboxylic acids and their partially or completely neutralized salts, monomeric polycarboxylic acids and hydroxycarboxylic acids and their salts. In some embodiments, salts of the above mentioned compounds include the ammonium and/or alkali metal salts, i.e. the lithium, sodium, and potassium salts, including sodium salts. Suitable polycarboxylic acids include acyclic, alicyclic, hetero-cyclic and aromatic carboxylic acids, wherein in some

embodiments, they can contain at least two carboxyl groups which are in each case separated from one another by, in some instances, no more than two carbon atoms.

**[0280]** In some embodiments, the cleaning compositions of the present invention contain at least one chelating agent. Suitable chelating agents include, but are not limited to copper, iron and/or manganese chelating agents and mixtures thereof. In embodiments in which at least one chelating agent is used, the cleaning compositions of the present invention comprise from about 0.1% to about 15% or even from about 3.0% to about 10% chelating agent by weight of the subject cleaning composition.

**[0281]** In some still further embodiments, the cleaning compositions provided herein contain at least one deposition aid. Suitable deposition aids include, but are not limited to, polyethylene glycol, polypropylene glycol, polycarboxylate, soil release polymers such as polytelephthalic acid, clays such as kaolinite, montmorillonite, atapulgit, illite, bentonite, halloysite, and mixtures thereof.

**[0282]** As indicated herein, in some embodiments, anti-redeposition agents find use in some embodiments of the present invention. In some embodiments, non-ionic surfactants find use. For example, in automatic dishwashing embodiments, non-ionic surfactants find use for surface modification purposes, in particular for sheeting, to avoid filming and spotting and to improve shine. These non-ionic surfactants also find use in preventing the re-deposition of soils. In some embodiments, the anti-redeposition agent is a non-ionic surfactant as known in the art (See e.g., EP 2 100 949). In some embodiments, the non-ionic surfactant can be ethoxylated nonionic surfactants, epoxy-capped poly(oxyalkylated) alcohols and amine oxides surfactants.

**[0283]** In some embodiments, the cleaning compositions of the present invention include one or more dye transfer inhibiting agents. Suitable polymeric dye transfer inhibiting agents include, but are not limited to, polyvinylpyrrolidone polymers, polyamine N-oxide polymers, copolymers of N-vinylpyrrolidone and N-vinylimidazole, polyvinylloxazolidones and polyvinylimidazoles or mixtures thereof. In embodiments in which at least one dye transfer inhibiting agent is used, the cleaning compositions of the present invention comprise from about 0.0001% to about 10%, from about 0.01% to about 5%, or even from about 0.1% to about 3% by weight of the cleaning composition.

**[0284]** In some embodiments, silicates are included within the compositions of the present invention. In some such embodiments, sodium silicates (e.g., sodium disilicate, sodium metasilicate, and crystalline phyllosilicates) find use. In some embodiments, silicates are present at a level of from about 1% to about 20%. In some embodiments, silicates are present at a level of from about 5% to about 15% by weight of the composition.

**[0285]** In some still additional embodiments, the cleaning compositions of the present invention also contain dispersants. Suitable water-soluble organic materials include, but are not limited to the homo- or co-polymeric acids or their salts, in which the polycarboxylic acid comprises at least two carboxyl radicals separated from each other by not more than two carbon atoms.

**[0286]** In some further embodiments, the enzymes used in the cleaning compositions are stabilized by any suitable technique. In some embodiments, the enzymes employed herein are stabilized by the presence of water-soluble sources of calcium and/or magnesium ions in the finished compositions

that provide such ions to the enzymes. In some embodiments, the enzyme stabilizers include oligosaccharides, polysaccharides, and inorganic divalent metal salts, including alkaline earth metals, such as calcium salts, such as calcium formate. It is contemplated that various techniques for enzyme stabilization will find use in the present invention. For example, in some embodiments, the enzymes employed herein are stabilized by the presence of water-soluble sources of zinc (II), calcium (II) and/or magnesium (II) ions in the finished compositions that provide such ions to the enzymes, as well as other metal ions (e.g., barium (II), scandium (II), iron (II), manganese (II), aluminum (III), Tin (II), cobalt (II), copper (II), nickel (II), and oxovanadium (IV)). Chlorides and sulfates also find use in some embodiments of the present invention. Examples of suitable oligosaccharides and polysaccharides (e.g., dextrans) are known in the art (See e.g., WO 07/145964). In some embodiments, reversible protease inhibitors also find use, such as boron-containing compounds (e.g., borate, 4-formyl phenyl boronic acid) and/or a tripeptide aldehyde find use to further improve stability, as desired.

**[0287]** In some embodiments, bleaches, bleach activators and/or bleach catalysts are present in the compositions of the present invention. In some embodiments, the cleaning compositions of the present invention comprise inorganic and/or organic bleaching compound(s). Inorganic bleaches include, but are not limited to perhydrate salts (e.g., perborate, percarbonate, perphosphate, persulfate, and persilicate salts). In some embodiments, inorganic perhydrate salts are alkali metal salts. In some embodiments, inorganic perhydrate salts are included as the crystalline solid, without additional protection, although in some other embodiments, the salt is coated. Any suitable salt known in the art finds use in the present invention (See e.g., EP 2 100 949).

**[0288]** In some embodiments, bleach activators are used in the compositions of the present invention. Bleach activators are typically organic peracid precursors that enhance the bleaching action in the course of cleaning at temperatures of 60° C. and below. Bleach activators suitable for use herein include compounds which, under perhydrolysis conditions, give aliphatic peroxyoxycarboxylic acids having preferably from about 1 to about 10 carbon atoms, in particular from about 2 to about 4 carbon atoms, and/or optionally substituted perbenzoic acid. Additional bleach activators are known in the art and find use in the present invention (See e.g., EP 2 100 949).

**[0289]** In addition, in some embodiments and as further described herein, the cleaning compositions of the present invention further comprise at least one bleach catalyst. In some embodiments, the manganese triazacyclononane and related complexes find use, as well as cobalt, copper, manganese, and iron complexes. Additional bleach catalysts find use in the present invention (See e.g., U.S. Pat. Nos. 4,246,612, 5,227,084, 4,810,410, WO 99/06521, and EP 2 100 949).

**[0290]** In some embodiments, the cleaning compositions of the present invention contain one or more catalytic metal complexes. In some embodiments, a metal-containing bleach catalyst finds use. In some embodiments, the metal bleach catalyst comprises a catalyst system comprising a transition metal cation of defined bleach catalytic activity, (e.g., copper, iron, titanium, ruthenium, tungsten, molybdenum, or manganese cations), an auxiliary metal cation having little or no bleach catalytic activity (e.g., zinc or aluminum cations), and a sequester having defined stability constants for the catalytic and auxiliary metal cations, particularly ethylenedi-

aminetetraacetic acid, ethylenediaminetetra (methylene-phosphonic acid) and water-soluble salts thereof are used (See e.g., U.S. Pat. No. 4,430,243). In some embodiments, the cleaning compositions of the present invention are catalyzed by means of a manganese compound. Such compounds and levels of use are well known in the art (See e.g., U.S. Pat. No. 5,576,282). In additional embodiments, cobalt bleach catalysts find use in the cleaning compositions of the present invention. Various cobalt bleach catalysts are known in the art (See e.g., U.S. Pat. Nos. 5,597,936 and 5,595,967) and are readily prepared by known procedures.

**[0291]** In some additional embodiments, the cleaning compositions of the present invention include a transition metal complex of a macropolycyclic rigid ligand (MRL). As a practical matter, and not by way of limitation, in some embodiments, the compositions and cleaning processes provided by the present invention are adjusted to provide on the order of at least one part per hundred million of the active MRL species in the aqueous washing medium, and in some embodiments, provide from about 0.005 ppm to about 25 ppm, more preferably from about 0.05 ppm to about 10 ppm, and most preferably from about 0.1 ppm to about 5 ppm, of the MRL in the wash liquor.

**[0292]** In some embodiments, transition-metals in the instant transition-metal bleach catalyst include, but are not limited to manganese, iron and chromium. MRLs also include, but are not limited to special ultra-rigid ligands that are cross-bridged (e.g., 5,12-diethyl-1,5,8,12-tetraazabicyclo [6.6.2]hexadecane). Suitable transition metal MRLs are readily prepared by known procedures (See e.g., WO 2000/32601, and U.S. Pat. No. 6,225,464).

**[0293]** In some embodiments, the cleaning compositions of the present invention comprise metal care agents. Metal care agents find use in preventing and/or reducing the tarnishing, corrosion, and/or oxidation of metals, including aluminum, stainless steel, and non-ferrous metals (e.g., silver and copper). Suitable metal care agents include those described in EP 2 100 949, WO 9426860 and WO 94/26859). In some embodiments, the metal care agent is a zinc salt. In some further embodiments, the cleaning compositions of the present invention comprise from about 0.1% to about 5% by weight of one or more metal care agent.

**[0294]** In some embodiments, the cleaning composition is a high density liquid (HDL) composition having a variant metalloprotease polypeptide protease. The HDL liquid laundry detergent can comprise a deterative surfactant (10%-40%) comprising anionic deterative surfactant (selected from a group of linear or branched or random chain, substituted or unsubstituted alkyl sulphates, alkyl sulphonates, alkyl alkoxyated sulphate, alkyl phosphates, alkyl phosphonates, alkyl carboxylates, and/or mixtures thereof); and optionally non-ionic surfactant (selected from a group of linear or branched or random chain, substituted or unsubstituted alkyl alkoxyated alcohol, for example a C<sub>8</sub>-C<sub>18</sub> alkyl ethoxyated alcohol and/or C<sub>6</sub>-C<sub>12</sub> alkyl phenol alkoxyates), optionally wherein the weight ratio of anionic deterative surfactant (with a hydrophilic index (HIC) of from 6.0 to 9) to non-ionic deterative surfactant is greater than 1:1.

**[0295]** The composition can comprise optionally, a surfactancy boosting polymer consisting of amphiphilic alkoxyated grease cleaning polymers (selected from a group of alkoxyated polymers having branched hydrophilic and hydrophobic properties, such as alkoxyated polyalkylenimines in the range of 0.05 wt %-10 wt %) and/or random

graft polymers (typically comprising of hydrophilic backbone comprising monomers selected from the group consisting of: unsaturated C<sub>1</sub>-C<sub>6</sub> carboxylic acids, ethers, alcohols, aldehydes, ketones, esters, sugar units, alkoxy units, maleic anhydride, saturated polyalcohols such as glycerol, and mixtures thereof; and hydrophobic side chain(s) selected from the group consisting of: C<sub>4</sub>-C<sub>25</sub> alkyl group, polypropylene, polybutylene, vinyl ester of a saturated C—C<sub>6</sub> mono-carboxylic acid, C<sub>1</sub>-C<sub>6</sub> alkyl ester of acrylic or methacrylic acid, and mixtures thereof).

**[0296]** The composition can comprise additional polymers such as soil release polymers (include anionically end-capped polyesters, for example SRP1, polymers comprising at least one monomer unit selected from saccharide, dicarboxylic acid, polyol and combinations thereof, in random or block configuration, ethylene terephthalate-based polymers and copolymers thereof in random or block configuration, for example Repel-o-tex SF, SF-2 and SRP6, Texcare SRA100, SRA300, SRN100, SRN170, SRN240, SRN300 and SRN325, Marloquest SL), anti-redeposition polymers (0.1 wt % to 10 wt %, include carboxylate polymers, such as polymers comprising at least one monomer selected from acrylic acid, maleic acid (or maleic anhydride), fumaric acid, itaconic acid, aconitic acid, mesaconic acid, citraconic acid, methylenemalonic acid, and any mixture thereof, vinylpyrrolidone homopolymer, and/or polyethylene glycol, molecular weight in the range of from 500 to 100,000 Da); cellulosic polymer (including those selected from alkyl cellulose, alkyl alkoxyalkyl cellulose, carboxyalkyl cellulose, alkyl carboxyalkyl cellulose examples of which include carboxymethyl cellulose, methyl cellulose, methyl hydroxyethyl cellulose, methyl carboxymethyl cellulose, and mixtures thereof) and polymeric carboxylate (such as maleate/acrylate random copolymer or polyacrylate homopolymer).

**[0297]** The composition can further comprise saturated or unsaturated fatty acid, preferably saturated or unsaturated C<sub>12</sub>-C<sub>24</sub> fatty acid (0 wt % to 10 wt %); deposition aids (examples for which include polysaccharides, preferably cellulosic polymers, poly diallyl dimethyl ammonium halides (DADMAC), and co-polymers of DADMAC with vinyl pyrrolidone, acrylamides, imidazoles, imidazolium halides, and mixtures thereof, in random or block configuration, cationic guar gum, cationic cellulose such as cationic hydroxyethyl cellulose, cationic starch, cationic polyacrylamides, and mixtures thereof).

**[0298]** The composition can further comprise dye transfer inhibiting agents examples of which include manganese phthalocyanine, peroxidases, polyvinylpyrrolidone polymers, polyamine N-oxide polymers, copolymers of N-vinylpyrrolidone and N-vinylimidazole, polyvinylloxazolidones and polyvinylimidazoles and/or mixtures thereof; chelating agents examples of which include ethylene-diamine-tetraacetic acid (EDTA); diethylene triamine penta methylene phosphonic acid (DTPMP); hydroxy-ethane diphosphonic acid (HEDP); ethylenediamine N,N'-disuccinic acid (EDDS); methyl glycine diacetic acid (MGDA); diethylene triamine penta acetic acid (DTPA); propylene diamine tetracetic acid (PDTA); 2-hydroxypyridine-N-oxide (HPNO); or methyl glycine diacetic acid (MGDA); glutamic acid N,N-diacetic acid (N,N-dicarboxymethyl glutamic acid tetrasodium salt (GLDA); nitrilotriacetic acid (NTA); 4,5-dihydroxy-m-benzenedisulfonic acid; citric acid and any salts thereof; N-hydroxyethylethylenediaminetri-acetic acid (HEDTA), triethylenetetraaminehexaacetic acid (TTHA),

N-hydroxyethyliminodiacetic acid (HEIDA), dihydroxyethylglycine (DHEG), ethylenediaminetetrapropionic acid (EDTP) and derivatives thereof.

[0299] The composition can further comprise enzymes (0.01 wt % active enzyme to 0.03 wt % active enzyme) selected from a group of acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageenases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1, 4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxxygenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoloxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases, pullulanases, reductases, rhamnogalacturonases, beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases, and any mixture thereof. The composition may comprise an enzyme stabilizer (examples of which include polyols such as propylene glycol or glycerol, sugar or sugar alcohol, lactic acid, reversible protease inhibitor, boric acid, a boric acid derivative, e.g., an aromatic borate ester, or a phenyl boronic acid derivative such as 4-formylphenyl boronic acid, peptides or formate).

[0300] The composition can further comprise silicone or fatty-acid based suds suppressors; heuing dyes, calcium and magnesium cations, visual signaling ingredients, anti-foam (0.001 wt % to about 4.0 wt %), and/or structurant/thickener (0.01 wt % to 5 wt %, selected from the group consisting of diglycerides and triglycerides, ethylene glycol distearate, microcrystalline cellulose, cellulose based materials, microfiber cellulose, biopolymers, xanthan gum, gellan gum, and mixtures thereof).

[0301] Suitable detergent surfactants also include cationic detergent surfactants (selected from a group of alkyl pyridinium compounds, alkyl quarternary ammonium compounds, alkyl quarternary phosphonium compounds, alkyl ternary sulphonium compounds, and/or mixtures thereof); zwitterionic and/or amphoteric detergent surfactants (selected from a group of alkanolamine sulfo-betaines); ampholytic surfactants; semi-polar non-ionic surfactants and mixtures thereof.

[0302] The composition can be any liquid form, for example a liquid or gel form, or any combination thereof. The composition may be in any unit dose form, for example a pouch.

[0303] In some embodiments, the cleaning composition is a high density powder (HDD) composition having a variant metalloprotease polypeptide protease. The HDD powder laundry detergent can comprise a detergent surfactant including anionic detergent surfactants (selected from a group of linear or branched or random chain, substituted or unsubstituted alkyl sulphates, alkyl sulphonates, alkyl alkoxyated sulphate, alkyl phosphates, alkyl phosphonates, alkyl carboxylates and/or mixtures thereof), non-ionic detergent surfactant (selected from a group of linear or branched or random chain, substituted or unsubstituted C<sub>8</sub>-C<sub>18</sub> alkyl ethoxylates, and/or C<sub>6</sub>-C<sub>12</sub> alkyl phenol alkoxyates), cationic detergent surfactants (selected from a group of alkyl pyridinium compounds, alkyl quarternary ammonium compounds, alkyl quarternary phosphonium compounds, alkyl ternary sulphonium compounds, and mixtures thereof), zwitterionic and/or amphoteric detergent surfactants (selected from a group of

alkanolamine sulfo-betaines); ampholytic surfactants; semi-polar non-ionic surfactants and mixtures thereof; builders (phosphate free builders [for example zeolite builders examples of which include zeolite A, zeolite X, zeolite P and zeolite MAP in the range of 0 wt % to less than 10 wt %]; phosphate builders [examples of which include sodium tri-polyphosphate in the range of 0 wt % to less than 10 wt %]; citric acid, citrate salts and nitrilotriacetic acid or salt thereof in the range of less than 15 wt %); silicate salt (sodium or potassium silicate or sodium meta-silicate in the range of 0 wt % to less than 10 wt %, or layered silicate (SKS-6)); carbonate salt (sodium carbonate and/or sodium bicarbonate in the range of 0 wt % to less than 10 wt %); and bleaching agents (photobleaches, examples of which include sulfonated zinc phthalocyanines, sulfonated aluminum phthalocyanines, xanthenes dyes, and mixtures thereof; hydrophobic or hydrophilic bleach activators (examples of which include dodecanoyl oxybenzene sulfonate, decanoyl oxybenzene sulfonate, decanoyl oxybenzoic acid or salts thereof, 3,5,5-trimethyl hexanoyl oxybenzene sulfonate, tetraacetyl ethylene diamine-TAED, and nonanoyloxybenzene sulfonate-NOBS, nitrile quats, and mixtures thereof; hydrogen peroxide; sources of hydrogen peroxide (inorganic perhydrate salts examples of which include mono or tetra hydrate sodium salt of perborate, percarbonate, persulfate, persphosphate, or persilicate); preformed hydrophilic and/or hydrophobic peracids (selected from a group consisting of percarboxylic acids and salts, percarbonic acids and salts, perimidic acids and salts, peroxymonosulfuric acids and salts) & mixtures thereof and/or bleach catalyst (such as imine bleach boosters examples of which include iminium cations and polyions; iminium zwitterions; modified amines; modified amine oxides; N-sulphonyl imines; N-phosphonyl imines; N-acyl imines; thiadiazole dioxides; perfluoroimines; cyclic sugar ketones and mixtures thereof; metal-containing bleach catalyst for example copper, iron, titanium, ruthenium, tungsten, molybdenum, or manganese cations along with an auxiliary metal cations such as zinc or aluminum and a sequester such as ethylenediaminetetraacetic acid, ethylenediaminetetra(methylenephosphonic acid) and water-soluble salts thereof).

[0304] The composition can further comprise enzymes selected from a group of acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageenases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1, 4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxxygenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoloxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases, pullulanases, reductases, rhamnogalacturonases, beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases and any mixture thereof.

[0305] The composition can further comprise additional detergent ingredients including perfume microcapsules, starch encapsulated perfume accord, hueing agents, additional polymers including fabric integrity and cationic polymers, dye lock ingredients, fabric-softening agents, brighteners (for example C.I. Fluorescent brighteners), flocculating

agents, chelating agents, alkoxyated polyamines, fabric deposition aids, and/or cyclodextrin.

**[0306]** In some embodiments, the cleaning composition is an automatic dishwashing (ADW) detergent composition having a metalloprotease of the present invention. The ADW detergent composition can comprise two or more non-ionic surfactants selected from a group of ethoxylated non-ionic surfactants, alcohol alkoxyated surfactants, epoxy-capped poly(oxyalkylated) alcohols, or amine oxide surfactants present in amounts from 0 to 10% by weight; builders in the range of 5-60% comprising either phosphate (mono-phosphates, di-phosphates, tri-polyphosphates or oligomeric-polyphosphates, preferred sodium tripolyphosphate-STPP or phosphate-free builders [amino acid based compounds, examples of which include MGDA (methyl-glycine-diacetic acid), and salts and derivatives thereof, GLDA (glutamic-N, Ndiacetic acid) and salts and derivatives thereof, IDS (iminodisuccinic acid) and salts and derivatives thereof, carboxy methyl inulin and salts and derivatives thereof and mixtures thereof, nitrilotriacetic acid (NTA), diethylene triamine penta acetic acid (DTPA), B-alaninediacetic acid (B-ADA) and their salts], homopolymers and copolymers of poly-carboxylic acids and their partially or completely neutralized salts, monomeric polycarboxylic acids and hydroxycarboxylic acids and their salts in the range of 0.5% to 50% by weight; sulfonated/carboxylated polymers (provide dimensional stability to the product) in the range of about 0.1% to about 50% by weight; drying aids in the range of about 0.1% to about 10% by weight (selected from polyesters, especially anionic polyesters optionally together with further monomers with 3 to 6 functionalities which are conducive to polycondensation, specifically acid, alcohol or ester functionalities, polycarbonate-, polyurethane- and/or polyurea-polyorganosiloxane compounds or precursor compounds thereof of the reactive cyclic carbonate and urea type); silicates in the range from about 1% to about 20% by weight (sodium or potassium silicates for example sodium disilicate, sodium meta-silicate and crystalline phyllosilicates); bleach-inorganic (for example perhydrate salts such as perborate, percarbonate, perphosphate, persulfate and persilicate salts) and organic (for example organic peroxyacids including diacyl and tetraacylperoxides, especially diperoxydodecanedioc acid, diperoxytetradecanedioc acid, and diperoxyhexadecanedioc acid); bleach activators-organic peracid precursors in the range from about 0.1% to about 10% by weight; bleach catalysts (selected from manganese triazacyclononane and related complexes, Co, Cu, Mn and Fe bispyridylamine and related complexes, and pentamine acetate cobalt(III) and related complexes); metal care agents in the range from about 0.1% to 5% by weight (selected from benzotriazoles, metal salts and complexes, and/or silicates); enzymes in the range from about 0.01 to 5.0 mg of active enzyme per gram of automatic dishwashing detergent composition (acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageenases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1, 4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxygenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoloxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases, pullulanases, reductases, rhamnolacturonases,

beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases, and any mixture thereof); and enzyme stabilizer components (selected from oligosaccharides, polysaccharides and inorganic divalent metal salts).

**[0307]** The metalloproteases are normally incorporated into the detergent composition at a level of from 0.000001% to 5% of enzyme protein by weight of the composition, or from 0.00001% to 2%, or from 0.0001% to 1%, or from 0.001% to 0.75% of enzyme protein by weight of the composition.

Metalloprotease Polypeptides of the Present Invention for Use in Animal Feed

**[0308]** In a further aspect of the invention, the metalloprotease polypeptides of the present invention can be used as a component of an animal feed composition, animal feed additive and/or pet food comprising a metalloprotease and variants thereof. The present invention further relates to a method for preparing such an animal feed composition, animal feed additive composition and/or pet food comprising mixing the metalloprotease polypeptide with one or more animal feed ingredients and/or animal feed additive ingredients and/or pet food ingredients. Furthermore, the present invention relates to the use of the metalloprotease polypeptide in the preparation of an animal feed composition and/or animal feed additive composition and/or pet food.

**[0309]** The term "animal" includes all non-ruminant and ruminant animals. In a particular embodiment, the animal is a non-ruminant animal, such as a horse and a mono-gastric animal. Examples of mono-gastric animals include, but are not limited to, pigs and swine, such as piglets, growing pigs, sows; poultry such as turkeys, ducks, chicken, broiler chicks, layers; fish such as salmon, trout, tilapia, catfish and carps; and crustaceans such as shrimps and prawns. In a further embodiment the animal is a ruminant animal including, but not limited to, cattle, young calves, goats, sheep, giraffes, bison, moose, elk, yaks, water buffalo, deer, camels, alpacas, llamas, antelope, pronghorn and nilgai.

**[0310]** In the present context, it is intended that the term "pet food" is understood to mean a food for a household animal such as, but not limited to, dogs, cats, gerbils, hamsters, chinchillas, fancy rats, guinea pigs; avian pets, such as canaries, parakeets, and parrots; reptile pets, such as turtles, lizards and snakes; and aquatic pets, such as tropical fish and frogs.

**[0311]** The terms "animal feed composition," "feedstuff" and "fodder" are used interchangeably and can comprise one or more feed materials selected from the group comprising a) cereals, such as small grains (e.g., wheat, barley, rye, oats and combinations thereof) and/or large grains such as maize or sorghum; b) by products from cereals, such as corn gluten meal, Distillers Dried Grain Solubles (DDGS) (particularly corn based Distillers Dried Grain Solubles (cDDGS), wheat bran, wheat middlings, wheat shorts, rice bran, rice hulls, oat hulls, palm kernel, and citrus pulp; c) protein obtained from sources such as soya, sunflower, peanut, lupin, peas, fava beans, cotton, canola, fish meal, dried plasma protein, meat and bone meal, potato protein, whey, copra, sesame; d) oils and fats obtained from vegetable and animal sources; e) minerals and vitamins.

### Metalloprotease Polypeptides of the Present Invention for Use in Textile Desizing

[0312] Also contemplated are compositions and methods of treating fabrics (e.g., to desize a textile) using a metalloprotease polypeptide of the present invention. Fabric-treating methods are well known in the art (see, e.g., U.S. Pat. No. 6,077,316). For example, the feel and appearance of a fabric can be improved by a method comprising contacting the fabric with a metalloprotease in a solution. The fabric can be treated with the solution under pressure.

[0313] A metalloprotease of the present invention can be applied during or after the weaving of a textile, or during the desizing stage, or one or more additional fabric processing steps. During the weaving of textiles, the threads are exposed to considerable mechanical strain. Prior to weaving on mechanical looms, warp yarns are often coated with sizing starch or starch derivatives to increase their tensile strength and to prevent breaking. A metalloprotease of the present invention can be applied during or after the weaving to remove these sizing starch or starch derivatives. After weaving, the metalloprotease can be used to remove the size coating before further processing the fabric to ensure a homogeneous and wash-proof result.

[0314] A metalloprotease of the present invention can be used alone or with other desizing chemical reagents and/or desizing enzymes to desize fabrics, including cotton-containing fabrics, as detergent additives, e.g., in aqueous compositions. An amylase also can be used in compositions and methods for producing a stonewashed look on indigo-dyed denim fabric and garments. For the manufacture of clothes, the fabric can be cut and sewn into clothes or garments, which are afterwards finished. In particular, for the manufacture of denim jeans, different enzymatic finishing methods have been developed. The finishing of denim garment normally is initiated with an enzymatic desizing step, during which garments are subjected to the action of proteolytic enzymes to provide softness to the fabric and make the cotton more accessible to the subsequent enzymatic finishing steps. The metalloprotease can be used in methods of finishing denim garments (e.g., a "bio-stoning process"), enzymatic desizing and providing softness to fabrics, and/or finishing process.

### Metalloprotease Polypeptides of the Present Invention for Use in Paper Pulp Bleaching

[0315] The metalloprotease polypeptides described herein find further use in the enzyme aided bleaching of paper pulps such as chemical pulps, semi-chemical pulps, kraft pulps, mechanical pulps or pulps prepared by the sulfite method. In general terms, paper pulps are incubated with a metalloprotease polypeptide of the present invention under conditions suitable for bleaching the paper pulp.

[0316] In some embodiments, the pulps are chlorine free pulps bleached with oxygen, ozone, peroxide or peroxyacids. In some embodiments, the metalloprotease polypeptides are used in enzyme aided bleaching of pulps produced by modified or continuous pulping methods that exhibit low lignin contents. In some other embodiments, the metalloprotease polypeptides are applied alone or preferably in combination with xylanase and/or endoglucanase and/or alpha-galactosidase and/or cellobiohydrolase enzymes.

### Metalloprotease Polypeptides of the Present Invention for Use in Protein Degradation

[0317] The metalloprotease polypeptides described herein find further use in the enzyme aided removal of proteins from animals and their subsequent degradation or disposal, such as feathers, skin, hair, hide, and the like. In some instances, immersion of the animal carcass in a solution comprising a metalloprotease polypeptide of the present invention can act to protect the skin from damage in comparison to the traditional immersion in scalding water or the defeathering process. In one embodiment, feathers can be sprayed with an isolated metalloprotease polypeptide of the present invention under conditions suitable for digesting or initiating degradation of the plumage. In some embodiments, a metalloprotease of the present invention can be used, as above, in combination with an oxidizing agent.

[0318] In some embodiments, removal of the oil or fat associated with raw feathers is assisted by using a metalloprotease polypeptide of the present invention. In some embodiments, the metalloprotease polypeptides are used in compositions for cleaning the feathers as well as to sanitize and partially dehydrate the fibers. In some other embodiments, the metalloprotease polypeptides are applied in a wash solution in combination with 95% ethanol or other polar organic solvent with or without a surfactant at about 0.5% (v/v).

[0319] In yet other embodiments, the disclosed metalloprotease polypeptides find use in recovering protein from plumage. The disclosed metalloprotease polypeptides may be used alone or in combination in suitable feather processing and proteolytic methods, such as those disclosed in PCT/EP2013/065362, PCT/EP2013/065363, and PCT/EP2013/065364, which are hereby incorporated by reference. In some embodiments, the recovered protein can be subsequently used in animal or fish feed.

### EXPERIMENTAL

[0320] The claimed invention is described in further detail in the following examples which are not in any way intended to limit the scope of the invention as claimed.

#### Example 1.1

##### Cloning of *Paenibacillus* sp. Metalloprotease PspPro3

[0321] A strain of *Paenibacillus* sp. was selected as a potential source for enzymes which may be useful for various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hours. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus* sp. strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus* sp. encodes a metalloprotease and the sequence of this gene, called PspPro3, is provided in SEQ ID NO: 1. The corresponding protein encoded by the PspPro3 gene is shown in SEQ ID NO: 2. At the N-terminus, the protein has a signal peptide with a length of 26 amino acids as

predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) Nature Methods, 8:785-786). The presence of a signal sequence suggests that PspPro3 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) Journal of Bacteriology, 173 (21): 6820-6825). The predicted mature region of PspPro3 protein is shown on SEQ ID NO: 3.

**[0322]** The nucleotide sequence of the PspPro3 gene isolated from *Paenibacillus* sp. is set forth as SEQ ID NO: 1. The sequence encoding the predicted native signal peptide is shown in italics:

```
ATGTTAATGAAAAAGTATGGGTTTCGCTTCTTGAGGAGCGATGTTATT
AGGGTCTGTAGCGTCTGGTGCATCAGCAGCGGAGAGTTCGGTTTCGGGGC
CGGCTCAGCTTACGCCAACCTTCATGCCGAACAATGGAAAGCACCTTCA
TCGGTATCGGGTATGACATCGTATGGAGCTATTTAAATCGGCAAAAGAA
AACGTTGCTGGGTACGGACAGCACCAGTGTCCGTGATCAATCCGTATCG
TAGATCGCACAAAGCGACAAATCCGGCGTGAGCCATTATCGGCTGAAGCAA
TATGTAACGGAATTCCCGTATATGGAGCTGAACAGACCATTATGTTGGG
CAAAATCCGGTGAAGTGACCTTTATCTGGGAGCCGTGATTACTGAGGATC
AGCAAGAAGAAGCTACGCAAGGTACAACCTCGAAAATCAGCGCTTCTGAA
CGGGTCCATACCCGATATCAGGAGGCAGCTACACGGGTTCAAGCCCTCC
TACCTCCGATGATACGATTCTAAAGATGCGGAGGAGCCAAGCAGTGTA
GCAAAGACACTTACTCCGAAGCAGCTAACAACGAAAAACGAGTTCTGTT
GAAAAGGACAAGCTCAGCCTTGAGAAAGCGGCTGACCTGAAAGATAGCAA
AATTGAAGCGGTGGAGGCAGAGCCAAACTCCATTGCCAAAATCGCCAACC
TGCAGCCTGAGGTAGATCCTAAAGCCGAACATATTTCTATGCGAAGGGC
GATGCATTGCAGCTGGTTTATGTGACTGAGGTTAATATTTTGCAGCCTGC
GCCGCTGCGTACACGCTACATCATTGACGCAATGATGGCAAAATCGTAT
CCAGTATGACATCATTAAAGGACGACAGGCACAGGCAAGGTGTACTC
GGTGATACAAAACATTCAACACTACTGCTTCCGGCAGCAGCTACCAGTT
AAGAGATACGACTCGCGGGAATGGAATCGTGACTTACACGGCTTCAACC
GTCAAAGCATCCCAGGTACGATCTGACCGATGCCGATAACGTATGGAAT
GATCCAGCCGGCTGGATGCCACGCTTATGCAGCCAAAACCTATGATTA
TTATAAGGAAAAGTTCAATCGCAACAGCATTGACGGACGAGGCCTGCAGC
TCGGTTCGACAGTTCATTACGGCAATCGTTACAACAACGCCTTCTGGAAC
GGCTCCCAAATGACTTATGGAGACGGAGACGGCACCACTTATTCGCTTT
TAGCGGTGATCCGGATGATGTTGGTTCATGAACTCACACACGGTGTACGG
AGTACTACTCCAATTTGGAATATACGGAGAATCCGGTGCCTTGAACGAG
GCCTTCCGGACATCATCGCAATGACATCCAGCGTAAAAACTGGCTTGT
AGGCGATGATATTTACAGCCACGCATTGCGGGTATGCACTTCGTTCTA
TGTTCAATCCTACGCTGTACGATCAACCGGATCACTATTCGAACTTGATC
AGAGGCAGCTCCGATAACGGCGGCTTATACGAACAGCGGTATTATAAA
```

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```
TAAAGCCTATTATCTGTTGGCACAGGGCGCACCTTCCATGGTGTAACTG
TCAATGGGATTGGCCCGATGCAGCGGTTCAAATTTACTACAGCGCCTTT
ACGAACTACCTGACTTCTTCTTCTGACTTCTCCAATGCACGTGATGCCGT
TGTACAAGCGGCAAAAGATCTCTACGGCGGAGCTCGGCACAAGTACCG
CAGCAGCCAAATCTTTTGTGCTGTAGGCGTTAAC
```

**[0323]** The amino acid sequence of the PspPro3 precursor protein is set forth as SEQ ID NO: 2. The predicted signal peptide is shown in italics, and the predicted pro-peptide is shown in underlined text:

```
MLMKVWVSLGGLLGSVAGSASAAESSVSGPAQLTPTFHAEQWKAPS
SVSGDDIVWSYLNQKKTLLGTDSTSVRDQFRIVDRSDKSGVSHYRLKQ
YVNGIPVYGAEQTIHVGKSGEVTSYLGAVITEDQOEEATQGTTPKISASE
AVHTAYQEAATRVQALPTSDDTISKDAEPEPSSVKDITYSEAANNKGTSSV
EKDKLSLEKAADLKDSEAVEAEPNISIAKIANLQPEVDPKAEIFYAKG
DALQLVYVTEVNILOPAPLRTRYIIDANDGKIVSQYDIINEATGTGKGLV
GDTKTFNTTASGSSYQLRDTTRNGIVTYTASNRSIPGTILTDADNVWN
DPAGVDAHAYAAKTYDYYKEKFNRSIDGRGLQLRSTVHYGNRYNNAFWN
GSQMTYGDGDGTTFIASGDPDVVGHETHGVTEYTSNLEYYGESGALNE
AFSDIIGNDIQRKNLVGDDIYTPRIAGDALRSMNPTLYDQPDHYSNLY
RGSSDNGGVHTNSGIINKAYYLLAQGGTFHGVTVNGIGRDAAVQIYYSAF
TNYLTSSSDFSNARDAVVQAAKLDYGASSAQATAAAKSFDAVGVN
```

**[0324]** The amino acid sequence of the predicted mature form of PspPro3 is set forth as SEQ ID NO: 3:

```
ATGTGKGLGDTKTFNTTASGSSYQLRDTTRNGIVTYTASNRSIPGTI
LTDADNVWNPAGVDAHAYAAKTYDYYKEKFNRSIDGRGLQLRSTVHYG
NRYNNAFWNNGSQMTYGDGDGTTFIASGDPDVVGHETHGVTEYTSNLEY
YGESGALNEAFSDIIGNDIQRKNLVGDDIYTPRIAGDALRSMNPTLYD
QPDHYSNLYRGSSDNGGVHTNSGIINKAYYLLAQGGTFHGVTVNGIGRDA
AVQIYYSAFTNYLTSSSDFSNARDAVVQAAKLDYGASSAQATAAAKSFDA
VGVN
```

### Example 1.2

#### Expression of *Paenibacillus* sp. Metalloprotease PspPro3

**[0325]** The DNA sequence of the propeptide-mature form of PspPro3 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Genaray (Shanghai, China), resulting in plasmid pGX085 (AprE-PspPro3) (FIG. 1.1). Ligation of the gene encoding the PspPro3 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *Bacillus subtilis* AprE signal sequence and the 5' end of the predicted PspPro3 native propeptide. The gene has an alternative start codon (GTG). As



shown in FIG. 1.1, pGX085(AprE-PspPro3) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature region of PspPro3 (SEQ ID NO: 4). The translation product of the synthetic AprE-PspPro3 gene is shown in SEQ ID NO: 5.

[0326] *B. subtilis* cells (degU<sup>ts</sup>32, ΔscoC) were transformed with the pGX085(AprE-PspPro3) plasmid and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>). The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to a 150 mL Q Sepharose High Performance column pre-equilibrated with the loading buffer above and PspPro3 was then eluted from the column via the loading buffer supplemented with a linear NaCl gradient from 0 to 0.7 M. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

[0327] The nucleotide sequence of the synthesized PspPro3 gene in plasmid pGX085(AprE-PspPro3) is depicted in SEQ ID NO: 4. The sequence encoding the predicted native signal peptide is shown in *italics* and the region encoding the three residue addition (AGK) is shown in **bold**:

*GTGAGAAGCAAAAAATTGTGGATCAGCTTGTGTTTGGCGTTAACGTTAAT*  
*CTTTACGATGGCGTTTCAGCAACATGAGCGCGCAGGCTGCTGGAAAAGCAG*  
 AATCATCAGTGTGACGACCGGCTCAGCTTACGCCGACGTTTCATGCAGAG  
 CAGTGGAAAGCACCGAGCAGCGTTAGCGGAGATGACATCGTGTGGAGCTA  
 CCTGAACAGACAGAAGAAAACGCTTCTTGGCACGGACAGCACGAGCGTCA  
 GAGACAGTTCAGAATCGTGGATAGAACAAGCGACAAAAGCGGCGTCAGC  
 CATTATAGACTGAAGCAGTATGTGAACGGAATCCCGGTTTATGGCGCAGA  
 ACAAACATCATGTGCGAAAGAGCGCGAAGTTACGAGCTATCTGGCGG  
 CGGTTATTACAGAGGACCAGCAAGAGGAGGCTACACAAGGCACGACCCG  
 AAAATTTACGATCAGAGGCGAGTTCATACGGCTACCAAGAAGCTGCAAC  
 GAGAGTTCAGCCCTGCCTACGTGATGATACATCAGCAAAGACGCTG  
 AGGAACCTAGCTCAGTTAGCAAGGACACGTATAGCGAAGCGCGAACCAAT  
 GGCAAGACGTCAAGCGTGGAAAAGACAAAGCTTCACTGGAGAAGGCCGC  
 TGATCTGAAAGACTCAAAGATCGAGGCTGTGGAAGCGGAACCGAATAGCA  
 TTGCAAGATTGCCAACCTGCAACCGGAGGTGGACCCGAAGCGGAGCTG  
 TATTCTACGCTAAAGCGATGCACTGCAACTGGTTTACGTACCGGAGGT  
 TAACATCCTGCAGCCGCGACCGCTTAGAACGAGATACATCATTGACGCGA  
 ACGACGGCAAGATCGTGGACGATACGACATTATCAACGAGGCCACGGGA  
 ACGGGCAAGGGAGTCTTGGCGACACGAAGACATTAATAACAACGGCTC  
 AGGCTCATCATACCGCTGAGAGACACGACGAGAGGCCAACGGAATCGTCA

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CGTACACGGCTAGCAATAGACAGAGCATTCCGGGCACAATCCTTACGGAC  
 GCAGACAATGTGTGGAATGACCCGGCAGGCGTGGACGCACATGCCTACGC  
 AGCGAAGACGTACGACTACTACAAGGAGAAGTTCAACAGAAAACAGCATCG  
 ACGGAAGAGGACTGCAACTTAGAAGCACGGTGCATTACGGCAACAGATAC  
 AACACGCTTTCTGGAACGGCAGCCAAATGACGTATGGAGACGGCGATGG  
 AACACGTTTATCGCATTCAGGGACCCCTGACGTTGTGGACATGAAC  
 TGACGCATGGAGTCACAGAATACACGAGCAATCTGGAGTATTACGGAGAA  
 TCAGGCGCACTTAATGAGGCCCTCAGCGACATCATCGAAAACGACATCCA  
 GAGAAAGAACTGGCTGGTTGGCGATGATATCTACACGCCGAGAATTGCGG  
 GCGACGCGCTGAGATCAATGAGCAACCTACGCTGTACGATCAGCCGGAT  
 CATTACAGCAACCTGTATAGAGGCTCAAGCGATAATGGCGGCGTGCATAC  
 AACACGCGCATCATCAACAAAGCCTATTATCTGCTGGCGCAAGGCGGCA  
 CATTCCATGGCGTTACAGTTAATGGCATTGGCAGAGACGAGCCGTCGAG  
 ATCTACTACAGCGCATTACGAATTACCTGACATCAAGCAGCGACTTTTC  
 AAATGCAAGAGATGCAGTGGTGCAGGCGGCTAAAGACCTTTATGGAGCTT  
 CAAGCGCTCAGGCCACAGCTGCGGCAAAAAGCTTCGACGCGGTTGGAGTG  
 AAT

[0328] The amino acid sequence of the PspPro3 precursor protein expressed from plasmid pGX085(AprE-PspPro3) is depicted in SEQ ID NO: 5. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) shown in **bold** and the predicted pro-peptide is shown in underlined text.:

*MRSKRLWISLLFALTLIFTMAFSNMSAQ**AGK**AESSVSGPAQLTPTFHAE*  
QWKAPSSVSGDDIVVSYLNRQKTLTLDSTSVRDFRIVDRSDKSGVS  
HYRLKQYVNGIPVYGAEQTIHVKSGEVTSYLGAVIDEDQEEATQGTTP  
KISASEAVHTAYQEAATRVQALPTSDDTISKDAEPEPSSVSKDITYEAANN  
GKTSVEKDKLSLEKAADLKD SKIEAVEAEPNSIAKIANLQPEVDPKAEI  
YFYAKGDALQLVYVTEVNILOPAPLRTRYIIDANDGKIVSQYDIINEATG  
 TGKVLGDTKTFNTTASGSSYQLRDTTRNGIVTYTASNRSIPGTILTD  
 ADNVWNPAGVDAHAYAAKTYDYKKEKFNRSIDGRGLQLRSTVHYGNRY  
 NNAFVNGSQMTYGDGDTTFIAFSGDDVVGHELTHGVTEYTSNLEYIGE  
 SGALNEAFSDIIGNDIQRKNLWVGDDIYTPRIAGDALRSMNPTLYDQPD  
 HYSNLYRGSDDNGGVHTNSGIINKAYYLLAQGGTFPHGVTVNGIGRDAAVQ  
 IYYSFTNYLTSSDFSNARDAVVQAADLYGASSAQATAAAKSFDAV  
 GVN

[0329] The amino acid sequence of the PspPro3 recombinant protein isolated from *Bacillus subtilis* culture was determined by tandem mass spectrometry, and shown below. It is the same as predicted and depicted in SEQ ID NO: 3.

ATGTGKGVLDGDTKTFNTTASGSSYQLRDTTRGNGIVTYTASNRSQIPGTI  
 LTDADNVVNDPAGVDAHAYAAKTYDYKKEKFNRSIDGRGLQLRSTVHYG  
 NRYNNAFWNGSQMTYIGDGDGTTFFIAFSGDPDVGHELTGVTYETSINLEY  
 YGESGALNEAFSDIIGNDIQRKNWLVGDDIYTPRIAGDALRSMSNPTLYD  
 QPDHYSNLYRGSSDNGGVHTNSGIINKAYYLLAQGGTFHGVTVNGIGRDA  
 AVQIYYSAFTNYLTSSSDFSNARDAVVQAADLYGASSAQATAAAKSFDA  
 VGVN

### Example 1.3

#### Proteolytic Activity of Metalloprotease PspPro3

**[0330]** The proteolytic activity of purified PspPro3 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ L of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ L of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ L of 5% Trichloroacetic Acid (TCA). Following equilibration (5 mM at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ L supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a Spectra-Max 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of duplicate assays, and the value varies no more than 5%. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (FIG. 1.2) indicates that PspPro3 is an active protease.

### Example 1.4

#### pH Profile of Metalloprotease PspPro3

**[0331]** With azo-casein as the substrate, the pH profile of PspPro3 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ L of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2  $\mu$ L diluted enzyme (250 ppm in Milli-Q H<sub>2</sub>O) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ L of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 1.3. Enzyme activity at each pH was reported as relative activity where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 1.3, the optimal pH of PspPro3 is 7.5, with greater than 70% of maximal activity retained between pH 5.5 and 9.

### Example 1.5

#### Temperature Profile of Metalloprotease PspPro3

**[0332]** The temperature profiles of PspPro3 were analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assay. The enzyme sample and azo-casein substrate were prepared as in Example 3. Prior to the reaction, 50  $\mu$ L of 1.5% azo-casein and 45  $\mu$ L Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ L PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ L of diluted PspPro3 (100 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ L of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 1.3. The activity was reported as relative activity where the activity at the optimal temperature was set to be 100%. The tested temperatures were 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of triplicate assays. The data in FIG. 1.4 suggest that PspPro3 showed an optimal temperature at 50° C., and retained greater than 70% of its maximal activity between 45° C and 60° C.

### Example 1.6

#### Cleaning Performance of Metalloprotease PspPro3 in Automatic Dishwashing (ADW) Conditions

**[0333]** The cleaning performance of PspPro3 in automatic dishwashing (ADW) conditions was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 or 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified PspPro3 were diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent (composition shown in Table 1.1) with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1), in the absence or presence of a bleach component (Peracid N,N-phthaloylaminoperoxycaproic acid-PAP). To initiate the reaction, 180  $\mu$ L of AT detergent buffered at pH 6 or 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ L of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ L of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm ( $A_{405}$ ) (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ L water. Following the addition of 180  $\mu$ L of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliter of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 for PspPro3 in AT detergent, in the absence or presence of bleach, is shown in FIGS. 5A and 5B, respectively.

TABLE 1.1

| Composition of AT dish detergent formula with bleach       |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| PAP (peracid N,N-phthaloylaminoperoxyacetic acid)          | 0.057                 |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

## Example 1.7

## Cleaning Performance of Metalloprotease PspPro3 in Laundry Conditions

**[0334]** The cleaning performance of PspPro3 protein in liquid laundry detergent was tested using EMPA-116 (cotton soiled with blood/milk/ink) microswatches (obtained from CFT Vlaardingen, The Netherlands) at pH 8.2 using a commercial detergent. Prior to the reaction, purified PspPro3 protein samples were diluted with a dilution solution (10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol) to the desired concentrations; and the commercial detergent (Tide®, Clean Breeze®, Proctor & Gamble, USA, purchased September 2011) was incubated at 95° C. for 1 hour to inactivate the enzymes present in the detergent. Proteolytic assays were subsequently performed to confirm the inactivation of proteases in the commercial detergent. The heat treated detergent was further diluted with 5 mM HEPES (pH 8.2) to a final concentration of 0.788 g/L. Meanwhile, the water hardness of the buffered liquid detergent was adjusted to 103 ppm (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1). The specific conductivity of the buffered detergent was adjusted to either 0.62 mS/cm (low conductivity) or 3.5 mS/cm (high conductivity) by adjusting the NaCl concentration in the buffered detergent. To initiate the reaction, 190 µl of either the high or low conductivity buffered detergent was added to a 96-MTP containing the EMPA-116 microswatches, followed by the addition of 10 µl of diluted enzyme (or the dilution solution as blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 20 min at 32° C. and 1150 rpm. After incubation, 150 µl of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 600 nm using a spectrophotometer, which indicates the protease activity on the model stain; and Net A<sub>600</sub> was subsequently calculated by subtracting the A<sub>600</sub> of the blank control from that of the enzyme. Dose response for the cleaning of EMPA-116 microswatches in liquid laundry detergent at high or low conductivity is shown in FIG. 1.6.

## Example 1.8

## Comparison of PspPro3 to Other Metalloproteases

**[0335]** A. Identification of Homologous Proteases

**[0336]** Homologs were identified by a BLAST search (Altschul et al., Nucleic Acids Res, 25:3389-402, 1997)

against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The mature protein amino acid sequence for PspPro3 (SEQ ID NO: 3) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 1.2A and 1.2B provide a list of sequences with the percent identity to PspPro3. The length in Table 1.2 refers to the entire sequence length of the homologous proteases.

TABLE 1.2A

| List of sequences with percent identity to PspPro3 protein identified from the NCBI non-redundant protein database |                |  |        |
|--|----------------|--|--------|
| Accession #  | PID to PspPro3 | Organism   | Length |
| ZP_10321515.1  | 55             | <i>Bacillus macauensis</i> ZFHKF-1                     | 552    |
| AAC43402.1   | 57             | <i>Alicyclobacillus acidocaldarius</i>                 | 546    |
| P00800   | 57             | <i>Bacillus thermoproteolyticus</i>                    | 548    |
| AAA22621.1   | 58             | <i>Geobacillus stearothermophilus</i>                  | 548    |
| ZP_01862236.1  | 59             | <i>Bacillus</i> sp. SG-1                               | 560    |
| YP_002884504.1   | 59             | <i>Exiguobacterium</i> sp. AT1b                        | 509    |
| AEI46285.1   | 60             | <i>Paenibacillus mucilaginosus</i> KNP414              | 525    |
| ZP_08093424  | 60             | <i>Planococcus donghaensis</i> MPA1U2                  | 553    |
| ZP_10324092.1  | 61             | <i>Bacillus macauensis</i> ZFHKF-1                     | 533    |
| YP_006792441.1   | 61             | <i>Exiguobacterium antarcticum</i> B7                  | 498    |
| AAK69076.1   | 63             | <i>Bacillus thuringiensis</i> serovar <i>finitimus</i> | 566    |
| NP_976992.1  | 64             | <i>Bacillus cereus</i> ATCC 10987                      | 566    |
| ZP_04321694  | 64             | <i>Bacillus cereus</i>                                 | 566    |
| BAA06144   | 64             | <i>Lactobacillus</i> sp.                               | 566    |
| ZP_10241029.1  | 78             | <i>Paenibacillus peoriae</i> KCTC 3763                 | 599    |
| YP_005073223   | 93             | <i>Paenibacillus terrae</i> HPL-003                    | 591    |
| YP_003872179   | 94             | <i>Paenibacillus polymyxa</i> E681                     | 592    |
| ZP_09775364  | 100            | <i>Paenibacillus</i> sp. <i>Aloe</i> -11               | 593    |

TABLE 1.2B

| List of sequences with percent identity to PspPro3 protein identified from the Genome Quest Patent database |                |                                    |        |
|---|----------------|------------------------------------|--------|
| Patent #  | PID to PspPro3 | Organism                           | Length |
| US20120107907-0184  | 57.88          | <i>Bacillus caldolyticus</i>       | 319    |
| US20120107907-0177  | 57.88          | <i>Bacillus caldolyticus</i>       | 544    |
| WO2012110563-0002   | 58.2           | <i>Bacillus caldolyticus</i>       | 319    |
| EP2390321-0176  | 58.52          | <i>Bacillus stearothermophilus</i> | 548    |
| US6518054-0002  | 59.22          | <i>Bacillus</i> sp.                | 316    |
| WO2004011619-0044   | 60.6           | Empty                              | 507    |
| WO2004011619-0047   | 62.14          | Empty                              | 532    |
| WO2004011619-0046   | 62.26          | Empty                              | 536    |
| WO2012110563-0004   | 63.02          | <i>Bacillus megaterium</i>         | 320    |
| JP2002272453-0003   | 63.67          | Empty                              | 562    |
| US8114656-0186  | 64.24          | <i>Bacillus brevis</i>             | 304    |
| WO2012110562-0005   | 64.52          | <i>Bacillus cereus</i>             | 320    |
| WO2007044993-0178   | 64.74          | <i>Bacillus thuringiensis</i>      | 566    |
| EP2178896-0184  | 65.38          | <i>Bacillus anthracis</i>          | 566    |
| WO2012110563-0005   | 65.48          | <i>Bacillus cereus</i>             | 320    |
| JP1995184649-0001   | 65.71          | <i>Lactobacillus</i> sp.           | 566    |
| US5962264-0004  | 65.81          | Empty                              | 566    |
| US20120107907-0185  | 66.13          | <i>Bacillus cereus</i>             | 317    |
| US8114656-0187  | 93.36          | <i>Bacillus polymyxa</i>           | 302    |
| JP2005229807-0019   | 93.38          | <i>Paenibacillus polymyxa</i>      | 566    |

**[0337]** B. Alignment of Homologous Protease Sequences

**[0338]** The amino acid sequence for mature PspPro3 (SEQ ID NO: 3) was aligned with thermolysin (P00800, *Bacillus*

*thermoproteolyticus*) and protease from *Paenibacillus* sp. Aloe-11 (ZP\_09775364) using CLUSTALW software (Thompson et al., Nucleic Acids Research, 22:4673-4680, 1994) with the default parameters. FIG. 1.7 shows the alignment of PspPro3 with these protease sequences.

#### [0339] C. Phylogenetic Tree

[0340] A phylogenetic tree for full length sequence of PspPro3 (SEQ ID NO: 2) was built using sequences of representative homologs from Tables 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. MolBiol. Evol. 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylogenetic tree printer software (<http://iubio.bio.indiana.edu/tree-app/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 1.8.

#### Example 1.9

##### Terg-o-Tometer Performance Evaluation of PspPro3

[0341] The wash performance of PspPro3 was tested in a laundry detergent application using a Terg-o-Tometer (Instrument Marketing Services, Inc, Fairfield, N.J.). The performance evaluation was conducted at 32° C. and 16° C. The soil load consisted of two of each of the following stain swatches: EMPA116 Blood, Milk, Ink on cotton (Test materials AG, St. Gallen, Switzerland), EMPA117 Blood, Milk, Ink on polycotton (Test materials AG, St. Gallen, Switzerland), EMPA112 Cocoa on cotton (Test materials AG, St. Gallen, Switzerland), and CFT C-10 Pigment, Oil, and Milk content

on cotton (Center for Testmaterials BV, Vlaardingen, Netherlands), plus extra white interlock knit fabric to bring the total fabric load to 40 g per beaker of the Terg-o-Tometer, which was filled with 1 L of deionized water. The water hardness was adjusted to 6 grains per gallon, and the pH in the beaker was buffered with 5 mM HEPES, pH 8.2. Heat inactivated Tide Regular HDL (Procter & Gamble), a commercial liquid detergent purchased in a local US supermarket, was used at 0.8 g/L. The detergent was inactivated before use by treatment at 92° C. in a water bath for 2-3 hours followed by cooling to room temperature. Heat inactivation of commercial detergents serves to destroy the activity of enzymatic components while retaining the properties of the non-enzymatic components. Enzyme activity in the heat inactivated detergent was measured using the Suc-AAPF-pNA assay for measuring protease activity. The Purafect® Prime HA, (Genencor Int'l) and PspPro3 proteases were each added to final concentrations of 0, 0.2, 0.5, 1, and 1.5 ppm. The wash time was 12 minutes. After the wash treatment, all swatches were rinsed for 3 minutes and machine-dried at low heat.

[0342] Four of each types of swatch were measured before and after treatment by optical reflectance using a Tristimulus Minolta Meter CR-400. The difference in the L, a, b values was converted to total color difference (dE), as defined by the CIE-LAB color space. Cleaning of the stains is expressed as percent stain removal index (% SRI) by taking a ratio between the color difference before and after washing, and comparing it to the difference of unwashed soils (before wash) to unsoiled fabric, and averaging the eight values obtained by reading two different regions of each washed swatch and is reported in Tables 1.9A and 1.9B as Average % SRI (dE) ±95CI. Table 1.9A summarizes the cleaning performance of PspPro3 at 32° C. and Table 1.9B at 16° C.

TABLE 1.9A

| Cleaning performance of PspPro3 at 32° C. |                   |              |            |              |                   |              |            |              |
|---|-------------------|--------------|------------|--------------|-------------------|--------------|------------|--------------|
| ppm enzyme                                | Average           | 95CI         | Average    | 95CI         | Average           | 95CI         | Average    | 95CI         |
|   | % SRI (dE)        | [% SRI (dE)] | % SRI (dE) | [% SRI (dE)] | % SRI (dE)        | [% SRI (dE)] | % SRI (dE) | [% SRI (dE)] |
| EMPA-116                                  |                   |              |            | EMPA-117     |                   |              |            |              |
|   | Purafect Prime HA |              | PspPro3    |              | Purafect Prime HA |              | SprPro3    |              |
| 0   | 0.19              | 0.01         | 0.19       | 0.01         | 0.17              | 0.01         | 0.17       | 0.01         |
| 0.2                                       | 0.27              | 0.02         | 0.27       | 0.02         | 0.25              | 0.03         | 0.30       | 0.02         |
| 0.5                                       | 0.28              | 0.03         | 0.31       | 0.01         | 0.30              | 0.03         | 0.31       | 0.02         |
| 1   | 0.30              | 0.01         | 0.32       | 0.02         | 0.35              | 0.02         | 0.34       | 0.03         |
| 1.5                                       | 0.31              | 0.02         | 0.31       | 0.01         | 0.37              | 0.01         | 0.37       | 0.03         |
| EMPA-112                                  |                   |              |            | CFT C-10     |                   |              |            |              |
|   | Purafect Prime HA |              | PspPro3    |              | Purafect Prime HA |              | PspPro3    |              |
| 0   | 0.11              | 0.03         | 0.11       | 0.03         | 0.07              | 0.01         | 0.07       | 0.01         |
| 0.2                                       | 0.11              | 0.05         | 0.18       | 0.04         | 0.12              | 0.01         | 0.11       | 0.01         |
| 0.5                                       | 0.13              | 0.04         | 0.17       | 0.03         | 0.15              | 0.01         | 0.16       | 0.01         |
| 1   | 0.18              | 0.03         | 0.19       | 0.04         | 0.17              | 0.01         | 0.21       | 0.01         |
| 1.5                                       | 0.19              | 0.03         | 0.18       | 0.04         | 0.18              | 0.01         | 0.23       | 0.01         |

TABLE 1.9B

| Cleaning performance of PspPro3 at 16° C. |                    |                    |                    |                    |                    |                    |                    |                    |
|---|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| ppm enzyme                                | Purafect Prime HA  |                    | PspPro3            |                    | Purafect Prime HA  |                    | PspPro3            |                    |
|   | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] |
|   | EMPA-116           |                    |                    |                    | EMPA-117           |                    |                    |                    |
| 0   | 0.15               | 0.02               | 0.15               | 0.02               | 0.13               | 0.01               | 0.13               | 0.01               |
| 0.2                                       | 0.19               | 0.02               | 0.20               | 0.03               | 0.15               | 0.02               | 0.15               | 0.02               |
| 0.5                                       | 0.20               | 0.02               | 0.19               | 0.02               | 0.21               | 0.02               | 0.20               | 0.02               |
| 1   | 0.24               | 0.04               | 0.21               | 0.02               | 0.22               | 0.02               | 0.20               | 0.01               |
| 1.5                                       | 0.19               | 0.02               | 0.25               | 0.04               | 0.23               | 0.03               | 0.20               | 0.01               |
|   | EMPA-112           |                    |                    |                    | CFT C-10           |                    |                    |                    |
| 0   | 0.08               | 0.03               | 0.08               | 0.03               | 0.04               | 0.08               | 0.04               | 0.08               |
| 0.2                                       | 0.12               | 0.02               | 0.09               | 0.01               | 0.06               | 0.12               | 0.06               | 0.09               |
| 0.5                                       | 0.08               | 0.02               | 0.11               | 0.02               | 0.08               | 0.08               | 0.08               | 0.11               |
| 1   | 0.11               | 0.02               | 0.10               | 0.03               | 0.08               | 0.11               | 0.09               | 0.10               |
| 1.5                                       | 0.13               | 0.02               | 0.11               | 0.03               | 0.11               | 0.13               | 0.10               | 0.11               |

Example 2.1

Cloning of Metalloprotease PspPro2 from *Paenibacillus* sp.

[0343] A strain of *Paenibacillus* sp. was selected as a potential source for enzymes which may be useful for various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus* sp. strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus* sp. encodes a metalloprotease and the sequence of this gene, called PspPro2, is provided in SEQ ID NO: 6. The corresponding protein encoded by the PspPro2 gene is shown in SEQ ID NO: 7. At the N-terminus, the protein has a signal peptide with a length of 24 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) Nature Methods, 8:785-786). The presence of a signal sequence suggests that PspPro2 is a secreted enzyme. The propeptide region of PspPro2 was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) Journal of Bacteriology, 173 (21): 6820-6825). The predicted mature region of PspPro2 is shown in SEQ ID NO: 8.

[0344] The nucleotide sequence of the PspPro2 gene isolated from *Paenibacillus* sp. is set forth as SEQ ID NO: 6. The sequence encoding the predicted native signal peptide is shown in italics:

*ATGAAAAAGTATGGGTTTCACTTCTTGAGGAGCGATGTTATTAGGGC*  
*TGTAGCACCAGGTGCATCAGCAGCAGAGCATTCTGTTCTGATCCTACTC*

-continued

AGCTAACACCGACCTTTCACGCCGAGCAATGGAGGCTCCTCCACGGTA  
 ACCGGCGACAATATTGTATGGAGCTATTTGAATCGACAAAAGAAAACCTT  
 ATTGAATACAGACAGCACCAGTGTGCGTGATCAGTTCGCCATCATTGATC  
 GTACAAGCGACAAATCCGGTGAAGCCATTATCGGCTCAAGCAATATGTA  
 AACGGGATCCCCGTATATGGGGCTGAACAGACCATTTCATGTGAACAACGC  
 CGGTAAGTAACCTCTTATTTGGGTGCTGTCAATTCAGAGGATCAGCAGC  
 AAGACCGGACCGAAGATACCACTCCAAAAATCAGCGGACTGAAGCCGTT  
 TATACCGCATATGCAGAAGCCGCTGCCCGGATTCAATCCTTCCCTTCCAT  
 CAATGATAGTCTTTCTGAGGCTAGTGAGGAACAAGGGAGTGAGAATCAAG  
 GCAATGAGATTCAAAAATTTGGGATTAAGAGCAGTGTAAAGTAATGACACT  
 TACGCAGAGGCGCATAACAACGTACTTTTAACCCCGTTGACCAAGCAGA  
 GCAAAGTTACATTGCCAAAATGCTAATCTGGAGCCAAGTGTAGAGCCCA  
 AAGCAGAATTATACATCTATCCAGATGGTGAAGACTACACGACTGGTTTAT  
 GTAACAGAGGTTAATATTCTTGAACCTGCGCCTCTGCGCACACGCTACTT  
 CATTGATGCGAAAACCGGCAAAATCGTATTCCAGTATGACATCCTCAACC  
 ACGCAACAGGCACCGGCCCGGGCGTGGATGGCAAAACAAAATCATTTACG  
 ACTACAGCTTCAGGCAACCGGTATCAGTTGAAAGACACGACTCGCAGCAA  
 TGGAAATCGTGACTTACACCGCTGGCAATCGCCAGACGACGCCAGGTACGA  
 TTTTGACCATACAGATAATGTATGGGAGGACCTGCGGCTGTTGATGCC  
 CATGCCATCGCCATTAACCTATGACTATTATAAGAATAAATTCGGTCCG  
 CGACAGTATTGATGGAGCTGGCATGCAAAATTCGTTCCAGAGTCCATTACG  
 GCAAAAAATATAACAATGCCTTCTGGAACGGCTCGCAAATGACCTACGGA  
 GACGGAGACGGGTCCACATTTACCTTCTTCAGCGCGATCCCAGTGTGCT  
 GGGGCATGAGCTCACCCACGGCGTACCGAGTTCACCTCCAATTTGGAGT

- continued

ATTATGGTGAGTCCGGTGCATGAAACGAAGCCTTCTCGGATATTATCGGT  
 AATGATATAGATGGCACCAGTTGGCTTCTTGGCGACGGCATTATACGCC  
 TAATATTCAGGCGACGCTCTGCGTTCCTGTCCGATCCTACACGATTCG  
 GCCAGCCGGATCACTACTCCAATTTCTATCCGGACCCCAACAATGATGAT  
 GAAGCGGGAGTCCATACGAACAGCGGTATTATCAACAAAGCCTATTATTT  
 GCTGGACAAGCGGTACGTCCTCATGGTAAACGGTAACTGGTATCGGAC  
 GCGAAGCGGCTGTATTCACTTACATAATGCCTTTACCAACTATTGACC  
 TCTACCTCCAACCTTCTTAACGCACGCGCTGCTGTTATACAGGCAGCCAA  
 GGATTTTATGGTCTGATTGCTGGCAGTAACAGTGTATTCAATCCT  
 TTGATGCGGTAGGAATCAAA

[0345] The amino acid sequence of the PspPro2 precursor protein is set forth as SEQ ID NO: 7. The predicted signal peptide is shown in italics, and the predicted pro-peptide is shown in underlined text:

*MKKVVVSLGGAMLLGAVAPGASAAEHSVPDPTQLTPTFHAEQWKAPSTV*  
TGDNIVWSYLNROKKTLLNLDSTSVRDQFRIIDRTSDKSGASHYRLKQYV  
NGIPVYGAEQTIHVNAGKVTSYLGAVIDEQQDATEDTTPKISATEAV  
YTAYAEAAARIQSFPINDSLSEASEEQGSENOGNEIQNIGIKSSVSNDT  
YAEAHNNVLLTPVDQAEQSYIAKIANLEPSVEPKAELYIYPDGETTRLVY  
VTEVNILEPAPLRTRYFIDAKTGKIVFOYDILNHATGTGRGVDGKTKSFT  
 TTASGNRYQLKDTTRSNGIVTYTAGNRQTPGTILTDTDNVWEDPAAVDA  
 HAYAIKTYDYKKNKFRGRSDIDGRGMQIRSTVHYGKKYNNAFWNGSQMTYG  
 DGDGSTFTFFSGDPDVVGHETHGVTEFTSNLEYEGESGALNEAFSDIIG  
 NDIDGTSWLLGDGIYTPNIPGDALRSLSDPTRFGQPDHYSNFYDPDNNDD  
 EGGVHTNSGIINKAYYLLAQGGTSHGVTVTGIGREAAVFIYNAFTNYLT  
 STSNFSNARA AVIQAAKDFYGADSLAVTSAIQSPDAVGIK

[0346] The amino acid sequence of the predicted mature form of PspPro2 is set forth as SEQ ID NO: 8.

ATGTGRGVDGKTKSFTTASGNRYQLKDTTRSNGIVTYTAGNRQTPGTI  
 LTDTDNVWEDPAAVDAHAYAIKTYDYKKNKFRGRSDIDGRGMQIRSTVHYG  
 KKYNNAFWNGSQMTYGDGDGSTFTFFSGDPDVVGHETHGVTEFTSNLEY  
 YGESGALNEAFSDIIGNDIDGTSWLLGDGIYTPNIPGDALRSLSDPTRFG  
 QPDHYSNFYDPDNNDDGGVHTNSGIINKAYYLLAQGGTSHGVTVTGIGR  
 EAAVFIYNAFTNYLTSNFSNARA AVIQAAKDFYGADSLAVTSAIQSF  
 DAVGIK

### Example 2.2

#### Expression of *Paenibacillus* sp. Metalloprotease PspPro2

[0347] The DNA sequence of the propeptide-mature form of PspPro2 was synthesized and inserted into the *Bacillus*

*subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX084 (AprE-PspPro2) (FIG. 2.1). Ligation of this gene encoding the PspPro2 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *Bacillus subtilis* AprE signal sequence and the 5' end of the predicted PspPro2 native propeptide. The gene has an alternative start codon (GTG). As shown in FIG. 2.1, pGX084(AprE-PspPro2) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PspPro2, (SEQ ID NO: 9). The translation product of the synthetic AprE-PspPro2 gene is shown in SEQ ID NO: 10.

[0348] The pGX084(AprE-PspPro2) plasmid was transformed into *B. subtilis* cells (degU<sup>32</sup>, AscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

[0349] The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5) and 1 mM CaCl<sub>2</sub> using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to a 150 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above, PspPro2 was eluted from the column with a linear salt gradient from 0 to 0.5 M NaCl in the loading buffer. The corresponding active fractions were collected, concentrated and buffer-exchanged again into the loading buffer described above. The sample was loaded onto a 20 ml DEAE Fast Flow column pre-equilibrated with the same loading buffer. PspPro2 was eluted from the column with a linear salt gradient from 0 to 0.3 M NaCl in the loading buffer. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses. The nucleotide sequence of the synthesized PspPro2 gene in plasmid pGX084 (AprE-PspPro2) is depicted in SEQ ID NO: 9. The sequence encoding the predicted native signal peptide is shown in italics and the oligo-nucleotide encoding the three residue addition (AGK) is shown in bold:

*GTGAGAAGCAAAAAATTGTGGATCAGCTTGTGTTTTCGCTTAACTAAT*  
*CTTTACGATGGCGTTCAGCAACATGAGCGCGCAGGCTGCTGGAAAAGCAG*  
 AGCATTAGTTCCTGACCCGACGCAACTTACACCGACATTCATGTGTAG  
 CAGTGGAAAGGCACCGAGCACGGTACCGGGCACAACATCGTGTGAGGCTA  
 CCTGAACAGACAGAAAAAGACGCTGCTGAACACGGACTCAACGAGCGTGA  
 GAGACCAGTTCAGAATCATCGACAGAACGAGCGACAAAGTCAAGCGCGTCA  
 CATTATAGACTGAAGCAGTACGTGAACGGCATCCGGTCTACGGAGCCGA  
 GCAAACGATCCATGTGAATAATGCGGGCAAAGTTACATCATACTGGGCG  
 CCGTCATCTCAGAAGACCAGCAGCAAGATGCAACGGAGGATACAACACCG  
 AAGATCAGCGCCACAGAAGCGGTCTATACGGCTTACGCCAAGCGGTGTC

-continued

AAGAATCCAGAGCTTCCCGTCAATTAATGACAGCCTGAGCGAAGCATCAG  
 AGGAACAAGGACGAGACCAAGGCAATGAAATCCAAAACATCGGCATC  
 AAGAGCAGCGTGTCAAACGACACGTATGCGGAGGCTCATAACAACGTTCT  
 GCTGACACCGGTGATCAGGCCGAACAGAGCTATATTGCAAAGATCGCGA  
 ATCTGGAGCCGTCAGTCGAGCCGAAGCCGAGCTGTATATCTATCCGGAC  
 GCGGAGACGACGAGACTGGTGTACGTTACGGAGGTCAACATCCTTGAGCC  
 TCGCCGCTGAGAAACAAGATACTTTATCGACGCCAAGACGGGCAAGATCG  
 TGTTTCAGTACGATATCTGAACCATGCGACGGGAACAGGCAGAGCGCTG  
 GACGGCAAAAACAAATCATTACGACAAACGCAAGCGGCAACAGATACCA  
 GCTGAAGGACACAAACAGATCAATGGCATCGTCACATACCGGCCGAA  
 ATAGACAGACGACGCCGGGAACGATTCGACGGATACAGATAACGTGTGG  
 GAAGATCCGCGCAGCTGTGATGCACATGCATACGCGATCAAGACGTACGA  
 CTAATAACAAGAAATTCGGAAGAGATTCATCGATGGAAGAGGCATGC  
 AAATCAGATCAACGGTTCATTATGGCAAAAAGTACAACAATGCCTTCTGG  
 AACGGCAGCCAAATGACATACGCGCATGGAGACGGCTCAACGTTTACATT  
 CTTTTAGGGCGACCCGGACGTCGTCGGCCATGAACTGACGCATGGCGTTA  
 CAGAGTTCACGAGCAACCTGGAGTATTACGGCGAATCAGGCGCACTGAAT  
 GAGGCTTTCAGCGACATCATTGGCAACGACATTGATGGCACATCATGGCT  
 GCTTGGCGACGGCATTACACACCTAACATTCGGGGCATGCACTGAGAA  
 GCCTGTGACACCTACGAGATTCGGCCAACTGACCATTACAGCAACTTC  
 TACCCGATCCTAATAACGATGATGAGGGCGAGTGCATACGAACAGCGG  
 CATTATCAACAAGCGTACTATCTGCTGGCACAAGGCGGAACGTCACATG  
 GAGTGACGGTGACAGGAATCGGCAGAGAGCGGCGAGTGTATCTACTAC  
 AACGCCCTTCAAACTACCTGACGAGCAGTCAAATTCAGCAACGCTAG  
 AGCGCGGTCATCCAGGCAGCAAGGACTTTTATGGAGCAGACTACTGG  
 CAGTTACGTGACCAATTCAGTCATTTCGACGCGAGTGGAAATTAAG

[0350] The amino acid sequence of the PspPro2 precursor protein expressed from plasmid pGX084(AprE-PspPro2) is depicted in SEQ ID NO: 10. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) is shown in **bold**, and the predicted pro-peptide is shown in underlined text:

*MRSKKLWISLLFALTLIFTMAFSNMSAQAGKAEHSVPDPTQLTPTFHAEL*  
QWKAPSTVTGDNIVWSYLNQRQKTLNLNTDSTSVRDQFRIIDRTSDKSGAS  
HYRLKQYVNGIPVYGAETIHVNNAGKVTSYLGAVISEDQQDATEDTTP  
KISATEAVYTAYAEAAARIQSFP SINDSLSEASEEQSENQNEIQNIGI  
KSSVSNDTYAEAHNNVLLTPVDQAEQSYIAKIANLEPSVEPKAELIYPD  
GETTRLVYVTEVNILEPAPLRTYFIDAKTGKIVFOYDILNHATGTGRGV  
 DGKTKSFTTTASGNRYQLKDTTRSNGIVTYTAGNRQTPGTILTDNDVW  
 EDPAAVDAHAYAIKTYDYKKNKFRDSDIDGRGMQIRSTVHYGKKYNNAFW

-continued

NGSQMTYGDGDGSTFTFFSGDPDVVGHETHGVTEFTSNLEYGESGALN  
 EAFSDIIGNDIDGTSWLLGDGIYTPNIPGDALRSLSDPFRGQPDHYSNF  
 YPDPNNDDEGGVHTNSGIINKAYYLLAQQGTSHGVTVTGIGREAAVFIYY  
 NAFTNYLTSTSNFSNARAIVIQAADFYGADSLAVTSAIQSPDAVGIK  
 [0351] The amino acid sequence of the recombinant Psp-Pro2 protein isolated from *Bacillus subtilis* culture was determined by tandem mass spectrometry, and shown below. It is the same as predicted and depicted in SEQ ID NO: 8.

ATGTGRGVGDKTKSFTTTASGNRYQLKDTTRSNGIVTYTAGNRQTPGTI  
 LTDTDNDVWEDPAAVDAHAYAIKTYDYKKNKFRDSDIDGRGMQIRSTVHYG  
 KKYNNAFWNGSQMTYGDGDGSTFTFFSGDPDVVGHETHGVTEFTSNLEY  
 YGESGALNEAFSDIIGNDIDGTSWLLGDGIYTPNIPGDALRSLSDPFRG  
 QPDHYSNFIYPPNNDDEGGVHTNSGIINKAYYLLAQQGTSHGVTVTGIGR  
 EAAVFIYYNAFTNYLTSTSNFSNARAIVIQAADFYGADSLAVTSAIQSF  
 DAVGIK

### Example 2.3

#### Proteolytic Activity of Metalloprotease PspPro2

[0352] The proteolytic activity of purified PspPro2 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ l of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ l of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ l of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ l supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a Spectra-Max 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of duplicate assays, and the value varies no more than 5%. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assays with azo-casein as the substrate (FIG. 2.2) indicate that PspPro2 is an active protease.

### Example 2.4

#### pH Profile of Metalloprotease PspPro2

[0353] With azo-casein as the substrate, the pH profile of PspPro2 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with

2  $\mu$ l diluted enzyme (500 ppm in Milli-Q H<sub>2</sub>O) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 2.3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 10 and 11. Each result was the mean of triplicate assays. As shown in FIG. 2.3, the optimal pH of PspPro2 is 7.5 with greater than 70% of its maximal activity retained between pH 5.5 and 9.5.

#### Example 2.5

##### Temperature Profile of Metalloprotease PspPro2

**[0354]** The temperature profile of PspPro2 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assay. The enzyme sample and azo-casein substrate were prepared as in Example 2.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted PspPro2 (200 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40, 50, 60, 70, 80 and 90° C. Each result was the mean of triplicate assays. The data in FIG. 2.4 suggest that PspPro2 showed an optimal temperature at 50° C., and retained greater than 70% of its maximal activity between 40 and 65° C.

#### Example 2.6

##### Cleaning Performance of Metalloprotease PspPro2 in Automatic Dishwashing (ADW) Conditions

**[0355]** The cleaning performance of PspPro2 protein in automatic dishwashing (ADW) conditions was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified PspPro2 protein samples were diluted with the dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1), in the presence of a bleach component (Peracid N,N-phthaloylaminoperoxycaproic acid-PAP) (AT detergent composition shown in Table 1). To initiate the reaction, 180  $\mu$ l of the AT detergent solution at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ l of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ l water. Following the addition of 180

$\mu$ l of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain. Dose response for cleaning of PA-S-38 microswatches at pH 6 and pH 8 for PspPro2 in AT detergent in the presence of bleach, is shown in FIGS. 2.5A and 2.5B, respectively.

TABLE 2.1

| Composition of AT dish detergent with bleach               |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| PAP (peracid N,N-phthaloylaminoperoxycaproic acid)         | 0.057                 |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired pH value (pH 6 or 8) by the addition of 0.9M citric acid.

#### Example 2.7

##### Cleaning Performance of Metalloprotease PspPro2 in Laundry Conditions

###### A. Cleaning Performance in Liquid Laundry Detergent

**[0356]** The cleaning performance of PspPro2 protein in liquid laundry detergent was tested using EMPA-116 (cotton soiled with blood/milk/ink) microswatches (obtained from CFT Vlaardingen, The Netherlands) at pH 8.2 using a commercial detergent. Prior to the reaction, purified PspPro2 protein samples were diluted with a dilution solution (10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol) to the desired concentrations; and the commercial detergent (Tide®, Clean Breeze®, Proctor & Gamble, USA, purchased September 2011) was incubated at 95° C. for 1 hour to inactivate the enzymes present in the detergent. Proteolytic assays were subsequently performed to confirm the inactivation of proteases in the commercial detergent. The heat treated detergent was further diluted with 5 mM HEPES (pH 8.2) to a final concentration of 0.788 g/L. Meanwhile, the water hardness of the buffered liquid detergent was adjusted to 103 ppm (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1). The specific conductivity of the buffered detergent was adjusted to either 0.62 mS/cm (low conductivity) or 3.5 mS/cm (high conductivity) by adjusting the NaCl concentration in the buffered detergent. To initiate the reaction, 190  $\mu$ l of either the high or low conductivity buffered detergent was added to a 96-MTP containing the EMPA-116 microswatches, followed by the addition of 10  $\mu$ l of diluted enzyme (or the dilution solution as blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 20 min at 32° C. and 1150 rpm. After incubation, 150  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 600 nm using a spectro-



photometer, which indicates the protease activity on the model stain; and Net  $A_{600}$  was subsequently calculated by subtracting the  $A_{600}$  of the blank control from that of the enzyme. Dose response for the cleaning of EMPA-116 microswatches in liquid laundry detergent at high or low conductivity is shown in FIG. 2.6A.

#### B. Cleaning Performance in Powder Laundry Detergent

[0357] The cleaning performance of PspPro2 protein in powder laundry detergent was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) using a commercial detergent. Prior to the reaction, purified PspPro2 protein samples were diluted with a dilution solution (10 mM NaCl, 0.1 mM  $\text{CaCl}_2$ , 0.005% TWEEN® 80 and 10% propylene glycol) to the desired concentrations. The powder laundry detergent (Tide®, Bleach Free, Proctor & Gamble, China, purchased in December 2011) was dissolved in water with 103 ppm water hardness ( $\text{Ca}^{2+}:\text{Mg}^{2+}=3:1$ ) to a final concentration of 2 g/L (with conductivity of 2.3 mS/cm-low conductivity) or 5 g/L (with conductivity of 5.5 mS/cm-high conductivity). The detergents of different conductivities were subsequently heated in a microwave to near boiling in order to inactivate the enzymes present in the detergent. Proteolytic activity was measured following treatment to ensure that proteases in the commercial detergent had been inactivated. To initiate the reaction, 190  $\mu\text{l}$  of either the high or low conductivity heat-treated detergent was added to a 96-MTP containing the PA-S-38 microswatches, followed by the addition of 10  $\mu\text{l}$  of

diluted enzyme (or the dilution solution as blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 15 minutes at 32° C. and 1150 rpm. After incubation, 150  $\mu\text{l}$  of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm using a spectrophotometer, which indicates the protease activity on the model stain; and Net  $A_{405}$  was subsequently calculated by subtracting the  $A_{405}$  of the blank control from that of the enzyme. Dose response for the cleaning of PA-S-38 microswatches in powder laundry detergent at high or low conductivity is shown in FIG. 2.6B.

#### Example 2.8

##### Comparison of PspPro2 to Other Metalloproteases

##### Identification of Homologous Proteases

[0358] Homologs were identified by a BLAST search (Altschul et al., Nucleic Acids Res, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The mature protein amino acid sequence for PspPro2 (SEQ ID NO: 8) is used as query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 2.2A and 2.2B provide a list of sequences with the percent identity to PspPro2. The length in Table 2.2 refers to the entire sequence length of the homologous proteases.

TABLE 2.2A

| List of sequences with percent identity to PspPro2 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PspPro2 | Organism  | Length |
| AAB02774.1   | 55             | <i>Geobacillus stearothermophilus</i>                       | 552    |
| AAA22623.1   | 56             | <i>Bacillus caldolyticus</i>                                | 544    |
| P00800   | 56             | <i>Bacillus thermoproteolyticus</i>                         | 548    |
| YP_003670279.1   | 57             | <i>Geobacillus</i> sp. C56-T3                               | 546    |
| BAD60997.1   | 57             | <i>Bacillus megaterium</i>                                  | 562    |
| ZP_02326503.1  | 58             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> BRL-230010 | 520    |
| ZP_08640523.1  | 58             | <i>Brevibacillus laterosporus</i> LMG 15441                 | 564    |
| YP_003597483.1   | 58             | <i>Bacillus megaterium</i> DSM 319                          | 562    |
| ZP_09069025.1  | 59             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 520    |
| YP_001373863.1   | 59             | <i>Bacillus cytotoxicus</i> NVH 391-98                      | 565    |
| ZP_04149724.1  | 59             | <i>Bacillus pseudomycooides</i> DSM 12442                   | 566    |
| CAA43589.1   | 60             | <i>Brevibacillus brevis</i>                                 | 527    |
| ZP_10738945.1  | 60             | <i>Brevibacillus</i> sp. CF112                              | 528    |
| ZP_04216147.1  | 60             | <i>Bacillus cereus</i> Rock3-44                             | 566    |
| ZP_10575942.1  | 61             | <i>Brevibacillus</i> sp. BC25                               | 528    |
| YP_002770810.1   | 62             | <i>Brevibacillus brevis</i> NBRC 100599                     | 528    |
| ZP_08511445.1  | 63             | <i>Paenibacillus</i> sp. HGF7                               | 525    |
| ZP_09077634.1  | 64             | <i>Paenibacillus elgii</i> B69                              | 524    |
| ZP_09071078.1  | 67             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 529    |
| YP_003872180.1   | 73             | <i>Paenibacillus polymyxa</i> E681                          | 587    |
| YP_005073223.1   | 78             | <i>Paenibacillus terrae</i> HPL-003                         | 591    |
| ZP_09775364.1  | 78             | <i>Paenibacillus</i> sp. <i>Aloe</i> -11                    | 593    |
| YP_003948511.1   | 80             | <i>Paenibacillus polymyxa</i> SC2                           | 592    |
| YP_005073224.1   | 94             | <i>Paenibacillus terrae</i> HPL-003                         | 595    |
| ZP_10241029.1  | 96             | <i>Paenibacillus peoriae</i> KCTC 3763                      | 599    |
| ZP_09775365.1  | 100            | <i>Paenibacillus</i> sp. <i>Aloe</i> -11                    | 580    |

TABLE 2.2B

| List of sequences with percent identity to PspPro2 protein identified from the Genome Quest database |                |                                       |        |
|--|----------------|---------------------------------------|--------|
| Patent #   | PID to PspPro2 | Organism                              | Length |
| JP2002272453-0002  | 57.01          | <i>Bacillus megaterium</i>            | 562    |
| US6518054-0001   | 57.19          | <i>Bacillus</i> sp.                   | 319    |
| EP2390321-0177   | 57.19          | <i>Bacillus caldolyticus</i>          | 544    |
| US20120107907-0176   | 57.19          | <i>Bacillus stearothermophilis</i>    | 548    |
| WO9520663-0003   | 57.51          | empty                                 | 319    |
| WO2012110562-0003  | 57.51          | <i>Geobacillus stearothermophilus</i> | 319    |
| WO2012110563-0002  | 57.51          | <i>Bacillus caldolyticus</i>          | 319    |
| WO2004011619-0056  | 57.51          | empty                                 | 546    |
| WO2004011619-0003  | 57.51          | empty                                 | 546    |
| JP2002272453-0003  | 57.64          | empty                                 | 562    |
| US6518054-0002   | 57.88          | <i>Bacillus</i> sp.                   | 316    |
| EP2178896-0184   | 58.15          | <i>Bacillus anthracis</i>             | 566    |
| WO2012110563-0004  | 58.28          | <i>Bacillus megaterium</i>            | 320    |
| JP1995184649-0001  | 58.79          | <i>Lactobacillus</i> sp.              | 566    |
| JP1994014788-0003  | 58.84          | empty                                 | 317    |
| US8114656-0178   | 59.42          | <i>Bacillus thuringiensis</i>         | 566    |
| WO2012110562-0005  | 59.49          | <i>Bacillus cereus</i>                | 320    |
| US5962264-0004   | 59.81          | empty                                 | 566    |
| US20120107907-0185   | 59.81          | <i>Bacillus cereus</i>                | 317    |
| US20120107907-0179   | 59.81          | <i>Bacillus cereus</i>                | 566    |
| WO2012110563-0005  | 60.13          | <i>Bacillus cereus</i>                | 320    |
| EP2390321-0186   | 60.33          | <i>Bacillus brevis</i>                | 304    |
| JP2005229807-0018  | 78.62          | <i>Paenibacillus polymyxa</i>         | 566    |
| EP2390321-0187   | 79.21          | <i>Bacillus polymyxa</i>              | 302    |

### B. Alignment of Homologous Protease Sequences

[0359] The amino acid sequence of mature PspPro2 (SEQ ID NO: 8) was aligned with thermolysin (P00800, *Bacillus thermoproteolyticus*) and protease from *Paenibacillus* sp. Aloe-11 (ZP\_09775365.1) sequences using CLUSTALW software (Thompson et al., *Nucleic Acids Research*, 22:4673-4680, 1994) with the default parameters. FIG. 2.7 shows the alignment of PspPro2 with these protease sequences.

### C. Phylogenetic Tree

[0360] A phylogenetic tree for precursor PspPro2 protein sequence (SEQ ID NO: 7) was built using sequences of representative homologs from Table 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. *Mol Biol. Evol.* 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylogenetic tree printer software (<http://iubio.bio.indiana.edu/tree-app/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 2.8.

#### Example 3.1

##### Cloning of *Paenibacillus Humicus* Metalloprotease PhuPro2

[0361] A strain (DSM18784) of *Paenibacillus humicus* was selected as a potential source of enzymes which may be useful in various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from

Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus humicus* strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus humicus* encodes a metalloprotease and the sequence of this gene, called PhuPro2, is provided in SEQ ID NO: 11. The corresponding protein encoded by the PhuPro2 gene is shown in SEQ ID NO: 12. At the N-terminus, the protein has a signal peptide with a length of 23 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) *Nature Methods*, 8:785-786). The presence of a signal sequence suggests that PhuPro2 is a secreted enzyme. The propeptide region was predicted based on its protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) *Journal of Bacteriology*, 173 (21): 6820-6825). The predicted mature region of PhuPro2 protein is shown in SEQ ID NO: 13. The nucleotide sequence of the PhuPro2 gene isolated from *Paenibacillus humicus* is set forth as SEQ ID NO: 11. The sequence encoding the predicted native signal peptide is shown in italics:

```

ATGAAAAAATGATTCCCTACTCTGCTCGGTACCGTATTGCTGCTTTCTTC
CGCTTCGCTGTGCTGCTGAATCGCAAGCCTCGGAGCGGCCGGAACTC
CCGGGGTCAGCGTCGTGAACAATCAGCTCGTGACTCAATTCATCGAGGCT
TCCAAGGATGCCAAGATTGTCCCGGGCTCTCCGAGGATAAAATCTGGGC
TTTCTCTGAAGGCCAGCAAGCAAGCTGGGTGTATCCGACGGGATGTA
AAACCTCGTTCTGATCCAGAAGAAGGAAGTCGATCCGACTTCGGGCGTC
GAGCATTTCGCCCTGCAGCAATATGTGAATGGCATCCCGGTATATGGCGG
TGACCAAACCATTCACATCGACAAGGCCGGCCAGGTTACGTCGTTCTGTAG
GAGCTGTTCTGCCGGCTCAAAATCAAATCACGGCAAAATCCAGCGTACCA
GCCATAAGCGCATCCGACGCTCTGGCTATCGCGGCAAGGAAGCCAGTTC
CCGCATCGCGGAGCTGGGAGCACAGGAGAAGACTCCGTCGGCTCAGCTGT
ACGTATATCCGGAAGGCAACGGGTCCGCTCGTCTACCAGACGGAAGTG
AATGTGCTTGAGCCGAGCCTCTGCGCACCCGCTATCTTATCGATGCGGC
CGACGGCCATATCGTCAGCAGTACGATCTGATCGAGACGGCCAGCCGTT
CGGGCACGGGCGTCTGGGCGACAATAAGACGTTCCAGACGACTCTTTCC
GGCAGCACGTACCAGCTGAAAGACACCACTCGCGGCAACGGCATCTACAC
CTACACAGCCAGCAATCGGACCAGGATCCGGGCACGCTGCTGACGGACG
CCGACAACGTATGGACGGATGGAGCCCGCTCGATGCCATACTTATGCC
GGAAAAGTATATGATTTCTACAAAACGAAGTTCGGACGCAACAGCCTCGA
CGGCAACGGCCTGCTGATCCGTTCTCGTCCACTACAGCAGCAGGTACA
ACAATGCCTTCTGGAACGGCACCCAGATTGTATTGCGCGACGGCGACGGC
TCGACGTTTCATTCGCTGTGGGCGATCTCGACGTGGTCCGGCCATGAGCT
GTCCCACGGAGTCATCGAGTACAGTCCAACTTCAATACCTCAATGAAT
CCGGGCGCTGAACGAGTCTATGCCGACGCTCTCGGCAACTCGATCCAG

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- continued

GCGAAAACTGGCTTATCGGCGACGATGCTATACGCTGGCATCTCCGG  
 AGATGCTCTCCGTTCCATGTCCAACCCGACGCTTTACGGGCGAGCCGACA  
 ACTATGCCAACCCGTATACGGGATCTTCCGACAACGGCGGCTTCATACG  
 AACAGCGGCATCACGAACAAAGCGTTCTACTGCTCGCCCAAGGCGGCAC  
 CCAGAACGGCGTTACCGTCGCGGCATCGGGCGGACGACGCGGTGAACA  
 TTTTCTACAACACAGTGGCCTATTACCTTACTTCCACTTCCAACCTCGCC  
 GCGGCGAAGAAGCCTCGATCCAGGCAGCCAAAGACCTGTACGGAACGGG  
 CTCCTCTTATGTACCTCGGTGACCAATGCATTAGAGCCGTAGGCCTG

**[0362]** The amino acid sequence of the PhuPro2 precursor protein is set forth as SEQ ID NO: 12. The predicted signal sequence is shown in italics, and the predicted propeptide is shown in underlined text:

MKKMIPTLLGTVLLSSASAVAESPSLGAAGTPGVSVVNNQLVTQFIEA  
SKDAKIVPGSSSEDKIWAFLGQQAKLGVSAADVKTSLFIQKKEVDPTSGV  
EHFRLQQYVNGIPVYGGDQTIHIDKAGQVTSFVGAFLPAQNQITAKSSVP  
AISASDALAIAAKEASSRIGELGAQKTPSAQLYVYPEGNSRLVYQTEV  
NVLEPQPLRTRYLIDAADGHIVQQYDLIETATGSGTGVLGDNKTFQTTLS  
 GSTYQLKDTTRNGIYTYTASNRTTIPGTLTDADNVWTDGAAVDAHTYA  
 GKVYDFYKTKFGRNSLDGNLLIRSSVHYSSRYNNAFWNGTQIVFGDGDG  
 STFIPLSGDLVDVVGHELHSHGVIEYTSNLQYLNESGALNESYADVLGNSIQ  
 AKNWLIGDDVYTPGISGDALRSMSNPTLYGQPDNYANRYTGSSDNGGVHT  
 NSGITNKAFYLLAQGGTQNGVTVAGIGRDAAVNI FYNTVAYYLTSTSNFA  
 AAKNASIQAAKDLYGTGSSYVTSVTNAPRAVGL

**[0363]** The amino acid sequence of the predicted mature form of PhuPro2 is set forth as SEQ ID NO: 13:

ATGSGTGVLGDNKTFQTTLSGSTYQLKDTTRNGIYTYTASNRTTIPGTL  
 LTDADNVWTDGAAVDAHTYAGKVYDFYKTKFGRNSLDGNLLIRSSVHYSS  
 RYNNAFWNGTQIVFGDGDGSTFIPLSGDLVDVVGHELHSHGVIEYTSNLQY  
 LNESGALNESYADVLGNSIQAKNWLIGDDVYTPGISGDALRSMSNPTLYG  
 QPDNYANRYTGSSDNGGVHTNSGITNKAFYLLAQGGTQNGVTVAGIGRDA  
 AVNI FYNTVAYYLTSTSNFAAKNASIQAAKDLYGTGSSYVTSVTNAPRA  
 VGL

### Example 3.2

#### Expression of *Paenibacillus humicus* s Metalloprotease PhuPro2

**[0364]** The DNA sequence of the propeptide-mature form of PhuPro2 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX150(AprE-PhuPro2) (FIG. 1). Ligation of this gene encoding the PhuPro2 protein into the digested vector resulted in the addition of three codons (Ala-

Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PhuPro2 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 1 was labeled pGX150 (AprE-PhuPro2). As shown in FIG. 3.1, pGX150(AprE-PhuPro2) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PhuPro2 (SEQ ID NO: 14). The translation product of the synthetic AprE-PhuPro2 gene is shown in SEQ ID NO: 15.

**[0365]** The pGX150 (AprE-PhuPro2) plasmid was then transformed into *B. subtilis* cells (degU<sup>H3</sup>32, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

**[0366]** The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to an 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above; and the active flow-through fractions were collected and concentrated. The sample was loaded onto a 320 ml Superdex 75 gel filtration column pre-equilibrated with the loading buffer described above containing 0.15 M NaCl. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

**[0367]** The nucleotide sequence of the synthesized PhuPro2 gene in plasmid pGX150(AprE-PhuPro2) is depicted in SEQ ID NO: 14. The sequence encoding the three residue addition (AGK) is shown in bold:

GTGAGAAGCAAAAAATTGTGGATCAGCTTGTGTTTTCGTTAACGTTAAT  
 CTTTACGATGGCGTTGAGCAACATGAGCGCGCAGGCT**GCTGGA**AAAGAAAT  
 CACCGAGCCTTGGCGTGCAGGAACACCGGGCGTTAGCGTTGTGAATAAC  
 CAACTGGTCACGCAGTTTATCGAAGCATCAAAGACGCGAAAAATTGTCCC  
 TGGATCAAGCGAAGATAAGATTTGGCATTCTCGAAGGCCAGCAAGCAA  
 AGCTTGGCGTCTCAGCTGCCGACGTGAAGACGAGCTTCTGTATCCAGAAG  
 AAGGAGGTTGACCCGACATCAGGCGTTGAGCACTTTAGACTGCAACAGTA  
 CGTCAACGGCATCCCGTTTATGGAGGCGATCAAACAATCCATATTGATA  
 AGGCAGGCCAGGTCAATCATTCTCGGAGCTGTCTGCGGCTCAGAAC  
 CAAATTACAGCAAAATCATCAGTTCCGGCAATTTAGCCTCAGACGCTCT  
 GGCAATCGCTGCCAAGGAGGCAAGCTCAAGAAATGGCGAACTGGGCGCAC  
 AAGAAAAGACACCGAGCGCCCAACTTTATGTCTATCCGGAGGCAACGGGA  
 AGCAGACTGGTGTACCAGACAGAGGTCAATGTTCTGGAGCCGCAACCGCT  
 GAGAACGAGATACCTTATCGATGCTGCGGATGGCCACATGTTTCAGCAAT  
 ACGACCTGATTGAGACAGCAACAGGAAGCGGAACGGGCGTGTGGGCGAC

- continued

AACAAGACGTTTTAGCGGCAGCACGTACCAACTTAAGGA  
 CACGACGAGAGGCAATGGCATTACACGTACACGGCCTCAAACAGAACGA  
 CAATCCAGGCACACTGCTGACGGATGACAGACAATGTTGGACGGACGGC  
 GCAGCAGTTGACGCACACAGTACGCCGGCAAGGTGTACGACTTTTACAA  
 GACGAGTTCCGCGAGAACAGCCTTGATGGAATGGACTGCTGATCAGAA  
 GCAGCGTCCACTACAGCAGAGATACAATAACGCCTTCTGGAACGGCACA  
 CAAATCGTCTTTGGCGATGGAGACGGATCAACATTCATCCCGCTGTCAGG  
 CGACCTGGACGTTGTGGGCCACGAGCTGAGCCACGGCGTCAATCGAGTACA  
 CGAGCAACCTGCAGTACTGAATGAAAGCGCGCACTGAACGAGTCATAT  
 GCTGATGTGCTTGGCAATAGCATCCAGGCCAAGAAGTGGCTTATCGGAGA  
 CGACGCTACACACCTGGCATCAGCGCGATGCTCTGAGAAGCATGAGCA  
 ATCTACACTTTACGGCCAACCGGACAACCTACGCAATAGATATACGGGC  
 AGCAGCGACAATGGCGCGTTATACAACTCAGGCATCACGAACAAGGC  
 GTTCTACCTGCTGGCACAGGAGGCACGCAAAACGGCGTTACAGTTGCGG  
 GCATGGCAGAGATGGCGCGTCAACATCTTCTACAACACAGTCGCCTAC  
 TACCTGACGAGCACGTCAAACCTCGCAGCGCAAAGAACGCATCAATTCA  
 AGCAGCAAAGGATCTGTACGGAACAGGAGCTCATATGTCACGTGAGTTA  
 CGAATGCGTTTAGAGCCGTCGGCCTTTAA

**[0368]** The amino acid sequence of the PhuPro2 precursor protein expressed from plasmid pGX150(AprE-PhuPro2) is depicted in SEQ ID NO: 15. The predicted signal sequence is shown in italics, the three residue addition (AGK) is shown in bold, and the predicted propeptide is shown in underlined text.

(SEQ ID NO: 15)

*MRSKKLWISLLFALTLIFTMAFSNMSAQ***AGK**ESPSLGAAGTPGVSVVNN  
QLVTPQIEASKDAKIVPGSSEDKIWFLEGGQAKLGVSAADVKTSFLIQK  
KEVDPTSGVEHFRLLQQYVNGIPVYGGDQTIHIDKAGQVTSFVGAVALPAQN  
QITAKSVPAISASDALAIAAKEASSRIGELGAQEKTPSAQLVYVPEGN  
SLRLVYQTEVNVLEPQLRTRYLIDAADGHIVQYDLIETATGSGTGLGD  
 NKTFFQTTLSGSTYQLKDTTRNGIYTYTASNRTTIPGTLTLDADNVWTDG  
 AAVDAHTYAGKVYDFYKTKFGRNSLDGNLLIRSVHYSSRYNNAFWNGT  
 QIVFGDGDGTFIPLSGDLVDVGHLSHGVI EYTSNLQYLNESGALNESY  
 ADVLGNISIQAKNWLIGDDVYTPGISGDALRSMSNPTLYGQPDNYANRYTG  
 SSDNGVHTNSGITNKAFYLLAQGGTQNGVTVAGIGRDAAVNI FYNTVAY  
 YLTSTSNFAAAKNASIQAAKLDLYGTGSSYVTSVTNAPRAVGL.

### Example 3.3

#### Proteolytic Activity of Metalloprotease PhuPro2

**[0369]** The proteolytic activity of purified metalloprotease PhuPro2 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Milli-

pore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ l of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ l of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ l of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ l supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a SpectraMax 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of triplicate assays.

**[0370]** The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (shown in FIG. 3.2) indicates that PhuPro2 is an active protease.

### Example 3.4

#### pH Profile of Metalloprotease PhuPro2

**[0371]** With azo-casein as the substrate, the pH profile of metalloprotease PhuPro2 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2 ml Milli-Q H<sub>2</sub>O diluted enzyme (125 ppm) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 3.3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 3.3, the optimal pH of PhuPro2 is 6, with greater than 70% of maximal activity retained between 5.5 and 8.5.

### Example 3.5

#### Temperature Profile of Metalloprotease PhuPro2

**[0372]** The temperature profile of metalloprotease PhuPro2 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assays. The enzyme sample and azo-casein substrate were prepared as in Example 3.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted enzyme (50 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 3.3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data

in FIG. 3.4 suggests that PhuPro2 showed an optimal temperature at 50° C., and retained greater than 70% of its maximum activity between 45 and 65° C.

#### Example 3.6

##### Cleaning Performance of Metalloprotease PhuPro2

**[0373]** The cleaning performance of PhuPro2 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified protease samples were diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1) (detergent composition shown in Table 3.1). To initiate the reaction, 180 µl of the AT detergent buffered at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20 µl of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100 µl of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200 µl water. Following the addition of 180 µl of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as the “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net A<sub>405</sub> was subsequently calculated by subtracting the A<sub>405</sub> of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 in AT dish detergent for PhuPro2 is shown in FIGS. 3.5A and 3.5B.

TABLE 3.1

| Composition of AT dish detergent                            |                       |
|---|-----------------------|
| Ingredient  | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                           | 0.143                 |
| Sodium citrate  | 1.86                  |
| Citric acid*  | varies                |
| Plurafac ® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate  | 0.006                 |
| Bayhibit ® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)             | 0.029                 |
| PEG 6000  | 0.043                 |
| PEG 1500  | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

#### Example 3.7

##### Comparison of PhuPro2 to Other Proteases

###### A. Identification of Homologous Proteases

**[0374]** Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res.*, 25:3389-402, 1997)

against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The predicted mature protein amino acid sequence for PhuPro2 (SEQ ID NO: 13) is used as the query sequences. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 3.2A and 3.2B provide a list of sequences with the percent identity to PhuPro2. The length in Table 3.2 refers to the entire sequence length of the homologous proteases.

TABLE 3.2A

| List of sequences with percent identity to PhuPro2 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PhuPro2 | Organism  | Length |
| P00800   | 59             | <i>Bacillus thermoproteolyticus</i>                         | 548    |
| YP_003872180.1   | 59             | <i>Paenibacillus polymyxa</i> E681                          | 587    |
| ZP_10575942.1  | 59             | <i>Brevibacillus</i> sp. BC25                               | 528    |
| ZP_02326602.1  | 60             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> BRL-230010 | 520    |
| ADM87306.1   | 61             | <i>Bacillus megaterium</i>                                  | 562    |
| ZP_09069025.1  | 61             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 520    |
| ZP_09069194.1  | 62             | <i>Paenibacillus larvae</i> subsp. <i>Larvae</i> B-3650     | 502    |
| ZP_10738945.1  | 63             | <i>Brevibacillus</i> sp. CF112                              | 528    |
| ZP_08511445.1  | 64             | <i>Paenibacillus</i> sp. HGF7                               | 525    |
| ZP_09077634.1  | 65             | <i>Paenibacillus elgii</i> B69                              | 524    |
| ZP_09775365.1  | 65             | <i>Paenibacillus</i> sp. <i>Aloe</i> -11                    | 580    |
| ZP_09775364.1  | 70             | <i>Paenibacillus</i> sp. <i>Aloe</i> -11                    | 593    |
| P29148   | 71             | <i>Paenibacillus polymyxa</i>                               | 590    |
| ZP_10241030.1  | 71             | <i>Paenibacillus peoriae</i> KCTC 3763                      | 593    |
| ZP_09071078.1  | 71             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 529    |
| YP_003872179.1   | 72             | <i>Paenibacillus polymyxa</i> E681                          | 592    |
| YP_005073223.1   | 72             | <i>Paenibacillus terrae</i> HPL-003                         | 591    |

TABLE 3.2B

| List of sequences with percent identity to PhuPro2 protein identified from the Genome Quest Patent database |                |                                       |        |
|---|----------------|---------------------------------------|--------|
| Patent ID #   | PID to PhuPro2 | Organism                              | Length |
| US20090208474-0030  | 59.22          | <i>Bacillus thermoproteolyticus</i>   | 316    |
| JP2002272453-0002   | 59.42          | <i>Bacillus megaterium</i>            | 562    |
| JP2006124323-0003   | 59.55          | <i>Bacillus thermoproteolyticus</i>   | 316    |
| US8114656-0183  | 59.87          | <i>Bacillus stearothermophilus</i>    | 316    |
| JP1989027475-0001   | 59.87          | <i>Bacillus subtilis</i>              | 316    |
| US20120009651-0002  | 59.87          | <i>Geobacillus caldoproteolyticus</i> | 548    |
| JP2002272453-0003   | 60.45          | empty                                 | 562    |
| WO2012110563-0004   | 60.77          | <i>Bacillus megaterium</i>            | 320    |
| EP2390321-0186  | 62.25          | <i>Bacillus brevis</i>                | 304    |
| JP2005229807-0018   | 71.85          | <i>Paenibacillus polymyxa</i>         | 566    |
| US8114656-0187  | 72.09          | <i>Bacillus polymyxa</i>              | 302    |

###### B. Alignment of Homologous Protease Sequences

**[0375]** The amino acid sequence of predicted mature PhuPro2 (SEQ ID NO: 13) protein was aligned with thermolysin (P00800, *Bacillus thermoproteolyticus*), and protease from *Paenibacillus terrae* HPL-003 (YP\_005073223.1) sequences using CLUSTALW software (Thompson et al., *Nucleic Acids*

Research, 22:4673-4680, 1994) with the default parameters. FIG. 3.6 shows the alignment of PhuPro2 with these protease sequences.

### C. Phylogenetic Tree

[0376] A phylogenetic tree for full length sequence of PhuPro2 (SEQ ID NO: 12) was built using sequences of representative homologs from Table 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. *Mol Biol. Evol.* 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylogenetic tree printer software (<http://iubio.bio.indiana.edu/tree-app/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 3.7.

#### Example 4.1

##### Cloning of *Paenibacillus ehimensis* Metalloprotease PehPro1

[0377] A strain (DSM11029) of *Paenibacillus ehimensis* was selected as a potential source of enzymes which may be useful in various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus ehimensis* strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus ehimensis* encodes a metalloprotease and the sequence of this gene, called PehPro1, is provided in SEQ ID NO: 16. The corresponding protein encoded by the PehPro1 gene is shown in SEQ ID NO: 17. At the N-terminus, the protein has a signal peptide with a length of 23 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) *Nature Methods*, 8:785-786). The presence of a signal sequence suggests that PehPro1 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) *Journal of Bacteriology*, 173 (21): 6820-6825). The predicted mature region of PehPro1 protein is shown in SEQ ID NO: 18. [0378] The nucleotide sequence of the PehPro1 gene isolated from *Paenibacillus ehimensis* is set forth as SEQ ID NO: 16. The sequence encoding the predicted native signal peptide is shown in *italics*:

*ATGTTAAAGATATGGGCATCGATTATTACAGGAGCATTTTGGCTCGGGAG  
CGTGCAAGGGGTGCAAGCTGCTCCACAAGATCAAGCTGCTCCCTTCGGAG  
GATTACCCCTCAATTGATTACCGGGGAAAGCTGGAGTGCGCCGCAAGGA  
GTATCGGGAGAGGAAAAATCTGGAAGTATCTCGAATCCAAGCAGGAAAG  
CTTCCAAATCGGCCAAACCGTTGATCTGAAAAAGCAATGAAAATTATCG  
GCCAAACGACCGACGAGAAAACGGGAACCAACGCATTACCGTCTACAGCAG*

- continued

TATGTGGGAGGCGTCCCGTATACGGCGGCGTACAAAACGATCCATGTCAA  
CAAAGAAGGACAAGTTACCTCGCTGATCGGCAGCCTGCTTCCCAGCCAGC  
AGCAGCAAGTTTCGAAAAGCTTGAATTCGCAAAATCAGCGAAGCGCAAGCC  
ATCGCCGTGGCCAGAAAGATACCGAGGCGCGCTCGGCAAGCTGGGTGA  
ACCGCAAAGACACCGGAAGCGGATCTGTACGTTTATTACACAACGGAC  
AACCGGTCTCGCTTATGTGACCGAGGTTAACGTTCTCGAACCGGAGGCA  
ATCCGGACCGCTACTTCATCAGCGCGAAGACGGCAGCATTTTATTCAA  
GTACGACATCCTCGCTCAGCTACAGGTACCGGAAAAGGCGTCTCGGAG  
ATACGAAATCGTTTACGACACGCAATCCGGCTCCACTTATCAATTGAG  
GATACGACCGCGGGCAAGGTATCGTCACTTACAGCGCTGGCAACCGGTC  
CTCTCTGCCGGGAACGCTGCTCACCAGCTCCAGCAATATTTGGAACGACG  
GCGCGGCGGTGATGCGCATGCCTATACCGCCAAAGTGTACGATTACTAT  
AAAAACAATTTGGCCGCAACAGCATTGACGGCAACGGCTTCCAGCTTAA  
ATCGACCGTGCCTATTCCTCCAGATACAAACAGCCTTCTGGAACGGTG  
TGCAATGGTGTACGGCGACGGCGACGGCGTAACTTCATTCCGTCTTCC  
GCCGATCCGGACGTCATCGGCCACGAATTGACCCACGGCGTTACGGAACA  
TACGGCCGGCCTGGAATACTACGGCGAATCCGGAGCGCTGAACGAATCGA  
TCTCCGATATTATCGGCAACCGGATCGACGGCAAAAACCTGGCTGATCGGC  
GACTTGATTTATACGCCGAATACTCCCGGGGACGCCCTCCGCTCTATGGA  
GAACCCCAAGCTGTATAACCAACCCGACCGCTATCAAGACCGCTATACGG  
GACCTTCGGATAACGGCGCGTGCATATTAACAGCGGTATCAACAACAAA  
GCCTTCTACCTGATCGCCCAAGGCGGCACGCACTATGGCGTCAACCGTGAA  
CGGGATCGGACCGGATCGCGCTGTGCAATTTTCTATGACGCCCTCATCA  
ATTACCTGACTCCAACCTCGAACTTCTCGCGATGCGCGCAGCAGCCATT  
CAAGCGGCAACCGACCTGTACGGAGCGAATCTTCTCAAGTAAACGCTGT  
CAAAAAGCGTATACTGCCGTCCGCGTGAAC

[0379] The amino acid sequence of the PehPro1 precursor protein is set forth as SEQ ID NO: 17. The predicted signal sequence is shown in *italics*, and the predicted propeptide is shown in underlined text:

*MLKRWASIIITGAFLLGSVQGVQAAPQDQAAPPFGGFTPQLITGESWSAPQG  
SGEEKIWKYLESKQESFQIGQTVDLKKQLKIIIGQTTDEKGTTHYRLQQ  
YVGGVPVYGGVQTIHVNKEGQVTSLIGSLLPDQQQVSKSLNSQISEAQA  
IAVAQKDTAAVGLGEPQKTPADLYVYLHNGQPVLAYVTEVNVLEPEA  
IRTRYFISAEDGSLFKYDILAHATGTGKGLVLDKTSFTTQSGSTYQLK  
DTRRQGIIVTYSAGNRRSLPGTLLTSSSNIWNDDGAAVDAHAYTAKVYDYD  
KNKFRNSIDGNGFQLKSTVHYSSRYNNAFWNQVMVYGDGDVTFIPIFS  
ADPDVIGHELTHGVTEHTAGLEYYGESGALNESISDIIGNAIDGKNWLI  
DLIYTPNTPGDALRSMENPKLYNQPDYQDRYTPGSDNGGVHINSGINNK*

-continued

AFYLIAQGGTHYGVTVNGIGRDAVQIFYDALINYLTPTSNFSAMRAAAI  
QAATDLYGANSSQVNAVKKAYTAVGVN

**[0380]** The amino acid sequence of the predicted mature form of PehPro1 is set forth as SEQ ID NO: 18:

ATGTGKGVLDGDKSFTTTSQSGSTYQLKDTTRGQIVTYSAGNRSSLPGTL  
LTSSNIWNDGAAVDAHAYTAKVYDYYKNKFGFRNSIDGNFQPKSTVHYS  
SRYNNAFWNGVQVMYGDGDGVTFIPFSADPDVIGHELTHGVTEHTAGLEY  
YGESGALNESISDIIGNAIDGKNWLDGLIYTPNTPGDALRSMENPKLYN  
QPDRYQDRYTGPSDNGGVHINSGINNKAFYLIAQGGTHYGVTVNGIGRDA  
AVQIFYDALINYLTPTSNFSAMRAAAIQAATDLYGANSSQVNAVKKAYTA  
VGVN

#### Example 4.2

##### Expression of *Paenibacillus ehimensis* Metalloprotease PehPro1

**[0381]** The DNA sequence of the propeptide-mature form of PehPro1 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, *Protein Expr Purif*, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX148(AprE-PehPro1) (FIG. 4.1). Ligation of this gene encoding the PehPro1 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PehPro1 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 1 was labeled pGX148 (AprE-PehPro1). As shown in FIG. 1, pGX148(AprE-PehPro1) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PehPro1 (SEQ ID NO: 19). The translation product of the synthetic AprE-PehPro1 gene is shown in SEQ ID NO: 20.

**[0382]** The pGX148(AprE-PehPro1) plasmid was then transformed into *B. subtilis* cells (degU<sup>ts</sup>32, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

**[0383]** The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to an 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above, PehPro1 was eluted from the column with a linear salt gradient from 0 to 0.3 M NaCl in the loading buffer. The corresponding active fractions were collected, concentrated and buffer-exchanged again into the loading buffer described above. The sample was loaded onto a 40 ml DEAE Fast Flow column pre-equilibrated with the

same loading buffer. PehPro1 was eluted from the column with a linear salt gradient from 0 to 0.15 M NaCl in the loading buffer. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

**[0384]** The nucleotide sequence of the synthesized PehPro1 gene in plasmid pGX148(AprE-PehPro1) is depicted in SEQ ID NO: 19. The sequence encoding the three residue addition (AGK) is shown in bold:

GTGAGAAGCAAAAATTGTGGATCAGCTTGTGTTTGCCTTAACGTTAAT  
CTTTACGATGGCGTTTACGCAACATGAGCGCGCAGGCT**GTGAAAAG**CAC  
CTCAAGATCAGGCAGCACCTTTTGGAGGCTTTACACCGCAACTTATCACA  
GGCGAATCATGGTCAGCACCGCAGGGCGTTTACGGCGAGAAAAGATCTG  
GAAGTACCTTGAGAGCAAGCAGGAGTCAATTTCAAATCGGCCAGACAGTCC  
ACCTGAAAAGCAACTGAAGATCATCGGCCAAACACCGACGAAAAGACG  
GGCAGCAGCATTATAGACTGCAACAATATGTTGGCGCGTGCCTGTTA  
TGGAGCGGTGCAACAATCCAGTGAACAAGGAAGGACAGGTCACGTCAC  
TGATCGGCAGCCTGCTGCCGATCAGCAGCAACAAGTCTCAAAGAGCCTG  
AACTCACAATAATAGCGAGGCACAAGCGATTGCAGTTGCACAAAAGGACAC  
GGAAGCAGCTGTCGGCAAGCTGGCGCAACCGCAAAAACACCTGAGGCTG  
ACCTTTACGCTCTACCTGCATAACCGCCAGCCGGTCTTTCGCTACGTTACG  
GAAGTTAACGTCCTGGAGCCGGAGGCATCAGAACGAGATACTTCATTAG  
CGCGGAGGATGGAAGCATTCTGTTAAGTACGATATCTTGCTCACGCGA  
CAGGCACAGGCAGGGCGTCTTGGCGACAAAAAGCTTACGACAACG  
CAGAGCGGATCAACGTACAGCTGAAAGATACAACAAGAGGACAAGGCAT  
CGTTACGTATTCAGCGGGCAATAGATCAAGCCTGCCGGGCACACTGCTGA  
CATCAAGCTCAAAACATTTGGAATGACGGCGCAGCAGTTGATGCCATGCG  
TACACAGCCAAGGTGTACGACTACTATAAGAACAAGTTTGGCAGAAATAG  
CATCAGCGAAATGGATTTCAACTTAAATCAACGGTGCCTACTCATCAA  
GATATAACAATGCGTTTTTGGAAACGGAGTGCAGATGGTCTACGGAGACGGC  
GACGGCGTGACATTTATTCCGTTTAGCGCCGACCCGGACGTGATTGGACA  
TGAAGTACACATGGAGTGACAGAGCATACGGCGGACTGGAATATTACG  
GCGAAAGCGGCGCACTGAACGAAAGCATCTCAGACATTATTGAAACGCA  
ATCGATGGCAAAAACCTGGCTGATTGGCGATCTGATTATACGCCGAATAC  
ACCGGGCGATGCACTGAGATCAATGGAGAATCCGAAGCTGTACAACCAAC  
CGGACAGATACCAAGATAGATACACAGGACCGTCAGACAACGGCGGAGTC  
CATATCAACAGCGGAATCAATAACAAGCCTTTTACCTGATCGCCCAAGG  
CGGAACGCCTATGGCGTTACAGTCAATGGCATCGGAAGAGATGCCCGAG  
TTCAGATTTTCTATGACGCGCTGATCAACTATCTGACGCTTACAAGCAAT  
TTCTCAGCAATGAGAGCCGAGCAATCCAAGCAGCCAGGATCTGTATGG  
AGCCAATTCATCAAGTTAATGCTGTTAAGAAGGCTTATACGGCAGTGG  
GAGTTAACTAA

**[0385]** The amino acid sequence of the PehPro1 precursor protein expressed from plasmid pGX148(AprE-PehPro1) is depicted in SEQ ID NO: 20. The predicted signal sequence is shown in italics, the three residue addition (AGK) is shown in bold, and the predicted pro-peptide is shown in underlined text.

*MRSKKLWISLLFALTLIFTMAFSNMSAQAAGKAPQDQAAAPFGGFTPOLIT*  
GESWSAPQGVSGEEKIKWYLESKQESFPQIGQTVLKKQLKIGQTTDEKT  
GTTHRYLQQYVGGVPVYGVQTIHVNKEGQVTSLIGSLLPDQQQVSKSL  
NSQISEAQAIAVAQKDTAAVKGKLGEPQKTPCADLYVYLHNGQPVLAYVT  
EVNVLPEAIRTRYFISAEEDSGSILFKYDILAHATGTGKGVLDGTSFTTT  
 QSGSTYQLKDTTRGQGIIVTYSAGNRSSLPGLTLLTSSSNIWNDGAAVDAHA  
 YTAKVYDYKKNKFRNSIDGNGFQLKSTVHYSSRYNNAFWNGVMVYGDG  
 DGVTFIPFSADPDVIGHELTHGVTEHTAGLEYYGESGALNESISDIIIGNA  
 IDGKNWLI GDLIYTPNTPGDALRSMENPKLYNQPDQRYDQRYTGPSDNGGV  
 HINSGINNKAFYLIAQGGTHYGVTVNIGRDAAVQIFYDALINYLTPTSN  
 FSAMRAAAIQAAATDLYGANSSQVNAVKKAYTAVGVN.

#### Example 4.3

##### Proteolytic Activity of Metalloprotease PehPro1

**[0386]** The proteolytic activity of purified metalloprotease PehPro1 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ l of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ l of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ l of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ l supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a SpectraMax 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of triplicate assays. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (shown in FIG. 4.2) indicates that PehPro1 is an active protease.

#### Example 4.4

##### pH Profile of Metalloprotease PehPro1

**[0387]** With azo-casein as the substrate, the pH profile of metalloprotease PehPro1 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH

was first mixed with 2  $\mu$ l Milli-Q H<sub>2</sub>O diluted enzyme (250 ppm) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 4.3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 4.3, the optimal pH of PehPro1 is 7, with greater than 70% of maximal activity retained between 5.5 and 9.5.

#### Example 4.5

##### Temperature Profile of Metalloprotease PehPro1

**[0388]** The temperature profile of metalloprotease PehPro1 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assays. The enzyme sample and azo-casein substrate were prepared as in Example 4.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted enzyme (100 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 4.3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data in FIG. 4.4 suggest that PehPro1 showed an optimal temperature at 70° C., and retained greater than 70% of its maximum activity between 60 and 75° C.

#### Example 4.6

##### Cleaning Performance of Metalloprotease PehPro1

**[0389]** The cleaning performance of PehPro1 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified protease samples were diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1) (detergent composition shown in Table 4.1). To initiate the reaction, 180  $\mu$ l of the AT detergent buffered at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ l of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ l water. Following the addition of 180  $\mu$ l of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the



resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as the “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net  $A_{405}$  was subsequently calculated by subtracting the  $A_{405}$  of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 for PehPro1 in AT detergent is shown in FIGS. 4.5A and 4.5B.

TABLE 4.1

| Composition of AT dish detergent                           |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

## Example 4.7

## Comparison of PehPro1 to Other Proteases

## A. Identification of Homologous Proteases

[0390] Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res.*, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The mature protein amino acid sequence for PehPro1 (SEQ ID NO: 18) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 4.2A and 4.2B provide a list of sequences with the percent identity to PehPro1. The length in Table 4.2 refers to the entire sequence length of the homologous proteases.

TABLE 4.2A

| List of sequences with percent identity to PehPro1 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PehPro1 | Organism  | Length |
| ZP_09077634.1  | 88             | <i>Paenibacillus elgii</i> B69                          | 524    |
| ZP_09071078.1  | 74             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650 | 529    |
| YP_003872179.1   | 74             | <i>Paenibacillus polymyxa</i> E681                      | 592    |
| P29148   | 73             | <i>Paenibacillus polymyxa</i>                           | 590    |
| P43263   | 68             | <i>Brevibacillus brevis</i>                             | 527    |
| ZP_09775365.1  | 68             | <i>Paenibacillus</i> sp. Aloe-11                        | 580    |
| ZP_10241029.1  | 67             | <i>Paenibacillus peoriae</i> KCTC 3763                  | 599    |
| ZP_10575942.1  | 66             | <i>Brevibacillus</i> sp. BC25                           | 528    |
| YP_002770810.1   | 67             | <i>Brevibacillus brevis</i> NBRC 100599                 | 528    |

TABLE 4.2A-continued

| List of sequences with percent identity to PehPro1 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PehPro1 | Organism                                    | Length |
| ZP_08640523.1  | 64             | <i>Brevibacillus laterosporus</i> LMG 15441 | 564    |
| YP_004646155.1   | 63             | <i>Paenibacillus mucilaginosus</i> KNP414   | 525    |
| ZP_08093424.1  | 60             | <i>Planococcus donghaensis</i> MPA1U2       | 553    |
| YP_003670279.1   | 59             | <i>Geobacillus</i> sp. C56-T3               | 546    |
| P00800   | 59             | <i>Bacillus thermoproteolyticus</i>         | 548    |

TABLE 4.2B

| List of sequences with percent identity to PehPro1 protein identified from the Genome Quest Patent database |                |                                    |        |
|---|----------------|------------------------------------|--------|
| Patent ID #   | PID to PehPro1 | Organism                           | Length |
| JP2005229807-0019   | 74.5           | <i>Paenibacillus polymyxa</i>      | 566    |
| US20120107907-0187  | 74.09          | <i>Bacillus polymyxa</i>           | 302    |
| US8114656-0186  | 68.21          | <i>Bacillus brevis</i>             | 304    |
| WO2004011619-0044   | 63.25          | empty                              | 507    |
| EP2390321-0185  | 62.9           | <i>Bacillus cereus</i>             | 317    |
| WO2012110563-0004   | 62.7           | <i>Bacillus megaterium</i>         | 320    |
| WO2012110563-0005   | 62.58          | <i>Bacillus cereus</i>             | 320    |
| JP1995184649-0001   | 62.5           | <i>Lactobacillus</i> sp.           | 566    |
| JP2005333991-0002   | 62.38          | empty                              | 562    |
| EP2178896-0184  | 62.18          | <i>Bacillus anthracis</i>          | 566    |
| JP1994014788-0003   | 61.94          | empty                              | 317    |
| EP2390321-0178  | 61.86          | <i>Bacillus thuringiensis</i>      | 566    |
| US6518054-0002  | 60.84          | <i>Bacillus</i> sp.                | 316    |
| US8114656-0176  | 60.13          | <i>Bacillus stearothermophilus</i> | 548    |
| US6103512-0003  | 59.81          | empty                              | 319    |
| US20120107907-0184  | 59.49          | <i>Bacillus caldoyticus</i>        | 319    |

## B. Alignment of Homologous Protease Sequences

[0391] The amino acid sequence of predicted mature PehPro1 (SEQ ID NO: 18) was aligned with thermolysin (P00800, *Bacillus thermoproteolyticus*) and protease from *Paenibacillus elgii* B69 (ZP\_09077634.1) using CLUSTALW software (Thompson et al., *Nucleic Acids Research*, 22:4673-4680, 1994) with the default parameters. FIG. 4.6 shows the alignment of PehPro1 with these protease sequences.

## C. Phylogenetic Tree

[0392] A phylogenetic tree for precursor protein PehPro1 (SEQ ID NO: 17) was built using sequences of representative homologs from Table 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. *Mol Biol. Evol.* 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylodendron-phylogenetic tree printer software (<http://iubio.bio.indiana.edu/treeapp/tree-print-form.html>) was used to display the phylogenetic tree shown in FIG. 4.7.

## Example 5.1

Cloning of *Paenibacillus barcinonensis* Metalloprotease PbaPro1

[0393] A strain (DSM15478) of *Paenibacillus barcinonensis* was selected as a potential source of enzymes which may

be useful in various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus barcinonensis* strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus barcinonensis* encodes a metalloprotease and the sequence of this gene, called PbaPro1, is provided in SEQ ID NO: 21. The corresponding protein encoded by the PbaPro1 gene is shown in SEQ ID NO: 22. At the N-terminus, the protein has a signal peptide with a length of 25 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) Nature Methods, 8:785-786). The presence of a signal sequence suggests that PbaPro1 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) Journal of Bacteriology, 173 (21): 6820-6825). The predicted mature region of PbaPro1 protein is shown in SEQ ID NO: 23.

**[0394]** The nucleotide sequence of the PbaPro1 gene isolated from *Paenibacillus barcinonensis* is set forth as SEQ ID NO: 21. The sequence encoding the predicted native signal peptide is shown in italics:

*ATGAAATTGACAAAATTATGCCAACAAATCTTG CAGGAGCTCTTTTGCT  
CACATCCCTGTCTCTGCAGCAGCAATGCCGTTATCTGACTCATCCATT  
CATTTGAGGGCCCTACACCTCCGAGGAGAGTATCTGTTGAACAACAAC  
CCGGACGAAATGATTTATAATTTCTTG CACAACAAGAGCAATTTCTGAA  
TGCCGACGTCAAAGGACAGCTCAAAATCATTAAACGCAACACAGACACT  
CCGGCATCAGACACTTTCGCTGAAGCAATACATCAAAGGTGTTCCGGTT  
TACGGCCGACAACAACAGTCCATCTGGACAAGAACGGAGCTGTAACCTC  
CGCACTCGGCAGTCTTCCGCAATTGAAGAACAGGCTGTTCCGAATGATG  
GCGTTC CCGCAATCAGTGCAGACGATGCCATCCGTGCCGCCGAGAATGAA  
GCCACCTCCCGTCTTGGAGAGCTTGGCGCACAGAGCTTGAGCCAAAGGC  
CGAATTAACATTTATCATCATGAAGATGACGGACAACCTACCTCGTTT  
ACATTACGGAAGTTAACGTGCTTGAGCCTTCCCGCTACGGACCAAATAT  
TTTATTAACGCCCTTGATGGAAGCATCGTATCTCAATACGATATTATCAA  
CTTTGCCACAGGCACCGGTACAGGCGTGATGATGATACAAAACACTGA  
CGACAACCTCAATCCGGCAGCACCATCAGCTGAAAGATACAACCTCGTGA  
AAAGGCATTCAAACCTATACTGCGAACAAATCGCTCCTCGTTC CAGGCAG  
CTTGCTACCAAGTTC AATAACGTATGGACAGACCGTG CAGCTGTAGATG  
CGCACGCCTATGCTGCCGCCACATATGACTTCTACAAAACAAATTC AAT  
CGCAACGGCATTGACGGAACCGGCTGTTGATTCGCTCTACAGTG CATT  
TGGCTCCAAC TATAAAAAACGCTTCTGGAACGGAGCACAGATTGTCTATG*

- continued

GAGATGGCGATGGCATCGAGTTCGGTCCCTCTCCGGTGATCTCGATGTT  
GTCGGACATGAATTGACACACGGGGTGATTGAATATACAGCCAATCTCGA  
ATATCGCAATGAGCCGGGTGCTTTAAACGAAGCTTTTGCCGACATTATGG  
GGAACACCATCGAAAAGCAAAAACCTGGCTGCTTGGCGACGGAATCTATACT  
CCAAACATTC CAGGTGATGCCCTGCGCTCGTTATCCGACCTACGCTGTA  
TAACCAGCCTGACAATAACAGTGATCGCTACACTGGCTCTCAGGATAATG  
GCGGTGTGCATATCAACAGCGGGATCATTAACAAGCATATTATCTTGCA  
GCCCAAGGCGGTACTCATAACGGGGTAACCGTTAGCGGCATCGGCCGGGA  
TAAAGCAGTACGTATTTCTATAGCACGCTGGTGAACCTCTGACGCCAA  
CCTCCAAATTTGCAGCAGCCAAAACAGCGACAATTCAGGCAGCCAAGGAC  
CTGTACGGTGCCAATTCGCTGAAGCTACGGCAATCACCAAAGCTTATCA  
AGCGGTAGGTTTG

**[0395]** The amino acid sequence of the PbaPro1 precursor protein is set forth as SEQ ID NO: 22. The predicted signal sequence is shown in italics, and the predicted pro-peptide is shown in underlined text:

*MKLTKIMPTLLAGALLLSLSSAAAMPLSDSSIPFEGPYTSEESILLNNN  
PDEMIYNFLAQOQOFLNADVKGLKLIKRNRTDTS GIRHFRLLKQYIKGVV  
YGAEQTIHLDKNGAVTSA LGLDLPPIEEQAVPNDGVP AISADDAIRAENE  
ATSR LGELGAPELEPKAELNIYHHEDDGQTYLVYITEVNVLEPSPLRTKY  
FINALDGSIVSQYDIINFATGTGTGVHGDTKLTLTTQSGSTYQLKDTTRG  
KGIQTYTANNRSSLPGSLSTSSNNVWTDRAAVDAHAYAAATYDFYKNKFN  
RNGIDGNLLIRSTVHYGSNYKNAFWNGAQIVYGDGDI EFGPFSGDLDV  
VGHELTHGVI EYTANLEYRNEP GALNEAFADIMGNTIESKNWLLGDGIYT  
PNIPGDALRSLSDPTLYNQPKYSDRYTGSQDNGGVHINSIINKAYYLA  
AQGGTHNGVTVSGIGRDKAVRIFYSTLVNLYLPTSKFAAAKTATIQA AKD  
LYGANSAEATAITKAYQAVGL*

**[0396]** The amino acid sequence of the predicted mature form of PbaPro1 is set forth as SEQ ID NO: 23:

ATGTGTGVHGDTKLTLTTQSGSTYQLKDTTRGKGIQTYTANNRSSLPGSL  
STSSNNVWTDRAAVDAHAYAAATYDFYKNKFN RNGIDGNLLIRSTVHYG  
SNYKNAFWNGAQIVYGDGDI EFGPFSGDLDV VGHELTHGVI EYTANLEY  
RNEP GALNEAFAD IMGNTIESKNWLLGDGI YTPNIPGDALRSLSDPTLYN  
QPKYSDRYTGSQDNGGVHINSIINKAYYLA AQGGTHNGVTVSGIGRDK  
AVRIFYSTLVNLYLPTSKFAAAKTATIQA AKDLYGANSAEATAITKAYQA  
VGL

### Example 5.2

#### Expression of *Paenibacillus barcinonensis* Metalloprotease PbaPro1

**[0397]** The DNA sequence of the propeptide-mature form of PbaPro1 was synthesized and inserted into the *Bacillus*

*subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX147(AprE-PbaPro1) (FIG. 5.1). Ligation of this gene encoding the PbaPro1 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PbaPro1 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 1 was labeled pGX147 (AprE-PbaPro1). As shown in FIG. 5.1, pGX147(AprE-PbaPro1) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PbaPro1 (SEQ ID NO: 24). The translation product of the synthetic AprE-PbaPro1 gene is shown in SEQ ID NO: 25.

[0398] The pGX147(AprE-PbaPro1) plasmid was then transformed into *B. subtilis* cells (degU<sup>Hv32</sup>, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

[0399] The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to an 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above; and the active flow-through fractions were collected and concentrated. The sample was loaded onto a 320 ml Superdex 75 gel filtration column pre-equilibrated with the loading buffer described above containing 0.15 M NaCl. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

[0400] The nucleotide sequence of the synthesized PbaPro1 gene in plasmid pGX147(AprE-PbaPro1) is depicted in SEQ ID NO: 24. The sequence encoding the three residue addition (AGK) is shown in bold:

```
GTGAGAAGCAAAAAATTGTGGATCAGCTTGTGTTGCGTTAACGTTAAT
CTTTACGATGGCGTTTCAGCAACATGAGCGCGCAGGCTGCTGGAAAAATGC
CTCTGTGAGCAGCAGCATTCCGTTTGAGGGCCGTACACATCAGAAGAA
AGCATCCTGCTGAACAACAACCCGGAGCAGATGATCTACAATTTCTCTGGC
ACAGCAGGAGCAGTTCCTGAACGCAGACGTGAAGGGCCAGCTGAAAAATCA
TCAAAGAAACACAGACACGAGCGGCATCAGACACTTCAGACTGAAGCAG
TACATCAAGGGCGTCCCGGTTTACGGCGCTGAGCAGACAATCCACCTGGA
CAAAAATGGCGCAGTGACGAGCGCACTGGAGATCTGCCGCCGATTGAAG
AGCAAGCAGTCCCGAACGATGGCGTTCGGCGATTAGCGCTGATGACGCT
ATCAGAGCCGGGAAAACGAAGCGACGTCAAGACTGGGAGAACTTGGCGC
ACCGGAACTTGAACCGAAGCGGAAGTGAACATCTATCACCACGAAGACG
ATGGACAGACGTACTGGTGTACATCACGGAGGTGAATGTGCTGGAGCCG
```

- continued

```
TCACCGCTGAGAACAAAATACCTTCATCAATGGCGCTGGATGGCAGCATCGT
TAGCCAATACGACATCATTAACTTCGCCACAGGCACGGGCACAGGCGTTC
ATGGCGACACAAAAACGCTTACGACAACACAGTCAGGCTCAACGTACCAG
CTGAAAGACACAAACAGAGGCAAGGGCATCCAGACGTATACAGCCAATAA
CAGAAGCTCACTTCGGGGTCACTGTCAACAGCAGCAATAATGTCTGGA
CGGACAGAGCTGCAGTGGACGCGCACGCGTATGCTCGGCCACGTACGAC
TTCTACAAGAACAAGTTCAACAGAAACGGCATTGATGGCAACGGCCTGCT
TATTAGAAGCAGGTCACACTACGGCTCAAACACTACAAGATGCGTTTTGGA
ACGGCGCCCAAATGTTTTATGGCGATGGAGACGGCATCGAGTTCGGACCT
TTTAGCGGCGACCTGGATGTGGTTCGGACATGAACTGACGCACGGCGTTAT
CGAGTATACGGCGAATCTGGAATACAGAATGAACCGGGCGCTCTGAATG
AGGCCTTCGGGATATCATGGGCAACACAATTGAGAGCAAAAACCTGGCTT
CTGGGCGACGGAATCTACAGCCGAACATTCCGGGAGATGCACTGAGATC
ACTGAGCGACCCCTACGCTGTACAACAGCCGGACAATAACAGCGACAGAT
ACACGGGATCAGAGCAATGGCGGCGTCCATATTAACCTCAGGCATCATC
AACAAAGCGTATTATCTGGCAGCTCAAGGGCGCACGCATAATGGCGTCAC
AGTTAGCGGAATCGGCAGAGACAAGCCGTCAGAATTTTCTACTCAACGC
TGGTGAACACTGACACCAGCAAGCAAGTTTTCAGCCGCCAAAACAGCC
ACGATTCAGGCAGCAAAGGACCTGTACGGAGCGAAGTCAAGCAGAGGCCAC
AGCGATTACGAAGGCTTATCAAGCCGTGGGACTGTAA
```

[0401] The amino acid sequence of the PbaPro1 precursor protein expressed from plasmid pGX147(AprE-PbaPro1) is depicted in SEQ ID NO: 25. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) is shown in **bold**, and the predicted pro-peptide is shown in underlined text.

```
MRSKKLWISLLFALTLIFTMAFSNMSAQAAAGKMPLSDSSIPFEGPYTSEE
SILLNNPNDEMIYNFLAQOQEQFLNADVKGQLKIIKRNTDTSGIRHFRLQ
YIKGVVYVYGAEQTIHLDKNGAVTSALGDLPPIEEEQAVPNDGVP AISADDA
IRAAENEATSRLGELGAPELEPKAELNIYHHEDDGQTYLVYITEVNVLEP
SPLRTKYFINALDGSIVSQYDIINFATGTGTGVHGDTKLTFTTQSGSTYQ
LKDTRGKGIQTYTANNRSLPGSLSTSNNVWTDRAAVDAHAYAAATYD
FYKKNFNRRNGIDGNLLIRSTVHYGSNYKNAFWNGAQIVYDGDGIEFGP
FSGDLVDVGHETHGVIEYTANLEYRNEP GALNEAFADIMGNTIESKNWL
LGDGIYTPNIPGDALRSLSDPTLYNQPKYSDRYTGSQDNGGVHINSGLI
NKAYYLAAQGGTHNGVTVSGIGRDKAVRIFYSTLVNYLTPTSKFAAAKTA
TTQAADLYGANSAEATAIKAYQAVGL
```

### Example 5.3

#### Proteolytic Activity of Metalloprotease PbaPro1

[0402] The proteolytic activity of purified metalloprotease PbaPro1 was measured in 50 mM Tris (pH 7), using azo-

casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ l of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ l of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ l of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ l supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a SpectraMax 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of triplicate assays.

[0403] The proteolytic activities are shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (shown in FIG. 5.2) indicates that PbaPro1 is an active protease.

#### Example 5.4

##### pH Profile of Metalloprotease PbaPro1

[0404] With azo-casein as the substrate, the pH profile of metalloprotease PbaPro1 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 5 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2  $\mu$ l Milli-Q H<sub>2</sub>O diluted enzyme (125 ppm) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 5.3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 5.3, the optimal pH of PbaPro1 is 8, with greater than 70% of maximal activity retained between 7 and 9.

#### Example 5.5

##### Temperature Profile of Metalloprotease PbaPro1

[0405] The temperature profiles of metalloprotease PbaPro1 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assays. The enzyme sample and azo-casein substrate were prepared as in Example 5.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted enzyme (50 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 5.3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40,

50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data in FIG. 5.4 suggest that PbaPro1 showed an optimal temperature at 50° C., and retained greater than 70% of its maximum activity between 45 and 55° C.

#### Example 5.6

##### Cleaning Performance of Metalloprotease PbaPro1

[0406] The cleaning performance of PbaPro1 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified protease samples were diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1) (detergent composition shown in Table 5.1). To initiate the reaction, 180  $\mu$ l of the AT detergent buffered at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ l of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ l water. Following the addition of 180  $\mu$ l of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as the “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net  $A_{405}$  was subsequently calculated by subtracting the  $A_{405}$  of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 in AT detergent for PbaPro1 is shown in FIGS. 5.5A and 5.5B.

TABLE 5.1

| Composition of AT dish detergent formula with bleach       |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| PAP (peracid N,N-phthaloylaminoperoxyacetic acid)          | 0.057                 |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

## Example 5.7

## Comparison of PbaPro1 to Other Proteases

**[0407]** A. Identification of Homologous Proteases

**[0408]** Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res.*, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The predicted mature protein amino acid sequence for PbaPro1 (SEQ ID NO: 23) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 5.2A and 5.2B provide a list of sequences with the percent identity to PbaPro1. The length in Table 5.2 refers to the entire sequence length of the homologous proteases.

TABLE 5.2A

| List of sequences with percent identity to PbaPro1 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PbaPro1 | Organism  | Length |
| AAB02774.1   | 56             | <i>Geobacillus stearothermophilus</i>                       | 552    |
| P00800   | 56             | <i>Bacillus stermoproteolyticus</i>                         | 548    |
| AAA22623.1   | 57             | <i>Bacillus caldolyticus</i>                                | 544    |
| YP_003670279.1   | 57             | <i>Geobacillus</i> sp. C56-T3                               | 546    |
| AAC43402.1   | 57             | <i>Alicyclobacillus acidocaldarius</i>                      | 546    |
| YP_003597483.1   | 57             | <i>Bacillus megaterium</i> DSM 319                          | 562    |
| ZP_08093424.1  | 57             | <i>Planococcus donghaensis</i> MPA1U2                       | 553    |
| ZP_08640523.1  | 59             | <i>Brevibacillus laterosporus</i> LMG 15441                 | 564    |
| ZP_04216147.1  | 59             | <i>Bacillus cereus</i> Rock3-44                             | 566    |
| YP_001373863.1   | 60             | <i>Bacillus cytotoxicus</i> NVH 391-98                      | 565    |
| YP_004646155.1   | 60             | <i>Paenibacillus mucilaginosus</i> KNP414                   | 525    |
| ZP_10738945.1  | 61             | <i>Brevibacillus</i> sp. CF112                              | 528    |
| CAA43589.1   | 63             | <i>Brevibacillus brevis</i>                                 | 527    |
| ZP_02326602.1  | 64             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> BRL-230010 | 520    |
| ZP_02326503.1  | 65             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 520    |
| ZP_09077634.1  | 66             | <i>Paenibacillus elgii</i> B69                              | 524    |
| ZP_08511445.1  | 68             | [ <i>Paenibacillus</i> sp. HGF7                             | 525    |
| ZP_09775364.1  | 70             | <i>Paenibacillus</i> sp. Aloe-11                            | 593    |
| YP_005073223.1   | 70             | <i>Paenibacillus terrae</i> HPL-003                         | 591    |
| ZP_10241030.1  | 70             | <i>Paenibacillus peoriae</i> KCTC 3763                      | 593    |
| YP_003948511.1   | 71             | <i>Paenibacillus polymyxa</i> SC2                           | 592    |

TABLE 5.2B

| List of sequences with percent identity to PbaPro1 protein identified from the Genome Quest Patent database |                |                               |        |
|---|----------------|-------------------------------|--------|
| Patent #  | PID to PbaPro1 | Organism                      | Length |
| JP2005333991-0002   | 56.91          |                               | 562    |
| WO2012110562-0007   | 56.96          | <i>Bacillus cereus</i>        | 320    |
| WO2012110562-0006   | 57.23          | <i>Bacillus megaterium</i>    | 320    |
| EP2390321-0178  | 57.23          | <i>Bacillus thuringiensis</i> | 566    |
| EP2390321-0184  | 57.56          | <i>Bacillus caldolyticus</i>  | 319    |
| WO2007044993-0184   | 57.56          | <i>Bacillus</i> sp.           | 319    |
| US20120107907-0177  | 57.56          | <i>Bacillus caldolyticus</i>  | 544    |
| CN102168095-0002  | 57.88          |                               | 319    |
| WO2012110562-0004   | 57.88          | <i>Bacillus caldolyticus</i>  | 319    |

TABLE 5.2B-continued

| List of sequences with percent identity to PbaPro1 protein identified from the Genome Quest Patent database |                |                                       |        |
|---|----------------|---------------------------------------|--------|
| Patent #  | PID to PbaPro1 | Organism                              | Length |
| WO2012110562-0003   | 57.88          | <i>Geobacillus stearothermophilus</i> | 319    |
| WO2004011619-0056   | 57.88          |                                       | 546    |
| JP1995184649-0001   | 57.88          | <i>Lactobacillus</i> sp.              | 566    |
| JP2010535248-0240   | 57.88          | <i>Bacillus anthracis</i>             | 566    |
| US6518054-0001  | 58.2           | <i>Bacillus</i> sp.                   | 319    |
| US6103512-0003  | 58.2           |                                       | 319    |
| WO2011163237-0001   | 58.2           | <i>Geobacillus stearothermophilus</i> | 548    |
| JP1994014788-0003   | 58.25          |                                       | 317    |
| US8114656-0185  | 58.9           | <i>Bacillus cereus</i>                | 317    |
| US20120107907-0179  | 58.9           | <i>Bacillus cereus</i>                | 566    |
| WO2012110563-0005   | 59.22          | <i>Bacillus cereus</i>                | 320    |
| WO2004011619-0044   | 59.6           |                                       | 507    |
| US20120107907-0186  | 63.25          | <i>Bacillus brevis</i>                | 304    |
| JP2005229807-0018   | 70.86          | <i>Paenibacillus polymyxa</i>         | 566    |
| EP2390321-0187  | 71.1           | <i>Bacillus polymyxa</i>              | 302    |
| JP2009511072-0203   | 71.1           | <i>Paenibacillus polymyxa</i>         | 302    |

## B. Alignment of Homologous Protease Sequences

**[0409]** The amino acid sequence of the predicted mature PbaPro1 (SEQ ID NO: 23) was aligned with Thermolysin (P00800, *Bacillus thermoproteolyticus*), and protease from *Paenibacillus polymyxa* SC2 (YP\_003948511.1) using CLUSTALW software (Thompson et al., *Nucleic Acids Research*, 22:4673-4680, 1994) with the default parameters. FIG. 5.6 shows the alignment of PbaPro1 with these protease sequences.

## C. Phylogenetic Tree

**[0410]** A phylogenetic tree for full length sequence of PbaPro1 (SEQ ID NO: 22) was built using sequences of representative homologs from Table 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. *Mol Biol. Evol.* 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylodendron-phylogenetic tree printer software (<http://iubio.bio.indiana.edu/treeapp/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 5.7.

## Example 6.1

Cloning of *Paenibacillus polymyxa* SC2 Metalloprotease PpoPro1

**[0411]** The nucleic acid sequence for the PpoPro1 gene was identified in the NCBI database (NCBI Reference Sequence: NC\_014622.1 from 4536397-4538175) and is provided in SEQ ID NO: 26. The corresponding protein encoded by the PpoPro1 gene is shown in SEQ ID NO: 27. At the N-terminus, the protein has a signal peptide with a length of 24 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) *Nature Methods*, 8:785-786). The presence of a signal sequence suggests that PpoPro1 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) *Journal of Bacteriology*, 173

(21): 6820-6825). The predicted mature region of PpoPro1 protein is shown in SEQ ID NO: 28.

**[0412]** The nucleotide sequence of the PpoPro1 gene identified from NCBI database is set forth as SEQ ID NO: 26. The sequence encoding the predicted native signal peptide is shown in italics:

*ATGAAAAAGTATGGGTTTCGCTTCTGGAGGAGCTATGTTATTAGGGTC*  
*TGTCGCGTCTGGTGCATCTGCGGAGAGTTCGTTTCGGGGCCAGCTCAGC*  
 TTACACCGACCTTCCACGCCGAGCAATGGAAAGCACCTACCTCGGTATCG  
 GGGGATGACATTGTATGGAGCTATTTAAATCGACAAAAGAAATCGTTGCT  
 GGGTGTGGATAGCTCCAGTGTACGTGAACAATTCGGAATCGTTGATCGCA  
 CAAGCGACAAATCCGGTGTAAAGCATTATCGACTGAAGCAGTATGTAAAC  
 GGAATCCCGTGTATGGAGCTGAACAACTATTCTATGTGGGCAAATCTGG  
 TGAGGTACCTCTTACTTAGGAGCGGTGTAATGAGGATCAGCAGGCAG  
 AAGCTACGCAAGGTACAACCCAAAAATCAGCGCTTCTGAAGCGGTCTAC  
 ACCGCATATAAAGAAGCAGCTGCACGGATTGAAGCCCTCCCTACCTCCGA  
 CGTACTATTTCTAAAGACCTGAGGAGCCAAAGCAGTGAAGTAAAGATA  
 CTTACGCCGAAGCAGCTAACAAACGAAAAACGCTTTCTGTTGATAAGGAC  
 GAGCTGAGTCTTGATCAGGCATCTGTCTGAAAGATAGCAAATGAAGC  
 AGTGAACCGAAGAAAAAGTTCCATTGCCAAATCGCTAATCTGCAGCCTG  
 AAGTAGATCCTAAAGCAGAACTCTACTACTACCCTAAGGGGGATGACCTG  
 CTGCTGGTTTATGTAACAGAAGTTAATGTTTTAGAACCTGCCCCACTGCG  
 TACCCGCTACATTATTGATGCCAATGACGGCAGCATCGTATTCCAGTATG  
 ACATCATTAATGAAGCGACAGGCACAGGTAAAGGTGTGCTTGGTGATTCC  
 AAATCGTTCACTACTACCGCTTCCGGCAGTAGCTACCAAGTAAAAGATAC  
 AACACGCGGTAACGGAATCGTGACTTACACGGCCTCCAACCGTCAAAGCA  
 TCCCAGGTACCATTTTGACAGATGCCGATAATGTATGGAATGATCCAGCT  
 GGTGTGGACGCCATCGCTATGCTGCTAAAACCTATGATTACTATAAAGC  
 CAAATTTGGACGCAACAGCATTGACGGACGCGGTCTGCAACTTCGTTCGA  
 CGGTCCATTACGGTAGTCGCTACAACAATGCCTTCTGGAACGGCTCCCAA  
 ATGACTTATGGAGATGGAGATGGTAGCACATTTATCGCCTTCAGCGGGGA  
 CCCCAGTAGTAGGACATGAACCTACGCATGGTGTACAGAGTATACTT  
 CGAATTTGGAATATTACGGAGAGTCCGGCGCATTGAATGAAGCTTCTCA  
 GACGTTATCGGGAATGACATTGACGCGAAAACTGGCTTGTAGGCGATGA  
 TATTTACACGCCAAACATTGACGGCGATGCCCTTCGCTCAATGTCCAATC  
 CAACCCGTGACGATCAACAGATCACTATTCCAACCTGTACAGAGGCAGC  
 TCCGATAACGGCGGTGTTACACCAACAGCGGTATTATCAATAAAGCTTA  
 CTACTTGTAGCACAAAGTGGTAATTTCCATGGCGTAACTGTAATGGAA  
 TTGGCCGTGATGACGGGTGCAAAATTTACTACAGTGCCTTTACGAACCTAC  
 CTGACTTCTTCTCCGACTTCTCCAACGCAGTGTGCTGTGATCCAAGC

- continued

CGCAAAAGATCTGTACGGGGCGAACTCAGCAGAAGCAACTGCAGCTGCCA  
 AGTCTTTTGACGCTGTAGGCGTAAACTAA

**[0413]** The amino acid sequence of the PpoPro1 precursor protein is set forth as SEQ ID NO: 27. The predicted signal sequence is shown in italics, and the predicted propeptide is shown in underlined text:

*MK*KVWVSLGGAMLLGSVASGASAESSVSGPAQLTPTFHAEQWKAPTSVS  
GDDIVWSYLNROKKSLLGVDSSSVREQPRIVDRSDKSGVSHYRLKQYVN  
 GIPVYGAEQTIHVKGSGEVTSYLGAVVNEDQQAEATQGTTPKISASEAVY  
 TAYKEAAARI EALPTSDDTISKDAEPPSVSKDTYAEAAANNEKTLVSDKD  
ELSLDQASVLKDSKIEAVEPEKSSIAKIANLQPEVDPKAELYYYPKGDDL  
 LLVVYVEVNVLEPAPLRTRYIIDANDGSIVFQYDIINEATGTGKGLGDS  
 KSFTTTASGSSYQLKDTTRNGIVTYTASNRSIPGTILTDADNVWNDPA  
 GVDHAHAYAAKTYDYYKAKFGRNSIDGRGLQLRSTVHYGSRYNNAFWNGSQ  
 MTYGDGDGSTFIAPSGDPDVVGHETHGVTEYTSNLEYGESGALNEAFS  
 DVI GNDIQRKNLWVGDDIYTPNIAGDALRSMNPTLYDQPDHYSNLYRGS  
 SDNNGVHTNSGIINKAYYLLAQGGNFHGVTVNIGRDAAVQIYYSFTNY  
 LTSSDFSNARA AVIQAAKDLYGANS AEATAAKSFDAVGVN

**[0414]** The amino acid sequence of the predicted mature form of PpoPro1 is set forth as SEQ ID NO: 28:

ATGTGKGLGDSKSFTTTASGSSYQLKDTTRNGIVTYTASNRSIPGTI  
 LTDADNVWNDPAGVDHAHAYA AKTYDYYKAKFGRNSIDGRGLQLRSTVHYG  
 SRYNNAFWNGSQMTYGDGDGSTFIAPSGDPDVVGHETHGVTEYTSNLEY  
 YGESGALNEAFSDVI GNDIQRKNLWVGDDIYTPNIAGDALRSMNPTLYD  
 QPDHYSNLYRGS SDNNGVHTNSGIINKAYYLLAQGGNFHGVTVNIGRDA  
 AVQIYYSFTNYLTSSDFSNARA AVIQAAKDLYGANS AEATAAKSFDA  
 VGVN

### Example 6.2

#### Expression of *Paenibacillus polymyxa* SC2 Metalloprotease PpoPro1

**[0415]** The DNA sequence of the propeptide-mature form of PpoPro1 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, *Protein Expr Purif*, 55:40-52, 2007) by Genaray (Shanghai, China), resulting in plasmid pGX138(AprE-PpoPro1) (FIG. 1). Ligation of this gene encoding the PpoPro1 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PpoPro1 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 6.1, labeled pGX138 (AprE-PpoPro1) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted

propeptide and mature regions of PpoPro1 (SEQ ID NO: 29). The translation product of the synthetic AprE-PpoPro1 gene is shown in SEQ ID NO: 30.

[0416] The pGX138(AprE-PpoPro1) plasmid was then transformed into *B. subtilis* cells (degU<sup>H32</sup>, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

[0417] The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to an 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above, PpoPro1 was eluted from the column with a linear salt gradient from 0 to 0.25 M NaCl in the loading buffer. The corresponding active fractions were collected and concentrated. The sample was loaded onto a 320 ml Superdex 75 gel filtration column pre-equilibrated with the loading buffer described above containing 0.15 M NaCl. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

[0418] The nucleotide sequence of the synthesized PpoPro1 gene in plasmid pGX138(AprE-PpoPro1) is depicted in SEQ ID NO: 29. The sequence encoding the three residue addition (AGK) is shown in bold:

```
GTGAGAAGCAAAAATTTGGATCAGCTTGTGTTGCGTTAACGTTAAT
CTTTACGATGGCGTTTCAGCAACATGAGCGCGCAGGCTGCTGGAAAGAAT
CATCAGTGTCCAGGACCGGCTCAGCTTACACCGACATTTACGCAGAACAA
TGGAAGGCTCCGACGTCAGTTTCAGGAGACGACATCGTGTGGAGCTACCT
GAATAGACAGAAGAAAAGCCTGCTGGGAGTGGATAGCAGCAGCGTCAGAG
AGCAGTTTCAAGTCTGTTGACAGAAGCAGCGACAAAAGCGGAGTCAGCCAT
TATAGACTGAAGCAGTACGTGAATGGCATCCCGGTTTATGGCGCAGAGCA
GACAATTCATGTTGGCAAGAGCGGAGAAGTACAAGCTATCTGGGCGCTG
TGGTCAATGAAGATCAACAAGCCGAGGCTACACAGGAACACCGCGAA
ATTAGCGCTCAGAGGAGCTACACGGCGTACAAAGAAGCGGCTGCAAG
AATCGAAGCCCTGCCGACATCAGACGATACAATTTCAAAGATGCGGAGG
AGCCGAGCTCAGTTAGCAAGGATACATACGCGGAAGCCGCAACAATGAG
AAAACACTGAGCGTGGACAAGGACGAGCTGTCACTTGTATCAGGCTAGCGT
CCTTAAAGACAGCAAGATCGAGGCGGTTGAGCCTGAAAAGTCAATCAATTG
CGAAAATCGCAATCTGCAACCTGAAGTCGACCCGAGGCGGAAGTGTAC
TACTACCGAAAAGCGATGACCTGCTTCTGGTGTACGTCACGGAAGTGAA
CGTCCTGGAACCGGCACCGCTGAGAACAAGATACATCATCGACGCGAACG
ACGGAAGCATCGTCTCCAGTATGACATTATCAACGAAGCAACGGGAACG
GGCAAGGCGTTCTTGGAGACTCAAAGAGCTTACGACAACGGCTTCAGG
```

- continued

```
AAGCAGCTACCAGCTGAAAAGACACGACGAGAGGAAACCGAATCGTCACAT
ATACGGCGTCAAACAGACAAAGCATCCCTGGCCACAATCGTGACGGATGCT
GACAACGTTTGAATGATCCGGCTGGCGTGATGCCATGCTTATGCGGC
AAAAACGTATGACTATTACAAGGCGAAGTTCGGCAGAAATTCATCGATG
GCAGAGGACTGCAGCTTAGAAGCACGGTGCACCTACGGATCAAGATATAAC
AATGCCTTCTGGAACGGCAGCCAGATGACATACGGAGACGGAGATGGAAG
CACATTTATTGCATTACGCGGCGACCCTGATGTGGTTGGCCATGAGCTGA
CGCATGGCGTTACAGAATATACAGCAATCTTGAATACTACGGCGAGTCA
GGCGCTCTGAACGAGGCATTTAGCGATGTTATCGGCAATGACATCCAGAG
AAAAAATGGCTGGTGGGCGACGATATTACACGCCTAATATCGCTGGCG
ATGCCCTTAGATCAATGTCAAACCCGACGCTGTATGATCAGCCTGACCAC
TACTCAAACCTGTATAGAGGCTCATCAGATAACGGAGGCGTCCATACGAA
TAGCGGCATCATTAAACAGGCATATTATCTTCTGGCCAGGGCGGCAATT
TTCATGGAGTGACGGTTAATGGAATTGGAAGAGACGCGCCGTCGAATC
TACTACAGCGCTTTCACGAACCTTACATCAAGCTCAGACTTTAGCAA
TGCCAGAGCTGCTGTTATCCAGGCGAGGAGGATCTTTACGGCGCAACT
CAGCCGAAGCTACGCGCGAGCTAAATCATTTGATGCGTGGCGTTAAT
```

[0419] The amino acid sequence of the PpoPro1 precursor protein expressed from plasmid pGX138(AprE-PpoPro1) is depicted in SEQ ID NO: 30. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) is shown in **bold**, and the predicted pro-peptide is shown in underlined text.

```
MRSKKLWISLLFALTLIFTMAFSNMSAQAAAGKESSVSGPAQLTPTFHAEQ
WKAPTSVSGDDIVWSYLNROKKSLLGVDSSSVREQFRIVDRSDKSGVSH
YRLKQYVNGIPVYGAEQTIHVKGSGEVTSYLGAVVNEDQQAEATQGTTPK
ISASEAVYTAYKEAAARIEALPTSDDTISKDAEEPSVSKDYAEAAANNE
KTLSDVKDELSDQASVLKDSKIEAVEPEKSSIAKIANLQPEVDPKAELY
YYPKGDLLLLVYVTEVNVLEPAPLRTRYIIDANDGSIVFQYDINEATGT
GKGVLGDSKSFTTTASGSSYQLKDTTRNGIVTYTASNRSQIPGTLTDA
DNVWNPAGVDAHAYAAKTYDYKAKFGRNSIDGRGLQLRSTVHYGSRYN
NAFWNGSQMTYGDGDSFTFIAPSGDPDVGHEALTHGVTEYTSNLEYGES
GALNEAFSDVIGNDIQRKNWLVGDDIYTPNIAGDALRSMNPTLYDQPDH
YSNLRYGSSDNGGVHTNSGIINKAYYLLAQGGFNHGVTVNGIGRDAAVQI
YYSAPTNYLTSSSDFSNARAIVIQAAKDLYGANSAEATAAKSFDVAVGN
```

### Example 6.3

#### Proteolytic Activity of Metalloprotease PpoPro1

[0420] The proteolytic activity of purified PpoPro1 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in

100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ L of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ L of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ L of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ L supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a Spectra-Max 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of duplicate assays, and the value varies no more than 5%. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (FIG. 6.2) indicates PpoPro1 is an active protease.

#### Example 4

##### pH Profile of Metalloprotease PpoPro1

[0421] With azo-casein as the substrate, the pH profile of PpoPro1 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ L of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2  $\mu$ L diluted enzyme (250 ppm in Milli-Q H<sub>2</sub>O) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ L of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 6.3. Enzyme activity at each pH was reported as relative activity where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 6.3, the optimal pH of PpoPro1 is about 7, with greater than 70% of maximal activity retained between 5.5 and 8.5.

#### Example 6.5

##### Temperature Profile of Metalloprotease PpoPro1

[0422] The temperature profile of PpoPro1 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assay. The enzyme sample and azo-casein substrate were prepared as in Example 6.3. Prior to the reaction, 50  $\mu$ L of 1.5% azo-casein and 45  $\mu$ L Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ L PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ L of diluted PpoPro1 (100 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ L of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 6.3. The activity was reported as relative activity where the activity at the optimal temperature was set to be 100%. The tested temperatures were 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data in FIG.

6.4 suggests that PpoPro1 showed an optimal temperature at 50° C., and retained greater than 70% of its maximum activity between 40 and 55° C.

#### Example 6.6

##### Cleaning Performance of Metalloprotease PpoPro1

[0423] The cleaning performance of PpoPro1 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 or 8 using a model automatic dishwashing (ADW) detergent (AT detergent). Prior to the reaction, purified PpoPro1 was diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent (composition shown in Table 6.1) with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1), in the presence of a bleach component ((Peracid N,N-phthaloylaminoperoxyacetic acid-PAP). To initiate the reaction, 180  $\mu$ L of AT detergent buffered at pH 6 or 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ L of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ L of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ L water. Following the addition of 180  $\mu$ L of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliter of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net  $A_{405}$  was subsequently calculated by subtracting the  $A_{405}$  of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 for PpoPro1 in AT dish detergent, in the presence of bleach, is shown in FIGS. 6.5A and 6.5B.

TABLE 6.1

| Composition of AT dish detergent formula with bleach       |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| PAP (peracid N,N-phthaloylaminoperoxyacetic acid)          | 0.057                 |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.



## Example 6.7

## Comparison of PpoPro1 to Other Metalloproteases

**[0424]** Identification of Homologous Proteases

**[0425]** Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res*, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The predicted mature protein amino acid sequence for PpoPro1 (SEQ ID NO: 28) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 6.2A and 6.2B provide a list of sequences with the percent identity to PpoPro1. The length in Table 6.2 refers to the entire sequence length of the homologous proteases.

TABLE 6.2A

| List of sequences with percent identity to PpoPro1 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PpoPro1 | Organism  | Length |
| P00800   | 56             | <i>Bacillus thermoproteolyticus</i>                         | 548    |
| ZP_08640523.1  | 57             | <i>Brevibacillus laterosporus</i> LMG 15441                 | 564    |
| AAA22623.1   | 57             | <i>Bacillus caldolyticus</i>                                | 544    |
| ZP_08093424.1  | 59             | <i>Planococcus donghaensis</i> MPA1U2                       | 553    |
| ZP_10738945.1  | 60             | <i>Brevibacillus</i> sp. CF112                              | 528    |
| CAA43589.1   | 62             | <i>Brevibacillus brevis</i>                                 | 527    |
| ZP_02326503.1  | 62             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> BRL-230010 | 520    |
| YP_005495105.1   | 63             | <i>Bacillus megaterium</i> WSH-002                          | 562    |
| YP_001373863.1   | 64             | <i>Bacillus cytotoxicus</i> NVH 391-98                      | 565    |
| ZP_04310163.1  | 64             | <i>Bacillus cereus</i> BGSC 6E1                             | 581    |
| BAA06144.1   | 64             | <i>Lactobacillus</i> sp.]                                   | 566    |
| ZP_08511445.1  | 65             | <i>Paenibacillus</i> sp. HGF7                               | 525    |
| ZP_04216147.1  | 65             | <i>Bacillus cereus</i> Rock3-44                             | 566    |
| ZP_09071078.1  | 68             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 524    |
| ZP_09077634.1  | 69             | <i>Paenibacillus elgii</i> B69                              | 595    |
| YP_005073224.1   | 79             | <i>Paenibacillus terrae</i> HPL-003                         | 599    |
| ZP_10241029.1  | 80             | <i>Paenibacillus peoriae</i> KCTC 3763                      | 591    |
| YP_005073223.1   | 93             | <i>Paenibacillus terrae</i> HPL-003                         | 591    |
| ZP_10241030.1  | 95             | <i>Paenibacillus peoriae</i> KCTC 3763                      | 593    |
| ZP_09775364.1  | 95             | <i>Paenibacillus</i> sp. Aloe-11                            | 593    |
| YP_003872179.1   | 97             | <i>Paenibacillus polymyxa</i> E681                          | 592    |
| YP_003948511.1   | 100            | <i>Paenibacillus polymyxa</i> SC2                           | 592    |

TABLE 6.2B

| List of sequences with percent identity to PpoPro1 protein identified from the Genome Quest Patent database |                |                            |        |
|---|----------------|----------------------------|--------|
| Patent #  | PID to PpoPro1 | Organism                   | Length |
| US20120107907-0187  | 97.34          | <i>Bacillus polymyxa</i>   | 302    |
| US5962264-0004  | 65.48          | empty                      | 566    |
| WO2012110563-0005   | 65.16          | <i>Bacillus cereus</i>     | 320    |
| JP1994070791-0002   | 64.52          | empty                      | 317    |
| WO2012110562-0005   | 64.19          | <i>Bacillus cereus</i>     | 320    |
| WO2012110563-0004   | 63.34          | <i>Bacillus megaterium</i> | 320    |
| JP2002272453-0002   | 61.98          | <i>Bacillus megaterium</i> | 562    |
| WO2004011619-0047   | 61.49          | empty                      | 532    |
| EP2390321-0186  | 62.58          | <i>Bacillus brevis</i>     | 304    |

TABLE 6.2B-continued

| List of sequences with percent identity to PpoPro1 protein identified from the Genome Quest Patent database |                |                                       |        |
|---|----------------|---------------------------------------|--------|
| Patent #  | PID to PpoPro1 | Organism                              | Length |
| US6518054-0002  | 59.22          | <i>Bacillus</i> sp.                   | 316    |
| US6518054-0001  | 58.52          | <i>Bacillus</i> sp.                   | 319    |
| US20120107907-0176  | 58.52          | <i>Bacillus stearothermophilus</i>    | 548    |
| JP2005229807-0019   | 93.05          | <i>Paenibacillus polymyxa</i>         | 566    |
| WO2012110562-0003   | 58.2           | <i>Geobacillus stearothermophilus</i> | 319    |
| WO2004011619-0044   | 59.27          | empty                                 | 507    |
| EP2390321-0185  | 66.13          | <i>Bacillus cereus</i>                | 317    |
| JP1995184649-0001   | 65.71          | <i>Lactobacillus</i> sp.              | 566    |
| EP2178896-0184  | 65.38          | <i>Bacillus anthracis</i>             | 566    |

**[0426]** Alignment of Homologous Protease Sequences

**[0427]** The amino acid sequence of predicted mature PpoPro1 (SEQ ID NO: 28) was aligned with thermolysin (P00800, *Bacillus thermoproteolyticus*) and protease from *Paenibacillus polymyxa* SC2 (YP\_003948511.1) using CLUSTALW software (Thompson et al., *Nucleic Acids Research*, 22:4673-4680, 1994) with the default parameters. FIG. 6.6 shows the alignment of PpoPro1 with these protease sequences.

**[0428]** Phylogenetic Tree

**[0429]** A phylogenetic tree for precursor PpoPro1 (SEQ ID NO: 27) was built using sequences of representative homologs from Tables 6.2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. *Mol Biol. Evol.* 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylodendron-phylogenetic tree printer software (<http://iubio.bio.indiana.edu/tree-app/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 6.7.

## Example 7.1

Cloning of *Paenibacillus* Hunanensis Metalloprotease PhuPro1

**[0430]** A strain (DSM22170) of *Paenibacillus humanensis* was selected as a potential source of enzymes which may be useful in various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus humanensis* strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the genes identified after annotation in *Paenibacillus humanensis* encodes a metalloprotease and the sequence of this gene, called PhuPro1, is provided in SEQ ID NO: 31. This gene has an alternative start codon (TTG). The corresponding protein encoded by the PhuPro1 gene is shown in SEQ ID NO: 32. At the N-terminus, the protein has a signal peptide with a length of 23 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) *Nature Methods*, 8:785-786). The pres-

ence of a signal sequence suggests that PhuPro1 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) Journal of Bacteriology, 173 (21): 6820-6825). The predicted mature region of PhuPro1 protein is shown in SEQ ID NO: 33.

[0431] The nucleotide sequence of the PhuPro1 gene isolated from *Paenibacillus hunanensis* is set forth as SEQ ID NO: 31. The sequence encoding the predicted native signal peptide is shown in italics:

*TTGAAAAAACAGTTGGTCTTTACTTGCAGGTAGCTTGCCTCGTTGGTGC*  
*TACAACGTC*CGCTTTTCGACAGCAAGCAAAATGATCTGGCACCACCTCGGTG  
ATTACACGCCAAAATGATTACGCAAGCAACAGGCATCCTGGCGCTAGT  
GGCGATGCTAAAGTATGGAAGTCTCTGGAGAAGCAAAAACGTACCATCGT  
AACCGATGATGCAGCTTCTGCTGATGTGAAGGAATTGTTTGGATCACAA  
AACGTCGAATCCGATTCTCAAACCGGTACAGAGCACTATCGCTGAACAA  
ACCTTTAAAGGCATCCAGTCTATGGCGCAGAGCAACACTGCACCTTGA  
CAATCCGGCAATGTATCTCTGTACATGGGTCAGGTTGTTGAGGATGTGT  
CCGCTAAACTGGAAGCTCCGATTCCAAAAAGGCGTAACTGAGGATGTA  
TAGCTTCGGATACGAAAAATGATCTGGTAACACCAGAAATCAGCGCTTC  
TCAAGCCATCTCGATTGCTGAAAAGGATGCAGCTTCCAAAATCCGCTCC  
TCGGCGAAGCACAAAAACGCCAGAAGCGAAGCTGTATATCTACGCTCCT  
GAGGATCAAGCAGCACGCTCGGCTTATGTGACAGAAGTAAACGTACTGGA  
GCCATCTCCGCTCGCTACTCGCTATTTGTAGATGCAAAAACAGGTTCGA  
TCCTGTCCAAATATGATCTGATTGAGCATGCAACAGGTACAGGTAAGGG  
GTACTGGGTGATACCAAGTCTTCACTGTAGGTACTCCGGTCTTCTCTA  
TGTGATGACTGATAGCACGCGTGGAAAAGGTATCCAAACCTACACGGCGT  
CTAACCGCACATCACTGCCAGGTAGCACTGTAACGAGCAGCAGCAGCACA  
TTTAACGATCCAGCATCTGTGATGCCATGCGTATGCACAAAAAGTATA  
TGATTTCTACAAATCCAACTTTAACCGCAACAGCATCGACGGTAATGGTC  
TGGCTATCCGCTCCACTACGCACATTTCCACACGTTATAACAATGCGTTCT  
TGGAATGGTTCCCAATGGTATACGGTGATGGCGATGGTTCCGAATTCAT  
CGCATTCTCCGGCGACCTTGACGTAGTAGGTCACGAGCTGACACACGGTG  
TAACCGAGTACACAGCGAACCTGGAATACTATGGTCAATCCGGTGCACGT  
AACGAATCCATTTCCGATATCTTTGGTAACACAATCGAAGTAAAACTG  
GATGGTAGGCGATGCGATCTACACACCAGGCGTATCCGGCGATGCTCTTC  
GCTACATGGATGATCCAAACAAAAGGTGGACAACAGCGCGTATGGCAGAT  
TACAACAACACAAGCGCTGATAATGGCGGTGATACACAAAACAGTGGTAT  
CCCCGAATAAAGCATACTACTTGTGTCGACAGGGTGGCACAATTTGGCGGTG  
TAAATGTAACAGGTATCGGTCGTCGCAAGCGATCCAGATCGTTTACCGT  
GCACTAACATACTACCTGACATCCACATCTAACTTCTCGAAGTACCGTTCT

- continued

TGCAATGGTGCAAGCATCTACAGACCTGTACGGTGCAAACCTACACAAA

CAACAGCGGTGAAAAACTCGCTGAGCGCAGTAGGCATTAAC

[0432] The amino acid sequence of the PhuPro1 precursor protein is set forth as SEQ ID NO: 32. The predicted signal sequence is shown in italics, and the predicted pro-peptide is shown in underlined text:

*MKKT*VGLLLAGSLLVGATTSAFAAEANDLAPLDYTPKLITQATGITGAS  
GDAKVWKFLEKQKRTIVTDDAASADVKELFEITKRQSDSQTGTEHYRLNQ  
TFKGIPVYGAEQTLHFDKSGNVSLYMGQVVEDVSAKLEASDSKKGVTEDV  
YASDTKNLDLVTPEISASQAISIAEKDAASKIGSLGEAOKTPEAKLYIYAP  
EDQAARLLAYVTEVNVLEPSPLRTRYFVDAKTGSILFQYDLIEHATGTGKG  
VLGDTKSFTVGTSGSSYVMTDSTRGKGIQTYTASNRTSLPGSTVTSST  
FNDPASVDAHAYAQKVYDFYKSNFNRN SIDGNGLAIRSTHYSRYNNAF  
WNGSQMVYGDGDSQFIAPSGDLVDVGHETHGVTEYTANLEYGQSGAL  
NESISDIFGNTIEGKNWVGDAIYTPGVSGDALRYMDDPTKGGQPARMAD  
YNNTSADNGGVHTNSGIPNKAYLLAQGGTFGGVNVGTIGRSQAIQIVYR  
ALTYLTSSTNSFNYSAMVQASTDLYGANSTQTTAVKNSLSAVGIN

[0433] The amino acid sequence of the predicted mature form of PhuPro1 is set forth as SEQ ID NO: 33:

ATGTGKGVLGDTKSFTVGTSGSSYVMTDSTRGKGIQTYTASNRTSLPGST  
VTSSTFNDPASVDAHAYAQKVYDFYKSNFNRN SIDGNGLAIRSTHYS  
TRYNNAFWNGSQMVYGDGDSQFIAPSGDLVDVGHETHGVTEYTANLEY  
YGQSGALNESISDIFGNTIEGKNWVGDAIYTPGVSGDALRYMDDPTKGG  
QPARMADYNNTSADNGGVHTNSGIPNKAYLLAQGGTFGGVNVGTIGRSQ  
AIQIVYRALTYLTSSTNSFNYSAMVQASTDLYGANSTQTTAVKNSLSA  
VGIN

## Example 7.2

### Expression of *Paenibacillus hunanensis* Metalloprotease PhuPro1

[0434] The DNA sequence of the propeptide-mature form of PhuPro1 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX149(AprE-PhuPro1) (FIG. 7.1). Ligation of this gene encoding the PhuPro1 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PhuPro1 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 1, labeled pGX149 (AprE-PhuPro1) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PhuPro1 (SEQ ID NO: 34). The translation product of the synthetic AprE-PhuPro1 gene is shown in SEQ ID NO: 35.

[0435] The pGX149(AprE-PhuPro1) plasmid was then transformed into *B. subtilis* cells (degU<sup>H32</sup>, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

[0436] The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius Stedim). After filtering, this sample was applied to a 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above; and the active flow-through fractions were collected and concentrated via 10K Amicon Ultra for further analyses.

[0437] The nucleotide sequence of the synthesized PhuPro1 gene in plasmid pGX149(AprE-PhuPro1) is depicted in SEQ ID NO: 34. The sequence encoding the three residue addition (AGK) is shown in bold:

GTGAGAAGCAAAAATTGTGGATCAGCTTGTGTTGCGTTAACGTTAAT  
 CTTTACGATGGCGTTCAGCAACATGAGCGCGCAGGCT**GCTGGAAA**AGCAG  
 AAGCTAATGATCTTGCCCGCTTGGCGATTATACACCGAAGCTTATTACA  
 CAGGCAACGGGAATTACAGCGCATCAGCGGATGCGAAGGTGTGGAAAGTT  
 CCTGGAGAAGCAGAAGAGAACGATTGT CACGGACGACGCCAAGCGCGG  
 ATGTCAAGGAGCTGTTTCGAGATCAGCAAGAGACAGAGCGATAGCCAGACG  
 GGAACGGAGCATTACAGACTGAACAGACGTTCAAGGGCATTCCGGTCTA  
 CGGAGCTGAACAAACGCTGCATTTTGATAAAAGCGGCAACGTCTCACGTG  
 ACATGGGCCAAGTCGTTGAGGACGTTAGCGCCAAACTTGAGGCTAGCGAC  
 AGCAAGAAAGCGGTACAGAAGATGTCTACGCGTCAGACACGAAAAACGA  
 CCTGGTTACACCGGAAATCTCAGCTTACAGGCCATCTCAATTGCGAGAGA  
 AAGACGCAGCGTCAAAAATCGGCTCAGTGGCGAGGCTCAGAAAAACCGG  
 GAGGCGAAACTTTACATCTACGCCCTGAGGACAGGCTGCGAGACTGGC  
 TTACGTGACAGAAGTTAATGTGCTGGAGCCGTCACCGCTTAGAACGAGAT  
 ATTTCTGGACGCAAGACGGGCAGCATTTCTGTTTCAGTACGATCTTATC  
 GAACACCGCAGCAGGCACAGAAAGGGAGTTCTGGGAGACACAAAAGCTT  
 CACGGTTGGCACGTCAGGCAGCAGCTACGTGATGACAGACAGCAGCAGAG  
 GCAAGGGCATTCAAACGTATACAGCGAGCAACAGAAACAGCCTGCCGGGA  
 AGCACAGTCACGAGCTCATCATCAACGTTTAAATGACCCGGCCTCAGTGG  
 TGCTCACGCATACCGCGAGAAAGTGTACGACTTCTACAAAAGCAACTTCA  
 ATAGAAACAGCATCGACGGAACGGCCTTGCGATCAGAAGCAGCAGCAGC  
 TACAGCAACAAGATACAACAACGCCCTTCTGGAACGGCAGCCAAATGGTTTA  
 CGGCGATGGCGCAGGATCACAGTTTATCGCATTTGACGGAGACCTGGACG  
 TCGTTGGCCATGAGCTGACACATGGCGTTACGGAGTACACAGCAAACTCG  
 GAATACTATGGCCAGTCAGCGCCCTTAAACGAGAGCATCAGCGACATTTT

-continued

TGGCAATACGATCGAAGGAAGAACTGGATGGTCGGCGACGCAATCTACA  
 CACCGGGCGTTTCAGGCGATGCACTGAGATATATGGACGACCCGACAAAG  
 GGCGGACAGCCGGCCAGAATGGCGGATTACAATAATACGTACAGAGATA  
 CGGCGCGCTGCATACAATAAGCGGCATCCCTAACAAAGCATATTACCTGC  
 TTGCGCAAGGAGGAACATTGGCGCGCTGAATGTTACGGGCATTGGCAGA  
 TCACAAGCGATTGATCGTTTACAGAGCGCTGACGTACTACCTTACGAG  
 CACGAGCAATTTTAGCAACTACAGAAGCGCAATGGTGCAGGCAAGCAGG  
 ATCTGTATGGCGCAAATCAACACAAACGACGGCGGTCAAGAATAGCCTT  
 TCAGCAGTGGGCATTAATACTAA

[0438] The amino acid sequence of the PhuPro1 precursor protein expressed from plasmid pGX149(AprE-PhuPro1) is depicted in SEQ ID NO: 35. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) is shown in **bold**, and the predicted pro-peptide is shown in underlined text.

*MRSKKLWISLLFALTLIFTMAFSNMSAQ**AGK**AEANDLAPLDYTPKLIIT*  
QATGITGASGDAKVVWKFLEKQKRTIVTDDAASADVKELEIITKROSDSQT  
GTEHYRLNQTFKGIPVYGAEQTLHFDKSGNVSLYMGQVVEDVSAKLEASD  
SKKGVTEDEVYASDTKNDLVTPEISASQAISIAEKDAASKIGSLGEAQKTP  
EAKLYIYAPEDQARLAYVTEVNVLEPSPLRTRYFVDAKTSILFQYDLI  
EHATGTGKVLGDTKSFTVGTSGSSVMTDSTRGKGIQTYTASNRTSLPG  
 STVTSSTSTFNDPASVDAHAYAQKVYDFYKSNFNRSIDGNGLAIRSTTH  
 YSTRYNNAFWNGSQMVYGDGDSQFIAFSGDLDDVVGHELTHGVTEYTANL  
 EYYGQSGALNESISDIFGNTIEGKNWVGDALYTPGVSGDALRYMDDPTK  
 GGQPARMADYNNTSADNGGVHTNSGIPNKAYYLLAQQGTFGGVNVVTGIGR  
 SQAIQIVYRALTYYLSTSNFNSYRSAMVQASTDLYGANSTQTTAVKNSL  
 SAVGIN

### Example 7.3

#### Proteolytic Activity of Metalloprotease PhuPro1

[0439] The proteolytic activity of purified metalloprotease PhuPro1 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50 μl of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50 μl of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100 μl of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120 μl supernatant was transferred to a new 96-MTP, and

absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a SpectraMax 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of triplicate assays. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (shown in FIG. 7.2) indicates that PhuPro1 is an active protease.

#### Example 7.4

##### pH Profile of Metalloprotease PhuPro1

**[0440]** With azo-casein as the substrate, the pH profile of metalloprotease PhuPro1 was studied in 12.5 mM acetate/Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2  $\mu$ l Milli-Q H<sub>2</sub>O diluted enzyme (125 ppm) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 7.3, the optimal pH of PhuPro1 is about 6, with greater than 70% of maximal activity retained between 5 and 8.

#### Example 7.5

##### Temperature Profile of Metalloprotease PhuPro1

**[0441]** The temperature profile of metalloprotease PhuPro1 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assays. The enzyme sample and azo-casein substrate were prepared as in Example 7.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted enzyme (50 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 7.3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data in FIG. 7.4 suggests that PhuPro1 showed an optimal temperature at 60° C., and retained greater than 70% of its maximum activity between 45 and 65° C.

#### Example 7.6

##### Cleaning Performance of Metalloprotease PhuPro1

**[0442]** The cleaning performance of PhuPro1 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified protease samples were diluted with a dilution solution containing 10 mM NaCl, 0.1

mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1) (detergent composition shown in Table 7.1). To initiate the reaction, 180  $\mu$ l of the AT detergent buffered at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ l of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ l water. Following the addition of 180  $\mu$ l of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as the “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net  $A_{405}$  was subsequently calculated by subtracting the  $A_{405}$  of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 in AT detergent for PhuPro1 is shown in FIGS. 7.5A and 7.5B.

TABLE 7.1

| Composition of AT detergent                                |                       |
|--|-----------------------|
| Ingredient   | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                          | 0.143                 |
| Sodium citrate   | 1.86                  |
| Citric acid*   | varies                |
| Plurafac® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate   | 0.006                 |
| Bayhibit® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)            | 0.029                 |
| PEG 6000   | 0.043                 |
| PEG 1500   | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

#### Example 7.7

##### Comparison of PhuPro1 to Other Proteases

##### [0443] A. Identification of Homologous Proteases

**[0444]** Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res*, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The predicted mature protein amino acid sequence for PhuPro1 (SEQ ID NO: 33) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 7.2A and 7.2B provide a list of sequences with the percent identity to PhuPro1. The length in Table 7.2 refers to the entire sequence length of the homologous proteases.

TABLE 7.2A

| List of sequences with percent identity to PhuPro1 protein identified from the NCBI non-redundant protein database |                |   |        |
|--|----------------|---|--------|
| Accession #  | PID to PhuPro1 | Organism  | Length |
| P00800   | 55             | <i>Bacillus thermoproteolyticus</i>                     | 548    |
| AAB02774.1   | 55             | <i>Geobacillus stearothermophilus</i>                   | 552    |
| EJS73098.1   | 56             | <i>Bacillus cereus</i> BAG2X1-3                         | 566    |
| BAD60997.1   | 56             | <i>Bacillus megaterium</i>                              | 562    |
| ZP_04216147.1  | 57             | <i>Bacillus cereus</i> Rock3-44                         | 566    |
| YP_893436.1  | 56             | <i>Bacillus thuringiensis</i> str. Al Hakam             | 566    |
| ZP_08640523.1  | 58             | <i>Brevibacillus laterosporus</i>                       | 564    |
| ZP_09069194.1  | 59             | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650 | 502    |
| YP_002770810.1   | 60             | <i>Brevibacillus brevis</i>                             | 528    |
| ZP_08511445.1  | 61             | <i>Paenibacillus</i> sp. HGF7                           | 525    |
| P43263   | 61             | <i>Brevibacillus brevis</i>                             | 527    |
| ZP_09775365.1  | 62             | <i>Paenibacillus</i> sp. Aloe-11                        | 580    |
| ZP_09077634.1  | 66             | <i>Paenibacillus elgii</i> B69                          | 524    |
| P29148   | 68             | NPPE_PAPEO  | 590    |
| ZP_09775364.1  | 69             | <i>Paenibacillus</i> sp. Aloe-11                        | 593    |
| ZP_10241030.1  | 69             | <i>Paenibacillus peoriae</i> KCTC 3763                  | 593    |
| YP_005073223.1   | 69             | <i>Paenibacillus terrae</i> HPL-003                     | 591    |

TABLE 7.2B

| List of sequences with percent identity to PhuPro1 protein identified from the Genome Quest Patent database |                |                                       |        |
|---|----------------|---------------------------------------|--------|
| Patent ID #   | PID to PhuPro1 | Organism                              | Length |
| WO2012110562-0003   | 56.23          | <i>Geobacillus stearothermophilus</i> | 319    |
| US6518054-0001  | 56.55          | <i>Bacillus</i> sp.                   | 319    |
| JP2002272453-0002   | 56.69          | <i>Bacillus megaterium</i>            | 562    |
| US20090123467-0184  | 56.73          | <i>Bacillus anthracis</i>             | 566    |
| US6103512-0003  | 56.87          |                                       | 319    |
| EP0867512-0002  | 56.96          |                                       | 316    |
| WO2012110562-0005   | 57.1           | <i>Bacillus cereus</i>                | 320    |
| WO2012110563-0005   | 58.06          | <i>Bacillus cereus</i>                | 320    |
| US20120107907-0187  | 68.44          | <i>Bacillus polymyxa</i>              | 302    |

**[0445]** B. Alignment of Homologous Protease Sequences

**[0446]** The amino acid sequence of predicted mature PhuPro1 (SEQ ID NO: 33) protein was aligned with Proteinase T (P00800, *Bacillus thermoproteolyticus*), and protease from *Paenibacillus terrae* HPL-003 (YP\_005073223.1) using CLUSTALW software (Thompson et al., Nucleic Acids Research, 22:4673-4680, 1994) with the default parameters. FIG. 7.6 shows the alignment of PhuPro1 with these protease sequences.

**[0447]** C. Phylogenetic Tree

**[0448]** A phylogenetic tree for full length sequence of PhuPro1 (SEQ ID NO: 2) was built using sequences of representative homologs from Table 2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. Mol Biol. Evol. 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of diver-

gence between the sequences. The phylodendron-phylogenetic tree printer software (<http://iubio.bio.indiana.edu/tree-app/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 7.7.

## Example 7.8

## Terg-o-Tometer Performance Evaluation of PhuPro1

**[0449]** The wash performance of PhuPro1 was tested in a laundry detergent application using a Terg-o-Tometer (Instrument Marketing Services, Inc, Fairfield, N.J.). The performance evaluation was conducted at 32° C. and 16° C. The soil load consisted of two of each of the following stain swatches: EMPA116 Blood, Milk, Ink on cotton (Test materials AG, St. Gallen, Switzerland), EMPA117 Blood, Milk, Ink on poly-cotton (Test materials AG, St. Gallen, Switzerland), EMPA112 Cocoa on cotton (Test materials AG, St. Gallen, Switzerland), and CFT C-10 Pigment, Oil, and Milk content on cotton (Center for Testmaterials BV, Vlaardingen, Netherlands), plus extra white interlock knit fabric to bring the total fabric load to 40 g per beaker of the Terg-o-Tometer, which was filled with 1 L of deionized water. The water hardness was adjusted to 6 grains per gallon, and the pH in the beaker was buffered with 5 mM HEPES, pH 8.2. Heat inactivated Tide Regular HDL (Proctor & Gamble), a commercial liquid detergent purchased in a local US supermarket, was used at 0.8 g/L. The detergent was inactivated before use by treatment at 92° C. in a water bath for 2-3 hours followed by cooling to room temperature. Heat inactivation of commercial detergents serves to destroy the activity of enzymatic components while retaining the properties of the non-enzymatic components. Enzyme activity in the heat inactivated detergent was measured using the Suc-AAPF-pNA assay for measuring protease activity. The Purafect® Prime HA, (Genencor Int'l) and PhuPro1 proteases were each added to final concentrations of 1 ppm. A control sample with no enzyme was included. The wash time was 12 minutes. After the wash treatment, all swatches were rinsed for 3 minutes and machine-dried at low heat.

**[0450]** Four of each type of swatch were measured before and after treatment by optical reflectance using a Tristimulus Minolta Meter CR-400. The difference in the L, a, b values was converted to total color difference (dE), as defined by the CIE-LAB color space. Cleaning of the stains is expressed as percent stain removal index (% SRI) by taking a ratio between the color difference before and after washing, and comparing it to the difference of unwashed soils (before wash) to unsoiled fabric, and averaging the eight values obtained by reading two different regions of each washed swatch. Cleaning performances of PhuPro1 and Purafect® Prime HA proteases at 32° C. are shown in Tables 7.8A and FIG. 7.8A and at 16° C. are shown in Table 7.8B and FIG. 7.8B.

TABLE 7.8A

| Cleaning performance of PhuPro1 at 32° C. |                    |                    |                    |                    |                    |                    |                    |                    |
|---|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| ppm enzyme                                | Purafect Prime HA  |                    | PhuPro1            |                    | Purafect Prime HA  |                    | PhuPro1            |                    |
|   | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] |
| EMPA-116                                  |                    |                    |                    | EMPA-117           |                    |                    |                    |                    |
| 0   | 0.25               | 0.02               | 0.25               | 0.02               | 0.19               | 0.02               | 0.19               | 0.02               |
| 0.2                                       | 0.31               | 0.02               | 0.31               | 0.01               | 0.31               | 0.03               | 0.32               | 0.04               |
| 0.5                                       | 0.34               | 0.02               | 0.33               | 0.03               | 0.34               | 0.02               | 0.37               | 0.02               |
| 1   | 0.35               | 0.03               | 0.36               | 0.02               | 0.38               | 0.03               | 0.42               | 0.03               |
| 1.5                                       | 0.36               | 0.02               | 0.37               | 0.03               | 0.35               | 0.03               | 0.43               | 0.03               |
| EMPA-112                                  |                    |                    |                    | CFT C-10           |                    |                    |                    |                    |
| 0   | 0.15               | 0.03               | 0.15               | 0.03               | 0.07               | 0.01               | 0.07               | 0.01               |
| 0.2                                       | 0.17               | 0.04               | 0.14               | 0.02               | 0.11               | 0.01               | 0.15               | 0.01               |
| 0.5                                       | 0.19               | 0.02               | 0.19               | 0.04               | 0.13               | 0.01               | 0.16               | 0.03               |
| 1   | 0.20               | 0.03               | 0.22               | 0.03               | 0.17               | 0.01               | 0.17               | 0.01               |
| 1.5                                       | 0.24               | 0.03               | 0.25               | 0.04               | 0.17               | 0.02               | 0.20               | 0.02               |

TABLE 7.8B

| Cleaning performance of PhuPro1 at 16° C. |                    |                    |                    |                    |                    |                    |                    |                    |
|---|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| ppm enzyme                                | Purafect Prime HA  |                    | PhuPro1            |                    | Purafect Prime HA  |                    | PhuPro1            |                    |
|   | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] | Average % SRI (dE) | 95CI [% SRI] (dE)] |
| EMPA-116                                  |                    |                    |                    | EMPA-117           |                    |                    |                    |                    |
| 0   | 0.14               | 0.02               | 0.14               | 0.02               | 0.12               | 0.01               | 0.12               | 0.01               |
| 0.2                                       | 0.19               | 0.02               | 0.17               | 0.03               | 0.17               | 0.02               | 0.14               | 0.03               |
| 0.5                                       | 0.22               | 0.03               | 0.28               | 0.04               | 0.20               | 0.03               | 0.22               | 0.01               |
| 1   | 0.24               | 0.02               | 0.26               | 0.02               | 0.20               | 0.01               | 0.24               | 0.04               |
| 1.5                                       | 0.23               | 0.03               | 0.26               | 0.03               | 0.23               | 0.02               | 0.25               | 0.02               |
| EMPA-112                                  |                    |                    |                    | CFT C-10           |                    |                    |                    |                    |
| 0   | 0.09               | 0.03               | 0.09               | 0.03               | 0.07               | 0.01               | 0.07               | 0.01               |
| 0.2                                       | 0.07               | 0.01               | 0.09               | 0.02               | 0.08               | 0.02               | 0.06               | 0.01               |
| 0.5                                       | 0.11               | 0.02               | 0.12               | 0.03               | 0.10               | 0.01               | 0.09               | 0.01               |
| 1   | 0.11               | 0.02               | 0.12               | 0.02               | 0.13               | 0.01               | 0.15               | 0.01               |
| 1.5                                       | 0.13               | 0.03               | 0.19               | 0.03               | 0.13               | 0.01               | 0.11               | 0.01               |

## Example 8.1

Cloning of *Paenibacillus amylolyticus*  
Metalloprotease PamPro1

**[0451]** A strain (DSM11747) of *Paenibacillus amylolyticus* was selected as a potential source of enzymes which may be useful in various industrial applications. Genomic DNA for sequencing was obtained by first growing the strain on Heart Infusion agar plates (Difco) at 37° C. for 24 hr. Cell material was scraped from the plates and used to prepare genomic DNA with the ZF Fungal/Bacterial DNA miniprep kit from Zymo (Cat No. D6005). The genomic DNA was used for genome sequencing. The entire genome of the *Paenibacillus amylolyticus* strain was sequenced by BaseClear (Leiden, The Netherlands) using the Illumina's next generation sequencing technology. After assembly of the data, contigs were annotated by BioXpr (Namur, Belgium). One of the

genes identified after annotation in *Paenibacillus amylolyticus* encodes a metalloprotease and the sequence of this gene, called PamPro1, is provided in SEQ ID NO: 36. The corresponding protein encoded by the PamPro1 gene is shown in SEQ ID NO: 37. At the N-terminus, the protein has a signal peptide with a length of 25 amino acids as predicted by SignalP version 4.0 (Nordahl Petersen et al. (2011) Nature Methods, 8:785-786). The presence of a signal sequence suggests that PamPro1 is a secreted enzyme. The propeptide region was predicted based on protein sequence alignment with the *Paenibacillus polymyxa* Npr protein (Takekawa et al. (1991) Journal of Bacteriology, 173 (21): 6820-6825). The predicted mature region of PamPro1 protein is shown in SEQ ID NO: 3.

**[0452]** The nucleotide sequence of the PamPro1 gene isolated from *Paenibacillus amylolyticus* is set forth as SEQ ID NO: 36. The sequence encoding the predicted native signal peptide is shown in italics:

ATGAAATTCGCCAAAGTTATGCCAACAAATCTGGAGGAGCTCTTTTGCT  
 CGCTCCGATATCTCTGCTACTGCAGCTCCAGTGCTGATCAATCCATTC  
 CACTTCAGGCCCTTATGCCTCTGAGGGGGTATTCCATTGAACAGTGG  
 ACAGATGACACTATCTTTAATATCTTTGGACAGCAGGAACAATTTCTGAA  
 TTCCGATGTGAAATCCCAGCTCAAATGTCAAAGAAACACAGATACAT  
 CTGGCGTAAGACACTTCCCGCTGAAACAGTATATTAAGGTATCCCGGTT  
 TATGGTGCAGAACAGACGGTCCACCTGGACAAAACCGGAGCCGTGAGCTC  
 CGCACTTGGCGATCTTCCACCGATTGAAGAGCAGGCCATTCCGAATGATG  
 GTGTAGCCGAGATCAGCGGAGAAGACGCGATCCAGATTGCAACCGAAGAA  
 GCAACCTCCCGGATTGGAGAGCTTGGTGCCGCGAAATCACGCCTCAAGC  
 TGAATTGAACATCTATCATCATGAAGAAGATGGTCAGACATATCTGGTTT  
 ACATTACGGAAGTAAACGTAAGTGAACCTGCCCTCTACGGACCAATAT  
 TTCATTAACGCAGTGGATGGCAGTATCGTATCCAGTTTGACCTCATTAA  
 CTTGCTACTGGAACAGGTACAGGTGTAAGTACTCGGTGATACAAAACCTGA  
 CAACCAACCAATCCGGCAGCACCTTCCAACGAAAGACACCACTCGTGGC  
 AATGGCATCCAAACGTATACGGCAACCAATGGCTCCTCACCTGCCTGGTAG  
 CTTGCTTACAGATTCCGATAATGTATGAGCCGATCGTGCAGGTGTAGATG  
 CTCATGCTCATGCCGCTGCTACGTATGATTCTACAAAACAAATTC AAC  
 CGTAACGGTATTAATGGTAAACGGATTGTTGATCAGATCAACCGTGACTA  
 CGGCTCAATTACAATAACGCCTTCTGGAACGGGCACAGATTGTCTTTG  
 GTGACGGAGATGGAACGATGTTCCGATCCCCTGTCTGGTGATCTGGATGT  
 GTGGGTCATGAATTGACGCATGGTGTATTGAATATACAGCCAATCTGGA  
 ATATCGCAATGAACAGGTGCCTCAATGAAGCCTTTGCCGATATTTTCG  
 GTAATACGATCCAAAGCAAAACTGGCTGCTCGGTGATGATATCTACACA  
 CCTAACACTCCAGGAGATGCGCTGCGCTCCCTCTCCAACCTACATTGTA  
 TGGTCAACCTGACAAATACAGCGATCGCTACACAGGCTCACAGGACAACG  
 GCGGTGTCCATATCAACAGTGGTATCATCAATAAAGCCTATTTCTTGTCT  
 GCTCAAGGCGGAACACATAATGGTGTGACTGTTACCGGAATCGGCCGGA  
 TAAAGCATCCAGATTTTCTACAGCACACTGGTGAACCTACCTGACACCAA  
 CGTCCAAATTTGCCGCTGCCAAAACAGCTACCATTCAAGCAGCCAAAGAT  
 CTGTACGGAGCAACTCCCGCTGAAGCTACTGCTATTACAAAGCATATCA  
 AGCTGTAGGCCTG

**[0453]** The amino acid sequence of the PamPro1 precursor protein is set forth as SEQ ID NO: 37. The predicted signal sequence is shown in italics, and the predicted propeptide is shown in underlined text:

MKFAKVMPTILGGALLLASVSSATAAPVSDQSIPLQAPYASEGGIPLNSG  
TTDDTIFNYLGQEQFLNSDVKSQQLKIVKRNDDTSGVRHFRKQYIKGIPV  
YGAEQTVHLDKGTAVSSALGLDLPPIEEQAI PNDGVAEISGEDAIQIATEE

- continued

ATSRIGELGAAEITPQAELENIYHHEEDGQTYLVYITEVNVLEPAPLRTKY  
FINAVDGSIVSQFDLINEFATGTGTGVLGDTKLTTLTTSQSGSTFQLKDTTRG  
 NGIQTYTANNGSSLPGSLTDSDNVWTDRAQVDAHAHAATYDFYKKNFN  
 RINGINGNLLIRSTVHYGSNYNNAFWNGAQIVFGDGDGTMFRSLSGDLDV  
 VGHELTHGVI EYTANLEYRNEP GALNEAFADI FGNTIQSKNWL LGDDIYT  
 PNTPGDALRSLSNPTLYGQPKYSDRYTGSQDNGGVHINSIINKAYFLA  
 AQGGTHNGVTVTGIGRDKAIQIFYSTLVNYLTPTSKFAAAKTATIQAAD  
 LYGATSAEATAITKAYQAVGL

**[0454]** The amino acid sequence of the predicted mature form of PamPro1 is set forth as SEQ ID NO: 38:

ATGTGTGVLGDTKLTTLTTSQSGSTFQLKDTTRNGIQTYTANNGSSLPGSL  
 LTDSDNVWTDRAQVDAHAHAATYDFYKKNFN RINGINGNLLIRSTVHYG  
 SNYNNAFWNGAQIVFGDGDGTMFRSLSGDL DVVGHELTHGVI EYTANLEY  
 RNEP GALNEAFADI FGNTIQSKNWL LGDDIYT PNTPGDALRSLSNPTLYG  
 QPKYSDRYTGSQDNGGVHINSIINKAYFLA AQGGTHNGVTVTGIGRDK  
 AIQIFYSTLVNYLTPTSKFAAAKTATIQAADLYGATSAEATAITKAYQA  
 VGL

### Example 8.2

#### Expression of *Paenibacillus amylolyticus* Metalloprotease PamPro1

**[0455]** The DNA sequence of the propeptide-mature form of PamPro1 was synthesized and inserted into the *Bacillus subtilis* expression vector p2JM103BBI (Vogtentanz, Protein Expr Purif, 55:40-52, 2007) by Generay (Shanghai, China), resulting in plasmid pGX146(AprE-PamPro1) (FIG. 1). Ligation of this gene encoding the PamPro1 protein into the digested vector resulted in the addition of three codons (Ala-Gly-Lys) between the 3' end of the *B. subtilis* AprE signal sequence and the 5' end of the predicted PamPro1 native propeptide. The gene has an alternative start codon (GTG). The resulting plasmid shown in FIG. 8.1, labeled pGX146 (AprE-PamPro1) contains an AprE promoter, an AprE signal sequence used to direct target protein secretion in *B. subtilis*, and the synthetic nucleotide sequence encoding the predicted propeptide and mature regions of PamPro1 (SEQ ID NO: 39). The translation product of the synthetic AprE-PamPro1 gene is shown in SEQ ID NO: 40.

**[0456]** The pGX146(AprE-PamPro1) plasmid was then transformed into *B. subtilis* cells (degU<sup>Hy32</sup>, ΔscoC) and the transformed cells were spread on Luria Agar plates supplemented with 5 ppm Chloramphenicol and 1.2% skim milk (Cat#232100, Difco). Colonies with the largest clear halos on the plates were selected and subjected to fermentation in a 250 ml shake flask with MBD medium (a MOPS based defined medium, supplemented with additional 5 mM CaCl<sub>2</sub>).

**[0457]** The broth from the shake flasks was concentrated and buffer-exchanged into the loading buffer containing 20 mM Tris-HCl (pH 8.5), 1 mM CaCl<sub>2</sub> and 10% propylene glycol using a VivaFlow 200 ultra filtration device (Sartorius

Stedim). After filtering, this sample was applied to an 80 ml Q Sepharose High Performance column pre-equilibrated with the loading buffer above; and the active flow-through fractions were collected and concentrated. The sample was loaded onto a 320 ml Superdex 75 gel filtration column pre-equilibrated with the loading buffer described above containing 0.15 M NaCl. The corresponding active purified protein fractions were further pooled and concentrated via 10K Amicon Ultra for further analyses.

[0458] The nucleotide sequence of the synthesized PamPro1 gene in plasmid pGX146(AprE-PamPro1) is depicted in SEQ ID NO: 39. The sequence encoding the three residue addition (AGK) is shown in bold:

GTGAGAAGCAAAAAATTGTGGATCAGCTTGTGTTGCGTTAACGTTAAT  
 CTTTACGATGGCGTTTCAGCAACATGAGCGCGCAGGCT**GCTGGAAA**AGCTC  
 CGGTTAGCGACCAGTCAATCCCTCTTCAAGCACCGTATGCCAGCGAAGGA  
 GGCATTCCGCTTAACAGCGCACGAGCACACGATTTTCAATTACCTGGG  
 CCAACAGGAGCAGTTCCTGAACAGCGACGTCAAGAGCCAGCTGAAGATCG  
 TCAAAAGAAACACAGACACATCAGGCGTGAGACACTTCAGACTGAAGCAA  
 TACATCAAGGCATCCCGGTTTATGGCGCTGAACAAACGGTTCACCTGGA  
 CAAAACAGGCGCAGTTTTCATCAGCACTGGGAGATCTGCCGCCGATTGAAG  
 AGCAAGCAATCCCGAATGATGGAGTTGCGGAAATAGCGGCGAGGATGCA  
 ATCCAATCGCGACGGAGGAGGCTACATCAAGAATTGGAGAACTTGGCGC  
 AGCGGAGATTACACCGCAGGCTGAACATCATTACCATGAGGAAG  
 ACGGCCAGACGTACTCGTTTACATTACGGAAGTGAACGTGCTGGAACCG  
 GCACCTCTGAGAACAAAGTACTTTATCAACCGGTTGACGGCAGCATCGT  
 CTCACAGTTCGACCTGATTAACCTTCGCCACGGGAACAGGAACGGGGCTTC  
 TTGGAGACACAAAGACGCTGACGACGACGAGTCAAGCAGCACATTCCAG  
 CTGAAGGACACAAAGAGGCAACGGCATCCAAACGTACACGGCGAACA  
 TGGATCATCACTGCCGGGCTCAGTGTGACGGATTAGATAACGTGTGGA  
 CGGATAGAGCTGGCGTTGACGCGCATGCTCAGCTGTGCGACGTACGAC  
 TTCTACAAGAACAAGTTCAACAGAAACGGCATTAAACGGAAATGGCCTGCT  
 GATCAGAAGCACGGTGCATTATGGCTCAAACCTACAACACGCTTTTGGGA  
 ACGGCGCACAGATCGTGTGCGGACGCGATGGCACAATGTTTAGAAGC  
 CTGTGAGGACCTGGATGTGGTGGGCGACGAACTGACGCACGGCGTGAT  
 CGAGTATACGGCGAACCTTGAATATAGAAACGAGCCGGGAGCACTGAATG  
 AGGCGTTCGCGGACATTTTCGGCAACACAATCCAGAGCAAAAACCTGGCTG  
 CTGGGCGACGATATCTATACACCGAACACACCGGGCGATGCACTGAGATC  
 ACTGTCAAATCCGACGCTGTATGGCCAACCGGATAAGTACTCAGACAGAT  
 ATACGGCGAGCAAGACAATGGCGCGTTCACATCAACTCAGGCATCATC  
 AACAAAGGCTTACTTCTTGGCGCCAAAGGAGGAACACATAACGGCGTTAC  
 AGTTACAGGCATGGCAGAGACAAGGCGATCCAGATCTTTTACAGCACGC  
 TGGTGAACCTGACACCTACGTCAAAGTTTGGCGCAGCGAAAACAGCA

- continued

ACAATTACGGCGCTAAAGACCTGTACGGAGCGACATCAGCCGAGGCCAC  
 AGCAATTACAAAAGCATATCAAGCAGTTGGCCCTTTAA

[0459] The amino acid sequence of the PamPro1 precursor protein expressed from plasmid pGX146(AprE-PamPro1) is depicted in SEQ ID NO: 40. The predicted signal sequence is shown in *italics*, the three residue addition (AGK) is shown in **bold**, and the predicted pro-peptide is shown in underlined text.

*MRSKKLWISLLFALTLIFTMAFSNMSAQAA**AGK**APVSDQSIPLQAPYASEG*  
GIPLNSGTDITFNILGQQEQFLNSDVKSQLKIVKRNTDTSQVGRHFRFKQ  
YIKGIPVYGAEQTVHLDKTVAVSSALGDLPPIEEQAIIPNDGVAEISGEDA  
IQIATEEATSRIIGELGAAEITPQAEELNIYHHEEDGQTYLVYITEVNVLEP  
APLRTKYFINAVDGSIVSQFDLINFATGTGTGVLGDTKTLTTTQSGSTFQ  
 LKDTTRNGIQTYTANNGSSLPGSLLTSDNVWTDRAQVDAHAHAATYD  
 FYKNFNRNGINGNLLIRSTVHYGSNYNNAFWNGAQIVFGDGDGTMFRS  
 LSGDLLDVVGHETHGVIEYTANLEYRNEPAGALNEAFADIFGNTIQSKNWL  
 LGDDIYTPNTPGDALRSLSNPTLYGQPKYSDRYTGSQDNGGVHINSGLI  
 NKAYFLAAQGGTHNGVTVTGIGRDKAIQIFYSTLVNLYLPTSKFAAAKTA  
 TTQAAKLDLYGATSAEATAITKAYQAVGL

### Example 8.3

#### Proteolytic Activity of Metalloprotease PamPro1

[0460] The proteolytic activity of purified metalloprotease PamPro1 was measured in 50 mM Tris (pH 7), using azo-casein (Cat#74H7165, Megazyme) as a substrate. Prior to the reaction, the enzyme was diluted with Milli-Q water (Millipore) to specific concentrations. The azo-casein was dissolved in 100 mM Tris buffer (pH 7) to a final concentration of 1.5% (w/v). To initiate the reaction, 50  $\mu$ l of the diluted enzyme (or Milli-Q H<sub>2</sub>O alone as the blank control) was added to the non-binding 96-well Microtiter Plate (96-MTP) (Corning Life Sciences, #3641) placed on ice, followed by the addition of 50  $\mu$ l of 1.5% azo-casein. After sealing the 96-MTP, the reaction was carried out in a Thermomixer (Eppendorf) at 40° C. and 650 rpm for 10 min. The reaction was terminated by adding 100  $\mu$ l of 5% Trichloroacetic Acid (TCA). Following equilibration (5 min at the room temperature) and subsequent centrifugation (2000 g for 10 min at 4° C.), 120  $\mu$ l supernatant was transferred to a new 96-MTP, and absorbance of the supernatant was measured at 440 nm ( $A_{440}$ ) using a SpectraMax 190. Net  $A_{440}$  was calculated by subtracting the  $A_{440}$  of the blank control from that of enzyme, and then plotted against different protein concentrations (from 1.25 ppm to 40 ppm). Each value was the mean of triplicate assays. The proteolytic activity is shown as Net  $A_{440}$ . The proteolytic assay with azo-casein as the substrate (shown in FIG. 8.2) indicates that PamPro1 is an active protease.

### Example 8.4

#### pH Profiles of Metalloprotease PamPro1

[0461] With azo-casein as the substrate, the pH profiles of metalloprotease PamPro1 were studied in 12.5 mM acetate/



Bis-Tris/HEPES/CHES buffer with different pH values (ranging from pH 4 to 11). To initiate the assay, 50  $\mu$ l of 25 mM acetate/Bis-Tris/HEPES/CHES buffer with a specific pH was first mixed with 2  $\mu$ l Milli-Q H<sub>2</sub>O diluted enzyme (125 ppm) in a 96-MTP placed on ice, followed by the addition of 48  $\mu$ l of 1.5% (w/v) azo-casein prepared in H<sub>2</sub>O. The reaction was performed and analyzed as described in Example 8.3. Enzyme activity at each pH was reported as the relative activity, where the activity at the optimal pH was set to be 100%. The pH values tested were 4, 5, 6, 7, 8, 9, 10 and 11. Each value was the mean of triplicate assays. As shown in FIG. 8.3, the optimal pH of PamPro1 is about 8, with greater than 70% of maximal activity retained between 7 and 9.5.

#### Example 8.5

##### Temperature Profile of Metalloprotease PamPro1

**[0462]** The temperature profile of metalloprotease PamPro1 was analyzed in 50 mM Tris buffer (pH 7) using the azo-casein assays. The enzyme sample and azo-casein substrate were prepared as in Example 8.3. Prior to the reaction, 50  $\mu$ l of 1.5% azo-casein and 45  $\mu$ l Milli-Q H<sub>2</sub>O were mixed in a 200  $\mu$ l PCR tube, which was then subsequently incubated in a Peltier Thermal Cycler (BioRad) at desired temperatures (i.e. 20–90° C.) for 5 min. After the incubation, 5  $\mu$ l of diluted enzyme (50 ppm) or H<sub>2</sub>O (the blank control) was added to the substrate mixture, and the reaction was carried out in the Peltier Thermal Cycle for 10 min at different temperatures. To terminate the reaction, each assay mixture was transferred to a 96-MTP containing 100  $\mu$ l of 5% TCA per well. Subsequent centrifugation and absorbance measurement were performed as described in Example 8.3. The activity was reported as the relative activity, where the activity at the optimal temperature was set to be 100%. The tested temperatures are 20, 30, 40, 50, 60, 70, 80, and 90° C. Each value was the mean of duplicate assays (the value varies no more than 5%). The data in FIG. 8.4 suggest that PamPro1 showed an optimal temperature at about 50° C., and retained greater than 70% of its maximum activity between 45 and 55° C.

#### Example 8.6

##### Cleaning Performance of Metalloprotease PamPro1

**[0463]** The cleaning performance of PamPro1 was tested using PA-S-38 (egg yolk, with pigment, aged by heating) microswatches (CFT-Vlaardingen, The Netherlands) at pH 6 and 8 using a model automatic dishwashing (ADW) detergent. Prior to the reaction, purified protease samples were diluted with a dilution solution containing 10 mM NaCl, 0.1 mM CaCl<sub>2</sub>, 0.005% TWEEN® 80 and 10% propylene glycol to the desired concentrations. The reactions were performed in AT detergent with 100 ppm water hardness (Ca<sup>2+</sup>:Mg<sup>2+</sup>=3:1) (detergent composition shown in Table 8.1). To initiate the reaction, 180  $\mu$ l of the AT detergent buffered at pH 6 or pH 8 was added to a 96-MTP placed with PA-S-38 microswatches, followed by the addition of 20  $\mu$ l of diluted enzymes (or the dilution solution as the blank control). The 96-MTP was sealed and incubated in an incubator/shaker for 30 min at 50° C. and 1150 rpm. After incubation, 100  $\mu$ l of wash liquid from each well was transferred to a new 96-MTP, and its absorbance was measured at 405 nm (referred here as the “Initial performance”) using a spectrophotometer. The remaining wash liquid in the 96-MTP was discarded and the microswatches were rinsed once with 200  $\mu$ l water. Following the

addition of 180  $\mu$ l of 0.1 M CAPS buffer (pH 10), the second incubation was carried out in the incubator/shaker at 50° C. and 1150 rpm for 10 min. One hundred microliters of the resulting wash liquid was transferred to a new 96-MTP, and its absorbance measured at 405 nm (referred here as the “Wash-off”). The sum of two absorbance measurements (“Initial performance” plus “Wash-off”) gives the “Total performance”, which measures the protease activity on the model stain; and Net A<sub>405</sub> was subsequently calculated by subtracting the A<sub>405</sub> of the “Total performance” of the blank control from that of the enzyme. Dose response in cleaning the PA-S-38 microswatches at pH 6 and pH 8 in AT dish detergent for PamPro1 is shown in FIGS. 5A and 5B.

TABLE 8.1

| Composition of AT dish detergent formula with bleach        |                       |
|---|-----------------------|
| Ingredient  | Concentration (mg/ml) |
| MGDA (methylglycinediacetic acid)                           | 0.143                 |
| Sodium citrate  | 1.86                  |
| Citric acid*  | varies                |
| PAP (peracid N,N-phthaloylaminoperoxyacetic acid)           | 0.057                 |
| Plurafac ® LF 18B (a non-ionic surfactant)                  | 0.029                 |
| Bismuthcitrate  | 0.006                 |
| Bayhibit ® S (Phosphonobutantricarboxylic acid sodium salt) | 0.006                 |
| Acusol™ 587 (a calcium polyphosphate inhibitor)             | 0.029                 |
| PEG 6000  | 0.043                 |
| PEG 1500  | 0.1                   |

\*The pH of the AT formula detergent is adjusted to the desired value (pH 6 or 8) by the addition of 0.9M citric acid.

#### Example 8.7

##### Comparison of PamPro1 to Other Proteases

###### A. Identification of Homologous Proteases

**[0464]** Homologs were identified by a BLAST search (Altschul et al., *Nucleic Acids Res*, 25:3389-402, 1997) against the NCBI non-redundant protein database and the Genome Quest Patent database with search parameters set to default values. The predicted mature protein amino acid sequence for PamPro1 (SEQ ID NO: 38) was used as the query sequence. Percent identity (PID) for both search sets is defined as the number of identical residues divided by the number of aligned residues in the pairwise alignment. Tables 8.2A and 8.2B provide a list of sequences with the percent identity to PamPro1. The length in Table 8.2 refers to the entire sequence length of the homologous proteases.

TABLE 8.2A

| List of sequences with percent identity to PamPro1 protein identified from the NCBI non-redundant protein database |                |  |        |
|--|----------------|--|--------|
| Accession #  | PID to PamPro1 | Organism                                   | Length |
| P23384   | 56             | <i>Bacillus caldolyticus</i>               | 544    |
| P00800   | 56             | <i>Bacillus thermoproteolyticus</i>        | 548    |
| ZP_08640523.1  | 57             | <i>Brevibacillus laterosporus</i>          | 564    |
|  |                | LMG 15441                                  |        |
| BAA06144.1   | 57             | <i>Lactobacillus</i> sp.                   | 566    |
| YP_003872180.1   | 58             | <i>Paenibacillus polymyxa</i> E681         | 587    |
| ZP_04149724.1  | 59             | <i>Bacillus pseudomycoloides</i> DSM 12442 | 566    |

TABLE 8.2A-continued

| List of sequences with percent identity to PamPro1 protein identified from the NCBI non-redundant protein database |         |   |        |
|--|---------|---|--------|
| Accession #  | PID to  |   | Length |
|  | PamPro1 | Organism  |        |
| EJR46541.1   | 60      | <i>Bacillus cereus</i> VD107                                | 566    |
| YP_001373863.1   | 60      | <i>Bacillus cytotoxicus</i> NVH 391-98                      | 565    |
| ZP_10738945.1  | 61      | <i>Brevibacillus</i> sp. CF112                              | 528    |
| YP_004646155.1   | 61      | <i>Paenibacillus mucilaginosus</i> KNP414                   | 525    |
| ZP_02326602.1  | 62      | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> BRL-230010 | 520    |
| P43263   | 63      | <i>Brevibacillus brevis</i>                                 | 527    |
| ZP_09775365.1  | 64      | <i>Paenibacillus</i> sp. Aloe-11                            | 580    |
| ZP_09077634.1  | 65      | <i>Paenibacillus elgii</i> B69                              | 529    |
| ZP_09071078.1  | 68      | <i>Paenibacillus larvae</i> subsp. <i>larvae</i> B-3650     | 529    |
| ZP_08511445.1  | 69      | <i>Paenibacillus</i> sp. HGF7                               | 525    |
| YP_005073223.1   | 70      | <i>Paenibacillus terrae</i> HPL-003                         | 591    |
| YP_003948511.1   | 71      | <i>Paenibacillus polymyxa</i> SC2                           | 592    |
| ZP_10241030.1  | 71      | <i>Paenibacillus peoriae</i> KCTC 3763                      | 593    |

TABLE 8.2B

| List of sequences with percent identity to PamPro1 protein identified from the Genome Quest Patent database |         |                                       |        |
|---|---------|---------------------------------------|--------|
| Patent #  | PID to  |                                       | Length |
|   | PamPro1 | Organism                              |        |
| US7335504-0030  | 56.63   | <i>Bacillus thermoproteolyticus</i>   | 316    |
| US20120107907-0184  | 56.91   | <i>Bacillus caldoyticus</i>           | 319    |
| JP2006124323-0003   | 56.96   | <i>Bacillus thermoproteolyticus</i>   | 316    |
| JP1993199872-0001   | 56.96   | <i>Bacillus</i> sp.                   | 316    |
| JP1997000255-0001   | 56.96   | empty                                 | 548    |
| US6518054-0001  | 57.23   | <i>Bacillus</i> sp.                   | 319    |
| US20120107907-0176  | 57.23   | <i>Bacillus stearothermophilus</i>    | 548    |
| US8114656-0183  | 57.28   | <i>Bacillus stearothermophilus</i>    | 316    |
| US20120009651-0002  | 57.28   | <i>Geobacillus caldoproteolyticus</i> | 548    |
| JP2011103791-0020   | 57.28   | <i>Geobacillus stearothermophilus</i> | 552    |
| WO2012110562-0006   | 57.88   | <i>Bacillus megaterium</i>            | 320    |
| EP2390321-0178  | 57.88   | <i>Bacillus thuringiensis</i>         | 566    |
| US6518054-0002  | 57.93   | <i>Bacillus</i> sp.                   | 316    |
| WO2012110562-0007   | 58.25   | <i>Bacillus cereus</i>                | 320    |
| JP1995184649-0001   | 58.52   | <i>Lactobacillus</i> sp.              | 566    |
| EP2178896-0184  | 58.52   | <i>Bacillus anthracis</i>             | 566    |
| EP2390321-0195  | 59.55   | <i>Bacillus cereus</i>                | 317    |
| WO2012110563-0005   | 59.87   | <i>Bacillus cereus</i>                | 320    |
| US20080293610-0186  | 63.25   | <i>Bacillus brevis</i>                | 304    |
| JP2005229807-0018   | 71.19   | <i>Paenibacillus polymyxa</i>         | 566    |
| US8114656-0187  | 71.43   | <i>Bacillus polymyxa</i>              | 302    |

### B. Alignment of Homologous Protease Sequences

[0465] The amino acid sequence of the predicted mature PamPro1 (SEQ ID NO: 38) was aligned with thermolysin (P00800, *Bacillus thermoproteolyticus*), and protease from *Paenibacillus peoriae* KCTC 3763 (YP\_005073223.1) using CLUSTALW software (Thompson et al., Nucleic Acids

Research, 22:4673-4680, 1994) with the default parameters. FIG. 8.6 shows the alignment of PamPro1 with these protease sequences.

### C. Phylogenetic Tree

[0466] A phylogenetic tree for full length sequences of PamPro1 (SEQ ID NO: 37) was built using sequences of representative homologs from Table 8.2A and the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. Mol Biol. Evol. 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylogenetic tree printer software (<http://iubio.bio.indiana.edu/treeapp/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 8.7.

#### Example 9

#### Comparison of the Various *Paenibacillus* Metalloproteases with Other Bacterial Metalloprotease Homologs

### A. Alignment of Homologous Protease Sequences

[0467] The amino acid sequence of the predicted mature sequences for the *Paenibacillus* proteases described in Examples 1.1 to 8.7 were aligned with related bacterial metalloproteases using CLUSTALW software (Thompson et al., Nucleic Acids Research, 22:4673-4680, 1994) with the default parameters. FIG. 9.1 shows the alignment of the various *Paenibacillus* metalloproteases with other bacterial metalloprotease homologs.

### B. Phylogenetic Tree

[0468] A phylogenetic tree for full length sequences of the metalloproteases aligned in FIG. 9.1 was created using the Neighbor Joining method (NJ) (Saitou, N.; and Nei, M. (1987). The neighbor-joining method: a new method for reconstructing Guide Trees. Mol Biol. Evol. 4, 406-425). The NJ method works on a matrix of distances between all pairs of sequences to be analyzed. These distances are related to the degree of divergence between the sequences. The phylogenetic tree printer software (<http://iubio.bio.indiana.edu/treeapp/treeprint-form.html>) was used to display the phylogenetic tree shown in FIG. 9.2, where one can observe the clustering of the sequences from *Paenibacillus* genus.

[0469] While preferred embodiments of the present invention have been shown and described herein, it will be obvious to those skilled in the art that such embodiments are provided by way of example only. Numerous variations, changes, and substitutions will now occur to those skilled in the art without departing from the invention. It should be understood that various alternatives to the embodiments of the invention described herein can be employed in practicing the invention. It is intended that the following claims define the scope of the invention and that methods and structures within the scope of these claims and their equivalents be covered thereby.

## SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 68

<210> SEQ ID NO 1

<211> LENGTH: 1785

<212> TYPE: DNA

<213> ORGANISM: Paenibacillus sp.

<220> FEATURE:

<221> NAME/KEY: misc\_feature

<222> LOCATION: (1)..(1785)

<223> OTHER INFORMATION: nucleotide sequence of the PspPro3 gene isolated from Paenibacillus sp.

<400> SEQUENCE: 1

```

atgttaatga aaaaagtatg ggtttogctt cttggaggag cgatgttatt agggctctgta    60
gcgtctggty catcagcagc ggagagtcc  gtttcggggc  cggctcagct tacgccaacc    120
ttccatgccg aacaatggaa agcaccttca tcggtatcgg gtgatgacat cgtatggagc    180
tatttaaatc ggcaaaagaa aacgttgctg ggtacggaca gcaccagtgt ccgatgatcaa    240
ttcogtatcg tagatcgcac aagcgacaaa tccggcgtga gccattatcg gctgaagcaa    300
tatgtaaacy gaattcccgt atatggagct gaacagacca ttcattgtgg caaatccggt    360
gaagtgcact cttatctggg agccgtgatt actgaggatc agcaagaaga agctacgcaa    420
ggtacaactc cgaaaatcag cgcttctgaa gcggtccata ccgcatatca ggaggcagct    480
acacgggttc aagccctccc tacctccgat gatacgattt ctaaagatgc ggaggagcca    540
agcagtgtaa gcaaagacac ttactccgaa gcagctaaca acggaaaaac gagttctgtt    600
gaaaaggaca agctcagcct tgagaaagcg gctgacctga aagatagcaa aattgaagcg    660
gtggaggcag agccaaaact cattgocaaa atcgccaacc tgcagcctga ggttagatcct    720
aaagccgaac tatatttcta tgcaagggc gatgcattgc agctggttta tgtgactgag    780
gttaatatatt tcagcctgc gccctgctg acacgctaca tcattgacgc caatgatggc    840
aaaaatcgat cccagtatga catcattaat gaagcgacag gcacaggcaa aggtgtactc    900
ggtgatacca aacattcaa cactactgct tccggcagca gctaccagtt aagagatagc    960
actcgcggga atggaatcgt gacttacagc gcctccaacc gtcaaagcat cccaggtacg   1020
atcctgaccg atgccgataa cgtatggaat gatccagccg gcgtggatgc ccacgcttat   1080
gcagcAAAA cctatgatta ttataaggaa aagttcaatc gcaacagcat tgacggacga   1140
ggcctgcagc tccgttcgac agttcattac ggcaatcgtt acaacaacgc cttctggaac   1200
ggctccaaa tgacttatgg agacggagac ggcaaccatc ttatcgcttt tagcggtgat   1260
ccgatgtag ttggtcatga actcaacac ggtgttacgg agtatacttc caatttgaa   1320
tattacggag aatccggtyc gttgaacgag gccttctcgg acatcatcgg caatgacatc   1380
cagcgtaaaa actggcttgt aggcgatgat atttacacgc cacgcattgc gggatgatgca   1440
cttcgttcta tgtccaatcc tacgctgtac gatcaaccgg atcactattc gaacttgatc   1500
agaggcagct ccgataacgg cggcgttcat acgaacagcg gtattataaa taaagcctat   1560
tatctgttgg cacaaggcgg caccttccat ggtgtaactg tcaatgggat tggcccgcat   1620
gcagcgggtc aaatttacta cagcgccttt acgaactacc tgacttcttc ttctgacttc   1680
tccaatgcac gtgatgccgt tgtacaagcg gcaaaagatc tctacggcgc gagctcggca   1740
caagctaccg cagcagccaa atcttttgat gctgtaggcg ttaac   1785

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<210> SEQ ID NO 2  
 <211> LENGTH: 595  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus sp.  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(595)  
 <223> OTHER INFORMATION: amino acid sequence of the PspPro3 precursor protein

<400> SEQUENCE: 2

Met Leu Met Lys Lys Val Trp Val Ser Leu Leu Gly Gly Ala Met Leu  
 1 5 10 15  
 Leu Gly Ser Val Ala Ser Gly Ala Ser Ala Ala Glu Ser Ser Val Ser  
 20 25 30  
 Gly Pro Ala Gln Leu Thr Pro Thr Phe His Ala Glu Gln Trp Lys Ala  
 35 40 45  
 Pro Ser Ser Val Ser Gly Asp Asp Ile Val Trp Ser Tyr Leu Asn Arg  
 50 55 60  
 Gln Lys Lys Thr Leu Leu Gly Thr Asp Ser Thr Ser Val Arg Asp Gln  
 65 70 75 80  
 Phe Arg Ile Val Asp Arg Thr Ser Asp Lys Ser Gly Val Ser His Tyr  
 85 90 95  
 Arg Leu Lys Gln Tyr Val Asn Gly Ile Pro Val Tyr Gly Ala Glu Gln  
 100 105 110  
 Thr Ile His Val Gly Lys Ser Gly Glu Val Thr Ser Tyr Leu Gly Ala  
 115 120 125  
 Val Ile Thr Glu Asp Gln Gln Glu Glu Ala Thr Gln Gly Thr Thr Pro  
 130 135 140  
 Lys Ile Ser Ala Ser Glu Ala Val His Thr Ala Tyr Gln Glu Ala Ala  
 145 150 155 160  
 Thr Arg Val Gln Ala Leu Pro Thr Ser Asp Asp Thr Ile Ser Lys Asp  
 165 170 175  
 Ala Glu Glu Pro Ser Ser Val Ser Lys Asp Thr Tyr Ser Glu Ala Ala  
 180 185 190  
 Asn Asn Gly Lys Thr Ser Ser Val Glu Lys Asp Lys Leu Ser Leu Glu  
 195 200 205  
 Lys Ala Ala Asp Leu Lys Asp Ser Lys Ile Glu Ala Val Glu Ala Glu  
 210 215 220  
 Pro Asn Ser Ile Ala Lys Ile Ala Asn Leu Gln Pro Glu Val Asp Pro  
 225 230 235 240  
 Lys Ala Glu Leu Tyr Phe Tyr Ala Lys Gly Asp Ala Leu Gln Leu Val  
 245 250 255  
 Tyr Val Thr Glu Val Asn Ile Leu Gln Pro Ala Pro Leu Arg Thr Arg  
 260 265 270  
 Tyr Ile Ile Asp Ala Asn Asp Gly Lys Ile Val Ser Gln Tyr Asp Ile  
 275 280 285  
 Ile Asn Glu Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys  
 290 295 300  
 Thr Phe Asn Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Arg Asp Thr  
 305 310 315 320  
 Thr Arg Gly Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser  
 325 330 335  
 Ile Pro Gly Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro



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| 65   | 70 | 75 | 80 |
|--|----|----|----|
| Phe Asn Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr<br>85 90 95        |    |    |    |
| Val His Tyr Gly Asn Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln<br>100 105 110     |    |    |    |
| Met Thr Tyr Gly Asp Gly Asp Gly Thr Thr Phe Ile Ala Phe Ser Gly<br>115 120 125     |    |    |    |
| Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr<br>130 135 140     |    |    |    |
| Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala<br>145 150 155 160 |    |    |    |
| Phe Ser Asp Ile Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val<br>165 170 175     |    |    |    |
| Gly Asp Asp Ile Tyr Thr Pro Arg Ile Ala Gly Asp Ala Leu Arg Ser<br>180 185 190     |    |    |    |
| Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu<br>195 200 205     |    |    |    |
| Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile<br>210 215 220     |    |    |    |
| Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe His Gly<br>225 230 235 240 |    |    |    |
| Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr<br>245 250 255     |    |    |    |
| Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser Asn Ala<br>260 265 270     |    |    |    |
| Arg Asp Ala Val Val Gln Ala Ala Lys Asp Leu Tyr Gly Ala Ser Ser<br>275 280 285     |    |    |    |
| Ala Gln Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn<br>290 295 300     |    |    |    |

&lt;210&gt; SEQ ID NO 4

&lt;211&gt; LENGTH: 1803

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic; nucleotide sequence of the synthesized PspPro3 gene in plasmid pGX085(AprE- PspPro3)

&lt;400&gt; SEQUENCE: 4

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg    60
gcgttcagca acatgagcgc gcaggctgct ggaaaagcag aatcatcagt gtcaggaccg    120
gctcagctta cgccgacggt tcatgcagag cagtggaaag caccgagcag cgttagcgga    180
gatgacatcg tgtggagcta cctgaacaga cagaagaaaa cgcttcttgg cacggacagc    240
acgagcgtca gagaccagtt cagaatcgtg gatagaacaa gcgacaaaag cggcgtcagc    300
cattatagac tgaagcagta tgtgaacgga atcccggttt atggcgcaga acaacaatc    360
catgtcggaa agagcggcga agttacgagc tatctgggcy cggttattac agaggaccag    420
caagaggagg ctacacaagg cacgacaccg aaaatttcag catcagaggc agttcatacg    480
gcctaccaag aagctgcaac gagagttcaa gccctgccta cgtcagatga tacaatcagc    540
aaagacgctg aggaacctag ctcagttagc aaggacacgt atagcgaagc cgccaacaat    600
ggcaagacgt caagcgtgga aaaagacaag ctttcactgg agaaggccgc tgatctgaaa    660

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gactcaaaga tcgaggctgt ggaagcggaa cgaatagca ttgcaaagat tgccaacctg 720
caaccggagg tggaccggaa ggcggagctg tatttctacg ctaaaggcga tgcactgcaa 780
ctggtttacg tcacggaggt taacatcctg cagccggcac cgcttagaac gagatacatc 840
attgacgcga acgacggcaa gatcgtgagc cagtacgaca ttatcaacga ggccacggga 900
acgggcaagg gagtcccttg cgacacgaag acattcaata caacggcctc aggctcatca 960
taccagctga gagacacgac gagaggcaac ggaatcgtca cgtacacggc tagcaataga 1020
cagagcattc cgggcacaat ccttacggac gcagacaatg tgtggaatga cccggcaggc 1080
gtggacgcac atgcctacgc agcgaagacg tacgactact acaaggagaa gttcaacaga 1140
aacagcatcg acggaagagg actgcaactt agaagcacgg tgcattacgg caacagatac 1200
aacaacgctt tctggaacgg cagccaaatg acgtatggag acggcgatgg aacaacgttt 1260
atcgattctc caggcgaccg tgacgttggt ggacatgaac tgacgcatgg agtcacagaa 1320
tacacgagca atctggagta ttacggagaa tcaggcgcac ttaatgaggc cttcagcgac 1380
atcatcggaa acgacatcca gagaaagaac tggctggttg gcgatgatat ctacacgccg 1440
agaattgctg gcgacgcgct gagatcaatg agcaacccta cgctgtacga tcagccggat 1500
cattacagca acctgtatag aggctcaagc gataatggcg gcgtgcatac aaacagcggc 1560
atcatcaaca aagcctatta tctgtggcg caaggcggca cattccatgg cgttacagtt 1620
aatggcattg gcagagacgc agccgtgcag atctactaca gcgcattcac gaattactg 1680
acatcaagca gcgacttttc aatgcaaga gatgcagtgg tgcaggcggc taaagacctt 1740
tatggagctt caagcgctca ggccacagct gcggcaaaaa gcttcgacgc ggttgagtg 1800
aat 1803

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&lt;210&gt; SEQ ID NO 5

&lt;211&gt; LENGTH: 601

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Synthetic; amino acid sequence of the PspPro3 precursor protein expressed from plasmid pGX085(AprE- PspPro3)

&lt;400&gt; SEQUENCE: 5

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10           15
Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
                20           25           30
Ala Glu Ser Ser Val Ser Gly Pro Ala Gln Leu Thr Pro Thr Phe His
                35           40           45
Ala Glu Gln Trp Lys Ala Pro Ser Ser Val Ser Gly Asp Asp Ile Val
                50           55           60
Trp Ser Tyr Leu Asn Arg Gln Lys Lys Thr Leu Leu Gly Thr Asp Ser
65           70           75           80
Thr Ser Val Arg Asp Gln Phe Arg Ile Val Asp Arg Thr Ser Asp Lys
                85           90           95
Ser Gly Val Ser His Tyr Arg Leu Lys Gln Tyr Val Asn Gly Ile Pro
                100          105          110
Val Tyr Gly Ala Glu Gln Thr Ile His Val Gly Lys Ser Gly Glu Val
115          120          125

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|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Thr | Ser | Tyr | Leu | Gly | Ala | Val | Ile | Thr | Glu | Asp | Gln | Gln | Glu | Glu | Ala | 130 | 135 | 140 |     |
| Thr | Gln | Gly | Thr | Thr | Pro | Lys | Ile | Ser | Ala | Ser | Glu | Ala | Val | His | Thr | 145 | 150 | 155 | 160 |
| Ala | Tyr | Gln | Glu | Ala | Ala | Thr | Arg | Val | Gln | Ala | Leu | Pro | Thr | Ser | Asp | 165 | 170 | 175 |     |
| Asp | Thr | Ile | Ser | Lys | Asp | Ala | Glu | Glu | Pro | Ser | Ser | Val | Ser | Lys | Asp | 180 | 185 | 190 |     |
| Thr | Tyr | Ser | Glu | Ala | Ala | Asn | Asn | Gly | Lys | Thr | Ser | Ser | Val | Glu | Lys | 195 | 200 | 205 |     |
| Asp | Lys | Leu | Ser | Leu | Glu | Lys | Ala | Ala | Asp | Leu | Lys | Asp | Ser | Lys | Ile | 210 | 215 | 220 |     |
| Glu | Ala | Val | Glu | Ala | Glu | Pro | Asn | Ser | Ile | Ala | Lys | Ile | Ala | Asn | Leu | 225 | 230 | 235 | 240 |
| Gln | Pro | Glu | Val | Asp | Pro | Lys | Ala | Glu | Leu | Tyr | Phe | Tyr | Ala | Lys | Gly | 245 | 250 | 255 |     |
| Asp | Ala | Leu | Gln | Leu | Val | Tyr | Val | Thr | Glu | Val | Asn | Ile | Leu | Gln | Pro | 260 | 265 | 270 |     |
| Ala | Pro | Leu | Arg | Thr | Arg | Tyr | Ile | Ile | Asp | Ala | Asn | Asp | Gly | Lys | Ile | 275 | 280 | 285 |     |
| Val | Ser | Gln | Tyr | Asp | Ile | Ile | Asn | Glu | Ala | Thr | Gly | Thr | Gly | Lys | Gly | 290 | 295 | 300 |     |
| Val | Leu | Gly | Asp | Thr | Lys | Thr | Phe | Asn | Thr | Thr | Ala | Ser | Gly | Ser | Ser | 305 | 310 | 315 | 320 |
| Tyr | Gln | Leu | Arg | Asp | Thr | Thr | Arg | Gly | Asn | Gly | Ile | Val | Thr | Tyr | Thr | 325 | 330 | 335 |     |
| Ala | Ser | Asn | Arg | Gln | Ser | Ile | Pro | Gly | Thr | Ile | Leu | Thr | Asp | Ala | Asp | 340 | 345 | 350 |     |
| Asn | Val | Trp | Asn | Asp | Pro | Ala | Gly | Val | Asp | Ala | His | Ala | Tyr | Ala | Ala | 355 | 360 | 365 |     |
| Lys | Thr | Tyr | Asp | Tyr | Tyr | Lys | Glu | Lys | Phe | Asn | Arg | Asn | Ser | Ile | Asp | 370 | 375 | 380 |     |
| Gly | Arg | Gly | Leu | Gln | Leu | Arg | Ser | Thr | Val | His | Tyr | Gly | Asn | Arg | Tyr | 385 | 390 | 395 | 400 |
| Asn | Asn | Ala | Phe | Trp | Asn | Gly | Ser | Gln | Met | Thr | Tyr | Gly | Asp | Gly | Asp | 405 | 410 | 415 |     |
| Gly | Thr | Thr | Phe | Ile | Ala | Phe | Ser | Gly | Asp | Pro | Asp | Val | Val | Gly | His | 420 | 425 | 430 |     |
| Glu | Leu | Thr | His | Gly | Val | Thr | Glu | Tyr | Thr | Ser | Asn | Leu | Glu | Tyr | Tyr | 435 | 440 | 445 |     |
| Gly | Glu | Ser | Gly | Ala | Leu | Asn | Glu | Ala | Phe | Ser | Asp | Ile | Ile | Gly | Asn | 450 | 455 | 460 |     |
| Asp | Ile | Gln | Arg | Lys | Asn | Trp | Leu | Val | Gly | Asp | Asp | Ile | Tyr | Thr | Pro | 465 | 470 | 475 | 480 |
| Arg | Ile | Ala | Gly | Asp | Ala | Leu | Arg | Ser | Met | Ser | Asn | Pro | Thr | Leu | Tyr | 485 | 490 | 495 |     |
| Asp | Gln | Pro | Asp | His | Tyr | Ser | Asn | Leu | Tyr | Arg | Gly | Ser | Ser | Asp | Asn | 500 | 505 | 510 |     |
| Gly | Gly | Val | His | Thr | Asn | Ser | Gly | Ile | Ile | Asn | Lys | Ala | Tyr | Tyr | Leu | 515 | 520 | 525 |     |
| Leu | Ala | Gln | Gly | Gly | Thr | Phe | His | Gly | Val | Thr | Val | Asn | Gly | Ile | Gly |     |     |     |     |



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|   |   |     |     |
|---|---|-----|-----|
| 530   | 535   | 540 |     |
| Arg Asp Ala Ala Val   | Gln Ile Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu |     |     |
| 545   | 550   | 555 | 560 |
| Thr Ser Ser Ser Asp Phe Ser Asn Ala Arg Asp Ala Val Val Gln Ala |   |     |     |
|   | 565   | 570 | 575 |
| Ala Lys Asp Leu Tyr Gly Ala Ser Ser Ala Gln Ala Thr Ala Ala Ala |   |     |     |
|   | 580   | 585 | 590 |
| Lys Ser Phe Asp Ala Val Gly Val Asn                             |   |     |     |
|   | 595   | 600 |     |

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<210> SEQ ID NO 6
<211> LENGTH: 1770
<212> TYPE: DNA
<213> ORGANISM: Paenibacillus sp.
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1770)
<223> OTHER INFORMATION: nucleotide sequence of the PspPro2 gene
isolated from Paenibacillus sp.

<400> SEQUENCE: 6
atgaaaaaag tatgggtttc acttcttggg gagcgcgatgt tattaggggc tgtagcacca 60
ggtgcatcag cagcagagca ttctgttctc gatcctactc agctaacacc gacctttcac 120
gccgagcaat ggaaggtcc ttccacggta accggcgaca atattgtatg gagctatttg 180
aatcgacaaa agaaaacctt attgaatata gacagcacca gtgtgctga tcagttccgc 240
atcattgatc gtacaagcga caaatccggt gcaagccatt atcggctcaa gcaatatgta 300
aacgggatcc ccgtatatgg ggctgaacag accattcatg tgaacaacgc cggtaaagta 360
acctcttatt tgggtgctgt catttcagag gatcagcagc aagacgcgac cgaagatacc 420
actcaaaaa tcagcgcgac tgaagccgtt tataccgcat atgcagaagc cgctgccccg 480
attcaatcct tcccttccat caatgatagt ctttctgagg ctagtgagga acaagggagt 540
gagaatcaag gcaatgagat tcaaaacatt gggattaaaa gcagtgtgtaag taatgacact 600
tacgcagagg cgcataacaa cgtactttta acccccgttg accaagcaga gcaaagtac 660
attgcacaaa ttgctaactc ggagccaagt gtagagccca aagcagaatt atacatctat 720
ccagatggtg agactacacg actggtttat gtaacagagg ttaatattct tgaacctgcg 780
cctctgcgca cacgctactt cattgatgcg aaaaccggca aaatcgtatt ccagtatgac 840
atcctcaacc acgcaacagg caccggccgc ggcgtggatg gcaaaacaaa atcatttacg 900
actacagctt caggcaaccg gtatcagttg aaagacacga ctgcagcaa tggaaatcgtg 960
acttacaccg ctggcaatcg ccagacgacg ccaggtaaga ttttgaccga tacagataat 1020
gtatgggagg accctgcggc tgttgatgcc catgcctacg ccattaaaac ctatgactat 1080
tataagaata aattcggtcg cgacagtatt gatggacgtg gcatgcaaat tcgttcgaca 1140
gtccattacg gcaaaaaata taacaatgcc ttctggaacg gctcgcaaat gacctacgga 1200
gacggagacg ggtccacatt taccttcttc agcggcgatc ccgatgtcgt ggggcatgag 1260
ctcaccacag gcgtaaccga gttcacctcc aatttggagt attatggtga gtcgggtgca 1320
ttgaacgaag ccttctcgga tattatcggg aatgatatag atggcaccag ttggcttctt 1380
ggcgacggca tttatacgcc taatattcca ggcgacgctc tgcgttcctt gtcgatcct 1440
acacgattcg gccagccgga tcaactactc aatttctatc cggaccccaa caatgatgat 1500
    
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gaaggcggag tccatacgaa cagcgggtatt atcaacaaag cctattatatt gctggcacia 1560
ggcggtagct cccatggtgt aacggtaact ggtatcggac gcgaagcggc tgtattcatt 1620
tactacaatg cctttaccaa ctatttgacc tctacctcca acttctctaa cgcacgcgct 1680
gctgttatac aggcagccaa ggatttttat ggtgctgatt cgctggcagt aaccagtgt 1740
attcaatcct ttgatgcggt aggaatcaaa 1770

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<210> SEQ ID NO 7
<211> LENGTH: 590
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus sp.
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(590)
<223> OTHER INFORMATION: amino acid sequence of the PspPro2 precursor
protein

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<400> SEQUENCE: 7

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Met Lys Lys Val Trp Val Ser Leu Leu Gly Gly Ala Met Leu Leu Gly
1 5 10 15
Ala Val Ala Pro Gly Ala Ser Ala Ala Glu His Ser Val Pro Asp Pro
20 25 30
Thr Gln Leu Thr Pro Thr Phe His Ala Glu Gln Trp Lys Ala Pro Ser
35 40 45
Thr Val Thr Gly Asp Asn Ile Val Trp Ser Tyr Leu Asn Arg Gln Lys
50 55 60
Lys Thr Leu Leu Asn Thr Asp Ser Thr Ser Val Arg Asp Gln Phe Arg
65 70 75 80
Ile Ile Asp Arg Thr Ser Asp Lys Ser Gly Ala Ser His Tyr Arg Leu
85 90 95
Lys Gln Tyr Val Asn Gly Ile Pro Val Tyr Gly Ala Glu Gln Thr Ile
100 105 110
His Val Asn Asn Ala Gly Lys Val Thr Ser Tyr Leu Gly Ala Val Ile
115 120 125
Ser Glu Asp Gln Gln Gln Asp Ala Thr Glu Asp Thr Thr Pro Lys Ile
130 135 140
Ser Ala Thr Glu Ala Val Tyr Thr Ala Tyr Ala Glu Ala Ala Ala Arg
145 150 155 160
Ile Gln Ser Phe Pro Ser Ile Asn Asp Ser Leu Ser Glu Ala Ser Glu
165 170 175
Glu Gln Gly Ser Glu Asn Gln Gly Asn Glu Ile Gln Asn Ile Gly Ile
180 185 190
Lys Ser Ser Val Ser Asn Asp Thr Tyr Ala Glu Ala His Asn Asn Val
195 200 205
Leu Leu Thr Pro Val Asp Gln Ala Glu Gln Ser Tyr Ile Ala Lys Ile
210 215 220
Ala Asn Leu Glu Pro Ser Val Glu Pro Lys Ala Glu Leu Tyr Ile Tyr
225 230 235 240
Pro Asp Gly Glu Thr Thr Arg Leu Val Tyr Val Thr Glu Val Asn Ile
245 250 255
Leu Glu Pro Ala Pro Leu Arg Thr Arg Tyr Phe Ile Asp Ala Lys Thr
260 265 270
Gly Lys Ile Val Phe Gln Tyr Asp Ile Leu Asn His Ala Thr Gly Thr

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| 275 |     |     |     |     | 280 |     |     |     |     | 285 |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Gly | Arg | Gly | Val | Asp | Gly | Lys | Thr | Lys | Ser | Phe | Thr | Thr | Thr | Ala | Ser |
| 290 |     |     |     |     | 295 |     |     |     |     | 300 |     |     |     |     |     |
| Gly | Asn | Arg | Tyr | Gln | Leu | Lys | Asp | Thr | Thr | Arg | Ser | Asn | Gly | Ile | Val |
| 305 |     |     |     |     | 310 |     |     |     |     | 315 |     |     |     |     | 320 |
| Thr | Tyr | Thr | Ala | Gly | Asn | Arg | Gln | Thr | Thr | Pro | Gly | Thr | Ile | Leu | Thr |
|     |     |     |     | 325 |     |     |     |     | 330 |     |     |     |     | 335 |     |
| Asp | Thr | Asp | Asn | Val | Trp | Glu | Asp | Pro | Ala | Ala | Val | Asp | Ala | His | Ala |
|     |     |     | 340 |     |     |     |     | 345 |     |     |     |     | 350 |     |     |
| Tyr | Ala | Ile | Lys | Thr | Tyr | Asp | Tyr | Tyr | Lys | Asn | Lys | Phe | Gly | Arg | Asp |
|     |     | 355 |     |     |     | 360 |     |     |     |     |     | 365 |     |     |     |
| Ser | Ile | Asp | Gly | Arg | Gly | Met | Gln | Ile | Arg | Ser | Thr | Val | His | Tyr | Gly |
|     | 370 |     |     |     |     | 375 |     |     |     |     | 380 |     |     |     |     |
| Lys | Lys | Tyr | Asn | Asn | Ala | Phe | Trp | Asn | Gly | Ser | Gln | Met | Thr | Tyr | Gly |
| 385 |     |     |     |     | 390 |     |     |     |     | 395 |     |     |     |     | 400 |
| Asp | Gly | Asp | Gly | Ser | Thr | Phe | Thr | Phe | Phe | Ser | Gly | Asp | Pro | Asp | Val |
|     |     |     |     | 405 |     |     |     |     | 410 |     |     |     |     | 415 |     |
| Val | Gly | His | Glu | Leu | Thr | His | Gly | Val | Thr | Glu | Phe | Thr | Ser | Asn | Leu |
|     |     |     | 420 |     |     |     |     | 425 |     |     |     |     |     | 430 |     |
| Glu | Tyr | Tyr | Gly | Glu | Ser | Gly | Ala | Leu | Asn | Glu | Ala | Phe | Ser | Asp | Ile |
|     |     | 435 |     |     |     |     | 440 |     |     |     |     | 445 |     |     |     |
| Ile | Gly | Asn | Asp | Ile | Asp | Gly | Thr | Ser | Trp | Leu | Leu | Gly | Asp | Gly | Ile |
|     | 450 |     |     |     |     | 455 |     |     |     |     | 460 |     |     |     |     |
| Tyr | Thr | Pro | Asn | Ile | Pro | Gly | Asp | Ala | Leu | Arg | Ser | Leu | Ser | Asp | Pro |
| 465 |     |     |     |     | 470 |     |     |     |     | 475 |     |     |     |     | 480 |
| Thr | Arg | Phe | Gly | Gln | Pro | Asp | His | Tyr | Ser | Asn | Phe | Tyr | Pro | Asp | Pro |
|     |     |     |     | 485 |     |     |     |     | 490 |     |     |     |     | 495 |     |
| Asn | Asn | Asp | Asp | Glu | Gly | Gly | Val | His | Thr | Asn | Ser | Gly | Ile | Ile | Asn |
|     |     |     | 500 |     |     |     |     | 505 |     |     |     |     | 510 |     |     |
| Lys | Ala | Tyr | Tyr | Leu | Leu | Ala | Gln | Gly | Gly | Thr | Ser | His | Gly | Val | Thr |
|     |     | 515 |     |     |     |     | 520 |     |     |     |     | 525 |     |     |     |
| Val | Thr | Gly | Ile | Gly | Arg | Glu | Ala | Ala | Val | Phe | Ile | Tyr | Tyr | Asn | Ala |
|     |     | 530 |     |     |     | 535 |     |     |     |     | 540 |     |     |     |     |
| Phe | Thr | Asn | Tyr | Leu | Thr | Ser | Thr | Ser | Asn | Phe | Ser | Asn | Ala | Arg | Ala |
| 545 |     |     |     |     | 550 |     |     |     |     | 555 |     |     |     |     | 560 |
| Ala | Val | Ile | Gln | Ala | Ala | Lys | Asp | Phe | Tyr | Gly | Ala | Asp | Ser | Leu | Ala |
|     |     |     |     | 565 |     |     |     |     | 570 |     |     |     |     | 575 |     |
| Val | Thr | Ser | Ala | Ile | Gln | Ser | Phe | Asp | Ala | Val | Gly | Ile | Lys |     |     |
|     |     |     | 580 |     |     |     |     | 585 |     |     |     |     | 590 |     |     |

<210> SEQ ID NO 8  
 <211> LENGTH: 306  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus sp.  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(306)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature form of PspPro2

<400> SEQUENCE: 8

|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Ala | Thr | Gly | Thr | Gly | Arg | Gly | Val | Asp | Gly | Lys | Thr | Lys | Ser | Phe | Thr |
| 1   |     |     |     | 5   |     |     |     |     |     | 10  |     |     |     | 15  |     |
| Thr | Thr | Ala | Ser | Gly | Asn | Arg | Tyr | Gln | Leu | Lys | Asp | Thr | Thr | Arg | Ser |

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|   |     |  |    |     |    |     |
|---|-----|--|----|-----|----|-----|
|   | 20  |  | 25 |     | 30 |     |
| Asn Gly Ile Val Thr Tyr Thr Ala Gly Asn Arg Gln Thr Thr Pro Gly |     |  |    |     |    |     |
|   | 35  |  | 40 |     | 45 |     |
| Thr Ile Leu Thr Asp Thr Asp Asn Val Trp Glu Asp Pro Ala Ala Val |     |  | 55 |     | 60 |     |
|   | 50  |  |    |     |    |     |
| Asp Ala His Ala Tyr Ala Ile Lys Thr Tyr Asp Tyr Tyr Lys Asn Lys |     |  | 70 |     | 75 | 80  |
|   | 65  |  |    |     |    |     |
| Phe Gly Arg Asp Ser Ile Asp Gly Arg Gly Met Gln Ile Arg Ser Thr |     |  | 85 |     | 90 | 95  |
|   |     |  |    |     |    |     |
| Val His Tyr Gly Lys Lys Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln |     |  |    | 105 |    | 110 |
|   | 100 |  |    |     |    |     |
| Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Thr Phe Phe Ser Gly |     |  |    | 120 |    | 125 |
|   | 115 |  |    |     |    |     |
| Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Phe |     |  |    | 135 |    | 140 |
|   | 130 |  |    |     |    |     |
| Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala |     |  |    | 150 |    | 155 |
|   | 145 |  |    |     |    | 160 |
| Phe Ser Asp Ile Ile Gly Asn Asp Ile Asp Gly Thr Ser Trp Leu Leu |     |  |    | 165 |    | 170 |
|   |     |  |    |     |    | 175 |
| Gly Asp Gly Ile Tyr Thr Pro Asn Ile Pro Gly Asp Ala Leu Arg Ser |     |  |    | 180 |    | 185 |
|   |     |  |    |     |    | 190 |
| Leu Ser Asp Pro Thr Arg Phe Gly Gln Pro Asp His Tyr Ser Asn Phe |     |  |    | 195 |    | 200 |
|   |     |  |    |     |    | 205 |
| Tyr Pro Asp Pro Asn Asn Asp Asp Glu Gly Gly Val His Thr Asn Ser |     |  |    | 215 |    | 220 |
|   | 210 |  |    |     |    |     |
| Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Ser |     |  |    | 230 |    | 235 |
|   | 225 |  |    |     |    | 240 |
| His Gly Val Thr Val Thr Gly Ile Gly Arg Glu Ala Ala Val Phe Ile |     |  |    | 245 |    | 250 |
|   |     |  |    |     |    | 255 |
| Tyr Tyr Asn Ala Phe Thr Asn Tyr Leu Thr Ser Thr Ser Asn Phe Ser |     |  |    | 260 |    | 265 |
|   |     |  |    |     |    | 270 |
| Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Phe Tyr Gly Ala |     |  |    | 275 |    | 280 |
|   |     |  |    |     |    | 285 |
| Asp Ser Leu Ala Val Thr Ser Ala Ile Gln Ser Phe Asp Ala Val Gly |     |  |    | 290 |    | 295 |
|   |     |  |    |     |    | 300 |
| Ile Lys   |     |  |    |     |    |     |
| 305   |     |  |    |     |    |     |

<210> SEQ ID NO 9  
 <211> LENGTH: 1794  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic: nucleotide sequence of the synthesized PspPro2 gene in plasmid pGX084 (AprE-PspPro2)

<400> SEQUENCE: 9

|   |     |
|---|-----|
| gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg | 60  |
| gcgttcagca acatgagcgc gcaggctgct ggaaaagcag agcattcagt tctgacccg  | 120 |
| acgcaactta caccgacatt tcatgctgag cagtggaagg caccgagcac ggtcacgggc | 180 |
| gacaacatcg tgtggagcta cctgaacaga cagaaaaaga cgctgctgaa cacggactca | 240 |
| acgagcgtga gagaccatt cagaatcatc gacagaacga gcgacaagtc aggcgcgtca  | 300 |

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cattatagac tgaagcagta cgtgaacggc atcccggctc acggagccga gcaaaccgatc 360
catgtgaata atgcgggcaa agttacatca tacctggggc ccgatcatctc agaagaccag 420
cagcaagatg caacggaggga tacaacaccg aagatcagcg ccacagaagc ggtctatcag 480
gcttacgccc aagcggctgc aagaatccag agcttcccgt caattaatga cagcctgagc 540
gaagcatcag aggaacaagg cagcgagAAC cagggcaatg aaatccaaaa catcggcac 600
aagagcagcg tgtcaaacga cacgtatgcg gaggtcata acaacgttct gctgacaccg 660
gtcgatcagg ccgaacagag ctatatgca aagatcgcga atctggagcc gtcagtcgag 720
ccgaaggccc agctgtatat ctatccggac ggcgagacga cgagactggt gtacgttacg 780
gaggtcaaca tccttgagcc tgcgcgctg agaacaagat actttatcga cgccaagacg 840
ggcaagatcg tgtttcagta cgatacctg aaccatgcga cgggaacagg cagaggcgtg 900
gacggcaaaa caaatcatt cacgacaacg gcaagcggca acagatacca gctgaaggac 960
acaacaagat caaatggcat cgtcacatc acggccggaa atagacagac gacgccggga 1020
acgattctga cggatacaga taactgtgg gaagatccgg cagcagttga tgcacatgca 1080
tacggatca agacgtacga ctactacaag aacaattcg gaagagattc aatcgatgga 1140
agaggcatgc aatcagatc aacggttcat tatggcaaaa agtacaacaa tgccttctgg 1200
aacggcagcc aatgacata cggcgatgga gacggctcaa cgtttacatt cttttcaggc 1260
gacccggacg tcgctggcca tgaactgacg catggcgta cagagttcac gagcaactg 1320
gagtattacg gcaatcagg cgcactgaat gaggtttca ggcacatcat tggcaacgac 1380
attgatggca catcatggct gcttggcgac ggcatttaca cacctaacat tccggcgat 1440
gcaactgaga gcctgtcaga ccctacgaga ttcggccaac ctgaccatta cagcaacttc 1500
taccggatc ctaataacga tgaatggggc ggagtgcata cgaacagcgg cattatcaac 1560
aaagcgtact atctgctggc acaaggcggc acgtcacatg gagtgacggt gacaggaatc 1620
ggcagagagg cggcagtggt tatctactac aacgccttca caaactacct gacgagcagc 1680
tcaaatttca gcaacgctag agcggcggtc atccaggcag caaaggactt ttatggagca 1740
gactcactgg cagttacgtc agcaattcag tcattcgacg cagttggaat taag 1794

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&lt;210&gt; SEQ ID NO 10

&lt;211&gt; LENGTH: 598

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Synthetic: amino acid sequence of the PspPro2 precursor protein expressed from plasmid pGX084 (AprE-PspPro2)

&lt;400&gt; SEQUENCE: 10

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Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10           15

Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20           25           30

Ala Glu His Ser Val Pro Asp Pro Thr Gln Leu Thr Pro Thr Phe His
35           40           45

Ala Glu Gln Trp Lys Ala Pro Ser Thr Val Thr Gly Asp Asn Ile Val
50           55           60

Trp Ser Tyr Leu Asn Arg Gln Lys Lys Thr Leu Leu Asn Thr Asp Ser
65           70           75           80

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Thr Ser Val Arg Asp Gln Phe Arg Ile Ile Asp Arg Thr Ser Asp Lys  
85 90 95

Ser Gly Ala Ser His Tyr Arg Leu Lys Gln Tyr Val Asn Gly Ile Pro  
100 105 110

Val Tyr Gly Ala Glu Gln Thr Ile His Val Asn Asn Ala Gly Lys Val  
115 120 125

Thr Ser Tyr Leu Gly Ala Val Ile Ser Glu Asp Gln Gln Gln Asp Ala  
130 135 140

Thr Glu Asp Thr Thr Pro Lys Ile Ser Ala Thr Glu Ala Val Tyr Thr  
145 150 155 160

Ala Tyr Ala Glu Ala Ala Ala Arg Ile Gln Ser Phe Pro Ser Ile Asn  
165 170 175

Asp Ser Leu Ser Glu Ala Ser Glu Glu Gln Gly Ser Glu Asn Gln Gly  
180 185 190

Asn Glu Ile Gln Asn Ile Gly Ile Lys Ser Ser Val Ser Asn Asp Thr  
195 200 205

Tyr Ala Glu Ala His Asn Asn Val Leu Leu Thr Pro Val Asp Gln Ala  
210 215 220

Glu Gln Ser Tyr Ile Ala Lys Ile Ala Asn Leu Glu Pro Ser Val Glu  
225 230 235 240

Pro Lys Ala Glu Leu Tyr Ile Tyr Pro Asp Gly Glu Thr Thr Arg Leu  
245 250 255

Val Tyr Val Thr Glu Val Asn Ile Leu Glu Pro Ala Pro Leu Arg Thr  
260 265 270

Arg Tyr Phe Ile Asp Ala Lys Thr Gly Lys Ile Val Phe Gln Tyr Asp  
275 280 285

Ile Leu Asn His Ala Thr Gly Thr Gly Arg Gly Val Asp Gly Lys Thr  
290 295 300

Lys Ser Phe Thr Thr Thr Ala Ser Gly Asn Arg Tyr Gln Leu Lys Asp  
305 310 315 320

Thr Thr Arg Ser Asn Gly Ile Val Thr Tyr Thr Ala Gly Asn Arg Gln  
325 330 335

Thr Thr Pro Gly Thr Ile Leu Thr Asp Thr Asp Asn Val Trp Glu Asp  
340 345 350

Pro Ala Ala Val Asp Ala His Ala Tyr Ala Ile Lys Thr Tyr Asp Tyr  
355 360 365

Tyr Lys Asn Lys Phe Gly Arg Asp Ser Ile Asp Gly Arg Gly Met Gln  
370 375 380

Ile Arg Ser Thr Val His Tyr Gly Lys Lys Tyr Asn Asn Ala Phe Trp  
385 390 395 400

Asn Gly Ser Gln Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Thr  
405 410 415

Phe Phe Ser Gly Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly  
420 425 430

Val Thr Glu Phe Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala  
435 440 445

Leu Asn Glu Ala Phe Ser Asp Ile Ile Gly Asn Asp Ile Asp Gly Thr  
450 455 460

Ser Trp Leu Leu Gly Asp Gly Ile Tyr Thr Pro Asn Ile Pro Gly Asp  
465 470 475 480

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Ala Leu Arg Ser Leu Ser Asp Pro Thr Arg Phe Gly Gln Pro Asp His  
485 490 495

Tyr Ser Asn Phe Tyr Pro Asp Pro Asn Asn Asp Asp Glu Gly Gly Val  
500 505 510

His Thr Asn Ser Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln  
515 520 525

Gly Gly Thr Ser His Gly Val Thr Val Thr Gly Ile Gly Arg Glu Ala  
530 535 540

Ala Val Phe Ile Tyr Tyr Asn Ala Phe Thr Asn Tyr Leu Thr Ser Thr  
545 550 555 560

Ser Asn Phe Ser Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp  
565 570 575

Phe Tyr Gly Ala Asp Ser Leu Ala Val Thr Ser Ala Ile Gln Ser Phe  
580 585 590

Asp Ala Val Gly Ile Lys  
595

<210> SEQ ID NO 11  
<211> LENGTH: 1599  
<212> TYPE: DNA  
<213> ORGANISM: Paenibacillus humicus  
<220> FEATURE:  
<221> NAME/KEY: misc\_feature  
<222> LOCATION: (1)..(1599)  
<223> OTHER INFORMATION: nucleotide sequence of the PhuPro2 gene  
isolated from Paenibacillus humicus

<400> SEQUENCE: 11

```

atgaaaaaaaa tgattcctac tctgctcggt accgtattgc tgetttcttc cgettcogct    60
gtcgtgctg aatcgccaag cctcggagcg gccggaactc cgggggtcag cgtcgtgaac    120
aatcagctcg tgactcaatt catcgaggct tocaaggatg ccaagattgt cccgggctct    180
tccgaggata aaatctgggg tttccttgaa ggccagcaag caaagctggg tgtatccgca    240
gcggatgtaa aaacctcggt cctgatccag aagaaggaag tccgatccgac ttcgggcgctc    300
gagcatttcc gctgcagca atatgtgaat ggcatcccgg tatatggcgg tgaccaaacc    360
attcacatcg acaaggccgg ccaggttacg tcgttcgtag gagctgttct gccggctcaa    420
aatcaaatca cggcaaaatc cagcgtacca gccataagcg catccgacgc tctggctatc    480
gcggcgaagg aagccagttc ccgcatcgcc gagctgggag cacaggagaa gactccgctc    540
gctcagctgt acgtatatcc ggaaggcaac gggtcgctgc tcgtctacca gacggaagtg    600
aatgtgcttg agccgcagcc tctgcgcacc cgctatctta tcgatgcggc cgacggccat    660
atcgtgcagc agtacgatct gatcgagacg gcgaccgggt cgggcacggg cgtgctgggc    720
gacaataaga cgttccagac gactctttcc ggcagcacgt accagctgaa agacaccact    780
cgcggaacg gcactctaac ctacacagcc agcaatcgga ccacgattcc gggcacgctg    840
ctgacggacg ccgacaacgt atggacggat ggagccgccc tcgatgccc tacttatgcc    900
ggaaaagtat atgatttcta caaaacgaag ttcggacgca acagcctcga cggcaacggc    960
ctgctgatcc gttcctcggt ccaactacagc agcaggtaca acaatgcctt ctggaacggc   1020
accagattg tattcggcga cggcgacggc tcgacgttca ttccgctgctc gggcgtctc   1080
gacgtggctg gccatgagct gtcccacgga gtcacgagc acacgtccaa ccttcaatac   1140
ctcaatgaat cggcgcgct gaacgagctc tatgccgacg tcctcggcaa ctcgatccag   1200

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gcgaaaaact ggcttatcgg cgacgatgtc tatacgctg gcatctcgg agatgcttc 1260
cgttccatgt ccaacccgac gctttacggg cagccggaca actatgcaa ccgtatacg 1320
ggatcttccg acaacggcgg cgttcatacg aacagcggca tcacgaacaa agcgttctac 1380
ctgctcgccc aaggcggcac ccagaacggc gttaccgtcg ccggcatcgg gcgcgacgca 1440
gccgtgaaca ttttctacaa cacagtggcc tattacctta cttccacttc caacttcgcc 1500
gcggcgaaga acgcctcgat ccaggcagcc aaagacctgt acggaacggg ctctcttat 1560
gtcacctcgg tgaccaatgc attcagagcc gtaggcctg 1599

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<210> SEQ ID NO 12
<211> LENGTH: 533
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus humicus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(533)
<223> OTHER INFORMATION: amino acid sequence of the PhuPro2 precursor
protein

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<400> SEQUENCE: 12

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```

Met Lys Lys Met Ile Pro Thr Leu Leu Gly Thr Val Leu Leu Leu Ser
1           5           10           15
Ser Ala Ser Ala Val Ala Ala Glu Ser Pro Ser Leu Gly Ala Ala Gly
20           25           30
Thr Pro Gly Val Ser Val Val Asn Asn Gln Leu Val Thr Gln Phe Ile
35           40           45
Glu Ala Ser Lys Asp Ala Lys Ile Val Pro Gly Ser Ser Glu Asp Lys
50           55           60
Ile Trp Ala Phe Leu Glu Gly Gln Gln Ala Lys Leu Gly Val Ser Ala
65           70           75           80
Ala Asp Val Lys Thr Ser Phe Leu Ile Gln Lys Lys Glu Val Asp Pro
85           90           95
Thr Ser Gly Val Glu His Phe Arg Leu Gln Gln Tyr Val Asn Gly Ile
100          105          110
Pro Val Tyr Gly Gly Asp Gln Thr Ile His Ile Asp Lys Ala Gly Gln
115          120          125
Val Thr Ser Phe Val Gly Ala Val Leu Pro Ala Gln Asn Gln Ile Thr
130          135          140
Ala Lys Ser Ser Val Pro Ala Ile Ser Ala Ser Asp Ala Leu Ala Ile
145          150          155          160
Ala Ala Lys Glu Ala Ser Ser Arg Ile Gly Glu Leu Gly Ala Gln Glu
165          170          175
Lys Thr Pro Ser Ala Gln Leu Tyr Val Tyr Pro Glu Gly Asn Gly Ser
180          185          190
Arg Leu Val Tyr Gln Thr Glu Val Asn Val Leu Glu Pro Gln Pro Leu
195          200          205
Arg Thr Arg Tyr Leu Ile Asp Ala Ala Asp Gly His Ile Val Gln Gln
210          215          220
Tyr Asp Leu Ile Glu Thr Ala Thr Gly Ser Gly Thr Gly Val Leu Gly
225          230          235          240
Asp Asn Lys Thr Phe Gln Thr Thr Leu Ser Gly Ser Thr Tyr Gln Leu
245          250          255

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Lys Asp Thr Thr Arg Gly Asn Gly Ile Tyr Thr Tyr Thr Ala Ser Asn  
                   260                                  265                                  270  
 Arg Thr Thr Ile Pro Gly Thr Leu Leu Thr Asp Ala Asp Asn Val Trp  
                   275                                  280                                  285  
 Thr Asp Gly Ala Ala Val Asp Ala His Thr Tyr Ala Gly Lys Val Tyr  
                   290                                  295                                  300  
 Asp Phe Tyr Lys Thr Lys Phe Gly Arg Asn Ser Leu Asp Gly Asn Gly  
 305                                  310                                  315                                  320  
 Leu Leu Ile Arg Ser Ser Val His Tyr Ser Ser Arg Tyr Asn Asn Ala  
                   325                                  330                                  335  
 Phe Trp Asn Gly Thr Gln Ile Val Phe Gly Asp Gly Asp Gly Ser Thr  
                   340                                  345                                  350  
 Phe Ile Pro Leu Ser Gly Asp Leu Asp Val Val Gly His Glu Leu Ser  
                   355                                  360                                  365  
 His Gly Val Ile Glu Tyr Thr Ser Asn Leu Gln Tyr Leu Asn Glu Ser  
                   370                                  375                                  380  
 Gly Ala Leu Asn Glu Ser Tyr Ala Asp Val Leu Gly Asn Ser Ile Gln  
 385                                  390                                  395                                  400  
 Ala Lys Asn Trp Leu Ile Gly Asp Asp Val Tyr Thr Pro Gly Ile Ser  
                   405                                  410                                  415  
 Gly Asp Ala Leu Arg Ser Met Ser Asn Pro Thr Leu Tyr Gly Gln Pro  
                   420                                  425                                  430  
 Asp Asn Tyr Ala Asn Arg Tyr Thr Gly Ser Ser Asp Asn Gly Gly Val  
                   435                                  440                                  445  
 His Thr Asn Ser Gly Ile Thr Asn Lys Ala Phe Tyr Leu Leu Ala Gln  
                   450                                  455                                  460  
 Gly Gly Thr Gln Asn Gly Val Thr Val Ala Gly Ile Gly Arg Asp Ala  
 465                                  470                                  475                                  480  
 Ala Val Asn Ile Phe Tyr Asn Thr Val Ala Tyr Tyr Leu Thr Ser Thr  
                   485                                  490                                  495  
 Ser Asn Phe Ala Ala Ala Lys Asn Ala Ser Ile Gln Ala Ala Lys Asp  
                   500                                  505                                  510  
 Leu Tyr Gly Thr Gly Ser Ser Tyr Val Thr Ser Val Thr Asn Ala Phe  
                   515                                  520                                  525  
 Arg Ala Val Gly Leu  
                   530

<210> SEQ ID NO 13  
 <211> LENGTH: 303  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus humicus  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(303)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature  
                                   form of PhuPro2

<400> SEQUENCE: 13

Ala Thr Gly Ser Gly Thr Gly Val Leu Gly Asp Asn Lys Thr Phe Gln  
 1                  5                                  10                                  15  
 Thr Thr Leu Ser Gly Ser Thr Tyr Gln Leu Lys Asp Thr Thr Arg Gly  
                   20                                  25                                  30  
 Asn Gly Ile Tyr Thr Tyr Thr Ala Ser Asn Arg Thr Thr Ile Pro Gly  
                   35                                  40                                  45

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Thr Leu Leu Thr Asp Ala Asp Asn Val Trp Thr Asp Gly Ala Ala Val  
 50 55 60  
 Asp Ala His Thr Tyr Ala Gly Lys Val Tyr Asp Phe Tyr Lys Thr Lys  
 65 70 75 80  
 Phe Gly Arg Asn Ser Leu Asp Gly Asn Gly Leu Leu Ile Arg Ser Ser  
 85 90 95  
 Val His Tyr Ser Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Thr Gln  
 100 105 110  
 Ile Val Phe Gly Asp Gly Asp Gly Ser Thr Phe Ile Pro Leu Ser Gly  
 115 120 125  
 Asp Leu Asp Val Val Gly His Glu Leu Ser His Gly Val Ile Glu Tyr  
 130 135 140  
 Thr Ser Asn Leu Gln Tyr Leu Asn Glu Ser Gly Ala Leu Asn Glu Ser  
 145 150 155 160  
 Tyr Ala Asp Val Leu Gly Asn Ser Ile Gln Ala Lys Asn Trp Leu Ile  
 165 170 175  
 Gly Asp Asp Val Tyr Thr Pro Gly Ile Ser Gly Asp Ala Leu Arg Ser  
 180 185 190  
 Met Ser Asn Pro Thr Leu Tyr Gly Gln Pro Asp Asn Tyr Ala Asn Arg  
 195 200 205  
 Tyr Thr Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile  
 210 215 220  
 Thr Asn Lys Ala Phe Tyr Leu Leu Ala Gln Gly Gly Thr Gln Asn Gly  
 225 230 235 240  
 Val Thr Val Ala Gly Ile Gly Arg Asp Ala Ala Val Asn Ile Phe Tyr  
 245 250 255  
 Asn Thr Val Ala Tyr Tyr Leu Thr Ser Thr Ser Asn Phe Ala Ala Ala  
 260 265 270  
 Lys Asn Ala Ser Ile Gln Ala Ala Lys Asp Leu Tyr Gly Thr Gly Ser  
 275 280 285  
 Ser Tyr Val Thr Ser Val Thr Asn Ala Phe Arg Ala Val Gly Leu  
 290 295 300

&lt;210&gt; SEQ ID NO 14

&lt;211&gt; LENGTH: 1629

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic: nucleotide sequence of the synthesized PhuPro2 gene in plasmid pGX150(AprE- PhuPro2)

&lt;400&gt; SEQUENCE: 14

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg      60
gcggttcagca acatgagcgc gcaggctgct ggaaaagaat caccgagcct tggcgtgca      120
ggaacaccgg gcgtagcgt tgtgaataac caactggcga cgcagttcat cgaagcatca      180
aaagacgcga aaattgtccc tggatcaagc gaagataaga tttgggcatt tctggaaggc      240
cagcaagcaa agcttggcgt ctcagctgcc gacgtgaaga cgagcttctc gatccagaag      300
aaggagggtt acccgacatc aggcgttgag cacttttagac tgcaacagta cgtcaacggc      360
atcccggttt atggaggcga tcaaacatc catattgata aggcaggcca ggtcacatca      420
ttcgtcggag ctgtcctgcc ggctcagaac caaattacag caaatatc agttccggca      480
atctcagcct cagacgctct ggcaatcgct gccaaaggagg caagctcaag aattggcgaa      540
  
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ctgggcgcac aagaaaagac accgagcgcc caactttatg tctatccgga gggcaacgga 600
agcagactgg tgtaccagac agaggtcaat gttctggagc cgcaaccgct gagaacgaga 660
taccttatcg atgctgcgga tggccacatt gttcagcaat acgacctgat tgagacagca 720
acaggaagcg gaacggcggt gctggggcgc aacaagacgt ttcagacaac acttagcggc 780
agcacgtacc aacttaagga cacgacgaga ggcaatggca tttacacgta cacggcctca 840
aacagaacga caatcccagg cacactgctg acggatgcag acaatgtttg gacggacggc 900
gcagcagttg acgcacacac gtacgccggc aaggtgtacg acttttacia gacgaagttc 960
ggcagaaaca gccttgatgg aaatggactg ctgatcagaa gcagcgtcca ctacagcagc 1020
agatacaata acgccttctg gaacggcaca caaatcgtct ttggcgatgg agacggatca 1080
acattcatcc cgctgtcagg cgacctggac gttgtggggc acgagctgag ccacggcgtc 1140
atcgagtaca cgagcaacct gcagtacctg aatgaaagcg gcgcaactgaa cgagtcatat 1200
gctgatgtgc ttggcaatag catccaggcc aagaactggc ttatcggaga cgacgtctac 1260
acacctggca tcagcggcga tgctctgaga agcatgagca atcctacact ttacggccaa 1320
ccggacaact acgcaatag atatacgggc agcagcgaca atggcggcgt tcatacaaac 1380
tcaggcatca cgaacaagcg gttctacctg ctggcacagg gaggcacgca aaacggcggt 1440
acagttgctg gcattggcag agatgcggcc gtcaacatct tctacaacac agtcgcctac 1500
tacctgacga gcacgtcaaa ctctgcagcg gcaaagaacg catcaattca agcagcaaag 1560
gatctgtacg gaacaggcag ctcatatgtc acgtcagtta cgaatgcggt tagagccgtc 1620
ggcctttaa 1629

```

&lt;210&gt; SEQ ID NO 15

&lt;211&gt; LENGTH: 542

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic: amino acid sequence of the PhuPro2 precursor protein expressed from plasmid pGX150(AprE- PhuPro2)

&lt;400&gt; SEQUENCE: 15

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10           15
Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20           25           30
Glu Ser Pro Ser Leu Gly Ala Ala Gly Thr Pro Gly Val Ser Val Val
35           40           45
Asn Asn Gln Leu Val Thr Gln Phe Ile Glu Ala Ser Lys Asp Ala Lys
50           55           60
Ile Val Pro Gly Ser Ser Glu Asp Lys Ile Trp Ala Phe Leu Glu Gly
65           70           75           80
Gln Gln Ala Lys Leu Gly Val Ser Ala Ala Asp Val Lys Thr Ser Phe
85           90           95
Leu Ile Gln Lys Lys Glu Val Asp Pro Thr Ser Gly Val Glu His Phe
100          105          110
Arg Leu Gln Gln Tyr Val Asn Gly Ile Pro Val Tyr Gly Gly Asp Gln
115          120          125
Thr Ile His Ile Asp Lys Ala Gly Gln Val Thr Ser Phe Val Gly Ala
130          135          140

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Val Leu Pro Ala Gln Asn Gln Ile Thr Ala Lys Ser Ser Val Pro Ala  
 145 150 155 160  
 Ile Ser Ala Ser Asp Ala Leu Ala Ile Ala Ala Lys Glu Ala Ser Ser  
 165 170 175  
 Arg Ile Gly Glu Leu Gly Ala Gln Glu Lys Thr Pro Ser Ala Gln Leu  
 180 185 190  
 Tyr Val Tyr Pro Glu Gly Asn Gly Ser Arg Leu Val Tyr Gln Thr Glu  
 195 200 205  
 Val Asn Val Leu Glu Pro Gln Pro Leu Arg Thr Arg Tyr Leu Ile Asp  
 210 215 220  
 Ala Ala Asp Gly His Ile Val Gln Gln Tyr Asp Leu Ile Glu Thr Ala  
 225 230 235 240  
 Thr Gly Ser Gly Thr Gly Val Leu Gly Asp Asn Lys Thr Phe Gln Thr  
 245 250 255  
 Thr Leu Ser Gly Ser Thr Tyr Gln Leu Lys Asp Thr Thr Arg Gly Asn  
 260 265 270  
 Gly Ile Tyr Thr Tyr Thr Ala Ser Asn Arg Thr Thr Ile Pro Gly Thr  
 275 280 285  
 Leu Leu Thr Asp Ala Asp Asn Val Trp Thr Asp Gly Ala Ala Val Asp  
 290 295 300  
 Ala His Thr Tyr Ala Gly Lys Val Tyr Asp Phe Tyr Lys Thr Lys Phe  
 305 310 315 320  
 Gly Arg Asn Ser Leu Asp Gly Asn Gly Leu Leu Ile Arg Ser Ser Val  
 325 330 335  
 His Tyr Ser Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Thr Gln Ile  
 340 345 350  
 Val Phe Gly Asp Gly Asp Gly Ser Thr Phe Ile Pro Leu Ser Gly Asp  
 355 360 365  
 Leu Asp Val Val Gly His Glu Leu Ser His Gly Val Ile Glu Tyr Thr  
 370 375 380  
 Ser Asn Leu Gln Tyr Leu Asn Glu Ser Gly Ala Leu Asn Glu Ser Tyr  
 385 390 395 400  
 Ala Asp Val Leu Gly Asn Ser Ile Gln Ala Lys Asn Trp Leu Ile Gly  
 405 410 415  
 Asp Asp Val Tyr Thr Pro Gly Ile Ser Gly Asp Ala Leu Arg Ser Met  
 420 425 430  
 Ser Asn Pro Thr Leu Tyr Gly Gln Pro Asp Asn Tyr Ala Asn Arg Tyr  
 435 440 445  
 Thr Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Thr  
 450 455 460  
 Asn Lys Ala Phe Tyr Leu Leu Ala Gln Gly Gly Thr Gln Asn Gly Val  
 465 470 475 480  
 Thr Val Ala Gly Ile Gly Arg Asp Ala Ala Val Asn Ile Phe Tyr Asn  
 485 490 495  
 Thr Val Ala Tyr Tyr Leu Thr Ser Thr Ser Asn Phe Ala Ala Ala Lys  
 500 505 510  
 Asn Ala Ser Ile Gln Ala Ala Lys Asp Leu Tyr Gly Thr Gly Ser Ser  
 515 520 525  
 Tyr Val Thr Ser Val Thr Asn Ala Phe Arg Ala Val Gly Leu  
 530 535 540

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<210> SEQ ID NO 16
<211> LENGTH: 1581
<212> TYPE: DNA
<213> ORGANISM: Paenibacillus ehimensis
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1581)
<223> OTHER INFORMATION: nucleotide sequence of the PehPro1 gene
        isolated from Paenibacillus ehimensis

<400> SEQUENCE: 16

atgttaaaag tatgggcatc gattattaca ggagcatttt tgctcgggag cgtgcaaggg    60
gtgcaagctg ctccacaaga tcaagctgct cccttcggag gattcacccc tcaattgatt    120
accggggaaa gctggagtgc gccgcaagga gtatcgggag aggaaaaaat ctggaagtat    180
ctcgaatcca agcagaaaag cttccaaatc ggccaaaccg ttgatctgaa aaagcaattg    240
aaaattatcg gccaaacgac cgacgagaaa acgggaacca cgcattaccg tctacagcag    300
tatgtgggag gcgctcccgt atacggcggc gtacaaaacga tccatgtcaa caaagaagga    360
caagttacct cgctgatcgg cagcctgctt cccgaccagc agcagcaagt ttcgaaaagc    420
ttgaattcgc aaatcagcga agcgcaagcc atcgccgtgg cccagaaaaga taccgaggcc    480
gccgtcggca agctgggtga accgcaaaag acaccggaag cggatctgta cgtttattta    540
cacaacggac aaccggctct cgcttatgtg accgaggtta acgttctcga accggaggca    600
atccggacgc gctacttcat cagcgccgaa gacggcagca ttttattcaa gtacgacatc    660
ctcgctcacg ctacaggtac cggaaaaggg gtgctcggag atacgaaatc gttcacgacc    720
acgcaatccg gctccactta tcaattgaag gatacgacgc gggggcaagg tatcgtcact    780
tacagcgctg gcaaccggtc ctctctgccg ggaacgctgc tcaccagctc cagcaatatt    840
tggaacgacg gcgcgcggtt cgatgcgcat gcctataccg ccaaaagtga cgattactat    900
aaaaaacaat ttggccgcaa cagcattgac ggcaacggct tccagcttaa atcgaccgtg    960
cactattcct ccagatacaa caacgccttc tggaacggtg tgcaaatggt gtacggcgac   1020
ggcgacggcg taaccttcat tccgttctcc gccgatccgg acgtcatcgg ccacgaattg   1080
acccacggcg ttacggaaac tacggccggc ctggaatact acggcgaatc cggagcgctg   1140
aacgaatcga tctccgatat tatcggcaac gcgatcgacg gcaaaaactg gctgatcggc   1200
gacttgattt atacggcga tactcccggg gacgcccctc gctctatgga gaacccaag   1260
ctgtataacc aaccgcaccg ctatcaagac cgctatacgg gaccttccga taacggcggc   1320
gtgcatatta acagcgggat caacaacaaa gccttctacc tgatcgccca aggcggcacc   1380
cactatggcg tcaccgtgaa cgggatcggg cgcgatcggg ctgtgcaaat tttctatgac   1440
gccctcatca attacctgac tccaacttcg aacttctcgg cgatgcgcgc agcagccatt   1500
caagcggcaa ccgacctgta cggagcgaat tcttctcaag taaacgctgt caaaaaagcg   1560
tatactgccg tcggcgtgaa c                                     1581

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<210> SEQ ID NO 17
<211> LENGTH: 527
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus ehimensis
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(527)
<223> OTHER INFORMATION: amino acid sequence of the PehPro1 precursor

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protein

<400> SEQUENCE: 17

Met Leu Lys Val Trp Ala Ser Ile Ile Thr Gly Ala Phe Leu Leu Gly  
 1 5 10 15

Ser Val Gln Gly Val Gln Ala Ala Pro Gln Asp Gln Ala Ala Pro Phe  
 20 25 30

Gly Gly Phe Thr Pro Gln Leu Ile Thr Gly Glu Ser Trp Ser Ala Pro  
 35 40 45

Gln Gly Val Ser Gly Glu Glu Lys Ile Trp Lys Tyr Leu Glu Ser Lys  
 50 55 60

Gln Glu Ser Phe Gln Ile Gly Gln Thr Val Asp Leu Lys Lys Gln Leu  
 65 70 75 80

Lys Ile Ile Gly Gln Thr Thr Asp Glu Lys Thr Gly Thr Thr His Tyr  
 85 90 95

Arg Leu Gln Gln Tyr Val Gly Gly Val Pro Val Tyr Gly Gly Val Gln  
 100 105 110

Thr Ile His Val Asn Lys Glu Gly Gln Val Thr Ser Leu Ile Gly Ser  
 115 120 125

Leu Leu Pro Asp Gln Gln Gln Gln Val Ser Lys Ser Leu Asn Ser Gln  
 130 135 140

Ile Ser Glu Ala Gln Ala Ile Ala Val Ala Gln Lys Asp Thr Glu Ala  
 145 150 155 160

Ala Val Gly Lys Leu Gly Glu Pro Gln Lys Thr Pro Glu Ala Asp Leu  
 165 170 175

Tyr Val Tyr Leu His Asn Gly Gln Pro Val Leu Ala Tyr Val Thr Glu  
 180 185 190

Val Asn Val Leu Glu Pro Glu Ala Ile Arg Thr Arg Tyr Phe Ile Ser  
 195 200 205

Ala Glu Asp Gly Ser Ile Leu Phe Lys Tyr Asp Ile Leu Ala His Ala  
 210 215 220

Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Thr Thr  
 225 230 235 240

Thr Gln Ser Gly Ser Thr Tyr Gln Leu Lys Asp Thr Thr Arg Gly Gln  
 245 250 255

Gly Ile Val Thr Tyr Ser Ala Gly Asn Arg Ser Ser Leu Pro Gly Thr  
 260 265 270

Leu Leu Thr Ser Ser Ser Asn Ile Trp Asn Asp Gly Ala Ala Val Asp  
 275 280 285

Ala His Ala Tyr Thr Ala Lys Val Tyr Asp Tyr Tyr Lys Asn Lys Phe  
 290 295 300

Gly Arg Asn Ser Ile Asp Gly Asn Gly Phe Gln Leu Lys Ser Thr Val  
 305 310 315 320

His Tyr Ser Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Val Gln Met  
 325 330 335

Val Tyr Gly Asp Gly Asp Gly Val Thr Phe Ile Pro Phe Ser Ala Asp  
 340 345 350

Pro Asp Val Ile Gly His Glu Leu Thr His Gly Val Thr Glu His Thr  
 355 360 365

Ala Gly Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ser Ile  
 370 375 380



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Met Glu Asn Pro Lys Leu Tyr Asn Gln Pro Asp Arg Tyr Gln Asp Arg  
 195 200 205

Tyr Thr Gly Pro Ser Asp Asn Gly Gly Val His Ile Asn Ser Gly Ile  
 210 215 220

Asn Asn Lys Ala Phe Tyr Leu Ile Ala Gln Gly Gly Thr His Tyr Gly  
 225 230 235 240

Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Phe Tyr  
 245 250 255

Asp Ala Leu Ile Asn Tyr Leu Thr Pro Thr Ser Asn Phe Ser Ala Met  
 260 265 270

Arg Ala Ala Ala Ile Gln Ala Ala Thr Asp Leu Tyr Gly Ala Asn Ser  
 275 280 285

Ser Gln Val Asn Ala Val Lys Lys Ala Tyr Thr Ala Val Gly Val Asn  
 290 295 300

<210> SEQ ID NO 19  
 <211> LENGTH: 1611  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic; nucleotide sequence of the synthesized PehPro1 gene in plasmid pGX148(AprE- PehPro1)

<400> SEQUENCE: 19

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg    60
gcgttcagca acatgagcgc gcaggctgct ggaaaagcac ctcaagatca ggcagcacct    120
tttgaggct ttacaccgca acttatcaca ggcgatcat ggtcagcacc gcagggcgctt    180
tcaggcgagg aaaagatctg gaagtacctt gagagcaagc aggagtcatt tcaaatcggc    240
cagacagtcg acctgaaaaa gcaactgaag atcatcggcc aaacaacgga cgaaaagacg    300
ggcacgacgc attatagact gcaacaatat gttggcggcg tgccggttta tggaggcgtg    360
caaaacatcc acgtgaacaa ggaaggacag gtcacgtcac tgatcggcag cctgctgccg    420
gatcagcagc aacaagtctc aaagagcctg aactcacaaa ttagcggaggc acaagcgatt    480
gcagttgcac aaaaggacac ggaagcagct gtcggcaagc tgggcaaac gcaaaaaaca    540
cctgaggctg acctttacgt ctacctgeat aacggccagc cggtccttgc gtacgttacg    600
gaagttaacg tgctggagcc ggaggccatc agaacgagat acttcattag cgcggaggat    660
ggaagcattc tgtttaagta cgatattctt gctcacgcga caggcacagg caagggcgtc    720
cttggcgaca caaaaagctt cacgacaacg cagagcggat caacgtacca gctgaaagat    780
acaacaagag gacaaggcat cgttacgtat tcagcgggca atagatcaag cctgccgggc    840
acactgctga catcaagctc aacatttgg aatgacggcg cagcagttga tgcccatgcg    900
tacacagcca aggtgtacga ctactataag aacaagtttg gcagaaatag catcgacgga    960
aatggatttc aacttaaatc aacggtgcac tactcatcaa gatataacaa tgcgttttgg   1020
aacggagtgc agatggteta cggagacggc gacggcgtga ctttattcc gtttagcgcc   1080
gaccgggacg tgattggaca tgaactgaca catggagtga cagagcatac ggcgggactg   1140
gaatattacg gcgaaagcgg cgcactgaac gaaagcatct cagacattat tggaaacgca   1200
atcgatggca aaaactggct gattggcgat ctgatttata cgccgaatac accgggcgat   1260
gcactgagat caatggagaa tccgaagctg tacaaccaac cggacagata ccaagataga   1320
    
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tacacaggac cgtcagacaa cggcggagtc catatcaaca gcggaatcaa taacaaagcc 1380
ttttactga tcgccaagc cggaaacgcac tatggcgta cagtcaatgg catcggaaga 1440
gatgccgcag ttcagatttt ctatgacgcg ctgatcaact atctgacgcc tacaagcaat 1500
ttctcagcaa tgagagccgc agcaatccaa gcagccacgg atctgtatgg agccaattca 1560
tcacaagtta atgctgttaa gaaggcttat acggcagtgg gagttaacta a 1611

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&lt;210&gt; SEQ ID NO 20

&lt;211&gt; LENGTH: 536

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic; amino acid sequence of the PehPro1 precursor protein expressed from plasmid pGX148(AprE- PehPro1)

&lt;400&gt; SEQUENCE: 20

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1 5 10 15
Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20 25 30
Ala Pro Gln Asp Gln Ala Ala Pro Phe Gly Gly Phe Thr Pro Gln Leu
35 40 45
Ile Thr Gly Glu Ser Trp Ser Ala Pro Gln Gly Val Ser Gly Glu Glu
50 55 60
Lys Ile Trp Lys Tyr Leu Glu Ser Lys Gln Glu Ser Phe Gln Ile Gly
65 70 75 80
Gln Thr Val Asp Leu Lys Lys Gln Leu Lys Ile Ile Gly Gln Thr Thr
85 90 95
Asp Glu Lys Thr Gly Thr Thr His Tyr Arg Leu Gln Gln Tyr Val Gly
100 105 110
Gly Val Pro Val Tyr Gly Gly Val Gln Thr Ile His Val Asn Lys Glu
115 120 125
Gly Gln Val Thr Ser Leu Ile Gly Ser Leu Leu Pro Asp Gln Gln Gln
130 135 140
Gln Val Ser Lys Ser Leu Asn Ser Gln Ile Ser Glu Ala Gln Ala Ile
145 150 155 160
Ala Val Ala Gln Lys Asp Thr Glu Ala Ala Val Gly Lys Leu Gly Glu
165 170 175
Pro Gln Lys Thr Pro Glu Ala Asp Leu Tyr Val Tyr Leu His Asn Gly
180 185 190
Gln Pro Val Leu Ala Tyr Val Thr Glu Val Asn Val Leu Glu Pro Glu
195 200 205
Ala Ile Arg Thr Arg Tyr Phe Ile Ser Ala Glu Asp Gly Ser Ile Leu
210 215 220
Phe Lys Tyr Asp Ile Leu Ala His Ala Thr Gly Thr Gly Lys Gly Val
225 230 235 240
Leu Gly Asp Thr Lys Ser Phe Thr Thr Thr Gln Ser Gly Ser Thr Tyr
245 250 255
Gln Leu Lys Asp Thr Thr Arg Gly Gln Gly Ile Val Thr Tyr Ser Ala
260 265 270
Gly Asn Arg Ser Ser Leu Pro Gly Thr Leu Leu Thr Ser Ser Ser Asn
275 280 285
Ile Trp Asn Asp Gly Ala Ala Val Asp Ala His Ala Tyr Thr Ala Lys

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| 290 |     |     | 295 |     |     | 300 |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Val | Tyr | Asp | Tyr | Tyr | Lys | Asn | Lys | Phe | Gly | Arg | Asn | Ser | Ile | Asp | Gly |
| 305 |     |     |     |     | 310 |     |     |     |     | 315 |     |     |     |     | 320 |
| Asn | Gly | Phe | Gln | Leu | Lys | Ser | Thr | Val | His | Tyr | Ser | Ser | Arg | Tyr | Asn |
|     |     |     |     | 325 |     |     |     |     |     | 330 |     |     |     |     | 335 |
| Asn | Ala | Phe | Trp | Asn | Gly | Val | Gln | Met | Val | Tyr | Gly | Asp | Gly | Asp | Gly |
|     |     |     | 340 |     |     |     |     | 345 |     |     |     |     |     | 350 |     |
| Val | Thr | Phe | Ile | Pro | Phe | Ser | Ala | Asp | Pro | Asp | Val | Ile | Gly | His | Glu |
|     |     | 355 |     |     |     |     | 360 |     |     |     |     |     | 365 |     |     |
| Leu | Thr | His | Gly | Val | Thr | Glu | His | Thr | Ala | Gly | Leu | Glu | Tyr | Tyr | Gly |
|     |     | 370 |     |     |     | 375 |     |     |     |     |     | 380 |     |     |     |
| Glu | Ser | Gly | Ala | Leu | Asn | Glu | Ser | Ile | Ser | Asp | Ile | Ile | Gly | Asn | Ala |
| 385 |     |     |     |     | 390 |     |     |     |     | 395 |     |     |     |     | 400 |
| Ile | Asp | Gly | Lys | Asn | Trp | Leu | Ile | Gly | Asp | Leu | Ile | Tyr | Thr | Pro | Asn |
|     |     |     | 405 |     |     |     |     |     | 410 |     |     |     |     |     | 415 |
| Thr | Pro | Gly | Asp | Ala | Leu | Arg | Ser | Met | Glu | Asn | Pro | Lys | Leu | Tyr | Asn |
|     |     |     | 420 |     |     |     |     | 425 |     |     |     |     |     | 430 |     |
| Gln | Pro | Asp | Arg | Tyr | Gln | Asp | Arg | Tyr | Thr | Gly | Pro | Ser | Asp | Asn | Gly |
|     |     | 435 |     |     |     |     |     | 440 |     |     |     |     | 445 |     |     |
| Gly | Val | His | Ile | Asn | Ser | Gly | Ile | Asn | Asn | Lys | Ala | Phe | Tyr | Leu | Ile |
|     |     | 450 |     |     |     | 455 |     |     |     |     | 460 |     |     |     |     |
| Ala | Gln | Gly | Gly | Thr | His | Tyr | Gly | Val | Thr | Val | Asn | Gly | Ile | Gly | Arg |
| 465 |     |     |     |     | 470 |     |     |     |     | 475 |     |     |     |     | 480 |
| Asp | Ala | Ala | Val | Gln | Ile | Phe | Tyr | Asp | Ala | Leu | Ile | Asn | Tyr | Leu | Thr |
|     |     |     |     | 485 |     |     |     |     | 490 |     |     |     |     |     | 495 |
| Pro | Thr | Ser | Asn | Phe | Ser | Ala | Met | Arg | Ala | Ala | Ala | Ile | Gln | Ala | Ala |
|     |     |     | 500 |     |     |     |     | 505 |     |     |     |     |     | 510 |     |
| Thr | Asp | Leu | Tyr | Gly | Ala | Asn | Ser | Ser | Gln | Val | Asn | Ala | Val | Lys | Lys |
|     |     | 515 |     |     |     |     | 520 |     |     |     |     |     | 525 |     |     |
| Ala | Tyr | Thr | Ala | Val | Gly | Val | Asn |     |     |     |     |     |     |     |     |
|     |     | 530 |     |     |     | 535 |     |     |     |     |     |     |     |     |     |

&lt;210&gt; SEQ ID NO 21

&lt;211&gt; LENGTH: 1563

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Paenibacillus barcinonensis

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1)..(1563)

&lt;223&gt; OTHER INFORMATION: nucleotide sequence of the PbaProl gene isolated from Paenibacillus barcinonensis

&lt;400&gt; SEQUENCE: 21

```

atgaaattga ccaaaattat gccacaatt cttgcaggag ctcttttgct cacatccctg    60
tcctctgcag cagcaatgcc gttatctgac tcattccattc catttgaggg ccctacacc    120
tccgaggaga gtattctggt gaacaacaac cgggacgaaa tgatttataa ttttcttgca    180
caacaagagc aattttctgaa tgccgacgctc aaaggacagc tcaaaatcat taaacgcaac    240
acagacactt ccggcatcag acactttcgt ctgaagcaat acatcaaagg tgttccgggt    300
tacggcgcag acaaacgat ccatctggac aagaacggag ctgtaacttc cgcactcggc    360
gatcttccgc caattgaaga acaggctggt ccgaatgatg gcgttcccgc aatcagtgca    420
gacgatgcca tccgtgccgc cgagaatgaa gccacctccc gtcttgaga gcttgcgca    480

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ccagagcttg agccaaaggc cgaattaaac atttatcatc atgaagatga cggacaaacc 540
tacctcgttt acattacgga agttaacgtg cttgagcctt ccccgctacg gaccaaatat 600
tttattaacg cccttgatgg aagcatcgta tctcaatagc atattatcaa ctttgccaca 660
ggcaccggta caggcgtgca tggtgatacc aaaacactga cgacaactca atccggcagc 720
acctatcagc tgaagatac aactcgtgga aaaggcattc aaacctatac tgcgaacaat 780
cgctcctcgc ttccaggcag cttgtctacc agttccaata acgtatggac agaccgtgca 840
gctgtagatg cgcacgccta tgctgccgcc acatatgact tctacaaaaa caaattcaat 900
cgcaacggca ttgacggaaa cgggctgttg attcgctcta cagtgcatta tggctccaac 960
tataaaaaag ccttctggaa cgggacacag attgtctatg gagatggcga tggcatcgag 1020
ttcggccctt tctccggtga tctcgatgtt gtcggacatg aattgacaca cggggtgatt 1080
gaatatacag ccaatctcga atatcgcaat gagccgggtg ctttaaacga agcttttgcc 1140
gacattatgg ggaacaccat cgaaagcaaa aactggctgc ttggcgacgg aatctatact 1200
ccaaacattc caggtgatgc cctgcgctcg ttatccgacc ctacgctgta taaccagcct 1260
gacaaatata gtgatcgcta cactggctct caggataatg gcggtgtgca tatcaacagc 1320
gggatcatta acaaagcata ttatcttgca gcccaaggcg gtactcataa cggggtaac 1380
gtagcggca tcggccggga taaagcagta cgtatcttct atagcacgct ggtgaactac 1440
ctgacgcaa cctccaaatt tgcagcagcc aaaacagcga caattcaggc agccaaggac 1500
ctgtacgggtg ccaattccgc tgaagctacg gcaatcacca aagcttatca agcggtaggt 1560
ttg 1563

```

&lt;210&gt; SEQ ID NO 22

&lt;211&gt; LENGTH: 521

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Paenibacillus barcinonensis

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1)..(521)

&lt;223&gt; OTHER INFORMATION: amino acid sequence of the PbaPro1 precursor protein

&lt;400&gt; SEQUENCE: 22

```

Met Lys Leu Thr Lys Ile Met Pro Thr Ile Leu Ala Gly Ala Leu Leu
1           5           10          15
Leu Thr Ser Leu Ser Ser Ala Ala Ala Met Pro Leu Ser Asp Ser Ser
20          25          30
Ile Pro Phe Glu Gly Pro Tyr Thr Ser Glu Glu Ser Ile Leu Leu Asn
35          40          45
Asn Asn Pro Asp Glu Met Ile Tyr Asn Phe Leu Ala Gln Gln Glu Gln
50          55          60
Phe Leu Asn Ala Asp Val Lys Gly Gln Leu Lys Ile Ile Lys Arg Asn
65          70          75          80
Thr Asp Thr Ser Gly Ile Arg His Phe Arg Leu Lys Gln Tyr Ile Lys
85          90          95
Gly Val Pro Val Tyr Gly Ala Glu Gln Thr Ile His Leu Asp Lys Asn
100         105         110
Gly Ala Val Thr Ser Ala Leu Gly Asp Leu Pro Pro Ile Glu Glu Gln
115         120         125
Ala Val Pro Asn Asp Gly Val Pro Ala Ile Ser Ala Asp Asp Ala Ile

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| 130  | 135 | 140 |
|--|-----|-----|
| Arg Ala Ala Glu Asn Glu Ala Thr Ser Arg Leu Gly Glu Leu Gly Ala<br>145 150 155 160 |     |     |
| Pro Glu Leu Glu Pro Lys Ala Glu Leu Asn Ile Tyr His His Glu Asp<br>165 170 175     |     |     |
| Asp Gly Gln Thr Tyr Leu Val Tyr Ile Thr Glu Val Asn Val Leu Glu<br>180 185 190     |     |     |
| Pro Ser Pro Leu Arg Thr Lys Tyr Phe Ile Asn Ala Leu Asp Gly Ser<br>195 200 205     |     |     |
| Ile Val Ser Gln Tyr Asp Ile Ile Asn Phe Ala Thr Gly Thr Gly Thr<br>210 215 220     |     |     |
| Gly Val His Gly Asp Thr Lys Thr Leu Thr Thr Thr Gln Ser Gly Ser<br>225 230 235 240 |     |     |
| Thr Tyr Gln Leu Lys Asp Thr Thr Arg Gly Lys Gly Ile Gln Thr Tyr<br>245 250 255     |     |     |
| Thr Ala Asn Asn Arg Ser Ser Leu Pro Gly Ser Leu Ser Thr Ser Ser<br>260 265 270     |     |     |
| Asn Asn Val Trp Thr Asp Arg Ala Ala Val Asp Ala His Ala Tyr Ala<br>275 280 285     |     |     |
| Ala Ala Thr Tyr Asp Phe Tyr Lys Asn Lys Phe Asn Arg Asn Gly Ile<br>290 295 300     |     |     |
| Asp Gly Asn Gly Leu Leu Ile Arg Ser Thr Val His Tyr Gly Ser Asn<br>305 310 315 320 |     |     |
| Tyr Lys Asn Ala Phe Trp Asn Gly Ala Gln Ile Val Tyr Gly Asp Gly<br>325 330 335     |     |     |
| Asp Gly Ile Glu Phe Gly Pro Phe Ser Gly Asp Leu Asp Val Val Gly<br>340 345 350     |     |     |
| His Glu Leu Thr His Gly Val Ile Glu Tyr Thr Ala Asn Leu Glu Tyr<br>355 360 365     |     |     |
| Arg Asn Glu Pro Gly Ala Leu Asn Glu Ala Phe Ala Asp Ile Met Gly<br>370 375 380     |     |     |
| Asn Thr Ile Glu Ser Lys Asn Trp Leu Leu Gly Asp Gly Ile Tyr Thr<br>385 390 395 400 |     |     |
| Pro Asn Ile Pro Gly Asp Ala Leu Arg Ser Leu Ser Asp Pro Thr Leu<br>405 410 415     |     |     |
| Tyr Asn Gln Pro Asp Lys Tyr Ser Asp Arg Tyr Thr Gly Ser Gln Asp<br>420 425 430     |     |     |
| Asn Gly Gly Val His Ile Asn Ser Gly Ile Ile Asn Lys Ala Tyr Tyr<br>435 440 445     |     |     |
| Leu Ala Ala Gln Gly Gly Thr His Asn Gly Val Thr Val Ser Gly Ile<br>450 455 460     |     |     |
| Gly Arg Asp Lys Ala Val Arg Ile Phe Tyr Ser Thr Leu Val Asn Tyr<br>465 470 475 480 |     |     |
| Leu Thr Pro Thr Ser Lys Phe Ala Ala Ala Lys Thr Ala Thr Ile Gln<br>485 490 495     |     |     |
| Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser Ala Glu Ala Thr Ala Ile<br>500 505 510     |     |     |
| Thr Lys Ala Tyr Gln Ala Val Gly Leu<br>515 520                                     |     |     |

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<211> LENGTH: 303  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus barcinonensis  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(303)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature form of PbaPro1

<400> SEQUENCE: 23

Ala Thr Gly Thr Gly Thr Gly Val His Gly Asp Thr Lys Thr Leu Thr  
 1 5 10 15  
 Thr Thr Gln Ser Gly Ser Thr Tyr Gln Leu Lys Asp Thr Thr Arg Gly  
 20 25 30  
 Lys Gly Ile Gln Thr Tyr Thr Ala Asn Asn Arg Ser Ser Leu Pro Gly  
 35 40 45  
 Ser Leu Ser Thr Ser Ser Asn Asn Val Trp Thr Asp Arg Ala Ala Val  
 50 55 60  
 Asp Ala His Ala Tyr Ala Ala Ala Thr Tyr Asp Phe Tyr Lys Asn Lys  
 65 70 75 80  
 Phe Asn Arg Asn Gly Ile Asp Gly Asn Gly Leu Leu Ile Arg Ser Thr  
 85 90 95  
 Val His Tyr Gly Ser Asn Tyr Lys Asn Ala Phe Trp Asn Gly Ala Gln  
 100 105 110  
 Ile Val Tyr Gly Asp Gly Asp Gly Ile Glu Phe Gly Pro Phe Ser Gly  
 115 120 125  
 Asp Leu Asp Val Val Gly His Glu Leu Thr His Gly Val Ile Glu Tyr  
 130 135 140  
 Thr Ala Asn Leu Glu Tyr Arg Asn Glu Pro Gly Ala Leu Asn Glu Ala  
 145 150 155 160  
 Phe Ala Asp Ile Met Gly Asn Thr Ile Glu Ser Lys Asn Trp Leu Leu  
 165 170 175  
 Gly Asp Gly Ile Tyr Thr Pro Asn Ile Pro Gly Asp Ala Leu Arg Ser  
 180 185 190  
 Leu Ser Asp Pro Thr Leu Tyr Asn Gln Pro Asp Lys Tyr Ser Asp Arg  
 195 200 205  
 Tyr Thr Gly Ser Gln Asp Asn Gly Gly Val His Ile Asn Ser Gly Ile  
 210 215 220  
 Ile Asn Lys Ala Tyr Tyr Leu Ala Ala Gln Gly Gly Thr His Asn Gly  
 225 230 235 240  
 Val Thr Val Ser Gly Ile Gly Arg Asp Lys Ala Val Arg Ile Phe Tyr  
 245 250 255  
 Ser Thr Leu Val Asn Tyr Leu Thr Pro Thr Ser Lys Phe Ala Ala Ala  
 260 265 270  
 Lys Thr Ala Thr Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser  
 275 280 285  
 Ala Glu Ala Thr Ala Ile Thr Lys Ala Tyr Gln Ala Val Gly Leu  
 290 295 300

<210> SEQ ID NO 24  
 <211> LENGTH: 1587  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic; nucleotide sequence of the synthesized PbaPro1 gene in plasmid pGX147(AprE- PbaPro1)

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&lt;400&gt; SEQUENCE: 24

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg    60
gcgttcagca acatgagcgc gcaggctgct ggaaaaatgc ctctgtcaga cagcagcatt    120
ccgtttgagg gcccgctacac atcagaagaa agcatcctgc tgaacaacaa cccggacgag    180
atgatctaca atttctggc acagcaggag cagttcctga acgcagacgt gaagggccag    240
ctgaaaatca tcaaaagaaa cacagacacg agcggcatca gacacttcag actgaagcag    300
tacatcaagg gcgtcccggg ttacggcgct gagcagacaa tccacctgga caaaaatggc    360
gcagtgacga gcgcacttgg agatctgccg ccgattgaag agcaagcagt cccgaacgat    420
ggcgttccgg cgattagcgc tgatgacgct atcagagccg cggaaaacga agcgacgtca    480
agactgggag aacttgccgc accggaactt gaaccgaagg cggaaactgaa catctatcac    540
cacgaagacg atggacagac gtacctggtg tacatcacgg aggtgaatgt gctggagccg    600
tcaccgctga gaacaaaata cttcatcaat gcgctggatg gcagcatcgt tagccaatac    660
gacatcatta acttcgccac aggcacgggc acaggcgctc atggcgacac aaaaacgctt    720
acgacaacac agtcaggctc aacgtaccag ctgaaagaca caacaagagg caagggcadc    780
cagacgtata cagccaataa cagaagctca cttccgggct cactgtcaac aagcagcaat    840
aatgtctgga cggacagagc tgcagtggac gcgcacgctg atgctgcggc cactgacgac    900
tttacaaga acaagttcaa cagaaacggc attgatggca acggcctgct tattagaagc    960
acggctccact acggctcaaa ctacaagaat gcgttttgga acggcgccca aattgtttat   1020
ggcgatggag acggcatcga gttcggacct tttagcggcg acctggatgt ggtcggacat   1080
gaactgacgc acggcgttat cgagtatacg gcgaatctgg aatacagaaa tgaaccgggc   1140
gctctgaatg aggccttcgc ggatatcatg ggcaacacaa ttgagagcaa aaactggctt   1200
ctgggcgacg gaatctacac gccgaacatt ccgggagatg cactgagatc actgagcgcac   1260
cctacgctgt acaaccagcc ggacaaatac agcgacagat acacgggatc acaggacaat   1320
ggcgcgctcc atattaactc aggcacatc aacaaagcgt attatctggc agctcaaggc   1380
ggcagcagata atggcgctcag agttagcggg atcggcagag acaaggccgt cagaattttc   1440
tactcaacgc tggatgaacta cctgacaccg acaagcaagt ttgcagccgc caaacagcc   1500
acgattcagg cagcaaaagga cctgtacgga gcgaactcag cagaggccac agcgattacg   1560
aaggcttate aagccgtggg actgtaa                                     1587

```

&lt;210&gt; SEQ ID NO 25

&lt;211&gt; LENGTH: 528

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Synthetic; amino acid sequence of the PbaPro1 precursor protein expressed from plasmid pGX147(AprE-PbaPro1)

&lt;400&gt; SEQUENCE: 25

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10          15

Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
                20          25          30

Met Pro Leu Ser Asp Ser Ser Ile Pro Phe Glu Gly Pro Tyr Thr Ser
          35          40          45

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Glu Glu Ser Ile Leu Leu Asn Asn Asn Pro Asp Glu Met Ile Tyr Asn  
 50 55 60

Phe Leu Ala Gln Gln Glu Gln Phe Leu Asn Ala Asp Val Lys Gly Gln  
 65 70 75 80

Leu Lys Ile Ile Lys Arg Asn Thr Asp Thr Ser Gly Ile Arg His Phe  
 85 90 95

Arg Leu Lys Gln Tyr Ile Lys Gly Val Pro Val Tyr Gly Ala Glu Gln  
 100 105 110

Thr Ile His Leu Asp Lys Asn Gly Ala Val Thr Ser Ala Leu Gly Asp  
 115 120 125

Leu Pro Pro Ile Glu Glu Gln Ala Val Pro Asn Asp Gly Val Pro Ala  
 130 135 140

Ile Ser Ala Asp Asp Ala Ile Arg Ala Ala Glu Asn Glu Ala Thr Ser  
 145 150 155 160

Arg Leu Gly Glu Leu Gly Ala Pro Glu Leu Glu Pro Lys Ala Glu Leu  
 165 170 175

Asn Ile Tyr His His Glu Asp Asp Gly Gln Thr Tyr Leu Val Tyr Ile  
 180 185 190

Thr Glu Val Asn Val Leu Glu Pro Ser Pro Leu Arg Thr Lys Tyr Phe  
 195 200 205

Ile Asn Ala Leu Asp Gly Ser Ile Val Ser Gln Tyr Asp Ile Ile Asn  
 210 215 220

Phe Ala Thr Gly Thr Gly Thr Gly Val His Gly Asp Thr Lys Thr Leu  
 225 230 235 240

Thr Thr Thr Gln Ser Gly Ser Thr Tyr Gln Leu Lys Asp Thr Thr Arg  
 245 250 255

Gly Lys Gly Ile Gln Thr Tyr Thr Ala Asn Asn Arg Ser Ser Leu Pro  
 260 265 270

Gly Ser Leu Ser Thr Ser Ser Asn Asn Val Trp Thr Asp Arg Ala Ala  
 275 280 285

Val Asp Ala His Ala Tyr Ala Ala Ala Thr Tyr Asp Phe Tyr Lys Asn  
 290 295 300

Lys Phe Asn Arg Asn Gly Ile Asp Gly Asn Gly Leu Leu Ile Arg Ser  
 305 310 315 320

Thr Val His Tyr Gly Ser Asn Tyr Lys Asn Ala Phe Trp Asn Gly Ala  
 325 330 335

Gln Ile Val Tyr Gly Asp Gly Asp Gly Ile Glu Phe Gly Pro Phe Ser  
 340 345 350

Gly Asp Leu Asp Val Val Gly His Glu Leu Thr His Gly Val Ile Glu  
 355 360 365

Tyr Thr Ala Asn Leu Glu Tyr Arg Asn Glu Pro Gly Ala Leu Asn Glu  
 370 375 380

Ala Phe Ala Asp Ile Met Gly Asn Thr Ile Glu Ser Lys Asn Trp Leu  
 385 390 395 400

Leu Gly Asp Gly Ile Tyr Thr Pro Asn Ile Pro Gly Asp Ala Leu Arg  
 405 410 415

Ser Leu Ser Asp Pro Thr Leu Tyr Asn Gln Pro Asp Lys Tyr Ser Asp  
 420 425 430

Arg Tyr Thr Gly Ser Gln Asp Asn Gly Gly Val His Ile Asn Ser Gly  
 435 440 445

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Ile Ile Asn Lys Ala Tyr Tyr Leu Ala Ala Gln Gly Gly Thr His Asn  
 450 455 460

Gly Val Thr Val Ser Gly Ile Gly Arg Asp Lys Ala Val Arg Ile Phe  
 465 470 475 480

Tyr Ser Thr Leu Val Asn Tyr Leu Thr Pro Thr Ser Lys Phe Ala Ala  
 485 490 495

Ala Lys Thr Ala Thr Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn  
 500 505 510

Ser Ala Glu Ala Thr Ala Ile Thr Lys Ala Tyr Gln Ala Val Gly Leu  
 515 520 525

<210> SEQ ID NO 26  
 <211> LENGTH: 1779  
 <212> TYPE: DNA  
 <213> ORGANISM: Paenibacillus polymyxa SC2  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(1779)  
 <223> OTHER INFORMATION: nucleotide sequence of the PpoProl gene  
 identified from NCBI database

<400> SEQUENCE: 26

```

atgaaaaaag tatgggtttc gcttcttggg ggagctatgt tattaggggc tgtcgcgtct 60
gggtcatctg cggagagtgc cgtttcgggg ccagctcagc ttacaccgac cttccacgcc 120
gagcaatgga aagcacctac ctcggtatcg ggggatgaca ttgtatggag ctatttaaata 180
cgacaaaaga aatcgttgcg ggggtgtggat agctccagtg tacgtgaaca attccgaatc 240
gttgatcgca caagcgacaa atccggtgta agccattatc gactgaagca gtagttaaac 300
ggaattcccc tgtatggagc tgaacaaact attcatgtgg gcaaatctgg tgaggtcacc 360
tcttacttag gagcgggtgt taatgaggat cagcaggcag aagctacgca aggtacaact 420
ccaaaaatca gcgcttctga agcgggtctac accgcatata aagaagcagc tgcacggatt 480
gaagccctcc ctacctccga cgatactatt tctaaagacg ctgaggagcc aagcagtgta 540
agtaaagata cttaccgcca agcagctaac aacgaaaaaa cgctttctgt tgataaggac 600
gagctgagtc ttgatcaggc atctgtcctg aaagatagca aaattgaagc agtggaaacca 660
gaaaaaagtt ccattgcca aatcgcta atctgcagcctg aagtagatcc taaagcagaa 720
ctctactact accctaaggg ggatgacctg ctgctggttt atgtaacaga agttaatggt 780
ttagaacctg cccactgcg taccgcgtac attattgatg ccaatgacgg cagcatcgta 840
ttccagtatg acatcattaa tgaagcgaca ggcacaggta aaggtgtgct tggtgattcc 900
aaatcgttca ctactaccgc ttccggcagt agctaccagt taaaagatac aacacgcggt 960
aacggaatcg tgacttacac ggcctccaac cgtcaaagca tcccaggtag cattttgaca 1020
gatgccgata atgtatggaa tgatccagct ggtgtggacg cccatgcgta tgctgctaaa 1080
acctatgatt actataaagc caaatttggg cgcaacagca ttgacggacg cggctctgcaa 1140
cttcgttcga cggctcatta cggtagtcgc tacaacaatg ccttctggaa cggctcccaa 1200
atgacttatg gagatggaga tggtagcaca tttatcgctc tcagcgggga ccccgatgta 1260
gtaggacatg aacttacgca tgggtgcaca gagtatactt cgaatttggg atattacgga 1320
gagtcggcgc cattgaatga agctttctca gacgttatcg ggaatgacat tcagcgcaaa 1380
aactggcttg taggcgatga tatttacagc ccaaacattg caggcgatgc ccttcgctca 1440

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atgtccaatc caacctgta cgatcaacca gatcactatt ccaacctgta cagaggcagc 1500
tccgataacg gcggtgttca caccaacagc ggtattatca ataaagctta ctacttgтта 1560
gcacaagggtg gtaatttcca tggcgtaact gtaaatggaa ttggccgtga tgcagcggtg 1620
caaatctact acagtgcctt tacgaactac ctgacttctt cttccgactt ctccaacgca 1680
cgtgctgctg tgatccaagc cgcaaaagat ctgtacgggg cgaactcagc agaagcaact 1740
gcagctgcca agtcttttga cgctgtaggc gtaaactaa 1779

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<210> SEQ ID NO 27
<211> LENGTH: 592
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus polymyxa SC2
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(592)
<223> OTHER INFORMATION: amino acid sequence of the PpoPro1 precursor
protein

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```

<400> SEQUENCE: 27

```

```

Met Lys Lys Val Trp Val Ser Leu Leu Gly Gly Ala Met Leu Leu Gly
 1           5           10          15
Ser Val Ala Ser Gly Ala Ser Ala Glu Ser Ser Val Ser Gly Pro Ala
          20           25           30
Gln Leu Thr Pro Thr Phe His Ala Glu Gln Trp Lys Ala Pro Thr Ser
          35           40           45
Val Ser Gly Asp Asp Ile Val Trp Ser Tyr Leu Asn Arg Gln Lys Lys
          50           55           60
Ser Leu Leu Gly Val Asp Ser Ser Ser Val Arg Glu Gln Phe Arg Ile
          65           70           75           80
Val Asp Arg Thr Ser Asp Lys Ser Gly Val Ser His Tyr Arg Leu Lys
          85           90           95
Gln Tyr Val Asn Gly Ile Pro Val Tyr Gly Ala Glu Gln Thr Ile His
          100          105          110
Val Gly Lys Ser Gly Glu Val Thr Ser Tyr Leu Gly Ala Val Val Asn
          115          120          125
Glu Asp Gln Gln Ala Glu Ala Thr Gln Gly Thr Thr Pro Lys Ile Ser
          130          135          140
Ala Ser Glu Ala Val Tyr Thr Ala Tyr Lys Glu Ala Ala Ala Arg Ile
          145          150          155          160
Glu Ala Leu Pro Thr Ser Asp Asp Thr Ile Ser Lys Asp Ala Glu Glu
          165          170          175
Pro Ser Ser Val Ser Lys Asp Thr Tyr Ala Glu Ala Ala Asn Asn Glu
          180          185          190
Lys Thr Leu Ser Val Asp Lys Asp Glu Leu Ser Leu Asp Gln Ala Ser
          195          200          205
Val Leu Lys Asp Ser Lys Ile Glu Ala Val Glu Pro Glu Lys Ser Ser
          210          215          220
Ile Ala Lys Ile Ala Asn Leu Gln Pro Glu Val Asp Pro Lys Ala Glu
          225          230          235          240
Leu Tyr Tyr Tyr Pro Lys Gly Asp Asp Leu Leu Leu Val Tyr Val Thr
          245          250          255
Glu Val Asn Val Leu Glu Pro Ala Pro Leu Arg Thr Arg Tyr Ile Ile
          260          265          270

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Asp Ala Asn Asp Gly Ser Ile Val Phe Gln Tyr Asp Ile Ile Asn Glu  
 275 280 285

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Ser Lys Ser Phe Thr  
 290 295 300

Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly  
 305 310 315 320

Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile Pro Gly  
 325 330 335

Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Gly Val  
 340 345 350

Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys Ala Lys  
 355 360 365

Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr  
 370 375 380

Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln  
 385 390 395 400

Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Ile Ala Phe Ser Gly  
 405 410 415

Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr  
 420 425 430

Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala  
 435 440 445

Phe Ser Asp Val Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val  
 450 455 460

Gly Asp Asp Ile Tyr Thr Pro Asn Ile Ala Gly Asp Ala Leu Arg Ser  
 465 470 475 480

Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu  
 485 490 495

Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile  
 500 505 510

Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Asn Phe His Gly  
 515 520 525

Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr  
 530 535 540

Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser Asn Ala  
 545 550 555 560

Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser  
 565 570 575

Ala Glu Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn  
 580 585 590

<210> SEQ ID NO 28  
 <211> LENGTH: 304  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus polymyxa SC2  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(304)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature  
 form of PpoPro1

<400> SEQUENCE: 28

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Ser Lys Ser Phe Thr  
 1 5 10 15

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Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly  
 20 25 30  
 Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile Pro Gly  
 35 40 45  
 Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Gly Val  
 50 55 60  
 Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys Ala Lys  
 65 70 75 80  
 Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr  
 85 90 95  
 Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln  
 100 105 110  
 Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Ile Ala Phe Ser Gly  
 115 120 125  
 Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr  
 130 135 140  
 Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala  
 145 150 155 160  
 Phe Ser Asp Val Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val  
 165 170 175  
 Gly Asp Asp Ile Tyr Thr Pro Asn Ile Ala Gly Asp Ala Leu Arg Ser  
 180 185 190  
 Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu  
 195 200 205  
 Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile  
 210 215 220  
 Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Asn Phe His Gly  
 225 230 235 240  
 Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr  
 245 250 255  
 Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser Asn Ala  
 260 265 270  
 Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser  
 275 280 285  
 Ala Glu Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn  
 290 295 300

&lt;210&gt; SEQ ID NO 29

&lt;211&gt; LENGTH: 1800

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic: nucleotide sequence of the synthesized PpoProl gene in plasmid pGX138(AprE-PpoProl)

&lt;400&gt; SEQUENCE: 29

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg      60
gcgttcagca acatgagcgc gcaggctgct ggaaaagaat catcagtgtc aggaccggct      120
cagcttacac cgacatttca cgcagaacaa tggaaggctc cgacgtcagt ttcaggagac      180
gacatcgtgt ggagctacct gaatagacag aagaaaagcc tgctggggagt ggatagcagc      240
agcgtcagag agcagttcag aatcgttgac agaacgagcg acaaaaagcgg agtcagccat      300
tatagactga agcagtacgt gaatggcatc ccggtttatg gcgcagagca gacaattcat      360

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gttggaaga gcggagaagt cacaagctat ctgggcgctg tggccaatga agatcaacaa 420
gccgaggcta cacagggaac aacgcgaaa attagcgctt cagaggcagt ctacacggcg 480
tacaaagaag cggtgcaag aatcgaagcc ctgccgacat cagacgatac aatttcaaaa 540
gatgaggagg agccgagctc agttagcaag gatacatagc cggaagccgc aaacaatgag 600
aaaaactga gcgtggacaa ggacgagctg tcaactgacg aggctagcgt ccttaaagac 660
agcaagatcg aggccgttga gcctgaaaag tcatcaattg cgaataatcg caatctgcaa 720
cctgaagtgc acccgaaggc ggaactgtac tactaccega aaggcgatga cctgcttctg 780
gtgtacgtca cggaagtga cgtcctggaa cggcaccgc tgagaacaag atacatcatt 840
gacgcgaacg acggaagcat cgtcttccag tatgacatta tcaacgaagc aacgggaacg 900
ggcaaaggcg ttcttgagga ctcaaagagc ttcacgacaa cggttcagg aagcagctac 960
cagctgaaag acacgacgag aggaaacgga atcgtcacat atacggcgtc aaacagacaa 1020
agcatccctg gcacaatcct gacggatgct gacaacgttt ggaatgatcc ggctggcgtg 1080
gatgcccatg cttatcgccg aaaaacgtat gactattaca aggcgaagtt cggcagaaat 1140
tcaatcgatg gcagaggact gcagcttaga agcagcgtgc actacggatc aagatataac 1200
aatgccttct ggaacggcag ccagatgaca tacggagacg gagatggaag cacatttatt 1260
gcattcagcg gcgaccctga tgtggttggc catgagctga cgcattggct tacagaatat 1320
acgagcaatc ttgaatacta cggcgagtca ggcgctctga acgaggcatt tagcagtggt 1380
atcggaatg acatccagag aaaaaactgg ctggtgggcg acgatattta cagcctaatt 1440
atcgctggcg atgcccttag atcaatgtca aaccgacgc tgtatgatca gcctgaccac 1500
tactcaaacc tgtatagagg ctcatcagat aacggaggcg tccatacga tagcggcatt 1560
attaacaagc catattatct tctggcccag ggcggcaatt ttcattggag gacggttaat 1620
ggaattggaa gagacgcagc cgtccaaatc tactacagcg ctttcacgaa ctaccttaca 1680
tcaagctcag actttagcaa tgccagagct gctgttatcc aggcagcga ggatctttac 1740
ggcgccaact cagccgaagc tacggccgca gctaaatcat ttgatgcagt gggcgttaat 1800

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&lt;210&gt; SEQ ID NO 30

&lt;211&gt; LENGTH: 600

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Synthetic: amino acid sequence of the PpoPro1 precursor protein expressed from plasmid pGX138 (AprE-PpoPro1)

&lt;400&gt; SEQUENCE: 30

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10           15

Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20           25           30

Glu Ser Ser Val Ser Gly Pro Ala Gln Leu Thr Pro Thr Phe His Ala
35           40           45

Glu Gln Trp Lys Ala Pro Thr Ser Val Ser Gly Asp Asp Ile Val Trp
50           55           60

Ser Tyr Leu Asn Arg Gln Lys Lys Ser Leu Leu Gly Val Asp Ser Ser
65           70           75           80

Ser Val Arg Glu Gln Phe Arg Ile Val Asp Arg Thr Ser Asp Lys Ser

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| 85  |     |     |     | 90  |     |     |     | 95  |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Gly | Val | Ser | His | Tyr | Arg | Leu | Lys | Gln | Tyr | Val | Asn | Gly | Ile | Pro | Val |
|     | 100 |     |     |     |     |     |     |     | 105 |     |     |     | 110 |     |     |
| Tyr | Gly | Ala | Glu | Gln | Thr | Ile | His | Val | Gly | Lys | Ser | Gly | Glu | Val | Thr |
|     | 115 |     |     |     |     |     | 120 |     |     |     |     |     | 125 |     |     |
| Ser | Tyr | Leu | Gly | Ala | Val | Val | Asn | Glu | Asp | Gln | Gln | Ala | Glu | Ala | Thr |
|     | 130 |     |     |     |     |     | 135 |     |     |     |     |     | 140 |     |     |
| Gln | Gly | Thr | Thr | Pro | Lys | Ile | Ser | Ala | Ser | Glu | Ala | Val | Tyr | Thr | Ala |
|     | 145 |     |     |     | 150 |     |     |     |     | 155 |     |     |     |     | 160 |
| Tyr | Lys | Glu | Ala | Ala | Ala | Arg | Ile | Glu | Ala | Leu | Pro | Thr | Ser | Asp | Asp |
|     |     |     | 165 |     |     |     |     |     |     | 170 |     |     |     | 175 |     |
| Thr | Ile | Ser | Lys | Asp | Ala | Glu | Glu | Pro | Ser | Ser | Val | Ser | Lys | Asp | Thr |
|     |     | 180 |     |     |     |     |     |     | 185 |     |     |     | 190 |     |     |
| Tyr | Ala | Glu | Ala | Ala | Asn | Asn | Glu | Lys | Thr | Leu | Ser | Val | Asp | Lys | Asp |
|     | 195 |     |     |     |     |     | 200 |     |     |     |     |     | 205 |     |     |
| Glu | Leu | Ser | Leu | Asp | Gln | Ala | Ser | Val | Leu | Lys | Asp | Ser | Lys | Ile | Glu |
|     | 210 |     |     |     |     | 215 |     |     |     |     | 220 |     |     |     |     |
| Ala | Val | Glu | Pro | Glu | Lys | Ser | Ser | Ile | Ala | Lys | Ile | Ala | Asn | Leu | Gln |
|     | 225 |     |     |     | 230 |     |     |     |     | 235 |     |     |     |     | 240 |
| Pro | Glu | Val | Asp | Pro | Lys | Ala | Glu | Leu | Tyr | Tyr | Tyr | Pro | Lys | Gly | Asp |
|     |     |     | 245 |     |     |     |     |     | 250 |     |     |     |     | 255 |     |
| Asp | Leu | Leu | Leu | Val | Tyr | Val | Thr | Glu | Val | Asn | Val | Leu | Glu | Pro | Ala |
|     | 260 |     |     |     |     |     |     |     | 265 |     |     |     | 270 |     |     |
| Pro | Leu | Arg | Thr | Arg | Tyr | Ile | Ile | Asp | Ala | Asn | Asp | Gly | Ser | Ile | Val |
|     | 275 |     |     |     |     |     | 280 |     |     |     |     |     | 285 |     |     |
| Phe | Gln | Tyr | Asp | Ile | Ile | Asn | Glu | Ala | Thr | Gly | Thr | Gly | Lys | Gly | Val |
|     | 290 |     |     |     |     | 295 |     |     |     |     | 300 |     |     |     |     |
| Leu | Gly | Asp | Ser | Lys | Ser | Phe | Thr | Thr | Thr | Thr | Ala | Ser | Gly | Ser | Tyr |
|     | 305 |     |     |     | 310 |     |     |     |     |     | 315 |     |     |     | 320 |
| Gln | Leu | Lys | Asp | Thr | Thr | Arg | Gly | Asn | Gly | Ile | Val | Thr | Tyr | Thr | Ala |
|     |     |     | 325 |     |     |     |     |     | 330 |     |     |     |     |     | 335 |
| Ser | Asn | Arg | Gln | Ser | Ile | Pro | Gly | Thr | Ile | Leu | Thr | Asp | Ala | Asp | Asn |
|     |     |     | 340 |     |     |     |     |     | 345 |     |     |     |     | 350 |     |
| Val | Trp | Asn | Asp | Pro | Ala | Gly | Val | Asp | Ala | His | Ala | Tyr | Ala | Ala | Lys |
|     | 355 |     |     |     |     |     | 360 |     |     |     |     |     | 365 |     |     |
| Thr | Tyr | Asp | Tyr | Tyr | Lys | Ala | Lys | Phe | Gly | Arg | Asn | Ser | Ile | Asp | Gly |
|     | 370 |     |     |     |     | 375 |     |     |     |     | 380 |     |     |     |     |
| Arg | Gly | Leu | Gln | Leu | Arg | Ser | Thr | Val | His | Tyr | Gly | Ser | Arg | Tyr | Asn |
|     | 385 |     |     |     | 390 |     |     |     |     | 395 |     |     |     |     | 400 |
| Asn | Ala | Phe | Trp | Asn | Gly | Ser | Gln | Met | Thr | Tyr | Gly | Asp | Gly | Asp | Gly |
|     |     |     | 405 |     |     |     |     |     | 410 |     |     |     |     | 415 |     |
| Ser | Thr | Phe | Ile | Ala | Phe | Ser | Gly | Asp | Pro | Asp | Val | Val | Gly | His | Glu |
|     |     | 420 |     |     |     |     |     |     | 425 |     |     |     | 430 |     |     |
| Leu | Thr | His | Gly | Val | Thr | Glu | Tyr | Thr | Ser | Asn | Leu | Glu | Tyr | Tyr | Gly |
|     |     | 435 |     |     |     |     | 440 |     |     |     |     |     | 445 |     |     |
| Glu | Ser | Gly | Ala | Leu | Asn | Glu | Ala | Phe | Ser | Asp | Val | Ile | Gly | Asn | Asp |
|     | 450 |     |     |     |     | 455 |     |     |     |     | 460 |     |     |     |     |
| Ile | Gln | Arg | Lys | Asn | Trp | Leu | Val | Gly | Asp | Asp | Ile | Tyr | Thr | Pro | Asn |
|     | 465 |     |     |     | 470 |     |     |     |     | 475 |     |     |     |     | 480 |
| Ile | Ala | Gly | Asp | Ala | Leu | Arg | Ser | Met | Ser | Asn | Pro | Thr | Leu | Tyr | Asp |
|     |     |     | 485 |     |     |     |     |     |     | 490 |     |     |     |     | 495 |

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Gln Pro Asp His Tyr Ser Asn Leu Tyr Arg Gly Ser Ser Asp Asn Gly  
 500 505 510

Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu  
 515 520 525

Ala Gln Gly Gly Asn Phe His Gly Val Thr Val Asn Gly Ile Gly Arg  
 530 535 540

Asp Ala Ala Val Gln Ile Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu Thr  
 545 550 555 560

Ser Ser Ser Asp Phe Ser Asn Ala Arg Ala Ala Val Ile Gln Ala Ala  
 565 570 575

Lys Asp Leu Tyr Gly Ala Asn Ser Ala Glu Ala Thr Ala Ala Ala Lys  
 580 585 590

Ser Phe Asp Ala Val Gly Val Asn  
 595 600

<210> SEQ ID NO 31  
 <211> LENGTH: 1641  
 <212> TYPE: DNA  
 <213> ORGANISM: Paenibacillus hunanensis  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(1641)  
 <223> OTHER INFORMATION: nucleotide sequence of the PhuProl gene  
 isolated from Paenibacillus hunanensis

<400> SEQUENCE: 31

```

ttgaaaaaa cagttggtct ttacttgca ggtagcttgc tcgttggtgc tacaacgtcc 60
gctttcgcag cagaagcaaa tgatctggca ccaactcggg attacacgcc aaaattgatt 120
acgcaagcaa caggcatcac tggcgctagt ggcgatgcta aagtatggaa gttcctggag 180
aagcaaaaac gtaccatcgt aaccgatgat gcagcttctg ctgatgtgaa ggaattgttt 240
gagatcacia aacgtcaatc cgattctcaa accggtacag agcactatcg cctgaaccaa 300
acctttaaag gcatcccagt ctatggcgca gagcaaacac tgcacttga caaatccggc 360
aatgatctc tgtacatggg tcaggttgtt gaggatgtgt ccgctaaact ggaagcttcc 420
gattcaaaaa aaggcgtaac tgaggatgta tacgcttcgg atacgaaaaa tgatctggta 480
acaccagaaa tcagcgcttc tcaagccatc tcgattgctg aaaaggatgc agcttccaaa 540
atcggctccc tcggcggaag caaaaaaacg ccagaagcga agctgtatat ctacgctcct 600
gaggatcaag cagcactctt ggcttatgtg acagaagtaa acgtactgga gccatctcgg 660
ctgcgtactc gctatthtgt agatgcaaaa acaggttcga tcctgttcca atatgatctg 720
attgagcatg caacaggtag aggtaaaggg gtactgggtg ataccaagtc cttcactgta 780
ggtacttccg gttcttccca tgtgatgact gatagcacgc gtggaaaagg tatccaaacc 840
tacacggcgt ctaaccgcac atcactgcca ggtagcactg taacgagcag cagcagcaca 900
tttaacgatc cagcatctgt cgatgoccat gcgtatgcac aaaaagtata tgatttctac 960
aaatccaact ttaaccgcaa cagcatcgac ggtaatggtc tggctatccg ctccactacg 1020
cactattcca cacgtataaa caatgcgttc tggaatggtt cccaaatggg atacggtgat 1080
ggcgatgggt cgcaattcat cgcattctcc ggcgacctg acgtagtagg tcacgagctg 1140
acacacgggt taaccgagta cacagcgaac ctggaatact atggtcaatc cggtgactg 1200
aacgaatcca tttcggatat ctttggtaac acaatcgaag gtaaaaactg gatggtaggc 1260
    
```

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gatgcatct acacaccagg cgtatccggc gatgctcttc gctacatgga tgatccaaca 1320
aaaggtggac aaccagcgcg tatggcagat tacaacaaca caagcgctga taatggcggt 1380
gtacacacaa acagtgggat cccgaataaa gcatactact tgctggcaca ggggtggcaca 1440
tttggcggtg taatgtaac aggtatcggt cgctcgcaag cgatccagat cgtttaccgt 1500
gcactaacat actacctgac atccacatct aacttctcga actaccgttc tgcaatggtg 1560
caagcatcta cagacctgta cggtgcaaac tctacacaaa caacagcggt gaaaaactcg 1620
ctgagcgcag taggcattaa c 1641

```

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<210> SEQ ID NO 32
<211> LENGTH: 547
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus hunanensis
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(547)
<223> OTHER INFORMATION: amino acid sequence of the PhuPro1 precursor
protein

```

```

<400> SEQUENCE: 32

```

```

Met Lys Lys Thr Val Gly Leu Leu Leu Ala Gly Ser Leu Leu Val Gly
1           5           10          15
Ala Thr Thr Ser Ala Phe Ala Ala Glu Ala Asn Asp Leu Ala Pro Leu
20          25          30
Gly Asp Tyr Thr Pro Lys Leu Ile Thr Gln Ala Thr Gly Ile Thr Gly
35          40          45
Ala Ser Gly Asp Ala Lys Val Trp Lys Phe Leu Glu Lys Gln Lys Arg
50          55          60
Thr Ile Val Thr Asp Asp Ala Ala Ser Ala Asp Val Lys Glu Leu Phe
65          70          75          80
Glu Ile Thr Lys Arg Gln Ser Asp Ser Gln Thr Gly Thr Glu His Tyr
85          90          95
Arg Leu Asn Gln Thr Phe Lys Gly Ile Pro Val Tyr Gly Ala Glu Gln
100         105        110
Thr Leu His Phe Asp Lys Ser Gly Asn Val Ser Leu Tyr Met Gly Gln
115        120        125
Val Val Glu Asp Val Ser Ala Lys Leu Glu Ala Ser Asp Ser Lys Lys
130        135        140
Gly Val Thr Glu Asp Val Tyr Ala Ser Asp Thr Lys Asn Asp Leu Val
145        150        155        160
Thr Pro Glu Ile Ser Ala Ser Gln Ala Ile Ser Ile Ala Glu Lys Asp
165        170        175
Ala Ala Ser Lys Ile Gly Ser Leu Gly Glu Ala Gln Lys Thr Pro Glu
180        185        190
Ala Lys Leu Tyr Ile Tyr Ala Pro Glu Asp Gln Ala Ala Arg Leu Ala
195        200        205
Tyr Val Thr Glu Val Asn Val Leu Glu Pro Ser Pro Leu Arg Thr Arg
210        215        220
Tyr Phe Val Asp Ala Lys Thr Gly Ser Ile Leu Phe Gln Tyr Asp Leu
225        230        235        240
Ile Glu His Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys
245        250        255

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Ser Phe Thr Val Gly Thr Ser Gly Ser Ser Tyr Val Met Thr Asp Ser  
 260 265 270  
 Thr Arg Gly Lys Gly Ile Gln Thr Tyr Thr Ala Ser Asn Arg Thr Ser  
 275 280 285  
 Leu Pro Gly Ser Thr Val Thr Ser Ser Ser Ser Thr Phe Asn Asp Pro  
 290 295 300  
 Ala Ser Val Asp Ala His Ala Tyr Ala Gln Lys Val Tyr Asp Phe Tyr  
 305 310 315 320  
 Lys Ser Asn Phe Asn Arg Asn Ser Ile Asp Gly Asn Gly Leu Ala Ile  
 325 330 335  
 Arg Ser Thr Thr His Tyr Ser Thr Arg Tyr Asn Asn Ala Phe Trp Asn  
 340 345 350  
 Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly Ser Gln Phe Ile Ala  
 355 360 365  
 Phe Ser Gly Asp Leu Asp Val Val Gly His Glu Leu Thr His Gly Val  
 370 375 380  
 Thr Glu Tyr Thr Ala Asn Leu Glu Tyr Tyr Gly Gln Ser Gly Ala Leu  
 385 390 395 400  
 Asn Glu Ser Ile Ser Asp Ile Phe Gly Asn Thr Ile Glu Gly Lys Asn  
 405 410 415  
 Trp Met Val Gly Asp Ala Ile Tyr Thr Pro Gly Val Ser Gly Asp Ala  
 420 425 430  
 Leu Arg Tyr Met Asp Asp Pro Thr Lys Gly Gly Gln Pro Ala Arg Met  
 435 440 445  
 Ala Asp Tyr Asn Asn Thr Ser Ala Asp Asn Gly Gly Val His Thr Asn  
 450 455 460  
 Ser Gly Ile Pro Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr  
 465 470 475 480  
 Phe Gly Gly Val Asn Val Thr Gly Ile Gly Arg Ser Gln Ala Ile Gln  
 485 490 495  
 Ile Val Tyr Arg Ala Leu Thr Tyr Tyr Leu Thr Ser Thr Ser Asn Phe  
 500 505 510  
 Ser Asn Tyr Arg Ser Ala Met Val Gln Ala Ser Thr Asp Leu Tyr Gly  
 515 520 525  
 Ala Asn Ser Thr Gln Thr Thr Ala Val Lys Asn Ser Leu Ser Ala Val  
 530 535 540  
 Gly Ile Asn  
 545

<210> SEQ ID NO 33  
 <211> LENGTH: 304  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus hunanensis  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(304)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature  
 form of PhuProl

<400> SEQUENCE: 33

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Thr  
 1 5 10 15  
 Val Gly Thr Ser Gly Ser Ser Tyr Val Met Thr Asp Ser Thr Arg Gly  
 20 25 30



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Lys Gly Ile Gln Thr Tyr Thr Ala Ser Asn Arg Thr Ser Leu Pro Gly  
 35 40 45

Ser Thr Val Thr Ser Ser Ser Ser Thr Phe Asn Asp Pro Ala Ser Val  
 50 55 60

Asp Ala His Ala Tyr Ala Gln Lys Val Tyr Asp Phe Tyr Lys Ser Asn  
 65 70 75 80

Phe Asn Arg Asn Ser Ile Asp Gly Asn Gly Leu Ala Ile Arg Ser Thr  
 85 90 95

Thr His Tyr Ser Thr Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln  
 100 105 110

Met Val Tyr Gly Asp Gly Asp Gly Ser Gln Phe Ile Ala Phe Ser Gly  
 115 120 125

Asp Leu Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr  
 130 135 140

Thr Ala Asn Leu Glu Tyr Tyr Gly Gln Ser Gly Ala Leu Asn Glu Ser  
 145 150 155 160

Ile Ser Asp Ile Phe Gly Asn Thr Ile Glu Gly Lys Asn Trp Met Val  
 165 170 175

Gly Asp Ala Ile Tyr Thr Pro Gly Val Ser Gly Asp Ala Leu Arg Tyr  
 180 185 190

Met Asp Asp Pro Thr Lys Gly Gly Gln Pro Ala Arg Met Ala Asp Tyr  
 195 200 205

Asn Asn Thr Ser Ala Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile  
 210 215 220

Pro Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe Gly Gly  
 225 230 235 240

Val Asn Val Thr Gly Ile Gly Arg Ser Gln Ala Ile Gln Ile Val Tyr  
 245 250 255

Arg Ala Leu Thr Tyr Tyr Leu Thr Ser Thr Ser Asn Phe Ser Asn Tyr  
 260 265 270

Arg Ser Ala Met Val Gln Ala Ser Thr Asp Leu Tyr Gly Ala Asn Ser  
 275 280 285

Thr Gln Thr Thr Ala Val Lys Asn Ser Leu Ser Ala Val Gly Ile Asn  
 290 295 300

<210> SEQ ID NO 34  
 <211> LENGTH: 1671  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic: nucleotide sequence of the  
 synthesized PhuPro1 gene in plasmid pGX149(AprE- PhuPro1)

<400> SEQUENCE: 34

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg      60
gcgttcagca acatgagcgc gcaggctgct ggaaaagcag aagctaatga tcttgccccg      120
cttggcgatt atacaccgaa gcttattaca caggcaacgg gaattacagg cgcacacaggc      180
gatgcaaggt tgtggaagt cctggagaag cagaagagaa cgattgtcac ggacgacgcc      240
gcaagcgagg atgtcaagga gctgttcgag atcacgaaga gacagagcga tagccagacg      300
ggaacggagc attacagact gaaccagacg ttcaagggca ttccggctcta cggagctgaa      360
caaacgctgc attttgataa aagcggcaac gtctcactgt acatgggcca agtcgttgag      420
    
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gacgtagcg ccaaacttga ggctagcgac agcaagaaag gcgtcacaga agatgtctac 480
gcgtcagaca cgaaaaacga cctggttaca cggaaatct cagcttcaca ggccatctca 540
attgcagaga aagacgcagc gtcaaaaatc ggctcactgg gcgaggctca gaaaacgccc 600
gaggcgaaac ttacatcta cgcccctgag gaccaggctg cgagactggc ttacgtgaca 660
gaagttaatg tgctggagcc gtcacogctt agaacgagat atttctgga cgaaaagacg 720
ggcagcattc tgtttcagta cgatcttacc gaacacgcca caggcacagg aaagggagtt 780
ctgggagaca caaaaagctt cacggttggc acgtcaggca gcagctactg gatgacagac 840
agcacgagag gcaagggcat tcaaacgtat acagcgagca acagaacaag cctgccggga 900
agcacagtcg cgagctcacc atcaacgttt aatgaccggg cctcagtgga tgctcacgca 960
tacggcgaga aagtgtacga cttctacaaa agcaacttca atagaaacag catcgacgga 1020
aacggccttg cgatcagaag cacgacgcac tacagcaca gatacaaca cgccttctgg 1080
aacggcagcc aatggttta cggcgatggc gacggatcac agtttatcgc atttagcgga 1140
gacctggacg tcggtggcca tgagctgaca catggcgcta cggagtacac agcaaacctg 1200
gaatactatg gccagtcagg cgcccttaac gagagcatca gcgacatttt tggcaatagc 1260
atcgaaggaa agaactggat ggtcggcgac gcaatctaca caccgggctt ttcaggcgat 1320
gcaactgagat atatggacga cccgacaaa ggccggacagc cggccagaat ggcggattac 1380
aataaacgtg cagcagataa cggcgcgctg catacaata gcggcatccc taacaaagca 1440
tattacctgc ttgcgcaagg aggaacattt ggcggcgctg atgttacggg cattggcaga 1500
tcacaagcga ttcagatcgt ttacagagcg ctgacgtact accttacgag cacgagcaat 1560
tttagcaact acagaagcgc aatggtgcag gcaagcacgg atctgtatgg cgcaaatcca 1620
acacaaaacga cggcggtcaa gaatagcctt tcagcagtgg gcattaacta a 1671

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&lt;210&gt; SEQ ID NO 35

&lt;211&gt; LENGTH: 556

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

<223> OTHER INFORMATION: Synthetic: amino acid sequence of the PhuPro1 precursor protein expressed from plasmid pGX149(AprE- PhuPro1)

&lt;400&gt; SEQUENCE: 35

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1           5           10          15
Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20          25          30
Ala Glu Ala Asn Asp Leu Ala Pro Leu Gly Asp Tyr Thr Pro Lys Leu
35          40          45
Ile Thr Gln Ala Thr Gly Ile Thr Gly Ala Ser Gly Asp Ala Lys Val
50          55          60
Trp Lys Phe Leu Glu Lys Gln Lys Arg Thr Ile Val Thr Asp Asp Ala
65          70          75          80
Ala Ser Ala Asp Val Lys Glu Leu Phe Glu Ile Thr Lys Arg Gln Ser
85          90          95
Asp Ser Gln Thr Gly Thr Glu His Tyr Arg Leu Asn Gln Thr Phe Lys
100         105         110
Gly Ile Pro Val Tyr Gly Ala Glu Gln Thr Leu His Phe Asp Lys Ser
115         120         125

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Gly Asn Val Ser Leu Tyr Met Gly Gln Val Val Glu Asp Val Ser Ala  
 130 135 140

Lys Leu Glu Ala Ser Asp Ser Lys Lys Gly Val Thr Glu Asp Val Tyr  
 145 150 155 160

Ala Ser Asp Thr Lys Asn Asp Leu Val Thr Pro Glu Ile Ser Ala Ser  
 165 170 175

Gln Ala Ile Ser Ile Ala Glu Lys Asp Ala Ala Ser Lys Ile Gly Ser  
 180 185 190

Leu Gly Glu Ala Gln Lys Thr Pro Glu Ala Lys Leu Tyr Ile Tyr Ala  
 195 200 205

Pro Glu Asp Gln Ala Ala Arg Leu Ala Tyr Val Thr Glu Val Asn Val  
 210 215 220

Leu Glu Pro Ser Pro Leu Arg Thr Arg Tyr Phe Val Asp Ala Lys Thr  
 225 230 235 240

Gly Ser Ile Leu Phe Gln Tyr Asp Leu Ile Glu His Ala Thr Gly Thr  
 245 250 255

Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Thr Val Gly Thr Ser  
 260 265 270

Gly Ser Ser Tyr Val Met Thr Asp Ser Thr Arg Gly Lys Gly Ile Gln  
 275 280 285

Thr Tyr Thr Ala Ser Asn Arg Thr Ser Leu Pro Gly Ser Thr Val Thr  
 290 295 300

Ser Ser Ser Ser Thr Phe Asn Asp Pro Ala Ser Val Asp Ala His Ala  
 305 310 315 320

Tyr Ala Gln Lys Val Tyr Asp Phe Tyr Lys Ser Asn Phe Asn Arg Asn  
 325 330 335

Ser Ile Asp Gly Asn Gly Leu Ala Ile Arg Ser Thr Thr His Tyr Ser  
 340 345 350

Thr Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly  
 355 360 365

Asp Gly Asp Gly Ser Gln Phe Ile Ala Phe Ser Gly Asp Leu Asp Val  
 370 375 380

Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr Thr Ala Asn Leu  
 385 390 395 400

Glu Tyr Tyr Gly Gln Ser Gly Ala Leu Asn Glu Ser Ile Ser Asp Ile  
 405 410 415

Phe Gly Asn Thr Ile Glu Gly Lys Asn Trp Met Val Gly Asp Ala Ile  
 420 425 430

Tyr Thr Pro Gly Val Ser Gly Asp Ala Leu Arg Tyr Met Asp Asp Pro  
 435 440 445

Thr Lys Gly Gly Gln Pro Ala Arg Met Ala Asp Tyr Asn Asn Thr Ser  
 450 455 460

Ala Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Pro Asn Lys Ala  
 465 470 475 480

Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe Gly Gly Val Asn Val Thr  
 485 490 495

Gly Ile Gly Arg Ser Gln Ala Ile Gln Ile Val Tyr Arg Ala Leu Thr  
 500 505 510

Tyr Tyr Leu Thr Ser Thr Ser Asn Phe Ser Asn Tyr Arg Ser Ala Met  
 515 520 525

-continued

Val Gln Ala Ser Thr Asp Leu Tyr Gly Ala Asn Ser Thr Gln Thr Thr  
 530 535 540

Ala Val Lys Asn Ser Leu Ser Ala Val Gly Ile Asn  
 545 550 555

<210> SEQ ID NO 36  
 <211> LENGTH: 1563  
 <212> TYPE: DNA  
 <213> ORGANISM: Paenibacillus amylolyticus  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(1563)  
 <223> OTHER INFORMATION: nucleotide sequence of the PamPro1 gene  
 isolated from Paenibacillus amylolyticus

<400> SEQUENCE: 36

```

atgaaattcg ccaaagttaa gccacaatt cttggaggag ctcttttctg cgcttcgta 60
tcctctgcta ctgcagctcc agtgtctgat caatccattc cacttcaggc cccttatgcc 120
tctgaggggg gtattccatt gaacagtgga acagatgaca ctatctttaa ttatcttga 180
cagcagggaac aatttctgaa ttccgatgtg aaatcccagc tcaaaattgt caaaagaaac 240
acagatacat ctggcgtaag aaccttccgc ctgaaacagt atattaaagg tatcccgggt 300
tatggtgtag aacagacggt ccacctggac aaaaccggag ccgtgagctc cgcacttggc 360
gatcttccac cgattgaaga gcaggccatt ccgaatgatg gtgtagccga gatcagcgga 420
gaagacgcga tccagattgc aaccgaagaa gcaacctccc ggattggaga gcttgggtgcc 480
gcggaatca cgctcaagc tgaattgaac atctatcatc atgaagaaga tggtcagaca 540
tatctgggtt acattacgga agtaaacgta ctggaacctg ccctctacg gaccaaatat 600
ttcattaacg cagtggatgg cagtatccta tcccagtttg acctcattaa ctctgctact 660
ggaacaggta cagggtgact cgggtgatac aaaaccctga caaccacca atccggtcagc 720
accttccaac tgaagacac cactcgtggc aatggcatcc aaacgtatac ggcaaacaaat 780
ggctcctcac tgctggttag cttgcttaca gattcggata atgtatggac cgatcgtgca 840
ggtgtagatg ctcatgctca tgccgctgct acgtatgatt tctacaaaaa caaattcaac 900
cgtaacggta ttaatggtaa cggattgttg atcagatcaa ccgtgcacta cggctccaat 960
tacaataacg ccttctgtaa cggggcacag attgtctttg gtgacggaga tggaacgatg 1020
ttccgatccc tgtctggtga tctggatggt gtgggtcatg aattgacgca tgggtttatt 1080
gaatatacag ccaatctgga atatcgcaat gaaccagggt cactcaatga agcctttgce 1140
gatattttcg gtaatacgat ccaaagcaaa aactggctgc tcggtgatga tatctacaca 1200
cctaacactc caggagatgc gctgctgccc ctctccaacc ctacattgta tggtaaacct 1260
gacaaataca gcgatcgcta cacaggctca caggacaacg gcggtgtcca tatcaacagt 1320
ggtatcatca ataaagccta tttccttgct gctcaaggcg gaacacataa tgggtgtgact 1380
gttaccggaa tcggccggga taaagcagtc cagatcttct acagcacact ggtgaactac 1440
ctgacaccaa cgtccaat tgcgctgccc aaaacagcta ccattcaagc agccaaagat 1500
ctgtacggag caacttccgc tgaagctact gctattacca aagcatatca agctgtaggc 1560
ctg 1563
    
```

<210> SEQ ID NO 37  
 <211> LENGTH: 521

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<212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus amylolyticus  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(521)  
 <223> OTHER INFORMATION: amino acid sequence of the PamProI precursor protein

<400> SEQUENCE: 37

```

Met Lys Phe Ala Lys Val Met Pro Thr Ile Leu Gly Gly Ala Leu Leu
 1           5           10          15
Leu Ala Ser Val Ser Ser Ala Thr Ala Ala Pro Val Ser Asp Gln Ser
          20          25          30
Ile Pro Leu Gln Ala Pro Tyr Ala Ser Glu Gly Gly Ile Pro Leu Asn
          35          40          45
Ser Gly Thr Asp Asp Thr Ile Phe Asn Tyr Leu Gly Gln Gln Glu Gln
          50          55          60
Phe Leu Asn Ser Asp Val Lys Ser Gln Leu Lys Ile Val Lys Arg Asn
 65          70          75          80
Thr Asp Thr Ser Gly Val Arg His Phe Arg Leu Lys Gln Tyr Ile Lys
          85          90          95
Gly Ile Pro Val Tyr Gly Ala Glu Gln Thr Val His Leu Asp Lys Thr
          100         105         110
Gly Ala Val Ser Ser Ala Leu Gly Asp Leu Pro Pro Ile Glu Glu Gln
          115         120         125
Ala Ile Pro Asn Asp Gly Val Ala Glu Ile Ser Gly Glu Asp Ala Ile
          130         135         140
Gln Ile Ala Thr Glu Glu Ala Thr Ser Arg Ile Gly Glu Leu Gly Ala
          145         150         155         160
Ala Glu Ile Thr Pro Gln Ala Glu Leu Asn Ile Tyr His His Glu Glu
          165         170         175
Asp Gly Gln Thr Tyr Leu Val Tyr Ile Thr Glu Val Asn Val Leu Glu
          180         185         190
Pro Ala Pro Leu Arg Thr Lys Tyr Phe Ile Asn Ala Val Asp Gly Ser
          195         200         205
Ile Val Ser Gln Phe Asp Leu Ile Asn Phe Ala Thr Gly Thr Gly Thr
          210         215         220
Gly Val Leu Gly Asp Thr Lys Thr Leu Thr Thr Thr Gln Ser Gly Ser
          225         230         235         240
Thr Phe Gln Leu Lys Asp Thr Thr Arg Gly Asn Gly Ile Gln Thr Tyr
          245         250         255
Thr Ala Asn Asn Gly Ser Ser Leu Pro Gly Ser Leu Leu Thr Asp Ser
          260         265         270
Asp Asn Val Trp Thr Asp Arg Ala Gly Val Asp Ala His Ala His Ala
          275         280         285
Ala Ala Thr Tyr Asp Phe Tyr Lys Asn Lys Phe Asn Arg Asn Gly Ile
          290         295         300
Asn Gly Asn Gly Leu Leu Ile Arg Ser Thr Val His Tyr Gly Ser Asn
          305         310         315         320
Tyr Asn Asn Ala Phe Trp Asn Gly Ala Gln Ile Val Phe Gly Asp Gly
          325         330         335
Asp Gly Thr Met Phe Arg Ser Leu Ser Gly Asp Leu Asp Val Val Gly
          340         345         350

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His Glu Leu Thr His Gly Val Ile Glu Tyr Thr Ala Asn Leu Glu Tyr  
 355 360 365  
 Arg Asn Glu Pro Gly Ala Leu Asn Glu Ala Phe Ala Asp Ile Phe Gly  
 370 375 380  
 Asn Thr Ile Gln Ser Lys Asn Trp Leu Leu Gly Asp Asp Ile Tyr Thr  
 385 390 395 400  
 Pro Asn Thr Pro Gly Asp Ala Leu Arg Ser Leu Ser Asn Pro Thr Leu  
 405 410 415  
 Tyr Gly Gln Pro Asp Lys Tyr Ser Asp Arg Tyr Thr Gly Ser Gln Asp  
 420 425 430  
 Asn Gly Gly Val His Ile Asn Ser Gly Ile Ile Asn Lys Ala Tyr Phe  
 435 440 445  
 Leu Ala Ala Gln Gly Gly Thr His Asn Gly Val Thr Val Thr Gly Ile  
 450 455 460  
 Gly Arg Asp Lys Ala Ile Gln Ile Phe Tyr Ser Thr Leu Val Asn Tyr  
 465 470 475 480  
 Leu Thr Pro Thr Ser Lys Phe Ala Ala Ala Lys Thr Ala Thr Ile Gln  
 485 490 495  
 Ala Ala Lys Asp Leu Tyr Gly Ala Thr Ser Ala Glu Ala Thr Ala Ile  
 500 505 510  
 Thr Lys Ala Tyr Gln Ala Val Gly Leu  
 515 520

<210> SEQ ID NO 38  
 <211> LENGTH: 303  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus amylolyticus  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(303)  
 <223> OTHER INFORMATION: amino acid sequence of the predicted mature  
 form of PamProl

<400> SEQUENCE: 38

Ala Thr Gly Thr Gly Thr Gly Val Leu Gly Asp Thr Lys Thr Leu Thr  
 1 5 10 15  
 Thr Thr Gln Ser Gly Ser Thr Phe Gln Leu Lys Asp Thr Thr Arg Gly  
 20 25 30  
 Asn Gly Ile Gln Thr Tyr Thr Ala Asn Asn Gly Ser Ser Leu Pro Gly  
 35 40 45  
 Ser Leu Leu Thr Asp Ser Asp Asn Val Trp Thr Asp Arg Ala Gly Val  
 50 55 60  
 Asp Ala His Ala His Ala Ala Ala Thr Tyr Asp Phe Tyr Lys Asn Lys  
 65 70 75 80  
 Phe Asn Arg Asn Gly Ile Asn Gly Asn Gly Leu Leu Ile Arg Ser Thr  
 85 90 95  
 Val His Tyr Gly Ser Asn Tyr Asn Asn Ala Phe Trp Asn Gly Ala Gln  
 100 105 110  
 Ile Val Phe Gly Asp Gly Asp Gly Thr Met Phe Arg Ser Leu Ser Gly  
 115 120 125  
 Asp Leu Asp Val Val Gly His Glu Leu Thr His Gly Val Ile Glu Tyr  
 130 135 140  
 Thr Ala Asn Leu Glu Tyr Arg Asn Glu Pro Gly Ala Leu Asn Glu Ala  
 145 150 155 160

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Phe Ala Asp Ile Phe Gly Asn Thr Ile Gln Ser Lys Asn Trp Leu Leu  
 165 170 175

Gly Asp Asp Ile Tyr Thr Pro Asn Thr Pro Gly Asp Ala Leu Arg Ser  
 180 185 190

Leu Ser Asn Pro Thr Leu Tyr Gly Gln Pro Asp Lys Tyr Ser Asp Arg  
 195 200 205

Tyr Thr Gly Ser Gln Asp Asn Gly Gly Val His Ile Asn Ser Gly Ile  
 210 215 220

Ile Asn Lys Ala Tyr Phe Leu Ala Ala Gln Gly Gly Thr His Asn Gly  
 225 230 235 240

Val Thr Val Thr Gly Ile Gly Arg Asp Lys Ala Ile Gln Ile Phe Tyr  
 245 250 255

Ser Thr Leu Val Asn Tyr Leu Thr Pro Thr Ser Lys Phe Ala Ala Ala  
 260 265 270

Lys Thr Ala Thr Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Thr Ser  
 275 280 285

Ala Glu Ala Thr Ala Ile Thr Lys Ala Tyr Gln Ala Val Gly Leu  
 290 295 300

<210> SEQ ID NO 39  
 <211> LENGTH: 1587  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic; nucleotide sequence of the  
 synthesized PamProl gene in plasmid pGX146(AprE- PamProl)

<400> SEQUENCE: 39

```

gtgagaagca aaaaattgtg gatcagcttg ttgtttgcgt taacgttaat ctttacgatg      60
gcgttcagca acatgagcgc gcaggctgct ggaaaagctc cggttagcga ccagtcaatc     120
cctottcaag caccgtatgc cagcgaagga ggcattccgc ttaacagcgg cacggacgac     180
acgattttca attacctggg ccaacaggag cagttcctga acagcgacgt caagagccag     240
ctgaagatcg tcaaaagaaa cacagacaca tcaggcgtga gacacttcag actgaagcaa     300
tacatcaagg gcatcccggg ttatggcgct gaacaaacgg ttcacctgga caaacaggc     360
gcagtttcat cagcactggg agatctgccc cggattgaag agcaagcaat cccgaatgat     420
ggagttgceg aaattagcgg cgaggatgca atccaaatcg cgacggagga ggctacatca     480
agaattggag aacttggcgc agcggagatt acaccgcagg ctgaactgaa catctatcac     540
catgaggaag acggccagac gtacctggtt tacattacgg aagtgaacgt gctggaaccg     600
gcacctctga gaacaaagta ctttatcaac gcggttgacg gcagcatcgt ctcacagttc     660
gacctgatta acttcgccac gggaacagga acgggcgctt ttggagacac aaagacgctg     720
acgacgacgc agtcaggcag cacattccag ctgaaggaca caacaagagg caacggcatc     780
caaacgtaca cggcgaacaa tggatcatca ctgccgggct cactgctgac ggattcagat     840
aacgtgtgga cggatagagc tggcgttgac gcgcatgctc acgctgctgc gacgtacgac     900
ttctacaaga acaagttcaa cagaaacggc attaacggaa atggcctgct gatcagaagc     960
acggtgcatt atggctcaaa ctacaacaac gctttttgga acggcgcaca gatcgtgttt    1020
ggcgacggcg atggcacaat gtttagaagc ctgtcaggag acctggatgt ggtgggccac    1080
gaactgacgc acggcgtgat cgagtatacg gcgaaccttg aatatagaaa cgagccggga    1140
    
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gcactgaatg aggcgttcgc ggacatttcc ggcaacacaaa tccagagcaa aaactggctg 1200
ctgggcgacg atatctatac accgaacaca cggggcgatg cactgagatc actgtcaaat 1260
ccgacgctgt atggccaacc ggataagtac tcagacagat atacgggcag ccaagacaat 1320
ggcggcgcttc acatcaactc aggcacatc aacaaggctt acttccttgc ggcccaagga 1380
ggaacacata acggcgcttac agttacagc attggcagag acaaggcgat ccagatcttt 1440
tacagcagc tggatgaacta cctgacacct acgtcaaagt ttgccgcagc gaaaacagca 1500
acaattcagg cggctaaaga cctgtacgga ggcacatcag ccgaggccac agcaattaca 1560
aaagcatatc aagcagttgg cctttaa 1587

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&lt;210&gt; SEQ ID NO 40

&lt;211&gt; LENGTH: 528

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: Synthetic: amino acid sequence of the PamPro1 precursor protein expressed from plasmid pGX146 (AprE- PamPro1)

&lt;400&gt; SEQUENCE: 40

```

Met Arg Ser Lys Lys Leu Trp Ile Ser Leu Leu Phe Ala Leu Thr Leu
1          5          10          15
Ile Phe Thr Met Ala Phe Ser Asn Met Ser Ala Gln Ala Ala Gly Lys
20          25          30
Ala Pro Val Ser Asp Gln Ser Ile Pro Leu Gln Ala Pro Tyr Ala Ser
35          40          45
Glu Gly Gly Ile Pro Leu Asn Ser Gly Thr Asp Asp Thr Ile Phe Asn
50          55          60
Tyr Leu Gly Gln Gln Glu Gln Phe Leu Asn Ser Asp Val Lys Ser Gln
65          70          75          80
Leu Lys Ile Val Lys Arg Asn Thr Asp Thr Ser Gly Val Arg His Phe
85          90          95
Arg Leu Lys Gln Tyr Ile Lys Gly Ile Pro Val Tyr Gly Ala Glu Gln
100         105         110
Thr Val His Leu Asp Lys Thr Gly Ala Val Ser Ser Ala Leu Gly Asp
115         120         125
Leu Pro Pro Ile Glu Glu Gln Ala Ile Pro Asn Asp Gly Val Ala Glu
130         135         140
Ile Ser Gly Glu Asp Ala Ile Gln Ile Ala Thr Glu Glu Ala Thr Ser
145         150         155         160
Arg Ile Gly Glu Leu Gly Ala Ala Glu Ile Thr Pro Gln Ala Glu Leu
165         170         175
Asn Ile Tyr His His Glu Glu Asp Gly Gln Thr Tyr Leu Val Tyr Ile
180         185         190
Thr Glu Val Asn Val Leu Glu Pro Ala Pro Leu Arg Thr Lys Tyr Phe
195         200         205
Ile Asn Ala Val Asp Gly Ser Ile Val Ser Gln Phe Asp Leu Ile Asn
210         215         220
Phe Ala Thr Gly Thr Gly Thr Gly Val Leu Gly Asp Thr Lys Thr Leu
225         230         235         240
Thr Thr Thr Gln Ser Gly Ser Thr Phe Gln Leu Lys Asp Thr Thr Arg
245         250         255
Gly Asn Gly Ile Gln Thr Tyr Thr Ala Asn Asn Gly Ser Ser Leu Pro

```





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<222> LOCATION: (3)..(4)  
 <223> OTHER INFORMATION: Xaa can be any naturally occurring amino acid

<400> SEQUENCE: 42

His Asp Xaa Xaa His  
 1 5

<210> SEQ ID NO 43  
 <211> LENGTH: 4  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Synthetic peptide  
 <220> FEATURE:  
 <221> NAME/KEY: MOD\_RES  
 <222> LOCATION: (1)..(1)  
 <223> OTHER INFORMATION: Succinyl at 5'-end  
 <220> FEATURE:  
 <221> NAME/KEY: MOD\_RES  
 <222> LOCATION: (4)..(4)  
 <223> OTHER INFORMATION: Para nitroanilide (pNA) at 3'-end

<400> SEQUENCE: 43

Ala Ala Pro Phe  
 1

<210> SEQ ID NO 44  
 <211> LENGTH: 306  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus sp.  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(306)  
 <223> OTHER INFORMATION: Paenibacillus\_sp\_Aloe-11

<400> SEQUENCE: 44

Asn Glu Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Thr  
 1 5 10 15  
 Phe Asn Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Arg Asp Thr Thr  
 20 25 30  
 Arg Gly Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile  
 35 40 45  
 Pro Gly Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala  
 50 55 60  
 Gly Val Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys  
 65 70 75 80  
 Glu Lys Phe Asn Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg  
 85 90 95  
 Ser Thr Val His Tyr Gly Asn Arg Tyr Asn Asn Ala Phe Trp Asn Gly  
 100 105 110  
 Ser Gln Met Thr Tyr Gly Asp Gly Asp Gly Thr Thr Phe Ile Ala Phe  
 115 120 125  
 Ser Gly Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr  
 130 135 140  
 Glu Tyr Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn  
 145 150 155 160  
 Glu Ala Phe Ser Asp Ile Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp  
 165 170 175  
 Leu Val Gly Asp Asp Ile Tyr Thr Pro Arg Ile Ala Gly Asp Ala Leu  
 180 185 190

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Arg Ser Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser
 195                200                205

Asn Leu Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser
 210                215                220

Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe
 225                230                235                240

His Gly Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile
                245                250                255

Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser
                260                265                270

Asn Ala Arg Asp Ala Val Val Gln Ala Ala Lys Asp Leu Tyr Gly Ala
                275                280                285

Ser Ser Ala Gln Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly
 290                295                300

Val Asn
 305

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<210> SEQ ID NO 45
<211> LENGTH: 316
<212> TYPE: PRT
<213> ORGANISM: B. thermoproteolyticus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(316)
<223> OTHER INFORMATION: B.thermoproteolyticus_P00800

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<400> SEQUENCE: 45

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Ile Thr Gly Thr Ser Thr Val Gly Val Gly Arg Gly Val Leu Gly Asp
 1                5                10                15

Gln Lys Asn Ile Asn Thr Thr Tyr Ser Thr Tyr Tyr Tyr Leu Gln Asp
                20                25                30

Asn Thr Arg Gly Asn Gly Ile Phe Thr Tyr Asp Ala Lys Tyr Arg Thr
 35                40                45

Thr Leu Pro Gly Ser Leu Trp Ala Asp Ala Asp Asn Gln Phe Phe Ala
 50                55                60

Ser Tyr Asp Ala Pro Ala Val Asp Ala His Tyr Tyr Ala Gly Val Thr
 65                70                75                80

Tyr Asp Tyr Tyr Lys Asn Val His Asn Arg Leu Ser Tyr Asp Gly Asn
                85                90                95

Asn Ala Ala Ile Arg Ser Ser Val His Tyr Ser Gln Gly Tyr Asn Asn
 100                105                110

Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly Gln
 115                120                125

Thr Phe Ile Pro Leu Ser Gly Gly Ile Asp Val Val Ala His Glu Leu
 130                135                140

Thr His Ala Val Thr Asp Tyr Thr Ala Gly Leu Ile Tyr Gln Asn Glu
 145                150                155                160

Ser Gly Ala Ile Asn Glu Ala Ile Ser Asp Ile Phe Gly Thr Leu Val
                165                170                175

Glu Phe Tyr Ala Asn Lys Asn Pro Asp Trp Glu Ile Gly Glu Asp Val
 180                185                190

Tyr Thr Pro Gly Ile Ser Gly Asp Ser Leu Arg Ser Met Ser Asp Pro
 195                200                205

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Ala Lys Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly Thr  
210 215 220

Gln Asp Asn Gly Gly Val His Ile Asn Ser Gly Ile Ile Asn Lys Ala  
225 230 235 240

Ala Tyr Leu Ile Ser Gln Gly Gly Thr His Tyr Gly Val Ser Val Val  
245 250 255

Gly Ile Gly Arg Asp Lys Leu Gly Lys Ile Phe Tyr Arg Ala Leu Thr  
260 265 270

Gln Tyr Leu Thr Pro Thr Ser Asn Phe Ser Gln Leu Arg Ala Ala Ala  
275 280 285

Val Gln Ser Ala Thr Asp Leu Tyr Gly Ser Thr Ser Gln Glu Val Ala  
290 295 300

Ser Val Lys Gln Ala Phe Asp Ala Val Gly Val Lys  
305 310 315

<210> SEQ ID NO 46  
 <211> LENGTH: 306  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus sp. Aloe-11  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(306)  
 <223> OTHER INFORMATION: ZP\_09775365.1\_P\_sp\_Aloe-11

<400> SEQUENCE: 46

Ala Thr Gly Thr Gly Arg Gly Val Asp Gly Lys Thr Lys Ser Phe Thr  
1 5 10 15

Thr Thr Ala Ser Gly Asn Arg Tyr Gln Leu Lys Asp Thr Thr Arg Ser  
20 25 30

Asn Gly Ile Val Thr Tyr Thr Ala Gly Asn Arg Gln Thr Thr Pro Gly  
35 40 45

Thr Ile Leu Thr Asp Thr Asp Asn Val Trp Glu Asp Pro Ala Ala Val  
50 55 60

Asp Ala His Ala Tyr Ala Ile Lys Thr Tyr Asp Tyr Tyr Lys Asn Lys  
65 70 75 80

Phe Gly Arg Asp Ser Ile Asp Gly Arg Gly Met Gln Ile Arg Ser Thr  
85 90 95

Val His Tyr Gly Lys Lys Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln  
100 105 110

Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Thr Phe Phe Ser Gly  
115 120 125

Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Phe  
130 135 140

Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala  
145 150 155 160

Phe Ser Asp Ile Ile Gly Asn Asp Ile Asp Gly Thr Ser Trp Leu Leu  
165 170 175

Gly Asp Gly Ile Tyr Thr Pro Asn Ile Pro Gly Asp Ala Leu Arg Ser  
180 185 190

Leu Ser Asp Pro Thr Arg Phe Gly Gln Pro Asp His Tyr Ser Asn Phe  
195 200 205

Tyr Pro Asp Pro Asn Asn Asp Asp Glu Gly Gly Val His Thr Asn Ser  
210 215 220

Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Ser

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225                230                235                240
His Gly Val Thr Val Thr Gly Ile Gly Arg Glu Ala Ala Val Phe Ile
                245                250                255

Tyr Tyr Asn Ala Phe Thr Asn Tyr Leu Thr Ser Thr Ser Asn Phe Ser
                260                265                270

Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Phe Tyr Gly Ala
                275                280                285

Asp Ser Leu Ala Val Thr Ser Ala Ile Gln Ser Phe Asp Ala Val Gly
                290                295                300

Ile Lys
305

<210> SEQ ID NO 47
<211> LENGTH: 304
<212> TYPE: PRT
<213> ORGANISM: P. terrae
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(304)
<223> OTHER INFORMATION: P_terrae_HPL-003_YP_005073223.

<400> SEQUENCE: 47
Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Asn
1      5      10      15
Thr Thr Gln Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly
20     25     30
Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Thr Ile Pro Gly
35     40     45
Thr Leu Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Gly Val
50     55     60
Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys Asp Lys
65     70     75     80
Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr
85     90     95
Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln
100    105   110
Met Thr Tyr Gly Asp Gly Asp Gly Thr Thr Phe Ile Ala Phe Ser Gly
115    120   125
Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr
130    135   140
Thr Ser Asn Leu Asp Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ser
145    150   155   160
Phe Ser Asp Ile Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val
165    170   175
Gly Asp Asp Ile Tyr Thr Pro Ser Ile Ala Gly Asp Ala Leu Arg Ser
180    185   190
Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu
195    200   205
Tyr Lys Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile
210    215   220
Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe His Asn
225    230   235   240
Val Thr Val Ser Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr
245    250   255

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Ser Ala Phe Thr Asn Tyr Leu Thr Ser Thr Ser Asn Phe Ser Asn Thr
      260                      265                      270

Arg Ala Ala Val Val Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser
      275                      280                      285

Ala Gln Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn
      290                      295                      300

<210> SEQ ID NO 48
<211> LENGTH: 301
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus elgii
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(301)
<223> OTHER INFORMATION: Paenibacillus_elgii_B69_ZP_090

<400> SEQUENCE: 48

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Thr
 1                      5                      10                      15

Thr Thr Gln Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly
      20                      25                      30

Gln Gly Ile Val Thr Tyr Ser Ala Gly Asn Arg Thr Ser Leu Pro Gly
 35                      40                      45

Ser Leu Leu Thr Ser Thr Asn Asn Ile Trp Asn Asp Gly Ser Ala Val
 50                      55                      60

Asp Ala His Ala Tyr Thr Gly Lys Val Tyr Asp Tyr Tyr Lys Asn Lys
 65                      70                      75                      80

Phe Gly Arg Asn Ser Ile Asp Gly Asn Gly Leu Gln Leu Lys Ser Thr
      85                      90                      95

Val His Tyr Ser Thr Arg Tyr Asn Asn Ala Phe Trp Asn Gly Val Gln
 100                      105                      110

Met Val Tyr Gly Asp Gly Asp Gly Val Thr Phe Arg Ser Phe Pro Ala
 115                      120                      125

Asp Pro Asp Val Ile Gly His Glu Leu Thr His Gly Val Thr Glu Ser
 130                      135                      140

Thr Ala Gly Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ser
 145                      150                      155                      160

Ile Ser Asp Ile Phe Gly Asn Ala Ile Glu Gly Lys Asn Trp Leu Ile
      165                      170                      175

Gly Asp Leu Ile Thr Leu Asn Ala Gly Ala Leu Arg Ser Met Glu Asn
 180                      185                      190

Pro Lys Leu Tyr Arg Gln Pro Asp Arg Tyr Gln Asp Arg Tyr Thr Gly
 195                      200                      205

Pro Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Asn Asn Lys
 210                      215                      220

Ala Phe His Leu Ile Ala Gln Gly Gly Thr His Tyr Gly Val Thr Val
 225                      230                      235                      240

Asn Gly Ile Gly Arg Ser Ala Ala Glu Gln Ile Phe Tyr Asp Ala Leu
      245                      250                      255

Thr His Tyr Leu Thr Pro Thr Ser Asn Phe Ser Ala Ile Arg Ala Ala
 260                      265                      270

Ala Ile Gln Ala Ala Thr Asp Ser Phe Gly Ala Asn Ser Ser Gln Val
 275                      280                      285

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Asp Ala Val Lys Lys Ala Tyr Asn Ala Val Gly Val Asn
 290                295                300

<210> SEQ ID NO 49
<211> LENGTH: 306
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus polymyxa SC2
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(306)
<223> OTHER INFORMATION: P_polymyxa_SC2

<400> SEQUENCE: 49

Asn Glu Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Ser Lys Ser
 1                    5                    10                    15

Phe Thr Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr
 20                    25                    30

Arg Gly Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile
 35                    40                    45

Pro Gly Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala
 50                    55                    60

Gly Val Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys
 65                    70                    75                    80

Ala Lys Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg
 85                    90                    95

Ser Thr Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly
 100                   105                   110

Ser Gln Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Ile Ala Phe
 115                   120                   125

Ser Gly Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr
 130                   135                   140

Glu Tyr Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn
 145                   150                   155                   160

Glu Ala Phe Ser Asp Val Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp
 165                   170                   175

Leu Val Gly Asp Asp Ile Tyr Thr Pro Asn Ile Ala Gly Asp Ala Leu
 180                   185                   190

Arg Ser Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser
 195                   200                   205

Asn Leu Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser
 210                   215                   220

Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Asn Phe
 225                   230                   235                   240

His Gly Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile
 245                   250                   255

Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser
 260                   265                   270

Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala
 275                   280                   285

Asn Ser Ala Glu Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly
 290                   295                   300

Val Asn
 305

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<210> SEQ ID NO 50
<211> LENGTH: 306
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus polymyxa SC2
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(306)
<223> OTHER INFORMATION: P_polymyxa_SC2_YP_003948511.1

<400> SEQUENCE: 50
Asn Glu Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Ser Lys Ser
1          5          10          15
Phe Thr Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr
20          25          30
Arg Gly Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile
35          40          45
Pro Gly Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala
50          55          60
Gly Val Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys
65          70          75          80
Ala Lys Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg
85          90          95
Ser Thr Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly
100         105         110
Ser Gln Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Ile Ala Phe
115        120        125
Ser Gly Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr
130        135        140
Glu Tyr Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn
145        150        155        160
Glu Ala Phe Ser Asp Val Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp
165        170        175
Leu Val Gly Asp Asp Ile Tyr Thr Pro Asn Ile Ala Gly Asp Ala Leu
180        185        190
Arg Ser Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser
195        200        205
Asn Leu Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser
210        215        220
Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Asn Phe
225        230        235        240
His Gly Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile
245        250        255
Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser
260        265        270
Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala
275        280        285
Asn Ser Ala Glu Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly
290        295        300
Val Asn
305

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<210> SEQ ID NO 51
<211> LENGTH: 304
<212> TYPE: PRT
<213> ORGANISM: P. terrae

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<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(304)
<223> OTHER INFORMATION: P_terrae_HPL-003_YP_005073223

<400> SEQUENCE: 51

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Asn
 1          5          10
Thr Thr Gln Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly
 20          25          30
Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Thr Ile Pro Gly
 35          40          45
Thr Leu Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Gly Val
 50          55          60
Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys Asp Lys
 65          70          75          80
Phe Gly Arg Asn Ser Ile Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr
 85          90          95
Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln
 100         105         110
Met Thr Tyr Gly Asp Gly Asp Gly Thr Thr Phe Ile Ala Phe Ser Gly
 115         120         125
Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr
 130         135         140
Thr Ser Asn Leu Asp Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ser
 145         150         155         160
Phe Ser Asp Ile Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val
 165         170         175
Gly Asp Asp Ile Tyr Thr Pro Ser Ile Ala Gly Asp Ala Leu Arg Ser
 180         185         190
Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu
 195         200         205
Tyr Lys Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile
 210         215         220
Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe His Asn
 225         230         235         240
Val Thr Val Ser Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr
 245         250         255
Ser Ala Phe Thr Asn Tyr Leu Thr Ser Thr Ser Asn Phe Ser Asn Thr
 260         265         270
Arg Ala Ala Val Val Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser
 275         280         285
Ala Gln Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn
 290         295         300

<210> SEQ ID NO 52
<211> LENGTH: 309
<212> TYPE: PRT
<213> ORGANISM: P. peoriae
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(309)
<223> OTHER INFORMATION: P_peoriae_KCTC

<400> SEQUENCE: 52

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Asp Ile Ile Asn Glu Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp
1      5      10      15
Thr Lys Ser Phe Thr Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Arg
20      25      30
Asp Thr Thr Arg Gly Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg
35      40      45
Gln Ser Ile Pro Gly Thr Ile Leu Thr Asp Ala Asp Asn Val Trp Asn
50      55      60
Asp Pro Ala Gly Val Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp
65      70      75      80
Tyr Tyr Lys Glu Lys Phe Asn Arg Asn Ser Ile Asp Gly Arg Gly Leu
85      90      95
Gln Leu Arg Ser Thr Val His Tyr Gly Asn Arg Tyr Asn Asn Ala Phe
100     105     110
Trp Asn Gly Ser Gln Met Thr Tyr Gly Asp Gly Asp Gly Thr Thr Phe
115     120     125
Ile Ala Phe Ser Gly Asp Pro Asp Val Val Gly His Glu Leu Thr His
130     135     140
Gly Val Thr Glu Tyr Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly
145     150     155     160
Ala Leu Asn Glu Ser Phe Ser Asp Ile Ile Gly Asn Asp Ile Gln Arg
165     170     175
Lys Asn Trp Leu Val Gly Asp Asp Ile Tyr Thr Pro Arg Ile Ala Gly
180     185     190
Asp Ala Leu Arg Ser Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp
195     200     205
His Tyr Ser Asn Leu Tyr Arg Gly Ser Ser Asp Asn Gly Gly Val His
210     215     220
Thr Asn Ser Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly
225     230     235     240
Gly Thr Phe His Gly Val Thr Val Asn Gly Ile Gly Arg Asp Ala Ala
245     250     255
Val Gln Ile Tyr Tyr Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser
260     265     270
Asp Phe Ser Asn Ala Arg Asp Ala Val Val Gln Ala Ala Lys Asp Leu
275     280     285
Tyr Gly Ala Ser Ser Ala Gln Ala Thr Ala Ala Ala Lys Ala Phe Asp
290     295     300
Ala Val Gly Val Asn
305

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<210> SEQ ID NO 53
<211> LENGTH: 316
<212> TYPE: PRT
<213> ORGANISM: Bacillus thermoproteolyticus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(316)
<223> OTHER INFORMATION: 1KEI.A

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<400> SEQUENCE: 53

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Ile Thr Gly Thr Ser Thr Val Gly Val Gly Arg Gly Val Leu Gly Asp
1      5      10      15
Gln Lys Asn Ile Asn Thr Thr Tyr Ser Thr Tyr Tyr Tyr Leu Gln Asp

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Ala Ala Tyr Asp Ala Ala Ala Val Asp Ala His Tyr Tyr Ala Gly Arg  
65 70 75 80

Thr Tyr Asp Tyr Tyr Lys Ala Thr Phe Asn Arg Asn Ser Ile Asn Asp  
85 90 95

Ala Gly Ala Pro Leu Lys Ser Thr Val His Tyr Gly Ser Arg Tyr Asn  
100 105 110

Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly  
115 120 125

Val Thr Phe Thr Ser Leu Ser Gly Gly Ile Asp Val Ile Gly His Glu  
130 135 140

Leu Thr His Ala Val Thr Glu Tyr Ser Ser Asp Leu Ile Tyr Gln Asn  
145 150 155 160

Glu Ser Gly Ala Leu Asn Glu Ala Ile Ser Asp Val Phe Gly Thr Leu  
165 170 175

Val Glu Tyr Tyr Asp Asn Arg Asn Pro Asp Trp Glu Ile Gly Glu Asp  
180 185 190

Ile Tyr Thr Pro Gly Lys Ala Gly Asp Ala Leu Arg Ser Met Ser Asp  
195 200 205

Pro Thr Lys Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly  
210 215 220

Thr Gly Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys  
225 230 235 240

Ala Ala Tyr Leu Leu Ala Asn Gly Gly Thr His Tyr Gly Val Thr Val  
245 250 255

Asn Gly Ile Gly Lys Asp Lys Val Gly Ala Ile Tyr Tyr Arg Ala Asn  
260 265 270

Thr Gln Tyr Phe Thr Gln Ser Thr Thr Phe Ser Gln Ala Arg Ala Gly  
275 280 285

Leu Val Gln Ala  
290

<210> SEQ ID NO 56  
 <211> LENGTH: 317  
 <212> TYPE: PRT  
 <213> ORGANISM: B. thuringiensis  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(317)  
 <223> OTHER INFORMATION: B\_thuringiensis\_YP893436.1

<400> SEQUENCE: 56

Val Thr Gly Thr Asn Ala Val Gly Thr Gly Lys Gly Val Leu Gly Asp  
1 5 10 15

Thr Lys Ser Leu Asn Thr Thr Leu Ser Ala Ser Ser Tyr Tyr Leu Gln  
20 25 30

Asp Asn Thr Arg Gly Ala Thr Ile Phe Thr Tyr Asp Ala Lys Asn Arg  
35 40 45

Ser Thr Leu Pro Gly Thr Leu Trp Val Asp Ala Asp Asn Val Phe Asn  
50 55 60

Ala Ala Tyr Asp Ala Ala Val Asp Ala His Tyr Tyr Ala Gly Lys  
65 70 75 80

Thr Tyr Asp Tyr Tyr Lys Ala Thr Phe Asn Arg Asn Ser Ile Asn Asp  
85 90 95

Ala Gly Ala Pro Leu Lys Ser Thr Val His Tyr Gly Ser Arg Tyr Asn

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|   |     |     |
|---|-----|-----|
| 100   | 105 | 110 |
| Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly |     |     |
| 115   | 120 | 125 |
| Val Thr Phe Thr Ser Leu Ser Gly Gly Ile Asp Val Ile Gly His Glu |     |     |
| 130   | 135 | 140 |
| Leu Thr His Ala Val Thr Glu Tyr Ser Ser Asp Leu Ile Tyr Gln Asn |     |     |
| 145   | 150 | 155 |
| Glu Ser Gly Ala Leu Asn Glu Ala Ile Ser Asp Val Phe Gly Thr Leu |     |     |
|   | 165 | 170 |
| Val Glu Phe Tyr Asp Asn Arg Asn Pro Asp Trp Glu Ile Gly Glu Asp |     |     |
|   | 180 | 185 |
| Ile Tyr Thr Pro Gly Lys Ala Gly Asp Ala Leu Arg Ser Met Ser Asp |     |     |
|   | 195 | 200 |
| Pro Thr Lys Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly |     |     |
|   | 210 | 215 |
| Thr Gly Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys |     |     |
| 225   | 230 | 235 |
| Ala Ala Tyr Leu Leu Ala Asn Gly Gly Thr His Tyr Gly Val Thr Val |     |     |
|   | 245 | 250 |
| Asn Gly Ile Gly Lys Asp Lys Val Gly Ala Ile Tyr Tyr Arg Ala Asn |     |     |
|   | 260 | 265 |
| Thr Gln Tyr Phe Thr Gln Ser Thr Thr Phe Ser Gln Ala Arg Ala Gly |     |     |
|   | 275 | 280 |
| Leu Val Gln Ala Ala Thr Asp Leu Tyr Gly Ala Ser Ser Ala Glu Val |     |     |
|   | 290 | 295 |
| Ala Ala Val Lys Gln Ser Tyr Ser Ala Val Gly Val Asn             |     |     |
| 305   | 310 | 315 |

&lt;210&gt; SEQ ID NO 57

&lt;211&gt; LENGTH: 314

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: B. cereus

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: misc\_feature

&lt;222&gt; LOCATION: (1)..(314)

&lt;223&gt; OTHER INFORMATION: B\_cereus\_ZP04310163.1

&lt;400&gt; SEQUENCE: 57

|   |     |     |
|---|-----|-----|
| Thr Asn Ala Val Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser |     |     |
| 1   | 5   | 10  |
| Leu Asn Thr Thr Leu Ser Ala Ser Ser Tyr Tyr Leu Gln Asp Asn Thr |     |     |
| 20  | 25  | 30  |
| Arg Gly Ala Thr Ile Phe Thr Tyr Asp Ala Lys Asn Arg Ser Thr Leu |     |     |
| 35  | 40  | 45  |
| Pro Gly Thr Leu Trp Val Asp Ala Asp Asn Val Phe Asn Ala Ala Tyr |     |     |
| 50  | 55  | 60  |
| Asp Ala Ala Ala Val Asp Ala His Tyr Tyr Ala Gly Lys Thr Tyr Asp |     |     |
| 65  | 70  | 75  |
| Tyr Tyr Lys Ala Thr Phe Asn Arg Asn Ser Ile Asn Asp Ala Gly Ala |     |     |
|   | 85  | 90  |
| Pro Leu Lys Ser Thr Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe |     |     |
|   | 100 | 105 |
| Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly Val Thr Phe |     |     |
|   | 115 | 120 |
|   |     | 125 |

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Thr Ser Leu Ser Gly Gly Ile Asp Val Ile Gly His Glu Leu Thr His  
 130 135 140  
 Ala Val Thr Glu Tyr Ser Ser Asp Leu Ile Tyr Gln Asn Glu Ser Gly  
 145 150 155 160  
 Ala Leu Asn Glu Ala Ile Ser Asp Val Phe Gly Thr Leu Val Glu Phe  
 165 170 175  
 Tyr Asp Asn Arg Asn Pro Asp Trp Glu Ile Gly Glu Asp Ile Tyr Thr  
 180 185 190  
 Pro Gly Lys Ala Gly Asp Ala Leu Arg Ser Met Ser Asp Pro Thr Lys  
 195 200 205  
 Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly Thr Gly Asp  
 210 215 220  
 Asn Gly Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys Ala Ala Tyr  
 225 230 235 240  
 Leu Leu Ala Asn Gly Gly Thr His Tyr Gly Val Thr Val Asn Gly Ile  
 245 250 255  
 Gly Lys Asp Lys Val Gly Ala Ile Tyr Tyr Arg Ala Asn Thr Gln Tyr  
 260 265 270  
 Phe Thr Gln Ser Thr Thr Phe Ser Gln Ala Arg Ala Gly Leu Val Gln  
 275 280 285  
 Ala Ala Ala Asp Leu Tyr Gly Ala Ser Ser Ala Glu Val Ala Ala Val  
 290 295 300  
 Lys Gln Ser Tyr Ser Ala Val Gly Val Asn  
 305 310

<210> SEQ ID NO 58  
 <211> LENGTH: 317  
 <212> TYPE: PRT  
 <213> ORGANISM: Lactobacillus sp.  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(317)  
 <223> OTHER INFORMATION: Lactobacillus\_sp\_BAA06144.1

<400> SEQUENCE: 58

Val Thr Gly Thr Asn Ala Val Gly Thr Gly Lys Gly Val Leu Gly Asp  
 1 5 10 15  
 Thr Lys Ser Leu Asn Thr Thr Leu Ser Ala Ser Ser Tyr Tyr Leu Gln  
 20 25 30  
 Asp Asn Thr Arg Gly Ala Thr Ile Phe Thr Tyr Asp Ala Lys Asn Arg  
 35 40 45  
 Ser Thr Leu Pro Gly Thr Leu Trp Val Asp Ala Asp Asn Val Phe Asn  
 50 55 60  
 Ala Ala Tyr Asp Ala Ala Ala Val Asp Ala His Tyr Tyr Ala Gly Lys  
 65 70 75 80  
 Thr Tyr Asp Tyr Tyr Lys Ala Thr Phe Asn Arg Asn Ser Ile Asn Asp  
 85 90 95  
 Ala Gly Ala Pro Leu Lys Ser Thr Val His Tyr Gly Ser Lys Tyr Asn  
 100 105 110  
 Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly  
 115 120 125  
 Val Thr Phe Thr Ser Leu Ser Gly Gly Ile Asp Val Ile Gly His Glu  
 130 135 140

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Leu Thr His Ala Val Thr Glu Tyr Ser Ser Asp Leu Ile Tyr Gln Asn
145                      150                      155                      160

Glu Ser Gly Ala Leu Asn Glu Ala Ile Ser Asp Val Phe Gly Thr Leu
165                      170                      175

Val Glu Tyr Tyr Asp Asn Arg Asn Pro Asp Trp Glu Ile Gly Glu Asp
180                      185                      190

Ile Tyr Thr Pro Gly Lys Ala Gly Asp Ala Leu Arg Ser Met Ser Asp
195                      200                      205

Pro Thr Lys Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly
210                      215                      220

Thr Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys
225                      230                      235

Ala Ala Tyr Leu Leu Ala Asn Gly Gly Thr His Tyr Gly Val Thr Val
245                      250                      255

Asn Gly Ile Gly Lys Asp Lys Val Gly Ala Ile Tyr Tyr Arg Ala Asn
260                      265                      270

Thr Gln Tyr Phe Thr Gln Ser Thr Thr Phe Ser Gln Ala Arg Ala Gly
275                      280                      285

Leu Val Gln Ala Ala Ala Asp Leu Tyr Gly Ala Ser Ser Ala Glu Val
290                      295                      300

Ala Ala Val Lys Gln Ser Tyr Ser Ala Val Gly Val Asn
305                      310                      315

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<210> SEQ ID NO 59
<211> LENGTH: 317
<212> TYPE: PRT
<213> ORGANISM: Bacillus thermoproteolyticus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(317)
<223> OTHER INFORMATION: 1NPC.A

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<400> SEQUENCE: 59

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Val Thr Gly Thr Asn Lys Val Gly Thr Gly Lys Gly Val Leu Gly Asp
1                      5                      10                      15

Thr Lys Ser Leu Asn Thr Thr Leu Ser Gly Ser Ser Tyr Tyr Leu Gln
20                      25                      30

Asp Asn Thr Arg Gly Ala Thr Ile Phe Thr Tyr Asp Ala Lys Asn Arg
35                      40                      45

Ser Thr Leu Pro Gly Thr Leu Trp Ala Asp Ala Asp Asn Val Phe Asn
50                      55                      60

Ala Ala Tyr Asp Ala Ala Ala Val Asp Ala His Tyr Tyr Ala Gly Lys
65                      70                      75                      80

Thr Tyr Asp Tyr Tyr Lys Ala Thr Phe Asn Arg Asn Ser Ile Asn Asp
85                      90                      95

Ala Gly Ala Pro Leu Lys Ser Thr Val His Tyr Gly Ser Asn Tyr Asn
100                     105                     110

Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr Gly Asp Gly Asp Gly
115                     120                     125

Val Thr Phe Thr Ser Leu Ser Gly Gly Ile Asp Val Ile Gly His Glu
130                     135                     140

Leu Thr His Ala Val Thr Glu Asn Ser Ser Asn Leu Ile Tyr Gln Asn
145                     150                     155                     160

Glu Ser Gly Ala Leu Asn Glu Ala Ile Ser Asp Ile Phe Gly Thr Leu

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|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
|     | 165 |     | 170 |     | 175 |     |     |     |     |     |     |     |     |     |     |
| Val | Glu | Phe | Tyr | Asp | Asn | Arg | Asn | Pro | Asp | Trp | Glu | Ile | Gly | Glu | Asp |
|     |     |     | 180 |     |     |     |     | 185 |     |     |     |     | 190 |     |     |
| Ile | Tyr | Thr | Pro | Gly | Lys | Ala | Gly | Asp | Ala | Leu | Arg | Ser | Met | Ser | Asp |
|     |     | 195 |     |     |     |     | 200 |     |     |     |     | 205 |     |     |     |
| Pro | Thr | Lys | Tyr | Gly | Asp | Pro | Asp | His | Tyr | Ser | Lys | Arg | Tyr | Thr | Gly |
|     |     | 210 |     |     |     | 215 |     |     |     |     | 220 |     |     |     |     |
| Ser | Ser | Asp | Asn | Gly | Gly | Val | His | Thr | Asn | Ser | Gly | Ile | Ile | Asn | Lys |
| 225 |     |     |     | 230 |     |     |     |     |     | 235 |     |     |     |     | 240 |
| Gln | Ala | Tyr | Leu | Leu | Ala | Asn | Gly | Gly | Thr | His | Tyr | Gly | Val | Thr | Val |
|     |     |     | 245 |     |     |     |     |     | 250 |     |     |     |     | 255 |     |
| Thr | Gly | Ile | Gly | Lys | Asp | Lys | Leu | Gly | Ala | Ile | Tyr | Tyr | Arg | Ala | Asn |
|     |     | 260 |     |     |     |     |     | 265 |     |     |     |     | 270 |     |     |
| Thr | Gln | Tyr | Phe | Thr | Gln | Ser | Thr | Thr | Phe | Ser | Gln | Ala | Arg | Ala | Gly |
|     |     | 275 |     |     |     |     | 280 |     |     |     |     | 285 |     |     |     |
| Ala | Val | Gln | Ala | Ala | Ala | Asp | Leu | Tyr | Gly | Ala | Asn | Ser | Ala | Glu | Val |
|     | 290 |     |     |     |     | 295 |     |     |     |     | 300 |     |     |     |     |
| Ala | Ala | Val | Lys | Gln | Ser | Phe | Ser | Ala | Val | Gly | Val | Asn |     |     |     |
| 305 |     |     |     | 310 |     |     |     |     |     | 315 |     |     |     |     |     |

<210> SEQ ID NO 60  
 <211> LENGTH: 317  
 <212> TYPE: PRT  
 <213> ORGANISM: B. cytotoxicus  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(317)  
 <223> OTHER INFORMATION: B\_cytotoxicus\_YP001373863.1

<400> SEQUENCE: 60

|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Val | Thr | Gly | Thr | Asn | Ala | Val | Gly | Thr | Gly | Thr | Gly | Val | Leu | Gly | Asp |
| 1   |     |     |     | 5   |     |     |     |     | 10  |     |     |     |     | 15  |     |
| Lys | Lys | Ser | Ile | Asn | Thr | Thr | Leu | Ser | Gly | Ser | Thr | Tyr | Tyr | Leu | Gln |
|     |     | 20  |     |     |     |     | 25  |     |     |     |     |     | 30  |     |     |
| Asp | Asn | Thr | Arg | Gly | Ala | Gln | Ile | Phe | Thr | Tyr | Asp | Ala | Lys | Asn | Arg |
|     |     | 35  |     |     |     | 40  |     |     |     |     |     | 45  |     |     |     |
| Ser | Ser | Leu | Pro | Gly | Thr | Leu | Trp | Ala | Asp | Val | Asp | Asn | Ala | Phe | His |
|     |     | 50  |     |     |     | 55  |     |     |     |     | 60  |     |     |     |     |
| Ala | Lys | Tyr | Asp | Ala | Ala | Ala | Val | Asp | Ala | His | Tyr | Tyr | Ala | Gly | Val |
| 65  |     |     |     | 70  |     |     |     |     |     | 75  |     |     |     | 80  |     |
| Thr | Tyr | Asp | Tyr | Tyr | Lys | Asn | Thr | Phe | Asn | Arg | Asn | Ser | Ile | Asn | Asp |
|     |     |     | 85  |     |     |     |     | 90  |     |     |     |     |     | 95  |     |
| Ala | Gly | Ala | Ala | Leu | Lys | Ser | Thr | Val | His | Tyr | Gly | Ser | Asn | Tyr | Asn |
|     |     | 100 |     |     |     |     |     | 105 |     |     |     |     |     | 110 |     |
| Asn | Ala | Phe | Trp | Asn | Gly | Ser | Gln | Met | Val | Tyr | Gly | Asp | Gly | Asp | Gly |
|     |     | 115 |     |     |     |     | 120 |     |     |     |     | 125 |     |     |     |
| Val | Thr | Phe | Thr | Ser | Leu | Ser | Gly | Gly | Ile | Asp | Val | Ile | Gly | His | Glu |
|     |     | 130 |     |     |     |     | 135 |     |     |     |     | 140 |     |     |     |
| Leu | Thr | His | Ala | Val | Thr | Glu | Tyr | Ser | Ser | Asn | Leu | Ile | Tyr | Gln | Tyr |
| 145 |     |     |     |     | 150 |     |     |     |     | 155 |     |     |     |     | 160 |
| Glu | Ser | Gly | Ala | Leu | Asn | Glu | Ala | Ile | Ser | Asp | Ile | Phe | Gly | Thr | Leu |
|     |     |     | 165 |     |     |     |     |     | 170 |     |     |     |     | 175 |     |
| Val | Glu | Tyr | Tyr | Asp | Asn | Arg | Asn | Pro | Asp | Trp | Glu | Ile | Gly | Glu | Asp |
|     |     | 180 |     |     |     |     |     |     | 185 |     |     |     |     | 190 |     |



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Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg Tyr Thr Gly Ser Ser Asp  
 210 215 220  
 Asn Gly Gly Val His Thr Asn Ser Gly Ile Ile Asn Lys Ala Ala Tyr  
 225 230 235 240  
 Leu Leu Ala Asn Gly Gly Thr His Tyr Gly Val Thr Val Thr Gly Ile  
 245 250 255  
 Gly Gly Asp Lys Leu Gly Lys Ile Tyr Tyr Arg Ala Asn Thr Leu Tyr  
 260 265 270  
 Phe Thr Gln Ser Thr Thr Phe Ser Gln Ala Arg Ala Gly Leu Val Gln  
 275 280 285  
 Ala Ala Ala Asp Leu Tyr Gly Ser Gly Ser Gln Glu Val Ile Ser Val  
 290 295 300  
 Gly Lys Ser Phe Asp Ala Val Gly Val Gln  
 305 310

<210> SEQ ID NO 62  
 <211> LENGTH: 322  
 <212> TYPE: PRT  
 <213> ORGANISM: Bacillus sp. SG-1  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(322)  
 <223> OTHER INFORMATION: B\_sp\_SG-1\_ZP01858398.1

<400> SEQUENCE: 62

Val Ser Gly Thr Asp Gln Val Gly Thr Gly Lys Gly Val Leu Gly Asp  
 1 5 10 15  
 Thr Lys Ser Leu Asn Thr Thr Leu Ser Asn Gly Thr Tyr Tyr Leu Gln  
 20 25 30  
 Asp Asn Thr Arg Gly Gly Gly Ile Met Thr Tyr Asp Met Lys Asn Arg  
 35 40 45  
 Thr Phe Phe Pro Gln Phe Tyr Leu Pro Gly Ser Leu Trp Ser Asp Ala  
 50 55 60  
 Asp Asn Val Tyr Asn Gln Ala Tyr Asp Ala Ala Val Asp Ala His  
 65 70 75 80  
 Tyr Phe Ala Gly Ala Thr Phe Asp Tyr Tyr Lys Asp Val Phe Gly Arg  
 85 90 95  
 Asn Ser Tyr Asp Asn Lys Gly Thr Thr Ile Gln Ser Ser Val His Tyr  
 100 105 110  
 Ser Lys Asn Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln Met Val Tyr  
 115 120 125  
 Gly Asp Gly Asp Gly Thr Thr Phe Ile Pro Leu Ser Gly Gly Leu Asp  
 130 135 140  
 Val Val Ala His Glu Leu Thr His Ala Val Thr Asp Thr Ser Ser Asp  
 145 150 155 160  
 Leu Val Tyr Gln Asn Glu Ser Gly Ala Leu Asn Glu Ala Ile Ser Asp  
 165 170 175  
 Ile Phe Gly Thr Leu Val Glu Tyr His Glu Asn His Asn Pro Asp Phe  
 180 185 190  
 Glu Ile Gly Glu Asp Ile Tyr Thr Pro Asn Thr Pro Asn Asp Ala Leu  
 195 200 205  
 Arg Ser Met Ser Asp Pro Ala Lys Tyr Gly Asp Pro Asp His Tyr Ser  
 210 215 220  
 Val Arg Tyr Thr Gly Thr Gln Asp Asn Gly Gly Val His Ile Asn Ser

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225          230          235          240
Gly Ile Ile Asn Lys Gln Ala Tyr Leu Leu Ser Glu Gly Gly Thr His
          245          250          255
Tyr Gly Val Asn Val Thr Gly Ile Gly Arg Glu Lys Leu Gly Glu Ile
          260          265          270
Tyr Tyr Arg Met Asn Thr Val Tyr Leu Thr Ala Ser Ser Thr Phe Ser
          275          280          285
Gln Ala Arg Ser Ala Ala Val Gln Ala Ala Ser Asp Leu Tyr Gly Ser
          290          295          300
Asn Ser Pro Glu Val Gln Ser Val Asn Gln Ser Phe Asp Ala Val Gly
305          310          315          320
Ile Asn

<210> SEQ ID NO 63
<211> LENGTH: 306
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus peoriae
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(306)
<223> OTHER INFORMATION: PpePro1

<400> SEQUENCE: 63
Ala Thr Gly Thr Gly Arg Gly Val Asp Gly Val Thr Lys Ser Phe Thr
 1          5          10          15
Thr Thr Ala Ser Gly Asn Gly Tyr Gln Leu Lys Asp Thr Thr Arg Ser
          20          25          30
Asn Gly Ile Val Thr Tyr Thr Ala Asn Asn Arg Gln Thr Thr Pro Gly
 35          40          45
Thr Ile Met Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Ala Val
 50          55          60
Asp Ala His Ala Tyr Ala Ile Lys Thr Tyr Asp Tyr Tyr Lys Asn Lys
 65          70          75          80
Phe Gly Arg Asp Ser Ile Asp Gly Arg Gly Met Gln Ile Arg Ser Thr
          85          90          95
Val His Tyr Gly Lys Lys Tyr Val Asn Ala Phe Trp Asn Gly Ser Gln
          100          105          110
Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Thr Phe Phe Ser Gly
          115          120          125
Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Phe
          130          135          140
Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala
          145          150          155          160
Phe Ser Asp Ile Ile Gly Asn Asp Ile Asp Gly Ala Asn Trp Leu Leu
          165          170          175
Gly Asp Gly Ile Tyr Thr Pro Gly Ile Pro Gly Asp Ala Leu Arg Ser
          180          185          190
Leu Ser Asp Pro Thr Arg Phe Gly Gln Pro Asp His Tyr Ser Asn Phe
          195          200          205
Tyr Pro Asp Pro Asn Asn Asp Asp Glu Gly Gly Val His Thr Asn Ser
          210          215          220
Gly Ile Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Ser
225          230          235          240

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His Gly Val Lys Val Thr Gly Ile Gly Arg Glu Ala Ala Val Phe Ile  
 245 250 255

Tyr Tyr Asn Ala Phe Thr Asn Tyr Leu Thr Ser Thr Ser Asn Phe Ser  
 260 265 270

Asn Ala Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Phe Tyr Gly Ala  
 275 280 285

Asp Ser Leu Ala Val Thr Ser Ala Ile Lys Ser Phe Asp Ala Val Gly  
 290 295 300

Ile Lys  
 305

<210> SEQ ID NO 64  
 <211> LENGTH: 304  
 <212> TYPE: PRT  
 <213> ORGANISM: Paenibacillus polymyxa  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)..(304)  
 <223> OTHER INFORMATION: PpoPro2

<400> SEQUENCE: 64

Ala Thr Gly Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Ser Phe Thr  
 1 5 10 15

Thr Thr Ala Ser Gly Ser Ser Tyr Gln Leu Lys Asp Thr Thr Arg Gly  
 20 25 30

Asn Gly Ile Val Thr Tyr Thr Ala Ser Asn Arg Gln Ser Ile Pro Gly  
 35 40 45

Thr Leu Leu Thr Asp Ala Asp Asn Val Trp Asn Asp Pro Ala Gly Val  
 50 55 60

Asp Ala His Ala Tyr Ala Ala Lys Thr Tyr Asp Tyr Tyr Lys Ser Lys  
 65 70 75 80

Phe Gly Arg Asp Ser Val Asp Gly Arg Gly Leu Gln Leu Arg Ser Thr  
 85 90 95

Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln  
 100 105 110

Met Thr Tyr Gly Asp Gly Asp Gly Ser Thr Phe Ile Ala Phe Ser Gly  
 115 120 125

Asp Pro Asp Val Val Gly His Glu Leu Thr His Gly Val Thr Glu Tyr  
 130 135 140

Thr Ser Asn Leu Glu Tyr Tyr Gly Glu Ser Gly Ala Leu Asn Glu Ala  
 145 150 155 160

Phe Ser Asp Val Ile Gly Asn Asp Ile Gln Arg Lys Asn Trp Leu Val  
 165 170 175

Gly Asp Asp Ile Tyr Thr Pro Asn Ile Ala Gly Asp Ala Leu Arg Ser  
 180 185 190

Met Ser Asn Pro Thr Leu Tyr Asp Gln Pro Asp His Tyr Ser Asn Leu  
 195 200 205

Tyr Lys Gly Ser Ser Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile  
 210 215 220

Ile Asn Lys Ala Tyr Tyr Leu Leu Ala Gln Gly Gly Thr Phe His Gly  
 225 230 235 240

Val Ala Val Asn Gly Ile Gly Arg Asp Ala Ala Val Gln Ile Tyr Tyr  
 245 250 255

Ser Ala Phe Thr Asn Tyr Leu Thr Ser Ser Ser Asp Phe Ser Asn Ala

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                260                265                270
Arg Ala Ala Val Ile Gln Ala Ala Lys Asp Leu Tyr Gly Ala Asn Ser
    275                280                285

Ala Glu Ala Thr Ala Ala Ala Lys Ser Phe Asp Ala Val Gly Val Asn
    290                295                300

<210> SEQ ID NO 65
<211> LENGTH: 303
<212> TYPE: PRT
<213> ORGANISM: Paenibacillus terrae
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(303)
<223> OTHER INFORMATION: PtePro1

<400> SEQUENCE: 65

Ala Thr Gly Thr Gly Val Gly Val Leu Gly Asp Thr Lys Thr Phe Thr
 1      5      10      15

Thr Thr Gln Ser Gly Thr Gln Tyr Val Met Gln Asp Thr Thr Arg Gly
 20     25     30

Gly Gly Ile Val Thr Tyr Ser Ala Gly Asn Thr Gln Ser Leu Pro Gly
 35     40     45

Thr Leu Met Arg Asp Thr Asp Asn Val Trp Thr Asp Pro Ala Ala Val
 50     55     60

Asp Ala His Ala Tyr Ala Ala Val Val Tyr Asp Tyr Phe Lys Asn Asn
 65     70     75     80

Phe Asn Arg Asp Ser Leu Asp Gly Arg Gly Met Ala Ile Lys Ser Thr
 85     90     95

Val His Tyr Gly Ser Arg Tyr Asn Asn Ala Phe Trp Asn Gly Thr Gln
 100    105    110

Ile Ala Tyr Gly Asp Gly Asp Gly Thr Thr Phe Arg Ala Phe Ser Gly
 115    120    125

Asp Leu Asp Val Ile Gly His Glu Leu Thr His Gly Ile Thr Glu Lys
 130    135    140

Thr Ala Gly Leu Ile Tyr Gln Gly Glu Ser Gly Ala Leu Asn Glu Ser
 145    150    155    160

Ile Ser Asp Val Phe Gly Asn Thr Ile Gln Gly Lys Asn Trp Leu Ile
 165    170    175

Gly Asp Asp Ile Tyr Thr Pro Ser Ile Pro Gly Asp Ala Leu Arg Ser
 180    185    190

Met Glu Asn Pro Thr Leu Phe Asn Gln Pro Asp His Tyr Ser Asn Ile
 195    200    205

Tyr Arg Gly Ser Asp Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile
 210    215    220

Pro Asn Lys Ala Phe Tyr Leu Leu Ala Gln Gly Gly Thr His Arg Gly
 225    230    235    240

Val Ser Val Thr Gly Ile Gly Arg Gly Asp Ala Ala Lys Ile Val Tyr
 245    250    255

Lys Ala Leu Thr Tyr Tyr Leu Thr Ser Thr Ser Asn Phe Ala Ala Met
 260    265    270

Arg Gln Ala Ala Ile Ser Ser Ala Thr Asp Leu Phe Gly Ala Asn Ser
 275    280    285

Ala Gln Val Asn Ser Val Lys Ala Ala Tyr Ala Ala Val Gly Ile
 290    295    300

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<210> SEQ ID NO 66
<211> LENGTH: 304
<212> TYPE: PRT
<213> ORGANISM: Brevibacillus brevis
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(304)
<223> OTHER INFORMATION: BbrPro1

<400> SEQUENCE: 66

Val Thr Ala Thr Gly Lys Gly Val Leu Gly Asp Thr Lys Gln Phe Glu
 1                               5                10                15

Thr Thr Lys Gln Gly Ser Thr Tyr Met Leu Lys Asp Thr Thr Arg Gly
                20                25                30

Lys Gly Ile Glu Thr Tyr Thr Ala Asn Asn Arg Thr Ser Leu Pro Gly
 35                               40                45

Thr Leu Met Thr Asp Ser Asp Asn Tyr Trp Thr Asp Gly Ala Ala Val
 50                               55                60

Asp Ala His Ala His Ala Gln Lys Thr Tyr Asp Tyr Phe Arg Asn Val
 65                               70                75                80

His Asn Arg Asn Ser Tyr Asp Gly Asn Gly Ala Val Ile Arg Ser Thr
                85                90                95

Val His Tyr Ser Thr Arg Tyr Asn Asn Ala Phe Trp Asn Gly Ser Gln
                100               105               110

Met Val Tyr Gly Asp Gly Asp Gly Thr Thr Phe Leu Pro Leu Ser Gly
 115                120                125

Gly Leu Asp Val Val Ala His Glu Leu Thr His Ala Val Thr Glu Arg
 130                135                140

Thr Ala Gly Leu Val Tyr Gln Asn Glu Ser Gly Ala Leu Asn Glu Ser
 145                150                155                160

Met Ser Asp Ile Phe Gly Ala Met Val Asp Asn Asp Asp Trp Leu Met
                165                170                175

Gly Glu Asp Ile Tyr Thr Pro Gly Arg Ser Gly Asp Ala Leu Arg Ser
 180                185                190

Leu Gln Asp Pro Ala Ala Tyr Gly Asp Pro Asp His Tyr Ser Lys Arg
 195                200                205

Tyr Thr Gly Ser Gln Asp Asn Gly Gly Val His Thr Asn Ser Gly Ile
 210                215                220

Asn Asn Lys Ala Ala Tyr Leu Leu Ala Glu Gly Gly Thr His Tyr Gly
 225                230                235                240

Val Arg Val Asn Gly Ile Gly Arg Thr Asp Thr Ala Lys Ile Tyr Tyr
                245                250                255

His Ala Leu Thr His Tyr Leu Thr Pro Tyr Ser Asn Phe Ser Ala Met
 260                265                270

Arg Arg Ala Ala Val Leu Ser Ala Thr Asp Leu Phe Gly Ala Asn Ser
 275                280                285

Arg Gln Val Gln Ala Val Asn Ala Ala Tyr Asp Ala Val Gly Val Lys
 290                295                300

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<210> SEQ ID NO 67
<211> LENGTH: 300
<212> TYPE: PRT
<213> ORGANISM: Bacillus subtilis
<220> FEATURE:

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<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(300)
<223> OTHER INFORMATION: NprE

<400> SEQUENCE: 67

Ala Ala Thr Thr Gly Thr Gly Thr Thr Leu Lys Gly Lys Thr Val Ser
 1          5          10          15
Leu Asn Ile Ser Ser Glu Ser Gly Lys Tyr Val Leu Arg Asp Leu Ser
 20          25          30
Lys Pro Thr Gly Thr Gln Ile Ile Thr Tyr Asp Leu Gln Asn Arg Glu
 35          40          45
Tyr Asn Leu Pro Gly Thr Leu Val Ser Ser Thr Thr Asn Gln Phe Thr
 50          55          60
Thr Ser Ser Gln Arg Ala Ala Val Asp Ala His Tyr Asn Leu Gly Lys
 65          70          75          80
Val Tyr Asp Tyr Phe Tyr Gln Lys Phe Asn Arg Asn Ser Tyr Asp Asn
 85          90          95
Lys Gly Gly Lys Ile Val Ser Ser Val His Tyr Gly Ser Arg Tyr Asn
 100         105         110
Asn Ala Ala Trp Ile Gly Asp Gln Met Ile Tyr Gly Asp Gly Asp Gly
 115         120         125
Ser Phe Phe Ser Pro Leu Ser Gly Ser Met Asp Val Thr Ala His Glu
 130         135         140
Met Thr His Gly Val Thr Gln Glu Thr Ala Asn Leu Asn Tyr Glu Asn
 145         150         155         160
Gln Pro Gly Ala Leu Asn Glu Ser Phe Ser Asp Val Phe Gly Tyr Phe
 165         170         175
Asn Asp Thr Glu Asp Trp Asp Ile Gly Glu Asp Ile Thr Val Ser Gln
 180         185         190
Pro Ala Leu Arg Ser Leu Ser Asn Pro Thr Lys Tyr Gly Gln Pro Asp
 195         200         205
Asn Phe Lys Asn Tyr Lys Asn Leu Pro Asn Thr Asp Ala Gly Asp Tyr
 210         215         220
Gly Gly Val His Thr Asn Ser Gly Ile Pro Asn Lys Ala Ala Tyr Asn
 225         230         235         240
Thr Ile Thr Lys Ile Gly Val Asn Lys Ala Glu Gln Ile Tyr Tyr Arg
 245         250         255
Ala Leu Thr Val Tyr Leu Thr Pro Ser Ser Thr Phe Lys Asp Ala Lys
 260         265         270
Ala Ala Leu Ile Gln Ser Ala Arg Asp Leu Tyr Gly Ser Gln Asp Ala
 275         280         285
Ala Ser Val Glu Ala Ala Trp Asn Ala Val Gly Leu
 290         295         300

<210> SEQ ID NO 68
<211> LENGTH: 300
<212> TYPE: PRT
<213> ORGANISM: Bacillus subtilis
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(300)
<223> OTHER INFORMATION: NprE_variant

<400> SEQUENCE: 68

Ala Ala Thr Thr Gly Thr Gly Thr Thr Leu Lys Gly Lys Thr Val Ser

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| 1   | 5   | 10  | 15  |
|---|-----|-----|-----|
| Leu Asn Ile Ser Ser Glu Ser Gly Lys Tyr Val Leu Arg Asp Leu Ser | 20  | 25  | 30  |
| Lys Pro Thr Gly Thr Gln Ile Ile Thr Tyr Asp Leu Gln Asn Arg Glu | 35  | 40  | 45  |
| Tyr Asn Leu Pro Gly Thr Leu Val Ser Ser Thr Thr Asn Gln Phe Thr | 50  | 55  | 60  |
| Thr Ser Ser Gln Arg Ala Ala Val Asp Ala His Tyr Asn Leu Gly Lys | 65  | 70  | 80  |
| Val Tyr Asp Tyr Phe Tyr Gln Lys Phe Asn Arg Asn Ser Tyr Asp Asn | 85  | 90  | 95  |
| Lys Gly Gly Lys Ile Val Ser Ser Val His Tyr Gly Ser Arg Tyr Asn | 100 | 105 | 110 |
| Asn Ala Ala Trp Ile Gly Asp Gln Met Ile Tyr Gly Asp Gly Asp Gly | 115 | 120 | 125 |
| Ile Leu Phe Ser Pro Leu Ser Gly Ser Leu Asp Val Thr Ala His Glu | 130 | 135 | 140 |
| Met Thr His Gly Val Thr Gln Glu Thr Ala Asn Leu Asn Tyr Glu Asn | 145 | 150 | 155 |
| Gln Pro Gly Ala Leu Asn Glu Ser Phe Ser Asp Val Phe Gly Tyr Phe | 165 | 170 | 175 |
| Asn Asp Thr Glu Asp Trp Asp Ile Gly Glu Asp Ile Thr Ile Ser Gln | 180 | 185 | 190 |
| Pro Ala Leu Arg Ser Leu Ser Asn Pro Thr Lys Tyr Gly Gln Pro Asp | 195 | 200 | 205 |
| Asn Phe Lys Asn Tyr Lys Asn Leu Pro Asn Thr Pro Ala Gly Asp Tyr | 210 | 215 | 220 |
| Gly Gly Val His Thr Asn Ser Gly Ile Pro Asn Lys Ala Ala Tyr Asn | 225 | 230 | 235 |
| Thr Ile Thr Lys Ile Gly Val Asn Lys Ala Glu Gln Ile Tyr Tyr Arg | 245 | 250 | 255 |
| Ala Leu Thr Val Tyr Leu Thr Pro Ser Ser Thr Phe Lys Asp Ala Lys | 260 | 265 | 270 |
| Ala Ala Leu Ile Gln Ser Ala Arg Asp Leu Tyr Gly Ser Gln Asp Ala | 275 | 280 | 285 |
| Ala Ser Val Glu Ala Ala Trp Asn Ala Val Gly Leu                 | 290 | 295 | 300 |

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1. A polypeptide comprising an amino acid sequence having at least 60%, 80%, or 95% sequence identity to an amino acid sequence selected from the group consisting of SEQ ID NOs: 3, 8, 13, 18, 23, 28, 33 and 38.

2-4. (canceled)

5. The polypeptide of claim 1, wherein said polypeptide is derived from a member of the order Bacillales or is derived from a *Planococcus* species.

6. The polypeptide of claim 5, wherein said Bacillales member is a Paenibacillaceae family member or a *Paenibacillus* spp.

7-8. (canceled)

9. The polypeptide of claim 1, wherein said polypeptide has protease activity.

10. The polypeptide of claim 9, wherein said protease activity comprises casein hydrolysis, collagen hydrolysis, elastin hydrolysis, keratin hydrolysis, soy protein hydrolysis or corn meal protein hydrolysis.

11. The polypeptide of claim 1, wherein said polypeptide retains at least 50% of its maximal activity between pH 4.5 and 10 and/or between 30° C. and 70° C.

12. (canceled)

13. The polypeptide of claim 1, wherein said polypeptide has cleaning activity in a detergent composition.

14. The polypeptide of claim 13, wherein said detergent composition is selected from an ADW detergent composition, a laundry detergent composition, a liquid laundry detergent composition, and a powder laundry detergent composition.

15-18. (canceled)

**19.** The polypeptide of claim **1**, wherein said polypeptide is a recombinant polypeptide.

**20.** A composition comprising the polypeptide of claim **1**.

**21.** The composition of claim **20**, wherein said composition is a cleaning composition or a detergent composition.

**22.** (canceled)

**23.** The composition of claim **21**, wherein said detergent composition is selected from the group consisting of a laundry detergent, a fabric softening detergent, a dishwashing detergent, and a hard-surface cleaning detergent.

**24.** The composition of any of claim **20**, wherein said composition further comprises a surfactant; at least one calcium ion and/or zinc ion; at least one stabilizer; from about 0.001 to about 0.1 weight % of said polypeptide; at least one bleaching agent; at least one adjunct ingredient; and/or one or more additional enzymes or enzyme derivatives selected from the group consisting of acyl transferases, alpha-amylases, beta-amylases, alpha-galactosidases, arabinosidases, aryl esterases, beta-galactosidases, carrageenases, catalases, cellobiohydrolases, cellulases, chondroitinases, cutinases, endo-beta-1,4-glucanases, endo-beta-mannanases, esterases, exo-mannanases, galactanases, glucoamylases, hemicellulases, hyaluronidases, keratinases, laccases, lactases, ligninases, lipases, lipoxygenases, mannanases, oxidases, pectate lyases, pectin acetyl esterases, pectinases, pentosanases, peroxidases, phenoloxidases, phosphatases, phospholipases, phytases, polygalacturonases, proteases, pullulanases, reductases, rhamnogalacturonases, beta-glucanases, tannases, transglutaminases, xylan acetyl-esterases, xylanases, xyloglucanases, and xylosidases, additional metalloprotease enzymes and combinations thereof.

**25-31.** (canceled)

**32.** The composition of claim **20**, wherein said cleaning composition contains phosphate or is phosphate-free.

**33-34.** (canceled)

**35.** The composition of claim **20**, wherein said composition is a granular, powder, solid, bar, liquid, tablet, gel, or paste composition.

**36-38.** (canceled)

**39.** A method of cleaning, comprising contacting a surface or an item with a cleaning composition comprising the polypeptide of claim **1**.

**40-41.** (canceled)

**42.** The method of claim **39**, wherein said item is dishware or fabric.

**43-48.** (canceled)

**49.** A method for producing the polypeptide of claim **1** comprising:

a. stably transforming a host cell with an expression vector comprising a polynucleotide encoding the polypeptide of claim **1**;

b. cultivating said transformed host cell under conditions suitable for said host cell to produce said protease; and

c. recovering said protease.

**50-56.** (canceled)

**57.** A nucleic acid sequence comprising a nucleic acid sequence:

(i) having at least 70% identity to a sequence selected from the group consisting of SEQ ID NOs: 4, 9, 14, 19, 24, 29, 34 and 39, or

(ii) being capable of hybridizing to a probe derived from the polynucleotide sequence selected from the group consisting of SEQ ID NOs: 4, 9, 14, 19, 24, 29, 34 and 39, under conditions of intermediate to high stringency, or

(iii) being complementary to the polynucleotide sequence selected from the group consisting of SEQ ID NOs: 4, 9, 14, 19, 24, 29, 34 and 39.

**58.** A vector comprising the nucleic acid sequence of claim **57**.

**59.** A host cell transformed with the vector of claim **58**.

**60-61.** (canceled)

**62.** A textile processing, animal feed, leather processing, feather processing, or corn soy protein processing composition comprising the polypeptide of claim **1**.

**63-67.** (canceled)

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