



(51) International Patent Classification:

A61K 35/12 (2015.01) C12N 5/071 (2010.01)
A61P 27/16 (2006.01)

(21) International Application Number:

PCT/US2017/015379

(22) International Filing Date:

27 January 2017 (27.01.2017)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

62/288,958 29 January 2016 (29.01.2016) US

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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DJ, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT,

HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KH, KN, KP, KR, KW, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Declarations under Rule 4.17:

- as to applicant's entitlement to apply for and be granted a patent (Rule 4.17(ii))
- as to the applicant's entitlement to claim the priority of the earlier application (Rule 4.17(iii))

Published:

- with international search report (Art. 21(3))
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments (Rule 48.2(h))

(54) Title: EXPANSION AND DIFFERENTIATION OF INNER EAR SUPPORTING CELLS AND METHODS OF USE THEREOF

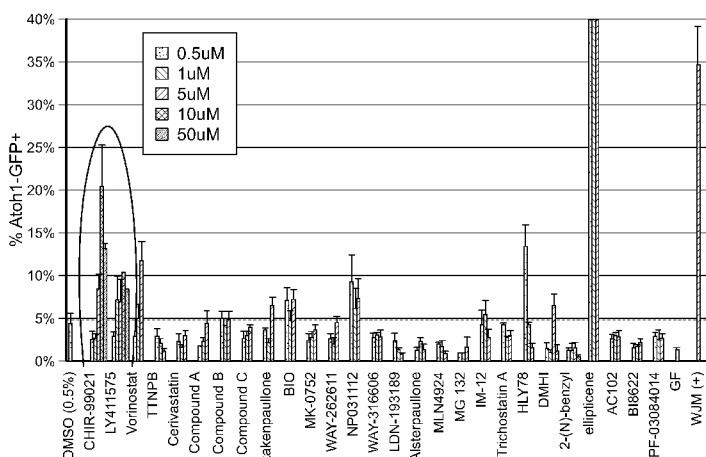


FIG. 5

(57) Abstract: This disclosure relates to methods for expanding inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) and differentiating inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) to inner ear hair cells (e.g., atonal homolog 1 (Atoh1)+ inner ear hair cells) and the use of the inner ear supporting cells and hair cells, e.g., for identifying candidate therapeutic compounds for the treatment of hearing loss and balance loss. Additionally, the methods described herein can be used in the treatment of a subject having hearing loss and balance loss that would benefit from increased proliferation and differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells).

WO 2017/132530 A1

Expansion and Differentiation of Inner Ear Supporting Cells and Methods of Use Thereof

CLAIM OF PRIORITY

This application claims the benefit of U.S. Provisional Application Serial No.
5 62/288,958, filed on January 29, 2016. The entire contents of the foregoing are
incorporated herein by reference.

TECHNICAL FIELD

Described herein are methods for expanding and differentiating inner ear
supporting cells (e.g., Lgr5+ inner ear supporting cells) from the sensory epithelium
10 of the inner ear of a mammal, and the use of the expanded cells, e.g., for identifying
candidate therapeutic compounds for the treatment of hearing loss or balance loss.
Additionally, the methods described herein can be used in the treatment of hearing
loss or balance loss that would benefit from increased proliferation of inner ear
supporting cells (e.g., Lgr5+ inner ear supporting cells), and differentiation thereof
15 into inner ear hair cells (e.g., atonal homolog 1 (Atoh1)+ inner ear hair cells).

BACKGROUND

Hearing impairment is a major health challenge estimated by the WHO to
affect over 5% of the world's population (360 million people, including 32 million
children). The sensory hair cells that detect sound and transmit their signal to the
20 brain via the auditory nerve are susceptible to damage. After loss, the hair cells are
never replaced^{1, 2}, and thus the number of cells, which is low (15,000 per ear) at the
start of postnatal life, only decreases with age, and the absence of cell replacement
leads to a high prevalence of acquired forms of deafness. Indeed, hair cell and
auditory nerve damage, typically caused by noise exposure, ototoxic drugs,
25 viral/bacterial infections, and aging accounts for more than 80% of all cases of
hearing loss³. In addition to hearing impairment, damage or loss of sensory hair cells
can cause balance impairment and diseases related to balance impairment, e.g., benign
paroxysmal positional vertigo (BPPV).

SUMMARY

30 The present disclosure is based, at least in part, on the discovery of methods
for expanding inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells, e.g.,

Lgr5⁺ cochlear or vestibular supporting cells) from the sensory epithelium of the inner ear of a mammal, and the use of the expanded cells, e.g., for identifying candidate therapeutic compounds for the treatment of hearing loss or balance loss. Additionally, described herein are methods for using screened compounds in the treatment of hearing loss or balance loss that would benefit from increased proliferation of inner ear supporting cells (e.g., Lgr5⁺ inner ear supporting cells) and/or increased numbers of inner ear hair cells (e.g., atonal homolog 1 (Atoh1)⁺ inner ear hair cells).

In a first aspect, the invention features a method of producing an expanded population of inner ear supporting cells. The method includes contacting a population of inner ear supporting cells with one or more agents selected from the group consisting of: (a) a retinoid receptor signaling activator; (b) a Wnt signaling activator set forth in Table A; (c) a bone morphogenetic protein (BMP) signaling inhibitor set forth in Table B; (d) a cyclin-dependent kinase (CDK) activator set forth in Table C; (e) an E box-dependent transcriptional activator set forth in Table D; (f) a Notch signaling activator set forth in Table E; (g) a histone deacetylase (HDAC) inhibitor set forth in Table F; (h) a protein degradation inhibitor set forth in Table G; (i) a PI3K-Akt signaling inhibitor set forth in Table H; and (j) a cAMP response element binding protein (CREB) activator set forth in Table I, in which the one or more agents are present in amounts sufficient to produce an expanded population of inner ear supporting cells.

In some embodiments of the first aspect of the invention, the Notch signaling activator is a Delta-like protein activator, a Jagged protein activator, a Notch activator, and/or a γ -secretase activator.

In some embodiments of the first aspect of the invention, the one or more agents are selected from the group consisting of: (a) a retinoid receptor signaling activator; (b) a Wnt signaling activator set forth in Table A; (c) a BMP signaling inhibitor set forth in Table B; (d) a CDK activator set forth in Table C; and (e) an E box-dependent transcriptional activator set forth in Table D.

In some embodiments of the first aspect of the invention, the expanded population of inner ear supporting cells is an expanded population of Lgr5⁺ inner ear supporting cells. In some embodiments, the expanded population of Lgr5⁺ inner ear supporting cells is an expanded population of Lgr5⁺ cochlear supporting cells. In

some embodiments, the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ vestibular supporting cells.

In a second aspect, the invention features a method of promoting differentiation of a population of inner ear supporting cells into a population of inner ear hair cells. The method includes contacting a population of inner ear supporting cells with one or more agents selected from the group consisting of: (a) a retinoid receptor signaling activator; (b) a Wnt signaling activator set forth in Table A; (c) a BMP signaling inhibitor set forth in Table B; (d) a CDK activator set forth in Table C; (e) an E box-dependent transcriptional activator set forth in Table D; (f) an HDAC inhibitor set forth in Table F; (g) a protein degradation inhibitor set forth in Table G; (h) a PI3K-Akt signaling inhibitor set forth in Table H; (i) a CREB activator set forth in Table I; and (j) a Notch signaling inhibitor set forth in Table J, in which the one or more agents are present in amounts sufficient to promote differentiation into a population of inner ear hair cells.

In some embodiments of the second aspect of the invention, the one or more agents is selected from the group consisting of: (a) a Wnt signaling activator set forth in Table A; (b) an E box-dependent transcriptional activator set forth in Table D; (c) an HDAC inhibitor set forth in Table F; (d) a protein degradation inhibitor set forth in Table G; and (e) a Notch signaling inhibitor set forth in Table J.

In some embodiments of the second aspect of the invention, the Notch signaling inhibitor is a Delta-like protein inhibitor, a Jagged protein inhibitor, a Notch inhibitor, and/or a γ -secretase inhibitor.

In some embodiments of the second aspect of the invention, the population of inner ear hair cells is a population of Atoh1+ inner ear hair cells. In some embodiments, the population of Atoh1+ inner ear hair cells is a population of Atoh1+ cochlear hair cells. In some embodiments, the population of Atoh1+ inner ear hair cells is a population of Atoh1+ vestibular hair cells.

In some embodiments of the first and second aspects of the invention, the retinoid receptor signaling activator is a retinoic acid receptor (RAR) agonist set forth in Table K and/or a retinoic X receptor (RXR) agonist set forth in Table K. In some embodiments, the RAR agonist is an RAR α agonist, an RAR β agonist, and/or an RAR γ agonist. In some embodiments, the RXR agonist is an RXR α agonist, an RXR β agonist, and/or an RXR γ agonist.

In some embodiments of the first and second aspects of the invention, the Wnt signaling activator is a glycogen synthase kinase-3 β (GSK-3 β) inhibitor, a Wnt activator, a Frizzled receptor activator, a lipoprotein receptor-related protein 5/6 (LRP5/6) activator, a Disheveled (Dvl) activator, an Axin inhibitor, a Dickkopf (Dkk) inhibitor, a secreted Frizzled-related protein (sFRP) inhibitor, a Groucho inhibitor, and/or a Wnt inhibitory protein (WIF) inhibitor.

In some embodiments of the first and second aspects of the invention, the BMP signaling inhibitor is a Noggin activator, a Chordin activator, a BMP receptor inhibitor, a SMAD1/5/8 inhibitor, a SMAD2/3 inhibitor, and/or a SMAD4 inhibitor.

In some embodiments of the first and second aspects of the invention, the CDK activator is a p27Kip1 inhibitor and/or a retinoblastoma protein (Rb) inhibitor.

In some embodiments of the first and second aspects of the invention, the E box-dependent transcriptional activator is an Atoh1 activator.

In some embodiments of the first and second aspects of the invention, the HDAC inhibitor is an HDAC class I inhibitor, an HDAC class II inhibitor, an HDAC class III inhibitor, and/or a pan-HDAC inhibitor. In some embodiments, the HDAC class III inhibitor is a SIRT1 inhibitor and/or a SIRT2 inhibitor.

In some embodiments of the first and second aspects of the invention, the protein degradation inhibitor is a proteasome inhibitor or a ubiquitin ligase inhibitor.

In some embodiments of the first and second aspects of the invention, the PI3K-Akt signaling inhibitor is an Akt inhibitor, a PI3K inhibitor, a PKC inhibitor, and/or a PDK1 inhibitor.

In some embodiments of the first and second aspects of the invention, the population of inner ear supporting cells is a population of Lgr5⁺ inner ear supporting cells. In some embodiments, the population of Lgr5⁺ inner ear supporting cells is a population of Lgr5⁺ cochlear supporting cells. In some embodiments, the population of Lgr5⁺ inner ear supporting cells is a population of Lgr5⁺ vestibular supporting cells.

In a third aspect, the invention features a transgenic mouse having two or more recombinant nucleic acid molecules stably integrated into the genome of the mouse. The two or more recombinant nucleic acid molecules include at least a first recombinant nucleic acid molecule that includes a first reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group

consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glast, and Lfng, and a second recombinant nucleic acid molecule that includes a second reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of Atoh1, Myo7a, Cdh23, Pcdh15, Myo6, Myo1c, Tmc1, and
5 Cav1.3, in which the first reporter gene is different from the second reporter gene.

In some embodiments of the third aspect of the invention, the inner ear supporting cell marker is Lgr5.

In some embodiments of the third aspect of the invention, the inner ear hair cell marker is Atoh1.

10 In some embodiments of the third aspect of the invention, the inner ear supporting cell marker is Lgr5 and the inner ear hair cell marker is Atoh1.

In some embodiments of the third aspect of the invention, the regulatory element of an inner ear supporting cell marker is an Lgr5 promoter.

In some embodiments of the third aspect of the invention, the regulatory
15 element of an inner ear hair cell marker is an Atoh1 enhancer. In some embodiments, the Atoh1 enhancer is operably linked to a promoter element, e.g., an SV40 promoter or a globin promoter.

In some embodiments of the third aspect of the invention, the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a
20 second fluorescent protein, wherein the first fluorescent protein is different from the second fluorescent protein.

In a fourth aspect, the invention features a cell isolated from the transgenic mouse of the third aspect of the invention. The cell includes the first recombinant nucleic acid molecule and the second recombinant nucleic acid molecule.

25 In some embodiments of the fourth aspect of the invention, the cell is isolated from the inner ear of the transgenic mouse.

In a fifth aspect of the invention, the invention features a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method
30 includes: (a) isolating a population of inner ear supporting cells from the mouse of any one of claims 29-36; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells; (c) administering a test compound to the expanded population of inner ear

supporting cells; (d) detecting the expression levels of the first reporter gene and the second reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the first reporter gene compared to the expression level of the first reporter gene in the absence of the test compound, and/or increases the expression level of the second reporter gene compared to the expression level of the second reporter gene in the absence of the test compound.

In some embodiments of the fifth aspect of the invention, the conditions sufficient to produce an expanded population of inner ear supporting cells include media, e.g., a mixture of DMEM and F12 media, e.g., a 1:1 mixture, in the presence of one or more growth factors, e.g., epidermal growth factor (EGF), basic fibroblast growth factor (bFGF), and/or insulin-like growth factor (IGF1). In some embodiments, the population of inner ear supporting cells is cultured for two or more days, e.g., between two days and ten days, e.g., between two days and five days.

In some embodiments of the fifth aspect of the invention, the conditions sufficient to produce an expanded population of inner ear supporting cells further include one or more agents selected from the group consisting of: (a) a retinoid receptor signaling activator; (b) a Wnt signaling activator set forth in Table A; (c) a BMP signaling inhibitor set forth in Table B; (d) a CDK activator set forth in Table C; (e) an E box-dependent transcriptional activator set forth in Table D; (f) a Notch signaling activator set forth in Table E or a Notch signaling inhibitor set forth in Table J; (g) an HDAC inhibitor set forth in Table F; (h) a protein degradation inhibitor set forth in Table G; (i) a PI3K-Akt signaling inhibitor set forth in Table H; and (j) a CREB activator set forth in Table I.

In some embodiments of the fifth aspect of the invention, the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

In some embodiments of the fifth aspect of the invention, the conditions sufficient to produce an expanded population of inner ear supporting cells include one or more agents set forth in Table 1.

In some embodiments of the fifth aspect of the invention, the conditions sufficient to produce an expanded population of inner ear supporting cells include

media, e.g., a 1:1 mixture of DMEM and F12, supplemented with N2, B27, EGF, bFGF, IGF1, CHIR99021, and VPA.

In some embodiments of the fifth aspect of the invention, the candidate agent is selected from a group consisting of a small molecule, a compound, a nucleic acid, a peptide, a polypeptide, a growth factor, and an epigenetic modifier.

In some embodiments of the fifth aspect of the invention, the population of inner ear supporting cells is isolated from the cochlea of the mouse by a method including first dissecting the organ of Corti, isolating sensory epithelium, and creating a single cell suspension. In some embodiments of the fifth aspect of the invention, the population of inner ear supporting cells is isolated from the vestibule of the inner ear of the mouse.

In some embodiments of the fifth aspect of the invention, the population of inner ear supporting cells is a population of Lgr5⁺ inner ear supporting cells.

In some embodiments of the fifth aspect of the invention, the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a second fluorescent protein, wherein the first fluorescent protein is different from the second fluorescent protein.

In some embodiments of the fifth aspect of the invention, the expression levels of the first reporter gene and the second reporter gene are protein expression levels.

In a sixth aspect, the invention features a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method includes: (a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that includes a reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glax, and Lfng; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells, wherein the conditions include one or more agents selected from the group consisting of: (i) a retinoid receptor signaling activator, (ii) a Wnt signaling activator set forth in Table A, (iii) a BMP signaling inhibitor set forth in Table B, (iv) a CDK activator set forth in Table C, (v) an E box-dependent transcriptional activator set forth in Table D, (vi) a Notch signaling activator set forth in Table E, (vii) an HDAC inhibitor set forth in Table F, (viii) a

protein degradation inhibitor set forth in Table G, (ix) a PI3K-Akt signaling inhibitor set forth in Table H, and (x) a CREB activator set forth in Table I; (c) administering a test compound to the expanded population of inner ear supporting cells; (d) detecting the expression level of the reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

In a seventh aspect, the invention features a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method includes: (a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that includes a reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of *Atoh1*, *Myo7a*, *Cdh23*, *Pcdh15*, *Myo6*, *Myo1c*, *Tmc1*, and *Cav1.3*; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells, wherein the conditions include one or more agents selected from the group consisting of: (i) a retinoid receptor signaling activator, (ii) a Wnt signaling activator set forth in Table A, (iii) a BMP signaling inhibitor set forth in Table B, (iv) a CDK activator set forth in Table C, (v) an E box-dependent transcriptional activator set forth in Table D, (vi) a Notch signaling activator set forth in Table E, (vii) an HDAC inhibitor set forth in Table F, (viii) a protein degradation inhibitor set forth in Table G, (ix) a PI3K-Akt signaling inhibitor set forth in Table H, and (x) a CREB activator set forth in Table I; (c) administering a test compound to the expanded population of inner ear supporting cells; (d) detecting the expression level of the reporter gene in the expanded population of inner ear cells in the presence of the test compound; and (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

In some embodiments of the sixth and seventh aspects of the invention, the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

In some embodiments of the sixth and seventh aspects of the invention, the conditions sufficient to produce an expanded population of inner ear supporting cells include one or more agents selected from the group consisting of: (a) a retinoid receptor signaling activator; (b) a Wnt signaling activator set forth in Table A; (c) a
5 BMP signaling inhibitor set forth in Table B; (d) a CDK activator set forth in Table C; and (e) an E box-dependent transcriptional activator set forth in Table D.

In some embodiments of the sixth and seventh aspects of the invention, the expanded population of inner ear supporting cells is an expanded population of Lgr5+ inner ear supporting cells. In some embodiments, the expanded population of Lgr5+
10 inner ear supporting cells is an expanded population of Lgr5+ cochlear supporting cells. In some embodiments, the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ vestibular supporting cells.

In some embodiments of the sixth and seventh aspects of the invention, the population of inner ear supporting cells is isolated from a human. In some
15 embodiments, the population of inner ear supporting cells is isolated from a mouse.

In some embodiments of the sixth and seventh aspects of the invention, the reporter gene encodes a fluorescent protein.

In an eighth aspect, the invention features a method of treating a subject having hearing loss or balance loss, in which the method includes administering to the
20 subject in need thereof one or both of: (a) a therapeutically effective amount of one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator
25 set forth in Table D; (vi) a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in Table F; (viii) a protein degradation inhibitor set forth in Table G; and (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and/or (b) a therapeutically effective amount of one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting
30 of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set

forth in Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set forth in Table I; and (x) a Notch signaling inhibitor set forth in Table J.

In some embodiments of the eighth aspect of the invention, the method
5 includes administering to the subject in need thereof one or both of: (a) a
therapeutically effective amount of one or more agents that promote proliferation of
inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor
signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP
signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; and
10 (v) an E box-dependent transcriptional activator set forth in Table D; and/or (b) a
therapeutically effective amount of one or more agents that promote differentiation of
inner ear supporting cells into inner ear hair cells selected from the group consisting
of: (i) a Wnt signaling activator set forth in Table A; (ii) an E box-dependent
transcriptional activator set forth in Table D; (iii) an HDAC inhibitor set forth in
15 Table F; (iv) a protein degradation inhibitor set forth in Table G; and (v) a Notch
signaling inhibitor set forth in Table J.

In some embodiments of the eighth aspect of the invention, the one or more
agents are administered systemically. In some embodiments of the eighth aspect of
the invention, the one or more agents are administered locally, e.g., to the ear of the
20 subject, e.g., transtympanically to the middle ear of the subject.

In some embodiments of the eighth aspect of the invention, the one or more
agents that promote proliferation of inner ear supporting cells are administered prior
to the one or more agents that promote differentiation of inner ear supporting cells
into inner ear hair cells.

In a ninth aspect, the invention features a method of treating a subject having
25 hearing loss or balance loss, in which the method includes: (a) contacting one or more
inner ear supporting cells, e.g., in vitro, with one or more agents that promote
proliferation of inner ear supporting cells selected from the group consisting of: (i) a
retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table
30 A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth
in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi)
a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in
Table F; (viii) a protein degradation inhibitor set forth in Table G; (ix) a PI3K-Akt

signaling inhibitor set forth in Table H; and (x) a CREB activator set forth in Table I; (b) optionally contacting the expanded population of inner ear supporting cells with one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of: (i) a retinoid receptor signaling
5 activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set forth in Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set forth in
10 Table I; and (x) a Notch signaling inhibitor set forth in Table J; and (c) administering the inner ear hair cells to the ear (e.g., the inner ear) of the subject.

In a tenth aspect, the invention features a method of treating a subject having hearing loss or balance loss, in which the method includes: (a) contacting one or more inner ear supporting cells, e.g., in vitro, with one or more agents that promote
15 proliferation of inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in
20 Table F; (viii) a protein degradation inhibitor set forth in Table G; (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and (x) a CREB activator set forth in Table I; and (b) administering the expanded population of inner ear supporting cells to the ear (e.g., the inner ear) of the subject in combination with, e.g., concurrently with or prior to administration of, one or more agents that promote differentiation of inner ear
25 supporting cells into inner ear hair cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set forth in
30 Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set forth in Table I; and (x) a Notch signaling inhibitor set forth in Table J.

In some embodiments of the eighth, ninth, and tenth aspects of the invention, (a) the one or more agents that promote proliferation of inner ear supporting cells is

selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; and (v) an E box-dependent transcriptional activator set forth in Table D; and (b) the one or more agents that
5 promote differentiation of inner ear supporting cells into inner ear hair cells is selected from the group consisting of: (i) a Wnt signaling activator set forth in Table A; (ii) an E box-dependent transcriptional activator set forth in Table D; (iii) an HDAC inhibitor set forth in Table F; (iv) a protein degradation inhibitor set forth in Table G; and (v) a Notch signaling inhibitor set forth in Table J.

10 In some embodiments of the eighth, ninth, and tenth aspects of the invention, the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

In some embodiments of the eighth, ninth, and tenth aspects of the invention, the inner ear supporting cells are Lgr5+ inner ear supporting cells.

15 In some embodiments of the eighth, ninth, and tenth aspects of the invention, the inner ear hair cells are Atoh1+ inner ear hair cells.

In some embodiments of the eighth, ninth, and tenth aspects of the invention, the subject has balance loss. In some embodiments of the eighth, ninth, and tenth aspects of the invention, the subject has hearing loss (e.g., sensorineural hearing loss).

20 In some embodiments, the hearing loss is the result of a genetic or congenital defect, trauma (e.g., physical trauma or noise-related insult trauma), aging, or chemical-induced ototoxicity. In some embodiments, the hearing loss is the result of an infection (e.g., a viral or a bacterial infection).

25 In some embodiments of the eighth, ninth, and tenth aspects of the invention, the subject is a human.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Methods and materials are described herein for use in the present invention; other, suitable methods and materials known in the art can also
30 be used. The materials, methods, and examples are illustrative only and not intended to be limiting. All publications, patent applications, patents, sequences, database entries, and other references mentioned herein are incorporated by reference in their

entirety. In case of conflict, the present specification, including definitions, will control.

Other features and advantages of the invention will be apparent from the following detailed description and figures, and from the claims.

5

Definitions

As used herein, the term “expanded population” of inner ear supporting cells refers to a population of cells including at least one more inner ear supporting cells, such that the quantity of inner ear supporting cells in the population is greater (e.g., at least 10% greater, at least 20% greater, at least 30% greater) than the number of inner ear supporting cells prior to administration of one or more agents as described herein (e.g., one or more agents that exhibit one or more activities, such as activation of retinoid receptor signaling, activation of Wnt signaling, inhibition of bone morphogenetic protein (BMP) signaling, activation of a cyclin-dependent kinase (CDK), activation of E box-dependent transcription, activation Notch signaling, inhibition of a histone deacetylase (HDAC), inhibition of protein degradation, inhibition of PI3K-Akt signaling, and activation of cAMP response element binding protein (CREB).

As used herein, the term “inner ear supporting cells” refers to non-sensory cells that reside between hair cells (e.g., cochlear supporting cells and vestibular supporting cells) and serve a diverse set of functions, such as maintaining an environment in the epithelium that enables hair cells to function and supporting the structural integrity of the sensory organs during sound stimulation and head movements.

As used herein, the term “inner ear hair cells” refers to sensory cells that reside within the organ of Corti in the cochlea or the vestibule of the osseous labyrinth of the inner ear. The inner ear hair cells are responsible for transmitting sounds waves as electrical signals to the brain. Damage to cochlear inner ear hair cells can result in decreased hearing sensitivity or hearing loss. Damage to vestibular inner ear hair cells can result in balance impairment or balance loss.

As used herein, a “retinoid receptor signaling activator” refers to a compound that binds and activates one or more retinoid receptors (e.g., RAR α , RAR β , RAR γ , RXR α , RXR β , and RXR γ), thereby affecting the transcriptional activity of a target

gene to which the activated retinoid receptor binds. A retinoid receptor signaling activator may be a pan-retinoid receptor activator or exhibit selectivity towards one or more retinoid receptors. Examples of activators of retinoid receptor signaling include, but are not limited to, compounds listed in Table K.

5 As used herein, a “Wnt signaling activator” refers to an agonist of the canonical Wnt signaling pathway. Agonists of this pathway further include Wnt proteins or other compounds that bind directly to the Frizzled and lipoprotein receptor-related protein 5/6 (LRP5/6) co-receptor proteins (e.g., a Frizzled receptor activator, a LRP5/6 activator), in manner that promotes an increase in the
10 concentration of β -catenin in the nucleus of a mammalian cell. Alternatively, a β -catenin/Wnt pathway agonist may function by inhibiting one or more secreted Frizzled-related proteins (sFRPs) (e.g., an sFRP inhibitor) or Wnt inhibitory protein (WIF) (e.g., a WIF inhibitor), which bind and sequester Wnt proteins from interacting with the endogenous Wnt co-receptors. Examples of Wnt signaling activators also
15 include, but are not limited to, a glycogen synthase kinase-3 β (GSK-3 β) inhibitor, a Wnt activator, a Disheveled (Dvl) activator, an Axin inhibitor, a Dickkopf (Dkk) inhibitor, and a Groucho inhibitor. GSK-3 β is a kinase that forms a complex with Axin, APC (Adenomatous polyposis coli), and β -catenin to prepare β -catenin for downstream degradation by the proteasome. Disheveled (Dvl) is an intracellular
20 protein that relays signals from activated Notch receptors to downstream effectors. Disheveled (Dvl) is recruited by the receptor Frizzled and prevents the constitutive destruction of β -catenin. Dickkopf (Dkk) is a secreted protein that acts to isolate the LRP5/6 co-receptor proteins, thus inhibiting Wnt signaling. Groucho is a protein that forms a complex with TLE in the nucleus to repress gene expression. Once β -catenin
25 enters the nucleus, it disrupts the Groucho/TLE complex to activate gene expression. A Wnt activator refers to a small molecule compound that activates Wnt signaling.

 Exemplary methods that can be used to determine the activity of a β -catenin/Wnt pathway agonist include, without limitation, monitoring the expression of a reporter gene under the control of a TCF/LEF family transcription factor, as well as
30 TOPFlash luciferase reporter assays, as described in US 2014/0044763. Examples of activators of Wnt signaling include, but are not limited to, compounds listed in Table A.

As used herein, a “bone morphogenic protein (BMP) signaling inhibitor” refers to an antagonist of the BMP signaling pathway. BMP is a member of the TGF β superfamily of ligands, and modulators of BMP signaling can be used in conjunction with the methods of the invention, e.g., to expand inner ear supporting cells or to differentiate inner ear supporting cells to inner ear hair cells. Inhibitors of the BMP signaling pathway include any proteins or small molecule compounds that inhibit a protein involved in the BMP signaling pathway. Examples of BMP signaling inhibitors include, but are not limited to, a Noggin activator, a Chordin activator, a BMP receptor inhibitor, a SMAD1/5/8 inhibitor, a SMAD2/3 inhibitor, and a SMAD4 inhibitor. Noggin and Chordin are antagonists that inhibit the binding of BMP to BMP receptors. BMP receptors transduce signals from BMPs. Activated BMP receptors recruit and phosphorylate transcription factors SMAD1/5/8. Phosphorylated SMAD1/5/8 interacts with SMAD4 to form a complex, which goes into the nucleus to regulate gene expression. BMP receptors transduce signals from activin or activin-like ligands. Activated BMP receptors recruit and phosphorylate transcription factors SMAD2/3. Phosphorylated SMAD2/3 interacts with SMAD4 to form a complex, which goes into the nucleus to regulate gene expression. Examples of BMP signaling inhibitors include, but are not limited to, compounds listed in Table B.

As used herein, a “cyclin-dependent kinase (CDK) activator” refers to an agonist of CDK. CDKs are a family of serome/threonine kinases involved regulating cell cycle. A CDK binds a regulatory protein cyclin to form the activated kinase. A CDK activator may be a protein or a small molecule compound that interacts with CDK to increase its activity, or a protein or a small molecule compound that interacts with a protein that interacts with CDK to indirectly increase CDK activity. A CDK activator may be an inhibitor of a CDK inhibitor (e.g., a p27Kip1 inhibitor) or a retinoblastoma protein (Rb) inhibitor. The protein p27Kip1 is a cell-cycle regulatory protein that interacts with cyclin-CDK2 complex and cyclin-CDK4 complex and inhibits cell cycle progression at G1. An Rb is a tumor suppressor protein that also inhibits cell cycle progression at G1.

As used herein, an “E box-dependent transcriptional activator” refers to a protein (e.g., a transcription factor) compound that binds to E box to activate the expression of the gene downstream of the E box. An E box refers to an enhancer box, which is a DNA response element that acts as a protein-binding site to regulate gene

expression. Transcription factors bind to an E box to initiate gene transcription. An example of an E box-dependent transcriptional activator is an Atoh1 activator, which up-regulates the expression of Atoh1. Atoh1 is a basic helix-loop-helix (bHLH) transcription factor involved in regulating neurosensory development in the ear.

5 As used herein, a “Notch signaling activator” refers to a protein or a small molecule compound that promotes the activation of Notch pathway function. The term “Notch pathway function” as used herein refers to a function mediated by the Notch signal transduction pathway including, but not limited to, nuclear translocation of the intracellular domain of Notch, nuclear translocation of RBP-Jκ or its
10 *Drosophila* homolog Suppressor of Hairless; activation of bHLH genes of the Enhancer of Split complex, e.g., Mastermind; activation of the HES-1 gene or the KBF2 (also referred to as CBF1) gene; inhibition of *Drosophila* neuroblast segregation; and binding of Notch to a Delta protein, a Jagged/Serrate protein, Fringe, Deltex or RBP-Jκ/Suppressor of Hairless, or homologs or analogs thereof. The Notch
15 signal transduction cascade and the phenotypes effected by Notch signaling are described, e.g., in Kopan et al., *Cell* 137:216-233 (2009) and Jarriault, et al., *Mol. Cell. Biol.* 18:7423-7431 (1998), the disclosures of each of which are incorporated herein by reference.

Examples of Notch agonists are described, e.g., in US 2014/0369973 and in
20 US 7,399,633, the disclosures of each of which are incorporated herein by reference. Exemplary Notch agonists include, without limitation, Notch proteins, as well as analogs, derivatives, and fragments thereof; other proteins that propagate the Notch signaling pathway, as well as analogs, derivatives, and fragments thereof; activating antibodies that stimulate Notch receptor activity and antigen-binding fragments
25 thereof that retain agonistic activity; nucleic acids encoding proteins that potentiate Notch signaling; as well as proteins, derivatives, and analogs thereof which bind to or otherwise interact with Notch proteins or other proteins in the Notch pathway such that Notch pathway activity is promoted. Such agonists include, but are not limited to, Notch proteins and derivatives thereof containing the Notch intracellular domain,
30 Notch nucleic acids encoding the foregoing, and proteins contacting the Notch-interacting domain of Notch ligands (e.g., the extracellular domain of Delta or Serrate).

Other Notch agonists include, but are not limited to, a Delta-like protein activator, a Jagged protein activator, a Notch activator, and a γ -secretase activator. Delta-like protein and Jagged protein are transmembrane proteins that interact with Notch receptors on adjacent cells to activate Notch signaling. Gamma-secretase is an enzyme that cleaves the part of the Notch receptor on the inside the inner leaflet of the cell membrane of the Notch receptor-expression cell. The cleavage by γ -secretase releases the intracellular domain of the Notch receptor, which then moves to the nucleus to regulate gene expression. A Notch activator refers to a small molecule compound that activates Notch signaling. Other Notch agonists include, but are not limited to, RBPJ κ /Suppressor of Hairless or Deltex. Fringe can additionally be used to enhance Notch activity, for example in conjunction with Delta protein. These proteins, fragments, and derivatives thereof can be recombinantly expressed and isolated or can be chemically.

As used herein, a “Notch signaling inhibitor” refers to a protein or a small molecule compound that inhibits Notch pathway function. The term “Notch pathway function” is described above. In some embodiments, a Notch signaling inhibitor may inhibit the activity of one or more proteins involved in the activation of Notch signaling. In some embodiments, a Notch signaling inhibitor may activate the activity of one or more proteins involved in the inhibition of Notch signaling. Notch signaling inhibitors include, but are not limited to, a Delta-like protein inhibitor, a Jagged protein inhibitor, a Notch inhibitor, and/or a γ -secretase inhibitor. Examples of Notch signaling inhibitors include, but are not limited to, compounds listed in Table J.

As used herein, a “histone deacetylase (HDAC) inhibitor” refers a compound that binds and inhibits one or more HDACs, thereby affecting the enzyme activity of the HDAC. An HDAC refers to any one of a family of enzymes that catalyze the removal of acetyl groups from the ϵ -amino groups of lysine residues at the N-terminus of a histone. Unless otherwise indicated by context, the term “histone” is meant to refer to any histone protein, including H1, H2A, H2B, H3, H4, and H5, from any species. Human HDAC proteins are separated into four classes: class I includes HDAC1, HDAC2, HDAC3, and HDAC8; class II includes HDAC4, HDAC5, HDAC7, and HDAC9; class III includes SIRT1, SIRT2, SIRT3, SIRT4, SIRT5, SIRT6, and SIRT7; and class IV includes HDAC11. An HDAC inhibitor may be a

pan-HDAC inhibitor or exhibit selectivity towards one or more HDACs. Examples of HDAC inhibitors include, but are not limited to, compounds listed in Table F.

As used herein, a “protein degradation inhibitor” refers to a compound that inhibits a protein degradation pathway, e.g., an ubiquitin-proteasome degradation pathway. A protein degradation inhibitor may inhibit the activity of one or more of the proteins involved in a protein degradation pathway, e.g., an ubiquitin-proteasome degradation pathway. For example, a protein degradation inhibitor may be a small molecule compound that inhibits the proteasome (e.g., a proteasome inhibitor), i.e., inhibits the 19S or 20S of the proteasome, or a small molecule compound that inhibits ubiquitin activating enzyme (E1), ubiquitin conjugating enzyme (E2), and/or ubiquitin ligase (E3) (e.g., an ubiquitin ligase inhibitor). In the ubiquitin-proteasome degradation pathway, proteins are marked for degradation by the proteasome by being linked to the co-factor ubiquitin. E1 first forms a thio-ester bond with ubiquitin. This reaction allows subsequent binding of ubiquitin to E2, which replaces E1. Finally, E3 ligase forms an isopeptide bond between the carboxy-terminus of ubiquitin and a lysine residue on the substrate protein. Numerous E3 ligases provide specificity in that each can modify only a subset of substrate proteins. Further specificity is achieved by post-translational modification of substrate proteins, including, but not limited to, phosphorylation. Examples of protein degradation inhibitors include, but are not limited to, compounds listed in Table G.

As used herein, a “PI3K-Akt signaling inhibitor” refers to a compound that inhibits one or more proteins involved in PI3K-Akt signaling. Akt is a serine/threonine kinase (also known as protein kinase B or PKB) that regulates diverse cellular functions, such as metabolism, growth, proliferation, survival, transcription, and protein synthesis. The Akt signaling cascade is activated by receptor tyrosine kinases, integrins, B and T cell receptors, cytokine receptors, G-protein-coupled receptors, and other stimuli that induce production of phosphatidylinositol (3,4,5) trisphosphates (PIP3) by phosphoinositide 3-kinase (PI3K). These lipids serve as plasma membrane docking sites for proteins that harbor pleckstrin-homology (PH) domains, including Akt and its upstream activator PDK1. At the membrane, PDK1 phosphorylates Akt, leading to partial activation of Akt. Phosphorylation of Akt by mTORC2 stimulates full enzymatic activity. Members of the PI3K-related kinase (PIKK) family, including DNA-PK, can also phosphorylate

Akt. Akt is dephosphorylated by protein phosphatase 2A (PP2A) and the PH-domain leucine-rich-repeat-containing protein phosphatases (PHLPP1/2). In addition, the tumor suppressor phosphatase and tensin homolog (PTEN) inhibits Akt activity by dephosphorylating PIP3. There are three highly related isoforms of Akt (Akt1, Akt2, and Akt3), which phosphorylate substrates containing the consensus phosphorylation motif RxRxxS/T. Examples of inhibitors of PI3K-Akt signaling include, but are not limited to, compounds that inhibit Akt (i.e., compounds that inhibit the activation of Akt), compounds that inhibit PI3K, compounds that inhibit PKC, and compounds that inhibit PDK1. Examples of PI3K-Akt signaling inhibitors include, but are not limited to, compounds listed in Table H.

As used herein, a “cAMP response element binding protein (CREB) activator” refers to a compound that binds and activates CREB or a compound that binds and activates a protein involved in the activation of CREB. In some embodiments, a CREB activator may increase the concentration of CREB. CREB is a transcription factor of the leucine zipper family of DNA binding proteins. CREB binds as a homodimer to the cAMP-responsive element (CRE), thereby increasing or decreasing the transcription of the downstream genes. Examples of proteins involved in the activation of CREB include, but are not limited to, PKA and Ca²⁺/calmodulin-dependent protein kinase. Genes whose regulated by CREB include, but are not limited to, c-fos, BDNF, tyrosine hydroxylase, neuropeptides (e.g., somatostatin, enkephalin, VGF, corticotropin-releasing hormone), and genes involved in the mammalian circadian clock (e.g., PER1, PER2). Examples of CREBs include, but are not limited to, CREB1, CREB2, CREB3, CREB5, CREB3-like protein 1 (CREB3L1), CREB3L2, CREB3L3, and CREB3L4. An example of a CREB activator is AC102 (see Table I).

As used herein, the term “balance loss” refers to a deficiency in the vestibular system or vestibular function of a subject that causes the subject to feel unsteady, for example, when standing or walking. Balance loss related to the ear also causes vertigo (spinning) and nausea. Diseases and disorders that are related to balance loss (i.e., caused by balance loss, or cause balance loss) include, but are not limited to, benign paroxysmal positional vertigo (BPPV), labyrinthitis (e.g., vestibular neuronitis, cochlear neuronitis), trauma (i.e., injury to the ear or skull, injury caused by surgery to the ear or skull), Ménière’s disease, perilymph fistula, superior canal

dehiscence syndrome, and bilateral vestibulopathy. Balance loss may also be caused by medication, stress, anxiety, and aging.

DESCRIPTION OF DRAWINGS

The patent or application file contains at least one drawing executed in color.
5 Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee.

FIG 1 is a timeline for the screening of Lgr5+ cells for drugs affecting proliferation.

10 FIG 2 is a timeline for the screening of Lgr5+ cells for drugs affecting differentiation.

FIG 3 is a graph depicting the percent of Lgr5-GFP+ cells from a screen for drugs that affect the proliferation of Lgr5-GFP cells. See Table 1 for a key to the compounds (WJM = CHIR 99021 and valproic acid).

15 FIG 4 is a graph depicting the number of Lgr5-GFP+ cells from a screen for drugs affecting proliferation of Lgr5-GFP cells. See Table 1 for a key to the compounds (WJM = CHIR 99021 and valproic acid).

FIG 5 is a graph depicting the percent of Atoh1-GFP+ cells from a screen for drugs that affect the differentiation of Atoh1-GFP+ cells. See Table 1 for a key to the compounds (WJM = CHIR 99021 and valproic acid).

20 FIG 6 is a schematic illustration of a plasmid for use in generating transgenic mice, in which mCherry fluorescent protein is under the control of an Atoh1 enhancer.

FIG 7 is structures of specific screening compounds Compound A (see WO 2009/100438), Compound B (see WO 2009/100438), Compound C (see WO 2009/100438), and BI8622 (from Peter et al., EMBO Mol Med. 2014 Dec; 6(12):
25 1525–1541).

DETAILED DESCRIPTION

Hair cells transduce sound via an apical stereociliary bundle that couples vibration-induced displacement to ion-channel gating. Damage and death of cochlear hair cells, which occurs in a high percentage of the population, is a cause of
30 widespread hearing loss due to the lack of a mechanism for hair cell replacement. And damage and death of vestibular hair cells can result in balance loss or impairment.

Lgr5, an epithelial stem cell marker, was recently shown to be expressed in supporting cells of the inner ear (e.g., cochlear supporting cells) that surround the hair

cells, and the Lgr5-expressing cells could be induced to proliferate when stimulated by Wnt in the normally post-mitotic cochlear sensory epithelium. Indeed, consistent with a progenitor role, supporting cells that expressed Lgr5 gave rise to new Lgr5-positive cells by propagation and to hair cells that were Lgr5-negative, whereas
5 supporting cells that did not express this receptor did not give rise to hair cells. Consistent with its role in upstream regulation of transcription factor Atoh1, which is a master regulator of hair cell differentiation, upregulation of Wnt also increased hair cell differentiation. This combination of the ability to divide in response to Wnt signaling and the potency to differentiate into hair cells suggested that Lgr5 cells were
10 acting as progenitor cells of the cochlear epithelium. Lgr5+ cells showed a limited capacity to regenerate spontaneously after damage, but their ability to divide and differentiate in response to Wnt stimulation and transdifferentiate in response to Notch inhibition was limited and only observed in neonatal animals.

Although these data supported a role of Lgr5+ cells as inner ear (e.g.,
15 cochlear) progenitor cells, and some expansion of the cells could be achieved by propagation as cochlear spheres, the heterogeneous cell populations suggested that other signaling pathways may be involved in stem cell expansion and hair cell differentiation. Furthermore, spontaneous regeneration capacity was lost after the first postnatal week, and changes in gene expression of these progenitors resulted in a
20 loss in sphere-forming capacity in the adult mouse cochlea. Efforts to replace hair cells have concentrated on supporting cell transdifferentiation to hair cells, but regenerating a functional cochlea or vestibule would require both stimulating these cells to divide and differentiating them to hair cells. Here, by employing a screen, we identified pathways and small molecules that promoted the proliferation and/or
25 differentiation of inner ear supporting cells (e.g., Lgr5-expressing inner ear supporting cells of the cochlea or the vestibule). These expanded inner ear cells can be used, e.g., for identifying compounds that can be used to treat hearing loss in mammals. Expanded inner ear cells (e.g., Lgr5+ inner ear cells) from transgenic animals comprising reporter genes for inner ear supporting cell and hair cell markers (e.g.,
30 Lgr5 and Atoh1 reporter genes, respectively) are particularly useful.

Compounds that Promote Proliferation and/or Differentiation of Inner Ear Supporting Cells

Examples of compounds that promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) include, but are not limited to, a retinoid receptor signaling activator (see, e.g., Table K); a Wnt signaling activator set forth in Table A; a bone morphogenetic protein (BMP) signaling inhibitor set forth in Table B; a cyclin-dependent kinase (CDK) activator set forth in Table C; an E box-dependent transcriptional activator set forth in Table D; a Notch signaling activator set forth in Table E; a histone deacetylase (HDAC) inhibitor set forth in Table F; a protein degradation inhibitor set forth in Table G; a PI3K-Akt signaling inhibitor set forth in Table H; and a cAMP response element binding protein (CREB) activator set forth in Table I.

Examples of compounds that promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., atonal homolog 1 (Atoh1)+ inner ear hair cells) include, but are not limited to, a retinoid receptor signaling activator (see, e.g., Table K); a Wnt signaling activator set forth in Table A; a BMP signaling inhibitor set forth in Table B; a CDK activator set forth in Table C; an E box-dependent transcriptional activator set forth in Table D; an HDAC inhibitor set forth in Table F; a protein degradation inhibitor set forth in Table G; a PI3K-Akt signaling inhibitor set forth in Table H; a CREB activator set forth in Table I; and a Notch signaling inhibitor set forth in Table J.

A number of compounds that support or promote the proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) and/or promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cell) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) are set forth in Table 1.

A number of compounds that support or promote the proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) are described herein, and include one or more of TTNPB, Compound A, Compound B, Compound C, 1-Azakenpaullone, BIO, WAY-316606, LDN-193189, and Alsterpaullone.

A number of compounds that promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cell) into to inner ear hair cells (e.g., Atoh1+ inner ear hair cells) are described herein, and include one or more of vorinostat, Compound A, Compound B, Compound C, 1-Azakenpaullone, BIO,

WAY-262611, NP031112, MG-132, IM-12, Trichostatin A, HLY78, and PF03084014.

In some embodiments of the invention, derivatives of the compounds listed in Tables A-K may also be used to promote the proliferation and/or expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells). A derivative of a compound listed in Tables A-K is a small molecule that differs in structure from the parent compound, but retains the ability to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells). A derivative of a compound may change its interaction with certain other molecules or proteins relative to the parent compound. A derivative of a compound may also include a salt, an adduct, or other variant of the parent compound. In some embodiments of the invention, any derivative of a compound described herein (e.g., any one compound of the compounds listed in Tables A-K) may be used instead of the parent compound. In some embodiments, any derivative of a compound listed in Tables A-I and K may be used in a method of producing an expanded population of inner ear supporting cells. In some embodiments, any derivative of a compound listed in Tables A-D and F-K may be used in a method of promoting differentiation of a population of inner ear supporting cells into a population of inner ear hair cells.

Table A

Compound	Target
CHIR-98023	GSK-3 β
CHIR-99021	GSK-3 β
CHIR-99030	GSK-3 β
Hymenialdisine	GSK-3 β
debromohymenialdisine	GSK-3 β
dibromocantherelline	GSK-3 β
Meridianine A	GSK-3 β
alsterpaullone	GSK-3 β
cazapauillone	GSK-3 β
Aloisine A	GSK-3 β
NSC 693868 (1 <i>H</i> -Pyrazolo[3,4- <i>b</i>]quinoxalin-3-amine)	GSK-3 β

Compound	Target
Indirubin-3'-oxime (Indirubin-3'-monoxime; 3-[1,3-Dihydro-3-(hydroxyimino)-2H-indol-2-ylidene]-1,3-dihydro-2H-indol-2-one)	GSK-3 β
A 1070722 (1-(7-Methoxyquinolin-4-yl)-3-[6-(trifluoromethyl)pyridin-2-yl]urea)	GSK-3 β
L803	GSK-3 β
L803-mts	GSK-3 β
TDZD8	GSK-3 β
NP00111	GSK-3 β
HMK-32	GSK-3 β
Manzamine A	GSK-3 β
Palinurin	GSK-3 β
Tricantin	GSK-3 β
IM-12 (3-(4-Fluorophenylethylamino)-1-methyl-4-(2-methyl-1H-indol-3-yl)-1H-pyrrole-2,5-dione)	GSK-3 β
NP031112	GSK-3 β
NP00111	GSK-3 β
NP031115	GSK-3 β
VP 2.51	GSK-3 β
VP2.54	GSK-3 β
VP 3.16	GSK-3 β
VP 3.35	GSK-3 β
HLY78 (4-Ethyl-5,6-Dihydro-5-methyl-[1,3]dioxolo[4,5-j]phenanthridine, 4-Ethyl-5-methyl-5,6-dihydro-[1,3]dioxolo[4,5-j]phenanthridine)	Axin
WAY-262611 (1-(4-(Naphthalen-2-yl)pyrimidin-2-yl)piperidin-4-yl)methanamine))	Dickkopf-1 (DKK1)
BHQ880	DKK1
NCI8642	DKK1
gallocyanine dyes	DKK1
Compounds 3-8 (Moore et al., <i>J.Med.Chem.</i> , 2009; 52:105)	secreted frizzled-related protein 1 (sFRP-1)
WAY-316606	sFRP-1

Table B

Compound	Target
A01 (Cao et al., <i>Scientific Reports</i> , 2014; 4:4965)	SMAD1/5/8
A17 (Cao et al., <i>Scientific Reports</i> , 2014; 4:4965)	SMAD1/5/8

Table C

Compound	Target
Cerivastatin (Baycol; Lipobay)	p27Kip1
Alsterpaullone 2-cyanoethyl	p27Kip1
SJ403	p27Kip1

5

Table D

Compound	Target
Compound A (See FIG. 7)	Atoh1
Compound B (See FIG. 7)	Atoh1
Compound C (See FIG. 7)	Atoh1
1-Azakenpaullone (Pyrido[3',2':2,3]azepino[4,5-b]indol- 6(5H)-one, 9-bromo-7,12-dihydro-)	Atoh1
2-(N)-benzyl ellipticine	Atoh1

Table E

Compound	Target
Delta/Serrate/Lag-2 peptide	Notch receptor

Table F

Compound	Target
Vorinostat (rINN; suberanilohydroxamic acid; suberoylanilide hydroxamic acid; SAHA (suberoyl+anilide+hydroxamic acid abbreviated); <i>N</i> - Hydroxy- <i>N'</i> -phenyloctanediamide; Zolinza [®])	HDAC class I (HDAC1, 2, 3, and 8) and HDAC class II (IIa: HDAC4, 5, 7, and 9; IIb: 6 and 10)
Trichostatin A (TSA; (2 <i>E</i> ,4 <i>E</i> ,6 <i>R</i>)-7-(4-(Dimethylamino)phenyl)- <i>N</i> - hydroxy-4,6-dimethyl-7-oxo-2,4-heptadienamido)	HDAC class I (HDAC1, 2, 3, and 8) and HDAC class II (IIa: HDAC4, 5, 7, and 9; IIb: 6 and 10)
belinostat (PXD101; Beleodaq)	HDAC

Compound	Target
Valproic acid (VPA; sodium valproate; Sodium 2-propylpentanoate)	HDAC
FK 228 (Depsipeptide; FR 901228; Romidepsin; Cyclo[(2Z)-2-amino-2-butenoyl-L-valyl-(3S,4E)-3-hydroxy-7-mercapto-4-heptenoyl-D-valyl-D-cysteinyl], cyclic (3-5) disulfide)	HDAC class I (HDAC1, 2, 3, and 8), HDAC4, and HDAC6
Sodium butyrate (Butanoic acid sodium salt; NaB)	HDAC
LMK 235 (<i>N</i> -[[6-(Hydroxy amino)-6-oxohexyl]oxy]-3,5-dimethylbenzamide)	HDAC4 and HDAC5
Scriptaid (<i>N</i> -Hydroxy-1,3-dioxo-1 <i>H</i> -benz[de]isoquinoline-2(3 <i>H</i>)-hexanamide)	HDAC
M 344 (4-(Diethylamino)- <i>N</i> -[7-(hydroxy amino)-7-oxoheptyl]benzamide)	HDAC
SBHA (<i>N,N</i> -Dihydroxy octanediamide; suberic bishydroxamate)	HDAC1 and HDAC3
CBHA (<i>m</i> -carboxycinnamic acid bishydroxamide)	HDAC1 and HDAC3
HMBA (hexamethylene bisacetamide).	HDAC
Tubacin (<i>N</i> -[4-[(2 <i>R</i> ,4 <i>R</i> ,6 <i>S</i>)-4-[(4,5-Diphenyl-2-oxazolyl)thio]methyl]-6-[4-(hydroxymethyl)phenyl]-1,3-dioxan-2-yl]phenyl]- <i>N'</i> -hydroxy octanediamide)	HDAC6
Sodium 4-phenylbutyrate (4-PB; sodium phenylbutyrate; 4-Phenylbutyric acid, sodium salt; 4-phenylbutyrate)	HDAC
MC 1568 (3-[5-(3-(3-Fluorophenyl)-3-oxopropen-1-yl)-1-methyl-1 <i>H</i> -pyrrol-2-yl]- <i>N</i> -hydroxy-2-propenamide)	HDAC class IIa (HDAC4, 5, 7, and 9)
Compound 9 (Mai et al., <i>J.Med.Chem.</i> , 2005; 48:3344)	HDAC class IIa (HDAC4, 5, 7, and 9)
Compound 24 (Mai et al., <i>J.Med.Chem.</i> , 2005; 48:3344)	HDAC class IIa (HDAC4, 5, 7, and 9)
TC-H 106 (<i>N</i> -(2-Aminophenyl)- <i>N</i> -(4-methylphenyl)heptanediamide; Pimelic Diphenylamide 106)	HDAC class I (HDAC1, 2, 3, and 8)
Pyroxamide (<i>N</i> -Hydroxy- <i>N'</i> -3-pyridinyloctanediamide)	HDAC1
NCH 51 (PTACH; 2-Methylpropanethioic acid <i>S</i> -[7-oxo-7-[(4-phenyl-2-thiazolyl)amino]heptyl] ester)	HDAC

Compound	Target
NCH 31	HDAC
PCI 34051 (<i>N</i> -Hydroxy-1-[(4-methoxyphenyl)methyl]-1 <i>H</i> -indole-6-carboxamide)	HDAC8
thiophene benzamide	HDAC1 and HDAC2
KD 5170 (<i>S</i> -[2-[6-[[[4-[3-(Dimethylamino)propoxy]phenyl]sulfonyl]amino]-3-pyridinyl]-2-oxoethyl]ethanethioic acid ester)	HDAC class I (HDAC1, 2, 3, and 8) and HDAC class II (IIa: HDAC4, 5, 7, and 9; IIb: 6 and 10)
TCS HDAC6 20b (2-Methylpropanethioic acid- <i>S</i> -[(6 <i>S</i>)-6-[[1,1-dimethylethoxy]carbonyl]amino]-7-oxo-7-(tricyclo[3.3.1.1 ^{3,7}]dec-1-ylamino)heptyl ester)	HDAC6
NSC 3852 (5-Nitroso-8-quinolinol)	HDAC
NSC69603	HDAC
NSC86371	HDAC
NSC305819	HDAC
CI 994 (<i>N</i> -acetyldinaline; Acetyldinaline; 4-(Acetylamino)- <i>N</i> -(2-aminophenyl)benzamide)	HDAC class I
LAQ824	HDAC class I
LBH589 (panobinostat; Farydak)	pan-HDAC
MS275 (SNDX-275; entinostat)	HDAC1-3
MGCD0103 (mocetinostat)	HDAC1-8 and 11
UF 010 (4-Bromo- <i>N</i> '-butylbenzohydrazide)	HDAC1-3
Cpd60	HDAC1-3
Romidepsin	HDAC1 and HDAC2
MS-27-275	HDAC
NaBu (n-butyrate)	HDAC
trapoxin	HDAC
Apicidin (Cyclo[(2 <i>S</i>)-2-Amino-8-oxodecanoyl-1-methoxy-L-tryptophyl-L-isoleucyl-(2 <i>R</i>)-2-piperidinecarbonyl])	HDAC
depudesin	HDAC
EX 527 (6-Chloro-2,3,4,9-tetrahydro-1 <i>H</i> -carbazole-1-carboxamide)	SIRT1
AGK 2 (2-Cyano-3-[[5-(2,5-dichlorophenyl)-2-furanyl]- <i>N</i> -5-quinolinyl-2-propenamide)	SIRT2

Compound	Target
AK 7 (<i>N</i> -(3-Bromophenyl)-3-[(hexahydro-1 <i>H</i> -azepin-1-yl)sulfonyl]benzamide)	SIRT2
SirReal2 (2-[(4,6-Dimethyl-2-pyrimidinyl)thio]- <i>N</i> -[5-(1-naphthalenylmethyl)-2-thiazolyl]acetamide)	SIRT2
Salermide (<i>N</i> -[3-[[2-Hydroxy-1-naphthalenyl)methylene]amino]phenyl]- α -methylbenzeneacetamide)	SIRT1 and SIRT2
Splitomicin (1,2-Dihydro-3 <i>H</i> -naphtho[2,1- <i>b</i>]pyran-3-one)	Sir2p (yeast form of SIRT1)

Table G

Compound	Target
MG132 (<i>Z</i> -LLL-al, <i>Z</i> -Leu-Leu-Leu-CHO; <i>N</i> -[(Phenylmethoxy)carbonyl]-L-leucyl- <i>N</i> -[(1 <i>S</i>)-1-formyl-3-methylbutyl]-L-leucinamide)	proteasome
MG262 (<i>Z</i> -Leu-Leu-Leu-B(OH) ₂)	proteasome
MG115 (<i>Z</i> -Leu-Leu- <i>N</i> va-CHO)	proteasome
<i>Z</i> -Leu-Leu-Phe-CHO (<i>Z</i> -LLF-CHO)	proteasome
<i>N</i> -Acetyl-leucyl-leucyl-norleucinal (Ac-Leu-Leu- <i>N</i> le-CHO)	proteasome
<i>N</i> -acetyl-leucyl-leucyl-methional (Ac-Leu-Leu-Met-CHO)	proteasome
<i>N</i> -benzyloxycarbonyl-isoleucyl- γ - <i>t</i> -butyl-glutamyl-alanyl-leucinal (<i>Z</i> -Ile-Glu(O <i>t</i> Bu)-Ala-Leu-CHO)	proteasome
<i>N</i> -benzyloxycarbonyl-leucyl-leucyl-leucinal (<i>Z</i> -Leu-Leu-Leu-CHO),	proteasome
<i>N</i> -benzyloxycarbonyl-leucyl-leucyl-tyrosyl α -keto aldehyde (<i>Z</i> -Leu-Leu-Tyr-COCHO)	proteasome
<i>N</i> -benzyloxycarbonyl-leucyl-leucyl-phenylalanyl (<i>Z</i> -Leu-Leu-Phe-CHO)	proteasome
<i>N</i> -benzyloxycarbonyl-leucyl-leucyl-leucyl boronic acid (<i>Z</i> -Leu-Leu-Leu-B(OH) ₂)	proteasome
Bortezomib (PS-341; Velcade; Neomib; Bortecad)	proteasome
Lactacystin ((<i>2R,3S,4R</i>)-3-Hydroxy-2-[(1 <i>S</i>)-1-hydroxy-2-methylpropyl]-4-methyl-5-oxo-2-pyrrolidinecarboxy- <i>N</i> -acetyl-L-cysteine thioester)	proteasome

Compound	Target
Disulfiram (Antabuse and Antabus)	proteasome
Epigallocatechin-3-gallate (Epigallocatechin gallate; EGCG)	proteasome
Salinosporamide A	proteasome
Carfilzomib (Kyprolis)	proteasome
epoxomicin	proteasome
Ixazomib (Ninlaro; MLN2238)	proteasome
ixazomib citrate (MLN9708)	proteasome
PS-341	proteasome
VLX1500 (b-AP15)	proteasome
clasto-Lactacystin beta Lactone	proteasome
Gliotoxin (Aspergillin; (3 <i>R</i> ,5 <i>aS</i> ,6 <i>S</i> ,10 <i>aR</i>)-2,3,5 <i>a</i> ,6-Tetrahydro-6-hydroxy-3-(hydroxymethyl)-2-methyl-10 <i>H</i> -3,10 <i>a</i> -epidithiopyrazino[1,2- <i>a</i>]indole-1,4-dione)	proteasome
AM 114 (3,5-Bis-[benzylidene-4-boronic acid]-1-methylpiperidin-4-one)	proteasome
PSI (<i>N</i> -[(Phenylmethoxy)carbonyl]-L-isoleucyl-L- α -glutamyl- <i>tert</i> -butyl ester- <i>N</i> -[(1 <i>S</i>)-1-formyl-3-methylbutyl]-L-alaninamide)	proteasome
Oprozomib (ONX 0912)	proteasome
Delanzomib (CEP-18770)	proteasome
BI8622	Huwei (E3 ubiquitin ligase)
BI8626	Huwei (E3 ubiquitin ligase)

Table H

Compound	Target
MLN4929 (Pevonedistat)	Akt
API-2 (Triciribine; NSC 154020; TCN; 1,5-Dihydro-5-methyl-1- β -D-ribofuranosyl-1,4,5,6,8-pentaazaacenaphthylen-3-amine; Akt/protein kinase B signaling inhibitor-2)	Akt
API-1 (4-Amino-5,8-dihydro-5-oxo-8- β -D-ribofuranosyl-pyrido[2,3- <i>a</i>]pyrimidine-6-carboxamide)	Akt

Compound	Target
GSK 690693 (4-[2-(4-Amino-1,2,5-oxadiazol-3-yl)-1-ethyl-7-[(3 <i>S</i>)-3-piperidinylmethoxy]-1 <i>H</i> -imidazo[4,5- <i>c</i>]pyridin-4-yl]-2-methyl-3-butyn-2-ol)	Akt
10-DEBC hydrochloride (10-[4'-(<i>N,N</i> -Diethylamino)butyl]-2-chlorophenoxazine hydrochloride)	Akt
FPA124 (Dichloro[(2 <i>Z</i>)-2-[(4-oxo-4 <i>H</i> -1-benzopyran-3-yl)methylene]hydrazinecarbothioamide copper complex)	Akt
SC66 ((2 <i>E</i> ,6 <i>E</i>)-2,6- <i>Bis</i> (4-pyridinylmethylene)cyclohexanone)	Akt
LY 294002 hydrochloride (2-(4-Morpholinyl)-8-phenyl-4 <i>H</i> -1-benzopyran-4-one hydrochloride)	PI3K
wortmannin	PI3K
PI 103	PI3K
Quercetin (2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-4 <i>H</i> -1-benzopyran-4-one)	PI3K and PKC
PHT 427 (4-Dodecyl- <i>N</i> -1,3,4-thiadiazol-2-yl-benzenesulfonamide)	Akt and PDK1
GSK 2334470 ((3 <i>S</i> ,6 <i>R</i>)-1-[6-(3-Amino-1 <i>H</i> -indazol-6-yl)-2-(methylamino)-4-pyrimidinyl]- <i>N</i> -cyclohexyl-6-methyl-3-piperidinecarboxamide)	PDK1
Fisetin (2-(3,4-Dihydroxyphenyl)-3,7-dihydroxy-4 <i>H</i> -1-benzopyran-4-one)	PI3K, Akt
OSU 03012 (2-Amino- <i>N</i> -[4-[5-(2-phenanthrenyl)-3-(trifluoromethyl)-1 <i>H</i> -pyrazol-1-yl]phenyl]acetamide)	Akt and PDK1
PIT 1 (<i>N</i> -[[3-Chloro-2-hydroxy-5-nitrophenyl]amino]thioxomethyl]benzamide)	Akt

Table I

Compound	Target
AC102 (6-fluoro-9-methyl- β -carboline; 6F9M β C)	CREB

Table J

Compound	Target
LY411575 (LSN-411575; Compound 5; benzeneacetamide; N-[(1S)-2-[[[(7S)-6,7-dihydro-5-methyl-6-oxo-5H-dibenz[b,d]azepin-7-yl]amino]-1-methyl-2-oxoethyl]-3,5-difluoro- α -hydroxy-(α S)-); N2-[(2S)-2-(3,5-Difluorophenyl)-2-hydroxyethanoyl]-N1-[(7S)-5-methyl-6-oxo-6,7-dihydro-5H-dibenzo[b,d]azepin-7-yl]-L-alaninamide)	γ -secretase
L-685458 (<i>(5S)</i> -(<i>tert</i> -Butoxy carbonylamino)-6-phenyl-(<i>4R</i>)-hydroxy-(<i>2R</i>)-benzylhexanoyl)-L-leucyl-L-phenylalaninamide; LY-685458; GSI-X)	γ -secretase
DBZ (Dibenzazepine; YO-01027; GSI-XX, deshydroxy LY-411575; N-[(1S)-2-[[[(7S)-6,7-Dihydro-5-methyl-6-oxo-5H-dibenz[b,d]azepin-7-yl]amino]-1-methyl-2-oxoethyl]-3,5-difluorobenzeneacetamide)	γ -secretase
MRK560 (<i>N</i> -[<i>cis</i> -4-[(4-Chlorophenyl)sulfonyl]-4-(2,5-difluorophenyl)cyclohexyl]-1,1,1-trifluoromethanesulfonamide)	γ -secretase
MRK-003	γ -secretase
MK-0752	γ -secretase
Compound W (CW; 3,5-Bis(4-nitrophenoxy)benzoic acid) (Okochi et al., <i>J.Biol.Chem.</i> , 2006; 281:7890; Ford et al., <i>J Neurosci Meth.</i> , 2008; 168:465-474)	γ -secretase
Compound E (GSI-XXI) (Olsauskas-Kuprys et al., <i>Onco Targets Ther.</i> , 2013; 6:943)	γ -secretase
BMS 2289948 (4-chloro-N-(2,5-difluorophenyl)-N-((1R)-{4-fluoro-2-[3-(1H-imidazol-1-yl)propyl]phenyl}ethyl)benzenesulfonamide hydrochloride)	γ -secretase
BMS-433796 (<i>(S)</i> -2-((<i>S</i>)-2-(3,5-difluorophenyl)-2-hydroxyacetamido)- <i>N</i> -((<i>S,Z</i>)-3-methyl-4-oxo-4,5-dihydro-3 <i>H</i> -benzo[d][1,2]diazepin-5-yl)propanamide)	γ -secretase
IN973	γ -secretase
Flurbiprofen bi(<i>(R)</i> -Flurbiprofen; tarenflurbil; Flurizan; (<i>R</i>)-2-Fluoro- α -methyl[1,1'-biphenyl]-4-acetic acid)	γ -secretase

Compound	Target
JLK2, JLK4, JLK6, JLK7 (7-Amino-4-chloro-3-methoxy-1 <i>H</i> -2-benzopyran)	γ -secretase
Begacestat (GSI-953; 5-Chloro- <i>N</i> -[(1 <i>S</i>)-3,3,3-trifluoro-1-(hydroxymethyl)-2-(trifluoromethyl)propyl]-2-thiophenesulfonamide)	γ -secretase
DFK167	γ -secretase
PF-0308414	γ -secretase

Table K

Compound	Target
TTNBP (RO 13-7410, arotinoid acid, AGN 191183)	RAR
ATRA	RAR
9-cis RA	RAR
CD271 (6-(4-Methoxy-3-tricyclo[3.3.1.1 ^{3,7}]dec-1-ylphenyl)-2-naphthalenecarboxylic acid)	RAR
CD336	RAR
CD-394	RAR
CD437 (6-3-(1-adamantyl)-4-hydroxyphenyl)-2 naphthanoic acid) (6-(4-Hydroxy-3-tricyclo[3.3.1.1 ^{3,7}]dec-1-ylphenyl)-2-naphthalenecarboxylic acid)	RAR
CD666 (<i>E</i>)-4-(1-hydroxy-1-(5,6,7,8-tetrahydro-5,5,8,8-tetramethyl-2-naphthyl)-2-propenyl)benzoic acid)	RAR
CD1530 (4-(6-Hydroxy-7-tricyclo[3.3.1.1 ^{3,7}]dec-1-yl-2-naphthalenyl)benzoic acid)	RAR
CD2019 (6-(3-(1-methylcyclohexyl)-4-methoxyphenyl)-2 naphthanoic acid)	RAR
CD2247	RAR
CD2081	RAR
CD2314	RAR
CD2325 (4-[(<i>E</i>)-2-(3-(1-adamantyl)-4-hydroxyphenyl)-1-propenyl]benzoic acid)	RAR
CD2425	RAR
CD2503	RAR
CD2665	RAR
BMS-270394 (enantiomer of BMS-189961) (3-Fluoro-4-[(<i>R</i>)-2-hydroxy-2-(5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-naphthalen-2-yl)-acetylaminol-benzoic acid)	RAR
BMS-189961 (3-Fluoro-4-[2-hydroxy-2-(5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-naphthalen-2-yl)-acetylaminol]-benzoic acid)	RAR

Compound	Target
6-[3-(adamantan-1-yl)-4-(prop-2-ynyloxy)phenyl]naphthalene-2-carboxylic acid	RAR
5-[(E)-3-oxo-3-(5,5,8,8-tetrahydronaphthalene-2-yl)propenyl]thiophene-2-carboxylic acid	RAR
Palovarotene (4[(1E)-2-[5,6,7,8-Tetrahydro-5,5,8,8-tetramethyl-3-(1H-pyrazol-1-ylmethyl)-2-naphthalenyl]-ethenyl]-benzoic acid; R667; CLM-001, RG667)	RAR
CH-55 (4-[(E)-3-(3,5-Di-tert-butyl-phenyl)-3-oxo-propenyl]-benzoic acid)	RAR
Docosahexaenoic acid (DHA; (4Z,7Z,10Z,13Z,16Z,19Z)-4,7,10,13,16,19-Docosahexaenoic acid)	RXR
CD 3254 (3-[4-Hydroxy-3-(5,6,7,8-tetrahydro-3,5,5,8,8-pentamethyl-2-naphthalenyl)phenyl]-2-propenoic acid)	RXR
9 <i>cis</i> -RA	RXR
3- <i>cis</i> -retinoic acid (Accutane; isotretinoin; 13- <i>cis</i> -Retinoic acid)	RXR
LG 100754 ((2E,4E,6Z)-3-Methyl-7-(5,6,7,8-tetrahydro-5,5,8,8-tetramethyl-3-propoxy-3-naphthalenyl)-2,4,6-octatrienoic acid)	RXR
SR 11237 (BMS 649; 4-[2-(5,6,7,8-Tetrahydro-5,5,8,8-tetramethyl-2-naphthalenyl)-1,3-dioxolan-2-yl]-benzoic acid)	RXR
Fluorobexarotene (2-Fluoro-4-[1-(5,6,7,8-tetrahydro-3,5,5,8,8-pentamethyl-2-naphthalenyl)ethenyl]benzoic acid)	RXR
LGD1069 (4-[1-(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthyl)ethenyl]benzoic acid)	RXR
LG100268 (6-[1-(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydronaphthalen-2-yl)cyclopropyl]nicotinic acid)	RXR
LG100754 (2E,4E,6Z)-3-Methyl-7-(5,6,7,8-tetrahydro-5,5,8,8-tetramethyl-3-propoxy-2-naphthalenyl)-2,4,6-Octatrienoic acid)	RXR
Compounds 1-11 (Wagner et al., <i>J.Med.Chem.</i> , 2009; 52:5950)	RXR
HX 630 (4-(7,8,9,10-Tetrahydro-7,7,10,10-tetramethylbenzo[<i>b</i>]naphtho[2,3- <i>f</i>][1,4]thiazepin-12-yl)-benzoic acid)	RXR
HX 640	RXR
HX 600	RXR

Compound	Target
TZ335	RXR
Adapalene (6-(4-Methoxy-3-tricyclo[3.3.1.1 ^{3,7}]dec-1-ylphenyl)-2-naphthalenecarboxylic acid, 6-[3-(1-Adamantyl)-4-methoxyphenyl]-2-naphthoic acid; CD-271; Differin)	RXR
Bexarotene (4-[1-(5,6,7,8-Tetrahydro-3,5,5,8,8-pentamethyl-2-naphthalenyl)ethenyl]benzoic acid, LGD-1069; SR-11247; targretin; TRG)	RXR
Retinoic acid (ATRA; Tretinoin; Vitamin A acid; all- <i>trans</i> -Retinoic acid)	RXR
4-[N-methanesulfonyl-N-(5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-2-naphthyl)amino]benzoic acid	RXR
6-[N-ethyl-N-(3-isopropoxy-4-isopropylphenyl)amino]nicotinic acid (NEt-3IP)	RXR
6-[N-ethyl-N-(3-isobutoxy-4-isopropylphenyl)amino]nicotinic acid (NEt-3IB)	RXR
PA024	RXR
AGN 194204	RXR
CNX-013-B2	RXR
UAB30	RXR
IRX4204	RXR

In some embodiments, the one or more agents used to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) include a combination of agents, in which the agents target two or more (e.g., three, four, five, or more) of the following pathways, proteins, and DNA response elements: the retinoid receptor signaling pathway, the Wnt signaling pathway, the BMP signaling pathway, the CDK signaling pathway, the Notch signaling pathway, the protein degradation pathway, the PI3K-Akt signaling pathway, the cAMP-dependent pathway, histone deacetylase (HDAC), and/or E box DNA response element.

In some embodiments, the one or more agents used to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) include a combination of agents (e.g., a combination two agents each selected from the compounds set forth in Tables A-K, wherein the two agents are different

from each other; a combination of three agents each selected from the compounds set forth in Tables A-K, wherein the three agents are different from each other; a combination of four agents each selected from the compounds set forth in Tables A-K, wherein the four agents are different from each other; and a combination of five agents each selected from the compounds set forth in Tables A-K, wherein the five agents are different from each other).

For example, if a combination of two agents each selected from the compounds set forth in Tables A-K is used to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells), the two agents in the combination may be selected based on the combinations listed in Table L.

Table L

Combination	Agent 1 chosen from	Agent 2 chosen from	Combination	Agent 1 chosen from	Agent 2 chosen from
1	Table A	Table A	34	Table D	Table G
2	Table A	Table B	35	Table D	Table H
3	Table A	Table C	36	Table D	Table I
4	Table A	Table D	37	Table D	Table J
5	Table A	Table E	38	Table D	Table K
6	Table A	Table F	39	Table E	Table E
7	Table A	Table G	40	Table E	Table F
8	Table A	Table H	41	Table E	Table G
9	Table A	Table I	42	Table E	Table H
10	Table A	Table J	43	Table E	Table I
11	Table A	Table K	44	Table E	Table J
12	Table B	Table B	45	Table E	Table K
13	Table B	Table C	46	Table F	Table F
14	Table B	Table D	47	Table F	Table G
15	Table B	Table E	48	Table F	Table H
16	Table B	Table F	49	Table F	Table I
17	Table B	Table G	50	Table F	Table J
18	Table B	Table H	51	Table F	Table K

Combination	Agent 1 chosen from	Agent 2 chosen from	Combination	Agent 1 chosen from	Agent 2 chosen from
19	Table B	Table I	52	Table G	Table G
20	Table B	Table J	53	Table G	Table H
21	Table B	Table K	54	Table G	Table I
22	Table C	Table C	55	Table G	Table J
23	Table C	Table D	56	Table G	Table K
24	Table C	Table E	57	Table H	Table H
25	Table C	Table F	58	Table H	Table I
26	Table C	Table G	59	Table H	Table J
27	Table C	Table H	60	Table H	Table K
28	Table C	Table I	61	Table I	Table I
29	Table C	Table J	62	Table I	Table J
30	Table C	Table K	63	Table I	Table K
31	Table D	Table D	64	Table J	Table J
32	Table D	Table E	65	Table J	Table K
33	Table D	Table F	66	Table K	Table K

Alternatively, the invention also contemplates methods that use one or more proteins involved in the retinoid receptor signaling pathway, the Wnt signaling pathway, the BMP signaling pathway, the CDK signaling pathway, the Notch signaling pathway, the protein degradation pathway, the PI3K-Akt signaling pathway, and/or the cAMP-dependent pathway to down-regulate or up-regulate the pathway to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells).

In some embodiments of the methods, RAR and/or RXR may be used to up-regulate retinoid receptor signaling,

In some embodiments of the methods, RSPO, Norrin, Wnt3a, and/or Wnt5a may be used to up-regulate the Wnt signaling pathway.

In some embodiments of the methods, Noggin and/or Chordin may be used to down-regulate the BMP signaling pathway.

In some embodiments of the methods, CDKs and/or cyclins may be used to up-regulate the CDK signaling pathway.

In some embodiments of the methods, Delta/Serrate/Lag-2 peptide and/or Notch receptors may be used to up-regulate the Notch signaling pathway.

5 In some embodiments of the methods, the level of ubiquitin may be decreased to down-regulate the protein degradation pathway.

In some embodiments of the methods, the level of Akt, PI 3-kinase, and/or PDK1 may be decreased to down-regulate the PI3K-Akt signaling pathway.

10 In some embodiments of the methods, the level of CREB protein may be increased to up-regulate cAMP-dependent pathway.

In some embodiments of the methods, the level of one or more transcription factors that bind to the E box DNA response element may be increased to up-regulate E box-dependent transcription.

15 In some embodiments of the methods, the level of histone deacetylase (HDAC) may be decreased to down-regulate HDAC activity.

Alternatively, the invention also contemplates methods that use one or more growth factors to down-regulate or up-regulate one or more of the following pathways: retinoid receptor signaling pathway, the Wnt signaling pathway, the BMP signaling pathway, the CDK signaling pathway, the Notch signaling pathway, the protein degradation pathway, the PI3K-Akt signaling pathway, and/or the cAMP-dependent pathway, in order to promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) or to promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells).

25 In some embodiments of the methods, the growth factors include, but are not limited to, epidermal growth factor (EGF), basic fibroblast growth factor (bFGF), and/or insulin-like growth factor (IGF1).

Transgenic Animals and Methods of Use

30 In one aspect, the invention provides non-human transgenic animals having two or more (e.g., two, three, or four or more) recombinant nucleic acid molecules stably integrated into their genome. The two or more recombinant nucleic acid molecules include at least a first recombinant nucleic acid molecule that comprises a

first reporter gene (e.g., a fluorescent marker) under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glis3, and Lfng (e.g., Lgr5), and a second recombinant nucleic acid molecule that comprises a second reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of Atoh1, Myo7a, Cdh23, Pcdh15, Myo6, Myo1c, Tmc1, and Cav1.3 (e.g., Atoh1), wherein the first reporter gene is different from the second reporter gene.

The invention also contemplates using various genetic engineering techniques to generate one or more reporters in a cell or a transgenic animal. In some embodiments, various genetic engineering techniques can be used to generate two or more reporters in a cell or a transgenic animal. Examples of genetic engineering techniques include, but are not limited to, techniques that use the CRISPR/Cas system, techniques that use the Cre recombinase-loxP recombination system, techniques that use the Cre-Lox recombination system, techniques that use the Flp-*FRT* recombination system, and techniques that use the RMCE (recombinase-mediated cassette exchange) system,

In some embodiments, the inner ear supporting cell marker is Lgr5 and the inner ear hair cell marker is Atoh1. In some embodiments, the regulatory element of an inner ear supporting cell marker is an Lgr5 promoter. In some embodiments, the regulatory element of an inner ear hair cell marker is an Atoh1 enhancer. In some embodiments, the Atoh1 enhancer is operably linked to an SV40 promoter or a globin promoter.

In some embodiments, the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a second fluorescent protein, in which the first fluorescent protein is different from the second fluorescent protein.

In some embodiments, the expression of Lgr5 results in expression of the first fluorescent marker (Lgr5 reporter transgene). In some embodiments, the expression of Atoh1 results in expression of the second fluorescent marker (Atoh1 reporter transgene). In preferred embodiments a mouse is obtained that contains the Lgr5 reporter protein and Atoh1 reporter protein transgenes in all of its somatic and germ cells. The first and second markers should be distinguishable from each other. In some embodiments, the first and second markers produce green and red fluorescence

in the cells. Although fluorescent markers are exemplified herein, other markers (reporter genes) can also be used; Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β -galactosidase, or acetylcholinesterase; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; examples of bioluminescent materials include luciferase, luciferin, and aequorin. Numerous others are known in the art. In some embodiments, one of the markers is green fluorescent protein or a derivative thereof, fluorescent proteins (e.g., green fluorescent protein (GFP), cyan fluorescent protein (CFP), red fluorescent protein (RFP), mCherry, Tag-RFP, etc.), luciferase which is a luminescent reporter (Ranella, Firefly, etc.), chomogenic (beta-Gal, etc.), etc. See e.g., Pollock et al., Trends in Cell Biology 9:57 (1999). Useful fluorescent proteins also include mutants and spectral variants of these proteins which retain the ability to fluoresce. See e.g., Shaner et al., Nat. Biotech. 22:1567 (2004), Tag-RFP (Shaner, N. C. et al., 2008 Nature Methods, 5(6), 545-551), Other fluorescent proteins that can be used in the methods described include, but are not limited to, AcGFP, AcGFP1, AmCyan, copGFP, CyPet, dKeima-Tandem, DsRed, dsRed-Express, DsRed-Monomer, DsRed2, AmCyan1, AQ143, AsRed2, Azami Green, Azurite, BFP, Cerulean, CFP, CGFP, Citrine, dTomato, dTomato-Tandem, EBFP, EBFP2, ECFP, EGFP, Emerald, EosFP, EYFP, GFP, mBanana, mCerulean, mCFP, mCherry, mCitrine, mECFP, mEmerald, mGrape1, mGrape2, mHoneydew, Midori-Ishi Cyan, mKeima, mKO, mOrange, mOrange2, mPlum, mRaspberry, mRFPI, mRuby, mStrawberry, mTagBFP, mTangerine, mTeal, mTomato, mTurquoise, mWasabi, PhiYFP, ReAsH, Sapphire, Superfolder GFP, T- HcRed-Tandem, HcRed1, JRed, Katuska, Kusabira Orange, Kusabira Orange2, mApple, Sapphire, TagCFP, TagGFP, TagRFP, TagRFP-T, TagYFP, tdTomato, Topaz, TurboGFP, Venus, YFP, YPet, ZsGreen, and ZsYellow1, all of which are known in the art, e.g., described in the literature or otherwise commercially available.

A “transgenic animal” is a non-human animal, such as a mammal, generally a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene as described herein. Other examples of transgenic animals include non-human primates, sheep, dogs, cows, goats, chickens, amphibians, and the like. A “transgene” is exogenous DNA that is integrated into the genome of a cell from which

a transgenic animal develops and thus remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. Knock-in animals, which include a gene insertion, are included in the definition of transgenic animals.

5 A “Lgr5 reporter transgene” or “Atoh1 reporter transgene” as used herein refers to a construct that features a coding sequence for a reporter protein inserted downstream of an Lgr5 or Atoh1 promoter, so as to result in expression of the reporter protein in cells expressing Lgr5 or Atoh1. The promoter drives expression of the reporter protein, and transcription is stopped by a polyadenylation signal. The
10 transgene is generally integrated into or occurs in the genome of the cells of a transgenic animal. Thus an Lgr5/Atoh1 transgenic animal as described herein is one in which at least one copy of an Lgr5 reporter transgene and at least one copy of an Atoh1 reporter transgene have been introduced into a cell of the animal, e.g., an embryonic cell of the animal, prior to development of the animal. A line of transgenic
15 animals (e.g., mice, rats, guinea pigs, hamsters, rabbits, or other mammals) can be produced bearing an Lgr5 reporter transgene and an Atoh1 reporter transgene in some or (preferably) all of their cells. Methods known in the art for generating such transgenic animals would be used, e.g., as described below.

 Methods known in the art for producing transgenic animals can be used to
20 generate an animal, e.g., a mouse, which bears an Lgr5 reporter transgene or an Atoh1 reporter transgene (see, e.g., Barker et al., *Nature*. 2007 Oct 25;449(7165):1003 and Lumpkin et al., *Gene Expr Patterns*. 2003 Aug;3(4):389, both of which are incorporated herein in their entirety). Such animals can be crossed to produce offspring that are homozygous for both the Lgr5 reporter transgene and Atoh1
25 reporter transgene, i.e., that have the Lgr5 reporter transgene and Atoh1 reporter transgene integrated into the genome.

 For example, in one embodiment, a suitable vector including a sequence encoding the Lgr5 reporter transgene or Atoh1 reporter transgene is introduced into a cell, e.g., a fertilized oocyte or an embryonic stem cell. Such cells can then be used to
30 create non-human transgenic animals in which said sequences have been introduced into their genome. These animals can then in turn be bred with other transgenic animals that express a recombinase, e.g., under the control of an Lgr5 or Atoh1

promoter that will turn on expression of the reporter protein in a specific cell or tissue, or at a specific time in development.

5 Methods for generating transgenic animals, particularly animals such as mice, via embryo manipulation and electroporation or microinjection of pluripotent stem cells or oocytes, are known in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, U.S. Patent No. 4,873,191, U.S.S.N. 10/006,611, “Transgenic Mouse Methods and Protocols (Methods in Molecular Biology),” Hofker and van Deursen, Editors (Humana Press, Totowa, N.J., 2002); and in “Manipulating the Mouse Embryo,” Nagy et al., Editors (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 2002), which are incorporated herein by reference in their
10 entirety. Methods similar to those used to create transgenic mice can be used for production of other transgenic animals.

In general, in the present methods, a transgenic mouse as described herein is made by injecting a vector made as described herein into the pronucleus of a fertilized
15 mouse oocyte (e.g., an oocyte from a mouse with an *Lgr5* reporter gene knocked in, see Barker et al., *Nature*. 2007 Oct 25;449(7165):1003) and used for generation of a transgenic mouse with the *Lgr5* reporter gene and *Atoh1* reporter transgene expressed in all cells, using standard transgenic techniques, e.g., as described in “Transgenic Mouse Methods and Protocols (Methods in Molecular Biology),” Hofker and van
20 Deursen, Editors (Humana Press, Totowa, N.J., 2002); U.S. Patent Nos. 4,736,866 and 4,870,009, U.S. Patent Nos. 4,873,191 and 6,791,006, and in Hogan, “Manipulating the Mouse Embryo,” Nagy et al., Editors (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 2002). The reporter genes can be maintained and expressed in all cells, e.g., on plasmids or stably integrated into the
25 genome, using standard molecular techniques.

A transgenic founder *Lgr5/Atoh1* animal can be identified based upon the presence of the *Lgr5* reporter transgene and *Atoh1* reporter transgene in its genome, for example by detecting the presence or expression of the reporter sequences or proteins in tissues or cells of the animals. A transgenic founder animal can then be
30 used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying the *Lgr5* reporter transgene and *Atoh1* reporter transgene can further be bred to other transgenic animals carrying other transgenes.

Vectors

The mice described herein can be made using vectors, e.g., expression vectors, containing a nucleic acid encoding the Lgr5 reporter transgene or Atoh1 reporter transgene as described herein. As used herein, the term “vector” refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked and can include a plasmid, cosmid or viral vector. The vector can be capable of autonomous replication or it can integrate into a host DNA. Viral vectors include, e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses.

A vector can include a nucleic acid encoding a Lgr5 reporter protein or Atoh1 reporter protein in a form suitable for expression of the nucleic acid in a host cell. Preferably the recombinant expression vector includes one or more regulatory sequences operatively linked to the nucleic acid sequence to be expressed. The term “regulatory sequence” includes promoters, enhancers and other expression control elements (e.g., polyadenylation signals). Regulatory sequences include those which direct constitutive expression of a nucleotide sequence, as well as tissue-specific regulatory and/or inducible sequences. The design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, and the like. The expression vectors of the invention can be introduced into host cells to thereby produce the Lgr5 reporter and Atoh1 reporter, encoded by nucleic acids as described herein.

The recombinant expression vectors described herein can be designed for expression of the Lgr5 reporter and Atoh1 reporter proteins in prokaryotic or eukaryotic cells. For example, polypeptides of the invention can be expressed in *E. coli*, insect cells (e.g., using baculovirus expression vectors), yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, “Gene Expression Technology: Methods in Enzymology 185,” Academic Press, San Diego, CA (1990). Alternatively, the recombinant expression vector can be transcribed and translated in vitro, for example using T7 promoter regulatory sequences and T7 polymerase.

In some embodiments, the Atoh1 enhancer described in Example 5 is used to drive expression of a reporter gene.

Cells

In another aspect, the invention provides isolated cells that include a nucleic acid molecule as described herein, e.g., a nucleic acid molecule encoding an Lgr5 reporter protein or Atoh1 reporter protein within a recombinant expression vector, or a nucleic acid molecule containing sequences that allow it to homologously recombine into a specific site of the host cell's genome. The terms "host cell" and "recombinant host cell" are used interchangeably herein. Such terms refer not only to the particular subject cell that was contacted with a nucleic acid molecule (e.g., a vector as described herein), but to the progeny or potential progeny of such a cell that also contain the nucleic acid molecule. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein so long as they also contain the nucleic acid molecule.

A host cell can be any prokaryotic or eukaryotic cell. For example, the cell can be a bacterial cell such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO), HEK, or COS cells). Other suitable host cells are known to those skilled in the art. Where the vector is a viral vector that can be produced from recombinant cells, e.g., retroviral vectors, the cells can be those that produce the viral vector.

Vector DNA can be introduced into host cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. In some embodiments, naked DNA is simply applied to a cell. Where the vector is a viral vector, known infection protocols can be used.

For example, retroviral vectors can be used, e.g., as described in Robertson et al., *Nature* 323:445-448 (1986). Retroviruses generally integrate into the host genome with no rearrangements of flanking sequences, which is not always the case when DNA is introduced by microinjection or other methods.

Cells of the present invention also include those cells obtained from the transgenic animals described herein, e.g., cells from the tissues of those animals, that contain the nucleic acid molecule.

Identity of Sequences

5 To determine the percent identity of two amino acid sequences, or of two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). The length of a reference sequence aligned for
10 comparison purposes is at least 80% of the length of the reference sequence, and in some embodiments is at least 90%, 95% or 100%. The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the
15 molecules are identical at that position. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

The comparison of sequences and determination of percent identity between
20 two sequences can be accomplished using a mathematical algorithm. For the present methods, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch ((1970) J. Mol. Biol. 48:444-453) algorithm, which has been incorporated into the GAP program in the GCG software package (available on the world wide web at gcg.com), using the default parameters, i.e., a Blossum 62
25 scoring matrix with a gap penalty of 12, a gap extend penalty of 4, and a frameshift gap penalty of 5.

Methods for constructing transgenes useful in the present methods are known in the art; see, e.g., Sambrook and Russell, "Molecular Cloning: A Laboratory Manual," Cold Spring Harbor Laboratory Press; 3rd Labman edition (January 15, 2001); and Ausubel et al., Eds., "Short Protocols in Molecular Biology," Current
30 Protocols; 5 edition (November 5, 2002). In some embodiments, commercially-available vectors can be used in constructing the nucleic acid molecules described herein, e.g., pC4M-Fv2E (available from Ariad Pharmaceuticals, Cambridge, MA).

Methods of Screening

Included herein are methods for screening test compounds, such as polypeptides, polynucleotides, inorganic or organic large or small molecule test compounds (e.g., compounds listed in Tables A-K) to identify agents useful in the treatment of hearing loss associated with a loss of cochlear hair cells (e.g., cochlear hair cells in the inner ear).

The present disclosure provides a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method includes: (a) isolating a population of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) from the transgenic mouse described herein; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells; (c) administering a test compound to the expanded population of inner ear supporting cells; (d) detecting the expression levels of the first reporter gene and the second reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the first reporter gene compared to the expression level of the first reporter gene in the absence of the test compound, and/or increases the expression level of the second reporter gene compared to the expression level of the second reporter gene in the absence of the test compound.

In some embodiments, the conditions sufficient to produce an expanded population of inner ear supporting cells comprise one or more agents set forth in Table 1.

The present disclosure also provides a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method includes: (a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that comprises a reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glaxt, and Lfng; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded

population of inner ear supporting cells, wherein the conditions comprise one or more agents selected from the group consisting of: (i) a retinoid receptor signaling activator, (ii) a Wnt signaling activator set forth in Table A, (iii) a BMP signaling inhibitor set forth in Table B, (iv) a CDK activator set forth in Table C, (v) an E box-
5 dependent transcriptional activator set forth in Table D, (vi) a Notch signaling activator set forth in Table E, (vii) an HDAC inhibitor set forth in Table F, (viii) a protein degradation inhibitor set forth in Table G, (ix) a PI3K-Akt signaling inhibitor set forth in Table H, and (x) a CREB activator set forth in Table I; (c) administering a test compound to the expanded population of inner ear supporting cells; (d) detecting
10 the expression level of the reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

15 The present disclosure also provides a method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, in which the method includes: (a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that comprises a reporter gene under the control of a regulatory
20 element of an inner ear hair cell marker selected from the group consisting of *Atoh1*, *Myo7a*, *Cdh23*, *Pcdh15*, *Myo6*, *Myo1c*, *Tmc1*, and *Cav1.3*; (b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells, wherein the conditions comprise one or more agents selected from the group consisting of: (i) a retinoid receptor
25 signaling activator, (ii) a Wnt signaling activator set forth in Table A, (iii) a BMP signaling inhibitor set forth in Table B, (iv) a CDK activator set forth in Table C, (v) an E box-dependent transcriptional activator set forth in Table D, (vi) a Notch signaling activator set forth in Table E, (vii) an HDAC inhibitor set forth in Table F, (viii) a protein degradation inhibitor set forth in Table G, (ix) a PI3K-Akt signaling
30 inhibitor set forth in Table H, and (x) a CREB activator set forth in Table I; (c) administering a test compound to the expanded population of inner ear supporting cells; (d) detecting the expression level of the reporter gene in the expanded population of inner ear cells in the presence of the test compound; and (e) selecting as

a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

5 The present disclosure also provides methods of using hair cells to screen for ototoxins. The present disclosure also provides methods of identifying one or more compounds (e.g., compounds listed in Tables A-K) that exhibit protective properties against hair cell damage caused by ototoxins. In some embodiments, ototoxins include therapeutic drugs including antineoplastic agents, salicylates, quinines, and aminoglycoside antibiotics, contaminants in foods or medicinals, and environmental
10 or industrial pollutants. The present disclosure also provides methods of using hair cells to identify synaptic connectivity. In some embodiments, the hair cells (e.g., Atoh1+ inner ear hair cells) used in these methods may be isolated from a mammal (e.g., a mouse or a human). In some embodiments, the hair cells (e.g., Atoh1+ inner ear hair cells) used in these methods may be differentiated from inner ear supporting
15 cells (e.g., Lgr5+ inner ear supporting cells), as described by the methods provided herein.

As used herein, “small molecules” refers to small organic or inorganic molecules of molecular weight below about 3,000 Daltons. In general, small molecules useful for the invention have a molecular weight of less than 3,000 Daltons
20 (Da). The small molecules can be, e.g., from at least about 100 Da to about 3,000 Da (e.g., between about 100 to about 3,000 Da, about 100 to about 2500 Da, about 100 to about 2,000 Da, about 100 to about 1,750 Da, about 100 to about 1,500 Da, about 100 to about 1,250 Da, about 100 to about 1,000 Da, about 100 to about 750 Da, about 100 to about 500 Da, about 200 to about 1500, about 500 to about 1000, about 300 to
25 about 1000 Da, or about 100 to about 250 Da).

The test compounds (e.g., compounds listed in Tables A-K and Table 1) can be, e.g., natural products or members of a combinatorial chemistry library. A set of diverse molecules should be used to cover a variety of functions such as charge, aromaticity, hydrogen bonding, flexibility, size, length of side chain, hydrophobicity,
30 and rigidity. Combinatorial techniques suitable for synthesizing small molecules are known in the art, e.g., as exemplified by Obrecht and Villalgorido, Solid-Supported Combinatorial and Parallel Synthesis of Small-Molecular-Weight Compound Libraries, Pergamon-Elsevier Science Limited (1998), and include those such as the

“split and pool” or “parallel” synthesis techniques, solid-phase and solution-phase techniques, and encoding techniques (see, for example, Czarnik, *Curr. Opin. Chem. Bio.* 1:60-6 (1997)). In addition, a number of small molecule libraries are commercially available. A number of suitable small molecule test compounds are listed in U.S. Patent No. 6,503,713, incorporated herein by reference in its entirety.

Libraries screened using the methods of the present invention can comprise a variety of types of test compounds (e.g., compounds listed in Tables A-K and Table 1). A given library can comprise a set of structurally related or unrelated test compounds. In some embodiments, the test compounds are peptide or peptidomimetic molecules. In some embodiments, the test compounds are nucleic acids.

In some embodiments, the test compounds and libraries thereof can be obtained by systematically altering the structure of a first test compound, e.g., a first test compound that is structurally similar to a known natural binding partner of the target polypeptide, or a first small molecule identified as capable of binding the target polypeptide, e.g., using methods known in the art or the methods described herein, and correlating that structure to a resulting biological activity, e.g., a structure-activity relationship study. As one of skill in the art will appreciate, there are a variety of standard methods for creating such a structure-activity relationship. Thus, in some instances, the work may be largely empirical, and in others, the three-dimensional structure of an endogenous polypeptide or portion thereof can be used as a starting point for the rational design of a small molecule compound or compounds. For example, in one embodiment, a general library of small molecules is screened, e.g., using the methods described herein.

In some embodiments, a test compound (e.g., a compound listed in Tables A-K and Table 1) is applied to a test sample, e.g., a cell or living tissue or organ, e.g., an eye, and one or more effects of the test compound is evaluated. In a cultured or primary cell for example, the ability of the test compound to increase expression of a supporting cell marker (e.g., *Lgr5*) and/or a hair cell marker (e.g., *Atoh1*).

In some embodiments, the test sample is, or is derived from (e.g., a sample taken from) an *in vivo* model of a disorder as described herein. For example, an animal model, e.g., a rodent such as a rat, can be used.

Methods for evaluating each of these effects are known in the art. For example, ability to modulate expression of a protein can be evaluated at the gene or protein level, e.g., using quantitative PCR or immunoassay methods. In some embodiments, high throughput methods, e.g., protein or gene chips as are known in the art (see, e.g., Ch. 12, Genomics, in Griffiths et al., Eds. Modern genetic Analysis, 1999, W. H. Freeman and Company; Ekins and Chu, Trends in Biotechnology, 1999, 17:217-218; MacBeath and Schreiber, Science 2000, 289(5485):1760-1763; Simpson, Proteins and Proteomics: A Laboratory Manual, Cold Spring Harbor Laboratory Press; 2002; Hardiman, Microarrays Methods and Applications: Nuts & Bolts, DNA Press, 2003), can be used to detect an effect on expression of a supporting cell marker (e.g., Lgr5) and/or a hair cell marker (e.g., Atoh1).

A test compound (e.g., a compound listed in Tables A-K and Table 1) that has been screened by a method described herein and determined to increase expression of Lgr5 and/or Atoh1 can be considered a candidate compound. A candidate compound that has been screened, e.g., in an in vivo model of a disorder, e.g., an animal model of hearing loss, and determined to have a desirable effect on the disorder, e.g., on one or more symptoms of the disorder or on number of hair cells, can be considered a candidate therapeutic agent. Candidate therapeutic agents, once screened in a clinical setting, are therapeutic agents. Candidate compounds, candidate therapeutic agents, and therapeutic agents can be optionally optimized and/or derivatized, and formulated with physiologically acceptable excipients to form pharmaceutical compositions.

Thus, test compounds identified as "hits" (e.g., test compounds that increase expression of Lgr5 and/or Atoh1) in a first screen can be selected and systematically altered, e.g., using rational design, to optimize binding affinity, avidity, specificity, or other parameter. Such optimization can also be screened for using the methods described herein. Thus, in one embodiment, the invention includes screening a first library of compounds using a method known in the art and/or described herein, identifying one or more hits in that library, subjecting those hits to systematic structural alteration to create a second library of compounds structurally related to the hit, and screening the second library using the methods described herein.

Test compounds identified as hits can be considered candidate therapeutic compounds, useful in treating disorders associated with loss of cochlear hair cells (e.g., cochlear hair cells in the inner ear), as described herein, e.g., hearing loss. A

variety of techniques useful for determining the structures of “hits” can be used in the methods described herein, e.g., NMR, mass spectrometry, gas chromatography equipped with electron capture detectors, fluorescence and absorption spectroscopy. Thus, the invention also includes compounds identified as “hits” by the methods
5 described herein, and methods for their administration and use in the treatment, prevention, or delay of development or progression of a disorder described herein.

Test compounds identified as candidate therapeutic compounds can be further screened by administration to an animal model of a disorder associated with loss of cochlear hair cells (e.g., cochlear hair cells in the inner ear), as described herein. The
10 animal can be monitored for a change in the disorder, e.g., for an improvement in a parameter of the disorder, e.g., a parameter related to clinical outcome. In some embodiments, the parameter is hearing ability, and an improvement would be improved hearing response. In some embodiments, the subject is a human, e.g., a human with hearing loss, and the parameter is improved hearing.

15

Methods of treatment

In some embodiments, the present disclosure provides novel therapeutic strategies for treating hearing loss or balance loss associated with a loss of cochlear hair cells (e.g., cochlear hair cells in the inner ear) or vestibular hair cells, respectively
20 (i.e., conditions that would benefit from an increased proliferation and differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells)). In some embodiments, such strategies can promote an increase in the proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) and/or an increase in the differentiation of the inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells)
25 into inner ear hair cells (e.g., Atoh1+ inner ear hair cells), thereby promoting the expansion and differentiation of a target cell into a mature cell of the inner ear, e.g., an auditory hair cell. In some embodiments, the methods and compositions described herein promote differentiation of target cells (e.g., inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells)) to or towards mature cells of the inner ear, e.g.,
30 auditory hair cells (e.g., inner ear hair cells (e.g., Atoh1+ inner ear hair cells)) without promoting substantial cellular proliferation. In some embodiments, the methods and compositions described herein promote proliferation of target cells (e.g., inner ear

supporting cells (e.g., Lgr5+ inner ear supporting cells)) without promoting substantial cellular proliferation.

In some embodiments, the present invention can be used to treat hair cell loss and any disorder that arises as a consequence of cell loss in the ear, such as hearing impairments, deafness, and vestibular disorders, for example, by promoting differentiation (e.g., complete or partial differentiation) of one or more cells (e.g., inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells)) into one or more cells capable of functioning as sensory cells of the ear, e.g., hair cells (e.g., inner ear hair cells (e.g., Atoh1+ inner ear hair cells)).

In some embodiments, the hearing loss is sensorineural hearing loss, which can result from damage or malfunction of the cochlea, e.g., loss of or damage to the sensory epithelium resulting in loss of hair cells.

In some embodiments, the hearing loss can be for any reason, or as a result of any type of event. For example, because of a genetic or congenital defect; for example, a human subject can have been deaf since birth, or can be deaf or hard-of-hearing as a result of a gradual loss of hearing due to a genetic or congenital defect. In another example, the hearing loss can be a result of a traumatic event, such as a physical trauma to a structure of the ear, or a sudden loud noise, or a prolonged exposure to loud noises. For example, prolonged exposures to concert venues, airport runways, and construction areas can cause inner ear damage and subsequent hearing loss.

In some embodiments, hearing loss can be due to chemical-induced ototoxicity, wherein ototoxins include therapeutic drugs including antineoplastic agents, salicylates, quinines, and aminoglycoside antibiotics, contaminants in foods or medicinals, and environmental or industrial pollutants. In some embodiments, hearing loss can result from aging.

In some embodiments, the present disclosure provides methods of treating a subject having hearing loss or balance loss, in which:

(a) a therapeutically effective amount of one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi)

a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in Table F; (viii) a protein degradation inhibitor set forth in Table G; (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and (x) a CREB activator set forth in Table I; and/or

5 (b) a therapeutically effective amount of one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set forth in Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set forth in Table I; and (x) a Notch signaling inhibitor set forth in Table J, are administered to the subject, e.g., to the ear of a subject, to promote inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) proliferation and/or differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) (direct therapy).

In some embodiments, the disclosure provides methods of treating a subject having hearing loss or balance loss, in which:

20 (a) a therapeutically effective amount of one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; and (v) an E box-dependent transcriptional activator set forth in Table D; and/or

25 (b) a therapeutically effective amount of one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of: (i) a Wnt signaling activator set forth in Table A; (ii) an E box-dependent transcriptional activator set forth in Table D; (iii) an HDAC inhibitor set forth in Table F; (iv) a protein degradation inhibitor set forth in Table G; and (v) a Notch signaling inhibitor set forth in Table J, are administered to the subject, e.g., to the ear of a subject, to promote inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) proliferation and/or differentiation of inner ear supporting cells (e.g.,

Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells).

In some embodiments of the methods of treating a subject having hearing loss or balance loss, the one or more agents are administered systemically or to the ear of the subject, e.g., transtympanically to the middle ear of the subject. In some
5 embodiments, the one or more agents that promote proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) are administered prior to the one or more agents that promote differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair
10 cells).

The present disclosure also provides methods of treating a subject having hearing loss or balance loss by:

(a) contacting one or more inner ear supporting cells, e.g., in vitro, with one or more agents that promote proliferation of inner ear supporting cells selected from the
15 group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in Table F; (viii) a protein degradation inhibitor set forth in
20 Table G; (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and (x) a CREB activator set forth in Table I;

(b) optionally contacting the expanded population of inner ear supporting cells with one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of: (i) a retinoid receptor
25 signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set forth in Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set
30 forth in Table I; and (x) a Notch signaling inhibitor set forth in Table J; and

(c) administering the inner ear hair cells to the ear (e.g., the inner ear) of the subject.

The present disclosure also provides methods of treating a subject having hearing loss or balance loss by:

(a) contacting one or more inner ear supporting cells, e.g., in vitro, with one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) a Notch signaling activator set forth in Table E; (vii) an HDAC inhibitor set forth in Table F; (viii) a protein degradation inhibitor set forth in Table G; (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and (x) a CREB activator set forth in Table I; and

(b) administering the expanded population of inner ear supporting cells to the ear (e.g., the inner ear) of the subject in combination with, e.g., concurrently with or prior to administration of, one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; (v) an E box-dependent transcriptional activator set forth in Table D; (vi) an HDAC inhibitor set forth in Table F; (vii) a protein degradation inhibitor set forth in Table G; (viii) a PI3K-Akt signaling inhibitor set forth in Table H; (ix) a CREB activator set forth in Table I; and (x) a Notch signaling inhibitor set forth in Table J.

In some embodiments of the methods of treating a subject having a hearing loss or a balance loss described herein, (a) the one or more agents that promote proliferation of inner ear supporting cells is selected from the group consisting of: (i) a retinoid receptor signaling activator; (ii) a Wnt signaling activator set forth in Table A; (iii) a BMP signaling inhibitor set forth in Table B; (iv) a CDK activator set forth in Table C; and (v) an E box-dependent transcriptional activator set forth in Table D; and (b) the one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells is selected from the group consisting of: (i) a Wnt signaling activator set forth in Table A; (ii) an E box-dependent transcriptional activator set forth in Table D; (iii) an HDAC inhibitor set forth in Table F; (iv) a protein degradation inhibitor set forth in Table G; and (v) a Notch signaling inhibitor set forth in Table J.

In some embodiments, the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K. In some embodiments, the inner ear supporting cells are Lgr5+ inner ear supporting cells. In some embodiments, the inner ear hair cells are Atoh1+ inner ear hair cells.

5 In some embodiments of the methods of treating a subject described herein, the subject has a balanced loss. In some embodiments of the methods of treating a subject described herein, the subject has hearing loss (e.g., sensorineural hearing loss). In some embodiments, the hearing loss is the result of a genetic or congenital defect, trauma, aging, or chemical-induced ototoxicity.

10 In some embodiments of the methods of treating a subject described herein, the subject is a human.

In general, compounds and methods described herein can be used to generate hair cell growth (e.g., Atoh1+ inner ear hair cell growth) in the ear and/or to increase the number of hair cells in the ear (e.g., in the inner, middle, and/or outer ear). For example, the number of hair cells in the ear can be increased about 2-, 3-, 4-, 6-, 8-, or 10-fold, or more, as compared to the number of hair cells before treatment. This new hair cell growth can effectively restore or establish at least a partial improvement in the subject's ability to hear. For example, administration of an agent can improve hearing loss by about 5, 10, 15, 20, 40, 60, 80, 100% or more.

20 A number of compounds that support or promote the proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) and/or promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cell) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) are set forth in Table 1.

A number of compounds that support or promote the proliferation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) are described herein, and include one or more of TTNPB, Compound A, Compound B, Compound C, 1-Azakenpaullone, BIO, WAY-316606, LDN-193189, and Alsterpaullone.

A number of compounds that promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cell) into to inner ear hair cells (e.g., Atoh1+ inner ear hair cells) are described herein, and include one or more of vorinostat, Compound A, Compound B, Compound C, 1-Azakenpaullone, BIO, WAY-262611, NP031112, MG-132, IM-12, Trichostatin A, HLY78, and PF03084014.

Other examples of compounds that promote the proliferation and expansion of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) include, but are not limited to, a retinoid receptor signaling activator (see, e.g., Table K); a Wnt signaling activator set forth in Table A; a bone morphogenetic protein (BMP) signaling inhibitor set forth in Table B; a cyclin-dependent kinase (CDK) activator set forth in Table C; an E box-dependent transcriptional activator set forth in Table D; a Notch signaling activator set forth in Table E; a histone deacetylase (HDAC) inhibitor set forth in Table F; a protein degradation inhibitor set forth in Table G; a PI3K-Akt signaling inhibitor set forth in Table H; and a cAMP response element binding protein (CREB) activator set forth in Table I.

Other examples of compounds that promote the differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) into inner ear hair cells (e.g., Atoh1+ inner ear hair cells) include, but are not limited to, a retinoid receptor signaling activator (see, e.g., Table K); a Wnt signaling activator set forth in Table A; a BMP signaling inhibitor set forth in Table B; a CDK activator set forth in Table C; an E box-dependent transcriptional activator set forth in Table D; an HDAC inhibitor set forth in Table F; a protein degradation inhibitor set forth in Table G; a PI3K-Akt signaling inhibitor set forth in Table H; a CREB activator set forth in Table I; and a Notch signaling inhibitor set forth in Table J.

Where appropriate, following treatment, a human can be tested for an improvement in hearing or in other symptoms related to inner ear disorders. Methods for measuring hearing are well-known and include pure tone audiometry, air conduction, and bone conduction tests. These exams measure the limits of loudness (intensity) and pitch (frequency) that a human can hear. Hearing tests in humans include behavioral observation audiometry (for infants to seven months), visual reinforcement orientation audiometry (for children 7 months to 3 years) and play audiometry for children older than 3 years. Oto-acoustic emission testing can be used to test the functioning of the cochlea hair cells, and electro-cochleography provides information about the functioning of the cochlea and the first part of the nerve pathway to the brain. In some embodiments, treatment can be continued with or without modification or can be stopped.

Pharmaceutical Compositions

In some embodiments, one or more compounds for the promotion of proliferation and/or differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) as described herein can be formulated as a pharmaceutical composition. Pharmaceutical compositions containing one or more compounds as described herein can be formulated according to the intended method of administration.

One or more compounds for the promotion of proliferation and/or differentiation of inner ear supporting cells (e.g., Lgr5+ inner ear supporting cells) as described herein can be formulated as pharmaceutical compositions for direct administration to a subject. Pharmaceutical compositions containing one or more compounds can be formulated in a conventional manner using one or more physiologically acceptable carriers or excipients. For example, a pharmaceutical composition can be formulated for local or systemic administration, e.g., administration by drops (e.g., otic drops) or injection into the ear, insufflation (such as into the ear), intravenous, topical, or oral administration.

The nature of the pharmaceutical compositions for administration is dependent on the mode of administration and can readily be determined by one of ordinary skill in the art. In some embodiments, the pharmaceutical composition is sterile or sterilizable. The therapeutic compositions featured in the invention can contain carriers or excipients, many of which are known to skilled artisans. Excipients that can be used include buffers (for example, citrate buffer, phosphate buffer, acetate buffer, and bicarbonate buffer), amino acids, urea, alcohols, ascorbic acid, phospholipids, polypeptides (for example, serum albumin), EDTA, sodium chloride, liposomes, mannitol, sorbitol, water, and glycerol. The nucleic acids, polypeptides, small molecules, and other modulatory compounds featured in the invention can be administered by any standard route of administration. For example, administration can be parenteral, intravenous, subcutaneous, or oral.

A pharmaceutical composition can be formulated in various ways, according to the corresponding route of administration. For example, liquid solutions can be made for administration by drops into the ear, for injection, or for ingestion; gels or powders can be made for ingestion or topical application. Methods for making such formulations are well known and can be found in, for example, Remington: The

Science and Practice of Pharmacy, 22nd Ed., Allen, ed., Mack Publishing Co., Easton, Pa., 2012.

One or more of the compounds can be administered, e.g., as a pharmaceutical composition, directly and/or locally by injection or through surgical placement, e.g.,
5 to the inner ear. The amount of the pharmaceutical composition may be described as the effective amount or the amount of a cell-based composition may be described as a therapeutically effective amount. Where application over a period of time is advisable or desirable, the compositions of the invention can be placed in sustained released formulations or implantable devices (e.g., a pump).

10 Alternatively or in addition, the pharmaceutical compositions can be formulated for systemic parenteral administration by injection, for example, by bolus injection or continuous infusion. Such formulations can be presented in unit dosage form, for example, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or
15 emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, for example, sterile pyrogen-free water, before use.

In addition to the formulations described previously, the compositions can also
20 be formulated as a depot preparation. Such long acting formulations can be administered by implantation (e.g., subcutaneously). Thus, for example, the compositions can be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

25 Pharmaceutical compositions formulated for systemic oral administration can take the form of tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (for example, pregelatinised maize starch, polyvinylpyrrolidone or hydroxypropyl methylcellulose); fillers (for example, lactose, microcrystalline cellulose or calcium hydrogen
30 phosphate); lubricants (for example, magnesium stearate, talc or silica); disintegrants (for example, potato starch or sodium starch glycolate); or wetting agents (for example, sodium lauryl sulphate). The tablets can be coated by methods well known in the art. Liquid preparations for oral administration may take the form of, for

example, solutions, syrups or suspensions, or they may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (for example, sorbitol syrup, cellulose derivatives or hydrogenated edible fats); emulsifying agents (for example, lecithin or acacia); non-aqueous vehicles (for example, almond oil, oily esters, ethyl alcohol or fractionated vegetable oils); and preservatives (for example, methyl or propyl-p-hydroxybenzoates or sorbic acid). The preparations may also contain buffer salts, flavoring, coloring and sweetening agents as appropriate. Preparations for oral administration may be suitably formulated to give controlled release of the active compound.

In some embodiments, the pharmaceutical compositions described herein can include one or more of the compounds formulated according to any of the methods described above, and one or more cells obtained to the methods described herein.

15

EXAMPLES

The invention is further described in the following examples, which do not limit the scope of the invention described in the claims.

Example 1. Drug screening for agents that promote proliferation and differentiation

20

Two mouse strains were used to develop a screen for agents that promote the proliferation or differentiation of Lgr5-expressing cells. Each strain contained a fluorescent marker for the either the detection of Lgr5 expressing cells or Atoh1 expressing hair cells. Lgr5-EGFP-IRES-Cre-ER mice were used to monitor the proliferation of Lgr5+ cells (Barker, N. et al. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* **449**, 1003-1007 (2007)). This strain was then crossed with Rosa26-td-Tomato reporter mice (Madisen, L. et al. A robust and high-throughput Cre reporting and characterization system for the whole mouse brain. *Nat Neurosci* **13**, 133-140 (2010)) to create a mouse line that enabled lineage tracing of the cells that resulted from differentiated Lgr5-expressing cells. Atoh1-nGFP mice were used to identify differentiated hair cells (Lumpkin, E.A. et al. Math1-driven GFP expression in the developing nervous system of transgenic mice. *Gene Expr Patterns* **3**, 389-395 (2003)).

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Lgr5⁺ cells from both the Lgr5-GFP⁺ and Atoh1-GFP⁺ reporter mice were obtained as follows. Cochleae from neonatal mice (postnatal days 1-3) were dissected in HBSS and the organ of Corti was separated from the stria vascularis and the modiulus. The organs of Corti were then treated with Cell Recovery Solution (Corning) for 1 hour to separate cochlear epithelium from the underlying mesenchyme. Epithelia were then collected and treated with TrypLE (Life Technologies) for 15-20 minutes at 37° C. Single cells obtained by mechanical trituration were filtered (40 µm) and suspended in Matrigel for 3D culture. Matrigel is a reconstituted basement membrane preparation extracted from the Engelbreth-Holm-Swarm (EHS) mouse sarcoma, a tumor rich in extracellular matrix proteins, and is approximately 60% laminin, 30% collagen IV, and 8% entactin. The resulting cells were separately cultured in 24 well plates for 5 days at a concentration of 1 cochlea per well using 1:1 mixture of DMEM and F12, supplemented with Glutamax (GIBCO), N2, B27 (Invitrogen), EGF (50 ng/mL; Chemicon), bFGF (50 ng/mL; Chemicon), IGF1 (50 ng/mL; Chemicon) and small molecules including CHIR99021 (3 µM; LC Labs), VPA (1 mM; Sigma), pVc (100 µg/ml; Sigma), and 616452 (2 µM; Calbiochem). Media were changed every other day.

To assess the degree of proliferation of the Lgr5⁺ cells, 200 µl of cell dissociation solution was then added to each well of the Lgr5-GFP⁺ cells and incubated for 45 minutes and then TrypleE for 20 minutes. Cell cultures were then transferred to a 15 ml falcon tube for centrifugation. Supernatant was then removed and the cell culture was re-suspended in matrigel once again. Cells were distributed into a 96 well plate with approximately 5000 cells per well. The Lgr5-GFP⁺ cultures were then treated with DMEM/F12 media containing from nanomolar to micromolar concentrations, e.g., 0.001, 0.005 µM, 0.01 µM, 0.1 µM, 1 µM, 10 µM, 50 µM, or 100 µM, of a candidate drug for an additional 5 days (See FIG. 1). At that point the Lgr5-GFP⁺ cells were sorted using FACS and the fluorescence was measured (See FIG. 3 and 4).

To assess the degree of differentiation of the Lgr5⁺ cells into hair cells, the Atoh1-nGFP cells were cultured for 2 more days (a total of 7 days) in the DMEM/F12 media containing EGF, bFGF, IGF, pVc, VPA, and CHIR99021 (as above, a 1:1 mixture of DMEM and F12, supplemented with Glutamax (GIBCO), N2, B27 (Invitrogen), EGF (50 ng/mL; Chemicon), bFGF (50 ng/mL; Chemicon), IGF1

(50 ng/mL; Chemicon) and small molecules including CHIR99021 (3 μ M), VPA (1 mM), pVc (100 μ g/ml), and 616452 (2 μ M). Media were changed every other day. 4-hydroxytamoxifen (20 ng/ml) was added to cultures on day 0 for lineage tracing studies) (See FIG. 2). After the 7-day incubation, 200 μ l of cell dissociation solution was added to each well and incubated for 45 minutes and then TrypleE for 20 minutes. Cell cultures were then transferred to a 15 ml falcon tube for centrifugation. Supernatant was then removed and the cell culture was re-suspended in matrigel once again. Cells were distributed into a 96 well plate with approximately 5000 cells per well. The cells were then treated with DMEM/F12 media containing the candidate drug. After 10 days of incubating with the candidate drug, the cells were sorted by FACS and the fluorescence levels of the Atoh1-nGFP cells were measured (see FIG. 5). CHIR99021, a GSK-3 β inhibitor of the Wnt pathway, and LY411575 a γ -secretase inhibitor of the Notch pathway were used as positive controls for this screen (see WO2014159356 and FIG. 5). This method of screening was used to screen drugs, compounds, genes, and growth factors (see Table 1).

Table 1: Screening agents

Name	Target
CHIR-99021 (6-(2-(4-(2,4-dichlorophenyl)-5-(4-methyl-1H-imidazol-2-yl)pyrimidin-2-ylamino)ethylamino)nicotinonitrile hydrochloride)	Wnt (GSK3 β)
LY411575 (Benzeneacetamide, N-[(1S)-2-[[[(7S)-6,7-dihydro-5-methyl-6-oxo-5H-dibenz[b,d]azepin-7-yl]amino]-1-methyl-2-oxoethyl]-3,5-difluoro- α -hydroxy-, (α S)-)	Notch (γ -Secretase)
Vorinostat (N1-hydroxy-N8-phenyl-octanediamide)	pan-HDAC
TTNPB (Ro 13-7410) (Arotinoid Acid)	RAR
Cerivastatin (sodium salt) (3R,5S,6E)-7-[4-(4-fluorophenyl)-5-(methoxymethyl)-2,6-bis(propan-2-yl)pyridin-3-yl]-3,5-dihydroxyhept-6-enoic acid)	P27
Compound A (See FIG. 7)	ATOH1
Compound B (See FIG. 7)	ATOH1
Compound C (See FIG. 7)	ATOH1
1-Azakenpaullone (Pyrido[3',2':2,3]azepino[4,5-b]indol-6(5H)-one, 9-bromo-7,12-dihydro-)	ATOH1
(2'Z,3'E)-6-Bromoindirubin-3'-oxime (BIO)	Wnt (GSK3 β)
MK-0752 (3-((1r,4s)-4-(4-	Notch (γ -Secretase)

Name	Target
chlorophenylsulfonyl)-4-(2,5-difluorophenyl)cyclohexyl)propanoic acid)	
WAY-262611 ((1-(4-(Naphthalen-2-yl)pyrimidin-2-yl)piperidin-4-yl)methanamine)	Wnt (β -catenin)
NP031112 (Tideglusib) (4-benzyl-2-naphthalen-1-yl-1,2,4-thiadiazolidine-3,5-dione)	Wnt (GSK3 β)
WAY-316606 (5-(benzenesulfonyl)-N-piperidin-4-yl-2-(trifluoromethyl)benzenesulfonamide)	Wnt (sFRP-1)
LDN-193189 (4-(6-(4-(piperazin-1-yl)phenyl)pyrazolo[1,5-a]pyrimidin-3-yl)quinoline)	BMP receptor
Alsterpaullone, 2 Cyanoethyl (3-(9-nitro-6-oxo-7,12-dihydro-5H-indolo[3,2-d][1]benzazepin-2-yl)propanenitrile)	P27 (Kip1)
MLN4924 (pevonedistat) ([[(1S,2S,4R)-4-[4-[[[(1S)-2,3-dihydro-1H-inden-1-yl]amino]pyrrolo[2,3-d]pyrimidin-7-yl]-2-hydroxycyclopentyl]methyl sulfamate)	AKT
MG 132 (Carbobenzoxy-L-leucyl-L-leucyl-L-leucinal)	Proteasome Inhibitor
IM-12 (3-(4-Fluorophenylethylamino)-1-methyl-4-(2-methyl-1H-indol-3-yl)-1H-pyrrole-2,5-dione)	Wnt (GSK3 β)
Trichostatin A ((2E,4E,6R)-7-[4-(dimethylamino)phenyl]-N-hydroxy-4,6-dimethyl-7-oxohepta-2,4-dienamide)	pan-HDAC
HLY78 (4-Ethyl-5,6-Dihydro-5-methyl-[1,3]dioxolo[4,5-j]phenanthridine, 4-Ethyl-5-methyl-5,6-dihydro-[1,3]dioxolo[4,5-j]phenanthridine)	Wnt
DMHI (4-(6-(4-isopropoxyphenyl)pyrazolo[1,5-a]pyrimidin-3-yl)quinoline)	BMP1
2-(N)-benzyl ellipticene	ATOH1
AC102 (6-Fluor-9-methyl- β -carboline, see WO2015044434)	PKC/CREB
BI8622 (See FIG. 7)	HUWE1
PF-03084014 ((S)-2-(((S)-6,8-difluoro-1,2,3,4-tetrahydronaphthalen-2-yl)amino)-N-(1-(2-methyl-1-(neopentylamino)propan-2-yl)-1H-imidazol-4-yl)pentanamide)	Notch (γ -Secretase)

The results, shown in Figures 3-5 showed that a number of compounds screened were able to effectively induce proliferation or differentiation. Tables 2 and 3 list the top compounds for proliferation and differentiation, respectively.

CHIR99021	GSK3 β inhibitor
TTNPB	retinoid acid receptor agonist
Compound A	Atoh1 stimulating compound
Compound B	Atoh1 stimulating compound
Compound C	Atoh1 stimulating compound
1-Azakenpauillone	Atoh1 stimulating compound
BIO	GSK-3 β inhibitor
WAY-316606	sFRP-1 (secreted frizzled-related 1) inhibitor
LDN-193189	BMP receptor inhibitor
Alsterpauillone, 2 Cyanoethyl	p27 Kip1 inhibitor

CHIR99021	GSK3 β inhibitor
LY411575	gamma-secretase inhibitor
vorinostat	class I, II and IV HDAc inhibitor
Compound A	Atoh1 stimulating compound
Compound B	Atoh1 stimulating compound
Compound C	Atoh1 stimulating compound
1-Azakenpauillone	Atoh1 stimulating compound
BIO	GSK-3 β inhibitor
WAY-262611	dickkopf inhibitor
NP031112	GSK3 β inhibitor
MG-132	proteasome inhibitor
IM-12	GSK3 β inhibitor
Trichostatin A	class I and II HDAC inhibitor
HLY78	Wnt signal transduction activator
PF03084014	gamma-secretase inhibitor

Example 2. Drug screening for agents for the treatment of hearing loss associated with the loss of cochlear hair cells.

A novel transgenic mouse is made that contains two florescent reporters stably integrated into the genome of the mouse (Lgr5/Atoh1 reporter mice). The Lgr5/Atoh1 reporter mice are made by starting with oocytes from a transgenic mouse that has GFP under the control of a Lgr5 promoter (Barker et al., supra) and adding a plasmid comprising mCherry under the control of an Atoh1 enhancer and a promoter (e.g., an SV40 or globin minimal promoter) (see Fig. 6).

The sequence of the Atoh1 enhancer used in these constructs is as follows:

TCCAAGGTC CGCAATGAAGTTTGCATAACAAACGTTTGGCAGCTCCCTC
TCTCACACCCCATTAACAAGCTGTAACATATAGCTGCAGGTTGCTATAATC
TCATTAATATTTGGAACTTGAATATTGAGTATTTCTGAGCGCTCATTCC

CCATATGCCAGACCACTCCTGCCATGCTGACTGGTTCCTTTCTCTCCATTA
 TTAGCAATTAGCTTCTACCTTCCAAAGTCAGATCCAAGTATCTAAGATACT
 ACCAAAGGCATCAACTATGTATGCAAGTTAGGCATGCTTAATATCACCCA
 AACAAACAAAGAGTCAGCACTTCTTAAAGTAATGAAGATAGATAAATCG
 5 GGTTAGTTCTTTGGGACACCGCTGTTGTTTTCCAGAGTTTTTCTATACTTTA
 AGCAGCTTGTTTTATATTCTGTCTTTGCCCTCAGCCAGCTAACATTTTTATTT
 GTTGAGGGTTTTGGCTCACCACTTTTGGAACTTATTTGATTTACGGG
 GAGCTGAAGGAAGATTGTTTTTGGCAACAGGCAAGTTTAACACGTTCTTC
 ATGGGGCATTGCGAATGGCACATCTACCAGAAAGGGAGGGGGAGTAACT
 10 TCCTCGTGCTGAACCAGCAGGAGACCAGAGCTTTCCTGAGGTCTTCCTATT
 GATTTTAAAGATTTAAAAGTGAAGTTGTAATGTTATTGAAGTTT
 GTCTTGGAATATACATCTCCTCTGCTAACTTAAAAGTTCAAGAAAGGAAA
 GGAAAGAAATAGAACCCCTTGCTAACTACAACCTAGACTGAGAGGTGAA
 GATCGCGGGCAAAGACAGGTGGTCACTGAAACGTTTGCAGTTCTTTTCTTC
 15 CGAAGGCTTAGGACACAGGGTAAGGAGGAGCTAAAATAAAGCCGAGTGT
 ACGTTTAGTCTTCTCTGCACCCAGGCCTAGTGTCTCCCAGGCAAGGAGT
 CACCCCTTTGCTTCTGGCTCCTAACTGAAAAAGGCAAAGGGAGTGGAG
 AATGGGTAAATCCCAGGACACAGGGGAGAGGCAGGGGAGGAGAGAAGT
 CGGAGGAAGATAAAGGAAAGGACAGGAACCAAGAAGCGTGGGGGTAGTT
 20 TGCCGTAATGTGAGTGTTCCTTAATTAGAGAGCGGCTGACAATAGAGGGG
 CTGGCAGAGGCTCCTGGCCCCGGTGCAGGAGCGTCTGGAGCGGAGCACGCG
 CTGTCAGCTGGTGAGCGCACTCGCTTTCAGGCCGCTCCCCGGGGAGCTGA
 GCGGCCACATTTAACACCGTCGTCACCCTCCCCGGCCTCCTAACATCGGC
 CTCCTCCTCGTAGACAGCCTTGCTCGGCCCCCACCAGGCAGAGTTTACAGA
 25 AGCCAGAGCCTCTCGCCGTTCCCCCGCATTCGCCCGGG (SEQ ID NO:1)

The sequence of the Atoh1-mCherry plasmid is as follows:

GGTACCGAGCTCTTACGCGTGCTAGCCCCGGGCTCGAGATCTTCCAAGGTC
 CGGCAATGAAGTTTGCATAACAAACGTTTGGCAGCTCCCTCTCTCACACCC
 30 CATTAAACAAGCTGTAACATATAGCTGCAGGTTGCTATAATCTCATTAAATAT
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 ATCAACTATGTATGCAAGTTAGGCATGCTTAATATCACCCAAACAAACAA
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5 AACACCGTCGTCACCCTCCCCGGCCTCCTCAACATCGGCCTCCTCCTCGTA
GACAGCCTTGCTCGGCCCCCCACCGGCAGAGTTTACAGAAGCCAGAGCCT
CTCGCCGTTCCCCCGCATTCGCCCGGGTCTAGAATGGTGAGCAAGGGCGA
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45 GATCTCAAGAAGATCCTTTGATCTTTTCTACGGGTCTGACGCTCAGTGGA
ACGAAAACCTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATC
TTCACCTAGATCCTTTTAAATTAATAAATGAAGTTTTAAATCAATCTAAAGT
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50 GTCGTGTAGATAACTACGATACGGGAGGGCTTACCATCTGGCCCCAGTGC

TGCAATGATACCGCGAGACCCACGCTCACCGGCTCCAGATTTATCAGCAA
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 CGCCACATAGCAGAACTTTAAAAGTGCTCATCATTGGAAAACGTTCTTCG
 GGGCGAAAACCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTA
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 15 AGGGCGACACGGAAATGTTGAATACTCATACTCTTCCTTTTTCAATATTAT
 TGAAGCATTATCAGGGTTATTGTCTCATGAGCGGATACATATTTGAATGT
 ATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCGAAAAGT
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 TTACGCGCAGCGTGACCGCTACACTTGCCAGCGCCCTAGCGCCCGCTCCTT
 20 TCGCTTTCTTCCCTTCTTTCTCGCCACGTTTCGCCGGCTTTCCCGTCAAGC
 TCTAAATCGGGGGCTCCCTTTAGGGTTCGATTTAGTGCTTACGGCACCT
 CGACCCCAAAAACTTGATTAGGGTGATGGTTCACGTAGTGGGCCATCGC
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 GTGGACTCTTGTTCCAAACTGGAACAACACTCAACCCTATCTCGGTCTATT
 25 CTTTTGATTTATAAGGGATTTTGCCGATTTCCGGCCTATTGGTTAAAAAATG
 AGCTGATTTAACAAAAATTTAACCGCAATTTTAACAAAATATTAACGCTT
 ACAATTTGCCATTCGCCATTCAGGCTGCGCAACTGTTGGGAAGGGCGATC
 GGTGCGGGCCTCTTCGCTATTACGCCAGCCCAAGCTACCATGATAAGTAA
 GTAATATTAAGGTACGGGAGGTACTTGGAGCGGCCGCAATAAAATATCTT
 30 TATTTTCATTACATCTGTGTGTTGGTTTTTTGTGTGAATCGATAGTACTAAC
 ATACGCTCTCCATCAAAACAAAACGAAACAAAACAACTAGCAAAATAG
 GCTGTCCCAGTGCAAGTGCAGGTGCCAGAACATTTCTCTATCGATACAT
 A (SEQ ID NO:2)

35 Stem cells from this mouse are isolated from the inner ear, suspended in
 matrigel and cultured in DMEM/F12 media containing EFG, bFGF, IGF, pVc and
 VPACHIR in a 24 well plate for up to 5 days. After 5 days, 200 µl of cell dissociation
 solution is added to each well and incubated for 45 minutes and then TrypleE for 20
 minutes. Cell cultures are then transferred to a 15 ml falcon tube for centrifugation.
 40 Supernatant is then removed and the cell culture is re-suspended in matrigel once
 again. Cells are then distributed into a 96 well plate with approximately 5000 cells per
 well.

The cell culture is then treated with DMEM/F12 media containing a candidate
 drug for an addition 5-7 days. At that point the cells are sorted using FACS and the
 45 fluorescence of both markers is measured. Those candidate drugs that increase the

fluorescence of both markers in the transgenic mouse cells when compared to an untreated transgenic control cell are selected as candidate agents for the treatment of hearing loss associated with the loss of cochlear hair cells (e.g., cochlear hair cells in the inner ear). This method of screening is used to screen drugs, compounds, genes, or growth factors.

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OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

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WHAT IS CLAIMED IS:

1. A method of producing an expanded population of inner ear supporting cells, the method comprising contacting a population of inner ear supporting cells with one or more agents selected from the group consisting of:

- (a) a retinoid receptor signaling activator;
- (b) a Wnt signaling activator set forth in Table A;
- (c) a bone morphogenetic protein (BMP) signaling inhibitor set forth in Table B;
- (d) a cyclin-dependent kinase (CDK) activator set forth in Table C;
- (e) an E box-dependent transcriptional activator set forth in Table D;
- (f) a Notch signaling activator set forth in Table E;
- (g) a histone deacetylase (HDAC) inhibitor set forth in Table F;
- (h) a protein degradation inhibitor set forth in Table G;
- (i) a PI3K-Akt signaling inhibitor set forth in Table H; and
- (j) a cAMP response element binding protein (CREB) activator set forth in Table I,

wherein the one or more agents are present in amounts sufficient to produce an expanded population of inner ear supporting cells.

2. The method of claim 1, wherein the Notch signaling activator is a Delta-like protein activator, a Jagged protein activator, a Notch activator, and/or a γ -secretase activator.

3. The method of claim 1, wherein the one or more agents is selected from the group consisting of:

- (a) a retinoid receptor signaling activator;
- (b) a Wnt signaling activator set forth in Table A;
- (c) a BMP signaling inhibitor set forth in Table B;
- (d) a CDK activator set forth in Table C; and
- (e) an E box-dependent transcriptional activator set forth in Table D.

4. The method of any one of claims 1-3, wherein the expanded population of inner ear supporting cells is an expanded population of Lgr5⁺ inner ear supporting cells.

5. The method of claim 4, wherein the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ cochlear supporting cells.
6. The method of claim 4, wherein the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ vestibular supporting cells.
7. A method of promoting differentiation of a population of inner ear supporting cells into a population of inner ear hair cells, the method comprising contacting a population of inner ear supporting cells with one or more agents selected from the group consisting of:
 - (a) a retinoid receptor signaling activator;
 - (b) a Wnt signaling activator set forth in Table A;
 - (c) a BMP signaling inhibitor set forth in Table B;
 - (d) a CDK activator set forth in Table C;
 - (e) an E box-dependent transcriptional activator set forth in Table D;
 - (f) an HDAC inhibitor set forth in Table F;
 - (g) a protein degradation inhibitor set forth in Table G;
 - (h) a PI3K-Akt signaling inhibitor set forth in Table H;
 - (i) a CREB activator set forth in Table I; and
 - (j) a Notch signaling inhibitor set forth in Table J,wherein the one or more agents are present in amounts sufficient to promote differentiation into a population of inner ear hair cells.
8. The method of claim 7, wherein the one or more agents is selected from the group consisting of:
 - (a) a Wnt signaling activator set forth in Table A;
 - (b) an E box-dependent transcriptional activator set forth in Table D;
 - (c) an HDAC inhibitor set forth in Table F;
 - (d) a protein degradation inhibitor set forth in Table G; and
 - (e) a Notch signaling inhibitor set forth in Table J.

9. The method of claim 7 or 8, wherein the Notch signaling inhibitor is a Delta-like protein inhibitor, a Jagged protein inhibitor, a Notch inhibitor, and/or a γ -secretase inhibitor.
10. The method of any one of claims 7-9, wherein the population of inner ear hair cells is a population of atonal homolog 1 (Atoh1)+ inner ear hair cells.
11. The method of claim 10, wherein the population of Atoh1+ inner ear hair cells is a population of Atoh1+ cochlear hair cells.
12. The method of claim 10, wherein the population of Atoh1+ inner ear hair cells is a population of Atoh1+ vestibular hair cells.
13. The method of any one of claims 1-7 and 9-12, wherein the retinoid receptor signaling activator is a retinoic acid receptor (RAR) agonist set forth in Table K or a retinoic X receptor (RXR) agonist set forth in Table K.
14. The method of claim 13, wherein the RAR agonist is an RAR α agonist, an RAR β agonist, and/or an RAR γ agonist.
15. The method of claim 13, wherein the RXR agonist is an RXR α agonist, an RXR β agonist, and/or an RXR γ agonist.
16. The method of any one of claims 1-15, wherein the Wnt signaling activator is a glycogen synthase kinase-3 β (GSK-3 β) inhibitor, a Wnt activator, a Frizzled receptor activator, a lipoprotein receptor-related protein 5/6 (LRP5/6) activator, a Disheveled (Dvl) activator, an Axin inhibitor, a Dickkopf (Dkk) inhibitor, a secreted Frizzled-related protein (sFRP) inhibitor, a Groucho inhibitor, and/or a Wnt inhibitory protein (WIF) inhibitor.

17. The method of any one of claims 1-7 and 9-16, wherein the BMP signaling inhibitor is a Noggin activator, a Chordin activator, a BMP receptor inhibitor, a SMAD1/5/8 inhibitor, a SMAD2/3 inhibitor, and/or a SMAD4 inhibitor.
18. The method of any one of claims 1-7 and 9-17, wherein the CDK activator is a p27Kip1 inhibitor and/or a retinoblastoma protein (Rb) inhibitor.
19. The method of any one of claims 1-18, wherein the E box-dependent transcriptional activator is an Atoh1 activator.
20. The method of any one of claims 1, 2, and 4-19, wherein the HDAC inhibitor is an HDAC class I inhibitor, an HDAC class II inhibitor, an HDAC class III inhibitor, and/or a pan-HDAC inhibitor.
21. The method of claim 20, wherein the HDAC class III inhibitor is a SIRT1 inhibitor and/or a SIRT2 inhibitor.
22. The method of any one of claims 1, 2, and 4-21, wherein the protein degradation inhibitor is a proteasome inhibitor or a ubiquitin ligase inhibitor.
23. The method of any one of claims 1, 2, 4-7, and 9-22, wherein the PI3K-Akt signaling inhibitor is an Akt inhibitor, a PI3K inhibitor, a PKC inhibitor, and/or a PDK1 inhibitor.
24. The method of any one of claims 1-23, wherein the population of inner ear supporting cells is a population of Lgr5+ inner ear supporting cells.
25. The method of claim 24, wherein the population of Lgr5+ inner ear supporting cells is a population of Lgr5+ cochlear supporting cells.
26. The method of claim 24, wherein the population of Lgr5+ inner ear supporting cells is a population of Lgr5+ vestibular supporting cells.

27. A human cell comprising a first recombinant nucleic acid molecule that comprises a first reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glast, and Lfng, and a second recombinant nucleic acid molecule that comprises a second reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of Atoh1, Myo7a, Cdh23, Pcdh15, Myo6, Myo1c, Tmc1, and Cav1.3, wherein the first reporter gene is different from the second reporter gene.
28. The cell of claim 27, wherein the inner ear supporting cell marker is Lgr5.
29. The cell of claim 27, wherein the inner ear hair cell marker is Atoh1.
30. The cell of claim 28 or 29, wherein the inner ear supporting cell marker is Lgr5 and the inner ear hair cell marker is Atoh1.
31. The cell of any one of claims 28-30, wherein the regulatory element of an inner ear supporting cell marker is an Lgr5 promoter.
32. The cell of any one of claims 28-30, wherein the regulatory element of an inner ear hair cell marker is an Atoh1 enhancer.
33. The cell of claim 32, wherein the Atoh1 enhancer is operably linked to an SV40 promoter or a globin promoter.
34. The cell of any one of claims 27-33, wherein the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a second fluorescent protein, wherein the first fluorescent protein is different from the second fluorescent protein.

35. A transgenic mouse having two or more recombinant nucleic acid molecules stably integrated into the genome of the mouse, comprising at least a first recombinant nucleic acid molecule that comprises a first reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of *Lgr5*, *Sox2*, *p27*, *Prox1*, *FGFR3*, *Glast*, and *Lfng*, and a second recombinant nucleic acid molecule that comprises a second reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of *Atoh1*, *Myo7a*, *Cdh23*, *Pcdh15*, *Myo6*, *Myo1c*, *Tmc1*, and *Cav1.3*, wherein the first reporter gene is different from the second reporter gene.

36. The transgenic mouse of claim 35, wherein the inner ear supporting cell marker is *Lgr5*.

37. The transgenic mouse of claim 35, wherein the inner ear hair cell marker is *Atoh1*.

38. The transgenic mouse of claim 36 or 37, wherein the inner ear supporting cell marker is *Lgr5* and the inner ear hair cell marker is *Atoh1*.

39. The transgenic mouse of any one of claims 36-38, wherein the regulatory element of an inner ear supporting cell marker is an *Lgr5* promoter.

40. The transgenic mouse of any one of claims 36-38, wherein the regulatory element of an inner ear hair cell marker is an *Atoh1* enhancer.

41. The transgenic mouse of claim 40, wherein the *Atoh1* enhancer is operably linked to an SV40 promoter or a globin promoter.

42. The transgenic mouse of any one of claims 35-41, wherein the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a second fluorescent protein, wherein the first fluorescent protein is different from the second fluorescent protein.

43. A cell isolated from the transgenic mouse of any one of claims 35-42, comprising the first recombinant nucleic acid molecule and the second recombinant nucleic acid molecule.

44. The cell of claim 43, wherein the cell is isolated from the inner ear of the transgenic mouse.

45. A method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, the method comprising:

(a) isolating a population of inner ear supporting cells from the mouse of any one of claims 35-42;

(b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells;

(c) administering a test compound to the expanded population of inner ear supporting cells;

(d) detecting the expression levels of the first reporter gene and the second reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and

(e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the first reporter gene compared to the expression level of the first reporter gene in the absence of the test compound, and/or increases the expression level of the second reporter gene compared to the expression level of the second reporter gene in the absence of the test compound.

46. The method of claim 45, wherein the conditions sufficient to produce an expanded population of inner ear supporting cells comprise media, preferably a mixture of DMEM and F12 media, preferably a 1:1 mixture, in the presence of one or more growth factors, preferably epidermal growth factor (EGF), basic fibroblast growth factor (bFGF), and/or insulin-like growth factor (IGF1).

47. The method of claim 45 or 46, wherein the conditions sufficient to produce an expanded population of inner ear supporting cells further comprise one or more agents selected from the group consisting of:

- (a) a retinoid receptor signaling activator;
- (b) a Wnt signaling activator set forth in Table A;
- (c) a BMP signaling inhibitor set forth in Table B;
- (d) a CDK activator set forth in Table C;
- (e) an E box-dependent transcriptional activator set forth in Table D;
- (f) a Notch signaling activator set forth in Table E or a Notch signaling inhibitor

set forth in Table J;

- (g) an HDAC inhibitor set forth in Table F;
- (h) a protein degradation inhibitor set forth in Table G;
- (i) a PI3K-Akt signaling inhibitor set forth in Table H; and
- (j) a CREB activator set forth in Table I.

48. The method of claim 47, wherein the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

49. The method of claim 47 or 48, wherein the conditions sufficient to produce an expanded population of inner ear supporting cells comprise one or more agents set forth in Table 1.

50. The method of any one of claims 45-49, wherein the conditions sufficient to produce an expanded population of inner ear supporting cells comprise media, preferably a 1:1 mixture of DMEM and F12, supplemented with N2, B27, EGF, bFGF, IGF1, CHIR99021, and VPA.

51. The method of any one of claims 45-50, wherein the candidate agent is selected from a group consisting of a small molecule, a compound, a nucleic acid, a peptide, a polypeptide, a growth factor, and an epigenetic modifier.

52. The method of any one of claims 45-51, wherein the population of inner ear supporting cells is isolated from the cochlea of the mouse by a method comprising first dissecting the organ of Corti, isolating sensory epithelium, and creating a single cell suspension.

53. The method of any one of claims 45-52, wherein the population of inner ear supporting cells is a population of Lgr5+ inner ear supporting cells.

54. The method of any one of claims 45-53, wherein the first reporter gene encodes a first fluorescent protein and the second reporter gene encodes a second fluorescent protein, wherein the first fluorescent protein is different from the second fluorescent protein.

55. The method of any one of claims 45-54, wherein the expression levels of the first reporter gene and the second reporter gene are protein expression levels.

56. A method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, the method comprising:

(a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that comprises a reporter gene under the control of a regulatory element of an inner ear supporting cell marker selected from the group consisting of Lgr5, Sox2, p27, Prox1, FGFR3, Glax, and Lfng;

(b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells, wherein the conditions comprise one or more agents selected from the group consisting of:

- (i) a retinoid receptor signaling activator,
- (ii) a Wnt signaling activator set forth in Table A,
- (iii) a BMP signaling inhibitor set forth in Table B,
- (iv) a CDK activator set forth in Table C,

- (v) an E box-dependent transcriptional activator set forth in Table D,
 - (vi) a Notch signaling activator set forth in Table E,
 - (vii) an HDAC inhibitor set forth in Table F,
 - (viii) a protein degradation inhibitor set forth in Table G,
 - (ix) a PI3K-Akt signaling inhibitor set forth in Table H, and
 - (x) a CREB activator set forth in Table I;
- (c) administering a test compound to the expanded population of inner ear supporting cells;
- (d) detecting the expression level of the reporter gene in the expanded population of inner ear supporting cells in the presence of the test compound; and
- (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

57. A method for identifying a candidate agent for the treatment of hearing loss or balance loss associated with a loss of cochlear or vestibular hair cells, the method comprising:

(a) providing a population of inner ear supporting cells having a stably integrated recombinant nucleic acid molecule that comprises a reporter gene under the control of a regulatory element of an inner ear hair cell marker selected from the group consisting of *Atoh1*, *Myo7a*, *Cdh23*, *Pcdh15*, *Myo6*, *Myo1c*, *Tmc1*, and *Cav1.3*;

(b) maintaining the population of inner ear supporting cells under conditions sufficient to produce an expanded population of inner ear supporting cells, wherein the conditions comprise one or more agents selected from the group consisting of:

- (i) a retinoid receptor signaling activator,
- (ii) a Wnt signaling activator set forth in Table A,
- (iii) a BMP signaling inhibitor set forth in Table B,
- (iv) a CDK activator set forth in Table C,
- (v) an E box-dependent transcriptional activator set forth in Table D,
- (vi) a Notch signaling activator set forth in Table E,
- (vii) an HDAC inhibitor set forth in Table F,

- (viii) a protein degradation inhibitor set forth in Table G,
 - (ix) a PI3K-Akt signaling inhibitor set forth in Table H, and
 - (x) a CREB activator set forth in Table I;
- (c) administering a test compound to the expanded population of inner ear supporting cells;
- (d) detecting the expression level of the reporter gene in the expanded population of inner ear cells in the presence of the test compound; and
- (e) selecting as a candidate agent for the treatment of hearing loss or balance loss a test compound that increases the expression level of the reporter gene compared to the expression level of the reporter gene in the absence of the test compound.

58. The method of claim 56 or 57, wherein the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

59. The method of any one of claims 56-58, wherein the conditions sufficient to produce an expanded population of inner ear supporting cells comprise one or more agents selected from the group consisting of:

- (a) a retinoid receptor signaling activator;
- (b) a Wnt signaling activator set forth in Table A;
- (c) a BMP signaling inhibitor set forth in Table B;
- (d) a CDK activator set forth in Table C; and
- (e) an E box-dependent transcriptional activator set forth in Table D.

60. The method of any one of claims 56-59, wherein the expanded population of inner ear supporting cells is an expanded population of Lgr5+ inner ear supporting cells.

61. The method of claim 60, wherein the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ cochlear supporting cells.

62. The method of claim 60, wherein the expanded population of Lgr5+ inner ear supporting cells is an expanded population of Lgr5+ vestibular supporting cells.

63. The method of any one of claims 56-62, wherein the population of inner ear supporting cells is isolated from a human.

64. The method of any one of claims 56-62, wherein the population of inner ear supporting cells is isolated from a mouse.

65. The method of any one of claims 56-64, wherein the reporter gene encodes a fluorescent protein.

66. A method of treating a subject having hearing loss or balance loss, the method comprising administering to the subject in need thereof one or both of:

(a) a therapeutically effective amount of one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;
- (vi) a Notch signaling activator set forth in Table E;
- (vii) an HDAC inhibitor set forth in Table F;
- (viii) a protein degradation inhibitor set forth in Table G; and
- (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and/or

(b) a therapeutically effective amount of one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;

- (vi) an HDAC inhibitor set forth in Table F;
- (vii) a protein degradation inhibitor set forth in Table G;
- (viii) a PI3K-Akt signaling inhibitor set forth in Table H; and
- (ix) a Notch signaling inhibitor set forth in Table J.

67. The method of claim 66, wherein the method comprises administering to the subject in need thereof one or both of:

(a) a therapeutically effective amount of one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
 - (ii) a Wnt signaling activator set forth in Table A;
 - (iii) a BMP signaling inhibitor set forth in Table B;
 - (iv) a CDK activator set forth in Table C; and
 - (v) an E box-dependent transcriptional activator set forth in Table D;
- and/or

(b) a therapeutically effective amount of one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of:

- (i) a Wnt signaling activator set forth in Table A;
- (ii) an E box-dependent transcriptional activator set forth in Table D;
- (iii) an HDAC inhibitor set forth in Table F;
- (iv) a protein degradation inhibitor set forth in Table G; and
- (v) a Notch signaling inhibitor set forth in Table J.

68. The method of claim 66 or 67, wherein the one or more agents are administered systemically or to the ear of the subject, preferably transtympanically to the middle ear of the subject.

69. The method of any one of claims 66-68, wherein the one or more agents that promote proliferation of inner ear supporting cells are administered prior to the one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells.

70. A method of treating a subject having hearing loss or balance loss, the method comprising:

(a) contacting one or more inner ear supporting cells, optionally in vitro, with one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;
- (vi) a Notch signaling activator set forth in Table E;
- (vii) an HDAC inhibitor set forth in Table F;
- (viii) a protein degradation inhibitor set forth in Table G;
- (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and
- (x) a CREB activator set forth in Table I;

(b) optionally contacting the expanded population of inner ear supporting cells with one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;
- (vi) an HDAC inhibitor set forth in Table F;
- (vii) a protein degradation inhibitor set forth in Table G;
- (viii) a PI3K-Akt signaling inhibitor set forth in Table H;
- (ix) a CREB activator set forth in Table I; and
- (x) a Notch signaling inhibitor set forth in Table J; and

(c) administering the inner ear hair cells to the ear (optionally to the inner ear) of the subject.

71. A method of treating a subject having hearing loss or balance loss, the method comprising:

(a) contacting one or more inner ear supporting cells, optionally in vitro, with one or more agents that promote proliferation of inner ear supporting cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;
- (vi) a Notch signaling activator set forth in Table E;
- (vii) an HDAC inhibitor set forth in Table F;
- (viii) a protein degradation inhibitor set forth in Table G;
- (ix) a PI3K-Akt signaling inhibitor set forth in Table H; and
- (x) a CREB activator set forth in Table I; and

(b) administering the expanded population of inner ear supporting cells to the ear (preferably the inner ear) of the subject in combination with, preferably concurrently with or prior to administration of, one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C;
- (v) an E box-dependent transcriptional activator set forth in Table D;
- (vi) an HDAC inhibitor set forth in Table F;
- (vii) a protein degradation inhibitor set forth in Table G;
- (viii) a PI3K-Akt signaling inhibitor set forth in Table H;
- (ix) a CREB activator set forth in Table I; and
- (x) a Notch signaling inhibitor set forth in Table J.

72. The method of any one of claims 66-71, wherein:

(a) the one or more agents that promote proliferation of inner ear supporting cells is selected from the group consisting of:

- (i) a retinoid receptor signaling activator;
- (ii) a Wnt signaling activator set forth in Table A;
- (iii) a BMP signaling inhibitor set forth in Table B;
- (iv) a CDK activator set forth in Table C; and
- (v) an E box-dependent transcriptional activator set forth in Table D; and

(b) the one or more agents that promote differentiation of inner ear supporting cells into inner ear hair cells is selected from the group consisting of:

- (i) a Wnt signaling activator set forth in Table A;
- (ii) an E box-dependent transcriptional activator set forth in Table D;
- (iii) an HDAC inhibitor set forth in Table F;
- (iv) a protein degradation inhibitor set forth in Table G; and
- (v) a Notch signaling inhibitor set forth in Table J.

73. The method of any one of claims 66-72, wherein the retinoid receptor signaling activator is an RAR agonist set forth in Table K or an RXR agonist set forth in Table K.

74. The method of any one of claims 66-73, wherein the inner ear supporting cells are Lgr5+ inner ear supporting cells.

75. The method of any one of claims 66-74, wherein the inner ear hair cells are Atoh1+ inner ear hair cells.

76. The method of any one of claims 66-75, wherein the subject has balance loss.

77. The method of any one of claims 66-75, wherein the subject has hearing loss.

78. The method of claim 77, wherein the hearing loss is sensorineural hearing loss.

79. The method of claim 77 or 78, wherein the hearing loss is the result of a genetic or congenital defect, trauma, aging, or chemical-induced ototoxicity.

80. The method of any one of claims 66-79, wherein the subject is a human.

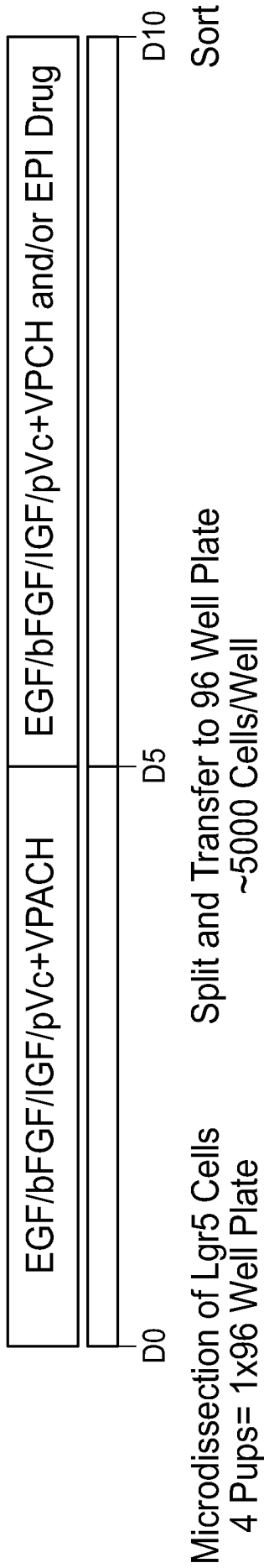


FIG. 1

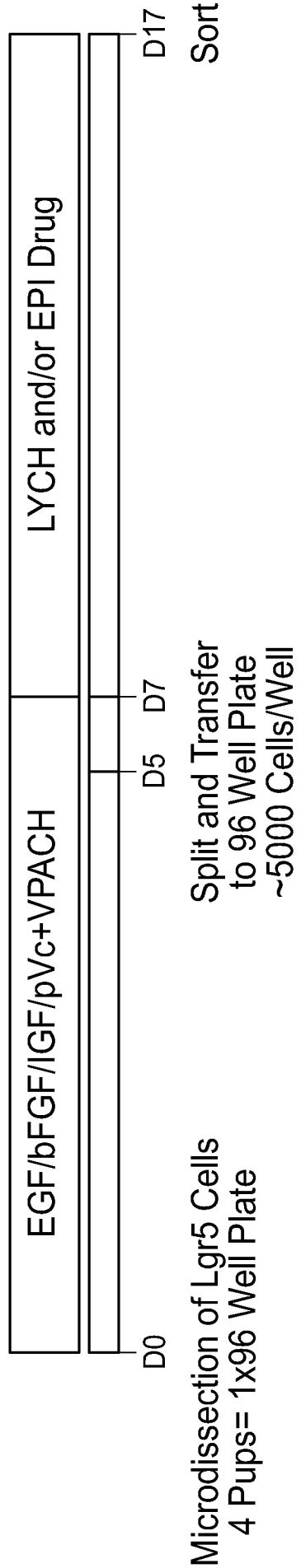


FIG. 2

Lgr5+Proliferation Pilot Screen

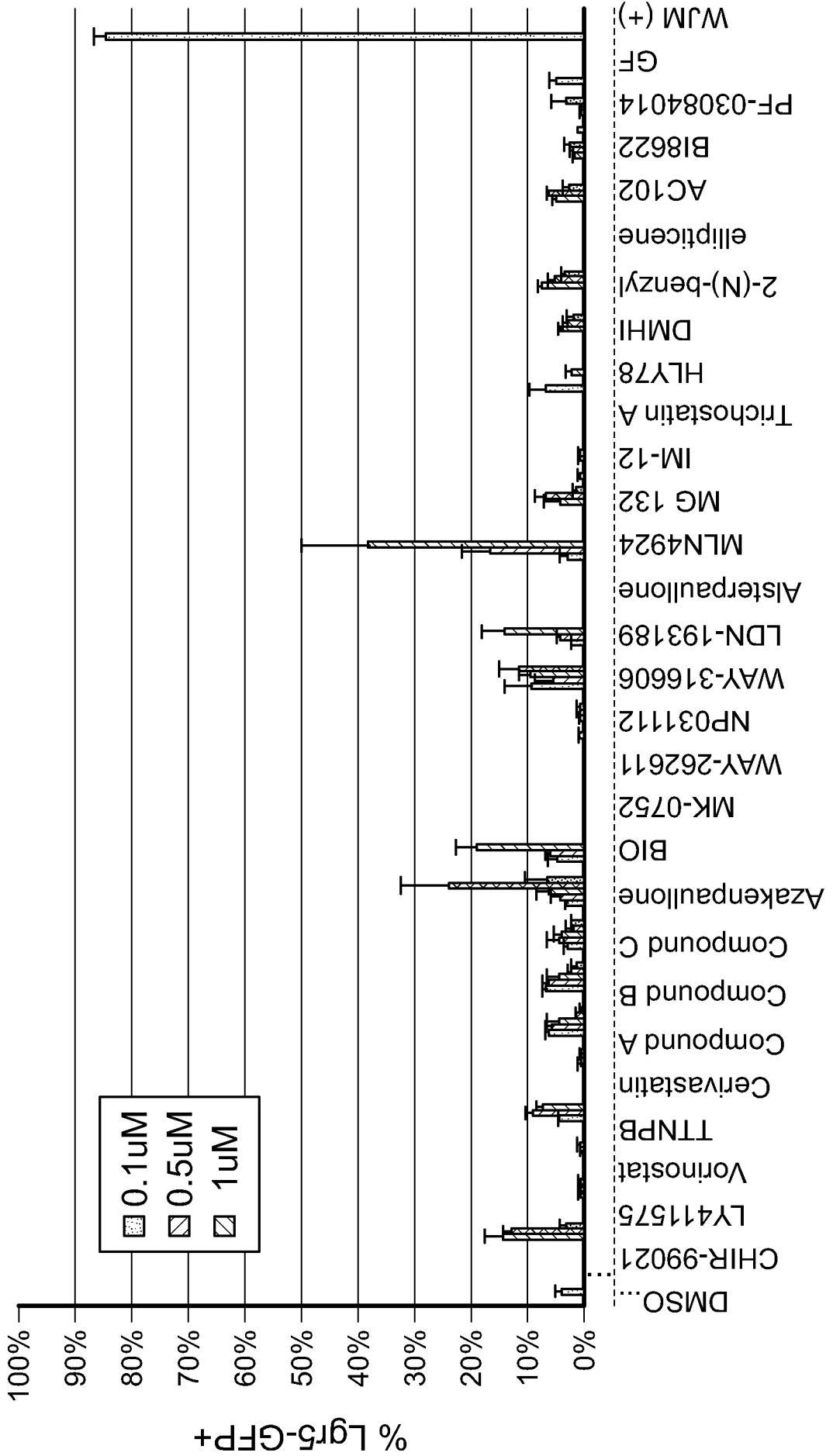


FIG. 3

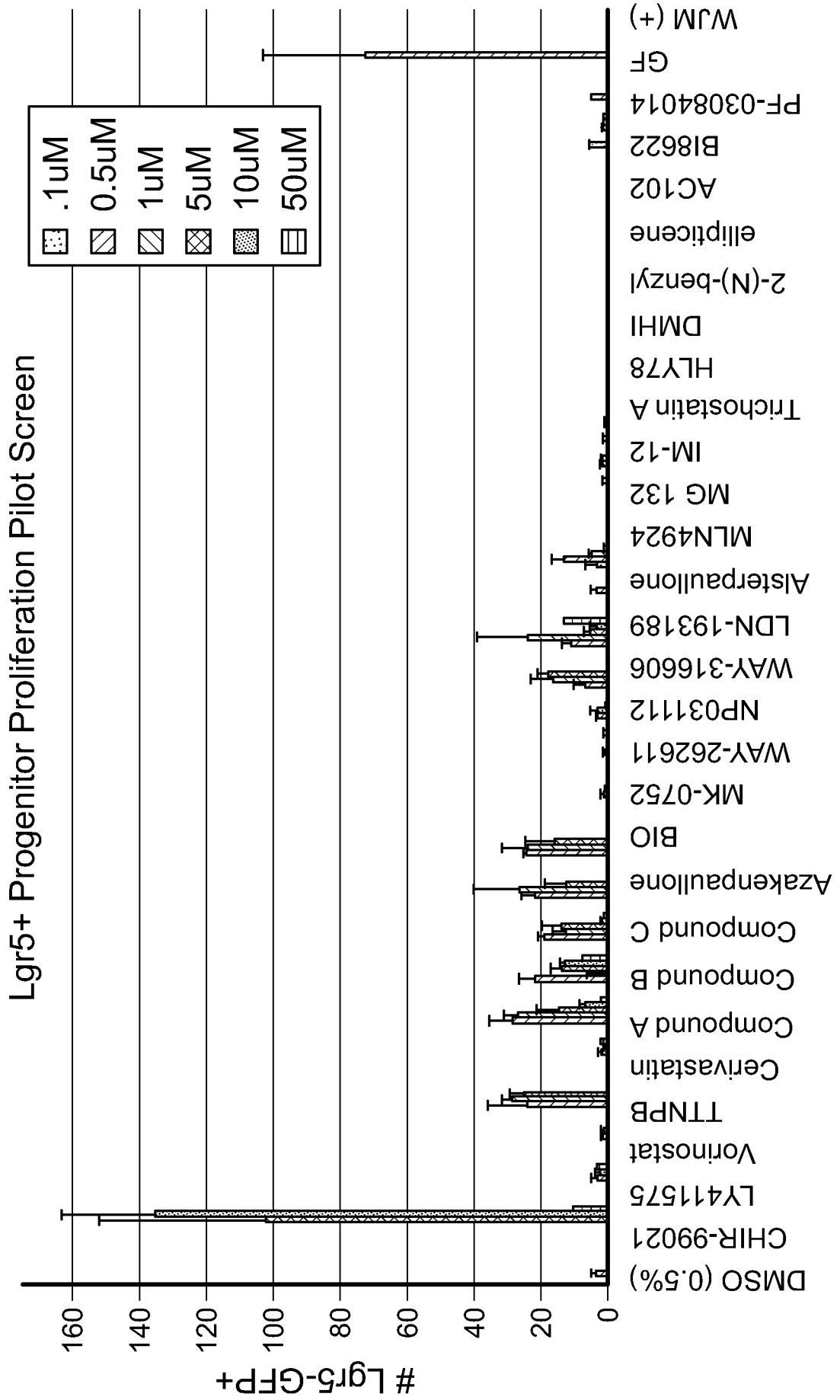


FIG. 4

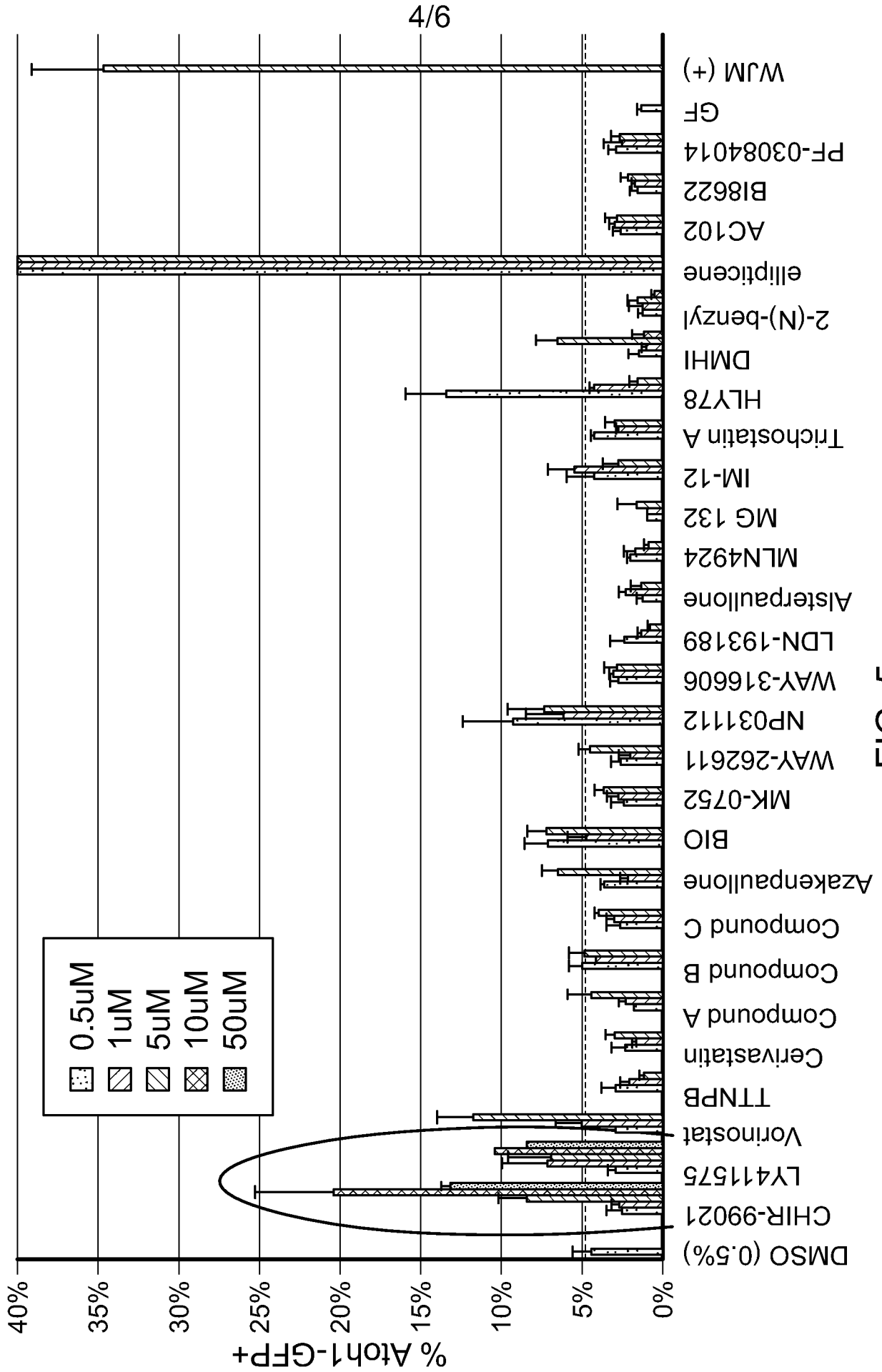


FIG. 5

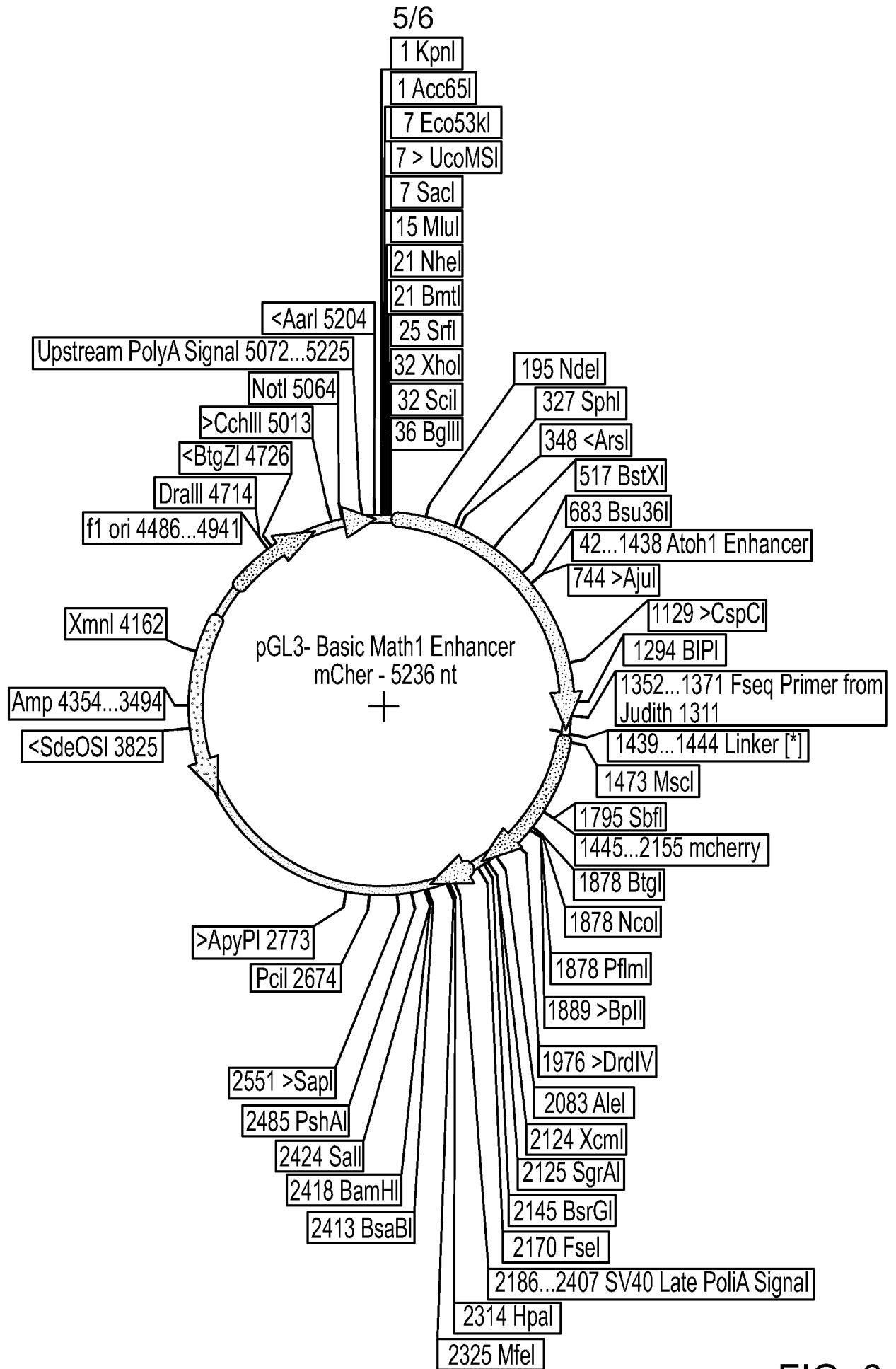
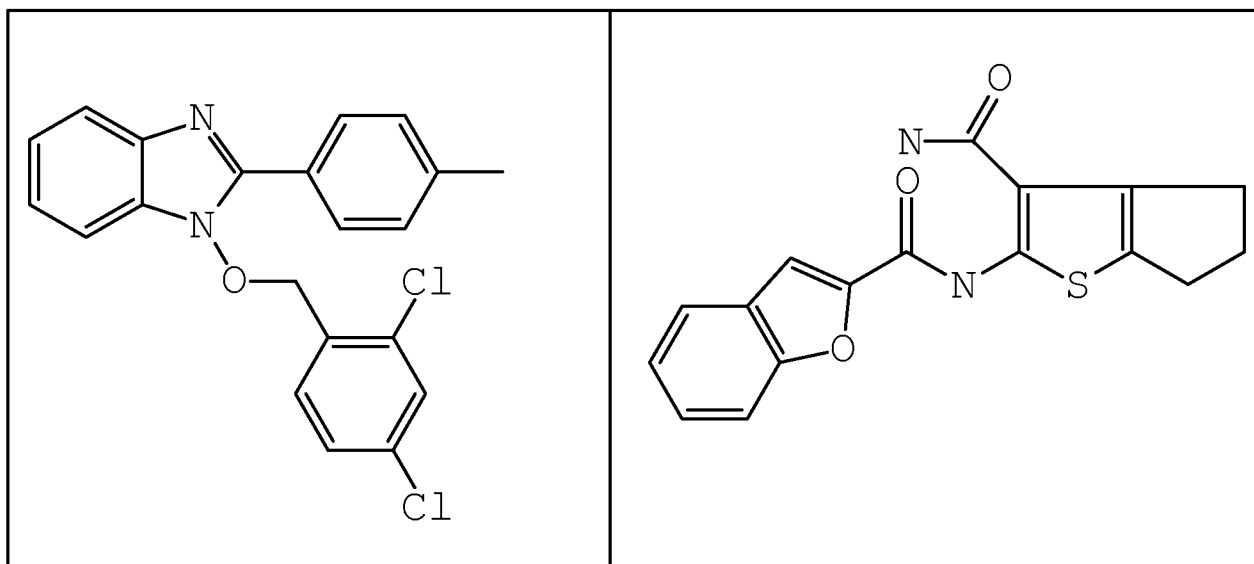
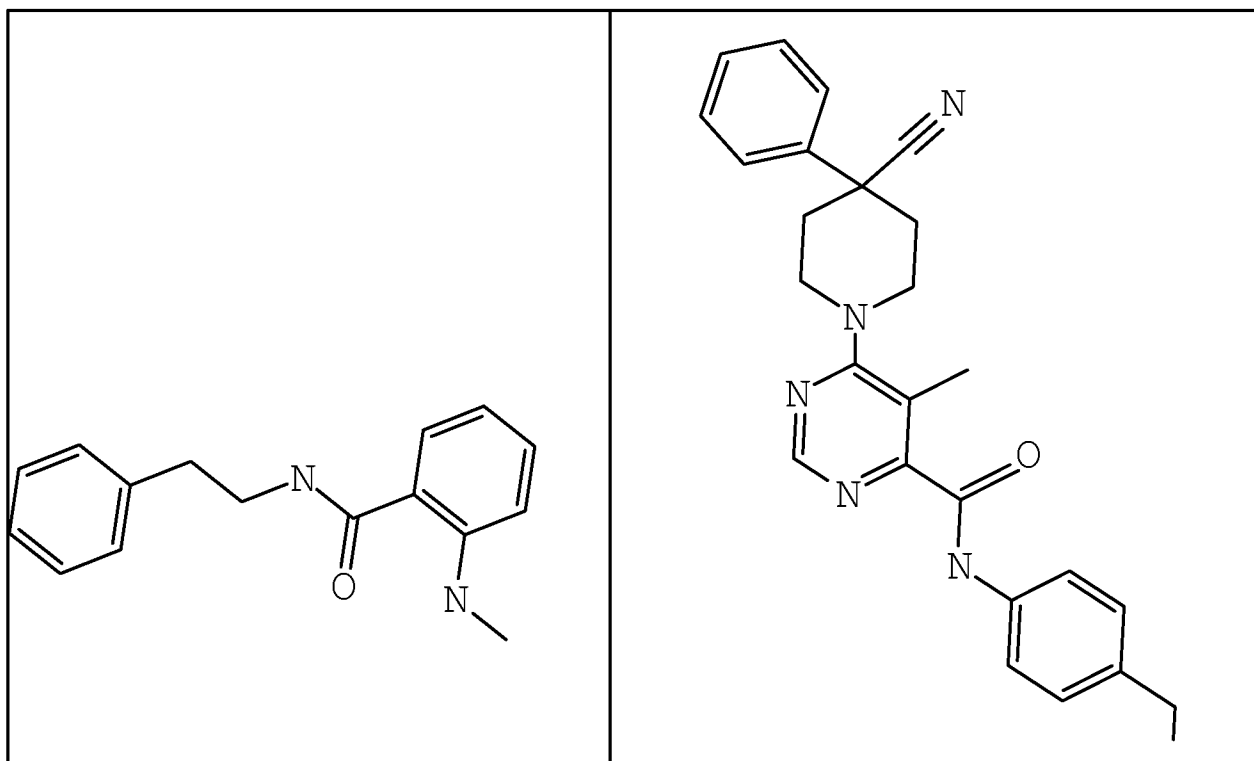


FIG. 6



Compound A

Compound B



Compound C

BI8622

FIG. 7

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/015379

A. CLASSIFICATION OF SUBJECT MATTER
 IPC(8) - A61K 35/12; A61P 27/16; C12N 5/071 (2017.01)
 CPC - A61K 9/0019; A61K 9/0046; C12N 15/8209 (2017.02)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 See Search History document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
 USPC - 424/93.21; 435/325; 800/14 (keyword delimited)

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
 See Search History document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2015/168149 A2 (MASSACHUSETTS EYE & EAR INFIRMARY) 05 November 2015 (05.11.2015) entire document	1, 4, 5, 7, 8, 66-68, 70, 71
--		-----
Y		2, 3, 6, 9
X	WO 2014/145205 A2 (ST. JUDE CHILDREN'S RESEARCH HOSPITAL) 18 September 2014 (18.09.2014) entire document	27-30, 35-38
X	US 2009/0232780 A1 (EDGE et al) 17 September 2009 (17.09.2009) entire document	57, 58
X	ROCCIO et al. "Cell cycle reactivation of cochlear progenitor cells in neonatal FUCCI mice by a GSK3 small molecule inhibitor," Sci. Rep. 08 December 2015 (08.12.2015), Vol. 5, Pgs. 1-11. entire document	56
Y	US 2015/0209406 A1 (MASSACHUSETTS EYE & EAR INFIRMARY) 30 July 2015 (30.07.2015) entire document	2, 3, 9
Y	LIN et al. "Hair cell damage recruited Lgr5-expressing cells are hair cell progenitors in neonatal mouse utricle," Front. Cell. Neurosci. 01 April 2015 (01.04.2015), Vol. 9:113, Pgs. 1-11. entire document	6

Further documents are listed in the continuation of Box C.

See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

25 April 2017

Date of mailing of the international search report

31 MAY 2017

Name and mailing address of the ISA/US

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Blaine R. Copenheaver

PCT Helpdesk: 571-272-4300
 PCT OSP: 571-272-7774

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/015379

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:

a. forming part of the international application as filed:

in the form of an Annex C/ST.25 text file.

on paper or in the form of an image file.

b. furnished together with the international application under PCT Rule 13ter.1(a) for the purposes of international search only in the form of an Annex C/ST.25 text file.

c. furnished subsequent to the international filing date for the purposes of international search only:

in the form of an Annex C/ST.25 text file (Rule 13ter.1(a)).

on paper or in the form of an image file (Rule 13ter.1(b) and Administrative Instructions, Section 713).

2. In addition, in the case that more than one version or copy of a sequence listing has been filed or furnished, the required statements that the information in the subsequent or additional copies is identical to that forming part of the application as filed or does not go beyond the application as filed, as appropriate, were furnished.

3. Additional comments:

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2017/015379

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. Claims Nos.: 10-26, 31-34, 39-55, 59-65, 69, 72-80
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- No protest accompanied the payment of additional search fees.